Assessing the role of pleiotropy in the evolution of animal color and behavior: a meta-analysis of

experimental studies

Sarah N. Ruckman[1], Eve A. Humphrey[2], Lily Muzzey[1], Ioanna Prantalou[1], Madison Pleasants[1], and

Kimberly A. Hughes[1]

[1]Department of Biological Science, Florida State University, Tallahassee, FL, USA

[2]Biology Department, Lincoln University, Lincoln University, PA, USA

Keywords: aggressive behavior, badge of status, coloration, condition dependence, pleiotropy

Abstract

Color varies in pattern and degree across the tree of life. In animals, genetic variation in color is

hypothesized to have pleiotropic effects on a variety of behaviors, due to shared dependence on

underlying biochemical pathways. Such pleiotropy can constrain the independent evolution of color and

behavior. Although associations between color and behavior have been reported, this relationship has

not yet been addressed across a broad taxonomic scale with a formal meta-analysis. We used a

phylogenetic meta-analytic approach to examine the relationship between individual variation in

aggressive behavior and variation in multiple colors. Seventy studies met our inclusion criteria

(vertebrates = 66; invertebrates = 4). After accounting for phylogeny and correcting for publication bias,

there was a positive association between measures of aggression and degree or area of coloration

(mean = 0.274, 95% CI = (0.041, 0.481)). However, this positive association was not influenced by type of

color or by several other variables that we tested. Because the data supports a positive association

between aggression and degree or area of coloration, irrespective of whether color is melanin-based,

carotenoid-based, or structural, we conclude that this pattern does not strongly support the melanin-

pleiotropy hypothesis. The relationship was also unaffected by moderators accounting for individual

condition, social rank, or age; thus, the results do not strongly support hypotheses that condition-

dependence accounts for relationships between color and aggressive behavior. We propose that the

moderate positive correlation between aggression and coloration across *Animalia* that we observed is

underlain by genetic covariation between behavior and color traits that serve as badges of status.

Introduction

A key goal of modern evolutionary biology is predicting if and how populations will evolve adaptively in response to environmental change. Predicting adaptive evolution requires knowing if there are constraints on how populations can respond to selection. The definition of adaptive constraint has been debated, but the unifying theme of these definitions is that populations are not always able to respond to selection in predicable ways (Blows and Walsh 2009; Walsh and Blows 2009). Pleiotropy, and the genetic covariance between traits that can result, is one potential cause of adaptive constraint. However, estimating genetic covariance in natural populations requires very large sample sizes and reliable information on relatedness which are currently available in few species. Trait covariance can also be environment-dependent, meaning many such estimates might be necessary to accurately predict responses to selection in variable environments.

Another approach is to use functional genetic information to predict pleiotropic effects and their population-level consequences. A well-known example is the hypothesis that, in animals, the biochemical pathways that regulate body coloration and behavior can overlap, leading to genetic covariance between color and behavior. Specifically, in vertebrates, melanic coloration arises from the binding of melanocortin agonists to the G-protein-coupled receptor MCR-1. Melanocortins (peptide hormones derived from the prohormone Proopiomelanocortin) also bind to other receptors that regulate diverse functions, including aggressive behavior (Ducrest et al. 2008). Therefore, one prediction of this hypothesis is that heritable variation in abundance or activity of melanocortins generates genetic correlation between body color and aggression; animals with higher levels of eumelanic (dark brown, black) color are expected to exhibit higher levels of aggression (Ducrest et al. 2008; Roulin and Ducrest 2011). We note that other kinds of changes in the melanocortin system, such as mutation in melanocortin receptors that are expressed both in the skin and the brain, could also produce these correlations (e.g., Reissmann and Ludwig 2013). In support of this hypothesis, studies have reported

correlations between melanin-based coloration and behavior, including aggressive behavior, in

vertebrates (e.g., Moore and Martin 2016; Dijkstra et al. 2017; Seddon and Hews 2017; Beck et al. 2018).

This hypothesis has recently been extended to include insects and other invertebrates (San-Jose and

Roulin 2018), where melanin is synthesized from dopamine. This invertebrate biosynthetic pathway

provides a plausible link between body coloration and behavior (Wittkopp and Beldade 2009; Massey

and Wittkopp 2016), and predicts correlations between traits that are similar to those seen in

vertebrates have been reported (San-Jose and Roulin 2018).

Other functional links between color and behavior have been hypothesized to arise from

carotenoid condition dependence, whereby animals in good condition can devote more energy to high

levels of aggression, and to carotenoid pigment synthesis, which is dependent on obtaining carotenoids

in the diet (Blount and McGraw 2008; Backström et al. 2015). Under this scenario, genetic variants

affecting foraging ability would simultaneously affect carotenoid color and aggressiveness, leading to

genetic correlations between these traits. However, carotenoid condition-dependence could also arise

from purely environmental covariance between aggression and color if, for example, individual variation

in foraging success arises from non-genetic variation (e.g., variation in habitat quality). Purely

environmental covariance between color and aggression would not impose the same kind of adaptive

constraint as would a genetic correlation.

A third hypothesis is that a broad range of colors can be indicators or "badges" of social status.

Badges of status are traits (e.g., color patches) that influence the outcome of aggressive encounters

(Rohwer 1975; Dawkins and Krebs 1978). Badges of status are not limited to specific colors; badges can

even lack color, as seen in the white forehead patch of collared flycatchers *Ficedula albicollis* (Pärt and

Qvarnström 1997). A meta-analysis of associations between dominance and plumage characteristics

(color, UV presence, and color patch size) in birds reported a positive correlation between dominance

and measures of coloration, irrespective of specific color; the authors interpreted this result as

supporting the badge of status hypothesis (Santos et al. 2011; but see Sánchez-Tójar et al. 2018). A

comparative analysis of competing bird species, found that dominant species have on average more

black than subordinate species; carotenoid and other colors were sometimes, but not always associated

with dominance (Kenyon and Martin 2023). The authors interpreted this result as also supporting the

badge of status hypothesis (Kenyon and Martin 2023). Such badges can be honest signals (reliable

predictors of aggressiveness), although the mechanism maintaining honesty in the signal has been

debated and could be different for each color or type of badge (Johnstone and Norris 1993; Tibbetts and

Dale 2004). In ruffs, *Philomachus pugnax,* a supergene relates color and aggression, where the genes

associated with color and behavior are located near each other and in linkage disequilibrium (Küpper et

al. 2016; Lamichhaney et al. 2016). In contrast, some badges of status might not vary genetically, but

might instead vary due to non-genetic sources. For example, some badges are plastic in expression and

can vary as a result of dominance interactions (e.g., Dey et al. 2014). As above, implications for adaptive

constraint depend upon the underlying cause of trait covariance.

Other hypotheses can be consistent with a positive association between aggression and color,

irrespective of the type of color. For example, although melanin is endogenously produced, production

and/or maintenance of melanic body color might be condition-dependent, although the empirical

evidence for this is mixed (Roulin 2016). Similar arguments can be made for structural colors. If

structural color depends on the condition of feathers or scales, then costs could be incurred in

maintaining or cleaning those structures. Structural color variation has been associated with condition

(e.g., McGraw et al. 2002) and mate quality (e.g., Siefferman and Hill 2003) in birds. If aggressive

behavior is also condition-dependent, then a general condition dependence might be responsible for

positive associations between color and aggressive behavior, irrespective of type of color.

If color and behavior are genetically linked via pleiotropy under any of the above scenarios,

independent expression and evolution of these traits can be constrained. This constraint has mainly

been discussed in relation to the melanocortin hypothesis, perhaps because melanocortin synthesis is

less likely to be condition dependent than other pigment-based colors (Ducrest et al. 2008; Roulin and

Ducrest 2011; San-Jose and Roulin 2018). However, counterexamples to the expected association

between darker color and higher aggression have been reported. Boerner and Krüger (2009) found that,

in the common buzzard (*Buteo buteo*), light colored males are more aggressive than darker colored

birds. In pied flycatchers (*Ficedula hypoleuca*), no significant relationship between male color and

aggressive behavior was found (Huhta and Alatalo 1993). Examples such as these suggest that species

(or populations within species) vary in the relationship between color and aggression. Counterexamples

also raise the possibility that the preponderance of studies reporting significant correlations (Ducrest et

al. 2008; Roulin and Ducrest 2011; San-Jose and Roulin 2018) reflects taxonomic or publication bias, or

that correlations arise from specific features of studies like the age, sex, or condition of the focal

animals. For example, a meta-analysis reported positive correlation between dominance and plumage

traits in birds; this correlation was unaffected by the type of plumage trait, but was influenced by the

assessment method (whether aggression was assessed by quantifying specific aggressive acts, or by an

indirect method such as distance between individuals; Santos et al. 2011). We know of no meta-analyses

of associations between coloration and behavior that extend across broader taxa, or that assess other

factors such as whether color is fixed or plastic during adulthood.

