1 **Original Article**

2 **Flower size evolution in the Southwest Pacific**

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Abstract

Background and Aims. Despite accelerating interest in island evolution, the general evolutionary trajectories of island flowers remain poorly understood. In particular the island rule, which posits that small organisms become larger and large organisms to become smaller after island colonization, while tested in various plant traits, has never been tested in flower size. Here, we provide the first test for the island rule in flower size for animal- and wind-pollinated flowers, and the first evidence for generalized *in-situ* evolution of flower size on islands.

Methods. Focusing on 10 archipelagos in the Southwest Pacific, we amassed a dataset comprising 131 independent colonization events, by pairing each island endemic to its closest mainland relative. We then tested for the island rule in flower size and for gigantism/dwarfism in floral display for animal- and wind-pollinated flowers.

Key results. Animal-pollinated flowers followed the island rule, while wind-pollinated flowers did not, instead showing evidence of gigantism. Results remained consistent after controlling for breeding system, mainland source pool, degree of taxonomic differentiation, taxonomic family, and island type.

Conclusions. While *in situ* evolution of flower size is widespread on islands in the Southwest Pacific, animal- and wind- pollinated flowers exhibited unexpected and markedly different evolutionary trajectories. Further studies are needed to understand the mechanisms behind these patterns.

Keywords

Island rule; flower size; island biogeography; plant evolution; Southwest Pacific

Introduction

Island flowers often differ from their mainland relatives, being typically dull-coloured and inconspicuous (Whittaker et al., 2023). They are often self-compatible and possess generalized morphologies that allow pollination by a wide array of pollinators (Baker, 1967; Carlquist, 1974; Barrett, 1996; Crawford et al., 2011; Burns, 2019). These differences suggest the existence of an island floral syndrome (i.e. a set of predictable differences between island and mainland flowers) which is still largely unexplored (Whittaker et al., 2023).

The island floral syndrome can arise from evolutionary processes and/or from insular assembly rules (i.e. patterns arising from biased colonization/establishment) (*sensu* Whittaker et al., 2023). At the moment, most components of the island floral syndrome are regarded as insular assembly rules (but see Hetherington-Rauth and Johnson, 2020). For example, plants capable of self-fertilization are overrepresented on islands as they can establish new populations more readily (Baker, 1967; Pannel et al., 2015; Burns, 2019). In turn, self-compatible plants typically have smaller flowers (Goodwillie et al., 2010). Whether components of the island floral syndrome can arise from in-situ evolutionary processes is still unclear (Bernardello et al., 2001; Newstrom and Robertson, 2005; Burns, 2019).

Directional evolutionary processes in island species traits can result either in overall trends toward gigantism or dwarfism (Burns et al., 2012; Kavanagh and Burns, 2014), or in the 'island rule', a pattern in which, after island colonization, small organisms become larger and large organisms to become smaller. While it remains one of the most controversial topics in biogeography (Van Valen, 1973; Lomolino, 2005; Meiri et al., 2008; Lomolino et al., 2012; Faurby and Svenning, 2016; Itescu et al., 2018; Rebouças et al., 2018; Lokatis and Jeschke, 2018; Benitez-Lopez et al., 2021), evidence for the island rule has recently been found for plant stature and leaf area on islands in the Southwest Pacific (Biddick et al., 2019). Yet, whether other plant traits conform to an island-rule-like pattern remains unclear.

Islands are known for their paucity of pollinators (Burns, 2019; Whittaker et al.,

2023). Changes in pollinator abundance between islands and the mainland could generate

directional selective pressures in floral display (i.e. flower size · number of flowers per

individual), causing animal-pollinated floral displays to increase/decrease and wind-

pollinated floral displays to remain unchanged (Parachnowitsch and Kessler, 2010; Caruso et

al., 2019; Brunet et al., 2021). Pollen limitation could intensify selection for traits promoting

outcrossing through increased floral display, potentially leading to larger flowers (Harder et

al., 2010; Thomann et al., 2013; Panique and Caruso, 2020). Alternatively, pollinator decline

and mate limitation could select for selfing mechanisms, leading to smaller floral displays

and smaller flowers in animal-pollinated species (Elle and Carney, 2003; Thomann et al.,

2013).

Flower size of both animal- and wind-pollinated species could alternatively follow the

island rule as a by-product of selection acting on flower-correlated traits. In particular, leaf

area and seed size are known to covary with sepal and corolla size and were shown to evolve

predictably on islands (Niklas, 1994; Burns et al., 2012; Biddick et al., 2019; E-Vojtkó et al.,

2022; Whittaker et al., 2023). As such, flower size might not be under direct selection, but

follow the island rule because of its allometric relationships with leaf area and seed size.

Under this hypothesis, both animal- and wind-pollinated island flowers should differ from

their mainland relatives, and these patterns should mirror those of leaf area and seed size.

Different plant lineages can respond independently to insular selective pressures. For

example, the genetic, morphological and evolutionary history of certain taxonomic families

make them more likely to evolve generalized floral structures or to break down mechanisms

of self-incompatibility (Goldberg et al., 2010; Minelli, 2018; Cossard et al., 2021). Similarly,

plants with different breeding systems are typically characterized by different flowers (Krizek

and Anderson, 2013; Salisbury et al., 2017) and might respond to insular conditions in a

distinct way. Additionally, evolutionary pressures will vary depending on the abiotic and

biotic characteristics of the islands themselves (MacArthur and Wilson, 1967; Whittaker et

al., 2023). Consequently, to test for general patterns of flower size evolution on islands, it is

vital to include multiple plant lineages with different pollination modes, evolutionary

histories and breeding systems that occur across multiple archipelagos.

In this study, we used a phylogenetic comparative approach to test whether island

animal-pollinated and wind-pollinated flowers evolve toward gigantism/dwarfism or follow

the island rule. Specifically, using a newly derived dataset representing 131 independent

colonization events on 10 archipelagos in the Southwest Pacific, we asked two questions: I)

Does floral display of animal-pollinated species increase/decrease, while floral display of

wind-pollinated species remain unchanged? II) Do both animal-pollinated and wind-

pollinated flowers evolve to follow the island rule, and does this pattern mirror those of leaf

area and seed size?

Methods

Study area

We focused on the endemic flora of 10 archipelagos distributed across 25° of latitude (29°04'

S - 54°37' S) around New Zealand (Fig 1). The island groups can roughly be divided into the

sub-tropical archipelagos of Lord Howe, Norfolk, Kermadec, and Three Kings, and the cool-

temperate/subantarctic archipelagos of Chatham, Snares, Antipodes, Auckland, Campbell and

Macquarie. All island systems are oceanic in origin, with faunas and floras derived by

overwater dispersal from New Zealand and Australia or vicariance (Chown et al., 1998;

Rutledge et al., 2003; Pendlebury and Barnes-Keoghan, 2007; Lord, 2015; Fitzgerald, 2020).

