reconcILS: A gene tree-species tree reconciliation

algorithm that allows for incomplete lineage sorting

Sarthak Mishra[1]*, Megan L. Smith[2], Matthew W. Hahn[1,3]

[1]Department of Computer Science, Indiana University, Bloomington, Indiana 47405

[2]Department of Biological Sciences, Mississippi State University, Starkville MS 39762

[3]Department of Biology, Indiana University, Bloomington, Indiana 47405

*Corresponding author. E-mail: samishr@iu.edu

Abstract

Reconciliation algorithms provide an accounting of the evolutionary history of individual gene trees given a species tree. Many reconciliation algorithms consider only duplication and loss events (and sometimes horizontal transfer), ignoring effects of the coalescent process, including incomplete lineage sorting (ILS). Here, we present a new heuristic algorithm for carrying out reconciliation that accurately accounts for ILS by treating it as a series of nearest neighbor interchange (NNI) events. For discordant branches of the gene tree identified by last common ancestor (LCA) mapping, our algorithm recursively chooses the optimal history by comparing the cost of duplication and loss to the cost of NNI and loss. We demonstrate the accuracy of our new method, which we call `reconcILS`, using a new simulation engine (`dupcoal`) that can accurately generate gene trees produced by the interaction of duplication, loss, and ILS. Despite being a heuristic method, we show that `reconcILS` is much more accurate than models that ignore ILS, and at least as accurate or better than leading methods that can model ILS, while also being able to handle much larger datasets. We demonstrate the use of `reconcILS` by applying it to a dataset of 23 primate genomes, highlighting its accuracy compared to standard methods in the presence of large amounts of ILS.

Introduction

The sequencing of a large number of genomes from many different species has highlighted the dynamic nature of genes: they are often duplicated, lost, or moved between species via introgression (no gain of an extra copy) or horizontal gene transfer (gain of an extra copy). One of the main goals of evolutionary and comparative genomics has been to quantify this dynamism, both in terms of the number of events of each type and their timing. A popular approach for quantifying both measures is gene tree-species tree reconciliation (reviewed in Boussau and Scornavacca 2020). Given a gene tree topology and a species tree topology, reconciliation algorithms attempt to infer the evolutionary events necessary to explain any discordance in the respective topologies. Many different reconciliation algorithms have been developed, most often considering only duplication and loss (so-called "DL" models; e.g. Goodman et al. 1979; Page 1994; Guigo et al. 1996; Page and Charleston 1997; Chen et al. 2000; Zmasek and Eddy 2001; Durand et al. 2006), but also sometimes considering duplication, transfer, and loss ("DTL" models; e.g. Hallett and Lagergren 2001; Hallett et al. 2004; Bansal et al. 2012; Stolzer et al. 2012; Szöllősi et al. 2013; Jacox et al. 2016; Morel et al. 2020).

Despite the success—and widespread use—of DL and DTL reconciliation algorithms, they do not consider gene tree discordance due to the coalescent process. The coalescent is a model that describes how sequences within a population are related (Kingman 1982; Hudson 1983; Tajima 1983), and is relevant even when considering sequences from different species because these must also coalesce in shared ancestral populations. Importantly, gene trees and species trees can be discordant solely due to coalescence, in a process often called incomplete lineage sorting (ILS) or deep coalescence (Maddison 1997). Standard DL and DTL algorithms will incorrectly infer one duplication and three losses for every discordance due only to ILS (Hahn 2007), resulting in misleading results when applying reconciliation methods to trees with ILS (e.g. Stull et al. 2021). Duplication, loss, and coalescence can also interact in complex and non-intuitive ways (Rasmussen and Kellis 2012; Mallo et al. 2014; Li et al. 2021), making it almost impossible to simply exclude gene trees discordant due to ILS from an analysis.

A few different approaches to reconciliation have been used to accommodate

discordance due to coalescence. An early approach allowed short branches in the species

tree—which can lead to ILS—to be collapsed (Vernot et al. 2008; Stolzer et al. 2012). The

use of non-binary species trees means that such algorithms will not mistake ILS alone for

duplication and loss; however, they do not account for all effects of coalescence on

incorrect inferences of duplication and loss (see below). Chan et al. (2017) introduced a

reconciliation algorithm that included duplication, loss, transfer, and ILS, but it assumed

that new duplicates were completely linked to their parental locus (i.e. no recombination

between them). Given that the majority of new gene duplicates are not completely linked

(Schrider and Hahn 2010), such an assumption limits the number of genes that can be

analyzed. Wu et al. (2014) developed a parsimony-based algorithm (DLCpar) that models

duplication, loss, and ILS; this approach is based on the probabilistic three-tree model of

Rasmussen and Kellis (2012), but runs faster and without the need to specify duplication

and loss rates ahead of time.

Here, we introduce a new heuristic method for finding the most parsimonious

reconciliation of a gene tree and a species tree in the presence of duplication, loss, and ILS.

Our goal is to improve upon existing methods by increasing speed and accuracy, as well as

by incorporating more coalescent processes (i.e. ILS is not the only cause of discordance

due to the coalescent). In addition, although DLCpar has been shown to perform well on

smaller trees (Wu et al. 2014; Du et al. 2021), it is not straightforward to include its

algorithm within other tools that also have reconciliation steps (e.g. Thomas et al. 2017;

Zhang et al. 2020) or to run it on larger datasets. We hope that the algorithm introduced

here will be easily adaptable to such scenarios. We demonstrate the use of this new

algorithm by applying it to 23 whole genomes from primates, a dataset that has previously

been shown to contain a large amount of discordance due to ILS (Vanderpool et al. 2020).

Figure 1. The multilocus multispecies coalescent model generates full gene trees. *a) A gene tree at the parent locus (blue) is drawn from the species tree (outlined in black) via the multispecies coalescent (MSC) model. A duplication event (red star) is drawn from a birth-death model over the species tree, in this case occurring in the population ancestral to species B and C. b) A gene tree at the daughter locus is drawn from the MSC. The duplication is placed on this gene tree at the time at which it occurred in the species tree (shown as a dotted line), choosing one of the two possible branches at random. The inserted DNA now takes on the gene tree branch labeled in red. c)-g) Possible ways that the parent and daughter loci coalesce to form the full gene tree. In all panels the star and red branch below it come from the daughter gene tree, and the blue tree comes from the parent gene tree. The coalescent process joining the two trees is shown as a red branch above the star. The trees in the bottom row are simply unfolded views of the full gene trees.*

New Approaches

Simulation using `dupcoal`

We begin with a description of the simulation program introduced here, `dupcoal`,
as it allows us to explain the biological model underlying both the simulations and

inferences. The explanation also makes clear which common processes are not modeled by

DL and DTL methods. Our simulations are based on the multilocus multispecies coalescent

(MLMSC) model of Li et al. (2021). This model treats each individual locus as evolving via

the multispecies coalescent (MSC) model, in which discordance between individual gene

trees and the species tree can arise only via incomplete lineage sorting. The MLMSC further

models relationships among these loci via duplication and coalescence, in addition to losses

at any locus.

To illustrate how duplications and losses occur, and how relationships among loci

are generated in dupcoal, we use a simplified example (a full description of the

simulation algorithm can be found at https://github.com/meganlsmith/dupcoal). As a first

step, a gene tree is generated via the MSC at the "parent" locus (Figure 1a); this tree exists

regardless of whether subsequent duplications occur. Duplications and losses are then

generated via a birth-death process on the species tree, with each event occurring at a

specific time on a specific branch of the species tree (one such duplication is represented as

a star in Figure 1a). For each duplication event, an additional gene tree is generated via the

MSC at a "daughter" locus (Figure 1b). This tree represents the history of the locus at which

the duplicated DNA will be inserted, and exists with or without this duplication event. In

other words, each locus has its own gene tree topology, and any sequence duplicated into a

locus will follow its topology. For now, we assume that this locus is unlinked from the

parent locus, so that the topologies are completely independent. The mutation representing

the duplication event is then placed on the gene tree at the daughter locus at the time at

which it occurred on the species tree—if multiple branches of the daughter gene tree exist

at this point in time on the relevant branch, the mutation is assigned to one at random (star

in Figure 1b). Regardless of where exactly the mutation occurs on the daughter gene tree,

the duplication now takes on the topology and branch lengths of the tree below this point

(red branch in Figure 1b). Losses are also simulated on the species tree and placed on each

gene tree in a similar manner (though no losses are shown in Figure 1, for simplicity).