Here, we describe a meta-analysis of the (within-species) relationship between body color

variation and aggressive behavior across a broad taxonomic scale. The meta-analysis included

vertebrate and invertebrate taxa and controlled for effects of phylogeny on statistical inference. We

investigated the possibility that publication bias has influenced the patterns reflected in the published

literature. In addition, we evaluated whether the relationship between color and aggression is

moderated by the type of color class (e.g. eumelanic, carotenoid, or structural), whether coloration is

fixed or varies plastically during adulthood, life stage and sex of the focal animals, type of population

studied (wild, domestic, lab-reared, or wild caught and then lab tested), type of aggressive act measured

(direct or indirectly measured), geographic origin of the species or source population of the focal

animals, and whether or not social rank, age, and the condition of the animal were controlled or

measured. We were especially interested in whether associations were moderated by color type

because the melanocortin pleiotropy hypothesis predicts specifically that variation in eumelanin-based

colors should be associated with aggression. By contrast, condition-dependence has most often been

discussed in relation to carotenoid pigmentation (and predicts a positive association between

carotenoid pigmentation and aggression). Consequently, positive associations between color and

aggression that are restricted to melanic or carotenoid colors would provide direct support for those

hypotheses. Both the badge-of-status hypothesis and general condition-dependence predict that

associations should be found between aggression and color, but that these associations should not be

limited to melanin- or carotenoid-based colors. The general condition dependence mechanism depends

upon both color and aggression being condition dependent, leading us to predict that controlling for

condition should moderate the relationship between color and behavior. By contrast, the badge-of-

status hypothesis does not, *a priori*, make that same prediction.

Methods

Data Collection: We defined aggression as any variable that measured antagonistic behaviors (e.g.,

biting, or chasing) toward a conspecific (of same sex and age) or mirror image. We narrowed our search

to conspecifics because sexes may differ in both behavior and coloration as well as how they respond to

differing individuals (e.g., Horth 2003). The colors included those regulated by melanin-, carotenoid-,

and pterin-based pigments. We also included colors produced by structural variation in skin, scales,

feathers, and cuticles (e.g., blue, purple, and white colors produced by iridophores in some fish; Schartl

et al. 2016). Each color classification per species was listed and justified with citations in Supplemental

Table S1.

We searched Web of Science using the search terms "color" AND "aggression" OR "aggressive"

(Figure 1). This yielded 751 papers as of September 2020. We then filtered the list by removing papers

with human subjects (n = 197). We removed human subjects because the variation in human

pigmentation is not due to the same mechanisms that have been proposed to lead to pleiotropic effects

on behavior (Deng and Xu 2017; San-Jose and Roulin 2018). We also removed any papers that did not

measure both individual color and aggression (n = 324), which is necessary to determine an association.

This yielded 230 papers. Finally, we removed papers if they did not include any of the following: natural

color variation within the same species, individual measures of both color and aggression against a con-

specific (same age, sex, and color), report relevant statistics for effect size calculations, and analyze the

data in comparison between color morphs. This yielded 70 papers (159 effect size estimates) that were

included in the phylogenetic meta-analysis.

From each paper, we collected the species name, color (how color was categorized or quantified

in the original study, (e.g., white vs. tan, area of eumelanic pigmentation, etc.), location of the color

(e.g., total body color or eye color), type of color class (e.g. eumelanin or pheomelanin, carotenoid,

structural, pteridine, or unknown), and the measure of aggression used (e.g., direct aggression such as

bites, or indirect such as proximity). We also collected the life stage (adult vs juvenile) and sex of the

focal individuals, location of the study (wild, lab-reared, domesticated, wild individuals measured in the

lab), vertebrate or invertebrate taxa, plasticity of pigment expression (plastic or fixed), seasonality of

pigment trait (year-round, breeding, or non-breeding), geographic location of focal or source

population, if the condition of the animal was controlled and how (e.g., weight, length, or weight and

length), if social rank was considered in the study design and how (e.g., uncontrolled, dummy used,

isolated, or recording used), if the age of the animal was controlled and how (e.g., same age or covariate

in analysis) and whether the study was an observational or experimental study. Finally, we recorded the

measure of association between color and aggression (means/standard deviations of discrete groups, t-,

F-, or χ^2-test statistics with the associated p-values and degrees of freedom, or a correlation coefficient),

and the sample sizes for each measure of association.

Effect Size Calculations: For a standardized effect size, when possible, we used reported Pearson's

correlation coefficients between color and aggression, which we then converted to Fisher Z statistics.

The Fisher Z transformation is recommended to normalize the sampling distribution of correlation

coefficient estimates when sample sizes are small and produces less biased results (Silver and Dunlap

1987; Berry and Mielke 2000). The Fisher Z transformation also widens the distribution around 0, which

is useful because Markov Chain Monte Carlo's (MCMC) have difficulty producing accurate estimates

when the true value of the mean is very close to zero (Lipsey and Wilson 2001; Hadfield 2010).

187   When studies did not report correlation coefficients, we used reported F or χ^2 statistics and

converted these into correlation coefficient estimates using methods described in Nakagawa et al.

(2007) and Lipsey and Wilson (2001). When studies analyzed categorical data using a t test or reported

only means and standard deviations for discrete groups, we could not calculate the product-moment

correlation coefficient (Pearson's r) and instead calculated the biserial correlation coefficient (Jacobs

and Viechtbauer 2017). The biserial-correlation coefficient is comparable to the product-moment

correlation and can therefore be used in the same meta-analysis (Jacobs and Viechtbauer, 2017). We

then converted both Pearson's and biserial correlation estimates to Fisher Z using the R package

DescTools and the command FisherZ (Signorell 2021). Biserial-correlation coefficients are sometimes

calculated to be greater than 1 or less than -1 (Pustejovsky 2014); values outside this range are

undefined under the Fisher Z transformation (Fisher 1915; Silver and Dunlap 1987). This was the case

with five positive values in our dataset; these five positive values were reasonably close to 1, total range

= -0.77 to 1.33. We therefore converted the positive values to 0.99, as appropriate, as recommended by

Pustejovsky (2014).

Phylogenetic Meta-Analytic Model: To account for the non-independence of the data due to

evolutionary history and the relationship between species, we constructed a phylogenetic tree of all the

species (70 in total, 66 vertebrates, and 4 invertebrates across 9 classes; Supplemental Figure S1). We

used an ultrametric tree that was fully resolved to the species level, which we obtained from

TimeTree.org (Hedges et al. 2006; Kumar et al. 2017). We then rooted the tree with the anemone

Phymactis clematis as the outgroup because it falls outside of Bilateria, to which all species in our

dataset belong. We used the R packages *ape, phytools,* and *TreeTools* to obtain the relatedness matrix,

root the tree, change the edge lengths from 0 to 0.00001, and plot the tree (Paradis et al. 2004; Revell

2012; Smith and Wickham 2019).

We used the R package *MCMCglmm* to perform the meta-analysis (Hadfield 2010). We chose an

expanded prior with a Cauchy distribution that mirrored the Fisher Z distribution (Adams 2008). We ran

each model for 2,000,000 interactions and removed 1,000,000 steps as the burnin with a thin of 1,000.

Once we determined the models with the lowest DIC values, we reran the analysis using 5,000,000

iterations with burnin of 2,500,000 and a thin value of 1,000; these parameters always produced

convergence and final values that were in the Fisher Z distribution bounded by [-2.64, 2.64]. All credible

intervals reported are based on the last 2,500,000 iterations of the *MCMCglmm*. Prior to all analysis, we

removed one data point that was the sole value associated with the color class pteridine (Robertson and

Rosenblum 2010).

Random and Mixed Models: We first evaluated a model containing only random effects of species,

study, the weights associated with each study (calculated as the inverse of the standard error for the

Fisher Z for each study), and the phylogenetic tree. Weights were added as a variance-covariance matrix

using the "us" option; phylogenetic information was incorporated as a relatedness matrix with the

"pedigree" option, which assumes a Brownian-motion model of evolution (Hadfield 2010; Nakagawa

and Santos 2012). We removed the random effects using backward elimination to confirm the

importance of each random effect. We found that the model with the lowest DIC value was that which

included all random effects (see Results). Retaining all random effects, we then assessed whether any

moderators improved model fit compared to the random-effects-only model (hereafter, "random

effects model"). We tested each of the following moderators one at a time, and in all two-way

combinations: type of color classification, plasticity, sex, life stage, vertebrate or invertebrate, location

of the study, seasonality of the color, geographic location of the study population, observational vs.

experimental studies, if condition of the animal was controlled and how, if social rank was controlled

and how, if age of focal animals was controlled and how, and type of aggressive acts (direct or indirect).

We also tested for two-way interactions between pairs of moderators that we deemed biologically

likely: color class by plasticity, color class by sex, and sex by plasticity. We compared these models using

DIC values. For the best-fitting models, we computed medians and 95% confidence intervals of the

effect size for each moderator using the R package *emmeans* (Lenth et al. 2021).

