Developmental tinkering of gene regulation facilitated super rapid adaptive radiation

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Abstract

Developmental shifts in gene regulation underlying key innovations remain largely uncharacterised at short evolutionary timescales. Here we investigate the gene expression and alternative splicing landscape of trophic innovations in the fastest vertebrate adaptive radiation: cichlid fishes from Lake Victoria. Using whole-transcriptomes of the oral and pharyngeal jaws from two life stages of species adapted to divergent trophic niches, we show that gene and isoform expression were fine-tuned during development to generate specialised species-specific eco-morphologies in adults. This is a striking contrast to mammals where tissue-specific gene expression is conserved across species even after 90 million years of evolution and suggests that gene regulatory programs have evolved rapidly in Lake Victoria cichlids. We show that gene expression during development is highly modular and this developmental modularity may have facilitated trophic diversification. Furthermore, we discovered that retrotransposon mediated exonisation of a craniofacial development gene, KAZNB, has contributed to the evolution of a novel trophic niche. This advocates for the importance of exonisation and the splicing machinery in rapidly expanding the coding capacity of the genome <15,000 years. We also identified a major transcription factor of craniofacial remodelling in Darwin's finches, ALX3, to have contributed to jaw divergence in Lake Victoria cichlids. Overall, our results suggest that developmental tinkering of gene regulation is a major facilitator of key innovations and diversification during adaptive radiation.

Introduction

Since King and Wilson (1975) concluded that variation in protein sequence alone cannot explain the phenotypic differences between human and chimpanzees, transcriptional regulation has been shown to play a fundamental role in phenotypic evolution and diversification (Brawand et al. 2011; Barbosa-Morais et al. 2012; Merkin et al. 2012). However, few studies have focused on recently diverged species, where regulatory changes are hypothesised to dominate phenotypic divergence, as they can emerge more rapidly from standing genetic variation and relaxed selection than de novo

protein-coding variation (Singh et al. 2017; El Taher et al. 2020; Richards et al. 2021; Singh et al 2021; Singh and Ahi 2022; Carruthers et al. 2022). Even fewer studies have looked at regulatory changes during development in recent and rapid bursts of diversification (Cardoso-Moreira et al 2019).

Cases of adaptive radiation, where species diversify rapidly in tight connection with ecology, provide promising model systems to study the role of gene regulation in rapid evolutionary diversification (Abzhanov et al. 2006; Gillespie et al. 2020; De-Kayne et al. 2022). The radiations of cichlid fishes in the Great East African lakes are such model systems where rapid diversification proceeded along many ecological and morphological axes (reviewed in Kocher 2004). The Lake Victoria (LV) cichlid radiation comprising ~500 species is arguably one of the most spectacular of these radiations, having emerged after the complete desiccation of the lake <15,000 years ago (Johnson et al. 2000; Seehausen 2002). Lake Victoria cichlids evolved diverse trophic adaptations facilitated by the reshuffling of ancestral standing genetic variation as ecological opportunity arose (Meier et al. 2023). The functionally decoupled oral and pharyngeal jaws of cichlids is hypothesised to be a key innovation and basis of their evolutionary success that enabled rapid trophic diversification (Fryer and Iles 1972; Parsons et al. 2011). It was recently found that cichlids dominate the biomass of Lake Victoria not because they arrived first as the lake filled up, but because they had ecologically versatile trophic morphology (Ngoepe et al. 2023).

Many studies have investigated the genetic architecture and myriad signalling pathways underlying trophic morphologies in teleosts and more specifically in cichlids (Albertson et al. 2005; Albertson and Kocher 2006; Parsons and Albertson 2009; Roberts et al. 2011; Parsons et al. 2014; Ahi 2016; Singh et al. 2021). In addition to gene expression variation, alternative splicing of pre-mRNA is also thought to contribute to transcriptional diversity and act as substrate for rapid adaptive evolution (Singh et al. 2017). Through alternative splicing a single gene can be altered into many mRNA isoforms which can potentially be translated into different proteins, boosting transcriptomic and proteomic diversity (Kim et al. 2007; Nilsen and Graveley 2010). Alternative splicing can provide

rapid paths to the evolution of phenotypic diversity (Singh and Ahi 2022). Although it is debated to which extent AS events are contributing to proteomic and phenotypic diversity (Tress et al. 2017; Blencowe 2017). Splicing processes such as exonisation, where exons evolve from retained introns can also rapidly generate protein-coding and regulatory variation (Keren et al. 2010; Sorek 2007). While it is generally thought that intron retention is rare in animals compared to plants, Singh et al (2017) found higher intron retention in cichlid fishes than humans. The relationship of alternative splicing for adaptive diversification and speciation is a research topic of increasing interest in animals and plants (Singh et al. 2017; Bush et al 2017; Smith et al. 2018; Smith et al. 2020; Jacobs and Elmer 2021; Wright et al. 2022), but more research is needed to illuminate the role of alternative splicing during adaptive radiation, and speciation in general (Singh and Ahi 2022).

Recent work has shown that ontogenetic diet shifts, which are common in animals (Sánchez-Hernández et al. 2019), can alter the trajectory of adaptive radiations in the presence of ecological opportunity (Ten Brink and Seehausen 2022). Little attention has been directed towards the role of ontogenetic developmental changes during adaptive radiation. In cichlids, there is some evidence that key shape changes in cichlid jaw cartilage and bone occur during early development (Albertson and Kocher 2006; Fujimura and Okada 2008) and that specialised morphologies are already distinct between trophic niches at the onset of exogenous feeding (Singh et al. 2017). But most studies on the transcriptional basis of trophic divergence across multiple species have focused on one stage (El Taher et al. 2020), and not much is known about the transcriptional patterns and mechanisms underlying the larval development of alternative trophic morphologies and further ontogenetic changes towards adulthood.

Here we leveraged the comparative framework to investigate the differences and commonalities between two postembryonic key life stages in cichlids: the end of the larval stage: stage 26 (Fujimura and Okada 2007) and the adult stage. The end of the larval development (stage 26) is marked by the complete absorption of the yolk sac into the body cavity and denotes the onset of independent foraging as the larvae are released from the mouthbrooding mother's mouth. The adult stage (marked by the

onset of breeding and the emergence of male nuptial coloration) stands for full maturity of the entire phenotype. To identify developmental genes and regulatory mechanisms involved in species differentiation and trophic adaptation, we explored the transcriptional landscape underlying the trophic adaptations of the oral jaw apparatus (OJA) and pharyngeal jaw apparatus (PJA) (Fig. 1 B and C) in the exceptionally young radiation of Lake Victoria cichlids (Fig. 1).

Results

Genome-guided transcriptome assembly and annotation

To analyse the transcriptional landscape of LV cichlids adapted to different diets we generated whole mRNA transcriptomes from oral and pharyngeal jaws of five Lake Victoria species (*Haplochromis thereuterion* (Ht), *Gaurochromis hiatus* (Gh), *Mbipia lutea* (Ml), *Neochromis omnicaeruleus* (No) and *Paralabidochromis sauvagei* (Ps)) across two developmental stages (stage 26-yolk sac absorption into body cavity and adulthood-onset of male nuptial colouration) (Figure 1). Each stage for each species was represented by five biological replicates. Tissues from two individuals were pooled for each biological replicate due to the small size of the larvae. Our total sample set comprised 140 whole transcriptomes. After quality filtering, we retained an average of 3-9 million reads per sample. Of these reads 68%-82% could be mapped to the reference genome (*O. niloticus*; NCBI accession number: GCA_001858045.3) in the larval samples and 75%-82% in the adult samples (Supplementary File 1). We chose to map to *O. niloticus* because it is the best annotated reference genome and high quality transcript annotations were important for our analyses. The final merged annotation of all samples included 37,334 genes and the transcript annotation 59,849 transcripts.

Figure 1: Overview of study design and broad patterns of gene expression. (A) Map of Lake Victoria (B)
Study species and their respective trophic niches. Photographs of Oral jaw apparatus (OJA) (C) and
pharyngeal jaw apparatus (PJA) (D). Normalised gene expression PCA plot for larvae (E) and adults (G)
based on 37,334 genes. (F) and (H) show dendrograms of the Euclidean distance based on the gene
expression matrix in. Oral jaw apparatus (OJA; black), pharyngeal jaw apparatus (PJA; white), H.
thereuterion (Ht; dark orange), G. hiatus (Gh; bright orange), M. lutea (Ml; bright blue), N. omnicaeruleus
(No; dark blue) and P. sauvagei (Ps; pink).

Transcriptome wide patterns of gene and isoform expression

We quantified the gene expression of 37,334 genes across all samples. To obtain an overview of global gene expression patterns we conducted a principal component analysis (PCA) based on normalised expression of 37,334 genes. The PCA shows a clear separation of expression profiles of the two jaw tissue sets for both life stages but the separation is less prominent in adults (Larvae: PC1 (13.4%), Adults: PC1 (6.1%))(Fig. 1 E and G). We also quantified expressions of 59,849 transcripts to see how isoform expression compared to the gene expression. The isoform expression PCA showed similar patterns as the larval gene expression profiles, with a clear distinction between the two jaw apparatuses but less differentiation among species (PC1 (11.4%)) (Fig. 2 A). Isoform expression clustering in adults is more species specific but less variation is explained by PC1 (5.6%) and PC2 (4.4%) when compared to the gene expression PCA (Fig. 2 C).

Additionally we analysed the expression data using hierarchical clustering analysis (Figure 1F-H). In the clustering analysis of the larvae, the expression of the two jaws is clearly distinct and the OJA clustering follows the trophic niche classification, while PJA is species specific (Fig. 1 F). The clustering of gene expression in adults is strictly species-specific, regardless of jaw tissue (Fig. 1 H). The hierarchical cluster analysis of the isoform expression matrix shows a slight difference in the species patterning of the larval PJA compared to the hierarchical clustering of the gene expression matrix (Fig. 2 B), while the adult patterning stays the same (Fig. 2 D).

To place the gene expression patterns into a wider phylogenetic context, we included data from individuals of *Tropheops tropheops* (Tt) and *Astatotilapia burtoni* (Ab) at both stages as outgroups to extend the comparison to two more divergent species from Lake Malawi and the rivers around Lake Tanganyika respectively. The PCA as well as the hierarchical cluster analysis based on Euclidean distance shows a clear separation of expression profiles of *A. burtoni* and *T. tropheops*, from the Lake Victoria species (Fig. S1).

Figure 2: PCA plot for larvae (A) and adults (B) based on 59,849 normalised isoform expression counts.
(B) and (D) show dendrograms of the Euclidean distance for larvae (B) and (D) adults. Oral jaw apparatus
(OJA), pharyngeal jaw apparatus (PJA), H. thereuterion (Ht), G. hiatus (Gh), M. lutea (Ml), N.
omnicaeruleus (No) and P. sauvagei (Ps).