When loss events occur, they are placed on branches of gene trees (either parent or

daughter trees), with all branches below such mutations removed. We use both a forward

algorithm (the birth-death process) and a backward algorithm (the coalescent) in order to

accurately model the full gene tree containing all duplications, losses, and coalescent

events.

The process described thus far in our example has produced two gene trees—one at

the parent locus (Figure 1a) and one at the daughter locus (Figure 1b). Either or both of

these two trees may be concordant (as shown) or discordant due to ILS. Because gene

duplication is fundamentally a process by which an existing haplotype in a population at

one locus is inserted into another location, the inserted DNA at the new daughter locus

carries with it a history of coalescence with all of the other haplotypes at the parent locus.

This coalescent process joining the two trees can cause additional discordance, even among

gene trees that individually cannot be discordant (e.g. when the species tree has only two

species). We illustrate this additional key coalescent process with an example in Figure 1.

Figures 1c-g show the five outcomes possible when joining the parent and daughter

trees shown in Figures 1a and 1b. Figure 1c is the simplest outcome, as the daughter tree

coalesces with the parent tree on the same branch on which the mutation occurred (i.e. the

one leading to species C); this is the tree that is assumed to be produced by duplication

under standard DL reconciliation. Figures 1d and 1e represent cases in which the daughter

tree joins the parent tree on a different branch, each of which would require one nearest

neighbor interchange (NNI) move to recover the tree topology in Figure 1c. Finally, Figures

1f and 1g represent more extreme cases, in which coalescence between parent and

daughter further up the tree requires two NNI moves to recover the topology in Figure 1c.

It should be emphasized here that all of the topologies presented in Figure 1 involve

only a single duplication and no losses, and neither the parent tree nor the daughter tree is

itself discordant due to ILS. In fact, the examples in Figures 1d and 1e are not technically

due to ILS either, since coalescence is occurring in the same population of the species tree

(the ancestral population of species B and C). This same dynamic in the ancestral

population also arises when the species tree only contains two species—this is why

approaches that simply collapse internal branches of a species tree will not eliminate

incorrect reconciliations due to coalescence. In addition, note that, in the scenario shown

here, the trees in Figures 1d and 1e are equiprobable with the topology in Figure 1c,

making even probabilistic reconciliation difficult. These simple examples demonstrate that
the vagaries of the coalescent process lead to a thicket of different tree topologies.

Parent and daughter trees can also be discordant if any of the locus gene trees do
not match the species tree. In such cases, a further complication can arise: "hemiplasy" or
"copy-number hemiplasy" (Avise and Robinson 2008; Rasmussen and Kellis 2012; Hahn
and Nakhleh 2016). Copy-number hemiplasy occurs when either a duplication or a loss
mutation occurs on a discordant branch of a gene tree (i.e. a branch that does not exist in
the species tree). While loss events can cause copy-number hemiplasy when they occur on
a discordant parent tree (even in the absence of duplication), duplication events can only
cause copy-number hemiplasy on discordant daughter trees. Such events are not simulated
by SimPhy (Mallo et al. 2016), a popular platform for carrying out gene tree simulations
with duplications and losses, but are simulated by `dupcoal`. Copy-number hemiplasy can
cause further complications for reconciliation, and so are an important phenomenon to
include (see Results).

Together, the processes described here make up the steps by which our simulator
can generate full gene trees: i.e. ones with the parent and all daughter trees joined together.

Reconciliation using `reconcILS`

Overview of main algorithm

Our reconciliation approach uses nearest-neighbor interchange (NNI)
rearrangements to model ILS and coalescence events. NNI has been used previously in
reconciliation algorithms to correct gene trees that may have been inferred incorrectly (e.g.
Chen et al. 2000; Chaudhary et al. 2011; Nguyen et al. 2013). Here, we use NNI because the
effects of ILS on gene trees can be modeled as a series of NNI rearrangements (Than and
Rosenberg 2013). For three species undergoing ILS, there are two possible discordant
topologies, each of which is one NNI move away from the species tree topology (e.g. Figure
2). For four lineages undergoing ILS, there are 14 possible discordant topologies, each of
which are between one and three NNI moves away from the species tree topology. (Note
that it is the number of lineages undergoing ILS at a specific point on the tree that matters,

not the total number of tips in a tree.) Our algorithm takes advantage of this relationship

between NNI and ILS in order to reconcile gene trees, while also considering duplication

and loss.

Figure 2. Summary of `reconcILS`. *a) The initial step generates an LCA mapping function*

between the gene tree and the species tree. Here, nodes G_1 and G_2 in the gene tree both map to

node S_1 in the species tree; we therefore refer to S_1 as a multiply mapped node. Additionally,

G_1 and G_2 together define a discordant branch, K_1, in the gene tree. For clarity, all mappings

involving tip nodes have been excluded. b) The algorithm attempts to reconcile the discordant

branch using NNI and loss. Here, reconciliation can be carried out with 1 NNI and 0 losses. c)

The algorithm attempts to reconcile the discordant branch using duplication and loss. Here,

reconciliation can be carried out with 1 duplication and 3 losses. `reconcILS` must decide

between the reconciliations in b) and c) for every discordant branch.

An overview of `reconcILS` is given in Algorithm 1 (see Supplementary Materials).

Our approach combines multiple sub-approaches that are commonly used in DL

reconciliation and other phylogenetic inferences. As a first step, we use last common

ancestor (LCA) mapping (Zmasek and Eddy 2001) to create a mapping function between

each node in G and a node in S (Figure 2a). We refer to each of these pairwise relationships

(arrows in Figure 2a) as "mappings" from here on.

Unlike in standard DL methods, LCA mapping here does not represent a final

reconciliation. Instead, this step identifies cases in which multiple nodes in the gene tree

map to an identical node in the species tree. For instance, in Figure 2a nodes G_1 and G_2 in

the gene tree both map to node S_1 in the species tree. We therefore call S_1 a "multiply

mapped" node. Additionally, we refer to the branch in the gene tree bracketed by nodes G_1

and G_2 as a "discordant" branch (labeled K_1 in Figure 2a); such branches do not exist in the

species tree. The focus of our method is in determining whether discordant branches

should be resolved via NNI and loss (Figure 2b) or via duplication and loss (Figure 2c).

Choosing among events (NNI, duplication, and loss) is carried out greedily for each branch,

with no consideration of a globally optimal set of events among all discordant branches in

the gene tree. Consequently, after choosing a set of events for one branch (given specified

costs), the process repeats itself until there are no multiply mapped nodes and no

discordant branches. Duplications implied by multiply mapped tip nodes in the species tree

are inferred via standard methods; inferences of losses are explained in the Methods.

As with all parsimony-based reconciliations, reconcILS requires a cost for each

event type: one for duplications (C_D), one for losses (C_L), and one for NNIs (C_I).

Reconciliations can be very sensitive to costs, such that maximum parsimony solutions

under one set of costs can be very different than under another set of costs. Although there

are few ways to determine costs independently (but see Discussion), there are some

approaches for finding the Pareto-optimal costs within a reconciliation context (e.g.

Libeskind-Hadas et al. 2014; Mawhorter et al. 2019). Here, as in most commonly used

reconciliation algorithms, we allow the user to set costs. The default costs within

reconcILS are set to $C_D = 1.1$, $C_L = 1$, and $C_I = 1$.

Labeling a reconciled gene tree

A full reconciliation of a gene tree, G, and species tree, S, will have reconciled all

multiply mapped nodes, leaving no discordant branches. The total number of duplications,

losses, and NNIs required for this reconciliation is the sum of each event type across all
branches; these numbers are reported by `reconcILS`. We also aim to label the gene tree
with these events—identifying where each event occurred in history—as this is an
important output of reconciliation algorithms. As `reconcILS` traverses the species tree
finding optimal local reconciliations for each branch, it also records the number and
location of all events. Labeling therefore consists of placing these events on the gene tree.
Traditional DL reconciliation algorithms label gene tree nodes as either   (for duplication)
or   (for speciation); they may also label the location of losses,   . Here, we additionally aim
to label the NNI events induced by ILS (I).