We restricted analyses to these island systems so that our set of island taxa would be

derived from a few well-known mainland floras, sharing similar ecological, geological and

evolutionary histories. Chatham, Snares, Antipodes, Auckland, Campbell, Three Kings,

Kermadec and Macquarie were included as most of their endemic species originated from the

New Zealand mainland. Lord Howe and Norfolk islands were included as most of their

endemic species originated from Australia. We included endemics from 2 mainland sources

to increase the number of taxonomic families present and to account for the possibility that

endemics originating from very different mainland source pools (i.e. New Zealand and

Australia) would respond differently to insular environments.

Species selection

To test flower size evolution in animal- and wind-pollinated species, we compiled a list of

island/mainland pairs of taxa. Each mainland taxon is the sister group of the island taxon so

that each pair represents a phylogenetically independent comparison.

We first compiled a list of endemic species and subspecies on each archipelago using

the most recent data available (Oliver, 1948; Godley, 1969; Taylor, 1971; Johnson and

Campbell, 1975; Meurk, 1977; Sykes, 1977; Godley, 1989; Wright, 1983; Green, 1994;

Meurk et al., 1994; Sykes and West, 1996; Amey et al., 2007; De Lange et al., 2011; Lord et

al., 2013; De Lange, 2014; De Salas and Baker, 2015). Because botanical sources varied

widely in age, all species names were cross-checked for synonymy, keeping the New Zealand

Plant Conservation Network (2023) and the Atlas of Living Australia (2023) as taxonomic

references.

We then identified the mainland sister taxon of each island taxon using published

molecular phylogenies. We searched Google Scholar for phylogenies using the species or

genus name and the Boolean operator "AND" along the term "phylogen*" (e.g. *Coprosma*

AND phylogen*). When multiple phylogenies were available, we selected the phylogeny that

used the most comprehensive genetic dataset, used either Bayesian inference or Maximum

likelihood for phylogenetic reconstruction, and showed the strongest support at each node

(Supplementary 1). Species names were standardized following the New Zealand Plant

Conservation Network (2023) and Atlas of Living Australia (2023). When molecular

phylogenies were unavailable, sister taxa were identified based on morphological evidence.

As without phylogenies we could not identify the closest relative with certainty, multiple

mainland species within the genus were selected and their floral traits averaged. If the genus

comprised less than five species on the mainland (e.g. genus *Streblus*), all species were

included as mainland relatives. If the genus comprised more than five species on the

mainland (e.g. genus *Exocarpos*), we included as mainland relatives up to five species based

on morphological affinities. (Supplementary 2). We excluded island taxa if mainland taxa

could not be identified using either phylogeny or taxonomy.

Trait measurements

Each island/mainland pair was categorized as animal-pollinated or wind-pollinated, and as

monomorphic (including hermaphroditic, monoecious, andromonoecious, and

gynomonoecious species) or dimorphic (including dioecious and gynodioecious species)

using McGlone and Richardson (2023), Atlas of Living Australia (2023), Flora of New South

Wales Online (2023) and the New Zealand Plant Conservation Network (2023).

To test our first hypothesis, floral display was defined as flower number · flower size.

For flower number, we counted the number of flowers per individual. When unavailable, we

used the number of flowers per inflorescence as an estimate of floral display. This can be a

reliable estimate as inflorescences typically behave as autonomous modules among which

resources cannot be reallocated (Herrera, 1991; Torices and Mendez, 2014; Dai et al., 2018).

To test our first and second hypotheses, flower size of animal-pollinated flowers was

measured using the unit of pollinator attraction (Faegri and Van der Pijl, 1979), i.e. the floral

trait bearing the primary attractiveness function (e.g. petals), so that each trait had a

comparable biological function across families. For radially symmetric flowers with fused

petals and petal lobes (e.g. Plantaginaceae, genus *Veronica*), we characterized lobe length,

lobe width, or corolla diameter. Tube length was collected but not included in the analysis as

it is not a measure of advertisement but of accessibility (Dafni, 1992). For radially symmetric

flowers with free petals, we characterized petal length, petal width or corolla diameter. For

zygomorphic flowers, we used the length and width of the organs most involved in

advertisement (e.g. banner and wing of Fabaceae flowers). For the Myrtaceae family, we

characterized both stamen length and petal length, as in this family the stamens are often

involved in pollinator attraction (Vasconcelos et al., 2019). For Asteraceae, we collected ray

floret length, disk diameter or capitulum diameter. For dimorphic taxa, we always used

measurements from male flowers, as we expect the male perianth to be the one under stronger

selective pressures (Barbot et al., 2023).

Flower size of wind-pollinated species was measured the sterile parts of flowers, as

these traits are known to correlate with leaf area and seed size (Burns et al., 2012; Biddick et

al., 2019; E-Vojtkó et al., 2022). For radially symmetric flowers with fused petals (e.g.

Plantaginaceae, genus *Plantago*), we characterized lobe length, lobe width, or corolla

diameter. For radially symmetric flowers with free petals (e.g. Arecaceae) we characterized

petal length or corolla diameter. For flowers with undifferentiated sepals and petals (e.g.

Juncaceae), we used tepal length. For Poaceae, we used glume, lemma, palea or lodicule

length. For Cyperaceae, we used glume length. For dimorphic taxa (e.g. Rubiaceae, genus

Coprosma) we characterized both male and female flowers.

To test whether the evolutionary trajectory of island flowers mirrors the patterns of

their putatively correlated traits, we measured leaf area and seed dry mass for all species.

These two traits were selected because, among the studied traits of island plants, they are the

ones most clearly under direct selective pressures (Kavanagh and Burns, 2014; Biddick et al.,

2019; Whittaker et al., 2023; Ciarle et al., 2024). Leaf shape was approximated as an ellipse

so that leaf area was calculated from leaf length and width using the equation:

$$A = \frac{Ll}{2} \times \frac{Lw}{2} \times \pi$$

where A is the estimated leaf area, Ll is leaf length and Lw is leaf width. Seed mass was

estimated from seed length using the equation:

$$m = 0.1329sl^{2.2243}$$

where m is the estimated seed mass, sl is the seed length and the two numeric values are

constants. The equation was parametrized based on a linear model with both variables log-

transformed, a sample size of 695 species with directly measured seed mass (74% of which

are native species) across 297 genera and 100 families, and a fit of R squared = 0.865 (Sarah

Richardson, personal communication, May 20, 2020).

