`reconcILS`: **A gene tree-species tree reconciliation**

algorithm that allows for incomplete lineage sorting

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Abstract

Reconciliation algorithms infer the evolutionary history of individual gene trees given a species tree. Many reconciliation algorithms consider only duplication and loss events (and sometimes horizontal transfer), ignoring effects of the coalescent process, including incomplete lineage sorting (ILS). Here, we present a new heuristic algorithm for carrying out reconciliation that accurately accounts for ILS by treating it as a series of nearest neighbor interchange (NNI) events. For discordant branches of the gene tree identified by last common ancestor (LCA) mapping, our algorithm recursively chooses the optimal history by comparing the cost of duplication and loss to the cost of NNI and loss. We demonstrate the accuracy of our new method, which we call `reconcILS`, using a new simulation engine (`dupcoal`) that generates gene trees produced by the interaction of duplication, loss, and ILS under the MSC-DL model. Despite being a heuristic method, `reconcILS` is much more accurate than models that ignore ILS, and at least as accurate or better than leading methods that can model ILS, while also able to handle much larger datasets. We demonstrate the use of `reconcILS` by applying it to a dataset of 23 primate genomes, highlighting its accuracy compared to standard methods in the presence of large amounts of ILS.

Introduction

The sequencing of a large number of genomes from many different species has highlighted the dynamic nature of genes: they are often duplicated, lost, or moved between species via introgression or horizontal gene transfer. One of the main goals of evolutionary and comparative genomics has been to quantify this dynamism, both in terms of the number of events of each type and their timing. A popular approach for quantifying both measures is gene tree-species tree reconciliation (reviewed in Boussau and Scornavacca 2020). Given a gene tree topology and a species tree topology, reconciliation algorithms attempt to infer the evolutionary events necessary to explain any discordance in the respective topologies. Many different reconciliation algorithms have been developed, most often considering only duplication and loss (so-called "DL" models; e.g. Goodman et al. 1979; Page 1994; Guigo et al. 1996; Page and Charleston 1997; Chen et al. 2000; Zmasek and Eddy 2001; Durand et al. 2006), but also sometimes considering duplication, transfer, and loss ("DTL" models; e.g. Hallett and Lagergren 2001; Hallett et al. 2004; Bansal et al. 2012; Stolzer et al. 2012; Szöllősi et al. 2013; Jacox et al. 2016; Morel et al. 2020).

Despite the success—and widespread use—of DL and DTL reconciliation algorithms, they do not consider gene tree discordance due to the coalescent process. The coalescent is a model that describes how sequences within a population are related (Kingman 1982; Hudson 1983; Tajima 1983), and is relevant even when considering sequences from different species because these must also coalesce in shared ancestral populations. Importantly, gene trees and species trees can be discordant solely due to coalescence, in a process often called incomplete lineage sorting (ILS) or deep coalescence (Maddison 1997). Standard DL and DTL algorithms will incorrectly infer one duplication and three losses for every discordance due only to ILS (Hahn 2007), resulting in misleading results when applying reconciliation methods to trees with ILS (e.g. Stull et al. 2021). Duplication, loss, and coalescence can also interact in complex and non-intuitive ways (Rasmussen and Kellis 2012; Mallo et al. 2014; Li et al. 2021), making it almost impossible to simply exclude gene trees discordant due to ILS from an analysis.

A few different approaches to reconciliation have been used to accommodate

discordance due to coalescence. An early approach allowed short branches in the species

tree—which can lead to ILS—to be collapsed (Vernot et al. 2008; Stolzer et al. 2012). The

use of non-binary species trees means that such algorithms will not mistake ILS alone for

duplication and loss; however, they do not account for all effects of coalescence on

incorrect inferences of duplication and loss (see below). Chan et al. (2017) introduced a

reconciliation algorithm that included duplication, loss, transfer, and ILS, but it assumed

that new duplicates were completely linked to their parental locus (i.e. no recombination

between them). Given that the majority of new gene duplicates are not completely linked

(Schrider and Hahn 2010), such an assumption limits the number of genes that can be

analyzed. Rasmussen and Kellis (2012) introduced a probabilistic model considering

duplication, loss, and ILS and an associated reconciliation algorithm. While promising,

DLCoalRecon is highly parameterized, requiring users to specify rates of duplication and

loss, population sizes, and more prior to inference, and is computationally intensive. Wu et

al. (2014) developed a parsimony-based algorithm (DLCpar) based on this same model that

runs faster and without the need to specify duplication and loss rates ahead of time.

Here, we introduce a new heuristic method for finding the most parsimonious

reconciliation of a gene tree and a species tree in the presence of duplication, loss, and ILS.

Our goal is to improve upon existing methods by increasing speed and accuracy, as well as

by incorporating more coalescent processes (i.e. ILS is not the only cause of discordance

due to the coalescent—see section below on "Complex coalescence"). In addition, although

DLCpar has been shown to perform well on smaller trees (Wu et al. 2014; Du et al. 2021), it

is not straightforward to run it on larger datasets or include its algorithm within other

tools that also have reconciliation steps (e.g. Thomas et al. 2017; Zhang et al. 2020). We

hope that the algorithm introduced here will be easily adaptable to such scenarios. We

demonstrate the use of this new algorithm by applying it to 23 whole genomes from

primates, a dataset that has previously been shown to contain a large amount of

discordance due to ILS (Vanderpool et al. 2020).

Figure 1. The MSC-DL model generates full gene trees. *a) A gene tree at the parent locus*

(blue) is drawn from the species tree (outlined in black) via the multispecies coalescent (MSC)

model. A duplication event (red star) is drawn from a birth-death model, in this case

occurring in the population ancestral to species B and C. b) A gene tree at the daughter locus

is drawn from the MSC. The duplication is placed on this gene tree at the time at which it

occurred in the species tree (shown as a dotted line), choosing one of the two possible

branches at random. The inserted DNA now takes on the gene tree branch labeled in red. c)-g)

Possible ways that the parent and daughter loci coalesce to form the full gene tree. In all

panels the star and red branch below it come from the daughter gene tree, and the blue tree

comes from the parent gene tree. The coalescent process joining the two trees is shown as a

red branch above the star. The trees in the bottom row are simply unfolded views of the full

gene trees.

New Approaches

The multispecies coalescent model with duplication and loss

We begin with a description of the multispecies coalescent model with duplication and loss (MSC-DL). Our model is very similar to the multilocus multispecies coalescent (MLMSC) model of Li et al. (2021). The description of the MLMSC model and an algorithm for generating trees under this model were first given in Li et al. (2021), with a mathematical model and further algorithmic updates in Li et al. (2024). The Appendix goes over the conceptual and algorithmic differences between the MSC-DL and MLMSC. Below, we describe the most important biological parts of our model, leaving algorithmic details to the next section. This explanation is intended to make clearer which common biological processes are modeled by the MSC-DL, but not by standard DL and DTL methods.