Some of the moderators listed above were not available in all the studies in our data set:

condition, social rank, and age of focal animals. We accounted for this in two ways. First, as described

above, we used a moderator that indicated whether the feature was controlled for or used as a

covariate in the study. For example, social rank could be uncontrolled, controlled by using unfamiliar

animals, or controlled by using a mirror test or video. For each of these three moderators, we also asked

if including only those studies that controlled for the moderator produced substantially different results.

Because we were particularly interested in any difference in effect size between different color types,

we also included color class as an additional moderator in these "subset" models.

Finally, we investigated whether effect sizes that were reported for "unknown" color classes

were potentially modulated by the melanocortin pathway. Recent studies suggest that, at least in fish,

the melanocortin system can regulate non-melanin-based colors (reviewed in Cal et al. 2017). To

determine if expanding the color classes regulated by the melanocortin system affected the results, we

recoded all unknown color classes as eumelanin and re-ran the mixed effects model with color class as a

moderator.

Heterogeneity: Ecology and evolutionary biology meta-analyses are likely to exhibit high levels of

heterogeneity due to differences in taxa and experimental methods across studies (Gurevitch and

Hedges 1999; Senior et al. 2016). We used a method developed by Nakagawa and Santos (2012) to

quantify the proportion of the total variance due to phylogeny (termed "phylogenetic signal" or H^2) their

Equation 26, and the heterogeneity due to study (I_s^2) and species (I_a^2) using Equations 24 and 25 of that

paper, respectively. To make the values more easily interpretable, we reported the heterogeneity values

as percentages, rather than proportions. Percentages close to 0 indicate low heterogeneity while

percentages close to 1 are considered high heterogeneity. The phylogenetic signal was left as a

proportion.

Publication Bias: Publication bias due to selective reporting of results can influence both the estimated

magnitude and reliability of the overall effect size estimate (Rosenthal 1979; Gurevitch and Hedges

1999). We investigated the possibility of publication bias by visualizing study asymmetry using contour

enhanced funnel plots and by testing for asymmetry using a modified Egger's test as described in

Nakagawa and Santos (2012). This test regresses the meta-analytic residuals on the precision of each

effect size estimate (inverse of standard error). These residuals, unlike the weighted effect sizes

themselves, are independent, and thus satisfy the assumptions of the test. We corrected for the effect

of publication bias using the trim-and-fill method implemented in the R package *meta* (Duval and

Tweedie 2000; Schwarzer 2007). This method restores funnel plot symmetry by iteratively removing

studies with large positive residuals and imputing missing effect size estimates. Because we used meta-

analytic residuals for this analysis, the resulting estimates of the mean effect size estimates are the

adjustment required to restore the funnel plot symmetry (Duval and Tweedie 2000). All analyses were

conducted in R version 4.3.1 (R Core Team 2023). Data and scripts for all analyses are available in a

GitHub repository (https://github.com/sruckman/meta-analysis).

Results

Full data set analysis

We calculated 159 effect sizes from 70 studies that met our inclusion criteria. The random

effects model with the lowest DIC value included all random effects (phylogenetic tree, species, study,

and weights; Supplemental Table S2). This model had a mean posterior effect size of 0.251 (95% credible

interval = (0.018, 0.458); Pearson's correlation mean = 0.246 and 95% credible interval = (0.018, 0.428)),

indicating support for a positive association between the intensity of color and aggression (Figure 2).

In the model that included all of the random effects, phylogenetic signal accounted for 21.0% of

the variation in the dataset (Supplemental Table S2, Figure 2). However, when we removed the

phylogeny, species, and study sequentially, the only term that caused substantially poorer fit when

removed was the study term (Spiegelhalter et al. 2002; Supplemental Table S2). Removing only the

phylogenetic signal or removing the tree alone did not result in a poorer fit when compared to the full

random effects model. This suggests that the "phylogenetic" signal arises because some taxonomic

groups are represented by one or a very few studies (e.g., Amphibia, see Figure 2). We therefore

interpret this signal as mainly reflecting variation among studies, not a true phylogenetic pattern.

None of the fixed effect moderators we investigated improved the fit of the full random effects

model; that is, adding any moderator increased the DIC value. Some of the mixed effects models

exhibited DIC values that were close to that of the full random effects model (Supplemental Table S2

and Supplemental Table S3). For example, a mixed model that included type of aggressive act as a

moderator had a DIC value that was very slightly higher than that of the full random effects model (-

262.442, Supplemental Table S3). No other model had a lower DIC value than these two. Critically, no

models that included color class (eumelanin, pheomelanin, carotenoid, structural, or unknown colors) as

a moderator, either alone or in combination with any other moderator, ever achieved a DIC value below

-258. Consequently, there is no evidence in this data set for color-type differences in the association

between intensity of color and aggression.

We did find evidence of a publication bias for the full random effects model. In this model, the

intercept for the modified Egger's regression was significantly different from zero (intercept ± SE:

0.777±0.3815, t-value = 2.03, p-value = 0.0433). The trim and fill method added zero imputed values to

the original 159 effect size estimates and indicated an adjustment of 0.0233 to the Fisher Z values to

restore funnel plot symmetry (Figure 3). After this adjustment, the overall mean Fisher Z value

increased, but still indicated a positive association between color and aggressive behavior (adjusted

mean = 0.274 and 95% credible interval = (0.041, 0.481); Pearson's correlation coefficient adjusted

mean = 0.267, and 95% credible interval = (0.041, 0.447); Figure 2).

To determine if the melanocortin pleiotropy hypothesis would be better supported if we

reclassified "unknown" color types as "eumelanic", we ran the mixed effects model that included color

class as a moderator on this recoded data set. This "recoded" mixed model had a similar posterior effect

distribution (mean = 0.261, 95% credible interval: (-0.020, 0.559)). As in the original data set, adding the

color moderator did not improve the fit (DIC Value for recoded color model: -259.414, Supplemental

Table S3). Notably, the mixed model did not provide a better fit than the random effects model for

either data set, suggesting that the relationship between color intensity and aggression does not vary

based on the type of color, as predicted by the melanocortin pleiotropy hypothesis.

Analysis of data subsets

Results did not differ substantially in analyses of data subsets that included only those studies

that controlled for animal condition, social rank, or age. As in analyses of the full data set, the random

effects model was a better fit than a model that included color type as a moderator for all the data

subsets (Supplemental Table S4). The mean posterior effect sizes for the relationship between color and

aggression was also similar in magnitude to the mean estimate of the full data set, ranging from 0.177 to

0.307 (Supplemental Table S4). Fewer studies controlled for animal age (beyond juvenile/adult)

compared to those that controlled for social rank or condition. Analysis of the subset that did control for

age (16 studies) or for age, condition, and social rank (9 studies), also produced similar mean posterior

effect size estimates, but in these cases, the credible interval for the mean effect size did overlap zero

(Supplemental Table S4).

Discussion

The meta-analysis of 70 published studies indicated a positive association between measures of

aggression and measures indicating more colorful individuals. For simplicity, we used the term "colorful"

to refer to variation in hue, intensity, or area as used in the original studies. This positive relationship

held after correcting for publication bias. No moderator that we evaluated improved the fit of the meta-

analytic model to the data, indicating a robust relationship between color and aggression. Critically, we

found no evidence that this relationship depended on the type of color (eumelanic, phaeomelanic,

carotenoid, or structural), counter to predictions of the melanin-pleiotropy and carotenoid-condition-

dependence hypotheses. This result was unchanged when we recoded colors of unknown cause, but for

which a melanocortin-based mechanism is plausible. Both the badge-of-status and general condition-

dependence hypotheses are consistent with this pattern. However, under general condition-

dependence, we expect the effect size to be sensitive to moderators indicating whether and how

condition was accounted for in the study design, or to moderators that are plausibly associated with

condition, such as social rank and age. None of these moderators explained variation in effect sizes in

our analyses, and none affected the (lack of) dependence of effect size on type of color.

It is possible that different mechanisms underly similar correlations between aggression and

different kinds of coloration. For example, it is possible that covariation between melanin-based colors

and aggression in the relevant studies in our data set is indeed regulated by variation in the

melanocortin pathway, and that covariation between aggression and carotenoid color is regulated by

condition dependence that was not accounted for by any of the proxies for condition that we included

in our analyses. However, the badge of status hypothesis is a more parsimonious explanation for these

patterns (Tibbetts and Dale 2004; Santos et al. 2011).

Potential caveats to this conclusion include failure of moderators to improve the fit can result

from including studies with low sample size and high variation, or from including few studies assessing

the moderators of interest (Ginzburg and Jensen 2004; Lajeunesse 2009). For key moderators, this study

had robust sample sizes. For example, in the condition category, 101 measures derived from studies that

accounted for animal condition in some way, while 58 measures did not. In the subset of data that

included only studies accounting for condition in some way, 34 measures accounted for both length and

weight, 20 for weight only, and 47 for length only. In addition, the low heterogeneity observed in the

pure random-effects model suggests that power was not severely compromised by low sample size or

high uncertainty in effect size estimates. Nevertheless, some combinations of moderators (e.g., color

class, social rank, and age) were represented by few effect sizes in our data set (Supplemental Table S4).