Genes involved in species and trophic differentiation in Lake Victoria cichlids

To identify genes that contribute to species differences in oral and pharyngeal jaws of LV cichlids we conducted differential expression analysis for all pairwise species comparisons for each jaw and developmental stage. Of the 37,334 genes, 5,743 - 10,024 (15.4%-26.9%) showed significant (adjusted p-value < 0.05) differential expression (DE) between at least one pairwise species comparison in all tissues and at both developmental stages (Table S1). Between 591 - 1,664 (1.6% - 4.5%) genes were differentially expressed in more than 50% of the comparisons investigated. Interestingly, no gene was differentially expressed across all comparisons.

To identify genes that play a role in trophic adaptation of the jaw apparatuses we primarily focused on comparing the carnivorous species (Ht, Gh) to the herbivorous species (Ml, No). We identified the intersection set of genes that were differentially expressed across all four comparisons (Gh vs Ml, Gh vs No, Ht vs Ml, Ht vs No) (Fig. 3), and discovered 237 genes that were differentially expressed in larval OJA across all four herbivore vs carnivore comparisons and 159 genes were differentially expressed in the larval PJA of all four comparisons (Fig. 3 A and B respectively). In adult OJA comparisons we identified an overlap of 128 genes that were consistently differentially expressed and 107 genes were differentially expressed in the PJA (Fig. 3 C and D). Of these gene intersection sets between carnivores versus herbivores 79% - 96% shared the same direction of differential expression in all stages and jaw modules (Table S2). Such genes that were upregulated in carnivorous species and downregulated in herbivorous species or vice versa are interesting genes for further investigation in shaping divergent trophic morphologies.

Figure 3: Differentially expressed genes between jaws of herbivorous and carnivorous Lake Victoria cichlids. Upset plots of overlapping differentially expressed genes for species comparisons including the carnivorous and herbivorous species. Points below bars show overlapping genes between indicated pairwise comparisons or unique sets of genes for single points. Light gray bars indicate an overlap between all pairwise comparisons. The total sum of differentially expressed genes for one pairwise comparison results from the sum of indicated bars. Upset plots are shown for larval OJA (A), larval PJA (B), adult OJA (C) and adult PJA (D). Venn diagrams show the overlap in larval and adult differentially expressed genes for OJA (E) and PJA (F). Oral jaw apparatus (OJA), pharyngeal jaw apparatus (PJA), H. thereuterion (Ht), G. hiatus (Gh), M. lutea (Ml) and N. omnicaeruleus (No).

We generated expression heatmaps with hierarchical clustering of all the aforementioned shared genes for each jaw tissue and developmental stage that were differentially expressed in all pairwise comparisons between carnivores and herbivores (Larvae: OJA: 273, PJA: 159; Adults: OJA: 128, PJA: 107; grey bars in Fig. 2 A - D). We also included the expression of these genes from omnivorous LV species *P. sauvagei*. The heatmap clustering placed *P. sauvagei* closer to the carnivorous *G. hiatus* and *H. thereuterion* for the oral and pharyngeal jaw gene expression at the larval (Fig. S2 A & B) and adult stages (Fig. S2 C and D). Additionally, we conducted Gene Ontology (GO) enrichment analysis to identify the biological processes that DE genes were involved in for these different stage and tissue comparisons (carnivorous versus herbivorous). We found that larval OJA genes were associated with jaw development related terms such as 'roof of mouth development' (*sox9b*) and 'collagen catabolic process' (*mmp2*); and also more general functions like 'netrin-activated signalling pathway' involved in axon extension and cell migration, 'regulation of lipid catabolic process' and 'actin filament severing' (Fig. 4 F, for full list see: Supplementary File 3). Genes differentially expressed in the larval PJA showed enrichment for GO terms involved in functions like 'regulation of cell shape' (*stat3*, *amotl2b*) and 'skeletal muscle satellite cell migration' (*rhocb*) (Supplementary File 3). In adult OJA, genes were significantly associated with the 'the positive regulation of canonical wnt signalling pathway' (*bambia*, *fam53b*) as well as 'wnt signalling pathway' (*bambia*, *bcl9l*, *fam53b*). Interestingly, genes differentially expressed in the adult PJA showed a significant enrichment in 'regulation of alternative mRNA splicing, via spliceosome' (*rbfox1*) (Full list in Supplementary File 3).

Differentially expressed genes spanning both life stages

Comparing the intersection sets of differentially expressed genes between carnivorous and herbivorous species at both life stages showed an overlap of 39 genes in the OJA (25 downregulated and 13 upregulated in carnivores compared to herbivores; initial DEG sets: larval OJA: 237; adult OJA: 128) (Fig. 3 E) and 27 genes in the PJA (18 downregulated and eight upregulated in carnivores compared to herbivores; initial DEG sets: larval PJA: 159; adult PJA: 107) (Fig. 3 F) (Supplementary File 2) across the life stages.

One gene that was consistently downregulated in the two carnivorous species as well as the omnivorous species in both tissues at both stages was *kaznb*, also called *Kazrin* or periplakin-interacting protein b (Fig. 4 A and D). We validated *kazbn* expression using qPCR analyses and found it supports the RNA-seq results (Fig. 4 B and E). A more detailed inspection of the *kaznb* gene found that the differentially expressed transcript of interest is absent in the reference genome's *kaznb* annotation. This transcript is spanning an intronic region in the *O. niloticus* reference and is not linked to any other annotated exons of *kaznb*. This observation suggests that exonization of introns is a mechanism through which both novel coding variation (new exon) and novel regulatory variation (new mRNA isoform) have evolved in the radiating modern haplochromine cichlids in Lake Victoria.

Figure 4: Genes involved in regulating herbivorous vs carnivorous jaw adaptations in Lake Victoria
cichlids. Selected parts of heatmaps (dashed lines show cuts in heatmap) showing differentially
expressed genes in comparisons including carnivorous versus herbivorous species for larval oral jaw
apparatus (A) and adult oral jaw apparatus (D) respectively (Full heatmaps in the supplementary (Fig. S3
A - D)). Genes printed in black were either used in the qPCR validation analysis and/or have been
significantly enriched for development and morphology related GO terms. Important jaw development
associated GO terms are listed in (C) for larval OJA tissue and (F) for adult OJA tissue. The bar plots
show qPCR validation of cmah and kaznb gene in larval (B) and kaznb and kcne4 in adult jaw tissues (E).
Oral jaw apparatus (OJA), pharyngeal jaw apparatus (PJA), H. thereuterion (Ht), G. hiatus (Gh), M. lutea
(Ml), N. omnicaeruleus (No) and P. sauvagei (Ps).

Gene expression modularity of the two jaws

Evolutionary modularity in gene regulation can contribute to morphological

diversification and specialisation through the rewiring of gene regulatory networks

(Wagner et al. 2007). We investigated gene expression differences between the oral

and pharyngeal jaw apparatuses in herbivorous versus carnivorous species, to see

whether the underlying gene regulatory architecture behind these functionally

independent jaw modules differs between trophic niches. At stage 26 larvae, 1,889

(1768+121) genes were found to be differentially expressed between oral and

pharyngeal jaw apparatus in all pairwise species comparisons, while at the adult stage

only 209 (121+88) genes were consistently differentially expressed between the two

jaws (Fig. 5 A; Fig. S3 A and B). One hundred and twenty-one of these genes were

shared between both developmental stages, of which 68 were upregulated and 49

downregulated in the OJA vs PJA across all species and stages (Fig. 5 A). Four of

these genes showed different directions of expression between the larval and the adult

stage. Three of those four genes *EPHB3*, *SLC9A3* and *tbx3* were upregulated in larval

PJA and downregulated in adult PJA. One gene, with the opposite expression pattern

was unannotated. Several genes of the *Hox* family (*hoxa4a*, *HOXA4*, *hoxa5a*, *hoxb3a*,

hoxc5a, *hoxc4a*) show a decreased expression in the OJA at both stages.

GO enrichment of the 121 overlapping differentially expressed genes between the OJA

and the PJA of all carnivores and herbivores for both life stages showed significant

enrichment for genes involved in 'cranial skeletal system development', regulation of

muscle contraction', 'multicellular organism development' and 'regulation of

transcription, DNA-templated' (Fig. 5 B). Genes that were associated with the GO term

'cranial skeletal system development' include the genes *ALX3* aristaless-like homeobox

3, *prrx1a*, paired related homeobox 1a, *dlx4b*, distal-less homeobox 4b and *nkx3-2*, NK3

homeobox 2 (Fig. 5 B).

Figure 5: The Venn diagram shows the overlap in larval and adult differentially expressed genes in the
oral versus the pharyngeal jaw apparatus and their direction of expression (upregulated genes are
indicated by the upward facing arrow in green, downregulated genes by the downward facing arrow in
red; direction indicated for OJA vs PJA) (A). Selected GO terms are listed in (B).

Validation of expression patterns using qPCR

To validate our findings concerning differentially expressed genes between carnivores and herbivores we conducted qPCR analysis for five candidate genes for trophic divergence at the stage 26 larvae and four candidate genes for trophic divergence at the adult stage, which were found to be differentially expressed in both jaw apparatuses in our DGE analysis. For the larval stage these included *cmah*, *ggt5a*, *granulin*, *kaznb* and *mpc1* and for the adult stage *kaznb*, *kcne4*, *mcl1b*, *slc5a8*. All of these differentially expressed genes showed the same direction of expression between the carnivores and the herbivores in our DGE analysis. While not all of the qPCR differences were significant, they followed similar trends as our DGE analysis results (Fig. S4). *cmah* was consistently upregulated in the DGE analysis in both tissues for all carnivorous species at the larval stage (Fig. 4 A; Fig. S4 A), while the qPCR analysis for *cmah* in both larval tissues showed a significant difference in expression only between *N. omnicaeruleus* and *H. thereutherion* in the OJA and both herbivores compared to *H. thereuterion* in the PJA (Fig. 4 B). The differential expression analysis of *mpc1* showed an upregulation in larval herbivores in the PJA, which can be seen in *N. omnicaeruleus* in the qPCR analysis, but the difference is not significant (Fig. S4 A). The gene expression of *kaznb* was upregulated in herbivores versus carnivores in all tissues and all stages in the DGE analysis. The qPCR analysis confirmed this pattern in both larval tissues, but at the adult stage, relative mRNA expression in the OJA of *kaznb* could not be detected, while in the PJA, a significant difference was detected only between *H. thereuterion* and *N. omnicaeruleus* (Fig. 4 E). The gene *slc5a8* did also not show significant differences between carnivorous and herbivorous species in relative mRNA expression in the adult OJA, despite it being upregulated in herbivorous species in the differential gene expression analysis (Fig. S4 B).