The MSC-DL model treats each individual locus (i.e. non-recombing position in the genome) as evolving via the multispecies coalescent (MSC) model, in which discordance can arise only via incomplete lineage sorting. The MSC-DL further models relationships among these loci via duplication and coalescence, in addition to losses at any locus. In particular, the MSC-DL specifies:

   i.   Every locus in the genome has a history (sometimes called a "locus tree") that follows the species tree.

  ii.   Each locus in the genome has a gene tree topology drawn from the species tree according to the MSC model. This gene tree (or "genealogy") exists at this locus whether or not a duplication or loss has occurred.

iii. Duplication events copy genetic material—i.e. DNA sequence—from one locus (the
"parent" locus) and insert it at another locus (the "daughter" locus). At the parent
locus, duplicated genetic material descends from a lineage that existed at the time of
duplication, and so must coalesce above this point with other lineages at the parent
locus according to the MSC (Figure 1a). At the daughter locus, this material inserts
into (is inherited by) a randomly chosen lineage of the gene tree at the time of
duplication (e.g. lineage C in Figure 1b), following the gene tree of this lineage after
this point. Nucleotide mutations occur in the sequence that was duplicated, allowing
us to reconstruct gene trees connecting one or more loci.

iv. Parent and daughter loci can be any recombination distance apart. If they are
completely linked they share the same gene tree topology, while if they are unlinked
each locus has an independent topology.

v. Duplications and losses are generated at rates μ and λ via a birth-death process,
starting at the original locus. Each duplication event adds a sub-tree from a new
daughter locus, consisting only of lineages descending from the duplication (see
point iii). Losses remove descendant branches of gene trees below their occurrence
(see point vii).

vi. Duplications and losses can occur on any gene tree lineage at any locus that contains
genetic material descended from the original parent locus, regardless of whether
they were the original parent or daughter locus.

vii. Loss events do not remove a locus, but rather do not allow lineages descended from
a loss to extend to the present (Supplementary Figure S1). These lineages are
therefore not present in the full gene tree.

viii. The gene tree at each daughter locus coalesces with the gene tree at its parent locus
via the MSC (Figure 1c-g; Supplementary Figure S2). Note that even if parent and
daughter loci are completely linked, they still must coalesce together at the parent
locus—linkage only determines whether the two marginal gene trees have the same
topology or not.

ix. The "full" gene tree consists of the combined genealogies at all parent and daughter
loci together, including all duplication and loss events.

Simulation using `dupcoal`

We created a new simulator—`dupcoal`—for generating gene trees under the MSC-
DL model. This simulator was originally needed because it was not possible to track events
using the simulator of Li et al. (2021; the only one available at the time), and to evaluate the
performance of `reconcILS` we require detailed output. `dupcoal` generates data under
the MSC-DL model, which differs in some ways from the MLMSC model (see Appendix). Our
algorithm works as follows (code can be found at
https://github.com/meganlsmith/dupcoal):

1. Generate the original parent gene tree from the species tree according to the MSC
model.
2. Simulate duplications along the branches of existing gene trees (only once for any
tree) at a rate λ.
3. For each duplication, generate a daughter tree at a new locus according to the MSC
model. Then, place the duplication on the daughter tree by randomly selecting a
lineage available at the time of duplication on the appropriate branch of the species
tree (Figure 1b). Discard all other lineages from the daughter locus, and truncate the
sub-tree at the time of duplication.
4. Return to step 2 and recursively generate new duplications (and subsequent daughter
trees) on each new daughter tree. Proceed to step 5 when no additional duplications
have been generated on the last daughter tree.
5. Coalesce all trees together. Begin at the present and work backwards in time. First,
attempt to coalesce a daughter tree with its parent, which could itself be a duplicated
tree. If coalescence fails to occur before the tree originates, attempt to coalesce the
daughter tree with the parent of its parent, and so on. Repeat until all trees have been
joined together.
6. Generate losses on the final, full gene family tree at a rate μ.

In general, this algorithm is similar to the updated "MLMSC-II" simulator of Li et al.
(2024) and is capable of generating copy-number hemiplasy, along with several other
interactions between ILS, duplication, and loss not modeled by DL and DTL approaches

(see next section). However, our algorithm differs in a few ways from those of Li et al.

(2021; 2024); these differences are discussed in the Appendix.

While our simulator does not currently include the ability to simulate linked

duplications, introgression, or horizontal gene transfers, it is straightforward to add these

types of events in future iterations of our software. Linked duplications simply require that

the topology of the daughter gene tree is correlated with the topology of the parent gene

tree (proportionally to recombination distance), while both introgression and transfer can

be accommodated by drawing gene trees from a different locus history (e.g. a history of

introgression) in step 1 or step 3.

Complex coalescence: ILS, unsorted homologs, and copy-number hemiplasy

The process we are modeling—the interaction of duplication, loss, and

coalescence—can be quite complex, leading to both conceptual and linguistic confusion.

Here we briefly attempt to clarify some important terms that have been used in different

ways by different researchers. While we try to avoid introducing new language, we do want

to clearly define how terms are being used in this paper and their relation to previous uses.

Incomplete lineage sorting (ILS). ILS occurs when two or more lineages do not

coalesce in the population in which they co-exist. When this happens, these lineages enter

an ancestral population, at which time they may again coalesce, either with each other or

with additional lineages that are present. If ILS results in lineages from the same original

population coalescing first with a lineage from a different population, gene trees will be

discordant with the species tree. This process can result in discordant gene trees at the

parent and daughter loci considered here.

Importantly for the MSC-DL model, the probability of ILS for two lineages is

normally expressed as e^{-T}, where T represents the length of time during which the lineages

can coalesce in the same population, and is usually taken as the length of the relevant

branch in the species tree (see Rosenberg 2002; Bryant and Hahn 2020). However, when

considering ILS between a pair of duplicates, the amount of time available for coalescence

before ILS has occurred is limited to the time between the duplication event and the next

speciation event (going backward in time). This time is always shorter than length of

the entire branch of the species tree on which the duplication occurred, which can lead to

ILS even when all branches of the species tree are long. The importance of this distinction

will hopefully become clearer in the next sub-section.

Unsorted homology. Gene duplication is fundamentally a process by which an

existing haplotype in a population at one locus is inserted into another location. Because of

this, the inserted DNA at the new daughter locus carries with it a history of coalescence

with all the other haplotypes at the parent locus, and the full gene tree will reflect this

history. In other words, all duplication events also involve coalescence events. Mallo et al.

(2014) described a common phenomenon that can occur when duplicates coalesce

together, which they called "unsorted homology." The general idea is that standard DL/DTL

methods assume that duplication along a lineage also requires the joining of parent and

daughter trees along that same lineage; if this does not occur, there is unsorted homology.

As an example, duplication of a gene along the C lineage (Figure 1b) is assumed to

only lead to gene trees with the two copies in species C as sister to one another (Figure 1c).

Any alternative joining of these trees will lead DL/DTL methods to incorrect

reconciliations, which will in turn lead to incorrect homology assignments (i.e. orthology

and paralogy). However, due to the coalescent process, many other trees can be formed

(Figure 1d-g). Because of the mismatch between the lineage containing the duplicate and

the lineage to which it coalesces in Figures 1d-g, all of these trees would be considered

unsorted homologs. Note that, in the scenario shown here, the trees in Figures 1d and 1e

are equiprobable with the topology in Figure 1c, making unsorted homology the most likely

outcome.

It should be emphasized that all of the topologies presented in Figure 1 involve only

a single duplication and no losses, and neither the parent tree nor the daughter tree is itself

discordant due to ILS. In fact, the example in Figure 1d is not technically due to ILS either,

since coalescence occurs in the same population of the species tree in which the duplication

appeared (the ancestral population of species B and C).

This example highlights an additional complication of unsorted homology: it can

occur with or without ILS, and even when the species tree only contains two species—and

therefore neither parent nor daughter gene trees can themselves be discordant.

Supplementary Figure S3 shows examples of unsorted homology in a two-species tree with

ILS, while Supplementary Figure S4 shows an example without ILS. One can see that there

is very little time for coalescence (i.e. T) before ILS occurs in Supplementary Figure S3, as it

is the timing of duplication relative to speciation, not the length of the branch in the species

tree, that determines the probability of ILS. Some approaches to reduce the effects of ILS on

reconciliation have required the presence of "apparent duplications" (Lafond et al. 2014) or

"observable duplications" (Scornavacca et al. 2011)—essentially, duplication nodes that

have more than one copy of the gene in a descendant species. While such approaches do

prevent the inference of duplication when none has occurred, as shown by the examples of

unsorted homology here (Figure 1d-f, Supplementary Figure S3, Supplementary Figure S4),

they do not ensure the correct identification of duplication nodes, nor do they prevent

excess losses from being inferred.

Hemiplasy. Gene trees at any locus in the genome can be discordant with the species

tree. In such cases, the term "hemiplasy" (Avise and Robinson 2008) describes the

incorrect inference of homoplasy that can arise when mutations occur on a discordant

branch of a gene tree (i.e. an internal branch that does not exist in the species tree; see

Figure 1 in Hahn and Nakhleh 2016). In the context of either duplication or loss, such

mutations must occur on a discordant branch of a gene tree to lead to "copy-number

hemiplasy" (CNH). Loss events can cause CNH when they occur either on a discordant

parent tree (Supplementary Figure S5a) or a discordant daughter tree (Supplementary

Figure S5b). In contrast, duplication events can only cause CNH on discordant daughter

trees (Supplementary Figure S5c,d).

