3D-Printable Non-invasive Head Immobilization System for Non-Human Primates

Elia Shahbazi [1], Drew Nguyen [1], Tyler Swedan [2], Timothy Ma [3], Rosa Lafer-Sousa [1], Alvin Dinh [1], Reza Azadi [1], Amy M. Ryan [1], Arash Afraz [1]

1 Laboratory of Neuropsychology, National Institutes of Health, National Institute of Mental Health, Bethesda, MD
2 Goucher College, Towson, MD
3 Center for Neural Science, New York University, New York, NY

Abstract

A critical contribution to understanding the primate brain is our ability to directly record, manipulate, or image the brain in awake, behaving monkeys. A challenge with carrying out these studies is that typically the monkey's head must be fixed in place. While non-invasive head immobilization systems (NHIS) have been demonstrated to be effective, they have not fully replaced the use of surgically applied headposts. Here, we introduce a novel NHIS, with an option for voluntary engagement, for macaques using a widely available resource: the 3D printer. We designed customized 3D-printable head-immobilization masks for monkeys and a docking system using standard CT scans and user-friendly open-source software packages (FLoRIN, Blender, Python). We examined the efficacy and stability of our mask system for collecting eye fixation data by measuring trial-by-trial gaze precision and accuracy in two mask-immobilized monkeys and three headposted monkeys, including a within-subjects comparison. Our results demonstrate that monkeys stabilized by the mask maintain gaze precision and accuracy comparable with their headposted counterparts. While the headpost outperformed the mask, the difference in precision was 0.03° of visual angle, and the difference in accuracy was less than 0.2°, well within the acceptable range for most use-cases. Our process allows for the production of 3D-printable masks based on CT or MRI scans with the flexibility to incorporate design modifications for experimental equipment and resizing, potentially serving as a promising and more easily accessible alternative to headposts and other NHISs for primate neuroscience research.

Introduction

Nonhuman primates, like us, use vision as their primary sensory modality to navigate the world, create memories, and make decisions. Given the functional and anatomical similarity between human and nonhuman primate visual systems (Van Essen et al, 2001; Orban et al, 2004; Lafer-Sousa, et al, 2016; Lu et al, 2023), nonhuman primates have long-served as a valuable model for the human visual system. Studies in nonhuman primates enable the execution of

experimental paradigms that are not feasible in humans, leveraging the most precise and
advanced tools available to neuroscience to measure and perturb neural activity and behavior.
While it is possible to carry out some neurophysiology studies in unrestrained monkeys, as is
done in certain behavioral studies of monkeys with experimentally induced brain lesions (Vaidya
et al., 2018) or those utilizing emerging technologies of wireless neural recording, still very much
in their infancy (Dal Monte, et al., 2022; Testard, et al., 2024), the majority of experimental
paradigms involving awake, behaving monkeys require that the head be absolutely still.
Research that involves eye-tracking and fixation, electrophysiological recording (Wang et al.
2023; Roussy et al. 2022; Hallum et al. 2017; Lemon, 1984), optogenetic perturbation (Shabazi
et al., 2024; Azadi et al. 2023b), or functional imaging (Goense, et al., 2010; Lafer-Sousa &
Conway, 2013; Tanigawa, et al., 2010) typically necessitate continuous immobilization with
minimal head movement for reliable results.

The foundational work of Evarts (1968) and Mountcastle et al. (1975) marked the advent of
long-term head fixation techniques. They employed stainless steel bolts anchored in the skull to
affix an acrylic head cap. Innovations from various laboratories (Betelak et al. 2001; Evarts
1968;Foeller and Tychsen 2002) have sought to eliminate the acrylic cap. Building on these
advancements, Adams et al. (2007) introduced a titanium headpost that is more securely
anchored to the skull. This contemporary approach has been rapidly adopted and is now
prevalent in most research laboratories. Further surgical innovations have been pursued, such
as a low profile halo head fixation system (Isoda et al., 2005; Pigarev, Saalmann, and
Vidyasagar 2009; Azimi et al, 2016), but have not been widely adopted.

Despite advancements in headpost design, the approach of surgically attaching a device to the
skull inherently carries risks and disadvantages. First, undergoing surgery presents health risks
due to anesthesia and postoperative recovery, particularly with respect to infections.
Unfortunately, intracranial infection from the headpost is a persistent risk, requiring continual
monitoring, cleaning, and maintenance by researchers. Even with this monitoring, headposts
can still fail due to infection, necrosis, or bone softening around the headpost and the high
torque applied during experiments. Second, these risks are disadvantages to the research
project and timeline, because surgery requires extensive physical and personnel resources,
cost, and recovery time as the headpost integrates into the bone.

Given these disadvantages and risks, multiple laboratories have independently developed
noninvasive approaches to head immobilization. Alternatives have been used in fMRI imaging
(Srihasam et al., 2010; Hadj-Bouziane et al., 2014; Drucker et al., 2014), electrophysiology
neuronal recordings (Drucker et al., 2014; Amemori et al., 2015) eye fixation tasks (Drucker et
al., 2014 Amemori et al., 2015; Machado and Nelson, 2011; De Luna et al., 2014; Rima et al.,
2024) and electroencephalographical (EEG) recordings (Nakamura et al., 2020). Most of these
head fixation techniques use thermoplastic frames to mold around the precise shape and
contours of the monkey's head to create a customized helmet (Machado and Nelson, 2011;
Drucker et al., 2014; De Luna et al., 2014; Slater et al., 2016; Nakamura et al., 2020), which is a
similar approach used in human neuroimaging. Most of the research groups lightly sedate the
animal for the molding process (Machado and Nelson, 2011; Drucker et al., 2014; Nakamura, et
al., 2020), although others create a 3D-printed model of the monkey's head from MRI scans
and use that to mold a thermoplastic frame (Slater et al., 2016), or initially create a mold on a
doll's head and then conduct minor modifications on the awake monkey (De Luna et al., 2014).
Other alternative head fixation approaches use negative air pressure (Srihasam et al., 2010;

Hadj-Bouziane et al., 2014), aquaplast moldable plastic (Amemori et al., 2015), and 3D-printed
chin rests (Rima, et al., 2024).

