1 Ecological diversification in an adaptive radiation of plants: the role of de novo mutation and
2 introgression
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Abstract
Adaptive radiations are characterized by rapid ecological diversification and speciation events,
leading to fuzzy species boundaries between ecologically differentiated species. Adaptive
radiations are therefore key systems for understanding how species are formed and maintained,
including the role of de novo mutations vs. pre-existing variation in ecological adaptation and the
genome-wide consequences of hybridization events. For example, adaptive introgression, where
beneficial alleles are transferred between lineages through hybridization, may fuel diversification
in adaptive radiations and facilitate adaptation to new environments. In this study, we employed
whole-genome resequencing data to investigate the evolutionary origin of hummingbird-
pollinated flowers and to characterize genome-wide patterns of phylogenetic discordance and
introgression in *Penstemon* subgenus *Dasanthera*, a small and diverse adaptive radiation of
plants. We found that magenta hummingbird-adapted flowers have apparently evolved twice
from ancestral blue-violet bee-pollinated flowers within this radiation. These shifts in flower
color are accompanied by a variety of inactivating mutations to a key anthocyanin pathway
enzyme, suggesting that independent de novo loss-of-function mutations underlie parallel
evolution of this trait. Although patterns of introgression and phylogenetic discordance were
heterogenous across the genome, a strong effect of gene density suggests that, in general, natural
selection opposes introgression and maintains genetic differentiation in gene-rich genomic
regions. Our results highlight the importance of both de novo mutation and introgression as
sources of evolutionary change and indicate a role for de novo mutation in driving parallel
evolution in adaptive radiations.

Introduction

Adaptive radiation – the proliferation of ecological roles and adaptations in different species within a lineage – is an important process considered to play a central role in the generation of biodiversity on Earth (Simpson 1953; Schluter 2000; Givnish 2015). This phenomenon, which often coincides with an increase in the rate of lineage diversification (and is thus termed a "rapid" or "explosive" adaptive radiation) encompasses a number of well-known and charismatic taxonomic groups, including Darwin's finches, *Anolis* lizards, and cichlid fishes. Sustained interest in adaptive radiations from evolutionary biologists has, over time, revealed several genomic features characteristic of rapidly radiating clades. Principal among these is an abundance of phylogenetic discordance which, in depicting species' complicated evolutionary histories, may more closely resemble a web than a bifurcating tree. This discordance may be chiefly caused by incomplete lineage sorting (ILS), which is expected to be abundant – potentially overwhelmingly so – when speciation occurs rapidly (Pamilo and Nei 1988; Maddison 1997; Rosenberg 2002; Whitfield and Lockhart 2007). Rapid adaptive radiations are also commonly characterized by introgression between closely related species that have not had much time to develop barriers to reproduction. Given that introgressed genetic material may confer fitness effects and can be subject to strong selection, particularly in functionally important genomic regions (Moran et al. 2021), signatures of introgression are expected to be distributed heterogeneously across the genome (Harrison and Larson 2016; Martin and Jiggins 2017). Introgression may provide a source of genetic diversity in adaptive radiations (Edelman and Mallet 2021), and although its importance in this context has historically been somewhat controversial (Seehausen 2004), many recent efforts have highlighted the myriad ways in which introgression may effectively broaden the genomic substrate for adaptation through novel combinations of haplotypes (e.g., McGee et al. 2020; Ferreira et al. 2021; Ronco et al. 2021; De-Kayne et al. 2022; Suvorov et al. 2022; Cicconardi et al. 2023; Marcionetti and Salamin 2023).

In addition to widespread ILS and introgression, radiations also often feature patterns of repeated evolution, which may be either the result of the adaptive introgression of beneficial alleles or independent (convergent) adaptation in disparate lineages to similar ecological conditions. While adaptive divergence progresses in adaptive radiations through complex interplays of standing genetic variation, introgression, and de novo mutation, it is standing or pre-existing variation (often introduced through introgression) that is often implicated as the most important source of genetic variation underlying repeated or parallel evolution (Berner and Salzburger 2015; Haenel et al. 2019; Marques et al. 2019; Marques et al. 2022). Yet, it is clear that the repeated generation of similar phenotypes in ecologically similar habitats – a hallmark of adaptive radiation – is also driven by de novo mutation. For example, both processes appear to be important in the radiation of hares (genus *Lepus*), in which the repeated evolution of nonwhite winter coats has proceeded through both adaptive introgression (Jones et al. 2018; Giska et al. 2019) and independent loss-of-function (LOF) mutations at the *Agouti* pigmentation gene (Jones et al. 2020). Whether adaptation more often involves de novo vs. standing variation informs on whether trait evolution is constrained by mutation events or by the opportunity for hybridization between differentiated lineages (Marques et al. 2019). Empirical data from diverse lineages across the tree of life are needed to identify common genetic features of adaptive radiations.

The genus *Penstemon* is an adaptive radiation of North American angiosperms which exhibits an astonishing array of phenotypic and ecological diversity across its nearly 300 described species.

