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Male germ cell-associated kinase (MAK) is required for axoneme formation during ciliogenesis in zebrafish photoreceptors

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Summary statement (up to 30 words)

Male germ cell-associated kinase (MAK) is a cilium-associated serine/threonine kinase that promotes axoneme development during ciliogenesis in zebrafish photoreceptors to ensure intracellular protein transport and photoreceptor survival.

Abstract

Vertebrate photoreceptors are highly specialized retinal neurons and have cilium-derived membrane organelles called outer segments (OS), which function as platforms for phototransduction. Male germ cell-associated kinase (MAK) is a cilium-associated serine/threonine kinase, and its genetic mutation causes photoreceptor degeneration in mice and retinitis pigmentosa in humans. However, the role of MAK in photoreceptors is not fully understood. Here, we report that zebrafish *mak* mutants show rapid photoreceptor degeneration during embryonic development. In *mak* mutants, both cone and rod photoreceptors completely lack OSs and undergo apoptosis. Interestingly, zebrafish *mak* mutants fail to generate axonemes during photoreceptor ciliogenesis, whereas basal bodies and transition zones are specified. These data suggest that MAK plays a specific role in axoneme development in zebrafish, in contrast to mouse *Mak* mutants, which have elongated photoreceptor axonemes. Furthermore, the kinase activity of MAK is critical in ciliary axoneme development and photoreceptor survival. Thus, MAK is required for ciliogenesis and OS formation in zebrafish photoreceptors to ensure intracellular protein transport and photoreceptor survival.

Introduction

The vertebrate photoreceptor is highly compartmentalized to form specialized structures related to phototransduction (Fain et al., 2010). The outer segment (OS) is a specialized cilium of the photoreceptor, in which multiple photoreceptive membrane discs are regularly stacked to accommodate phototransduction molecules such as rhodopsin and opsins (Bachmann-Gagescu and Neuhauss, 2019; Chen et al., 2021). The inner segment (IS) is a mitochondria-enriched region between the OS and the nucleus. Finally, photoreceptors have a specialized synaptic structure in the most basal region, which forms the outer plexiform layer (OPL) and mediates transmission of electric signals to horizontal cells and bipolar cells (Mercer and Thoreson, 2011).

Upon photoactivation in vertebrate photoreceptors, opsins or rhodopsin absorb light energy and change their conformation, which activates Transducin, and then cGMP phosphodiesterase 6 (PDE6). PDE6 catalyzes the hydrolysis of a second messenger, cGMP. Reduction of the cGMP concentration promotes dissociation of cGMP from a cyclic nucleotide binding channel (CNG), leading to the closure of CNG, which hyperpolarizes photoreceptor membranes. All of these phototransduction molecules are specifically expressed in photoreceptors and properly transported into the OS, which is essential for photoreceptor functions (Athanasiou et al., 2018; Gulati and Palczewski, 2023; Karan et al., 2008; Wang and Deretic, 2014). Indeed, mislocalization of opsins leads to photoreceptor degeneration, although detailed mechanisms are not fully understood (Alfinito and Townes-Anderson, 2002; Lopes, 2010; Rohrer et al., 2005).

The photoreceptor OS is a highly specialized primary cilium, which consists of stacked membrane discs around a microtubule-based backbone called the axoneme (Bachmann-Gagescu and Neuhauss, 2019; Chen et al., 2021). The axoneme is anchored from the basal body in the IS of photoreceptors and extended apically through the connecting cilium. The connecting cilium bridges the IS and the OS, through which phototransduction molecules are transported from the ER, then to the Golgi and into the OS. Importantly, the connecting cilium is equivalent to the transition zone in other cell types and functions as a gating system (Park and Leroux, 2022), through which OS resident proteins are sorted and transported to

the OS by the intraflagellar transport (IFT) complex (Klena and Pigino, 2022) and BBSome,

composed of eight Bardet-Biedl syndrome (BBS) proteins (Tian et al., 2023).

The process of ciliary development is called ciliogenesis, which consists primarily of

three steps: centriole (basal body) docking to the apical plasma membrane, establishment of

a transition zone, and ciliary axoneme extension (Baehr et al., 2019). A number of factors

are indispensable for ciliogenesis. For example, CC2D2A is required by basal body

maturation (Gorden et al., 2008). BBS1 is required for ciliary assembly and maintenance

(Davis, 2007). RP1 has microtubule extension activity that promotes axoneme extension (Liu

et al., 2004). Because cilia work as "rail tracks" in IFT-mediated trafficking, disruption of

cilia affects IFT and further causes photoreceptor degeneration, suggesting that the structural

and functional integrity of the connecting cilium is indispensable for photoreceptor functions

and maintenance. Although many other factors reportedly regulate photoreceptor cilia

integrity (Cideciyan et al., 2007; den Hollander et al., 2006; Gao J., 2002; Gorden et al., 2008;

Kakakhel et al., 2020; Rao et al., 2015; Zhu et al., 2021), the underlying mechanisms are not

fully understood.

Male germ cell-associated kinase (MAK) belongs to the MAK/ICK/MOK

serine/threonine kinase family (Chen et al., 2013). MAK was originally identified by cross-

hybridization with tyrosine kinase v-ros in rat testicular cells, so it was considered to be a

spermatogenesis regulator (Matsushime et al., 1990). Furthermore, MAK functions as a

coactivator of androgen receptors (ARs) to promote AR-mediated signaling, which is

associated with prostate tumorigenesis (Ma et al., 2006; Wang and Kung, 2012; Xia et al.,

2002). In addition to the role of MAK in male reproductive organs, MAK is expressed in

developing rod photoreceptors (Blackshaw et al., 2004). In *Mak* knockout mice,

photoreceptors show abnormally elongated axonemes and malformation of membrane discs

in the OS, leading to photoreceptor degeneration (Omori et al., 2010). In humans, *MAK*

mutations cause retinitis pigmentosa (RP), in which patients progressively lose their vision

(Ozgul et al., 2011; Stone et al., 2011). However, human patients carrying *MAK* mutations

do not show elongation of the photoreceptor layer under optical coherence tomography (OCT)

scanning (Stone et al., 2011), which may be different from photoreceptor phenotypes of *Mak*

knockout mice. Thus, it is important how *mak* mutations affect ciliary regulation in

photoreceptors among vertebrate species.

In this study, we found that zebrafish photoreceptors of *mak* mutants degenerate during

embryonic development. In *mak* mutants, both rods and cones undergo apoptosis, although

their degeneration processes differ. Interestingly, *mak* mutants fail to form axonemes in

photoreceptor cilia, whereas basal bodies and transition zones are specified, which is the

opposite of mouse *Mak* knockout photoreceptors, with elongated ciliary axonemes.

Furthermore, both cones and rods completely lack OSs in *mak* mutants, leading to ectopic

distribution of opsins. MAK is essential to form axonemes and OSs in photoreceptors. Finally,

we confirmed that MAK kinase activity is critical for axoneme formation and photoreceptor

survival. Thus, MAK is essential for ciliogenesis, OS formation, and photoreceptor survival.

Results

Zebrafish *pday* mutants carry a genetic mutation in *mak* gene and show photoreceptor degeneration

Zebrafish *payday* (*pday*) mutants display no visual response during the embryonic stage (Muto et al., 2005). We selected *pday* homozygous mutant embryos using the optokinetic response (OKR) assay and examined their phenotypes. At 6 dpf, *pday* mutants showed no apparent morphological defect in body structure (Fig. 1A), but did not survive until adulthood, suggesting a lethal mutation. However, under intensive feeding conditions, some mutants did survive until 2 months post-fertilization (mpf), and displayed scoliosis, cardiac edema, and smaller body size (Fig. 1A).

Next, we mapped the *pday* mutant locus with simple sequence-length polymorphism (SSLP) markers (Knapik et al., 1998; Shimoda et al., 1999). The *pday* mutation was mapped between two SSLP markers z23011 (1/186 miosis) and z13695 (3/186 mitosis), so the mutant gene is restricted to a genomic region between 8.517 and 9.123 Mb on chromosome 24. In this genomic region, 10 genes including *mak* were annotated in the zebrafish genomic database (GRCz10, Ensemble release 80). Since *Mak* mutations cause photoreceptor degeneration in mice (Omori et al., 2010) and humans (Ozgul et al., 2011; Stone et al., 2011), we focused on the *mak* gene. First, we designed a new polymorphic marker, called Mak-N1, located around 20 bp upstream of the first exon of the *mak* gene. The recombination rate of Mak-N1 for *pday* mutation was 0 of 186 meiosis, suggesting that *mak* is a strong candidate for the *pday* mutant gene. Next, we cloned *mak* cDNA prepared from wild-type embryos, and found that *mak* cDNA consists of 15 exons (Fig. 1B) and that there are two isoforms of *mak* mRNAs. One has and the other lacks exon 14 due to alternative splicing. Then, we focused on the longest isoform, which has the exon 14 and it was annotated as the full-length isoform on the zebrafish genomic database (mak-201, GRCz11, Ensemble release 110). We amplified *mak* full-length cDNA from *pday* mutant mRNA by PCR. However, we did not recover *pday* mutant full-length cDNA, but only a partial cDNA that covers the C-terminal coding region from the middle of exon 7 to exon 15. Sequencing this partial cDNA fragment revealed no mutation in the coding region from exons 8 to 15 in *pday* mutants. Next, we

amplified six DNA fragments from the *pday* mutant genome by PCR, each of which covers
six coding exons (exons 2 – 7). Sequencing of these fragments revealed that a non-sense
mutation occurs in exon 4 of the *mak* gene in *pday* mutants, which causes a premature
termination codon at 70R (Fig. 1B). These data suggest that the *pday* mutant gene encodes
MAK.

Next, we examined retinal phenotypes of wild-type and *pday* mutant retinas at 3,
4, 5, and 6 dpf (Fig. 1C, 1C'). In wild-type retinas at 3 dpf, photoreceptors differentiate to
form the outer nuclear layer (ONL). The OS is formed in the apical region of the ONL (Fig.
1C'). In *pday* mutants, the ONL was formed; however, densely labeled nuclei were observed
in the basal region of the ventro-central ONL (Fig. 1C, 1C'), and the OS was markedly
reduced (Fig. 1C'). In wild-type photoreceptors at 4 dpf, the OS was elongated. However, in
pday mutants, the ONL was decreased in thickness (Fig. 1C, 1C'), and no OS-like structure
was observed in the apical region (Fig. 1C'). In addition, there were pyknotic nuclei (Fig.
1C') and bubble-like structures (Fig. 1C, 1C'), mainly in the dorso-central ONL. In wild-
type embryos at 5 dpf, the ONL was divided into two subnuclear layers (Fig. 1C'), the apical
and basal regions of which contain nuclei of Blue/Green/Green cones and rods/UV cones,
respectively (D'Orazi et al., 2020; Oel et al., 2020). However, in *pday* mutants, the dorso-
central ONL drastically decreased in thickness (Fig. 1C) and contained flattened nuclei (Fig.
1C, 1C'). In wild-type embryos at 6 dpf, photoreceptors showed more mature shapes in the
ONL. However, in *pday* mutants, most photoreceptor nuclei disappeared in the dorso-central
retina (Fig. 1C), so there are very flat nuclei in the outermost region of the retina, which are
difficult to define as the ONL or horizontal cells (Fig. 1C'). Interestingly, in *pday* mutants,
ONL thickness was less affected in the retinal region more ventral to the optic nerve head,
even at 6 dpf (Fig. 1C), although the OS is absent and bubble-like structures are occasionally
observed (Fig. 1C). Thus, the sensitivity of photoreceptor degeneration differs between the
dorso-central region and the ventral region of the neural retina. In addition, other retinal
layers, namely the inner nuclear layer (INL), the inner plexiform layer (IPL), and the retinal
ganglion cell layer (RGCL) seemed to be intact in *pday* mutants from 3 to 6 dpf (Fig. 1C),
suggesting photoreceptor-specific degeneration in *pday* mutants.

To confirm photoreceptor degeneration in mutant retinas at 6pf, we introduced a

transgenic line *Tg[crx:eGFP]*, which expresses eGFP in photoreceptor precursors and

mature rod and cone photoreceptors under control of the *cone-rod homeobox* (*crx*) promoter

(Suzuki et al., 2013) (Fig. 1D). In wild-type retinas, eGFP signals were detected in the ONL

at 6 dpf. However, in *pday* mutant retinas, eGFP signals disappeared in the dorso-central

retina at 6 dpf (Fig. 1D). Measurements of the thickness of eGFP-positive areas revealed that

photoreceptor degeneration is more severe in the dorso-central retina than in the ventral retina

at 6 dpf (Fig. 1E). Thus, photoreceptors undergo degeneration in *pday* mutants.

