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Impact of climate change on leafhopper vectors of phytoplasmas in North America

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ABSTRACT

Climate change significantly contributes to shifts in the geographical range of pests and diseases.

Leafhoppers (Hemiptera: Cicadellidae), known vectors of phytoplasmas pathogens, are linked to the

transmission of more than 600 diseases affecting a thousand plant species worldwide. Despite this,

the potential effects of climate change on leafhopper vectors of phytoplasmas remain a critical

knowledge gap. To address this gap, our study investigated the potential impact of climate change

on 14 species of Nearctic leafhoppers associated with phytoplasmas. Using the MaxEnt species

distribution algorithm and other ecological niche modeling techniques, we assessed *(i)* the expected

species richness under current climate conditions and four future scenarios and *(ii)* the

environmental niche similarity among these species across these scenarios. Our projections suggest

that the eastern region of North America holds the potential for the highest species diversity, a trend

expected to persist across all future scenarios, gradually expanding eastward. Notably, our findings

indicate the increasing suitability of northern Canada for more leafhopper species. Network analysis

further revealed a remarkable similarity in environmental niches among most leafhopper species.

Moreover, across the four future scenarios, there is a tendency for an increase in this similarity.

Altogether, our study underscores the potential persistent presence of Nearctic leafhoppers in their

current habitats while pointing to a shift toward northern North America in future scenarios. These

findings have significant implications for sustainable pest management practices, prompting a

necessary discussion on strategies to mitigate climate change and pests migration impact on

agricultural systems.

INTRODUCTION

Climate change significantly contributes to alterations in species distributions, prompting shifts in their geographic ranges[1,2]. Over the past six decades, crop pests and diseases have notably moved toward northern latitudes globally, progressing at approximately 2.7 km annually since 1960[3]. This phenomenon is primarily attributed to global warming and trade dynamics[3]. However, the response of these organisms to the ongoing environmental changes is intricate and conditional upon various determinants, notably their geographic origins[4–6]. Tropical pest species, for instance, typically exhibit a restricted tolerance to temperature variation and tend to live close to their maximum thermal limits[4,6]. In contrast, temperate species contend with a broader spectrum of climate extremities, including temperature fluctuations[4]. As a result, projections suggest a potential reduction in the suitable habitats for tropical pest species with the anticipated progression of climate warming[4,7]. Conversely, a counterintuitive trend is expected for temperate species, given their adaptations to adapt to more diverse climatic events[3,4].

Recently, the efforts to assess the impacts of climate change on biological systems have intensified[8]. Species distribution modeling (SDM), also known as ecological niche modeling (ENM), has risen as a crucial tool to explore the potential effects of climate change on the geographic distribution of organisms[9]. However, within entomology, research efforts have predominantly centered around specific orders, such as Lepidoptera and Diptera, while less attention has been dedicated to others like Hemiptera[8]. It is, therefore, essential to increase research focus on neglected insect orders like Hemiptera to understand the impact of climate change on a wide array of insect distribution.

Leafhoppers (Hemiptera: Cicadellidae) constitute a herbivorous group known for their varied feeding habits across vascular plants[10]. With an estimated 21,351 species described globally[11], these insects exhibit a remarkable characteristic: about 90% of their genera are endemic to specific geographical regions[12]. The Nearctic region, spanning from Mexico to the Arctic, stands out as a leafhopper hotspot, with nearly 3,000 described species showcasing a high level of

endemism and diversity[11–17]. Nearctic leafhoppers are well-adapted to various climatic conditions,

surviving from harsh winters to scorching summers, finely synchronizing their life cycles with these

climatic patterns[13–16]. This adaptation suggests that environmental factors such as temperature and

continental winds could primarily influence their distribution.

Increasing temperatures favor Nearctic population growth, as higher temperatures decrease

the species' life cycle duration[18,19]. Additionally, continental winds facilitate their dispersal into

northern regions during the spring and summer[20–22]. Consequently, it is plausible to anticipate that

ongoing global warming and other climatic changes may offer Nearctic leafhoppers opportunities to

expand their potential geographical range.

In addition to the damage associated with herbivory, leafhoppers are also vectors of

devastating plant pathogens. Over 100 leafhopper species carry phytoplasmas linked to more than

600 diseases affecting a thousand plant species worldwide[10,23–25]. In the Nearctic region,

leafhoppers exhibit oligophagous or polyphagous feeding behaviors, demonstrating a broad

consumption pattern across diverse host plants that can serve as main hosts or reservoirs of

phytoplasmas[13,14,16,26]. In this region, 38 leafhopper species have been identified as vectors of

phytoplasmas affecting horticultural, fruit, and field crops[27,28]. However, recent findings indicate a

tripling in the incidence of phytoplasma diseases in the Nearctic over the past decade[29]. This trend

suggests several potential factors: (*i*) advancements in molecular diagnostic methods and novel

detection technologies for phytoplasmas[30,31]; (*ii*) a rise in the abundance of leafhopper vectors[32];

(*iii*) the probable existence of yet-to-be-identified highly efficient vectors[19,33–35]; and (*iv*) an

expansion of the geographic range of leafhopper vectors[20].

Nonetheless, a significant research gap still exists concerning investigating the potential

impacts of climate change on the phytoplasma-vectoring Nearctic leafhoppers. Our work addresses

this gap by examining the potential impact of climate change on 14 species of leafhopper vectors

(or potentially be) of phytoplasma affecting berries and other small fruits across North America like

Graphocephala spp., *Erythroneura* spp., and *Empoasca fabae* (Harris) (Fig. 1)[27,28,33,34,36],

employing ENM-based methods. The study is particularly urgent given the scarcity of research on

the dynamics of these leafhopper communities in North America and the impact of climate change

in the short and long term.

METHODS

In this study, our objective was to assess environmental similarity and potential leafhopper

species richness under current and future climate scenarios using ENM-based methods. The

analyses were conducted using the Java version of the machine learning algorithm Maximum

Entropy Modeling of Species Geographic Distributions (MaxEnt, v. 3.4.1)[37], the wallace shiny app

(v. 2.0.5)[38], SDMtoolbox (Pro v. 0.9.1)[39], ArcGIS Pro (v. 3.1.2), and R packages described further.

We chose MaxEnt as our modeling algorithm due to its high statistical performance, mainly when

dealing with various species occurrence records datasets[9,40,41].

