Comparing random and timed mating of mice in the experimental paradigm

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Abstract

Mus Musculus has provided scientists a well-adapted animal model for the developmental studies. There

is one on one co-relation between molecular determinants such as genes between humans and mice

hence making them superior model to study gene expression and to do translation research. Swiss albino

mice strain of house mice was used in this study with comparisons of efficiency of pregnancies during

two different strategies of mating; i.e., more traditional random/blind mating and more recent timed

mating in experimental paradigms to get staged mice embryos. The workflow required for the blind

mating was less labor intensive and required minimal maintenance both time and human resource and

was easy to execute. On the other hand, timed mating strategy was ideal for getting staged embryos that

was efficient in giving pregnancies and was ideal for less equipped labs but however required more

maintenance and human resource. The results of comparisons of weight gain with pregnancies was

similar in two groups where some un-pregnant mice also showed average or above average weight gain

with time. The z-test analysis for difference between two proportions was employed to calculate

efficiency of pregnancies in the two groups and p value was found to be 0.06 which suggested that was

no significant difference between the two groups however, this might be due to small sample size that

was possible in our small setting. The accurate embryonic staging was required in the experiments

corresponding to the peak of neurogenesis in mice and embryos were fixed and sectioned and later

studied with immunohistochemistry. The timed matings showed to be superior regarding total

pregnancies and having accurate stages and is recommended in labs even if in smaller settings and

prevent less animal sacrifice.

Introduction

The scientific name of house mice is *Mus Musculus* and it belongs to kingdom Animalia, phylum chordate,

class Mammalia and order Rodentia. Mice are preferred in Genetics and Molecular biology research

because they show great similarity with humans with respect to primordial genetic, physiological and

anatomical components (1). To investigate human health almost 59% animals used in biomedical research

are mice (2). Short life cycle, short gestation period and high breeding efficiency make them an excellent

animal models in laboratory settings (3). Moreover, mice are small, easy to handle and can be transported

easily. They are used to study human diseases and human genetics because they are phylogenetically

related to humans (4). Gestation periods of mice is between 18 to 22 days (5). The developmental biology

studies require accurate staging of embryos that can help with comparative developmental studies

between mice with mice or mice with different species known as evo-devo approach (6). Almost 95% of

the genes of human and mice are similar so it is an excellent model to do gene expression studies involving

transcriptional analysis (7). To get staged embryos mice may be mated randomly or at specific stages of

estrous cycle. We anticipated to compare both strategies in smaller settings with respect to efficiency of

pregnancies and accurately staged embryos for downward analysis. It looked apparent that the mating

on the day of ovulation; i.e., at the proestrous-estrous and visualization of plug the next day will lead to

more efficient pregnancies and also accurate staging if pregnant. The estrous cycle of female mice consists

of four stages namely proestrous, estrous, metestrous and diestrous (8). There are various methods to

evaluate the estrous cycle of mice that include visual observation (9), vaginal cytology (10), histology of

reproductive organs (8), vaginal wall impedance (11) and biochemistry of the urine (12, 13). We employed

visual observations of the vagina and vaginal cytology combinations and set out on this comparative study.

Material and Method:

53   Animal acquisition

54   The collaboration letter was signed between the Department of Biochemistry and Department of

55   Pharmacy to do the animal work. The Islamia University of Bahawalpur's Ethical committee approved the

56   use of animals in the work via letter no. 3990/ORIC. We acquired albino house mice, BALB/c that were

57   enrolled in the study. We enrolled mice n=15 blind/random matings experiment and n=15 in second

58   experiment of timed matings in mice. Male to female ratio was 1:2.

59   Housing of the animals

We set up two experiments different experiments scheduled at different time periods to get timed

embryos at different embryonic days (from E12.5-E18.5). In the first experiment we took 10 female mice

in a separate cage and housed 5 stud males mice each in a separate cage to increase their sperm level,

fertility and also to avoid their aggressive behavior (14). We used wooden cages that were custom made

and had a proper fresh water supply with adjusted bottles. The pelleted food was given which was made

up of ground ingredients. We used fine wooden scrap for the cage beddings. The cages were placed in the

air-conditioned environment to maintain the optimum temperatures in summers. In second   experiment

we used the same setting and numbers but were analysed for estrous stage.