While there has been a consistent push for noninvasive head fixation techniques, there has not
been widespread adoption of the previously described approaches, and headposts remain the
predominant method used across research groups. Perhaps this is because the previously
mentioned methods still pose some challenges. First, most options use materials that are not
commonly used in a primate neuroscience laboratory, such as thermoplastics and plaster, thus
requiring both time and training to acquire and learn how to use them. For researchers already
familiar with or even trained in surgical approaches, this learning curve can be perceived as
cumbersome and time-consuming. Secondly, every mask has to be created anew as monkeys
age and grow. These systems thus are wanting for easy reproducibility to accommodate
changes in the monkey's head size over time, presenting an obstacle to consistent long-term
use. Third, thermoplastics are shaped by heating and molding, which limits the precision of fine
anatomical details, especially for intricate surfaces like those of the face or head. Finally,
thermoplastics readily degrade and lose their shape over time.

Our head fixation system uses an emerging technology that is increasingly becoming prevalent
in primate neuroscience laboratories or within their larger institutions: the 3D printer. For
example, Rima and colleagues (2024) recently used 3D printing to create a chin-rest system.
Here, we expand this concept to create a 3D-printed mask that can fully immobilize the
monkey's head. Utilizing CT scans of the monkey's head and our custom open-source software,
FLoRIN (Shahbazi et al. 2018), we engineered a 3D-printable mask system. The approach can
also be carried out using anatomical MRI scans. This mask system allows for hi-precision fit,
long-term durability, and custom modifications to accommodate experiment-specific needs (e.g.
eye-tracking equipment, reward delivery mechanisms, and physiology experiment hardware).
The mask can also be readily modified and reprinted to accommodate changes in head shape
over time. Similar to Slater et al. (2016), our system comprises a front piece (the mask), which
can be used alone for voluntary engagement, and a back piece, which can be attached for full
head-immobilization.

After developing this system and demonstrating the feasibility of the production pipeline, we
sought to evaluate the efficacy of our mask system for primate neuroscience research. As with
the previously discussed alternatives to headposts, we measured our timeline with respect to
training monkeys for mask use and then tested whether monkeys wearing masks could match
the precision of headposted monkeys on eye fixation tasks. Finally, we report on the success of
long-term data collection using our system.

Methods

All procedures were conducted in accordance with and approved by the National Institute of
Mental Health Animal Care and Use and Committee guidelines.

Subjects

We designed masks for two adult male rhesus macaques named Nd (5 years old, approx. 8 kg)
and Gr (8 years old, approx. 14 kg). Nd had previously been implanted with an Opto-Array
(Blackrock Neurotech; Rajalingham et al, 2021) over cortical area V4, which had been virally

transduced with the depolarizing opsin C1V1, resulting in a cereport pedestal (Blackrock
Neurotech) mounted on top of the head, offset from the midline, following procedures similar to
those detailed in Azadi et al (2023a). Gr was previously implanted with two chronic multi-
electrode Utah Array in the central IT cortex, following similar procedures as in Kar, et al.
(2019). This resulted in a custom titanium pedestal housing 4 headstage ports mounted on top
of the head, slightly offset from the midline.

Nd and Gr are the two monkeys in our population who had traditional headposts that failed, in
part inspiring the development of our mask project. For both monkeys, while they had the
headpost, they were trained in head fixation by securing the headpost to their testing chair.
Furthermore, they went through our established training protocol to learn to fixate on targets on
a computer screen for liquid reward. Nd did not advance far enough in his training to collect eye
fixation data. However, while Gr had a headpost, we collected many sessions of data over the
course of more than 6 months. Because Gr previously had a headpost and fixation data had
been collected, we had the opportunity to conduct a within-subject analysis on precision of eye
fixation between a headposted monkey and that same monkey whilst it used the mask.

To test the precision of eye fixation while wearing the mask, we compared Nd and Gr's eye
fixation precision with two other adult males who were similarly trained on an eye fixation task
but had a traditional surgically applied headpost, Ph (7 years old, approx. 8.5kg) and Kr (5 years
old, approx. 6.8kg). Ph and Kr were implanted with both an Opto-Array over central IT cortex,
virally transduced with the depolarizing opsin C1V1, as well as a titanium headpost, again
following procedures detailed in Azadi et al. (2023a). This resulted in an acrylic headcap on the
midline housing both a cereport pedestal and the caudally located headpost in Kr, and two
separate acrylic islands housing the cereport and headpost in Ph.

Mask Creation

Making a 3D base model

The first step of mask creation is the generation of a 3D base model of the monkey's head. To
generate the 3D base model for our mask, we use a structural scan of the monkey's head,
which can be a standard CT or T1-weighted MRI. For Nd and Gr, CT scans were used because
the animals had existing implants that were not MR-compatible. For each scan, prior to
sedation, animals were premedicated with glycopyrrolate (0.015mg/kg) and ketoprofen
(1mg/kg). The animals were then sedated with ketamine (10mg/kg) and dexmedetomidine
(0.02mg/kg) and positioned in the scanner. We scanned monkeys on an Epica Vimago HU CT
scanner using a 0.2mm resolution available at the NIH Neurophysiology Imaging Facility.
Following the scan, monkeys were recovered with revertidine (0.02mg/kg).

We then employed FLoRIN, open course software originally developed for reconstructing
neurons and membranes from high-resolution microscopy images (Joesch et al. 2016; Shahbazi

2018, shahbazi et al. 2018), to reconstruct the monkey's head, creating a precise 3D base
model for designing the monkey's mask, as shown in Fig 1a.

Our custom FLoRIN pipeline is a key innovation in our approach. First, FLoRIN can quickly and
easily transform any standard volumetric structural scan into a highly detailed 3D reconstruction
of the animal's head, including the soft tissue structures of the face. Second, FLoRIN's 3D
modeling accurately reflects the current physical state of the monkey's head, including potential
surgical implants. Third, FLoRIN provides compatibility with Python. This, in conjunction with our
choice of software for 3D modeling, Blender, allows for the use of Python scripting to convert a
series of scan data into 3D models of masks with ease and efficiency.