Consequently, if condition is affected by adult age, as seems likely, current literature might be

inadequate to evaluate the general-condition-dependence hypothesis. We therefore suggest that future

studies control for animal age, body condition, and social rank (where applicable).

We did observe considerable variation in effect sizes among studies, and the among-study

variation was the main contributor to heterogeneity in our analysis. In contrast, species and

phylogenetic relatedness did not explain additional heterogeneity. For example, the most extreme

effect sizes (both positive and negative) are found within Actinopterygii (Figure 2). The strongest

negative association derived from a single study of orange color intensity in female convict cichlids

(Beeching et al. 1998). In that study, females with the most orange color displayed the lowest level of

aggression toward a stimulus female. The most positive associations between color and aggression

occurred in a study of plastic eye color in juvenile guppies (*Poecilia reticulata*; Martin and Hengstebeck

1981). Fish with darker eye color engaged in and won more aggressive encounters than the light-eyed

fish. Losers in these encounters also lightened their eye color, consistent with some studies of traits

deemed to be badges of status (Dey et al. 2014).

In our data set, effect sizes within studies tended to be tightly clustered. This clustering suggests

that even closely-related species can differ appreciably in the associations we measured. Alternately,

this pattern could be driven by differences in methodologies, as was found in a meta-analysis of birds

(Santos et al. 2011). We categorized studies based on whether measures of aggression were direct or

indirect (as in Santos et al. 2011), and on whether the general methodology was observational or

experimental; neither of these categorizations was associated with variation in effect size. We do

recommend that future studies ideally take an experimental approach and that they include direct

measures of aggression when possible.

Of the four hypotheses explaining consistent associations between animal coloration and

aggression, our results are most parsimoniously explained by the badge of status hypothesis. This

hypothesis proposes that aggressiveness, fighting ability, or dominance status is honestly reflected by

variation in a trait that is perceptible to conspecifics (Rohwer 1975; McGraw et al. 2003; Tibbetts and

Dale 2004). While many studies suggest that melanocortin-based genetic pleiotropy can constrain the

joint evolution of color and behavior, whether the badge-of-status mechanism should impose such

constraints has received less attention. Under the badge-of-status hypothesis, the correlation between

color and aggression could be caused by pleiotropy (Santos et al. 2011; Küpper et al. 2016; Lamichhaney

et al. 2016; Sánchez-Tójar et al. 2018). The supergene regulating feather coloration and social status in

ruffs is a prime example (Küpper et al. 2016; Lamichhaney et al. 2016). The relationship between head

stripe color and aggression in white-throated sparrows is another likely example of genetically-based

covariation that arises from the badge of status mechanism (Lowther 1961; Knapton and Falls 1983;

Tuttle 2003; Tuttle et al. 2016; Hedrick et al. 2018). Variation in a badge of status could also be

regulated by non-genetic variation in resource availability or acquisition, variation in exposure to disease

or parasites, or non-genetic maternal effects (Rohwer 1975; Dawkins and Krebs 1978). However, like for

other condition-dependent mechanisms, our analysis does not support this interpretation for the overall

trend we observed: a moderate positive correlation between coloration and behavior across *Animalia*.

We therefore propose that this moderate correlation is underlain by genetic covariation between

behavior and color traits that serve as badges of status. A key question remains: is this genetic

correlation underlain by the same mechanism across different color types? This could occur, for

example, if genetic variation in condition regulates both behavior and color, irrespective of the color of

the badge. Alternatively, do different mechanisms, e.g. melanocortin pleiotropy in the case of

eumelanin color, and genetic variation in condition for carotenoid colors. Studies of the basis of genetic

covariance variety of systems will be needed to answer this question.

To date, few studies have attempted to partition covariation between color and behavior into

heritable and non-heritable components, and most of these have been in systems characterized by

discrete color polymorphisms (e.g., Tuttle 2003; Küpper et al. 2016; Lamichhaney et al. 2016). In

tractable organisms (mainly short-lived invertebrates), artificial selection or experimental evolution

could be used to measure genetic correlation between traits and thus, directly assess whether color-

behavior associations impose constraints on evolutionary change. Many free-living populations (mainly

vertebrates) have been the subject of long-term observational study, and now have pedigree data that

could allow genetic correlations between color and behavior to be estimated (Clutton-Brock and

Pemberton 2004; Blondel et al. 2006; Charmantier et al. 2006; Foerster et al. 2007; McAdam et al.

2007). These kinds of data will become increasingly available as more long-term studies incorporate

genomic analyses of relatedness. Consequently, both experimental and non-experimental studies in a

variety of species will be necessary to understand the extent to which color-behavior correlations

influence evolutionary dynamics.

Figures and Tables:

Figure 1: PRISMA Diagram for Papers Examined. Solid arrows indicate papers that moved on to the next

step and dashed arrows indicate papers that were removed from the analysis. First, we identified papers

using Web of Science and found 751 papers. Next, we screened these papers for human studies and

those that clearly did not examine color and aggression. This removed 521 papers. Finally, we assessed

the 230 papers based on our inclusion criteria and removed any studies that did not meet all of the

inclusion criteria. We found that 70 papers met our inclusion criteria and were analyzed in our meta-

analysis.

Figure 2: The effect size and 95% credible intervals for all studies listed by family. The study names are

on the left side of the graph and any study with a dash then a number is a second effect size from the

same study. The colors of the points indicate the color class, while the shape of the point indicates if the

trait was plastic or non-plastic. The points at the top of the graph are the mean effect size and 95%

credible intervals for the random effects model. The black triangle and line through it indicate the model

mean with 95% credible intervals before publication bias correction. Black diamond and line indicate

model means with 95% credible intervals after publication bias correction.

Figure 3: Funnel Plot of Random Effects Only Model. No points were added to restore plot symmetry.

Acknowledgements:

We thank the Hughes Lab, Emily DuVal, and David Houle for their helpful comments throughout the

data collection process and for valuable suggestions that improved this manuscript.

Funding Sources:

This work was funded by the National Science Foundation Grant # DEB-1740466.

Author Contribution:

SNR: Conceptualization, Methods development/experimental design, Data Collection, Data Analysis,

Data Visualization, Model Analysis, Coding Simulation, Supervision, Writing – original draft and review &

editing. EAH: Conceptualization, Methods development/experimental design, Data Collection, Writing –

original draft and review & editing. LM, IP, and MP: Data Collection. KAH: Conceptualization, Methods

development/experimental design, Supervision, Writing – original draft and review & editing

Literature Cited

Adams, D. C. 2008. Phylogenetic Meta-Analysis. Evolution 62:567–572.

Backström, T., M. Heynen, E. Brännäs, J. Nilsson, and C. Magnhagen. 2015. Dominance and stress

signalling of carotenoid pigmentation in Arctic charr (Salvelinus alpinus): Lateralization effects?

Physiol. Behav. 138:52–57.

Beck, M. L., S. Davies, and K. B. Sewall. 2018. Urbanization alters the relationship between coloration

and territorial aggression, but not hormones, in song sparrows. Anim. Behav. 142:119–128.

Beeching, S. C., S. H. Gross, H. S. Bretz, and E. Hariatis. 1998. Sexual dichromatism in convict cichlids: the

ethological significance of female ventral coloration. Anim. Behav. 56:1021–1026.

Berry, K. J., and P. W. Mielke. 2000. A Monte Carlo Investigation of the Fisher Z Transformation for

Normal and Nonnormal Distributions. Psychol. Rep. 87:1101–1114. SAGE Publications Inc.

Blondel, J., D. W. Thomas, A. Charmantier, P. Perret, P. Bourgault, and M. M. Lambrechts. 2006. A Thirty-

464   Year Study of Phenotypic and Genetic Variation of Blue Tits in Mediterranean Habitat Mosaics.

465   BioScience 56:661–673.

466   Blount, J. D., and K. J. McGraw. 2008. Signal Functions of Carotenoid Colouration. Pp. 213–236 *in* G.

467   Britton, S. Liaaen-Jensen, and H. Pfander, eds. Carotenoids: Volume 4: Natural Functions.

468   Birkhäuser, Basel.

469   Blows, M., and B. Walsh. 2009. Spherical Cows Grazing in Flatland: Constraints to Selection and

470   Adaptation. Pp. 83–101 *in* J. van der Werf, H.-U. Graser, R. Frankham, and C. Gondro, eds.

471   Adaptation and Fitness in Animal Populations: Evolutionary and Breeding Perspectives on

472   Genetic Resource Management. Springer Netherlands, Dordrecht.

473   Boerner, M., and O. Krüger. 2009. Aggression and fitness differences between plumage morphs in the

474   common buzzard (Buteo buteo). Behav. Ecol. 20:180–185.