All traits were extracted from the Floras of New Zealand and Australia series (Allan,

1961; Moore and Edgar, 1970; Webb et al., 1988; Edgar et al., 2010) and additional online

sources (New Zealand Plant Conservation Network, 2023; Atlas of Living Australia, 2023;

and New South Wales Flora Online, 2023). We searched the primary literature when data

were missing. When online data was unavailable, we manually measured floral traits from

herbaria in the Botany collection of Te Papa Tongarewa Museum of New Zealand. For each

species trait, 5 to 10 specimens with multiple, well-preserved flowers were sampled. Three to

five measurements were then taken for each specimen and averaged. When possible, to take

into account temporal and spatial variability, specimens of different ages and from different

regions of New Zealand were selected for each species.

Statistical analyses

In this study we used two types of trait data: herbaria measurements and data retrieved from

other sources. To test for significant differences between data types, we randomly selected 30

species for which data was available online and measured them in the herbarium collection.

These data were then compared using a Wilcoxon signed-rank test.

For all analyses, we used the same trait for all species within a family (e.g. capitulum

diameter for all Asteraceae). When a trait value was missing in a species, we used another

trait in its place. To make sure this did not alter the results significantly, we tested whether

the replacing trait correlated with the original one for all species of that family by using a

Pearson's correlation test (Berry and Feldman, 1985). A cut-off value of 0.7 was chosen, and

only traits above the cut-off value were considered as possible replacements (Table S4).

To test whether floral display (i.e. flower size · number of flowers per inflorescence)

of island species increased/decreased in size, we computed a floral display log ratio (fdLR)

for each island/mainland pair of animal-pollinated plants as log(mean island floral

display/mean mainland floral display). We then used a one-sided t-test to test whether fdLR

was significantly above or below zero.

To test whether flower size of both animal- and wind-pollinated species followed the

island rule, we computed a flower size log ratio (fsLR) as log(mean island flower size/mean

mainland flower size). We then conducted two major axis regressions using fsLR as the

dependent variable and log(mean mainland flower size) as the independent variable. Slopes

significantly lower than zero would be evidence for the island rule. Slopes indistinguishable

from zero would denote morphological isometry (Lomolino et al., 2013). Intercepts above or

below zero would be evidence for gigantism or dwarfism respectively (Burns 2022). To avoid

having the independent variable also as part of the dependent variable, the analysis was

repeated without the fsLR ratio, with log(mean island flower size) as the dependent variable

and log(mean mainland flower size) as the independent variable. Slopes significantly lower

than one would be evidence for the island rule, while slopes indistinguishable from one

would denote morphological isometry.

To test whether the evolutionary trajectories of flower size mirror the patterns of leaf

area and seed mass, we computed a leaf area log ratio (laLR) as log(mean island leaf

area/mean mainland leaf area), and a seed mas log ratio (smLR) as log(mean island seed

mass/mean mainland seed mass). We then calculated Pearson's correlations among fsLR,

laLR and smLR for animal- and wind-pollinated species.

Mixed effect models were used to control for potential confounding factors. Because

flower size may differ between plants with different breeding strategies, the breeding system

was included as a fixed factor with two categories: monomorphic and dimorphic. Because

flowers from different mainlands may respond differently to insular systems, mainland source

was included as a fixed factor with two categories: Australia and New Zealand. Because we

expect fully differentiated species to exhibit more pronounced insular trends compared to

partially differentiated species (i.e. subspecies), taxonomic differentiation was included as a

fixed effect with two categories: partially differentiated and fully differentiated. To control

for phylogenetic conservatism, taxonomic families were included as a random effect. To

avoid singularity, only taxonomic families with three or more island/mainland pairs were

included. To control for environmental and ecological differences among islands, islands

were included as a random effect. Since flower size may differ between different flower

morphologies of animal-pollinated species, flower type was included as a fixed effect with

three categories: actinomorphic flowers with fused petals; actinomorphic flowers with free

petals, and zigomorphic flowers (Supplementary 2).

All analyses were conducted in the R environment version 4.3.1 (R Core Team,

2022). Reduced major axis regressions were conducted using the R package *lmodel2*

(Legendre and Oksanen, 2018). Mixed effect models were conducted using the R packages

nlme (Pinhero et al., 2017) and lme4 (Bates et al., 2009). When used as the independent

variable, mainland flower size was log-transformed to avoid heteroskedasticity. *ggplot2* was

used for data visualization (Wickham et al., 2016).

Results

We identified 155 endemic island species. From these, we could compile island/mainland

pairs for 149 species belonging to 43 families and representing 131 phylogenetically

independent colonization events. We identified 91 animal-pollinated pairs and 40 wind-

pollinated pairs. Data for 104 pairs was gathered from online sources, while data for the

remaining 27 pairs was manually measured from herbaria specimens (Supplementary 1).

Results of the Wilcoxon signed-rank test indicated no significant differences between

herbarium measurements and data obtained from other sources ($V = 177.5$, p-value $= 0.39$).

For 5 families, we retrieved data for multiple floral traits. Pearson's correlation revealed all

within-family traits to strongly correlate (Pearson's correlation coefficient > 0.7,

Supplementary 3, Table S1).

Floral display in animal-pollinated species exhibited no trend toward gigantism or

dwarfism ($n = 75$, LR mean $= -0.011$, $t = -0.36$, p-value $= 0.72$) (Fig 2a). Similarly, floral

display in wind-pollinated species exhibited no trend toward gigantism or dwarfism ($n = 34$,

LR mean $= 0.023$, $t = 0.44$, p-value $= 0.66$) (Fig 2b). Flower number alone exhibited no trend

as well (Supplementary 3, Table S2).

Flowers of animal-pollinated plants did not get larger ($n = 87$, LR mean $= -0.005$, $t = $

0.15, p-value $= 0.82$) but followed the island rule, with a slope lower than zero ($s = -6.87$,

95% confidence interval $= -19.78, -3.82$; p-value < 0.01) (Fig 2b). Instead, flowers of wind-

pollinated species evolved toward gigantism ($n = 39$, LR mean $= 0.060$, $t = 3.28$, p-value $= $

0.002) and did not follow the island rule ($s = -0.01$, 95% confidence interval $= -0.13, 0.10$; p-

value $= 0.81$) (Fig 2c).

These patterns did not mirror those of leaf area and seed mass. fsLR and laLR did not

correlate for animal- (cor = 0.18, p-value = 0.21) and wind-pollinated species (cor = -0.01, p-

value = 0.90). Similarly, fsLR and smLR did not correlate for animal- (cor = 0.16, p-value =

0.22) and wind-pollinated species (cor = 0.23, p-value = 0.22).