Many *Penstemon* species are interfertile and form hybrids in nature, fueling speculation that
interspecific hybridization has played a major role in the diversification of the group (Straw
1955; Wolfe et al. 2006; Wessinger et al. 2016). *Penstemon* is also a recent and rapid radiation,
estimated to have originated 1.419-6.416 MYA (Wolfe et al. 2021), and this rapid speciation has
been highlighted as a potential major source of genealogical discordance (due to ILS) in addition
to discordance caused by interspecific hybridization (Wolfe et al. 2006; Wessinger et al. 2016).
Within *Penstemon*, the subgenus *Dasanthera* (hereafter, "*Dasanthera*") includes nine
ecologically diverse species and represents an adaptive radiation in microcosm. This group
includes several species which hybridize in nature, making it an attractive system in which to
study the importance of introgression and de novo mutation on adaptive divergence. A primary
axis of variation in *Dasanthera* is floral pollination syndrome, wherein two hummingbird-
adapted species (*P. newberryi* and *P. rupicola*) have evolved from the ancestral state of
hymenopteran-adapted flowers (Datwyler and Wolfe 2004; Wilson et al. 2004; Kimball 2008).
This mirrors a genus-wide trend involving the repeated evolution of specialist hummingbird-
adapted flowers from more general hymenopteran-adapted progenitors (Wilson et al. 2007). The
hummingbird-adapted species *P. newberryi* and *P. rupicola* each form hybrid zones with the
hymenopteran-adapted, high-elevation *P. davidsonii*; in each case the hummingbird-adapted
species occupies a lower elevation zone, and hybrids are formed and persist at intermediate
elevations in the Cascades and Sierra Nevada Mountains where the parent species' distributions
overlap. Previous studies indicate that *P. newberryi* and *P. rupicola* are not sister species,
suggesting parallel divergence in elevational tolerance and pollinator adaptation within
Dasanthera (Stone and Wolfe 2021; Stone et al. 2023). An alternative scenario is that
hummingbird pollination syndrome in *Dasanthera* could have a single evolutionary origin shared
between species through introgression. Such a scenario has occurred in other plant groups; for
example, the introgression of a *MaMyb2* allele has led to repeated origins of red-flowered
phenotypes in the *Mimulus aurantiacus* species complex (Stankowski and Streisfeld 2015; Short
and Streisfeld 2023).
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Floral syndrome divergence in *Dasanthera* involves changes in floral morphology
(hummingbird-pollinated: long, narrow flowers with exserted stamens and styles; hymenopteran-
pollinated: short, wide flowers with inserted stamens and styles), and perhaps most strikingly, the
evolution of bright magenta flowers in hummingbird-pollinated species from ancestral
hymenopteran-pollinated blue-violet flowers. Whereas little is currently known about the
underlying genetic mechanisms for floral morphological shifts, the evolution of red flowers tends
to involve predictable changes to the genetic pathway for anthocyanin production. In other
groups within *Penstemon*, shifts towards redder floral hues often involve the functional
inactivation of the anthocyanin pathway enzyme Flavonoid 3′,5′-hydroxylase (F3′5′H)
(Wessinger and Rausher 2014; Wessinger and Rausher 2015). F3′5′H belongs to the functionally
diverse superfamily of cytochrome P450-dependent monooxygenases. This enzyme catalyzes
hydroxylation of the B-ring of anthocyanins, leading to the production of blue-colored
delphinidin-based pigments (Tanaka and Brugliera 2013). The degeneration of *F3′5′h* prohibits
the production of delphinidin and has been shown in *Penstemon* to result in flowers that produce
red pelargonidin-based anthocyanins; importantly, the evolution of red flowers in multiple
Penstemon species appears to be the result of repeated de novo LOF mutations to the *F3′5′h*
coding sequence (Wessinger and Rausher 2014; Wessinger and Rausher 2015).
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In this study we used whole-genome resequencing data to trace the evolutionary history and
source of genetic variation for hummingbird-pollinated flowers in *Dasanthera* and characterize
genome-wide patterns of introgression. Signatures of introgression were uneven across the
genome, and were generally lower in gene-rich regions, suggesting that even in this very recent
radiation, introgression often may be maladaptive. We found strong support for independent
evolutionary origins of magenta floral pigments in the two hummingbird-pollinated species.
These parallel shifts in floral pigment production are accompanied by distinct degenerative
mutations to the *F3'5'h* coding sequence. While introgression is prevalent within *Dasanthera*, we
found little evidence that the two hummingbird-pollinated species share a history of
introgression; this suggests that despite the importance of introgression across *Dasanthera*
generally, it is de novo mutation, and not adaptive introgression, that likely fuels parallel shifts to
hummingbird pollination in this adaptive radiation.
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Methods
Study system and species sampling
We generated whole-genome resequencing data for 18 individual plants, each from a unique
locality, representing seven of the nine total *Dasanthera* species (Table S.1). We included
multiple samples for each of the species in the Cascades/Sierra Nevada Mountain clade (*P.*
cardwellii, P. davidsonii, P. fruticosus, P. newberryi, P. rupicola), and a single individual each
of the Northern Rocky Mountain exclusive taxa *P. montanus* and *P. lyallii* for use as outgroups,
following results from previous phylogenetic inquiries into the group (Stone and Wolfe 2021;
Stone et al. 2023). We extracted DNA from silica-dried leaf tissues using a modified CTAB
protocol and performed whole-genome Illumina library preparations and sequencing to ~10x
depth using 150-bp paired-end reads generated on the NovaSeq (Illumina Inc.) platform at Duke
University. Raw Illumina reads were quality-filtered with fastp (Chen et al. 2018), enabling auto-
detection of adapters, limiting read length to 30 bp, filtering out unpaired reads, enabling base
correction for overlapping reads, and enabling poly-x trimming on 3' ends of reads. Final
sequence quality was verified with fastqc (Andrews 2010) and multiqc (Ewels et al. 2016).
Quality filtered reads were mapped to the annotated *P. davidsonii* reference genome (Ostevik et
al. 2023) with bwa-mem (Li 2013), and reads with low mapping quality (q<20) were removed.
Duplicated reads were marked and removed with samtools markdup (Li et al. 2009), and
overlapping paired end reads were clipped with bamutil clipOverlap (Jun et al. 2015).
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Filtered reads were then used to call genotypes to produce an "all sites" .vcf file using bcftools
v1.15.1 (Li 2011). Variant sites were soft-filtered for minimum genotype-call quality (20),
minimum and maximum allelic depths, which were chosen based on genome-wide averages of
coverage depth (3 and 60, respectively), minimum individual allelic depth (2), and missingness
(50%). Sites failing the minimum genotype-call quality filter were changed to match the
reference allele (changed to invariant sites); otherwise, filters produced missing data for sites or
individual genotypes. The resulting filtered all sites .vcf was then used to generate consensus
whole-genome sequences and individual gene coding sequences (CDS).
166
Species relationships
We inferred species trees in three ways to compare genomic sampling strategies and analysis
methods. First, we used custom python scripts to split whole-genome sequences into non-
overlapping 10kb windows, removing windows with more than 75% missing data (Ns) for any

sample. We then estimated "window trees" for each genomic window with IQ-TREE 2 v2.2.0.3
(Nguyen et al. 2015), enabling the ModelFinder (-m MFP) option (Kalyaanamoorthy et al. 2017)
to select the best substitution model for each window. We then used these window trees to infer a
species tree with ASTRAL-III v.5.7.8 (Zhang et al. 2018), using the full annotation option (-t 2).
Second, for each sample, we used gffread v0.12.7 (Pertea and Pertea 2020) and the *P. davidsonii*
reference genome annotations to generate fasta files with spliced exons (CDS), retaining the
single longest isoform for each transcript. We then used a custom python script to filter
individuals with >50% missing data for each CDS. Gene trees were then estimated for each CDS
and a species tree was constructed in ASTRAL in the same manner as for the window
alignments. For both of the ASTRAL species trees, we also calculated gene- and site
concordance factors (gCF, sCF; Minh et al. 2020) in IQ-TREE, using as input the window or
gene trees and sequence alignments used to generate the species tree. Third, we estimated a
concatenated ML species tree in IQ-TREE, specifying the GTR+I+R model, estimating rates for
each of the 12 largest scaffolds, and performing 1000 UltraFast Bootstrap (UFBoot) replicates
(Hoang et al. 2018).
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Floral pigment production
We determined the identity of anthocyanidin pigments produced in floral tissues, which are
responsible for floral color in *Penstemon*. We collected intact flowers from voucher specimens
that were preserved in an herbarium press at the time of collection (samples were collected in
2017 and 2018). We used an isoamyl alcohol extraction protocol outlined in Harborne (1984)
and separated pigments using TLC on Cellulose F glass plates (Millipore, Burlington, MA). We
compared extracted anthocyanidin pigments to a multi-anthocyanidin standard solution
containing pelargonidin, cyanidin, and delphinidin each in concentrations of ~0.013 mg/mL, as
described in Stevens et al. (2023).
196
Genome-wide average signatures of introgression
We investigated genome-wide signatures of introgression using Dsuite v0.5 (Malinsky et al.
2021). Specifically, we used Dsuite to calculate genome-wide D statistics (ABBA-BABA) for all
possible trios of samples and to implement and visualize the "f-branch" metric f_b (Malinsky et al.
2018) using the 10kb window ASTRAL species tree as the guide tree and *P. montanus* as the
outgroup.
203
Genomic landscape of divergence, discordance, and introgression
We examined patterns of differentiation among species and patterns of phylogenetic discordance
across the genome. We calculated average F_{ST} (Weir and Cockerham 1984) and average
nucleotide distance within (π) and between (d_{xy}) species in 10kb and 50kb non-overlapping
windows with pixy v1.2.7 (Korunes and Samuk 2021), using the filtered all sites .vcf files as
input. We used bedtools v2.27.1 (Quinlan and Hall 2010) to calculate genic fractions (the
proportion of sites that are coding sequence) per window. We identified d_{xy} outliers between two
species pairs that form hybrid zones: *P. davidsonii-P. newberryi*, and *P. davidsonii-P. rupicola*.
Highly differentiated genomic regions in the face of gene flow may be especially resistant to
introgression and could play a key role in the formation or maintenance of species boundaries.
We first filtered all sliding windows with more missing counts than count comparisons, and then
identified outliers as 10kb sliding windows with average d_{xy} values exceeding the genome-wide

mean by three standard deviations (Z-score > 3). We then identified shared outliers as sliding
windows with outlier d_{xy} values in both species pairs.

