Comprehensive benchmark of integrative strategies for

analyzing microbiome-metabolome relationships

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Abstract

Background
The exponential growth of high-throughput sequencing technologies was an incredible
opportunity for researchers to combine different -omics within computational frameworks. In
particular metagenomics and metabolomics data have gained an increasing interest due to
their implication in many complex diseases. However, currently, no standard seems to
emerge for jointly integrating both microbiome and metabolome datasets within statistical
models.
Results
Thus, in this paper we comprehensively benchmarked fifteen different integrative methods to
link microorganisms and metabolites. Methods evaluated in this paper cover most of the
researcher's goals such as global associations, data summarization, individual associations
and feature selection. Through an extensive simulation study and an application to real gut
microbial datasets, we highlighted the best approaches for unraveling complementary
biological processes involved between the two omics. We provided general guidelines for
practitioners depending on the scientific question and the data at-hand.
Conclusion
In summary, we argue that this paper constitutes a promising avenue for establishing
research standards when mutually analyzing metagenomics and metabolomics data, while
providing foundations for future methodological developments.
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Background

The recent development of high-throughput sequencing technologies has permitted the generation of omics data at an exponential scale. Combining different high dimensional biological datasets within computational models represents a wonderful opportunity for researchers to better understand the underlying biological mechanisms involved in diseases [1]. In particular, the microorganism-metabolite interactions have gained an increasing interest due to their potential involvement in a large set of traits. It has been demonstrated that shifts in the microbiome-metabolome interactions have important implications on individual health [2, 3]. Indeed, recent studies for cardio-metabolic diseases [4] or autism spectrum disorders [5] have shown that pathoetiology could be explained by a complex interplay between microbes and host metabolites [6] or by disruptions in the microbiota-derived metabolite processes [7]. Thus, efficiently incorporating microbiome and metabolome data within statistical frameworks offers critical insights on the complex relationships occurring between diet or lifestyle factors on the microbe-metabolite recomposition and remains an important challenge in order to adequately identify hence target biological pathways [8]. However, the tremendous amount of available statistical models makes the choice of the right method a daunting task for many researchers.

The statistical joint integration of microbiome and metabolome data can be achieved with different integrative strategies. Standard workflows include various types of analysis, each addressing a specific biological question [2]. Briefly, traditional pipelines include the detection of global associations, data summarization, individual associations and identification of core features. Firstly, researchers are often interested in determining whether a global association is occurring between the two omics. For example, one can look for a global change in metabolome levels due to a microbial recomposition induced by a specific diet or lifestyle [2]. Consistent with recent reports, testing for global associations can be performed using multivariate methods such as the Mantel test [9] or the multivariate microbiome regression-based kernel association test (MMiRKAT) [10]. This step frequently

precedes the application of subsequent analyses such as data summarization methods or

the identification of core features [2]. Then, following approaches used for single omics, a

common research objective is to summarize information contents in the two omics,

facilitating the visualization and interpretation of large scale biological data [1]. The presence

of two types of omics allows the exploitation of the intra- and inter- correlation existing

between features of the two datasets. Application of data summarization methods including

Canonical Correlation Analysis (CCA) [11], Partial Least Square (PLS) [12], Redundancy

Analysis (RDA) [13] or more recently Multi-Omics Factor Analysis (MOFA2) [14] is an

important step in order to uncover features explaining a large proportion of data variability.

Indeed, applications of data summarization methods have allowed the identification of

taxonomic groups or metabolites involved in Type 2 diabetes [15]. However, both global

association and data summarization methods fail to provide individual relationships between

one or several microorganisms and metabolites. This aspect remains central to highlight core

features involved in a particular biological context. As an illustration, methods for detecting

individual associations may prove relevant for the identification of bacterial genus associated

with dietary-impacted metabolites [2]. One strategy is to compute a measure of association

between each metabolite-microbiota pair, using either a correlation or a regression model.

Although easily implementable and interpretable, these approaches suffer from lack of power

induced by the number of models fitted, limiting result transferability. An alternative way is to

employ univariate or multivariate feature selection methods to adequately identify key actors

at a large scale. The least absolute shrinkage and selection operator (LASSO) is a method

initially developed to improve predictability while proceeding to feature selection [16]. Indeed,

the LASSO is able to set coefficients to zero, hence facilitating identification of core features.

Consistently with this idea, sparse CCA (sCCA) [17] or sparse Partial Least Square (sPLS)

[18] are multivariate penalized methods summarizing data variability while proceeding to

feature selection. However, due to the complex structure of both microbiome and

metabolome data, standard methods fall short of providing unbiased associations, limiting

the biological interpretation of results.