Importantly, there is no standard way to label ILS events on a tree, largely because
there is no exact map between gene tree nodes and species tree nodes under ILS. A branch
undergoes NNI (and ILS), not a node, so it is not clear which node flanking such a branch
should be chosen to be labelled. This labeling problem therefore differs fundamentally
from labeling under DL and DTL algorithms because of the lack of a one-to-one map
between nodes in the two trees. Here, we have created our own labeling system for ILS, one
that we think is both understandable and easy to work with.

Figure 3. Labeling reconciled gene trees. *a) One possible reconciliation of the gene tree*
from Figure 1f labeled by `reconcILS`*. The single ILS node ( ) has been labeled with a branch*
that has been rearranged by NNI (   ), the number of NNIs it has undergone ( ), and the fact
that it also represents a duplication ( ). b) The gene tree from Figure 1d labeled by
`reconcILS`*. This tree is correctly labeled, but note that it is the same topology as in panel c*
(see Methods for further explanation). c) The gene tree from Figure 1e labeled by
`reconcILS`*. Note that the program has labeled the bottom-most node flanking the*
discordant branch with   and , although that is not the true history. d) A gene tree with only

duplications and losses (L) labeled by reconcILS. *Internal branches on a tree can also be*
labeled with L, though none are shown here.

Our solution for placing the I label on nodes of gene trees in reconcILS is to
denote both a daughter branch affected and the number of NNI moves that were associated
with this branch. A few examples are shown in Figure 3, which also helps to illustrate why
these choices were made. Figure 3a labels the gene tree from Figure 1f—with more
complete tip names—with output from reconcILS. As can be seen, this tree has one node
(denoted with a small black dot for clarity) labeled with both duplication and ILS. Further,
the labeling tells you which daughter branch was reconciled with NNI (C_1), and how many
NNI moves were required to reconcile it (2). Without such information, simply labeling the
node with I would not have distinguished histories involving coalescent events along the A
branch from those along the C_1 branch. Further details of this labeling are provided in the
Methods, and caveats are covered in the Discussion.

reconcILS also labels duplications and losses in the absence of ILS. Figure 3d
shows how both individual duplications and losses are labelled: observe that neither
requires a label with a specified number of events (like the I label), nor does either need to
specify on which branch they act. reconcILS places the label L on any node in the implied
gene tree that is not represented by a gene copy; for example, generic tip nodes A, C, and B
in Figure 3d do not exist in the input gene tree itself, but are labelled with an L.
reconcILS can do this because every duplication event is represented internally by
generating a new copy of the species tree below the duplicated node—if losses occur on
these implied trees, we can track their timing as well. Note that internal nodes can also be
labelled with an L, so that the timing of loss events must be inferred by comparing the state
of parent and daughter nodes (i.e. losses on internal branches will lead the descendant
internal node and all of its daughter nodes to be labelled with an L). Finally, any node
without a label can be inferred to be a speciation node.

Results

Accuracy and advantages of `dupcoal`

We assessed the accuracy of `dupcoal` by first simulating 1000 gene trees along a 40-tip species tree with no duplication and no loss (Supplementary Figure S1). These simulations therefore only model ILS as a cause of discordance. We compared the degree of discordance in these simulations to ones carried out using the same species tree with SimPhy (after appropriate branch-length scaling; see Methods). We found a very high correlation between concordance factors on each branch calculated from gene trees produced by the two methods (ρ=0.99, P=1.13 x 10^{-30}), indicating that we are accurately simulating ILS. We also carried out simulations in `dupcoal` with both duplication and loss, again comparing our results to SimPhy (after scaling duplication and loss rates appropriately; see Methods). Again, the average number of duplications and losses per simulated gene tree (n=1000) were highly similar and not significantly different between the two methods: 1.81 vs. 1.77 duplications and 1.78 vs. 1.84 losses for `dupcoal` and SimPhy, respectively (P=0.13 for duplications and P=0.29 for losses; t-tests), indicating that we are accurately simulating duplications and losses in `dupcoal`.

To highlight additional features contained within `dupcoal`, we also tracked the number of simulated gene trees in which copy-number hemiplasy occurred (this is a standard output reported by `dupcoal`). Out of 1000 gene trees simulated on the 40-tip tree with duplication, loss, and ILS, 246 contained at least one hemiplasy event. As a straightforward way to demonstrate the effect of such events, we separately inferred the number of duplications and losses on gene trees with and without copy-number hemiplasy using ETE3 (Huerta-Cepas et al. 2016). While ETE3 implements a DL reconciliation algorithm that does not model any type of ILS (see next section), this contrast shows the effects of hemiplasy above and beyond the effects of ILS. As expected, both the number of inferred duplications (1.7 excess duplications; P=6.3 x 10^{-6}, t-test) and the number of inferred losses (5.4 excess losses; P=6.7 x 10^{-7}, t-test) were higher among the trees containing hemiplasy. These results emphasize the importance of having a simulation engine that can simulate hemiplasy.

Accuracy and advantages of `reconcILS`

As a simple test of DL reconciliation algorithms—which do not allow for ILS—we
used the DL method implemented in ETE3 (Huerta-Cepas et al. 2016). We initially ran
simulations in `dupcoal` without duplication or loss (i.e. ILS only); this allows us to see
incorrectly inferred duplications and losses more easily. As expected, on a species tree with
only 3 tips, ETE3 always inferred 1 extra duplication and 3 extra losses on gene trees with
discordance due to ILS (not shown). On a larger species tree with 40 tips, ETE3 inferred a
large excess of duplications and losses when only ILS is acting (9.5 duplications and 65.0
losses per gene tree, on average). On datasets simulated on a 40-tip tree with duplication,
loss, and ILS, the accuracy of ETE3 is low for losses (ρ=-0.04) and fairly good for
duplications (ρ=0.73) (Supplementary Figure S2).

In contrast, `reconcILS` accurately infers the numbers of duplications and losses
(Figure 4a and 4b), as well as ILS events (Figure 4c) on the 40-tip tree. The correlation
between inferred and simulated duplications and losses were high (ρ=0.95 and 0.65,
respectively). We also observed a high correlation between the inferred and simulated
number of NNI events needed to reconcile a gene tree (ρ=0.76). ILS immediately followed
by loss will not leave a signal in the resulting gene trees, leading to a slight undercounting
of NNI events caused by ILS.

As we did with ETE3 in the previous section, we also contrasted the number of
duplications and losses inferred by `reconcILS` on simulated gene trees with and without
copy-number hemiplasy. Unfortunately, `reconcILS` also does not deal perfectly with
hemiplasy, though in a slightly different way than ETE3. Rather than overestimating both
the number of duplications and losses, `reconcILS` appears to generally convert true ILS
events into losses. This occurs because (for instance) a duplication on a discordant
daughter gene tree branch will duplicate a non-sister pair of lineages (e.g. *A* and *C*), which
`reconcILS` interprets as a duplication and a loss (of *B*, in this case) rather than as a
duplication and an NNI event. Consistent with this, we observed very few excess
duplications in the simulations with hemiplasy (0.26 extra duplications, on average), but

significantly more losses (1.45; P=3.8 x 10^{-26}, t-test) and significantly fewer NNI events

(1.66 P=3.5 x 10^{-7}, t-test) compared to simulations that did not have hemiplasy.

347
Figure 4. Accuracy of `reconcILS` on data simulated from the 40-tip tree. *a)*

Number of inferred versus true duplication events. b) Number of inferred versus true loss

events. c) Number of inferred versus true ILS events. Circle size is proportional to the number

of reconciliations with each value, with jitter applied for clarity. Orange line represent best-fit

regression (as calculated in Seaborn; Waskom 2021), plus confidence intervals. The dashed

line represents y=x. See Supplementary Figure S2 for results from ETE3 on the same data.

