Title: The potassium channel subunit $K_V1.8$ (*Kcna10*) is essential for the distinctive outwardly rectifying conductances of type I and II vestibular hair cells

Hannah R. Martin[1], ORCID 0000-0003-2028-5798
Anna Lysakowski[2], ORCID 0000-0001-6259-0294
Ruth Anne Eatock[1], ORCID 0000-0001-7547-2051

[1]University of Chicago, Department of Neurobiology
[2]University of Illinois at Chicago, Department of Anatomy and Cell Biology

Author for correspondence:
Ruth Anne Eatock
Dept of Neurobiology, J251
University of Chicago
Chicago, IL
eatock@uchicago.edu

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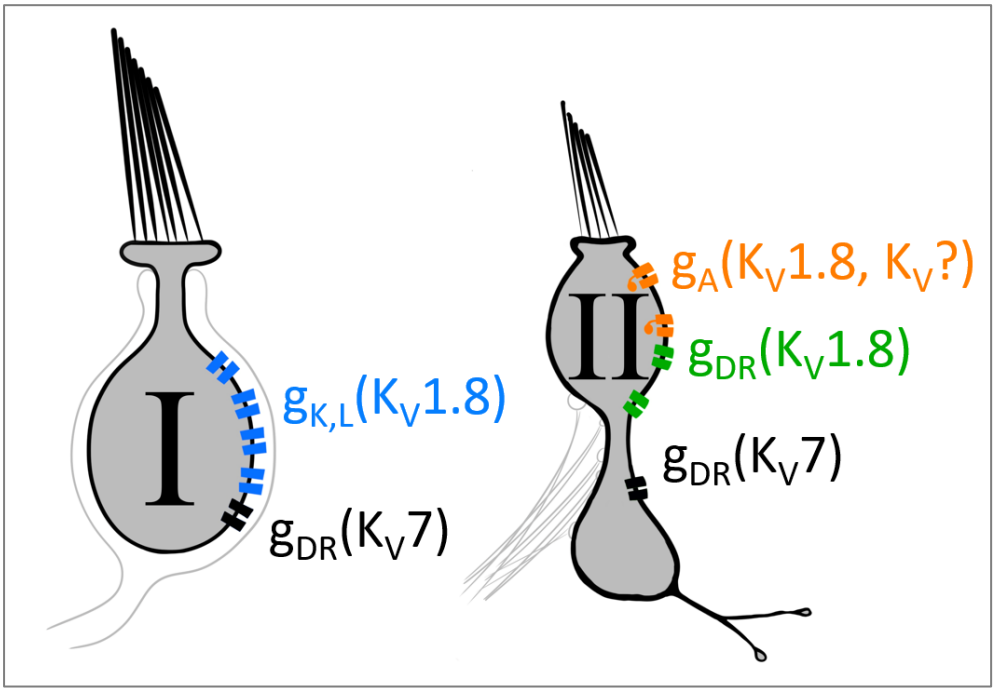
Abstract

In amniotes, head motions and tilt are detected by two types of vestibular hair cells (HCs) with strikingly different morphology and physiology. Mature type I HCs express a large and very unusual potassium conductance, $g_{K,L}$, which activates negative to resting potential, confers very negative resting potentials and low input resistances, and enhances an unusual non-quantal transmission from type I cells onto their calyceal afferent terminals. Following clues pointing to $K_V1.8$ (KCNA10) in the Shaker K channel family as a candidate $g_{K,L}$ subunit, we compared whole-cell voltage-dependent currents from utricular hair cells of $K_V1.8$-null mice and littermate controls. We found that $K_V1.8$ is necessary not just for $g_{K,L}$ but also for fast-inactivating and delayed rectifier currents in type II HCs, which activate positive to resting potential. The distinct properties of the three $K_V1.8$-dependent conductances may reflect different mixing with other K_V subunits that are reported to be differentially expressed in type I and II HCs. In $K_V1.8$-null HCs of both types, residual outwardly rectifying conductances include K_V7 (KCNQ) channels.

Current clamp records show that in both HC types, $K_V1.8$-dependent conductances increase the speed and damping of voltage responses. Features that speed up vestibular receptor potentials and non-quantal afferent transmission may have helped stabilize locomotion as tetrapods moved from water to land.

Graphical abstract



Abbreviations:

g_A, A-type (inactivating) K_V conductance in type II HCs

g_{DR}, delayed rectifier K^+ conductance

$g_{K,L}$, low-voltage-activated K^+ conductance in type I HCs

HC, hair cell

K_V, voltage-gated K^+ conductance

Introduction

The receptor potentials of hair cells (HCs) are strongly shaped by large outwardly rectifying K^+ conductances that are differentially expressed according to HC type. Here we report that a specific voltage-gated K^+ (K_V) channel subunit participates in very different K_V channels dominating the membrane conductances of type I and type II HCs in amniote vestibular organs.

Type I HCs occur only in amniote vestibular organs. Their most distinctive features are that they are enveloped by a calyceal afferent terminal (Wersäll, 1956; Lysakowski and Goldberg, 2004) and that they express $g_{K,L}$ (Correia and Lang, 1990; Rennie and Correia, 1994; Rüsch and Eatock, 1996a): a large non-inactivating conductance with an activation range from –100 to –60 mV, far more negative than other "low-voltage-activated" K_V channels. HCs are known for their large outwardly rectifying K+ conductances, which repolarize membrane voltage following a mechanically evoked perturbation and in some cases contribute to sharp electrical tuning of the hair cell membrane. $g_{K,L}$ is unusually large and unusually negatively activated, and therefore strongly attenuates and speeds up the receptor potentials of type I HCs (Correia et al., 1996; Rüsch and Eatock, 1996b). In addition, $g_{K,L}$ augments non-quantal transmission from type I HC to afferent calyx by providing open channels for K^+ flow into the synaptic cleft (Contini et al., 2012, 2017, 2020; Govindaraju et al., 2023), increasing the speed and linearity of the transmitted signal (Songer and Eatock, 2013).

Type II HCs have compact afferent synaptic contacts (boutons) where the receptor potential drives quantal release of glutamate. They have fast-inactivating (A-type, g_A) and delayed rectifier (g_{DR}) conductances that are opened by depolarization above resting potential (V_{rest}).

The unusual properties of $g_{K,L}$ have long attracted curiosity about its molecular nature. $g_{K,L}$ has been proposed to include "M-like" K_V channels in the K_V7 and/or erg channel families (Kharkovets et al., 2000; Hurley et al., 2006; Holt et al., 2007). The $K_V7.4$ subunit was of particular interest because it contributes to the low-voltage-activated conductance, $g_{K,n}$, in cochlear outer hair cells, but was eventually eliminated as a $g_{K,L}$ subunit by experiments on $K_V7.4$-null mice (Spitzmaul et al., 2013).

Several observations suggested the $K_V1.8$ (KCNA10) subunit as an alternative candidate for $g_{K,L}$. $K_V1.8$ is highly expressed in vestibular sensory epithelia (Carlisle et al., 2012), particularly hair cells (Lee et al., 2013; Scheffer et al., 2015; McInturff et al., 2018), with slight expression elsewhere (skeletal muscle, Lee et al., 2013; kidney, Yao, 2002). $K_V1.8^{-/-}$ mice show absent or delayed vestibular-evoked potentials, the synchronized activity of afferent nerve fibers sensitive to fast linear head motions (Lee et al., 2013). Unique among K_V1 channels, $K_V1.8$ has a cyclic nucleotide binding domain (Lang et al., 2000) with the potential to explain $g_{K,L}$'s known cGMP dependence (Behrend et al., 1997; Chen and Eatock, 2000).

Our comparison of whole-cell currents and immunohistochemistry in type I HCs from $K_V1.8^{-/-}$ and $K_V1.8^{+/+,+/-}$ mouse utricles confirmed that $K_V1.8$ expression is necessary for $g_{K,L}$. More surprisingly, $K_V1.8$ expression is also required for A-type and delayed rectifier conductances of type II HCs. In both HC types, eliminating the $K_V1.8$-dependent major conductances revealed a smaller delayed rectifier conductance involving K_V7 channels. Thus, the distinctive outward rectifiers that produce such different receptor potentials in type I and II HCs both include $K_V1.8$ and K_V7 channels.

Results

We compared whole-cell voltage-activated K^+ currents in type I and type II hair cells from homozygous knockout ($K_V1.8^{-/-}$) animals and their wildtype ($K_V1.8^{+/+}$) or heterozygote ($K_V1.8^{+/-}$) littermates. We immunolocalized $K_V1.8$ subunits in the utricular epithelium and pharmacologically characterized the residual K^+ currents of $K_V1.8^{-/-}$ animals. Current clamp experiments demonstrated the impact of $K_V1.8$-dependent currents on passive membrane properties.

We recorded from three utricular zones: lateral extrastriola (LES), striola, and medial extrastriola (MES) (Fig. 3A.1); striolar and extrastriolar zones have many structural and functional differences and give rise to afferents with different physiology (reviewed in Goldberg, 2000; Eatock and Songer, 2011). Recordings are from 412 type I and II HCs (53% LES, 30% MES, 17% striola) from mice between postnatal day (P) 5 and P370. We recorded from such a wide age range to test for developmental or senescent changes in the impact of the null mutation. Above P18, we did not see substantial changes in K_V channel properties, as reported (González-Garrido et al., 2021).

$K_V1.8^{-/-}$ animals appeared to be healthy and to develop and age normally, as reported (Lee et al., 2013), and hair cells were healthy (see Methods for criteria).

$K_V1.8$ is necessary for $g_{K,L}$ in type I hair cells

The large low-voltage activated conductance, $g_{K,L}$, in $K_V1.8^{+/+,+/-}$ type I hair cells produces distinctive whole-cell current responses to voltage steps, as highlighted by our standard type I voltage protocol (Fig. 1A). From a holding potential within the $g_{K,L}$ activation range (here –74 mV), the cell is hyperpolarized to –124 mV, negative to E_K and the activation range, producing a large inward current through open $g_{K,L}$ channels that rapidly decays as the channels deactivate. We use the large transient inward current as a hallmark of $g_{K,L}$. The hyperpolarization closes all channels, and then the activation function is probed with a series of depolarizing steps, obtaining the maximum conductance from the peak tail current at –44 mV (Fig. 1A). We detected no difference between the Boltzmann parameters of $g_{K,L}$ G-V curves from $K_V1.8^{+/-}$ and $K_V1.8^{+/+}$ type I HCs.

For a similar voltage protocol, $K_V1.8^{-/-}$ type I HCs (Fig. 1B) produced no inward transient current at the step from –74 mV to –124 mV and much smaller depolarization-activated currents during the iterated steps, even at much more positive potentials. Figure 1C compares the conductance-voltage (G-V, activation) curves fit to tail currents (Eq. 1; see insets in Fig. 1A-B): the maximal conductance (g_{max}) of the $K_V1.8^{-/-}$ HC was over 10-fold smaller (Fig. 1C.1), and the curve was positively shifted by >40 mV (Fig. 1C.2). Figure 1D shows the G-V Boltzmann fit parameters for type I HCs from mice >P12, an age at which type I HCs normally express $g_{K,L}$ (Rüsch et al., 1998).

Suppl. Fig. 1 shows how G-V parameters of outwardly rectifying currents in type I HCs changed from P5 to P360. In $K_V1.8^{+/+,+,-}$ mice, the parameters transitioned over the first 15-20 postnatal days from values for a conventional delayed rectifier, activating positive to resting potential, to $g_{K,L}$ values, as previously described (Rüsch et al., 1998; Géléoc et al., 2004; Hurley et al., 2006). Between P5 and P10, some type I HCs have not yet acquired the physiologically defined conductance, $g_{K,L}$. No effects of $K_V1.8$ deletion were

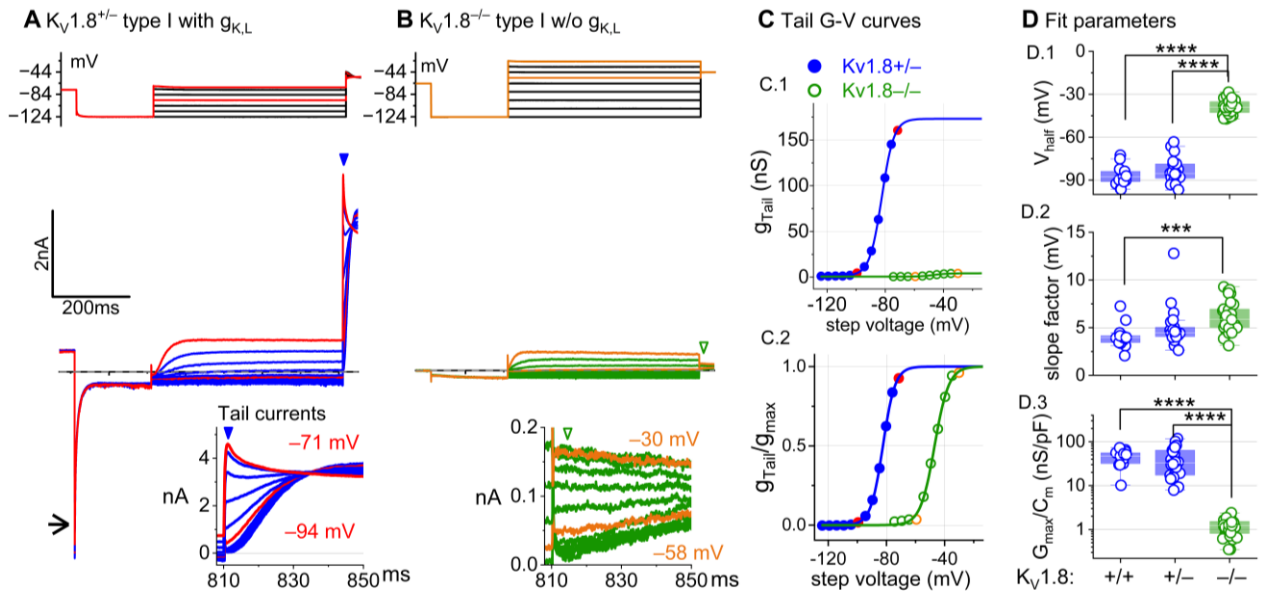


Figure 1. $K_V1.8^{-/-}$ type I hair cells lacked $g_{K,L}$, the dominant conductance in mature $K_V1.8^{+/+,+/-}$ type I HCs. Representative voltage-evoked currents in **(A)** a P22 $K_V1.8^{+/-}$ type I HC and **(B)** a P29 $K_V1.8^{-/-}$ type I HC. (A) *Arrow,* transient inward current that is a hallmark of $g_{K,L}$. *Arrowheads,* tail currents, magnified in *insets*. For steps positive to the midpoint voltage, tail currents are very large. As a result, K^+ accumulation in the calyceal cleft reduces driving force on K^+, causing currents to decay rapidly, as seen in A (Lim et al., 2011). Note that the voltage protocol (top) in B extends to more positive voltages. **(C)** Activation (G-V) curves from tail currents in A and B; symbols, data; curves, Boltzmann fits (Eq. 1). **(D)** Fit parameters from mice >P12 show big effect of $K_V1.8^{-/-}$ and no difference between $K_V1.8^{+/-}$ and $K_V1.8^{+/+}$. *Asterisks* (here and elsewhere): *, p<0.05; **, p<0.01; ***, p<0.001; and ****, p<0.0001. *Line,* median; *Box,* interquartile range; *Whiskers*, outliers. See Table 1 for statistics.

seen in the delayed rectifier currents of immature type I HCs (Suppl. Fig. 1B), showing that they were not immature forms of the $K_V1.8$-dependent $g_{K,L}$ channels.

