Can biocrust moss hide from climate change? Fine-scale habitat sheltering improves summer-stress resistance in *Syntrichia caninervis*

Theresa A. Clark[1,2,3], Alexander Russell[1], Joshua L. Greenwood[1], Dale Devitt[1], Daniel Stanton[2], Lloyd R. Stark[1]

[1]School of Life Sciences, University of Nevada, Las Vegas, NV
[2]Department of Evolution, Ecology & Behavior, University of Minnesota, Twin Cities, MN
[3]Corresponding author

Can biocrust moss hide from climate change?

ABSTRACT

Premise: Mosses provide many ecosystem functions and are the most vulnerable of biocrust organisms to climate change due to their sensitive water relations stressed by summer aridity. Given their small size, moss stress resistance may be more dependent on fine-scale habitat than macroscale (climate and community), but this sheltering role of habitat (i.e. habitat buffering) has never been compared to macroclimate and may have important implications for predicting critical biocrust refugia in changing climates.

Methods: We located three populations of a keystone biocrust moss, *Syntrichia caninervis*, spanning 1200-m of altitude comprising three plant communities (elevation-plant zones) in the Mojave Desert. We stratified 96 microsites along three habitat aridity gradients: elevation-zone, topography (aspect), and microhabitat (shrub proximity). We estimated summer photosynthetic stress (F_v/F_m) and aridity exposure (macroclimate, irradiance, and shade).

Results: Microsite aridity exposure varied greatly revealing exposed and buffered microhabitats in all elevation-zones. Moss stress did not differ by elevation zone despite the extensive macroclimate gradient, failing to support the high-elevation refugia hypothesis. Instead, stress was lowest on N-facing slopes and microhabitats with higher shrub shading, while the importance of (and interactions between) topography, irradiance, and shade varied by elevation zone.

Conclusions: We demonstrate fine-scale habitat buffering is physiologically more protective than high-elevation climate, and thus, may allow some mosses to hide from the brunt of climate change in widespread microrefugia throughout their current ranges. Our findings support a scale-focused vulnerability paradigm: microrefugia may be more important than macrorefugia for bolstering biocrust moss resistance to summer climate stress.

Keywords: biological soil crust; chlorophyll fluorescence; climate change; desert bryophytes; desiccation tolerance; habitat buffering; Mojave Desert; NevCAN; *Syntrichia caninervis*; water relations

Introduction

Biocrusts (i.e., biological soil crusts) are diverse networks of cyanobacteria, fungi, algae,
lichens, mosses, and liverworts weaving together the upper few centimeters of topsoil in arid
and semiarid environments globally (Belnap 2003). Where sufficient moisture or shade exist,
mosses are often the larger members of these soil communities and drive many biocrust
ecosystem services (Bowker et al. 2013) mediated by their large (typically $1 - 10$ cm^2),
absorbent colonies or "patches" that can increase water infiltration and retention (Lafuente et
al. 2018), buffer temperature (Xiao et al. 2015), increase soil fertility (Belnap 2003), shelter
microbiota (Abed et al. 2019, Fisher et al. 2020), store carbon (Elbert et al. 2012), and prevent
erosion (Stovall et al. 2022). However, biocrust mosses are predicted to have more dramatic
responses to climate change than most other poikilohydric biocrust species because of their
requirements for higher shade and moisture during hydration periods (i.e., "hydroperiods")
when the plants are metabolically active (He et al. 2016, Rodriguez-Caballero et al. 2018, Weber
et al. 2018, Ladrón de Guevara & Maestre 2022). Such sensitivity in biocrust mosses has been
documented in multiple field and laboratory experiments simulating extreme climate stress
(e.g., rapid drying events), which has caused stress, tissue damage, or lethality (e.g. Brinda et al.
2011, Stark et al. 2011, Reed et al. 2012, Zhang et al. 2016, Greenwood et al. 2019, Coe et al.
2020). Moreover, ecophysiological models predict that climatic change may greatly reduce
biocrust moss biomass via reduced productivity and increased mortality (Coe and Sparks 2014).
Collectively, such impacts on biocrust moss resilience and productivity may reduce their
diversity and ecosystem services in changing climates (Ladrón De Guevara & Maestre 2022).
High-elevation habitat such as mountains and plateaus offer cooler temperatures and
higher humidity than lower elevations in drylands and thus are predicted to offer oasis to many
plants as climate warms and becomes drier (Kelly & Goulden 2008). Such macroclimate
gradients along broad altitudinal ranges drive zonation in the dominant vegetation community
(elevation zones). High-elevation communities are already documented to support greater
moss abundance and diversity in several drylands with extensive aridity-elevation gradients
exceeding hundreds of meters (e.g. Nash et al. 1977, Seppelt et al. 2016, Clark 2020). Such
elevation zone gradients may, therefore, function as high-elevation refugia for dryland mosses,
assuming species have time and sufficient propagules to migrate upward (Zanatta et al. 2020).
An alternative or co-occurring possibility is that mosses could "hide from climate
change" in sheltered microrefugia that offer optimal microhabitat within a species' present
range (Ashcroft 2010). The microscale of dryland moss colonies or "patches" are often less than
5 cm^2 (Nash et al. 1977, Clark 2020) such that fine-scale habitat structure within sites may be
equally or more important to moss resiliency than macroscale climate gradients. Fine-scale
habitat such as pole-facing slopes and rock surfaces and shade vegetation have been shown to
reduce local extremes in temperature, incident radiation, and evaporative demand adjacent to
dryland mosses (Alpert 1988, He et al. 2016, Li et al. 2018, Ladrón de Guevara & Maestre 2022).
The habitat buffering hypothesis predicts that such sheltered microhabitats will "buffer", or
reduce, climate stress for many organisms (Williams et al. 2008, Scheffers et al. 2014, Shi et al.
2016). Moreover, mosses are small enough to be sheltered by multiple scales of habitat in
additive or interactive ways, which we will call *multiscale* habitat buffering. In drylands, there is
a prevalence of topographically complex, multiscaled habitat for biocrust; for example a habitat

that is shaded by fine-scale vegetation on a north-facing slope within a drainage basin (e.g., Williams et al. 2013, Pietrasiak et al. 2014, Bowker et al. 2016). There is potential that such scales of habitat structure may interact to create strongly sheltered biocrust habitats in future climates.

Predicting the most important scales of habitat buffering to biocrust mosses in changing climates can begin with studying present-day stress and mortality patterns across scales of habitat structure relevant to these mosses. Scales known to influence their distributions include macro-, meso-, and microscale environmental features (Bowker et al. 2016). The mesoscale includes within-site topographical variation on the scale of meters where changes in aspect, slope, and hydrological position can alter soil-water dynamics important to biocrust water availability, such as location relative to a drainage. Microscale variation can include microtopographical shading driven by proximity and azimuth to surrounding vegetation and rocks. Such studies will elucidate the types of refugia, macro- or microrefugia (Ashcroft 2010), that are more probable for small, photosynthetic organisms. Measuring climate stress estimates using gas exchange or chlorophyll fluorescence, will also provide spatially explicit data that can be used to strengthen predictions for future moss abundance distributions in drylands (Coe & Sparks 2014).

We sought to test the high-elevation refugia and habitat buffering hypotheses in the biocrust moss, *Syntrichia caninervis* Mitten, a member of the acrocarpous moss family, Pottiaceae. This keystone biocrust species is understood to be one of the most broadly distributed and ecologically important biocrust mosses in western North America, North Africa, and Asia (e.g., Coe et al. 2012, Seppelt et al. 2016, Ros et al. 1999, Zhang & Zhang 2019). With a broad geographical and altitudinal distribution in the American Southwest (BFNA 2007), *S. caninervis* provides an ideal model species to study summer stress resistance along multiscaled gradients in habitat structure. The Mojave Desert is the most climatically extreme part the species' North American range where resident populations likely exist at or near physiological thresholds of precipitation minima and temperature maxima (e.g. Stark et al. 2009)**.** Summer presents the highest risk for moss mortality when combinations of extreme desiccation (i.e., cellular water potentials < -400 mPa) interrupted by small rain events have been shown to prevent *S. caninervis* from achieving positive carbon balance during summer hydroperiods (Coe et al. 2012b). Such carbon deficits can lead to full-patch mortality and suggest *S. caninervis* may be increasingly threatened by continued climate change. Current climate trends and predictions in this most arid North American desert include smaller summer rain events and increased drought intensity and variability (Seager et al. 2007, Seager & Vecchi 2010, Zhang et al. 2021).

Objectives

Our specific objectives in the Desert National Wildlife Refuge (DNWR) along a ~1200-m elevation zone gradient in Nevada were to: (1) locate 96 *S. caninervis* microsites spanning three nested scales of habitat structure by surveying three elevation zones (macroscale), three topography zones per site (mesoscale), and three microhabitat types per zone (microscale), (2) estimate moss habitat buffering at each microsite (shade time and potential insolation), (3) use in-situ chlorophyll fluorescence to estimate end-of-summer moss photosynthetic stress and mortality, and (4) elucidate the most important scale of habitat structure by testing for stress

patterns by habitat scale and modeling the relationship between habitat buffering proxies (elevation, potential insolation, and shade) and summer stress in *S. caninervis*. This natural experiment allowed us to test relationships between three scales of habitat structure and habitat buffering with summer stress resistance in three elevation zone populations of a biocrust moss to gather evidence to either support or reject the high-elevation and habitat buffering hypotheses in one of the harshest environments for which this species occurs globally, the Mojave Desert. Results have important implications for predicting and effectively monitoring the response of this and similar biocrust species to future climate change in drylands by using scale-appropriate and spatially explicit ecological models (Gignac 2001, Dunning et al. 1995).

MATERIALS AND METHODS

Elevation zone sites

The Desert National Wildlife Refuge (DNWR) is a large (6,430 km^2) topographically and biologically diverse basin and range landscape in the eastern Mojave Desert of southern Nevada. DNWR is home to the Sheep Range EPSCoR-NevCAN transect (*Established Program to Stimulate Competitive Research - Nevada Ecohydrological Climate Assessment Network;* Mensing et al. 2013), a set of five climate stations spanning 2000 m of elevation and located in one of the five Mojave elevation zones (**Fig. 1a**), all of which have been floristically characterized without mention of bryophytes (Ackerman 2003, NCCP 2018). Regional soils are limestone-derived, highly calcareous, and range from low-organic to relatively high organic content at high elevations.

