Tmc7 deficiency causes acrosome biogenesis defects and male infertility in mice

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Abstract

Transmembrane channel-like (Tmc) proteins are a highly conserved ion channel family consisting of eight members (TMC1–TMC8) in mammals. TMC1/2 are components of the mechanotransduction channel in hair cells, and mutations of TMC1/2 cause deafness in humans and mice. However, the physiological roles of other TMC proteins remain largely unknown. Here, we show that Tmc7 is specifically expressed in the testis and that it is required for acrosome biogenesis during spermatogenesis. *Tmc7*$^{-/-}$ mice exhibited complete male infertility due to abnormal sperm morphology, similar to human oligo-astheno-teratozoospermia. We further demonstrate that Tmc7 is colocalized with Gm130 at the cis-Golgi region in round spermatids. Tmc7 deficiency leads to aberrant Golgi morphology and impaired fusion of Golgi-derived vesicles to the developing acrosome. Moreover, upon loss of Tmc7 Golgi pH and ion homeostasis is impaired and ROS levels are increased, which in turn causes Golgi and endoplasmic reticulum (ER) stress. Taken together, these results suggest that Tmc7 is required to maintain Golgi pH and ion homeostasis, which is needed for acrosome biogenesis. Our findings unveil a novel role for Tmc7 in acrosome biogenesis during spermiogenesis.

Keywords: Transmembrane channel-like (Tmc) proteins; infertility; spermiogenesis; acrosome biogenesis

Introduction

Spermiogenesis is the post-meiotic phase of male germ cell development during which the haploid spermatids differentiate into mature spermatozoa[1]. Mouse spermiogenesis consists of 16 steps characterized by morphological changes in acrosome formation, nuclear condensation and elongation, flagellum formation, and cytoplasm removal[2]. Acrosome biogenesis is one of the key events of spermiogenesis, and it begins with the initial stage of spermatid development. The acrosome is a specialized organelle with a cap-like structure covering the anterior part of the mature spermatozoon head and is indispensable for the fertilization process[3]. Although acrosome biogenesis is thought to be derived from the Golgi complex in early spermatids, growing evidence suggests that the endoplasmic reticulum (ER) and lysosomes are involved in acrosome biogenesis[4]. Defects in acrosome biogenesis can result in subfertility or infertility.

Acrosome biogenesis is generally classified into four successive phases, namely the Golgi, cap, acrosome, and maturation phases[5]. A number of ER and Golgi-associated proteins have been shown to be involved in acrosome biogenesis. Heat shock protein 90B1 (Hsp90b1) is an ER molecular chaperone that has been shown to be a testis-specific protein, and Hsp90b1 deficiency leads to a globozoospermia-syndrome like phenotype [6]. Gm130 (Golgi matrix protein 130) localize on the cis surface of the Golgi apparatus, and loss of Gm130 causes acrosome malformations and results in male infertility[7]. Gopc (Golgi-associated PDZ and coiled-coil motif-containing) is a trans-Golgi-associated protein that plays a role in vesicle transport from the Golgi apparatus. Gopc deficiency leads to vesicle transport failure, which in turn results in malformation of the acrosome and to infertility[8]. Other vesicle

transport proteins – including Pick1, Ica1l, and Vps13b – are involved in proacrosomal vesicle trafficking during acrosome formation[9,10]. Recent evidence suggests that autophagy is involved in proacrosomal vesicle fusion and transport to form the acrosome, and disruption of Atg7[11,12] or Atg5 causes aberrant acrosome formation and male infertility or subfertility. Despite these findings, further studies are needed to better understand the complexity of acrosome biogenesis.

Transmembrane channel-like (Tmc) proteins are highly conserved ion channel-like proteins, with eight family members (Tmc1–Tmc8) identified in humans and mice [13-16]. Tmc proteins are predicted to have 6–10 transmembrane domains, and all protein members contain a conserved 120 amino acid Tmc domain[15]. The single-particle cryo-electron microscopy structure shows that Tmc1 assembles as dimers and that each monomer contains 10 transmembrane domains with four domains (S4–S7) that line the channel pore[17]. Tmc1 and 2 are mechano-electrical transducer (MET) channels in cochlear hair cells, and mutations in Tmc1/2 cause deafness in humans and mice[18] by disrupting hair cell mechanoelectrical transduction, and *Tmc* gene therapy can restore auditory function and balance in mice[19,20]. Germline mutations in *Tmc6* and *Tmc8* are associated with epidermodysplasia verruciformis, which leads to increased susceptibility to HPV infection [14,15]. Among the eight Tmc protein members, Tmc1 and Tmc2 are well studied due to their role in hearing conduction, and understanding the physiological and functional roles of other Tmc family members remains a major challenge.

In this study, we found that Tmc7 begins to be expressed in diplotene spermatocytes and that it is expressed at high levels in round spermatids in mouse testes. By generating *Tmc7*

conventional knockout mice (*Tmc7*$^{-/-}$), we demonstrate that Tmc7 deficiency leads to aberrant Golgi morphology and impaired fusion of Golgi-derived vesicles to the acrosome, thus resulting in male infertility with an oligo-astheno-teratozoospermia (OAT) phenotype. Our findings provide new insights into the physiological role of Tmc family members in spermiogenesis.

Results

Tmc7 expression is confined to the testis, and deletion of Tmc7 leads to male infertility

To explore the roles of Tmc family members during spermatogenesis, we analyzed the expression of Tmc1–Tmc8 using the mouse developmental transcriptome (E-MTAB-6798)[21]. Among the eight Tmc family proteins, we found that only Tmc5 and Tmc7 exhibited elevated expression in the testis during development (Fig. 1a). Tmc7 caught our great interest because Tmc7 was expressed at higher levels than Tmc5 in the testis during development. We next analyzed Tmc7 expression in different tissues using single-cell transcriptomic data from the Mouse Cell Atlas database[22]. We found that Tmc7 was predominantly expressed in the testis when compared with all other analyzed tissues (Fig. 1b). Further, we analyzed Tmc7 in different stages of germ cell development using single-cell sequencing data of male-specific tissues and organs in mice and humans[23]. Interestingly, Tmc7 began to be expressed in diplotene spermatocytes and was expressed at high levels in round spermatids in mouse testes (Fig. 1c, d). The Tmc7 expression pattern was confirmed by RT-PCR (Fig. 1e, f), and Tmc7 was again found to be highly expressed in round spermatids of human testis (Fig. S1a, b). The distinct expression pattern in the testis indicates the potential role of Tmc7 in

spermatogenesis.

We generated *Tmc7*-deficient mice (*Tmc7*$^{-/-}$) by partial deletion of exon 2 (the first 241 bp) using the CRISPR/Cas9 system (Fig. 1g). *Tmc7*$^{-/-}$ mice were confirmed by PCR-based genotyping and immunoblotting of the separated cytoplasmic and membrane testis proteins (Fig. S1c, d). We then examined the phenotypes and found that *Tmc7*-deficient mice were viable and had a normal appearance and exhibited a normal metabolic rate (Fig. S1e-j). However, *Tmc7*-deficient male mice showed smaller testes (Fig. 1h, i) and a reduced testis/body weight ratio (Fig. 1j) compared to control mice. Moreover, *Tmc7*$^{-/-}$ male mice exhibited complete infertility (Table S1). No difference in the number of tubules per testicle or in the number of Sertoli cells was seen when comparing *Tmc7*$^{-/-}$ and WT mice; Sertoli cell were identified by immunofluorescence staining of the Sertoli cell marker Sox9 (Fig. S1k, i). These findings suggest that Tmc7 is essential for spermatogenesis and thus for male fertility.