Differential Alternative Splicing underlying adaptation

To investigate the role of alternative splicing in trophic adaptation in Lake Victoria herbivorous and carnivorous species we conducted pairwise species comparisons to detect differentially used isoforms. The numbers of differentially used isoforms ranged

from 671 to 980 across all comparisons and stages (Table S3). Further investigations revealed quite small numbers of significantly alternatively used isoforms between the two trophic niches at each stage within the OJA and PJA. In total we found 56 isoforms to be differentially used between carnivorous and herbivorous species in the OJA at the larval stage and 43 isoforms in the PJA at the larval stage. At the adult stage we found 75 isoforms in the OJA to be differentially used and 78 isoforms in the PJA at the adult stage (Fig. S5 A, B, C, D). The intersection sets of differentially used isoforms between the stages per tissue reveals even less overlap with 20 isoforms between the OJA of larvae and adults and seven between the PJA (Fig. S5, E and F).

To visualise differential isoform usage, we chose the median PSI (percentage or proportion spliced in) values, describing the relative abundance of each isoform, for differentially spliced transcripts between our carnivorous and herbivorous comparisons (Fig. S6; Supplementary File 4). Two isoforms of *fgfr2* (fibroblast growth factor receptor 2) were differentially used in the OJA (Fig. 6 A & D) at both stages and the PJA of adults (Fig. S6) but showed a trophic signal only in the larval OJA tissue, with only *fgfr2.1* being expressed in the herbivores while both isoforms were used in the carnivores. In the adult OJA a gene called *frzb* (frizzled related protein) showed a clear distinction between the carnivorous and herbivorous species, whereby in the herbivores only one isoform is used (*frzb.1*), while in the adult carnivorous and the omnivorous OJA tissues the other isoform (*frzb.2*) shows the higher median PSI value (Fig. 6 E). *frzb* also is significantly differentially used in our comparisons of larval OJA tissue and showed up in the GO enrichment, however the pattern is more species-specific, with only *frzb.1* being abundant in larval Gh (carnivorous) and Ps (omnivorous) OJA tissue. The PJA of larvae showed the most distinct isoforms to be differentially used compared to the other three tissues (Fig. S6 B). Another gene that showed very similar patterns in the OJA tissues of both stages and the adult PJA is *col6a1*, collagen type VI alpha 1 chain, associated with maintenance of tissue integrity. The isoforms are more equally used in the carnivorous species, while in the herbivores *col6a1.1* shows a higher rate of usage than *col6a1.2* (Fig. 6 C & F).

Figure 6: Median PSI values for isoforms, which were differentially used in carnivorous versus herbivorous comparisons in larval (A, B, C) or adult (D, E, F) OJA and PJA. Depicted are three selected genes (fgfr2, frzb and col6a1) and their respective median PSI values in the respective tissue. Oral jaw apparatus (OJA), H. thereuterion (Ht), G. hiatus (Gh), M. lutea (Ml), N. omnicaeruleus (No).

GO enrichment analysis of genes that were alternatively spliced showed *frzb* and *fgfr2*

to be significantly enriched in the OJA at both stages, as well as the PJA of the adult

stage (Supplementary File 5). *frzb*, was associated with 'chondrocyte proliferation' and

fgfr2 with 'fibroblast growth factor receptor signalling pathway' in larval and 'chondrocyte

proliferation', 'peptidyl-tyrosine phosphorylation' and 'peptidyl-tyrosine modification' in

the adult OJA as well as 'positive regulation of cell proliferation' and 'transmembrane

receptor protein tyrosine' in adult PJA GO enrichment analysis.

Contrasting differential gene expression and alternative splicing

To investigate if genes that were differentially expressed were also alternatively spliced

we looked at the intersection set between the two analyses. Interestingly, there was little

to no overlap between differentially expressed and alternatively spliced genes between

herbivores and carnivores. Only one transcript was both, differentially expressed as well as differentially spliced in the OJA of larvae, but was not annotated. In larval OJA different transcripts associated with three genes (*col6a1*, *efna1a* and *si:ch211-219a4.3*) were differentially spliced and differentially expressed. This suggests that gene expression and alternative splicing perform non-redundant functions.

Discussion

Variation in gene regulation is associated with phenotypic diversity and adaptation (King and Wilson 1975) and it is more likely to be important during short evolutionary timescales because new beneficial protein-coding and structural mutations take more time to evolve and establish. While the role of gene regulation has been studied in the context of adaptation and speciation (Mack and Nachman 2017, Singh et al. 2017, El Taher et al. 2020, Pavey et al. 2010), its developmental and ontogenetic dynamics are not well understood. The extremely young and ecologically diverse species flock of haplochromine cichlids from Lake Victoria in East Africa represents a highly suitable model to test the relative importance of different levels of gene regulation during development across different species. Here we compared the gene regulatory landscape of Lake Victoria cichlid species adapted to divergent trophic niches at two post-embryonic life stages and found evidence for substantial transcriptional rewiring and modularity during development between species adapted to divergent trophic niches.

Developmental transcriptional dynamics of cichlid jaws

Studying natural populations that have undergone adaptive diversification can provide insights into how organisms adapt to changing environments or new ecological opportunities. It can help solve the evolutionary puzzle as to why some lineages diversify rapidly, while others do not, and which genetic and molecular mechanisms underlie these processes. We analysed the transcriptional landscape of oral and pharyngeal jaws in Lake Victoria cichlids and found a strong signal of gene and isoform expression conservation at the tissue level in larvae (Fig. 1 E & F, Fig. 2). Unlike the

pharyngeal jaws, which are used for processing food, oral jaw samples are clustered by trophic niche specialisation at the larval stage. Oral jaws are used to capture food and morphological differences in species-specific adult morphologies already arise during early juvenile development (from stage 26 onwards) (Fujimura and Okada 2008). This suggests that the gene regulatory pathways that are involved in trophic differentiation of oral jaws are active early in development, in preparation for when the larvae leave their mouthbrooding mother's buccal cavity to start feeding independently. In contrast to the larvae, jaw gene and isoform expression in adults was species-specific (Fig. 1 G & H, Fig. 2). So oral and pharyngeal jaws of one species were more similar to each other than to the same tissue in a closely related species. This is different to what has been observed in adult mammals where gene expression is conserved in deep evolutionary time ~100 Ma (Merkin et al. 2012). It suggests that gene expression is evolving rapidly in cichlid jaws, and this may have allowed this lineage to rapidly evolve versatile trophic adaptations. Overall our results suggest that gene regulation was fine-tuned during development in species adapted to different trophic niches from being tissue specific in larvae and species-specific in adults.

Cichlid oral and pharyngeal jaw expression decoupling versus conservation

It has been proposed that evolutionary decoupling (i.e. modularity) of jaws played an important role in cichlid diversification (Liem 1973). However, this hypothesis has mostly been tested in form and function (Liem 1973; Hulsey et al. 2006; Conith and Albertson 2021), or gene regulation at one developmental stage (Singh et al 2021) and little is known about the role of development and post-larval ontogeny underlying cichlid jaw modularity. As pointed out by Conith and Albertson (2021) it is important to study integration/modularity at different taxonomic and developmental levels. Our data found that two jaw apparatuses are developmentally modular, in that the morphological trajectories according to the trophic niche are set early in life and later the transcriptional maintenance of the tissues, including response to environmental influences via gene expression, becomes more species-specific (Fig. 1 F & H). The two jaw apparatuses become more integrated (i.e. clustered by similarity) in gene expression towards

adulthood, pointing to a shift from modular to more integrated along the life history in Lake Victoria cichlids. Hence, ontogenetic fine-tuning of gene expression is important for building species-specific jaws in cichlids, at least at an early stage of adaptive radiation.

Developmentally changes in gene regulation were also reflected in the number of differentially expressed genes between OJA and PJA across the stages (Fig. S3 A & B) and in the ratio of genes shared across species (Fig. 5). A subset of genes was differentially expressed between the two jaws at both stages in all species (a total of 121 genes - 6,8% of the shared genes for larvae but 63.7% for adults) and showed the same direction of expression (except four genes). This points to a conserved function of these 121 genes at both life stages. Several *Hox* genes were part of this conserved gene set as *Hox* genes play an important role in the anteroposterior patterning of organisms and are known to be not present in the first pharyngeal arch, the mandibular arch (MA), forming the jaws in gnathostomes (Kuratani 2004; Ahi 2016). Recently it has also been shown that genes of the *Hox* family are not only expressed during development but also in adult tissues, contributing to bone regeneration (Rux and Wellik 2017; Bradaschia-Correa et al. 2019). Members of the ALX transcription factor family are major players in skeletogenesis in vertebrates (Khor and Ettensohn 2020). While the exact role of *ALX* genes during development is not completely understood, they are known for their association with cranial disorders (frontonasal dysplasia; FND). We found that alx3, which is known to be involved in cranial skeletal system development, was upregulated in oral jaws of LV cichlid fishes. In zebrafish (*Danio rerio*), *Alx* genes are enriched in frontonasal neural crest cells (NCC) at early stages of development, playing a role in shaping skeletal elements of the anterior neurocranium (Mitchell et al. 2021) . While *Alx3* has been repeatedly lost in chicken (*Gallus gallus*), a frog (*Xenopus tropicalis*) and lizard (*Anolis carolinensis*) species, its role might be compensated for by Alx1 and/or Alx4 (McGonnell et al. 2011). An alx1 haplotype has been associated with beak shape diversification across the adaptive radiation of Darwin's finches (Lamichhaney et al. 2015). Beak shape in finches has evolved in response to selection

pressures to adapt to different food sources. Our findings suggest the ALX transcription factors may be key players in trophic diversification during adaptive radiation across vertebrates.

Gene expression divergence in herbivores vs carnivores

Trophic diversification is a hallmark of adaptive radiation. While the transition from carnivory to herbivory seems limited to only some fish genera, including cichlids and carps, the repeated evolution of herbivory is even rarer but can be observed in modern haplochromines (He et al. 2015; McGee 2020; Singh et al. 2022). Unravelling the underpinnings of the evolution of herbivory and carnivory is key to understanding how trophic diversity shaped cichlid radiations. Singh et al. (2022) found that in Tropheini from Lake Tanganyika, the repeated trophic transitions between herbivory and carnivory are probably based on repeated cases of separation and secondary contact, but the molecular driver of these transitions may be regulatory variation rather than coding mutations. As regulatory changes can evolve more rapidly than coding mutations, investigating the role of gene regulation is especially interesting in very young adaptive radiations.