The lowest elevation desert scrubland site (890 m, N36.435345, W115.355850; "Low-scrubland") and surrounding landscape are characterized by an open salt basin interrupted by shallow, calcareous drainages with gentle slopes and occasional steep ravines 1 – 2 m deep. Excluding drainages, the soil is covered almost entirely by desert pavement (e.g. Pietrasiak et al. 2014) with well-spaced shrubs (>2 m apart) and (**Fig. 2a**). The blackbrush-Joshua tree (*Coleogyne ramosissima and Yucca brevifolia*) elevation zone site (1680 m, N36.51723, W115.16191; "Mid-shrubland") is situated in the center of an intermountain basin divided by drainages that range from ~1 – 3 m deep. The ground is nearly covered by desert pavement, moderately spaced shrubs (<2 m apart), and widely spaced succulents (**Fig. 2a**). The pinyon-juniper woodland site (2065 m, N36.572808, W115.204060; "High-woodland") is at the base of the Sheep Mountains on one of the deeply divided ridges with steep, rocky slopes (>3 m tall, ~10° - 15°). The soil is nearly covered by loose gravel with a dense community of short and tall shrubs and the dominant well-spaced pygmy conifers (*Pinus monophyla* and *Juniperus osteosperma*; **Fig. 2a**). The Montane site (2320 m, N36.590255, W115.214166) has an open-canopy of *Pinus ponderosa* and well-spaced shrubs (>3m apart). The highest-elevation Subalpine site (3015 m, N36.657641 W115.200777) in the NevCAN transect is a nearly closed-canopy mixed-conifer forest (*Abies* concolor, *A. lasiocarpa*, *Picea englemannii*, and *Pinus longaeva*) with calcareous, rocky, organic soils (Ackerman 2003).

Climate metrics

To compare mean annual and summer climate where we located *S. caninervis* along the NevCAN-DNWR elevation zone transect, we acquired NevCAN daily means for temperature, humidity, irradiance, and precipitation from 2011 – 2018 (DRI, 2020). We calculated the 7-yr mean annual air and soil temperature, percent relative humidity (RH), wind speed, and soil moisture. We calculated summer 2017 climate means to include Mojave hot-season months preceding our moss tissue collection, which took place at the beginning of November (6/1/17 – 11/6/17; **Table 1a**).

Aridity exposure survey: elevation zones, topography zones, & microhabitats

To determine the macroclimate exposure of *S. caninervis* in DNWR across the three habitat gradients, we surveyed for species occurrence within a 1-km radius of each elevation zone climate tower (**Fig. 1a & b**); when we found the species in a elevation zone, we surveyed three topographical exposure zones (40 x 10 m plots) selecting the most northerly-facing, southerly-facing, and flat terrain nearest the tower. Selected topography zones varied in their hydrological positions, including association with drainages, uplands, or mountain ridges (**Fig. 1c**). Within each topography zone, we surveyed for moss occurrence in three microhabitat shade classes ("*microhabitat types*" hereafter): (1) high-shade *canopy habitats* partially or fully under shrub canopies, (2) low-shade *interspace habitats* ≥ 0.5 m from the outer edge of the shrub canopy (the "canopy dripline"), and (3) intermediate-shade *dripline habitats* located within a 0.5 m marginal band from the shrub dripline; **Fig. 1 d**). Any vegetation located to the north (cardinal 315° – 45°) of moss microsites was ignored in habitat type assignment because it did not contribute to moss shade.

Moss microsite selection – We systematically selected 12 microsites per topography zone, attempting to find 4 of each microhabitat type per topography zone, but because this was often not possible (see **Fig. 1c**), the resulting proportion of microhabitats at each elevation zone is a coarse measure of habitat frequency (**S1**). We selected only undisturbed microhabitats having the shrub canopy intact (i.e., not dead or broken off). We systematically centered a 20 x 20 cm quadrat over the highest-density patch of *S. caninervis* in each microhabitat (>3 cm^2 of *S. caninervis* cover), orienting the quadrat with sides parallel to a North-South axis (**Fig. 1d**). Four shoots were sampled from each quadrat for microscopic species verification; however, mid-way in the study, we removed two microsites after finding (via lab culture) two *Syntrichia* species (*caninervis* and *ruralis* sensu lato) were unknowingly intermixed and possibly measured in the stress assay (Clark 2020; **Fig. 1a**).

Habitat buffering metrics

Sheltered (i.e. "buffered") microhabitats for dryland mosses are generally thought to be those with higher humidity, lower temperature, and reduced insolation compared to neighboring local habitats (Alpert 1988). Although such habitat buffering can be measured directly with microenvironmental sensors, large replication (n=96) and sensor installation in delicate biocrust systems is highly invasive; therefore, we measured buffering proxies (e.g. slope, aspect, and shade) for the three scales of habitat structure (utilizing only NevCAN climate tower sensors at each of the NevCAN sites) as follows.

Macroscale climate buffering – Elevation zone buffering was estimated at the
landscape scale by calculating the mean difference in macroclimate relative to the low-
elevation Creosote elevation zone where conditions are most stressful for mosses (i.e. most
arid and hot). For example, the elevation zone temperature buffer was calculated as the mean
annual decrease in daily temperature between the creosote elevation zone and the higher
elevation zones.

Mesoscale topographical buffering – We measured aspect (compass cardinal direction)
and slope (Suunto PM-5 hand-held clinometer) of the 3 x 3 m^2 area surrounding each microsite
for use in calculating potential direct incident radiation (PDIR). Within elevation zones, we
calculated mesoscale topographical exposure as the mean of microsite PDIR in each topography
zone. PDIR was estimated with a complex formula that incorporates microsite elevation,
latitude, slope, and folded aspect (McCune & Keon 2002, McCune 2007). We observed *S.*
caninervis ground cover to be greatest on the N side of shrubs in this ecosystem and thus
modified the PDIR equation for use with a N-S rather than NE-SW axis as described in McCune
(2007). To estimate PDIR buffering across and within elevation zones, we calculated differences
in mean mesoscale topographical exposure. For example, the maximum topographical zone
buffer was simply the difference in mean PDIR between topography zones with the highest and
lowest mean PDIRs. Mean elevation zone PDIR was simply the average of all topography zone
PDIR at each elevation zone site.

Microscale shade buffering – To precisely measure fine-scale shade time of microsites,
we developed a photographic method using the smartphone app, *Sun Seeker© Solar AR*
(Augmented Reality) Tracker (Sydney, Australia), which maps onto the camera view the annual
solar window from sunset to sunrise for a given location (e.g., moss microsite). Seven photos
taken at any time of the year circumscribe all geographical-time-referenced shade objects from
the vantage point of the moss (**Fig. 2**). For each photo, the area of shade objects (i.e.,
vegetation and topography) inside the solar window was scored using a shade class from 0 (no
shade) to 4 (75 – 100% shaded; see classes in **Fig. 2**). For each microsite, the resulting seven
shade classes were divided by the total possible score of 28 shade points to yield an annual
shade time percentage. Our novel microscale shade metric estimates the percentage of the
year a microhabitat is shaded by vegetation and/or topography; we used this sensor-free
buffering metric to test differences in mean shade at various scales of habitat structure.

Field sampling & tissue prep

To measure the summer stress signal of *S. caninervis*, we collected shoots when patches
were fully desiccated in late fall (November 6 – 13, 2017; *Permit #84555-17-019 U.S. Fish and*
Wildlife Service National Refuge System Research & Monitoring), which is still considered the
Mojave warm season. This dormant, dry tissue preserved the summer/warm-season stress
signal because we collected prior to any fall precipitation during which some recovery could
begin. We collected two to three shoots for every 2-cm^2 patch area per quadrat using fine
forceps (**Fig. 3a-b**). Collections were stored dark and dry at 20°C with 20 – 30% relative
humidity, conditions which should not exacerbate stress in this species (Guo & Zhao 2018).
Before the stress assay, shoots were hydrated with distilled water on a microscope slide and
the upper 2 mm were cut and retained to target the living apical "green zone" (Stark 2017) (**Fig.**
3b). Green zones were swirled in a drop of water to remove debris, and placed vertically into a

rosette formation on a wetted 7-mm filter paper disc creating a *"moss bouquet"* (**Fig. 3b**). The number of shoots in a moss bouquet (5 – 23) depended on shoot size to maximize microsite representation and standardize leaf area for chlorophyll fluorescence.

Summer-stress resistance assay (Fv/Fm)

Efficiency of photosystem II (PSII) photosynthesis is commonly measured as a stress proxy in plants via the non-invasive technique, chlorophyll fluorescence (Papadatos et al. 2017), convenient for measurements on small moss species with limited biomass or long-term monitoring (Proctor 2009). We refer to Fv/Fm as a stress proxy because in desiccation tolerant mosses, Fv/Fm responds sensitively and negatively in a near linear fashion to the level of desiccation and light stress induced on species in carefully controlled experiments (e.g. Proctor et al. 2007, Proctor 2001). In almost all cases, the lower the Fv/Fm when such a moss is rehydrated (30 min – 1 hr after desiccation), the more stressful were the conditions during the previous hydration/desiccation cycles (e.g. Coe et al. 2020). This reduced Fv/Fm can be viewed as a photoprotective acclimation to light and low water potentials both of which stress the electron transport chain and induce a photoprotective response (Papadatos et al. 2017). Long-term acclimation to high light and desiccation is the most parsimonious explanation for reduced potential photosynthetic efficiency (Fv/Fm) in desiccation tolerant mosses (when measured on fully hydrated material) regardless of the photoprotective response, which usually is assumed to involve the D1 repair cycle and slowly reversible forms of nonphotochemical quenching (Proctor et al. 2007). Therefore, our photosynthetic stress proxy, Fv/Fm, will be called "stress" in figures and the Discussion.

Our Fv/Fm measurements were performed with a Hansatech FMS-2 modulated fluorometer (Norfolk, England) and modified leaf clips hollowed to accommodate taller shoots (**Fig. 3b & c**). Hydrated moss bouquets were each placed on a folded chemical-wipe and carefully positioned into leaf clips, then placed in a tray of distilled water to maintain full-turgor throughout the 20-min dark acclimation (a period needed to close all active PSII reaction centers). Including 10 min to clean and prep bouquets, we measured chlorophyll fluorescence at 30-min post-rehydration, a standard photosynthetic reactivation timepoint for assessing the photosynthetic stress or vitality of mosses prior to significant recovery from the preceding desiccation event (e.g., Munzi et al. 2019, Hamerlynck et al. 2000, Ekwealor et al. 2021).