***Tmc7* deficiency leads to severe defects in spermiogenesis**

To determine the cause of infertility in *Tmc7*$^{-/-}$ mice, we examined the composition of cell types in seminiferous tubules and cauda epididymis sections using hematoxylin staining in mouse testes at different developmental stages. In seminiferous tubules, the populations of germ cells were comparable between wild type (WT) and *Tmc7*$^{-/-}$ mice at postnatal day (PD)14 and PD21. Intriguingly, the number of elongating spermatids in *Tmc7*$^{-/-}$ mice decreased significantly compared with WT mice at PD28 (Fig. S2a). We next examined the testis and cauda epididymis sections of 9-week-old WT and *Tmc7*$^{-/-}$ mice. We observed spermiogenesis defects in *Tmc7*$^{-/-}$ mice as evidenced by the reduced number of elongating

spermatids, the increased number of sloughed germ cells, and large vacuoles in the seminiferous tubules and cauda epididymis (Fig. 2a, b). The vacuolation became more serious with increasing age (Fig. S2a). The total number of sperm in the cauda epididymis was drastically decreased in *Tmc7*$^{-/-}$ mice (~0.15 × 10^6 cells) compared to wild type (WT) mice (~8.5 × 10^6 cells) (Fig. 2c). The morphological assessment of spermatozoa from the caudal epididymis revealed that a large proportion of spermatozoa (~85%) in *Tmc7*$^{-/-}$ mice exhibited abnormal head morphology, similar to human OAT (Fig. 2d, e). In parallel with this, periodic acid Schiff (PAS) staining indicated that a large number of spermatids failed to differentiate into elongated spermatids with sickle-shaped heads and instead exhibited abnormal head morphology. The total number of spermatids began to significantly decrease at steps 9–12 of spermiogenesis (Fig. 2f, g), and we performed Mito-tracker staining and found shortened or disorganized mitochondrial sheaths in *Tmc7*$^{-/-}$ spermatozoa collected from the cauda epididymis (Fig. 2h). Meanwhile, no motile or progressive sperm were found in the *Tmc7*$^{-/-}$ mice (Fig. 2i, j). Moreover, meiotic progression was not affected in *Tmc7*$^{-/-}$ mice as evidenced by immunofluorescence staining of the meiosis markers Sycp3 (red) and γH2ax (green) in the seminiferous tubules of PD20 WT and *Tmc7*$^{-/-}$ mice (Fig. S3a). Overall, we concluded that Tmc7 ablation results in defects in spermiogenesis and in OAT-like phenotypes.

Acrosome biogenesis is impaired starting from the Golgi phase in *Tmc7*$^{-/-}$ spermatids

Acrosome biogenesis is one of the key events of spermiogenesis. To investigate whether Tmc7 is required for the proper formation of the acrosome, FITC-labelled peanut agglutinin (PNA) was used to assess the acrosomal status of spermatids in 9-week-old WT and *Tmc7*$^{-/-}$

mice. The spermatids of WT mice had normal structures in the Golgi, cap, acrosome, and maturation phases, while spermatids in *Tmc7*$^{-/-}$ mice exhibited abnormal acrosome morphology as early as the Golgi phase. Fragmented and malformed acrosomes were observed in all four phases of spermatids in the *Tmc7*$^{-/-}$ mice (Fig. 3a). Absent or fragmented acrosomes were also found in *Tmc7*$^{-/-}$ spermatids collected from the cauda epididymis (Fig. S4a). To better understand the morphologic defects of the acrosome in greater detail, transmission electron microscopy (TEM) was performed to analyze the acrosome structure in WT and *Tmc7*$^{-/-}$ mice. We found that large vesicles accumulated adjacent to developing acrosomes in all phases of acrosome biogenesis, starting from the Golgi phase (Fig. 3b, left panel) in the spermatids of *Tmc7*$^{-/-}$ mice. Additionally, abnormal acrosome structures and defective nuclear condensation were observed in *Tmc7*$^{-/-}$ spermatids (Fig. 3b, right panels). Further investigation revealed that knockout of *Tmc7* resulted in Golgi apparatus fragmentation and disorganization, as determined by TEM and immunofluorescence staining of the Golgi marker Gm130 (Fig. 3b-c). The aspect ratio (height over width) of the Golgi apparatuses in *Tmc7*$^{-/-}$ spermatids (R = ~2.9) based on TEM images were also significantly smaller than in WT spermatids (R = ~4.9) (Fig. 3d). Meanwhile, we found that the Golgi body failed to correctly orient toward the acrosome (Fig. S4b). We further performed immunoblotting to measure Golgi stress markers, Hsp47 and Tfe3 in 9-week-old *Tmc7*$^{-/-}$ testes compared with WT testes. The results showed that the protein levels of Hsp47 and Tfe3 were significantly increased upon loss of Tmc7, indicating the occurrence of Golgi stress (Fig. 3e). Gopc is localized in the trans-Golgi network and is involved in the transport and fusion of the proacrosomal vesicles with the acrosome[8]. By immunostaining for Gopc and PNA, we

found that Gopc staining (red) was colocalized with PNA (green) in WT spermatids. In contrast, colocalized Gopc and PNA spermatids were significantly decreased in *Tmc7*$^{-/-}$ mice (Fig. 3f). Together these findings suggest that Tmc7 is required for proacrosomal vesicle trafficking and fusion during acrosome biogenesis.

Tmc7 localizes to the cis-Golgi, and this is required for maintaining Golgi integrity

To further delineate the role of Tmc7 in acrosome formation, we next sought to determine the subcellular localization of Tmc7 in the testis. We performed immunostaining and found that Tmc7 was predominantly expressed in the perinuclear regions of pachytene spermatocytes and round spermatids in WT testes but not in *Tmc7*$^{-/-}$ testes, suggesting the specificity of the antibody (Fig. 4a). By immunofluorescence co-staining of Tmc7 with the cis-Golgi marker Gm130, the trans-Golgi marker Tgn46, and PNA, we found that Tmc7 colocalized closely with Gm130 but not with Tgn46 (Fig. 4b, c, Fig. S5a), indicating its localization to the cis-Golgi apparatus. Further evaluation showed that Tmc7 and PNA were closely associated but not colocalized (Fig. 4d, Fig. S5b). In parallel, we carried out co-immunoprecipitation with Tmc7 antibody coupled with immunoblotting in mouse testes and found that Tmc7 was able to pull down Gm130, the Gm130-interacting protein P115, and Grasp65 (Fig. 4e), further confirming that Tmc7 is localized to the cis-Golgi apparatus.

Gm130 is reported to be involved in maintaining the function and integrity of the Golgi apparatus[7]. To gain more insight into the role of Tmc7 in the Golgi apparatus, we performed immunostaining against Gm130, P115, and Grasp65 in the testes of 9-week-old WT and *Tmc7*$^{-/-}$ mice. The results showed that the protein levels of Gm130, P115, and Grasp65 were

significantly reduced in *Tmc7*$^{-/-}$ testes compared to WT testes (Fig. 4f, g, Fig. S5c-e). Immunoblotting further verified the reduced levels of Gm130, P115, and Grasp65 upon loss of Tmc7 (Fig. 4h). These results suggest that Tmc7 colocalizes to Gm30 in the cis-Golgi and that this is required for maintaining Golgi integrity.

Tmc7 depletion impairs cellular homeostasis and leads to spermatid apoptosis

Golgi pH and ion homeostasis is essential for proper function, including membrane trafficking, protein glycosylation, and protein sorting[24], and the Golgi apparatus has been shown to be another intracellular Ca^{2+} store in addition to the endoplasmic reticulum (ER)[25]. *Tmc1* mutation leads to reduced Ca^{2+} permeability, thus triggering hair cell apoptosis and subsequent deafness[26]. We measured intracellular Ca^{2+} by flow cytometry in WT and *Tmc7*$^{-/-}$ germ cells isolated from PD30 testes. The results showed that the intracellular Ca^{2+} concentration was significantly reduced in *Tmc7*$^{-/-}$ germ cells compared to WT germ cells (Fig. 5a). In contrast, intracellular pH was increased in *Tmc7*$^{-/-}$ germ cells compared to WT germ cells (Fig. 5b). These results suggest that the pH and ion homeostasis was impaired. We further measured the reactive oxygen species (ROS) levels and observed that ROS levels were significantly increased upon loss of Tmc7 (Fig. 5c) In addition, we examined ER stress-related proteins by immunoblotting and found that p-eif2α, Chop, and Grp78 were significantly increased in 9-week-old *Tmc7*$^{-/-}$ testes compared with WT testes, indicating the occurrence of ER stress (Fig. 5d). Moreover, we carried out TUNEL staining in the seminiferous tubules and cauda epididymis of 9-week-old WT and *Tmc7*$^{-/-}$ mice. The TUNEL-positive cells in seminiferous tubules and cauda epididymis were significantly

increased in *Tmc7*$^{-/-}$ mice compare to WT mice (Fig. 5e). The expression of apoptosis-related proteins was next measured by immunoblotting, and depletion of Tmc7 increased the levels of cleaved caspase-3 and Bax and decreased the levels of Bcl2 (Fig. 5f). These results support the hypothesis that Tmc7 is required to maintain Golgi pH and ion homeostasis and that this is needed for acrosome biogenesis.