Species selection

We selected 14 species that are known or potentially be vectors of phytoplasmas in North

America's berries, grapevines, and other small fruits: *Amplicephalus* (formerly *Endria*), *inimicus*

(Say), *Ceratagallia* (formerly *Aceratagallia*) *humilis* (Oman), *Colladonus geminatus* (Van Duzee),

Edwardsiana rosae (Linnaeus), *Empoasca fabae* (Harris), *Erythroneura comes* (Say), *Erythroneura*

vitis (Harris), *Erythroneura ziczac* (Walsh), *Exitianus exitiosus* (Uhler), *Graphocephala fennahi*

(Young), *Jikradia olitoria* (Say), *Macrosteles quadrilineatus* (formerly *fascifrons*) (Forbes),

Paraphlepsius irroratus (Say), and *Scaphoideus titanus* (Ball). In this specific community, there is

an absence of documented reports indicating that three species (*Erythroneura comes*, *Erythroneura*

vitis, and *Jikradia olitoria*) act as vectors of phytoplasmas under field conditions[28,33,34,36,42]. Despite

this absence, we retained these three species due to their recognized economic importance.

Furthermore, 12 out of 14 are exclusively present in the Nearctic region. Meanwhile, *Empoasca*

fabae extends its occurrence to the Neotropics[43], while *Scaphoideus titanus*, introduced to Europe in

1950, subsequently spread in this continent due to human trade activities[44].

Occurrence records

We obtained species occurrence records (latitude, longitude) for North America from openly

available online repositories, including the Global Biodiversity Information Facility (GBIF,

accessed on 9 June 2023; https://doi.org/10.15468/dl.h3x8mf), which also includes records from

INaturalist platform. We also included recent literature on leafhoppers' field surveys[19,32,34,35,45].

These records were collected after 1970 to ensure relevance to the environmental variables used in

our models (1970-2000) while avoiding historical records that may not accurately reflect current

species occurrence sites[46,47]. We removed duplicate records within the same pixel to enhance data

quality and eliminate those with incomplete information. We also applied spatial thinning to reduce

spatial bias, retaining one record for each 5 km², corresponding to the spatial resolution used in our

models. The final number of occurrence records for each species were 300 for *Amplicephalus*

inimicus, 33 for *Ceratagallia humilis*, 33 for *Colladonus geminatus*, 66 for *Edwardsiana rosae*, 852

for *Empoasca fabae*, 71 for *Erythroneura comes*, 189 for *Erythroneura vitis*, 110 for *Erythroneura*

ziczac, 578 for *Exitianus exitiosus*, 912 for *Graphocephala fennahi*, 2376 for *Jikradia olitoria*, 277

for *Macrosteles quadrilineatus*, 280 for *Paraphlepsius irroratus*, and 94 for *Scaphoideus titanus*

(Fig. 2).

Current environmental variables and climate change scenarios

For our current distribution model, we downloaded 19 world bioclimatic variables (v. 2.1)

covering the period of 1970-2000 at a spatial resolution of approximately 5 km²[48]. These variables

are derived from monthly temperature and precipitation data, providing insights into annual trends,

seasonality, and extreme environmental conditions. Given that these variables can be highly

correlated, potentially affecting model performance due to multicollinearity[49], we conducted a

Pearson pair-wise correlation analysis[46]. This analysis allowed us to identify and subsequently

remove variables with correlations exceeding a threshold of r ≥ |0.7|, as values beyond this

threshold can distort model estimation (Supplementary Table S1)[49]. Thus, five bioclimatic variables

were selected: annual mean temperature (bio_1), mean temperature diurnal range (bio_2),

temperature annual range (bio_7), annual precipitation (bio_12), and precipitation seasonality

(bio_15).

For the climate change scenarios, we used the global circulation model GISS-E2-1-G[50]

under the four shared socioeconomic pathway scenarios, SSP1-2.6, SSP2-4.5, SSP3-7.0, and SSP5-

8.5 for the period 2041-2060 at a spatial resolution of 5 km^2 (available at

https://www.worldclim.org/data/cmip6/cmip6_clim2.5m.html).

The GISS-E2-1-G model represents an updated version of the GISS-E2 developed by the

National Aeronautics and Space Administration (NASA) Goddard Institute for Space Studies

(GISS). This updated model incorporates significant improvements in the representation of climate

variability[50]. One notable improvement of this version is the model's ability to depict the

temperature response to increased carbon dioxide. In this iteration, the response is marginally more

significant than in previous versions, indicating a more accurate and realistic representation of

climate dynamics[50].

The SSPs, as featured in the Intergovernmental Panel on Climate Change (IPCC) sixth

assessment report, offer global narratives that consider responses to climate change. They are

associated with various challenges for mitigation and adaptation, ranging from 1 to 5, where 1

represents low, 2 medium, 3 high, 4 low for mitigation and high for adaptation, and 5 high for

mitigation and low for adaptation[51]. These SSPs are coupled with global circulation models (GCM)

scenarios (previously known as Representative Concentration Pathways – RCP). In our study, we

adopted four GCM scenarios: 2.6 (low), 4.5 (intermediate), 7.0 (high), and 8.5 (very high). The

SSP1-2.6 scenario limits global warming by 2100 to 2 °C, while the SSP2-4.5, SSP3-7.0, and

SSP5-8.5 scenarios project values of 3 °C, 4 °C, and over 4 °C, respectively[51].

In the context of North America, predictive models indicate significant shifts in the annual

mean temperature, demonstrating an increase in maximum values from 28.6 ℃ to 31 ℃

(Supplementary Figure S1). Moreover, the annual precipitation patterns suggest a uniform decrease

in the west compared to the east in all scenarios. In contrast, precipitation seasonality is predicted to

show the opposite way (Supplementary Figure S1).

We used the raster package in R[52] to extract bioclimatic variables for each SSP.

Subsequently, we created a North American shapefile, and all layers were cropped to align with this

shapefile. This cropping procedure was conducted using ArcGIS.

