The identification of extensive samples of motor units in human muscles reveals diverse effects of neuromodulatory inputs on the rate coding

Simon Avrillon[1], François Hug[2, 3], Roger Enoka[4], Arnault H. Caillet[1], Dario Farina[1]

[1] Department of Bioengineering, Faculty of Engineering, Imperial College London, London W12 7TA, UK
[2] Université Côte d'Azur, LAMHESS, Nice 06200, France
[3] The University of Queensland, School of Biomedical Sciences, St Lucia QLD 4072, QLD, Australia
[4] Department of Integrative Physiology, University of Colorado Boulder, Boulder, CO, USA

Correspondence:
Simon Avrillon
Imperial College London, London, UK
e-mail: s.avrillon@imperial.ac.uk

Dario Farina
Imperial College London, London, UK
e-mail: d.farina@imperial.ac.uk

Conflict of interests:
The authors declare no competing financial interests.

Funding resources.
Simon Avrillon is supported by the French National Research Agency through Nantes Excellence Trajectory (NExT, ANR-16-IDEX-0007). Simon Avrillon and Dario Farina are supported by the BBSRC, "Neural Commands for Fast Movements in the Primate Motor System" (BB/V00896X). François Hug is supported by the UCAJEDI Investments in the Future project managed by the French National Research Agency (ANR-15-IDEX-01) and the French National Research Agency (Neuromotor, ANR-24-CE17-5805). Dario Farina is supported by the European Research Council Synergy Grant NaturalBionicS (contract #810346) and the EPSRC Transformative Healthcare, NISNEM Technology (EP/T020970).

Abstract:

Movements are performed by motoneurons transforming synaptic inputs into an activation signal that controls muscle force. The control signal emerges from interactions between ionotropic and neuromodulatory inputs to motoneurons. Critically, these interactions vary across motoneuron pools and differ between muscles. To provide the most comprehensive framework to date of motor unit activity during isometric contractions, we identified the firing activity of extensive samples of motor units in the Tibialis Anterior (129±44 per participant; n=8) and the Vastus Lateralis (130±63 per participant; n=8) muscles during isometric contractions of up to 80% of maximal force. From this unique dataset, the rate coding of each motor unit was characterised as the relation between its instantaneous firing rate and the applied force, with the assumption that the linear increase in isometric force reflects a proportional increase in the net synaptic excitatory inputs received by the motoneuron. This relation was characterised with a natural logarithm function that comprised two stages. The initial stage was marked by a steep acceleration of firing rate, which was greater for low- than medium- and high-threshold motor units. The second stage comprised a linear increase in firing rate, which was greater for high- than medium- and low-threshold motor units. Changes in firing rate were largely non-linear during the ramp-up and ramp-down phases of the task, but with significant prolonged firing activity only evident for medium-threshold motor units. Contrary to what is usually assumed, our results demonstrate that the firing rate of each motor unit can follow a large variety of trends with force across the pool. From a neural control perspective, these findings indicate how motor unit pools use gain control to transform inputs with limited bandwidths into an intended muscle force.

Introduction:

Human muscles are versatile effectors producing forces that span several orders of magnitude. They allow humans to perform a broad range of motor tasks with the same limbs, such as a surgeon closing an incision or a climber who grasps supports on a cliff. This versatility relies on a unique structure that converts neural inputs into muscle force; that is, the motor unit, which comprises an alpha motoneuron and the muscle fibres it innervates (Sherrington, 1925; Heckman and Enoka, 2012). The nervous system controls muscle force by modulating the number of active motor units (recruitment) and their firing rates [rate coding; (Enoka and Duchateau, 2017)]. This control strategy involves projecting a substantial level of common excitatory synaptic inputs to motoneurons (Farina et al., 2014a; Farina and Negro, 2015; Negro et al., 2016a) that recruits them in a fixed order following the size principle (Henneman, 1957).

Although we have a clear understanding of the general strategies used to control muscle force, we still lack a full picture of the detailed organisation of the firing activities of motor units. One of the key challenges in the field is the ability to identify the concurrent firing activity of many motor units spanning the range of recruitment thresholds observed in human muscles (Farina and Holobar, 2016). Indeed, much of our knowledge on the modulation of motor unit activity still derives from animal preparations (Hounsgaard et al., 1988; Bennett et al., 1998b; Lee and Heckman, 2000) or from serial recordings of a few concurrent motor units in humans (Desmedt and Godaux, 1977b; Bigland-Ritchie et al., 1983; Oya et al., 2009; Kirk et al., 2016), mainly identified over narrow force ranges (Monster and Chan, 1980; Fuglevand et al., 2015; Johnson et al., 2017; Revill and Fuglevand, 2017).

Animal studies have shown two stages in the rate coding of individual motor units to increase muscle force (amplification and rate limiting) in response to linear increases in the net excitatory synaptic inputs (Bennett et al., 1998a; Lee and Heckman, 2000; Powers and Binder, 2001; Binder et al., 2020). These stages represent the different responses of motoneurons to ionotropic inputs due to modulation of their intrinsic properties by metabotropic inputs acting on the somato-dendritic surfaces of motoneurons through several ion channels (Powers and Binder, 2001; Heckman and Enoka, 2012; Binder et al., 2020). Similar characteristics in firing activity have been observed in humans (Fuglevand et al., 2015; Revill and Fuglevand, 2017; Beauchamp et al., 2023), although these observations have been limited to small samples of motor units (typically < 40) with firing activities experimentally decoded over a narrow range

of submaximal forces [typically <30% maximal voluntary contraction (MVC) force; (Fuglevand et al., 2015; Revill and Fuglevand, 2017; Beauchamp et al., 2023)]. It is difficult to infer from these studies a control scheme that is generalisable to an entire motor unit pool and to all contraction levels (De Luca, 1985).

The combination of arrays of electromyographic (EMG) electrodes with modern source-separation algorithms set the path for the identification of populations of active motor units - and their motoneurons - in humans (Farina and Holobar, 2016; Negro et al., 2016b). Recent studies have identified the concurrent firing activity of up to 60 motor units per contraction and per muscle by increasing the density of electrodes (Muceli et al., 2022; Caillet et al., 2023b). Here, we further extended this approach by tracking motor units across contractions at target forces that ranged from 10 to 80 %MVC, based on the unique spatial distribution of motor unit action potentials across the array of surface electrodes (Farina et al., 2008; Martinez-Valdes et al., 2017). We were able to identify up to ~200 unique active motor units per muscle and per participant in two human muscles *in vivo*, yielding extensive samples of motor units that are representative of the motoneuron pools (Caillet et al., 2023a). With this approach, we described the non-linear transformation of the net excitatory synaptic inputs into firing activity for individual motoneurons (Heckman and Enoka, 2012; Binder et al., 2020). We specifically focused on the non-linearities between the changes in firing rate and force: amplification, saturation, and hysteresis (Hounsgaard et al., 1988; Powers and Heckman, 2017; Binder et al., 2020). The results provided a framework picture of the rate coding of motor units during large and slow increases and decreases in an applied isometric force.

Having access to the rate coding of motoneuron pools allowed us to understand how ionotropic and neuromodulatory inputs combine to modulate force, confirming some details of hypothetical force-control schemes in humans. For example, experimental (Wei et al., 2014; Naufel et al., 2019) and computational (Powers and Heckman, 2017) studies have proposed a gain-control mechanism driven by neuromodulatory inputs to motoneurons. One common conclusion of these studies is that pools of motor units involved in fine motor tasks have a low gain, whereas additional motor units activated during more forceful tasks exhibit a progressively increase in gain (Wei et al., 2014; Powers and Heckman, 2017; Naufel et al., 2019; Binder et al., 2020). However, this conclusion is based on results from motor units in animals (Powers and Binder, 2001) and humans (Wei et al., 2014; Goodlich et al., 2022; Henderson et al., 2022) while experimentally modulating inputs, or by fitting a non-linear

model predicting muscle activation from excitatory synaptic inputs at multiple contraction levels (Naufel et al., 2019). In contrast, we addressed this question by identifying the concurrent activity of hundreds of motor units spanning most of the range of recruitment thresholds in vivo in humans.

Results:

<u>Identification and tracking of individual motor units:</u>

16 participants performed either isometric dorsiflexion (n = 8) or knee extension tasks (n = 8) while we recorded the EMG activity of the tibialis anterior (TA - dorsiflexion) or the vastus lateralis (VL – knee extension) with four arrays of 64 surface electrodes (256 electrodes per muscle). The motoneuron pools of these two muscles of the lower limb receive a large part of common input (Laine et al., 2015; Negro et al., 2016a), constraining the recruitment of their motor units in a fixed order across tasks. They are therefore good candidates for an accurate description of rate coding. Moreover, we wanted to determine whether differences in rate coding observed between proximal and distal muscles in the upper limb (De Luca et al., 1982) were also present in the lower limb.