$K_V1.8^{-/-}$ type I HCs had a much smaller residual delayed rectifier that activated positive to resting potential, with V_{half} ~–40 mV and g_{max} density of 1.3 nS/pF. No additional K^+ conductance activated up to +40 mV, and G-V parameters did not change much with age from P5 to P370. We characterize this $K_V1.8$-independent delayed rectifier later. A much larger non-$g_{K,L}$ delayed rectifier conductance ("$g_{DR,I}$") was reported in our earlier publication on mouse utricular type I HCs (Rüsch et al., 1998). This current was identified as that remaining after "blocking" $g_{K,L}$ with 20 mM external Ba^{2+}. Our new data suggest that there is no large non-$g_{K,L}$ conductance, and that instead high Ba^{2+} positively shifted the apparent voltage dependence of $g_{K,L}$.

$K_V1.8$ strongly affects type I passive properties and responses to current steps.

While the cells of $K_V1.8^{-/-}$ and $K_V1.8^{+/-}$ epithelia appeared healthy, type I hair cells had smaller membrane capacitances (C_m): 4-5 pF in $K_V1.8^{-/-}$ type I HCs, ~20% smaller than $K_V1.8^{+/-}$ type I HCs (~6 pF) and ~30% smaller than $K_V1.8^{+/+}$ type I HCs (6-7 pF; Table 2). C_m scales with surface area, but soma sizes were unchanged by deletion of $K_V1.8$ (Suppl. Table 2). Instead, C_m may be higher in $K_V1.8^{+/+}$ cells because of $g_{K,L}$ for two reasons. First, highly expressed trans-membrane proteins (see discussion of $g_{K,L}$ channel density in Chen and Eatock, 2000) can affect membrane thickness (Mitra et al., 2004), which is inversely proportional to specific C_m. Second, $g_{K,L}$ could contaminate estimations of capacitive current, which is

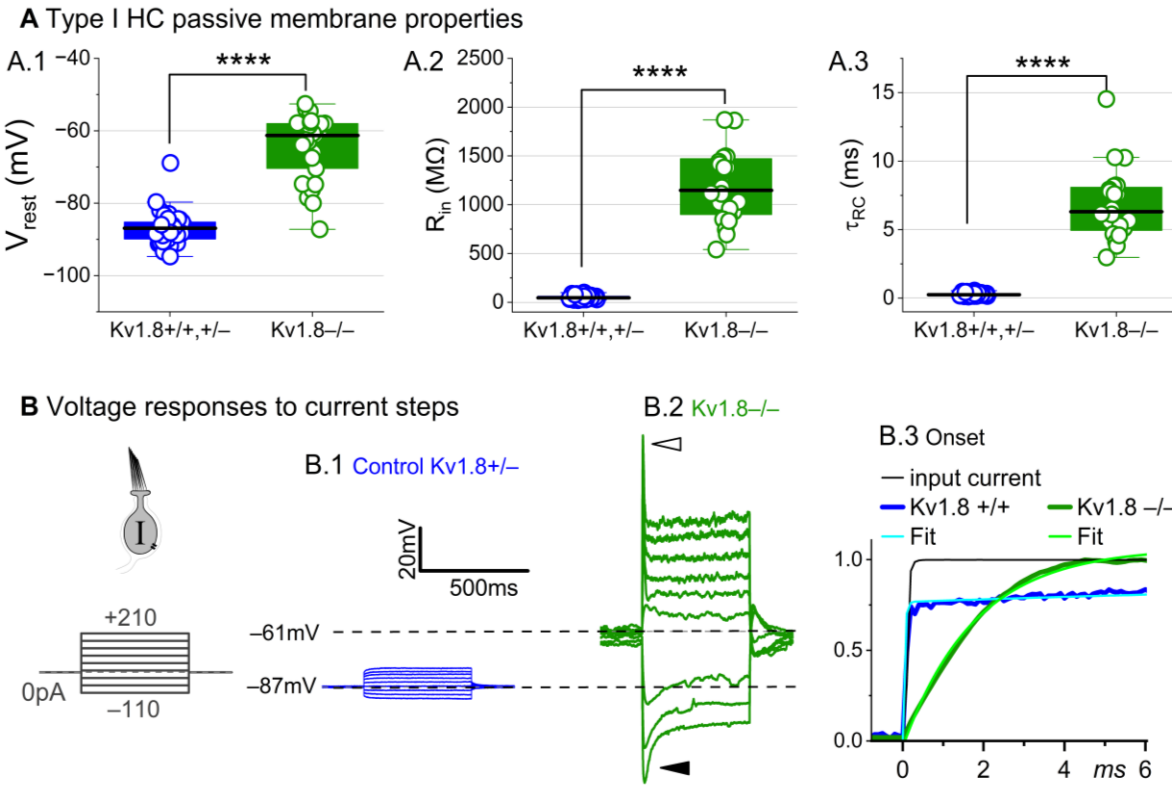


Figure 2. $K_V1.8^{-/-}$ type I hair cells had much longer membrane charging times and higher input resistances (voltage gains) than $K_V1.8^{+/+,+/-}$ type I HCs. (A) $g_{K,L}$ strongly affects passive membrane properties: (**A.1**) V_{rest} , (**A.2**) R_{in}, input resistance, and (**A.3**) membrane time constant, τ_{RC} = (R_{input} * C_m). See Table 2 for statistics. **(B)** Current clamp responses to the same scale from (B.1) $K_V1.8^{+/-}$ and (B.2) $K_V1.8^{-/-}$ type I cells, both P29. *Filled arrowhead (B.2),* sag indicating I_H activation. *Open arrowhead*, Depolarization rapidly decays as I_{DR} activates. B.3, The 1st 6 ms of voltage responses to 170-pA injection is normalized to steady-state value; overlaid curves are double-exponential fits ($K_V1.8^{+/+}$, τ 40 µs and 2.4 ms) and single-exponential fits ($K_V1.8^{-/-}$, τ 1.1 ms) .

calculated from the decay time constant of transient current evoked by a small voltage step *outside* the operating range of any ion channels. $g_{K,L}$ has such a negative operating range that, even for V_m negative to –90 mV, some $g_{K,L}$ channels are voltage-sensitive and could add to capacitive current.

Basolateral conductances help set the resting potential and passive membrane properties that regulate the time course and gain of voltage responses to small currents. To examine the effect of $K_V1.8$ on these properties, we switched to current clamp mode and measured resting potential (V_{rest}), input resistance (R_{in}, equivalent to voltage gain for small current steps, △V/△I), and membrane time constant (τ_{RC}). In $K_V1.8^{-/-}$ type I HCs, V_{rest} was much less negative (Fig. 2A.1), R_{in} was greater by ~20-fold (Fig. 2A.2), and membrane charging times were commensurately longer (Fig. 2A.3).

The differences between the voltage responses of $K_V1.8^{+/+,+/-}$ and $K_V1.8^{-/-}$ type I HCs are expected from the known impact of $g_{K,L}$ on V_{rest} and R_{in} (Correia and Lang, 1990; Ricci et al., 1996; Rüsch and Eatock, 1996b; Songer and Eatock, 2013). The large K^+-selective conductance at V_{rest} holds V_{rest} close to E_K (K^+ equilibrium potential) and minimizes gain (△V/△I), such that voltage-gated conductances are negligibly affected by the input current and the cell produces approximately linear, static responses to iterated current steps. For $K_V1.8^{-/-}$ type I HCs, with their less negative V_{rest} and larger R_{in}, positive current steps evoked a fast initial depolarization (Fig. 2B.2), activating residual delayed rectifiers and repolarizing the

Table 1. *Type I hair cell K_V activation voltage dependence and kinetics. Mean ± SEM (number of cells). g is effect size, Hedge's g. KWA is Kruskal-Wallis ANOVA.*

Zone	$K_V1.8$	Tail $V_{1/2}$, mV [a]	Tail S, mV [b]	Tail G_{max}, nS [c]	Tail G_{max}/C_m, nS/pF [d]	Age (median, range)
Extrastriola	+/+	–85 ± 2 (12)	4.3 ± 0.4 (12)	270 ± 40 (11)	47 ± 8 (11)	22, 14-287
	+/–	–83 ± 1 (40)	5.2 ± 0.3 (40)	210 ± 20 (40)	37 ± 4 (40)	19, 13-259
	–/–	–40.2 ± 0.9 (26)	5.7 ± 0.3 (26)	5.4 ± 0.3 (26)	1.11 ± 0.08 (26)	45, 14-277
Striola	+/+	–87 ± 3 (6)	4.3 ± 0.3 (6)	310 ± 70 (6)	41 ± 7 (6)	40, 15-59
	+/–	–88 ± 2 (3)	4.7 ± 0.9 (3)	270 ± 60 (3)	44 ± 6 (3)	19, 14-20
	–/–	–38 ± 1 (13)	6.2 ± 0.4 (13)	6.5 ± 0.6 (13)	1.5 ± 0.1 (13)	202, 14-370

[a] –/– vs +/+: 2-way ANOVA, p < 1E-9, g 7.7; –/– vs +/–: 2-way ANOVA, p < 1E-9, g 6.8
[b] –/– vs +/+: 2-way ANOVA, p = 8.4E-4, g 1.2
[c] –/– vs +/+: 2-way ANOVA, p < 1E-9, g 3.7; –/– vs +/–: 2-way ANOVA, p < 1E-9, g 2.1
[d] –/– vs +/+: 2-way ANOVA, p < 1E-9, g 3.6; –/– vs +/–: 2-way ANOVA, p < 1E-9, g 2.0

Table 2. *Type I hair cell* p*assive membrane properties. Mean ± SEM (number of cells). g is effect size, Hedge's g. KWA is Kruskal-Wallis ANOVA.*

Zone	*$K_V1.8$*	*V_{rest}, mV* [a, b]	R_{input}, MΩ [c]	τ_{RC}, ms [d]	C_m (pF) [e]	Age (median, range)
Extrastriola	+/+	–84 ± 3 (6)	44 ± 6 (6)	0.24 ± 0.03 (6)	6.1 ± 0.4 (13)	19.5, 14-287
	+/–	–88.0 ± 0.7 (28)	55 ± 5 (24)	0.32 ± 0.03 (23)	5.8 ± 0.2 (44)	21, 16-29
	–/–	–63 ± 2 (15)	1400 ± 100 (15)	6.4 ± 0.6 (15)	5 ± 0.2 (27)	45, 14-202
Striola	+/+	–87 ± 2 (4)	50 ± 8 (4)	0.30 ± 0.04 (4)	7.4 ± 0.7 (7)	43, 40-59
	+/–	–87 ± 3 (3)	38 ± 8 (2)	0.21 ± 0.01 (2)	5.9 ± 0.6 (3)	19, 19-20
	–/–	–74 ± 5 (5)	1000 ± 300 (4)	4.2 ± 1.0 (4)	4.4 ± 0.2 (14)	202, 24-370

[a] striolar –/– vs ES –/–: 2-way ANOVA, p = 0.006, g 1.2; striolar –/– vs striolar +/+,+/–: 2-way ANOVA, p = 0.005, g 1.7
[b] –/– vs +/+: 2-way ANOVA, p < 1E-9, g 2.3; –/– vs +/–: 2-way ANOVA, p < 1E-9, g 3.4
[c] –/– vs +/+: 2-way ANOVA, p < 1E-9, g 3.1; –/– vs +/–: 2-way ANOVA, p < 1E-9, g 3.9
[d] –/– vs +/+: 2-way ANOVA, p < 1E-9, g 2.7; –/– vs +/–: 2-way ANOVA, p < 1E-9, g 3.4
[e] –/– vs +/+: 2-way ANOVA, p = 3E-7, g 1.5; –/– vs +/–: 2-way ANOVA, p = 1.3E-4, g 1.0; +/– vs +/+: 2-way ANOVA, p = 0.048, g 0.6

membrane toward E_K. Negative current steps evoked an initial "sag" (Fig. 2B.2), a hyperpolarization followed by slow repolarization as the HCN1 channels open (Rüsch and Eatock, 1996b; Horwitz et al., 2011).

Overall, comparison of the $K_V1.8^{+/+,+/-}$ and $K_V1.8^{-/-}$ responses shows that with $K_V1.8$ ($g_{K,L}$), the voltage response of the type I hair cell is smaller but better reproduces the time course of the input current.