However, there is also a broader definition of copy-number hemiplasy in use, which

includes any duplication or loss that does not fix in all descendant lineages, regardless of

underlying gene tree topologies (Rasmussen and Kellis 2012; Wu et al. 2014). While this

definition of CNH encompasses the stricter one given above, it also includes cases such as

the one shown in Figure 1 (and Supplementary Figure S4), which the stricter definition

does not include. Because the mutation in Figure 1 occurs in the common ancestral

population of species B and C, but was only inherited by lineage C, this scenario would be

considered to be hemiplasy by the broader definition. A similar event occurs in
Supplementary Figure S4, even though there is no ILS, and no gene tree discordance is
possible. Importantly, unlike the strict definition of CNH (and unlike unsorted homology),
there is not necessarily a genealogical signal left by events under this broader definition of
CNH. For instance, if the duplication event in Figure 1a occurred a few generations later—
after the split between species B and C, along a tip lineage—the scenario would not be
considered hemiplasy by this broader definition, but would still produce the same range of
gene trees (i.e. Figure 1c-g). (This would now involve ILS, however.)

Regardless of the exact definitions and terms used, many of these important
processes are not modeled by common simulators (e.g. SimPhy does not simulate either
type of hemiplasy) and are not considered by common reconciliation algorithms (e.g.
DLCpar assumes that neither type of hemiplasy occurs). `Dupcoal` simulates all of these
phenomena—as does MLMSC-II—but also reports the occurrence of each in generating the
underlying gene trees.

Reconciliation using `reconcILS`

Overview of main algorithm

Our reconciliation approach uses nearest-neighbor interchange (NNI)
rearrangements to model ILS and coalescence events, and to avoid invoking unnecessary
duplications and losses. An NNI move on a rooted tree takes the three lineages defined by a
single internal branch, for instance $(A,(B,C))$, and rearranges them into the two alternative
trees, either $(B,(A,C))$ or $(C,(B,A))$. Any two bifurcating trees with the same set of labels at
the tips can always be transformed one into the other by a series of NNI rearrangements
(Robinson 1971).

NNI has been used previously in reconciliation algorithms to correct gene trees that
may have been inferred incorrectly (e.g. Chen et al. 2000; Chaudhary et al. 2011; Górecki
and Eulenstein 2011; Nguyen et al. 2013). Here, we use NNI because the effects of ILS on
gene trees can be modeled as a series of NNI rearrangements (Than and Rosenberg 2013),
not because there is necessarily a biological link between NNI and ILS. For instance, for

three species undergoing ILS, there are two possible discordant topologies, each of which is

one NNI move away from the species tree topology (e.g. Figure 2). For four lineages

undergoing ILS, there are 14 possible discordant topologies, each of which are between one

and three NNI moves away from the species tree topology. (Note that it is the number of

lineages undergoing ILS at a specific point on the tree that determines the NNI distance

between trees, not the total number of tips in a tree.) Our algorithm takes advantage of this

connection between NNI and ILS in order to reconcile gene trees, but can also

accommodate gene tree error using the same NNI events (see Results).

Figure 2. Summary of `reconcILS`*. a) The initial step generates an LCA mapping function*

between the gene tree and the species tree. Here, nodes G_1 and G_2 in the gene tree both map to

node S_1 in the species tree; we therefore refer to S_1 as a multiply mapped node. Additionally,

G_1 and G_2 together define a discordant branch, K_1, in the gene tree. For clarity, all mappings

involving tip nodes have been excluded. b) The algorithm attempts to reconcile the discordant

branch using NNI and loss. Here, reconciliation can be carried out with 1 NNI and 0 losses. c)

The algorithm attempts to reconcile the discordant branch using duplication and loss. Here,

reconciliation can be carried out with 1 duplication and 3 losses. `reconcILS` *must decide*

between the reconciliations in b) and c) for every discordant branch.

An overview of `reconcILS` is given in Algorithm 1 (see Supplementary Materials).

Our approach combines multiple sub-approaches that are commonly used in DL

reconciliation and other phylogenetic inferences. As a first step, we use the last common

ancestor (LCA) algorithm (Zmasek and Eddy 2001) to create a mapping function between

each node in a gene tree, G, and nodes in a species tree, S (Figure 2a). We refer to each of

these pairwise relationships (arrows in Figure 2a) as "mappings" from here on.

Unlike in standard DL methods, the LCA mapping here does not represent a final

reconciliation. Instead, this step identifies cases in which multiple nodes in the gene tree

map to an identical node in the species tree. For instance, in Figure 2a nodes G_1 and G_2 in

the gene tree both map to node S_1 in the species tree. We therefore call S_1 a "multiply

mapped" node. Additionally, we refer to the branch in the gene tree bracketed by nodes G_1

and G_2 as a "discordant" branch (labeled K_1 in Figure 2a); such branches do not exist in the

species tree. The focus of our method is in determining whether discordant branches

should be resolved via NNI and loss (Figure 2b) or via duplication and loss (Figure 2c).

Choosing among events (NNI, duplication, and loss) is carried out greedily for each branch,

with no consideration of a globally optimal set of events among all discordant branches in

the gene tree. Consequently, after choosing a set of events for one branch (given specified

costs), the process repeats itself until there are no multiply mapped nodes and no

discordant branches. Duplications implied by multiply mapped tip nodes in the species tree

are inferred via standard DL methods; inferences of losses are explained in the Methods.

As with all parsimony-based reconciliations, `reconcILS` requires a cost for each

event type: one for duplications (C_D), one for losses (C_L), and one for NNIs (C_I).

Reconciliations can be very sensitive to costs, such that maximum parsimony solutions

under one set of costs can be very different than under another set of costs. Although there

are few ways to determine costs independently (but see Discussion), there are some

approaches for finding the Pareto-optimal costs within a reconciliation context (e.g.

Libeskind-Hadas et al. 2014; Mawhorter et al. 2019). Here, as in most commonly used

reconcilation algorithms, we allow the user to set costs. The default costs within

`reconcILS` are set to $C_D = 1.1$, $C_L = 1$, and $C_I = 1$.

Labeling a reconciled gene tree

A full reconciliation of a gene tree, G, and species tree, S, will have reconciled all

multiply mapped nodes, leaving no discordant branches. The total number of duplications,

losses, and NNIs required for this reconciliation is the sum of each event type across all

branches; these numbers are reported by `reconcILS`. We also aim to label the gene tree

with these events—identifying where each event occurred in history—as this is an

important output of reconciliation algorithms. As `reconcILS` traverses the species tree

finding optimal local reconciliations for each branch, it also records the number and

location of all events. Labeling therefore consists of placing these events on the gene tree.

Traditional DL reconciliation algorithms label gene tree nodes as either D (for duplication)

or S (for speciation); they may also label the location of losses, L. Here, we additionally aim

to label the NNI events induced by ILS (I).

Importantly, there is no standard way to label ILS events on a tree, largely because

there is no exact map between gene tree nodes and species tree nodes under ILS. A branch

undergoes NNI (and ILS), not a node, so it is not clear which node flanking such a branch

should be chosen to be labelled. This labeling problem therefore differs fundamentally

from labeling under DL and DTL algorithms because of the lack of a one-to-one map

between nodes in the two trees. Here, we have created our own labeling system for ILS, one

that we think is both understandable and easy to work with.

Figure 3. Labeling reconciled gene trees. a) One possible reconciliation of the gene tree

from Figure 1f labeled by `reconcILS`*. The single ILS node (I) has been labeled with a branch*

that has been rearranged by NNI (C_1), the number of NNIs it has undergone (2), and the fact

that it also represents a duplication (D). b) The gene tree from Figure 1d labeled by

`reconcILS`. *This tree is correctly labeled, but note that it is the same topology as in panel c*

(see Methods for further explanation). c) The gene tree from Figure 1e labeled by

`reconcILS`. *Note that the program has labeled the bottom-most node flanking the*

discordant branch with D and I, although that is not the true history. d) A gene tree with only

duplications and losses (L) labeled by `reconcILS`. *Internal branches on a tree can also be*

labeled with L, though none are shown here.