We also compared the results from `reconcILS` with another reconciliation method

that allows for ILS, DLCpar (Wu et al. 2014). DLCpar was written in Python 2, and—

possibly because we had to run it in a virtual environment—could not run on the larger

gene trees produced by the 40-tip tree. Therefore, we compared the performance of

`reconcILS` and DLCpar on 1000 gene trees simulated on the 3-tip species tree. Overall,

the accuracy of both methods was high for duplication (`reconcILS`, ρ=0.89; DLCpar,

ρ=0.89; Supplementary Figure S3a). The correlation between simulated and inferred ILS

events was ρ=   for `reconcILS` and ρ=   for DLCpar (Supplementary Figure S3b). It

is important to note that DLCpar infers the number of ILS events, rather than the number of

NNI rearrangements that occur within such an event (which are what is inferred by

`reconcILS`). To ensure that a fair comparison can be made, results reported for DLCpar

therefore are simulated vs. inferred ILS events. In both cases the methods are doing well in

their respective inferences, but with slightly higher accuracy for `reconcILS`.

There is quite a difference between reconcILS and DLCpar in the inference of
losses (correlations of ρ=0.67 and ρ=0.27, respectively; Supplementary Figure S3c). Upon
closer inspection, the lower accuracy in DLCpar is due to many loss events of one particular
type not being counted. Specifically, DLCpar does not count losses that occur above the root
of the gene tree being considered. This means that, for instance, in gene trees with the
topology (B,C), DLCpar does not count the loss that occurred along the A branch (assuming
the species tree in Figure 1). We confirmed this behavior on slightly larger trees in DLCpar,
as well as in all trees in ETE3—no losses above the root of a gene tree are considered. The
difference in counting of losses among algorithms arises because reconcILS explicitly
traverses the species tree, while the other methods do not. This enables our method to
track losses that occur above the root of the input gene tree, but below the root of the input
species tree. However, there may be some cases in which the behavior of DLCpar and ETE3
is preferred. For instance, since gene families must originate on some branch of the species
tree (e.g. Richter et al. 2018), forcing reconciliation algorithms to infer losses on branches
that pre-date this appearance would be incorrect. It is not clear which approach to counting
losses one should prefer when analyzing real data.

Finally, we assessed the runtime and memory requirements of reconcILS
compared to ETE3 on a single node of the Indiana University Research Desktop, running an
Intel Xeon processor with 100 GB of RAM (DLCpar can only be run in Python 2, which
makes comparisons highly unfair). On a 3-species tree, both programs are extremely fast
(average time of 0.015 seconds per gene tree for both; Supplementary Figure S4a) and both
use very little memory (average peak memory per gene tree of 0.17 MB for reconcILS
and 0.075 MB for ETE3; Supplementary Figure S4b). Both programs remain very fast on
data produced from a 40-species tree, though the faster speed of ETE3 is also evident (5.02
seconds per gene tree for reconcILS and 1.60 seconds for ETE3; Supplementary Figure
S4a). Regardless, this means that the entire 1000-gene-tree simulated dataset on the 40-
species tree took less than 84 minutes to analyze using reconcILS on a moderately
powerful computer. Similarly, the memory requirements of reconcILS increase on the
40-tip tree (16.88 MB per gene tree for reconcILS and 2.93 MB for ETE3; Supplementary
Figure S4a), but are not onerous for modern computers. As the 40-species tree can contain

many more events (duplication, loss, or ILS), we also plotted the average time and memory

required for trees containing different numbers of total events (Supplementary Figure

S4c,d). Both programs show linear increases in time and memory with the number of

events, which is promising for analyzing larger datasets with larger species trees.

Analysis of primate genomes

To further demonstrate the power and accuracy of `reconcILS`, we applied it to a

dataset of gene trees inferred from 23 primate genomes (Vanderpool et al. 2020; Smith et

al. 2022). These genomes all come from the suborder Haplorhini, which includes all

primates except the lemurs and lorises (suborder Strepsirhini). Among these genomes, we

analyzed two sets of gene trees that were previously inferred: First, we considered 1,820

single-copy orthologs. This dataset is not expected to contain any duplicates, though it does

contain losses. However, because there are high levels of ILS among the primates sampled

here, the dataset serves as an ideal comparator for reconciliation approaches with and

without ILS. Second, 11,555 trees containing both orthologs and paralogs were used to

infer duplications, losses, and ILS among the primates. We used both `reconcILS` and

ETE3 to analyze the two datasets, assigning inferred events to each branch of the primate

tree.

Single-copy ortholog dataset

Results obtained from analysis of only the single-copy orthologs are shown

graphically in Figure 5a (see Supplementary Figure S5a for numerical values on each

branch). The pattern that stands out from this graph is that ETE3 is vastly overestimating

the number of duplications and, especially, the number of losses. Because ETE3 does not

model ILS, all such events must be reconciled by one duplication (placed above the

discordant branch) and three losses (placed below the discordant branch; Hahn 2007). We

can see this effect in the data by looking at the correlation between the gene concordance

factor (gCF) estimated for a branch with the number of duplicates inferred on its parent

branch. Gene concordance factors quantify the fraction of loci that contain the same branch

as the species tree; when single-copy genes are analyzed, low gCFs can indicate

discordance due to ILS (Baum 2007; Lanfear and Hahn 2024). The correlation between

gCFs for these data (estimated in Smith et al. 2022) and duplications inferred by ETE3 on

parent branches is ρ=-0.56. Lower gCFs lead to higher numbers of duplicates, as expected

when ILS is not accounted for—this problem is not unique to ETE3, but should be expected

for all standard DL and DTL approaches. Such errors can lead to incorrect inferences of

bursts of duplication and loss.

Figure 5. Results from primates. *a) Analysis of 1,820 single-copy orthologs. Both*

`reconcILS` *and ETE3 were used to analyze the data, with duplication, NNI, and loss events*

mapped to each branch of the species tree. On each branch, the bar plots show results from

`reconcILS` *on top, with results from ETE3 mirrored below. ETE3 cannot infer NNI events, so*

this bar is always set to 0. b) Analysis of 11,555 gene trees containing both orthologs and

paralogs. Results are presented in the same manner as in panel a. However, note that the

height of all bars is normalized within a panel, so that heights of bars within a tree can be

compared among branches, but the heights of bars between the trees cannot be compared.

Supplementary Figure S5 shows the absolute counts of all events from both panels and

Supplementary Figure S6 shows species tree with branch lengths in substitution units rather

than as a cladogram, as shown here.

Using the same reasoning, we can assess the accuracy of `reconcILS` by examining

the correlation between NNI events inferred by our algorithm and gCF values for single-

copy genes. High NNI counts indicate more ILS, and therefore this measure should be

negatively correlated with gCF values for the same branch. Indeed, the correlation between

NNI counts and gCF values is ρ=-0.88, demonstrating the high accuracy of `reconcILS`.

However, `reconcILS` does sometimes infer a small number of duplications (Figure 5a;

Supplementary Figure S5a). While it is possible that some of the single-copy genes we

analyzed represent so-called pseudoorthologs—and therefore do contain actual

duplication events—these are expected to be rare (Smith and Hahn 2022). Instead, these

false duplicates are likely due to ILS among four or more lineages. Recall that, although our

algorithm has been developed to account for rearrangements due to the coalescent process,

ILS among four lineages can result in gene trees that are three NNI moves away from the

species tree; ILS among more lineages will produce trees that are even further away.

Therefore, multiple NNI events are sometimes simply more costly than duplication and

loss, even when ILS is the true cause of discordance.

Full gene tree dataset

We also examined patterns of ILS, duplication, and loss for the entire primate dataset. The results from the full set of gene trees (Figure 5b; Supplementary Figure S5b) largely mirror the results from the single-copy orthologs (which are a subset of the entire dataset). Part of the apparent similarity is simply due to normalizing the two datasets separately, but several non-graphical factors also contribute to the pattern. First, although there are many true duplicates in the full dataset, the vast majority of gene trees only have events on tip branches: 7,693 out of 11,555 gene trees have duplicates specific to one species (Smith et al. 2022). Another ~1,000 trees have duplicates specific to two species. Combined with the 1,820 single-copy genes, it becomes clear that there are in fact relatively few trees with duplicates on internal branches even in the full dataset.