Results did not vary when mean island flower size (log-transformed) was used as the

dependent variable instead of fsLR. Results remained robust after controlling for breeding

system, mainland source, degree of taxonomic differentiation, phylogenetic conservatism (i.e.

taxonomic families), and islands (Supplementary 3, Table S3). In animal-pollinated species,

fsLR did not vary significantly among mainland sources, taxonomic families, and degrees of

taxonomic differentiation, but did so among breeding systems and islands (Supplementary 3,

Table S4). In wind-pollinated species, fsLR did not vary significantly between breeding

system, mainland source, degree of taxonomic differentiation, taxonomic families, and

islands (Supplementary 3, Table S4).

Discussion

We provided the first evidence for generalized *in-situ* evolution of flower size on islands and

the first test for the island rule in flower size. Different pollination modes followed different

evolutionary trajectories. We found no evidence for increased/decreased floral display in

animal-pollinated species. Instead, flower size of animal-pollinated species followed the

island rule. Conversely, flower size of wind-pollinated species became consistently larger on

islands. However, neither of these patterns mirrored those of leaf area and seed mass.

The floral display of animal-pollinated species exhibited no recognizable pattern,

becoming neither larger nor smaller. Resources allocated to flowers need to increase

reproductive success, which is pollinator-driven. Floral displays are also subjected to non-

pollinator selective pressures such as predators and environmental stresses, which determine

flower maintenance costs. If pollinators decline, but reproductive success still increases at a

higher rate than maintenance costs, selection will favour increased pollinator attraction. Only

when pollinators become too scarce and/or mating partners are limited will selection favour

individuals that rely on abiotic vectors/self-fertilization (Maciel et al., 2020; Roddy et al.,

2021). Consequently, depending on the biotic and abiotic factors of islands, pollinator paucity

could result in showy flowers, inconspicuous flowers, or neither. Our results indicate that, if

pollinators do decline on the islands of the Southwest Pacific, differences in pollinator

pressures between islands and the mainland might be negligible and/or countered by

differences in flower maintenance costs.

Flower size of animal-pollinated flowers followed the island rule. Flower size is

known to correlate with other plant traits, including leaf area (Niklas, 1994; Burns et al.,

2012; Bawa et al., 2019; Roddy et al., 2021; E☐Vojtkó et al., 2022). However, results

indicate that flower size of island species evolved independently of leaf area and seed mass.

Thus, it is unlikely that selection acting on these traits caused correlated evolution in flower

size. Biddick and Burns (2021) showed how a null model only accounting for evolutionary

drift can provide a parsimonious explanation for the island rule. As a mechanistic explanation

for why some plant traits obey the island rule and others do not is still lacking (Biddick et al.,

2019; Whittaker et al., 2023), it could be beneficial to look outside mechanisms of selection

to explain why flower size of animal-pollinated species follows the island rule.

Results for animal-pollinated species varied significantly between breeding systems

(Figure S1). Globally, pollinator selection acts more strongly on dimorphic species (Geber et

al., 2012). Larger flowers in males of dimorphic animal-pollinated island plants may

therefore reflect evolution in a depauperate environment where every possibility of attracting

pollinators needs to be exploited (Godley, 1979). Additionally, differences in breeding

systems in animal- and wind-pollinated species might be explained by considering that, while

females allocate more to reproduction in dimorphic animal-pollinated plants, in wind-

pollinated species the reproductive costs of males may exceed or match those of females,

potentially causing plants with different pollination modes to follow different evolutionary

trajectories (Harris and Pannell, 2008; Liu et al., 2021; Roddy et al., 2021; but see Midgley,

2022).

Flower size of wind-pollinated species did not follow the island rule but evolved to be

consistently larger on islands. As with animal-pollinated species, results indicate that flower

size evolved independently of leaf area and seed mass. Why wind-pollinated flowers would

evolve toward gigantism on islands is thus puzzling.

First, wind-pollinated plants typically benefit from producing many small, uniovulate,

inexpensive flowers. This is because numerous flowers can sample more of the airstream and

maximize pollen capture and seed production (Sakai and Sakai, 1995; Friedman and Barrett,

2011). According to this model, producing larger, more expensive flowers would reduce

maternal fitness. Larger flowers could be selected only if island plants possessed more

resources available for flower and seed production than their mainland relatives.

Second, seed gigantism is a hallmark of New Zealand's outlying islands (Kavanagh

and Burns, 2014; Biddick et al., 2019) and larger seeds were shown to promote smaller

flowers. Larger seeds can lead to increased within-fruit sibling competition. One of the

strategies adopted by plant species to counter sibling competition is to reduce the number of

seeds per fruit (Bawa, 2016). While this potentially reduces fitness, it also reduces flower and

pollinator investment, allowing plants to produce small flowers that are wind-pollinated or

that rely on generalist pollinators (Bawa et al., 2019). In other words, as seed size increases,

flower size of wind-pollinated plants should decline, which is not what we observe. Again,

perhaps insular wind-pollinated plants allocate resources differently than their mainland

counterparts, promoting consistent positive flower/seed correlations. The reason behind it

though remains unclear.

Unclear is also why animal- and wind-pollinated species should exhibit different

island syndromes. Differences in pollination agents may cause the two groups to follow

different evolutionary trajectories. But beyond pollination mode, the two groups seem rather

heterogeneous. For example, they both contain animal- and wind-dispersed species spanning

multiple plant families, with several families featuring in both groups. It is therefore unclear

why traits performing functions unrelated to pollination (e.g. seed dispersal) should evolve

according to pollinator-selected traits. This difference may just be an idiosyncrasy of our

system. Further studies should be conducted on other island systems to test this possibility.

Burns (2019) proposed that flower size may follow the island rule because of flower-

pollinators size-coupling. Considering that size-coupling is an important aspect of plant-

pollinator mutualism in specialized flowers (Dalsgaard et al. 2009; Maglianesi et al. 2014;

Biddick and Burn, 2018), a reduction in pollinators' diversity and size range could select for

reduced flower diversity, causing specialized flowers to follow the island rule (Hiraiwa and

Ushimaru, 2017; Burns, 2019). While intriguing, we were not able to test this hypothesis as

most of our species possessed open, radial, generalist flowers that would likely not comply

with size changes in pollinators (Faure et al., 2022). As pollinator specialists are rare on

islands (Olesen et al., 2002; Traveset et al., 2016; Wang et al., 2020), testing this hypothesis

would likely require a global dataset.