Our solution for placing the *I* label on nodes of gene trees in `reconcILS` is to

denote both a daughter branch affected and the number of NNI moves that were associated

with this branch. A few examples are shown in Figure 3, which also helps to illustrate why

these choices were made. Figure 3a labels the gene tree from Figure 1f—with more

complete tip names—with output from `reconcILS`. As can be seen, this tree has one node

(denoted with a small black dot for clarity) labeled with both duplication and ILS. Further,

the labeling tells you which daughter branch was reconciled with NNI (C_1), and how many

NNI moves were required to reconcile it (2). Without such information, simply labeling the

node with *I* would not have distinguished histories involving coalescent events along the *A*

branch from those along the C_1 branch. Further details of this labeling are provided in the

Methods, and caveats are covered in the Discussion.

`reconcILS` also labels duplications and losses in the absence of ILS. Figure 3d

shows how both individual duplications and losses are labelled: observe that neither

requires a label with a specified number of events (like the *I* label), nor does either need to

specify on which branch they act. `reconcILS` places the label *L* on any node in the implied

gene tree that is not represented by a gene copy; for example, generic tip nodes *A*, *C*, and *B*

in Figure 3d do not exist in the input gene tree itself, but are labelled with an *L*.

`reconcILS` can do this because every duplication event is represented internally by

generating a new copy of the species tree below the duplicated node—if losses occur on

these implied trees, we can track their timing as well. Note that internal nodes can also be

labelled with an *L*, so that the timing of loss events must be inferred by comparing the state
of parent and daughter nodes (i.e. losses on internal branches will lead the descendant
internal node and all of its daughter nodes to be labelled with an *L*). Finally, any node
without a label can be inferred to be a speciation node.

Results

Accuracy and advantages of `dupcoal`

We assessed the accuracy of `dupcoal` by first simulating 1000 gene trees along a
40-tip species tree with no duplication and no loss (Supplementary Figure S6). These
simulations therefore only model ILS as a cause of discordance. We compared the degree of
discordance in these simulations to ones carried out using the same species tree with
SimPhy (after appropriate branch-length scaling; see Methods). We found a very high
correlation between concordance factors on each branch calculated from gene trees
produced by the two methods (ρ=0.99, *P*=3.53 x 10^{-31}), indicating that we are accurately
simulating ILS. We also carried out simulations in `dupcoal` with both duplication and
loss, again comparing our results to SimPhy (after scaling duplication and loss rates
appropriately; see Methods). Again, the average number of duplications and losses per
simulated gene tree (*n*=1000) were highly similar and not significantly different between
the two methods: 3.82 vs. 3.84 duplications and 3.08 vs. 3.12 losses for `dupcoal` and
SimPhy, respectively (*P*=0.88 for duplications and *P*=0.64 for losses; *t*-tests), indicating that
we are accurately simulating duplications and losses in `dupcoal`.

To highlight additional features contained within `dupcoal`, we also tracked the
number of simulated gene trees in which copy-number hemiplasy occurred (this is a
standard output reported by `dupcoal`). Out of 1000 gene trees simulated on the 40-tip
tree with duplication, loss, and ILS, 511 contained at least one (strict) hemiplasy event. As a
straightforward way to demonstrate the effect of such events, we separately inferred the
number of duplications and losses on gene trees with and without copy-number hemiplasy
using ETE3 (Huerta-Cepas et al. 2016). While ETE3 implements a DL reconciliation

algorithm that does not model any type of ILS (see next section), this contrast shows the

effects of hemiplasy above and beyond the effects of ILS. As expected, both the number of

inferred duplications (2.61 excess duplications; P=6.44 x 10^{-11}, t-test) and the number of

inferred losses (8.31 excess losses; P=1.59 x 10^{-12}, t-test) were higher among the trees

containing hemiplasy. These results emphasize the importance of having a simulation

engine that can simulate hemiplasy.

Accuracy and advantages of `reconcILS`

As a simple test of DL reconciliation algorithms—which do not allow for ILS—we

used the DL method implemented in ETE3 (Huerta-Cepas et al. 2016). We initially ran

simulations in `dupcoal` without duplication or loss (i.e. ILS only); this allows us to see

incorrectly inferred duplications and losses more easily. As expected, on a species tree with

only 3 tips, ETE3 always inferred 1 extra duplication and 3 extra losses on gene trees with

discordance due to ILS (not shown). On a larger species tree with 40 tips, ETE3 inferred a

large excess of duplications and losses when only ILS is acting (9.5 duplications and 34.4

losses per gene tree, on average). On datasets simulated on a 40-tip tree with duplication,

loss, and ILS, the accuracy of ETE3 is low for losses (ρ=0.23) but is still fairly good for

duplications (ρ=0.79) (Supplementary Figure S7).

In contrast, `reconcILS` accurately infers the numbers of duplications and losses

(Figure 4a and 4b), as well as ILS events (Figure 4c) on the 40-tip tree. The correlation

between inferred and simulated duplications and losses were high (ρ=0.95 and 0.63,

respectively). We also observed a high correlation between the inferred and simulated

number of NNI events needed to reconcile a gene tree (ρ=0.67). ILS immediately followed

by loss will not leave a signal in the resulting gene trees, leading to a slight undercounting

of NNI events caused by ILS. It should also be noted that all inferences here are carried out

with the true gene trees, as we have not included any (error-prone) steps to infer the trees

from sequence data. Later, we relax this assumption.

We also examined results from `reconcILS` when using different costs (the

numbers in the previous paragraph all used the default costs: $C_D = 1.1$, $C_L = 1$, and $C_I = 1$).

When increasing the cost of duplication and loss ($C_D = 4$, $C_L = 3$, and $C_I = 1$), the results

remained largely the same, with high correlations between inferred and true duplication,

loss, and NNI: ρ=0.92, 0.61, and 0.69, respectively. The same is true when using the default

costs from DLCpar ($C_D = 2$, $C_L = 2$, and $C_I = 1$): all correlations remain high (ρ=0.98, 0.64,

and 0.68, respectively). However, when increasing the cost of ILS ($C_D = 1$, $C_L = 1$, and $C_I = 5$),

5), the results are much worse, closely resembling those from ETE3: ρ=0.79, 0.23, and 0.25,

respectively. This match between `reconcILS` and ETE3 should not be surprising, as

having a high cost of ILS is similar to ignoring ILS.

***Figure 4. Accuracy of `reconcILS` on data simulated from the 40-tip tree.** a)*

Number of inferred versus true duplication events. b) Number of inferred versus true loss

events. c) Number of inferred versus true ILS events. Circle size is proportional to the number

of reconciliations with each value, with jitter applied for clarity. Orange line represent best-fit

regression (as calculated in Seaborn; Waskom 2021), plus confidence intervals. The dashed

line represents y=x. See Supplementary Figure S7 for results from ETE3 on the same data.

As we did with ETE3 in the previous section, we also contrasted the number of

duplications and losses inferred by `reconcILS` on simulated gene trees with and without

copy-number hemiplasy (using default costs). Unfortunately, `reconcILS` also does not

deal perfectly with strict hemiplasy, though in a slightly different way than ETE3. Rather

than overestimating both the number of duplications and losses, `reconcILS` appears to

generally infer a true ILS event as a loss event instead. This occurs because (for instance) a

duplication on a discordant daughter gene tree branch will duplicate a non-sister pair of

lineages (e.g. *A* and *B*), which `reconcILS` interprets as a duplication and a loss (of *C*, in

this case) rather than as a duplication and an NNI event (Supplementary Figure S5).