On the one hand, because of the sequencing technology, metagenomics data

highlight hard-to-analyze characteristics [19, 20]. Indeed, it is now globally accepted that

microbiome datasets are over-dispersed, zero-inflated, highly correlated, and compositional.

Without adequate transformation the inherent compositionality of the data makes the

application of standard methods incorrect, leading to inconsistent results [19–21]. On the

other hand, metabolomics data shares some of these features, exhibiting over-dispersion

and high correlation structures [21]. Thus, combining these two omics together within

statistical frameworks requires particular attention. Approaches to deal with compositional

data either as an outcome or explanatory variable have already been proposed [20, 22, 23],

covering applications of global association methods, data summarization, individual

associations or identification of core features. Conventional strategies include utilization of

standard methods after suitable data transformations or purely compositional approaches

[24–27]. Subsequently, determining which strategy is the best depending on the research

question remains an open problem with major implications for practitioners.

Despite recent efforts to integrate microbiome and metabolome within unified tools

[28], to our knowledge there is no systematic framework to evaluate integrative methods to

link microbiome with metabolome datasets; constantly pushing researchers to make their

choice without any robust comparison. Thus, in this paper, we comprehensively

benchmarked fifteen different integrative methods to link microorganisms and metabolites,

covering most of the researcher's aims, such as global associations, data summarization,

individual associations, or feature selection (Figure 1). Our extensive simulation studies

provide insightful lessons on the strengths and limits of methods commonly encountered in

practice. Then, we applied best methods to real data on the gut microbiome and metabolome

for Konzo disease [29], highlighting a complex interplay between the two omics occurring at

different scales. Finally, we provide general guidelines and avenues for future

methodological developments, depending on the data at-hand and the research aims.

Results

SIMULATION SETUP AND BENCHMARKED METHODS

Taking advantage of the "Normal to Anything" (NORtA) framework, we generated synthetic

microbiome and metabolome datasets mimicking complex data structures and relationships

(See Methods). We produced two simulation settings, a low dimensional and a high

dimensional setting, both representing different scenarios commonly encountered in practice

(Figure 1A). We therefore compared fifteen integrative methods depending on the research

question (Figure 1B). Methods were presented as follows. Firstly, in the *global associations*

subsection we compared the Mantel test and MMiRKAT with respect to the Type-I error rate

and power. Then, in the *data summarization* subsection we evaluated four different models

including CCA, PLS, RDA and MOFA2, regarding their capability to recapitulate data

variability across latent factors. Third, in the *individual associations* subsection we compared

three strategies for performing regression-based approaches between compositional

covariates and metabolites, the clr-linear model, the log-contrast and MiRKAT, respectively.

Approaches were evaluated based on the Type-I error rate and power. Fourth, in

subsections *univariate feature-selection for compositional predictors*, *univariate feature-

selection for compositional outcomes* and *multivariate feature-selection* we compared

approaches for identifying core microbes and metabolites, leveraging both univariate and

multivariate feature selection strategies. For univariate frameworks, depending on the nature

of the response, several models were considered. Indeed, when microorganisms are the

explanatory variables, we compared three approaches, the clr-LASSO, the clr-MLASSO and

CODA-LASSO [23]. Consistently, when microorganisms are the response variables, we

evaluated the LASSO, MLASSO, and the sparse Dirichlet regression [27]. Nonetheless, for

multivariate feature selection models, we considered sCCA and sPLS. Approaches were

evaluated based on sparsity and reliability. Details on the methods and their related

performance metrics were provided in the Methods section. Finally, in order to highlight

complementary biological insights provided by methods, best approaches were illustrated in

the *real-data application* subsection, exploiting metagenomics and metabolomics data from

Konzo disease.

GLOBAL ASSOCIATIONS

A common question in practice for researchers is to find global associations between two

omics datasets [2]. Thus, we compared two multivariate methods detecting associations

occurring at the global level between microbiome and metabolome, the Mantel test [9] and

MMiRKAT [10], respectively. Since these two methods provide frequentist statistical

frameworks i.e., p-values, we systematically evaluated their performance with respect to

Type-I error rate control and power (See Methods). Firstly, when applying on the ILR

transformed microbiome data, the Mantel test provides a good control of Type-I error rate in

the high dimensional scenario while MMiRKAT highlights a slightly more conservative

behavior (Figures 2A-2B). Secondly, MMiRKAT exhibits strikingly higher power than the

Mantel test under our high dimensional scenario. Indeed, at the 0.05 significance threshold

MMiRKAT reaches on average 97% of power against 22% for the Mantel test (Figures 2C).