Model development, selection, and evaluation

MaxEnt is a correlative maximum entropy-based algorithm that estimates species

probability of presence or habitat suitability, ranging from 0 (unsuitable) to 1 (highly suitable),

using species occurrence records and background data associated with the environmental variables

used in the modeling[41]. We used the MaxEnt java (version 3.4.1) and the ENMeval package[53]. We

randomly selected 10000 background points to represent environmental conditions across North

America for model calibration. We utilized Maxent's' clamping' function to prevent model

extrapolations beyond the environmental range. We configured two main sets for model

performance and transferability testing: feature class (FC) and regularization multiplier (RM). FC

involves mathematical transformations of the environmental variables into predictors: linear (L),

quadratic (Q), product (P), and hinge (H), as well as their combinations. RM penalizes model

complexity and reduces the number of parameters[54]. In our approach, we tested five FC

combinations (L, LQ, H, LQH, LQHP) and RM values ranging from 0 to 5 with a step of 0.5.

We employed two occurrence partition methods, spatial partition block and random k-fold,

to perform cross-validation during model training and testing. The spatial partition method divides

localities into four groups based on latitudinal and longitudinal lines, creating spatial rectangles.

The random k-fold method randomly assigns localities to one of the 'k' groups (k = 4). In both

approaches, we conducted cross-validation by generating predictions with different training sets and

evaluated model performance by projecting outcomes for the omitted sites[54].

We initially ran 100 models for each species using the feature class (FC) and regularization

multiplier (RM) mentioned earlier, combining partition methods, and rendering 2800 models.

However, we found that the random k-fold method produced better results. For some species, the

spatial partition block method could not be implemented due to a limited number of localities[38].

Consequently, we retained the random k-fold method for training and testing data. We selected the

best model from the Area Under the Receiver Operating Characteristic Curve (AUC) and True Skill

Statistics.

The AUC is a standard metric for assessing model performance. It ranges from 0 to 1 and

measures the model's ability to rank positive records higher than negative ones across the suitability

range. Values closer to 1 indicate excellent model performance. However, for presence-only models

like MaxEnt, interpreting AUC has some limitations, especially when there is a lack of absence

data[55]. Additionally, when occurrence records are fewer than 100, AUC may not accurately estimate

model complexity[56]. We also used True Skill Statistics (TSS) to address these limitations.

TSS assesses model performance by considering sensitivity and specificity at a binary

threshold (unsuitable/suitable). TSS values range from -1 to 1, with values closer to one indicating

excellent sensitivity and specificity. High TSS (> 0.5) values mean a high proportion of real

presences, and current absences are correctly predicted. Conversely, values at or below zero suggest

errors in predicting presence (commission) or predicting absence (omission)[46,57].

Projections

We transformed asci (cloglog output) maps into raster files using ArcGIS. These maps were

then classified into binary outputs, unsuitable and suitable, for each species based on the maximum

test sensitivity plus specificity cloglog threshold (MaxTSS values in Table 1). Sensitivity and

specificity represent the proportions of correctly predicted presences and absences, respectively.

Subsequently, we overlaid the binary maps to calculate species richness under current and future

scenarios. We used the 'biodiversity measurement' function available in SDMtoolbox. The final

output is a map showing the sum of unique species per pixel, representing the total number of

species in each grid cell.

Environmental similarity

We assessed environmental similarity using the final species occurrence records and the five

environmental variables employed in our models under current and four future scenarios. We

conducted a principal component analysis (PCA) to reduce the original environmental space's

dimensionality. Subsequently, we calculated species occurrence density along the first two

components of the PCA and performed similarity tests.

In the similarity test, both species' environments were randomly shifted within the

background extent, and 'null' overlaps were calculated from these simulations. If the observed

overlap was greater than 95% of the simulated overlaps (p-value < 0.05), the test indicates that the

two species' environments were more similar than expected by chance (a one-tailed test). We

conducted tests by pairing two leafhopper species at each iteration until all 91 possible

combinations among the 14 species were evaluated for each climate scenario. Subsequently, we

created a matrix containing the similarity p-values (supplementary Table S2).

The similarity p-values matrix was further transformed into a binary matrix, where '0'

represents random, and '1' indicates species with environment similarity (supplementary Table S2).

Using network analysis, we identified species with higher and lower similarity. For each network

scenario, we calculated the network density. Density is the proportion of observed ties or

connections to a network's maximum and possible relations. Thus, density is a ratio that can range

from 0 to 1. The closer to 1 the density is, the more interconnected the network[58]. These analyses

and tests were performed using the wallace interface to compute the tests, the packages ade4[59],

igraph[60], ecospat[61], and Corel Painter (Essential 7, ON, CAD) for data illustration.

RESULTS

Models performance and variables contribution

Our predictive models demonstrated high discriminant performance, as evidenced by AUC values ranging from 0.883 to 0.980 and TSS values spanning from 0.629 to 0.864 (Table 1). Our relative contribution analysis highlighted that temperature-related variables substantially shaped the models for most of the studied species (Fig. 3). However, some exceptions were observed for *Erythroneura comes*, *Erythroneura vitis*, *Graphocephala fennahi*, and *Jikradia olitoria*, where precipitation-related variable had a more substantial influence on the models. Interestingly, in the case of *Scaphoideus titanus*, both sets of variables - temperature and precipitation - contributed almost equally to the model (Fig. 3).

Current species richness

The richness map obtained in this study reveals that the eastern part of North America, encompassing southern Ontario and Québec—identified as an anticipated hotspot for Nearctic leafhoppers—exhibits the potential for the highest species diversity (Fig. 4). Another minor hotspot was observed in the southwestern of the United States. These regions typically feature a temperate climate with elevated humidity levels. In contrast, species diversity tends to decrease in drier areas like the grasslands of Canada and the U.S., which include per-arid, arid, and semi-arid zones. Consequently, northern regions of Canada and the U.S. show a lower species diversity (Fig. 4).

Future projections

To predict the impact of climate change on leafhopper richness, future scenarios SSP1-2.6 (Fig. 5a), 2-4.5 (Fig. 5b), 3-7.0 (Fig. 5c), and 5-8.5 (Fig. 5d) were evaluated, finding that under the four scenarios, suitable areas for species richness will tend to increase (Table 2). The regions previously identified with the highest species richness, hosting all 14 species, are projected to retain

similar patterns in all four future scenarios (Fig. 5). We observed a considerable increase in this

richness, gradually extending towards the east (Fig. 5). A new hotspot emerged in the southern parts

of Alberta and British Columbia, Canada (Fig. 5). Additionally, an expansion in species richness is

evident in the southeast and northeast parts of the continent. In line with current conditions, our

models predict lower species diversity in arid zones compared to regions with higher humidity (Fig.