475   Cal, L., P. Suarez-Bregua, J. M. Cerdá-Reverter, I. Braasch, and J. Rotllant. 2017. Fish pigmentation and

476   the melanocortin system. Comp. Biochem. Physiol. A. Mol. Integr. Physiol. 211:26–33.

477   Charmantier, A., C. Perrins, R. H. McCleery, and B. C. Sheldon. 2006. Evolutionary Response to Selection

478   on Clutch Size in a Long-Term Study of the Mute Swan. Am. Nat. 167:453–465. The University of

479   Chicago Press.

480   Clutton-Brock, T. H., and J. M. Pemberton (eds). 2004. Soay sheep: dynamics and selection in an island

481   population. Cambridge University Press, Cambridge England; New York.

482   Dawkins, R., and J. R. Krebs. 1978. Animal signals: information or manipulation. Behav. Ecol. Evol.

483   Approach 2:282–309.

484   Deng, L., and S. Xu. 2017. Adaptation of human skin color in various populations. Hereditas 155:1.

485   Dey, C. J., J. Dale, and J. S. Quinn. 2014. Manipulating the appearance of a badge of status causes

486   changes in true badge expression. Proc. R. Soc. B Biol. Sci. 281:20132680. Royal Society.

Dijkstra, P. D., S. M. Maguire, R. M. Harris, A. A. Rodriguez, R. S. DeAngelis, S. A. Flores, and H. A.

Hofmann. 2017. The melanocortin system regulates body pigmentation and social behaviour in

a colour polymorphic cichlid fish†. Proc. R. Soc. B Biol. Sci. 284:20162838. Royal Society.

Ducrest, A.-L., L. Keller, and A. Roulin. 2008. Pleiotropy in the melanocortin system, coloration and

behavioural syndromes. Trends Ecol. Evol. 23:502–510.

Duval, S., and R. Tweedie. 2000. Trim and Fill: A Simple Funnel-Plot–Based Method of Testing and

Adjusting for Publication Bias in Meta-Analysis. Biometrics 56:455–463.

Fisher, R. A. 1915. Frequency Distribution of the Values of the Correlation Coefficient in Samples from an

Indefinitely Large Population. Biometrika 10:507–521. [Oxford University Press, Biometrika

Trust].

Foerster, K., T. Coulson, B. C. Sheldon, J. M. Pemberton, T. H. Clutton-Brock, and L. E. B. Kruuk. 2007.

Sexually antagonistic genetic variation for fitness in red deer. Nature 447:1107–1110. Nature

Publishing Group.

Ginzburg, L. R., and C. X. J. Jensen. 2004. Rules of thumb for judging ecological theories. Trends Ecol.

Evol. 19:121–126.

Gurevitch, J., and L. V. Hedges. 1999. Statistical Issues in Ecological Meta-Analyses. Ecology 80:1142–

1149.

Hadfield, J. D. 2010. MCMC Methods for Multi-Response Generalized Linear Mixed Models: The

MCMCglmm R Package. J. Stat. Softw. 33:1–22.

Hedges, S. B., J. Dudley, and S. Kumar. 2006. TimeTree: a public knowledge-base of divergence times

among organisms. Bioinformatics 22:2971–2972.

Hedrick, P. W., E. M. Tuttle, and R. A. Gonser. 2018. Negative-Assortative Mating in the White-Throated

Sparrow. J. Hered. 109:223–231.

Horth, L. 2003. Melanic body colour and aggressive mating behaviour are correlated traits in male

mosquitofish (Gambusia holbrooki). Proc. R. Soc. Lond. B Biol. Sci. 270:1033–1040. Royal

Society.

Huhta, E., and R. V. Alatalo. 1993. Plumage colour and male-male interactions in the pied flycatcher.

Anim. Behav. 45:511–518.

Jacobs, P., and W. Viechtbauer. 2017. Estimation of the biserial correlation and its sampling variance for

use in meta-analysis. Res. Synth. Methods 8:161–180.

Johnstone, R. A., and K. Norris. 1993. Badges of status and the cost of aggression. Behav. Ecol. Sociobiol.

32:127–134.

Kenyon, H. L., and P. R. Martin. 2023. Color as an Interspecific Badge of Status: A Comparative Test. Am.

Nat. 202:433–447. The University of Chicago Press.

Knapton, R. W., and J. B. Falls. 1983. Differences in parental contribution among pair types in the

polymorphic White-throated Sparrow. Can. J. Zool. 61:1288–1292. NRC Research Press.

Kumar, S., G. Stecher, M. Suleski, and S. B. Hedges. 2017. TimeTree: A Resource for Timelines, Timetrees,

and Divergence Times. Mol. Biol. Evol. 34:1812–1819.

Küpper, C., M. Stocks, J. E. Risse, N. dos Remedios, L. L. Farrell, S. B. McRae, T. C. Morgan, N. Karlionova,

P. Pinchuk, Y. I. Verkuil, A. S. Kitaysky, J. C. Wingfield, T. Piersma, K. Zeng, J. Slate, M. Blaxter, D.

B. Lank, and T. Burke. 2016. A supergene determines highly divergent male reproductive morphs

in the ruff. Nat. Genet. 48:79–83. Nature Publishing Group.

Lajeunesse, M. J. 2009. Meta-Analysis and the Comparative Phylogenetic Method. Am. Nat. 174:369–

381. The University of Chicago Press.

Lamichhaney, S., G. Fan, F. Widemo, U. Gunnarsson, D. S. Thalmann, M. P. Hoeppner, S. Kerje, U.

Gustafson, C. Shi, H. Zhang, W. Chen, X. Liang, L. Huang, J. Wang, E. Liang, Q. Wu, S. M.-Y. Lee, X.

Xu, J. Höglund, X. Liu, and L. Andersson. 2016. Structural genomic changes underlie alternative

reproductive strategies in the ruff (Philomachus pugnax). Nat. Genet. 48:84–88. Nature

Publishing Group.

Lenth, R., H. Singmann, J. Love, P. Buerkner, and M. Herve. 2021. Estimated Marginal Means, aka Least-

Squares Means.

Lipsey, M. W., and D. B. Wilson. 2001. Practical meta-analysis. Sage Publications, Inc, Thousand Oaks,

CA, US.

Lowther, J. K. 1961. Polymorphism in the white-throated sparrow, zonotrichia albicollis (gmelin). Can. J.

Zool. 39:281–292. NRC Research Press.

Martin, F. D., and M. F. Hengstebeck. 1981. Eye colour and aggression in juvenile guppies, *Poecilia*

reticulata peters (Pisces: Poeciliidae). Anim. Behav. 29:325–331.

Massey, J. H., and P. J. Wittkopp. 2016. The Genetic Basis of Pigmentation Differences Within and

Between Drosophila Species. Curr. Top. Dev. Biol. 119:27–61.

McAdam, A. G., S. Boutin, A. K. Sykes, and M. M. Humphries. 2007. Life histories of female red squirrels

and their contributions to population growth and lifetime fitness. Écoscience 14:362–369. Taylor

& Francis.

McGraw, K. J., J. Dale, and E. A. Mackillop. 2003. Social environment during molt and the expression of

melanin-based plumage pigmentation in male house sparrows (Passer domesticus). Behav. Ecol.

Sociobiol. 53:116–122.

McGraw, K. J., E. A. Mackillop, J. Dale, and M. E. Hauber. 2002. Different colors reveal different

information: how nutritional stress affects the expression of melanin- and structurally based

ornamental plumage. J. Exp. Biol. 205:3747–3755.

Moore, M. P., and R. A. Martin. 2016. Intrasexual selection favours an immune-correlated colour

ornament in a dragonfly. J. Evol. Biol. 29:2256–2265.

Nakagawa, S., N. Ockendon, D. O. Gillespie, B. J. Hatchwell, and T. Burke. 2007. Assessing the function of

house sparrows bib size using a flexible meta-analysis method. Behav. Ecol. 18:831. Oxford

University Press, Great Britain.

Nakagawa, S., and E. S. A. Santos. 2012. Methodological issues and advances in biological meta-analysis.

Evol. Ecol. 26:1253–1274.

Paradis, E., J. Claude, and K. Strimmer. 2004. APE: Analyses of Phylogenetics and Evolution in R language.

Bioinformatics 20:289–290.

Pärt, T., and A. Qvarnström. 1997. Badge size in collared flycatchers predicts outcome of male

competition over territories. Anim. Behav. 54:893–899.

Pustejovsky, J. E. 2014. Converting from d to r to z when the design uses extreme groups,

dichotomization, or experimental control. Psychol. Methods 19:92–112. US: American

Psychological Association.

R Core Team. 2023. R: A Language and Environment for Statistical Computing. R Foundation for

Statistical Computing, Vienna, Austria.

Reissmann, M., and A. Ludwig. 2013. Pleiotropic effects of coat colour-associated mutations in humans,

mice and other mammals. Semin. Cell Dev. Biol. 24:576–586.