Mask Design

Blender, a software that allows sequences of 3D modeling operations to be exported as code,
facilitates mask creation. In Blender, we design a surface with strategically placed cutouts for
the eyes, nose, mouth, ears, and top of the head, as well as supports for facial bone structures
like the cheekbones, lower jaw, and brow ridge. The designed surface is then conformed to the
3D base model of the monkey's head using the shrinkwrap modifier, maintaining a default
distance of 1mm from the monkey's face. Following fitting to the base model of the monkey's
head, the mask's cutouts and supports are then refined and adjusted as needed. The model
code can be modified to accommodate diverse experimental hardware devices as needed. In
particular, our design leaves the top of the head unobstructed, facilitating the use of commonly
implanted devices for optogenetics, electrophysiology, or other cranial chambers and
headstages. Figure 1b shows the 3D model of the mask with the 3D base model of the
monkey's head, and Fig 1c shows the 3D printed mask. Figure 1d illustrates the mask in use
with a liquid reward delivery system.

In order to attach the mask to the primate testing chair (Crist Instrument Co., Inc.), we used
Blender to add connecting components to the 3D model of the mask and to design a 3D-
printable medial-diapason-shaped connector (Figure 1e), which mounts on a standard
adjustable head post holder (Crist Instrument Co., Inc.). Figure 1f shows the 3D printed mask
and medial-diapason-shaped connector attached to the primate chair, which is assembled using
nuts, bolts, and washers.

To enable full head immobilization, we used Blender to design a back piece that can attach to
the mask from behind and secure the head in the mask (Figure 1g). The back piece design
comprises a buttressed curved head support that makes contact along the skull's occipital crest,
as well as a handle and connectors that enable attachment to the mask with nuts, bolts, and
washers. During use, a foam pad is attached to the head support and covered with an
impermeable disposable shoe cover.

The back piece and connectors are fully customizable based on the monkey's head position,
implants, and chair system. Figure 1h shows the full 3D-printed mask assembly mounted on a
standard head post holder (Crist Instrument Co).

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Figure 1 - Overview of NHIS Design and Implementation: a) 3D base model (head reconstruction)
using FLoRIN based on a standard CT scan of the monkey's head, providing a detailed and accurate
model of the subject's facial anatomy. b) Design of the NHIS using Blender, illustrating the mask's
structure and various connecting components. c) 3D-printed mask d) Illustrative depiction of the mask in
use with a reward delivery system and without the back piece. e) Design of the medial-diapason-shaped
connector piece with mask. f) 3D printed mask and medial-diapason connector assembled and mounted
on a vertical primate chair (Crist Instrument Co.). g) Design of the back piece for full head-immobilization.
h) 3D printed full mask system assembly: includes the mask, back piece, medial-diapason-shaped
connector piece, attached to a primate head post holder (Crist Instrument Co.), assembled with nuts,
bolts, and washers.

3D Printing and Post-processing

For this project, we used the Stratasys F370 3D printer, which offers a print precision of up to
0.127mm (0.005 in.). The material selected for printing was Acrylonitrile Styrene Acrylate (ASA),
known for its high durability and versatility in 3D printing applications. ASA exhibits high impact

resistance, robust mechanical performance, and strong resistance to UV rays, water, and
various chemicals (Carlota 2023). Post-printing, we sanded the mask surface to ensure a
smooth surface and double-coated the mask with a resin seal to ensure it could not absorb
contaminants and could be properly disinfected between uses. The resin seal also increases
durability.

Training phase

Once we had a mask ready to introduce to a monkey, we created a training plan to help the
monkeys learn to place their head in the mask while seated in their testing chair. The animals
were on water control so that we could control the timing and amount of fluid intake. Each
monkey's weight and hydration level were monitored closely and maintained throughout training
and data collection. For training, we first desensitized the monkey to the mask, familiarizing
them with the object itself by holding the mask in front of the monkey while they were seated in
the testing chair. In this session we also paired mask presentation with food and liquid reward to
create a positive association. In the following training sessions, we brought the mask closer to
the monkey's face and threaded a straw through the mask or fruit held with long tweezers, so
that the monkey would learn to associate moving its head toward the mask with receiving a
reward. We then progressed to providing continuous liquid reward while the monkey tolerated
affixed mask placement and long-term wear. Within four days, both monkeys willingly engaged
with the mask for extended periods to receive juice and water.

The next phase of training was to introduce the back piece, which would allow for full head
immobilization ("full mask system"). For this phase, we first placed the affixed mask on the
monkey for continuous liquid reward and then progressively desensitized the monkey to the
presence of the back piece through to full attachment of the back piece.

After this step, Nd voluntarily (using the mask without the back piece) began maintaining eye
fixation on a computer display in exchange for continuous liquid rewards, while Gr immediately
began data collection using the back piece. In sum, Nd and Gr required ten and six days of
initial exposure to the mask, respectively, before they were sufficiently acclimatized, allowing us
to collect data. Figure 2 delineates this behavioral training timeline for mask system
stabilization, providing a detailed overview of each acclimatization and training process step.

Figure 2—Behavioral training with mask: The training process using our 3D-printed mask system
began with familiarizing the monkeys with the mask and employing positive reinforcement for placing their
head in the mask. Monkeys were then introduced to the back piece for full head immobilization (in the 6th
session for Nd and the 3rd session for Gr). In the 8th session, Nd commenced visual fixation training for
continuous rewards without the back piece (voluntary configuration). After two sessions data collection
commenced (10th session) in the voluntary configuration (data presented in results section 1).
Subsequently, data collection with the full mask system began (data presented in results section 2). Gr
immediately began data collection with the full mask system (6th session, data presented in results
section 2). The timelines for Nd and Gr are depicted in blue and orange, respectively.

Data Collection and Analysis: Comparison with Traditional Headpost

Voluntary Head Immobilization with Mask System vs Traditional Headpost

We first conducted a comparative analysis of visual fixation data (eye-tracking) using two
distinct restraint systems on two monkeys: one with a traditional headpost (Monkey Ph) and the
other using our non-invasive mask-based system, without the back piece (Monkey Nd).
Crucially, entry into the mask immobilization system was entirely voluntary for monkey Nd—at
no point during this session was the locking back piece of the mask engaged. This aspect of the
study design was implemented to assess the effectiveness and compliance of the mask-based
system under its least restrictive condition. These data were collected in Nd as soon as initial
training with the mask system was complete (see Figure 2 training plot).

The monkeys performed a simple 13-target fixation task. The animals were positioned 57 cm
from a calibrated 32" screen (120 Hz, 1920x1080 IPS LCD, Cambridge Research System Ltd).