Our analysis encompasses only a portion of the Southwest Pacific. First, the study

does not include any tropical islands. Ecologically, tropical systems might operate differently

than non-tropical ones. For instance, while pollination networks seem to be structurally

similar in tropical and non-tropical areas (Vizentin-Bugoni et al., 2018), many plant traits

follow a latitudinal gradient, including flower morphological diversity (Currie and Paquin,

1987; Chen et al., 2017; Chartier et al., 2021). Additionally, tropical systems are

underrepresented in the literature (Culumber et al., 2019). Secondly, our study focuses on

small, isolated archipelagos with relatively small species pools. As the ecological and

evolutionary dynamics of islands vary markedly with area and isolation (MacArthur and

Wilson, 1967; Whittaker et al., 2023), these results may not apply to larger and less isolated

archipelagos.

In this study, we accounted for phylogenetic relatedness through paired taxonomic

comparisons and by adding taxonomic family as a random effect in our mixed models. To

make sure that flower size evolution was widespread among flowering plants we included

multiple families and two mainland source pools. However, accounting for relatedness using

a systematic approach, such as a phylogenetic generalized least square model, would have

likely yielded more accurate results. This was not possible as a well-resolved phylogeny for

the investigated species is not currently available.

In this study, we focused on many island/mainland pairs of species, spanning several

families. However, many of the families included in the analysis were only represented by

one or a few species, making it difficult to evaluate within family trends. Moreover, focusing

on island/mainland pairs implies that we did not account for intraspecific variance. It is

possible that accounting for such variance would alter the observed patterns. Finally, while

our study focused on flower size, future research should aim for a multi-trait approach

capable of encompassing the plant island syndrome in its entirety.

In conclusion, we provided the first test for the island rule in flowers, and the first

evidence for generalized *in-situ* evolution of flower size on islands. Results indicate that,

while *in situ* evolution of flower size is widespread on islands in the Southwest Pacific,

species belonging to different pollination modes exhibit markedly different evolutionary

trajectories. These results combine with a growing body of literature investigating the island

syndrome in plants (Burns, 2019; Schrader et al., 2021; Delavaux et al., 2021; Meudt et al.,

2021; Zizka et al., 2022; Whittaker et al., 2023; Ciarle, 2024), and provide a much-needed

insight into the evolution of plants on islands.

References

Allan, H. H. (1961). *Flora of New Zealand.* New Zealand Government Printer Publications.

Amey, J., Lord, J. M., & de Lange, P. (2007). First record of a vascular plant from the Bounty

Islands: *Lepidium oleraceum* (nau, Cook's scurvy grass) (Brassicaceae). *New Zealand*

Journal of Botany, 45(1), 87–90.

Atlas of Living Australia. (2023). Atlas of living Australia. https://www.ala.org.au/

Baker, H. G. (1967). Support for Baker's law-as a rule. *Evolution*, *21*(4), 853-856.

Barbot, E., Dufaÿ, M., & De Cauwer, I. (2023). Sex-specific selection patterns in a dioecious

insect-pollinated plant. *Evolution*, 77(7), 1578–1590.

Barrett, S. C. H. (1996). The reproductive biology and genetics of island

plants. *Philosophical Transactions of the Royal Society of London. Series B:*

Biological Sciences, *351*(1341), 725-733.

Bates, D., Maechler, M., Bolker, B., Walker, S., Christensen, R. H. B., Singmann, H., Dai,

B., Scheipl, F., Grothendieck, G, Fox, J., Bauer, A., Krivitsky. P. N., Tanaka, E., &

Green, P. (2009). Package 'lme4'. *URL http://lme4. r-forge. r-project. org*.

Bawa, K. S. (2016). Kin selection and the evolution of plant reproductive traits. *Proceedings*

of the Royal Society B: Biological Sciences, *283*(1842), 20160789.

Bawa, K. S., Ingty, T., Revell, L. J., & Shivaprakash, K. N. (2019). Correlated evolution of

flower size and seed number in flowering plants (monocotyledons). *Annals of*

Botany, *123*(1), 181-190.

Benítez-López, A., Santini, L., Gallego-Zamorano, J., Milá, B., Walkden, P., Huijbregts, M.

A., & Tobias, J. A. (2021). The island rule explains consistent patterns of body size

evolution in terrestrial vertebrates. *Nature Ecology & Evolution*, *5*(6), 768-786.

Bernardello, G., Anderson, G. J., Stuessy, T. F., & Crawford, D. J. (2001). A survey of floral

traits, breeding systems, floral visitors, and pollination systems of the angiosperms of

the Juan Fernández Islands (Chile). *The Botanical Review*, 67, 255-308.

Berry, W. D., & Feldman, S. (1985). Multiple regression in practice. *SAGE publications*.

Biddick, M., & Burns, K. C. (2018). Phenotypic trait matching predicts the topology of an

insular plant–bird pollination network. *Integrative Zoology*, *13*(3), 339-347.

Biddick, M., Hendriks, A., & Burns, K. C. (2019). Plants obey (and disobey) the island

rule. *Proceedings of the National Academy of Sciences*, 116(36), 17632-17634.

Biddick, M., & Burns, K. C. (2021). A simple null model predicts the island rule. *Ecology*

Letters, *24*(8), 1646-1654.

Brunet, J., Flick, A. J., & Bauer, A. A. (2021). Phenotypic selection on flower color and

floral display size by three bee species. *Frontiers in plant science*, *11*, 587528.

Burns, K. C., Herold, N., & Wallace, B. (2012). Evolutionary size changes in plants of the

south☐west Pacific. *Global Ecology and Biogeography*, *21*(8), 819-828.

Burns, K. C. (2019). *Evolution in isolation: the search for an island syndrome in plants*.

Cambridge University Press.

Burns, K. C. (2022). The paradox of island evolution. *Journal of Biogeography*, *49*(2), 248-

253.

Carlquist S. (1974). *Island biology*. Columbia University Press.

Caruso, C. M., Eisen, K. E., Martin, R. A., & Sletvold, N. (2019). A meta-analysis of the

agents of selection on floral traits. *Evolution*, *73*(1), 4-14.

Chartier, M., von Balthazar, M., Sontag, S., Löfstrand, S., Palme, T., Jabbour, F., Sauquet,

H., & Schönenberger, J. (2021). Global patterns and a latitudinal gradient of flower

disparity: perspectives from the angiosperm order Ericales. *New Phytologist*, *230*(2),

821-831.

Chen, S. C., Cornwell, W. K., Zhang, H. X., & Moles, A. T. (2017). Plants show more flesh

in the tropics: Variation in fruit type along latitudinal and climatic

gradients. *Ecography*, *40*(4), 531-538.