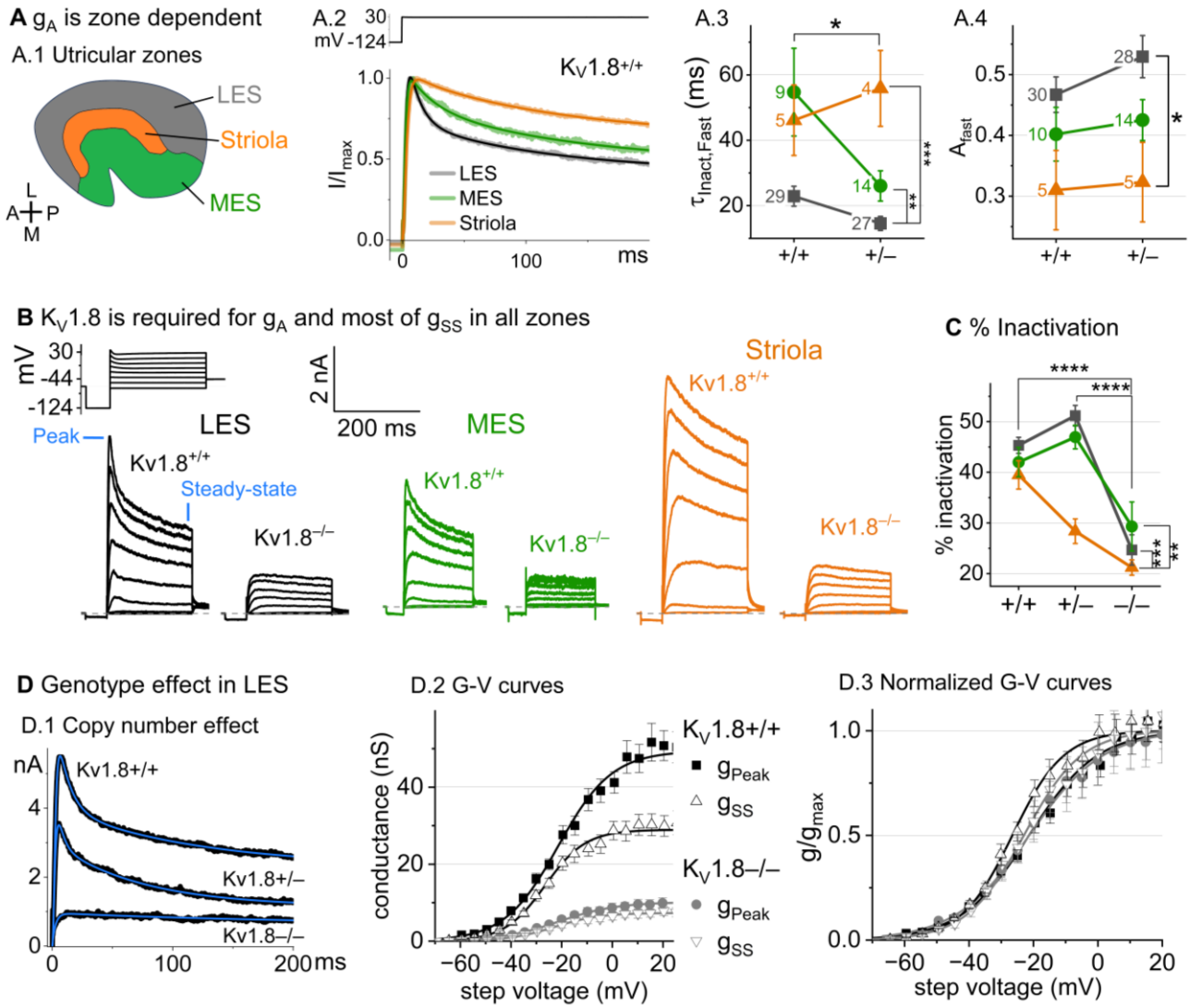


Figure 3. $K_V1.8^{-/-}$ type II HCs in all zones of the sensory epithelium lacked the major rapidly inactivating conductance, g_A, and had less delayed rectifier conductance. Activation and inactivation varied with epithelial zone and genotype. (**A**) g_A inactivation time course varied across zones. (**A.1**) Zones of the utricular epithelium. (**A.2**) Normalized currents evoked by steps from –124 mV to +30 mV with overlaid fits of Eq. 3. (**A.3**) $\tau_{Inact,Fast}$ was faster in $K_V1.8^{+/-}$ than $K_V1.8^{+/+}$ HCs , and faster in LES than other zones. Brackets show post hoc pairwise comparisons between two zones (vertical brackets) and horizontal brackets compare two genotypes; see Table 3 for statistics. (**A.4**) Fast inactivation was a greater fraction of total inactivation in LES than striola.
(**B**) Exemplars; ages, *left to right*, P312, P53, P287, P49, P40, P154.
(**C**) % inactivation at 30 mV was much lower in $K_V1.8^{-/-}$ than $K_V1.8^{+/-}$ and $K_V1.8^{+/+}$, and lower in striola than LES and MES. Interaction between zone and genotype was significant (Table 3).
(**D**) Exemplar currents and G-V curves from LES type II HCs show a copy number effect. (**D.1**) Currents for examples of the 3 genotypes evoked by steps from –124 mV to +30 mV fit with Eq. 3. (**D.2**) Averaged peak and steady-state conductance-voltage datapoints from LES cells (+/+, n=37; –/–, n=20) were fit with Boltzmann equations (Eq. 1) and normalized by g_{max} in (**D.3**). See Table 4 for statistics.

$K_V1.8$ is necessary for both inactivating and non-inactivating K_V currents in type II hair cells

Type II HCs also express $K_V1.8$ mRNA (McInturff et al., 2018; Orvis et al., 2021). Although their outwardly-rectifying conductances (g_A and g_{DR}) differ substantially in voltage dependence and size from $g_{K,L}$, both conductances were strongly affected by the null mutation: g_A was eliminated and the delayed rectifier

was substantially smaller. Below we describe g_A and g_{DR} in $K_V1.8^{+/+,+/-}$ type II HCs and the residual outward rectifying current in $K_V1.8^{-/-}$ type II HCs.

$K_V1.8^{+/+,+/-}$ type II HCs. Most (81/84) $K_V1.8^{+/+,+/-}$ type II HCs expressed a rapidly-activating, rapidly-inactivating A-type conductance (g_A). We define A current as the outwardly rectifying current that inactivates by over 30% within 200 ms. g_A was more prominent in extrastriolar zones, as reported (Holt et al. 1999, Weng and Correia 1999).

We compared the activation and inactivation time course and inactivation prominence for 200-ms steps from –124 mV to ~30 mV. Outward currents fit with Eq. 3 yielded fast inactivation time constants ($\tau_{Inact,\ Fast}$) of ~30 ms in LES (Fig. 3A.2). $\tau_{Inact,\ Fast}$ was faster in LES than in MES or striola (Fig. 3A.3) and fast inactivation was a larger fraction of the total inactivation in LES than striola (~0.5 vs. 0.3, Fig. 3A.4).

To show voltage dependence of activation, we generated G-V curves for peak currents (sum of A-current and delayed rectifier) and steady-state currents measured at 200 ms, after g_A has mostly inactivated (Figure 3D.2). $K_V1.8^{+/-}$ HCs had smaller currents than $K_V1.8^{+/+}$ HCs, reflecting a smaller g_{DR} (Fig. 3D) and faster fast inactivation (Fig. 3A.3). As discussed later, these effects may relate to effects of the $K_V1.8$ gene dosage on the relative numbers of different $K_V1.8$ heteromers.

For $K_V1.8^{+/+}$ and $K_V1.8^{+/-}$ HCs, the voltage dependence as summarized by V_{half} and slope factor (S) was similar. Relative to g_{SS}, g_{Peak} had a more positive V_{half} (~–21 vs. ~–26) and greater S (~12 vs. ~9, Fig. 3D, Table 4). Because g_{Peak} includes channels with and without fast inactivation, the shallower g_{Peak}-V curve may reflect a more heterogeneous channel population. Only g_{Peak} showed zonal variation, with more positive V_{half} in LES than striola (~–20 mV vs. ~–24 mV, Fig. 3D, Table 4). We later suggest that variable subunit composition may drive zonal variation in g_{Peak}.

$K_V1.8^{-/-}$ type II HCs from all zones were missing g_A and 30-50% of g_{DR} (Fig. 3B-D). The residual delayed rectifier (1.3 nS/pF) had a more positive V_{half} than g_{DR} in $K_V1.8^{+/+,+/-}$ HCs (~–20 mV *vs.* ~–26 mV, Fig. 3D.2). We refer to the $K_V1.8$-dependent delayed rectifier component as $g_{DR}(K_V1.8)$ and to the residual, $K_V1.8$-*in*dependent delayed rectifier component as $g_{DR}(K_V7)$ because, as we show later, it includes K_V7 channels.

Supplemental Figure 3A shows the development of $K_V1.8$-dependent and independent K_V currents in type II HCs with age from P5 to over P300. In $K_V1.8^{+/+,+/-}$ type II HCs, g_A was present at all ages with a higher % inactivation after P18 than at P5-P10 (Suppl. Fig. 3A.4). g_{Peak} did not change much above P12 except for a compression of conductance density from P13 to P370 (partial correlation coefficient = –0.4, p = 2E-5, Suppl. Fig. 3A.3).

We saw small rapidly inactivating outward currents in a minority of $K_V1.8^{-/-}$ type II HCs (23%, 7/30), all >P12 and extrastriolar (Suppl. Fig. 4). These currents overlapped with g_A in percent inactivation, inactivation kinetics, and activation voltage dependence but were very small. As discussed later, we suspect that these currents flow through homomers of inactivating K_V subunits that in control hair cells join with $K_V1.8$ subunits and confer inactivation on the heteromeric conductance.

$K_V1.8$ affects type II passive properties and responses to current steps.

In type II HCs, absence of $K_V1.8$ did not change V_{rest} (Fig. 4A.1) because g_A and g_{DR} both activate positive to rest, but significantly increased R_{in} and τ_{RC} (Fig. 4A.2-A.3).

Table 3. *Type II hair cell K_V currents: Activation and inactivation time course at +30 mV. Mean ± SEM. g is effect size, Hedge's g. KWA is Kruskal-Wallis ANOVA.*

Zone	$K_V1.8$	τ_{Act} at 30 mV, ms [a, b]	$\tau_{Inact,Fast}$ at 30 mV, ms [c, d]	Fast inactivation prominence [e]	Inactivation % [f, g]	N cells	Age (median, range)
LES	+/+	2.11 ± 0.09	23 ± 3	0.46 ± 0.03	45 ± 2	30	46, 14-312
	+/–	1.64 ± 0.09	15 ± 2	0.53 ± 0.03	51 ± 2	27	29, 13-280
	–/–	4.4 ± 0.5	*NA*	*NA*	25 ± 3	21	128, 15-355
MES	+/+	2.8 ± 0.5	50 ± 10	0.40 ± 0.04	42 ± 3	9	94, 22-296
	+/–	2.2 ± 0.2	90 ± 60	0.42 ± 0.03	47 ± 2	15	24, 13-52
	–/–	10 ± 7	*NA*	*NA*	29 ± 5	10	84, 28-355
Striola	+/+	2.7 ± 0.3	50 ± 10	0.31 ± 0.07	39 ± 3	5	45, 40-287
	+/–	2.9 ± 0.4	300 ± 200	0.3 ± 0.06	28 ± 2	5	19, 14-30
	–/–	7 ± 2	*NA*	*NA*	22 ± 2	6	202, 29-298

[a] –/– vs +/+: KWA, p = 0.0048, g 0.6; –/– vs +/–: KWA, p = 2.3E-7, g 0.6
[b] Striola vs LES: KWA, p = 5.7E-4, g 1.0
[c] +/– vs +/+: KWA, p = 0.027, g 0.2
[d] LES vs MES: KWA, p = 0.0018, g 0.3; LES vs Striola: KWA, p = 1.9E-4, g 0.8
[e] LES vs Striola: 2-way ANOVA, p = 0.0041, g 0.7
[f] –/– vs +/+: 2-way ANOVA, p < 1E-9, g 1.7; –/– vs +/–: 2-way ANOVA, p < 1E-9, g 1.8
[g] Striola vs LES: 2-way ANOVA, p = 3.4E-5, g 0.9; Striola vs MES: 2-way ANOVA, p = 0.0011, g 1.0; Interaction between genotype and Zone: 2-way ANOVA, p = 0.026

Table 4. *Type II hair cell K_V currents: Activation voltage dependence. Mean ± SEM. g is effect size, Hedge's g. KWA is Kruskal-Wallis ANOVA.*

Zone	*$K_V1.8$*	*Peak $V_{1/2}$, mV* [a]	Peak S, mV [b, c]	A-type g_{max}/C_m, nS/pF [d]	SS V_{half}, mV [e]	SS S, mV [f]	SS g_{max}/C_m, nS/pF [g, h]	N cells	Age (median, range)
LES	+/+	–19.8 ± 0.6	11.8 ± 0.4	4.0 ± 0.3	–25.0 ± 0.5	8.7 ± 0.3	7.1 ± 0.8	37	46, 14-312
	+/–	–19.8 ± 0.8	12.8 ± 0.4	3.8 ± 0.3	–26.8 ± 0.8	8.7 ± 0.3	4.9 ± 0.4	35	29, 13-280
	–/–	–18 ± 1	11.7 ± 0.4	0.37 ± 0.05	–19 ± 1	12.1 ± 0.5	1.8 ± 0.2	20	128, 15-355
MES	+/+	–22 ± 1	11 ± 0.7	4.1 ± 0.7	–26 ± 1	8.3 ± 0.5	9 ± 1	11	94, 22-296
	+/–	–21 ± 1	11.8 ± 0.4	3.6 ± 0.5	–27 ± 1	9.0 ± 0.3	5.9 ± 0.7	16	24, 13-52
	–/–	–19 ± 1	10.8 ± 0.6	0.6 ± 0.1	–20 ± 1	10.7 ± 0.7	2.5 ± 0.3	15	84, 28-355
Striola	+/+	–24 ± 1	9.6 ± 0.5	5 ± 1	–26.6 ± 0.9	8.2 ± 0.4	12 ± 1	7	45, 40-287
	+/–	–25 ± 2	9.4 ± 0.4	2.6 ± 0.6	–28 ± 2	8.2 ± 0.3	10 ± 2	6	19, 14-30
	–/–	–21.3 ± 0.9	10.3 ± 0.5	0.7 ± 0.1	–21.7 ± 0.8	10.5 ± 0.6	3.9 ± 0.5	8	202, 29-298

[a] Striola vs LES: 2-way ANOVA, p = 0.00116, g 0.9
[b] Striola vs MES: 2-way ANOVA, p = 0.016, g 0.8; Striola vs LES: 2-way ANOVA, p = 7.5E-6, g 1.2
[c] –/– vs +/–: 2-way ANOVA, p = 0.036, g 0.5
[d] –/– vs +/+: Welch ANOVA, p < 1E-9, g 2.3; –/– vs +/–: Welch ANOVA, p < 1E-9, g 2.3
[e] –/– vs +/+: 2-way ANOVA, p < 1E-9, g 1.4; –/– vs +/–: 2-way ANOVA, p < 1E-9, g 1.6
[f] –/– vs +/+: 2-way ANOVA, p < 1E-9, g 1.4; –/– vs +/–: 2-way ANOVA, p = 4.5E-7, g 1.1
[g] –/– vs +/+: Welch ANOVA, p < 1E-9, g 1.6; –/– vs +/–: Welch ANOVA, p < 1E-9, g 1.3; +/+ vs +/–: Welch ANOVA, p = 0.007, g 1.6
[h] Striola vs LES: 1-way ANOVA, p = 0.001, g 0.9); Striola vs MES: 1-way ANOVA, p = 0.01, g 0.8

222 Positive current steps evoked an initial depolarizing transient in both $K_V1.8^{+/+}$ and $K_V1.8^{-/-}$ type II HCs,
223 but the detailed time course differed (Fig. 4B). Both transient and steady-state responses were larger in