This difference is however mitigated in the low dimensional setting, where the two methods

exhibit comparable performances (Figures S1-S2) . Importantly, the distance kernel choice

seems to strongly impact the Mantel test power, from single to double, while MMiRKAT

power remains stable across data transformations (Figure 2C). These results were confirmed

in our low dimensional scenario and considering different data normalizations (Figures S3-

S15). Interestingly, when the Mantel test was considered, the Canberra distance exhibits the

lowest powers, while no clear distinction could be observed between Euclidean and

Manhattan distance kernels (Figure 2C). This result suggests the Canberra distance as the

poorest choice when using the Mantel test. Collectively, our results suggest comparable

performance for the two methods under the low dimensional setting regarding both Type-I

error rate and power. However, in the high dimensional scenario MMiRKAT is the most

powerful method to find global associations. In addition the method is robust to data

transformation and distance kernels.

DATA SUMMARIZATION

Instead of measuring one global association, one can be interested in recapitulating

information contained within the two datasets through latent factors, accounting for the

between- within-correlation [30] . Thus, we compared Canonical Correlation Analysis (CCA)

[11], Regression PLS (PLS-Reg) [12], Canonical PLS (PLS-Can) [12], Redundancy Analysis

(RDA) [13], and Multi-Omic Factor Analysis (MOFA2) [14] in our two scenarios with respect

to their capability to summarize explained variance through their components (See Methods).

Generally, regardless of the considered data normalization, in our two scenarios, MOFA2

was the best method, exhibiting larger explained variances, with a modest variability

compared to PLS-Reg, PLS-Can, CCA, and RDA (Figure 2D; Figures S16-S19). Indeed,

when ILR transformed microbiome data were considered, in our high dimensional scenario,

MOFA2 exhibited an average of explained variance of 86% (sd = 1.37) compared to 44% (sd

= 4.35), 14% (sd = 2.03), 21% (sd = 2.34), and 22% (sd = 0.76) for PLS-Reg, PLS-Can, CCA

and RDA, respectively. Surprisingly, except for MOFA2 and the PLS-Reg, where the

explained variances increase (64% to 86% and 41% to 44%, respectively), all the remaining

methods exhibit a smaller explained variance in the high dimensional scenario compared to

the low dimensional setting. Aligned with this result, we investigated the behavior of each

method with respect to the number of associated features and the effect size and found

positive associations in both cases across all methods (Figures S20-S21). Importantly,

method performances may vary depending on the considered data transformation (Figure

2D; Figures S16-S19). Our results pointed to MOFA2 as the best model to summarize data

variability through latent factors. Finally, our findings suggested that the method is versatile

and robust under scenarios commonly encountered in practice.

INDIVIDUAL ASSOCIATIONS

Studying the relationship between metabolites and microorganisms may represent an

important challenge in order to account for the compositionality induced by microbiome

datasets. Indeed, the perfect correlation brought by the compositionality makes the

application of standard methods incorrect. This is particularly true when microbiota are

incorporated as covariates [19, 22]. We therefore compared three equivalent strategies in

order to study the global effect of microorganisms on one particular metabolite, the Log-

contrast model [22], MiRKAT [10] and a linear regression on the CLR transformed

microbiome (referred to as clr-lm), respectively. Methods were evaluated with respect to their

capability to adequately control false positives while maintaining a good power (See

Methods). Globally, under the null hypothesis, the three methods adequately controlled the

Type-I error rate, with the linear log-contrast model exhibiting a slightly conservative behavior

across the two scenarios (Figures 3A-3B). Then, under the alternative hypothesis, the linear

log-contrast model offers a higher power than MiRKAT or the clr-lm model, on average twice

larger across the data transformations considered in the high dimensional setting (Figure

3C). This result was also confirmed when comparing the log-contrast model to Spearman's

or Pearson's correlations, while MiRKAT or the clm-lm model do not exhibit clear advantage

(Figure S22). Indeed, at a 0.05 significance threshold, the log-contrast model offers 52% of

power against 29% for MiRKAT and clr-lm, and 29% and 21% for Pearson's and Spearman's

correlations, respectively. This result was confirmed in our low dimensional setting, where

smaller discrepancies can be observed (Figure 3C). However, consistent with results

observed for MMiRKAT, MiRKAT provided a stable power and a good control of Type-I error

rate across data normalizations (Figure S23). Importantly, when evaluating individual

association methods for compositional outcomes, we found no clear superiority of the

Dirichlet regression or the linear regression on the CLR transformed microbiome data over

Spearman's or Pearson's correlations in our low dimensional setting (Figure S24).