68   Experiment 1: Random or blind mating

69   1.   Initially, female mice were labeled by using green and black permanent makers. One, two, three

70   and four black bands on the tail of mice represent mice number 1, 2, 3 and 4 respectively while

full black tail show mice number 5. Similarly one, two, three and four green bands represent

mice number 6, 7, 8 and 9 respectively. Further full green tail was identified as mice number 10.

2. Body weight of all female mice was measured in the start of the experiment. Then all female

mice were shifted to the male cages in1:2 male to female mice ratios. Vaginal plug was checked

in next day early in the morning. The female mice in which vaginal plugs were observed were

separated from the males and were separated.

3. Weight of all female mice were then recorded on the daily basis (whenever possible).

4. At the desired embryonic day after the separation the mice were dissected to collect the

embryos (the day of plug was considered 0.5 embryonic day) and adult tissues.

Experiment 2: Timed mating

1. Method of labeling the mice in this trial was same as above and the weights of each female mice

were also recorded on daily basis (whenever possible).

2. To evaluate estrous stage of mice we used vaginal cytology protocol (as per Byers et al. 2012 (8)

using saline dropper method and but the smear staining was done by crystal violet staining and

visualized by light microscopye) combined with visual observations under light was done (8).

3. Female mice that were in their pro-estrous to estrous stages were set for mating.

4. The next day, in the early morning, the female in which vaginal plug was visible were separated

and dissected at the required embryonic day to get staged embryos (the day of plug was

considered 0.5 embryonic day).

5. The remaining female mice were also mated using same strategy until all mice were mated and

dissected respectively.

Dissection of mice:

On the desired embryonic day the mice were dissected to collect the embryos from the pregnant

dams. For downward experimentation the embryos were collected either in RNA later or fixed in

formalin and some embryos were also snap frozen. We did this for future work.

Statistical analysis

Two proportion Z-test was used to calculate the efficiency of pregnancies in the two independent

groups and p values were calculated.

Immunohistochemistry

The formalin fixed paraffin embedding and sectioning was done and later stained by

Immunohistochemistry using established (15) and improved protocols. DAPI (4',6-diamidino-2-

phenylindole) a counterstain that binds to A/T rich DNA bases in the grooves is used where it marks

the complete architecture and cellular organisation in the tissues. Embryonic tissues at various stages

were stained.

Results

Weight dynamics and pregnancies

Weight gain is one of the hallmarks of pregnancy generally but in a small animal like mice it sometimes is

not relevant. Here, we analysed weight dynamics in both the experiments oin the first 12 days in mice

(the starting embryonic day at which embryos were anticipated to be collected) and was found to be not

related to the induction of pregnancy. The weight readings considerably increase in pregnant mice after

day 12 of pregnancy and would give false indication if weight dynamics were matched of E12.5 and E18.5

mice hence the period was kept constant. Female mice were also plotted for total weight gain calculated

from the initial weight at the start of trial and before dissection and was plotted (see figure 1). Mice in

both groups showed weight gain ranging from 4g to 18g but found to be not co-related directly with

pregnancy (see tables 1, 2 and 3). However, mice with higher than 6g of net weight gain corresponded

well with pregnancies.

Weight gain was observed in each mice, though it did not reflected true pregnancy (Table 1).

Mice No	Initial weight (g)		Final weight before dissection (g)		Net weight gain (g)	
	Exp1	Exp2	Exp1	Exp2	Exp1	Exp2
1	34	22	38	28	4	6
2	32	14	36	20	4	6
3	32	26	36	30	4	4
4	34	26	42	42	8	18
5	24	14	38	20	14	6
6	26	28	34	28	8	0
7	Paralyzed	Died				
8	24	30	34	40	10	10
9	24	24	32	32	8	8
10	26	22	36	40	10	18

118   **Table1:** Correlation of mice with weight during the two experiments

Figure no. 1: A, Weight dynamics in female mice during blind mating trial from Day1 to Day12. **B,** Total weight gain in mice throughout the blind mating trial. **C,** Weight dynamics in female mice in timed mating trial from Day1 to Day12. **D,** and Total weight gain in mice in timed mating trial

Blind mating trial and number of pregnancies

(Blind mating experiment of 10 female mice with 5 stud male mice took place from 16-05-22 to 20-06-

22) (see table 2). The vaginal plug was observed once and female mice were separated but it led to false

pregnancies or/and non-pregnant mice in more than half of female mice that were sacrificed. In total we

got 40% pregnant rate (see table 2).