In each trial, a fixation target (0.2 degrees in size) was presented for 500 ms seconds at one of
13 locations on the computer display (Figure 3a). Target locations were uniformly distributed
across the screen (Figure 3a). The most peripheral target location was approximately 14
degrees from the visual field center (10 degrees away both horizontally and vertically). The
monkeys were tasked with continuously fixating on each target during the presentation window.
Compliance was reinforced through liquid rewards via a sipper tube. Data were collected using
custom NIMH MonkeyLogic scripts on a Dell computer. Eye tracking was carried out with an
Eyelink 1000 Plus (SR Research), at a frequency 1000 Hz.

To assess the efficacy of our voluntary mask immobilization system compared to the traditional
headpost method, we compared eye tracking precision and accuracy of gaze fixations (Figure
3b). Both values were calculated on a per-trial basis: precision was defined as the mean
distance (in degrees of visual angle) between each gaze position sample and the mean gaze
position for that trial, and accuracy was defined as the mean distance (in degrees of visual
angle) between each gaze position sample and the fixation target for that trial. Eye tracking
samples were taken from the 500ms target fixation period in each trial, and trials in which
monkeys broke fixation were not taken into account. We then performed a Mann-Whitney U test
to compare the distributions of Nd and Ph's trial-by-trial gaze precisions, as well as their trial-by-
trial gaze accuracies. This non-parametric test was chosen because the data did not meet the
assumptions of normality (Shapiro-Wilk test), and the two datasets had unequal sample sizes.
The test evaluates whether the distributions of gaze metrics for the two conditions are
significantly different.

Next we assessed the stability of gaze accuracy over the course of the session while Nd was
using the mask in the voluntary configuration. This analysis was important because, in the
voluntary configuration, Nd could pull his face back at any time during the session to take a
break, and then place his face back in the mask to continue data collection without recalibration.
We wanted to know whether such disengagement and reengagement events impacted our gaze
accuracy measurements. We fit a linear model to Nd's gaze accuracy data to determine
whether there was any correlation between trial index and Nd's trial-by-trial gaze accuracy. A
positive correlation would indicate increasing instability over the course of a session, while
negative or no correlation would indicate stable fixation across the session.

Full Immobilization with Mask System vs Traditional Headpost

In order to test the relative efficacy of the full-immobilization mode of the mask system (back
piece engaged) versus a standard headpost in a typical experimental setup, we analyzed
fixation data acquired in two monkeys using the full-immobilization mask system (Nd & Gr) and
three monkeys with traditional headposts (Ph, Gr, & Kr) while they performed a cortical
perturbation detection (CPD) task (Azadi et al., 2023b), as part of ongoing research projects.
Note that Gr was tested with both immobilization systems (mask and headpost), enabling a
within-subject comparison. Gr, being a particularly large animal (14 kg), also provided a robust
test of the mask system's durability. At this stage of data collection, all four monkeys had been
trained to perform the CPD task, with two of the monkeys (Ph & Kr) tasked with detecting
optogenetic stimulation delivered to central IT, one monkey (Nd) tasked with detecting

optogenetic stimulation delivered to visual area V4, and the fourth monkey (Gr) tasked with
detecting electrical microstimulation delivered to central IT. Each trial in the CPD task was
initiated when the monkey fixated a central fixation target (0.4 degrees in diameter) for 500 ms.
Then, an image was displayed for 1 second, during which, in half of the trials randomly selected,
cortical stimulation was delivered for 200 ms halfway through the image presentation. After the
image disappeared, the monkey was rewarded for correctly identifying whether the trial
contained cortical stimulation by saccading to one of two targets at the top and bottom of the
screen. For the purposes of the present study, fixation data were extracted from a random
selection of these CPD task sessions, which were collected over many months. For each
monkey, five sessions were analyzed.

We compared the efficacy of our mask system in its full-immobilization configuration against the
headpost during long-term data collection while monkeys performed the CPD task, by analyzing
the trial-by-trial gaze precision and accuracy of fixations (Figure 4). We considered gaze
position samples collected during the image presentation phase of each trial, throughout which
central fixation was required. Trials in which fixation was broken at any point after trial initiation
were not taken into account. Precision and accuracy were calculated using the same method as
above.,. For each gaze fixation metric (precision and accuracy), we performed a linear mixed-
effects model analysis using MATLAB's fitlme function. The model included a fixed effect for
condition (mask or headpost) and a random intercept for subject (monkey) to account for
between-subject variability. The headpost condition was set as the reference category. The
gaze fixation metric (either precision or accuracy) for each trial was used as the dependent
variable, and we tested for significance using t-statistics and p-values for the fixed effects.
Model coefficients were estimated using restricted maximum likelihood (REML). The same
analysis was performed for gaze precision and gaze accuracy. This approach allowed us to
examine the difference in gaze fixation metrics between the mask and headpost conditions,
while controlling for individual differences in baseline gaze fixation.

Because we collected data in monkey Gr using both the mask and headpost head-
immobilization methods, we were able to carry out a within-subject analysis. We used a Mann-
Whitney U test to compare the distribution of trial-by-trial gaze precisions, as well as the trial-by-
trial gaze accuracies, between the mask and headpost conditions. This non-parametric test was
chosen because the data did not meet the assumptions of normality (Shapiro-Wilk test), and the
two conditions had unequal sample sizes. The test evaluates whether the distributions of gaze
metrics for the two conditions are significantly different.

Results

Voluntary Head Immobilization with Mask System vs Traditional Headpost

In the initial comparative analysis between a traditional headpost and our mask system without
the back piece engaged, Nd (voluntarily mask-immobilized) completed 365 trials over 1 session
and Ph (headposted) completed 673 trials over 1 session. Figure 3 shows the trial-by-trial
distributions of the two monkeys' gaze precision and accuracy. The difference in mean gaze
precision between Nd and Ph was 0.12° of visual angle, and the difference in mean gaze
accuracy was 0.060° (Nd measurement - Ph measurement). These results indicate that the
precision and accuracy of gaze fixations were slightly better in the headpost condition. While
these differences were small in magnitude, they were significant. The Mann-Whitney U test
indicated a significant difference in gaze precision between the mask (median = 0.2010°, IQR =
0.1402°) and headpost (median = 0.1029°, IQR = 0.0403°) conditions, U = 32,229, p < 0.001.
Similarly, the test revealed a significant difference in gaze accuracy between the mask (median
= 0.43° degrees, IQR = 0.2765°) and headpost (median = 0.3501°, IQR = 0.2000°) conditions, U
= 93,744, p < 0.001. Overall, the results show a statistically significant but slight decrease in eye
tracking performance for Nd (mask-stabilized) compared to Ph (headpost-stabilized).