Collectively, our results suggest that in order to study the global impact of microorganisms on

individual metabolites, the linear log-contrast model represents the best method compared to

competitor approaches, providing higher power and a suitable control of the Type-I error

rate.

UNIVARIATE FEATURE-SELECTION FOR COMPOSITIONAL PREDICTORS

Feature selection methods have gained increasing interest from researchers in order to identify a subset of microbiota associated with a variable of interest [31]. However, due to the compositionality induced by microbiome data, traditional methods have been shown to lead to incorrect results [19]. Thus, we compared univariate feature selection methods accounting for compositional predictors, CODA-LASSO [23], clr-LASSO [23] and clr-MLASSO, respectively. Firstly, we evaluated whether methods were able to provide sparse sets of microorganisms across our two scenarios. In our low dimensional setting, CODA-LASSO highlighted sparser selections, showing average sparsities of 2% against 9% and 14% for clr-LASSO and clr-MLASSO. This result was consistent in our high dimensional setting, where CODA-LASSO showed stable sparsities, while the sparsity of clr-LASSO and clr-MLASSO greatly improves (Figures 4A-4D; CODA-LASSO=2%; clr-LASSO=5%; clr-MLASSO=11%). This result suggests that CODA-LASSO tends to provide a stable sparsity across our two scenarios, selecting only a small proportion of the total microorganism-metabolite interactions compared to the two other methods. Then, we assessed how accurate the methods are in order to find true associations. In the low dimensional scenario, clr-LASSO offered slightly higher classification performances, showing average F1-Scores of 43%, compared to 35% and 30% for CODA-LASSO and clr-MLASSO, respectively (Figure 4A). Nonetheless, in the high dimensional scenario, CODA-LASSO provided higher F1-Scores than clr-LASSO or clr-MLASSO, with accurate classification rates on average 1.40 times higher (Figure 4D). Collectively, our results point to CODA-LASSO as a good trade-off between sparsity and classification performances to accurately select sparse subset of microbiota associated with metabolites.

UNIVARIATE FEATURE-SELECTION FOR COMPOSITIONAL OUTCOMES

Finding a subset of metabolites associated with microbiota may bring important insights into the underlying biological mechanisms involved between the two omics. Thus, consistently

with the previous subsection, we systematically compared three different methods taking into

account compositional outcomes with respect to sparsity and F1-Score, the sparse Dirichlet

regression [27], LASSO and MLASSO of the CLR transformed microbiome data. Firstly, in

the low dimensional setting, the LASSO offered strikingly sparser solutions, showing sparsity

scores of 8% compared to 40% and 18% for the sparse Dirichlet regression and MLASSO,

respectively (Figure 4C). Except for the sparse Dirichlet regression, where the sparsity was

multiplied by roughly 2 between the two scenarios, LASSO and MLASSO exhibit sparser

selection in the high dimensional setting compared to the low dimensional scenario (Figure

4D). This result suggests that standard methods applied on the CLR transformed

microbiome data seems to provide sparse and consistent solutions across our scenarios.

Moreover, regardless of the scenario considered, F1-Scores remained low across methods

suggesting poor method performances to accurately classify associations between

microorganisms and metabolites (Figures 4C-4D). However, it is worth mentioning that high

F1-Scores achieved by the sparse Dirichlet regression in the low dimensional scenario may

be explained by weak sparsity scores. Taken together, our results point to poor performance

of methods to select accurately metabolites associated with microorganisms; where standard

methods applied on the CLR transformed microbiome data correspond to a better trade-off

between sparsity and classification performances than a purely compositional penalized

method.

MULTIVARIATE FEATURE-SELECTION

Instead of analyzing each feature independently, exploiting information shared across two

omics may represent an interesting avenue to select the most contributive features [32].