Consistent with this, we observed no excess duplications in the simulations with hemiplasy

(0 extra duplications, on average), but significantly more losses (1.36; P=2.2 x 10^{-25}, t-test)

and significantly fewer NNI events (3.63; P=4.1 x 10^{-13}, t-test) compared to simulations that

did not have hemiplasy.

For completeness, we compared `reconcILS` to ETE3 under two additional

conditions. First, we used the same 40-tip tree topology as above to simulate gene trees

with duplication and loss, but very little ILS (Supplementary Table 1). Under these

conditions `reconcILS` remained highly accurate for inferences of duplication and loss

(ρ=0.99 and 0.85, respectively), though the inferences of very rare ILS events (much fewer

than 1 per tree) meant that the accuracy of these inferences was low (ρ=0.08). As expected

under these conditions, ETE3 was highly accurate in its inferences of duplications and

losses (ρ=0.99 and 0.94, respectively), with even greater accuracy than `reconcILS` for

losses. Second, we added error to the trees generated by the original 40-tip tree with

duplication, loss, and ILS (Methods). At the lowest level of error (mean of 5 errors per tree),

both `reconcILS` and ETE3 show slight reductions in accuracy (Supplementary Table 1).

As more error is added, both methods continue to become less accurate. At extreme levels

of error (mean of 15 errors per tree) `reconcILS` and ETE3 have almost equivalent

accuracy, with only a slight advantage in inferring loss for `reconcILS` (Supplementary

Table 1).

We also compared the results from `reconcILS` with another reconciliation method

that allows for ILS, DLCpar (Wu et al. 2014). DLCpar was written in Python 2, and—

possibly because we had to run it in a virtual environment—could not run on the larger

gene trees produced by the 40-tip tree. Therefore, we compared the performance of

`reconcILS` and DLCpar on 1000 gene trees simulated on both a 3-tip and a 16-tip species

tree (we still had to remove all gene trees with 25 or more tips, as these would not run in

DLCpar). On the 3-tip tree, the accuracy of both methods was high for duplication

(`reconcILS`, ρ=0.97; DLCpar, ρ=0.97; Supplementary Figure S8a). The correlation

between simulated and inferred ILS events was also high for both (`reconcILS`, ρ=0.85;

DLCpar, ρ=0.76; Supplementary Figure S8b). All results were highly similar on the 16-tip

tree, using the default costs from both programs (Supplementary Table 2). It is important

to note that DLCpar infers the number of ILS events, rather than the number of NNI

rearrangements that occur within such an event (as in `reconcILS`). To ensure that a fair

comparison can be made, results reported for DLCpar are simulated vs. inferred ILS events,

rather than NNI events. In both cases the methods are doing well in their respective

inferences, but with slightly higher accuracy for `reconcILS`.

There is quite a difference between `reconcILS` and DLCpar in the inference of loss

on the 3-tip tree (correlations of ρ=0.84 and ρ=0.48, respectively; Supplementary Figure

S8c). Upon closer inspection, the lower correlation in DLCpar is due to many loss events of

one particular type not being counted, which are much more common on the 3-tip tree;

indeed, on the 16-tip tree the correlations are highly similar (Supplementary Table 2).

Specifically, DLCpar does not count losses that occur above the root of the gene tree being

considered. This means that, for instance, in gene trees with the topology (B,C), DLCpar

does not count the loss that occurred along the A branch (assuming the species tree in

Figure 1). We confirmed this behavior on larger trees in DLCpar, as well as in all trees in

ETE3—no losses above the root of a gene tree are considered. The difference in counting of

losses among algorithms arises because `reconcILS` explicitly traverses the species tree,

while the other methods do not. This enables our method to track losses that occur above

the root of the input gene tree, but below the root of the input species tree. However, there

may be some cases in which the behavior of DLCpar and ETE3 is preferred. For instance,

since gene families must originate on some branch of the species tree (e.g. Richter et al.

2018), forcing reconciliation algorithms to infer losses on branches that pre-date this

appearance would be incorrect. It is not clear which approach to counting losses one

should prefer when analyzing real data.

Finally, we assessed the runtime and memory requirements of `reconcILS`

compared to ETE3 on a single node of the Indiana University Research Desktop, running an

Intel Xeon processor with 100 GB of RAM (DLCpar can only be run in Python 2, which

makes comparisons highly unfair). On a 3-species tree, both programs are extremely fast

(average time of 0.0086 seconds per gene tree for reconcILS and 0.013 seconds per gene

tree for ETE3; Supplementary Figure S9a) and both use very little memory (average peak

memory per gene tree of 0.075 MB for `reconcILS` and 0.054 MB for ETE3;

Supplementary Figure S9b). Both programs remain very fast on data produced from a 40-

species tree, though the faster speed of ETE3 is also evident (4.9 seconds per gene tree for

`reconcILS` and 1.3 seconds for ETE3; Supplementary Figure S9a). Regardless, this means

that the entire 1000-gene-tree simulated dataset on the 40-species tree took less than 84

minutes to analyze using `reconcILS` on a moderately powerful computer. Similarly, the

memory requirements of `reconcILS` increase on the 40-tip tree (18.79 MB per gene tree

for `reconcILS` and 3.19 MB for ETE3; Supplementary Figure S9a), but are not onerous for

modern computers. As the 40-species tree can contain many more events (duplication, loss,

or ILS), we also plotted the average time and memory required for trees containing

different numbers of total events (Supplementary Figure S9c,d). Both programs show

approximately linear increases in time and memory with the number of events, which is

promising for analyzing larger datasets with larger species trees, though we have not

tested it on much larger trees.

Analysis of primate genomes

To further demonstrate the power and accuracy of `reconcILS`, we applied it to a

dataset of gene trees inferred from 23 primate genomes (Vanderpool et al. 2020; Smith et

al. 2022). These genomes all come from the suborder Haplorhini, which includes all

primates except the lemurs and lorises (suborder Strepsirhini). Among these genomes, we

analyzed two sets of gene trees that were previously inferred. First, we considered 1,820

single-copy orthologs. This dataset is not expected to contain any duplicates, though it does

contain losses. However, because there are high levels of ILS among the primates sampled

here (Vanderpool et al. 2020), the dataset serves as an ideal comparator for reconciliation

approaches with and without ILS. Second, 11,555 trees containing both orthologs and

paralogs were used to infer duplications, losses, and ILS among the primates. We used both

`reconcILS` and ETE3 to analyze the two datasets, assigning inferred events to each

branch of the primate tree.

Single-copy ortholog dataset

Results obtained from analysis of only the single-copy orthologs are shown

graphically in Figure 5a (see Supplementary Figure S10a for numerical values on each

branch). While this is a somewhat contrived example—a set of only single-copy genes

would not be analyzed on its own using reconciliation methods—we think the comparison

with the full dataset is highly informative. For instance, the pattern that stands out from

this graph is that ETE3 is vastly overestimating the number of duplications and, especially,

the number of losses. Because ETE3 does not model ILS, all such events must be reconciled

by one duplication (placed above the discordant branch) and three losses (placed below

the discordant branch; Hahn 2007). We can see this effect in the data by looking at the

correlation between the gene concordance factor (gCF) estimated for a branch with the

number of duplicates inferred on its parent branch. Gene concordance factors quantify the

fraction of loci that contain the same branch as the species tree; when single-copy genes

are analyzed, low gCFs can indicate discordance due to ILS or other factors, like gene tree

estimation error (Baum 2007; Lanfear and Hahn 2024). The correlation between gCFs for

these data (estimated in Smith et al. 2022) and duplications inferred by ETE3 on parent

branches is ρ=-0.56. Lower gCFs lead to higher numbers of duplicates, as expected when

ILS is not accounted for—this problem is not unique to ETE3, but should be expected for all

standard DL and DTL approaches. Such errors can lead to incorrect inferences of bursts of

duplication and loss.

