The decoding of extensive samples of motor units in human muscles reveals the rate coding of entire motoneuron pools.

Simon Avrillon[1], François Hug[2, 3], Roger Enoka[4], Arnault H. Caillet[1], Dario Farina[1]

[1] Department of Bioengineering, Faculty of Engineering, Imperial College London, London W12 7TA, UK
[2] Université Côte d'Azur, LAMHESS, Nice 06200, France
[3] The University of Queensland, School of Biomedical Sciences, St Lucia QLD 4072, QLD, Australia
[4] Department of Integrative Physiology, University of Colorado Boulder, Boulder, CO, USA

Correspondence:
Dario Farina
Imperial College London, London, UK
e-mail: d.farina@imperial.ac.uk

Conflict of interests:
The authors declare no competing financial interests.

Funding resources.
François Hug is supported by the French government, through the UCAJEDI Investments in the Future project managed by the National Research Agency (ANR) with the reference number ANR-15-IDEX-01. Dario Farina is supported by the European Research Council Synergy Grant NaturalBionicS (contract #810346), the EPSRC Transformative Healthcare, NISNEM Technology (EP/T020970), and the BBSRC, "Neural Commands for Fast Movements in the Primate Motor System" (NU-003743).

Abstract:

To advance our understanding of the neural control of muscle, we decoded the firing activity of extensive samples of motor units in the Tibialis Anterior (129±44 per participant; n=8) and the Vastus Lateralis (130±63 per participant; n=8) during isometric contractions of up to 80% of maximal force. From this unique dataset, we characterised the rate coding of each motor unit as the relation between its instantaneous firing rate and the muscle force, with the assumption that the linear increase in isometric force reflects a proportional increase in the net synaptic excitatory inputs received by the motoneuron. This relation was characterised with a natural logarithm function that comprised two phases. The initial phase was marked by a steep acceleration of firing rate, which was greater for low- than medium- and high-threshold motor units. The second phase comprised a linear increase in firing rate, which was greater for high- than medium- and low-threshold motor units. Changes in firing rate were largely non-linear during the ramp-up and ramp-down phases of the task, but with significant prolonged firing activity only evident for medium-threshold motor units. Contrary to what is usually assumed, our results demonstrate that the firing rate of each motor unit can follow a large variety of trends with force across the pool. From a neural control perspective, these findings indicate how motor unit pools use gain control to transform inputs with limited bandwidths into an intended muscle force.

Significance statement:

Movements are performed by motoneurons transforming synaptic input into an activation signal that controls muscle force. The control signal is not linearly related to the net synaptic input, but instead emerges from interactions between ionotropic and neuromodulatory inputs to motoneurons. Critically, these interactions vary across motoneuron pools and differ between muscles. We decoded the activity of hundreds of motor units from electromyographic signals recorded in vivo in humans to derive the most comprehensive framework to date of motor unit activity during isometric contractions that spanned the entire operating range of two human leg muscles. Our results address several key gaps in knowledge in this field and, importantly, characterise the activity of entire motor unit populations for each muscle.

Introduction:

Human muscles are versatile effectors producing forces that span several orders of magnitude. They allow humans to perform extremely diverse motor tasks with the same limbs, like a surgeon closing an incision, and a climber who grasps supports on a cliff. This versatility relies on a unique structure that converts neural inputs into muscle force; that is, the motor unit, which comprises an alpha motoneuron and the muscle fibres it innervates (1, 2). The nervous system controls muscle force by modulating the number of active motor units (recruitment) and their firing rates (rate coding) (3). This control strategy involves projecting a substantial level of common synaptic inputs to motoneurons (4-6) that recruits them in a fixed order following the size principle (7).

While we have a clear understanding of the general mechanisms of force control, we still lack a full picture of the detailed organisation of the firing activities of motor units. This is challenged by our inability, so far, to decode the concurrent firing activity of the full spectrum of motor units spanning the entire range of recruitment thresholds in human muscles (8). Indeed, much of our knowledge on the modulation of motor unit activity still derives from animal preparations (9-11) or from serial recordings of motor units in humans, mainly decoded over narrow force ranges (12-14).

Animal studies have shown two stages in the rate coding of individual motor units to increase muscle force (amplification and rate limiting) in response to linear increases in the net excitatory synaptic inputs (11, 15-17). These stages correspond to different responses of motoneurons to ionotropic inputs due to modulation of their intrinsic properties by metabotropic inputs acting on the somato-dendritic surfaces of motoneurons through several ion channels (2, 16, 17). Similar characteristics in firing activity have been observed in humans (12, 13, 18), although these observations have been limited to small samples of motor units (typically < 40) with firing activities decoded over a narrow range of submaximal forces (typically <30% maximal voluntary contraction (MVC) force) (12, 13, 18). It is therefore difficult to infer from these studies a control scheme that is generalisable to entire pool and to all contraction levels.