Thus, we compared three methods taking advantage of both intra- and inter-correlation

occurring between features of the two datasets, the regression sparse PLS, the canonical

sparse PLS [18] and the sparse CCA [17], respectively. Firstly, in our low dimensional setting

the regression sPLS seems to provide high levels of sparsity compared to the two other

methods (Figure 4C). Indeed, the method tends to select about 34% of total features

compared to 23% or 26% for sCCA or canonical sPLS. This pattern was also observed in our

high dimensional setting, even if an increase of sparsity between the two scenarios has to be

noted (Figures 4C-4F). This result aligns with a too high number of selected features, since

our simulation setup maximally assumes a 10% of associated features. Then, we

investigated whether methods were able to accurately discriminate contributive features from

uninformative ones. In our low dimensional scenario, the regression sPLS offered higher F1-

Scores, showing average values of 76% compared to 70% and 60% for the canonical sPLS

and sCCA, respectively (Figures 4C). This result was confirmed in the high dimensional

scenario, even if lower scores across the three methods have to be noted (Figures 4F). For

example, the average F1-Score for the regression sPLS decreased by 63%, while for the

canonical sPLS and sCCA, the decrease is of 53% and 69%, respectively, consistent with

lower classification performance as the dimensionality increases. Then, we investigated

whether methods are sensitive to data transformation. Interestingly, we found that in the low

dimensional scenario CLR transformation offered higher sparsity scores showing equivalent

F1-Scores across methods, while in the high dimensional setting absence of microbiome

data transformation slightly improved both sparsity and F1-Scores (Figure S25). Finally, our

results align with regression sPLS as the preferred choice for selecting features accounting

for between and within omics correlation. However, our findings point to modest levels of

sparsity across the methods suggesting poor method specificity with inconsistencies of

method results across data transformation.

REAL-DATA APPLICATION

Our systematic evaluation of strategies to jointly analyze microbiome and metabolome data

has permitted the illustration of the best methods depending on the research question. Thus,

through an application on metabolomics and metagenomics data of the Konzo disease [29],

we applied the more appropriate approaches to highlight different biological patterns

occurring between microorganisms and metabolites. We presented the exact workflow in the

Konzo data analysis section and Figure S26. Firstly, we used the Mantel test and found a

significant global association between the two omics (Spearman's permutation p-value <=

9.9e-5). Then we applied MOFA2 and found that through the fifteen first latent factors, the

model explains 50% and 40% of microbiome and metabolome variability, respectively

(Figure S27). Moreover, the top-10 most contributing features on the first factor highlighted

relevant microbiota or metabolites associated with intestinal health. For example, MOFA2

identifies the *2,3-Dihydroxy-2-methylbutanoic acid*, a fatty-acid which has been

demonstrated to be related to lipid metabolism pathways [33] (Figure 5A). Similarly,

Faecalibacterium prausnitzii was identified as the most strongly associated microbiota,

exhibiting a highly negative contribution (Figure 5B). This microbiota has already been

shown to be involved in gut health [34, 35]. Subsequently we used the sPLS regression and

were able to identify 249 metabolites and 70 microorganisms significantly contributing to the

two first components, where clear clusters of microbiota could be observed (Figure 5C).

Consistently with our benchmark, we used the log-contrast regression in order to identify

metabolites significantly impacted by microbial communities and found that out of the 249

metabolites, 193 are significantly associated with microbial communities (Bonferroni adjusted

p-values <= 2e-04). Then applying CODA-LASSO we detected 234 metabolites with at least

one interaction with microorganisms. Interestingly, every microorganism has been selected

at least once across the 234 metabolites, with an average of 35 microbiota associated

(Figure 5D). For example, the *2,3-Dihydroxy-2-methylbutanoic acid*, previously identified by

MOFA2, is associated with 8 microorganisms, mostly involved in gastrointestinal health

(Figure 5E). Finally, we checked whether microorganisms exhibit consistent effects across

metabolites and we observed 5 microbiota highlighting important variability in their effect

(Figure 5F). This result was confirmed at a larger scale by network analysis from log-contrast

regression and CODA-LASSO (Figures S28-S29). Our results from metagenomics and

metabolomics data from Konzo disease highlight complementary biological interactions

between microorganisms and metabolites, where different microbial dynamics seems to be

involved.

Discussion

The integration of microbiome and metabolome datasets within statistical frameworks has

become an important resource for researchers in order to comprehensively understand the

underlying biological mechanisms involved in diseases. Indeed, recent studies in

inflammatory bowel disease [36] or cardiometabolic traits [4] have highlighted that

pathoetiology may result in disruptions of interactions between microorganisms and host-

metabolites interplay or shifts in the microbial-derived metabolite levels. Understanding these

interactions represent therefore a critical avenue for unraveling the biology of complex

phenotypes. However, currently, there are no standards on how to integrate these two omics

together, pushing researchers to constantly reinvent the wheel. Thus, deciding which method

fits best for a specific biological question remains a daunting task, critically limiting the result

interpretations and replicability. In this paper, we extensively benchmarked fifteen existent

integrative methods to study microbiome-metabolome interactions covering most of the

researcher aims: global associations, data summarization, individual associations, and

feature selection (Figure 1). Based on a comprehensive simulation study and a real data

application, we highlighted best methods depending on the research question and data at-

hand, providing important insights about statistical good practices (Table 1) and avenues for

future methodological developments (Table 2).