Both rods and cones degenerate in *pday* mutants

In human *MAK*-associated RP patients, rods degenerate more prominently (Ozgul et al., 2011;

Stone et al., 2011). To evaluate rod and cone degeneration separately, we generated double

transgenic lines, *Tg[rho:NLS-eGFP; gnat2:NLS-tdTomato*], to visualize rods and cones with

eGFP and tdTomato fluorescence, respectively, and introduced them into *pday* mutants (Fig.

2-S1A). In wild-type retinas, eGFP expression in rods was strong and dense in the ventral

ONL but relatively weak and sparse in the dorso-central ONL at 3 dpf (Fig. S1A). On the

other hand, eGFP expression was very faint in *pday* mutant retinas under the same conditions.

However, when the laser intensity was increased, eGFP expression was detected (Fig. S1A),

suggesting that rods are maintained in 3 dpf *pday* mutant retinas, although the rhodopsin

promoter does not effectively drive *eGFP* mRNA or else translation of *eGFP* mRNA to eGFP

protein was affected in *pday* mutants. Therefore, in later experiments using *Tg[rho:NLS-*

eGFP], we applied higher laser intensity to *pday* mutant scanning, and compared rod

phenotypes between *pday* mutants and their wild-type siblings.

Since photoreceptor degeneration is more severe in the dorso-central retina,

compared with the ventral retina at 6 dpf (Fig. 1D, 1E), we visualized morphology of rods

and cones using *Tg[rho:NLS-eGFP; gnat2:NLS-tdTomato*] at 3, 4, 5, and 6 dpf (Fig. 2A, 2B),

and focused on the dorsal and ventral ONL regions (Fig. 2C-F). First, we examined rods (Fig.

2A). In wild-type retinas at 3 dpf, rods were sparse in the dorsal ONL (Fig. 2C), but densely

produced in the ventral ONL (Fig. 2D). In *pday* mutants, many rods show very round nuclei

in both the dorsal and ventral ONL (Fig. 2C and 2D). Interestingly, eGFP signals were

predominantly localized in nuclei of wild-type rods because eGFP was tagged with a nuclear

localization sequence (NLS); however, eGFP signals were often excluded from the round

nuclei of *pday* mutant rods, suggesting apoptotic-like pyknotic nuclei of *pday* mutant rods.

After 4 dpf, in wild-type retinas, rods progressively increased in number and aligned along

the dorsal ONL. Their nuclei are positioned in the most basal region of the ONL, and rods

extend a process to form the OS in the apical region of the ONL (Fig. 2C). On the other hand,

rods are more densely differentiated in the ventral ONL (Fig. 2D). However, in *pday* mutants

at 4 dpf, rods almost disappeared in both the dorsal and ventral ONL. Only a few rods were

retained, but displayed pyknotic nuclei or were excluded from the ONL (Fig. 2C and 2D). In

pday mutants at 5 and 6 dpf, rods disappeared in most regions of the ONL (Fig. 2C and 2D),

except in the peripheral retina adjacent to the ciliary marginal zone (CMZ) where new

photoreceptors continued to be produced by retinal stem cells. So, rods are eliminated soon

after they are specified from the CMZ. To understand the spatio-temporal pattern of rod

degeneration, we counted the number of rod nuclei along the DV axis of the retina (Fig. S1B).

The number of rods progressively decreased in the central ONL of *pday* mutant retinas and

were completely eliminated, except in the peripheral region near the CMZ area at 5 and 6

dpf.

Next, we examined cones at 3, 4, 5 and 6 dpf (Fig. 2B). In wild-type retinas at 3

dpf, cones differentiated uniformly in the dorsal ONL (Fig. 2E); however, cone density

became progressively more sparse toward the CMZ region in the ventral ONL (Fig. 2F), a

pattern complimentary to that of rods (Fig. 2D). In *pday* mutant retinas, cones normally

differentiated to form a spatial pattern similar to that of wild-type retinas (Fig. 2E, 2F).

However, there were tdTomato-negative rounded nuclei in both the dorsal and ventral ONL

(Fig. 2E and 2F), whose positions are identical to pyknotic-like rod nuclei (Fig. 2C and 2D).

These data confirm that rods abnormally invaded the cone cell layer in *pday* mutants at 3 dpf.

After 4 dpf, in wild-type retinas, cones form two subnuclear layers in the dorso-central ONL

(Fig. 2E). However, cone density was complimentary to rod density and became

progressively more sparse toward the CMZ region in the ventral ONL (Fig. 2F). In *pday*

mutants at 4 dpf, segregation of two cone subnuclear layers was incomplete in the dorso-

central ONL (Fig. 2E). Furthermore, cones were sparse and shortened in the ventral ONL

(Fig. 2F). In *pday* mutants at 5 dpf, cones were shortened in the dorsal ONL (Fig. 2E).

Interestingly, cone density was increased toward the CMZ in the ventral ONL of *pday*

mutants (Fig. 2F), where rods failed to be maintained (Fig. 2D). In *pday* mutants at 6 dpf,

cones were extremely fattened and there were gaps between them in the dorsal ONL (Fig.

2E), indicating that cones are eliminated. Furthermore, cones were similarly flattened in the

ventro-central ONL (Fig. 2F), whereas cones were uniformly produced and less flattened

toward the CMZ of the ventral ONL (Fig. 2F). Therefore, in *pday* mutants, a substantial

fraction of cones was retained in the central retina at 5 dpf, even though their shape became

flattened. In *pday* mutants at 6 dpf, cones eventually disappeared, especially in the dorso-

central ONL.

Taken together, these data suggest that rods undergo acute apoptosis-like cell death

in *pday* mutants from 3 to 4 dpf (Fig. 2G). On the other hand, cones undergo progressive

shrinkage after 4 dpf and eventually disappear by 6 dpf in *pday* mutants (Fig. 2G), although

cone degeneration is less severe in the peripheral region of the ventral ONL.

Both rods and cones undergo apoptosis in *pday* mutants

Condensed rod nuclei in *pday* mutants at 3dpf (Fig. 2C, 2D) are reminiscent of apoptosis, in

which chromatin condensation is a hallmark (Oberhammer et al., 1994). We performed TdT-

mediated dUTP-nick-end labeling (TUNEL) to *pday* mutant retinas at 3, 4, and 5 dpf (Fig.

3A, 3A', Fig. S2A). In wild-type retinas, the number of TUNEL-positive cells was

continuously very low in the ONL (Fig. 3A, Fig. S2A). In *pday* mutant retinas at 3 dpf,

TUNEL signals were detected prominently in the central and ventral ONL (Fig. 3A'), which

is consistent with our observation that rod pyknotic nuclei were detected at 3 dpf (Fig. 2C,

2D), suggesting that these early TUNEL-positive cells are likely to be rods. In *pday* mutant

retinas at 4 dpf, TUNEL signals increased prominently in the dorsal and central ONL (Fig.

3A'). In *pday* mutant retinas at 5 dpf, TUNEL signals were detected prominently in the dorsal

and central ONL (Fig. 3A'). Interestingly, TUNEL-positive cells were positioned in a very

thin central ONL of 5 dpf *pday* mutant retinas (Fig. 3A'), suggesting that these late TUNEL-

positive cells are likely to be cones. Number of TUNEL-positive cells was higher in *pday*

mutant ONL than wild-type ONL at 3 dpf, but the difference was not significant. However,

the number of TUNEL-positive cells was the highest in *pday* mutant ONL at 4 dpf and was

still significantly higher in *pday* mutant ONL at 5 dpf, compared with wild-type ONL (Fig.

S2A).

Next, we examined whether cones undergo apoptosis in *pday* mutants. First, we

carried out double labeling of 5-dpf, wild-type and *pday* mutant retinas with anti-zebrafish

rhodopsin (Vihtelic et al., 1999) and zpr1 (Larison and Bremiller, 1990) antibodies, which

label rod OSs and double-cone-type photoreceptors (red/green cones), respectively (Fig. 3B).

Consistent with data of *Tg[rho:NLS-eGFP; gnat2:NLS-tdTomato]* (Fig. 2), rods were only

detected in the peripheral region of the dorsal and ventral ONL (Fig. 3B'). On the other hand,

cones were observed in the peripheral region of the dorsal and ventral ONL and as very

flattened cells in the central ONL (Fig. 3B'). Next, we carried out TUNEL of 5-dpf, wild-

type and *pday* mutant retinas combined with zpr1 antibody labeling (Fig. 3C). Many TUNEL-

positive cells were observed in dorsal, central and ventral ONL (Fig. 3C'). A significant

fraction of these TUNEL-positive cells was associated with zpr1 signals (Fig. 3C'). Third,

we carried out TUNEL in 5-dpf, wild-type and *pday* mutant retinas combined with anti-

rhodopsin antibody labeling (Fig. 3D). Many TUNEL-positive cells were observed in dorsal,

central and ventral ONL (Fig. 3D'). However, rhodopsin signals were detected only in the

CMZ of dorsal and ventral ONL (Fig. 3D'), but not in the central ONL. Many TUNEL signals

were not associated with rhodopsin signals in the central ONL (Fig. 3D'). Consistent with

our observation that almost all flattened photoreceptors in the central ONL at 5 dpf are cones

(Fig. 2), these data suggest that flattened cones undergo apoptosis at or before 5 dpf. TUNEL

analysis also revealed that the number of TUNEL signals in other retinal layers, INL, and

RGCL, was not significantly different between *pday* mutants and wild-type siblings at stages

3, 4, or 5 dpf (Fig. S2B, S2C), although TUNEL number in the INL was higher in *pday*

mutants than in wild-type siblings at 4 dpf (Fig. S2B), suggesting that apoptosis is specific

to photoreceptors.

To confirm that both rods and cones undergo apoptosis in *pday* mutants, we

combined a transgenic line *Tg[hsp:mCherry-Bcl2]*, which expresses an anti-apoptotic protein

Bcl2 (Nishiwaki and Masai, 2020), with *pday* mutants and evaluated cell survival by labeling

with zpr1 (Larison and Bremiller, 1990) and zpr3 (Gao et al., 2022) antibodies, which

visualize double cone photoreceptors and rhodopsin/green opsin, respectively. Bcl2

overexpression significantly rescued survival of both cones and rods in *pday* mutants at 6 dpf

(Fig. 3E, 3F; Fig. S2D, S2E) even along the DV axis (Fig. S2F), suggesting that both cones

and rods undergo apoptosis in *pday* mutants.

Although rods and cones survived in *pday* mutants that overexpressed Bcl2, zpr3

signals fail to be localized in the OS, but spread in the plasma membrane throughout the cell

body, including the basal synaptic region (Fig. 3F'), suggesting that green opsin and

rhodopsin are mislocalized in Bcl2-overexpressing *pday* mutant photoreceptors. Furthermore,

photoreceptor nuclear shape was different between Bcl2-overexpressing *pday* mutants and

wild-type siblings (Fig. S2G). In addition, Bcl2 overexpression did not restore OKR of *pday*

mutants (Fig. S2H). These data suggest that MAK activity is primarily required for visual

functions of photoreceptors, disruption of which may subsequently cause Bax-mediated

apoptosis.

mak is responsible for _pday_ mutant phenotypes

To validate whether the *mak* gene is responsible for *pday* mutant phenotypes, we generated

a zebrafish transgenic line *Tg[crx:eGFP-MAK]*, which expresses the N-terminal eGFP-

tagged, full-length isoform of MAK under control of the *crx* promotor. A transgenic line

Tg[crx:eGFP] was used as a control. Labeling with zpr1 antibody revealed that

Tg[crx:eGFP-MAK] rescued cone photoreceptor degeneration in *pday* mutants at 6 dpf,

whereas *Tg[crx:eGFP]* did not (Fig. 4A, 4A', and 4B). Furthermore, *Tg[crx:eGFP-MAK]*

restored the visual activity of *pday* mutants at 6 dpf, whereas *Tg[crx:eGFP]* did not (Fig.

4C). Thus, *mak* is responsible for *pday* mutant phenotypes: photoreceptor degeneration, and

visual response defects.