The combination of arrays of electromyographic (EMG) electrodes with modern source-separation algorithms set the path for the decoding of populations of active motor units - and their motoneurons - in humans (8, 19). Recent studies have identified the concurrent firing

activity of up to 60 motor units per contraction and per muscle by increasing the density of

electrodes (20, 21). Here, we further extended this approach by tracking motor units across

contractions at target forces that ranged from 10 to 80 %MVC, based on the unique spatial

distribution of motor unit action potentials across the array of surface electrodes (22, 23). We

were able to identify up to ~200 unique active motor units per muscle and per participant in two

human muscles *in vivo*, yielding extensive samples of motor units that are representative of the

entire motoneuron pools (24). With this approach, we described the non-linear transformation

of the net excitatory synaptic inputs into firing activity for individual motoneurons (2, 17). We

specifically focused on the non-linearities between the changes in firing rate and force:

amplification, saturation, and hysteresis (9, 17, 25). The results provided a framework picture

of the rate coding of motor units during large increases and decreases in an applied force.

Having access to the rate coding of entire motoneuron pools allowed us to understand how

ionotropic and neuromodulatory inputs combine to modulate force, confirming some details of

hypothetical force-control schemes in humans. For example, experimental (26, 27) and

computational (25) studies have proposed a gain-control mechanism driven by

neuromodulatory inputs to motoneurons. One common conclusion of these studies is that pools

of motor units involved in fine motor tasks have a low gain, whereas additional motor units

activated during more forceful tasks exhibit a progressively increase in gain (17, 25-27).

However, this conclusion is based on results from motor units in animals (16) and humans (26,

28, 29) while experimentally modulating inputs, or by fitting a non-linear model predicting

muscle activation from excitatory synaptic inputs at multiple contraction levels (27). In contrast,

we addressed this question by decoding the concurrent activity of hundreds of motor units

spanning the entire range of recruitment thresholds in vivo in humans.

Results:

<u>Identification and tracking of individual motor units:</u>

16 participants performed either isometric dorsiflexion (n = 8) or knee extension tasks (n = 8) while we recorded the EMG activity of the tibialis anterior (TA - dorsiflexion) or the vastus lateralis (VL – knee extension), with four arrays of 64 surface electrodes (256 electrodes per muscle). The experimental tasks consisted of isometric contractions involving a ramp-up, a plateau, and a ramp-down phase. The ramp-up and ramp-down phases were relatively slow compared with contractile speeds observed during activities of daily living. They were performed at a constant pace of 5 %MVC.s^{-1} and the force plateau was maintained for either 10-s (70-80 %MVC), 15-s (50-60 %MVC) or 20-s (10-40 %MVC) (Figure 1A). The level of force during the plateau ranged between 10 and 80 %MVC, with increments of 10 %MVC (randomised order). A source-separation algorithm (8, 19) was applied to the EMG signals to extract motor unit pulse trains, from which we automatically identified the series of discharge times. All identified motor unit firings were visually inspected and manually edited when necessary (30). Because the signal was stationary during the plateau, we were able to estimate reliable separation vectors for large samples of motor units with source-separation algorithms (8, 19) (Figure 1A). The average number of motor units identified in each contraction per participant was 42 ± 24 (25th–75th percentile: 24 – 53, up to 95) motor units from the TA and 33 ± 15 (25th–75th percentile: 23 – 47, up to 71) motor units from the VL. The datasets from all contraction levels were merged to obtain a sample of unique motor units per muscle and participant spanning the full range of recruitment thresholds (1st–99th percentile: 0.9-73.4% MVC; see below).

To estimate the proportion of the EMG signal represented by the identified motor units, a synthetic EMG signal was reconstructed from the firing activity. To do so, the discharge times were used as triggers to segment the differentiated EMG signals over a window of 25 ms and estimate the averaged action potential waveforms for each motor unit (Figure 1A). The action potentials were then convolved with the discharge times to obtain trains of action potentials, and all the trains of the identified motor units were summed to reconstruct the synthetic EMG signal. Finally, the ratio between the powers of synthetic and original EMG signals was calculated (Figure 1B). This ratio reached 69.3 ± 17.3% (25th–75th percentile: 59.3 – 83.6%, up to 94.2%) for TA and 55.2 ± 19.5% (25th–75th percentile: 50.0 – 71.9%, up to 86.5%) for VL

(Figure 1C), meaning that most of the recorded EMG signals were successfully decomposed into motor unit activity.

To investigate motor unit activity across large force ranges, motor units were tracked between contractions using their unique spatial distribution of action potentials (23) (Figure 1D). This method was validated beforehand using both simulated EMG signals and two-source validation with simultaneous recordings of firing activity from intramuscular and surface EMG signals (Figure S1). On average, we tracked $67.1 \pm 10.0\%$ (25th–75th percentile: $53.9 - 80.1\%$) of the motor units between consecutive contraction levels (10% increments, e.g., between 10% and 20% MVC) for TA and $57.2 \pm 5.1\%$ (25th–75th percentile: $46.6 - 68.3\%$) of the motor units for VL (Figure S2). There are two explanations for the inability to track all motor units across consecutive contraction levels. First, some motor units are recruited at higher targets only. Second, it is challenging to track small motor units beyond a few contraction levels due to signal cancellation (31, 32). In total, we identified 129 ± 44 motor units (25th–75th percentile: $100 - 164$, up to 194) per participant in TA and 130 ± 63 motor units (25th–75th percentile: $103 - 173$, up to 199) per participant in VL.