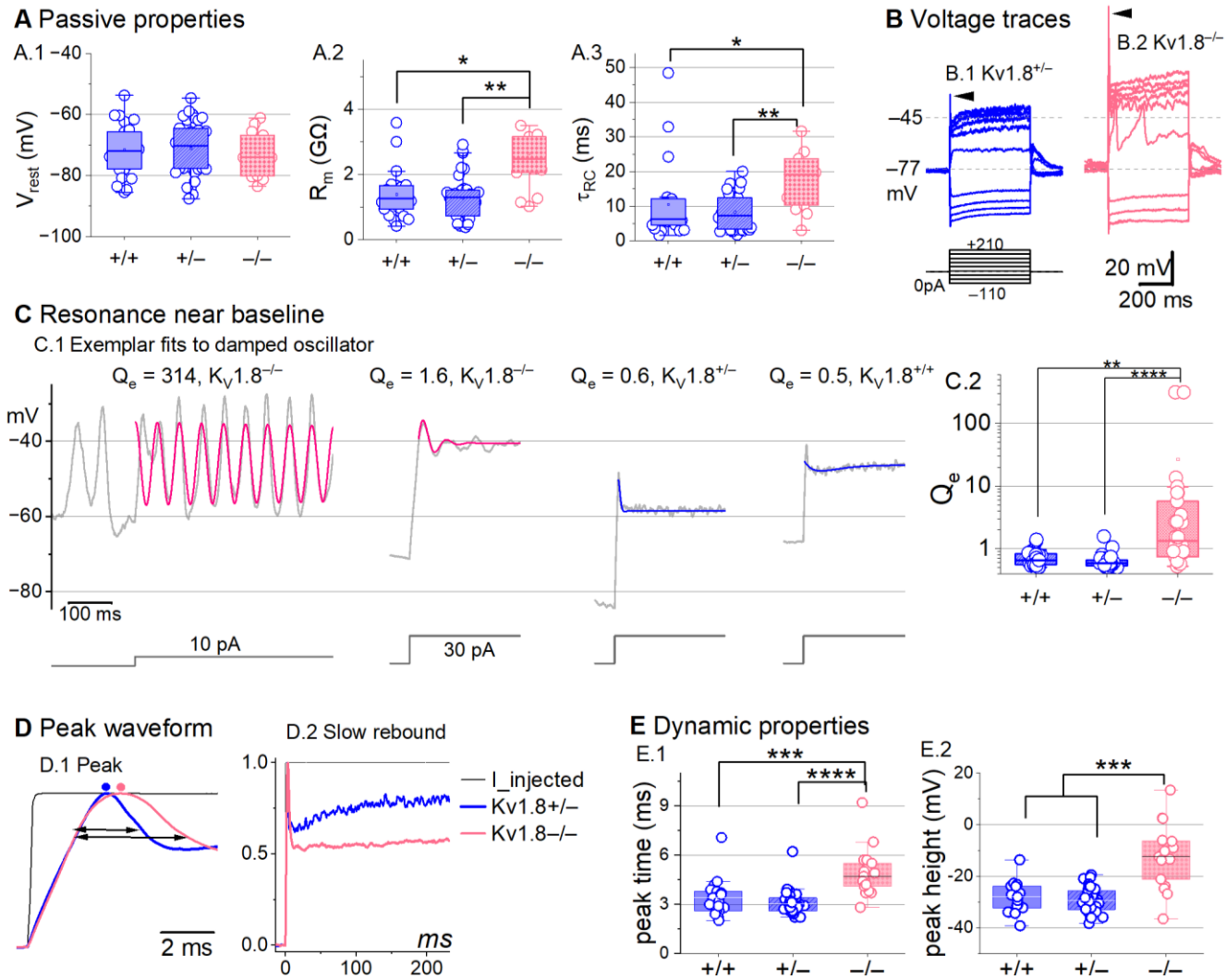


Figure 4. $K_V1.8^{-/-}$ type II hair cells had larger, slower voltage responses and more electrical resonance. **(A)** Passive membrane properties near resting membrane potential: A.1) Resting potential. R_{input} (A.2) and τ_{RC} (A.3) were obtained from single exponential fits to voltage responses < 15 mV. See Table 5 for statistics.
(**B**) Exemplar voltage responses to iterated current steps (*bottom)* illustrate key changes in gain and resonance with $K_V1.8$ knockout. (**B.1**) $K_V1.8^{+/-}$ type II HC (P24, LES) and (**B.2**) $K_V1.8^{-/-}$ type II HC (P53, LES). *Arrowheads,* depolarizing transients.
(**C**) Range of resonance illustrated for $K_V1.8^{-/-}$ type II HCs (*left, pink curves fit to Eq. 5*) and controls (*right, blue fits*). (**C.1**) *Resonant frequencies, left to right:* 19.6, 18.4, 34.4, 0.3 Hz. Leftmost cell resonated spontaneously (before step). (**C.2**) Tuning quality (Q_e; Eq. 6) was higher for $K_V1.8^{-/-}$ type II HCs (KWA, p = 0.0064 vs. $K_V1.8^{+/+}$; p = 7E-8 vs. $K_V1.8^{+/-}$).
(**D**) $K_V1.8^{-/-}$ type II HCs had higher, slower peaks and much slower rebound potentials in response to large (170-pA) current steps. (**D.1**) Normalized to show initial depolarizing transient (*filled circles*, times of peaks; *horizontal arrows*, peak width at half-maximum). (**D.2**) Longer time scale to highlight how null mutation reduced post-transient rebound.
(**E**) In $K_V1.8^{-/-}$ HCs, depolarizing transients evoked by a +90-pA step were slower to peak (**E.1**) and (**E.2**) larger.

$K_V1.8^{-/-}$, consistent with their larger R_{in} values.

Absence of $K_V1.8$ increased the incidence of sharp electrical resonance in type II HCs. Electrical resonance, which manifests as ringing responses to current steps, can support receptor potential tuning (Ashmore, 1983; Fettiplace, 1987; Hudspeth and Lewis, 1988; Ramanathan and Fuchs, 2002). Larger R_{in} values made $K_V1.8^{-/-}$ type II HCs more prone to electrical resonance; Figure 4C.1 shows a striking example.

Table 5. *Type II hair cell passive membrane properties. Mean ± SEM (number of cells). g is effect size, Hedge's g. KWA is Kruskal-Wallis ANOVA.*

Zone	$K_V1.8$	V_{rest}, mV	R_{input}, GΩ [a]	τ_{RC}, ms [b]	Peak height, mV, 170 [c]	Peak time, ms [d]	C_m, pF	Age (median, range)
Extrastriola	+/+	–71 ± 2 (19)	1.4 ± 0.2 (16)	11 ± 3 (16)	–20 ± 2 (15)	2.5 ± 0.2 (15)	4.7 ± 0.2 (50)	45, 16-312
	+/–	–71 ± 2 (34)	1.2 ± 0.1 (27)	9 ± 1 (27)	–20 ± 1 (30)	2.44 ± 0.08 (30)	4.6 ± 0.1 (52)	27, 13-280
	–/–	–76 ± 2 (9)	2.3 ± 0.3 (7)	16 ± 3 (7)	2 ± 6 (7)	3.6 ± 0.3 (7)	4.6 ± 0.2 (35)	53, 15-154
Striola	+/+	–73.1 ± 1.0 (6)	1.4 ± 0.1 (6)	9 ± 1 (6)	–20 ± 2 (5)	2.7 ± 0.1 (5)	4.6 ± 0.2 (7)	45, 40-224
	+/–	–71 ± 3 (5)	1.4 ± 0.3 (6)	7 ± 2 (6)	–20 ± 2 (6)	2.3 ± 0.1 (6)	4.8 ± 0.2 (6)	19, 19-30
	–/–	–68 ± 2 (6)	3.0 ± 0.7 (6)	26 ± 10 (6)	2 ± 7 (4)	4 ± 1 (4)	4.4 ± 0.3 (7)	178, 29-298

[a] –/– vs +/+: KWA, p = 0.015, g 1.2; –/– vs +/–: KWA, p = 0.002, g 1.5
[b] –/– vs +/+: KWA, p = 0.016, g 0.7; –/– vs +/–: KWA, p = 0.008, g 1.2
[c] –/– vs +/+: KWA, p = 0.006, g 2.1; –/– vs +/–: KWA, p = 2E-4, g 2.6
[d] –/– vs +/+: 2-way ANOVA, p <1E-9, g 1.3; –/– vs +/–: 2-way ANOVA, p < 1E-9, g 1.9

Median resonance quality (Q_e, sharpness of tuning) was greater in $K_V1.8^{-/-}$ (1.33, n=26) than $K_V1.8^{+/+}$ (0.66, n=23) or $K_V1.8^{+/-}$ (0.59, n=44) type II HCs.

$K_V1.8$ affected the time course of the initial peak in response to much larger current injections (Fig. 4D-E). Fast activation of g_A in control type II HCs rapidly repolarizes the membrane and then inactivates, allowing the constant input current to progressively depolarize the cell, producing a slow rebound (Fig 4D.2). This behavior has the potential to counter mechanotranduction adaptation (Vollrath and Eatock, 2003).

$K_V1.8$ immunolocalized to basolateral membranes of both type I and II HCs

If $K_V1.8$ is a pore-forming subunit in the $K_V1.8$-dependent conductances $g_{K,L}$, g_A, and g_{DR}, it should localize to hair cell membranes. Figure 5 compares $K_V1.8$ immunoreactivity in $K_V1.8^{+/+}$ and $K_V1.8^{-/-}$ utricles, showing specific immunoreactivity along the basolateral membranes of both hair cell types in $K_V1.8^{+/+}$ utricles. To identify hair cell type and localize the hair cell membrane, we used antibodies against $K_V7.4$ (KCNQ4), an ion channel densely expressed in the calyceal "inner-face" membrane next to the synaptic cleft (Hurley et al., 2006; Lysakowski et al., 2011), producing a cup-like stain around type I HCs (Fig. 5A). $K_V1.8$ immunoreactivity was present in hair cell membrane apposing $K_V7.4$-stained calyx inner face in $K_V1.8^{+/+}$ utricles (Fig. 5A.1, A.2) and not in $K_V1.8^{-/-}$ utricles (Fig. 5A.3).

In other experiments, we used antibodies against calretinin (Calb2), a cytosolic calcium binding protein expressed by many type II HCs and also by striolar calyx-only afferents (Desai et al., 2005; Lysakowski et al., 2011) (Fig. 5B). A hair cell is type II if it is calretinin-positive (Fig. 5B.1) or if it lacks a $K_V7.4$-positive or

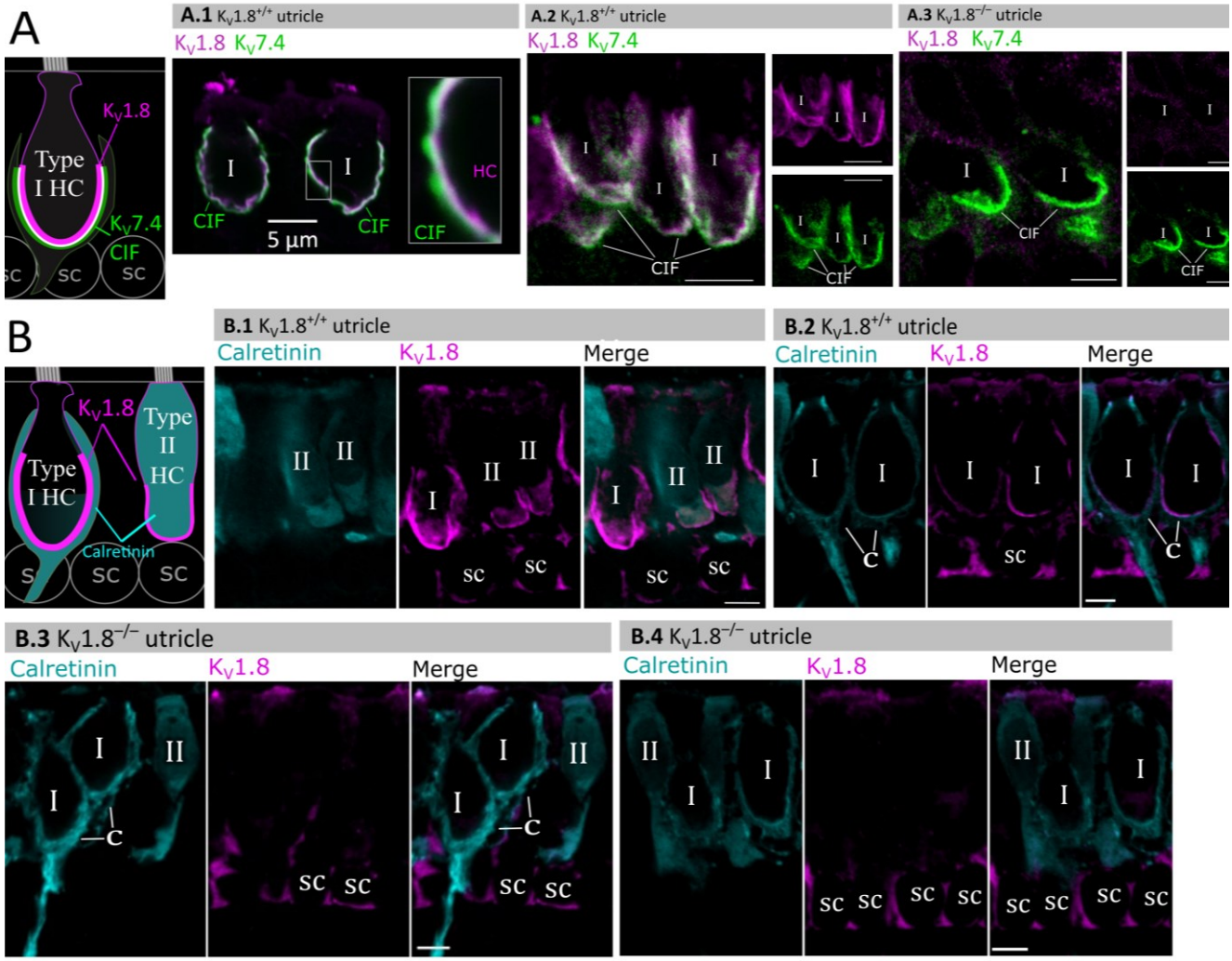


Figure 5. **Type I and type II HC basolateral membranes show specific immunoreactivity to Kv1.8 antibody (magenta).** Antibodies for $K_V7.4$ (A, green) and calretinin (B, cyan) were used as counterstains for calyx membrane (Kv7.4), type II HC cytoplasm (calretinin) and cytoplasm of striolar calyx-only afferents (calretinin). (**A**) *Left,* Cartoon showing $K_V7.4$ on the calyx inner face membrane (CIF) and $K_V1.8$ on the type I HC membrane. SC, supporting cell nuclei. *A.1-3*, Adult mouse utricle sections. $K_V7.4$ antibody labeled calyces on two $K_V1.8$-positive type I HCs (**A.1**), four $K_V1.8$-positive type I HCs (**A.2**), and two $K_V1.8$-negative type I HCs from a $K_V1.8^{-/-}$ mouse (**A.3**). (**B**) *Left,* Cartoon showing cytoplasmic calretinin stain in calyx-only striolar afferents and most type II HCs, and $K_V1.8$ on membranes of both HC types. In wildtype utricles, $K_V1.8$ immunolocalized to basolateral membranes of type I and II HCs (**B.1**). $K_V1.8$ immunolocalized to type I HCs in striola (**B.2**). Staining of supporting cell (SC) membranes by Kv1.8 antibody was non-specific, as it was present in $K_V1.8^{-/-}$ tissue (**B.3, B.4**).

calretinin-positive calyceal cup (Fig. 5A.2, 5B.3, rightmost cells). Hair cell identification was confirmed with established morphological indicators: for example, type II HCs tend to have basolateral processes (feet) (Pujol et al., 2014) and, in the extrastriola, more apical nuclei than type I HC.

Previously, Carlisle et al. (2012) reported $K_V1.8$-like immunoreactivity in many cell types in the inner ear. In contrast, Lee et al. (2013) found that gene expression reporters indicated expression only in hair cells and some supporting cells. Here, comparison of control and null tissue showed selective expression of HC membranes, and that some supporting cell staining is non-specific.