Discussion

Among the eight Tmc protein members, Tmc1 and Tmc2 are localized to the site of MET channels at the tips of the shorter stereocilia of cochlear hair cells[27,28], and mutations in *Tmc1* and *Tmc2* cause deafness. However, Tmc1 and Tmc2 expressed in heterologous cell lines remain in the ER and fail to localize to the plasma membrane[18]. Tmc4 is a voltage-dependent chloride channel that is predominantly expressed in the taste buds in the posterior region of the tongue[29]. Tmc6 and Tmc8 are localized to the ER and play crucial roles in zinc transport[30]. However, the physiological function of Tmc7 has not been defined. In this study, we demonstrated that Tmc7 was predominantly expressed in testes (Fig. 1e), and by generating knockout mice lacking *Tmc7* we found that Tmc7 deficiency resulted in complete male infertility with an OAT-like phenotype. Interestingly, we found that Tmc7 co-localized with Gm130, but not with Tgn46 (Fig. 4b, c), as illustrated by the immunofluorescence co-staining assay, indicating that Tmc7 located on the cis side of the Golgi apparatus.

Our results demonstrate the critical role of Tmc7 during the formation of the acrosome. The acrosome is thought to be derived from the Golgi apparatus, so we speculated that Tmc7 deficiency might cause defects in acrosome formation because Tmc7 localized on the Golgi

apparatus. We observed fragmented and malformed acrosomes starting from the Golgi phase in *Tmc7*$^{-/-}$ spermatids as analyzed by TEM (Fig. 3b). Additionally, proacrosomal vesicles were found to accumulate around the acrosomes (Fig. 3b), and spermatids with colocalized Gopc and PNA were significantly decreased in *Tmc7*$^{-/-}$ mice (Fig. 3f), suggesting that Golgi-derived vesicles failed to fuse with developing acrosomes. In accordance with our findings, other proteins that localize on the Golgi apparatus have also been reported to be involved in acrosome biogenesis. Gm130 deficiency results in acrosome biogenesis failure and male infertility because proacrosomal vesicles are unable to fuse to form acrosomes[30]. Our findings showed that Tmc7 was co-localized with Gm130 and that Tmc7 depletion significantly reduced the protein level of Gm130. Gopc is localized on the trans-Golgi region, and its deficiency leads to defective fusion of Golgi-derived transport vesicles to the acrosomal cap and to a complete lack of acrosomes[8]. Colocalized Gopc and PNA spermatids were significantly decreased in *Tmc7*$^{-/-}$ spermatids compared to WT spermatids. Further, Golga3, Pick1, and several other Golgi proteins are also required for acrosome biogenesis[31]. Our work, combined with previous studies, broadens our understanding of role the Golgi apparatus plays in the formation of acrosomes.

Our findings highlight the role of Tmc7 in maintaining Golgi and cellular homeostasis. Upon Tmc7 depletion, the Golgi apparatus undergoes significant disorganization and is unable to correctly orient toward the acrosome. The mechanism behind this is that intracellular Ca^{2+} concentrations decreased while the pH value was increased in *Tmc7*$^{-/-}$ germ cells compared to WT germ cells, suggesting that Tmc7 is essential for maintaining the pH and ion homeostasis of the Golgi apparatus. Our findings further revealed that Tmc7

deficiency led to increased ROS levels, Golgi and ER stress, ultimately resulting in apoptosis of spermatids. The Na^+/H^+ exchanger Nhe8 is involved in the maintenance of intracellular pH and ion homeostasis, and Nhe8 deficiency leads to Golgi-derived vesicles failing to form large acrosomal granules and the acrosomal cap[32]. Slc9a3 is another Na^+/H^+ exchanger, and it is expressed in the acrosomal region of spermatids and loss of Slc9a3 causes severe acrosomal defects[33]. These studies, together with our current findings, support the hypothesis that pH and ion homeostasis of the Golgi apparatus is needed for the proper progression of acrosome biogenesis.

In conclusion, our findings show that Tmc7 localizes to the cis-Golgi region in spermatids and that this is essential for acrosome biogenesis and for male fertility. In the absence of Tmc7, Golgi pH and ion homeostasis is disrupted, and this leads to Golgi disorganization and failure to correctly orientate to the acrosome. Subsequently, Golgi-derived proacrosomal vesicles are unable to fuse with the developing acrosome and acrosome biogenesis is severely impaired, and thus Tmc7-deficent male mice are completely infertile. Our study highlights the physiological role of TMC family members in spermatogenesis.

Materials and methods

Generation of *Tmc7* knockout mice

Tmc7 conventional knockout mice in the C57BL/6J background were obtained from Cyagen Bioscience (China). Exon 2 was selected as the target site, and Cas9 mRNA and single guide RNA were co-injected into fertilized eggs for knockout mouse production. The pups were genotyped by PCR using the following primers: F1:5’-AGA TGG CTT AGT GGG TGC CTT

AGT-3'; R1:5'TGA TGG ACA GAG AGA TGA TGA ATG G-3'; F3: 5'-CAG TCC CAG GTT CAG TTC AGT GAG-3'. In WT mice, the primer pair amplified a 513 bp PCR product, while in *Tmc7*$^{-/-}$ mice it amplified an 891 bp product. Mice were housed under a controlled illumination regime (light for 12 h) at 21°C–22°C and 60% ± 10% relative humidity. Food and water were freely available, and all animal experiments were conducted in accordance with the guidelines of the Animal Care and Use Committee at Shandong University (ECSBMSSDU2022-2-113).

Metabolic assessment in mice

A Columbus Instruments Comprehensive Lab Animal Monitoring System CLAMS6 was used for the experiment. Nine-week-old WT and *Tmc7*$^{-/-}$ male mice (n = 4) were weighed and had the metabolic rates measured over a period of 48 hours. The rate of oxygen consumption, RER (breathing entropy), energy expenditure, and food intake were automatically recorded every eight minutes. These data were analyzed with a 12 h dark:12 h light cycle at 22–23 °C.

RNA isolation and PCR

The tissues were homogenized in TRIzol (Invitrogen, 15596-026) with a grinder. Following centrifugation ,the RNA in the supernatant was obtained by precipitating with isopropanol and washing with ethanol. A total of 1 μg RNA was reverse-transcribed into cDNA with HiScript III RT SuperMix for RT-PCR (+gDNA wiper) (Vazyme, R223-01) according to the manufacturer's instructions. The cDNA was used to detect the expression of Tmc7 with Taq Master Mix (Vazyme, P112-03) with the following primers: F – AGC CAA TCC ATC CGC AAG TAT; R – TGC TGC TG TAC CGT TTC CAC. The products were analyzed by agarose gel electrophoresis.

Sperm collection

The cauda epididymis was collected and placed in an EP tube containing PBS. Sperm were released for 30 min in an atmosphere of 5% CO_2 at 37 °C. The suspension was diluted at 1:50 and transferred to a hemocytometer for counting. For the anther experiments, the PBS with sperm was spread on the slide and fixed with 4% paraformaldehyde (Servicebio, G1101). The slides were stored at –80°C after drying.

Histological analysis

For the morphology analysis of the sperm from the caudal epididymis, sperm smears were directly stained with hematoxylin and sealed with neutral balsam. The tissue was fixed in 4% paraformaldehyde overnight, then dehydrated and embedded in paraffin. Paraffin-embedded tissues were sectioned at 4 μm. Tissue slides were deparaffinized in xylene and dehydrated in a series of ethanol. The tissues were then stained by hematoxylin or with periodic acid staining using a PAS Stain Kit (Abcam, ab150680) according to the manufacturer's instructions. Finally, tissue slides were rehydrated in a series of ethanol and xylene and sealed with neutral balsam. An Olympus BX53 microscope system was used for imaging.