Figure 5. Results from primates. *a) Analysis of 1,820 single-copy orthologs. Both*
`reconcILS` and ETE3 were used to analyze the data, with duplication, NNI, and loss events
mapped to each branch of the species tree. On each branch, the bar plots show results from
`reconcILS` on top, with results from ETE3 mirrored below. ETE3 cannot infer NNI events, so
this bar is always set to 0. b) Analysis of 11,555 gene trees containing both orthologs and
paralogs. Results are presented in the same manner as in panel a. However, note that the
height of all bars is normalized within a panel, so that heights of bars within a tree can be

 compared among branches, but the heights of bars between the trees cannot be compared.

Supplementary Figure S10 shows the absolute counts of all events from both panels and

Supplementary Figure S11 shows species tree with branch lengths in substitution units rather

than as a cladogram, as shown here. Trees were drawn with ggtree (Yu et al. 2017).

Using the same reasoning, we can assess the accuracy of `reconcILS` by examining

the correlation between NNI events inferred by our algorithm and gCF values for single-

copy genes. High NNI counts indicate more ILS, and therefore this measure should be

negatively correlated with gCF values for the same branch. Indeed, the correlation between

NNI counts and gCF values is ρ=-0.83, demonstrating the high accuracy of `reconcILS` in

quantifying ILS (or in correcting for gene tree error). However, `reconcILS` does

sometimes infer a small number of duplications (Figure 5a; Supplementary Figure S10a).

While it is possible that some of the single-copy genes we analyzed represent so-called

pseudoorthologs—and therefore do contain actual duplication events—these are expected

to be rare (Smith and Hahn 2022). Instead, these false duplicates are likely due to ILS

among four or more lineages. Recall that, although our algorithm has been developed to

account for rearrangements due to the coalescent process, ILS among four lineages can

result in gene trees that are three NNI moves away from the species tree; ILS among more

lineages will produce trees that are even further away. Therefore, multiple NNI events may

sometimes simply be equally or more costly than duplication and loss, even when ILS is the

true cause of discordance (`reconcILS` will report the duplication+loss solution to a

reconciliation when there are equally optimal reconciliations).

Full gene tree dataset

We also examined patterns of ILS, duplication, and loss for the entire primate

dataset. The results from the full set of gene trees (Figure 5b; Supplementary Figure S10b)

largely mirror the results from the single-copy orthologs (which are a subset of the entire

dataset). Part of the apparent similarity is simply due to normalizing the two datasets

separately, but several non-graphical factors also contribute to the pattern. First, although

there are many true duplicates in the full dataset, the vast majority of gene trees only have

events on tip branches: 7,693 out of 11,555 gene trees have duplicates specific to one

species (Smith et al. 2022). Another ~1,000 trees have duplicates specific to two species.

Combined with the 1,820 single-copy genes, it becomes clear that there are in fact

relatively few trees with duplicates on internal branches even in the full dataset.

Second, errors in inferring duplications and losses due to the coalescent process still

dominate the results from standard DL reconciliation (i.e. ETE3), possibly even more so

than in the single-copy dataset. Because ILS (and introgression) can affect all gene trees,

trees with duplicates can experience twice as much discordance when there are twice as

many branches. One example of this can be seen among the Papionini (macaques, baboons,

geladas, and mandrills): ETE3 has to infer >6,000 losses on each of the tip-branches in this

clade, and more than 10,000 losses on most of the internal branches (Figure 5b;

Supplementary Figure S10b). There are also many thousands of duplicates inferred on

internal branches of this subclade, though these species have the same number of total

protein-coding genes as all the others analyzed here (Vanderpool et al. 2020). Although the

inferred duplications and losses in this group were also relatively higher in the single-copy

dataset, it is clear that even a small number of true duplicates can have an outsized effect

on methods that cannot account for ILS.

The analysis using `reconcILS` is also highly similar between the two datasets,

again largely because most duplicates are confined to tip branches. `reconcILS` is able to

deal with discordance due to coalescence, even in the presence of duplication: the

correlation between NNI events inferred on each branch of the species tree from the single-

copy orthologs and the full dataset is $\rho = 0.99$. Although the numbers of events are of

course larger with the larger dataset (Supplementary Figure S10b), the effect of ILS on any

given branch of the species tree is experienced by all gene trees. Given its ability to deal

with ILS, duplication events across the primate tree are more reliably inferred with

`reconcILS`.

Discussion

We have introduced a new reconciliation algorithm and associated software

package (`reconcILS`) that considers discordance due to duplication, loss, and

coalescence. To better demonstrate the accuracy of our method, we have also developed a

new simulation program (`dupcoal`) that generates gene trees while allowing these

processes to interact under the MSC-DL model. Our results demonstrate that `reconcILS`

is a fast and accurate approach to gene tree reconciliation, one that offers multiple

advantages over alternative approaches, even those that also consider ILS (e.g. DLCpar; Wu

et al. 2014). Nevertheless, there are still several caveats to consider, as well as future

directions to explore.

Despite our demonstration of good performance in practice, our method is heuristic:

we have no theoretical proof that our method finds most parsimonious reconciliations.

While such proofs often require strict assumptions—e.g. the proofs for DL algorithms

assume there is no ILS—they are very helpful in providing confidence under the specified

conditions. However, given the high accuracy of `reconcILS` demonstrated with both

simulations and empirical data from single-copy genes, we hope the heuristic nature of our

algorithm is not a barrier to its utility. Future theoretical work on this or similar

approaches using NNIs (e.g. Górecki and Eulenstein 2011) would of course be of benefit to

the field. Under any such model, it must also be recognized that errors in gene tree

reconstruction will have a detrimental effect on the results (e.g. Supplementary Table 1).

Both theoretical work and new algorithms must take such limits into account.

As mentioned earlier, there is often ambiguity in how to label nodes in gene trees

affected by ILS and duplication (e.g. Figures 3b and 3c). This is due in part to the nature of

ILS—it is not clear which node to label because there is not a one-to-one map between

gene tree nodes and species tree nodes—and in part to the intrinsic lack of information

contained only within tree topologies when coalescence and duplication interact. The

difficulty in labeling is not confined to this work: DLCpar does not label nodes, and

RecPhyloXML, a format for representing reconciled trees, does not include a way to label

ILS events at all (Duchemin et al. 2018).

Nevertheless, labeling gene trees serves multiple important functions, and is often

interpreted as biological truth. For example, multiple methods use inferred gene tree labels

to identify orthologous genes (genes related by speciation events; Fitch 1970). The general

approach taken is to label nodes using reconciliation or similar approaches, treating any

node not labeled with a D as a speciation node; genes whose common ancestor is a

speciation node are subsequently defined as orthologs. Both standard DL reconciliation

and so-called "overlap" methods (e.g. Huerta-Cepas et al. 2007; Emms and Kelly 2019) will

consistently label the top-most node flanking a discordant branch with a duplication event.

This means that, in cases like the one shown in Figure 3b, these methods will label the

wrong node with a D, and subsequently identify the wrong pair of genes as orthologs.

(Figure 3c would be labeled correctly; recall that this outcome has an equal probability to

the one in Figure 3b in the scenario considered here.) Cases such as these have been

referred to as "unsorted homologs" (Mallo et al. 2014), and will occur any time duplication

and coalescence interact, either before or after a speciation event. They can also occur even

if only two species are analyzed (e.g. Supplementary Figures S3 and S4), which means that

approaches that simply collapse short branches to avoid ILS will also be affected. Although

`reconcILS` currently only labels one node with a D, in the future we could allow for

multiple possible labelings in order to emphasize the inherent ambiguity of some labels.

The algorithm used by `reconcILS` deals, in part, with an additional interaction

between duplication, loss, and coalescence: hemiplasy. Hemiplasy is the phenomenon by

which a single mutational event on a discordant gene tree will be interpreted as multiple

events ("homoplasy") when reconciled to the species tree (Avise and Robinson 2008; Hahn

and Nakhleh 2016). In analyses of nucleotide substitutions, generic binary traits, and even

continuous traits, hemiplasy requires that mutations have occurred on a discordant branch

of a gene tree (Mendes and Hahn 2016; Guerrero and Hahn 2018; Hibbins et al. 2023). In

the context of either duplication or loss, such mutations would have to occur on a

discordant branch of a daughter gene tree to lead to copy-number hemiplasy. The

simulation engine introduced here, `dupcoal`, simulates hemiplasy events and reports any

gene trees affected. Our results show that these events lead to increased error in

reconciliation, and are therefore an important process to include. Interestingly, such events

have little effect on species tree inference when paralogs are used for this purpose (Li et al.