To explore how topological discordance varies across the genome, we first calculated normalized
Robinson-Foulds (RF) distances (Robinson and Foulds 1981) between each 10kb window tree
and the species tree using the R package phangorn v2.10.0 (Schliep 2011). In addition, we
examined topological discordance affecting three main internal branches of the species tree using
TWISST (Martin and Van Belleghem 2017). TWISST quantifies the proportion (or "weighting")
of different tree topologies across the genome. For this approach, we used scripts provided by the
developers (available at https://github.com/simonhmartin/twisst), using the 10kb window trees as
input. We summed topology weights for all trees consistent with the following internal branches
of the species tree: the branch leading to *P. newberryi*, *P. cardwellii*, and *P. rupicola* (termed
IB1); the branch leading to *P. newberryi* and *P. cardwellii* (IB2), and the branch leading to *P.*
davidsonii and *P. fruticosus* (IB3). In each case, *P. montanus* served as the outgroup.

Although the *D* statistic is a powerful metric to detect genome-wide patterns of introgression, it
is less useful to identify specific introgressed genomic regions because it is a poor estimator of
the amount of introgression and it has a high degree of variance when applied to genomic
windows of smaller sizes (Martin et al. 2015). For this reason, to explore signatures of
introgression along the genome, we employed the f_{dM} statistic, which is specifically designed to
estimate introgression between pairs of taxa in genomic windows along chromosomes, and, like
many statistics related to the *f4*-ratio, performs better than the *D* statistic (exhibits less variance
and is a better estimator of the amount of introgression) when applied to small genomic windows
(Malinsky et al. 2015; Malinsky et al. 2021). We estimated f_{dM} in 10,000 SNP windows, sliding
every 2,500 SNPs. For each of these f_{dM} windows, we also calculated the genic fraction to assess
the relationship between genic fraction and f_{dM}. We conducted these calculations for all triplets
of taxa consistent with the species tree.

Introgression of F3'5'h
We examined several lines of evidence to assess whether *F3'5'h* – one of the key genetic
components underlying transitions to hummingbird pollination syndrome – could be shared
between red-flowered *Dasanthera* through introgression. First, we calculated f_{dM} in sliding
windows along the genome to investigate signatures of introgression between *P. newberryi* (P2)
and *P. rupicola* (P3), setting *P. cardwellii* as P1. Here we estimated f_{dM} at four different window
sizes: (1) in 10,000 SNP windows, sliding every 2,500 SNPs, (2) in 5,000 SNP windows, sliding
every 1,250 SNPs, (3) in 1,000 SNP windows, sliding every 250 SNPs, and (4) in 500 SNP
windows, sliding every 125 SNPs. If *F3'5'h* is introgressed, values of f_{dM} should be elevated
compared to both genome-wide estimates and the local genomic region.

Because *F3'5'h* is potentially pseudogenized in our magenta-flowered samples (leading to
relaxed selection against indels), we re-called variants for reads mapping to this genomic region
while additionally calling and normalizing indels with bcftools. We performed downstream
variant calling and filtering as described for genome-wide variant calling steps. We then aligned
the CDS for each sample with muscle v.5.1 (Edgar 2004) and re-estimated the gene tree for
F3'5'h in IQ-TREE, performing 1000 UFBoot replicates and 1000 bootstrap replicates for the
SH-like approximate likelihood ratio test (SH-aLRT) to assess branch support (Guindon et al.

2010). If *F3′5′h* is introgressed, this gene tree should be discordant with the species tree and
place *P. newberryi* and *P. rupicola* as sister.
264
We examined *F3′5′h* nucleotide and amino acid sequence alignments to identify the positions of
potential indels and nonsynonymous mutations. *F3′5′h* coding sequences in other hummingbird-
adapted *Penstemon* species exhibit a variety of functional inactivating mutations, including
premature stop codon mutations, frameshift mutations, and large deletions (Wessinger and
Rausher 2015). We searched for similar LOF mutations present in our magenta-flowered
samples. Likewise, mutations in any of the highly conserved domains, which included the heme-
binding domain and six substrate recognition sites (SRSs) putatively involved in substrate
contact (Gotoh 1992), would likely signal a dysfunctional enzyme. If *F3′5′h* is introgressed, our
magenta-flowered samples should share the same loss-of-function mutation(s), be they through
structural changes (e.g., indels) or through mutations in conserved domains or SRSs.
275
276   **Results**
277   *Whole genome data confirm species relationships and suggest two separate origins of*
278   *hummingbird pollination in Dasanthera*
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280   Our whole genome sequencing of 18 *Dasanthera* samples produced ~13x average coverage per
281   sample across approximately 70% of the eight main scaffolds (assumed to represent the eight
282   *Penstemon* chromosomes). All information pertaining to read quantities, genome-wide coverage,
283   and per-scaffold coverage can be found in Table S.2.
284
All three species tree inference methods produced very similar topologies and support the same
main relationships among species within the Cascades and Sierra Nevada Mountains clade.
These results corroborate previous inferences made with restriction enzyme-based approaches
(Stone and Wolfe 2021; Stone et al. 2023). While the 10kb window ASTRAL-based species tree
and the concatenated ML tree produced identical topologies, the CDS ASTRAL-based tree
differed in the placement of one branch, placing *P. rupicola* from Cle Elum Lake in central
Washington as sister to the remainder of the *P. rupicola* clade, instead of the sample from Castle
Lake in northern California (Figure S.1). From this point forward we refer to the 10kb window
ASTRAL tree as the species tree (Figure 1). Despite agreement among alternative species tree
topologies and high bootstrap values, quartet support and concordance factors (gCFs and SCFs)
were relatively low (Figure 1). This suggests conflicting phylogenetic signal among window
trees (due to ILS and/or introgression) at many branches, especially for relationships deeper in
the tree. While the overall level of discordance was relatively high, the most strongly supported
internal branch (with the least discordance) is the branch placing *P. newberryi* and *P. cardwellii*
as sister (Figure 1, IB2: quartet support = 50%, gCF = 18.6%, sCF = 58.8%), suggesting strongly
that the two hummingbird-pollinated species (*P. newberryi* and *P. rupicola*) are not sister
species.
302
Anthocyanin pigment data for the ten available samples confirm that the magenta flowers
associated with bird pollination in *P. newberryi* and *P. rupicola* contain pink cyanidin-based
anthocyanin pigments only, whereas bee-adapted species produce blue-delphinidin-based
pigments, sometimes in combination with cyanidin (results summarized in Table S.3.). The only
exception is the *P. newberryi* var. *berryi* individual that produced both cyanidin and delphinidin.