Immunofluorescence

We performed immunofluorescence of germ cells in the testes using two methods, including testis slides and squashed seminiferous tubules. The squashed seminiferous tubules were prepared according to a previous report[34]. The testis slides were deparaffinized in xylene and rehydrated in a series of ethanol. Antigen retrieval was performed by heating in EDTA Antigen Retrieval Solution (pH 9.0) (ZSGB-BIO, ZLI-9096), and nonspecific binding sites were blocked with EZ-Buffers M 1X Block BSA in PBS (Sangon, C520035-0250). The slides

were incubated with primary antibody overnight at 4℃. Primary antibodies included anti-Tmc7 (1:200 dilution, Abcam, ab191521), anti-Sox9 (1:500 dilution, Sigma, AB5535), anti-Gopc (1:200 dilution, Proteintech, 12163-1-AP), anti-Sycp3 (1:500 dilution, Santa Cruz, sc-33195), anti-H2ax (1:500 dilution, Millipore, 05-636-1), anti-Gm130 (1:100 dilution, 610822, BD Biosciences), anti-P115 (1:200 dilution, Proteintech, 13509-1-AP), and anti-Grasp65 (1:200 dilution, Proteintech, 10747-2-AP). The next day they were incubated with the secondary antibody (1:500 dilution, Thermo, A-11070 and A-11012) for 1 h at room temperature. The Alexa Fluor 488 conjugates of lectin PNA (1:500 dilution, Thermo, L21409), anti-Gm130 (1:200 dilution, Proteintech, CL488-11308), and anti-Tgn46 (1:200 dilution, Proteintech, CL488-13573) were used as secondary antibodies. For the staining of Mito-track (1:5000 dilution, Beyotime, C1048), the sperm released from the caudal epididymis were directly stained for 30 min at 37 °C. The sperm were then spread on the slide and observed. A TUNEL assay was performed using a one-step TUNEL Apoptosis Assay Kit (KeyGEN BioTECH, KGA7072) according to the manufacturer's instructions. Briefly, the slide was deparaffinized and antigen retrieval was performed. dUTP was used to label the cells, and the nuclei were stained with DAPI (Abcam, ab285390). Fluorescent images were captured on a Dragonfly 200 confocal microscopy system (Andor Technology).

Western blotting

The membrane and cytoplasmic proteins of the testis were obtained using a membrane protein extraction kit (Abmart, A1008) according to the manufacturer's instructions. Total protein in the testis was obtained by grinding in a tissue grinder in RIPA lysis buffer (Beyotime, P0013B), and the concentration of protein was measured using a BCA protein assay kit

(Beyotime, P0009). The proteins were separated by SDS-PAGE and then transferred to a PVDF membrane. The membranes were blocked in 5 % non-fat milk and then incubated with primary antibodies overnight at 4 °C. Primary antibodies included anti-Tmc7 (Dia-an Biotech), anti-Atp1a1 (Proteintech, 14418-1-AP), anti-Gapdh (Proteintech, 60004-1-1g), anti-Cleaved-Caspase3 (Cell Signaling Technology, 9661), anti-Caspase3 (Cell Signaling Technology, 9662), anti-Bax (Cell Signaling Technology, 41162), anti-Bcl2 (Cell Signaling Technology, 17447), anti-α-Tubulin (Proteintech,11224-1-AP), anti-P-Eif2α (ABclonal, AP0745), anti-Eif2α (ABclonal, A0764), anti-Grp78 (Proteintech, 11587-1-AP), anti-Chop (Proteintech, 15204-1-AP), anti-P115 (Proteintech, 13509-1-AP), and anti-Grasp65 (Proteintech, 10747-2-AP). All antibodies were diluted 1:1000. The next day, the membrane was incubated with horseradish peroxidase-conjugated secondary antibody for 1 hour, and the protein was detected using an ECL system (Vazyme, E412-02). The quantitation was performed in Image J.

Chromosome spreading

Meiotic nuclear spreading was performed as previously reported[35]. Briefly, spermatocytes released from seminiferous tubules were incubated in hypotonic solution for 1 hour and then transferred to 100 mM sucrose. The suspension was spread on the glass slides with 1% PFA and 0.15% Triton X100, and dried slides were stored at –80°C or were used for the immunofluorescence staining.

Transmission electron microscopy

The testes of 9-week-old WT and *Tmc7*$^{-/-}$ mice (n = 3) were used for the TEM analysis. The testes were washed in PBS three times and then fixed in 1% O_SO_4 in 0.1 M PBS (pH 7.4)

375 buffer at 4°C overnight. The testes were then fixed in 1% O_SO_4 at room temperature for 1
376 hour. The samples were then washed with water three times for 15 min each and dehydrated
377 in a graded ethanol series (50%, 70%, 95%, and 100%). The samples were finally embedded
378 with resin, cut into 80 nm sections, and viewed with a JEM-1400 transmission electron
379 microscope.

380 **Flow cytometry**

381 The testes of PD30 WT and *Tmc7*$^{-/-}$ mice (n = 3) were used to obtain germ cells. The tunica
382 albuginea was removed and placed into PBS with 120 U/ml Type I Collagenase (Gibco,
383 17100-017) on a shaker at 37°C and 220 rpm for 10 min. The supernatant was filtered through
384 a 70 μm cell filter screen into a new tube. The sediment was digested again with 0.25%
385 trypsin-EDTA (Gibco, 25200–072) on a shaker at 37°C and 220 rpm for 10 min and then
386 neutralized by serum albumin and filtered. The germ cells were obtained by centrifuging
387 twice at 1000 × g for 5 min and then removing the supernatant.

388 The levels of Ca^{2+}, pH, and ROS were measured using fluorescent probes according to the
389 manufacturer's protocols (Beyotime, S1056, S1006, S0033S). The probes were all diluted
390 1:2000. Germ cells were stained with the probes at 37°C for 30 min, then washed with PBS
391 three times and analyzed by flow cytometry (Gallios, US).

392 **Co-immunoprecipitation**

393 Testes of 9-week-old WT mice were ground in a tissue grinder in RIPA Lysis Buffer
394 (Beyotime, P0013B) to obtain the total protein. Tmc7 (Dia-an Biotech) and IgG (Beyotime,
395 A7016) antibodies were incubated with the proteins overnight. The next day, they were
396 incubated with protein A/G magnetic beads (MCE, HY-K0202) for 2 hours. The beads were

wash and boiled for 10 min in 1× SDS loading buffer. Western blotting was used for the analysis of input and immunoprecipitated samples with anti-P115 and anti-Grasp65 antibodies.

Statistical analysis

GraphPad Prism 9 was used to analyze the data, and the results are shown as the mean ± standard deviation (S.D.). Statistical significance was determined using two-sided Student's t-tests or two-way analysis of variance (ANOVA). *$p < 0.05$, **$p < 0.01$, and ***$p < 0.001$ indicate the level of statistical significance, and ns stands for not significant.

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Author Contributions

J.W., Y.Y, performed most experiments, and data analysis, and writing the manuscript. Z.W., L.Y., performed public database sequence data analysis. C.Q., Y.G., L.Y., helped with animal and cell experiment. F.G., G.Lu., Z.C., X.Z., is involved technical, material support and guided the experiment. H.L., Z.L. Supervised the project and revised the manuscript. Zhaojian Liu was the lead contact for the study. Final approval: All authors.

Conflict of interest

The authors declare no competing interests.