2024).

Unfortunately, strict copy-number hemiplasy events as defined above are still not

accurately inferred by `reconcILS`: we have found that our algorithm consistently turns

NNIs into losses when hemiplasy occurs. While the total number of events does not seem to

increase, it is not ideal that NNIs are mistaken for losses. However, `reconcILS` does deal

accurately with copy-number hemiplasy under the broader definition in use (see section on

"Complex coalescence…"), and `dupcoal` also simulates and records such events. In

contrast, DLCpar assumes that this type of hemiplasy does not occur, and SimPhy does not

simulate this type of hemiplasy.

The algorithm used by `reconcILS` deals with ILS differently than does DLCpar.

While DLCpar treats ILS as an event, `reconcILS` considers NNIs as events induced by ILS.

It is not clear that either approach should necessarily be preferred on first principles. In

simple cases involving three species and a single discordant branch (e.g. Figure 2), ILS and

NNI are more or less equivalent, though it should be noted that the process of ILS can also

lead to concordant trees and is not synonymous with discordance. However, in cases with

more than one discordant branch there is no simple relationship between the two. For

instance, in a 4-tip tree undergoing ILS, lineages may undergo ILS across two branches, but

result in between 0 and 3 NNI events (Than and Rosenberg 2013). We think that directly

modeling NNIs—as is done in `reconcILS`—is a natural way to express the events needed

to reconcile a gene tree with a species tree, including discordance due to gene tree error. In

fact, NNI and other moves have been used previously in reconciliation algorithms to correct

gene trees that may have been inferred incorrectly (e.g. Chen et al. 2000; Chaudhary et al.

2011; Górecki and Eulenstein 2011; Nguyen et al. 2013). Though these algorithms

minimize DL reconciliation costs using NNIs, they could also have been used within

`reconcILS` for each local reconciliation step. Regardless of the exact algorithm used, the

utility of such methods also provides a motivation for using `reconcILS` to obtain more

accurate inferences of duplications and losses in the presence of estimation error, not just

ILS.

One relatively straightforward improvement that could be made to `reconcILS` is
in the cost of different events. Costs must be specified in `reconcILS` ahead of time, but it
is difficult to determine biologically realistic costs for duplication, loss, and ILS. One
approach to determining duplication and loss costs would be to infer the number of such
events using an alternative, non-reconciliation approach (e.g. Mendes et al. 2020), setting
the cost in `reconcILS` to be inversely related to the inferred number of events of each
type. Varying rates of duplication and loss can also be inferred along different branches of
the species tree, and so the costs of these events could vary commensurately along the tree.
Similarly, we can envision a simple way to scale the costs of NNI with the length of species
tree branches: this is because the probability of discordance due to ILS is inversely
proportional to internal branch lengths on a species tree (Hudson 1983). We could
therefore make NNI events on longer branches of the species tree more costly, which would
properly weight the relative probability of such events locally on the tree. Regardless of the
method used, researchers should ensure that biological conclusions are qualitatively and
quantitatively similar across a range of costs.

There are also several extensions that could be made to `dupcoal`, our new
simulation engine. Currently, `dupcoal` draws a daughter gene tree from the multispecies
coalescent (MSC) at random: this implicitly assumes that the parent and daughter trees are
completely unlinked. It is straightforward, however, to draw the daughter tree at a given
recombination distance from the parent tree, ranging from fully linked to fully unlinked. In
addition, though the software currently draws gene trees from the MSC, we could also
include introgression as a source of discordance by drawing trees from the multispecies
network coalescent (Meng and Kubatko 2009; Yu et al. 2011). In this case, users would
simply input a species network to simulate from, rather than a species tree. We should also
note the differences between the current version of `dupcoal` and alternative simulators.
As mentioned previously, SimPhy (Mallo et al. 2016) does not simulate hemiplasy of any
kind, as it uses the same three-tree model of Rasmussen and Kellis (2012) and Wu et al.
(2014). However, this method does include gene transfers, which are not simulated by
`dupcoal`. The MLMSC-II simulator of Li et al. (2024) simulates hemiplasy under a similar
model to the one used here; `dupcoal` is therefore most similar to this method, with a few

differences (see Appendix). Despite these minor differences, both methods are capable of

generating a wide variety of gene trees impacted by duplication, loss, coalescence, and the

interactions of these processes. Unlike MLMSC-II, `dupcoal` records the exact counts of

several different event types, a necessity when evaluating the performance of

reconciliation approaches.

Finally, we envision multiple ways in which the `reconcILS` algorithm could be

used in conjunction with other methods. For instance, ASTRAL-Pro (Zhang et al. 2020)

infers a species tree from gene trees containing paralogs by first labeling nodes in each

gene tree as either duplication or speciation. The authors of this software rightfully

recognize the problems with labeling discussed here, but nevertheless the labeling step

might benefit from methods that include ILS. Likewise, GRAMPA (Thomas et al. 2017)

allows for reconciliation of polyploidy events, but does not include coalescence as a source

of discordance; its algorithm could easily accommodate this extension. There are also

multiple approaches that can be used to correct gene trees via reconciliation (e.g. El-

Mabrouk and Ouangraoua 2017; Christensen et al. 2020). These and many other tasks

might benefit from approaches that model duplication, loss, and coalescence in a single

framework.

Methods

Building and testing `dupcoal`

The simulator `dupcoal` is written in Python 3 and uses functions from DendroPy

(Sukumaran and Holder 2010) to generate trees under the MSC and to manipulate parent

and daughter trees. It also uses functions from NumPy (Harris et al. 2020) to draw

coalescent times, duplication times, and loss times from exponential distributions.

To test the accuracy of `dupcoal` we simulated gene trees along a species tree with

40 tips (Supplementary Figure S6). The species tree topology was drawn at random using

the program ape (Paradis and Schliep 2018), with the "Grafen" method for drawing branch

lengths (scaled up by 10); this tree was used as input to both `dupcoal` and SimPhy. The

amount of ILS in this tree, as measured by gene concordance factors (Minh et al. 2020a),

ranged from 29.1 to 99.6; in comparison, the values estimated from the primate tree in

Vanderpool et al. (2020) ranged from 27 to 96, so the two are quite similar. Because

`dupcoal` specifies branch lengths in coalescent units, while SimPhy uses time, branch

lengths were converted by assuming a population size of N=10 and a generation time of 1

year. Simulations were run on this tree with only ILS (setting duplication and loss rates to

$\lambda = 0$ and $\mu = 0$, respectively). When using non-zero duplication and loss rates (see next

paragraph), they were scaled to the appropriate branch lengths to make them comparable

in `dupcoal` and SimPhy. Comparisons between `dupcoal` and SimPhy with no duplication

and loss were carried out by calculating gene concordance factors (gCFs) on the species

tree using IQ-TREE2 (Minh et al. 2020a).

To test the accuracy of inferences using `reconcILS` and ETE3, the same 40-taxon

tree was used for simulations in `dupcoal` with ILS, duplication, and loss ($\lambda = 0.03$ and $\mu =$

0.03). These parameter values were chosen so that the simulations produced a large

number of duplications and losses to assess accuracy with, not because they are

biologically motivated. To add error, we specified three levels of gene tree errors, with a

Poisson number of errors drawn for each tree. The three levels corresponded to a mean of

5, 10, or 15 errors per gene tree; all errors were modeled as an NNI rearrangement.

Changes on each branch of each tree were drawn independently and sequentially, such that

the same branch could be rearranged multiple times. We also carried out simulations on

the 40-tip tree with very low levels of ILS by scaling all branch lengths up by 1000; to avoid

extremely large gene trees under this scenario, we also scaled the duplication and loss

rates down by 10.