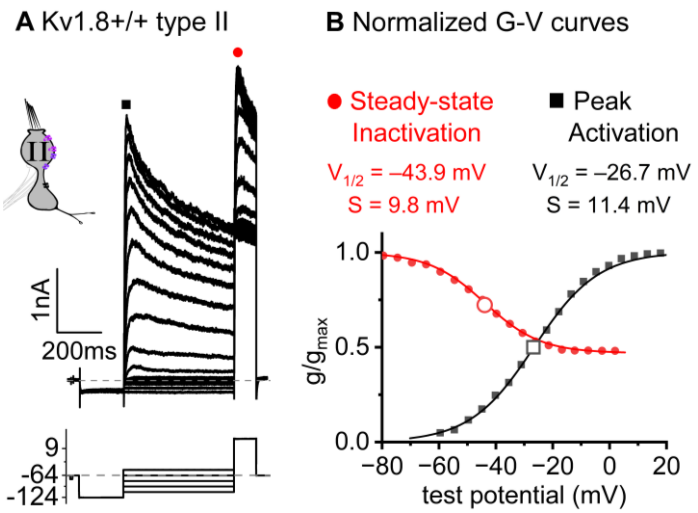


Figure 6. Inactivation curve of g_A in extrastriolar type II HCs.
(**A**) Modified voltage protocol measured accumulated steady-state inactivation in the tail potential. 100 µM ZD7288 in bath prevented contamination by HCN current. (**B**) Voltage dependence of g_A's steady-state inactivation (h_∞ curve) and peak activation are consistent with $K_V1.4$ heteromers. H_∞ curves have overlapped normalized activation (peak conductance, black squares) and inactivation (tail conductance, red circles) G-V data points. *Curves,* Boltzmann fits (Eq. 1). *Average fit parameters* from $K_V1.8^{+/+,+/-}$ type II HCs, P40-P210, median P94. Inactivation: V_{half}, –42 ±2 mV (n=11); S, 11 ± 1 mV. Activation: V_{half}, –23 ± 1 mV (n=11); S, 11.2 ± 0.4 mV.

$K_V1.4$ may also contribute to g_A

Results with the $K_V1.8$ knockout suggest that type II hair cells have an inactivating K_V1 conductance that includes $K_V1.8$ subunits. $K_V1.8$, like most K_V1 subunits, does not show fast inactivation as a heterologously expressed homomer (Lang et al., 2000; Ranjan et al., 2019; Dierich et al., 2020), nor do the $K_V1.8$-dependent channels in type I HCs, as we show, and in cochlear inner hair cells (Dierich et al., 2020). K_V1 subunits without intrinsic inactivation can produce rapidly inactivating currents by associating with $K_V\beta1$ (KCNB1) or $K_V\beta3$ subunits. $K_V\beta1$ is present in type II HCs alongside $K_V\beta2$ (McInturff et al., 2018; Jan et al., 2021; Orvis et al., 2021), which does not confer rapid inactivation (Dwenger et al., 2022).

Another possibility is that in type II HCs, $K_V1.8$ subunits heteromultimerize with $K_V1.4$ subunits—the only K_V1 subunits which, when expressed as a homomer, have complete N-type (fast) inactivation (Stühmer et al., 1989). Multiple observations support this possibility. $K_V1.4$ has been linked to g_A in pigeon type II HCs (Correia et al., 2008) and is the second-most abundant K_V1 transcript in mammalian vestibular HCs, after $K_V1.8$ (Scheffer et al., 2015). $K_V1.4$ is expressed in type II HCs but not type I HCs (McInturff et al., 2018; Orvis et al., 2021), and is not found in striolar HCs (Jan et al., 2021; Orvis et al., 2021), where even in type II HCs, inactivation is slower and less extensive (Fig. 3A).

Functional heteromers form between $K_V1.4$ and other $K_V1.x$ and/or $K_V\beta1$ (Imbrici et al., 2006; Correia et al., 2008; Al-Sabi et al., 2011). Although $K_V1.4$ and $K_V1.8$ heteromers have not been studied directly, g_A's inactivation time course ($\tau_{Fast,Inac}$ of ~30 ms +30 mV, Fig. 3A) and voltage dependence (V_{half} –41 mV, Fig. 6B) are consistent with these other $K_V1.4$-containing heteromers.

K_V7 channels contribute a small delayed rectifier in type I and type II hair cells

In $K_V1.8^{-/-}$ HCs, absence of $I_{K,L}$ and I_A revealed smaller delayed rectifier K^+ currents that, unlike $I_{K,L}$, activated positive to resting potential and, unlike I_A, lacked fast inactivation. Candidate channels include members of the K_V7 (KCNQ, M-current) family, which have been identified previously in rodent vestibular HCs (Kharkovets et al., 2000; Rennie et al., 2001; Hurley et al., 2006; Scheffer et al., 2015).

We test for K_V7 contributions in $K_V1.8^{-/-}$ type I HCs, $K_V1.8^{-/-}$ type II HCs, and $K_V1.8^{+/+,+/-}$ type II HCs of multiple ages by applying XE991 at 10 µM (Fig. 7A), a dose selective for K_V7 channels (Brown et al., 2002) and close to the IC_{50} (Alexander et al., 2019). In $K_V1.8^{-/-}$ HCs of both types, 10 µM XE991 blocked about

286 half of the residual K_V conductance (Fig. 7B.1), consistent with K_V7 channels conducting most or all of the
287 non-$K_V1.8$ delayed rectifier current. In all tested HCs (P8-355, median P224), the XE991-sensitive
288 conductance did not inactivate substantially within 200 ms at any voltage, consistent with $K_V7.2$, 7.3, 7.4,
289 and 7.5 currents (Wang, 1998; Kubisch et al., 1999; Schroeder et al., 2000; Jensen et al., 2007; Xu et al.,
290 2007). We refer to this component as $g_{DR}(K_v7)$. The voltage dependence and Gmax density (G_{max}/C_m) of
291 $g_{DR}(K_v7)$ were comparable across HC types and genotypes (Figure 7B.2-4). Although $K_V7.4$ was not
292 detectable in HCs during immunostaining (Fig. 5), $K_V7.4$ has been shown in type I HCs with immunogold
293 labeling (Kharkovets et al., 2000; Hurley et al., 2006).

294 These results are consistent with similar K_V7 channels contributing a relatively small delayed rectifier
295 in both HC types. In addition, the similarity of XE991-sensitive currents of $K_V1.8^{+/+}$ and $K_V1.8^{-/-}$ type II HCs
296 indicates that knocking out $K_V1.8$ did not cause general effects on ion channel expression. We did not test
297 XE991 on $K_V1.8^{+/+,+/-}$ type I HCs because $g_{K,L}$ runs down in ruptured patch recordings (Rüsch and Eatock,
298 1996a; Chen and Eatock, 2000; Hurley et al., 2006), which could contaminate the XE991-sensitive
299 conductance obtained by subtraction.

300 In one striolar $K_V1.8^{-/-}$ type I HC, XE991 also blocked a small conductance that activated negative to
301 rest (Suppl. Fig. 5A-B). This conductance (V_{half} ~= –100 mV, Suppl. Fig. 5C) was detected only in $K_V1.8^{-/-}$
302 type I HCs from the striola (5/23 vs. 0/45 extrastriolar). The V_{half} and $\tau_{deactivation}$ were similar to values
303 reported for $K_V7.4$ channels in cochlear HCs (Wong et al., 2004; Dierich et al., 2020). This very negatively
304 activating K_V7 conductance coexisted with the larger more positively activating K_V7 conductance (Suppl.
305 Fig. 5C) and was too small (<0.5 nS/pF) to contribute significantly to $g_{K,L}$ (~10-100 nS/pF, Fig. 1D).

306

307 Other channels

308 While our data are consistent with $K_V1.8$- and K_V7- containing channels carrying most of the outward-
309 rectifying current in mouse utricular hair cells, there is evidence in other preparations for additional
310 channels, including K_V11 (KCNH, Erg) channels in rat utricular type I hair cells (Hurley et al., 2006) and BK
311 (KCNMA1) channels in rat utricle and rat and turtle semicircular canal hair cells (Schweizer et al., 2009;
312 Contini et al., 2020).

313 BK is expressed in mouse utricular hair cells (McInturff et al., 2018; Jan et al., 2021; Orvis et al., 2021).
314 However, Ca^{2+}-dependent currents have not been observed in mouse utricular HCs, and we found little to
315 no effect of the BK-channel blocker iberiotoxin at a dose (100 nM) well beyond the IC_{50}: percent blocked
316 at –30 mV was 2 ± 6% (3 $K_V1.8^{-/-}$ type I HCs); 1 ± 5% (5 $K_V1.8^{+/+,+/-}$ type II HCs); 7% and 14% (2 $K_V1.8^{-/-}$ type
317 II HCs). We also did not see N-shaped I-V curves typical of many Ca^{2+}-dependent K^+ currents. In our
318 ruptured-patch recordings, Ca^{2+}-dependent BK currents and erg channels may have been eliminated by
319 wash-out of the hair cells' small Ca_V currents (Bao et al., 2003) or cytoplasmic second messengers (Hurley
320 et al., 2006).

321 To check whether the constitutive $K_V1.8$ knockout has strong non-specific effects on channel
322 trafficking, we examined the summed HCN and fast inward rectifier currents (I_H and I_{Kir}) at –124 mV, and
323 found them similar across genotypes (Suppl. Fig. 6). The $g_{K,L}$ knockout allowed identification of zonal
324 differences in I_H and I_{Kir} in type I HCs, previously examined in type II HCs (Masetto and Correia, 1997; Levin
325 and Holt, 2012). In type I HCs from both control and null utricles, I_H and I_{Kir} were less prevalent in striola
326 than extrastriola, and, when present, the combined inward current was smaller (Suppl. Fig. 6B).

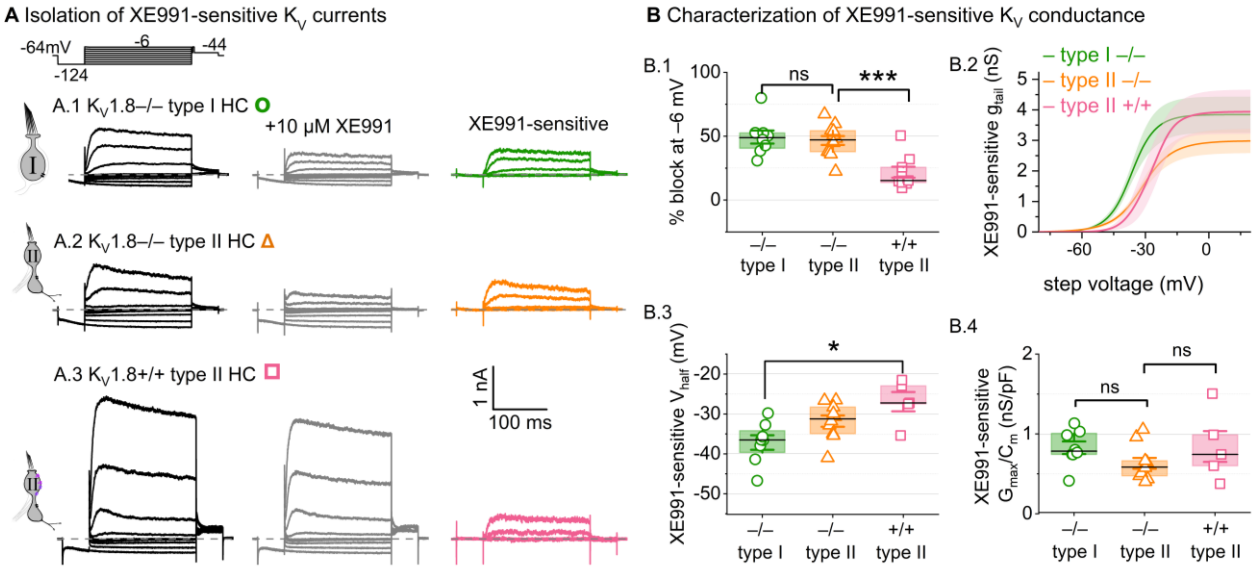


Figure 7. A K_V7-selective blocker, XE991, reduced residual delayed rectifier currents in $K_V1.8^{-/-}$ type I and II HCs.
(A) XE991 (10 µM) partly blocked similar delayed rectifier currents in type I and type II $K_V1.8^{-/-}$ HCs and a type II $K_V1.8^{+/+}$ HC.
(B) Properties of XE991-sensitive conductance, $g_{DR}(K_V7)$. (**B.1**) % Block of steady-state current. (**B.2**) tail G-V curves for $K_V1.8^{-/-}$ type I HCs (n=8), $K_V1.8^{-/-}$ type II HCs (9), and $K_V1.8^{+/+}$ type II HCs (5); mean ± SEM. (**B.3**) V_{half} was less negative in $K_V1.8^{+/+}$ type II than $K_V1.8^{-/-}$ type I HC (p = 0.01, KWA). (**B.4**) Conductance density was similar in all groups (ANOVA, non-significant at 40% power (*left*), 20% power (*right*).

Discussion

We have shown that constitutive knockout of $K_V1.8$ eliminated $g_{K,L}$ in type I HCs, and g_A and much of g_{DR} in type II HCs. $K_V1.8$ immunolocalized specifically to the basolateral membranes of type I and II HCs. We conclude that $K_V1.8$ is a pore-forming subunit of $g_{K,L}$, g_A, and part of g_{DR} [$g_{DR}(K_V1.8)$]. We suggest that fast inactivation of g_A may arise from heteromultimerization of non-inactivating $K_V1.8$ subunits and inactivating $K_V1.4$ subunits. Finally, we showed that a substantial component of the residual delayed rectifier current in both type I and type II HCs comprises K_V7 channels.

$K_V1.8$ is expressed in hair cells from mammalian cochlea (Dierich et al., 2020), avian utricle (Scheibinger et al., 2022), and zebrafish (Erickson and Nicolson, 2015). Our work suggests that in anamniotes, which lack type I cells and $g_{K,L}$, $K_V1.8$ contributes to g_A and g_{DR}, which are widespread in vertebrate HCs (reviewed in Meredith and Rennie, 2016). $K_V1.8$ expression has not been detected in rodent brain but is reported in the pacemaker nucleus of weakly electric fish (Smith et al., 2018).

$K_V1.8$ subunits may form homomultimers to produce $g_{K,L}$ in type I hair cells

Recent single-cell expression studies on mouse utricles (McInturff et al., 2018; Jan et al., 2021; Orvis et al., 2021) have detected just one K_V1 subunit, $K_V1.8$, in mouse type I HCs. Given that $K_V1.8$ can only form multimers with K_V1 family members, and given that $g_{K,L}$ channels are present at very high density (~150

per μm^2 in rat type I, Chen and Eatock, 2000), it stands to reason that most or all of the channels are $K_V1.8$ homomers. Other evidence is consistent with this proposal. $g_{K,L}$ (Rüsch and Eatock, 1996a) and heterologously expressed $K_V1.8$ homomers in oocytes (Lang et al., 2000) are non-inactivating and blocked by millimolar Ba^{2+} and 4-aminopyridine and >10 mM TEA. Unlike channels with $K_V1.1$, $K_V1.2$, and $K_V1.6$ subunits, $g_{K,L}$ is not sensitive to 10 nM α-dendrotoxin (Rüsch and Eatock, 1996a). $g_{K,L}$ and heterologously expressed $K_V1.8$ channels have similar single-channel conductances (~20 pS for $g_{K,L}$ at positive potentials, Chen and Eatock, 2000; 11 pS in oocytes, Lang et al., 2000). $g_{K,L}$ is inhibited—or positively voltage-shifted—by cGMP (Behrend et al., 1997; Chen and Eatock, 2000), presumably via the C-terminal cyclic nucleotide binding domain of $K_V1.8$.