MAK is localized in the connecting cilium and promotes axoneme development during ciliogenesis

To examine the spatio-temporal profile of *mak* mRNA expression during embryonic development, we performed whole-mount *in situ* hybridization of zebrafish embryos from the 1-cell stage to 72 hpf (Fig. 5A). *mak* mRNA is ubiquitously expressed from 1- to 4-cell stages, indicating maternal mRNA expression. *mak* mRNA expression disappeared at the 50%-epiboly stage, but was detected in Kupffer's vesicle, a ciliated embryonic organ that is important for left-right asymmetry patterning (Essner et al., 2005), at the tail-bud stage. At 24 hpf, *mak* mRNA is ubiquitously expressed in the central nervous system, and especially strongly expressed in the epiphysis and retina. At 48 hpf, *mak* mRNA is prominently expressed in the brain, the retina, and the pectoral fin bud. At 72 hpf, *mak* mRNA is highly expressed in the brain, including the optic tectum and the photoreceptor layer of the retina. Thus, *mak* mRNA is expressed in various cell types in zebrafish embryos, including Kupffer's vesicle and retinal and pineal photoreceptors, in which ciliation plays a key role in supporting these cell functions (Essner et al., 2005; Liu et al., 2004; Sukumaran and Perkins, 2009; Zhai et al., 2014). In addition, zebrafish mutants with cilia defects tend to develop scoliosis (Gray et al., 2021; Latour et al., 2020; Wang et al., 2022), which was observed in *pday* mutants at 2 mpf (Fig. 1A). Thus, Mak regulates cilium-associated mechanisms in the photoreceptors in zebrafish.

Next, we examined MAK protein localization in zebrafish photoreceptors by tracking the eGFP signal in *Tg[crx:eGFP-MAK]* retinas. eGFP-MAK is localized in a stem-like structure inside the OS of live photoreceptors (Fig. 5B). Next, we examined double labeling of *Tg[crx:eGFP-MAK]* retinas with anti-GFP antibody and anti-acetylated α-tubulin antibody. The latter mainly labels the ciliary axoneme. eGFP-MAK was co-localized to the ciliary axoneme visualized by an anti-acetylated α-tubulin antibody (Fig. 5C, Fig. S3). Thus, MAK is a ciliary protein in photoreceptors.

Next, to examine the ciliary structure in *pday* mutant photoreceptors, we used anti-γ tubulin, anti-Eyes Shut homolog (EYS), and anti-acetylated α tubulin antibodies, which visualize the basal body, the transition zone, and the axoneme of the cilium, respectively

(Malicki and Johnson, 2017; Messchaert et al., 2018). Double labeling of 3-dpf, wild-type

and *pday* mutant retinas with anti-acetylated α tubulin and anti-EYS antibodies revealed that

EYS signals were normally detected, but acetylated α tubulin signals were absent in *pday*

mutant photoreceptors (Fig. 6A). Next, double labeling of 3-dpf, wild-type and *pday* mutant

retinas with anti-γ tubulin and anti-EYS antibodies revealed that both γ tubulin and EYS

signals were normally associated in *pday* mutant photoreceptors (Fig. 6B). These data

indicate that the ciliary axoneme is absent in 3-dpf *pday* mutant photoreceptors, whereas the

basal body and the transition zone seem to be normal, suggesting that the ciliary axoneme is

the primary target of MAK.

The OS is considered to be a specialized cilium of photoreceptors (Bachmann-Gagescu

and Neuhauss, 2019; Chen et al., 2021). Arl13b is a Joubert syndrome protein, which is

localized in the primary cilium and mutated in zebrafish *scorpion* (*sco*) mutants (Duldulao et

al., 2009). In addition, GFP-tagged Arl13b visualizes cilia in zebrafish (Bachmann-Gagescu

et al., 2011; Borovina et al., 2010). We established two zebrafish transgenic lines,

Tg[gnat2:arl13b-tdTomato; rho:arl13b-eGFP], which visualize the OSs of cones and rods,

respectively, in wild-type retinas (Fig. 6C). However, both Arl13b-tdTomato and Arl13b-

GFP expression were absent in 3-dpf *pday* mutant photoreceptors, suggesting that the OS

fails to be formed in both cones and rods of *pday* mutants. Thus, OS formation is

compromised in *pday* mutant photoreceptors, which may result from the absence of the

ciliary axoneme.

To confirm ciliary phenotypes in *pday* mutant photoreceptors, we applied electron

microscopy (EM) to examine the ultrastructure of wild-type and *pday* mutant photoreceptors

at 3 dpf (Fig. 6D, 6E). In wild-type photoreceptors at 3 dpf, photoreceptors show columnar

shapes and their OSs started to form (Fig. 6D). On the other hand, OSs were not observed in

pday mutant photoreceptors (Fig. 6E). Higher magnification EM images detected a basal

body, which is a pair of centrioles near the apical domain underneath the OS, as well as a

long extension from the basal body, which corresponds to the transition zone and the

axoneme, in wild-type photoreceptors (Fig. 6D'). However, a basal body and only a short

extension from the basal body, which is likely to correspond to only the transition zone, were

observed in *pday* mutant photoreceptors (Fig. 6E'). Taken together, these data suggest that
the OS and the ciliary axoneme are absent in *pday* mutant photoreceptors, whereas the basal
body and transition zone are normally formed. Thus, MAK regulates photoreceptor
ciliogenesis by promoting axonemal formation.

Phototransduction molecules fail to be transported to the OS in *pday* mutant
photoreceptors
Phototransduction molecules such as rhodopsin and opsins are transported to the OS through
the cilium (Bachmann-Gagescu and Neuhauss, 2019; Chen et al., 2021). Therefore, we
examined distributions of opsins and rhodopsin in *pday* mutant photoreceptors using zpr3
(Gao et al., 2022) and 1D4 (Yin et al., 2012) antibodies, which label green opsin/rhodopsin
and red opsin in zebrafish, respectively. In 3 dpf wild-type photoreceptors, green
opsin/rhodopsin and red opsin were normally localized in the OS (Fig. 6F, 6G). However, in
pday mutants, these opsins were mislocalized to the IS, plasma membrane, and the basal
synaptic region (Fig. 6F, 6G). It is likely that OS formation defects cause the failure in
transport of phototransduction molecules to the OS in zebrafish *mak* mutants.

Kinase activity of MAK is essential for photoreceptor survival and ciliary axoneme
formation
To understand how MAK regulates photoreceptor survival and ciliary axoneme formation,
we focused on kinase-associated motifs (Wang and Kung, 2012) of MAK and conducted site-
directed mutagenesis to silence kinase activity (Fig. 7A). Like human and mouse MAK, the
kinase domain is located at the N-terminal region of zebrafish MAK, which covers amino
acids 4 to 285 of the 633-aa full-length protein (Fig. S4). It was reported that KR (32-33 aa)
and TDY (157-159 aa) are required for kinase activity as an ATP-binding motif and an
autophosphorylation site, respectively (Wang and Kung, 2012). We designed two kinase-
dead mutants of zebrafish MAK, namely AR MAK and ADF MAK, by replacing KR and
TDY in wild-type zebrafish MAK with AR and ADF, respectively (Fig. 7A). We also
prepared DM MAK, which carries both AR and ADF mutant motifs. *In vitro* kinase assay

using MAK proteins purified from an *E. coli.* expression system revealed that all three
kinase-dead MAK mutants, AR MAK, ADF MAK, and DM MAK, failed to phosphorylate
a MAK mock substrate, myelin basic protein (MBP), indicating that MAK kinase activity is
silenced in either mutant, AR MAK, ADF MAK, or DM MAK (Fig. 7B).

Next, we generated zebrafish transgenic lines, *Tg[crx:eGFP-AR MAK]*,
Tg[crx:eGFP-ADF MAK], and *Tg[crx:eGFP-DM MAK]*, by introducing kinase-dead mutant
motifs into the wild-type MAK-expressing construct, *Tg[crx:eGFP-WT MAK]*. First, we
examined subcellular localization of these three kinase-dead mutant MAK proteins in
zebrafish photoreceptors by labeling these transgenic wild-type retinas with anti-GFP
antibody (Fig. 7C). eGFP-WT MAK was localized in a stem-like pattern similar to the
axoneme of photoreceptor cilium. However, eGFP-AR MAK was localized in a spot-like
region near the proximal region of photoreceptor cilium, which seems to correspond to either
the basal body or the transition zone. Thus, ATP-binding activity is important for proper
localization of MAK in the axoneme of photoreceptor cilium. eGFP-ADF MAK was likely
to be localized in the axoneme of photoreceptor cilium, but its expression level was low,
suggesting that GFP-ADF MAK may be unstable. GFP-DM MAK showed a low expression
level as well, which made it difficult to define its subcellular localization. These data suggest
that MAK kinase activity influences its localization in the axoneme of photoreceptor cilium.

Next, using zpr1 antibody, we examined whether MAK kinase activity is required for
survival of double-cone photoreceptors. We combined transgenic lines, *Tg[crx:eGFP-AR*
MAK], *Tg[crx:eGFP-ADF MAK]*, and *Tg[crx:eGFP-DM MAK]*, with the *pday* mutants. In
contrast to *Tg[crx:eGFP-WT MAK]*, all three kinase-dead mutant MAK transgenes fail to
rescue survival of double-cone photoreceptors in the dorso-central ONL at 6 dpf (Fig. 7D),
suggesting that MAK kinase activity is important for photoreceptor survival.

To determine whether MAK kinase activity is required for ciliary axoneme formation,
we examined ciliary axonemes in *pday* mutants by labeling *pday* mutants combined with
Tg[crx:eGFP-AR MAK], *Tg[crx:eGFP-ADF MAK]*, and *Tg[crx:eGFP-DM MAK]* with anti-
acetylated α tubulin antibody. In contrast to *Tg[crx:eGFP-WT MAK]*, all three kinase-dead
mutant MAK transgenes failed to rescue axoneme formation at 6 dpf (Fig. 7E). Thus, MAK

kinase activity is critical for ciliary axoneme development and photoreceptor survival (Fig.

8).

Discussion

In this study, we showed that zebrafish *mak* mutants show progressive degeneration of photoreceptors during embryonic development. In *mak* mutants, both rods and cones undergo apoptosis; however, in contrast to rods, cones do not display an acute apoptotic phenotype such as condensed nuclei, but rather a progressive shrinkage of cell volume. In *mak* mutants, photoreceptor cilia completely lack axonemes, whereas basal bodies and transition zones seem to be specified. Since a photoreceptor cilium is essential for IFT-mediated protein transport to the OS, it is likely that IFT-mediated ciliary transport is affected in *mak* mutants. Consistently, the OS is absent in *mak* mutants. OS resident proteins, rhodopsin and opsins, are abnormally distributed in photoreceptors. Finally, we confirmed that the kinase activity of MAK is essential for axoneme formation and photoreceptor maintenance. Thus, MAK is essential for photoreceptor ciliogenesis, especially axoneme formation, in zebrafish.

In zebrafish, photoreceptor precursors undergo one round of asymmetric cell division to produce two daughter cones after 60 hpf, in which apically localized centrosomes start ciliogenesis (Zolessi et al., 2021). The primary cilium is generated by either extracellular or intracellular pathways in vertebrate photoreceptors (Breslow and Holland, 2019). In either case, the first step of ciliogenesis is the removal of CP110 from a distal appendage of the mother centriole, which enables ARL13b and components of IFT-A and IFT-B to assemble at the apical surface of a distal appendage of the mother centriole in a rab8-dependent manner (Breslow and Holland, 2019; Ge et al., 2022). The second step is formation of the transition zone by the transport of MKS and NPHP module components (Park and Leroux, 2022). The third step is extension of the axoneme through IFT-mediated ciliary transport, which transports and adds α-/β-tubulin dimers to the plus end of the microtubule comprising the axoneme (Brouhard and Rice, 2018; Ge et al., 2022; Nakayama and Katoh, 2018).

In humans and other animal models, including zebrafish and mice, many genetic mutations showing defects in photoreceptor ciliogenesis have been identified (Chen et al., 2021). In zebrafish mutants of transition zone components, which are classified into three modules, MKS, NPHP, and core holding module, the gating mechanism of the transition zone, which selectively transports phototransduction molecules such as opsins into the OS,

is compromised, resulting in ectopic distribution of these OS-resident proteins outside the

OS (Wang et al., 2022). However, the axoneme itself is formed in almost all of these

transition-zone-defective mutants (Wang et al., 2022). In zebrafish mutants of BBS

components, which cooperate with IFT to regulate intra-ciliary transport (Tian et al., 2023),

OS non-resident proteins are mislocalized into the OS, but axoneme and the OS are formed

(Masek et al., 2022). Furthermore, in zebrafish *arl13b* mutants, with compromised cargo

release in the OS, the axoneme and the OS are reduced in size, but still formed (Song et al.,

2016). Thus, most ciliogenesis-defective mutants show formation of axoneme structure in

photoreceptors. However, there are a few exceptions. In zebrafish mutants of an IFT-B

component, IFT88 (also named *oval*), Traf3ip1 (also named *elipsa*), and a Kinesin II family

protein Kif3a, the axoneme and the OS are totally absent in photoreceptors (Krock and

Perkins, 2008; Omori et al., 2008; Sukumaran and Perkins, 2009; Zhu et al., 2021).