DLCpar is written in Python 2 and could not be run in reasonable time on the 40-

taxon tree. We therefore also simulated data on a 3-taxon tree with topology

(A:1.5,(B:0.5,C:0.5):1), where branch lengths are measured in coalescent units and

duplication and loss rates were the same as on the larger tree. We also simulated data on a

16-taxon tree (based on the fungal tree in Butler et al. 2009), though DLCpar was unable to

analyze any gene tree with 25 or more tips. All reported results are therefore only from

those trees that could be analyzed by both methods. The default costs ($C_D = 1.1$, $C_L = 1$,

$C_I = 1$) were used for all analyses of simulated data for all methods, except where noted.

Note that it is the relative value of costs that matters for analysis, and the absolute values

do not correspond to any biological parameter.

Implementation of `reconcILS`

Inputs and outputs

Given a rooted, bifurcating species tree, S, and a rooted, bifurcating gene tree, G, as

input, `reconcILS` outputs the number of duplications, losses, and NNIs required to

reconcile G and S given event costs. Inferred events are all also assigned to nodes in the

gene tree.

Traversing trees

After generating an initial LCA mapping between a gene tree and the species tree,

`reconcILS` performs a preorder traversal of the species tree. For each species tree node,

the algorithm checks how many gene tree nodes are mapped to it. If there are fewer than 2

mappings, the algorithm asks whether speciation or loss can be inferred. As is standard in

reconciliation, species tree nodes mapped to one gene tree node imply a speciation event at

that gene tree node—these do not incur a cost. Nodes mapped to zero gene tree nodes

imply at least one loss in the gene tree. Our program asks whether the parent node of a

node with 0 mappings itself has 0 mappings. If not, a loss is inferred; if so, no event is

recorded, because the loss must have occurred further up the tree. If there are 2 or more

mappings the species tree node is multiply mapped, and a function to carry out "local

reconciliation" is called. Local reconciliation is described in more detail in the next section,

but here can simply be described as determining the events that have generated the

multiple mappings to a single species tree node. It should also be noted that if a duplication

node is inferred for a gene tree, both sub-trees induced by the duplication are further

locally reconciled (Algorithm 1); this means that `reconcILS` can detect cases in which

multiple independent losses have occurred along the same lineage (e.g. Supplementary

Figure 1).

If there are only two adjacent nodes in the gene tree mapped to a node in the species

tree (i.e. one discordant branch, as in Figure 2), then the method proceeds as described

below. However, when there are more than two nodes mapped to an internal species tree

node—and therefore more than one discordant branch in the gene tree—we must decide

which branch to reconcile with the multiply mapped node first. We have found cases where

the order in which the branches are resolved matters, with only one of the choices leading

to an optimal reconciliation. Algorithm 2 (see Supplementary Materials) describes an

algorithm for choosing which discordant branch to locally reconcile first. This algorithm

calculates the "bipartition cost" of each possible branch to reconcile. We calculate the

bipartition cost of a branch by examining the two trees induced by an NNI move applied to

the branch. For each of these new topologies, we calculate costs using two measures, both

of which are applied independently to the two sub-trees extending from the root of these

topologies (denoted by the red vertical line in Supplementary Figure 12b,c). First, we count

the number of bipartitions present in these sub-trees but not present in the species tree

("bipartition" cost in Supplementary Figure 12); second, we count the number of lineages

missing relative to the node in the species tree the root of the topology is mapped to ("lost"

cost in Supplementary Figure 12). The bipartition cost of a single branch therefore requires

eight values to be calculated, four for each of the trees created by NNI (e.g. Supplementary

Figure 12b). Such calculations are simple and can be carried out quickly, and by choosing

the branch with the lowest bipartition cost we can more accurately proceed with

reconciliation.

Once reconciliation for a single multiply mapped node in the species tree is carried

out (i.e. it is no longer multiply mapped), our algorithm begins again: a new round. In each

round we generate an LCA mapping function and continue traversing the species tree,

counting gene tree nodes mapped to each species tree node.

Choosing among events

In order for our algorithm to more efficiently and optimally choose among different

evolutionary events (e.g. Figure 2b vs 2c), we introduce the concept of a "local

reconciliation" for a multiply mapped node. Here, a species tree node is locally reconciled if

the number of gene tree nodes mapped to it, m, is reduced. For internal nodes of the

species tree, this, by definition, also reduces the number of discordant branches in the gene

tree.

The use of local reconciliation has several advantages. Most importantly, it provides

a criterion for moving forward: we can determine if a reconciliation has been achieved

within a single round of the algorithm. It also makes it clear that there are some scenarios

that can only be reconciled using duplication and not NNI (e.g. a multiply mapped tip node),

in which case choosing events is straightforward. In a single round of our algorithm, we

attempt to locally reconcile a single multiply mapped node using both NNI+loss and

duplication+loss (Figure 2).

Note that, given this definition, a species tree node can be locally reconciled by NNI

while still remaining multiply mapped; e.g., if $m = 3$ is reduced to $m = 2$, the node still

requires more reconciliation. In this case our algorithm will carry out up to m NNI moves

within a single round, as long as each one results in a local reconciliation. For example,

starting with $m = 3$, the first NNI move will locally reconcile a node if one of the two NNI

topologies reduces the number of mappings to the species tree (we have never observed a

case where both NNI topologies reduce this number). In this case, $m = 2$, and we now carry

out another NNI move on the updated gene tree. Only one move needs to be examined at

this point, however, since the back-NNI rearrangement would return us to the starting

state. The cost of all of these NNI moves is then compared to the cost of duplication and loss

at the end of the round. Duplication+loss naturally allows for more than one of each type of

event for a local reconciliation, and is therefore not handled differently when $m > 2$.

Finally, at the end of a round, `reconcILS` will sum the cost of all events in order to

choose the optimal local reconciliation. If there are equally optimal reconciliations,

`reconcILS` will report the duplication+loss solution.

Labeling events

Our approach for labeling ILS on reconciled gene trees is described above in the

"New Approaches" section. Here, we consider two important caveats to this approach, one

perhaps more obvious than the other. First, a specific labeling is associated with a specific

reconciliation. That is, if we had found a different optimal reconciliation for the tree in

Figure 3a (for instance, one duplication and two losses), we would have had different

labeled events. Therefore, a labeling is always reported in the context of a particular

reconciliation.

Second, as mentioned, for some ILS events it is not always clear which of the two

nodes flanking a discordant branch to label with I: this is why there is no exact map

between gene tree node labels and species tree nodes in the presence of ILS. For a

discordant branch, we have arbitrarily set `reconcILS` to always choose the node closest

to the tips as I. In some cases, this choice should not matter very much. Consider the toy

reconciliation in Figure 2b—should we have placed the NNI event on node G_1 or on node

G_2? In this case the choice does not seem to imply any difference in evolutionary histories.

In contrast, consider the reconciliations shown in Figure 3b and 3c—these represent the

output of `reconcILS` applied to the gene trees in Figure 1d and 1e, respectively. While we

have left the daughter tree branch in red for clarity, it is important to note that these trees

are identical topologically, which is why `reconcILS` has labelled them in exactly the same

way. The algorithm has no way to "see" the red branches, and has consequently simply

picked the bottom-most node in both cases as the one to label as having undergone ILS and

duplication, even though this is not precisely correct in Figure 3c. At the moment, this is a

limitation of `reconcILS` (and all methods attempting to label gene trees with ILS events),

but we mention other possible solutions in the Discussion.

Primate data

We used two sets of gene trees that were previously constructed in Smith et al.

(2022) using IQ-TREE2 (Minh et al. 2020b), based on data from Vanderpool et al. (2020).

The first set consists of 1,820 single-copy orthologs; this set was denoted as "Single-copy

clusters" in Smith et al. (2022). The second set consists of 11,555 trees containing both

orthologs and paralogs; this set was denoted "All paralogs" in Smith et al. (2022). We

randomly resolved all polytomies. When using `reconcILS`, costs were set to $C_D = 1.1$,

$C_L = 1$, and $C_I = 1$.

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Data availability

`reconcILS` is available at https://github.com/smishra677/reconcILS. `dupcoal` is available at https://github.com/meganlsmith/dupcoal.

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