To understand the role of gene expression in trophic adaptation we compared expression patterns in pairwise species comparisons between herbivorous and carnivorous species. A small number of shared differentially expressed genes across all species pairs indicates highly species-specific gene regulatory programs in Lake Victoria cichlids. The majority of genes (79% - 96%) that were differentially expressed between all carnivores vs herbivores pairs has the same direction of expression, suggesting a role of these genes in repeatedly contributing to carnivorous and herbivorous trophic adaptations. Several of these genes were associated with Wnt signalling, such as the *cmah* gene that was higher expressed in carnivores than herbivores at the larval stage (Fig. 4 A & B). *Cmah* encodes an enzyme that regulates Wnt signalling (Nystedt et al. 2009). The Wnt/β-catenin signalling pathway plays a crucial role in skeletogenesis of trophic structures, especially in remodelling and growth

of fish jaws (reviewed in Ahi 2016). Higher expression of Wnt in Lake Malawi cichlids, was also correlated with a more steeply descending profile and proposed to lead to early variation and increased ossification in craniofacial development (Parsons et al. 2014). Herbivory vs carnivory genes in OJA at the larval stage were enriched for GO terms like 'roof of mouth development' (Fig. 4 C) that was associated with genes *sox9b*, which is downregulated in the carnivorous species. *Sox9b* is a transcription factor involved in chondrogenesis and has been shown to be important for craniofacial development (Yan et al. 2005). The repression of *sox9b* expression through activation of the Ahr pathway resulted in the development of a shortened jaw (Xiong et al. 2008). The differential regulation of the Ahr pathway has been shown to play a key role in the trophic divergence of the OJA in Arctic charr (Ahi et al. 2015). In adults, *slc5a8* was upregulated in the jaws of herbivores. The role of *slc5a8* in skeletal development and morphogenesis is unclear, however, it has been shown to be a regulator of Wnt/β -catenin signalling in mammals (Hu et al. 2016). In the OJA of adults, significant GO enrichment terms included 'wnt signalling pathway' and 'positive regulation of canonical Wnt signalling' (Fig. 4 F) associated with the genes *fam53b* and *bcl9l*. *Fam53b* plays a pivotal role in regulating the Wnt pathway (Kizil et al. 2014); *bcl9l* has been linked to tooth development in mice (Cantù et al. 2017) and craniofacial malformations in zebrafish (Cantù et al. 2018). Both these genes showed a lower expression in carnivores versus the herbivores.

Besides genes related to the canonical Wnt/β-catenin signalling pathway we found *ggt5* to be downregulated in the jaw tissues of larval herbivores. *Ggt5* is a direct downstream target of *runx2*, a major osteogenic transcription factor (Tarkkonen et al. 2017). It is likely that increased expression of *ggt5* in skeletal tissue enhances bone resorption by induction of osteoclastogenesis which has also been observed in another member of the same gene family in mice (Hiramatsu et al. 2007). Thus, the lower expression of *ggt5a* in both jaws of herbivorous species may indicate differences in bone resorption level between the trophic niches. Interestingly, another gene with reduced expression in both jaws of herbivores, *granulin*, is also known for its key role in the stimulation of

osteoclastogenesis through activation of RANK signalling (Oh et al. 2015). Taken together expression results for *ggt5a* and *granulin* may imply a distinct bone resorption process between carnivores and herbivores at the larval stage.

In adults, the most consistent expression differences in both jaws (validated by RNAseq and qPCR) between the trophic niches were observed for *kcne4* showing reduced expression in herbivores. The role of *kcne4* in skeletal development has remained unclear, however, it is known that *kcne4* directly interacts with calmodulin (CaM) (Ciampa et al. 2011), the ubiquitous Ca2+-transducing protein with an important role in jaw skeletogenesis (reviewed by Ahi 2016). The signal mediated by CaM has been already demonstrated as a key molecular mechanism contributing to variation in jaw structures among closely related species of birds and fish (Abzhanov et al. 2006; Parsons et al. 2009; Gunter et al. 2014). Interestingly, *kcne4* is identified as the only ion channel gene which showed an upregulated expression after mechanical loading in rats (Mantila Roosa et al. 2011) which is accompanied with increased bone formation. This could imply an increased bone formation in herbivores as has also been suggested by Albertson et al. (2012) when comparing the maxilla of a cichlid specialised in scraping tough, filamentous algae to a closely related more pelagic planktivorous species. The maxilla of the herbivore species is revealed to contain more bone and to be more resistant to bending, compared to the planktivorous species (Albertson et al. 2012).

Alternative splicing contributed to trophic divergence of jaws

Similar to differentially expressed genes, alternatively spliced genes showed stage-specific and tissue-specific patterns. Some genes were alternatively spliced in more than one tissue and stage. One of those genes that was significantly enriched in the GO analysis with the biological process term 'chondrocyte proliferation' (larval OJA and adult OJA) was *frzb*, frizzled related protein. While the two isoforms of this gene showed a species-specific usage pattern in the larval OJA, a trophic niche specific pattern in usage in the adult OJA is apparent (Fig. 6 B & E). Kamel et al. (2013) showed

that *frbz* plays an important role in craniofacial development and palate morphogenesis, with a knockdown of *frzb* in zebrafish causing the complete loss of the lower jaw. *frzb* also enhances the diffusion of Wnt by blocking short-range effects and increasing long-range gene responses (Rochard et al. 2016), making it a modulator of Wnt signaling and playing a role in the convergent-extension of the jaw skeleton, meaning that it might be an underlying molecular reason for the protrusion of the jaw in carnivores. A gene with different isoform usage between the OJA of larval herbivores and carnivores was *fgfr2*, fibroblast growth factor receptor 2 (Fig. 6 A & D). Mutations in *fgfr2* have been associated with Crouzon syndrome, causing craniosynostosis (the premature fusion of sutures) and is also linked, among other features, to a short hypoplastic maxilla and mandibular prognathism (Reardon et al. 1994; Glaser et al. 2000), potentially linking differences in isoform usage to a steeper jaw profile in herbivores. In cichlids *fgfr2* has also been found to be involved in tooth development (Hulsey et al. 2016). Overall these results suggest that the alternative usage of spliced isoforms played a role in the rapid evolution of jaws adapted to different food sources in Lake Victoria cichlids. Alternative splicing has been shown to evolve faster than gene expression in deeply divergent vertebrates (Barbosa-Morais et al. 2012). Since the long evolutionary divergence time between those species is not comparable to the very young Lake Victoria cichlids studied here, alternative splicing seems to already play an important role in formation of adaptive morphologies at very early stages of evolutionary divergence.

Exonization generated novel variation for the evolution of herbivory

Exonization, the process of non-coding intronic regions becoming protein-coding exons, is a mechanism of genome evolution that can generate new gene isoforms (Sorek 2007). Although most exonization events are believed to be molecular noise, it provides an efficient way for generating new evolutionary solutions from the original genome. As long as the original isoform maintains functionality, the new isoform can avoid selection and either evolve a novel function or be purged from the population (Schmitz and Brosius 2011). Because exonization can rapidly evolve from standing variation, it has the potential to be an important generator of adaptive isoforms during diversification.

When we compared the jaw transcriptomes of herbivorous and carnivorous cichlids in
our study, we found that only a small fraction of genes to be differentially expressed
between all carnivores and herbivores across both stages within each jaw and just a
few of those were annotated (Supplementary File 2). One of these was a transcript of
kaznb, also called Kazrin, encodes periplakin interacting protein b, and is involved in
several cellular functions such as cytoskeletal organisation, epidermal differentiation,
and cell adhesion (e.g. Sevilla et al. 2008; Cho et al. 2011). The expressed transcript
falls into an intronic region in the *O. niloticus* reference and was consistently expressed
in the same direction across all tissues and stages with lower expression in the
carnivorous (and the omnivorous) species and higher expression in herbivores. We
were able to validate the expression profile of the *kaznb* gene in the larval tissues via
qPCR analyses (Fig. 4 B) and to some extent in the pharyngeal jaws of the adult tissues
(Fig. 4 E). A study in *Xenopus laevis* larvae concentrating on the function of *kaznb*
showed that it plays an essential role in craniofacial development, through controlling
neural crest cell (NCC) establishment (Cho et al. 2011). When depleting *X. laevis*
embryos for *kaznb* in the developing head region with morpholino oligo injection that
blocked *kaznb* translation, Cho et al. (2011) detected eye and head defects, with
smaller ceratohyal and ceratobranchial cartilages and eyes on the treated sides,
suggesting an important role in craniofacial development. This seems to be an
interesting gene which has not been suggested so far to be relevant in shaping
divergent cichlid trophic morphologies, at least to our knowledge. Future studies should
address a possible link to size differences between herbivorous and carnivorous jaw
structures.

It has been previously shown that specialised herbivorous and carnivorous
haplochromine cichlids had significantly higher intron retention in their mRNA than
omnivorous species (Singh et al. 2017). Some retained introns may represent exonised
introns that are translated produce a novel protein with new domains. As exonization
provides a relatively rapid way to generate protein coding variation and potentially
evolutionary novelty, it could play a previously underappreciated role in adaptive
radiation.

Distinct roles of gene expression and alternative splicing during adaptive radiation

A number of studies have empirically investigated the differing roles of differential gene expression and alternative splicing in phenotypic adaptation and found them to play non-redundant roles. For example, it was found that the overlap of differentially expressed and differentially spliced genes was not greater than expected by chance in *Drosophila melanogaster* adapted to extreme temperatures (Jakšić and Schlötterer 2016) and differentially spliced genes were involved in different processes compared to differentially expressed genes in arctic charr (Jacobs and Elmer 2021). In the butterfly *Bicyclus anynana* more than half of the genes that were differentially expressed were also differentially spliced but none of the genes associated with seasonal plasticity were differentially expressed and alternatively spliced (Steward et al. 2022). Similarly a recent study on the Eda haplotype in sticklebacks found only a small overlap (six genes) of gene expression and alternative splicing, but a lot of differentially spliced genes (no significant enrichment) were involved in the same biological processes as the differentially expressed ones (Rodríguez-Ramírez et al. 2023).

In our study, even though almost no genes that were both differentially expressed and differentially spliced, we did find several GO terms involved in morphological structure formation, even if not related to the exact same biological processes. The alternative splicing GO enrichment showed involvement in 'chondrocyte proliferation' (L OJA, A OJA and A PJA) while the differential expression GO enrichment was associated with terms like 'roof of mouth development' (L OJA) and 'Wnt signalling pathway' (A OJA). Even though different genes were regulated by these two gene regulatory mechanisms, the same developmental pathways were being targeted by both alternative splicing and differential gene expression to shape trophic morphologies. This is different to what has been found in the much older Lake Tanganyika cichlids where differentially expressed genes are involved in housekeeping processes while differentially spliced genes are involved in trophic morphologies (Singh et al. 2017). Furthermore, the distinction between which genes are regulated by alternative splicing or gene expression

divergence is likely affected by the genetic architecture (i.e. polygenic or simple) of the adaptive phenotype and the evolutionary age of the species.