5). Altogether, our results indicate that, under the four climate change scenarios investigated,

northern Canada is expected to become more hospitable to a more significant number of species

than under the current scenario.

Environmental similarity

Our network analysis revealed that most leafhopper species present a similar environmental

niche with a density value of 0.467 (Fig. 6). Notably, *Colladonus geminatus*, *Edwardsiana rosae,*

and *Exitianus exitiosus* showed a remarkable environmental similarity with other 4 and 3 species,

respectively, indicating a more specialized environment. In contrast, *Amplicephalus inimicus*,

Empoasca fabae, *Erythroneura* spp., *Macrosteles quadrilineatus*, *Paraphlepsius irroratus*, and

Scaphoideus titanus exhibited a higher degree of environmental similarity with most of the species

in our study, suggesting a broader ecological environment (Fig. 6). These findings underscore the

existence of distinct environmental patterns within the leafhopper species studied here.

Under four future scenarios, the environmental similarity tended to increase between

leafhoppers species suggesting more interconnected networks as indicated by the increase of

density values from 0.467 under the current scenario to 0.523 under SSP1-2.6 (Fig. 6a), 0.533 under

SSP2-4.5 (Fig. 6b), 0.571under SSP3-7.0 (Fig. 6c), and 0.542 under SSP5-8.5 (Fig. 6d). In all

scenarios, leafhopper species tended to present environmental similarity, including those with a

more restricted environment under the current scenario like *Colladonus geminatus* and

Edwardsiana rosae. An exception was registered for *Exitianus exitiosus*, which only shares an

ecological environment with a maximum of three species. Overall, our results underscore the

potential of leafhopper species to occupy a more similar environment niche under a warming world

scenario.

DISCUSSION

Our study examined the environmental niche similarity and distribution of 14 Nearctic

leafhopper species across North America under current conditions and four future climate scenarios.

We found that 11 out of 14 leafhopper species share a similar environmental niche with more

species. Meanwhile, *Colladonus geminatus*, *Edwardsiana rosae*, and *Exitianus exitiosus* show more

specialized niches. Interestingly, a similar trend appears to grow across climate change scenarios.

Our prediction points to warm and humid regions in North America as potential leafhopper species

hotspots, with fewer species expected to occur in arid areas. Under all climate scenarios, we

observed an expansion of the suitable regions and species overlap, especially in the northern area of

the continent. Our findings suggest a positive link between future climates and the potential

occurrence and niche similarity of North America's Nearctic leafhopper community potential

phytoplasma vector.

According to our models, temperature significantly influences the distribution of most

leafhopper species investigated in this study. In the Nearctic region, temperature is the primary

factor driving leafhopper distribution. Historical periods of high temperatures are thought to trigger

large-scale dispersal of leafhopper species into northern areas[12,16]. During winter, some leafhopper

species undergo temperature-related diapause in their egg, nymph, or adult stages[14,16,62], while

migratory species like *Empoasca fabae* overwinter in southern regions[20]. As temperatures rise in

early spring, leafhopper populations start to increase from both diapausing stages and the arrival of

migratory species due to continental-scale winds[21,22], with populations peaking in warmer

months[19,20,32]. Leafhopper populations in the Nearctic region exhibit shorter life cycles in warmer

conditions[18], resulting in more yearly generations than colder ones[20,62]. Yet, precipitation also plays

a role in species occurrence, particularly noted in our models for *Erythroneura comes*, *Erythroneura*

vitis, *Graphocephala fennahi*, and *Jikradia olitoria*. Heavy precipitation, especially during early

nymphal stages, can cause species mortality[63]. In addition, high humidity indirectly impacts species'

performance by affecting their plant hosts, as noted for *Erythroneura* spp., which tends to increase

populations due to weakened host plants due to moisture[62].

Upon conducting an environmental niche analysis, we show substantial environmental

similarity among most leafhopper species. As previously mentioned, historical dispersion patterns

of leafhoppers indicate movements from regions characterized by higher diversity and warmer

climates to regions with lower diversity and cooler climates, such as between the Neotropical and

Nearctic regions[12,16,64]. Consequently, approximately 90% of leafhopper genera exhibit endemism

within their respective zoogeographical regions, demonstrating a marked inclination toward host

specificity and limited dispersal rates[12]. This endemism potentially elucidates the environmental

similarity observed among the species under study, given that the majority, 12 out of 14, exclusively

inhabit the Nearctic region.

Recent surveys of leafhoppers reinforce these observations, highlighting increased species

richness and potential overlaps in the regions examined. For example, Saguez *et al.*[45] recorded

17,496 leafhopper specimens across Canadian vineyards in British Columbia, Québec, and Ontario.

These specimens spanned 54 genera and 110 species, with the genera *Empoasca*, *Erythroneura*, and

Macrosteles being widespread. Plante *et al.*[19] captured 33,007 leafhopper specimens from

strawberry fields in Québec, which included 61 genera and 118 species, with *Empoasca*,

Macrosteles, and *Hebata* as prevalent genera.

In contrast, three species, *Colladonus geminatus*, *Edwardsiana rosae*, and *Exitianus*

exitiosus, displayed a reduced level of niche similarity, suggesting a more specialized

environmental niche within regions characterized by lower species overlap. Intriguingly, two

species, *Colladonus geminatus* and *Exitianus exitiosus*, exhibit distribution and abundance within

grassland habitats[63,65], which are recognized for harboring reduced species diversity compared to

humid environments[12,63].

Our assessment of species richness has uncovered a geographical trend for the species

evaluated: areas with higher humidity and moisture generally support a greater variety of species,

with numbers dwindling in drier regions. This prediction aligns with prior observations in southern

Ontario Québec[63] and New Hampshire[13], regions known for high biodiversity. These findings

suggest a complex relationship between environmental conditions, habitat preferences of different

leafhopper species, and the resulting distribution of these species.