A major novel property of $g_{K,L}$ is that it activates 30-60 mV negative to type II $K_V1.8$ conductances and most other low-voltage-activated K_V channels (Ranjan et al., 2019). The very negative activation range is a striking difference between $g_{K,L}$ and known homomeric $K_V1.8$ channels. Heterologously expressed homomeric $K_V1.8$ channels have an activation V_{half} of –10 to 0 mV (*X. laevis* oocytes, Lang et al., 2000; CHO cells, Dierich et al., 2020). In cochlear inner HCs, currents attributed to $K_V1.8$ (by subtraction of other candidates) have a near-zero activation V_{half} (–4 mV, Dierich et al., 2020).

Possible factors in the unusually negative voltage dependence of $g_{K,L}$ include:

1) elevation of extracellular K^+ by the enveloping calyceal terminal, unique to type I HCs (Lim et al., 2011; Contini et al., 2012; Spaiardi et al., 2020; Govindaraju et al., 2023). High K^+ increases conductance though $g_{K,L}$ channels (Contini et al., 2020), perhaps through K^+-mediated relief of C-type inactivation (López-Barneo et al., 1993; Baukrowitz and Yellen, 1995). We note, however, that $g_{K,L}$ is open at rest even in neonatal mouse utricles cultured without innervation (Rüsch et al., 1998) and persists in dissociated type I HCs (Chen and Eatock, 2000; Hurley et al., 2006).

2) The high density of $g_{K,L}$ (~50 nS/pF in striolar $K_V1.8^{+/+}$ HCs) implies close packing of channels, possibly represented by particles (12-14 nm) seen in freeze-fracture electron microscopy of the type I HC membrane (Gulley and Bagger-Sjöbäck, 1979; Sousa et al., 2009). Such close channel packing might hyperpolarize *in situ* voltage dependence of $g_{K,L}$, as proposed for $K_V7.4$ channels in outer hair cells (Perez-Flores et al., 2020). Type I HC-specific partners that may facilitate this close packing include ADAM11 (McInturff et al., 2018), which clusters presynaptic $K_V1.1$ and $K_V1.2$ to enable ephaptic coupling at a cerebellar synapse (Kole et al., 2015).

3) Modulation by accessory subunits. Type I HCs express $K_V\beta1$ (McInturff et al., 2018; Orvis et al., 2021), an accessory subunit that can confer fast inactivation and hyperpolarize activation V_{half} by ~10 mV. $K_V\beta1$ might interact with $K_V1.8$ to shift voltage dependence negatively. Arguments against this possibility include that $g_{K,L}$ lacks fast inactivation (Rüsch and Eatock, 1996a; Hurley et al., 2006; Spaiardi et al., 2017) and that cochlear inner hair cells co-express $K_V1.8$ and $K_V\beta1$ (Liu et al., 2018) but their $K_V1.8$ conductance has a near-0 V_{half} (Dierich et al., 2020).

$K_V1.8$ subunits may combine with different subunits to produce g_A and $K_V1.8$-dependent g_{DR} in type II HCs

The $K_V1.8$-dependent conductances of type II HCs vary in their fast and slow inactivation. In not showing fast inactivation (Lang et al., 2000; Ranjan et al., 2019; Dierich et al., 2020), heterologously expressed $K_V1.8$ subunits resemble most other K_V1 family subunits, with the exception of $K_V1.4$ (for comprehensive review, see Ranjan et al., 2019). $K_V1.4$ is a good candidate to provide fast inactivation based on immunolocalization and voltage dependence (Figs. 4, 6). We suggest that g_A and $g_{DR}(K_V1.8)$ are $K_V1.8$-

containing channels that may include a variable number of $K_V1.4$ subunits and $K_V\beta2$ and $K_V\beta1$ accessory subunits.

$K_V1.4$-$K_V1.8$ heteromeric assembly could account for several related observations. The faster $\tau_{Inact,Fast}$ in $K_V1.8^{+/-}$ relative to $K_V1.8^{+/+}$ type II HCs (Fig. 3A.3, Suppl. Fig. 2A.1) could reflect an increased ratio of $K_V1.4$ to $K_V1.8$ subunits and therefore more N-terminal inactivation domains per heteromeric channel. Zonal variation in the extent and speed of N-type inactivation (Fig. 3A) might arise from differential expression of $K_V1.4$. The small fast-inactivating conductance in ~20% of extrastriolar $K_V1.8^{-/-}$ type II HCs (Suppl. Fig. 4) might flow through $K_V1.4$ homomers.

Fast inactivation may also receive contribution from $K_V\beta$ subunits. $K_V\beta1$ is expressed in type II HCs (McInturff et al., 2018; Jan et al., 2021; Orvis et al., 2021), and, together with $K_V1.4$, has been linked to g_A in pigeon vestibular HCs (Correia et al., 2008). $K_V\beta2$, also expressed in type II HCs (McInturff et al., 2018; Orvis et al., 2021), does not confer fast inactivation.

We speculate that g_A and $g_{DR}(K_V1.8)$ have different subunit composition: g_A may include heteromers of $K_V1.8$ with other subunits that confer rapid inactivation, while $g_{DR}(K_V1.8)$ may comprise homomeric $K_V1.8$ channels, given that they do not have N-type inactivation.

$K_V1.8$ relevance for vestibular function

In both type I and type II utricular HCs, $K_V1.8$-dependent channels strongly shape receptor potentials in ways that promote temporal fidelity rather than electrical tuning (Lewis, 1988), consistent with the utricle's role in driving reflexes that compensate for head motions as they occur. This effect is especially pronounced for type I HCs, where the current-step evoked voltage response reproduces the input with great speed and linearity (Fig. 2).

$g_{K,L}$ dominates passive membrane properties in mature $K_V1.8^{+/+,+/-}$ type I HCs such that $K_V1.8^{-/-}$ type I HCs are expected to have receptor potentials with higher amplitudes but lower low-pass corner frequencies, closer to those of type II HCs and immature HCs of all types (Correia et al., 1996; Rüsch and Eatock, 1996a; Songer and Eatock, 2013). In $K_V1.8^{-/-}$ epithelia, we expect the lack of a large basolateral conductance open at rest to reduce the speed and gain of non-quantal transmission, which depends on K^+ ion efflux from the type I HC to change electrical and K^+ potentials in the synaptic cleft (Govindaraju et al., 2023). In hair cells, K^+ enters the mechanosensitive channels of the hair bundle from the K^+-rich apical endolymph and exits through basolateral potassium conductances into the more conventional low-K+ perilymph. For the type I-calyx synapse, having in the hair cell a large, non-inactivating K^+ conductance open across the physiological range of potentials avoids channel gating time and allows for instantaneous changes in current into the cleft and fast afferent signaling (Pastras et al., 2023).

In contrast, mature type II HCs face smaller synaptic contacts and have $K_V1.8$-dependent currents that are not substantially activated at resting potential. They do affect the time course and gain of type II HC responses to input currents, speeding up depolarizing transients, producing a repolarizing rebound during the step, and reducing resonance.

Type I and II vestibular hair cells are closely related, such that adult type II HCs acquire type I-like properties upon deletion of the transcription factor Sox2 (Stone et al., 2021). In normal development of the two cell types, the Kcna10 gene generates biophysically distinct and functionally different ion channels, presenting a natural experiment in functional differentiation of sensory receptor cells.

Materials and methods

Preparation

All procedures for handling animals followed the NIH Guide for the Care and Use of Laboratory Animals and were approved by the Institutional Animal Care and Use Committees of the University of Chicago and the University of Illinois Chicago. Most mice belonged to a transgenic line with a knockout allele of *Kcna10* (referred to here as $K_V1.8^{-/-}$). Our breeding colony was established with a generous gift of such animals from Sherry M. Jones and Thomas Friedman. These animals are described in their paper (Lee et al., 2013). Briefly, the Texas A&M Institute for Genomic Medicine generated the line on a C57BL/6;129SvEv mixed background by replacing Exon 3 of the *Kcna10* gene with an IRES-bGeo/Purocassette. Mice in our colony were raised on a 12:12h light-dark cycle with access to food and water *ad libitum*.

Semi-intact utricles were prepared from ~150 male and ~120 female mice, postnatal days (P) 5-375, for same-day recording. Hair cell K_V channel data were pooled across sexes as most results did not appear to differ by sex; an exception was that $g_{K,L}$ had a more negative V_{half} in males (Suppl. Table 1), an effect not clearly related to age, copy number, or other properties of the activation curve.

Preparation, stimulation, and recording methods followed our previously described methods for the mouse utricle (Vollrath and Eatock, 2003). Mice were anesthetized through isoflurane inhalation. After decapitation, each hemisphere was bathed in ice-cold, oxygenated Liebowitz-15 (L15) media. The temporal bone was removed, the labyrinth was cut to isolate the utricle, and the nerve was cut close to the utricle. The utricle was treated with proteinase XXIV (100 μg/mL, ~10 mins, 22°C) to facilitate removal of the otoconia and attached gel layer and mounted beneath two glass rods affixed at one end to a coverslip.

Electrophysiology

We used the HEKA Multiclamp EPC10 with Patchmaster acquisition software, filtered by the integrated HEKA filters: a 6-pole Bessel filter at 10 kHz and a second 4-pole Bessel filter at 5 kHz, and sampled at 10-100 kHz. Recording electrodes were pulled (PC-100, Narishige) from soda lime glass (King's Precision Glass R-6) wrapped in paraffin to reduce pipette capacitance. Internal solution contained (in mM) 135 KCl, 0.5 $MgCl_2$, 3 MgATP, 5 HEPES, 5 EGTA, 0.1 $CaCl_2$, 0.1 Na-cAMP, 0.1 LiGTP, 5 $Na_2CreatinePO_4$ adjusted to pH 7.25 and ~280 mmol/kg by adding ~30 mM KOH. External solution was Liebowitz-15 media supplemented with 10 mM HEPES (pH 7.40, 310 ± 10 mmol/kg). Recording temperature was 22-25°C. Pipette capacitance and membrane capacitance transients were subtracted during recordings with Patchmaster software. Series resistance (8-12 MΩ) was measured after rupture and compensated 60-80% with the amplifier, to final values of ~2 MΩ. Potentials are corrected for remaining (uncompensated) series resistance and liquid junction potential of ~+4 mV, calculated with LJPCalc software (Marino et al., 2014).

$K_V1.8^{-/-}$ hair cells appeared healthy in that cells had resting potentials negative to –50 mV, cells lasted a long time (20-30 minutes) in ruptured patch recordings, membranes were not fragile, and extensive blebbing was not seen. Type I HCs with $g_{K,L}$ were transiently hyperpolarized to –90 mV to close $g_{K,L}$ enough to increase R_{input} above 100 MΩ, as needed to estimate series resistance and cell capacitance. The average resting potential, V_{rest}, was –87 mV ± 1 (41), similar to the calculated E_K of –86.1 mV, which is not surprising given the large K^+ conductance of these cells. V_{rest} is likely more positive *in vivo*, where lower endolymphatic Ca^{2+} increases standing inward current through MET channels.

Voltage protocols to characterize K_V currents differed slightly for type I and II HCs. In standard protocols, the cell is held at a voltage near resting potential (–74 mV in type I and –64 mV in type II), then jumped to –124 mV for 200 ms in type I HCs in order to fully deactivate $g_{K,L}$ and 50 ms in type II HCs in order to remove baseline inactivation of g_A. The subsequent iterated step depolarizations lasted 500 ms in type I HCs because $g_{K,L}$ activates slowly (Wong et al., 2004) and 200 ms in type II HCs, where K_V conductances activate faster. The 50-ms tail voltage was near the reversal potential of HCN channels (–44 mV in mouse utricular hair cells, Rüsch et al., 1998) to avoid HCN current contamination.

G-V (activation) parameters for control type I cells may be expected to vary across experiments on semi-intact (as here), organotypically cultured and denervated (Rüsch et al.,1998), or dissociated-cell preparations, reflecting variation in retention of the calyx (Discussion) and voltage step durations (Wong et al., 2004) which elevate K^+ concentration around the hair cell. Nevertheless, the values we obtained for type I and type II HCs resemble values recorded elsewhere, including experiments in which extra care was taken to avoid extracellular K^+ accumulation (Spaiardi et al., 2017, 2020). The effects of K^+ accumulation on $g_{K,L}$'s steady-state activation curves are small because the operating range is centered on E_K and can be characterized with relatively small currents (Fig. 1A).

Pharmacology

Drug-containing solutions were locally with BASI Bee Hive syringes at a final flow rate of 20 µL/min and a dead time of ~30 s. Global bath perfusion was paused during drug perfusion and recording, and only one cell was used per utricle. Aliquots of test agents in solution were prepared, stored at –20°C, and thawed and added to external solution on the recording day (see Key Resources Table).

Analysis

Data analysis was performed with software from OriginLab (Northampton, MA) and custom MATLAB scripts using MATLAB fitting algorithms.

Fitting voltage dependence and time course of conductances

G-V curves. Current was converted to conductance (G) by dividing by driving force ($V - E_K$; E_K calculated from solutions). For type I HCs, tail G-V curves were generated from current 1 ms after the end of the iterated voltage test step. For type II HCs, peak G-V curves were generated from peak current during the step and steady-state G-V curves were generated from current 1 ms before the end of a 200 ms step. Sigmoidal voltage dependence of G-V curves was fit with the first-order Boltzmann equation (Eq. 1).

$$G(V) = G_{min} + \frac{G_{max}}{1 + e^{\frac{V_{half} - V}{S}}} \qquad (1)$$

V_{half} is the midpoint and S is the slope factor, inversely related to curve steepness near activation threshold.

Activation time course of type II HCs. For type II HCs lacking fast inactivation, outward current activation was fit with Eq. 2.

$$I(t) = I_{SS} * \left(1 - e^{-\frac{t}{\tau_w}}\right)^n + I_0 \qquad (2)$$

I_{SS} is steady-state current, τ_w is activation time constant, n is the state factor related to the number of closed states (typically constrained to 3), and I_o is baseline current.

To measure activation and inactivation time course of g_A, we used Eq. 3 to fit outward K^+ currents evoked by steps from –125 mV to above –50 mV (Rothman and Manis, 2003b).

$$I(t) = I_{max} * \left(1 - e^{-\frac{t}{\tau_w}}\right)^n * (1 - Z * (f * \left(1 - e^{-\frac{t}{\tau_{zf}}}\right) + (1 - f) * \left(1 - e^{-\frac{t}{\tau_{zs}}}\right)) + I_0 \quad (3)$$

Z is total steady-state inactivation ($0 \leq Z < 1$ means incomplete inactivation, which allows the equation to fit non-inactivating delayed rectifier currents), f is the fraction of fast inactivation relative to total inactivation, I_{max} is maximal current, τ_{zf} and τ_{zs} are the fast and slow inactivation time constants. We chose to compare fit parameters at 30 ± 2 mV (91), where fast and slow inactivation were consistently separable and g_A was maximized. In most $K_V1.8^{-/-}$ and some striolar $K_V1.8^{+/+,+/-}$ cells, where fast inactivation was absent and adjusted R^2 did not improve on a single-exponential fit by >0.01, we constrained f in Eq. 3 to 0 to avoid overfitting.