The experimental tasks comprised isometric contractions with a ramp-up, a plateau, and a ramp-down. The ramp-up and ramp-down phases were performed slowly compared with contractile speeds observed during activities of daily living. They were performed at a constant rate of 5 %MVC.s^{-1} and the force plateau was maintained for either 10-s (70-80 %MVC), 15-s (50-60 %MVC), or 20-s (10-40 %MVC) (Figure 1A). The target during the plateau ranged from 10 to 80 %MVC in increments of 10 %MVC (randomised order).

A source-separation algorithm (Farina and Holobar, 2016; Negro et al., 2016b) was applied to the EMG signals to extract motor unit pulse trains, from which discharge times were automatically identified. All identified motor unit firings were visually inspected and manually edited when necessary (Avrillon et al., 2023). Because the signal was stationary during the plateau, it was possible to estimate reliable separation vectors for large samples of motor units with source-separation algorithms [Figure 1A; (Farina and Holobar, 2016; Negro et al., 2016b)]. The average number of motor units identified in each contraction per participant was 42 ± 24 (25th–75th percentile: 24 – 53, up to 95; Table 1) motor units from the TA and 33 ± 15 (25th–75th percentile: 23 – 47, up to 71; Table 1) motor units from the VL. The datasets from all target forces were merged into a sample of unique motor units per muscle and participant that spanned most of the operating range of recruitment thresholds observed in humans (1st–99th percentile: 0.9-73.4% MVC; see below).

167 Table 1: Mean ± standard deviation (range) for the number of motor units across the eight target
168 forces and two muscles

	10%	20%	30%	40%	50%	60%	70%	80%
TA	38±25 (1-73)	45±24 (21-83)	50±27 (24-95)	49±26 (24-93)	45±27 (15-93)	40±25 (14-80)	37±26 (14-84)	34±19 (9-60)
VL	44±17 (17-63)	46±20 (19-71)	42±20 (10-67)	34±18 (3-56)	32±15 (5-53)	26±13 (3-38)	26±14 (1-38)	20±13 (1-37)

TA, tibialis anterior, VL, vastus lateralis

The proportion of the EMG signal represented by the identified motor units was estimated by
reconstructing a synthetic EMG signal from the firing activity. To do so, the discharge times
were used as triggers to segment the differentiated EMG signals over a window of 25 ms that
yielded averaged action potential waveforms for each motor unit (Figure 1A). The action
potentials were then convolved with the discharge times to obtain trains of action potentials,
and all the trains of the identified motor units were summed to reconstruct the synthetic EMG
signal. The ratio between the powers of synthetic and experimental EMG signals was calculated
(Figure 1B): it was $69.3 \pm 17.3\%$ (25th–75th percentile: $59.3 – 83.6\%$, up to 94.2%) for TA and
$55.2 \pm 19.5\%$ (25th–75th percentile: $50.0 – 71.9\%$, up to 86.5%) for VL (Figure 1C). These
values indicate that most of the recorded surface EMG signals were successfully decomposed
into motor unit activity.

Motor units were tracked between contractions using their unique spatial distribution of action
potentials [Figure 1D; (Martinez-Valdes et al., 2017)]. This method was validated by using both
simulated EMG signals and two-source validation with simultaneous recordings of firing
activity from intramuscular and surface EMG signals (Figure S1). On average, we tracked 67.1
$\pm 10.0\%$ (25th–75th percentile: $53.9 – 80.1\%$) of the motor units between consecutive target
forces (10% increments, e.g., between 10% and 20% MVC) for TA and $57.2 \pm 5.1\%$ (25th–75th
percentile: $46.6 – 68.3\%$) of the motor units for VL (Figure S2). There are two explanations for
the inability to track all motor units across consecutive target forces: i) some motor units are
recruited at higher targets only; ii) it is challenging to track small motor units beyond a few
target forces due to a lower signal-to-noise ratio when larger motor units are recruited, or signal
cancellation (Keenan et al., 2005; Farina et al., 2014b). In total, we identified 129 ± 44 motor
units (25th–75th percentile: $100 – 164$, up to 194) per participant in TA and 130 ± 63 motor units
(25th–75th percentile: $103 – 173$, up to 199) per participant in VL.

The accuracy of the tracking method was further tested by confirming the uniqueness of each

motor unit within the identified sample, assuming that each motor unit had a unique

representation across the array of surface electrodes (Farina et al., 2008). This was

accomplished by calculating the root mean-square error (RMSE) between their action potentials

across the electrodes relative to those of the rest of the motor units (Figure 1E). The RMSE

between the action potentials of the same motor unit tracked across contractions was calculated

as a reference and compared with the 5th percentile of the distribution of RMSE between motor

units (Figure 1F). The reference value was typically less than the 5th percentile of the RMSE

calculated with the action potentials of the other motor units (TA: 5.6 ± 2.4 vs. $8.0 \pm 1.5\%$;

$p<0.001$; VL: $6.2 \pm 2.8\%$ vs. $8.5 \pm 1.6\%$; $p<0.001$). Inspection of the data for each motor unit

revealed that 92.1% (TA) and 87.0% (VL) of the motor units had a lower RMSE between target

forces compared with the other motor units (<5th percentile). Of note, motor units with the

highest reference values (>95th percentile) were considered as outliers due to tracking errors

and were excluded from the subsequent analyses (Figure 1F). We excluded 34 motor units from

TA and 28 from VL.

Figure 1. Identification of motor units in two human muscles. A. We used a blind source separation (BSS) algorithm to decompose the overlapping activity of motor units (MU) into spike trains during a force-matching trapezoidal task (red trace). B. We reconstructed synthetic EMG signals by summing the trains of action potentials from all the identified motor units and interpreting the remaining EMG signal as the part of the signal not explained by the decomposition. C. We calculated the ratio between the powers of synthetic and original EMG signals to estimate the proportion of the signal variance explained by the decomposition. Each data point indicates the average value for one participant. D. We estimated the uniqueness of each identified motor unit within the pool by calculating the root-mean-square error (RMSE) between the distributions of action potentials of the same motor unit across contractions (two panels on the left, reference value in E), and between motor units (left vs. right panels, distribution of RMSE between motor units in

yellow in E). F. Each motor unit was unique within the pool when the RMSE between its distributions of action potentials across target forces (reference value) was less than the 5th percentile of the distribution of RMSE with the rest of the motor units. Motor units considered as outliers in F (red data points) were removed from the analysis due to potential errors in tracking between contractions. Each data point is a motor unit, the box represents 25th-75th percentiles of the distribution of data, and the black line shows the median. The horizontal thick line denotes a statistical difference between reference values and 5th percentiles for each muscle.

<u>Non-linear rate coding during the ramp-up phase – amplification and rate limiting:</u>

The input-output function for each motoneuron was characterised as the relation between its instantaneous firing rate and the applied force during the ramp-up phase of the contractions (Figure 2A). The linear increase in force was assumed to reflect a proportional increase in the net synaptic excitatory inputs received by the motoneurons, as proposed previously (Fuglevand et al., 1993; Revill and Fuglevand, 2017). Thus, any deviation from a linear increase in firing rate with respect to a linear increase in force presumably reflects the influence of neuromodulatory inputs on motoneuron gain (Johnson et al., 2017; Revill and Fuglevand, 2017; Beauchamp et al., 2023), or a saturation of the motoneuron firing rate (Fuglevand et al., 2015; Revill and Fuglevand, 2017).

The association between the firing rate and the force was characterised by comparing three curve fits: i) linear - an increase in firing rate proportional to the increase in the net synaptic excitatory input, ii) rising exponential - an initial acceleration of firing rates followed by full saturation (De Luca and Contessa, 2012; Fuglevand et al., 2015; Revill and Fuglevand, 2017), and iii) natural logarithm - an initial acceleration of firing rate followed by a slower constant increase in firing rate that reflects a rate limiting effect [Figure 2A; (Bennett et al., 1998a; Lee and Heckman, 2000; Powers et al., 2012)]. Of note, this analysis was performed on each unique motor unit with the data pooled across all contractions (Figure 2A). The result was that 69.5% of the motor units in TA and 72.3% of those in VL had an input-output function better fitted by a natural logarithm (Figure 2B, inset panels).

The input-output functions of the motor units were then compared based on the distribution of recruitment thresholds (1st–99th percentile: 0.9-73.4% MVC). Motor units were clustered into three groups with equal ranges of recruitment thresholds: low- (0 to 25% MVC), medium- (25 to 50% MVC), and high- (50 to 75% MVC) threshold motor units (Figure 2B). A significant percentage of input-output functions for low threshold motor units was better fitted with a rising

exponential function (12.5% for the TA, 30.9% for the VL), highlighting a steep acceleration of firing rate followed by a full saturation. This pattern was not observed in medium- and high-threshold motor units (0% for TA and VL). In contrast, a significant percentage of input-output functions of high threshold motoneurons was better fitted with a linear function for TA (39.6%), although the input-output functions of high threshold motor units from VL were still better fitted with natural logarithms (87.9%) than linear functions (12.1%).