Flowers of this taxon are not nearly as apparently specialized to bird pollination: the anthers and
stigma are inserted within the flower, and the corolla tube may appear blue-violet in color, rather
than magenta. Overall, these results align with a previous study reporting a single *P. newberryi*
sample produced cyanidin and a single *P. montanus* sample produced both cyanidin and
delphinidin (Scogin and Freeman 1987).
313
Because the production of cyanidin is biochemically simpler than the production of delphinidin,
P. rupicola and *P. newberryi* (with the exception of the var. *berryi* sample) have experienced a
loss of delphinidin production, relative to the bee-pollinated species. Taking the species tree at
face value, these results would suggest one of the following scenarios: (1) parallel losses of the
ability to produce delphinidin (resulting in magenta flowers) in *P. newberryi* and *P. rupicola*, (2)
a single loss of delphinidin production along the branch leading to *P. newberryi*, *P. cardwellii*,
and *P. rupicola*, followed by at least one reversal in *P. cardwellii* (and additionally introgression
into *P. newberryi* var. *berryi*, or a second reversal in this taxon), or (3) a single loss of
delphinidin production, which is shared by *P. newberryi* and *P. rupicola* through either
hemiplasy (ILS) or introgression.
324
Coding sequence variation at F3'5'h suggests separate LOF de novo mutations are responsible
for the repeated evolution of magenta flowers
Nucleotide sequence alignments of the *F3'5'h* gene sequence revealed that a diversity of LOF
mutations to the coding sequence accompany loss of delphinidin production in *P. newberryii* and
P. rupicola. We identified two distinct large deletions (~500 bp) present in two *P. newberryi*
samples from Eldorado and Plumas National Forests but absent in all other samples of *P.*
newberryi and *P. rupicola* (Figure 2c). These deletions either partially or entirely remove the
second exon, undoubtedly abolishing the activity of F3′5′H (Seitz et al. 2007). A third *P.*
newberryi sample from Castle Lake shares a 4-bp insertion with the sample from Plumas which
produces a premature stop codon (Figure 2c; Figure S.2). All three of these samples only
produce cyanidin in their floral tissues (Table S.3). The *F3'5'h* sequence for *P. newberryi* var.
berryi lacks inactivating mutations, consistent with observed delphinidin production in its floral
sample. Two *P. rupicola* samples from Mt. Rainier and Rattlesnake Ridge shared a 4-bp
insertion that results in premature stop codons. We were unable to obtain pigment data for the *P.*
rupicola sample from Rattlesnake Ridge, but the sample from Mt. Rainier produces only
cyanidin in floral tissues (Table S.3). The remaining two *P. rupicola* samples are less clear. One
of these samples (from Cle Elum Lake) produces only cyanidin in flowers and is heterozygous
for sites that cause amino acid substitutions in SRS1 and the hydroxylation activity site (Figure
S.2) – these replacement mutations could reduce functionality of F3′5′H. The *P. rupicola* sample
from Castle Lake has no suggestive functional mutations in *F3'5'h*, but we were unable to extract
anthocyanidins due to poor tissue quality.
346
Values of f_{dM} assessing introgression between *P. newberryi* and *P. rupicola* surrounding the
F3'5'h locus were near zero and were not significantly elevated above the scaffold-wide mean
(one-tailed t-test: *p*-value = 0.18), suggesting *F3'5'h* is unlikely to have been shared between the
two species through introgression (Figure 2b). This pattern is apparent regardless of the genomic
window size used (Figure S.3). The maximum likelihood *F3'5'h* gene tree was discordant from
the species tree, but not in a manner consistent with introgression of between *P. newberryi* and
P. rupicola (Figure 2a). However, we note that many internal branches of the tree were quite

poorly supported, likely due to low information content. In summary, we identified several distinct LOF mutations in *F3′5′h*. Because LOF mutations are difficult to reverse through gene repair, we find a scenario involving a single loss of delphinidin followed by reversals through gene repair (in *P. cardwellii* and *P. newberryi* var. *berryi*) to be highly unlikely. Further, we find no evidence for a history of introgression at the *F3′5′h* locus. Our data are consistent with parallel, independent LOF mutations as the mechanism behind the repeated evolution of magenta flowers in *Dasanthera*.

Introgression is prevalent in Dasanthera, but does not suggest a history of gene flow between hummingbird-pollinated species

Our analysis of genome-wide introgression found that many taxa exhibit a history of introgression and admixture, consistent with previous inquiries into the group (Stone and Wolfe 2021; Stone et al. 2023). 64% of ABBA-BABA tests for all trios of samples in our dataset were significantly different from zero (after a Bonferroni correction at $\alpha = 0.05$), providing strong support for widespread gene flow between species (Table S.4). The f-branch statistics corroborated these results and identified individual samples with elevated levels of introgression with another taxon, suggesting population-specific admixture that in some cases may be recent (Figure 3). Included among these results are (1) the *P. rupicola* sample from Cle Elum Lake, with introgression from *P. fruticosus*, (2) the *P. rupicola* sample from Castle Lake, with introgression from *P. newberryi*, and (3) the *P. davidsonii* sample from Crater Lake, with introgression from the clade containing *P. newberryi*, *P. cardwellii*, and *P. rupicola* (Figure 3). In each of these examples, the taxa in question are geographically close enough to exchange genes. As such, these may represent lineages that have experienced relatively recent bouts of introgression. Additionally, we identified significant signatures of introgression between *P. davidsonii* and both *P. newberryi* and *P. rupicola*, a result consistent with the presence of longstanding hybrid zones between these species.

While our *F3′5′h* sequence analyses suggest independent parallel evolution of magenta flowers in the two hummingbird-adapted species, the repeated evolution of other floral traits that make up the hummingbird syndrome (e.g., narrow floral tubes, increased nectar, elongated reproductive organs) might have evolved through adaptive introgression. In this scenario, we might expect to see a prevailing genomic signal of admixture between *P. newberryi* and *P. rupicola*. Contrary to this, our f-branch results find only limited support for introgression between *P. newberryi* and *P. rupicola*. Introgression between these species appears to be limited to samples collected together at the same site near Castle Lake in the Trinity Mountains of northern California. This region represents the northernmost and southernmost extent of the distributions of *P. newberryi* and *P. rupicola* respectively. Genome-wide estimates of the D statistic suggest a slightly positive but statistically insignificant signature of introgression between *P. newberryi* and *P. rupicola* (D = 0.003, p = 0.409; Table S.5). Moreover, f_{dM} estimated in windows across the genome fail to identify any clear signatures of introgression between these species (Figure 2d). After removing the two samples from Castle Lake, genome-wide estimates of the D statistic suggest a significant but weak signature of introgression between *P. rupicola* and *P. cardwellii* (D = -0.037, p = $2.30e^{-16}$).

Genome-wide patterns of genomic variation indicate selection against introgression

Genome-wide patterns of genetic divergence and discordance in *Dasanthera* are strongly shaped
by local gene density (Figures S.4-S.7). We found that both genetic distance (d_{xy}) and
differentiation (F_{ST}) between species are significantly positively associated with gene density
(Table S.6), a pattern that is evident in both 10kb and 50kb windows (Figure S.8). Conversely,
normalized Robinson-Foulds distance (a measure of topological discordance) of trees calculated
from genomic windows relative to the species tree was negatively related to gene density, such
that RF distance decreases (window trees become more similar to the species tree) as genic
fraction increases (Figure 4). The topology weights for the main internal branches of the species
tree were higher in genomic regions with a larger proportion of coding sequence (Figure 4),
suggesting a higher fraction of taxa have data conforming to the species tree in gene dense
regions. Moreover, topology weights for the internal branch supporting *P. newberryi* and *P.*
cardwellii as sister taxa (IB2) were consistently higher across the genome compared to the
topology weights for the other two internal branches. This pattern is consistent with metrics of
branch support (quartet weights and concordance factors) that find stronger support overall for
IB2 than for either of the other internal branches (Figure 4).
414
In general, we found that introgression is lower in more gene-dense regions of the genome
(Figure S.9), consistent with selection against introgression. For five of the eight possible
interspecific comparisons, there was a significant negative relationship between f_{dM} and genic
fraction. Curiously, this negative relationship was absent for all three comparisons including *P.*
fruticosus: two comparisons showed no relationship, and one comparison showed a significantly
positive relationship. Two species pairs are known to form hybrid zones in secondary contact: *P.*
davidsonii-P. newberryi and *P. davidsonii-P. rupicola*. We found strong and significant negative
relationships between f_{dM} and genic fraction in both cases. This signal was stronger in *P.*
davidsonii-P. newberryi (regression coefficient = -0.119; R^2 = 0.115) than in *P. davidsonii-P.*
rupicola (regression coefficient = -0.065; R^2 = 0.054) and in both cases is consistent with
selection against introgression. We additionally searched for genomic windows (10kb sliding
intervals) exhibiting exceptionally high levels of genetic differentiation. We found a total of 70
outlier d_{xy} windows that were present in both species pairs (shared outlier windows). These
shared outlier windows were distributed across each of the eight largest scaffolds in the *P.*
davidsonii reference genome (Figure S.10). Given their high degree of divergence in the face of
hybridization, these regions may be particularly resistant to gene flow and may represent
genomic loci underlying reproductive barriers in both hybrid zones.
432
Discussion
Independent loss-of-function mutations contribute to repeated adaptation in an adaptive
radiation
Shifts from blue-violet flowers to magenta flowers in two *Dasanthera* species are most likely the
result of independent LOF mutations to a gene in the anthocyanin biosynthesis pathway, *F3'5'h*.
First, this gene is required for the production of delphinidin-based pigments and non-functional
alleles are present in the magenta-flowered species that lack delphinidin pigments. This result
strongly suggests shifts to magenta flowers involve LOF alleles at this gene. Second, distinct
LOF alleles are found in *P. newberryi* and *P. rupicola*. Finally, we found no signal of
introgression between the two magenta-flowered species near the *F3'5'h* gene, based on the
genealogy and values of f_{dM} surrounding the gene. These findings highlight the role of de novo
mutation in the repeated evolution of adaptive phenotypes in an adaptive radiation of plants,