When evaluating global association methods, our results have pointed to important

lessons for practitioners. Indeed, MMiRKAT represents the most promising method

compared to the Mantel test, showing higher power and robustness to data transformations

and distance kernels (Figure 2C). We argue this aspect is particularly relevant since

choosing the right data transformation or distance metric may represent an important

challenge for practitioners. Moreover, MMiRKAT has the possibility to adjust for confounding

factors which is an appealing feature for most phenotypes where bias can be induced by

certain individual characteristics, such as age, sex or lifestyle [3, 4]. However, one limitation

of MMiRKAT compared to the Mantel test is its incapability to deal with scenarios with a

larger number of features than individuals. We therefore recommend filtering out features

based on a feature selection approach or to use the Mantel test in order to have a crude idea

about the global association occurring between the two omics. Importantly, when using the

Mantel test, our results suggest that the Canberra distance on metabolome data is the

poorest choice for detecting global associations across all our scenarios (Figures 2B; Figures

S1-S15). Thus, applying Euclidean distance on transformed microbiome data while applying

Euclidean or Manhattan distances on metabolites should constitute the default usage for

most cases.

Data reduction is often used by practitioners in order to summarize information

through a small number of components. Having an efficient method which recapitulates

variability across two omics is critical for facilitating subsequent analyses such as

visualization or clustering [1]. We considered four different methods exhibiting specific

features to summarize omics information and found that in addition to being robust to data

normalization, MOFA2 is the best method to recapitulate data variability. In our high

dimensional setting MOFA2 explains about 80% of metabolome variance when ILR

normalization was considered and remains stable across alpha and CLR transformations

(Figure 2D, Figure S16). This result may be explained by the capability of the method to

capture complex relationships, as suggested by [37]. Thus, we recommend using MOFA2

when researchers want to achieve efficient data reduction. We then applied MOFA2 to our

Konzo dataset and found important microbiota and metabolites involved in biologically

relevant pathways of gut health, while preserving a great portion of data variability (Figure

5A-5B). For example, MOFA2 identifies *Faecalibacterium prausnitzii* as the most negatively

contributive microorganisms on the first factor (Figure 5B). Previous studies have shown that

Faecalibacterium prausnitzii levels are strongly associated with anti-inflammatory metabolite

quantities involved in intestinal health [34, 35]. Similarly, MOFA2 found *2,3-Dihydroxy-2-*

methylbutanoic acid with the strongest positive correlation on the first factor, a fatty-acid

which has been demonstrated to be related to lipid metabolism pathways [33] (Figure 5A).

409   In practice another important question for researchers is to determine the relationship

between microbial communities with a variable of interest [29, 38]. However, the underlying

compositional structure of microbiome data is an important challenge for model performance.

In this paper we have compared three methods accounting for the compositionality of

predictors with different strategies, a linear regression applied on the CLR transformed

microbiome data, MiRKAT, and the log-contrast model. Compared to correlations, these

methods can incorporate confounding factors which is an important feature in practice. Our

main finding is that regardless of the method considered here, better performances are

achieved compared to correlations, still widely used in practice [5]. However, the linear log-

contrast offers higher power across our simulation scenarios compared to MiRKAT and the

linear regression (Figure 3C). Also, one important advantage of the log-contrast model over

MiRKAT or the linear regression is to not require a choice of a particular data normalization,

which can represent an important challenge for most researchers. This is particularly

important since the CLR transformation has been shown to provide still-correlated features

while sub-compositionally incoherent, limiting result transferability [23, 24]. This result

highlights the need for new compositional data transformations, keeping the original number

of features while linearly independent (Table 2). Hopefully, MiRKAT performance is robust

across data transformations, with stable power and suitable Type-I error rate control (Figure

3C, Figure S23). Additionally, one main difference of the log-contrast or the linear regression

over MiRKAT is to provide individual contribution of each microbe. We therefore strongly

recommend to use the log-contrast regression when evaluating the association between

microorganisms and metabolites. Consistently, out of the 249 metabolites selected by the

regression sPLS, the log-contrast model highlights 193 metabolites with significant

associations with microbiota in the Konzo dataset. Interestingly, we found that

microorganisms exhibit heterogeneous effects across metabolites suggesting different

microbial dynamics possibly involved in the disease (Figure S28). Similarly to MDiNe [39],

where authors provided a mechanistic framework to study differential microbial co-

occurrence networks, additional work is needed to link microbiome and metabolome from a

dynamic perspective at large scale (Table 2; Ongoing work). We argue this aspect is

particularly critical in order to pinpoint the underlying biological mechanisms hence facilitating

precision medicine applications [40, 41].