A Hansatech script automated our measurement of the dark-acclimated metrics, basal (Fo) and maximum (Fm) fluorescence, during a 0.8-s saturation pulse of 3000 μmol/m^2/s. Using variable fluorescence (Fv = Fm – Fo), Fv/Fm was then derived, which estimates potential maximum quantum efficiency of photosystem II (PSII) photochemistry of dark-acclimated plants (Baker 2008). Fv/Fm provides a universal physiological indicator of climate stress in plants that is more sensitive and integrative than chlorophyll content (Murchie & Lawson 2013) and requires less tissue and time than gas exchange analysis.

Analysis

Analyses were performed in *R* (Version 1.447 and 4.2.3; R-Core-Development-Team 2023) using the *tidyverse* package (Wickham et al. 2019) and statistical packages denoted as (*package::function*). We used an exploratory approach with hypothesis testing in which we

consider the ecological significance of statistical patterns when *P* is < 0.05 and opt out of arbitrary family corrections for testing our small set of environmental factors (Gotelli and Ellison 2013).

Shade buffering & moss stress vs multiscale habitat structure – Although our design involved three categorical habitat predictors of moss stress and shade buffering (elevation zone, topography zone, and microhabitat type), using three-way ANOVA's to test patterns in mean shade buffering and moss stress were not possible due to missing factor levels (i.e. no S-facing zone in the Low-scrubland and no interspace microhabitats in several topography zones (**Fig. 1c, S1**). Alternatively, we conducted six univariate ANOVA's: three stress tests and three shade buffering tests, one for each habitat factor. We calculated the effect sizes, Eta-squared (equivalent to R^2 in one-way ANOVA) and Cohen's *f* (Lakens 2013). When heteroskedasticity was present (determined with the Fligner-Killeen test; Fligner & Killeen 1976), we opted out of transformations, which are known to yield nonsensical predictions of proportion data and hinder interpretability (Warton and Hui 2011). Instead, we used robust Welch's denominator degrees of freedom corrections appropriate for the approximate normality, unbalanced design, and heteroskedasticity present (*stats::oneway.test*; Welch 1951). Post-hoc tests for Welch's ANOVA's were nonparametric Games-Howell multiple comparisons (*rstatix:: games_howell_test*; Kassambara 2023, Ruxton & Beauchamp 2008).

Habitat buffering proxies as predictors of moss stress – To explore the relationship between the three scales of habitat buffering (elevation, PDIR, and shade) and summer moss stress, we fit a linear multiple regression model for Fv/Fm (**S2**). Despite small variance inflation factors of 2.07, 1.17, 2.19, for the three predictors, respectively, our model testing procedure revealed instability in parameter estimates and p-values when elevation was included in the model (and no interaction terms were included). Therefore, the final reduced model included only PDIR, shade, and their interaction, centering the explanatory variables to remove structural multicollinearity. Diagnostic residual plots revealed adequate fit (**S3**); however, we acknowledge that beta regression is optimal for modeling such a two-category ratio (Douma & Weedon 2018) and should be used for prediction-focused studies.

Elevation zone models – With elevation zone management in mind, we performed a set of hypothesis tests focused on each elevation zone site to explore whether our spatially efficient (i.e. nested) sampling design (**Fig. 1**) of meso-scale topography zones (within-site plots) and micro-scale shade measurements (within-plot microsite quadrats) could explain significant patterns in moss stress. (Note: although ideal, testing these relationships in a single ANOVA model including all elevation zones was not possible due to the multicollinearity between elevation zone (elevation) and shade (as discussed in the habitat buffering model above).

Alternatively, we ran three separate ANOVAs (i.e. ANCOVAs) to test additive and interactive relationships between topography zone and percent shade time (centered before analysis) with moss stress for each elevation zone. ANOVAs were made robust to (a) the unbalanced design (**Fig. 1c**) using Type II sums of squares (Langsrud 2003, Logan 2010) and to (b) unequal variance using a Huber-white heteroskedasticity-corrected covariance matrix (HCCM; White 1980; Long & Ervin 2000) for each model (*car::Anova,* Fox & Weisberg 2019). Post-hoc pairwise comparisons for regression slopes (by topography zone) were not conducted (when the interaction term was significant) because we believe sample sizes greater than 11 –

1 12 quadrats per topography zone should be collected to produce more accurate fine-scale
2 relationships. The OLS coefficient of determination (R^2) is shown for each final model as an
3 effect size metric; the overall F-test for each HCCM-corrected model was derived from a Wald
4 test comparing the intercept model to the respective HCCM model including only significant
5 terms (*lmtest::waldtest*; Zeileis & Hothorn 2002).

RESULTS

7   Results are numbered by objective and statistics are printed in figures if not shown here.
8 We refer to our three scales of aridity gradient sampling (i.e., elevation zones, topography
9 zones, and microhabitat types, **Fig. 1**) as the three scales of habitat structure or habitat
10 buffering, depending on context.

Aridity exposure across elevation zones (Objective 1)

After extensive surveying within a 1-km radius of each NevCAN climate tower, we
located *S. caninervis* biocrust in the lower three sites of the DNWR eco-hydrological gradient
spanning 1,180 m of elevation from 890 to 2070 m, while a closely related species, *Syntrichia*
ruralis sensu lato, was found primarily at higher elevations (**Fig. 1a**); we will refer to these
sampling sites as the Low-scrubland, Mid-shrubland, and High-woodland, respectively (**Fig. 1b**).
We located *S. caninervis* on North-facing, South-facing, and flat topography zones at the Mid-
shrubland and High-woodland, but it was absent from southerly facing slopes at the Low-
scrubland (also absent from upland flats) and was only found on shallow drainage flats and
northerly-facing slopes (**Fig. 1c**). In the Low and Mid-elevation sites, *S. caninervis* occurred in all
three microhabitat types, but in the High-woodland, shrub interspace habitat did not support
high-density *S. caninervis*. Relative habitat frequency changed with elevation: shrub canopy
habitat increased, interspace habitat decreased, and shrub-dripline frequency remained similar
from low to high elevation (**S1, Fig. 1c**). Canopy microhabitats were the most frequent habitat
type supporting high-density *S. caninervis* biocrust across elevation zones (54/94 microsites,
S1).

Habitat buffering across three scales of habitat structure (Objective 2)

Macroscale climate buffering – Mean daily macroclimate differed substantially by
elevation zone (**Table 1a**) with higher elevations having cooler temperatures, higher humidity
and precipitation, and slower wind speeds. Relative to the Low-scrubland, the High-woodland
site was climatically buffered in four metrics: (1) 6.5 times more precipitation, (2) reduced
mean daily air and soil temperatures by -7.4°C and -9.0°C, respectively, and (3) increased mean
relative humidity by +8% (**Table 1a**).

Mesoscale topographical buffering – The nine topography zones collectively represent
the mesoscale exposure gradient for *S. caninervis*-dominant biocrust at each elevation zone site
and varied in their hydrological position by elevation zone: the Low-scrubland site was in an
ephemeral drainage, the Mid-shrubland included a drainage and upland flat, and the High-
woodland traversed a mountain ridge (**Fig. 1c**). Of the nine topography zones, there was a 22%
reduction from the highest mean PDIR of the S-facing slope of the High-woodland (1044
kJ/cm²/yr) to the lowest mean PDIR of the N-facing slope of the High-woodland (817 kJ/cm²/yr),
which created a maximum mean topography zone buffer of 227 kJ/cm²/yr (**Table 1b, Fig. 1c**).

Notably, the Low-scrubland had the lowest average PDIR (of 24 microsites) due to its relatively
steep 16° N-facing slope.
Microscale shade buffering – Percent annual shade time ("shade" hereafter) across the
94 microsites ranged from 21 – 96% and averaged 64 ± 2% (SE) such that most microsites were
shaded over half of the year. Elevation zone significantly explained 48% of shade variation
($^{Welch}F_{2, 52} = 40.6$, P < 0.0001); mean shade in the High-woodland was 1.9x greater than in the
Low-scrubland (**Fig. 4a**). Mean shade differed little by topography zone when pooling microsites
across elevation zones ($^{Welch}F_{2, 58} = 2.7$, P = 0.074, R^2 = 0.05, **Fig. 4b**), while microhabitat type
explained 67% of variation in shade, which increased 2.5-fold from interspace to canopy
habitats, on average ($^{Welch}F_{2, 44} = 44.0$, P < 0.0001, **Fig. 4c**).

Summer Stress Proxy (Fv/Fm) by elevation zone (Objective 3)

Nearly all *S. caninervis* patches showed visible signs of stress in their darkened, sun-
pigmented leaves (**Fig. 5**). The sample distribution of Fv/Fm was left-skewed with a mean of
0.455 ± 0.148 (SE) and ranged from 0.130 at the Mid-shrubland to 0.737 at the Mid-shrubland
(**Fig. 6**). Of the 94 mosses, ~23% were near normal levels (Fv/Fm >0.6) leaving 77% stressed (see
Discussion). Dividing the typical physiological range of Fv/Fm (0 – 0.85) into four stress
categories, we report 7% severely stressed, 25% moderately stressed, and 42% mildly stressed,
and 26% unstressed (**Fig. 6**). The seven samples with Fv/Fm < 0.2 (in the lower 25th percentile;
Fig. 6) were from Flat-zones at all three elevation zones. These outliers included two interspace,
three dripline, and one canopy microhabitat and had moderate annual shading ranging from 43
– 68%, except for one interspace microsite at the Low-scrubland that was shaded only 28% of
the year. Signs of extreme stress in these samples were evident in chlorotic leaf tips (**Fig. 5b-c**).