Second, errors in inferring duplications and losses due to the coalescent process still dominate the results from standard DL reconciliation (i.e. ETE3), possibly even more so than in the single-copy dataset. Because ILS (and introgression) can affect all gene trees, trees with duplicates can experience twice as much discordance when there are twice as many branches. One example of this can be seen among the Papionini (macaques, baboons, geladas, and mandrills): ETE3 has to infer >6,000 losses on each of the tip-branches in this clade, and more than 10,000 losses on most of the internal branches (Figure 5b; Supplementary Figure S5b). There are also many thousands of duplicates inferred on internal branches of this subclade, though these species have the same number of total protein-coding genes as all the others analyzed here (Vanderpool et al. 2020). Although the inferred duplications and losses in this group were also relatively higher in the single-copy dataset, it is clear that even a small number of true duplicates can have an outsized effect on methods that cannot account for ILS.

The analysis using reconcILS is also highly similar between the two datasets, again largely because most duplicates are confined to tip branches. reconcILS is able to deal with discordance due to the coalescent, even in the presence of duplication: the correlation between NNI events inferred on each branch of the species tree from the single-copy orthologs and the full dataset is $\rho = 0.99$. Although the numbers of events are of

490 course larger with the larger dataset (Supplementary Figure S5b), the effect of ILS on any
491 given branch of the species tree is experienced by all gene trees. Given its ability to deal
492 with ILS, duplication events across the primate tree are more reliably inferred with
493 `reconcILS`.

Discussion

We have introduced a new reconciliation algorithm and associated software
package (`reconcILS`) that considers discordance due to duplication, loss, and
coalescence. To better demonstrate the accuracy of our method, we have also developed a
new simulation program (`dupcoal`) that generates gene trees while allowing these
processes to interact. Our results demonstrate that `reconcILS` is a fast and accurate
approach to gene tree reconciliation, one that offers multiple advantages over alternative
approaches, even those that also consider ILS (e.g. DLCpar; Wu et al. 2014). Nevertheless,
there are still several caveats to consider, as well as future directions to explore.

Despite our demonstration of good performance in practice, our method is heuristic:
we have no theoretical proof that our method finds most parsimonious reconciliations.
While such proofs often require strict assumptions—e.g. the proofs for DL algorithms
assume there is no ILS—they are very helpful in providing confidence under the specified
conditions. However, given the high accuracy of `reconcILS` demonstrated with both
simulations and empirical data from single-copy genes, we hope the heuristic nature of our
algorithm is not a barrier to its utility.

As mentioned earlier, there is often ambiguity in how to label nodes in gene trees
affected by ILS and duplication (e.g. Figures 3b and 3c). This is due in part to the nature of
ILS—it is not clear which node to label because there is not a one-to-one map between
gene tree nodes and species tree nodes—and in part to the intrinsic lack of information
contained only within tree topologies when coalescence and duplication interact. The
difficulty in labeling is not confined to this work: DLCpar does not label nodes, and
RecPhyloXML, a format for representing reconciled trees, does not include a way to label
ILS events at all (Duchemin et al. 2018).

Nevertheless, labeling gene trees serves multiple important functions, and is often

interpreted as biological truth. For example, multiple methods use inferred gene tree labels

to identify orthologous genes (genes related by speciation events; Fitch 1970). The general

approach taken is to label nodes using reconciliation or similar approaches, treating any

node not labeled with a D as a speciation node; genes whose common ancestor is a

speciation node are subsequently defined as orthologs. Both standard DL reconciliation

and so-called "overlap" methods (e.g. Huerta-Cepas et al. 2007; Emms and Kelly 2019) will

consistently label the top-most node flanking a discordant branch with a duplication event.

This means that, in cases like the one shown in Figure 3b, these methods will label the

wrong node with a D, and subsequently identify the wrong pair of genes as orthologs.

(Figure 3c would be labeled correctly; recall that this outcome has an equal probability to

the one in Figure 3b in the scenario considered here.) Cases such as these have been

referred to as "unsorted homologs" (Mallo et al. 2014), and will occur any time duplication

and coalescence interact, either before or after a speciation event. They can also occur even

if only two species are analyzed (e.g. if only species B and C were considered in Figures 3b

and 3c), which means that approaches that simply collapse short branches to avoid ILS will

also be affected. Although `reconcILS` currently only labels one node with a D, in the

future we could allow for multiple possible labelings in order to emphasize the inherent

ambiguity of some labels.

The algorithm used by `reconcILS` deals, in part, with an additional interaction

between duplication, loss, and coalescence: hemiplasy. Hemiplasy is the phenomenon by

which a single mutational event on a discordant gene tree will be interpreted as multiple

events ("homoplasy") when reconciled to the species tree (Avise and Robinson 2008; Hahn

and Nakhleh 2016). In analyses of nucleotide substitutions, generic binary traits, and even

continuous traits, hemiplasy requires that mutations have occurred on a discordant branch

of a gene tree (Mendes and Hahn 2016; Guerrero and Hahn 2018; Hibbins et al. 2023). In

the context of either duplication or loss, such mutations would have to occur on a

discordant branch of a gene tree to lead to copy-number hemiplasy. The simulation engine

introduced here, `dupcoal`, simulates hemiplasy events and reports any gene trees

affected. Our results show that these events lead to increased error in reconciliation, and

are therefore an important process to include. Interestingly, such events have little effect
on species tree inference when paralogs are used for this purpose (Li et al. 2024).

Unfortunately, most copy-number hemiplasy events as defined above are still not
accurately inferred by `reconcILS`: we have found that our algorithm consistently turns
NNIs into losses when hemiplasy occurs. While the total number of events does not seem to
increase, it is not ideal that NNIs are mistaken for losses. However, there is also a broader
definition of copy-number hemiplasy in use, one that `reconcILS` does deal with
accurately (and is also simulated by `dupcoal`). This broader definition includes any
duplication or loss that does not fix in all descendant lineages, regardless of underlying
gene tree topologies (Rasmussen and Kellis 2012; Wu et al. 2014). While this definition
encompasses the stricter one given above, it also includes cases such as the one shown in
Figure 1, which the stricter definition does not include. Because the mutation in that figure
occurs in the common ancestral population of species B and C, but was only inherited by
lineage C, this scenario would be considered to be hemiplasy by the broader definition
(although it technically does not even involve ILS if the outcome resembles the gene trees
in Figure 1c-e, since coalescence is happening in the same population). The number of each
type of event in such cases is accurately inferred using `reconcILS`; DLCpar assumes that
this type of hemiplasy does not occur, and SimPhy does not simulate this type of hemiplasy,
either. Note, however, that if the duplication event (the star in Figure 1a) occurred a few
generations later—after the split between species B and C, along a tip lineage—the
scenario would not be considered hemiplasy by this broader definition, but would still
produce the same range of gene trees (i.e. Figure 1c-g). (This would now involve ILS,
however.) Regardless of the exact definitions and terms used, it should be clear that
duplication, loss, and coalescence can interact to produce a wide array of topologies not
considered by most reconciliation approaches.

The algorithm used by `reconcILS` deals with ILS differently than does DLCpar.
While DLCpar treats ILS as an event, `reconcILS` considers NNIs as events induced by ILS.
It is not clear that either approach should necessarily be preferred on first principles. In
simple cases involving three species and a single discordant branch (e.g. Figure 2), ILS and
NNI are more or less equivalent, though it should be noted that the process of ILS can also

lead to concordant trees and is not synonymous with discordance. However, in cases with
more than one discordant branch there is no simple relationship between the two. For
instance, in a 4-tip tree undergoing ILS, lineages may undergo ILS across two branches, but
result in between 0 and 3 NNI events (Than and Rosenberg 2013). We think that directly
modeling NNI—as is done in `reconcILS`—is a natural way to express the events needed
to reconcile a gene tree with a species tree.