which is in contrast to a growing body of research suggesting that parallel evolution in adaptive
radiations is often derived from the adaptive introgression of pre-existing genetic variation
(Colosimo et al. 2005; Schluter and Conte 2009; Berner and Salzburger 2015; Marques et al.
2019). It is worth considering that the type of mutation – in this case, one conferring loss-of-
function – may strongly influence the frequency of de novo mutation in driving parallel
phenotypic evolution. LOF mutations are an abundant source of variation in natural populations
and may act as hotspots for repeated evolutionary transitions (Martin and Orgogozo 2013; Xu
and Guo 2020). For example, LOF mutations are particularly prevalent in the evolution of plant-
pollinator interactions; the loss of scent compounds, associated with transitions to hummingbird
pollination and self-pollination, has repeatedly and independently evolved in several plant
groups, including in *Petunia* (Segatto et al. 2014; Amrad et al. 2016), *Capsella* (Sas et al. 2016),
and monkeyflowers (Peng et al. 2017; Liang et al. 2023). While there are many examples of LOF
mutations conferring adaptive benefit, their occurrence is often limited to genes with few
pleiotropic effects, or to regulatory elements that affect gene expression only in specific tissues
(Xu and Guo 2020). Although transcription factors in the flavonoid pigment pathway have been
shown to generally have higher rates of molecular evolution than the structural genes (like
F3′5′h) they regulate (Wheeler et al. 2022), LOF mutations to *F3′5′h* seem to incur minimal
pleiotropic effects, perhaps explaining their apparent preponderance. *F3′5′h* functions to produce
tri-hydroxylated flavonoid compounds including delphinidin. Yet tri-hydroxylated flavonoids
seem to be restricted to floral tissue – many species that have delphinidin-producing flowers
produce di-hydroxylated flavonoids (products of the upstream gene *F3′h*) in vegetative tissues
(Wessinger and Rausher 2012). Furthermore, several plant species have evolved red floral
pigments through LOF mutations in *F3′5′h* (Smith and Rausher 2011; Ishiguro et al. 2012)
including other species of *Penstemon* (Wessinger and Rausher 2015), and many other
angiosperm taxa lack this gene entirely (Wessinger and Rausher 2012), suggesting the
defunctionalization of *F3′5′h* can underlie adaptation without deleterious side-effects. While
transitions to magenta flowers in *P. newberryi* and *P. rupicola* appear to involve parallel de novo
mutations, this does not preclude a role for adaptive introgression fueling other aspects of the
hummingbird floral syndrome. However, we found no clear evidence of gene flow between *P.*
newberryi and *P. rupicola* genome-wide or near the *F3′5′h* locus. While these results do not rule
out the potential for adaptive introgression of other floral traits associated with hummingbird
pollination syndrome (e.g., exserted stigma and anthers), at this time we have no evidence to
support this hypothesis.
478
It is also worth considering that the degree of color shift we observe in *Dasanthera* is not as
severe as in other *Penstemon* taxa that have transitioned to hummingbird pollination. Most other
hummingbird-pollinated *Penstemon* species produce pelargonidin-based anthocyanins (Scogin
and Freeman 1987; Wessinger and Rausher 2015), whereas the hummingbird-adapted
Dasanthera species produce the more-hydroxylated cyanidin. This difference is observable by
the naked eye, such that flowers with cyanidin are pink-magenta, while flowers with
pelargonidin are red. It is possible that the absence of pelargonidin in *Dasanthera* flowers
reflects a more generalized floral phenotype that is thus less specialized to hummingbird
pollination. Although hummingbirds are frequent visitors to *P. newberryi*, they are only present
at low and mid-elevations throughout their range; a diverse community of potential pollinators
visit both *P. newberryi* and *P. davidsonii*, which suggests a somewhat generalist phenotype in *P.*
newberryi and explains why hybridization is so common between these species (Kimball 2008).

And, while *P. newberryi* has many characteristics of hummingbird specialization, other features,
including small quantities of nectar (Scogin and Freeman 1987; personal observation) and the
aforementioned magenta corolla suggest it has not fully transitioned to a specialized
hummingbird-pollination phenotype. The same may be true for *P. rupicola*, where (to the
authors' knowledge) the only pollinator observation study found that while hummingbirds do
visit *P. rupicola* flowers, the most common floral visitors to *P. rupicola* were pollen-collecting
halictid bees, which are unlikely to be effective pollinators (Datwyler 2001). Conversely to *P.*
newberryi, however, *P. rupicola* produces large quantities of nectar (personal observation),
consistent with specialization to hummingbird pollination. In both cases, the apparent lack of
complete pollinator specialization likely plays a large role in facilitating interspecific gene flow,
an apparently common phenomenon in *Dasanthera*.
502
Patterns of discordance and introgression are consistent with selection driving divergence
We found widespread signatures of introgression, corroborating previous inquiries into
hybridization dynamics in *Dasanthera* (Stone and Wolfe 2021; Stone et al. 2023). However,
patterns of introgression and phylogenetic discordance were heterogeneous across the genome.
The genomic landscape of discordance and introgression are consistent with selection driving
species divergence and opposing introgression in the *Dasanthera* radiation. Trees inferred from
genomic windows tended to be more similar to the species tree topology in regions of higher
gene density (Figure 4). This relationship between discordance and gene content is consistent
with stronger purifying selection acting in gene dense regions: purifying selection acts to reduce
effective population size hence reducing discordance caused by ILS, and acts to oppose
introgression, a further source of discordance. Moreover, gene-dense regions exhibited greater
differentiation and reduced signals of introgression, consistent with reduced gene flow in gene-
rich regions. We expect that levels of differentiation and introgression are also shaped by
variation in local recombination rate, a near-universal pattern in population genetic datasets
(Begun and Aquadro 1992; Cruickshank and Hahn 2014; Martin and Jiggins 2017; Wolf and
Ellegren 2017). Specifically, high recombination regions should experience reduced genetic
differentiation and tolerate higher levels of introgression. Unfortunately, we currently lack a
recombination map for the *P. davidsonii* reference genome. In a more distantly related
Penstemon species (*P. barbatus*), gene-dense regions tend to have higher recombination rates,
resulting in an overall negative relationship between population differentiation and
recombination rate (Wessinger et al. 2023).
524
The two hummingbird-adapted species in our study both hybridize with the bee-adapted species
P. davidsonii in the Cascades and Sierra Nevada Mountains. These sets of species pairs provide
an opportunity to examine whether similar genomic barriers to gene flow accompany parallel
divergence in floral syndrome. For each species pair, we found strongly differentiated genomic
regions (Figure S.10). Such localized genomic regions of elevated divergence are expected when
there is strong divergent selection in the face of gene flow (Charlesworth et al. 1997; Feder and
Nosil 2010). These genomic loci appear resistant to introgression and may potentially be
involved in local adaptation to divergent habitats or reproductive isolation. One caveat is that
these outlier loci do not truly represent independent regions of elevated divergence, because they
include the same samples of *P. davidsonii* in both contrasts. Future genomic and phenotypic
analyses of these replicate hybrid zones will be better suited to determine whether these genomic
regions represent parallel loci of elevated divergence, and will begin to address whether patterns

of phenotypic variation and selection within hybrid zones show similar and/or predictable
patterns.
539

Conclusion

Penstemon subgenus *Dasanthera* represents a microcosm adaptive radiation, with its nine
species exhibiting a diversity of ecological niches and floral pollination strategies. Species
boundaries remain porous in *Dasanthera*, allowing for interspecific hybridization and
highlighting the potential for the adaptive introgression of ecologically important traits. In this
study we focused on the source of genetic variation for repeated origins of a novel flower color
in hummingbird-pollinated species, as we currently lack information on the genetic basis for
other ecological traits that have diversified in the group. Although our data suggests parallel de
novo mutations underlie repeated flower color evolution, it is plausible that adaptive
introgression has fueled other aspects of the radiation because evidence for introgression is
prevalent in *Dasanthera*. We suspect that both parallel de novo mutations and introgression have
contributed to diversification, as inferred in other rapidly radiating clades.