In the voluntary configuration, Nd could pull his face back from the mask at any time during the
session, and then return his face to the mask to continue data collection. We did not recalibrate
the eye-tracker between these events. In order to assess the impact of such disengagement
and re-engagement events, we analyzed Nd's gaze accuracy over time and found it to be
consistent throughout the session (Figure 3c): Linear regression showed no significant
correlation between trial index and Nd's trial-by-trial gaze accuracy (slope = 0.00019, R^2 =
0.0100, p = 0.55). These results show that disengagement and re-engagement events did not
impact our gaze accuracy measurements.

Figure 3–Accuracy and precision of visual fixation eye-tracking data collected using voluntary head-immobilization configuration of the mask NHIS vs traditional headpost: 2 monkeys (Nd, mask-restrained, blue; and Ph, headposted, red) fixated on 13 targets throughout the testing procedure, receiving a liquid reward, recorded using an eye tracker. a) Fixation targets and mean fixation positions for monkeys Nd and Ph. X-axis and y-axis show horizontal and vertical degrees of visual angle on the computer screen, and error bars show ±1 SD. b) Trial-by-trial gaze precision and accuracy for monkeys Nd (n = 365 trials) and Ph (n = 763) during the 13-target fixation task. Y-axes show mean gaze precision and accuracy in degrees of visual angle for the left and right plots, respectively. Means are indicated with a black marker. Black outlines show boxplot statistics: boxes represent interquartile range (IQR) with median, and whiskers represent 1.5*IQR from the quartiles. The Mann-Whitney U test indicated a significant difference in gaze precision between the mask (median = 0.2010°, IQR = 0.1402°) and headpost (median = 0.1029°, IQR = 0.0403°) conditions, U = 32,229, p < 0.001. Similarly, the test revealed a significant difference in gaze accuracy between the mask (median = 0.43°, IQR = 0.2765°) and headpost (median = 0.3501°, IQR = 0.2000°) conditions, U = 93,744, p < 0.001. Precision and accuracy were significantly better in the headpost condition compared to the mask condition. c) Gaze

accuracy for monkey Nd across trials in 13-target fixation task. Y-axis shows gaze accuracy in degrees of
visual angle, and x-axis shows trial number. Linear regression showed no significant correlation between
trial index and gaze accuracy for Nd (slope = 0.00019, R^2 = 0.0100, p = 0.55).

Full Immobilization with Mask System vs Traditional Headpost

In the subsequent analysis comparing standard headposts and the full-immobilization mode of
the mask system (front and back piece engaged) during long-term data collection in a typical
experimental set-up (Figure 4), across the 5 sessions per monkey that we analyzed, Nd (fully
mask-immobilized) completed 8064 trials, Ph (headposted) completed 8100 trials, and Kr
(headposted) completed 6282 trials. Gr, the within-subject comparison monkey, completed 7244
trials across 5 sessions while fully mask-immobilized, and completed 5113 trials across another
5 sessions while headposted. On average, monkeys who were immobilized using our mask
system completed more trials than monkeys who were headposted. Additionally, long-term data
collection using the mask was successful; over 8 months, Nd completed approximately 100,000
trials, and Gr completed 30,000 trials over the course of 3 months while using the mask.

Linear mixed-effects models revealed small but significant effects of head-stabilization method
on gaze precision and accuracy. The mean gaze precision in the headpost condition was
estimated to be 0.1096° (SE = 0.0184°), t(34,801) = 5.94, p < 0.001. Gaze precision in the mask
condition was significantly worse compared to the headpost condition, with a difference of
0.0337° (SE = 0.0010°), t(34,801) = 32.52, p < 0.001. The 95% confidence interval for the effect
of the mask condition was [0.0317°, 0.0358°]. The average gaze accuracy in the headpost
condition was estimated at 0.1832° (SE = 0.0379°), t(34,801) = 4.83, p < 0.001. Gaze accuracy
in the mask condition was significantly worse than in the headpost condition, with an estimated
difference of 0.0914° (SE = 0.0024°), t(34,801) = 37.73, p < 0.001. The 95% confidence interval
for the effect of the Mask condition was [0.0866°, 0.0961°].
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For our within-subject comparison in Gr, there was a difference of 0.033° in mean gaze
precision and a 0.089° difference in mean gaze accuracy when using the mask compared to a
traditional headpost (mask measurement - headpost measurement). While these differences
were small in magnitude, they were significant. The Mann-Whitney U test indicated a significant
difference in gaze precision between the mask (median = 0.143°, IQR = 0.070°) and headpost
(median = 0.117°, IQR = 0.051°) conditions, U = 12,331,258, p < 0.001. Similarly, the test
revealed a significant difference in gaze accuracy between the mask (median = 0.237°, IQR =
0.133°) and headpost (median = 0.161°, IQR = 0.072°) conditions, U = 8,252,961, p < 0.001.
Precision and accuracy were significantly better in the headpost condition compared to the
mask condition.