The accuracy of the tracking method was further tested by confirming the uniqueness of each motor unit within the identified sample, assuming that each motor unit must have a unique representation of their action potentials across the array of surface electrodes (22). This was accomplished by calculating the root mean-square error (RMSE) between their action potentials across the electrodes and the action potentials of the rest of the motor units (Figure 1E). The RMSE between the action potentials of the same motor unit tracked across contractions was calculated as a reference and compared with the 5th percentile of the distribution of RMSE between motor units (Figure 1F). This reference value typically less than the 5th percentile of the RMSE calculated with the action potentials of the other motor units (TA: 5.6 ± 2.4 vs. $8.0 \pm 1.5\%$; $p<0.001$; VL: $6.2 \pm 2.8\%$ vs. $8.5 \pm 1.6\%$; $p<0.001$). Inspection of data for each motor unit revealed that 92.1% (TA) and 87.0% (VL) of the motor units had a lower RMSE between contractions than with the other motor units ($<5^{th}$ percentile). Of note, motor units with the highest reference values ($>95^{th}$ percentile) were considered as outliers due to tracking errors and were excluded from the subsequent analyses (Figure 1F). As a result of this approach, a total of 34 motor units were excluded from TA and 28 from VL.

Figure 1. Identification of extensive samples of motor units in two human muscles. A. We used a blind source separation (BSS) algorithm to decompose the overlapping activity of motor units (MU) into trains of discharge times during a force-matching trapezoidal task. B. We reconstructed synthetic EMG signals by summing the trains of action potentials from all the identified motor units and interpreting the remaining EMG signal as the part of the signal not explained by the decomposition. C. We calculated the ratio between the powers of synthetic and original EMG signals to estimate the proportion of the signal variance explained by the decomposition. D. We estimated the uniqueness of each identified motor unit within the pool by calculating the root-mean-square error (RMSE) between the distributions of action potentials of the same motor unit across contractions (two panels on the left, reference value in E), and between motor units (left vs. right panels, distribution of RMSE between motor units in yellow in E). F. We considered each motor unit

to be unique within the pool when the RMSE between its distributions of action potentials across contractions

(reference value) was less than the 5th percentile of the distribution of RMSE with the rest of the motor units

across contraction levels. Motor units considered as outliers in F (red data points) were removed from the

analysis due to potential errors in tracking between contractions. Each data point is a motor unit, the box

represents 25th-75th percentiles of the distribution of data, and the black line is the median. The horizontal

thick line denotes a statistical difference between reference values and 5th percentiles for each muscle.

<u>Non-linear rate coding during the linear increase in force – amplification and rate limiting:</u>

The input-output function for each motoneuron was characterised as the relation between its

instantaneous firing rate and the muscle force (Figure 2A). The linear increase in isometric force

was assumed to reflect a proportional increase in the net synaptic excitatory inputs received by

the motoneurons, as proposed previously (13, 33). Thus, any deviation from a linear increase

in firing rate with respect to a linear increase in force should reflect the influence of

neuromodulatory inputs on motoneuron gain (13, 14, 18), or a saturation of the motoneuron

firing rate (12, 13). Therefore, the association between the firing rate and the force was

characterised by comparing three curve fits: i) a linear fit, which would indicate an increase in

firing rate proportional to the increase in the net synaptic excitatory input, ii) a rising

exponential fit, which would correspond to an initial acceleration of firing rates followed by

full saturation (12, 13, 34), and iii) a natural logarithm fit, which would denote an initial

acceleration of firing rate followed by a slower constant increase in firing rate that reflects a

rate limiting effect (11, 15, 35) (Figure 2A). Of note, this analysis was performed on each unique

motor unit with the data pooled across all contractions (Figure 2A). The result was that 69.5%

of the motor units in TA and 72.3% of those in VL had an input-output function better fitted by

a natural logarithm (Figure 2B, inset panels).

The input-output functions of the motor units were then compared based on the distribution of

recruitment thresholds (1st–99th percentile: 0.9-73.4% MVC). We clustered motor units into

three groups with equal ranges of recruitment thresholds: low- (0 to 25% MVC), medium- (25

to 50% MVC), and high- (50 to 75% MVC) threshold motor units (Figure 2B). A significant

percentage of input-output functions of low threshold motor units was better fitted with a rising

exponential function (12.5% for the TA, 30.9% for the VL), highlighting a steep acceleration

of firing rate followed by a full saturation. This pattern was not observed in medium- and high-

threshold motor units (0% for TA and VL). In contrast, a significant percentage of input-output

functions of high threshold motoneurons was better fitted with a linear function for TA (39.6%),

although the input-output functions of high threshold motor units from VL were still better fitted with natural logarithms (87.9%) than linear functions (12.1%).

We further described the first stage of rate coding by estimating the acceleration of firing rate (Figure 2C; Figure S3). The initial acceleration of the firing rate, likely due to the amplification of synaptic inputs by persistent inward currents (11), was calculated as the value of the first derivative of the natural logarithm function at the recruitment threshold (Figure 2C). These values were compared between motor units using a linear mixed effect model, with *Muscle* (TA; VL) and *Threshold* (low; medium; high) as fixed effects and *Participant* as a random effect. There was a significant effect of *Muscle* (F=30.4; p<0.001), *Threshold* (F=24.7; p<0.001), and a significant interaction *Muscle × Threshold* (F=4.1; p=0.017). The initial acceleration of firing rate in TA was greater for the low- (1.0 ± 0.7 pps^2; p<0.001) and the high- (1.0 ± 0.3 pps^2; p=0.032) threshold motor units than for medium-threshold motor units (0.6 ± 0.3 pps^2), with no difference between the low- and high-threshold motor units (p=0.999) (Figure 2C). Similarly, the initial acceleration of firing rate was greater for low-threshold motor units (0.6 ± 0.6 pps^2) than for medium- (0.4 ± 0.2 pps^2; p<0.001) and the high-threshold motor units (0.4 ± 0.2 pps^2; p=0.013) in VL, with no difference between the medium- and high-threshold motor units (p=0.990) (Figure 2C). The initial acceleration of firing rate was also greater for the low- and high-threshold motor units from TA comparing with VL (p<0.001 for both).