Interestingly, in IFT88 (*oval*) and Traf3ip1 (*elipsa*) mutants, a single spot-like γ-tubulin or

centrin signal is detected in the apical region of photoreceptors, similar to zebrafish *mak*

mutants, suggesting that basal bodies are formed in both mutants (Omori et al., 2008;

Sukumaran and Perkins, 2009; Tsujikawa and Malicki, 2004). Thus, it is likely that MAK

cooperates with anterograde IFT complexes including IFT88 to promote axoneme formation

during ciliogenesis.

In this study, we show that overexpression of wild-type MAK, using the

photoreceptor-specific *crx* promoter recovers formation of axonemes, leading to survival of

functional photoreceptors, in *mak* mutants. However, overexpression of kinase-mutant forms

of MAK, AR MAK and ADF MAK, which disrupt the ATP-binding motif of the kinase

domain and autophosphorylation sites, fail to rescue defects in axoneme and cell survival, so

phosphorylation events on MAK kinase substrates are necessary for axoneme formation.

Interestingly, GFP-tagged AR MAK is localized in a single spot area near the apical region

of photoreceptors, which seems to correspond to the basal body or transition zone, whereas

GFP-tagged, wild-type MAK is located along the proximo-distal axis of the axoneme. One

possibility is that a target protein or proteins of MAK-mediated phosphorylation exist near

the basal body or transition zone, and that these target proteins move with MAK along the

axoneme after their phosphorylation by MAK. Another possibility is that MAK interacts with

adaptor proteins that are localized in the basal body or transition zone to support MAK

substrate phosphorylation for ciliogenesis. At present, a molecular network of MAK

phosphorylation substrates and adaptor proteins that facilitate MAK substrate

phosphorylation is unknown in zebrafish; however, since the IFT complex is assembled and

transported through the transition zone in the first step of ciliogenesis (Breslow and Holland,

2019), IFT complex components may be promising candidates to promote MAK substrate

phosphorylation.

MAK genes share a highly conserved amino acid sequence between zebrafish and

humans (51.38 % amino acid identity), especially the N-terminal kinase domain (88.65 %

amino acid identity). However, in contrast to zebrafish *mak* mutants, the axoneme is not

shortened, but is elongated in mouse *Mak* mutants (Omori et al., 2010). It is an interesting

question why *mak* gene knockout induces such opposite phonotypes in zebrafish and mice.

From a phylogenetic point of view, MAK belongs to the MAK/ICK/MOK serine/threonine

kinase family, all members of which share a highly conserved kinase domain (Chen et al.,

2013; Miyata and Nishida, 1999). CILK1 (previously known as ICK) is a member of this

kinase family and is now classified as a paralog of MAK. Indeed, human CILK1 shows a

highly similar amino acid sequence in the kinase domain to that of zebrafish MAK (81.5%

amino acid identity). Interestingly, in *Cilk1* mouse mutants, the photoreceptor ciliary

axoneme is shortened compared with that of wild-type controls (Chaya et al., 2014).

Furthermore, mouse CILK1 interacts with IFT-B components, including IFT88 (Nakamura

et al., 2020). Since a CILK1 homologous gene has not been identified in the zebrafish

genomic database, zebrafish MAK and mouse CILK1 may share a common regulatory

pathway for axoneme formation during ciliogenesis. In the future, it will be important to

investigate whether zebrafish MAK cooperates with IFT-B to regulate axoneme formation

during ciliogenesis.

Another interesting finding is that there is a difference in degeneration of rods and

cones in *mak* mutants. In *mak* mutants, the ONL progressively degenerates from 3 dpf and

disappears by 6 dpf. Using zebrafish transgenic lines, which visualize rods and cones

separately, we found that rods frequently show pyknotic nuclei, which are commonly observed in apoptosis, at 3 dpf in *mak* mutants, and that the number of rods is markedly decreased at 4 dpf. Since TUNEL signals increase to a maximum at 4 dpf and then decrease in *mak* mutant ONL, it is likely that most TUNEL-positive photoreceptors at 3 and 4 dpf in *mak* mutants are rods. On the other hand, we did not observe typical, round, pyknotic nuclei in cones in *mak* mutants. Rather, mutant cones progressively shorten after 4 dpf, becoming extremely flattened at 5 dpf, and finally disappearing by 6 dpf. Although we observed TUNEL signals in flattened cone photoreceptors in the central retina at 5 dpf, the degenerative process of cones seems to be different from that of rods in *mak* mutants.

Recently, it was reported that photoreceptor degeneration in ciliogenesis-defective zebrafish *Tulp1* mutants is associated with pathological features of ferroptosis (Jia et al., 2022), which is a new form of cell death linked to iron-dependent accumulation of lipid peroxides (Yu et al., 2017), suggesting that ciliogenesis defects may trigger non-apoptotic cell death mechanisms. However, we found that overexpression of anti-apoptotic protein, Bcl2, effectively suppresses degeneration in both rods and cones, suggesting that in *mak* mutants, both depend on Bax-mediated apoptosis. Interestingly, early pyknotic nuclei in presumptive rods and a later progressive flatten-shaped shrinkage of the photoreceptor layer was also observed in zebrafish *oval* and *elipsa* mutants, which show severe defects in axoneme formation by genetic mutation of IFT88 and Traf3ip1, respectively (Bahadori et al., 2003; Doerre and Malicki, 2002; Sukumaran and Perkins, 2009; Wang et al., 2022). It is an interesting question why such a difference in degeneration occurs between rods and cones in zebrafish mutants showing defects in axoneme formation. Recent metabolic transcriptome analysis combined with scRNA-seq revealed that cellular metabolism of rods and cones depends on aerobic glycolysis and oxidative phosphorylation, respectively, but that cones shift toward glycolysis in the early pathological stage of retinitis pigmentosa, suggesting that there is an intrinsic difference of metabolic regulation between rods and cones (Lee et al., 2023). In general, ciliopathy is the common important topic beyond photoreceptor degeneration, but it is still unknown how ciliogenesis defects activate the cell death pathway.

Zebrafish *mak* mutants provide a good model to study ciliogenesis-linked cell death

mechanisms.

Materials and Methods

Ethics statement

Zebrafish were maintained at 28.5 ℃, on a 14h light/10h dark cycle, following the standard

procedure (Westerfield, 2000). Collected embryos were cultured in E3 embryo medium (5

mM NaCl, 0.17 mM KCl, 0.33 mM $CaCl_2$, 0.33 mM $MgSO_4$) containing 0.003% 1-phenyl-

2-thiouera (PTU) to prevent pigmentation and 0.01% methylene blue to prevent fungal

growth. All experiments were performed on zebrafish embryos between the one-cell stage

and 6 dpf prior to sexual differentiation. Therefore, sexes of embryos could not be

determined. All zebrafish experiments were performed under the OIST Animal Care and Use

Program based on the Guide for the Care and Use of Laboratory Animals by the National

Research Council of the Nation Academies and has been approved by the Association for

Assessment and Accreditation of Laboratory Animal Care (AAALAC). The OIST

Institutional Animal Care and Use Committee approved all experimental protocols (Protocol:

ACUP-2023-016, ACUP -2023-017, ACUP -2023-018, ACUP -2023-019, ACUP -2023-

020).

Zebrafish strains and maintenance

Okinawa wild-type (Oki) was used as a wild-type strain to maintain all mutant and transgenic

strains. The zebrafish mutant, *pday*[s351], was originally isolated by Herwig Baier lab (Muto et

al., 2005). Mapping of the *pday*[s351] mutation was carried out in the genetic background of

WIK. Zebrafish transgenic lines, *Tg[gnat2:NLS-tdTomato]*[oki070] and *Tg[rho: NLS-eGFP]*[oki071],

were used to visualize cone and rod nuclei, respectively. *Tg[hsp:mCherry-Bcl2]*[oki029]

(Nishiwaki and Masai, 2020) was used to express mCherry-tagged zebrafish Bcl2 under

control of the zebrafish *heat-shock 70* (*hsp70*) promoter. *Tg[crx:eGFP]*[oki072], *Tg[crx:eGFP-

WT MAK]*[oki073], *Tg[crx:eGFP-AR MAK]*[oki074] and *Tg[crx:eGFP-ADF MAK]*[oki075], and

Tg[crx:eGFP-DM MAK][oki076] were used to express eGFP-tagged wild-type and kinase-

deficient mutants MAK, respectively. *Tg[gnat2:arl13b-tdTomato]*[oki077] and *Tg[rho:arl13b-

eGFP]*[oki078] were used to visualize cone and rod OSs, respectively.

Mapping and cloning of the *pday* mutant gene

pday^{+/-} fish maintained in *oki* wild-type background were outcrossed with *WIK* wild-type strain to generate F1 *pday*^{+/-} fish with an *oki* and *WIK* trans-heterozygous wild-type background. These F1 *pday*^{+/-} male and female fish were crossed to produce F2 embryos. F2 *pday*^{-/-} embryos were selected by measuring OKR at 5 dpf and stored in methanol at -80 °C. Genomic DNA was extracted from individual F2 *pday*^{-/-} embryos. First, two DNA pools of 20 wild-type and 20 homozygous mutant embryos, respectively, were applied to PCR-mediated amplification of SSLP markers (Knapik et al., 1998; Shimoda et al., 1999) to examine which chromosome hosts the *pday* mutation. Furthermore, 93 F2 *pday*^{-/-} embryos (186 meiosis) were used to restrict the cytogenetic position of the *pday* mutation using SSLP markers mapped on the linked chromosome. After we restricted the *pday* mutation within the genomic region flanked between two SSLP markers, z23011 and z13695, which were annotated at positions 8.517 and 9.123 Mb on chromosome 24 (zebrafish genomic database, GRCz10, Ensemble release 80), we designed a new SSLP marker, namely Mak-N1, which was located around 20 bp upstream of the first exon of the *mak* gene, and confirmed no recombination of Mak-N1 in all 93 F2 *pday*^{-/-} embryos (186 meiosis). Sequence information on forward and reverse primers of z23011, z13695, and Mak-N1 are provided in the appendix.

To confirm that *pday* gene encodes MAK, full-length *mak* cDNAs were amplified by PCR using mRNA prepared from wild-type and *pday*[s351] homozygous embryos at 6 dpf. However, only a partial cDNA fragment that corresponds to the C-terminal coding region covering from the middle of exons 7 to 15 was amplified by PCR from *pday* homozygous mutant mRNA. We sequenced the full-length cDNA of wild type and the C-terminal partial cDNA of *pday* homozygous mutants and found no amino acid change in the MAK coding region from exons 8 to 15 between wild-type and *pday*^{-/-} cDNA. Next, we amplified six DNA fragments from *the pday*^{-/-} genome, each of which covers six N-terminal coding exons (the exon 2 - 7), respectively. Sequencing of these six DNA fragments revealed a non-sense mutation in exon 4 of the *pday*^{-/-} genome. To confirm that the *pday* mutant gene encodes MAK, we combined *pday* mutants with the transgenic line *Tg[crx:eGFP-WT MAK]*[oki073], and

confirmed that expression of eGFP-tagged MAK in photoreceptor precursors rescues
photoreceptor degeneration in *pday* mutants.

Generation of DNA expression constructs and their transgenic lines
Tol2[crx: membrane targeted YFP (MYFP)] was used as a template to make the expression
construct *Tol2[crx: eGFP-MAK]. Tol2[crx: MYFP]* was constructed using the To2kit
multiple gateway-based construction system (Kwan et al., 2007), whose Tol2 vector
backbone contains *cmcl2:CFP* transgenesis marker (Suzuki et al., 2013). *mak* cDNA
corresponding to a full-length isoform annotated on the zebrafish genomic database (mak-
201, GRCz11, Ensemble release 110), was amplified by PCR using a mRNA pool of 3-dpf,
wild-type embryos and was subcloned into pCR TOPO vector (Invitrogen). After the eGFP
cDNA fragment was tagged in-frame to the *mak* full-length cDNA at the N-terminus, the
MYFP region of *Tol2[crx: MYFP]* was replaced with the DNA fragment encoding eGFP-
tagged *mak* cDNA at the *BamHI* and *ClaI* sites, to make the expression construct
pTol2[crx:eGFP-WT MAK]. The *pTol2[crx:eGFP-WT MAK]* was used as a template in the
site-directed mutagenesis for making three kinase-dead MAK constructs, *Tg[crx:eGFP-AR*
MAK], *Tg[crx:eGFP-ADF MAK]* and *Tg[crx:eGFP-DM MAK]*.