Moss stress (Fv/Fm) vs three scales of habitat structure & buffering (Objective 4)

Habitat type ANOVAs – Contradicting our expectations, *S. caninervis* Fv/Fm did not
differ on average by elevation zone ($^{Welch}F_{2,59} = 1.6$, P = 0.204, **Fig. 4d**). Pooling topographical
zones across sites, topography zone explained 21% of variation in Fv/Fm with the N-facing zone
supporting healthier mosses, on average, than the S-facing or flat zones ($^{Welch}F_{2,54} = 12.7$, P <
0.0001, **Fig. 4e**). Microhabitat type explained 23% of variation in Fv/Fm with healthier mosses
under shrub canopies than in drip lines or interspaces ($^{Welch}F_{2,45} = 16.6$, P < 0.0001, **Fig. 4f**).
Habitat buffering model – The full regression model including all multiscale buffering
variables (elevation, PDIR, shade, and their interactions) was statistically significant explaining
55.4% of variance in Fv/Fm ($F_{7,84} = 14.9$, P < 0.0001; note this model should only be used for
predictive purposes due to multicollinearity; **S2**). The highly significant reduced model (without
elevation zone elevation) had acceptably stable beta estimates for PDIR, shade, and their
interaction providing a model that can be reliably used for mechanistic inference ($F_{3,88} = 23.5$, P
< 0.0001, R^2 = 0.444, **Fig. 7**). Specifically, shade and PDIR were positively related to Fv/Fm (**Fig.
7b**); however, the full model indicated the strength of this relationship increased by elevation
zone elevation (**Fig. 7a**).
Elevation zone models (topography zone and shade) – Fv/Fm was not related to shade
at the Low-scrubland, but the two topography zones explained 57% of variation in which mean
Fv/Fm was higher on the N-facing zone than on the Flat zone ($^{Wald}F_{1,22} = 26.9$, P < 0.0001, **Fig.**

8c). At the Mid-shrubland, shade and topography and their interaction explained 67% of variation in Fv/Fm in which the Flat zone appears more positively related to shade than the N- and S-facing zones (*slopes not tested*; [Wald]$F_{5,28}$ = 12.5, P < 0.0001, **Fig. 8b**). At the High-woodland, only shade was positively related to Fv/Fm and explained 52% of variation ([Wald]$F_{1,32}$ = 57.6, P < 0.0001; **Fig. 8a**).

DISCUSSION

Fv/Fm summer stress interpretation

Our Mojave Desert stress assay confirmed the extreme resiliency of *S. caninervis* to present-day summer climate as we found no patch mortality in the 94 *S. caninervis* microsites studied (i.e. all Fv/Fm > 0, **Fig. 6**) despite the extreme weather of summer 2017 (**Table 1**, 2017), which included record-breaking temperatures (NOAA, 2020), low relative humidity, many small rain events, and over 45 days without rain. Moreover, only 33% of the *S. caninervis* microsites showed signs of moderate to severe photosynthetic stress as suggested by low Fv/Fm values relative to the maximum potential PSII efficiency of plants, ~0.85 (**Fig. 6**). While over 50% of microsites had a moderate to extreme photosynthetic stress signal with severely stressed individuals present at all three elevation zones (**Fig. 6**). Fv/Fm can be confidently used as a stress proxy for desiccation tolerant mosses and revealed a much larger variation in photosynthetic summer stress and photoprotective acclimation than could be detected visually. The appearance of sampled patches ranged from obviously damaged shoots with reddish-orange leaf tips (severely stressed) to those lacking visible damage but darkly sun-pigmented from acclimatory zeaxanthins (Ekwealor et al. 2021; **Fig. 5**).

Reduced Fv/Fm in desiccation tolerant mosses may be caused by many stress factors including rapid drying, low water potential, high temperatures, excess light, and UV damage (Takács et al. 1999, Proctor 2001, Proctor et al. 2007, Greenwood et al. 2019, Ekwealor et al. 2021) all of which are likely acute or chronic stressors in these Mojave mosses especially during hydroperiods when metabolically active following rare precipitation events. Part of their impressive resiliency arises from being in the dry and dormant state most of the year during which they can tolerate extreme heat and extended drought (Stark 2017). When fully desiccated, *S. caninervis* can tolerate intense solar loading of 120°C (248°F) – two times greater than typical Mojave summer air temperatures (Stark 2005, Stark et al. 2009).

Historically called photoinhibition, reduced Fv/Fm has been described as a state of compromised efficiency in the light reactions of photosynthesis thought to result from deactivated PSII reaction centers and/or acclimatory changes in nonphotochemical quenching (NPQ; Demmig-Adams & Adams 2006). Specifically, there are three possible mechanisms of reduced Fv/Fm in desiccation tolerant bryophytes including rapid light acclimation through NPQ, and two longer-term acclimation responses including PSII repair cycling and slowly reversible NPQ (ql) which Muller et al. (2001) states are the result of prolonged, severe light stress. Regardless of the cellular mechanism, reduced Fv/Fm is usually interpreted as a photoprotective acclimation response to climate stress in mosses undergoing desiccation-rehydration cycles (Proctor & Smirnoff 2011, Proctor & Smirnoff 2000). Severely low Fv/Fm (< 0.2) can precede mortality (e.g. Coe et al. 2020), however, some experiments have shown

recovery from such low Fv/Fm (e.g., Cruz de Carvalho 2011, Ekwealor et al. 2021). Further research is needed to determine in what climate scenarios does low Fv/Fm lead to reduced productivity or survival in bryophytes (e.g. Hájek & Vicherová 2013). Regardless of long-term resiliency potential, the large variation we observed in moss stress resistance presented a strong sample for testing the high-elevation and habitat buffering hypotheses.

High-elevation refugia hypothesis

We had anticipated that the cooler, wetter climate and mountain topography at the High-woodland (**Table 1a**) would reduce summer moss stress relative to the Low-scrubland, but found no such evidence to support high-elevation buffering that could indicate potential for future high-elevation refugia in DNWR (**Fig. 4a**, **Fig. 6**). The lack of signal for high-elevation buffering along a 1200-m macroclimate gradient is surprising given the evident positive relationship between moss abundance and elevation along this aridity-stress gradient, the latter of which has been documented in other drylands for which floristics or community analyses have been conducted (Nash et al. 1977, Seppelt et al. 2016, Clark 2012).

In contrast, we found support for the habitat buffering hypothesis at all three elevation zones in which meso- and/or microscale habitat structure and buffering were positively related to summer stress resistance in *S. caninervis*. Mesoscale topography varied primarily by hydrological position and exposure (PDIR) while vegetation shading varied as a function of three microhabitat types related to shade shrub proximity (**Fig. 1c**). To our knowledge, this is the first study to link natural (i.e. nonexperimental) fine-scale habitat buffering to physiologically significant stress variation in a dryland moss. Future implications of these findings include the potential that the most resilient mosses may not simply be those at high elevations in drylands, but could be scattered throughout all elevation zones within species' current ranges.

Habitat buffering hypothesis

We next discuss how the fine-scale within-site habitat structure and buffering factors explored were related to moss stress in univariate and multivariate (i.e. multiscale buffering) frameworks.

Fine-scale habitat buffering– Supporting the habitat buffering hypothesis, high-shade canopy microhabitats were the most common habitat type for *S. caninervis* across elevation zones (**Appendix S1; see Supplemental Data with this article**) and harbored mosses with the highest Fv/Fm, thereby appearing the most stress resistant. Beneath these shrub canopies, mosses were shaded over 60% of the year regardless of elevation zone (**Fig. 4c**, **Fig. 4f**). Furthermore, our novel shade buffering proxy, percent annual shade time (**Fig. 2**), appeared most important to summer stress resistance at the Mid-shrubland and High-woodland, where shade was positively related to Fv/Fm within each mesoscale topography zone (**Fig. 8a-b**). However, we found no evidence for a shade-stress relationship at the Low-scrubland (**Fig. 8c**). The lack of a shade signal in this most arid and exposed elevation zone contradicted our expectation. Possibly, shade was overpowered by other habitat buffers; on the steep N-facing zone, topography buffering may overpower the buffering effect of shade, and hydrological buffering by the drainage basin may overpower shade buffering on the Flat zone where

additional soil saturation may increase moss recovery time regardless of the shade level (**Fig. 1c**). Nonetheless, the patterns we present for fine-scale habitat buffering at the two higher elevation zones suggest that most shrub microhabitats in the Mojave Desert can provide an impressively high and physiologically meaningful shade buffer for *S. caninervis* regardless of shrub species and elevation zone.

In addition to microscale, mesoscale habitat buffering also appears important to moss stress resistance in DNWR (**Fig. 8**). We found small differences in topographical exposure (6-14% deviance in PDIR) are related to moss summer stress resistance and that, overall, northerly exposures appear to reduce stress even if slopes are gentle. For example, topography buffering appeared most important to stress at the Low-scrubland where mosses on the N-facing slope had significantly lower stress with exposure reduced by only 14% relative to the flat drainage basin (**Fig. 1c**, **Fig. 8c**). Mesoscale exposure may also be important at the High-woodland, but topographical exposure was confounded with shade here – the steep, N-facing zone was also the most shaded zone (**Fig. 8a**). Such ecological confoundedness is often unavoidable – greater shade on northerly-facing slopes is driven by topographical slope shading and higher vegetation density (Pelletier et al. 2017). However, we suggest that disentangling confounded fine-scale habitat features is less important for climate change vulnerability monitoring; determining the contribution of fine-scale vs. macroscale habitat structure to moss stress is perhaps a most critical first step to inform scales for monitoring and predicting future impacts on biocrust.

We found relationships that suggest mesoscale topography can drive unique buffering patterns for biocrust mosses that involve aspect and hydrological position that appear to influence biocrust moss distribution. For example, on the highly exposed S-facing slopes of the Low-scrubland. A mere 8% PDIR buffer (**Table 1c**) was associated with an absence of moss on our reference S-facing slope at the Low-scrubland. In fact, no mosses were found on any S-facing slopes in this elevation zone, suggesting a niche limitation for *S. caninervis* on soil surfaces receiving >1000 kJ/cm^2/yr in low elevation Mojave Desert (**Table 1c**, **Fig. 1c**). Niche boundaries in response to such small differences in exposure shed light on the sensitivity of these poikilohydric organisms and corroborate prior warnings of their potential vulnerability to shifts in climate (Ladrón de Guevara and Maestre 2022).