One relatively straightforward improvement that could be made to `reconcILS` is
in the cost of different events. Costs must be specified in `reconcILS` ahead of time, but it
is difficult to determine biologically realistic costs for duplication, loss, and ILS. One
approach to determining duplication and loss costs would be to infer the number of such
events using an alternative, non-reconciliation approach (e.g. Mendes et al. 2020), setting
the cost in `reconcILS` to be inversely related to the inferred number of events of each
type. Varying rates of duplication and loss can also be inferred along different branches of
the species tree, and so the costs of these events could vary commensurately along the tree.
Similarly, we can envision a simple way to scale the costs of NNI with the length of species
tree branches: this is because the probability of discordance due to ILS is inversely
proportional to internal branch lengths on a species tree (Hudson 1983). We could
therefore make NNI events on longer branches of the species tree more costly, which would
properly weight the relative probability of such events locally on the tree.

There are also several extensions that could be made to `dupcoal`, our new
simulation engine. Currently, `dupcoal` draws a daughter gene tree from the multispecies
coalescent (MSC) at random: this implicitly assumes that the parent and daughter trees are
completely unlinked. It should be straightforward, however, to draw the daughter tree at a
given recombination distance from the parent tree, ranging from fully linked to fully
unlinked. In addition, though the software currently draws gene trees from the MSC, we
could also include introgression as a source of discordance by drawing trees from the
multispecies network coalescent (Meng and Kubatko 2009; Yu et al. 2011). In this case,
users would simply input a species network to simulate from, rather than a species tree.
We should also note the differences between the current version of `dupcoal` and
alternative simulators. As mentioned previously, SimPhy (Mallo et al. 2016) does not

simulate hemiplasy of any kind, as it uses the same three-tree model of Rasmussen and

Kellis (2012) and Wu et al. (2014). However, this method does include gene transfers,

which are not simulated by dupcoal. The MLMSC simulator of Li et al. (2021) simulates

both hemiplasy and transfers. dupcoal is therefore most similar to this method, with two

small differences. First, dupcoal does not change the rate of duplication and loss above

speciation events to account for ancestral polymorphism (cf. Gillespie and Langley 1979);

instead, users can simply extend all nodes $2N$ generations into the past to do so (where N

is the effective population size). Second, dupcoal does not account for the possibility of

further events occurring while duplications and losses are still polymorphic in a

population. In order to streamline the simulation process, we have chosen to ignore this

squared term.

Finally, we envision multiple ways in which the reconcILS algorithm could be

used in conjunction with other methods. For instance, ASTRAL-Pro (Zhang et al. 2020)

infers a species tree from gene trees containing paralogs by first labeling nodes in each

gene tree as either duplication or speciation. The authors of this software rightfully

recognize the problems with labeling discussed here, but nevertheless the labeling step

might benefit from methods that include ILS. Likewise, GRAMPA (Thomas et al. 2017)

allows for reconciliation of polyploidy events, but does not include coalescence as a source

of discordance; its algorithm could easily accommodate this extension. There are also

multiple approaches that can be used to correct gene trees via reconciliation (e.g. El-

Mabrouk and Ouangraoua 2017; Christensen et al. 2020). These and many other tasks

might benefit from approaches that model duplication, loss, and coalescence in a single

framework.

Methods

Building and testing dupcoal

The simulator dupcoal is written in Python 3 and uses functions from DendroPy

(Sukumaran and Holder 2010) to generate trees under the MSC and to manipulate parent

and daughter trees. It also uses functions from NumPy (Harris et al. 2020) to draw

coalescent times, duplication times, and loss times from exponential distributions.

To test the accuracy of `dupcoal` we simulated gene trees along a species tree with

40 tips (Supplementary Figure S1). The species tree topology was drawn at random using

the program ape (Paradis and Schliep 2018), with the "Grafen" method for drawing branch

lengths (scaled up by 10); this tree was used as input to both `dupcoal` and SimPhy.

Because `dupcoal` specifies branch lengths in coalescent units, while SimPhy uses time,

branch lengths were converted by assuming a population size of $N=10$ and a generation

time of 1 year. Simulations were run on this tree with only ILS (setting duplication and loss

rates to $\lambda = 0$ and $\mu = 0$, respectively). When using positive duplication and loss rates (see

next paragraph), they were scaled to the appropriate branch lengths to make them

comparable in `dupcoal` and SimPhy. Comparisons between `dupcoal` and SimPhy with no

duplication and loss were carried out by calculating gene concordance factors (gCFs) on the

species tree using IQ-TREE2 (Minh et al. 2020a).

To test the accuracy of inferences using `reconcILS` and ETE3, the same 40-taxon

tree was used for simulations in `dupcoal` with ILS, duplication, and loss ($\lambda = 0.03$ and

$\mu = 0.03$). DLCpar is written in Python 2 and could not be run in reasonable time on the 40-

taxon tree. We therefore also simulated data on a 3-taxon tree with topology

$(A{:}1.5,(B{:}0.5,C{:}0.5){:}1)$, where branch lengths are measured in coalescent units and

duplication and loss rates were the same as on the larger tree. The same costs were used

for all analyses of simulated data for all methods ($C_D = 1.1, C_L = 1, C_I = 1$).

Implementation of `reconcILS`

Inputs and outputs

Given a rooted, bifurcating species tree, S, and a rooted, bifurcating gene tree, G, as

input, `reconcILS` outputs the minimum number of duplications, losses, and NNIs

required to reconcile G and S given event costs. Inferred events are all also assigned to

nodes in the gene tree.

 Traversing trees

After generating an initial LCA mapping function between a gene tree and the

species tree, `reconcILS` performs a preorder traversal of the species tree. For each

species tree node, the algorithm checks how many gene tree nodes are mapped to it. If

there are fewer than 2 mappings, the algorithm invokes a function that asks whether

speciation or loss can be inferred. Nodes with 1 mapping are speciation events—these do

not incur a cost. Nodes that have 0 mappings represent losses. Our function asks whether

the parent node of a node with 0 mappings itself has 0 mappings. If not, a loss is inferred; if

so, no event is recorded, because the loss must have occurred further up the tree. If there

are 2 or more mappings the species tree node is multiply mapped, and a function to carry

out "local reconciliation" is called. Local reconciliation is described in more detail in the

next section, but here can simply be described as determining the events that have

generated the multiple mappings to a single species tree node.

If there are only two adjacent nodes in the gene tree mapped to a node in the species

tree (i.e. one discordant branch, as in Figure 2), then the method proceeds as described

below. However, when there are more than two nodes mapped to an internal species tree

node—and therefore more than one discordant branch in the gene tree—we must decide

which branch to reconcile with the multiply mapped node first. We have found cases where

the order in which the branches are resolved matters, with only one of the choices leading

to an optimal reconciliation. Algorithm 2 (see Supplementary Materials) describes an

algorithm for choosing which discordant branch to locally reconcile first. This algorithm

calculates the "bipartition cost" of each possible branch—the number of bipartitions and

losses induced by NNI moves applied to each branch (Supplementary Figure S7). Such a

calculation can be carried out quickly, and by choosing the branch with the lowest

bipartition cost, we can more accurately proceed with reconciliation.

Once reconciliation for a single multiply mapped node in the species tree is carried

out (i.e. it is no longer multiply mapped), our algorithm begins again: a new round. In each

round we generate an LCA mapping function and continue traversing the species tree,

counting gene tree nodes mapped to each species tree node.

 Choosing among events

In order for our algorithm to more efficiently and optimally choose among different

evolutionary events (e.g. Figure 2b vs 2c), we introduce the concept of a "local

reconciliation" for a multiply mapped node. Here, a species tree node is locally reconciled if

the number of gene tree nodes mapped to it, m, is reduced. For internal nodes of the

species tree, this, by definition, also reduces the number of discordant branches in the gene

tree.

The use of local reconciliation has several advantages. Most importantly, it provides

a criterion for moving forward: we can determine if a reconciliation has been achieved

within a single round of the algorithm. It also makes it clear that there are some scenarios

that can only be reconciled using duplication and not NNI (e.g. a multiply mapped tip node),

in which case choosing events is straightforward. In a single round of our algorithm, we

attempt to locally reconcile a single multiply mapped node using both NNI+loss and

duplication+loss (Figure 2).