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Fig 1. Tmc7 expression is confined to the testis, and its deletion leads to male infertility. (a) Heatmap analysis of the expression of Tmc family proteins at embryonic day (E)10–E18 and different postnatal days (P0, P3, PD14, PD28, and PD63) in mouse testis using the Evo-devo mammalian organ database (http://apps.Kaessmannlab.org/evodevoapp/). (b) The analysis of Tmc7 expression in different tissues using the Mouse Cell Atlas database (https://bis.zju.edu.cn/MCA/). (c, d) T-Distributed Stochastic Neighbor Embedding (t-SNE) plot and violin plot showing the expression of Tmc7 in different mouse germ cells using the Male Health Atlas (http://malehealthatlas.cn/). SSC1-3: Spermatogonial stem cells I-III; Progenitor: spermatogonia progenitor cell; A1–4: A1–4 spermatogonia; In: Intermediate spermatogonia; B: Type B spermatogonia; PI: Preleptotene; L: Leptotene; Z: Zygotene; P: pachytene; D: Diplotene; M: Metaphase; RS: Round spermatid. E: Elongating spermatid. (e) The expression of *Tmc7* in different adult mouse tissues was measured by RT-PCR, including the heart, liver, spleen, lung, kidney, brain, ovary, and testis. *Gapdh* was used as the control. (f) The expression of *Tmc7* at different postnatal times (PD1, PD7, PD9, PD12, PD24, PD33, and 2 months) was measured by RT-PCR. *Gapdh* was used as the control. (g) Schematic

illustrating the generation of *Tmc7*$^{-/-}$ mice using CRISPR/Cas9. (h) Testes from 9-week-old WT and *Tmc7*$^{-/-}$ mice. (i) Testicular weights of 9-week-old WT and *Tmc7*$^{-/-}$ mice. Four adult mice of each genotype were used. (j) Testis/body weight ratio of 9-week-old WT and *Tmc7*$^{-/-}$ mice. Four mice of each genotype were used. Images are representative of at least three independent experiments. The results are presented as the mean ± S.D., and two-sided unpaired Student's t-tests were used to calculate the P-values (*P < 0.05, **P < 0.01, ***P < 0.001).

Fig 2. Tmc7 deficiency leads to severe defects in spermiogenesis. (a, b) Hematoxylin staining in testis and caudal epididymis sections of 9-week-old WT and *Tmc7*$^{-/-}$ male mice. Scale bar, 50 μm. (c) The total number of sperm in the caudal epididymis from 9-week-old WT and *Tmc7*$^{-/-}$ male mice. Three mice were used. (d) Hematoxylin-eosin staining of the spermatozoa isolated from the caudal epididymis in 9-week-old WT and *Tmc7*$^{-/-}$ male mice. H: head, F: flagella. Scale bar, 10 μm, 2 μm. (e) The percentage of spermatozoa with abnormal heads in the caudal epididymis from WT and *Tmc7*$^{-/-}$ male mice. Three mice were used. (f) PAS and hematoxylin staining in 9-week-old WT and *Tmc7*$^{-/-}$ mouse seminiferous tubules. The numbers represent the stages of spermatogenesis. P: pachytene spermatocyte, PL: preleptotene spermatocyte, L: leptotene spermatocyte, Z: zygotene spermatocyte, D: diplotene spermatocyte, RS: round spermatid, ES: elongating spermatid. Scale bar, 5 μm. (g) Quantification of spermatid numbers in seminiferous tubules from spermatid stages 1–16. N = 15 tubules per stage and per genotype; 3 mice per genotype were used and 5 tubules per stage and per mouse were counted. (h) Mitochondrial sheath staining by Mito-tracker in sperm collected from the caudal epididymis. Scale bar, 5 μm. (i, j) The percent of progressive and

motile spermatozoa in 9-week-old WT and *Tmc7*$^{-/-}$ mice. Three mice were used. Images are representative of at least three independent experiments. The results are presented as the mean ± S.D., and two-sided unpaired Student's t-tests were used to calculate the P-values. (*P < 0.05, **P < 0.01, ***P < 0.001).

Figure 3. Acrosome biogenesis is impaired starting from the Golgi phase in *Tmc7*$^{-/-}$ spermatids. (a) Immunofluorescence staining of PNA (green) in the phases of acrosome biogenesis in 9-week-old WT and *Tmc7*$^{-/-}$spermatids. Nuclei were stained with DAPI (blue). The phases of acrosome biogenesis and corresponding spermiogenesis steps are the Golgi phase (steps 1–3), cap phase (steps 4–7), acrosome phase (steps 8–12), and maturation phase (steps 13–16). (b) TEM images of testicular sections of 9-week-old WT and *Tmc7*$^{-/-}$ mice. The red star represents the abnormal plasma membrane space and accumulated vesicles. N: nucleus, Ac: acrosome; Apx: acroplaxome; pm: plasma membrane. G: Golgi complex; Scale bar, 2 μm. (c) Immunofluorescence staining with the antibody against Gm130 in spermatids from the testes of 9-week-old WT and *Tmc7*$^{-/-}$ mice. Scale bar, 2 μm. (d) Calculation of the aspect ratio (height [X] over width [Y]) of 9-week-old WT and *Tmc7*$^{-/-}$ Golgi apparatuses based on TEM images. N = 27, and 9 tubules in each testis were counted from 3 mice. (e) Western blot analysis of protein levels of Golgi stress- associated proteins (Hsp47, Tfe3) in 9-week-old WT and *Tmc7*$^{-/-}$ testes (*n* = 3 per group). Image J was used to quantify the protein level. (f) Immunofluorescence staining of Gopc (red) and PNA (green) and the quantitative analysis of Gopc and PNA lectin colocalization in squashed tubules of 9-week-old WT and *Tmc7*$^{-/-}$ mice. Nuclei were stained with DAPI (blue). Scale bar, 10 μm. Four mice were used, and 200 cells were counted per mouse. Images are representative of at least three independent

experiments. The results are presented as the mean ± S.D., and two-sided unpaired Student's t-tests were used to calculate the P-values. (*P < 0.05, **P < 0.01, ***P < 0.001).

Figure 4. Tmc7 localizes to the cis-Golgi, and this is required for maintaining Golgi integrity. (a) Immunohistochemical staining with an antibody against Tmc7 in seminiferous tubules of 9-week-old mouse testes. Scale bar, 50 μm. (b) Immunofluorescence staining of Tmc7 (red) and Gm130 (green, a marker of the cis-Golgi), with DAPI indicating the nuclei in seminiferous tubules of 9-week-old WT and *Tmc7*$^{-/-}$ testis. Scale bar, 10 μm. Co-localization was quantified by Image J, and 20 cells were analyzed for the Pearson's R-value. (c) Immunofluorescence staining of Tmc7 (red) and Tgn46 (green, a marker of the trans-Golgi), with DAPI indicating the nuclei in the seminiferous tubule. Scale bar, 10 μm. (d) Immunofluorescence staining of Tmc7 (red) and PNA (green, a marker of the acrosome), with DAPI indicating the nuclei in the seminiferous tubule. Scale bar, 10 μm. (e) Western blot was used to analyze the results of co-immunoprecipitation and showed the interaction between Tmc7 and Gm130, P115, and Grasp65 in 9-week-old WT testes. (f, g) Immunofluorescence staining and quantification of Gm130 (green), P115 (red) and Grasp65 (red) in testicular sections of 9-week-old WT and *Tmc7*$^{-/-}$ mice. Scale bar, 50 μm. (h) Western blots of the protein levels of Gm130 and the Gm130-interacting proteins P115 and Grasp65 in 9-week-old WT and *Tmc7*$^{-/-}$ testes. Image J was used to quantify the protein level. Images are representative of at least three independent experiments. The results are presented as the mean ± S.D., and two-sided unpaired Student's t-tests were used to calculate the P-values. (*P < 0.05, **P < 0.01, ***P < 0.001).

Fig 5. Tmc7 depletion impairs cellular homeostasis and leads to spermatid apoptosis.

(a) The Ca^{2+} levels were measured by flow cytometry in WT and *Tmc7*$^{-/-}$ germ cells isolated from PD30 testes. (b) The pH levels were measured by flow cytometry in WT and *Tmc7*$^{-/-}$ germ cells isolated from PD30 testes. (c) The ROS levels were measured by flow cytometry in WT and *Tmc7*$^{-/-}$ germ cells isolated from PD30 testes. (d) Western blot analysis of protein levels of ER chaperone-associated proteins (P-Eif2α, Eif2α, Chop, and Grp78) in 9-week-old WT and *Tmc7*$^{-/-}$ testes. Image J was used to quantify the protein level. Each value was detected in three mice of each group. (e) TUNEL assay and the number of TUNEL-positive cells per tubule in seminiferous tubules and cauda epididymis of 9-week-old WT and *Tmc7*$^{-/-}$ male mice. Scale bar, 50 μm. N = 60, 20 tubules in each testis and cauda epididymis were counted, and three mice were used. (f) Apoptosis-related proteins (Cleaved-caspase3, Caspase3, Bax, Bcl2) were detected by western blotting in 9-week-old WT and *Tmc7*$^{-/-}$ testes. Image J was used to quantify the protein level. Images are representative of at least three independent experiments. The results are presented as the mean ± S.D., and two-sided unpaired Student's t-tests were used to calculate the P-values. (*P < 0.05, **P < 0.01, ***P < 0.001).