Revell, L. J. 2012. phytools: an R package for phylogenetic comparative biology (and other things).

Methods Ecol. Evol. 3:217–223.

Robertson, J. M., and E. B. Rosenblum. 2010. Male territoriality and 'sex confusion' in recently adapted

lizards at White Sands. J. Evol. Biol. 23:1928–1936.

Rohwer, S. 1975. The Social Significance of Avian Winter Plumage Variability. Evolution 29:593–610.

[Society for the Study of Evolution, Wiley].

Rosenthal, R. 1979. The file drawer problem and tolerance for null results. Psychol. Bull. 86:638–641.

American Psychological Association, US.

Roulin, A. 2016. Condition-dependence, pleiotropy and the handicap principle of sexual selection in

melanin-based colouration. Biol. Rev. 91:328–348.

Roulin, A., and A.-L. Ducrest. 2011. Association between melanism, physiology and behaviour: A role for

the melanocortin system. Eur. J. Pharmacol. 660:226–233.

Sánchez-Tójar, A., S. Nakagawa, M. Sánchez-Fortún, D. A. Martin, S. Ramani, A. Girndt, V. Bókony, B.

Kempenaers, A. Liker, D. F. Westneat, T. Burke, and J. Schroeder. 2018. Meta-analysis challenges

a textbook example of status signalling and demonstrates publication bias. eLife 7:e37385. eLife

Sciences Publications, Ltd.

San-Jose, L. M., and A. Roulin. 2018. Toward Understanding the Repeated Occurrence of Associations

between Melanin-Based Coloration and Multiple Phenotypes. Am. Nat. 192:111–130. The

University of Chicago Press.

Santos, E. S. A., D. Scheck, and S. Nakagawa. 2011. Dominance and plumage traits: meta-analysis and

metaregression analysis. Anim. Behav. 82:3–19.

Schartl, M., L. Larue, M. Goda, M. W. Bosenberg, H. Hashimoto, and R. N. Kelsh. 2016. What is a

vertebrate pigment cell? Pigment Cell Melanoma Res. 29:8–14.

Schwarzer, G. 2007. meta: an R package for meta-analysis. R News 7:40–45.

Seddon, R. J., and D. K. Hews. 2017. Correlates of melanization in multiple high- and low-elevation

populations of the lizard, Sceloporus occidentalis: Behavior, hormones, and parasites. J. Exp.

Zool. Part Ecol. Integr. Physiol. 327:481–492.

Senior, A. M., C. E. Grueber, T. Kamiya, M. Lagisz, K. O'Dwyer, E. S. A. Santos, and S. Nakagawa. 2016.

Heterogeneity in ecological and evolutionary meta-analyses: its magnitude and implications.

Ecology 97:3293–3299.

Siefferman, L., and G. E. Hill. 2003. Structural and melanin coloration indicate parental effort and

reproductive success in male eastern bluebirds. Behav. Ecol. 14:855–861.

Signorell, A. 2021. DescTools: Tools for Descriptive Statistics.

Silver, N. C., and W. P. Dunlap. 1987. Averaging correlation coefficients: Should Fisher's z transformation

be used? J. Appl. Psychol. 72:146–148.

Smith, M. R., and H. Wickham. 2019. TreeTools: create, modify and analyse phylogenetic trees.

Spiegelhalter, D. J., N. G. Best, B. P. Carlin, and A. Van Der Linde. 2002. Bayesian measures of model

complexity and fit. J. R. Stat. Soc. Ser. B Stat. Methodol. 64:583–639.

Tibbetts, E. A., and J. Dale. 2004. A socially enforced signal of quality in a paper wasp. Nature 432:218–

222. Nature Publishing Group.

Tuttle, E. M. 2003. Alternative reproductive strategies in the white-throated sparrow: behavioral and

genetic evidence. Behav. Ecol. 14:425–432.

Tuttle, E. M., A. O. Bergland, M. L. Korody, M. S. Brewer, D. J. Newhouse, P. Minx, M. Stager, A. Betuel, Z.

A. Cheviron, W. C. Warren, R. A. Gonser, and C. N. Balakrishnan. 2016. Divergence and

Functional Degradation of a Sex Chromosome-like Supergene. Curr. Biol. 26:344–350.

Walsh, B., and M. W. Blows. 2009. Abundant Genetic Variation + Strong Selection = Multivariate Genetic

Constraints: A Geometric View of Adaptation. Annu. Rev. Ecol. Evol. Syst. 40:41–59.

Wittkopp, P. J., and P. Beldade. 2009. Development and evolution of insect pigmentation: Genetic

mechanisms and the potential consequences of pleiotropy. Semin. Cell Dev. Biol. 20:65–71.

References Only in Online Materials

Adams, D. C. 2008. Phylogenetic Meta-Analysis. Evolution 62:567–572.

Backström, T., M. Heynen, E. Brännäs, J. Nilsson, and C. Magnhagen. 2015. Dominance and stress

signalling of carotenoid pigmentation in Arctic charr (Salvelinus alpinus): Lateralization effects?

Physiol. Behav. 138:52–57.

Beck, M. L., S. Davies, and K. B. Sewall. 2018. Urbanization alters the relationship between coloration

and territorial aggression, but not hormones, in song sparrows. Anim. Behav. 142:119–128.

Beeching, S. C., S. H. Gross, H. S. Bretz, and E. Hariatis. 1998. Sexual dichromatism in convict cichlids: the

ethological significance of female ventral coloration. Anim. Behav. 56:1021–1026.

Berry, K. J., and P. W. Mielke. 2000. A Monte Carlo Investigation of the Fisher Z Transformation for

Normal and Nonnormal Distributions. Psychol. Rep. 87:1101–1114. SAGE Publications Inc.

Blondel, J., D. W. Thomas, A. Charmantier, P. Perret, P. Bourgault, and M. M. Lambrechts. 2006. A Thirty-

Year Study of Phenotypic and Genetic Variation of Blue Tits in Mediterranean Habitat Mosaics.

BioScience 56:661–673.

Blount, J. D., and K. J. McGraw. 2008. Signal Functions of Carotenoid Colouration. Pp. 213–236 *in* G.

Britton, S. Liaaen-Jensen, and H. Pfander, eds. Carotenoids: Volume 4: Natural Functions.

Birkhäuser, Basel.

Blows, M., and B. Walsh. 2009. Spherical Cows Grazing in Flatland: Constraints to Selection and

Adaptation. Pp. 83–101 *in* J. van der Werf, H.-U. Graser, R. Frankham, and C. Gondro, eds.

Adaptation and Fitness in Animal Populations: Evolutionary and Breeding Perspectives on

Genetic Resource Management. Springer Netherlands, Dordrecht.

Boerner, M., and O. Krüger. 2009. Aggression and fitness differences between plumage morphs in the

common buzzard (Buteo buteo). Behav. Ecol. 20:180–185.

Cal, L., P. Suarez-Bregua, J. M. Cerdá-Reverter, I. Braasch, and J. Rotllant. 2017. Fish pigmentation and

the melanocortin system. Comp. Biochem. Physiol. A. Mol. Integr. Physiol. 211:26–33.

Charmantier, A., C. Perrins, R. H. McCleery, and B. C. Sheldon. 2006. Evolutionary Response to Selection

on Clutch Size in a Long-Term Study of the Mute Swan. Am. Nat. 167:453–465. The University of

Chicago Press.

Clutton-Brock, T. H., and J. M. Pemberton (eds). 2004. Soay sheep: dynamics and selection in an island

population. Cambridge University Press, Cambridge England; New York.

Dawkins, R., and J. R. Krebs. 1978. Animal signals: information or manipulation. Behav. Ecol. Evol.

Approach 2:282–309.

Deng, L., and S. Xu. 2017. Adaptation of human skin color in various populations. Hereditas 155:1.

Dey, C. J., J. Dale, and J. S. Quinn. 2014. Manipulating the appearance of a badge of status causes

changes in true badge expression. Proc. R. Soc. B Biol. Sci. 281:20132680. Royal Society.

Dijkstra, P. D., S. M. Maguire, R. M. Harris, A. A. Rodriguez, R. S. DeAngelis, S. A. Flores, and H. A.

Hofmann. 2017. The melanocortin system regulates body pigmentation and social behaviour in

a colour polymorphic cichlid fish†. Proc. R. Soc. B Biol. Sci. 284:20162838. Royal Society.

Ducrest, A.-L., L. Keller, and A. Roulin. 2008. Pleiotropy in the melanocortin system, coloration and

behavioural syndromes. Trends Ecol. Evol. 23:502–510.

Duval, S., and R. Tweedie. 2000. Trim and Fill: A Simple Funnel-Plot–Based Method of Testing and

Adjusting for Publication Bias in Meta-Analysis. Biometrics 56:455–463.