Multiscale habitat buffering– We also found strong support for the multiscale habitat buffering hypothesis using multivariate models which revealed microsites for which two scales of habitat structure and buffering could predict moss stress resistance. At the Mid-shrubland, 67% of moss stress was predicted by mesoscale topography zone, microhabitat shade, and their interaction (**Fig. 8b**). Similarly, 43% of stress across all elevation zones was related to mesoscale PDIR, microscale shade, and their interaction (**Fig. 7**). There may also be an interaction between elevation zone and shade in which the importance of shade increases with elevation (**Fig. 7**). Overall, these models suggest that within-site fine-scale habitat buffering can have additive and interactive effects on moss stress at multiple scales and that potential exists for interactions with the macroscale, which could likely be in response to macroclimate (driven by elevation and shading by the dominant vegetation community)**.** Specifically, it appears that effects of fine-scale shade are likely to vary based on topographical exposure, hydrological position, and elevation zone (or site) – thus we urge future researchers to anticipate such complexities in understanding moss climate stress resistance (**S2**).

In the Mojave Desert and other drylands, sheltered fine-scale habitats like N-facing slopes and soil beneath vegetation have been shown to reduce extremes in soil temperature, soil moisture, and moss thermal loading, but these studies do not link the habitat types or buffering to moss stress (Breshears et al. 1998, Bowker et al. 2000, Thompson et al. 2005, Kidron 2009). The only two studies that have linked natural variation to dryland moss photosynthetic stress also found microhabitat variation to be important, but did not compare its signal to macroscale environmental variation as we have done. The first study by Alpert (1988) used infrared gas analysis to compare carbon balance (i.e. the carbon offset between respiration and photosynthesis) of mosses cycling in and out of desiccation, and found healthier carbon budgets in N-facing, higher shaded habitats compared to S-facing, lower-shaded rock mosses in semi-arid California. The second study by Yin et al. (2017), experimentally altered shade environments in the temperate Gurbantunggut Desert of China. They measured healthier *S. caninervis* patches (e.g. greater Fv/Fm and antioxidant concentrations) under native shrub canopies compared to exposed patches after shrub-removal treatments. Our findings corroborate these studies and emphasize the potential that fine scale habitat buffering will be most relevant to these small plants if climate conditions persist or worsen.

Can biocrust mosses hide from climate change?

Many studies predict that climate change will reduce dryland moss biomass and thus the functional roles of these plants in future drylands, which may cause ecosystem-wide consequences while potentially accelerating desertification (e.g., Coe & Sparks 2014, Rodriguez-Caballero et al. 2018, Ladrón de Guevara and Maestre 2022). Predicting such climate change responses in mosses is more complex than for other plants given their unique poikilohydric physiologies and fine-scale habitats, which require different temporal (e.g. intermittent hydroperiods) and spatial (i.e., microhabitat) scales of study than for large, homiohydric tracheophytes (He et al. 2016, Ladrón de Guevara and Maestre 2022). The shelter offered by shrubs and north-facing slopes in our study are examples of such fine-scale spatial complexity. These habitat features may have important implications for climate change presenting the likelihood that (1) microrefugia created by shade shrubs and northerly slopes will best protect biocrust mosses in future climates, and that (2) such fine-scale microrefugia will likely be more important to moss long-term resilience than macrorefugia existing within or outside species ranges.

Increased drought or erratic summer precipitation is predicted to increasingly stress sensitive moss hydration cycling (e.g. Coe et al. 2012). We warn that these hydrological stressors may strengthen the moss-nurse shrub dependency presenting an additional factor of vulnerability for biocrust mosses, as some shrub species may not be resistant to climate change. However, high-shade microhabitats in our study were not restricted to a particular shrub species by elevation zone, but rather, included over thirty species. This lack of shrub specificity should reduce future vulnerability in biocrust mosses due to the prevalence of diverse and abundant shrub habitat. We must warn, however, that *S. caninervis* establishment was most frequent under shorter shrubs with low-lying canopies (<0.5 m) such as *Ambrosia dumosa*, *Coleogyne ramosissima*, and *Artemisia* spp. at the Low-scrubland, Mid-shrubland, and High-woodland, respectively. Therefore, there is risk that large patches of biocrust moss may not survive if such low-lying dominant shrub species experience die-back (e.g., Ladrón de

Guevara & Maestre 2022). Nonetheless, the previously mentioned niche shrub diversity may help alleviate impacts across many other shrub microhabitats.

Furthermore, unlike tracheophytes whose distributions are usually constrained to narrower elevational bands and single continents (Patiño & Vanderpoorten 2018), many moss species including in *S. caninervis* have broad ecological amplitude (BFNA 2007, eFloras 2023, María Ros et al. 1999), which is thought to be a function of their ability to exploit similarly buffered microhabitats existing across vastly different environments (Ladrón de Guevara & Maestre 2022) often cross-continental or disjunct locations (e.g. Brooks et al. 2023, Burge et al. 2016, Carter et al. 2016). The broad elevational and fine-scale niche we observed in *S. caninervis* in DNWR exemplifies such a pattern in which many shrub species appear to offer similarly buffered microhabitats distributed across at least three unique elevation zones. Therefore, we foresee potential that the diversity of shade shrubs spanning multiple elevation zones and plant communities in the Mojave Desert may offer widespread and common microrefugia for biocrust mosses in future climates and may serve to minimize climate impacts on moss frequency, abundance, and ecological functions in biocrust.

CONCLUSIONS

We used a natural experiment to test the habitat buffering and high-elevation refugia hypotheses for a broadly distributed, keystone biocrust moss, *Syntrichia caninervis*, within a climatically extreme part of its global distribution – the Mojave Desert. By characterizing its diverse microhabitat, aridity exposure, and photosynthetic stress resistance, we found evidence for physiologically significant summer habitat buffering in reduced-exposure microhabitats created by meso- and microscales of topography and vegetation sometimes interacting with each other or with macroscale elevation. These buffered microsites were distributed across all three elevation zones in the species' current range, failing to support the high-elevation buffering hypothesis while strongly supporting the fine-scale (within-site) habitat buffering hypothesis. We conclude that many of these buffered microsites appear as candidate microrefugia potentially capable of "hiding" this species from future climatic extremes via sheltering by shrubs and N-facing slopes. Contrary to most climate change experiments predicting low moss resiliency to increasing aridity and altered precipitation patterns in drylands, our findings present the possibility that at least one ecologically critical biocrust moss may be more prepared for a changing climate than previously thought, as long as associated shrub mortality does not significantly accelerate.

Acknowledgments

We thank the doctoral committee of TAC (Drs. Daniel Thompson, Sandra Catlins, Dale Devitt, and Peter Nelson) for assistance in the development of this University of Nevada Las Vegas (UNLV) dissertation project. We thank Amy Sprunger for logistical support in field research at the Desert National Wildlife Refuge. We thank Brian Bird for infrastructural support working within the NevCAN Network. Field sampling, lab work, and data management were aided by many volunteers to which we are grateful. The research was supported by the UNLV Summer Doctoral Research Fellowship and the American Bryological and Lichenological Society

Author Contributions

TAC conceptualized this dissertation research under advisership by LRS and DD. TAC performed all analyses, made graphics, and wrote the original draft. TAC and AR performed field investigations. TAC, AR, and JG performed the laboratory investigation. All authors contributed to manuscript review and editing.

Data Availability

Data and R code used in this manuscript are openly available on GitHub at https://github.com/TreesaClark/Syntrichia_summer_stress.

Supporting Information

Additional supporting information may be found online in the Supporting Information section at the end of the article.

S1. Microhabitat type relative frequency by elevation zone site.

S2. Correlation and regression table for the full habitat buffering model.

S3. Residual diagnostic plots for the reduced habitat buffering regression model.

Literature Cited

Abed, R. M. M., A. Tamm, C. Hassenrück, A. N. Al-Rawahi, E. Rodríguez-Caballero, S. Fiedler, S. Maier, and B. Weber. 2019. Habitat-dependent composition of bacterial and fungal communities in biological soil crusts from Oman. *Scientific Reports* 9: 1–16.

Ackerman, T. L. 2003. A flora of the Desert National Wildlife Range, Nevada. *Mentzelia: The Journal of the Nevada Native Plant Society* 7.

Alpert, P. 1985. Distribution Quantified by Microtopography in an Assemblage of Saxicolous Mosses. *Vegetatio* 64: 131–139.

Alpert, P. 1988. Survival of a desiccant-tolerant moss, Grimmia laevigata, beyond its observed microdistributional limit. *Journal of Bryology* 15: 219–227.

Ashcroft, M. B. 2010. Identifying refugia from climate change. *Journal of Biogeography* 37: 1407–1413.

Baker, N. R. 2008. Chlorophyll Fluorescence: A Probe of Photosynthesis In Vivo. *Annual Reviews in Plant Biology* 59: 89–113.

Belnap, J., S. L. Phillips, and M. E. Miller. 2004. Pulse events and arid ecosystems: Response of desert biological soil crusts to alterations in precipitation frequency. *Oecologia* 141: 306–316.

Belnap, J. 2003. The World at Your Feet- Desert Biological Soil Crusts. *Frontiers in Ecology and the Environment* 1: 181–189.

BFNA Editorial Committee ed. . 2007. Bryophyte Flora of North America: Bryophytes North of Mexico. Volumes 27. Oxford University Press, New York.

Bowker, M. A., L. R. Stark, D. N. McLetchie, and B. D. Mishler. 2000. Sex Expression, Skewed Sex Ratios, and Microhabitat Distribution in the Dioecious Desert Moss Syntrichia caninervis (Pottiaceae). *American Journal of Botany* 87: 517–526.

Bowker, M. A., J. Belnap, B. Büdel, C. Sannier, N. Pietrasiak, D. J. Eldridge, and V. Rivera-Aguilar. 2016. Controls on Distribution Patterns of Biological Soil Crusts at Micro- to Global Scales. *In* B. Weber, B. Büdel, and J. Belnap [eds.], Biological Soil Crusts: An Organizing Principle in Drylands, 173–197. Springer International Publishing, Switzerland.

Bowker, M. A., F. T. Maestre, and R. L. Mau. 2013. Diversity and Patch-Size Distributions of Biological Soil Crusts Regulate Dryland Ecosystem Multifunctionality. *Ecosystems* 16: 923–933.