We further described the first stage of rate coding by estimating the acceleration of firing rate (Figure 2C; Figure S3). The initial acceleration of the firing rate, likely due to the amplification of synaptic inputs by persistent inward currents (Lee and Heckman, 2000), was calculated as the value of the first derivative of the natural logarithm function at the recruitment threshold (Figure 2C). These values were compared between motor units using a linear mixed effect model, with *Muscle* (TA; VL) and *Threshold* (low; medium; high) as fixed effects and *Participant* as a random effect. There was a significant effect of *Muscle* (F=30.4; p<0.001), *Threshold* (F=24.7; p<0.001), and a significant interaction *Muscle × Threshold* (F=4.1; p=0.017). The initial acceleration of firing rate in TA was greater for the low- (1.0±0.7 pps^2; p<0.001) and the high- (1.0±0.3 pps^2; p=0.032) threshold motor units than for medium-threshold motor units (0.6±0.3 pps^2), with no difference between the low- and high-threshold motor units (p=0.999; Figure 2C). Similarly, the initial acceleration of firing rate was greater for low-threshold motor units (0.6±0.6 pps^2) than for medium- (0.4±0.2 pps^2; p<0.001) and high-threshold motor units (0.4±0.2 pps^2; p=0.013) in VL, with no difference between the medium- and high-threshold motor units (p=0.990; Figure 2C). The initial acceleration of firing rate was greater for the low- and high-threshold motor units in TA compared with VL (p<0.001 for both).

We also tested whether the distribution of initial accelerations of firing rate observed across the entire motor unit pool was generalisable to all the participants. Participants with either fewer than 20 motor units or with no motor units recruited below 5%MVC were excluded from this analysis (TA: n=4; VL: n=3). The function representing the relation between recruitment thresholds and initial acceleration were accurately fitted using the non-linear least-square fitting method (TA; adjusted R-squared: 0.77; VL; adjusted R-squared: 0.80; green lines on Figure 2C). We then fitted the same non-linear functions for each participant; the adjusted R-squared values increased to 0.86 ± 0.04 for the TA and 0.84 ± 0.11 for the VL (inset panels in Figure 2C).

Figure 2. Non-linear rate coding of motor units. A. The relation between firing rate (pulses per second, pps) and the applied force during the ramp-up phase of the contraction was determined by concatenating the instantaneous firing rates for each motor unit (grey data points) recorded over all the contractions where it was identified, as shown here for one motor unit (coloured data points for contractions at 20, 40, and 60%MVC). The derived relations were then fitted with three different functions: linear (green), rising exponential (dark red), and natural logarithm (yellow), to characterise the input-output function of each motoneuron. B. The motor units were grouped according to their best fit. The graphs show the distribution of these groups as a function of recruitment thresholds (RT) for each muscle. The inset panels depict the

percentage of motor units (MU) in each group. C. The initial acceleration of firing rate was derived from

force-firing rate relation fitted with the natural logarithm (f(force) =a*ln(force)+b; yellow trace) and its first

derivative (f(force)=a/force; dark red trace). The right panels show the distribution of initial acceleration

values relative to recruitment threshold (RT) for all participants (n = 328 motor units for TA and n = 393

motor units for VL). Each data point indicates a motor unit. The horizontal thick lines denote a statistical

difference between the motor units groups (low-threshold = Blue; medium-threshold = Grey; high-threshold

= Pink). The green line depicts the non-linear fits of these relations for the Tibialis Anterior and the Vastus

Lateralis. Similar fits were observed for all the participants (inset panels).

The second stage of rate coding involved a linear increase in firing rate after the initial

acceleration phase. This stage was analysed using the average firing rate during the successive

force plateaus for each tracked motor unit (Figure 3). Specifically, the increase in firing rate

between consecutive contraction levels (i.e., change in force equal to 10% MVC) was compared

with a linear mixed effect model with *Muscle* (TA; VL) and *Threshold* (low; medium; high) as

fixed effects and *Participant* as a random effect. There were significant effects for *Muscle*

(F=39.7; p<0.001) and *Threshold* (F=145.3; p<0.001), and a significant interaction *Muscle ×*

Threshold (F=37.7; p<0.001). The rate of increase was significantly greater for TA (3.5 ± 1.7

pps; 25th-75th percentile: 2.4 – 4.1 pps) than for VL (2.0 ± 1.1 pps; 25th-75th percentile: 1.3 – 2.6

pps. p<0.001). The rate of increase was significantly greater in TA for high-threshold than for

low- (p<0.001) and medium- (p<0.001) threshold motor units and for medium- than for low-

threshold motor units (p<0.001). In contrast, the rate of increase in VL was greater for high-

(p<0.001) and medium- (p<0.001) threshold motor units than for low-threshold motor units,

with no difference between high- and medium-threshold motor units in VL (p=0.993).

The linear functions for each participant are shown in Figure 3. There was a significant positive

association between the rate of increase in firing rate and the recruitment threshold for all the

participants (Pearson's r: TA: 0.71±0.14 pps and VL:0.57±0.22 pps; all p<0.022), except in one

participant for each muscle (TA: r=0.17; p=0.272; VL: r= 0.12; p=0.411). Most motor units

(92.0% in TA and 80.9% in VL) increased firing rates by more than one pulse per second

between contractions up to the maximal force tested or until tracking was not possible.

Figure 3. Motor unit firing rates across contraction levels. The left column shows average firing rate (pulses per second, pps) during the force plateaus for each tracked motor unit across contraction levels for all participants from TA (n = 998 motor units; A) and VL (n = 1016 motor units; B). Each data point indicates one motor unit, and each line connects the firing rates of this motor unit across contractions. The colour scale identifies the three groups of motor units based on recruitment threshold: low (blue), medium (grey), and high (pink). The middle column depicts the change in firing rates between contractions separated by 10% to 70% of the maximal voluntary contraction level (MVC) of force. The right column shows the relation between the rate of increase in firing rate between successive target forces (e.g., between 10% and 20%MVC) and the recruitment threshold of the motor unit. These relations were fitted with a linear function (coloured lines in the inset panels) for each participant. The horizontal thick line denotes a statistical difference between motor units grouped by recruitment thresholds.

Differences between ramp-up and ramp-down - hysteresis:

Neuromodulatory inputs can prolong excitatory synaptic inputs and produce unequal recruitment and derecruitment thresholds; that is, a hysteresis. The left columns in Figure 4 show the relation between recruitment and derecruitment thresholds within the motor unit pools (Figure 4). The relation for most participants was better characterised with a non-linear than a linear function (TA: 6 out of 8 participants, adjusted R-squared >0.92); VL: 7 out of 8 participants, adjusted R-squared >0.83). The difference between recruitement and

derecruitment thresholds was compared with a linear mixed effect model with *Muscle* (TA; VL) and *Threshold* (low; medium; high) as fixed effects and *Participant* as a random effect. There was a significant effect of *Threshold* (F=83.0; p<0.001) and a significant interaction *Muscle* × *Threshold* (F=30.1; p<0.001), but no effect of *Muscle* (F=2.5; p<0.137). We further tested for each group of motor units whether the difference between the two thresholds was significantly different from 0 (absence of hysteresis). The result was that only medium-threshold motor units had a significant positive hysteresis (derecruitment threshold was lower) in both muscles (TA: -3.8 ± 7.2 pps; p = 0.013; VL: -3.1 ± 6.0 pps; p = 0.026), whereas low-threshold motor units from both muscles (TA: -1.5 ± 4.8; VL: -1.0 ± 3.2 pps; p>0.221) did not exhibit significant hysteresis. Although high-threshold motor units from TA followed the same trend (-1.7 ± 6.0 pps; p<0.437), high-threshold motor units from VL exhibited a negative hysteresis (VL: +3.2 ± 6.7 pps; p=0.039), which indicated that derecruitment threshold was greater than the recruitment threshold.

We also compared the type of the best fit between motor unit firing rate and force between the ramp-up and ramp-down phases of the isometric contraction. Most motor units (TA: 60.2%; VL: 76.7%) were best fit with the same relation during the two phases. Although the percentage of associations better fitted with linear functions increased for the ramp-down phase for the TA (from 25.4% to 41.9% of motor units), there was no change between ramps for the VL (from 12.8% to 19.4% of motor units). Thus, most motor units kept a non-linear relation during the ramp-down phase.

Figure 4. Hysteresis between recruitment and derecruitment thresholds. The left column depicts the relations between the recruitment and derecruitment thresholds of each motor unit from TA (A) and VL (B). These relations were fitted for each participant (coloured lines) using either non-linear or linear regressions. The values below the dashed red line (recruitment threshold = derecruitment threshold) show a positive hysteresis between recruitment and derecruitment thresholds, the values above a negative hysteresis. The right column shows the difference between the recruitment and derecruitment thresholds, with negative values showing a positive hysteresis with recruitment threshold greater than derecruitment threshold and the positive values indicating the converse (a negative hysteresis). Each data point is a motor unit. The asterisk denotes a statistical difference between the hysteresis values for motor units grouped according to recruitment threshold (low = Blue; medium = Grey; high = Pink) and the absence of a hysteresis (dashed horizontal line).