For *Peak* G-V relations, peak conductance was taken from fitted curves (Eqs. 2 and 3). To construct '*Steady-state'* G-V relations, we used current at 200 ms (6 ± 1 % (94) greater than steady-state estimated from fits to Eq. 3 (Fig. 3C-D)).

Percent inactivation was calculated at 30 mV with Eq. 4 :

$$\%\ Inactivation = (I_{Peak} - I_{SS})/I_{Peak} \quad (4)$$

I_{Peak} is maximal current, and I_{SS} is current at the end of a 200 ms voltage step.

The electrical resonance of type II HCs was quantified by fitting voltage responses to current injection steps (Songer and Eatock, 2013). We fit Eq. 5, a damped sinusoid, to the voltage trace from half-maximum of the initial depolarizing peak until the end of the current step.

$$V(t) = V_{ss} + V_p * e^{-\frac{t}{\tau_e}} * \sin(2\pi f_e - \theta) \quad (5)$$

V_{SS} is steady-state voltage, V_p is the voltage of the peak response, τ_e is the decay time constant, f_e is the fundamental frequency, and θ is the phase angle shift.

Quality factor, Q_e, was calculated with Eq. 6 (Crawford and Fettiplace, 1981).

$$Q_e = [(\pi f_e \tau_e)^2 + 0.25]^{1/2} \quad (6)$$

Statistics

We give means ± SEM for normally-distributed data, and otherwise, median and range. Data normality was assessed with the Shapiro-Wilk test for n<50 and the Kolmogorov-Smirnov test for n>50. To assess homogeneity of variance we used Levene's test. With homogeneous variance, we used two-way ANOVA for genotype and zone with the posthoc Tukey's test. When variance was non-homogeneous, we used one-way Welch ANOVA with the posthoc Games-Howell test. For data that were not normally distributed, we used the non-parametric one-way Kruskal-Wallis ANOVA (KWA) with posthoc Dunn's test. Effect size is Hedge's g (g). For age dependence, we used partial correlation coefficients controlling for genotype and

547 zone. Statistical groups may have different median ages, but all have overlapping age ranges. In figures,
548 asterisks represent p-value ranges as follows: *, $p<0.05$; **, $p<0.01$; ***, $p<0.001$; ****, $p<0.0001$.
549

550 ## Immunohistochemistry

551 Mice were anesthetized with Nembutal (80 mg/kg), then perfused transcardially with 40mL of
552 physiological saline containing heparin (400 IU), followed by 2 mL/g body weight fixative (4%
553 paraformaldehyde, 1% picric acid, and 5% sucrose in 0.1 M phosphate buffer at pH 7.4, sometimes with
554 1% acrolein). Vestibular epithelia were dissected in phosphate buffer, and tissues were cryoprotected in
555 30% sucrose-phosphate buffer overnight at 4°C. Otoconia were dissolved with Cal-Ex (Fisher Scientific) for
556 10 min. Frozen sections (35 µm) were cut with a sliding microtome. Immunohistochemistry was
557 performed on free-floating sections. Tissues were first permeabilized with 4% Triton X-100 in PBS for 1 h
558 at room temperature, then incubated with 0.5% Triton X-100 in a blocking solution of 0.5% fish gelatin
559 and 1% BSA for 1 h at room temperature. Sections were incubated with 2-3 primary antibodies for 72 h
560 at 4°C and with 2-3 secondary antibodies. Sections were rinsed with PBS between and after incubations
561 and mounted on slides in Mowiol (Calbiochem).
562

Table 6. Key Resources Table

Antibody	Source	Catalog Number	Lot Number	Dilution
Rabbit anti-K_V1.8	Alomone	APC-157	0102	1:200 or 1:400
Mouse anti-K_V1.4	NeuroMab	P15385	5HK-05	1:400
Mouse IGg2a-conjugated anti-Tuj1	Covance	MMS-435P	B205808	1:300
Goat anti-calretinin	Millipore	AB1550	9669	1:600
Mouse IgG1-conjugated anti-K_V7.4	NeuroMab	2HK-65		1:200
Donkey anti-Rabbit Secondary Antibody, Alexa Fluor 594	Invitrogen	A21207	8652	1:200
Donkey anti-Goat 488 nm	Invitrogen	A21125	1920483	1:200
Goat anti-Mouse IgG1 594	Invitrogen	A11055	1869589	1:200
Chemicals and Peptides	Source	Catalog Number	Diluent	
Iberiotoxin	Alomone	STI-400	Water	
XE991 dihydrochloride	Sigma	X2254	Water	
ZD7288	Tocris	APN18035-2	Water	
Bovine serum albumin	Fisher	BP671	water	

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567 # Data Availability

568 Data used in this study are available on Dryad (https://doi.org/10.5061/dryad.37pvmcvrw).
569

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Author Contributions

HRM designed and performed experiments, analyzed data, and wrote the paper; RAE helped design experiments, analyze data, and write the paper; AL performed immunohistochemistry experiments and reviewed and edited the manuscript.

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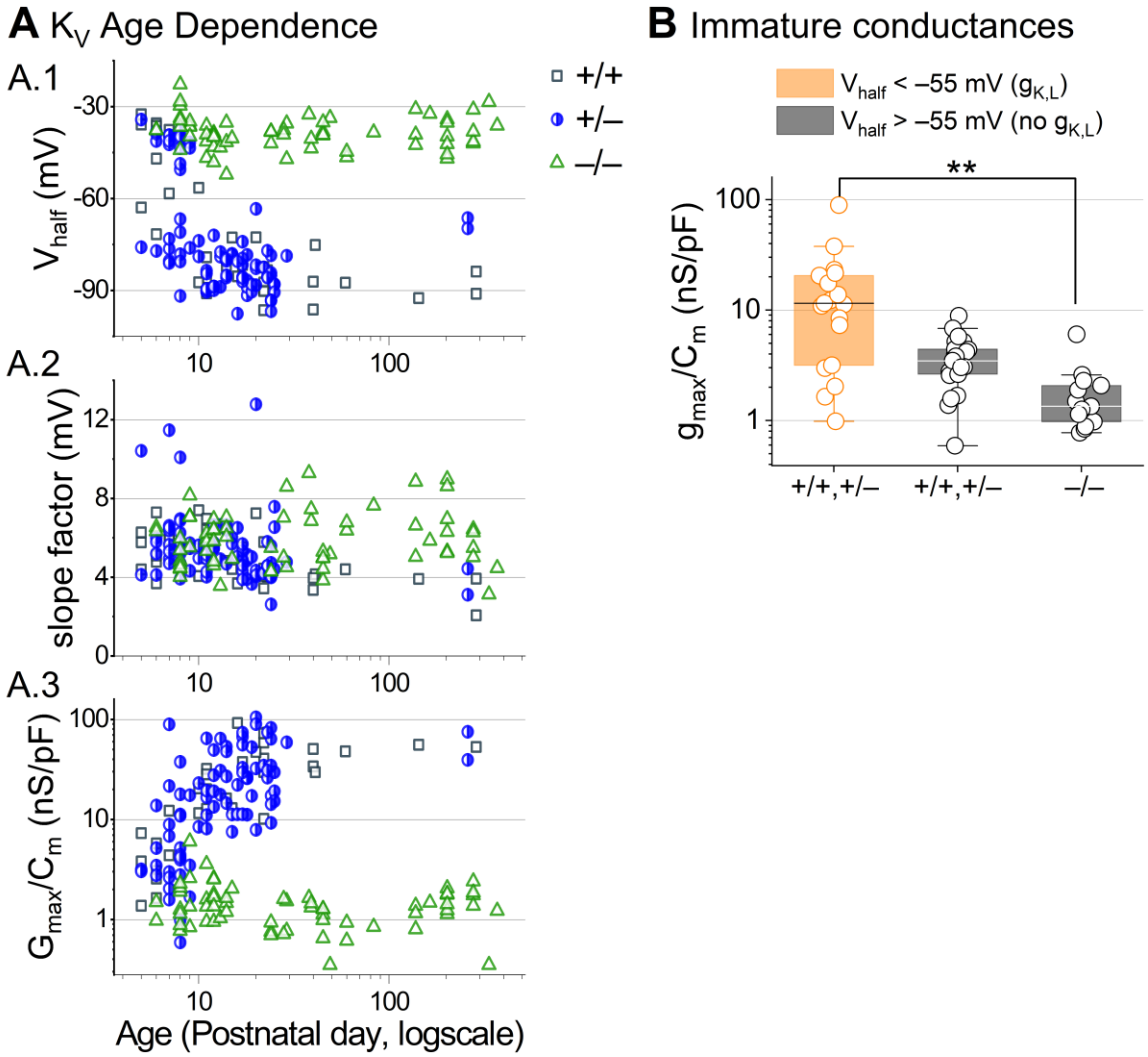
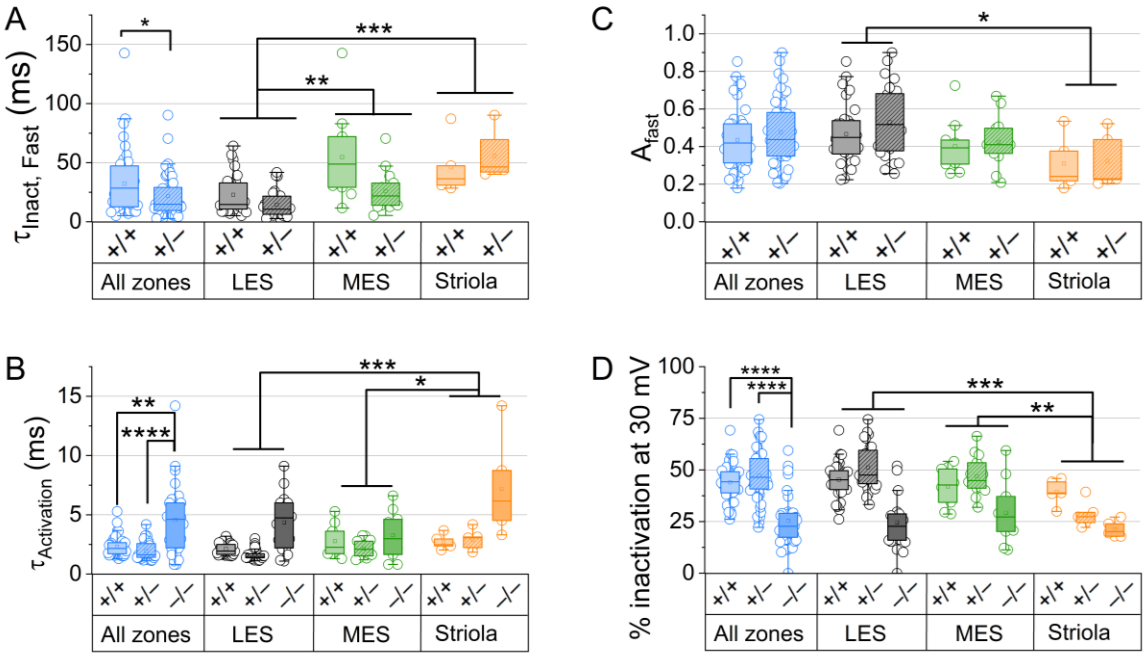
Supplemental Figures



Supplemental Figure 1. Developmental changes in type I HC K_V conductances.
(**A**) Parameters from Boltzmann fits of tail G-V relations for type I HCs plotted against age.
(**B**) Conductance density is similar in young (P5-P10) type I HCs that lack $g_{K,L}$. $g_{K,L}$ is defined here as having a V_{half} negative to –55 mV. $K_V1.8^{+/+,+/-}$ *with* $g_{K,L}$, 17 ± 5 nS/pF (19); $K_V1.8^{+/+,+/-}$ *without* $g_{K,L}$, 3.7 ± 0.4 nS/pF (22); $K_V1.8^{-/-}$, 1.8 ± 0.4 nS/pF (13). $K_V1.8^{+/+,+/-}$ *with* $g_{K,L}$ vs. $K_V1.8^{-/-}$: p = 0.007, KWA, g 1.0.



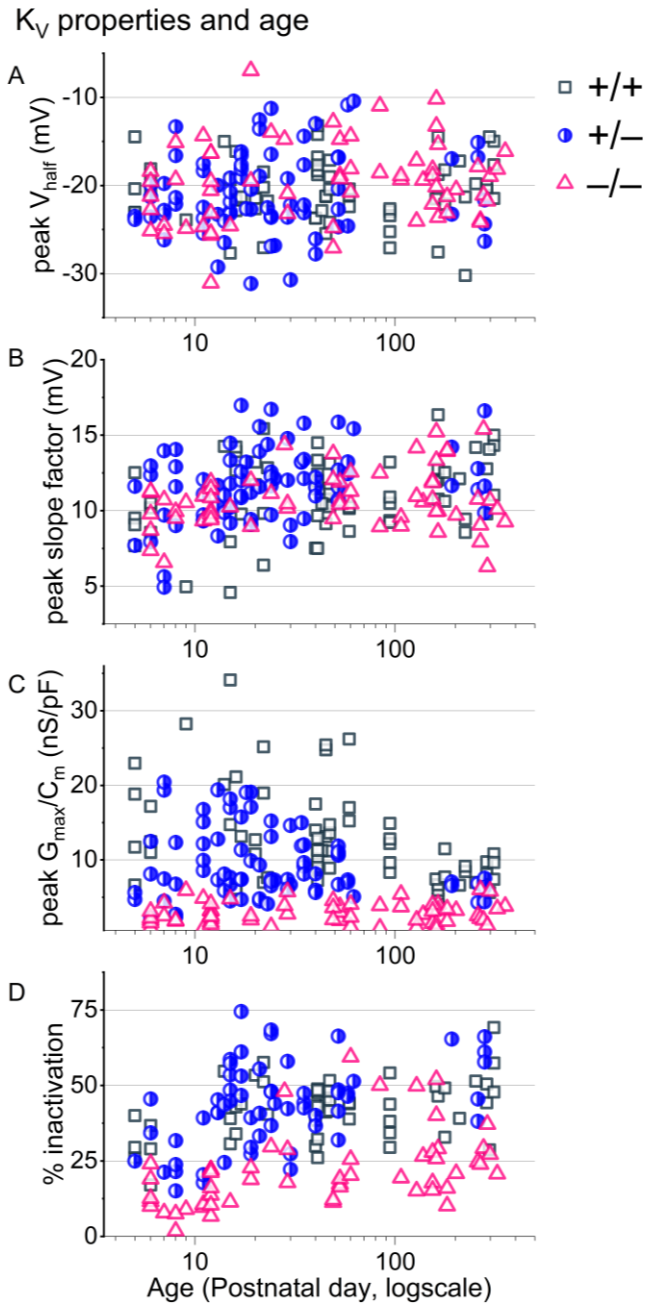
Supplemental Figure 2. **For type II HCs older than P12, K_V conductance activation and inactivation differed across zones and genotypes.**

(**A**) $\tau_{inact,Fast}$ at 30 mV was fastest in LES in $K_V1.8^{+/+}$ and $K_V1.8^{+/-}$ HCs, and faster in $K_V1.8^{+/-}$ than $K_V1.8^{+/+}$ HCs (see Table 3 for p-values).

(**B**) Fast inactivation was a larger fraction of the total in LES than striola.