Under the four climate change scenarios studied, our network analysis revealed a notable

rise in species similarity, hinting at a likely increase of species overlap within their suitable habitats

in forthcoming scenarios[6]. Various indicators emphasize the impact of rising temperatures on the

phytoplasma vectors studied here, leading to larger distribution areas for these species. For

example, the northward shift of the migratory *Empoasca fabae* in North America has sped up over

the last 62 years, a change linked to warmer temperatures[20]. This earlier arrival has notably

intensified infestations, potentially favoring this species in the North American region[20]. A recent

survey conducted in Québec's strawberry fields identified 11 leafhopper species for the first time in

the province. Ten of these species were the farthest observed towards the east in Canada, marking a

significant expansion[19]. This survey also noted a considerable increase in leafhopper species

composition compared to data from 14 years ago in vineyards[45] and 10 years ago in blueberry

fields[35], with 53 new species registered in 2021 and 2022[19]. These findings strongly imply a

growing overlap among leafhopper species in North America due to their expanding occurrence

ranges, aligning with the trends observed through our predictive models.

We expect a significant expansion of suitable habitats for various leafhopper species,

especially in northern areas. This trend mirrors similar trends observed in other groups of crop

pests, moving toward northern regions driven primarily by global warming[3]. This shift results in

broader ranges for species distribution and overwintering zones[3,66,67]. Additionally, multiple lines of

evidence support that most species in the Nearctic region are likely to adjust their geographic

distribution to occupy areas that meet their specific needs[1,4,6,68]. This adaptability stems from the

ability of Nearctic species to cope with seasonal temperatures and extreme variations, survive cold

winters, and succeed in hot summers, thus aligning their life cycles with these environmental

conditions[4].

Neotropical leafhopper species, in comparison, are less adaptable to warm climates due to

their specialization within a narrower temperature range and increased sensitivity to changes in

precipitation. For example, studies with *Macrosteles quadrilineatus* at 0 °C showed survival for 18

days but not reproduction, while at 35 °C, leafhoppers also survived 18 days and were able to

reproduce[18]. In contrast, the neotropical leafhopper *Daubulus maidis* (DeLong & Wolcott) has a

developmental temperature range between 20 °C and 30 °C[69], suggesting that a warming climate

might reduce suitable habitats for this species in its native South American range as temperatures

approach its upper limit[7].

Our study investigated the impact of abiotic factors on current and future potential

environmental niche similarity and species richness of 14 North American leafhopper potential

vectors of phytoplasmas. However, they do not account for biotic variables such as natural enemies

(parasitoids and predators) and the availability of hosts, the seasonal effect on area suitability, or the

effect of climate change on continental wind patterns. These factors, collectively, act as a biological

filter, influencing the actual niche of the species studied[16,46]. Moreover, many Nearctic leafhoppers

do not inhabit their entire potential range[13,16]. Therefore, our analyses should be viewed through the

lens of the environmental niche concept, focusing primarily on the climatic factors limiting species

distribution. This approach often extends beyond the realized niche[46].

IMPLICATIONS FOR SUSTAINABLE AGRICULTURE

Our study highlights significant challenges in managing Nearctic leafhoppers, particularly

their continued presence in current habitats and their potential movement towards northern regions

in North America. Although insecticide resistance in Nearctic leafhopper species has not been

studied in depth, new evidence suggests that the heavy reliance on insecticides for leafhopper

control in Canada could be becoming less effective at managing populations[19]. Moreover, the early

arrival of migratory leafhopper populations exacerbates plant damage[20], potentially leading to

increased insecticide usage and increasing the chances of developing insecticide resistance[19,70].

Additionally, the potential resilience of Nearctic leafhopper species under warmer climates

could amplify the spread of diseases they transmit. It is well-known that temperature fluctuations

moderate phytoplasma multiplication within leafhoppers[18], and higher temperatures accelerate

phytoplasma cycles within plants[71–73], increasing the chances of acquisition during vector feeding

activities. Given that species such as *Macrosteles quadrilineatus* and *Empoasca fabae* have already

tested positive for phytoplasma strains that affect multiple hosts[27,28,34], another anticipated effect of

climate change might be the expansion of host ranges for these phytoplasmas, previously limited by

the food sources available to these leafhoppers.

The changes in climate patterns will lead to increased Nearctic leafhopper activity,

migration, and interaction, which will negatively affect plant health. These results call for a

complete rethink of our control and integrated pest management strategies to meet these new

challenges. Our study emphasizes the need for further research to discover effective products to

manage Nearctic leafhopper populations. We must consider effective application techniques and the

insect's biology. Additionally, we should explore new methods of biological control and the

potential of biotechnology and emerging technologies to halt the spread of these pests and the

diseases they carry without harming the environment.

AUTHOR CONTRIBUTION

Conceptualization: A.A.S. and E.P.L.; Data analysis and presentation: A.A.S.; Writing full draft,

review, and editing: A.A.S., J.J., and E.P.L.; Supervision: E.P.L.

COMPETING INTERESTS

The authors declare no competing interests.

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DATA AVAILABILITY

The dataset and resources used in this study are openly available on websites upon user registration: GBIF (https://www.gbif.org/), INaturalist (https://www.inaturalist.org/), MaxEnt java (https://biodiversityinformatics.amnh.org/open_source/maxent/), Wallace app (https://wallaceecomod.github.io/), WorldClim climate data (https://www.worldclim.org/data/index.html). Supplementary tables and figures can be found in Zenodo following this doi: https://doi.org/10.5281/zenodo.10371910.

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REFERENCES

1. Pecl, G. T. *et al.* Biodiversity redistribution under climate change: impacts on ecosystems and human well-being. *Science* **355**, eaai9214; 10.1126/science.aai9214 (2017).

2. Kharouba, H. M., Lewthwaite, J. M. M., Guralnick, R., Kerr, J. T. & Vellend, M. Using insect natural history collections to study global change impacts: challenges and opportunities. *Phil. Trans. R. Soc. B* **374**, 20170405; 10.1098/rstb.2017.0405 (2019).

3. Bebber, D. P., Holmes, T. & Gurr, S. J. The global spread of crop pests and pathogens. *Glob.*

Ecol. Biogeogr. **23**, 1398–1407 (2014).

4. Harvey, J. A., Heinen, R., Gols, R. & Thakur, M. P. Climate change☐mediated temperature

extremes and insects: from outbreaks to breakdowns. *Glob. Chang. Biol.* **26**, 6685–6701 (2020).

5. Lehmann, P. *et al.* Complex responses of global insect pests to climate warming. *Front. Ecol.*

Environ. **18**, 141–150 (2020).

6. Araújo, M. B. *et al.* Heat freezes niche evolution. *Ecol. Lett.* **16**, 1206–1219 (2013).

7. Santana, P. A., Kumar, L., Da Silva, R. S., Pereira, J. L. & Picanço, M. C. Assessing the impact

of climate change on the worldwide distribution of *Dalbulus maidis* (DeLong) using MaxEnt.