Fisher, R. A. 1915. Frequency Distribution of the Values of the Correlation Coefficient in Samples from an

Indefinitely Large Population. Biometrika 10:507–521. [Oxford University Press, Biometrika

Trust].

Foerster, K., T. Coulson, B. C. Sheldon, J. M. Pemberton, T. H. Clutton-Brock, and L. E. B. Kruuk. 2007.

Sexually antagonistic genetic variation for fitness in red deer. Nature 447:1107–1110. Nature

Publishing Group.

Ginzburg, L. R., and C. X. J. Jensen. 2004. Rules of thumb for judging ecological theories. Trends Ecol.

Evol. 19:121–126.

Gurevitch, J., and L. V. Hedges. 1999. Statistical Issues in Ecological Meta-Analyses. Ecology 80:1142–

1149.

Hadfield, J. D. 2010. MCMC Methods for Multi-Response Generalized Linear Mixed Models: The

MCMCglmm R Package. J. Stat. Softw. 33:1–22.

Hedges, S. B., J. Dudley, and S. Kumar. 2006. TimeTree: a public knowledge-base of divergence times

among organisms. Bioinformatics 22:2971–2972.

Hedrick, P. W., E. M. Tuttle, and R. A. Gonser. 2018. Negative-Assortative Mating in the White-Throated

Sparrow. J. Hered. 109:223–231.

Horth, L. 2003. Melanic body colour and aggressive mating behaviour are correlated traits in male

mosquitofish (Gambusia holbrooki). Proc. R. Soc. Lond. B Biol. Sci. 270:1033–1040. Royal

Society.

Huhta, E., and R. V. Alatalo. 1993. Plumage colour and male-male interactions in the pied flycatcher.

Anim. Behav. 45:511–518.

Jacobs, P., and W. Viechtbauer. 2017. Estimation of the biserial correlation and its sampling variance for

use in meta-analysis. Res. Synth. Methods 8:161–180.

Johnstone, R. A., and K. Norris. 1993. Badges of status and the cost of aggression. Behav. Ecol. Sociobiol.

32:127–134.

Kenyon, H. L., and P. R. Martin. 2023. Color as an Interspecific Badge of Status: A Comparative Test. Am.

Nat. 202:433–447. The University of Chicago Press.

Knapton, R. W., and J. B. Falls. 1983. Differences in parental contribution among pair types in the

polymorphic White-throated Sparrow. Can. J. Zool. 61:1288–1292. NRC Research Press.

Kumar, S., G. Stecher, M. Suleski, and S. B. Hedges. 2017. TimeTree: A Resource for Timelines, Timetrees,

and Divergence Times. Mol. Biol. Evol. 34:1812–1819.

Küpper, C., M. Stocks, J. E. Risse, N. dos Remedios, L. L. Farrell, S. B. McRae, T. C. Morgan, N. Karlionova,

P. Pinchuk, Y. I. Verkuil, A. S. Kitaysky, J. C. Wingfield, T. Piersma, K. Zeng, J. Slate, M. Blaxter, D.

B. Lank, and T. Burke. 2016. A supergene determines highly divergent male reproductive morphs

in the ruff. Nat. Genet. 48:79–83. Nature Publishing Group.

Lajeunesse, M. J. 2009. Meta-Analysis and the Comparative Phylogenetic Method. Am. Nat. 174:369–

381. The University of Chicago Press.

Lamichhaney, S., G. Fan, F. Widemo, U. Gunnarsson, D. S. Thalmann, M. P. Hoeppner, S. Kerje, U.

Gustafson, C. Shi, H. Zhang, W. Chen, X. Liang, L. Huang, J. Wang, E. Liang, Q. Wu, S. M.-Y. Lee, X.

Xu, J. Höglund, X. Liu, and L. Andersson. 2016. Structural genomic changes underlie alternative

reproductive strategies in the ruff (Philomachus pugnax). Nat. Genet. 48:84–88. Nature

Publishing Group.

Lenth, R., H. Singmann, J. Love, P. Buerkner, and M. Herve. 2021. Estimated Marginal Means, aka Least-

Squares Means.

Lipsey, M. W., and D. B. Wilson. 2001. Practical meta-analysis. Sage Publications, Inc, Thousand Oaks,

CA, US.

Lowther, J. K. 1961. Polymorphism in the white-throated sparrow, zonotrichia albicollis (gmelin). Can. J.

Zool. 39:281–292. NRC Research Press.

Martin, F. D., and M. F. Hengstebeck. 1981. Eye colour and aggression in juvenile guppies, *Poecilia*

reticulata peters (Pisces: Poeciliidae). Anim. Behav. 29:325–331.

Massey, J. H., and P. J. Wittkopp. 2016. The Genetic Basis of Pigmentation Differences Within and

Between Drosophila Species. Curr. Top. Dev. Biol. 119:27–61.

McAdam, A. G., S. Boutin, A. K. Sykes, and M. M. Humphries. 2007. Life histories of female red squirrels

and their contributions to population growth and lifetime fitness. Écoscience 14:362–369. Taylor

& Francis.

720 McGraw, K. J., J. Dale, and E. A. Mackillop. 2003. Social environment during molt and the expression of

721   melanin-based plumage pigmentation in male house sparrows (Passer domesticus). Behav. Ecol.

722   Sociobiol. 53:116–122.

723 McGraw, K. J., E. A. Mackillop, J. Dale, and M. E. Hauber. 2002. Different colors reveal different

724   information: how nutritional stress affects the expression of melanin- and structurally based

725   ornamental plumage. J. Exp. Biol. 205:3747–3755.

726 Moore, M. P., and R. A. Martin. 2016. Intrasexual selection favours an immune-correlated colour

727   ornament in a dragonfly. J. Evol. Biol. 29:2256–2265.

728 Nakagawa, S., N. Ockendon, D. O. Gillespie, B. J. Hatchwell, and T. Burke. 2007. Assessing the function of

729   house sparrows bib size using a flexible meta-analysis method. Behav. Ecol. 18:831. Oxford

730   University Press, Great Britain.

731 Nakagawa, S., and E. S. A. Santos. 2012. Methodological issues and advances in biological meta-analysis.

732   Evol. Ecol. 26:1253–1274.

733 Paradis, E., J. Claude, and K. Strimmer. 2004. APE: Analyses of Phylogenetics and Evolution in R language.

734   Bioinformatics 20:289–290.

735 Pärt, T., and A. Qvarnström. 1997. Badge size in collared flycatchers predicts outcome of male

736   competition over territories. Anim. Behav. 54:893–899.

737 Pustejovsky, J. E. 2014. Converting from d to r to z when the design uses extreme groups,

738   dichotomization, or experimental control. Psychol. Methods 19:92–112. US: American

739   Psychological Association.

740 R Core Team. 2023. R: A Language and Environment for Statistical Computing. R Foundation for

741   Statistical Computing, Vienna, Austria.

742 Reissmann, M., and A. Ludwig. 2013. Pleiotropic effects of coat colour-associated mutations in humans,

743   mice and other mammals. Semin. Cell Dev. Biol. 24:576–586.

Revell, L. J. 2012. phytools: an R package for phylogenetic comparative biology (and other things).

Methods Ecol. Evol. 3:217–223.

Robertson, J. M., and E. B. Rosenblum. 2010. Male territoriality and 'sex confusion' in recently adapted

lizards at White Sands. J. Evol. Biol. 23:1928–1936.

Rohwer, S. 1975. The Social Significance of Avian Winter Plumage Variability. Evolution 29:593–610.

[Society for the Study of Evolution, Wiley].

Rosenthal, R. 1979. The file drawer problem and tolerance for null results. Psychol. Bull. 86:638–641.

American Psychological Association, US.

Roulin, A. 2016. Condition-dependence, pleiotropy and the handicap principle of sexual selection in

melanin-based colouration. Biol. Rev. 91:328–348.

Roulin, A., and A.-L. Ducrest. 2011. Association between melanism, physiology and behaviour: A role for

the melanocortin system. Eur. J. Pharmacol. 660:226–233.

Sánchez-Tójar, A., S. Nakagawa, M. Sánchez-Fortún, D. A. Martin, S. Ramani, A. Girndt, V. Bókony, B.

Kempenaers, A. Liker, D. F. Westneat, T. Burke, and J. Schroeder. 2018. Meta-analysis challenges

a textbook example of status signalling and demonstrates publication bias. eLife 7:e37385. eLife

Sciences Publications, Ltd.

San-Jose, L. M., and A. Roulin. 2018. Toward Understanding the Repeated Occurrence of Associations

between Melanin-Based Coloration and Multiple Phenotypes. Am. Nat. 192:111–130. The

University of Chicago Press.

Santos, E. S. A., D. Scheck, and S. Nakagawa. 2011. Dominance and plumage traits: meta-analysis and

metaregression analysis. Anim. Behav. 82:3–19.

Schartl, M., L. Larue, M. Goda, M. W. Bosenberg, H. Hashimoto, and R. N. Kelsh. 2016. What is a

vertebrate pigment cell? Pigment Cell Melanoma Res. 29:8–14.