Discussion:

No previous studies have identified to date the firing activity of >100 motor units spanning most of the spectrum of recruitment thresholds from the same individual, whether in animals or humans. According to previous estimates, the number of motor units detectable with surface EMG in human would be 200±61 for TA and 146±29 for VL (Duchateau and Enoka, 2022). We have therefore identified in this study most of the active motor units present in the recorded volume for each muscle (Figure 1), which has allowed us to infer the rate coding of pools of motor units during slow isometric contractions. Overall, we found that motor units within a pool exhibit distinct rate coding with changes in force level (Figure 2 and 3), which contrasts with the long-held belief that rate coding is similar across motor units from the same pool (Fuglevand et al., 1993; De Luca and Contessa, 2012).

The fast initial acceleration stage reflects the amplification of synaptic inputs through the activation of persistent inward currents via voltage-gated sodium and calcium channels (Bennett et al., 1998a; Lee and Heckman, 2000; Heckman and Enoka, 2012; Binder et al., 2020). Although noradrenergic and serotonergic synapses that generate persistent inward currents are widespread within motor unit pools (Maratta et al., 2015), the influence of the neuromodulatory inputs is greater during this phase for low- than for high-threshold motor units. This is presumably due to a larger ratio between the amplitude of inward currents and the input conductance in low threshold motoneurons (Huh et al., 2017). The level of recurrent and reciprocal inhibition has also probably increased with the increase in force during the ramp up, progressively blunting the effect of persistent inward currents for late-recruited motor units (Kuo et al., 2003; Hyngstrom et al., 2007; Revill and Fuglevand, 2017). This may also explain the larger percentage of high-threshold motor units with a linear fit for the firing rate/force relation (Figure 2), as the integration of larger inhibitory inputs should linearise the firing rate/force relation (Revill and Fuglevand, 2017).

The second stage of rate coding involves a slower linear increase in firing rate, as characterised by the second part of a logarithmic function (Figures 2-3). This stage corresponds to mechanisms previously referred to as 'rate limiting' (Heckman and Binder, 1993; Heckman and Enoka, 2012; Powers and Heckman, 2017) or 'saturation' (De Luca and Contessa, 2012; Fuglevand et al., 2015), and was more pronounced for low- than high-threshold motor units (Figure 3). This difference may be explained by smaller excitatory synaptic inputs onto low-

than high-threshold motoneurons (Powers and Binder, 2001; Heckman and Enoka, 2012), lower synaptic driving potential of the dendritic membrane (Powers and Binder, 2000; Cushing et al., 2005; Fuglevand et al., 2015), and longer and larger afterhyperpolarisation phase in low- than high-threshold motoneurons (Bakels and Kernell, 1993; Gardiner, 1993; Deardorff et al., 2013; Caillet et al., 2022).

Taken together, these results show how ionotropic and neuromodulatory inputs to motoneurons uniquely combine to generate distinct rate coding across the pool, even if a more direct manipulation of the sources of neuromodulatory and ionotropic inputs will be required to directly estimate their interactions. While the size of the motor unit determines its recruitment threshold, neuromodulatory inputs determine its gain. Thus, in a similar fashion as size imposes a fixed recruitment order (Henneman, 1957), neuromodulatory inputs determines a spectrum of variations in motor unit firing rates without the need to differentiate the ionotropic inputs to each motoneuron. [Figure 2C-3-S3; (Powers and Heckman, 2017; Bracklein et al., 2022; Chardon et al., 2023; Škarabot et al., 2023)]. Because high-threshold motor units exhibit a higher gain than low-threshold motor units (Figure 3), their firing rate could eventually reach similar or even greater values than that of low-threshold motor units during strong contractions (Gydikov and Kosarov, 1974; Moritz et al., 2005; Oya et al., 2009; Škarabot et al., 2023). This indicates that the onion skin principle (De Luca and Contessa, 2012, 2015) may not hold in all muscles and for all contraction forces (Gydikov and Kosarov, 1974; Moritz et al., 2005; Oya et al., 2009; Škarabot et al., 2023). In addition, rate coding patterns should also vary with the pattern of contractions, with fast contractions lowering the range of recruitment thresholds within motoneuron pools (Desmedt and Godaux, 1977a, 1979; van Bolhuis et al., 1997). The variability in rate coding observed here between motor units from the same pool could lead to small deviations from the size principle sometimes observed between pairs of units during isometric contractions with various patterns of force (Desmedt and Godaux, 1979; Marshall et al., 2022) or during the derecruitment phase (Bracklein et al., 2022).

It is also worth noting that the profile of rate coding mostly followed the same trend during the ramp-up and ramp-down phases of the contraction, but with prolonged firing activity only evident for medium-threshold motor units (Figure 4). This hysteresis is possible because of the bistability of the membrane potential mediated by inward currents from calcium channels, the two stable membrane states being the resting potential and a depolarised state that enables self-sustained firing (Hounsgaard et al., 1988; Lee and Heckman, 1998). The absence of statistically

significant hysteresis in low threshold motoneurons may be simply explained by their early recruitment during the first percentages of force, mathematically lowering the potential amplitude of their hysteresis. Alternatively, prominent outward currents may attenuate the impact of persistent inward currents on the prolongation of their firing activity (Powers and Heckman, 2015). Similarly, the absence of hysteresis (TA) and even a negative hysteresis (VL) in high-threshold motoneurons can be explained by a briefer membrane bistability due to a faster decay of inward currents from calcium channels, and a narrower range of membrane depolarisation over which these inward currents are activated (Lee and Heckman, 1998). This result must be confirmed with a more direct proxy of the net synaptic drive, such as the firing rate of a *reference* low-threshold motor neuron used in the delta F method (Gorassini et al., 1998), or the cumulative spike train of low-threshold motor neurons (Afsharipour et al., 2020).

The increase in firing rate was also significantly greater for TA motor units than for those in VL. This difference may reflect a varying balance between excitatory/inhibitory synaptic inputs and neuromodulation due to multiple spinal circuits (Heckman and Binder, 1993; Heckman et al., 2008; Johnson et al., 2017; Powers and Heckman, 2017; Chardon et al., 2023; Škarabot et al., 2023). Specifically, the strength of recurrent and reciprocal inhibitory inputs to motoneurons innervating VL and TA, and their proportional or inverse covariation with excitatory inputs, respectively, may explain the differences in rate limiting and maximal firing rates (Heckman and Binder, 1993; Heckman et al., 2008; Johnson et al., 2017; Powers and Heckman, 2017; Chardon et al., 2023; Škarabot et al., 2023). Thus, the motor units from the VL may receive more recurrent inhibition than those of distal muscles, though direct evidence of these differences remains to be found in humans (Windhorst, 1996). Interestingly, similar differences in rate coding were previously observed between proximal and distal muscles of the upper limb (De Luca et al., 1982). However, other muscles that serve different functions within the human body, such as muscles from the face, have different rate coding characteristics with much higher firing rates (Kirk et al., 2021). Future work should investigate those muscles and other to reveal the myriads of rate coding strategies in human muscles.

Our results on rate coding characteristics of the motor unit pool provide insight on force control strategies. The accurate control of force depends on the firing activity of the population of active motor units, which effectively filters out the synaptic noise of individual motor units and primarily transforms common synaptic inputs into muscle force (Dideriksen et al., 2012; Farina et al., 2014a; Farina and Negro, 2015). Because the bandwidth of muscle force is <10 Hz during

isometric contractions (Enoka and Farina, 2021), the activation of inward currents to low-threshold motoneurons at recruitment may enable them to promptly discharge at a rate (>8 pps) that transmits the effective common synaptic inputs (<8-10 Hz) without phase distortion (Dideriksen et al., 2012; Farina et al., 2014a; Farina and Negro, 2015). This mechanism facilitates the accuracy of force control.

Moreover, force generation can be described as the filtering of the cumulative firing activity of active motor units with the average twitch force of their muscle units (Farina and Negro, 2015; Thompson et al., 2018). From this perspective, at low forces, recruitment of new motor units has a higher impact on force modulation than rate coding since each recruitment impacts both the cumulative firing activity of the pool and the average twitch force. Rate coding only modulates the cumulative firing activity. Thus, the amplification of the firing rate of low-threshold motor units near their recruitment threshold instantly favours rate coding over the recruitment of additional motor units, which likely allows for smoother force control. Similarly, the progressive recruitment of motor units with a higher gain promotes faster changes in firing rates, which promotes force control accuracy across the full force range. On a different note, the steep increase in firing rate over the first percentages of the ramp-up may also enable the motor units to produce the required level of force despite having a more compliant muscle-tendon unit (Mazzo et al., 2021). Overall, our results may help to design future non-linear decoders that aim to predict muscle activation or muscle force from descending inputs recorded in the motor cortex, with the will to generalise their performance across movements (Naufel et al., 2019).