Pest Manag. Sci. **75**, 2706–2715 (2019).

8. Andrew, N. R. *et al.* Assessing insect responses to climate change: What are we testing for?

Where should we be heading? *PeerJ* **1**, e11; 10.7717/peerj.11 (2013).

9. Feng, X. *et al.* A checklist for maximizing reproducibility of ecological niche models. *Nat.*

Ecol. Evol. **3**, 1382–1395 (2019).

10. Weintraub, P. G. & Beanland, L. Insect vectors of phytoplasmas. *Annu. Rev. Entomol.* **51**, 91–

111 (2006).

11. Bartlett, C. R. *et al*. The diversity of the true hoppers (Hemiptera: Auchenorrhyncha) 501-590

(ed. Foottit, R. G. & Adler, P. H.) (John Wiley & Sons, Inc, 2017).

12. Nielson, M. W. & Knight, W. J. Distributional patterns and possible origin of leafhoppers

(Homoptera, Cicadellidae). *Rev. Bras. Zool.* **17**, 81–156 (2000).

13. Chandler, D. S. & Hamilton, K. G. A. Biodiversity and ecology of the leafhoppers (Hemiptera:

Cicadellidae) of New Hampshire. *Trans. Am. Entomol. Soc.* **143**, 773–971 (2017).

14. DeLong, D. M. The leafhoppers, or Cicadellidae, of Illinois (Eurymelinae Balcluthinae). *Bull.*

Ill. Nat. Hist. Sur. **24**, 97–376 (1948).

15. Hamilton, K. G. A. The species of the North American leafhoppers *Ceratagallia* Kirkaldy and

Aceratagallia Kirkaldy (Rhynchota: Homoptera: Cicadellidae). *Can. Entomol.* **130**, 427–490

(1998).

16. Hamilton, K. G. A. Leafhoppers (Homoptera: Cicadellidae) of the Yukon: Dispersal and

Endemism 337-375 (ed. Danks, H. V. & Downes, J. A.) (University of Chicago, 1997).

17. Pinedo￼Escatel, J. A. *et al.* Biogeographical evaluation and conservation assessment of

arboreal leafhoppers in the Mexican transition zone biodiversity hotspot. *Divers. Distrib.* **27**,

1051–1065 (2021).

18. Bahar, M. H., Wist, T. J., Bekkaoui, D. R., Hegedus, D. D. & Olivier, C. Y. Aster leafhopper

survival and reproduction, and aster yellows transmission under static and fluctuating

temperatures, using ddPCR for phytoplasma quantification. *Sci. Rep.* **8**, 227; 10.1038/s41598-

017-18437-0 (2018).

19. Plante, N. *et al.* Can leafhoppers help us trace the impact of climate change on agriculture?

biorxiv; 10.1101/2023.06.13.544773 (2023).

20. Baker, M. B., Venugopal, P. D. & Lamp, W. O. Climate change and phenology: *Empoasca*

fabae (Hemiptera: Cicadellidae) migration and severity of impact. *PLoS ONE* **10**, e0124915;

10.1371/journal.pone.0124915 (2015).

21. Taylor, R. A. J. & Reling, D. Preferred wind direction of long-distance leafhopper (*Empoasca*

fabae) migrants and its relevance to the return migration of small insects. *J. Anim. Ecol.* **55**,

1103-1114 (1986).

22. Taylor, R. A. J. & Shields, E. J. Revisiting potato leafhopper, *Empoasca fabae* (Harris),

migration: implications in a world where invasive insects are all too common. *Am. Entomol.* **64**,

44–51 (2018).

23. Bertaccini, A. *et al.* Revision of the 'Candidatus Phytoplasma' species description guidelines.

Int. J. Syst. Evol. Microbiol. **72**; 10.1099/ijsem.0.005353 (2022).

24. Maejima, K., Oshima, K. & Namba, S. Exploring the phytoplasmas, plant pathogenic bacteria.

J. Gen. Plant. Pathol. **80**, 210–221 (2014).

25. Weintraub, P. G., Trivellone, V. & Krüger, K. The biology and ecology of leafhopper

transmission of phytoplasmas 27-52 (ed. Bertaccini, A., Weintraub, P. G., Rao, G. P. & Mori,

N.) (Springer Singapore, 2019).

26. Romero, B., Olivier, C., Wist, T. & Prager, S. M. Oviposition behavior and development of

aster leafhoppers (Hemiptera: Cicadellidae) on selected host plants from the Canadian Prairies.

J. Econ. Entomol. **113**, 2695–2704 (2020).

27. Olivier, C. Y., Lowery, D. T. & Stobbs, L. W. Phytoplasma diseases and their relationships with

insect and plant hosts in Canadian horticultural and field crops. *Can. Entomol.* **141**, 425–462

(2009).

28. Almeida Santos, A., Jacques, J., Plante, N., Fournier, V. & Pérez-López, E. Leafhoppers as

vectors of phytoplasma diseases in Canadian berry crops: a review in the face of climate

change. *Ann. Entomol. Soc. Am.* saad038; 10.1093/aesa/saad038 (2023).

29. Brochu, A.-S., Dionne, A., Fall, M. L. & Pérez-López, E. A decade of hidden phytoplasmas

unveiled through citizen science. *Plant Dis.* **107**, 3389–3393 (2023).

30. Hammond, C. *et al.* Detection of blueberry stunt phytoplasma in Eastern Canada using cpn60-

based molecular diagnostic assays. *Sci. Rep.* **11**, 22118; 10.1038/s41598-021-01439-4 (2021).

31. Pérez-López, E., Omar, A. F., Al-Jamhan, K. M. & Dumonceaux, T. J. Molecular identification

and characterization of the new 16SrIX-J and cpn60 UT IX-J phytoplasma subgroup associated

with chicory bushy stunt disease in Saudi Arabia. *Int. J. Syst. Evol. Microbiol.* **68**, 518–522

(2018).

32. Olivier, C. Y., Séguin-Swartz, G., Galka, B. & Olfert, O. O. Aster yellows in leafhoppers and

field crops in Saskatchewan, Canada, 2001–2008. *Am. J. Plant Sci. Biotechnol.* **5**, 88–94

(2011).