Conclusion

Little is known about how gene expression and alternative splicing evolve at very short time scales as most studies published so far have focused on older lineages. Additionally the developmental dynamics of these two mechanisms is not well understood. Here we show that both differential expression and differential splicing are important regulatory processes for driving rapid morphological change. We also find the gene regulation at different life stages is distinct and future studies would benefit from including multiple developmental stages in their study design. Remarkably, we find that exonisation may be an important evolutionary mechanism for generating novel protein variation during rapid adaptive radiation. However, more research is needed to establish how common this mechanism is.

Material and Methods

Sampling

In this study we focus on five model species from Lake Victoria (*Gaurochromis hiatus* (*Gh*), *Haplochromis thereuterion* (*Ht*), *Mbipia lutea* (*Ml*), *Neochromis omnicaeruleus* (*No*) and *Paralabidochromis sauvagei* (*Ps*)). We picked those five species to try to reflect the broad spectrum of trophic specialisations in Lake Victoria. *G. hiatus* feeds on chironomid larvae as well as on bivalves and mollusks (Witte et al. 1982). *H. thereuterion* mostly feeds on insects like chironomid larvae, but a substantial portion of its nutrition is covered by collecting terrestrial insects from the water surface (Oijen et al. 1996). They both represent differentially specialised carnivores. *M. lutea* and *N. omnicaeruleus* are both mainly herbivorous but with different modes of feeding. While *M. lutea* feeds mainly on filamentous algae and also takes in some detritus , *N. omnicaeruleus* grazes off epilithic algae and diatoms (Seehausen et al. 1998). *P.*

sauvagei is an omnivore, feeding on epilithic algae, associated insects and sponges (Seehausen 1996). As an outgroup we included *Tropheops tropheops* (Tt), an herbivorous Lake Malawi cichlid, which mainly feeds on algae and scratches it from hard surfaces, but has also been reported to exclusively feed on plankton if abundant (Konings 1990). As a second outgroup species we selected *Astatotilapia burtoni* (Ab), a mostly riverine cichlid that is located in Lake Tanganyika and its surrounding waterways and can be classified as an omnivore, feeding on plants, aufwuchs, insects and fish remains (Muschick et al. 2012).

The *M. lutea* were collected from Makobe Island and the *G. hiatus* from the northern Mwanza Gulf. Both were collected by Ole Seehausen. The other samples, *N. omnicaeruleus*, *P. sauvagei* and *H. thereuterion* were obtained via aquarium trade. All fish were reared in the fish facility at University of Graz under standardised aquarium conditions regarding tank environment, diet and water parameters. Eggs were taken from the mouth of their mother 24 hours after fertilisation, by applying slight pressure onto their cheeks, and reared in a separate tank until the yolk sac was completely absorbed (stage 26, (Fujimura and Okada 2007)). The larvae were euthanized with an overdose of MS 222 and stored in RNAlater in a fridge for seven days and then moved to a -80°C freezer. Fish raised to the adult life stage were fed with a standardised diet of green spirulina flakes. When individuals started to show adult colouring and breeding behavior males were sacrificed with MS 222 and dissected. Bones were stored in RNAlater in a fridge for seven days and then frozen at -80°C.

RNA extraction

Per species and stage five biological replicates of oral and pharyngeal jaw apparatus samples were taken. In the case of the larvae the area of the OJA and of the PJA were sampled under a stereo microscope and submerged in a lysis buffer and 1-thioglycerol according to the protocol of the Promega RNA Reliaprep kit. This kit includes filter columns and a DNA digestion step. The sample in the lysis buffer was crushed with a ceramic bullet in a homogenizer (FastPrep-24, MP Biomedicals, Santa Ana, CA, USA), before further processing. Two individuals were pooled per sample to increase the

amount of RNA. For the adult species the bones were all cleaned from skin and muscle tissue and stored in RNAlater. As the whole set of bones would have been too much tissue input for the RNA extraction, only half of the bones were collected. The set of the bones is the same for all samples, but if they were collected from the left or right side was randomly decided by a simple randomization script in R (R Core Team 2020). RNA for the adults was extracted using TRIzol reagent (Thermo Fisher Scientific), following the protocol of the TRIzol protocol for RNA extraction with the inclusion of a DNAse step at the end (invitrogen). RNA quality was tested at a TapeStation 2200 with RNA Screen Tapes. It was tried to reach a RINe number of >= 7 for each sample.

Library preparation and sequencing

Library preparation was done with the TruSeq Stranded mRNA Sample Prep Kit using 24 indexing adapters and following the standard protocol. For the input RNA we tried to reach 1000 ng for each sample. The quality of the cDNA was tested on a TapeStation 2200 (Agilent) with D1000 Screen Tapes from the Agilent RK6 ScreenTape System. Sequencing was executed at the Vienna BioCenter Core Facilities on a HiSeq 2500 with paired end 125 cycles per read (2 x 125 cycles). Demultiplexing was conducted by the same facility.

Transcriptome assembly

For each of the samples we got an average of ten million paired-end reads. Sequence reads are available at NCBI sequence read archive (SRA) under the accession number PRJNA640176. After a quality check with Fastqc (v0.11.8) (FastQC 2015) and a trimming step with Trimmomatic (v0.3.9) (Bolger et al. 2014), only reads with a phred > 28 and a minimum length of 70 bp were retained. For each sample a median of about 5 million reads were obtained. For read alignment we used RNAstar (v2.7.3.a) (Dobin et al. 2013) with the *Oreochromis niloticus* genome reference from the University of Maryland (Conte et al. 2017) as guide reference, as it is very well annotated and this is important for accurate isoform reconstruction. After checking the mapping statistics with samtools idxstats (v1.9) (Danecek et al. 2021) and merging the single files for the same

species and jaw type with picard (v2.21.7) (Picard toolkit 2019) we used StringTie

(v2.0.6) (Pertea et al. 2015) without a reference to assemble the RNA-seq alignments

into potential transcripts. This was done separately for the single files (per fish) and the

merged files (per species and jaw). The single files for each biological replicate were

then merged according to species, to subsequently merge all the files per species,

tissue and stage. These repeated merging steps were conducted to reduce the

probability of false positives. To estimate the accuracy of the produced annotation files

we compared them with gffcompare (v0.11.2) (Pertea and Pertea 2020) to our reference

annotation. We filtered for monoexonic transcripts not contained in the reference and a

class code assigned by gffcompare, indicating 'possible polymerase run-on' fragments.

The maximum intron length was reduced to 200000 bp, which is the maximum intron

length found in the *O.niloticus* reference. Based on these annotations, the expression

estimates were generated with StringTie allowing no multimapping. From these

expression estimates, count matrices were produced using the prepDE.py script that

comes with StringTie (v2.0.6) to extract raw count data from StringTie results. The code

used for the whole analysis is stored in a github repository

(https://github.com/annaduenser/LV_RNAseq).

Differential gene expression analysis

We were not able to directly compare jaws in adults and larvae as we were not able to

apply an identical dissection and extraction process at both life stages due to the

minimal size of the larvae. We counteracted this shortcoming with not directly

comparing the data sets, but conducting the same species-pairwise comparisons and

further investigating the intersection set between those. This method should reveal

identical patterns in gene expression and alternative splicing between trophic niches, if

present. When looking at the Euclidean cluster dendrogram at the larval stage, that also

includes our outgroup, we see a clear distinction between the LV species and the latter.

When including species from LM and LT, a jaw tissue specific pattern is still visible for

LV as well as for LM (clustering next to LV), but not for LT (Singh in preparation)

suggesting that this separation is not solely caused by the inclusion of surrounding

muscle tissue. If anything, the surrounding tissue could potentially add to the trophic

niche clustering in the larval oral jaw apparatus. We used DESeq2 (Love et al. 2014) in
(R Core Team 2020) using RStudio (RStudio Team 2019) to detect differentially
expressed genes running comparisons between all species and tissues as well as
comparisons between the tissues within a species. DESeq2 uses raw read counts and
estimates variance-mean dependence based on a model that utilizes the negative
binomial distribution (Love et al. 2014). The cutoff for differentially expressed genes was
chosen at a false discovery rate of (p < 0.05).

GO term enrichment

GO terms for *Oreochromis niloticus* were acquired in R (R Core Team 2020) using
RStudio (RStudio Team 2019) via the biomaRt package (v2.46.1)(Durinck et al. 2005;
Durinck et al. 2009) in R from the Ensembl database. Gene set enrichment analysis
was conducted with the R program topGO (v2.36.0)(Alexa and Rahnenfuhrer 2019)
using the method *weight* to account for GO topology and a Fisher's exact test for the
enrichment analysis. We used our annotation to build the gene universe, where we then
tested the list of genes, overlapping between carnivorous and herbivorous species in
gene expression or alternative splicing analysis for gene set enrichment.

qPCR analysis

From four replicates of RNA samples of each jaw apparatus, stage and species, we
used 500 ng of RNA to synthesise first strand cDNA using the High Capacity cDNA
Reverse Transcription kit (Applied Biosystems). The cDNA from each sample was
diluted 1:10 times in nuclease-free water to be used for qPCR. For the adult stage, we
selected five candidate target genes; *kcne4*, *kaznb*, *mcl1b*, *rnf114* and *slc5a8*, and for
the larval stage we also chose five genes; *ggt5a*, *kaznb*, *granulin*, *faah* and *cmah* (Fig.
S5). We used *rpl18* as a reference gene, which we have already validated as a stably
expressed gene in both cichlid jaw tissues across different trophic niches (Ahi et al.
2019), to normalise our expression data for qPCR. The primers were designed at
conserved sequence regions of the coding sequences from all species using CLC
Genomic Workbench, version 7.5 (CLC Bio, Aarhus, Denmark) for alignment and

delineation of exon boundaries using *A. burtoni* annotated genome in the Ensembl

database (<http://www.ensembl.org>) (Zerbino et al. 2018). Primer Express 3.0 (Applied

Biosystems, CA, USA) and OligoAnalyzer 3.1 (Integrated DNA Technology) were used

to design the primers as described in details in our previous studies (Ahi et al. 2018; Ahi

et al. 2019) (Supplementary data for primer info). Maxima SYBR Green/ROX qPCR

Master Mix (2X) (Thermo Fisher Scientific, Germany) was used to generate qPCR

reactions. The amplification steps were conducted in 96 well-PCR plates on ABI 7500

real-time PCR System (Applied Biosystems). The primer efficiency calculation, qPCR

program, plate set-up and a dissociation step were performed as described previously

(Ahi et al. 2018; Ahi et al. 2019). To normalise the expression levels, the Cq value of

rpl18 was used as normalisation factor (Cq reference), and Cq of each target gene was

calculated accordingly (ΔCq target = Cq target - Cq reference). Relative expression

quantities (RQ) were calculated for the normalised values using E-ΔCq (Pfaffl 2001) and

then fold difference values were calculated by transformation of RQ values to

logarithmic values in order to conduct further statistical analysis (Bergkvist et al. 2010).