Materials and Methods:

Participants:

16 young individuals volunteered to participate either in the experiment on the Tibialis Anterior (n=8; age: 27 ± 3) or on the Vastus Lateralis (n=8; age: 27 ± 10). They had neither history of lower limb injury in the last six months before the experiments nor lower leg pain that would impair their ability to complete the experimental tasks. The study was reviewed and approved by Imperial College London (Study 18IC4685) and Comité de Protection des Personnes Ouest III (Study 23.00453.000166) and followed the standards of the declaration of Helsinki. Participants provided their informed written consent before starting the experimental session.

Experimental setup:

The two experimental sessions consisted of either a series of submaximal isometric ankle dorsiflexions or isometric knee extensions. Electromyographic (EMG) signals were recorded from either the Tibialis Anterior (TA) or the Vastus Lateralis (VL) muscles using four arrays of 64 surface electrodes for a total of 256 electrodes.

For the session of ankle dorsiflexions, participants sat on a massage table with the hips flexed at 45°, 0° being the hip neutral position, and the knees fully extended. The foot of the dominant leg (right in all participants) was fixed onto the pedal of an ankle dynamometer (OT Bioelettronica, Turin, Italy) positioned at 30° in the plantarflexion direction, 0° being the foot perpendicular to the shank. The thigh and the foot were fixed with inextensible Velcro straps. Force signals were recorded with a load cell (CCT Transducer s.a.s, Turin, Italy) connected in-series to the pedal using the same acquisition system as for the EMG recordings (EMG-Quattrocento; OT Bioelettronica, Italy).

For the session of knee extensions, participants sat on an instrumented chair with the hips flexed at 85°, 0° being the hip neutral position, and the knees flexed at 85°, 0° being the knees fully extended. The torso and the thighs were fixed to the chair with Velcro straps and the tibia were positioned against a rigid resistance connected to force sensors (Metitur, Jyvaskyla, Finland). The force signals were recorded using the same acquisition system as for the EMG recordings.

The experimental session began with a warm-up that consisted of submaximal isometric contractions from 50% to 80% of the subjective MVC. Then, participants performed two

maximal voluntary contractions (MVC) with two minutes of rest in between. The maximal

torque was considered as the highest torque value recorded over an average window of 250 ms.

The rest of the session consisted of a series of eight submaximal isometric contractions

performed at 10% to 80% of the MVC with 10 %MVC increments. The pattern of the

contractions followed a trapezoidal target displayed on a screen in real time with the force trace.

Ramp-up and ramp-down phases were performed at a constant pace of 5 %MVC.s^{-1}. The force

plateaus were maintained for either 10-s (70-80 %MVC), 15-s (50-60 %MVC) or 20-s (10-40

%MVC). The contractions were separated by 90 s of rest and their order was randomised.

<u>EMG recording:</u>

Surface EMG signals were recorded from the Tibialis Anterior (TA) or the Vastus Lateralis (VL)

using four two-dimensional arrays of 64 electrodes (GR04MM1305 for the TA; GR08MM1305

for the VL, 13x5 gold-coated electrodes with one electrode absent on a corner; interelectrode

distance: 4 and 8mm, respectively; OT Bioelettronica, Italy). The grids were positioned over

the muscle bellies to cover the largest surface while staying aways from the boundaries of the

muscle identified by manual palpation. Before placing the electrodes, the skin was shaved and

cleaned with an abrasive pad and water. A biadhesive foam layer was used to hold each array

of electrodes onto the skin, and conductive paste filled the cavities of the adhesive layers to

make skin-electrode contact. For two-source validation of the EMG decomposition, an

intramuscular linear array of 40 electrodes on a thin-film (platinum coated, interelectrode

distance: 1mm) was inserted into the TA in one participant at an approximate angle of 30°. The

insertion was guided with a portable ultrasound probe (Butterfly IQ+, Butterfly Network, USA).

Two bands damped with water were placed around the ankle as ground and reference electrodes.

EMG signals were recorded in monopolar derivation with a sampling frequency of 2048 Hz for

surface electrodes, 10240 Hz for intramuscular electrodes, amplified (x150), band-pass filtered

(10–500 Hz for surface; 100-4400 Hz for intramuscular), and digitized using a 400 channels

acquisition system with a 16-bit resolution (EMG-Quattrocento; OT Bioelettronica, Italy).

<u>HD-EMG decomposition</u>

The monopolar EMG signals were band-pass filtered between 20 and 500 Hz with a second-

order Butterworth filter, and channels with low signal-to-noise ratio or artifacts were discarded

after visual inspection. The EMG signals were decomposed into individual motor unit pulse

trains using convolutive blind-source separation (Negro et al., 2016b). EMG signals were first

extended by adding delayed versions of each channel, with an extension factor, i.e., number of

delayed versions, adjusted to reach a total of 1000 extended channels. The extended signals

were spatially whitened to make them uncorrelated and of equal power. Thereafter, a fixed-

point algorithm was applied to identify the sources of the EMG signals, i.e., the motor unit pulse

trains. In this algorithm, a contrast function $g(x) = \log(\cosh(x))$ and its derivatives were applied

to the extended and whitened EMG signals to iteratively optimise a separation vector that skew

the distribution of the values of the motor unit pulse trains toward 0, and thus maximise the

level of sparsity of the motor unit pulse train. The high level of sparsity matches the

physiological properties of motor units, with a relatively small number of discharges, or 1, per

second (< 50 discharge times/s during submaximal isometric contractions). The convergence

was reached once the level of sparsity did not substantially vary (with a tolerance fixed at 10^{-4})

when compared to the previous iteration. At this stage, the motor unit pulse train contained high

peaks (i.e., the discharge times of the identified motor unit) and lower values due to the activities

of neighbouring motor units and noise. High peaks were separated from lower values using the

MATLAB functions *findpeaks.m* and *kmeans.m* (with two classes). The peaks from the class

with the highest centroid were considered as the discharge times of the identified motor unit.

After this automatic identification of the discharge times, duplicates were automatically

removed. For this purpose, the pulse trains of pairs of motor units were first aligned using a

cross-correlation function to account for a potential delay due to the propagation time of action

potentials along the fibres. Then, two discharge times identified from these pulse trains were

considered as common when they occurred within a time interval of 0.5 ms, and two or more

motor units were considered as duplicates when they had at least 30% of their identified

discharge times in common (Holobar et al., 2010; Negro et al., 2016b). In principle, the limited

level of synchronisation between individual motor units results in a few simultaneous discharge

times between pairs of motor units. A threshold of 30% is therefore highly conservative and

ensure the removal of all motor units with a level of synchronisation well above physiological

values. When duplicates were identified, the motor unit with the lowest coefficient of variation

of the inter-spike intervals was retained for the analyses.

At the end of these automatic steps, all the motor unit pulse trains and identified discharge times

were visually inspected, and manual editing was performed to correct the false identification of

artifacts or the missed discharge times (Del Vecchio et al., 2020; Hug et al., 2021; Avrillon et

al., 2023). The manual editing consisted of i) removing the spikes causing erroneous discharge

rates (outliers), ii) adding the discharge times clearly separated from the noise, iii) recalculating

the separation vector, iv) reapplying the separation vector on the entire EMG signals, and v)

repeating this procedure until the selection of all the discharge times is achieved. The manual

editing of potential missed discharge times and falsely identified discharge times was never

immediately accepted. Instead, the procedure was consistently followed by the application of

the updated motor unit separation vector on the entire EMG signals to generate a new motor

unit pulse train. Then, the manual editing was only accepted when the silhouette value increased

or stayed well above the threshold of 0.9 quantified with the silhouette value (Negro et al.,

2016b). Only these motor units were retained for further analysis.

<u>Validation of the decomposition.</u>

EMG decomposition was validated using both simulation of EMG signals and two-source

validation.

For the simulation, we generated the series of discharge times of a group of 150 motor units in

an anatomical model entailing a cylindrical muscle volume with parallel fibres [see (Konstantin

et al., 2020) for a full description of the model], in which subcutaneous and skin layers separate

the muscle from the surface electrodes. We set the radius of the muscle to 15 mm and the

thicknesses of the subcutaneous and skin layers to 4 mm and 1 mm, respectively. The motor

unit action potentials were detected in the model by a grid of 64 circular surface electrodes with

a diameter of 1 mm arranged in 5 columns and 13 rows (inter-electrode distance: 4mm).

Following this, we estimated the rate of agreements between simulated and estimated series of

motor unit discharge times, calculated as the ratio between correctly identified discharge times

and the sum of correctly identified discharge times, missed discharge times, and falsely

identified discharge times.

For the two-source validation, EMG signals were simultaneously recorded with an

intramuscular linear array of 40 electrodes on a thin film and two grids of 64 surface electrodes.

After the decomposition of both intramuscular and surface EMG signals, identified motor units

were matched following the same procedure as for the elimination of duplicates. In short, the

motor unit pulse trains were first aligned using a cross-correlation function. Two motor units

identified with intramuscular and surface EMG decompositions were considered as matches

when they had at least 30% of their discharge times in common. We calculated the accuracy of

the decomposition using the rate of agreements between their series of discharge times.