33. Lenzi, P., Stoepler, T. M., McHenry, D. J., Davis, R. E. & Wolf, T. K. *Jikradia olitoria*

(Hemiptera: Cicadellidae) transmits the sequevar NAGYIIIβ phytoplasma strain associated with

North American grapevine yellows in artificial feeding assays. *J. Insect Sci.* **19**;

10.1093/jisesa/iey124 (2019).

34. Olivier, C. *et al.* Occurrence of phytoplasmas in leafhoppers and cultivated grapevines in

Canada. *Agric. Ecosyst. Environ.* **195**, 91–97 (2014).

35. Pagé, D. Acquisition de connaissances sur les phytoplasmes dans la culture du bleuetier en

corymbe. https://www.agrireseau.net/petitsfruits/documents/Rapport_IRDA_2013-06-

03_phytoplasme_bleuetier-final.pdf (2013).

36. Arocha-Rosete, Y. *et al.* Surveys reveal a complex association of phytoplasmas and viruses with

the blueberry stunt disease on Canadian blueberry farms. *Ann. Appl. Biol.* **174**, 142–152 (2019).

37. Phillips, S. J., Dudík, M. & Schapire, R. E. Maxent software for modeling species niches and

distributions (Version 3.4.1). http://biodiversityinformatics.amnh.org/open_source/maxent/

(2017).

38. Kass, J. M. *et al. wallace* 2: a shiny app for modeling species niches and distributions

redesigned to facilitate expansion via module contributions. *Ecography* **2023**, e06547;

10.1111/ecog.06547 (2023).

39. Brown, J. L. SDMtoolbox: a python-based GIS toolkit for landscape genetic, biogeographic and

species distribution model analyses. *Methods Ecol. Evol.* **5**, 694–700 (2014).

40. Elith, J. *et al.* Novel methods improve prediction of species' distributions from occurrence data.

Ecography **29**, 129–151 (2006).

41. Phillips, S. J., Anderson, R. P., Dudík, M., Schapire, R. E. & Blair, M. E. Opening the black

box: an open-source release of Maxent. *Ecography* **40**, 887–893 (2017).

42. Lee, I.-M. *et al.* New subgroup 16SrIII-Y phytoplasmas associated with false-blossom diseased

cranberry (*Vaccinium macrocarpon*) plants and with known and potential insect vectors in New

Jersey. *Eur. J. Plant. Pathol.* **139**, 399–406 (2014).

43. Ross, H. H. A survey of the *Empoasca fabae* complex (Hemiptera, Cicadellidae). *Ann. Entomol.*
Soc. Am. **52**, 304–316 (1959).

44. Bertin, S. *et al.* Diffusion of the Nearctic leafhopper *Scaphoideus titanus* Ball in Europe: a
consequence of human trading activity. *Genetica* **131**, 275–285 (2007).

45. Saguez, J. *et al.* Diversity and abundance of leafhoppers in Canadian vineyards. *J. Insect Sci.*
14, 73; 10.1093/jis/14.1.73 (2014).

46. Franklin, J. Mapping species distributions: spatial inference and prediction (ed. Franklin, J.)
(Cambrige University Press, 2010).

47. Jarnevich, C. S., Stohlgren, T. J., Kumar, S., Morisette, J. T. & Holcombe, T. R. Caveats for
correlative species distribution modeling. *Ecol. Inform.* **29**, 6–15 (2015).

48. Fick, S. E. & Hijmans, R. J. WorldClim 2: new 1␣km spatial resolution climate surfaces for
global land areas. *Int. J. Climatol.* **37**, 4302–4315 (2017).

49. Dormann, C. F. *et al.* Collinearity: a review of methods to deal with it and a simulation study
evaluating their performance. *Ecography* **36**, 27–46 (2013).

50. Kelley, M. *et al.* GISS␣E2.1: Configurations and Climatology. *J. Adv. Model. Earth Syst.* **12**,
e2019MS002025; 10.1029/2019MS002025 (2020).

51. Calvin, K. *et al.* Climate Change 2023: Synthesis Report. Contribution of Working Groups I, II
and III to the Sixth Assessment Report of the Intergovernmental Panel on Climate Change (ed.
Core Writing Team., Lee, H. & Romero, J.) (IPCC, Geneva, Switzerland, 2023).

52. Hijmans, R. J. raster: Geographic Data Analysis and Modeling. R package version 3.6-23.
https://CRAN.R-project.org/package=raster (2023).

53. Kass, J. M. *et al.* ENMeval 2.0: Redesigned for customizable and reproducible modeling of
species' niches and distributions. *Methods Ecol. Evol.* **12**, 1602–1608 (2021).

54. Elith, J. *et al.* A statistical explanation of MaxEnt for ecologists. *Divers. Distrib.* **17**, 43–57
(2011).

55. Lobo, J. M., Jiménez-Valverde, A. & Real, R. AUC: a misleading measure of the performance

of predictive distribution models. *Glob. Ecol. Biogeogr.* **17**, 145–151 (2008).

56. Warren, D. L. & Seifert, S. N. Ecological niche modeling in Maxent: the importance of model

complexity and the performance of model selection criteria. *Ecol. Appl.* **21**, 335–342 (2011).

57. Allouche, O., Tsoar, A. & Kadmon, R. Assessing the accuracy of species distribution models:

prevalence, kappa and the true skill statistic (TSS): Assessing the accuracy of distribution

models. *J. Appl. Ecol.* **43**, 1223–1232 (2006).

58. Luke, D. A User's Guide to Network Analysis in R (ed. Luke, D.) (Springer International

Publishing, 2015).

59. Dray, S. & Dufour, A.-B. The ade4 Package: Implementing the Duality Diagram for Ecologists.

J. Stat. Soft. **22**, 1-20; 10.18637/jss.v022.i04 (2007).

60. Csárdi, G. *et al.* igraph for R: R interface of the igraph library for graph theory and network

analysis. R package version 1.6.0, https://zenodo.org/records/8240644 (2023).

61. Di Cola, V. *et al.* ecospat: an R package to support spatial analyses and modeling of species

niches and distributions. *Ecography* **40**, 774–787 (2017).

62. Oman, P. W. The Nearctic leafhoppers (Homoptera: Cicadellidae): a generic classification and

checklist. *Mem. Entomol. Soc. Wash.* **3**, 1–253 (1949).