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Data Availability Statement

564 Raw sequencing data have been deposited into the NCBI Sequence Read Archive under the
565 accession number PRJNA1057825. Other data underlying this article, including processed .vcf
566 files, are available on figshare under doi: 10.6084/m9.figshare.24480499.

Figures

Figure 1. Sampling sites, species relationships, and anthocyanin pigmentation data for sampled *Dasanthera* taxa. Left: sampling localities for all ingroup samples included in this study. Right: 10kb genomic window ASTRAL species tree, where tips are individual samples as shown on the map. Numbers on nodes indicate ASTRAL quartet support / gene concordance factors / site concordance factors, respectively. All ASTRAL local posterior probabilities are 1. Black circles at select nodes denote focal internal branches. Magenta and blue-violet squares indicate floral anthocyanidins produced by each sample; pigment extraction was unsuccessful for samples without squares. Inset shows a simplified cartoon version of the anthocyanin biosynthesis pathway where Flavonoid 3',5'-Hydroxylase (F3'5'H) functions to convert the precursor of cyanidin (dihydroquercetin; DHQ) into the precursor of delphinidin (dihydromyricetin; DHM). Arrows indicate enzymatic reactions.

Figure 2. Patterns of genetic variation at the candidate flower color gene *F3′5′h*. (a) Maximum-

likelihood gene tree estimated for *F3′5′h* gene sequence. Numbers at nodes signify the

Shimodaira-Hasegawa approximate likelihood ratio test (SH-aLRT) support (%) / ultrafast

bootstrap support (%). Branches with no SH-aLRT support are collapsed. (b) Values of the

modified f_d introgression statistic (f_{dM}) for the triplet *P. cardwellii-P. newberryi-P. rupicola*,

calculated in 500 SNP windows, sliding every 250 SNPs. Positive values are consistent with

introgression between *P. newberryi* and *P. rupicola*, and negative values are consistent with

introgression between *P. cardwellii* and *P. rupicola*. f_{dM} is symmetrically distributed around zero

under the null hypothesis of no introgression. The red bar indicates a 400kb window surrounding

F3′5′h, the red dashed line indicates the scaffold-wide average f_{dM} value. (c) Visualization of

CDS for *F3′5′h* locus for a reference *P. cardwellii* (known functional copy) and all *P. newberryi*

and *P. rupicola* samples. Red arrows represent deletions. Green lines with black arrows represent

frameshift insertions. (d) Scaffold-by-scaffold values of f_{dM}, with the same parameters as in panel

B. Red lines are loess-smoothed f_{dM} values.

Figure 3. Patterns of introgression (f-branch) across the *Dasanthera* species tree. The f-branch statistic f_b is a genome-wide metric to help interpret a system of correlated f_4-ratio results, and is interpreted as excess allele (i.e., introgression) sharing between branches b (y-axis) and tips (x-axis) in the species tree. The color gradient represents the f_b score; grey boxes represent tests that are inconsistent with the species tree topology (tip is descended from sister branch of focal branch b). Statistically significant results (Z-score > 3) are indicated by an asterisk.

Figure 4. Patterns of topological discordance across the genome. Top panel: smoothed values for the proportion of sites in coding regions (genic fraction), and the normalized Robinson-Foulds distance between 10kb window trees and the species tree. Second panel: smoothed topology weights for 5-taxon window trees consistent with the three main internal branches of the species tree. Third panel: Smoothed site concordance factors (sCF) averaged over 100 quartets for 10kb window trees at the three main internal branches of the species tree. Bottom panel: Smoothed topology weights for the triplet: *P. newberryi* (*new*), *P. cardwellii* (*car*), *P. rupicola* (*rup*).

References

References

Amrad A, Moser M, Mandel T, De Vries M, Schuurink RC, Freitas L, Kuhlemeier C. 2016. Gain
and Loss of Floral Scent Production through Changes in Structural Genes during
Pollinator-Mediated Speciation. *Current Biology* 26:3303–3312.

Andrews S. 2010. FastQC: A Quality Control Tool for High Throughput Sequence Data. Available
from: http://www.bioinformatics.babraham.ac.uk/projects/fastqc/

Begun DJ, Aquadro CF. 1992. Levels of naturally occurring DNA polymorphism correlate with
recombination rates in D. melanogaster. *Nature* 356:519–520.

Berner D, Salzburger W. 2015. The genomics of organismal diversification illuminated by
adaptive radiations. *Trends in Genetics* 31:491–499.

Charlesworth B, Nordborg M, Charlesworth D. 1997. The effects of local selection, balanced
polymorphism and background selection on equilibrium patterns of genetic diversity in
subdivided populations. *Genet. Res.* 70:155–174.

Chen S, Zhou Y, Chen Y, Gu J. 2018. fastp: an ultra-fast all-in-one FASTQ preprocessor.
Bioinformatics 34:i884–i890.

Cicconardi F, Milanetti E, Pinheiro De Castro EC, Mazo-Vargas A, Van Belleghem SM, Ruggieri
AA, Rastas P, Hanly J, Evans E, Jiggins CD, et al. 2023. Evolutionary dynamics of genome
size and content during the adaptive radiation of Heliconiini butterflies. *Nat Commun*
14:5620.

Colosimo PF, Hosemann KE, Balabhadra S, Villarreal G, Dickson M, Grimwood J, Schmutz J,
Myers RM, Schluter D, Kingsley DM. 2005. Widespread Parallel Evolution in Sticklebacks
by Repeated Fixation of Ectodysplasin Alleles. *Science* 307:1928–1933.

Cruickshank TE, Hahn MW. 2014. Reanalysis suggests that genomic islands of speciation are due
to reduced diversity, not reduced gene flow. *Mol Ecol* 23:3133–3157.

Datwyler SL. 2001. Evolution and dynamics of hybridization in *Penstemon* subgenus *Dasanthera*
(Scrophulariaceae S.L.).

Datwyler SL, Wolfe AD. 2004. Phylogenetic Relationships and Morphological Evolution in
Penstemon subg. *Dasanthera* (Veronicaceae). *Systematic Botany* 29:165–176.

De-Kayne R, Selz OM, Marques DA, Frei D, Seehausen O, Feulner PGD. 2022. Genomic
architecture of adaptive radiation and hybridization in Alpine whitefish. *Nat Commun*
13:4479.

Edelman NB, Mallet J. 2021. Prevalence and adaptive impact of introgression. *Annu. Rev. Genet.*
55:265–283.

Edgar RC. 2004. MUSCLE: multiple sequence alignment with high accuracy and high throughput.
Nucleic Acids Research 32:1792–1797.

Ewels P, Magnusson M, Lundin S, Käller M. 2016. MultiQC: summarize analysis results for
multiple tools and samples in a single report. *Bioinformatics* 32:3047–3048.

Feder JL, Nosil P. 2010. The efficacy of divergence hitchhiking in generating genomic islands
during ecological speciation. *Evolution* 64:1729–1747.