Fig. 1

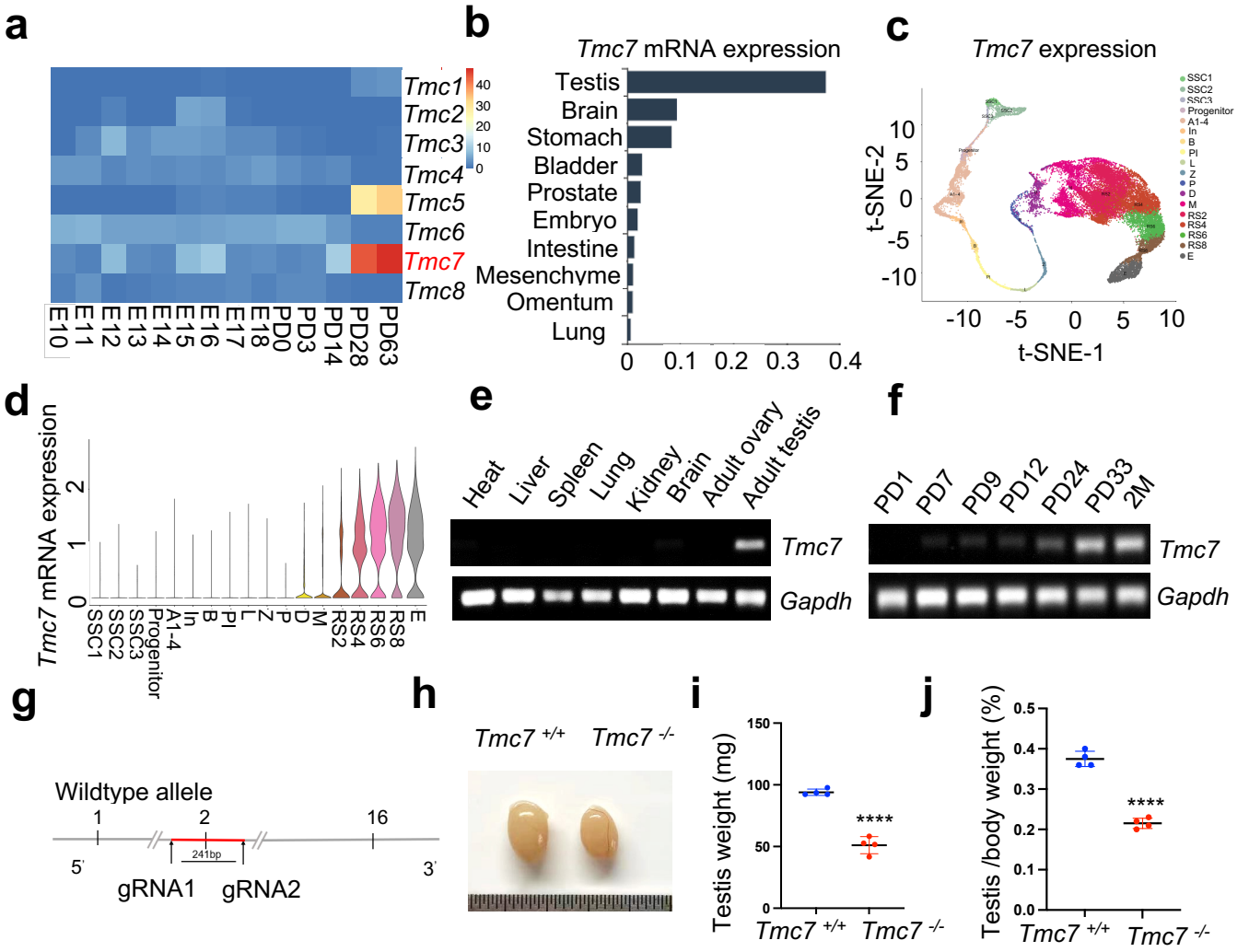

a
Tmc1
Tmc2
Tmc3
Tmc4
Tmc5
Tmc6
Tmc7
Tmc8
40
30
20
10
0
E10
E11
E12
E13
E14
E15
E16
E17
E18
PD0
PD3
PD14
PD28
PD63
b
Tmc7 mRNA expression
Testis
Brain
Stomach
Bladder
Prostate
Embryo
Intestine
Mesenchyme
Omentum
Lung
0
0.1
0.2
0.3
0.4
c
Tmc7 expression
t-SNE-2
t-SNE-1
10
5
0
-5
-10
SSC1
SSC2
SSC3
Progenitor
A1-4
In
B
Pl
L
Z
P
D
M
RS2
RS4
RS6
RS8
E
d
Tmc7 mRNA expression
0
1
2
SSC1
SSC2
SSC3
Progenitor
A1-4
In
B
Pl
L
Z
P
D
M
RS2
RS4
RS6
RS8
E
e
Heat
Liver
Spleen
Lung
Kidney
Brain
Adult ovary
Adult testis
Tmc7
Gapdh
f
PD1
PD7
PD9
PD12
PD24
PD33
2M
Tmc7
Gapdh
g
Wildtype allele
1
2
16
5'
3'
241bp
gRNA1
gRNA2
h
Tmc7 +/+
Tmc7 -/-
i
Testis weight (mg)
150
100
50
0
****
Tmc7 +/+
Tmc7 -/-
j
Testis /body weight (%)
0.5
0.4
0.3
0.2
0.1
0.0
****
Tmc7 +/+
Tmc7 -/-