Note that, given this definition, a species tree node can be locally reconciled by NNI

while still remaining multiply mapped; e.g., if $m = 3$ is reduced to $m = 2$, the node still

requires more reconciliation. In this case our algorithm will carry out up to m NNI moves

within a single round, as long as each one results in a local reconciliation. For example,

starting with $m = 3$, the first NNI move will locally reconcile a node if one of the two NNI

topologies reduces the number of mappings to the species tree (we have never observed a

case where both NNI topologies reduce this number). In this case, $m = 2$, and we now carry

out another NNI move on the updated gene tree. Only one move needs to be examined at

this point, however, since the back-NNI rearrangement would return us to the starting

state. The cost of all of these NNI moves is then compared to the cost of duplication and loss

at the end of the round. Duplication+loss naturally allows for more than one of each type of

event for a local reconciliation, and is therefore not handled differently when $m > 2$.

Finally, at the end of a round, `reconcILS` will sum the cost of all events in order to

choose the optimal local reconciliation. If there are equally optimal reconciliations,

`reconcILS` will report the duplication+loss solution.

 Labeling events

Our approach for labeling ILS on reconciled gene trees is described above in the

"New Approaches" section. Here, we consider two important caveats to this approach, one

perhaps more obvious than the other. First, a specific labeling is associated with a specific

reconciliation. That is, if we had found a different optimal reconciliation for the tree in

Figure 3a (for instance, one duplication and two losses), we would have had different

labeled events. Therefore, a labeling is always reported in the context of a particular

reconciliation.

Second, as mentioned, for some ILS events it is not always clear which of the two

nodes flanking a discordant branch to label with I: this is why there is no exact map

between gene tree node labels and species tree nodes in the presence of ILS. For a

discordant branch, we have arbitrarily set `reconcILS` to always choose the node closest

to the tips as I. In some cases, this choice should not matter very much. Consider the toy

reconciliation in Figure 2b—should we have placed the NNI event on node G_1 or on node

G_2? In this case the choice does not seem to imply any difference in evolutionary histories.

In contrast, consider the reconciliations shown in Figure 3b and 3c—these represent the

output of `reconcILS` applied to the gene trees in Figure 1d and 1e, respectively. While we

have left the daughter tree branch in red for clarity, it is important to note that these trees

are identical topologically, which is why `reconcILS` has labelled them in exactly the same

way. The algorithm has no way to "see" the red branches, and has consequently simply

picked the bottom-most node in both cases as the one to label as having undergone ILS and

duplication, even though this is not precisely correct in Figure 3c. At the moment, this is a

limitation of `reconcILS` (and all methods attempting to label gene trees with ILS events),

but we mention other possible solutions in the Discussion.

Primate data

We used two sets of gene trees that were previously constructed in Smith et al.

(2022) using IQ-TREE2 (Minh et al. 2020b), based on data from Vanderpool et al. (2020).

The first set consists of 1,820 single-copy orthologs; this set was denoted as "Single-copy

clusters" in Smith et al. (2022). The second set consists of 11,555 trees containing both

orthologs and paralogs; this set was denoted "All paralogs" in Smith et al. (2022). We

randomly resolved all polytomies. When using `reconcILS`, costs were set to $C_D = 2$,

$C_L = 2$, and $C_I = 1$.

Acknowledgements

We thank Yao-ban Chan for many helpful conversations about the MLMSC model, Jessica Wu for answering questions about DLCpar, and Ben Fulton for tips on best programming practices. Reviewers of a previous version of the manuscript also provided constructive comments. This work was supported by NSF grant DEB-1936187 and DBI-2146866 to MWH.

Data availability

`reconcILS` is available at https://github.com/smishra677/reconcILS. `dupcoal` is available at https://github.com/meganlsmith/dupcoal.

References

Avise, J. C., and T. J. Robinson. 2008. Hemiplasy: A new term in the lexicon of phylogenetics. Systematic Biology **57**:503-507.

Bansal, M. S., E. J. Alm, and M. Kellis. 2012. Efficient algorithms for the reconciliation problem with gene duplication, horizontal transfer and loss. Bioinformatics **28**:i283-i291.

Baum, D. A. 2007. Concordance trees, concordance factors, and the exploration of reticulate genealogy. Taxon **56**:417-426.

Boussau, B., and C. Scornavacca. 2020. Reconciling gene trees with species trees. Pp. 3.2:1-3.2:23 *in* C. Scornavacca, F. Delsuc, and N. Galtier, eds. Phylogenetics in the Genomic Era. No commercial publisher | Authors open access book.

Chan, Y.-b., V. Ranwez, and C. Scornavacca. 2017. Inferring incomplete lineage sorting, duplications, transfers and losses with reconciliations. Journal of Theoretical Biology **432**:1-13.

Chaudhary, R., J. G. Burleigh, and O. Eulenstein. 2012. Efficient error correction algorithms for gene tree reconciliation based on duplication, duplication and loss, and deep coalescence. BMC Bioinformatics **13**:S11.

Chen, K., D. Durand, and M. Farach-Colton. 2000. NOTUNG: A program for dating gene duplications and optimizing gene family trees. Journal of Computational Biology **7**:429-447.

Christensen, S., E. K. Molloy, P. Vachaspati, A. Yammanuru, and T. Warnow. 2020. Non-parametric correction of estimated gene trees using TRACTION. Algorithms for Molecular Biology **15**:1.

Du, H., Y. S. Ong, M. Knittel, R. Mawhorter, N. Liu, G. Gross, R. Tojo, R. Libeskind-Hadas, and Y.-C. Wu. 2021. Multiple optimal reconciliations under the duplication-loss-coalescence model. IEEE/ACM Transactions on Computational Biology and Bioinformatics **18**:2144-2156.

Duchemin, W., G. Gence, A.-M. Arigon Chifolleau et al. 2018. RecPhyloXML: a format for reconciled gene trees. Bioinformatics **34**:3646-3652.

Durand, D., B. V. Halldórsson, and B. Vernot. 2006. A hybrid micro-macroevolutionary

approach to gene tree reconstruction. Journal of Computational Biology **13**:320-335.

El-Mabrouk, N., and A. Ouangraoua. 2017. A general framework for gene tree correction

based on duplication-loss reconciliation. Pp. 8:1-8:14. WABI.

Emms, D. M., and S. Kelly. 2019. OrthoFinder: phylogenetic orthology inference for

comparative genomics. Genome Biology **20**:238.

Fitch, W. M. 1970. Distinguishing homologous from analogous proteins. Systematic Zoology

19:99-113.

Gillespie, J. H., and C. H. Langley. 1979. Are evolutionary rates really variable? Journal of

Molecular Evolution **13**:27-34.

Goodman, M., J. Czelusniak, G. W. Moore, A. E. Romero-Herrera, and G. Matsuda. 1979.

Fitting the gene lineage into its species lineage, a parsimony strategy illustrated by

cladograms constructed from globin sequences. Systematic Zoology **28**:132-163.

Guerrero, R. F., and M. W. Hahn. 2018. Quantifying the risk of hemiplasy in phylogenetic

inference. Proceedings of the National Academy of Sciences **115**:12787-12792.

Guigo, R., I. Muchnik, and T. F. Smith. 1996. Reconstruction of ancient molecular phylogeny.

Molecular Phylogenetics and Evolution **6**:189-213.

Hahn, M. W. 2007. Bias in phylogenetic tree reconciliation methods: implications for

vertebrate genome evolution. Genome Biology **8**:R141.

Hahn, M. W., and L. Nakhleh. 2016. Irrational exuberance for resolved species trees.

Evolution **70**:7-17.

Hallett, M., J. Lagergren, and A. Tofigh. 2004. Simultaneous identification of duplications

and lateral transfers. Pp. 347-356. Proceedings of the eighth annual international

conference on Research in computational molecular biology (RECOMB '04).

Hallett, M. T., and J. Lagergren. 2001. Efficient algorithms for lateral gene transfer

problems. Pp. 149-156. Proceedings of the fifth annual international conference on

Computational biology (RECOMB '01).

Harris, C. R., K. J. Millman, S. J. Van Der Walt et al. 2020. Array programming with NumPy.

Nature **585**:357-362.