Figure 4 - Accuracy and precision of visual fixation eye-tracking data using full head-immobilization configuration of the mask NHIS vs traditional headpost. Individual monkeys' trial-by-trial gaze precision (left) and accuracy (right) distributions during central fixation in a cortical perturbation detection (CPD) task. Data collected in monkeys head-immobilized with the mask NHIS (Nd, Gr) are shown in blue, and those head-immobilized with a headpost (Gr, Ph, Kr) are shown in red. Y-axis shows gaze precision (left) and accuracy (right) in degrees of visual angle. Means are indicated with a white marker. Black outlines show boxplot statistics: boxes represent interquartile range (IQR) with median, and whiskers represent 1.5*IQR from the quartiles. Data shown for all plots is from n= 34803 trials (Nd: 8064; Gr with mask: 7244; Gr with headpost: 5113; Ph: 8100; Kr: 6282). A linear mixed-effects model revealed a significant effect of head stabilization method on gaze precision and accuracy. The mean gaze precision in the headpost condition (the reference condition) was estimated to be 0.1096° (SE = 0.0184°), t(34,801) = 5.94, p < 0.001. Gaze precision in the mask condition was significantly higher (worse) compared to the Headpost condition, with a difference of 0.0337° (SE = 0.0010°), t(34,801) = 32.52, p < 0.001. The mean gaze accuracy in the headpost condition was estimated at 0.1832° (SE = 0.0379°), t(34,801) = 4.83, p < 0.001. Gaze accuracy in the mask condition was significantly higher than in the headpost condition, with an estimated difference of 0.0914° (SE = 0.0024°), t(34,801) = 37.73, p < 0.001. A Mann-Whitney U test was used to carry out within-subject pairwise comparisons between conditions for Gr, for both gaze precision and accuracy. The results indicated a significant difference in gaze precision between the mask (median = 0.143°, IQR = 0.070°) and headpost (median = 0.117°, IQR = 0.051°) conditions, U = 12,331,258, p < 0.001. The test revealed a significant difference in gaze accuracy between the mask (median = 0.237°, IQR = 0.133°) and headpost (median = 0.161°, IQR = 0.072°) conditions, U = 8,252,961, p < 0.001. Precision and accuracy were significantly better in the headpost condition compared to the mask condition.

Discussion

In our study, monkeys with their heads immobilized using a 3D-printed mask demonstrated eye-
tracking precision and accuracy similar to that of their headposted counterparts. While
headposted monkeys demonstrated significantly better gaze precision and accuracy during
fixation, the difference observed when using the NHIS in its least restrictive configuration was
just 0.12° for precision and 0.06° for accuracy (pairwise comparison), and in its most restrictive
configuration was 0.03° for precision and 0.18° for accuracy (mixed-effects model estimate). In
our within-subject comparison, the differences were similar: 0.03° for precision and 0.09° for
accuracy. Though statistically significant, this minimal loss in gaze precision and accuracy using
our NHIS relative to the traditional headpost is well-within the acceptable range for most use
cases. Typically, experiments carried out with head-fixed awake behaving monkeys require
monkeys to maintain fixation within a not less than 0.8° x 0.8° window, including studies of V1
neurons whose receptive fields tend to be small (Abhishek & Horwitz, 2022). Thus, the quality of
eye-tracking with NHIS head fixation is comfortably within the range of suitability for most
experimental needs.

Moreover, in this study, we used Eyelink 1000 (SR Research Ltd.), which has a precision of
$0.01°$ and accuracy down to $0.15°$ for tracking and recording gaze position under ideal
conditions. Most other video-based eye trackers commonly used in nonhuman primate
physiology experiments do not offer such high precision and accuracy. The performance of the
mask introduced in this study is comparable to, or even exceeds, some other popular systems
used in similar research. For example, the ViewPoint EyeTracker (Arrington Research, Inc.) has
a precision of $0.15°$ and an accuracy ranging from $0.25°$ to $1.0°$.

Furthermore, monkeys stabilized with the mask were able to perform as many if not more trials
than those stabilized using a headpost. As well, the monkeys were able to complete months of
data collection using the mask, demonstrating long-term suitability of the mask and the continual
cooperation of monkeys to engage with the mask. As to whether our NHIS can remain stable
with larger and stronger monkeys, we successfully used our NHIS with a relatively large adult
male who was 14kg. There were no complications from his use of the NHIS for 30,000 trials
over 3 months, thus demonstrating the mechanical robustness of our system. The comparable
performance observed between headposted monkeys and ones wearing our mask comes with a
range of benefits. First, reducing the need for surgery and using a non-invasive approach for
head immobilization aligns with the continual mission to refine methods and minimize the
potential for pain and stress in research animals, as outlined in the principles of the 3Rs
(Russell, Burch, and Hume, 1959). While the tools commonly used to measure and perturb
neural activity and behavior still require surgical interventions, the field is moving towards the
use of wireless devices (Dal Monte, et al., 2022; Testard, et al., 2024) which will reduce the
persistent risk of infection posed by implants that breach the skin barrier. Replacing headposts
with effective non-invasive head-immobilization systems goes hand-in-hand with these efforts.
Moreover, reducing the need for headpost placement surgery and recovery allows for a faster
transition to data collection. The mask system can be quickly produced in-house using 3D
printing technology, and the monkeys in our study learned to use the mask within 10 days.

Furthermore, the use of 3D printing to produce an NHIS, as presented here, offers several
advantages over the commonly used thermoplastic materials in most NHIS designs (Machado

and Nelson, 2011; Drucker et al., 2014; De Luna et al., 2014; Slater et al., 2016; Nakamura et
al., 2020). First, 3D printing allows for precise anatomical fit, better accommodating unique
individual differences than thermoplastics can. Thermoplastics are shaped by heating and
molding, limiting the precision of fine anatomical details, especially for intricate surfaces like the
face or head. Second, designing the mask in a 3D modeling software environment allows for
complex geometries that might improve comfort, stability, and fit beyond the capabilities of
molded thermoplastic. Third, such an approach enables design modifications to suit specific
experimental needs and equipment configurations (e.g., cranial implants, style of primate chair,
reward delivery system), enhancing overall utility, integration, and production. For example, the
design presented here affords additional space on the top of the head for the placement of
research-related implants, in contrast to full-head thermoplastic helmets. Fourth, 3D-printed
materials can be designed to withstand repeated use and cleaning, potentially outlasting
thermoplastic molds, which may degrade or lose shape over time. Finally, the mask can be
readily modified and re-printed without sedation or excessive downtime to accommodate
changes in head size due to animal growth, experimental needs, or to replace damaged parts.