Finally, we tested whether the distribution of initial accelerations of firing rate observed across the entire motor unit pool was generalisable to all the participants. Participants with either less than 20 motor units or with no motor units recruited below 5%MVC were excluded from this analysis (TA: n=4; VL: n=3). The function representing the relation between recruitment thresholds and initial accelerations were accurately fitted using the non-linear least-square fitting method (TA; adjusted R-squared: 0.77; VL; adjusted R-squared: 0.80; green lines on Figure 2C). We then fitted the same non-linear functions for each participant; the adjusted R-squared were increased to 0.86 ± 0.04 for the TA and 0.84 ± 0.11 for the VL (inset panels in Figure 2C).

Figure 2. Non-linear rate coding of motor units. A. We analysed the relation between motor unit firing rate (pulses per second, pps) and force with the assumption that a linear increase in force is proportional to a linear increase in net synaptic input. For this, we concatenated the values of instantaneous firing rate for each motor unit (grey data points) recorded over all the contractions where that motor unit was identified, as shown here for one motor unit (coloured data points for contractions at 20, 40, and 60%MVC). The force-firing rate relations were then fitted with three different functions: linear (green), rising exponential (dark red), and natural logarithm (yellow), to characterise the input-output function of each motoneuron. B. The motor units were grouped according to their best fit. We display the distribution of motor unit recruitment thresholds (RT) for each of these groups. The inset panels depict the percentage of motor units (MU) in each group. C. We

further analysed the rate coding of motor units by reporting the acceleration of firing rate. For this, we fitted the force-firing rate relation with the natural logarithm (f(force) =a*ln(force)+b; yellow trace) and computed its first derivative (f(force)=a/force; dark red trace). The right panels depict the relations between the initial acceleration of motor unit firing rates and their recruitment threshold (RT) for all participants, with a total of 328 and 393 motor units for TA and VL, respectively. Each data point is a motor unit. The horizontal thick line denotes a statistical difference between motor units grouped according to their recruitment thresholds (low = Blue; medium = Grey; high = Pink). The green line depicts the non-linear fits of these relations for the Tibialis Anterior and the Vastus Lateralis. Similar fits were observed for all the participants (inset panels).

The second stage of rate coding involved a linear increase in firing rate after the initial acceleration phase. This phase was analysed using the average firing rate during the successive force plateaus for each tracked motor unit (Figure 3). Specifically, the increase in firing rate between consecutive contraction levels (i.e., change in force equal to 10% MVC) was compared with a linear mixed effect model with *Muscle* (TA; VL) and *Threshold* (low; medium; high) as fixed effects and *Participant* as a random effect. There were significant effects for *Muscle* (F=39.7; p<0.001) and *Threshold* (F=145.3; p<0.001), and a significant interaction *Muscle × Threshold* (F=37.7; p<0.001). The rate of increase was significantly greater for TA (3.5 ± 1.7 pps; 25th-75th percentile: 2.4 – 4.1 pps) than for VL (2.0 ± 1.1 pps; 25th-75th percentile: 1.3 – 2.6 pps. p<0.001). The rate of increase was significantly greater in TA for high-threshold than for low- (p<0.001) and medium- (p<0.001) threshold motor units and for medium- than for low-threshold motor units (p<0.001). In contrast, the rate of increase in VL was greater for high- (p<0.001) and medium- (p<0.001) threshold motor units than for low-threshold motor units, with no difference between high- and medium-threshold motor units in VL (p=0.993).

The data for each participant was characterised with a linear function (Figure 3C). There was a significant positive association between the rate of increase in firing rate and the recruitment threshold for all the participants (Pearson's r: TA: 0.71±0.14 pps and VL:0.57±0.22 pps; all p<0.022), except in one participant for each muscle (TA: r=0.17; p=0.272; VL: r= 0.12; p=0.411). Most motor units (92.0% in TA and 80.9% in VL) increased firing rates by more than one pulse per second between contractions despite reaching the maximal force tested or until tracking was not possible.

Figure 3. Motor unit firing rates across contraction levels. We calculated the average firing rate (pulses per second, pps) during the force plateaus for each tracked motor unit for all participants from TA (n = 998 motor units; A) and VL (n = 1016 motor units; B). Each data point is a motor unit, and each line connects the firing rates of this motor unit across contractions. The colour scale identifies the three groups of motor units based on recruitment threshold: low (blue), medium (grey), and high (pink). The central panels depict the change in firing rates between contractions separated by 10% to 70% of the maximal voluntary contraction level (MVC). The right panels show the relation between the rate of increase in firing rate between successive levels of force (e.g., between 10% and 20%MVC) and the recruitment threshold of the motor unit. We fitted these relations for each participant with a linear function (coloured lines). The horizontal thick line denotes a statistical difference between motor units grouped according to their recruitment thresholds.