Ferreira MS, Jones MR, Callahan CM, Farelo L, Tolesa Z, Suchentrunk F, Boursot P, Mills LS, Alves
PC, Good JM, et al. 2021. The Legacy of Recurrent Introgression during the Radiation of
Hares. *Systematic Biology* 70:593–607.

Giska I, Farelo L, Pimenta J, Seixas FA, Ferreira MS, Marques JP, Miranda I, Letty J, Jenny H,
Hackländer K, et al. 2019. Introgression drives repeated evolution of winter coat color
polymorphism in hares. *Proceedings of the National Academy of Sciences* 116:24150–
24156.

Givnish TJ. 2015. Adaptive radiation versus 'radiation' and 'explosive diversification': why
conceptual distinctions are fundamental to understanding evolution. *New Phytol*
207:297–303.

Gotoh O. 1992. Substrate recognition sites in cytochrome P450 family 2 (CYP2) proteins
inferred from comparative analyses of amino acid and coding nucleotide sequences.
Journal of Biological Chemistry 267:83–90.

Guindon S, Dufayard J-F, Lefort V, Anisimova M, Hordijk W, Gascuel O. 2010. New Algorithms
and Methods to Estimate Maximum-Likelihood Phylogenies: Assessing the Performance
of PhyML 3.0. *Systematic Biology* 59:307–321.

Haenel Q, Roesti M, Moser D, MacColl ADC, Berner D. 2019. Predictable genome-wide sorting
of standing genetic variation during parallel adaptation to basic versus acidic
environments in stickleback fish. *Evolution Letters* 3:28–42.

Harborne JB. 1984. Methods of plant analysis. In: Phytochemical methods: a guide to modern
techniques of plant analysis. Dordrecht: Springer Netherlands. p. 1–36.

Harrison RG, Larson EL. 2016. Heterogeneous genome divergence, differential introgression,
and the origin and structure of hybrid zones. *Mol Ecol* 25:2454–2466.

Hoang DT, Chernomor O, Von Haeseler A, Minh BQ, Vinh LS. 2018. UFBoot2: Improving the
Ultrafast Bootstrap Approximation. *Molecular Biology and Evolution* 35:518–522.

Ishiguro K, Taniguchi M, Tanaka Y. 2012. Functional analysis of *Antirrhinum kelloggii* flavonoid
3'-hydroxylase and flavonoid 3',5'-hydroxylase genes; critical role in flower color and
evolution in the genus *Antirrhinum*. *J Plant Res* 125:451–456.

Jones MR, Mills LS, Alves PC, Callahan CM, Alves JM, Lafferty DJR, Jiggins FM, Jensen JD, Melo-
Ferreira J, Good JM. 2018. Adaptive introgression underlies polymorphic seasonal
camouflage in snowshoe hares. *Science* 360:1355–1358.

Jones MR, Mills LS, Jensen JD, Good JM. 2020. Convergent evolution of seasonal camouflage in
response to reduced snow cover across the snowshoe hare range. *Evolution* 74:2033–
2045.

Jun G, Wing MK, Abecasis GR, Kang HM. 2015. An efficient and scalable analysis framework for
variant extraction and refinement from population-scale DNA sequence data. *Genome*
Res. 25:918–925.

Kalyaanamoorthy S, Minh BQ, Wong TKF, Von Haeseler A, Jermiin LS. 2017. ModelFinder: fast
model selection for accurate phylogenetic estimates. *Nat Methods* 14:587–589.

Kimball S. 2008. Links between floral morphology and floral visitors along an elevational
gradient in a *Penstemon* hybrid zone. *Oikos* 117:1064–1074.

Korunes KL, Samuk K. 2021. PIXY : Unbiased estimation of nucleotide diversity and divergence in
the presence of missing data. *Mol Ecol Resour* 21:1359–1368.

Li H. 2011. A statistical framework for SNP calling, mutation discovery, association mapping and
population genetical parameter estimation from sequencing data. *Bioinformatics*
27:2987–2993.

Li H. 2013. Aligning sequence reads, clone sequences and assembly contigs with BWA-MEM.
Available from: http://arxiv.org/abs/1303.3997

Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, Marth G, Abecasis G, Durbin R, 1000
Genome Project Data Processing Subgroup. 2009. The Sequence Alignment/Map format
and SAMtools. *Bioinformatics* 25:2078–2079.

Liang M, Chen W, LaFountain AM, Liu Y, Peng F, Xia R, Bradshaw HD, Yuan Y-W. 2023. Taxon-
specific, phased siRNAs underlie a speciation locus in monkeyflowers. *Science* 379:576–
582.

Maddison WP. 1997. Gene trees in species trees. *Systematic Biology* 46:523–536.

Malinsky M, Challis RJ, Tyers AM, Schiffels S, Terai Y, Ngatunga BP, Miska EA, Durbin R, Genner
MJ, Turner GF. 2015. Genomic islands of speciation separate cichlid ecomorphs in an
East African crater lake. *Science* 350:1493–1498.

Malinsky M, Matschiner M, Svardal H. 2021. Dsuite - Fast *D* -statistics and related admixture
evidence from VCF files. *Molecular Ecology Resources* 21:584–595.

Malinsky M, Svardal H, Tyers AM, Miska EA, Genner MJ, Turner GF, Durbin R. 2018. Whole-
genome sequences of Malawi cichlids reveal multiple radiations interconnected by gene
flow. *Nat Ecol Evol* 2:1940–1955.

Marcionetti A, Salamin N. 2023. Insights into the Genomics of Clownfish Adaptive Radiation:
The Genomic Substrate of the Diversification. *Genome Biology and Evolution* 15.

Marques DA, Jones FC, Di Palma F, Kingsley DM, Reimchen TE. 2022. Genomic changes
underlying repeated niche shifts in an adaptive radiation. *Evolution* 76:1301–1319.

Marques DA, Meier JI, Seehausen O. 2019. A Combinatorial View on Speciation and Adaptive
Radiation. *Trends in Ecology & Evolution* 34:531–544.

Martin A, Orgogozo V. 2013. The loci of repeated evolution: a catalog of genetic hotspots of
phenotypic variation. *Evolution* 67:1235–1250.

Martin SH, Davey JW, Jiggins CD. 2015. Evaluating the Use of ABBA–BABA Statistics to Locate
Introgressed Loci. *Molecular Biology and Evolution* 32:244–257.

Martin SH, Jiggins CD. 2017. Interpreting the genomic landscape of introgression. *Current
Opinion in Genetics & Development* 47:69–74.

Martin SH, Van Belleghem SM. 2017. Exploring Evolutionary Relationships Across the Genome
Using Topology Weighting. *Genetics* 206:429–438.

McGee MD, Borstein SR, Meier JI, Marques DA, Mwaiko S, Taabu A, Kishe MA, O'Meara B,
Bruggmann R, Excoffier L, et al. 2020. The ecological and genomic basis of explosive
adaptive radiation. *Nature* 586:75–79.

Minh BQ, Hahn MW, Lanfear R. 2020. New Methods to Calculate Concordance Factors for
Phylogenomic Datasets. *Molecular Biology and Evolution* 37:2727–2733.

Moran BM, Payne C, Langdon Q, Powell DL, Brandvain Y, Schumer M. 2021. The genomic
consequences of hybridization. *eLife* 10:e69016.

Nguyen L-T, Schmidt HA, Von Haeseler A, Minh BQ. 2015. IQ-TREE: A Fast and Effective
Stochastic Algorithm for Estimating Maximum-Likelihood Phylogenies. *Molecular
Biology and Evolution* 32:268–274.

Ostevik KL, Alabady M, Zhang M, Rausher MD. 2023. Genome report: Whole genome sequence
and annotation of *Penstemon davidsonii*. Available from:
http://biorxiv.org/lookup/doi/10.1101/2023.08.31.555769

Pamilo P, Nei M. 1988. Relationships between Gene Trees and Species Trees. *Molecular Biology
and Evolution* 5:568–583.