The *EF1α* promoter of Tol2 transposon vector pT2AL200R150G (Urasaki et al.,
2006) was replaced with the zebrafish *gnat2* promoter (Iribarne et al., 2019) at the *XhoI* and
BamHI site to make *Tol2[gnat2:eGFP]*. Zebrafish *arl13b* cDNA was amplified by PCR
using an mRNA pool prepared from 4-dpf, wild-type embryos and subcloned into the pCR
TOPO vector (Invitrogen). After a tdTomato cDNA fragment was tagged in-frame to the
arl13b cDNA at the C-terminus, a DNA fragment of tdTomato tagged *arl13b* cDNA was
further subcloned into the modified pT2AL200R150G carrying the *gnat2* promoter at the
BamHI and *ClaI* sites, to make the expression construct *pTol2[gnat2:arl13b-tdTomato]*.
Similarly, the *EF1α* promoter of Tol2 transposon vector pT2AL200R150G was replaced
with the zebrafish *rhodopsin* promoter (1.1kb upstream of the transcription unit) at the *XhoI*
and *BamHI* site. A DNA fragment of C-terminal eGFP-tagged *arl13b* cDNA was prepared

and subcloned into the modified pT2AL200R150G carrying *rhodopsin* promoter at the
BamHI and *ClaI* sites, to make the expression construct *pTol2[rho:arl13b-eGFP]*.

The nuclear localization signal (NLS) was fused to cDNA encoding tdTomato or
eGFP at the N-terminus. DNA fragments of NLS-tdTomato and NLS-eGFP were subcloned
into the modified pT2AL200R150G carrying the zebrafish *gnat2* promoter and the zebrafish
rhodopsin promoter, respectively, at the *BamHI* and *ClaI* sites, to make the expression
constructs *pTol2[gnat2:NLS-tdTomato]* and *pTol2[rho:NLS-eGFP]*.

Next, these Tol2 expression construct plasmids and Tol2 transposase mRNA were
co-microinjected into one-cell-stage zebrafish fertilized eggs for generation of transgenic
lines, *Tg[crx:eGFP-WT MAK], Tg[crx:eGFP-AR MAK], Tg[crx:eGFP-ADF MAK],*
Tg[crx:eGFP-DM MAK], Tg[gnat2:arl13b-tdTomato], Tg[rho:arl13b-eGFP],
Tg[gnat2:NLS-tdTomato] and *Tg[rho:NLS-eGFP]*. Injected F0 fish expressing fluorescent
protein (eGFP or tdTomato) in the ONL were bred until adulthood and used to identify
founder fish, which produce fluorescent protein-expressing F1 progeny. These transgenic F1
embryos were raised and used to produce F2 generation for establishment of the transgenic
strain.

Histology

For plastic section, zebrafish embryos were anesthetized in 0.02% 3-aminobenzoic acid ethyl
ester (tricaine) and fixed in PBS (150mM NaCl, 10mM $PO4^{3-}$, pH 7.4) (Invitrogen) with 4%
paraformaldehyde (PFA) at 4 °C overnight. Samples were dehydrated with an ethanol
gradient before being embedded in JB4 resin (Polysciences). Samples were sectioned at 5
μm with a microtome Rotatif, NM335E (MICROM International GmbH) and stained with
0.02% toluidine blue for further analysis.

For TUNEL, samples were covered with the staining mixture at 37 °C for 1 h
following instructions of the In Situ Cell Death Detection Kit (Roche, 11684795910). After
the TUNEL reaction, nuclei were stained with 60 mM PO_4 buffer (PB) (42.6 mM NaH_2PO_4,
17.4 mM Na_2HPO_4, pH 7.3) containing Hoechst 33342 (Fujifilm, 346-07951) at 1:1000,

followed by washing with PB with 0.1% TritonX-100 (0.1% PBTx) for 3 x 5 min. Samples

were mounted with Fluoromount (Diagnostic BioSystems, K024) for further analysis.

For immunohistochemistry, 3-6-dpf embryos were fixed with 4 % PFA at 4 °C

overnight, and transferred to 30% sucrose in PB overnight before being embedded and frozen

in OCT compound. Cryosections was prepared at 7 μm with a cryostat (Cryostar NX70,

Thermo Scientific.), except at 20 μm for imaging cilia components. Cryosections were air-

dried for at least 2 h, then rehydrated in 0.1% PBTx for 3 x 5 min. Rehydrated sections were

immersed in blocking solution (10% goat serum in 0.1% PBTx) for 1 h at room temperature,

then applied to the primary antibody in the blocking solution at an appropriate dilution (see

appendix), and washed with 0.1% PBTx 3 x 10 min. Next, primary antibody-labeled sections

were applied to the fluorophore-conjugated secondary antibody at 1:500 and Hoechst 33342

(DOJINDO Laboratories, CAS23491-52-3) or 50 nM Sytox-Green (Molecular Probes) in

blocking solution for 1 h at room temperature, and then washed with 0.1% PBTx 3 x 10 min.

Sections were finally mounted with Fluoromount. Confocal images were scanned with

confocal laser-scanning microscopes, LSM780 (Carl Zeiss) or Fluoview FV3000 (Olympus,

Evident).

For EM analysis, 3-dpf embryos were fixed in PB with fixative (2.5% glutaraldehyde,

1% paraformaldehyde, 3% sucrose, and 30 mM HEPES) on ice for 2 h, followed by washing

with PB with 3% sucrose and 30 mM HEPES 3 x 5 min. Embryos were postfixed in 60 mM

PB with 1% OsO_4 for 1 h on ice, followed by washing with Milli-Q water three times.

Samples were dehydrated with a gradient series of ethanol and replaced by acetone. After

treatment with a gradient series of propylene oxide (PO), epoxy resin EPON812 (Nisshin-

EM, 3402) was applied to the samples before they were embedded in resin and solidified for

2 days at 60 °C. Ultra-thin sections at 50 nm were prepared with a Leica UC7 ultra-microtome

and processed with Reynolds' stain (Reynolds, 1963). Images were observed and captured

using a JOEL JEM-1230R transmission EM.

Bcl2 overexpression in *pday* mutant retinas

Transgenic line *Tg[hsp:mCherry-Bcl2]* (Nishiwaki and Masai, 2020) was combined with

pday mutants. Heat shock treatment was carried out by incubation of embryos at 39 °C for 1

h every 12 h after 1 dpf until 6 dpf.

***In situ* hybridization**

Whole-mount *in situ* hybridization was carried out using a published protocol (Thisse and

Thisse, 2008). A DNA fragment containing *mak* full-length cDNA combined with T7 RNA

polymerase promoter, which is located at the 3' end of the *mak* full-length cDNA, was

amplified by PCR. Digoxigenin (DIG)-labeled anti-sense *mak* RNA probe was generated by

in vitro transcription from this T7 promoter: *mak* cDNA fragment as the template using a

DIG RNA labeling kit (Roche, 11175025910). Zebrafish embryos from the 1-cell stage to 72

hpf were fixed with 4% PFA at 4 °C overnight, and were stored in 100 % methanol at -70 °C.

Samples were rehydrated from methanol to PBS, and then treated with protenase K (10

μg/mL) to increase permeability. After refixing with 4% PFA for 20 min, samples were

soaked with prehybridization buffer at 65 °C for 2 h, then incubated with hybridization buffer

containing *mak* antisense RNA probe at 65 °C overnight. Samples were washed with 2xSSC

with 0.1 % Tween20 for 5 min, room temperature, and then 0.2xSSC with 0.1 % Tween20 3

x 30 min at 65 °C. Washed samples were transferred to maleic acid buffer with 0.1%

Tween20 (MABT), and then to MABT with 2% blocking reagent (Roche, 11096176001) at

4°C overnight. Samples were incubated with anti-DIG-AP antibody (Roche, 11093274910)

at 1:6,000 in MABT for 1 h at room temperature, washed with MABT 3 x 15 min, and then

transferred to Tris-HCl buffer (pH 9.5) for 15 min. For alkaline phosphatase (AP) reaction,

samples were transferred to BM-Purple (Roche, 11442074001) until a blue signal developed,

and fixed with 4% PFA overnight at 4 °C. These samples were mounted in 70% glycerol-

PBS and stored at 4 °C for long-term preservation. Images of the embryos were observed

with a SteREO Discovery V12 dissection microscope (Carl Zeiss).

Live imaging

For live imaging using confocal LSM, embryos were anesthetized in 0.02% tricaine and

mounted on glass slides in appropriate positions and glued with 3% methylcellulose. An

upright confocal LSM, FV3000 (Olympus, Evident) was used for scanning fluorescent

signals in the photoreceptor OS of *Tg[crx:GFP-MAK]*.

***In vitro* kinase assay**

The wild-type zebrafish *mak* cDNA fragment was introduced into pGEX6p3 (GE Health

Care Life Science). Kinase-dead *mak* cDNA fragments were generated from pGEX6p3-

zMAK(wt) with primers for site-directed mutagenesis. Plasmids were transformed into *E.*

coli strain BL21-competent cells (TaKaRa Bio, TKR9126). Colonies were inoculated into

LB broth for starter cultures incubated at 37 °C overnight, followed by amplification cultures

containing 1% of the starters. Amplification cultures were cooled on ice once the O.D.

reached 0.6 and IPTG was added (TaKaRa Bio, TKR9030) to a final concentration of 1 mM,

and incubated at 20 °C overnight. Cultures were pelleted, washed with cold PBS, and

sonicated in 1% PBS TritonX-100 (PBSTx) with 1x protease inhibitor (Roche,

04693132001). Debris was then pelleted, and supernatant fractions were subjected to

Glutathione Sepharose 4B (Cyvita, 17-0756-01) for 1.5 h at 4 °C. After being washed with

cold PBS 3x, Sepharose beads were incubated with PreScission protease (Cyvita, 27084301)

in the solution at 4 °C to release MAK from the GST tag. Digested products were pelleted,

and supernatant fractions were used for *in vitro* kinase assays.

Each reaction for *in vitro* kinase assays is composed of 50 mM Tris-HCl (pH 7.0), 10

mM $MgCl_2$, 5 mM ATP (Thermo Scientific, R0441), 1 mM DTT (Thermo Scientific, 20290),

5 μg dephosphorylated myelin basic protein (Merck, 13-110), and purified MAK protein.

Reactions were incubated 1 h at 30 °C. Samples were mixed with SDS sample dye and

incubated for 20 min at 75 °C to stop the reactions. Samples were separated by SDS-PAGE

and transferred onto a PVDF membrane for later analyses. The membrane was blocked with

5% skim milk for 1 h at room temperature followed by probing with primary antibodies and

HRP-conjugated secondary antibodies. ECL substrate (Thermo Scientific, 34577) was

applied to visualize signals using a iBright FL1500 imager (Invitrogen).

Data quantification and statistical analysis

To qualify the thickness of photoreceptor layers, photoreceptor layers in the images were straightened with the straighten tool of ImageJ (Fig. S5A). A line dividing the photoreceptor layer into 10 parts was generated with Adobe Illustrator. The photoreceptor layer thickness at each decile was measured with ImageJ by following the conversion ratio of pixels to the actual length of the scale bar. To qualify photoreceptor numbers, photoreceptor layers in the images were straightened with ImageJ, and a reference line marking every 1/10 points was drawn with Adobe Illustrator. Cell numbers were manually counted. TUNEL signals in retinas were manually counted. The zpr1-positive area in the total retinal was calculated as previously described (Fig. S5B) (Nishiwaki et al., 2013). All statistical analyses were performed with GraphPad Prism 9.2.1. Significance levels, numbers of samples, and the details of analyses are indicated in each figure and figure legend.

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Data availability

N/A

Author contributions statement

Conceptualization: H-JC, IM; Methodology: H-JC, YN, IM; Software: H-JC, IM; Validation:

H-JC, YN, W-CC, IM; Formal analysis: H-JC, YN, IM; Investigation: H-JC, YN, W-CC,

IM; Resources: H-JC, YN, W-CC, IM; Data curation: H-JC, YN, W-CC, IM; Writing -

original draft: H-JC, IM; Writing - review & editing: H-JC, IM; Visualization: H-JC, YN,

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IM.

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Declaration of interests

The authors declare no competing interests.

Figure Legends

Fig. 1: Zebrafish *pday* mutants carry a non-sense mutation in the *mak* gene and show

photoreceptor degeneration.