Schwarzer, G. 2007. meta: an R package for meta-analysis. R News 7:40–45.

Seddon, R. J., and D. K. Hews. 2017. Correlates of melanization in multiple high- and low-elevation

populations of the lizard, Sceloporus occidentalis: Behavior, hormones, and parasites. J. Exp.

Zool. Part Ecol. Integr. Physiol. 327:481–492.

Senior, A. M., C. E. Grueber, T. Kamiya, M. Lagisz, K. O'Dwyer, E. S. A. Santos, and S. Nakagawa. 2016.

Heterogeneity in ecological and evolutionary meta-analyses: its magnitude and implications.

Ecology 97:3293–3299.

Siefferman, L., and G. E. Hill. 2003. Structural and melanin coloration indicate parental effort and

reproductive success in male eastern bluebirds. Behav. Ecol. 14:855–861.

Signorell, A. 2021. DescTools: Tools for Descriptive Statistics.

Silver, N. C., and W. P. Dunlap. 1987. Averaging correlation coefficients: Should Fisher's z transformation

be used? J. Appl. Psychol. 72:146–148.

Smith, M. R., and H. Wickham. 2019. TreeTools: create, modify and analyse phylogenetic trees.

Spiegelhalter, D. J., N. G. Best, B. P. Carlin, and A. Van Der Linde. 2002. Bayesian measures of model

complexity and fit. J. R. Stat. Soc. Ser. B Stat. Methodol. 64:583–639.

Tibbetts, E. A., and J. Dale. 2004. A socially enforced signal of quality in a paper wasp. Nature 432:218–

222. Nature Publishing Group.

Tuttle, E. M. 2003. Alternative reproductive strategies in the white-throated sparrow: behavioral and

genetic evidence. Behav. Ecol. 14:425–432.

Tuttle, E. M., A. O. Bergland, M. L. Korody, M. S. Brewer, D. J. Newhouse, P. Minx, M. Stager, A. Betuel, Z.

A. Cheviron, W. C. Warren, R. A. Gonser, and C. N. Balakrishnan. 2016. Divergence and

Functional Degradation of a Sex Chromosome-like Supergene. Curr. Biol. 26:344–350.

Walsh, B., and M. W. Blows. 2009. Abundant Genetic Variation + Strong Selection = Multivariate Genetic

Constraints: A Geometric View of Adaptation. Annu. Rev. Ecol. Evol. Syst. 40:41–59.

Wittkopp, P. J., and P. Beldade. 2009. Development and evolution of insect pigmentation: Genetic

mechanisms and the potential consequences of pleiotropy. Semin. Cell Dev. Biol. 20:65–71.

Random Effects Model

Actinopterygii

Horth 2003
Martin and Hengstebeck 1981 – 1
Martin and Hengstebeck 1981 – 2
Martin and Hengstebeck 1981 – 3
Martin and Hengstebeck 1981 – 4
Martin and Hengstebeck 1981 – 5
Martin and Hengstebeck 1981 – 6
Martin and Hengstebeck 1981 – 7
Martin and Hengstebeck 1981 – 8
Martin and Hengstebeck 1981 – 9
Beeching et al 1998 – 1
Beeching et al 1998 – 2
Beeching et al 1998 – 3
Beeching et al 1998 – 4
Barlow and Siri 1994
Lehtonen 2014 – 1
Lehtonen 2014 – 2
Balzarini et al 2017 – 1
Balzarini et al 2017 – 2
Renn et al 2012
Korzan et al 2008
Clement et al 2005
Dijkstra et al. 2009b
Dijkstra et al. 2009a
Rodriguez–Barreto et al 2019
Seaver and Hurd 2017 – 1
Seaver and Hurd 2017 – 2
Seaver and Hurd 2017 – 3
Seaver and Hurd 2017 – 4
Yong et al 2018 – 1
Yong et al 2018 – 2
Yong et al 2015 – 1
Yong et al 2015 – 2
Yong et al 2015 – 3
Tinghitella et al 2018 – 1
Tinghitella et al 2018 – 2
Tinghitella et al 2015 – 1
Tinghitella et al 2015 – 2
Baube 1997
Suter and Huntingford 2002
O'Connor et al. 1999

Amphibia

Reiter et al. 2014
Yang et al. 2018 – 1
Yang et al. 2018 – 2
Yang et al. 2018 – 3
Yang et al. 2018 – 4
Yang et al. 2018 – 5
Yang et al. 2018 – 6
Yang et al. 2018 – 7
Yang et al. 2018 – 8
Rudh et al. 2013
Crothers et al. 2011
Crothers and Cummings 2015

Reptilia

Deckel and Jevitts 1997 – 1
Deckel and Jevitts 1997 – 2
Deckel and Jevitts 1997 – 3
Deckel and Jevitts 1997 – 4
Andrews and Summers 1996 – 1
Andrews and Summers 1996 – 2
Bruinje et al 2019
Robertson and Rosenblum 2010
Seddon and Hews 2017
Seddon and Hews 2016 – 1
Seddon and Hews 2016 – 2
Seddon and Hews 2016 – 3
Seddon and Hews 2016 – 4
Seddon and Hews 2016 – 5
Seddon and Hews 2016 – 6
Seddon and Hews 2016 – 7
Taylor and Lattanzio 2016
McElroy et al. 2007
Brown and Lattanzio 2018
Yewers et al. 2016
Vercken and Clobert 2008 – 1
Vercken and Clobert 2008 – 2
Vercken and Clobert 2008 – 3
Titone et al 2018
Stapley 2006
Mafli et al. 2011

Aves

O'Connor et al. 2011 – 1
O'Connor et al. 2011 – 2
Nie et al. 2019 – 1
Nie et al. 2019 – 2
Karlsson et al. 2011 – 1
Karlsson et al. 2011 – 2
Karlsson et al. 2011 – 3
Karlsson et al. 2011 – 4
McDonald et al. 2001 – 1
McDonald et al. 2001 – 2
McDonald et al. 2001 – 3
McDonald et al. 2001 – 4
Huhta and Alatalo 1993 – 1
Huhta and Alatalo 1993 – 2
Huhta and Alatalo 1993 – 3
Huhta and Alatalo 1993 – 4
Huhta and Alatalo 1993 – 5
Huhta and Alatalo 1993 – 6
Dale and Slagsvold 1996
Korsten et al. 2007
Beck et al. 2015 – 1
Beck et al. 2015 – 2
Zinzow–Kramer et al. 2015 – 1
Zinzow–Kramer et al. 2015 – 2
Zinzow–Kramer et al. 2015 – 3
Zinzow–Kramer et al. 2015 – 4
Zinzow–Kramer et al. 2015 – 5
Zinzow–Kramer et al. 2015 – 6
Zinzow–Kramer et al. 2015 – 7
Zinzow–Kramer et al. 2015 – 8
Horton et al. 2014
Horton et al. 2012 – 1
Horton et al. 2012 – 2
Horton et al. 2012 – 3
Horton et al. 2012 – 4
Collins and Houtman 1999 – 1
Collins and Houtman 1999 – 2
Chaine and Lyon 2008 – 1
Chaine and Lyon 2008 – 2
Chaine and Lyon 2008 – 3
McGraw et al. 2003
Leitao et al. 2015 – 1
Leitao et al. 2015 – 2
Leitao et al. 2015 – 3
Leitao et al. 2015 – 4
McGraw et al. 2007
Jawor et al. 2004
Prkye et al. 2001 – 1
Prkye et al. 2001 – 2
Prkye et al. 2001 – 3
Prkye et al. 2001 – 4
Williams et al. 2012
Pryke and Griffith 2009
Leitao et al. 2019 – 1
Leitao et al. 2019 – 2
Leitao et al. 2019 – 3
Leitao et al. 2019 – 4
Barron et al. 2015
Rose and Soole 2020
Boerner and Kruger 2009 – 1
Boerner and Kruger 2009 – 2

Mammalia

Podberscek and Serpell 1996 – 1
Podberscek and Serpell 1996 – 2
Podberscek and Serpell 1996 – 3
Podberscek and Serpell 1996 – 4
Podberscek and Serpell 1996 – 5
Podberscek and Serpell 1996 – 6
Podberscek and Serpell 1996 – 7
Podberscek and Serpell 1996 – 8
Podberscek and Serpell 1996 – 9
Podberscek and Serpell 1996 – 10
Kim et al. 2010
Carola et al. 2014

Malacostraca

Franklin et al. 2019

Insecta

Hooper et al. 2006
Moore and Martin 2016
Petrocelli et al. 2015

Anthoza

Brace 1981 – 1
Brace 1981 – 2
Brace 1981 – 3

Colors:

Carotenoid
Eumelanin
Pheomelanin
Structural
Pteridine
Unknown

Shapes:

Plastic
Non–Plastic
Before Correction
After Correction

−4   −2   0   2   4

Fisher Z