Peng F, Byers KJRP, Bradshaw HD. 2017. Less is more: Independent loss-of-function *OCIMENE*
SYNTHASE alleles parallel pollination syndrome diversification in monkeyflowers
(*Mimulus*). *American J of Botany* 104:1055–1059.

Pertea G, Pertea M. 2020. GFF Utilities: GffRead and GffCompare. *F1000Research* 9.

Quinlan AR, Hall IM. 2010. BEDTools: a flexible suite of utilities for comparing genomic features.
Bioinformatics 26:841–842.

Robinson DF, Foulds LR. 1981. Comparison of phylogenetic trees. *Mathematical Biosciences*
53:131–147.

Ronco F, Matschiner M, Böhne A, Boila A, Büscher HH, El Taher A, Indermaur A, Malinsky M,
Ricci V, Kahmen A, et al. 2021. Drivers and dynamics of a massive adaptive radiation in
cichlid fishes. *Nature* 589:76–81.

Rosenberg NA. 2002. The Probability of Topological Concordance of Gene Trees and Species
Trees. *Theoretical Population Biology* 61:225–247.

Sas C, Müller F, Kappel C, Kent TV, Wright SI, Hilker M, Lenhard M. 2016. Repeated Inactivation
of the First Committed Enzyme Underlies the Loss of Benzaldehyde Emission after the
Selfing Transition in Capsella. *Current Biology* 26:3313–3319.

Schliep KP. 2011. phangorn: phylogenetic analysis in R. *Bioinformatics* 27:592–593.

Schluter D. 2000. The ecology of adaptive radiation. OUP Oxford

Schluter D, Conte GL. 2009. Genetics and ecological speciation. *Proc. Natl. Acad. Sci. U.S.A.*
106:9955–9962.

Scogin R, Freeman CE. 1987. Floral anthocyanins of the genus Penstemon: Correlations with
taxonomy and pollination. *Biochemical Systematics and Ecology* 15:355–360.

Seehausen O. 2004. Hybridization and adaptive radiation. *Trends in Ecology & Evolution*
19:198–207.

Segatto ALA, Cazé ALR, Turchetto C, Klahre U, Kuhlemeier C, Bonatto SL, Freitas LB. 2014.
Nuclear and plastid markers reveal the persistence of genetic identity: A new
perspective on the evolutionary history of *Petunia exserta*. *Molecular Phylogenetics and*
Evolution 70:504–512.

Seitz C, Ameres S, Forkmann G. 2007. Identification of the molecular basis for the functional
difference between flavonoid 3'-hydroxylase and flavonoid 3',5'-hydroxylase. *FEBS*
Letters 581:3429–3434.

Short AW, Streisfeld MA. 2023. Ancient hybridization leads to the repeated evolution of red
flowers across a monkeyflower radiation. *Evolution Letters* 7:293–304.

Simpson GG. 1953. The major features of evolution. Columbia University Press

Smith SD, Rausher MD. 2011. Gene Loss and Parallel Evolution Contribute to Species Difference
in Flower Color. *Molecular Biology and Evolution* 28:2799–2810.

Stankowski S, Streisfeld MA. 2015. Introgressive hybridization facilitates adaptive divergence in
a recent radiation of monkeyflowers. *Proc. R. Soc. B.* 282:20151666.

Stevens JTE, Wheeler LC, Williams NH, Norton AM, Wessinger CA. 2023. Predictive links
between petal color and pigment quantities in natural *Penstemon* hybrids. *Integrative*
and Comparative Biology:icad073.

Stone BW, Rodríguez-Peña RA, Wolfe AD. 2023. Testing hypotheses of hybrid taxon formation
in the shrubby beardtongues (*Penstemon* subgenus *Dasanthera*). *American Journal of*
Botany 110:e16118.

Stone BW, Wolfe AD. 2021. Phylogeographic analysis of shrubby beardtongues reveals range
expansions during the Last Glacial Maximum and implicates the Klamath Mountains as a
hotspot for hybridization. *Molecular Ecology* 30:3826–3839.

Straw RM. 1955. Hybridization, Homogamy, and Sympatric Speciation. *Evolution* 9:441–444.

Suvorov A, Kim BY, Wang J, Armstrong EE, Peede D, D'Agostino ERR, Price DK, Waddell PJ, Lang
M, Courtier-Orgogozo V, et al. 2022. Widespread introgression across a phylogeny of
155 Drosophila genomes. *Current Biology* 32:111-123.e5.

Tanaka Y, Brugliera F. 2013. Flower colour and cytochromes P450. *Phil. Trans. R. Soc. B*
368:20120432.

Weir BS, Cockerham CC. 1984. Estimating F-Statistics for the analysis of population structure.
Evolution 38:1358–1370.

Wessinger CA, Freeman CC, Mort ME, Rausher MD, Hileman LC. 2016. Multiplexed shotgun
genotyping resolves species relationships within the North American genus *Penstemon*.
American Journal of Botany 103:912–922.

Wessinger CA, Katzer AM, Hime PM, Rausher MD, Kelly JK, Hileman LC. 2023. A few essential
genetic loci distinguish Penstemon species with flowers adapted to pollination by bees
or hummingbirds. *PLoS Biol* 21:e3002294.

Wessinger CA, Rausher MD. 2012. Lessons from flower colour evolution on targets of selection.
Journal of Experimental Botany 63:5741–5749.

Wessinger CA, Rausher MD. 2014. Predictability and irreversibility of genetic changes
associated with flower color evolution in *Penstemon barbatus*. *Evolution* 68:1058–1070.

Wessinger CA, Rausher MD. 2015. Ecological Transition Predictably Associated with Gene
Degeneration. *Molecular Biology and Evolution* 32:347–354.

Wheeler LC, Walker JF, Ng J, Deanna R, Dunbar-Wallis A, Backes A, Pezzi PH, Palchetti MV,
Robertson HM, Monaghan A, et al. 2022. Transcription Factors Evolve Faster Than Their
Structural Gene Targets in the Flavonoid Pigment Pathway. *Molecular Biology and*
Evolution 39:msac044.

Whitfield JB, Lockhart PJ. 2007. Deciphering ancient rapid radiations. *Trends in Ecology &*
Evolution 22:258–265.

Wilson P, Castellanos MC, Hogue JN, Thomson JD, Armbruster WS. 2004. A multivariate search
for pollination syndromes among penstemons. *Oikos* 104:345–361.

Wilson P, Wolfe AD, Armbruster WS, Thomson JD. 2007. Constrained lability in floral evolution:
counting convergent origins of hummingbird pollination in *Penstemon* and *Keckiella*.
New Phytologist 176:883–890.

Wolf JBW, Ellegren H. 2017. Making sense of genomic islands of differentiation in light of
speciation. *Nat Rev Genet* 18:87–100.

Wolfe AD, Blischak PD, Kubatko LS. 2021. Phylogenetics of a Rapid, Continental Radiation:
Diversification, Biogeography, and Circumscription of the Beardtongues (*Penstemon*;
Plantaginaceae). Available from:
http://biorxiv.org/lookup/doi/10.1101/2021.04.20.440652

Wolfe AD, Randle CP, Datwyler SL, Morawetz JJ, Arguedas N, Diaz J. 2006. Phylogeny,
taxonomic affinities, and biogeography of *Penstemon* (Plantaginaceae) based on ITS and
cpDNA sequence data. *Am. J. Bot.* 93:1699–1713.

Xu Y-C, Guo Y-L. 2020. Less is more, natural loss-of-function mutation is a strategy for
adaptation. *Plant Communications* 1:100103.

Zhang C, Rabiee M, Sayyari E, Mirarab S. 2018. ASTRAL-III: polynomial time species tree
reconstruction from partially resolved gene trees. *BMC Bioinformatics* 19:153.