Chown, S. L., Gremmen, N. J. M., & Gaston, K. J. (1998). Ecological biogeography of

Southern Ocean islands: Species-area relationships, human impacts, and conservation.

The American Naturalist, 152(4), 562–575.

Ciarle, R. (2024). Trait correlation and the assembly of island plant communities: Evidence

from the Southwest Pacific. *Journal of Vegetation Science*, 35(4), e13291.

Ciarle, R., Burns, K. C., & Mologni, F. (2024). The Loss (and Gain) of Defensive

Adaptations in Island Plants and Animals: A Comparative Review. In *Ecology and*

Evolution of Plant-Herbivore Interactions on Islands, 69-93. Springer.

Cossard, G. G., Gerchen, J. F., Li, X., Cuenot, Y., & Pannell, J. R. (2021). The rapid

dissolution of dioecy by experimental evolution. *Current Biology*, 31(6), 1277-1283.

Crawford, D. J., Anderson, G. J., & Bernardello, G. (2011). The reproductive biology of

island plants. In *The biology of island floras*, 11-36.

Culumber, Z. W., Anaya-Rojas, J. M., Booker, W. W., Hooks, A. P., Lange, E. C., Pluer, B.,

Ramírez-Bullón, N., & Travis, J. (2019). Widespread biases in ecological and

evolutionary studies. *BioScience*, *69*(8), 631-640.

Currie, D. J., & Paquin, V. (1987). Large-scale biogeographical patterns of species richness

of trees. *Nature*, *329*(6137), 326-327.

Dafni, A. (1992). *Pollination ecology: a practical approach*. Oxford University Press.

Dai, C., Luo, W. J., Gong, Y. B., Liu, F., & Wang, Z. X. (2018). Resource reallocation

patterns within Sagittaria trifolia inflorescences following differential

pollination. *American Journal of Botany*, *105*(4), 803-811.

Dalsgaard, B., Martín González, A. M., Olesen, J. M., Ollerton, J., Timmermann, A.,

Andersen, L. H., & Tossas, A. G. (2009). Plant–hummingbird interactions in the West

Indies: floral specialisation gradients associated with environment and hummingbird

size. *Oecologia*, *159*, 757-766.

Delavaux, C. S., Weigelt, P., Dawson, W., Essl, F., van Kleunen, M., König, C., Pergl. J.,

Pysek. P., Stein. A., Winter. M., Taylor. A., Schultz T. A., Whittaker. R. J., Kreft. H.,

& Bever, J. D. (2021). Mycorrhizal types influence island biogeography of

plants. *Communications Biology*, *4*(1), 1128.

De Lange, P. J., Heenan, P. B., & Rolfe, J. R. (2011). *Checklist of vascular plants recorded*

from Chatham Islands. Department of Conservation, Wellington Hawke's Bay

Conservancy.

De Lange, P. J. (2014). The flora of Egeria Rock, Northern Kermadec Island Group.

Wellington Botanical Society Bulletin, 55, 41–49.

De Salas, M. F., & Baker, M. L. (2015). *A census of the vascular plants of Tasmania,*

including Macquarie Island. Tasmanian Herbarium, Tasmanian Museum and Art

Gallery.

E☐Vojtkó, A., Junker, R. R., de Bello, F., & Götzenberger, L. (2022). Floral and

reproductive traits are an independent dimension within the plant economic spectrum

of temperate central Europe. *New Phytologist*, *236*(5), 1964-1975.

Edgar, E., Connor, H. E., Sykes, W. R., & Dawson, M. I. (2010). Flora of New Zealand:

Gramineae. With Contributions by W. R. Sykes and M. I. Dawson: [Illustrations by S.

B. Malcolm, K. R. West and P. A. Brooke, with Photographs by P. N. Johnson].

Manaaki Whenua Press

Elle, E., & Carney, R. (2003). Reproductive assurance varies with flower size in Collinsia

parviflora (Scrophulariaceae). *American Journal of Botany*, *90*(6), 888-896.

Faegri, K., & Van der Pijl, L. (1979). A short history of the study of pollination ecology. *The*

Principles of pollination ecology, 1-77.

Faurby, S., & Svenning, J. C. (2016). Resurrection of the island rule: human-driven

extinctions have obscured a basic evolutionary pattern. *The American*

Naturalist, *187*(6), 812-820.

Faure, J., Martén-Rodríguez, S., Clark, J. L., & Joly, S. (2022). The level of pollination

specialization affects the relationship between the shape of flowers and the bills of

their hummingbird pollinators in Antillean Gesneriaceae. *International journal of*

plant sciences, *183*(3), 193-204.

Fitzgerald, N. B. (2020). *Vegetation change on subantarctic Macquarie Island (Doctoral*

dissertation). University of Tasmania.

Friedman, J., & Barrett, S. C. (2011). The evolution of ovule number and flower size in wind-

pollinated plants. *The American Naturalist*, *177*(2), 246-257.

Geber, M. A., Dawson, T. E., & Delph, L. F. (Eds.). (2012). *Gender and sexual dimorphism*

in flowering plants. Springer Science & Business Media.

Godley, E. J. (1969). Additions and corrections to the flora of the Auckland and Campbell

Islands. *New Zealand Journal of Botany*, 7(4), 336–348.

Godley, E. J. (1979). Flower biology in New Zealand. *New Zealand journal of botany*, *17*(4),

441-466.

Godley, E. J. (1989). The flora of Antipodes Island. *New Zealand Journal of Botany*, 27(4),

531–564.

Goldberg, E. E., Kohn, J. R., Lande, R., Robertson, K. A., Smith, S. A., & Igić, B. (2010).

Species selection maintains self-incompatibility. *Science*, *330*(6003), 493-495.

Goodwillie, C., Sargent, R. D., Eckert, C. G., Elle, E., Geber, M. A., Johnston, M. O., Kalisz,

S., Moeller, D. A., Ree, R. H., Vallejo-Marin, M., & Winn, A. A. (2010). Correlated

evolution of mating system and floral display traits in flowering plants and its

implications for the distribution of mating system variation. *New Phytologist*, *185*(1),

311-321.

Green, P. S. (1994). *Flora of Australia, volume 49, Oceanic Islands 1*. Australian

Government Printing Service.

Harder, L. D., & Johnson, S. D. (2005). Adaptive plasticity of floral display size in animal-

pollinated plants. *Proceedings of the Royal Society B: Biological Sciences*, *272*(1581),

2651-2657.

Harris, M. S., & Pannell, J. R. (2008). Roots, shoots and reproduction: sexual dimorphism in

size and costs of reproductive allocation in an annual herb. *Proceedings of the Royal*

Society B: Biological Sciences, *275*(1651), 2595-2602.