Breshears, D. D., J. W. Nyhan, C. E. Heil, and B. P. Wilcox. 1998. Effects of woody plants on microclimate in a semiarid woodland: Soil temperature and evaporation in canopy and intercanopy patches. *International Journal of Plant Sciences* 159: 1010–1017.

Brinda, J. C., C. Fernando, and L. R. Stark. 2011. Ecology of Bryophytes in Mojave Desert Biological Soil Crusts: Effects of Elevated CO2 on Sex Expression, Stress Tolerance, and Productivity in the Moss Syntrichia caninervis Mitt. *In* Z. Tuba, N. G. Slack, and L. R. Stark [eds.], Bryophyte Ecology and Climate Change, 169–189. Cambridge University Press, Cambridge.

Brinda, J. C., J. A. Jauregui-lazo, M. J. Oliver, and B. D. Mishler. 2021. Notes on the genus Syntrichia with a revised infrageneric classification and the recognition of a new genus Syntrichiadelphus (Bryophyta , Pottiaceae). *Phytologia* 103: 90–103.

Brooks, J. R., and J. A. Jauregui-Lazo. 2023. Syntrichia lithophila, a fantastic new amphitropical disjunct from Oregon, United States. *The Bryologist* 126: 221–225.

Burge, D. O., J. H. Thorne, S. P. Harrison, B. C. O'Brien, J. P. Rebman, J. R. Shevock, E. R. Alverson, et al. 2016. Plant Diversity and Endemism in the California Floristic Province. *Madroño* 63: 3–206.

Carter, B. E., B. Shaw, and A. J. Shaw. 2016. Endemism in the moss flora of North America. *American Journal of Botany* 103: 769–779.

Clark, T. A. 2020. Can desert mosses hide from climate change? The Ecophysiological Importance of Habitat Buffering & Water Relations to the Stress Response of a Keystone Biocrust Moss. University of Nevada Las Vegas.

Clark, T. A. 2012. Bryophyte Floristics and Ecology in Grand Canyon National Park.

Coe, K. K., J. Belnap, E. E. Grote, and J. P. Sparks. 2012. Physiological ecology of desert biocrust moss following 10 years exposure to elevated CO2: Evidence for enhanced photosynthetic thermotolerance. *Physiologia Plantarum* 144: 346–356.

Coe, K. K., J. Belnap, and J. P. Sparks. 2012. Precipitation-driven carbon balance controls survivorship of desert biocrust mosses. *Ecology* 93: 1626–1636.

Coe, K. K., J. L. Greenwood, M. L. Slate, J. C. Brinda, K. M. Fisher, B. D. Mishler, M. A. Bowker, et al. 2020. Strategies of Desiccation Tolerance vary across Life Phases in the Moss Syntrichia

caninervis. *American Journal of Botany* 108.2: 249–262.

Coe, K. K., and J. P. Sparks. 2014. Physiology-based prognostic modeling of the influence of changes in precipitation on a keystone dryland plant species. *Oecologia* 176: 933–942.

Cruz de Carvalho, R., C. Branquinho, and J. M. da Silva. 2011. Physiological consequences of desiccation in the aquatic bryophyte Fontinalis antipyretica. *Planta* 234: 195–205.

Demmig-Adams, B., and W. W. Adams. 2006. Photoprotection in an ecological context: The remarkable complexity of thermal energy dissipation. *New Phytologist* 172: 11–21.

Douma, J. C., and J. T. Weedon. 2019. Analysing continuous proportions in ecology and evolution: A practical introduction to beta and Dirichlet regression. *Methods in Ecology and Evolution* 10: 1412–1430.

DRI. 2020. Desert Research Institute, Western Regional Climate Center. *Nevada EPSCOR Sheep Range Transect Stations*. Website http://wrcc.dri.edu/SRtransect/ [accessed 1 March 2020].

Duning, J. B. 1995. Spatially explicit population models: current forms and future uses. *Ecological Applications* 5: 3–11.

eFloras. 2023. eflora Moss Flora of China, Volume 2. Missouri Botanical Garden and Harvard University Herbaria, St. Louis, MO and Cambridge, MA.

Ekwealor, J. T. B., T. A. Clark, O. Dautermann, A. Russell, S. Ebrahimi, L. R. Stark, K. K. Niyogi, and B. D. Mishler. 2021. Natural ultraviolet radiation exposure alters photosynthetic biology and improves recovery from desiccation in a desert moss. *Journal of Experimental Botany* 72: 4161–4179.

Elbert, W., B. Weber, S. Burrows, J. Steinkamp, B. Büdel, M. O. Andreae, and U. Pöschl. 2012. Contribution of cryptogamic covers to the global cycles of carbon and nitrogen. *Nature Geoscience* 5: 459–462.

Fisher, K., J. S. Jefferson, and P. Vaishampayan. 2020. Bacterial Communities of Mojave Desert Biological Soil Crusts Are Shaped by Dominant Photoautotrophs and the Presence of Hypolithic Niches. *Frontiers in Ecology and Evolution* 7: 1–11.

Fligner, M. A., and T. J. Killeen. 1976. Distribution-free two sample tests for scale. *Journal of American Statistical Association* 71: 210–213.

Fox, J., and S. Weisberg. 2019. An R Companion to Applied Regression. 3rd ed. Sage, Thousand Oaks, CA.

Gignac, L. D. 2001. Bryophytes as Indicators of Climate Change. *The Bryologist* 104: 410–420.

Gotelli, N. J., and A. M. Ellison. 2013. A Primer of Ecological Statistics. 2nd ed. Sinauer Associates, Inc., Sunderland, MA, USA.

Greenwood, J. L., L. R. Stark, and L. P. Chiquoine. 2019. Effects of rate of drying, life history phase, and ecotype on the ability of the moss, *Bryum argenteum*, to survive desiccation events and the influence on conservation and selection of material for restoration. *Frontiers in Ecology and Evolution* 7: 388.

Guo, Y., and Y. Zhao. 2018. Effects of storage temperature on the physiological characteristics and vegetative propagation of desiccation-tolerant mosses. *Biogeosciences* 15: 797–808.

Hájek, T., and E. Vicherová. 2014. Desiccation tolerance of Sphagnum revisited: a puzzle resolved. *Plant Biology* 16: 765–773.

Hamerlynck, E. P., Z. Tuba, Z. Csintalan, Z. Nagy, G. Henebry, and D. Goodin. 2000. Diurnal variation in photochemical dynamics and surface reflectance of the desiccation-tolerant

moss, Tortula ruralis. *Plant Ecology* 151: 55–63.

He, X., K. S. He, and J. Hyvönen. 2016. Will bryophytes survive in a warming world? *Perspectives in Plant Ecology, Evolution and Systematics* 19: 49–60.

Kassambara, A. 2023. rstatix: Pipe-Friendly Framework for Basic Statistical Tests. *R package version 0.7.2*. Website https://cran.r-project.org/package=rstatix.

Kelly, A. E., and M. L. Goulden. 2008. Rapid shifts in plant distribution with recent climate change. *Proceedings of the National Academy of Sciences* 105: 11823–11826.

Kidron, G. J. 2009. The effect of shrub canopy upon surface temperatures and evaporation in the Negev Desert. *Earth Surface Processes and Landforms* 34: 123–132.

Ladrón De Guevara, M. and F. T. Maestre. 2022. Ecology and responses to climate change of biocrust-forming mosses in drylands. Journal of Experimental Botany 73: 4380–4395.

Lafuente, A., M. Berdugo, M. Ladrón de Guevara, B. Gozalo, and F. T. Maestre. 2018. Simulated climate change affects how biocrusts modulate water gains and desiccation dynamics after rainfall events. *Ecohydrology* 11: 1–10.

Lakens, D. 2013. Calculating and reporting effect sizes to facilitate cumulative science: a practical primer for t-tests and ANOVAs. *Frontiers in Psychology* 4: 863.

Langsrud, Ø. 2003. ANOVA for unbalanced data: Use Type II instead of Type III sums of squares. *Statistics and Computing* 13: 163–167.

Li, X.-R., R.-L. Jia, Z.-S. Zhang, P. Zhang, and R. Hui. 2018. Hydrological response of biological soil crusts to global warming: A ten-year simulative study. *Global Change Biology* 24: 4960–4971.

Logan, M. 2010. Biostatistical Design and Analysis Using R. Wiley-Blackwell, Oxford, UK.

Long, J. S., and L. H. Ervin. 2000. Using Heteroscedasticity Consistent Standard Errors in the Linear Regression Model. *The American Statistician* 54: 217–224.

McCune, B., and D. Keon. 2002. Equations for potential annual direct incident radiation and heat load. *Journal of Vegetation Science* 13: 603–606.

McCune, B. 2007. Improved estimates of incident radiation and heat load using non-parametric regression against topographic variables. *Journal of Vegetation Science* 18: 751–754.

Mensing, S., S. Strachan, J. Arnone, L. Fenstermaker, F. Biondi, D. Devitt, B. Johnson, et al. 2013. A network for observing great basin climate change. *EOS Transactions American Geophysical Union* 94: 105–106.

Müller, P., X. P. Li, and K. K. Niyogi. 2001. Non-photochemical quenching. A response to excess light energy. *Plant Physiology* 125: 1558–1566.

Munzi, S., Z. Varela, and L. Paoli. 2019. Is the length of the drying period critical for photosynthesis reactivation in lichen and moss components of biological soil crusts? *Journal of Arid Environments* 166: 86–90.

Murchie, E. H., and T. Lawson. 2013. Chlorophyll fluorescence analysis: A guide to good practice and understanding some new applications. *Journal of Experimental Botany* 64: 3983–3998.

Nash, T. H., S. L. White, and J. E. Marsh. 1977. Lichen and Moss Distribution and Biomass in Hot Desert Ecosystems. *The Bryologist* 80: 470–479.

NCCP. 2018. Nevada Climate Change Portal. *The Nevada Climate-Ecohydrological Assessment Network (NevCAN)*. Website http://sensor.nevada.edu/NCCP/Default.aspx.

NOAA. 2017. Climate at a Glance: Divisional Rankings. *NOAA National Centers for Environmental Information*. Website https://www.ncdc.noaa.gov/cag/ [accessed 24 February 2020].

Papadatos, S., A. C. Charalambous, and V. Daskalakis. 2017. A pathway for protective quenching in antenna proteins of Photosystem II. *Scientific Reports* 7: 1–13.