<u>Proportion of the EMG signal represented by the decomposition.</u>

A synthetic EMG signal was reconstructed from the cumulative firing activity of the identified

motor units. First, EMG signals from the 256 electrodes were differentiated in the column

direction to obtain 236 single-differential EMG channels. Second, each differentiated EMG

signal was segmented over successive windows of 25 ms centred around the discharge times of

a motor unit. All the windows were averaged to estimate the average action potential waveform

of that motor unit over the 236 each channel. The action potentials were then convolved with

the series of discharge times to obtain trains of action potentials. These steps were repeated for

all the identified motor units, and all the trains of action potentials were summed to reconstruct

the synthetic EMG signals. At the end of this process, the ratio between the powers of synthetic

and original EMG signals was calculated.

<u>Motor unit tracking:</u>

As explained in the section that described the decomposition framework and in previous studies

(Francic and Holobar, 2021; Skarabot et al., 2023; Škarabot et al., 2023), a motor unit pulse

train results from the projection of extended and whitened EMG signals onto a separation

vector, optimised during a fixed-point algorithm. Discharge times are automatically identified

from this motor unit pulse train, before visually inspection and manual editing.

In this study, motor units were tracked by slightly adapting this process. First, EMG signals

from two successive contraction levels, say 10% and 20% MVC, were separately decomposed.

The motor unit pulse trains, the identified discharge times, and the associated separation vectors

were saved for each contraction level. Second, the EMG signals from the highest contraction

level (20% MVC) were projected onto the separation vectors identified in the lowest contraction

level (10% MVC). A new group of pulse trains was generated, which represented the firing

activity of all the motor unit identified at 10% MVC, but during the contraction performed at

20% MVC. When the high peaks were clearly separated from the noise, the discharge times

were automatically identified using the MATLAB functions *findpeaks.m* and *kmeans.m*. Third,

these new series of discharge times were matched with those that have been initially identified

at 20% MVC using the same approach as during two-source validation (>30% of common

discharge times, see above). This process ended with three groups of motor units: i) the motor

units only identified at 10% MVC, ii) the motor units identified at 10% and 20% MVC, and iii)

the motor units only identified at 20% MVC. These steps were repeated between all the

successive contraction levels, that is, from between 20% and 30% MVC to between 70% and
80% MVC. At the end of this iterative process, the following data were saved for each motor
unit across the contraction levels where they were tracked: the instantaneous firing rates and
force during ramp up and ramp down phases (Figure 2), the average firing rate during the
plateau (Figure 3), the recruitment threshold (Figure S3).

Motor unit uniqueness:

The accuracy of motor unit tracking was further tested by confirming the uniqueness of each
motor unit within the entire sample, assuming that each motor unit must have a unique
representation of their action potentials across the array of surface electrodes [Figure 1; (Farina
et al., 2008)]. This was accomplished by calculating the root mean-square error (RMSE)
between their action potentials across contractions and the action potentials of the rest of the
motor units. EMG signals from the 256 electrodes were differentiated in the column direction
to obtain 236 single-differential EMG channels. For each motor unit, the single-differential
EMG signals were segmented around the discharge times and averaged to identify an average
action potential waveforms for each channel. The action potential waveforms were
concatenated in a matrix of 236 rows (EMG channels) and 102 columns (time samples; 50 ms).
This process was repeated for each contraction level where the motor unit was tracked. A first
RMSE was calculated between the concatenated action potentials identified across contractions,
and this value was defined as a reference. Then, RMSE was calculated between the
concatenated action potentials of that motor unit and the concatenated action potentials of the
rest of the motor units. A reference value lower than the 5th percentile of the distribution of
RMSE calculated between motor units demonstrated that its distribution of action potentials
across contractions were more similar than with the rest of the motor units, proving their
uniqueness within the entire sample.

Input-output function of motoneurons:

The input-output function of each motor unit was characterised as the relation between its
instantaneous firing rate and the muscle force. The instantaneous firing rates and muscle forces
recorded during all the ramp-up phases where that motor unit was tracked were pooled. For
example, the motor unit displayed in Figure 2A was tracked from 20% to 60% of the MVC.
The relation between firing rates and force was characterised by comparing three curve fits:
• A linear fit: Firing rate(Force) = a(Force) + b
• A rising exponential: Firing rate(Force) = a(1 − e$^{(-Force/b)}$) + c

•   A natural logarithm: Firing rate(Force) = a * ln(Force) + b

The parameters of the functions were estimated with the MATLAB functions *fittype.m* and *fit.m*

using a nonlinear least-squares solver with a maximum of 1000 iterations. The best model of

the firing rate-force relation during the ramp up phase was the fit with the lowest Bayesian

information criterion (BIC). This criterion assesses the performance of a model by balancing

its goodness of fit with its complexity (number of parameters). The BIC was calculated as

follow:

$$BIC = n * \ln(sse / n) + p * \ln(n)$$

where n is the number of data samples on which the fit was estimated, sse is the sum of squares

error, and p is the number of parameters used in the model.

The input-output functions of motor units during the ramp-down phase were characterised using

the same approach, and we reported in the results whether the best fit changed between the

ramp-up and ramp-down phases.

<u>Rate coding of motor units:</u>

Most of motor units input-output functions were better characterised with a natural logarithm

function. This function determines two stages in rate coding: a steep acceleration of firing rate

followed by a slower linear increase in firing rate. Therefore, these two stages of the firing rate-

force relation were further analysed. To estimate the acceleration of firing rate during the ramp-

up phase, the first derivative of the firing rate-force relation fitted with a natural logarithm was

computed as:

Acceleration of firing rate(Force) = a / Force

where a is the parameter of the equation of the natural logarithm function. The initial

acceleration of firing rate was calculated as the value of the derivative at the recruitment

threshold of the motor unit. The relation between the initial acceleration of firing rate and the

recruitment threshold of all motor units across the pool was characterised for each muscle and

each participant using nonlinear least-squares solvers in MATLAB. The slow linear increase in

firing rate that followed the initial steep acceleration of firing rate was characterised by

comparing the average firing rate of motor units during the plateaus of force between successive

contraction levels (Figure 3). Specifically, the rate of increase in firing rate for an increment of

10 %MVC were calculated for each motor unit, and the relation between the rate of increase

and the recruitment threshold of all motor units across the pool was characterised for each

participant and each muscle using a linear fit in MATLAB. Pearson's r and p values of these relations were reported in the results.

Hysteresis between recruitment and derecruitment thresholds.

The relation between recruitment and derecruitment thresholds of all motor units across the pool were characterised for each participant and each muscle using non-linear and linear fits. As for the relations between the firing rate and force of each motor unit, the best fit was selected using the Bayesian information criterion. The differences between the recruitment and derecruitment thresholds (hysteresis) were calculated for each motor unit, with negative values indicating a hysteresis and a positive value indicating a reverse hysteresis.

Statistical analyses.

All statistical analyses were performed with RStudio (USA). Quantile-quantile plots and histograms were displayed to check the normality of the data distribution. If the distribution was determined not to be normal, they were transformed to remove the skew. All statistical analyses were performed using linear mixed effect models implemented in the R package *lmerTest* with the Kenward-Roger method to estimate the denominator degrees of freedom and the p-values. This method considers the dependence of data points (i.e., individual motor unit) in each participant. When necessary, multiple comparisons were performed using the R package *emmeans*, which adjusts the p-value using the Tukey or the Bonferroni method. The significance level was set at 0.05. Values are reported as mean ± standard deviation.

Initial accelerations of firing rate and rates of increase in firing rate between successive contraction levels were compared between motor units using a linear mixed effect model, with *Muscle* (TA; VL) and *Threshold* (low; medium; high) as fixed effects and *Participant* as a random effect.

The difference between motor units' recruitment and derecruitment thresholds were compared using a linear mixed effect model, with *Muscle* (TA; VL) and *Threshold* (low; medium; high) as fixed effects and *Participant* as a random effect. Whether the difference between the two thresholds were significantly different from 0 (absence of hysteresis) was further tested using function *contrast* in the package *emmeans*.

References:

Afsharipour B, Manzur N, Duchcherer J, Fenrich KF, Thompson CK, Negro F, Quinlan KA, Bennett DJ, Gorassini MA (2020) Estimation of self-sustained activity produced by persistent inward currents using firing rate profiles of multiple motor units in humans. J Neurophysiol 124:63-85.

Avrillon S, Hug F, Gibbs C, Farina D (2023) Tutorial on MUedit: An open-source software for identifying and analysing the discharge timing of motor units from electromyographic signals. bioRxiv:2023.2007.2013.548568.

Bakels R, Kernell D (1993) Matching between motoneurone and muscle unit properties in rat medial gastrocnemius. J Physiol 463:307-324.

Beauchamp JA, Pearcey GEP, Khurram OU, Chardon M, Wang YC, Powers RK, Dewald JPA, Heckman CJ (2023) A geometric approach to quantifying the neuromodulatory effects of persistent inward currents on individual motor unit discharge patterns. J Neural Eng 20.