Herrera, J. (1991). Allocation of reproductive resources within and among inflorescences of

Lavandula stoechas (Lamiaceae). *American Journal of Botany*, *78*(6), 789-794.

Hetherington-Rauth, M. C., & Johnson, M. T. (2020). Floral trait evolution of angiosperms

on Pacific Islands. *The American Naturalist*, *196*(1), 87-100.

Hiraiwa, M. K., & Ushimaru, A. (2017). Low functional diversity promotes niche changes in

natural island pollinator communities. *Proceedings of the Royal Society B: Biological*

Sciences, *284*(1846), 20162218.

Itescu, Y., Schwarz, R., Donihue, C. M., Slavenko, A., Roussos, S. A., Sagonas, K., Valakos.

E. D., Foufopoulos, J., Pafilis, P., & Meiri, S. (2018). Inconsistent patterns of body

size evolution in co☐occurring island reptiles. *Global Ecology and*

Biogeography, *27*(5), 538-550.

Johnson, P. N., & Campbell, D. J. (1975). Vascular plants of the Auckland Islands. *New*

Zealand Journal of Botany, 13(4), 665–720.

Kavanagh, P. H., & Burns, K. C. (2014). The repeated evolution of large seeds on

islands. *Proceedings of the Royal Society B: Biological Sciences*, *281*(1786),

20140675.

Krizek, B. A., & Anderson, J. T. (2013). Control of flower size. *Journal of experimental*

botany, *64*(6), 1427-1437.

Legendre, P., & Oksanen, M. J. (2018). Package 'lmodel2'. *See https://CRAN. R-project.*

org/package= lmodel2.

Liu, M., Korpelainen, H., & Li, C. (2021). Sexual differences and sex ratios of dioecious

plants under stressful environments. *Journal of Plant Ecology*, *14*(5), 920-933.

Lokatis, S., & Jeschke, J. M. (2018). The island rule: an assessment of biases and research

trends. *Journal of Biogeography*, *45*(2), 289-303.

Lomolino, M. V. (2005). Body size evolution in insular vertebrates: generality of the island

rule. *Journal of biogeography*, *32*(10), 1683-1699.

Lomolino, M. V., Sax, D. F., Palombo, M. R., & van der Geer, A. A. (2012). Of mice and

mammoths: evaluations of causal explanations for body size evolution in insular

mammals. *Journal of Biogeography*, *39*(5), 842-854.

Lomolino, M. V., van der Geer, A. A., Lyras, G. A., Palombo, M. R., Sax, D. F., & Rozzi, R.

(2013). Of mice and mammoths: generality and antiquity of the island rule. *Journal of*

Biogeography, *40*(8), 1427-1439.

Lord, J. M., Huggins, L., Little, L. M., & Tomlinson, V. R. (2013). Floral biology and flower visitors on subantarctic Campbell Island. *New Zealand Journal of Botany*, *51*(3), 168-180.

Lord, J. M. (2015). Patterns in floral traits and plant breeding systems on Southern Ocean Islands. AoB Plants, 7, plv095.

MacArthur, R. H., & Wilson, E. O. (1967). *The theory of island biogeography*. Princeton University Press.

Maciel, A. A., Cardoso, J. C., & Oliveira, P. E. (2020). On the low reproductive success of two Cyrtopodium species (Orchidaceae: Cyrtopodiinae): The relative roles of biotic and abiotic pollination. *Plant species biology*, *35*(1), 49-58.

Maglianesi, M. A., Blüthgen, N., Böhning-Gaese, K., & Schleuning, M. (2014). Morphological traits determine specialization and resource use in plant–hummingbird networks in the neotropics. *Ecology*, *95*(12), 3325-3334.

McGlone, M. S., & Richardson, S. J. (2023). Sexual systems in the New Zealand angiosperm flora. *New Zealand Journal of Botany*, *61*(4), 201-231.

Meiri, S., Cooper, N., & Purvis, A. (2008). The island rule: made to be broken? *Proceedings of the Royal Society B: Biological Sciences*, *275*(1631), 141-148.

Meudt, H. M., Albach, D. C., Tanentzap, A. J., Igea, J., Newmarch, S. C., Brandt, A. J., Lee. W. G., & Tate, J. A. (2021). Polyploidy on islands: its emergence and importance for diversification. *Frontiers in Plant Science*, 12, 637214.

Meurk, C. D. (1977). Alien plants in Campbell Island's changing vegetation. *Mauri Ora*, 5, 93–118.

Meurk, C. D., Foggo, M. N., & Wilson, J. B. (1994). The vegetation of subantarctic Campbell Island. *New Zealand Journal of Ecology,* 18(2), 123–168.

Midgley, J. J. (2022). The costs of reproduction in plants cannot differ between the

sexes. *Journal of Plant Ecology*, *15*(6), 1308-1311.

Minelli, A. (2018). *Plant evolutionary developmental biology: the evolvability of the*

phenotype. Cambridge University Press.

Moore, L. B., & Edgar, E. (1970). Flora of New Zealand. In Flora of New Zealand. Volume

2. *Indigenous Tracheophyta. Monocotyledones except Gramineae*. Manaaki Whenua

Press.

Newstrom, L., & Robertson, A. (2005). Progress in understanding pollination systems in New

Zealand. *New Zealand journal of botany*, *43*(1), 1-59.

New Zealand Plant Conservation Network. (2023). New Zealand Plant Conservation

Network. https://www.nzpcn.org.nz/

NSW Flora Online. (2023). NSW Flora Online.

https://plantnet.rbgsyd.nsw.gov.au/floraonline.htm

Niklas, K. J. (1994). *Plant allometry: the scaling of form and process*. University of Chicago

Press.

Olesen, J. M., Eskildsen, L. I., & Venkatasamy, S. (2002). Invasion of pollination networks

on oceanic islands: importance of invader complexes and endemic super

generalists. *Diversity and distributions*, *8*(3), 181-192.

Oliver, W. R. B. (1948). The flora of the Three Kings Islands. *Records of the Auckland*

Institute and Museum, 3(4/5), 211–238.

Panique, H., & Caruso, C. M. (2020). Simulated pollinator declines intensify selection on

floral traits that facilitate selfing and outcrossing in Impatiens capensis. *American*

Journal of Botany, *107*(1), 148-154.

Pannell, J. R., Auld, J. R., Brandvain, Y., Burd, M., Busch, J. W., Cheptou, P. O., Conner. J.

K., Goldberg. E. E., Grant. A., Grossenbacher D. L., Hovick. S. M., Igic. B., Kalisz.