(A) *pday* mutants at 6 dpf and 2 mpf. At 6 dpf, *pday* mutants show no morphological

differences from wild-type siblings. At 2 mpf, *pday* mutants show smaller body size with

less well-developed fins, scoliosis, and cardiac edema (open arrowhead). Scale bars: 200

μm.

(B) The zebrafish *mak* gene consists of 15 exons and encodes 633 amino acids with a

serine/threonine kinase domain. A non-sense mutation occurs in exon 4 of *mak* gene in

pday mutants, leading to a premature termination codon at 70R, resulting in a truncated

protein lacking most of serine/threonine kinase domain.

(C) Wild-type and *pday* mutant retinas at 3, 4, 5 and 6 dpf. The ONL in *pday* mutants shows

dense nuclear granules (yellow arrowheads), bubble-like structures (red arrowheads) and

reduced outer segments (yellow lines). In *pday* mutants, the dorso-central ONL is

progressively decreased in thickness after 4 dpf, becomes extremely flat at 5 dpf and

disappears at 6 dpf (red dotted lines). On the other hand, other retinal layers, INL, IPL,

RGCL are maintained in *pday* mutants.

(C') Higher-magnification images of the central ONL. The OS is drastically reduced in *pday*

mutants (red lines). Dense nuclear granules (yellow arrowheads) and bubble-like

structures (red arrowheads) are observed in *pday* mutants. The ONL thickness is

progressively reduced in *pday* mutants (yellow lines). HC, horizontal cells; PR,

photoreceptors.

(D) Wild-type and *pday* mutant retinas at 6 dpf carrying the transgenic line *Tg[crx:eGFP]*.

Nuclei were counterstained with Hoechst 33342 (blue). crx:eGFP (green) expression is

more severely decreased in the dorsal region (red dotted line) than in the ventral region

of *pday* mutant retinas.

(E) Histogram of thickness of eGFP-positive photoreceptors along the DV axis of the retina.

The thickness of photoreceptors is more severely decreased in the dorsal region than in

the ventral region of *pday* mutant retinas. Color bars and lines indicate mean ± SD.

Statistical difference is evaluated with 2-way ANOVA and Sidak's multiple comparison

tests, n=3 for each point. ** p<0.01, *** p<0.001, **** p<0.0001.

Scale bars: 200 μm (A), 50 μm (C), 10 μm (C'), 40 μm (D).

Fig. 2: Rods undergo apoptotic-like degeneration in *pay* mutants, whereas cones

progressively shrink.

(A-B) Wild-type and *pday* mutant retinas carrying the transgenic lines *Tg[rho:NLS-eGFP]*

and *Tg[gnat2:NLS-tdTomato]*, which visualize rods (green) and cones (red),

respectively. Nuclei were counterstained with Hoechst 33342 (blue). (A) Only green and

blue channel. (B) Only red and blue channel. Squares indicate the dorsal and ventral

ONL, shown in (C–F).

(C) The dorsal ONL of wild-type and *pday* mutant retinas with *Tg[rho:NLS-eGFP]*, which

visualizes rods (green). In *pday* mutants, rods have condensed, round nuclei (white

arrowheads) at 3 dpf, which resembles apoptosis. In *pday* mutants, rods are drastically

reduced in number at 4 dpf and disappeared at 5 and 6 dpf (asterisks), except in the

peripheral region near the dorsal CMZ.

(D) The ventral ONL of wild-type and *pday* mutant retinas with *Tg[rho:NLS-eGFP]*, which

visualizes rods (green). In *pday* mutants, rods have condensed, round nuclei (white

arrowheads) at 3 and 4 dpf. However, in *pday* mutants, rods disappeared at 5 and 6 dpf

(asterisks), except in the peripheral region near the ventral CMZ.

(E) The dorsal ONL of wild-type and *pday* mutant retinas with *Tg[gnat2: NLS-tdTomato]*,

which visualizes cones (red). In wild type, cones form monolayer at 3 dpf and two

subnuclear layers after 4 dpf. In *pday* mutants, cones form monolayer at 3 dpf; however,

tdTomato-negative nuclei are observed, whose position is identical to that of condensed

rod nuclei (white arrowheads). From 4 to 5 dpf, cones shrink progressively and become

flattened (yellow line). At 6 dpf, cones are extremely flattened and disappear, leaving

gaps (yellow arrowheads), except in the peripheral region near the dorsal CMZ.

(F) The ventral ONL of wild-type and *pday* mutant retinas with *Tg[gnat2: NLS-tdTomato]*,

which visualizes cones (red). In wild type, cones differentiate at 3 dpf but progressively

842 disappear toward the peripheral area of the ventral ONL, where rods densely differentiate.

843 In *pday* mutants, the ONL thickness is progressively decreased after 4 dpf. Interestingly,

844 after 5 dpf, the cone differentiating area is expanded toward the periphery of the ventral

845 ONL (asterisks), where rods disappear.

846 (G) Schematic drawing of cone and rod degeneration in *pday* mutants. Rods undergo acute

847 degeneration with apoptotic-like pyknotic nuclei during 3 - 4 dpf, whereas cones undergo

848 progressive shrinkage of cell volume during 4 - 6 dpf and disappear by 6 dpf.

850 **Fig. 3: Rods and cones undergo apoptosis in *pday* mutants.**

851 (A) TUNEL of wild-type and *pday* mutant retinas at 3, 4, and 5 dpf. TUNEL signals are

852 indicated in red. Nuclei are counterstained with Sytox-Green (green).

853 (A') Higher-magnification images of dorsal, central, and ventral ONL of *pday* mutant retinas,

854 which are indicated by white squares in (A). White arrowheads indicate TUNEL signals

855 (red). TUNEL signals are prominent in the central and ventral ONL at 3 dpf, and in the

856 dorsal and central ONL at 4 dpf. TUNEL signals are detected in a very thin central ONL

857 of *pday* mutants at 5 dpf, where only cones are maintained.

858 (B) Double labeling of 5-dpf wild-type and *pday* mutant retinas with zpr1 (red) and anti-

859 rhodopsin (green) antibodies.

860 (B') Higher-magnification images of dorsal, central, and ventral ONL of *pday* mutant retinas,

861 which are indicated by white squares in (B). Rhodopsin signals are observed only in the

862 peripheral region of the dorsal and ventral ONL (yellow arrowheads). zpr1 signals are

863 also detected in the peripheral ONL; however, zpr1-positive flattened cells are located

864 in the central ONL (white arrow).

865 (C) TUNEL (green) of 5-dpf wild-type and *pday* mutant retinas combined with zpr1 antibody

866 labeling (red).

867 (C') Higher-magnification images of dorsal, central, and ventral ONL of *pday* mutant retinas,

868 indicated by white squares in (C). TUNEL signals are detected in dorsal, central, and

869 ventral ONL, many of which are associated with zpr1 signals (white arrows). A few

870 TUNEL signals are not associated with zpr1 (white arrowheads).

(D) TUNEL (red) of 5-dpf wild-type and *pday* mutant retinas combined with anti-rhodopsin

antibody labeling (green).

(D') Higher-magnification images of dorsal, central, and ventral ONL of *pday* mutant retinas,

indicated by white squares in (D). Rhodopsin signals are detected in the peripheral region

of dorsal and ventral ONL (yellow arrowheads). TUNEL signals are detected in the

dorsal, central, and ventral ONL. TUNEL signals associated with rhodopsin signals are

observed only the peripheral region of the dorsal and ventral ONL (white arrowheads).

TUNEL signals are not associated with rhodopsin signals in central ONL and non-

peripheral region of dorsal and ventral ONL (white arrows).

(E, F) Wild-type and *pday* mutant retinas with and without the transgenic line *Tg[hsp:*

mCherry-Bcl2], labelled with zpr1 (E) and zpr3 (F) antibodies. Nuclei were

counterstained with Hoechst 33342 (blue). zpr1 and zpr3 signals are markedly reduced

in non-transgenic *pday* mutant retinas. However, after heat-shock treatment, zpr1 and

zpr3 signals in the ONL are recovered in *pday* mutant retinas with *Tg[hsp: mCherry-*

Bcl2] (white arrowheads).

(F') Higher-magnification images of the dorsal ONL indicated by white squares, shown in

(F). zpr3 signals are localized in the OS of wild type, but ectopically distributed outside

the OS in *pday* mutants.

Scale bars: 50 μm (A-D), 40 μm (E, F)

Fig. 4: Overexpression of MAK restores survival and visual function of *pday* mutant

photoreceptors.

(A) Labeling of wild-type and *pday* mutant retinas with zpr1 (red) and anti-eGFP (green)

antibodies carrying transgenic line *Tg[crx:eGFP]* or *Tg[crx:eGFP-MAK]*. Nuclei were

counterstained with Hoechst 33342 (blue). (A') indicates only red channel. zpr1 signals

are markedly decreased in the dorso-central region of *pday* mutant retinas carrying

transgene *Tg[crx:eGFP]* (white dotted lines), but maintained in *pday* mutant retinas

carrying transgene *Tg[crx:eGFP-MAK]*.

(B) The fraction of zpr1-positive area in the total retinal area in wild-type and *pday* mutants

with *Tg[crx:eGFP]* or *Tg[crx:eGFP-MAK]*. The fraction is markedly reduced in *pday*

mutants with *Tg[crx:eGFP]* (red bar), compared with wild-type siblings with

Tg[crx:eGFP] (blue bar). However, the fraction is recovered in *pday* mutants with

Tg[crx:eGFP-MAK] (purple bar), equivalent to wild-type with *Tg[crx:eGFP-MAK]*

(green bar). Bars and lines indicate mean ± SD. Statistical significance was evaluated

with ordinary one-way ANOVA and Turkey's multiple comparison tests; $*p<0.0332$,

$****p<0.0002$, ns: not significant.

(C) The fraction of OKR^+ and OKR^- embryos relative to total progeny produced by crosses

of $pday^{+/-}$ parent fish (blue), $pday^{+/-}$ parent fish carrying *Tg[crx:eGFP]* (red), and $pday^{+/-}$

parent fish carrying *Tg[crx:eGFP-MAK]* (green). 100% of embryos produced by crosses

of $pday^{+/-}$ parent fish carrying *Tg[crx:eGFP-MAK]* (green) are positive for OKR. Bars

and lines indicate mean ± SD. Statistical significance was evaluated with 2-way ANOVA

and Turkey's multiple comparison tests. $*** p<0.0002$, ns: not significant.

Scale bars: 20 μm (A, A')

Fig. 5: MAK is localized in axonemes of photoreceptor cilia.

(A) Whole-mount *in situ* hybridization of zebrafish wild-type embryos with *mak* RNA probe.

mak mRNA is ubiquitously expressed in one-cell- to four-cell stage, but disappears at the

50% epiboly stage. *mak* mRNA is expressed in a spot-like region at the most posterior

side of tail-bud-stage embryos, which corresponds to Kupffer's vesicle (red open

arrowhead). At 24 pf, *mak* mRNA is weakly expressed in the whole body, and strongly

expressed in retina and epiphysis (pineal eye) (red open arrowhead). At 48 hpf, *mak*

mRNA is expressed in the brain, including retina and pectoral fin. At 72 hpf, *mak* mRNA

is expressed in the optic tectum and retina. Retinal expression is restricted to the

photoreceptor layer (yellow arrowheads). kv, Kuffper's vesicle; re, retina; epi, epiphysis;

br, brain; fin, pectoral fin; Tc, optic tectum; PR, photoreceptor layer.

(B) Confocal live image of wild-type *Tg[crx: eGFP-MAK]* transgenic retina at 5 dpf. eGFP-

MAK is localized in the OS of photoreceptors (yellow arrowheads).