Bennett DJ, Hultborn H, Fedirchuk B, Gorassini M (1998a) Synaptic activation of plateaus in hindlimb motoneurons of decerebrate cats. J Neurophysiol 80:2023-2037.

Bennett DJ, Hultborn H, Fedirchuk B, Gorassini M (1998b) Short-term plasticity in hindlimb motoneurons of decerebrate cats. J Neurophysiol 80:2038-2045.

Bigland-Ritchie B, Johansson R, Lippold OC, Smith S, Woods JJ (1983) Changes in motoneurone firing rates during sustained maximal voluntary contractions. J Physiol 340:335-346.

Binder MD, Powers RK, Heckman CJ (2020) Nonlinear Input-Output Functions of Motoneurons. Physiology (Bethesda) 35:31-39.

Bracklein M, Barsakcioglu DY, Ibanez J, Eden J, Burdet E, Mehring C, Farina D (2022) The control and training of single motor units in isometric tasks are constrained by a common input signal. Elife 11.

Caillet AH, Phillips ATM, Farina D, Modenese L (2022) Mathematical relationships between spinal motoneuron properties. Elife 11.

Caillet AH, Phillips ATM, Farina D, Modenese L (2023a) Motoneuron-driven computational muscle modelling with motor unit resolution and subject-specific musculoskeletal anatomy. PLoS Comput Biol 19:e1011606.

Caillet AH, Avrillon S, Kundu A, Yu T, Phillips ATM, Modenese L, Farina D (2023b) Larger and Denser: An Optimal Design for Surface Grids of EMG Electrodes to Identify Greater and More Representative Samples of Motor Units. eNeuro 10.

Chardon MK, Wang YC, Garcia M, Besler E, Beauchamp JA, D'Mello M, Powers RK, Heckman CJ (2023) Supercomputers and Reverse Engineering of Motoneuron Firing Patterns. bioRxiv:2022.2012.2009.519818.

Cushing S, Bui T, Rose PK (2005) Effect of nonlinear summation of synaptic currents on the input-output properties of spinal motoneurons. J Neurophysiol 94:3465-3478.

De Luca CJ (1985) Control properties of motor units. J Exp Biol 115:125-136.

De Luca CJ, Contessa P (2012) Hierarchical control of motor units in voluntary contractions. J Neurophysiol 107:178-195.

De Luca CJ, Contessa P (2015) Biomechanical benefits of the Onion-Skin motor unit control scheme. J Biomech 48:195-203.

De Luca CJ, LeFever RS, McCue MP, Xenakis AP (1982) Behaviour of human motor units in different muscles during linearly varying contractions. J Physiol 329:113-128.

Deardorff AS, Romer SH, Deng Z, Bullinger KL, Nardelli P, Cope TC, Fyffe RE (2013) Expression
of postsynaptic Ca2+-activated K+ (SK) channels at C-bouton synapses in mammalian
lumbar -motoneurons. J Physiol 591:875-897.
Del Vecchio A, Holobar A, Falla D, Felici F, Enoka RM, Farina D (2020) Tutorial: Analysis of motor
unit discharge characteristics from high-density surface EMG signals. J Electromyogr
Kinesiol 53:102426.
Desmedt JE, Godaux E (1977a) Fast motor units are not preferentially activated in rapid
voluntary contractions in man. Nature 267:717-719.
Desmedt JE, Godaux E (1977b) Ballistic contractions in man: characteristic recruitment pattern
of single motor units of the tibialis anterior muscle. J Physiol 264:673-693.
Desmedt JE, Godaux E (1979) Voluntary motor commands in human ballistic movements. Ann
Neurol 5:415-421.
Dideriksen JL, Negro F, Enoka RM, Farina D (2012) Motor unit recruitment strategies and
muscle properties determine the influence of synaptic noise on force steadiness. J
Neurophysiol 107:3357-3369.
Duchateau J, Enoka RM (2022) Distribution of motor unit properties across human muscles. J
Appl Physiol (1985) 132:1-13.
Enoka RM, Duchateau J (2017) Rate Coding and the Control of Muscle Force. Cold Spring Harb
Perspect Med 7.
Enoka RM, Farina D (2021) Force Steadiness: From Motor Units to Voluntary Actions.
Physiology (Bethesda) 36:114-130.
Farina D, Negro F (2015) Common synaptic input to motor neurons, motor unit
synchronization, and force control. Exerc Sport Sci Rev 43:23-33.
Farina D, Holobar A (2016) Characterization of Human Motor Units From Surface EMG
Decomposition. Proceedings of the Ieee 104:353-373.
Farina D, Negro F, Dideriksen JL (2014a) The effective neural drive to muscles is the common
synaptic input to motor neurons. J Physiol 592:3427-3441.
Farina D, Merletti R, Enoka RM (2014b) The extraction of neural strategies from the surface
EMG: an update. J Appl Physiol (1985) 117:1215-1230.
Farina D, Negro F, Gazzoni M, Enoka RM (2008) Detecting the unique representation of motor-
unit action potentials in the surface electromyogram. J Neurophysiol 100:1223-1233.
Francic A, Holobar A (2021) On the Reuse of Motor Unit Filters in High Density Surface
Electromyograms Recorded at Different Contraction Levels. Ieee Access 9:115227-
115236.
Fuglevand AJ, Winter DA, Patla AE (1993) Models of recruitment and rate coding organization
in motor-unit pools. J Neurophysiol 70:2470-2488.
Fuglevand AJ, Lester RA, Johns RK (2015) Distinguishing intrinsic from extrinsic factors
underlying firing rate saturation in human motor units. J Neurophysiol 113:1310-1322.
Gardiner PF (1993) Physiological properties of motoneurons innervating different muscle unit
types in rat gastrocnemius. J Neurophysiol 69:1160-1170.
Goodlich BI, Horan SA, Kavanagh JJ (2022) Blockade of 5-HT(2) receptors suppresses rate of
torque development and motor unit discharge rate during rapid contractions. J
Neurophysiol 127:150-160.
Gorassini MA, Bennett DJ, Yang JF (1998) Self-sustained firing of human motor units. Neurosci
Lett 247:13-16.
Gydikov A, Kosarov D (1974) Some features of different motor units in human biceps brachii.
Pflugers Arch 347:75-88.

Heckman CJ, Binder MD (1993) Computer simulations of the effects of different synaptic input systems on motor unit recruitment. J Neurophysiol 70:1827-1840.

Heckman CJ, Enoka RM (2012) Motor unit. Compr Physiol 2:2629-2682.

Heckman CJ, Hyngstrom AS, Johnson MD (2008) Active properties of motoneurone dendrites: diffuse descending neuromodulation, focused local inhibition. J Physiol 586:1225-1231.

Henderson TT, Taylor JL, Thorstensen JR, Tucker MG, Kavanagh JJ (2022) Enhanced availability of serotonin limits muscle activation during high-intensity, but not low-intensity, fatiguing contractions. J Neurophysiol 128:751-762.

Henneman E (1957) Relation between size of neurons and their susceptibility to discharge. Science 126:1345-1347.

Holobar A, Minetto MA, Botter A, Negro F, Farina D (2010) Experimental analysis of accuracy in the identification of motor unit spike trains from high-density surface EMG. IEEE Trans Neural Syst Rehabil Eng 18:221-229.

Hounsgaard J, Hultborn H, Jespersen B, Kiehn O (1988) Bistability of alpha-motoneurones in the decerebrate cat and in the acute spinal cat after intravenous 5-hydroxytryptophan. J Physiol 405:345-367.

Hug F, Avrillon S, Del Vecchio A, Casolo A, Ibanez J, Nuccio S, Rossato J, Holobar A, Farina D (2021) Analysis of motor unit spike trains estimated from high-density surface electromyography is highly reliable across operators. J Electromyogr Kinesiol 58:102548.

Huh S, Siripuram R, Lee RH, Turkin VV, O'Neill D, Hamm TM, Heckman CJ, Manuel M (2017) PICs in motoneurons do not scale with the size of the animal: a possible mechanism for faster speed of muscle contraction in smaller species. J Neurophysiol 118:93-102.

Hyngstrom AS, Johnson MD, Miller JF, Heckman CJ (2007) Intrinsic electrical properties of spinal motoneurons vary with joint angle. Nat Neurosci 10:363-369.

Johnson MD, Thompson CK, Tysseling VM, Powers RK, Heckman CJ (2017) The potential for understanding the synaptic organization of human motor commands via the firing patterns of motoneurons. J Neurophysiol 118:520-531.

Keenan KG, Farina D, Maluf KS, Merletti R, Enoka RM (2005) Influence of amplitude cancellation on the simulated surface electromyogram. J Appl Physiol (1985) 98:120-131.

Kirk EA, Zero AM, Rice CL (2021) Firing rate trajectories of human occipitofrontalis motor units in response to triangular voluntary contraction intensity. Exp Brain Res 239:3661-3670.