Fig. 2

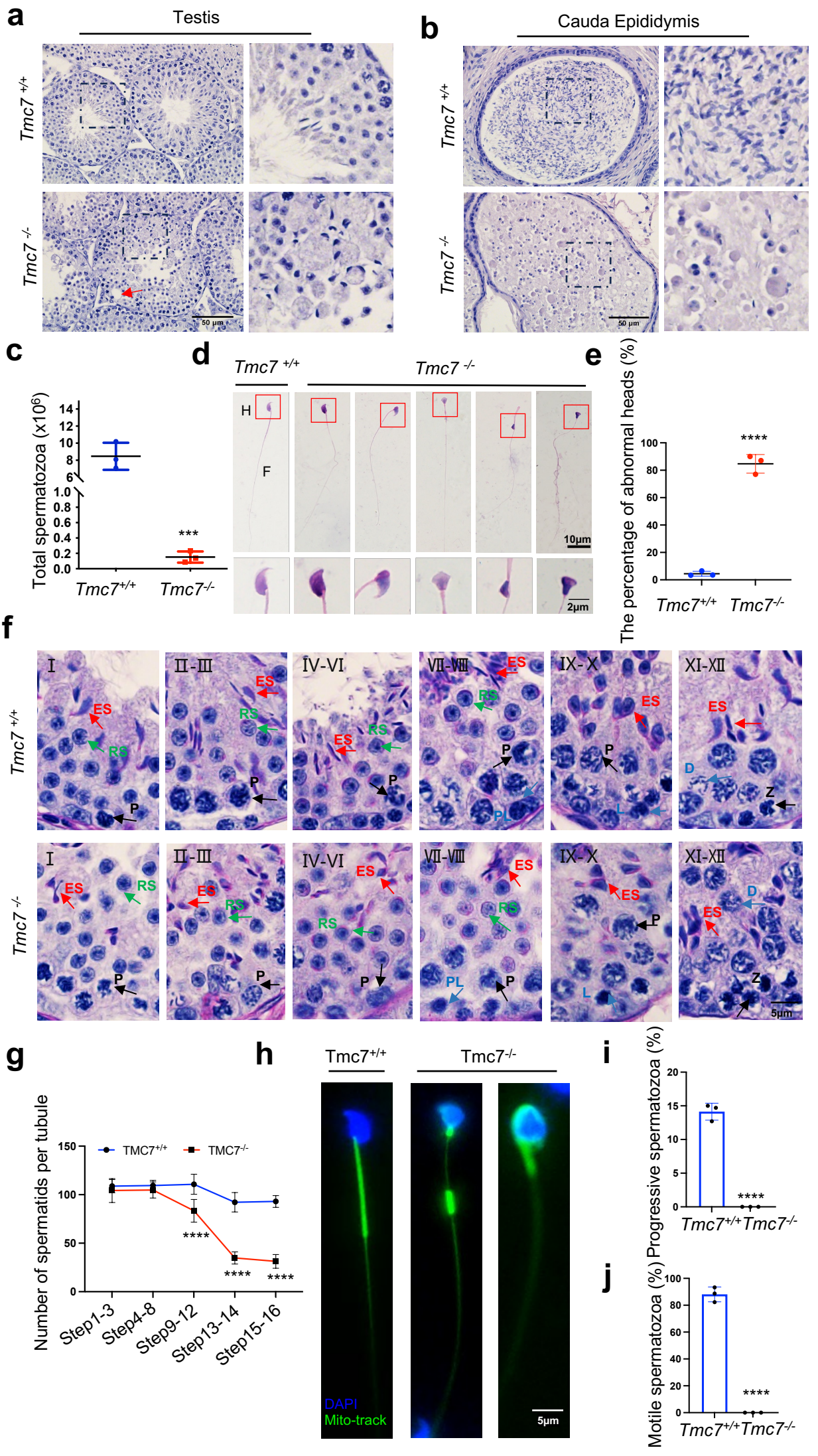

a
Testis
Tmc7 +/+
Tmc7 -/-
50 μm
b
Cauda Epididymis
Tmc7 +/+
Tmc7 -/-
50 μm
c
Total spermatozoa (x10^6)
14
12
10
8
6
1.0
0.8
0.6
0.4
0.2
0.0
***
Tmc7+/+
Tmc7-/-
d
Tmc7 +/+
Tmc7 -/-
H
F
10μm
2μm
e
The percentage of abnormal heads (%)
100
80
60
40
20
0
****
Tmc7+/+
Tmc7-/-
f
Tmc7 +/+
Tmc7 -/-
I
II-III
IV-VI
VII-VIII
IX-X
XI-XII
ES
RS
P
PL
L
D
Z
5μm
g
Number of spermatids per tubule
150
100
50
0
TMC7+/+
TMC7-/-
****
****
****
Step1-3
Step4-8
Step9-12
Step13-14
Step15-16
h
Tmc7+/+
Tmc7-/-
DAPI
Mito-track
5μm
i
Progressive spermatozoa (%)
20
15
10
5
0
****
Tmc7+/+
Tmc7-/-
j
Motile spermatozoa (%)
100
80
60
40
20
0
****
Tmc7+/+
Tmc7-/-

Fig. 3

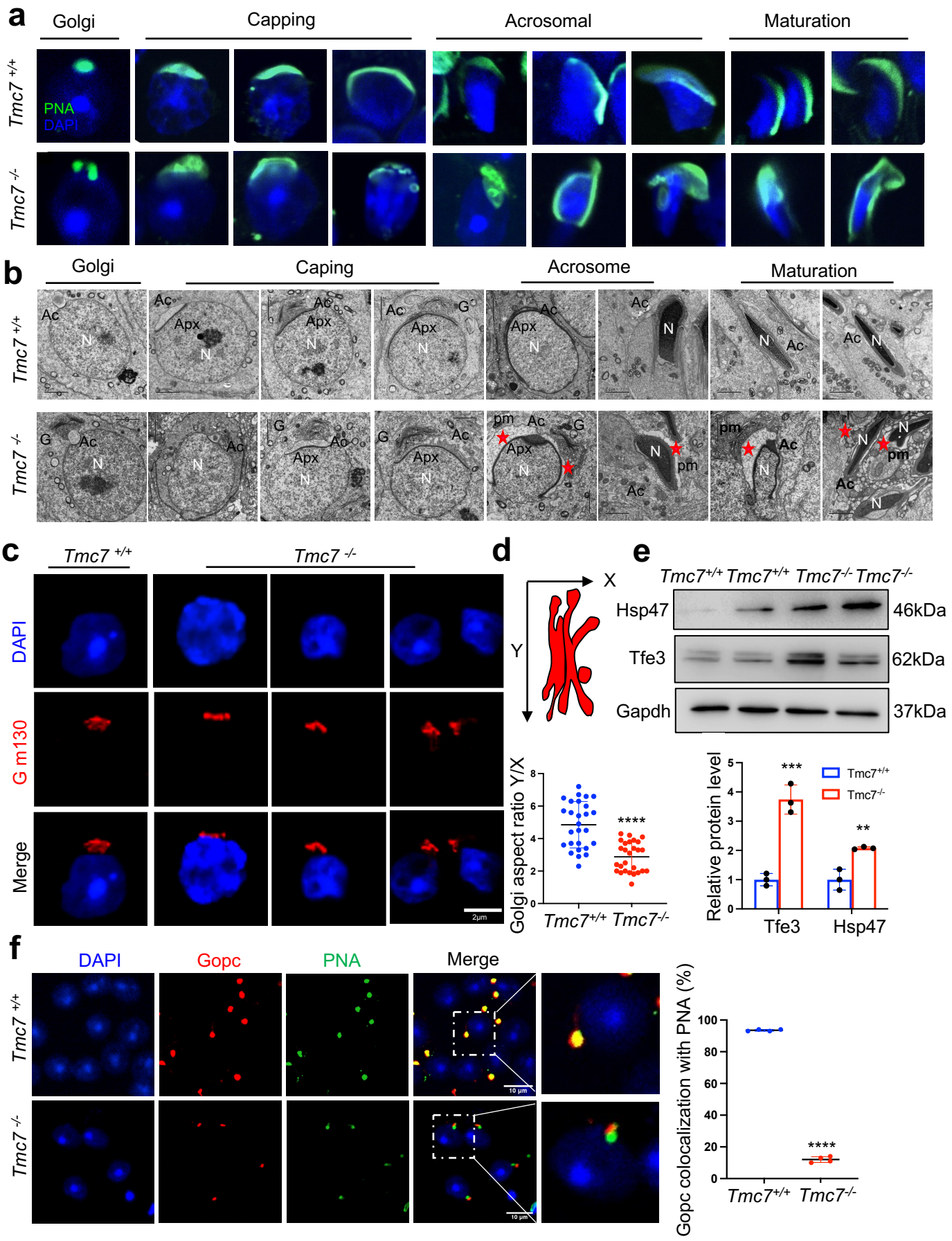

a
Golgi
Capping
Acrosomal
Maturation
Tmc7 +/+
Tmc7 -/-
PNA
DAPI
b
Golgi
Caping
Acrosome
Maturation
Tmc7 +/+
Tmc7 -/-
Ac
N
Apx
G
pm
c
Tmc7 +/+
Tmc7 -/-
DAPI
G m130
Merge
2μm
d
X
Y
Golgi aspect ratio Y/X
****
Tmc7+/+
Tmc7-/-
e
Tmc7+/+ Tmc7+/+ Tmc7-/- Tmc7-/-
Hsp47
Tfe3
Gapdh
46kDa
62kDa
37kDa
Relative protein level
***
**
Tmc7+/+
Tmc7-/-
Tfe3
Hsp47
f
DAPI
Gopc
PNA
Merge
Tmc7 +/+
Tmc7 -/-
10 μm
Gopc colocalization with PNA (%)
****
Tmc7+/+
Tmc7-/-