The statistical expression differences were identified using ANOVA statistical tests,

followed by Tukey's HSD post hoc tests.

Alternative splicing analysis

As the alternatively spliced isoforms can be a result of different types of splicing, it is not

as straightforward to assess but can be explored from different angles. Either from an

exon centric approach via differential exon usage (DEU), or with an isoform centric

approach, which can be subdivided by differential transcript expression (DTE) and

differential transcript usage (DTU). To get the most information about splicing patterns,

we use a combination of DEU and DTU analysis in this study. Alternative splicing

analysis was conducted with SUPPA2 (v2.3) (Trincado et al. 2018) to evaluate

differential transcript usage (DTU) between the samples. SUPPA2 takes the normalised

isoform counts of all replicates of a condition and calculates a relative abundance for

each transcript in each sample (PSI). To detect the difference in transcript usage

between conditions dPSI is computed (dPSI = PSI condition 2 - PSI condition 1)

(Trincado et al. 2018). By taking the dPSI comparisons of herbivore versus carnivore

species and selecting the isoforms that were significantly used differently. We illustrated

the difference in usage by computing the median PSI value over the five biological

replicates for each species for each transcript. Visualisation of the analysis was

conducted in R (R Core Team 2020) using RStudio (RStudio Team 2019).

Data availability

816 Sequence reads are available at NCBI sequence read archive (SRA) under the

817 accession number PRJNA640176.

819 The code for the analysis is available at the following github repository

820 (https://github.com/annaduenser/LV_RNAseq).

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References

Abzhanov A, Kuo WP, Hartmann C, Grant BR, Grant PR, Tabin CJ. 2006. The calmodulin pathway and evolution of elongated beak morphology in Darwin's finches. *Nature*. 442(7102):563–567. https://doi.org/10.1038/nature04843

Ahi EP. 2016. Signalling pathways in trophic skeletal development and morphogenesis: Insights from studies on teleost fish. *Developmental Biology*. 420(1):11–31. https://doi.org/10.1016/j.ydbio.2016.10.003

Ahi EP, Singh P, Duenser A, Gessl W, Sturmbauer C. 2019. Divergence in larval jaw gene expression reflects differential trophic adaptation in haplochromine cichlids prior to foraging. *BMC Evolutionary Biology*. 19(1):1–14. https://doi.org/10.1186/s12862-019-1483-3

Ahi EP, Singh P, Lecaudey LA, Gessl W, Sturmbauer C. 2018. Maternal mRNA input of growth and stress-response-related genes in cichlids in relation to egg size and trophic specialization. *EvoDevo*. 9(1):1–17. https://doi.org/10.1186/s13227-018-0112-3

Ahi EP, Steinhäuser SS, Pálsson A, Franzdóttir SR, Snorrason SS, Maier VH, Jónsson ZO. 2015. Differential expression of the aryl hydrocarbon receptor pathway associates with craniofacial polymorphism in sympatric Arctic charr. *EvoDevo*. 6(1):1–18. https://doi.org/10.1186/s13227-015-0022-6

Albertson RC, Cooper WJ, Mann KA. 2012. More than Meets the Eye: Functionally Salient Changes in Internal Bone Architecture Accompany Divergence in Cichlid Feeding Mode. *International Journal of Evolutionary Biology*. 2012:1–9. https://doi.org/10.1155/2012/538146

Albertson RC, Kocher TD. 2006. Genetic and developmental basis of cichlid trophic diversity. *Heredity*. 97(3):211–221. https://doi.org/10.1038/sj.hdy.6800864

Albertson RC, Streelman JT, Kocher TD, Yelick PC. 2005. From The Cover: Integration and evolution of the cichlid mandible: The molecular basis of alternate feeding strategies. *Proceedings of the National Academy of Sciences*. 102(45):16287–16292. https://doi.org/10.1073/pnas.0506649102

Alexa A, Rahnenfuhrer J. 2019. topGO: Enrichment analysis for gene ontology.

Barbosa-Morais NL, Irimia M, Pan Q, Xiong HY, Gueroussov S, Lee LJ, Slobodeniuc V, Kutter C, Watt S, Colak R, et al. 2012. The Evolutionary Landscape of Alternative Splicing in Vertebrate Species. *Science*. 338(6114):1587–1593. https://doi.org/10.1126/science.1230612

Bergkvist A, Rusnakova V, Sindelka R, Garda JMA, Sjögreen B, Lindh D, Forootan A,
Kubista M. 2010. Gene expression profiling - Clusters of possibilities. *Methods*.
50(4):323–335. https://doi.org/10.1016/j.ymeth.2010.01.009

Blencowe BJ. 2017. The Relationship between Alternative Splicing and Proteomic
Complexity. *Trends in Biochemical Sciences*. 42(6):407–408.
https://doi.org/10.1016/j.tibs.2017.04.001

Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: A flexible trimmer for Illumina
sequence data. *Bioinformatics*. 30(15):2114–2120.
https://doi.org/10.1093/bioinformatics/btu170

Bradaschia-Correa V, Leclerc K, Josephson AM, Lee S, Palma L, Litwa HP, Neibart SS,
Huo JC, Leucht P. 2019. Hox gene expression determines cell fate of adult periosteal
stem/progenitor cells. *Scientific Reports*. 9(1):1–13.
https://doi.org/10.1038/s41598-019-41639-7

Brawand D, Soumillon M, Necsulea A, Julien P, Csárdi G, Harrigan P, Weier M, Liechti
A, Aximu-Petri A, Kircher M, et al. 2011. The evolution of gene expression levels in
mammalian organs. *Nature*. 478(7369):343–348. https://doi.org/10.1038/nature10532

Bush SJ, Chen L, Tovar-Corona JM, Urrutia AO. 2017. Alternative splicing and the
evolution of phenotypic novelty. *Philosophical Transactions of the Royal Society B:*
Biological Sciences. 372(1713):1–7. https://doi.org/10.1098/rstb.2015.0474

Cantù C, Felker A, Zimmerli D, Prummel KD, Cabello EM, Chiavacci E,
Méndez-Acevedo KM, Kirchgeorg L, Burger S, Ripoll J, et al. 2018. Mutations in Bcl9
and Pygo genes cause congenital heart defects by tissue-specific perturbation of Wnt/β
-catenin signaling. *Genes and Development*. 32(21-22):1443–1458.
https://doi.org/10.1101/gad.315531.118

Cantù C, Pagella P, Shajiei TD, Zimmerli D, Valenta T, Hausmann G, Basler K, Mitsiadis
TA. 2017. A cytoplasmic role of Wnt/b-catenin transcriptional cofactors Bcl9, Bcl9l, and
Pygopus in tooth enamel formation. *Science Signaling*. 10(465):1–11.
https://doi.org/10.1126/scisignal.aah4598

Cardoso-Moreira M, Halbert J, Valloton D, Velten B, Chen C, Shao Y, Liechti A,
Ascenção K, Rummel C, Ovchinnikova S, et al. 2019. Gene expression across
mammalian organ development. *Nature*. 571(7766):505–509.
https://doi.org/10.1038/s41586-019-1338-5

Carruthers M, Edgley DE, Saxon AD, Gabagambi NP, Shechonge A, Miska EA, Durbin
R, Bridle JR, Turner GF, Genner MJ. 2022. Ecological Speciation Promoted by
Divergent Regulation of Functional Genes Within African Cichlid Fishes. *Molecular*
Biology and Evolution. 39(11):1–42. https://doi.org/10.1093/molbev/msac251

Cho K, Lee M, Gu D, Munoz WA, Ji H, Kloc M, Mccrea PD. 2011. Kazrin, and its binding
partners ARVCF- and delta-catenin, are required for Xenopus laevis craniofacial

development. *Developmental Dynamics*. 240(12):2601–2612.
https://doi.org/10.1002/dvdy.22721

Ciampa EJ, Welch RC, Vanoye CG, George AL. 2011. KCNE4 Juxtamembrane Region
Is Required for Interaction with Calmodulin and for Functional Suppression of KCNQ1.
Journal of Biological Chemistry. 286(6):4141–4149.
https://doi.org/10.1074/jbc.M110.158865

Conith AJ, Albertson RC. 2021. The cichlid oral and pharyngeal jaws are evolutionarily
and genetically coupled. *Nature Communications*. 12(1):1–11.
https://doi.org/10.1038/s41467-021-25755-5

Conte MA, Gammerdinger WJ, Bartie KL, Penman DJ, Kocher TD. 2017. A high quality
assembly of the Nile Tilapia (Oreochromis niloticus) genome reveals the structure of two
sex determination regions. *BMC Genomics*. 18(1):1–19.
https://doi.org/10.1186/s12864-017-3723-5

Danecek P, Bonfield JK, Liddle J, Marshall J, Ohan V, Pollard MO, Whitwham A, Keane
T, McCarthy SA, Davies RM, Li H. 2021. Twelve years of SAMtools and BCFtools.
GigaScience. 10(2):1–4. https://doi.org/10.1093/gigascience/giab008

De-Kayne R, Selz OM, Marques DA, Frei D, Seehausen O, Feulner, PGD. 2022.
Genomic architecture of adaptive radiation and hybridization in Alpine whitefish. *Nature
Communications*. 13(1):4479. https://doi.org/10.1038/s41467-022-32181-8

Dobin A, Davis CA, Schlesinger F, Drenkow J, Zaleski C, Jha S, Batut P, Chaisson M,
Gingeras TR. 2013. STAR: Ultrafast universal RNA-seq aligner. *Bioinformatics*.
29(1):15–21. https://doi.org/10.1093/bioinformatics/bts635

Durinck S, Moreau Y, Kasprzyk A, Davis S, De Moor B, Brazma A, Huber W. 2005.
BioMart and Bioconductor: A powerful link between biological databases and microarray
data analysis. *Bioinformatics*. 21(16):3439–3440.
https://doi.org/10.1093/bioinformatics/bti525

Durinck S, Spellman PT, Birney E, Huber W. 2009. Mapping identifiers for the
integration of genomic datasets with the R/Bioconductor package biomaRt. *Nature
Protocols*. 4(8):1184–1191. https://doi.org/10.1038/nprot.2009.97

El Taher A, Böhne A, Boileau N, Ronco F, Indermaur A, Widmer L, Salzburger W. 2020.
Gene expression dynamics during rapid organismal diversification in African cichlid
fishes. *Nature Ecology & Evolution*. https://doi.org/10.1038/s41559-020-01354-3

FastQC. 2015. https://qubeshub.org/resources/fastqc

Fryer G, Iles TD. 1972. The Cichlid Fishes of the Great Lakes of Africa: Their Biology
and Evolution. Oliver; Boyd, Edinburgh.