Kirk EA, Copithorne DB, Dalton BH, Rice CL (2016) Motor unit firing rates of the gastrocnemii during maximal and sub-maximal isometric contractions in young and old men. Neuroscience 330:376-385.

Konstantin A, Yu T, Le Carpentier E, Aoustin Y, Farina D (2020) Simulation of Motor Unit Action Potential Recordings From Intramuscular Multichannel Scanning Electrodes. IEEE Trans Biomed Eng 67:2005-2014.

Kuo JJ, Lee RH, Johnson MD, Heckman HM, Heckman CJ (2003) Active dendritic integration of inhibitory synaptic inputs in vivo. J Neurophysiol 90:3617-3624.

Laine CM, Martinez-Valdes E, Falla D, Mayer F, Farina D (2015) Motor Neuron Pools of Synergistic Thigh Muscles Share Most of Their Synaptic Input. J Neurosci 35:12207-12216.

Lee RH, Heckman CJ (1998) Bistability in spinal motoneurons in vivo: systematic variations in persistent inward currents. J Neurophysiol 80:583-593.

921   Lee RH, Heckman CJ (2000) Adjustable amplification of synaptic input in the dendrites of spinal
922   motoneurons in vivo. J Neurosci 20:6734-6740.

923   Maratta R, Fenrich KK, Zhao E, Neuber-Hess MS, Rose PK (2015) Distribution and density of
924   contacts from noradrenergic and serotonergic boutons on the dendrites of neck flexor
925   motoneurons in the adult cat. J Comp Neurol 523:1701-1716.

926   Marshall NJ, Glaser JI, Trautmann EM, Amematsro EA, Perkins SM, Shadlen MN, Abbott LF,
927   Cunningham JP, Churchland MM (2022) Flexible neural control of motor units. Nat
928   Neurosci 25:1492-1504.

929   Martinez-Valdes E, Negro F, Laine CM, Falla D, Mayer F, Farina D (2017) Tracking motor units
930   longitudinally across experimental sessions with high-density surface
931   electromyography. J Physiol 595:1479-1496.

932   Mazzo MR, Weinman LE, Giustino V, McLagan B, Maldonado J, Enoka RM (2021) Changes in
933   neural drive to calf muscles during steady submaximal contractions after repeated
934   static stretches. J Physiol 599:4321-4336.

935   Monster AW, Chan H (1980) Surface electromyogram potentials of motor units; relationship
936   between potential size and unit location in a large human skeletal muscles.
937   Experimental neurology 67:280-297.

938   Moritz CT, Barry BK, Pascoe MA, Enoka RM (2005) Discharge rate variability influences the
939   variation in force fluctuations across the working range of a hand muscle. J
940   Neurophysiol 93:2449-2459.

941   Muceli S, Poppendieck W, Holobar A, Gandevia S, Liebetanz D, Farina D (2022) Blind
942   identification of the spinal cord output in humans with high-density electrode arrays
943   implanted in muscles. Science advances 8:eabo5040.

944   Naufel S, Glaser JI, Kording KP, Perreault EJ, Miller LE (2019) A muscle-activity-dependent gain
945   between motor cortex and EMG. J Neurophysiol 121:61-73.

946   Negro F, Yavuz US, Farina D (2016a) The human motor neuron pools receive a dominant slow-
947   varying common synaptic input. J Physiol 594:5491-5505.

948   Negro F, Muceli S, Castronovo AM, Holobar A, Farina D (2016b) Multi-channel intramuscular
949   and surface EMG decomposition by convolutive blind source separation. J Neural Eng
950   13:026027.

951   Oya T, Riek S, Cresswell AG (2009) Recruitment and rate coding organisation for soleus motor
952   units across entire range of voluntary isometric plantar flexions. J Physiol 587:4737-
953   4748.

954   Powers RK, Binder MD (2000) Summation of effective synaptic currents and firing rate
955   modulation in cat spinal motoneurons. J Neurophysiol 83:483-500.

956   Powers RK, Binder MD (2001) Input-output functions of mammalian motoneurons. Rev Physiol
957   Biochem Pharmacol 143:137-263.

958   Powers RK, Heckman CJ (2015) Contribution of intrinsic motoneuron properties to discharge
959   hysteresis and its estimation based on paired motor unit recordings: a simulation study.
960   J Neurophysiol 114:184-198.

961   Powers RK, Heckman CJ (2017) Synaptic control of the shape of the motoneuron pool input-
962   output function. J Neurophysiol 117:1171-1184.

963   Powers RK, Elbasiouny SM, Rymer WZ, Heckman CJ (2012) Contribution of intrinsic properties
964   and synaptic inputs to motoneuron discharge patterns: a simulation study. J
965   Neurophysiol 107:808-823.

966   Revill AL, Fuglevand AJ (2017) Inhibition linearizes firing rate responses in human motor units:
967   implications for the role of persistent inward currents. J Physiol 595:179-191.

Sherrington CS (1925) Remarks on some aspects of reflex inhibition. Proc Biol Sci 97:519-545.

Skarabot J, Ammann C, Balshaw TG, Divjak M, Urh F, Murks N, Foffani G, Holobar A (2023)
Decoding firings of a large population of human motor units from high-density surface
electromyogram in response to transcranial magnetic stimulation. J Physiol 601:1719-
1744.

Škarabot J, Beauchamp JA, Pearcey GEP (2023) Human motor unit discharge patterns reveal
differences in neuromodulatory and inhibitory drive to motoneurons across
contraction levels. bioRxiv:2023.2010.2016.562612.

Thompson CK, Negro F, Johnson MD, Holmes MR, McPherson LM, Powers RK, Farina D,
Heckman CJ (2018) Robust and accurate decoding of motoneuron behaviour and
prediction of the resulting force output. J Physiol 596:2643-2659.

van Bolhuis BM, Medendorp WP, Gielen CC (1997) Motor unit firing behavior in human arm
flexor muscles during sinusoidal isometric contractions and movements. Exp Brain Res
117:120-130.

Wei K, Glaser JI, Deng L, Thompson CK, Stevenson IH, Wang Q, Hornby TG, Heckman CJ,
Kording KP (2014) Serotonin affects movement gain control in the spinal cord. J
Neurosci 34:12690-12700.

Windhorst U (1996) On the role of recurrent inhibitory feedback in motor control. Progress in
neurobiology 49:517-587.

Figure S1. Validation of electromyographic decomposition and motor unit tracking. We used two approaches to validate the electromyographic decomposition. First, we simulated a pool of 150 motor units producing forces from 10 to 80% of the maximal excitation. Electromyographic signals were simulated from either 16 intramuscular electrodes (black horizontal line, top panels) or 64 surface electrodes (dark red curves on the left of top panels). Each of the grey elliptical shapes represents the cross-sectional area of the simulated muscle. We identified motor units using blind-source separation, with their territories displayed in red in the top panels. We then calculated the rate of agreement between the series of discharge times identified from the decomposition and simulated. Each data point is a motor unit, and the horizontal black line depicts the

median. After this, we used the two-source validation approach, i.e., the comparison of discharge times identified from intramuscular and surface electromyographic signals. A participant performed trapezoidal isometric contractions with plateaus ranging from 10 to 80% of the maximal force while we recorded surface signals with a grid of 128 electrodes and intramuscular signals with 40 electrodes along a thin film that was inserted into the muscle with a hypodermic needle. Intramuscular and surface electromyographic signals were both decomposed and two motor units were matched between intramuscular and surface signals. We calculated the rate of agreement between the series of discharge times. It is worth noting that we identified the same number of discharge times with both methods. A rate of agreements less than 100% reflects misalignments of some discharge times with the surface signals due to the propagation of the action potentials through the tissue and potential overlapping action potentials at the surface of the skin. Finally, we validated the tracking method using the simulated signals, with high percentages highlighting the high accuracy of our approach.

Figure S2. Recruitment thresholds of motor units tracked across contractions. We tracked motor units across contractions using their unique distribution of action potentials within the grid of 256 electrodes. Then, we estimated their recruitment thresholds as the average of force over a window of 10 ms centred around the first discharge time. The recruitment thresholds, estimated for each contraction where the motor unit was identified, are displayed for each participant. A series of data points connected with a line of the same colour is a motor unit. TA. Tibialis Anterior. VL. Vastus Lateralis. MVC. Maximal voluntary contraction level P1. Participant #1.

Figure S3. Relations between force and firing rates fitted with a natural logarithm. We reported the relations between instantaneous motor units firing rates and the applied force. Then, we fitted these relations for each motor unit using a natural logarithm function (left panels). The non-linearities in the relations depict the influence of neuromodulatory inputs to motoneurons on the transformation between the synaptic inputs and the motor unit firing rates. To quantify the impact of the amplification effect of these neuromodulatory inputs, we estimated the acceleration of firing rate as the first derivative of the fitted firing rate/force curves (right panels). The fits are drawn over the range of forces where the motor unit was firing during the ramp-up phases of the contractions. Each line is a motor unit. MVC. Maximal voluntary contraction level.