(C) Labeling of 5-dpf, wild-type *Tg[crx: eGFPMAK]* transgenic retina with anti-acetylated
α-tubulin (red) and anti-eGFP (green) antibodies. Nuclei were counterstained with
Hoechst 33342 (blue). The right panel indicates higher magnification of the square shown
in the left panel. eGFP-MAK is localized in the axoneme (white open arrowheads, right
panel).
Scale bars: 200 μm (A), 5 μm (B, C left), and 1 μm (C, right).
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Fig. 6: The axoneme is absent, but the basal body and transition zone are formed in
***pday* mutant photoreceptors.**
(A) 3-dpf wild-type and *pday* mutant retinas with anti-acetylated α-tubulin (red) and anti-
Eys (green) antibodies. Nuclei were counterstained with Hoechst 33342 (blue). Bottom
panels are higher magnification of the ONL shown in top panels. In *pday* mutants,
acetylated α-tubulin signal is absent.
(B) 3-dpf wild-type and *pday* mutant retinas with anti-γ-tubulin (red) and anti-Eys (green)
antibodies. Nuclei were counterstained with Hoechst 33342 (blue). Bottom panels are
higher magnification of the ONL shown in top panels. In *pday* mutants, both γ-tubulin
and Eys signals are detected.
(C) 3-dpf wild-type and *pday* mutant retinas combined with the transgenic lines, *Tg[gnat2:*
arl13b-tdTomato] and *Tg[rho: ail13b-eGFP]*, which visualize cone OS (red) and rod
OS (green), respectively. Nuclei were counterstained with Hoechst 33342 (blue). Bottom
panels are higher magnification of the ONL. In *pday* mutants, arl13b signal is absent in
both cones and rods.
(D-E) EM analysis of wild-type (D) and *pday* mutant (E) photoreceptors at 3 dpf. Bottom
panels indicate higher-magnification (3000x) images of apical region of photoreceptors
in wild-type siblings (D') and *pday* mutants (E'). Bottom right panels are enlarged
images of the square region shown in the bottom left panels. In wild-type photoreceptors,
the OS starts to form (red asterisks). Basal body (red arrowheads) and ciliary axoneme
(yellow arrowheads) are observed at the basal area of the OS. In *pday* mutants, the OS

is absent; however, basal body (red arrowheads) and short ciliary extension from the

basal body (orange arrowheads) are observed.

(F-G) Labeling of 3-dpf, wild-type and *pday* mutant retinas with zpr3 antibody (F) and 1D4

antibody (G), which label rhodopsin/green opsin and red opsin, respectively. Nuclei

were counterstained with Hoechst 33342 (blue). Bottom panels are higher magnification

of the ONL. Both zpr3 and 1D4 signals are localized in the OS of wild-type

photoreceptors, but mislocalized in the apico-most region corresponding to the IS as well

as plasma membrane including synaptic area (arrowheads) in *pday* mutant

photoreceptors.

Scale bars: 5 μm (top panels of A, B, C; bottom panel of F, G), 1 μm (bottom panels of A,

B, C; D), and 20 μm (top panels of F, G)

Fig. 7: MAK kinase activity is required for MAK protein localization in the ciliary

axoneme, axoneme formation, and photoreceptor maintenance.

(A) A scheme of generation of three types of MAK kinase-dead mutants: AR MAK, ADF

MAK and DM MAK. Zebrafish MAK has a catalytic domain in the N-terminal region,

which contains an ATP binding motif, KR, and autophosphorylation site, TDY. AR MAK

and ADF MAK were generated by conversion of KR and TDY into AR and ADF,

respectively. DM MAK has both AR and ADF mutations.

(B) *In vitro* kinase assay of MAK using a substrate, MBP. Only wild-type MAK

phosphorylates MBP.

(C) Labeling of 6-dpf, wild-type ONL carrying the transgenic line, *Tg[crx:eGFP],*

Tg[crx:eGFP-WT MAK], Tg[crx:eGFP-AR MAK], Tg[crx:eGFP-ADF MAK], or

Tg[crx:eGFP-DM MAK], with anti-GFP antibody (green). Nuclei were counterstained

with Hoechst 33342 (blue). eGFP is ubiquitously localized in cytoplasmic area of

Tg[crx:eGFP] photoreceptors, but in the ciliary axoneme of *Tg[crx:eGFP-WT MAK]*

photoreceptors. However, eGFP is localized in a spot region of *Tg[crx:eGFP-AR MAK]*

photoreceptors, which corresponds to the basal area of the OS. eGFP expression is

observed in the ciliary axoneme, but reduced in *Tg[crx:eGFP-ADF MAK]* photoreceptors.

eGFP expression is markedly reduced in *Tg[crx:eGFP-DM MAK]* photoreceptors.

(D) Labeling of 6-dpf, wild-type and *pday* mutant retinas carrying the transgenic line,

Tg[crx:eGFP], Tg[crx:eGFP-WT MAK], Tg[crx:eGFP-AR MAK], Tg[crx:eGFP-ADF

MAK], or *Tg[crx:eGFP-DM MAK]*, with zpr1 (red) and anti-GFP (green) antibodies.

Nuclei were counterstained with Hoechst 33342 (blue). zpr1-positive photoreceptors are

degenerated in dorso-central retinas of *pday* mutants carrying *Tg[crx:eGFP]* (white

dotted line). However, photoreceptor degeneration is rescued only in *pday* mutants

carrying *Tg[crx:eGFP-WT MAK]*, but not in *pday* mutants carrying kinase-dead mutant

transgenes (white dotted line).

(E) Labeling of 3 dpf wild-type and *pday* mutant ONL carrying the transgenic line,

Tg[crx:eGFP], Tg[crx:eGFP-WT MAK], Tg[crx:eGFP-AR MAK], Tg[crx:eGFP-ADF

MAK], or *Tg[crx:eGFP-DM MAK]*, with anti-acetylated α-tubulin antibody (red). Nuclei

were counterstained with Hoechst 33342 (blue). Axoneme formation defects are rescued

only in *pday* mutants carrying *Tg[crx:eGFP-WT MAK]* (arrowheads), but not in *pday*

mutants carrying kinase-dead mutant transgenes (asterisks).

Scale bars: 5 μm (C, E) and 40 μm (D).

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Fig. 8: MAK is essential for ciliary axoneme formation in zebrafish photoreceptor

ciliogenesis.

(A) A possible role of MAK in ciliogenesis in zebrafish photoreceptors. During ciliogenesis,

MAK promotes axoneme extension following formation of the transition zone through

phosphorylation of its substrate proteins.

(B) Phenotypes of zebrafish *mak* mutant photoreceptors. In the absence of MAK, the

axoneme does not form, resulting in failure to form the OS and to transport

phototransduction molecules into the OS.

1010

Fig. S1: Visualization of rods and cones in *pday* mutants.

(A) Confocal images of wild-type (the left column) and *pday* mutant retinas (middle column) carrying the transgenes *Tg[rho:NLS-eGFP]* and *Tg[gnat2:NLS-tdTomato]*, which visualize rods (green) and cones (red), respectively. Nuclei were counterstained with Hoechst 33342 (blue). In *pday* mutant retinas, eGFP signals are weak. The right column indicates confocal scanning images of *pday* mutant retinas carrying the transgenes *Tg[rho:NLS-eGFP]* and *Tg[gnat2:NLS-tdTomato]*, with higher laser intensity, to enhance eGFP signals. We compare wild-type and *pday* mutant retinas in conventional scanning conditions and with higher laser intensity, respectively, in Figure 2. Scale bars: $40 \, \mu$m.

(B) Histogram of rod number of wild-type siblings (black) and *pday* mutants (red) retinas at 3, 4, 5 and 6 dpf. Rod number was counted in $1/10^{th}$ divided area of the ONL along the DV axis and analyzed with 2-way ANOVA and Sidak's multiple comparison tests, n=3 for wild-type, n=4 for *pday* mutants at 3 dpf; n=4 for wild-type, n=5 for *pday* mutants at 4 dpf; n=3 for wild-type, n=3 for *pday* mutants at 5 dpf; n=4 for wild-type, n=4 for *pday* mutants at 6 dpf. Bars and lines indicate mean ± SD. * $p<0.02$, ** $p<0.003$, **** $p<0.0001$, ns: not significant.

Fig. S2: Statistical analysis of TUNEL and Bcl2-mediated rescue of photoreceptor phenotypes in *pday* mutants.

(A) The number of TUNEL signals in wild-type and *pday* mutant ONL at 3, 4, and 5 dpf. TUNEL signals in the ONL are higher in *pday* mutants than in wild type at 3 dpf, although the difference is not significant. TUNEL signals in the ONL are the highest in *pday* mutants at 4 dpf and are still significantly higher in *pday* mutants at 5 dpf, compared with wild-type siblings. Bars and lines indicate mean ± SD. Statistical significance was evaluated with 2-way ANOVA and Sidak's multiple comparison tests; * $p<0.03$, **** $p<0.0001$, ns: not significant.

(B) The number of TUNEL signals in wild-type and *pday* mutant INL at 3, 4, and 5 dpf. TUNEL signals in the INL are higher in *pday* mutants than in wild type at 4 dpf, although the difference is not significant. Bars and lines indicate mean ± SD. Statistical significance was evaluated with 2-way ANOVA and Sidak's multiple comparison tests; ns: not significant.

(C) The number of TUNEL signals in wild-type and *pday* mutant RGCL at 3, 4, and 5 dpf. There is no significant difference between wild-type and *pday* mutants at any stage. Bars and lines indicate mean ± SD. Statistical significance was evaluated with 2-way ANOVA and Sidak's multiple comparison tests; ns: not significant.

(D) The fraction of zpr1-positive area in the total retinal in wild-type and *pday* mutants without and with mCherry-Bcl2 overexpression. The fraction of the zpr1-positive area is markedly reduced in *pday* mutants (red bar), compared with wild-type siblings (blue bar). However, the fraction of the zpr1-positive area is recovered in *pday* mutants with mCherry-Bcl2 overexpression (purple bar), equivalent to wild-type without (blue bar) and with mCherry-Bcl2 overexpression (green bar). Bars and lines indicate mean ± SD. Statistical significance was evaluated with 2-way ANOVA and Turkey's multiple comparison tests; *p<0.0332, **p<0.0021, ns: not significant.

(E) The fraction of the zpr3-positive area in the total retinal area in wild-type and *pday* mutants without and with mCherry-Bcl2 overexpression. The fraction of the zpr3-positive area is markedly reduced in *pday* mutants (red bar), compared with wild-type siblings (blue bar). However, the fraction of the zpr3-positive area is recovered and increased in *pday* mutants with mCherry-Bcl2 overexpression (purple bar), compared with wild-type without (blue bar) and with mCherry-Bcl2 overexpression (green bar), probably due to ectopic distribution of zpr3 signals outside the OS. Bars and lines indicate mean ± SD. Statistical significance was evaluated with 2-way ANOVA and Turkey's multiple comparison tests; **p<0.0021, ***p< 0.0002, ****p<0.0001, ns: not significant.

(F) Histogram of zpr1-positive cone thickness in *pday* mutant retinas with (blue) and without (red) overexpression of mCherry-Bcl2. The ONL is divided into 10th area along the DV axis. zpr1-positive cone thickness was measured at the interface between each

neighboring region and evaluated with 2-way ANOVA and Sidak's multiple comparison

tests, n=3 for *pday* mutants, n=4 for *pday* mutants with *Tg[hsp:mCherry-Bcl2]*. **

$p<0.003$, *** $p<0.001$, **** $p<0.0001$. Bars and lines indicate mean ± SD.

(G) Nuclear labeling of wild-type and *pday* mutant ONL overexpressing mCherry-Bcl2 with

Hoechst 33342. Nuclear shape in the ONL is slightly irregular in *pday* mutants

overexpressing Bcl2.

(H) The percentage of OKR+ and OKR- embryos produced by crosses of *pday* heterozygous

male and female fish without (red) and with the transgene *Tg[hsp:mCherry-Bcl2]* (blue).

There is no significant difference between the absence and the presence of

Tg[hsp:mCherry-Bcl2]. Statistical difference as evaluated with 2-way ANOVA and

Sidak's multiple comparison tests. Bars and lines indicate mean ± SD, ns: not significant.

Scale bars: 5 μm (G).

1080

Fig. S3: MAK is localized in ciliary axoneme.

Projection view of 3D confocal image of wild-type transgenic *Tg[crx: eGFP-MAK]* ONL

with anti-GFP and anti-acetylated tubulin antibodies. Nuclei were counterstained with

Hoechst 33342. Left panel indicates a radially sectioned projection view of the retina. The

eGFP-MAK signals overlap with anti-acetylated α-tubulin signals. Right panel indicates

tangentially sectioned projection view, which was digitally produced by rotating the radially

sectioned view using Imaris software (Bitplane). Scale bars: 5 μm.

1088

1089 **Fig. S4: Amino acid comparison between zebrafish, mouse, and human MAK.**

1090 Amino acid sequences of zebrafish, mouse and human MAK proteins are aligned using

1091 Clustal Omega. Grey shading indicates sequence mismatches. The kinase domain is indicated

1092 by an underline. The bottom table shows the percentage of amino acid identity in the full

1093 length protein, the kinase domain, and the C-terminal domain.

1094

1095 **Fig. S5 Measurement of ONL thickness, rod cell number, and the fraction of zpr1-**

1096 **positive area in total retinal area.**

(A) Images with retinas were straightened with Image J (Fuji). Straightened images were divided into 10 equal areas (A) with 9 deciles (D) between adjacent areas. The thickness of the ONL was measured at the deciles and cell numbers were counted in each area.