Our assessment of the mask system, as presented in this paper, has some potential limitations.
The sample was restricted to two non-naive monkeys, both of which had already undergone
fixation training. We anticipate naive monkeys will take longer to acclimate to the mask, possibly
extending the training process beyond the timeline we have outlined here (Fig 2). Also, the
efficacy of our system in providing stable intracranial recordings is yet to be shown. We hope
that sharing this method with the greater scientific community brings interest to shed more light
on these unaddressed aspects. In addition, the mask creation pipeline requires sedation,
access to an anatomical scanner (CT or MRI), and an overnight or longer 3D printing process,
as well as post-processing, before use. Future design iterations may minimize the post-
processing required, but the speed of available 3D printing technology and requisite sedated
scanning present a ceiling on the extent to which the mask system can be streamlined. Finally,
the scope of the present work does not evaluate the mask's suitability for acute
electrophysiology or functional imaging, as has been done for other NHIS systems (Srihasam,
et al., 2010; Amemori, et al., 2015; Drucker, et al., 2015; Slater, et al., 2016). We expect that
our system will perform at least as well as those systems, but further testing is needed.

Going forward, we plan to continue to monitor the efficacy of the mask system for data
collection, as well as incorporate and test additional options and functionality. For example, our
in-house CT scanner allows us to access scans for many monkeys, so we are currently
developing and testing an option to have a mask design based on average macaque 3D base
models (averaged within sex). This standardized ready-made mask would be suited for use in
experiments where stringent immobilization is not required. The average face mask could be
used and scaled to accommodate growth and slight changes in monkeys' face structure without
requiring sedation or scanning. We are also testing a double-layer design to allow for a softer,
padded material to form the interior surface of the mask to increase monkeys' comfort. Finally,
there is potential for our NHIS mask, without the back piece, to be placed in the front of the
monkey's home cage, with cage modifications or other approaches. The opportunity to collect
precise eye-tracking data in an unrestrained monkey trained to place its head in the mask,
unobstructed by cage bars from the eye-tracker or stimuli, would be an exciting potential to

further refine our experimental approaches for neuroscience research in awake and behaving
monkeys.

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Availability statement: The production pipeline code will be made publically available to
encourage widespread adoption.

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References:
Adams, D. L., Economides, J. R., Jocson, C. M., & Horton, J. C. (2007). A biocompatible
titanium headpost for stabilizing behaving monkeys. Journal of neurophysiology, 98(2),
993-1001.

Abhishek, D., Horwitz, G.D. (2022). Coding of chromatic spatial contrast by macaque V1
neurons eLife 11:e68133.

Amemori, Satoko, Ken-Ichi Amemori, Margaret L. Cantor, and Ann M. Graybiel. 2015. "A Non-
Invasive Head-Holding Device for Chronic Neural Recordings in Awake Behaving
Monkeys." *Journal of Neuroscience Methods* 240 (January):154–60.

Azadi, R., Bohn, S., Eldridge, M. A., & Afraz, A. (2023a). Surgical Procedure for Implantation of
Opto☐Array in Nonhuman Primates. *Current protocols*, *3*(3), e704.

Azadi, R., Bohn, S., Lopez, E., Lafer-Sousa, R., Wang, K., Eldridge, M. A., & Afraz, A. (2023b).
Image-dependence of the detectability of optogenetic stimulation in macaque
inferotemporal cortex. Current biology, 33(3), 581-588.

Azimi, K., Prescott, I. A., Marino, R. A., Winterborn, A., & Levy, R. (2016). Low profile halo head
fixation in non-human primates. Journal of neuroscience methods, 268, 23-30.

Betelak, K. F., E. A. Margiotti, M. E. Wohlford, and D. A. Suzuki. 2001. "The Use of Titanium
Implants and Prosthodontic Techniques in the Preparation of Non-Human Primates for
Long-Term Neuronal Recording Studies." *Journal of Neuroscience Methods* 112 (1): 9–20.

Carlota, V. 2023. "All You Need to Know About ASA for 3D Printing." 3Dnatives. May 15, 2023.
https://www.3dnatives.com/en/asa-3d-printing-170820204/.

Dal Monte, O., Fan, S., Fagan, N. A., Chu, C. C. J., Zhou, M. B., Putnam, P. T., ... & Chang, S.
W. (2022). "Widespread implementations of interactive social gaze neurons in the primate
prefrontal-amygdala networks." Neuron, 110(13), 2183-2197.

De Luna, P., Mustafar, M. F. B. M., & Rainer, G. (2014). A MATLAB-based eye tracking control
system using non-invasive helmet head restraint in the macaque. Journal of neuroscience
methods, 235, 41-50.

Drucker, Caroline B., Monica L. Carlson, Koji Toda, Nicholas K. DeWind, and Michael L. Platt.
2015. "Non-Invasive Primate Head Restraint Using Thermoplastic Masks." *Journal of*
Neuroscience Methods 253 (September):90–100.

Evarts, E. V. 1968. "Relation of Pyramidal Tract Activity to Force Exerted during Voluntary
Movement." *Journal of Neurophysiology*, January. https://doi.org/10.1152/jn.1968.31.1.14.

Foeller, Paul, and Lawrence Tychsen. 2002. "Eye Movement Training and Recording in Alert
Macaque Monkeys: 1. Operant Visual Conditioning; 2. Magnetic Search Coil and Head
Restraint Surgical Implantation; 3. Calibration and Recording." *Strabismus* 10 (1): 5–22.

Goense, J. B., Whittingstall, K., & Logothetis, N. K. (2010). Functional magnetic resonance
imaging of awake behaving macaques. Methods, 50(3), 178-188.

Hadj-Bouziane, F., Monfardini, E., Guedj, C., Gardechaux, G., Hynaux, C., Farnè, A., &
Meunier, M. (2014). "The helmet head restraint system: a viable solution for resting state
fMRI in awake monkeys." Neuroimage, 86, 536-543.

Hallum, Luke E., Christopher Shooner, Romesh D. Kumbhani, Jenna G. Kelly, Virginia García-
Marín, Najib J. Majaj, J. Anthony Movshon, and Lynne Kiorpes. 2017. "Altered Balance of
Receptive Field Excitation and Suppression in Visual Cortex of Amblyopic Macaque
Monkeys." *The Journal of Neuroscience: The Official Journal of the Society for*
Neuroscience 37 (34): 8216–26.

Isoda, Masaki, Ken-Ichiro Tsutsui, Narumi Katsuyama, Tomoka Naganuma, Naohiro Saito,
Yoshihito Furusawa, Hajime Mushiake, Masato Taira, and Jun Tanji. 2005. "Design of a
Head Fixation Device for Experiments in Behaving Monkeys." *Journal of Neuroscience*
Methods 141 (2): 277–82.