(**C**) τ_{Act} at 30 mV was slower in $K_V1.8^{-/-}$ than $K_V1.8^{+/+}$ and $K_V1.8^{+/-}$, and slower in striola than LES and MES.

(**D**) Percent inactivation at 30 mV was lowest in striola (zone effect), and lowest in $K_V1.8^{-/-}$ HCs (genotype effect).



Supplemental Figure 3. For type II HCs older than P12, K_V conductances were stable. (**A-C**) Parameters from Boltzmann fits of peak G-V relations and (**D**) % inactivation at +30 mV plotted against age from all zones. Overlaid curves are smoothing cubic β-splines. Note the seven extrastriolar $K_V1.8^{-/-}$ type II HCs with % inactivation >30%.

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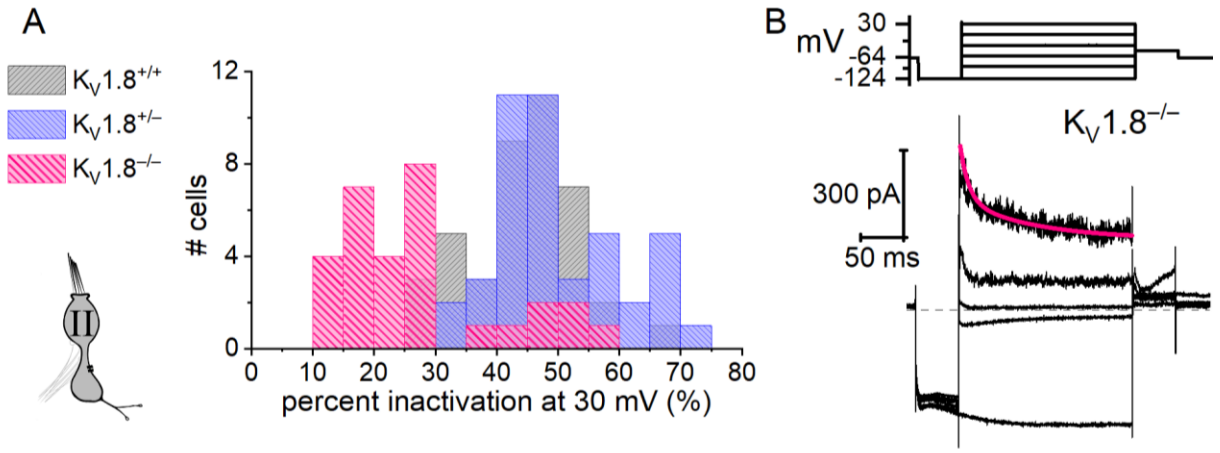

Supplemental Figure 4. A minority of extrastriolar $K_V1.8^{-/-}$ type II HCs had a very small fast-inactivating outward rectifier current.

(**A**) All extrastriolar $K_V1.8^{+/+,+/-}$ type II HCs inactivated by >30%. Most mature (>P12) extrastriolar $K_V1.8^{-/-}$ type II HCs inactivated by <30% but some inactivated by >30% (7/30, 23%) because they had fast inactivation (B).

(**B**) Exemplar residual fast inactivation ($\tau_{FastInact}$ = 10 ms at +30 mV). For the 7 cells in this group, $\tau_{FastInact}$ = 30 ± 6 ms, amplitude of fast inactivation = 310 ± 70 pA; activation peak V_{half} = –15 ± 2 mV and slope factor = 12.4 ± 0.9 mV. These parameters are similar to g_A but for the much smaller conductance (one-way ANOVAs).

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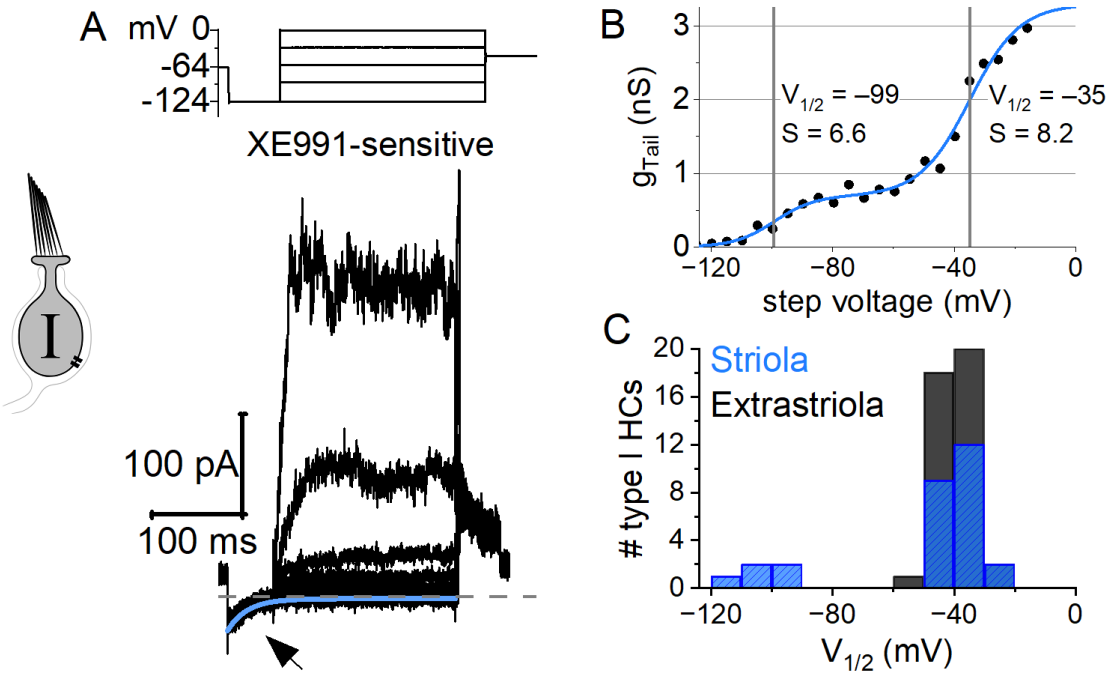

Supplemental Figure 5. A minority of striolar $K_V1.8^{-/-}$ type I HCs had a small low-voltage-activated outward rectifier current in addition to a more positively activating outward rectifier.

(**A**) Low-voltage-activated current from one cell was isolated by subtraction with 10 µM XE991 (P39), indicating that it was carried by K_V7 channels. Deactivation of XE991-sensitive current evoked by step from –64 mV to –124 mV *(arrow)* was fit with exponential decay (τ = 21 ms). (**B**) XE991-sensitive tail G-V curve of the XE991-blocked conductance (A) was fit with a sum of two Boltzmann equations: $G(V) = A_1/(1 + \exp((V_{half,1} - V)/S_1)) + A_2/(1 + \exp((V_{half,2} - V)/S_2))$.

(**C**) The low-voltage-activated $V_{half,1}$ component was only seen in striolar $K_V1.8^{-/-}$ type I HCs, and even there in the minority: 5/23; 22%; P6-P370). It was always seen together with a more positively activating outward rectifier. Boltzmann parameters, including (B): $A_1/(A_1+A_2)$ = 0.15 ± 0.04, $V_{half,1}$ = –106 ± 5 mV (n=5), S_1 = 3.8 ± 0.8 mV, $V_{half,2}$ = –41 ± 1 mV, S_2 = 7 ± 1 mV. Ages: P11, 39, 202, 202, 202. No extrastriolar type I HCs (0/45; P6-277) had a double activation tail G-V curve.

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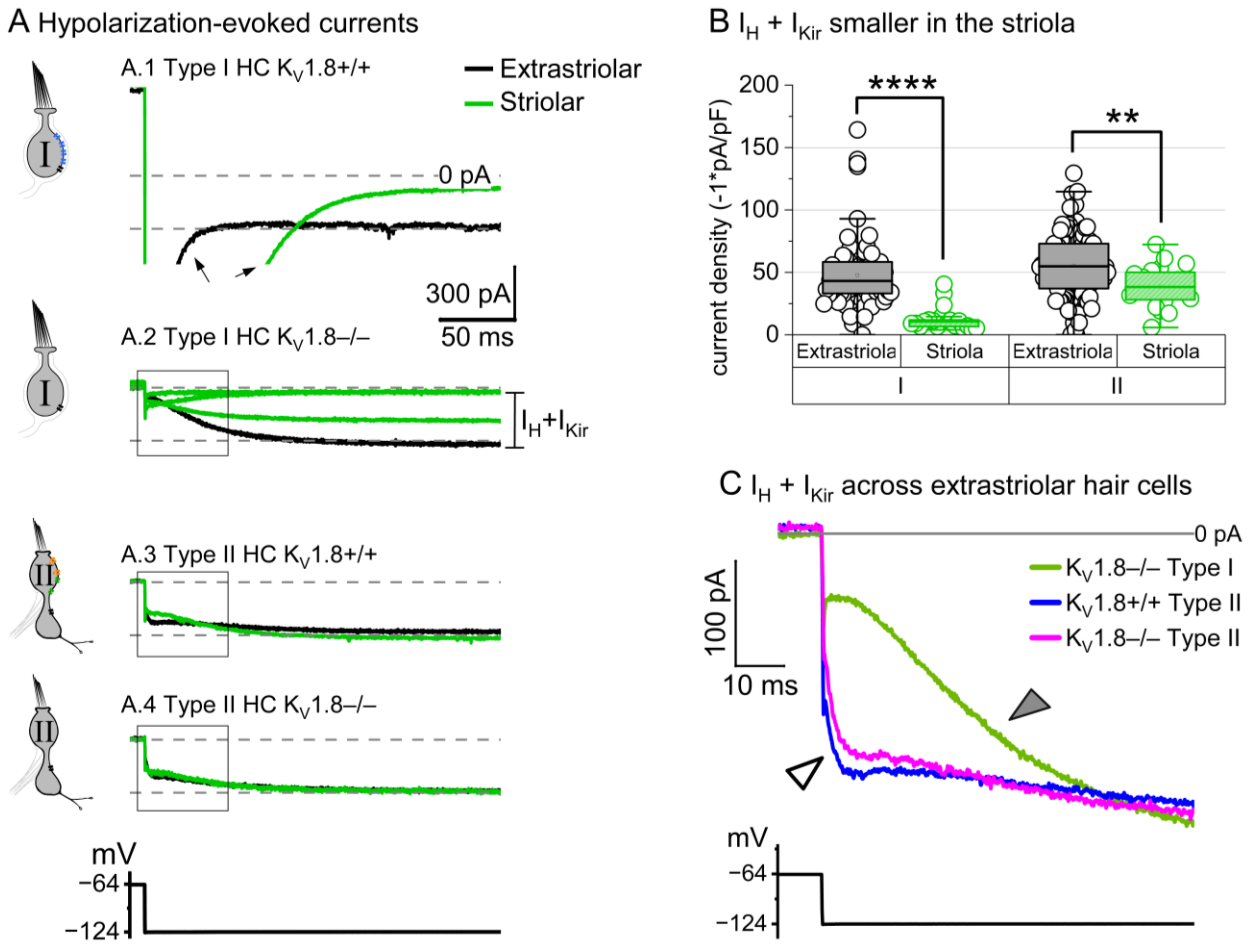

Supplemental Figure 6. No difference was detected in H (HCN) and Kir (fast inward rectifier) currents between $K_V1.8^{+/+}$ and $K_V1.8^{-/-}$ hair cells, consistent with a specific involvement of $K_V1.8$ in *Kcna10* expression.

(**A**) Hyperpolarizing voltage steps evoked I_{Kir} and I_{HCN} in $K_V1.8^{+/+,+/-,-/-}$ type I and II HCs. **A.1** *Arrows*, deactivation of $g_{K,L}$. **A.2-A.4** *Bracket,* the sum of I_H and I_{Kir} were measured as total inward current after 250 ms at –124 mV.

(**B**) Summed I_{KIR} and I_H density was the same across genotypes but smaller in striola than extrastriola (see Supplemental Table 3 for statistics).

(**C**) Inward currents seen at the onset of hyperpolarization, including I_{Kir}, were larger in type II HCs than type I. Magnification of boxed in inset from the extrastriolar cells in A.2-A.4. *Open arrowhead*, activation of fast inward rectifier, I_{Kir}; *filled arrowhead*, slower activation of I_{HCN}.

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Supplemental Table 1. *Test of sex differences in hair cell K_V channel data.*

Cell Type	Parameter	Subgroup	Male vs Female posthoc p-value	Test
Type I HC	Tail V_{half}	+/+,+/–	0.0023 ** [a]	Normal, homogeneous variance ANOVA: Genotype (2 levels), Sex (2 levels), Zone (2 levels), Genotype*Sex.
		–/–	0.57	
	Tail S	+/+,+/–	0.98	ANOVA: Genotype (2 levels), Sex (2 levels), Zone (2 levels), Genotype*Sex. Normal, homogeneous variance
		–/–	0.58	
	Tail $g_{Density}$	+/+,+/–	0.999	Normal, nonhomogeneous variance Welch ANOVA: Genotype*Sex
		–/–	0.936	
Type II HC	Peak V_{half}	+/+,+/–	0.95	Normal, homogeneous variance ANOVA: Genotype (2 levels), Sex (2 levels), Zone (2 levels), Genotype*Sex.
		–/–	0.28	
	Peak S	+/+,+/–	0.999	Normal, homogeneous variance ANOVA: Genotype (2 levels), Sex (2 levels), Zone (2 levels), Genotype*Sex.
		–/–	0.97	
	Peak $g_{Density}$	+/+,+/–	0.64	Normal, nonhomogeneous variance Welch ANOVA: Genotype*Sex
		–/–	0.43	
	% inactivation at 30 mV	+/+,+/–	0.98	Normal, homogeneous variance ANOVA: Genotype (2 levels), Sex (2 levels), Zone (2 levels), Genotype*Sex.
		–/–	0.82	

[a] g, 0.9. Male $K_V1.8^{+/+,+/-}$, –85 ± 1 mV (40) vs. Female $K_V1.8^{+/+,+/-}$, –79 ± 2 mV (12)

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Supplemental Table 2. *Soma size of type I HCs. The perimeter of cross-sections of basolateral cell bodies was manually measured from immunohistochemistry sections with sufficient autofluorescence in the 488 channel to distinguish cell morphology, and anti-calretinin to label type II hair cells and calyx-only afferents. Cells were measured from two female littermates.*

Cell Type	Genotype	Age	Weight (g)	Perimeter (µm)	Test
Type I HC	+/+	P117	24.9	47 ± 1 (19)	ANOVA: p=0.1, power = 0.07
	–/–	P117	25.2	46 ± 1 (28)	

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Supplemental Table 3. *Detected zonal but not genotype differences in hair cell I_{Kir} and I_H.*

Cell Type	Zone	I_H + I_{Kir} current density (-1*pA/pF)	$K_v1.8^{+/+,+/-}$ *vs* $K_v1.8^{-/-}$ p-value	ES *vs* Striola p-value	Test
Type I HC	ES Striola	48 ± 3 (78) 13.0 ± 2 (19)	0.3	4E-9 **** [a]	Non-normal, KWA
Type II HC	ES Striola	55 ± 2 (116) 39 ± 4 (20)	0.19 (0.25 power)	0.0058 ** [b]	Normal, homogeneous variance. 2-way ANOVA: Genotype (2 levels), Zone (2 levels)

[a] g 1.4
[b] g 0.6