S., Petanidou. T., Randle. A. M., Rubio de Casas, R., Pauw. A., Vamosi. J. C., &

Winn, A. A. (2015). The scope of Baker's law. *New Phytologist*, *208*(3), 656-667.

Parachnowitsch, A. L., & Kessler, A. (2010). Pollinators exert natural selection on flower

size and floral display in Penstemon digitalis. *New Phytologist*, *188*(2), 393-402.

Pendlebury, S., & Barnes-Keoghan, I. P. (2007). Climate and climate change in the sub-

Antarctic. *Papers and Proceedings of the Royal Society of Tasmania,* 141(1), 67–81.

Pinheiro, J., Bates, D., DebRoy, S., Sarkar, D., Heisterkamp, S., Van Willigen, B., &

Maintainer, R. (2017). Package 'nlme'. *Linear and nonlinear mixed effects models,*

version, *3*(1), 274.

R Core Team (2022). R: A language and environment for statistical

computing. R Foundation for Statistical Computing, Vienna, Austria.

URL https://www.R-project.org/.

Rebouças, R., da Silva, H. R., & Solé, M. (2018). Frog size on continental islands of the coast

of Rio de Janeiro and the generality of the Island Rule. *PLoS One*, *13*(1), e0190153.

Roddy, A. B., Martínez☐Perez, C., Teixido, A. L., Cornelissen, T. G., Olson, M. E., Oliveira,

R. S., & Silveira, F. A. (2021). Towards the flower economics spectrum. *New*

Phytologist, *229*(2), 665-672.

Rutledge, D. T., Wilson, G. W., & Whenua, M. (2003). Climate parameters for New

Zealand's Offshore Islands. *Landcare Research Contract Report*.

Sakai, S., & Sakai, A. (1995). Flower size-dependent variation in seed size: theory and a

test. *The American Naturalist*, *145*(6), 918-934.

Salisbury, P. A., Fripp, Y. J., Gurung, A. M., & Williams, W. M. (2017). Is floral structure a

reliable indicator of breeding system in the Brassicaceae? *Plos one*, *12*(3), e0174176.

Schrader, J., Wright, I. J., Kreft, H., & Westoby, M. (2021). A roadmap to plant functional

island biogeography. *Biological Reviews*, *96*(6), 2851-2870.

Sykes, W. R. (1977). Kermadec Islands flora: an annotated check list. *Manaaki Whenua*

Press.

Sykes, W. R., & West, C. J. (1996). New records and other information on the vascular flora

of the Kermadec Islands. *New Zealand Journal of Botany*, 34(4), 447–462.

Taylor, R. H. (1971). Influence of man on vegetation and wildlife of Enderby and Rose

Islands, Auckland Islands. *New Zealand Journal of Botany*, 9(2), 225–268.

Thomann, M., Imbert, E., Devaux, C., & Cheptou, P. O. (2013). Flowering plants under

global pollinator decline. *Trends in plant science*, *18*(7), 353-359.

Torices, R., & Méndez, M. (2014). Resource allocation to inflorescence components is highly

integrated despite differences between allocation currencies and sites. *International*

Journal of Plant Sciences, *175*(6), 713-723.

Traveset, A., Tur, C., Trøjelsgaard, K., Heleno, R., Castro⬜Urgal, R., & Olesen, J. M.

(2016). Global patterns of mainland and insular pollination networks. *Global Ecology*

and Biogeography, *25*(7), 880-890.

Van Valen, L. (1973). Patterns and the balance of nature. *Evolutionary Theory*, *1*, 31-49.

Vasconcelos, T. N., Prenner, G., & Lucas, E. J. (2019). A systematic overview of the floral

diversity in Myrteae (Myrtaceae). *Systematic Botany*, *44*(3), 570-591.

Vizentin-Bugoni, J., Maruyama, P. K., de Souza, C. S., Ollerton, J., Rech, A. R., & Sazima,

M. (2018). Plant-pollinator networks in the tropics: a review. In *Ecological networks*

in the tropics: an integrative overview of species interactions from some of the most

species-rich habitats on earth, pp 73-91. Springer Nature.

Wang, X., Wen, M., Qian, X., Pei, N., & Zhang, D. (2020). Plants are visited by more

pollinator species than pollination syndromes predicted in an oceanic island

community. *Scientific reports*, *10*(1), 13918.

Webb, C. J., Sykes, W. R., & Garnock-Jones, P. J. (1988). Flora of New Zealand. Volume

IV: *Naturalised pteridophytes, gymosperms, dicotyledons*. Botany Division, D.S.I.R.

Whittaker, R. J., Fernández-Palacios, J. M., & Matthews, T. J. (2023). *Island Biogeography:*

Geo-environmental Dynamics, Ecology, Evolution, Human Impact, and Conservation.

Oxford University Press.

Wickham, H., Chang, W., & Wickham, M. H. (2016). Package 'ggplot2'. *Create elegant data*

visualisations using the grammar of graphics. Version, *2*(1), 1-189.

Wright, A. E. (1983). Conservation status of the Three Kings Islands endemic flora in 1982.

Records of the Auckland Institute and Museum, 20, 175–184.

Zizka, A., Onstein, R. E., Rozzi, R., Weigelt, P., Kreft, H., Steinbauer, M. J., Bruelheide. H.,

& Lens, F. (2022). The evolution of insular woodiness. *Proceedings of the National*

Academy of Sciences, *119*(37), e2208629119.

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Ethics

This work did not require ethical approval from a human subject or animal welfare

committee.

Data availability Statement

Plant species lists, flower traits measured, and models used for the analysis are available in

the supplementary materials of this article.

Conflict of Interest Declaration

The authors declare no conflict of interest.

Supplementary data

Supplementary 1: List of Island-Mainland pairs and sources used as references.

Supplementary 2: List of floral traits used for the analysis.

Supplementary 3: Supplementary tables and figures.

Figure legends

Figure 1 Map of the studied archipelagos.

Figure 2 Floral display log ratio (fdLR) and flower size log ratio (fsLR) of animal- and wind-

pollinated species (y-axis) varying as a function of log mainland values (x-axis) for a)

animal-pollinated floral display, b) wind-pollinated floral display c) animal-pollinated flower

size and d) wind-pollinated flower size. Each point denotes an island/mainland pairing. Floral

display follows no recognisable pattern for species of either pollination mode. For animal-

pollinated species, the flower size slope is significantly smaller than 0, giving support for the

island rule. For wind-pollinated species, the flower size slope does not vary significantly

from 0, but most points are above the horizontal line, indicating a trend toward gigantism.

Opposing arrows denote the island rule. Single, upward arrows denote gigantism.

Fig. 1

Fig 2.