MICE No.	Pregnancy induced after blind mating	Stages in days	Adult tissues collected	Embryos collected
1	X	NA	√	NA
2	X	NA	X	NA
3	X	NA	X	NA
4	√	E 6.5	√	√ (whole oviducts)
5	√	E 12.5	√	√
6	√	E 13.5	√	√
7	Died	NA	X	NA
8	X	NA	√	NA
9	√	E 18.5	√	√
10	Died	NA	X	NA

Table 2: Random or blind matings and the success in pregnancies and collection of samples

Timed mating trial setup and number of pregnancies

(Timed mating experiment took place in two phases from 24-03-23 to 10-05-23 and later in 2024). As

above total of 10 female mice were mated with 5 stud male mice (Table 3)

Visual observations of the vagina in female mice are different at different estrous cycle stages and

hence maybe used to assess the cycle stage (Figure 2A). In pro-estrous stage the vaginal tissues and

surrounding tissues appear pink, moist and swollen with a wide opening and with wrinkles in dorsal

and ventral edges whereas, at estrous stage the vaginal tissues are relatively less swollen and are not

pink or moist and are ideal for mating. However, during metestrous stage, the vaginal opening is small

and has minor swelling and at diestrous no opening and swelling of vagina was observed (Figure 2A).

In addition, the vaginal cytology of mice at various estrous cycle stages may be done to determine

the estrous cycle stages (Figure 2B). The vaginal cytology at pro-estrous stage showed majority of

nucleated cells and small numbers of cornified cells whereas the estrous stage was revealed by large

numbers of cornified epithelial cells. The metestrous was marked with cornified epithelial cells plus

polymorphonuclear leukocytes with some nucleated epithelial cells and in diestrous, which lasts for

2 days and is the longest stage in estrous cycle there are more polymorphonuclear leukocytes (see

figure 2B). The combination of both the observations was done to identify female mice at pro-estrus

to estrous stage that lasts up to 15 hours and represent the day of ovulation and were set to mating

with stud males (see supplemental data). The female mice with vaginal plug were separated next day

to get high efficiency of pregnancy and accurately staged embryos. The number of pregnancies as

expected were quite high as anticipated and was 80% in this group (see table 3).

Proestrous   Estrous   Metestrous   Diestrous

Proestrous   Estrous   Metestrous   Diestrous

159 **Figure no. 2: A,** The shape of vagina changes at various stages in the mice estrous cycle through visual
160 observations, **B,** The stages are visible after crystal violet dye staining of the vaginal smear at various stages
161 in the mice estrous cycle

162
163
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MICE No.	Pregnancy induced after timed mating	Stages in days	Adult tissues collected	Embryos collected
1	√	E 14.5	√	√
2	√	E 7.5	√	√ (Whole embryos)

3	√	E 13.5	√	√
4	√	E 14.5	√	√
5	√	E 15.5	√	√
6	√	E 15.5	√	√
7	Died	NA		
8	√	E 13.5	√	√
9	√	E 13.5	X	√
10	X	NA	X	X

Table 3: Timed pregnant mice results showing more pregnancies and accurate embryonic stages

Two proportion z- test analysis to calculate statistical difference if any in rate of pregnancies in both groups

A two proportion z- test is used to calculate the true difference between two independent groups and to a given confidence interval. We used this analysis to calculate statistical difference in the rate of pregnancies in both groups. Our data was shown to have p value of 0.06 and correspond with the null hypothesis which states both groups are same hence there is no statistical difference in these two mating strategies in terms of efficiency of pregnancy in both groups (Figure 3). This may be due to small sample size in the two groups. This was unavoidable due to small laboratory setting and availability of mice.