Fujimura K, Okada N. 2007. Development of the embryo, larva and early juvenile of Nile
tilapia *Oreochromis niloticus* (Pisces: Cichlidae). Developmental staging system.

Development Growth and Differentiation. 49(4):301–324.
https://doi.org/10.1111/j.1440-169X.2007.00926.x

Fujimura K, Okada N. 2008. Shaping of the lower jaw bone during growth of Nile tilapia *Oreochromis niloticus* and a Lake Victoria cichlid Haplochromis chilotes: A geometric morphometric approach. *Development Growth and Differentiation*. 50(8):653–663.
https://doi.org/10.1111/j.1440-169X.2008.01063.x

Gillespie RG, Bennett GM, De Meester L, Feder J L, Fleischer R C, Harmon L J, Hendry AP, Knope ML, Mallet J, Martin C, Parent CE, Patton AH, Pfennig KS, Rubinoff D, Schluter D, Seehausen O, Shaw KL, Stacy E, Stervander M, Stroud JT, Wagner C, Wogan GO. 2020. Comparing Adaptive Radiations Across Space, Time, and Taxa. *Journal of Heredity*. 111(1):1–20. https://doi.org/10.1093/jhered/esz064

Glaser RL, Jiang W, Boyadjiev SA, Tran AK, Zachary AA, Van Maldergem L, Johnson D, Walsh S, Oldridge M, Wall SA, et al. 2000. Paternal origin of FGFR2 mutations in sporadic cases of Crouzon syndrome and Pfeiffer syndrome. American Journal of *Human Genetics*. 66(3):768–777. https://doi.org/10.1086/302831

Gunter HM, Koppermann C, Meyer A. 2014. Revisiting de Beer's textbook example of heterochrony and jaw elongation in fish: Calmodulin expression reflects heterochronic growth, and underlies morphological innovation in the jaws of belonoid fishes. *EvoDevo*. 5(1):1–13. https://doi.org/10.1186/2041-9139-5-8

He S, Liang XF, Li L, Sun J, Wen ZY, Cheng XY, Li AX, Cai WJ, He YH, Wang YP, Tao YX, Yuan XC. 2015. Transcriptome analysis of food habit transition from carnivory to herbivory in a typical vertebrate herbivore, grass carp *Ctenopharyngodon idella*. *BMC Genomics*. 16(1):1–14. https://doi.org/10.1186/s12864-015-1217-x

Hiramatsu K, Asaba Y, Takeshita S, Nimura Y, Tatsumi S, Katagiri N, Niida S, Nakajima T, Tanaka S, Ito M, et al. 2007. Overexpression of γ-Glutamyltransferase in Transgenic Mice Accelerates Bone Resorption and Causes Osteoporosis. *Endocrinology*. 148(6):2708–2715. https://doi.org/10.1210/en.2007-0215

Hu BS, Xiong SM, Li G, Li JP. 2016. Downregulation of SLC5A8 inhibits hepatocellular carcinoma progression through regulation of Wnt/β-catenin signaling. *Tumor Biology*. 37(10):13445–13453. https://doi.org/10.1007/s13277-016-5170-3

Hulsey CD, Fraser GJ, Meyer A. 2016. Biting into the genome to phenome map: Developmental genetic modularity of cichlid fish dentitions. *Integrative and Comparative Biology*. 56(3):373–388. https://doi.org/10.1093/icb/icw059

Hulsey CD, García de León FJ, Rodiles-Hernández R. 2006. Micro- and macroevolutionary decoupling of cichlid jaws: a test of Liem's key innovation hypothesis. *Evolution*. 60(10):2096–2109.
https://doi.org/10.1111/j.0014-3820.2006.tb01847.x

Jacobs A, Elmer KR. 2021. Alternative splicing and gene expression play contrasting
roles in the parallel phenotypic evolution of a salmonid fish. *Molecular Ecology*.
30(20):4955-4969. https://doi.org/10.1111/mec.15817

Jakšić AM, Schlötterer C. 2016. The interplay of temperature and genotype on patterns
of alternative splicing in Drosophila melanogaster. Genetics. 204(1):315–325.
https://doi.org/10.1534/genetics.116.192310

Johnson TC, Kelts K, Odada E. 2000. The Holocene history of Lake Victoria. *Ambio*.
29(1):2–11. https://doi.org/10.1579/0044-7447-29.1.2

Kamel G, Hoyos T, Rochard L, Dougherty M, Kong Y, Tse W, Shubinets V, Grimaldi M,
Liao EC. 2013. Requirement for frzb and fzd7a in cranial neural crest convergence and
extension mechanisms during zebrafish palate and jaw morphogenesis. *Developmental
Biology*. 381(2):423–433. https://doi.org/10.1016/j.ydbio.2013.06.012

Keren H, Lev-Maor G, Ast G. 2010. Alternative splicing and evolution: Diversification,
exon definition and function. *Nature Reviews Genetics*. 11(5):345–355.
https://doi.org/10.1038/nrg2776

Khor JM, Ettensohn CA. 2020. Transcription Factors of the Alx Family: Evolutionarily
Conserved Regulators of Deuterostome Skeletogenesis. *Frontiers in Genetics*. 11.
https://doi.org/10.3389/fgene.2020.569314

Kim E, Magen A, Ast G. 2007. Different levels of alternative splicing among eukaryotes.
Nucleic Acids Research. 35(1):125–131. https://doi.org/10.1093/nar/gkl924

King M-C, Wilson AC. 1975. Evolution at Two Levels in Humans and Chimpanzees.
Science. 188(4184):107–116.

Kizil C, Küchler B, Yan JJ, Özhan G, Moro E, Argenton F, Brand M, Weidinger G, Antos
CL. 2014. Simplet/Fam53b is required for Wnt signal transduction by regulating β
-catenin nuclear localization. *Development*. 141(18):3529–3539.
https://doi.org/10.1242/dev.108415

Kocher TD. 2004. Adaptive evolution and explosive speciation: The cichlid fish model.
Nature Reviews Genetics. 5(4):288–298. https://doi.org/10.1038/nrg1316

Konings A. 1990. Konings's book of cichlids and all the other fishes of lake malawi.
T.F.H. Publications.

Kuratani S. 2004. Evolution of the vertebrate jaw: Comparative embryology and
molecular developmental biology reveal the factors behind evolutionary novelty. *Journal
of Anatomy*. 205(5):335–347. https://doi.org/10.1111/j.0021-8782.2004.00345.x

Lamichhaney S, Berglund J, Almén MS, Maqbool K, Grabherr M, Martinez-Barrio A,
Promerová M, Rubin CJ, Wang C, Zamani N, Grant BR, Grant PR, Webster MT,
Andersson L. 2015. Evolution of Darwin's finches and their beaks revealed by genome
sequencing. *Nature*. 518(7539):371–375. https://doi.org/10.1038/nature14181

Liem KF. 1973. Evolutionary Strategies and Morphological Innovations: Cichlid Pharyngeal Jaws. Systematic Zoology. 22(4):425. https://doi.org/10.2307/2412950

Love MI, Huber W, Anders S. 2014. Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2. *Genome Biology.* 15(12):1–21. https://doi.org/10.1186/s13059-014-0550-8

Mack KL, Nachman MW. 2017. Gene Regulation and Speciation. *Trends in Genetics*. 33(1):68–80. https://doi.org/10.1016/j.tig.2016.11.003

Mantila Roosa SM, Liu Y, Turner CH. 2011. Gene expression patterns in bone following mechanical loading. *Journal of Bone and Mineral Research*. 26(1):100–112. https://doi.org/10.1002/jbmr.193

McGee MD, Borstein SR, Meier JI, Marques DA, Mwaiko S, Taabu A, Kishe MA, O'Meara B, Bruggmann R, Excoffier L, Seehausen O. 2020. The ecological and genomic basis of explosive adaptive radiation. *Nature*. 586(7827):75–79. https://doi.org/10.1038/s41586-020-2652-7

McGonnell IM, Graham A, Richardson J, Fish JL, Depew MJ, Dee CT, Holland PWH, Takahashi T. 2011. Evolution of the Alx homeobox gene family: Parallel retention and independent loss of the vertebrate Alx3 gene. *Evolution and Development*. 13(4), 343–351. https://doi.org/10.1111/j.1525-142X.2011.00489.x

Meier, JI, McGee MD., Marques DA., Mwaiko S, Kishe M, Wandera S, Neumann D, Mrosso H, Chapman LJ, Chapman CA, Kaufman L, Taabu-Munyaho A, Wagner CE, Bruggmann R, Excoffier L, Seehausen O. 2023. Cycles of fusion and fission enabled rapid parallel adaptive radiations in African cichlids. *Science*. 381(6665):eade2833. https://doi.org/10.1126/science.ade2833

Merkin J, Russell C, Chen P, Burge CB. 2012. Evolutionary Dynamics of Gene and Isoform Regulation in Mammalian Tissues. *Science*. 338(6114):1593–1599. https://doi.org/10.1126/science.1228186

Mitchell JM, Sucharov J, Pulvino AT, Brooks EP, Gillen AE, Nichols JT. 2021. The alx3 gene shapes the zebrafish neurocranium by regulating frontonasal neural crest cell differentiation timing. *Development*. 148(7). https://doi.org/10.1242/dev.197483

Muschick M, Indermaur A, Salzburger W. 2012. Convergent Evolution within an Adaptive Radiation of Cichlid Fishes. *Current Biology*. 22(24):2362–2368. https://doi.org/10.1016/j.cub.2012.10.048

Ngoepe N, Muschick M, Kishe MA, Mwaiko S, Temoltzin-Loranca Y, King L, Courtney Mustaphi C, Heiri O, Wienhues G, Vogel H, Cuenca-Cambronero M, Tinner W, Grosjean M, Matthews B, Seehausen O. 2023. A continuous fish fossil record reveals key insights into adaptive radiation. *Nature*. 622(7982):315–320. https://doi.org/10.1038/s41586-023-06603-6

Nilsen TW, Graveley BR. 2010. Expansion of the eukaryotic proteome by alternative splicing. *Nature*. 463(7280):457–463. https://doi.org/10.1038/nature08909

Nystedt J, Anderson H, Hirvonen T, Impola U, Jaatinen T, Heiskanen A, Blomqvist M, Satomaa T, Natunen J, Saarinen J, et al. 2009. Human CMP- N -Acetylneuraminic Acid Hydroxylase (CMAH) is a Novel Stem Cell Marker Linked to Stem Cell-Specific Mechanisms. *Stem Cells*. https://doi.org/10.1002/stem.250

Oh J, Kim JY, Kim HS, Oh JC, Cheon YH, Park J, Yoon KH, Lee MS, Youn BS. 2015. Progranulin and a five transmembrane domain-containing receptor-like gene are the key components in receptor activator of nuclear factor κB (RANK)-dependent formation of multinucleated osteoclasts. *Journal of Biological Chemistry*. 290(4):2042–2052. https://doi.org/10.1074/jbc.M114.608786

Oijen MJP van, Witte F. 1996. Taxonomical and ecological description of a species complex of zooplanktivorous and insectivorous cichlids from Lake Victoria. *Zoologische Verhandelingen*. 302(1):1- 56.