Joesch, Maximilian, David Mankus, Masahito Yamagata, Ali Shahbazi, Richard Schalek, Adi
Suissa-Peleg, Markus Meister, Jeff W. Lichtman, Walter J. Scheirer, and Joshua R. Sanes.
2016. "Reconstruction of Genetically Identified Neurons Imaged by Serial-Section Electron
Microscopy." *eLife* 5 (July). https://doi.org/10.7554/eLife.15015.

Kar, K., Kubilius, J., Schmidt, K. et al. "Evidence that recurrent circuits are critical to the ventral
stream's execution of core object recognition behavior." Nat Neurosci 22, 974–983 (2019).
https://doi.org/10.1038/s41593-019-0392-5

Lafer-Sousa, R., Conway, B. R., & Kanwisher, N. G. (2016). Color-biased regions of the ventral
visual pathway lie between face-and place-selective regions in humans, as in macaques.
Journal of Neuroscience, *36*(5), 1682-1697.

Lafer-Sousa, R., & Conway, B. R. (2013). Parallel, multi-stage processing of colors, faces and
shapes in macaque inferior temporal cortex. Nature neuroscience, 16(12), 1870-1878.

Lemon, Roger. 1984. *Methods for Neuronal Recording in Conscious Animals*. Vol. 4. IBRO
Handbook Series, Vol 4. Chichester, England: John Wiley & Sons.

Lu, X., Wang, Q., Li, X., Wang, G., Chen, Y., Li, X., & Li, H. (2023). Connectivity reveals
homology between the visual systems of the human and macaque brains. *Frontiers in*
Neuroscience, *17*, 1207340.

Machado, C. J., & Nelson, E. E. (2011). Eye☐tracking with nonhuman primates is now more

accessible than ever before. American journal of primatology, 73(6), 562-569.

Mountcastle, V. B., J. C. Lynch, A. Georgopoulos, H. Sakata, and C. Acuna. 1975. "Posterior
Parietal Association Cortex of the Monkey: Command Functions for Operations within
Extrapersonal Space." *Journal of Neurophysiology* 38 (4): 871–908.

Nakamura, T., Dinh, T. H., Asai, M., Nishimaru, H., Matsumoto, J., Takamura, Y., ... & Nishijo,
H. (2020). Non-invasive electroencephalographical (EEG) recording system in awake
monkeys. Heliyon, 6(5).

Orban, G. A., Van Essen, D., and Vanduffel, W. (2004). Comparative mapping of higher visual
areas in monkeys and humans. Trends Cogn. Sci. 8, 315–324. doi:
10.1016/j.tics.2004.05.009

Pigarev, I. N., Y. B. Saalmann, and T. R. Vidyasagar. 2009. "A Minimally Invasive and
Reversible System for Chronic Recordings from Multiple Brain Sites in Macaque Monkeys."
Journal of Neuroscience Methods 181 (2): 151–58.

Rima, S., Greilsamer, J., Haag, M., Cadena-Valencia, J., Sansonnens, M., Francovich, A., ... &
Schmid, M. C. (2024). "A chinrest-based approach to measure eye movements and
experimental task engagement in macaques with minimal restraint." Journal of
neuroscience methods, 408, 110173.

Roussy, Megan, Benjamin Corrigan, Rogelio Luna, Roberto A. Gulli, Adam J. Sachs, Lena
Palaniyappan, and Julio C. Martinez-Trujillo. 2022. "Stable Working Memory and
Perceptual Representations in Macaque Lateral Prefrontal Cortex during Naturalistic
Vision." *The Journal of Neuroscience: The Official Journal of the Society for Neuroscience*
42 (44): 8328–42.

Russell, W. M. S., Burch, R. L., & Hume, C. W. (1959). The principles of humane experimental
technique (Vol. 238). London: Methuen.

Shahbazi, Ali. 2018. *Computer Vision-Based Approaches to Neural Circuit Tracing at Scale*.
University of Notre Dame.

Shahbazi, Ali, Jeffery Kinnison, Rafael Vescovi, Ming Du, Robert Hill, Maximilian Joesch, Marc
Takeno, et al. 2018. "Flexible Learning-Free Segmentation and Reconstruction of Neural
Volumes." *Scientific Reports* 8 (1): 14247.

Shahbazi, E., Ma, T., Pernuš, M., Scheirer, W., & Afraz, A. (2024). Perceptography unveils the
causal contribution of inferior temporal cortex to visual perception. Nature Communications,
15(1), 3347.

Slater, Heather, Alice E. Milne, Benjamin Wilson, Ross S. Muers, Fabien Balezeau, David
Hunter, Alexander Thiele, Timothy D. Griffiths, and Christopher I. Petkov. 2016.
"Individually Customisable Non-Invasive Head Immobilisation System for Non-Human
Primates with an Option for Voluntary Engagement." *Journal of Neuroscience Methods* 269
(August):46–60.

Srihasam, Krishna, Kevin Sullivan, Tristram Savage, and Margaret S. Livingstone. 2010.
"Noninvasive Functional MRI in Alert Monkeys." *NeuroImage* 51 (1): 267–73.

Tanigawa, H., Lu, H. & Roe, A. Functional organization for color and orientation in macaque V4.

Nat Neurosci 13, 1542–1548 (2010). https://doi.org/10.1038/nn.2676

Testard, Camille, Sébastien Tremblay, Felipe Parodi, Ron W. DiTullio, Arianna Acevedo-Ithier,
Kristin L. Gardiner, Konrad Kording, and Michael L. Platt. "Neural signatures of natural
behaviour in socializing macaques." Nature 628, no. 8007 (2024): 381-390.

Vaidya, A. R., Pujara, M. S., Petrides, M., Murray, E. A., & Fellows, L. K. (2019). "Lesion studies
in contemporary neuroscience." Trends in cognitive sciences, 23(8), 653-671.

Van Essen DC, Lewis JW, Drury HA, Hadjikhani N, Tootell RB, Bakircioglu M, Miller MI (2001)
Mapping visual cortex in monkeys and humans using surface-based atlases. Vision Res 41:
1359–1378

Wang, S., Falcone, R., Richmond, B., & Averbeck, B. B. (2023). Attractor dynamics reflect
decision confidence in macaque prefrontal cortex. Nature Neuroscience, 26(11), 1970-
1980.

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