A

$$z = \frac{p_1 - p_2}{\sqrt{p(1-p)(1/n_1 + 1/n_2)}}$$

C

P1 denotes the proportion for population 1

P2 denotes the proportion for population 2

B

	Blind matings	Timed matings
Died	1	1
Pregnant	4	8
Not-pregnant	5	1
Mice number total	10	10
Rate of pregnancy	40%	80%
Total pregnant	12	
Total non pregnant	8	
Total died	2	
Sample proportion	60%	
Z- test	1.82	
p value	0.068759	

D

178 **Figure no. 3: A,** z test to calculate p values for two proportions analysis. **B-C,** P values calculated for
179 number of pregnancies between blind and time mating was done. **D,** 0.06 p value between two groups
180 support null hypothesis is correct.
181
182

183 **Embryonic Stages collected and downwards analysis:**

184 Embryonic stages were collected and were verified by characteristic feature of embryos at respective

185 stages (see figure 4).

E13.5 Embryo

E 14.5 Embryos

E15.5 embryos

Figure no. 4: The embryo stages collected during the experiment E13.5, E14.5 and E15.5 (Left to right.

The embryo at 13.5 stage (see Figure 4) has foot plate that is anterior and has posterior distal borders that

are sectioned. Hand plate has more clearly sectioned than foot plate. In hind limb only ankle region is

identifiable while in forelimb elbow and wrist are distinguishable. Furthermore pinna and whiskers are

appearing.

At E14.5 embryonic stage (see Figure 3) the separation of individual digits in the anterior footplate is a

unique feature of this stage. Although there isn't yet any discernible separation between the growing

toes and the rear footplate exhibits significant indentations.

At embryonic stage E15.5 (see Figure 4) there was complete interdigital indentation is the hand and foot

plates, and the vanishing of the webbing improves the delineation of each individual digit. The elbow and

knee are obviously bent. Hair follicles can be found in the cephalic region, but not along the margins of

the vibrissae. The anterior area of the back is gray, and the curvature of the back is markedly different

from the currently straight post-cranial vertebral axis.

Immunohistochemistry to confirm the staging of embryos

Downwards analysis for confirmation of embryonic stages was done using formalin fixation and paraffin

embedding followed by sectioning and immunohistochemistry. DAPI (4',6-diamidino-2-phenylindole)

staining was used to assess the stage by developing neuroepithelium which shows overall architecture of

all cell nuclei. The figure 5 shows that E12.5, E13.5 and E14.5 embryos showing developing

neuroepithelium where the formation and organization of layers confirmed the stage.

Figure no. 5: Immunohistochemistry of paraffin embedded sections of mice embryos showing developing

neuroepithelium at the peak of neurogenesis is shown that were acquired through the timed mating

experiment.

Discussion:

In the series of experiment done in this study we got contradicting results showing no/very little

dependency on weight gain with the successful pregnancies. Average net weight gain in female mice in

both random and blind matings was 4g regardless of pregnancy induction. However in the pregnant mice

females with net weight gain of higher than 6g correlated with higher proportions of pregnancies in both

groups. The two trials were done at different times but in the same settings. The pregnancy number was

two folds in timed matings as compared to random/blind mating which was evident from mice behaviour

and other features in addition to weight of female mice but not only weight gain. This was due to the

mating exactly at the day of ovulation which is at pro-estrous/estrous stages and increases chances of

pregnancy. In blind matings we were only sure of staging after dissecting out embryos and didn't always

find the required stage which maybe due to dropping out of vaginal plug and mating again however the

timed mating experiment took more time and was labour intensive corresponded well with efficient

pregnancies. The values of weight gains in both experiments were almost comparable but just on the

criterion of weight it is hard to judge the pregnancy and more so the exact stage we are looking for as was

our requirement in our study and hence were dependent on the observation of vaginal plug. In one study

the early pregnancy around E7.5 was found to be more related to significant weight gain compared with

non-pregnant mice (16) our data doesn't support it. The time taken to finish the first experiment was 4

weeks whereas timed mating took almost two months to complete due to staging confirmation prior to

sacrifice and required more housekeeping of mice and care for the animal requirements but inturn gave

effcient pregnancies and staging which was confirmed by downwards immunohistochemistry analysis

using DAPI stain.

The second question was whether we can statistically calculate difference between the proportions

of pregnancies in the two groups. For this we used, Two proportions Z-test and although looking quite

different in numbers the p value came out to be 0.06 which is not statistically significant and supports the

null hypothesis which states both groups have same rate of pregnancies. This may be due to small sample

size which was unevitable due to small setup and was unavoidable and might not reflect the true

difference. In previous work by Byers et al. (2012) and others (17,18) with more than 100 sample size and

mating variations and and some small group mating trials, there was some statistical difference in the two

groups which seem to be the true reflection of the strategies in large cohorts. Our numbers also reflect

this but states are different because of very small sample size. In bigger lab settings, with high resolution

microscopy random/blind mating might be continued to be used as traditionally used where staging of

embryos maybe confirmed by developmental hallmarks but in small setting, timed mating is relatively a

method of choice for staged embryos and efficient pregnancies and might be adopted.