Hibbins, M. S., L. C. Breithaupt, and M. W. Hahn. 2023. Phylogenomic comparative methods:
Accurate evolutionary inferences in the presence of gene tree discordance.
Proceedings of the National Academy of Sciences **120**:e2220389120.
Hudson, R. R. 1983. Testing the constant-rate neutral allele model with protein sequence
data. Evolution **37**:203-217.
Huerta-Cepas, J., H. Dopazo, J. Dopazo, and T. Gabaldón. 2007. The human phylome.
Genome Biology **8**:R109.
Huerta-Cepas, J., F. Serra, and P. Bork. 2016. ETE 3: Reconstruction, analysis, and
visualization of phylogenomic data. Molecular Biology and Evolution **33**:1635-1638.
Jacox, E., C. Chauve, G. J. Szöllősi, Y. Ponty, and C. Scornavacca. 2016. ecceTERA:
comprehensive gene tree-species tree reconciliation using parsimony. Bioinformatics
32:2056-2058.
Kingman, J. F. C. 1982. The coalescent. Stochastic Processes and their Applications **13**:235-
248.
Lanfear, R., and M. W. Hahn. in press. The meaning and measure of concordance factors.
Molecular Biology and Evolution.
Li, Q., Y.-b. Chan, N. Galtier, and C. Scornavacca. 2024. The effect of copy number hemiplasy
on gene family evolution. Systematic Biology **73**:355-374.
Li, Q., C. Scornavacca, N. Galtier, and Y.-B. Chan. 2021. The multilocus multispecies
coalescent: A flexible new model of gene family evolution. Systematic Biology **70**:822-
837.
Libeskind-Hadas, R., Y.-C. Wu, M. S. Bansal, and M. Kellis. 2014. Pareto-optimal phylogenetic
tree reconciliation. Bioinformatics **30**:i87-i95.
Maddison, W. P. 1997. Gene trees in species trees. Systematic Biology **46**:523-536.
Mallo, D., L. De Oliveira Martins, and D. Posada. 2014. Unsorted homology within locus and
species trees. Systematic Biology **63**:988-992.
Mallo, D., L. De Oliveira Martins, and D. Posada. 2016. *SimPhy*: Phylogenomic simulation of
gene, locus, and species trees. Systematic Biology **65**:334-344.
Mawhorter, R., and R. Libeskind-Hadas. 2019. Hierarchical clustering of maximum
parsimony reconciliations. BMC Bioinformatics **20**:612.

Mendes, F. K., and M. W. Hahn. 2016. Gene tree discordance causes apparent substitution
rate variation. Systematic Biology **65**:711-721.

Mendes, F. K., D. Vanderpool, B. Fulton, and M. W. Hahn. 2020. CAFE 5 models variation in
evolutionary rates among gene families. Bioinformatics **36**:5516-5518.

Meng, C., and L. S. Kubatko. 2009. Detecting hybrid speciation in the presence of incomplete
lineage sorting using gene tree incongruence: A model. Theoretical Population Biology
75:35-45.

Minh, B. Q., M. W. Hahn, and R. Lanfear. 2020a. New methods to calculate concordance
factors for phylogenomic datasets. Molecular Biology and Evolution **37**:2727-2733.

Minh BQ, Schmidt HA, Chernomor O, Schrempf D, Woodhams MD, Von Haeseler A, Lanfear
R. 2020b. IQ-TREE 2: New models and efficient methods for phylogenetic inference in
the genomic era. Molecular Biology and Evolution **37**:1530-1534.

Morel, B., A. M. Kozlov, A. Stamatakis, and G. J. Szöllősi. 2020. GeneRax: A tool for species-
tree-aware maximum likelihood-based gene family tree inference under gene
duplication, transfer, and loss. Molecular Biology and Evolution **37**:2763-2774.

Nguyen, T. H., V. Ranwez, S. Pointet, A.-M. A. Chifolleau, J.-P. Doyon, and V. Berry. 2013.
Reconciliation and local gene tree rearrangement can be of mutual profit. Algorithms
for Molecular Biology **8**:12.

Page, R. D. M. 1994. Maps between trees and cladistic analysis of historical associations
among genes, organisms, and areas. Systematic Biology **43**:58-77.

Page, R. D. M., and M. A. Charleston. 1997. From gene to organismal phylogeny: Reconciled
trees and the gene tree/species tree problem. Molecular Phylogenetics and Evolution
7:231-240.

Paradis, E., and K. Schliep. 2018. ape 5.0: an environment for modern phylogenetics and
evolutionary analyses in R. Bioinformatics **35**:526-528.

Rasmussen, M. D., and M. Kellis. 2012. Unified modeling of gene duplication, loss, and
coalescence using a locus tree. Genome Research **22**:755-765.

Richter, D. J., P. Fozouni, M. B. Eisen, and N. King. 2018. Gene family innovation,
conservation and loss on the animal stem lineage. eLife **7**:e34226.

Schrider, D. R., and M. W. Hahn. 2010. Lower linkage disequilibrium at CNVs is due to both

recurrent mutation and transposing duplications. Molecular Biology and Evolution

27:103-111.

Smith, M. L., and M. W. Hahn. 2022. The frequency and topology of pseudoorthologs.

Systematic Biology **71**:649-659.

Smith, M. L., D. Vanderpool, and M. W. Hahn. 2022. Using all gene families vastly expands

data available for phylogenomic inference. Molecular Biology and Evolution

39:msac112.

Stolzer, M., H. Lai, M. Xu, D. Sathaye, B. Vernot, and D. Durand. 2012. Inferring duplications,

losses, transfers and incomplete lineage sorting with nonbinary species trees.

Bioinformatics **28**:i409-i415.

Stull, G. W., X.-J. Qu, C. Parins-Fukuchi et al. 2021. Gene duplications and phylogenomic

conflict underlie major pulses of phenotypic evolution in gymnosperms. Nature Plants

7:1015-1025.

Sukumaran, J., and M. T. Holder. 2010. DendroPy: a Python library for phylogenetic

computing. Bioinformatics **26**:1569-1571.

Szöllősi, G. J., W. Rosikiewicz, B. Boussau, E. Tannier, and V. Daubin. 2013. Efficient

exploration of the space of reconciled gene trees. Systematic Biology **62**:901-912.

Tajima, F. 1983. Evolutionary relationships of DNA sequences in finite populations.

Genetics **105**:437-460.

Than, C. V., and N. A. Rosenberg. 2013. Mathematical properties of the deep coalescence

cost. IEEE/ACM Transactions on Computational Biology and Bioinformatics **10**:61-72.

Thomas, G. W. C., S. H. Ather, and M. W. Hahn. 2017. Gene-tree reconciliation with MUL-

trees to resolve polyploidy events. Systematic Biology **66**:1007-1018.

Vanderpool, D., B. Q. Minh, R. Lanfear et al. 2020. Primate phylogenomics uncovers multiple

rapid radiations and ancient interspecific introgression. PLoS Biology **18**:e3000954.

Vernot, B., M. Stolzer, A. Goldman, and D. Durand. 2008. Reconciliation with non-binary

species trees. Journal of Computational Biology **15**:981-1006.

Waskom, M. 2021. Seaborn: Statistical data visualization. Journal of Open Source Software

6:3021.

Wu, Y.-C., M. D. Rasmussen, M. S. Bansal, and M. Kellis. 2014. Most parsimonious

reconciliation in the presence of gene duplication, loss, and deep coalescence using

labeled coalescent trees. Genome Research **24**:475-486.

Yu, Y., C. Than, J. H. Degnan, and L. Nakhleh. 2011. Coalescent histories on phylogenetic

networks and detection of hybridization despite incomplete lineage sorting.

Systematic Biology **60**:138-149.

Zhang, C., C. Scornavacca, E. K. Molloy, and S. Mirarab. 2020. ASTRAL-Pro: Quartet-based

species-tree inference despite paralogy. Molecular Biology and Evolution **37**:3292-

3307.

Zmasek, C. M., and S. R. Eddy. 2001. A simple algorithm to infer gene duplication and

speciation events on a gene tree. Bioinformatics **17**:821-828.