Patiño, J., and A. Vanderpoorten. 2018. Bryophyte biogeography. *Critical Reviews in Plant Sciences* 0: 1–35.

Pietrasiak, N., R. E. Drenovsky, L. S. Santiago, and R. C. Graham. 2014. Biogeomorphology of a Mojave Desert landscape - Configurations and feedbacks of abiotic and biotic land surfaces during landform evolution. *Geomorphology* 206: 23–36.

Proctor, M. C. F., and N. Smirnoff. 2011. Ecophysiology of photosynthesis in bryophytes: major roles for oxygen photoreduction and non-photochemical quenching? *Physiologia Plantarum* 141: 130–140.

Proctor, M. C. F. 2009. Physiological Ecology. *In* B. Goffinet, and J. A. Shaw [eds.], Bryophyte Biology, 237–268. Cambridge University Press, Cambridge, United Kingdom and New York, New York.

Proctor, M. C. F., M. J. Oliver, A. J. Wood, L. R. Stark, N. L. Cleavitt, B. D. Mishler, P. Alpert, et al. 2007. Desiccation-tolerance in bryophytes: a review. *The Bryologist* 110: 595–621.

Proctor, M. C. F. 2001. Patterns of desiccation tolerance and recovery in bryophytes. *Plant growth regulation* 35: 147–156.

Proctor, M. C. F., and N. Smirnoff. 2000. Rapid recovery of photosystems on rewetting desiccation-tolerant mosses: chlorophyll fluorescence and inhibitor experiments. *Journal of experimental botany* 51: 1695–704.

R-Core-Development-Team. 2023. R: A language and environment for statistical computing.

Reed, S. C., K. K. Coe, J. P. Sparks, D. C. Housman, T. J. Zelikova, and J. Belnap. 2012. Changes to dryland rainfall result in rapid moss mortality and altered soil fertility. *Nature Climate Change* 2: 752–755.

Rodriguez-Caballero, E., J. Belnap, B. Büdel, P. J. Crutzen, M. O. Andreae, U. Pöschl, and B. Weber. 2018. Dryland photoautotrophic soil surface communities endangered by global change. *Nature Geoscience* 11: 185–189.

Ros, R. M., M. J. Cano, and J. Guerra. 1999. Bryophyte checklist of northern Africa. *Journal of Bryology* 21: 207--244.

Ruxton, G. D., and G. Beauchamp. 2008. Time for some a priori thinking about post hoc testing. *Behavioral Ecology* 19: 690–693.

Scheffers, B. R., D. P. Edwards, A. Diesmos, S. E. Williams, and T. a. Evans. 2014. Microhabitats reduce animal's exposure to climate extremes. *Global Change Biology* 20: 495–503.

Seager, R., M. Ting, I. Held, Y. Kushnir, J. Lu, G. Vecchi, H.-P. Huang, et al. 2007. Model projections of an imminent transition to a more arid climate in southwestern North America. *Science* 316: 1181–1184.

Seager, R., and G. A. Vecchi. 2010. Greenhouse warming and the 21st century hydroclimate of southwestern North America. *Proceedings of the National Academy of Sciences* 14: 21277–21282.

Seppelt, R., A. Downing, K. K. Coe, Y. Zhang, and J. Zhang. 2016. Bryophytes within Biological

Soil Crusts. *In* B. Weber, B. Büdel, and J. Belnap [eds.], Biological Soil Crusts: An Organizing Principle in Drylands, 101–120. Springer International Publishing, AG Switzerland.

Shi, H., Z. Wen, D. Paull, and M. Guo. 2016. A framework for quantifying the thermal buffering effect of microhabitats. *Biological Conservation* 204: 175–180.

Stark, L. R. 2017. Ecology of desiccation tolerance in bryophytes: A conceptual framework and methodology. *The Bryologist* 120: 130–165.

Stark, L. R. 2005. Phenology of patch hydration, patch temperature and sexual reproductive output over a four-year period in the desert moss Crossidium crassinerve. *Journal of Bryology* 27: 231–240.

Stark, L. R., J. C. Brinda, and N. D. McLetchie. 2011. Effects of increased summer precipitation and N deposition on Mojave Desert populations of the biological crust moss Syntrichia caninervis. *Journal of Arid Environments* 75: 457–463.

Stovall, M. S., A. C. Ganguli, J. W. Schallner, A. M. Faist, Q. Yu, and N. Pietrasiak. 2022. Can biological soil crusts be prominent landscape components in rangelands? A case study from New Mexico, USA. *Geoderma* 410: 115658.

Takács, Z., Z. Csintalan, L. Sass, E. Laitat, I. Vass, and Z. Tuba. 1999. UV-B tolerance of bryophyte species with different degrees of desiccation tolerance. *Journal of Photochemistry and Photobiology B: Biology* 48: 210–215.

Thompson, D. B., L. R. Walker, F. H. Landau, and L. R. Stark. 2005. The influence of elevation, shrub species, and biological soil crust on fertile islands in the Mojave Desert, USA. *Journal of Arid Environments* 61: 609–629.

Warton, D. I., and F. C. K. Hui. 2011. The arcsine is asinine: the analysis of proportions in ecology. *Ecology* 92: 3–10.

Welch, B. L. 1951. On the comparison of several mean values: an alternative approach. *Biometrika* 38: 330–336.

White, H. 1980. A Heteroskedastic-Consistent Covariance Matrix Estimator and a Direct Test of Heteroskedasticity. *Econometrica* 48: 817–838.

Wickham, H. 2016. ggplot2: Elegant Graphics for Data Analysis. Springer-Verlag, New York.

Wickham, H., M. Averick, J. Bryan, W. Chang, L. D. McGowan, R. François, G. Grolemund, et al. 2019. Welcome to the {tidyverse}. *Journal of Open Source Software* 4: 1686.

Williams, A. J., B. J. Buck, D. A. Soukup, and D. J. Merkler. 2013. Geomorphic controls on biological soil crust distribution: A conceptual model from the Mojave Desert (USA). *Geomorphology* 195: 99–109.

Williams, S. E., C. Moritz, L. P. Shoo, J. L. Isaac, A. a Hoffmann, G. Langham, C. Moritz, et al. 2008. Towards an Integrated Framework for Assessing the Vulnerability of Species to Climate Change. *PLoS biology* 6: e325.

Xiao, B., K. Hu, T. Ren, and B. Li. 2015. Moss-dominated biological soil crusts significantly influence soil moisture and temperature regimes in semiarid ecosystems. *Geoderma* 263: 35–46.

Yin, B. F., Y.-M. Zhang, and A. R. Lou. 2017. Impacts of the removal of shrubs on the physiological and biochemical characteristics of Syntrichia caninervis Mitt: In a temperate desert. *Scientific Reports* 7: 1–12.

Zanatta, F., R. Engler, F. Collart, O. Broennimann, R. G. Mateo, B. Papp, J. Muñoz, et al. 2020.

Bryophytes are predicted to lag behind future climate change despite their high dispersal capacities. *Nature Communications* 11.

Zeileis, A., and T. Hothorn. 2002. Diagnostic Checking in Regression Relationships. *R News* 2: 7–10.

Zhang, F., J. A. Biederman, M. P. Dannenberg, D. Yan, S. C. Reed, and W. K. Smith. 2021. Five decades of observed daily precipitation reveal longer and more variable drought events across much of the western United States. *Geophysical Research Letters* 48: 1–11.

Zhang, J., and Y. Zhang. 2019. Ecophysiological responses of the biocrust moss *Syntrichia caninervis* to experimental snow cover manipulations in a temperate desert of central Asia. *Ecological Research* 35: 198–207.

Zhang, Y., X. Zhou, B. Yin, and A. Downing. 2016. Sensitivity of the xerophytic moss *Syntrichia caninervis* to prolonged simulated nitrogen deposition. *Annals of Botany* 117: 1153–1161.

Table 1. Habitat buffering on three scales (elevation zones, topography zones, & microhabitats)

Table 1. Macroclimate measured by the NevCAN station near each elevation zone site supporting *S. caninervis* in DNWR. **(a)** Annual (2011 – 2018) and Summer (6/1/17 – 11/6/17) daily climate means ± (SD). The maximum elevation zone buffer (**Mean buffer**) is the maximum deviance between the three life-zone means for the respective period. **(b)** Mean ± (SD) potential direct incident radiation (*Mean PDIR*) for elevation zone sites and topography zones (see **Fig. 1**). The greatest topography zone buffer (**Mean buffer**) is the greatest mean deviance between all pairwise comparisons between topography zones for each row. Topography zones varied in their sample sizes (see Fig. 1c); SD = 0 occurred on two topography zones having nearly identical exposure across microsites.

		Elevation zone sites				
		All sites	Low-scrubland (creosote, 890 m)	Mid-shrubland (blackbrush, 1670 m)	High-woodland (pinyon-juniper, 2070 m)	
a) Climate metric		**Mean (SD)**				**Mean buffer**
Air temperature (°C)	Annual	15.0 (9)	18.9 (9)	13.8 (8)	12.3 (8)	(-) 6.6 °C
	Summer	22.6 (6)	26.9 (6)	21.3 (6)	19.5 (6)	(-) 7.4 °C
Soil temperature (°C)	Annual	17.3 (11)	21.8 (11)	16.2 (10)	13.9 (10)	(-) 7.9 °C
	Summer	26.0 (7)	31.1 (7)	24.9 (7)	22.1 (6)	(-) 9.0 °C
Relative Humidity (%)	Annual	32 (19)	29 (17)	34 (19)	35 (20)	(+) 6%
	Summer	26 (13)	21 (11)	28 (14)	29 (15)	(+) 8%
Wind Speed (m/s)	Annual	3.6 (1)	4.2 (2)	3.8 (1)	2.8 (1)	(-) 1.4 m/s
	Summer	3.6 (1)	4.2 (1)	3.8 (1)	2.8 (1)	(-) 1.4 m/s
Soil moisture (θ)	Annual	6.1 (6)	2.9 (4)	5.8 (6)	14.5 (5)	(+) 11.6 θ
Precipitation (mm)[2]	Annual	186 (52)	119 (41)	160 (54)	278 (85)	(+) 159 mm/yr
	Summer[2]	92	26	81	169	(+) 143 mm
b) Topography zone		**Mean PDIR (kJ/cm²/yr)**				**Mean buffer**
Topography zones pooled		943	905	975	949	(-) 70 kJ/cm²/yr
South-facing zone[1]		1022[1]	1061[1]	999 (8)	1044 (4)	(-) 45 kJ/cm²/yr
Flat terrain zone		990	985 (0)	1000 (0)	986 (2)	(-) 15 kJ/cm²/yr
North-facing zone		856	826 (16)	926 (8)	817 (29)	(-) 109 kJ/cm²/yr

Notes: Climate metrics were measured at 2-m height except for soil temperature and moisture, measured at 1 and 4 inches subsurface, respectively. [1]The S-facing slope at the Low-scrubland site was not included in mean calculations because no mosses were found there. [2]The standard deviation of total precipitation is annual variability, and thus summer 2017 total precipitation has no standard deviation.