63. Beirne, B. P. Leafhoppers (Homoptera: Cicadellidae) of Canada and Alaska. *Can. Entomol.* **88**,

1–180 (1956).

64. Krishnankutty, S. M., Dietrich, C. H., Dai, W. & Siddappaji, M. H. Phylogeny and historical

biogeography of leafhopper subfamily Iassinae (Hemiptera: Cicadellidae) with a revised tribal

classification based on morphological and molecular data. *Syst. Entomol.* **41**, 580–595 (2016).

65. Whitcomb, R. F., Kramer, J., Coan, M. E. & Hicks, A. L. Ecology and Evolution of leafhopper-

grass host relationships in North America (ed. Harris, K. F.) 121-178 (Springer New York,

1987).

66. Lawton, D. *et al.* Pest population dynamics are related to a continental overwintering gradient.

Proc. Natl. Acad. Sci. U.S.A. **119**, e2203230119; 10.1073/pnas.2203230119 (2022).

67. Bebber, D. P., Ramotowski, M. A. T. & Gurr, S. J. Crop pests and pathogens move polewards in

a warming world. *Nat. Clim. Change* **3**, 985–988 (2013).

68. Rossi, J.-P. & Rasplus, J.-Y. Climate change and the potential distribution of the glassy-winged

sharpshooter (*Homalodisca vitripennis*), an insect vector of *Xylella fastidiosa*. *Sci. Total*

Environ. **860**, 160375; 10.1016/j.scitotenv.2022.160375 (2023).

69. Van Nieuwenhove, G. A., Frías, E. A. & Virla, E. G. Effects of temperature on the development,

performance and fitness of the corn leafhopper *Dalbulus maidis* (DeLong) (Hemiptera:

Cicadellidae): implications on its distribution under climate change. *Agric. For. Entomol.* **18**, 1–

10 (2016).

70. Crossley, M. S. *et al.* Warmer temperatures trigger insecticide□associated pest outbreaks. *Pest*

Manag. Sci. ps.7832; 10.1002/ps.7832 (2023).

71. Salar, P., Charenton, C., Foissac, X. & Malembic-Maher, S. Multiplication kinetics of

Flavescence dorée phytoplasma in broad bean. Effect of phytoplasma strain and temperature.

Eur. J. Plant Pathol. **135**, 371–381 (2013).

72. Maggi, F., Galetto, L., Marzachì, C. & Bosco, D. Temperature-dependent transmission of

Candidatus phytoplasma asteris by the vector leafhopper *Macrosteles quadripunctulatus*

Kirschbaum. *Entomologia* **2**, 202; 10.4081/entomologia.2014.202 (2014).

73. Sabato, E. O., Landau, E. C., Barros, B. A. & Oliveira, C. M. Differential transmission of

phytoplasma and spiroplasma to maize caused by variation in the environmental temperature in

Brazil. *Eur. J. Plant Pathol.* **157**, 163–171 (2020).

Table 1. Selected model setup and performance metrics for 14 leafhopper species in North America. Feature class (FC) and regularization multiplier (RC) indicated the model setup in MaxEnt. Area under the operational curve (AUC) and true skill statistics (TSS) indicate model performance. MaxTSS refers to the probability threshold for binary conversion of the maps (unsuitable and suitable). SD = standard deviation (n = 4).

Species	FC	RM	AUC (SD)	TSS (SD)	MaxTSS
Amplicephalus inimicus	LQH	0.5	0.928 (0.012)	0.723 (0.030)	0.166
Ceratagallia humilis	H	0.5	0.980 (0.012)	0.828 (0.009)	0.066
Colladonus geminatus	LQH	1.5	0.918 (0.051)	0.700 (0.080)	0.057
Edwardsiana rosae	LQH	4.0	0.948 (0.018)	0.766 (0.081)	0.373
Empoasca fabae	L	0.5	0.883 (0.011)	0.706 (0.028)	0.341
Erythroneura comes	LQH	4.0	0.974 (0.013)	0.852 (0.077)	0.144
Erythroneura vitis	LQ	4.0	0.954 (0.006)	0.832 (0.017)	0.379
Erythroneura ziczac	LQH	1.5	0.939 (0.018)	0.738 (0.069)	0.222
Exitianus exitiosus	LQ	1.0	0.916 (0.007)	0.698 (0.005)	0.218
Graphocephala fennahi	H	4.0	0.939 (0.004)	0.840 (0.003)	0.247
Jikradia olitoria	LQ	0.5	0.888 (0.004)	0.816 (0.004)	0.219
Macrosteles quadrilineatus	LQHP	0.5	0.895 (0.017)	0.629 (0.051)	0.260
Paraphlepsius irroratus	LQHP	0.5	0.953 (0.011)	0.810 (0.054)	0.264
Scaphoideus titanus	LQ	0.5	0.979 (0.006)	0.864 (0.033)	0.050

639 **Figure 1.** Examples of Nearctic leafhopper genera: a) *Graphocephala* spp., b) *Erythroneura* spp.,

640 and c) *Empoasca fabae* (Harris). Joseph Moisan-De-Serres provided pictures from the Ministry of

641 Agriculture, Fisheries and Food – Québec (MAPAQ).

643   **Figure 2.** Occurrence records of 14 leafhopper species in North America after removing records with uncertainties and spatial thin (5 km).

Figure 3. The relative importance of the environmental variables for the current potential distribution models for 14 leafhopper species in North America.

648 **Figure 4.** Estimated leafhopper species richness based on 14 species potential distribution models

649 in North America.

651 **Figure 5.** Estimated leafhopper species richness based on 14 species potential distribution in North

652 America under four shared socioeconomic pathway scenarios [a) SSP1-2.6, b) SSP2-4.5, c) SSP3-

653 7.0, and d) SSP5-8.5) for 2041-2060.

Figure 6. A comparison of environmental similarity networks and their density among 14 leafhopper species in North America under current and four future scenarios: a) SSP1-2.6, b) SSP2-4.5, c) SSP3-7.0, and d) SSP5-8.5 for the period 2041-2060. The columns, represented in grey for the current scenario and orange for the four alternative scenarios, indicate the network density values. This metric reflects the proportion of observed connections relative to the network's maximum potential connections. Density is a ratio that ranges from 0 to 1, with higher values indicating a more interconnected network. The size of circles within the network graph represents the number of interactions between species, meaning larger circles signify more significant similarities among species.