Fig. 4

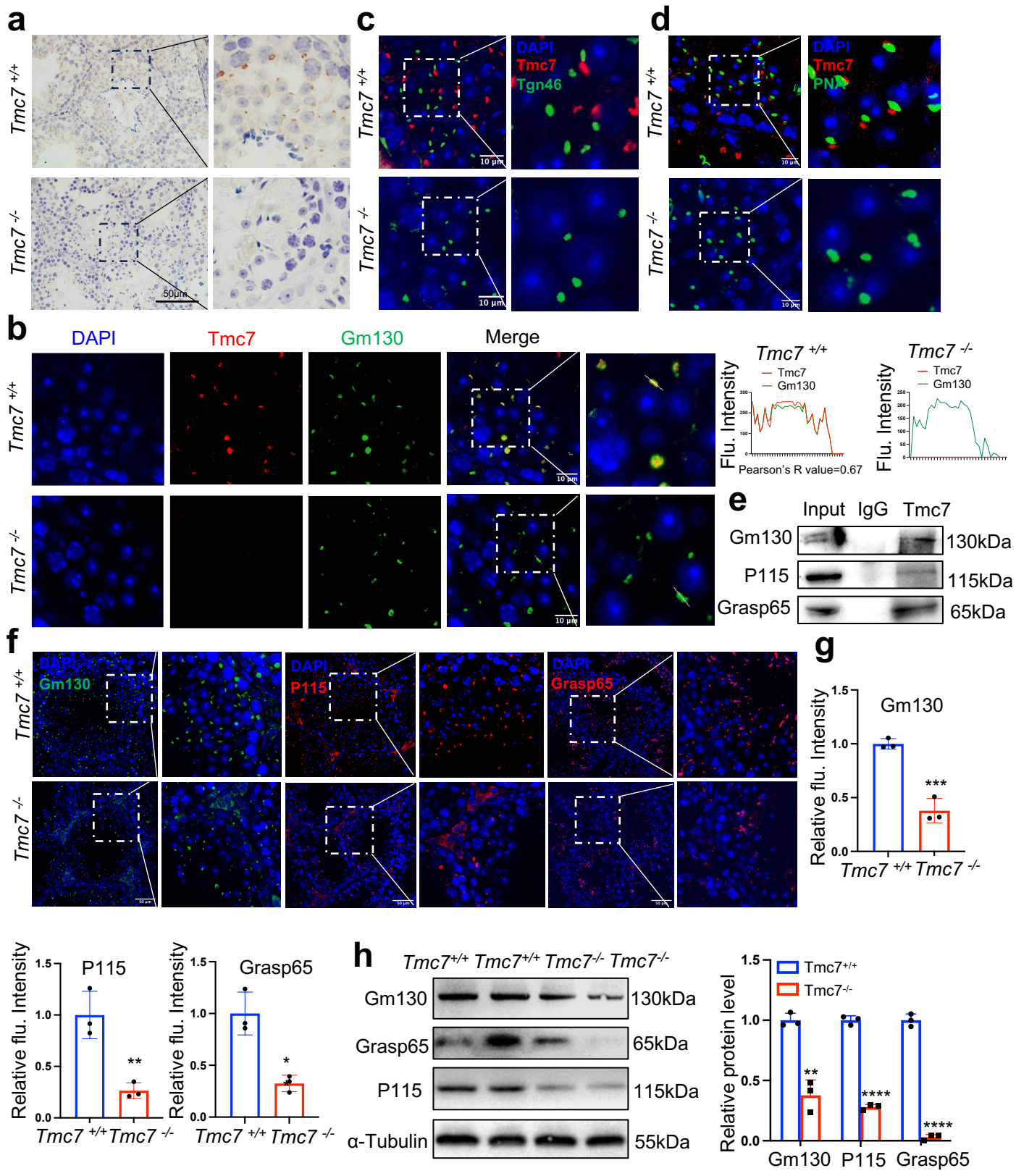
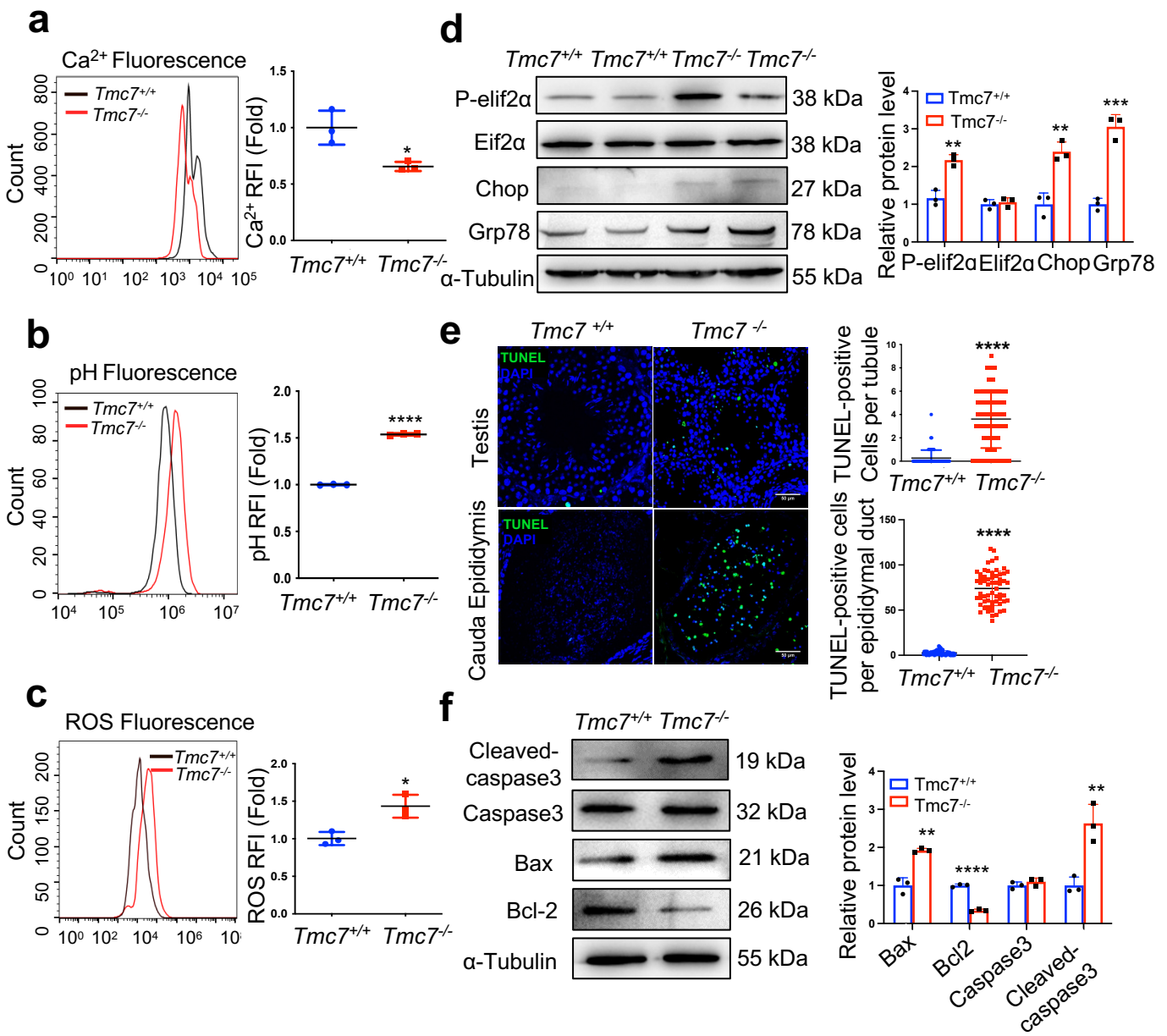

a
Tmc7 +/+
Tmc7 -/-
50μm
c
DAPI
Tmc7
Tgn46
10 μm
d
DAPI
Tmc7
PNA
b
DAPI
Tmc7
Gm130
Merge
Flu. Intensity
Pearson's R value=0.67
e
Input
IgG
Tmc7
Gm130
P115
Grasp65
130kDa
115kDa
65kDa
f
g
Relative flu. Intensity
***
h
α-Tubulin
55kDa
Relative protein level
**
****

Fig. 5


a
Ca2+ Fluorescence
Tmc7+/+
Tmc7-/-
Count
Ca2+ RFI (Fold)
b
pH Fluorescence
pH RFI (Fold)
c
ROS Fluorescence
ROS RFI (Fold)
d
P-elif2α
Eif2α
Chop
Grp78
α-Tubulin
38 kDa
27 kDa
78 kDa
55 kDa
Relative protein level
e
Testis
Cauda Epididymis
TUNEL
DAPI
TUNEL-positive Cells per tubule
TUNEL-positive cells per epididymal duct
f
Cleaved-caspase3
Caspase3
Bax
Bcl-2
19 kDa
32 kDa
21 kDa
26 kDa
Bcl2