Conclusions:

The timed matings gave us more pregnant mice at desired stages than mice with blind matings in terms

of numbers however statistically there was no significant difference between two groups and supports

null hypothesis. However, it might still be adopted in smaller setting to get effieient and accurate

staging of embryos.

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Supplementary Data:

A- Blind mating weight readings

Mice no	D1	D2	D3	D4	D5	D6	D7	D8	D9	D10	D11	D12	D13	D14	D15	D16	D17	D18
1	34		32	36	36	36		36	36	36	36	38			38	38	38	
2	32	32		34	32	34		36	34	34	36	36			36	36	36	
3	32	34		36	36	36		34	36	36	36	34			34	36	36	
4	34								38	38	36	38				40	42	
5	24								28	30	30	30				36	38	
6	26								30	28	28	30				32	34	
7	28								30	30	32	32				32	32	
8	24			32	28	28		32	32	32	32	34				36	34	
9	24								28	28	30	30			34	32	32	
10	26		28	30	30	30		30	32	32	36	34			36	34	36	

B- Timed mating weight readings

Mice no.	D1	D2	D3	D4	D5	D6	D7	D8	D9	D10	D11	D12	D13	D14

1	22			24	24			24	24	24	28	28		
2	14		14	14				18	18	18	20	20		
3	26		28	26	32			32	30	30	30	30		
4	26	32	34	36	38			42	42	40	42	42		
5	14		14	14				18	22	20	20	20		
6	28			30	30			30	30	28	28	28		
7	18													
8	30			30	30			36	38	38	38	40		
9	24	26	26	28	29	32	32	30	30	32	34	32	32	32
10	22	24	24	26	27	28	28	30	32	36	36	38	40	

C- Vaginal cytology and setting up of matings

Mice no	D1	D2	D3	D4	D5	D8
1	Diestrous	Diestrous	Diestrous	Estrous (Mating)		
2	Metestrous	Metestrous	Metestrous	Metestrous	Diestrous	Estrous (Mating)
3	Diestrous	Diestrous	Metestrous	Diestrous	Diestrous	Estrous (Mating)
4	Estrous (Mating)					
5	Metestrous	Metestrous	Diestrous	Diestrous	Metestrous	Proestrous(Mating)
6	Diestrous	Diestrous	Diestrous	Proestrous (Mating)		
7	Paralyzed					
8	Diestrous	Diestrous	Diestrous	Estrous(Mating)		
9	Diestrous	Diestrous	Estrous (mating)			
10	Diestrous	Metestrous	Proestrous (mating)			

Visual observations of vagina in estrous cycle

Proestrous   Estrous   Metestrous   Diestrous

Vaginal cytology using crystal violet stain in estrous cycle

Proestrous   Estrous   Metestrous   Diestrous

A — Weight dynamics with time during blind mating strategy

B — Total weight gain in mice throughout the blind mating trial

C — Weight dynamics in mice with time in blind mating strategy

D — Total weight gain in mice throughout the timed mating trial

A

$$z = \frac{p_1 - p_2}{\sqrt{p(1-p)(1/n_1 + 1/n_2)}}$$

C

P1 denotes the proportion for population 1

P2 denotes the proportion for population 2

B

	fixed settings	focal settings
Died		
Pregnant	6	4
Not pregnant	5	4
Insemination- total	11	10
Rate of pregnancy	55%	40%
New pregnant	13	
Total not pregnant	9	
Total died	1	
Transfer procedures	45%	
z score	1.87	
p value	0.003793	

D

The comparison of number of pregnant after diatery mating strategies

P value = 0.06

E13.5 Embryo

E 14.5 Embryos

E15.5 embryos

DAPI

| 20X | 20X | 10X |

E12.5 Mouse Neuroepithelium   E13.5 Mouse Neuroepithelium   E14.5 Mouse Neuroepithelium