Differences between ascending and descending levels of force - hysteresis:

Neuromodulatory inputs can prolong excitatory synaptic inputs and produce a hysteresis between the recruitment and the derecruitment thresholds. The left columns in Figure 4 show the relation between recruitment and derecruitment thresholds within the motor unit pools (Figure 4). The relation for most participants was better characterised with a non-linear than a linear regression (TA: 6 out of 8 participants, adjusted R-squared >0.92); VL: 7 out of 8 participants, adjusted R-squared >0.83), indicating potential differences in the level of

hysteresis between motor units of different size. The difference between the recruitement and the derecruitment threshold was compared with a linear mixed effect model with *Muscle* (TA; VL) and *Threshold* (low; medium; high) as fixed effects and *Participant* as a random effect. There was a significant effect of *Threshold* (F=83.0; p<0.001) and a significant interaction *Muscle × Threshold* (F=30.1; p<0.001), but no effect of *Muscle* (F=2.5; p<0.137). We further tested for each group of motor units whether the difference between the two thresholds was significantly different from 0 (absence of hysteresis). Thus, only medium-threshold motor units had a significant positive hysteresis in both muscles (TA: -3.8 ± 7.2 pps; p = 0.013; VL: -3.1 ± 6.0 pps; p = 0.026), whereas low-threshold motor units from both muscles (TA: -1.5 ± 4.8; VL: -1.0 ± 3.2 pps; p>0.221) did not exhibit significant hysteresis. Although high-threshold motor units from TA followed the same trend (-1.7 ± 6.0 pps; p<0.437), high-threshold motor units from VL exhibited a negative hysteresis (VL: +3.2 ± 6.7 pps; p=0.039), that indicated that derecruitment threshold was greater than the recruitment threshold.

We also compared the relation between motor unit firing rate and force between the ramp-up and ramp-down phases of the isometric contraction. Most motor units (TA: 60.2%; VL: 76.7%) kept the same relation during the two phases. Although the percentage of associations better fitted with linear functions increased for the ramp-down phase for the TA (from 25.4% to 41.9% of motor units), there was no change between ramps for the VL (from 12.8% to 19.4% of motor units). Thus, most motor units kept a non-linear relation during the ramp-down phase.

Figure 4. Hysteresis between recruitment and derecruitment thresholds. The left panels depict the relations between the recruitment and derecruitment thresholds of each motor unit from TA (A) and VL (B) across all participants. Each grey data point is a motor unit. These relations were fitted for each participant (coloured lines) using either non-linear or linear regressions. The values below the dashed red line (recruitment threshold = derecruitment threshold) show a positive hysteresis between recruitment and derecruitment thresholds, the values above a negative hysteresis. The right panels show the difference between the recruitment and derecruitment thresholds, with negative values showing a positive hysteresis with recruitment threshold greater than derecruitment threshold and the positive values indicating the converse (a negative hysteresis). Each data point is a motor unit. The asterisk denotes a statistical difference between the hysteresis values for motor units grouped according to recruitment threshold (low = Blue; medium = Grey; high = Pink) and the absence of a hysteresis (dashed horizontal line).

Discussion:

No previous studies have decoded to date the firing activity of >100 motor units spanning the full spectrum of recruitment thresholds, whether in animals or humans. According to previous estimates, the number of motor units in human would be 200±61 for TA and 146±29 for VL (36). We have therefore identified in this study most of the active motor units for each muscle (Figure 1), which has allowed us to infer the rate coding of entire pools of motor units. Overall, we found that motor units within a pool exhibit distinct rate coding with changes in force level (Figure 2 and 3), which contrasts with the long-held belief that rate coding is similar across motor units from the same pool.

The fast initial acceleration phase reflects the amplification of synaptic inputs through the activation of persistent inward currents via voltage-gated sodium and calcium channels (2, 11, 15, 17). Although noradrenergic and serotonergic synapses that generate persistent inward currents are widespread within motor unit pools (37), the influence of the neuromodulatory inputs is greater during this phase for low- than for high-threshold motor units. This is presumably due to a larger ratio between the amplitude of inward currents and the input conductance in low threshold motoneurons (38).

The second phase of rate coding involves a slower linear increase in firing rate, as characterised by the second part of a logarithmic function (Figures 2-3). This phase corresponds to mechanisms previously referred to as 'rate limiting' (2, 25, 39) or 'saturation' (12, 34), and was more pronounced for low- than high-threshold motor units (Figure 3). This difference may be explained by smaller excitatory synaptic inputs onto low- than high-threshold motoneurons (2, 16), lower synaptic driving potential of the dendritic membrane (12, 40, 41), and longer and larger afterhyperpolarisation phase in low- than high-threshold motoneurons (42-45).

Taken together, these results show how ionotropic and neuromodulatory inputs to motoneurons uniquely combine to generate distinct rate coding across the pool, following a similar pattern across the pool, but with a pace that depends on motor unit size. While the size of the motor unit determines its recruitment threshold, neuromodulatory inputs determine its gain. Thus, in a similar fashion as size imposes a fixed recruitment order (7), neuromodulatory inputs determines a spectrum of variations in motor unit firing rates without the need to differentiate the ionotropic inputs to each motoneuron. (Figure 2C-3-S3) (25, 46, 47). Because high-

threshold motor units exhibit a higher gain than low-threshold motor units (Figure 3), their
firing rate could eventually reach similar or even greater values than that of low-threshold motor
units during strong contractions (47-50). This indicates that the onion skin principle (34, 51)
may not hold in all muscles and for all contraction forces (47-50).