Figure 1. Distribution survey & sampling of populations across nested habitat gradients

Figure 1. A. Elevation range surveyed for *S. caninervis* in the Sheep Range of the Mojave Desert National Wildlife Refuge (DNWR). EPSCoR-NevCAN climate towers are shown in each elevation zone (see text). Final sample of 92 microsites (20 x 20 cm quadrats enumerated in **C**) spanned three aridity exposure gradients: elevation zones (**B**), topography zones (northerly, southerly, or flat transects, **C**), and microhabitat shade types (**D**). For each topography zone in **C**, the slope and microhabitat frequency are shown and missing squares indicate *S. caninervis* was not found for a given zone or microhabitat type. See **Figure 3** for example quadrats.

Figure 2. Sun Seeker© shade time method

Photo scoring classes: **0** (no shade), **1** (1 – 25%), **2** (25 - 50%), **3** (50 - 75%), **4** (> 75%)

Annual shade index = sum of 7 scored photos = 18/28 = 64% of the year shaded

Figure 2. Microsite annual shade time was quantified using seven photos taken on a smartphone using the *Sun Seeker© Solar AR Tracker* app. The phone was held approximately 1 cm above the center of each microsite quadrat (***center photo***) and the photos encompass the solar arc from 6 am to 8 pm (***red circles***), which collectively captures the annual solar window from summer solstice (***red lines***) to winter solstice (blue lines) for a single microsite. Photos are scored using a 5-scale percent shade class by assessing the area of all shade objects intersecting each 2-h solar window (***white dashed boxes and triangles***). Percent annual shade time is the sum of classes for all seven photos divided by 28, the maximum possible (i.e. for a habitat shaded 75-100% of the year). The microsite shown here is shaded 64% of the year ((4 + 4 + 1 + 2 + 1 + 2 + 4 = 18)/28 = 0.64 x 100). Note: the center photo in this figure showing the smartphone position is not located at the microsite of the illustrated solar window.

Figure 3. Sampling and stress assay methods for small biocrust moss

A. Moss microhabitat quadrat placement (20 x 20 cm microsites, dry moss)

Low-scrubland | Mid-shrubland | High-woodland

B. Sampling

Shoots ✹) plucked from all patches (dry).

C. Moss bouquet assembly

Washing & trimming field shoots (dissecting scope).

Shoots placed on leaf clip with moistened filter paper.

D. Summer stress assay (Fv/Fm)

Batch processing 24 leaf clips for dark adapting (close clips) prior to chlorophyll fluorescence stress assay.

Figure 3. Methods for microhabitat selection at three elevation zone sites showing moss abundance gradient (**A**), field shoot sampling (**B**), moss bouquet assembly (**C**), and the photosynthetic efficiency assay (**D**) to test summer stress in a small desert moss.

Figure 4. Shade buffering & moss stress across three scales of habitat structure.

Figure 4. Boxplots of shade time (top row) and summer stress (bottom row) measured in 92 *Syntrichia caninervis* microhabitats pooled by scale of habitat structure: elevation zone (**A & D**), topography zone (**B & E**), and microhabitat type (**C & F**). Overlays include raw data (black points) and respective means (white points) with SE bars. Welch's ANOVA results, R^2, and Cohen's F (*Eff*) are shown for each panel. Letters indicate familywise Games-Howell significant differences between groups ($P < 0.05$) in post-hoc testing. Denominator degrees of freedom in F-statistics do not reflect sample size due to Welch's heterogeneity correction.

Figure 5. Patch appearance & shoot mortality: healthy & stressed *S. caninervis* in three Mojave Desert elevation zones

Figure 5. *Syntrichia caninervis* patches (i.e. shoot colonies) by elevation zone site (***rows***) before sampling (**A**), in leaf clips during Fv/Fm assay (**B, C**). Healthy and mildly stressed shoots (***green arrows***) when wet (***A, Low & High sites***), often appeared green (**A**) or blackish-red from protective pigments (***A, Low site***). Severe stress can appear orange-green (***A, High site***). Dead (chlorotic) shoots appear orange (***A, red arrows***). **B.** Subset of variously stressed shoots (chlorophyll fluorescence stress metric, ***Fv/Fm, listed for each sample***) from topography-shrub microsites (***labels***) by elevation zone site. **C.** Orange leaf tips suggest stressed shoots while Fv/Fm reveals healthy photosynthetic efficiency (**Fig. 6**), showing the importance of photosynthetic stress assays over visual stress surveys.

 Figure 6. *Syntrichia* summer stress distribution by elevation zone site

2
Figure 6. Frequency histogram of biocrust moss (*Syntrichia caninervis*) microhabitats with
severe, moderate, and mild levels of photosynthetic stress (estimated by the chlorophyll
fluorescence metric, **Fv/Fm**) in 92 microsites colored by elevation zone site (see **Fig. 1**). Each bin
is open on its maximum value (i.e. the bin [0, 0.1) does not include 0.1). Four colored bands
indicate moss stress categories to aid climate vulnerability assessment in the Mojave Desert.
Healthy mosses vary in their maximum Fv/Fm, so we have broadened "healthy" to begin at 0.6
(*see text*).
10

Figure 7. Summer stress vs habitat buffering proxies (elevation, PDIR, shade)

2
3
Figure 7. A. Relationship of summer moss stress (lower Fv/Fm values indicate higher stress) of
92 *Syntrichia caninervis* microsites to habitat buffering proxies measured at three spatial scales:
macroclimate (**elevation zone site: *point colors***), mesoscale potential direct incident radiation
(***PDIR: circle size***), microscale shade time (***Shade.Index** in B*), and their interaction (**:**). The OLS
regression line for shade vs Stress is plotted with a 95% confidence interval in grey, and the
interaction of elevation with shade is illustrated by varying slopes (*not tested*) of three OLS best
fit lines plotted for each elevation zone. Elevation is shown graphically, but could not be
included in the final model due to its multicollinearity with shade (see full model in **S2**). **B.** OLS
regression results; residual plots are in **S3**.
13

Figure 8. Three elevation zone models and plots illustrate the linear relationship of summer moss stress of *Syntrichia caninervis* to topography (***Topog.***), annual shade time (***Shade***), and their interaction (***Intera.***). HCCM-ANCOVA results for each elevation zone (***panels***) include alpha significance for each term in the full model (**bold *P < 0.05***), R^2 for the reduced model (including only significant terms), and best fit lines with 95% C.I. (***grey bands***) from OLS regression. ***Note***: no mosses occurred on the S-facing slope of the Low-scrubland (see **Fig. 1c**).

APPENDIX

Supplement 1. Microhabitat type relative frequency by elevation zone site

Relative frequency (proportion by elevation zone site) of three microhabitat types (shrub proximity classes: canopy, dripline, interspace) supporting *Syntrichia caninervis* in the DNWR elevation zone sites. Surveying for microsite distribution was conducted within a 2-km radius of each elevation zone climate tower and should be representative of elevation zone patterns at large in the Mojave Desert.

Supplement 2. Correlation and regression table for full habitat buffering model
A. Pearson's correlations table between regression predictors in the following models. **B**. Full
OLS linear regression model of the relationship of summer moss stress (Fv/Fm) in *Syntrichia*
caninervis and three habitat buffering proxies: elevation zone elevation (***Elevation***), potential
direct incident radiation (***PDIR***), microscale shade time (***Shade.Index***), and their interactions.
Elevation and shade suffer from multicollinearity such that this model is appropriate only for
prediction rather than precise coefficient estimation (***see Analysis***). **C.** Corresponding ANOVA
model (Type I errors) results, which do not suffer from multicollinearity; residual plots
reasonably met assumptions and are not shown.

B Full regression model coefficients & standard errors

	Estimate	Std. Error
(Intercept)	0.29541280	1.68464416
Elevation	0.00191438	0.00141486
PDIR	0.27697979	1.77041616
Shade.Index	-0.02540937	0.09569381
Elevation:PDIR	-0.00231605	0.00147387
Elevation:Shade.Index	-0.00005336	0.00006671
PDIR:Shade.Index	0.02625171	0.10293684
Elevation:PDIR:Shade.Index	0.00006893	0.00007060

C ANOVA Table

	Df	Sum Sq	Mean Sq	F value	Pr(>F)	
Elevation	1	0.0402	0.0402	3.710	0.05747	.
PDIR	1	0.4068	0.4068	37.514	0.00000002808	***
Shade.Index	1	0.4585	0.4585	42.288	0.00000000536	***
Elevation:PDIR	1	0.0034	0.0034	0.314	0.57687	
Elevation:Shade.Index	1	0.1193	0.1193	11.006	0.00134	**
PDIR:Shade.Index	1	0.0956	0.0956	8.813	0.00390	**
Elevation:PDIR:Shade.Index	1	0.0103	0.0103	0.953	0.33172	
Residuals	84	0.9108	0.0108			

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Residual standard error: 0.1041 on 84 degrees of freedom
Multiple R-squared: 0.5546,   Adjusted R-squared: 0.5175
F-statistic: 14.94 on 7 and 84 DF, p-value: 0.000000000001615

Supplement 3. Residual diagnostic plots for reduced habitat buffering regression model
Residual diagnostic plots (*R stats::plot(Y~X1*X2)*) for the OLS (ordinary least squares) regression
model for moss stress (Fv/Fm) as a function of two habitat buffering predictors and their
interaction term: potential direct incident radiation (PDIR) and annual shade time. See *Analysis*
for details on model selection and **S2** for the full model. Model statistics and plot are in **Figure**
7.
7

8
9