Also, one important contribution of this work is to extensively evaluate feature

selection methods. This is particularly critical for researchers in order to accurately select

metabolites and microorganisms involved in a specific biological context. Our results point to

moderate performance of multivariate feature selection methods with inconsistent

performances across scenarios and the data transformations considered (Figures 4C-4F,

Figure S25). This result is also observed for univariate feature selection models with

compositional outcomes (Figures 4B-4E). The best performances are achieved for methods

with compositional predictors, with CODA-LASSO exhibiting stable sparsity results with good

classification performances (Figures 4A-4D). Thus, we recommend in practice to use CODA-

LASSO for scenarios with microbial predictors, while using the LASSO regression after CLR

transforming the microbiome data when these latter are the outcome. Then we applied both

regression sPLS and CODA-LASSO on the Konzo dataset. Regression sPLS has permitted

the detection of 249 metabolites and 70 microorganisms contributing the most to data

variability (Figure 5C). From these 249 metabolites, CODA-LASSO has subsetted the most

contributing features, highlighting different microbial dynamics of effects (Figures 5F; Figure

S29). Further investigations have shown that *Vescimonas fastidiosa* was the most interacting

microbiota, significantly connected to 138 metabolites. This result is aligned with the model

where microorganisms may be connected to a large set of metabolites. This complex

microbiome-metabolome crosstalk has been shown to be associated with diseases [6].

However, associations found may result in artifact signals since most feature selection

methods benchmarked in this paper suffer from lack of sparsity and reliability. This result is

aligned with previous reports where authors have shown poor performances of traditional

feature selection models [42]. Indeed, most penalized methods are mainly built upon cross-

validation where small perturbations in data may yield drastic changements in results.

Similarly to [42] extending sparse multivariate or univariate methods to knockoff framework

[43] or stability selection [44] should represent interesting avenues for improving both

sparsity and reliability for compositional data [45] (Table 2).

Although our simulation setup is able to realistically simulate microbiome and

metabolome data, our framework suffers from two limitations that we think it is important to

mention here. Firstly, the NORtA algorithm is limited in its capability to generate real

correlated compositional data. Indeed, as discussed by [46], simulating pure compositional

data from a Dirichlet distribution induced only a small correlation between features, which is

often unrealistic regarding the biology of the microbial communities and metabolites. We

therefore generated compositional microbiome data post-hoc, promoting the correlation,

zero-inflation and overdispersion characteristics over a purely compositional structure. This

"hard" compositionality disturbed the original data structure but has several advantages

especially in the data generating process (DGP). Indeed, through our simulation we are able

to control underlying parameters while providing a DGP-agnostic procedure, not promoting

one method over another. We argue that this aspect is central in order to provide systematic

objective method comparisons. Also, as a parametric framework the NORtA algorithm is

limited in its capability to simulate data with a higher number of microorganisms or

metabolites than the number of individuals. Thus, as initially mentioned for global association

methods, we suggest filtering out core elements using either an univariate or a multivariate

method before using models assuming a sample size bigger than the number of features.

To summarize, in this paper we provide an extensive benchmark of integrative

computational methods for incorporating metagenomics and metabolomics data. We hope

this work will represent a great opportunity for the multi-omics community in order to improve

research standards and practices. This aspect is central for scientific discovery and

reproducibility.

Conclusions

In summary, the present study provides to the multi-omics community one of the largest

comprehensive benchmarks of statistical frameworks to jointly integrate metagenomics and

metabolomics data. Through an extensive simulation study, we systematically compared

fifteen integrative approaches across most of the research questions encountered in

practice. We identified the best methods and illustrated their capability to highlight

complementary biological processes involved at different scales with an application to

microbiome and metabolome data for Konzo disease. Overall, our study provides a robust

and replicable comparative framework of integrative methods. We hope this work will serve

as a foundation for setting research standards and the development of new efficient

statistical models to mutually analyze metagenomics and metabolomics data.