It is also worth noting that the profile of rate coding mostly followed the same natural
logarithmic trend during the ramp-up and ramp-down phases of the contraction, but with
prolonged firing activity only evident for medium-threshold motor units (Figure 4). This
hysteresis is possible because of the bistability of the membrane potential mediated by inward
currents from calcium channels, the two stable membrane states being the resting potential and
a depolarised state that enables self-sustained firing (9, 52). The absence of statistically
significant hysteresis in low threshold motoneurons may be simply explained by their early
recruitment during the first percentages of force, mathematically lowering the potential
amplitude of their hysteresis. Alternatively, prominent outward currents may attenuate the
impact of persistent inward currents on the prolongation of their firing activity (53). Similarly,
the absence of hysteresis (TA) and even a negative hysteresis (VL) in high-threshold
motoneurons can be explained by a briefer membrane bistability due to a faster decay of inward
currents from calcium channels, and a narrower range of membrane depolarisation over which
these inward currents are activated (52).

The increase in firing rate was also significantly greater for TA motor units than for those in
VL. This difference may reflect a varying balance between excitatory/inhibitory synaptic inputs
and neuromodulation (14, 25, 39, 46, 47, 54). Specifically, the strength of recurrent and
reciprocal inhibitory inputs to motoneurons innervating VL and TA, and their proportional or
inverse covariation with excitatory inputs, respectively, may explain the differences in rate
limiting and maximal firing rates (14, 25, 39, 46, 47, 54). It is worth noting that similar
differences in rate coding were previously observed between proximal and distal muscles of the
upper limb (55).

Our results on rate coding characteristics of the entire motor unit pool provide insight on force
control strategies. The accurate control of force depends on the firing activity of the population
of active motor units, which effectively filters out the synaptic noise of individual motor units
and primarily transforms common synaptic inputs into muscle force (4, 5, 56). Because the
bandwidth of muscle force is <10 Hz during isometric contractions (57), the activation of

inward currents to low-threshold motoneurons at recruitment may enable them to promptly

discharge at a rate (>8 pps) that transmits the effective common synaptic inputs (<8-10 Hz)

without phase distortion (4, 5, 56). This mechanism facilitates the accuracy of force control.

Moreover, force generation can be described as the filtering of the cumulative firing activity of

active motor units with the average twitch force of their muscle units (5, 58). From this

perspective, at low forces, recruitment of new motor units has a higher impact on force

modulation than rate coding since each recruitment impacts both the cumulative firing activity

of the pool and the average twitch force. Rate coding only modulates the cumulative firing

activity. Thus, the amplification of the firing rate of low-threshold motor units near their

recruitment threshold instantly favours rate coding over the recruitment of additional motor

units, which likely allows for smoother force control. Similarly, the progressive recruitment of

motor units with a higher gain promotes faster changes in firing rates, which promotes force

control accuracy across the full force range. Overall, our results may help to design future non-

linear decoders that aim to predict muscle activation or muscle force from descending inputs

recorded in the motor cortex, with the will to generalise their performance across movements

(27).

Methods:

Detailed materials and methods are provided in supplementary files. Data and code are provided
at https://github.com/simonavrillon/MUdictionnary.

References:

1. C. S. Sherrington, Remarks on some aspects of reflex inhibition. *Proc Biol Sci* **97**, 519-545 (1925).
2. C. J. Heckman, R. M. Enoka, Motor unit. *Compr Physiol* **2**, 2629-2682 (2012).
3. R. M. Enoka, J. Duchateau, Rate Coding and the Control of Muscle Force. *Cold Spring Harb Perspect Med* **7** (2017).
4. D. Farina, F. Negro, J. L. Dideriksen, The effective neural drive to muscles is the common synaptic input to motor neurons. *J Physiol* **592**, 3427-3441 (2014).
5. D. Farina, F. Negro, Common synaptic input to motor neurons, motor unit synchronization, and force control. *Exerc Sport Sci Rev* **43**, 23-33 (2015).
6. F. Negro, U. S. Yavuz, D. Farina, The human motor neuron pools receive a dominant slow-varying common synaptic input. *J Physiol* **594**, 5491-5505 (2016).
7. E. Henneman, Relation between size of neurons and their susceptibility to discharge. *Science* **126**, 1345-1347 (1957).
8. D. Farina, A. Holobar, Characterization of Human Motor Units From Surface EMG Decomposition. *Proceedings of the Ieee* **104**, 353-373 (2016).
9. J. Hounsgaard, H. Hultborn, B. Jespersen, O. Kiehn, Bistability of alpha-motoneurones in the decerebrate cat and in the acute spinal cat after intravenous 5-hydroxytryptophan. *J Physiol* **405**, 345-367 (1988).
10. D. J. Bennett, H. Hultborn, B. Fedirchuk, M. Gorassini, Short-term plasticity in hindlimb motoneurons of decerebrate cats. *J Neurophysiol* **80**, 2038-2045 (1998).
11. R. H. Lee, C. J. Heckman, Adjustable amplification of synaptic input in the dendrites of spinal motoneurons in vivo. *J Neurosci* **20**, 6734-6740 (2000).
12. A. J. Fuglevand, R. A. Lester, R. K. Johns, Distinguishing intrinsic from extrinsic factors underlying firing rate saturation in human motor units. *J Neurophysiol* **113**, 1310-1322 (2015).
13. A. L. Revill, A. J. Fuglevand, Inhibition linearizes firing rate responses in human motor units: implications for the role of persistent inward currents. *J Physiol* **595**, 179-191 (2017).
14. M. D. Johnson, C. K. Thompson, V. M. Tysseling, R. K. Powers, C. J. Heckman, The potential for understanding the synaptic organization of human motor commands via the firing patterns of motoneurons. *J Neurophysiol* **118**, 520-531 (2017).
15. D. J. Bennett, H. Hultborn, B. Fedirchuk, M. Gorassini, Synaptic activation of plateaus in hindlimb motoneurons of decerebrate cats. *J Neurophysiol* **80**, 2023-2037 (1998).
16. R. K. Powers, M. D. Binder, Input-output functions of mammalian motoneurons. *Rev Physiol Biochem Pharmacol* **143**, 137-263 (2001).
17. M. D. Binder, R. K. Powers, C. J. Heckman, Nonlinear Input-Output Functions of Motoneurons. *Physiology (Bethesda)* **35**, 31-39 (2020).
18. J. A. Beauchamp *et al.*, A geometric approach to quantifying the neuromodulatory effects of persistent inward currents on individual motor unit discharge patterns. *J Neural Eng* **20** (2023).
19. F. Negro, S. Muceli, A. M. Castronovo, A. Holobar, D. Farina, Multi-channel intramuscular and surface EMG decomposition by convolutive blind source separation. *J Neural Eng* **13**, 026027 (2016).
20. S. Muceli *et al.*, Blind identification of the spinal cord output in humans with high-density electrode arrays implanted in muscles. *Science advances* **8**, eabo5040 (2022).
21. A. H. Caillet *et al.*, Larger and Denser: An Optimal Design for Surface Grids of EMG Electrodes to Identify Greater and More Representative Samples of Motor Units. *eNeuro* **10** (2023).

498   22.   D. Farina, F. Negro, M. Gazzoni, R. M. Enoka, Detecting the unique representation of
499   motor-unit action potentials in the surface electromyogram. *J Neurophysiol* **100**, 1223-
500   1233 (2008).
501   23.   E. Martinez-Valdes *et al.*, Tracking motor units longitudinally across experimental
502   sessions with high-density surface electromyography. *J Physiol* **595**, 1479-1496 (2017).
503   24.   A. H. Caillet, A. T. M. Phillips, D. Farina, L. Modenese, Motoneuron-driven
504   computational muscle modelling with motor unit resolution and subject-specific
505   musculoskeletal anatomy. *bioRxiv* 10.1101/2023.06.03.543552,
506   2023.2006.2003.543552 (2023).
507   25.   R. K. Powers, C. J. Heckman, Synaptic control of the shape of the motoneuron pool
508   input-output function. *J Neurophysiol* **117**, 1171-1184 (2017).
509   26.   K. Wei *et al.*, Serotonin affects movement gain control in the spinal cord. *J Neurosci* **34**,
510   12690-12700 (2014).
511   27.   S. Naufel, J. I. Glaser, K. P. Kording, E. J. Perreault, L. E. Miller, A muscle-activity-
512   dependent gain between motor cortex and EMG. *J Neurophysiol* **121**, 61-73 (2019).
513   28.   B. I. Goodlich, S. A. Horan, J. J. Kavanagh, Blockade of 5-HT(2) receptors suppresses
514   rate of torque development and motor unit discharge rate during rapid contractions. *J
515   Neurophysiol* **127**, 150-160 (2022).
516   29.   T. T. Henderson, J. L. Taylor, J. R. Thorstensen, M. G. Tucker, J. J. Kavanagh, Enhanced
517   availability of serotonin limits muscle activation during high-intensity, but not low-
518   intensity, fatiguing contractions. *J Neurophysiol* **128**, 751-762 (2022).
519   30.   S. Avrillon, F. Hug, C. Gibbs, D. Farina, Tutorial on MUedit: An open-source software
520   for identifying and analysing the discharge timing of motor units from
521   electromyographic signals. *bioRxiv* 10.1101/2023.07.13.548568,
522   2023.2007.2013.548568 (2023).
523   31.   K. G. Keenan, D. Farina, K. S. Maluf, R. Merletti, R. M. Enoka, Influence of amplitude
524   cancellation on the simulated surface electromyogram. *J Appl Physiol (1985)* **98**, 120-
525   131 (2005).
526   32.   D. Farina, R. Merletti, R. M. Enoka, The extraction of neural strategies from the surface
527   EMG: an update. *J Appl Physiol (1985)* **117**, 1215-1230 (2014).
528   33.   A. J. Fuglevand, D. A. Winter, A. E. Patla, Models of recruitment and rate coding
529   organization in motor-unit pools. *J Neurophysiol* **70**, 2470-2488 (1993).
530   34.   C. J. De Luca, P. Contessa, Hierarchical control of motor units in voluntary contractions.
531   *J Neurophysiol* **107**, 178-195 (2012).
532   35.   R. K. Powers, S. M. Elbasiouny, W. Z. Rymer, C. J. Heckman, Contribution of intrinsic
533   properties and synaptic inputs to motoneuron discharge patterns: a simulation study. *J
534   Neurophysiol* **107**, 808-823 (2012).
535   36.   J. Duchateau, R. M. Enoka, Distribution of motor unit properties across human muscles.
536   *J Appl Physiol (1985)* **132**, 1-13 (2022).
537   37.   R. Maratta, K. K. Fenrich, E. Zhao, M. S. Neuber-Hess, P. K. Rose, Distribution and
538   density of contacts from noradrenergic and serotonergic boutons on the dendrites of
539   neck flexor motoneurons in the adult cat. *J Comp Neurol* **523**, 1701-1716 (2015).
540   38.   S. Huh *et al.*, PICs in motoneurons do not scale with the size of the animal: a possible
541   mechanism for faster speed of muscle contraction in smaller species. *J Neurophysiol*
542   **118**, 93-102 (2017).
543   39.   C. J. Heckman, M. D. Binder, Computer simulations of the effects of different synaptic
544   input systems on motor unit recruitment. *J Neurophysiol* **70**, 1827-1840 (1993).
545   40.   S. Cushing, T. Bui, P. K. Rose, Effect of nonlinear summation of synaptic currents on
546   the input-output properties of spinal motoneurons. *J Neurophysiol* **94**, 3465-3478
547   (2005).