Parsons KJ, Albertson RC. 2009. Roles for Bmp4 and CaM1 in Shaping the Jaw: Evo-Devo and Beyond. *Annual Review of Genetics*. 43(1):369–388. https://doi.org/10.1146/annurev-genet-102808-114917

Parsons KJ, Cooper WJ, Albertson RC. 2011. Modularity of the Oral Jaws Is Linked to Repeated Changes in the Craniofacial Shape of African Cichlids. *International Journal of Evolutionary Biology*. 2011, 1–10. https://doi.org/10.4061/2011/641501

Parsons KJ, Trent Taylor A, Powder KE, Albertson RC. 2014. Wnt signalling underlies the evolution of new phenotypes and craniofacial variability in Lake Malawi cichlids. *Nature Communications*. 5(1):3629. https://doi.org/10.1038/ncomms4629

Pavey SA, Collin H, Nosil P, Rogers SM. 2010. The role of gene expression in ecological speciation. *Annals of the New York Academy of Sciences*. 1206:110–129. https://doi.org/10.1111/j.1749-6632.2010.05765.x

Pertea G, Pertea M. 2020. GFF Utilities: GffRead and GffCompare. F1000Research. 9:1–19. https://doi.org/10.12688/f1000research.23297.2

Pertea M, Pertea GM, Antonescu CM, Chang T-C, Mendell JT, Salzberg SL. 2015. StringTie enables improved reconstruction of a transcriptome from RNA-seq reads. *Nature Biotechnology*. 33:290. https://doi.org/10.1038/nbt.3122 http://10.0.4.14/nbt.3122

Pfaffl MW. 2001. A new mathematical model for relative quantification in real-time RT-PCR. *Nucleic Acids Research*. 29(9):45e–45. https://doi.org/10.1093/nar/29.9.e45

Picard toolkit. 2019. Broad Institute, GitHub repository.

R Core Team. 2020. R: A language and environment for statistical computing. Vienna, Austria: R Foundation for Statistical Computing. https://www.R-project.org/

Reardon W, Winter RM, Rutland P, Pulleyn LJ, Jones BM, Malcolm S. 1994. Mutations in the fibroblast growth factor receptor 2 gene cause Crouzon syndrome. *Nature Genetics*. 8(1):98–103. https://doi.org/10.1038/ng0994-98

Richards EJ, McGirr JA, Wang JR, St. John ME, Poelstra JW, Solano MJ, O'Connell DC, Turner BJ, Martin CH. 2021. A vertebrate adaptive radiation is assembled from an ancient and disjunct spatiotemporal landscape. *Proceedings of the National Academy of Sciences*. 118(20):e2011811118. https://doi.org/10.1073/pnas.2011811118

Roberts RB, Hu Y, Albertson RC, Kocher TD. 2011. Craniofacial divergence and ongoing adaptation via the hedgehog pathway. *Proceedings of the National Academy of Sciences of the United States of America*. 108(32):13194–13199. https://doi.org/10.1073/pnas.1018456108

Rochard L, Monica SD, Ling ITC, Kong Y, Roberson S, Harland R, Halpern M, Liao EC. 2016. Roles of Wnt pathway genes wls, wnt9a, wnt5b, frzb and gpc4 in regulating convergent-extension during zebrafish palate morphogenesis. *Development*. 143(14):2541–2547. https://doi.org/10.1242/dev.137000

Rodríguez-Ramírez CE, Hiltbrunner M, Saladin V, Walker S, Urrutia A, Peichel CL. 2023. Molecular mechanisms of Eda -mediated adaptation to freshwater in threespine stickleback. *Molecular Ecology*. 1–19. https://doi.org/10.1111/mec.16989

RStudio Team. 2019. RStudio: Integrated development environment for r . Boston, MA: RStudio, Inc. http://www.rstudio.com/

Rux DR, Wellik DM. 2017. Hox genes in the adult skeleton: Novel functions beyond embryonic development. *Developmental Dynamics*. 246(4):310–317. https://doi.org/10.1002/dvdy.24482

Sánchez-Hernández J, Nunn AD, Adams CE, Amundsen PA. 2019. Causes and consequences of ontogenetic dietary shifts: a global synthesis using fish models. *Biological Reviews*. 94(2):539–554. https://doi.org/10.1111/brv.12468

Schmitz J, Brosius J. 2011. Exonization of transposed elements: A challenge and opportunity for evolution. *Biochimie*, 93(11):1928–1934. https://doi.org/10.1016/j.biochi.2011.07.014

Seehausen O. 1996. Lake Victoria rock cichlids: taxonomy, ecology, and distribution. Verduyn Cichlids, Germany.

Seehausen O. 2002. Patterns in fish radiation are compatible with Pleistocene desiccation of Lake Victoria and 14 600 year history for its cichlid species flock. *Proceedings of the Royal Society B: Biological Sciences*. 269(1490):491–497. https://doi.org/10.1098/rspb.2001.1906

Seehausen O, Lippitsch E, Bouton N, Zwennes H. 1998. Mbipi, the rock-dwelling cichlids of Lake Victoria: description of three new genera and fifteen new species (Teleostei). *Ichthyological Exploration of Freshwaters*. 9(2):129–228.

Sevilla LM, Nachat R, Groot KR, Watt FM. 2008. Kazrin regulates keratinocyte cytoskeletal networks, intercellular junctions and differentiation. *Journal of Cell Science*, 121(21):3561–3569. https://doi.org/10.1242/jcs.029538

Singh P, Ahi EP, Sturmbauer C. 2021. Gene coexpression networks reveal molecular interactions underlying cichlid jaw modularity. *BMC Ecology and Evolution*. 21(1):62. https://doi.org/10.1186/s12862-021-01787-9

Singh P, Irisarri I, Dowdall JT, Thallinger GG, Svarda H, Lemmon EM, Lemmon AR, Koblmüller S, Meyer A, Sturmbauer C. 2022. Phylogenomics of trophically diverse cichlids disentangles processes driving adaptive radiation and repeated trophic transitions. Ecology and Evolution. 12(e9077):1–15. https://doi.org/10.1002/ece3.9077

Singh P, Ahi EP. 2022. The importance of alternative splicing in adaptive evolution. *Molecular Ecology*. https://doi.org/10.1111/mec.16377

Singh P, Börger C, More H, Sturmbauer C. 2017. The role of alternative splicing and differential gene expression in cichlid adaptive radiation. *Genome Biology and Evolution*. 9(10):2764–2781. https://doi.org/10.1093/gbe/evx204

Smith CCR, Rieseberg LH, Hulke BS, Kane NC. 2021. Aberrant RNA splicing due to genetic incompatibilities in sunflower hybrids. *Evolution*. 75(11):2747-2758. https://doi.org/10.1111/evo.14360

Smith CCR, Tittes S, Mendieta JP, Collier-zans E, Rowe HC, Rieseberg LH, Kane NC. 2018. Genetics of alternative splicing evolution during sunflower domestication. Proceedings of the National Academy of Sciences. 115(26):6768–6773. https://doi.org/10.1073/pnas.1803361115

Sorek R. 2007. The birth of new exons: Mechanisms and evolutionary consequences. *Rna*. 13(10):1603–1608. https://doi.org/10.1261/rna.682507

Steward RA, Jong MA de, Oostra V, Wheat CW. 2022. Alternative splicing in seasonal plasticity and the potential for adaptation to environmental change. *Nature Communications*. 13(1):1–12. https://doi.org/10.1038/s41467-022-28306-8

Tarkkonen K, Hieta R, Kytölä V, Nykter M, Kiviranta R. 2017. Comparative analysis of osteoblast gene expression profiles and Runx2 genomic occupancy of mouse and human osteoblasts in vitro. *Gene*. 626:119–131. https://doi.org/10.1016/j.gene.2017.05.028

Ten Brink H, Seehausen, O. 2022. Competition among small individuals hinders adaptive radiation despite ecological opportunity. *Proceedings of the Royal Society B: Biological Sciences*, 289(1971). https://doi.org/10.1098/rspb.2021.2655

Tress ML, Abascal F, Valencia A. 2017. Alternative Splicing May Not Be the Key to Proteome Complexity. *Trends in Biochemical Sciences*. 42(2):98–110. https://doi.org/10.1016/j.tibs.2016.08.008

Trincado JL, Entizne JC, Hysenaj G, Singh B, Skalic M, Elliott DJ, Eyras E. 2018. SUPPA2: Fast, accurate, and uncertainty-aware differential splicing analysis across multiple conditions. *Genome Biology*. 19(1):1–11. https://doi.org/10.1186/s13059-018-1417-1

Wagner GP, Pavlicev M, Cheverud JM. 2007. The road to modularity. *Nature Reviews Genetics*. 8(12):921–931. https://doi.org/10.1038/nrg2267

Witte F, Barel CDN, Hoogerhoud RJC. 1982. The Ecological Differentiation of Two Closely Resembling Haplochromis Species From Lake Victoria (H. Iris and H. Hiatus ; Pisces, Cichlidae). Netherlands Journal of Zoology. 33(3):283–305. https://doi.org/10.1163/002829683X00147

Wright CJ, Smith CWJ, Jiggins CD. 2022. Alternative splicing as a source of phenotypic diversity. *Nature Reviews Genetics*. https://doi.org/10.1038/s41576-022-00514-4

Xiong KM, Peterson RE, Heideman W. 2008. Aryl hydrocarbon receptor-mediated down-regulation of Sox9b causes jaw malformation in zebrafish embryos. *Molecular Pharmacology*. 74(6):1544–1553. https://doi.org/10.1124/mol.108.050435

Yan YL, Willoughby J, Liu D, Crump JG, Wilson C, Miller CT, Singer A, Kimmel C, Westerfield M, Postlethwait JH. 2005. A pair of Sox: Distinct and overlapping functions of zebrafish sox9 co-orthologs in craniofacial and pectoral fin development. *Development*. 132(5):1069–1083. https://doi.org/10.1242/dev.01674

Zerbino DR, Achuthan P, Akanni W, Amode MR, Barrell D, Bhai J, Billis K, Cummins C, Gall A, Girón CG, et al. 2018. Ensembl 2018. *Nucleic Acids Research*. 46(D1):D754–D761. https://doi.org/10.1093/nar/gkx1098