(B) Cryosections labeled with zpr1 antibody were scanned under a laser-scanning microscope (LSM510; Carl Zeiss, FV3000; Olympus). First, a one-section image containing the central retina per eye was selected from each individual sample. Using the "split channel" command of Image J (Fuji), confocal scanning RGB color images were split into either R, G or B channel. Only the zpr1 channel (green channel in this figure) is used to draw the outline of the total retinal area. Next, using the "threshold" command of Image J (Fuji), we converted zpr1-positive and -negative areas to a binary scale, 1 and 0, respectively. The number of pixels corresponding to 1 or 0 within the neural retina was determined. Using the "measurement" command of Image J (Fuji), the percentage of zpr1-positive areas relative to the total retinal area was calculated as the percentage of the number of 1 pixels relative to the number of 0+1 pixels. Means and standard deviations were calculated from data obtained for 2-5 sections from more than two embryos.

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 Appendix

 Antibodies

NAME	SPECIES	SOURCE	USAGE	REMARK
Zpr-1	Mouse mAb	ZIRC	IHC: 1:100	
Zpr-3	Mouse mAb	ZIRC	IHC: 1:100	recognize zebrafish green opsin and rhodopsin (Gao et al., 2022)
Anti-Rhodopsin	Mouse mAb, clone 1D4	Abcam, ab5417	IHC: 1:100	recognize zebrafish red opsin (Yin et al., 2012)
Anti-GFP	Rabbit pAb	Invitrogen, A11122	IHC: 1:100	
Anti-acetylated Tubulin	Mouse mAb, clone 6-11B-1	Sigma Aldrich, T7451	IHC: 1:250	
Anti-EYS	Rabbit pAb	Novus Bio, NBP1-90038	IHC: 1:300	pH 6.0 HIAR
Anti-gamma Tubulin	Mouse mAb, clone GTU88	Sigma Aldrich, T6557	IHC: 1:100	pH 6.0 HIAR
Anti-MAK	Rabbit pAb	Scrum Inc.	WB: 1:1000	custom ab
Anti-MBP	Mouse mAb, clone SKB3	Merck, 05-675	WB: 1:5000	
Anti-phospho-MBP	Mouse mAb, clone P12	Merck, 05-429	WB: 1:10000	
Anti-GST	Mouse mAb, clone B-14	Santa Cruz, sc-138	WB: 1:10000	
Anti-zebrafish rhodopsin	Rabbit pAb	ZFIN: ZDB-ATB-09058-5	IHC: 1:5000	Ref. Vihtelic and Hyde, 1999 (DOI: 10.1017/s0952523899163168)

 Primers

NAME	SEQUENCE (5' TO 3')	REMARK
z23011_Fw	TTGGGGAAAAAGTCTGCTTG	SSLP BSA
z23011_Rev	TCCAACATGTAAAAACCTGACAA	SSLP BSA
mak-N1_Fw	GTGCAAGGTGTGAATTAGAC	SSLP BSA
mak-N1_Rev	AACTTCCCGACTTCTGTAAC	SSLP BSA
mak_Fw3	ACAGTGGGATGGAGTGGCATCCCTT	mak cDNA cloning
mak_Rev3	TTTCCGAGCGTTGAAAGTAGTCC	mak cDNA cloning
mak-I4_Fw	TGATGTTCACCAGCCACACT	pday site sequence
mak-I3_Rev	TCCCAGTTTTGATGTTGCAT	pday site sequence
mak-E5_Fw	CACAAATGCCAAACCAGAGA	pday site sequence

mak-E4_Rev	TGGTGAAGCTAAAGGAGGTGA	pday site sequence
MAK-AR_Fw	AGTGGCCATCgcgAGAATGAAGAG	AR MAK mutagenesis
MAK-AR_Rev	CTCTTCATTCTcgcGATGGCCACT	AR MAK mutagenesis
MAK-ADF_Fw	cttcGTCTCAACACGATGGTATCGAG	ADF MAK mutagenesis
MAK-ADF_Rev	tctgcGTAAGGTGGCCGAGAGCG	ADF MAK mutagenesis
arl13b_Fw	GGAAATGTTCAATCTGATGGCGAACTG	arl13b cDNA cloning
arl13b_Rev	GTGTTAGGAAATGACATCATGAGTATCGC	arl13b cDNA cloning
T3_mak_Fw	CTAATTAACCCTCACTAAAGGATGAACCGTTACACGACACTCAAG	mak in situ probe
mak_T7_Rev	CTTAATACGACTCACTATAGGGTTAACGATGGCTTCCATATT	mak in situ probe
Z13695_Fw	TGTCGTAGGACATGGGGATT	BSA SSLP
Z13695_Rev	TCGCTGCTTCGTTCTCACTA	BSA SSLP

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Figure 1

Figure 2

Figure 3

Figure 4

Figure 5

Figure 6

Figure 7

Figure 8

Figure S1

Figure S2

anti-acetyl tubulin / anti-GFP

Tg[crx:eGFP-mak]

Figure S3

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                    1                                                                          80
Consensus           MNRYTTMKQLGDGTYGSVLMGKSNESGELVAIKRMKRKFYSWDECMNLREVKSLKKLNHANVIKLKEVIRENDHLYFVFE

zMAK                MNRYTTLKQLGDGTYGSVLMGKSNESGELVAIKRMKRKFYSWEECMNLREVKSLKKLNHANVVKLKEVIRENDHLYFVFE
mMAK                MNRYTTMKQLGDGTYGSVLMGKSNESGELVAIKRMKRKFYSWDECMNLREVKSLKKLNHANVIKLKEVIRENDHLYFVFE
hMAK                MNRYTTMRQLGDGTYGSVLMGKSNESGELVAIKRMKRKFYSWDECMNLREVKSLKKLNHANVIKLKEVIRENDHLYFIFE
                    ..................................................................................
                    81                                                                         160
Consensus           YMKENLYQLMKDRENKLFPESVIRNIMYQILQGLAFIHKHGFFHRDMKPENLLCMGPELVKIADFGLARELRSQPPYTDY

zMAK                YMKENLYQLMKDRENKMFTENEIRNIMFQVLSGLAFVHKHGFFHRDMKPENLLCMGPELVKIADFGLAREIRSRPPYTDY
mMAK                YMKENLYQLMKDR-NKLFPESVIRNIMYQILQGLAFIHKHGFFHRDMKPENLLCMGPELVKIADFGLARELRSQPPYTDY
hMAK                YMKENLYQLMKDR-NKLFPESVIRNIMYQILQGLAFIHKHGFFHRDMKPENLLCMGPELVKIADFGLARELRSQPPYTDY
                    ..................................................................................
                    161                                                                        240
Consensus           VSTRWYRAPEVLLRSSVYSSPIDVWAVGSIMAELYTLRPLFPGTSEVDEIFKICQVLGTPKKSDWPEGYQLASSMNFRFP

zMAK                VSTRWYRAPEVLLRSPVYSSPIDIWAVGCIMAELYTLRPLFPGNSEVDEIFKICQVLGTVKKSDWPEGHQLASAMNFRFP
mMAK                VSTRWYRAPEVLLRSSVYSSPIDVWAVGSIMAELYTFRPLFPGTSEVDEIFKICQVLGTPKKSDWPEGYQLASSMNFRFP
hMAK                VSTRWYRAPEVLLRSSVYSSPIDVWAVGSIMAELYMLRPLFPGTSEVDEIFKICQVLGTPKKSDWPEGYQLASSMNFRFP
                    ..................................................................................
                    241                                                                        320
Consensus           QCVPINLKTLIPNASNEAIQLMTEMLNWDPKKRPTASQALKHPYFQVGQVLGPSAHHLDSKQALHKQLQPLELKPSLQEV

zMAK                QCVPTPLKTLIPNATNEALDIMRDLLQWDPKKRPSAVKALRYPYFQVGQLLGPRPMTPDVRKAPVKAVLPSELRSEMQSV
mMAK                QCIPINLKTLIPNASSEAIQLMTEMLNWDPKKRPTASQALKHPYFQVGQVLGSSAHHLDTKQTLHKQLQPLEPKPSSSER
hMAK                QCVPINLKTLIPNASNEAIQLMTEMLNWDPKKRPTASQALKHPYFQVGQVLGPSSNHLESKQSLNKQLQPLESKPSLVEV
                    ..................................................................................
                    321                                                                        400
Consensus           DPKPLPDIIDQPAGPQPQPKNSHQPLQPIQPPQNLSQHDPPKQQGHEKPPQTLFPSIIKKIPGGENSTLGHKGGRRRWGQT

zMAK                SE---PVLPSQ--TKGQSRNSHQPLQQIPLPQSLTQTDTVA--------------LHTQKVVGGENSLMGLKNGRRRWGQT
mMAK                DPKPLPNILDQPAGPQPQPKQGHQPLQTIQPPQNTVTHPPPKQQGHQKPPQTMFPSIIKTIPVNSVSTLGHKGARRRWGQT
hMAK                EPKPLPDIIDQVVGQPQPKTSQQPLQPIQPPQNLSVQQPPKQQSQEKPPQTLFPSIVKNMPTKPNGTLSHKSGRRRWGQT
                    ..................................................................................
                    401                                                                        480
Consensus           IFKSGDSWDDIEDSDFGASHSKKPSMGACKEKRKKDSPFRFPDPGFSGSNHFKGENKKLPATSRSLKSDSELSTASTAKQ

zMAK                NMKPSDSWDDSDCSDTAASLSKKPSKGPEEEKTTPQPKEQKPLYTFST-------VTKLPNTSRLNQSDSSHQSTSSARQ
mMAK                VFKSGDSCDDIED-DLGASHSKKPSMEACKEK-KKESPFRFPDSGLPVSNHFKGENRNLHA-SVSLKSDPNLSTASTAKQ
hMAK                IFKSGDSWEELEDYDFGASHSKKPSMGVFKEKRKKDSPFRLPEPVPSGSNHSTGENKSLPA-VTSLKSDSELSTAPTSKQ
                    ..................................................................................
                    481                                                                        560
Consensus           YYLKQSRYLPGVNPKNVSLIASGKEINPHFWDNSQLFPKSLGPIGAALAFKRSNAEDSIIKPIEKLSCNEKFAEKLEDPQ

zMAK                HYLRQSRYMPGLISKNTQSLSSK-ESQQDFWDTSALMSKPLAPIGGAPMS-------------------------RISP
mMAK                YYLKQSRYLPGVNPKNVSLVAGGKDINSHSWNN-QLFPKSLGSMGADLSFKRSNAEDSIIKPIENLSCTGKSAEQLEDPQ
hMAK                YYLKQSRYLPGVNPKKVSLIASGKEINPHTWSN-QLFPKSLGPVGAELAFKRSNAEESIIKPIEKLSCNETFPEKLEDPQ
                    ..................................................................................
                    561                                                                        640
Consensus           GNLGSYTTYNQGGYIPSFLKKEVGSAGQRIQLAPLGAQASLDLSATADCKTGKAKPSKSKPSSTVSVNDNSEDDYTWKTK

zMAK                GNFVTTTYNLSGGYIPSFQKKEVGSVGQRIQLAPLGHQHALDLSATADCKTGKAKPSKSKPSSTVSVNDNSEDYDGWKRR
mMAK                GNLGSYTTYNQTGYMPSFLKKEVGSAGQRIQLAPLGASAS---------------------------------DYTWSTK
hMAK                GNLGSYATYNQSGYIPSFLKKEVQSAGQRIHLAPLNATAS--------------------------------EYTWNTK
                    ..................................................................................
                    641                                     685
Consensus           TGRGQFSGRTYNPTAKNSLNIVNRAQPVPSVHGRTDWVAKYGGHR

zMAK                AERTQLKGTSYSALGKNSANLITHAPSVQPVHGRIDWAAKYGSHR
mMAK                TGRGQFSGRTYNPTAKN-LNIVNRTQPVPSVHGRTDWVAKYGGHR
hMAK                TGRGQFSGRTYNPTAKN-LNIVNRAQPIPSVHGRTDWVAKYGGHR
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Amino acid sequence identity percentage

	Full length	Kinase domain	C'-terminus
zMAK vs mMAK	50% (323/646)	86.17% (243/282)	21.82% (79/362)
zMAK vs hMAK	51.38% (333/648)	88.65% (250/282)	21.97% (80/364)
mMAK vs hMAK	83.64% (542/648)	97.86% (275/281)	72.52% (264/364)
Among the three animals	47.22% (306/648)	85.10% (240/282)	17.03% (62/364)

Figure S4

Figure S5