548	41.	R. K. Powers, M. D. Binder, Summation of effective synaptic currents and firing rate modulation in cat spinal motoneurons. *J Neurophysiol* **83**, 483-500 (2000).

41.   R. K. Powers, M. D. Binder, Summation of effective synaptic currents and firing rate modulation in cat spinal motoneurons. *J Neurophysiol* **83**, 483-500 (2000).

42.   R. Bakels, D. Kernell, Matching between motoneurone and muscle unit properties in rat medial gastrocnemius. *J Physiol* **463**, 307-324 (1993).

43.   P. F. Gardiner, Physiological properties of motoneurons innervating different muscle unit types in rat gastrocnemius. *J Neurophysiol* **69**, 1160-1170 (1993).

44.   A. S. Deardorff *et al.*, Expression of postsynaptic Ca2+-activated K+ (SK) channels at C-bouton synapses in mammalian lumbar -motoneurons. *J Physiol* **591**, 875-897 (2013).

45.   A. H. Caillet, A. T. M. Phillips, D. Farina, L. Modenese, Mathematical relationships between spinal motoneuron properties. *Elife* **11** (2022).

46.   M. K. Chardon *et al.*, Supercomputers and Reverse Engineering of Motoneuron Firing Patterns. *bioRxiv* 10.1101/2022.12.09.519818, 2022.2012.2009.519818 (2023).

47.   J. Škarabot, J. A. Beauchamp, G. E. P. Pearcey, Human motor unit discharge patterns reveal differences in neuromodulatory and inhibitory drive to motoneurons across contraction levels. *bioRxiv* 10.1101/2023.10.16.562612, 2023.2010.2016.562612 (2023).

48.   A. Gydikov, D. Kosarov, Some features of different motor units in human biceps brachii. *Pflugers Arch* **347**, 75-88 (1974).

49.   C. T. Moritz, B. K. Barry, M. A. Pascoe, R. M. Enoka, Discharge rate variability influences the variation in force fluctuations across the working range of a hand muscle. *J Neurophysiol* **93**, 2449-2459 (2005).

50.   T. Oya, S. Riek, A. G. Cresswell, Recruitment and rate coding organisation for soleus motor units across entire range of voluntary isometric plantar flexions. *J Physiol* **587**, 4737-4748 (2009).

51.   C. J. De Luca, P. Contessa, Biomechanical benefits of the Onion-Skin motor unit control scheme. *J Biomech* **48**, 195-203 (2015).

52.   R. H. Lee, C. J. Heckman, Bistability in spinal motoneurons in vivo: systematic variations in persistent inward currents. *J Neurophysiol* **80**, 583-593 (1998).

53.   R. K. Powers, C. J. Heckman, Contribution of intrinsic motoneuron properties to discharge hysteresis and its estimation based on paired motor unit recordings: a simulation study. *J Neurophysiol* **114**, 184-198 (2015).

54.   C. J. Heckman, A. S. Hyngstrom, M. D. Johnson, Active properties of motoneurone dendrites: diffuse descending neuromodulation, focused local inhibition. *J Physiol* **586**, 1225-1231 (2008).

55.   C. J. De Luca, R. S. LeFever, M. P. McCue, A. P. Xenakis, Behaviour of human motor units in different muscles during linearly varying contractions. *J Physiol* **329**, 113-128 (1982).

56.   J. L. Dideriksen, F. Negro, R. M. Enoka, D. Farina, Motor unit recruitment strategies and muscle properties determine the influence of synaptic noise on force steadiness. *J Neurophysiol* **107**, 3357-3369 (2012).

57.   R. M. Enoka, D. Farina, Force Steadiness: From Motor Units to Voluntary Actions. *Physiology (Bethesda)* **36**, 114-130 (2021).

58.   C. K. Thompson *et al.*, Robust and accurate decoding of motoneuron behaviour and prediction of the resulting force output. *J Physiol* **596**, 2643-2659 (2018).