Figure 1 Overview of the simulation setup and integrative methods for analyzing

microbiome-metabolome relationships depending on the research question

(**A**) Correlated microbiome and metabolome data were generated using the "Normal to

Anything" framework (See Methods). Microbiome data were simulated considering a zero-

inflated negative binomial distribution, while metabolome datasets follow a negative binomial

distribution. For each dataset, proportions of associated features vary between 1% and 10%,

with association strengths randomly picked from a Gaussian distribution.

(**B**) Overview of the integrative methods related to the research question.

Figure 2 Performance of the multivariate methods for both global association and data

summarization in the high dimensional scenario.

When control of Type-I error rate is of interest, we are looking for methods providing

quantiles of observed p-values similar to quantiles of expected p-values, i.e., following the

diagonal line. In other words, the closer the dots to the straight line, the more the method

adequately controls the false positives. Similarly, for power, we are looking for methods

providing high powers. That is, detecting a significant association when we know there is an

association. Explained variance is the data variance contained through latent factors. See

Methods for details on performance metrics. **(A)** QQ-Plot of the Mantel test applied on the

ILR transformed microbiome and log transformed metabolome data, considering different

distance kernels for metabolites. Here we considered Spearman's method for computing the

global association between the two datasets. **(B)** QQ-Plot of MMiRKAT applied on the ILR

transformed microbiome and log transformed metabolome data, considering different

distance kernels for metabolites. Points below the straight line refer to a conservative

behavior in the result section. **(C)** Power of the Mantel test applied on the ILR transformed

microbiome and log transformed metabolome data, considering different distance kernels for

metabolites for both the Mantel test and MMiRKAT. P-values <= 0.05 were considered as

significant. **(D)** Proportion of explained variance for the data summarization methods

considering the log transformed metabolome and the the alpha transformed and ILR

transformed microbiome data.

Figure 3 Performance of the individual association methods for compositional

predictors

When control of Type-I error rate is of interest, we are looking for methods providing

quantiles of observed p-values similar to quantiles of expected p-values, i.e., following the

diagonal line. In other words, the closer the dots to the straight line, the more the method

adequately controls the false positive. Similarly, for power, we are looking for methods

providing high powers. That is, detecting a significant association when we know there is an

association. See Methods for details on performance metrics.

QQplots of the individual association methods in (**A**) the low dimensional scenario and in the

(**B**) high dimensional scenario. (**C**) Power of the individual association methods across our

two main scenarios. P-values <= 0.05 were considered as significant. For the clr-lm method,

p-values were combined using ACAT [47] in order to provide similar comparisons with the

log-contrast regression and MiRKAT (See Methods). For MiRKAT, we reported Type-I error

rate and power using the ILR transformed microbiome data and the log transformed

metabolites.

Figure 4 Performance of the feature selection methods for providing sparse and

reliable subset of elements across our two scenarios.

Method performance was evaluated with respect to sparsity and F1-Score. For the former,

we are looking for methods providing low values corresponding to a small proportion of

selected features, while for the latter, high values of F1-Scores correspond to better

classification performances (See Methods).

Performance of univariate feature selection methods considering microorganisms as

covariates under our **(A)** low dimensional and **(D)** high dimensional scenarios. For CODA-

LASSO under the high dimensional setting performances were calculated on 100 replicates.

Performance of univariate feature selection methods considering metabolites as covariates

under our **(B)** low dimensional and **(E)** high dimensional scenarios. For the sparse Dirichlet

regression under the high dimensional setting performances were calculated on 100

replicates. Performance of the multivariate feature selection methods considering the CLR

transformed microbiome and the log transformed metabolome under our **(C)** low dimensional

and **(F)** high dimensional scenarios.

Figure 5 Application of best strategies highlights complementary biological

interactions between microorganisms and metabolites in Konzo data

Top-10 most contributing **(A)** metabolites and **(B)** microbiota on the first factor as identified

by MOFA2. Positive correlations were identified by a +, while negative correlations were

identified with a - sign **(C)** Projection of metabolites (red) and microorganisms (blue) into the

2D regression sPLS space. Features with null loadings were removed from the analysis. **(D)**

Distribution of the number of significant microorganisms found by CODA-LASSO across the

subset of metabolites identified by the regression sPLS. **(E)** Log-contrast coefficients for the

2,3-Dihydroxy-2-methylbutanoic acid **(F)** Violin plot of the variance of log-contrast coefficients

through the subset of microorganisms identified by the regression sPLS. Red dots

correspond to outliers with high coefficient's variability.

Scientific Question	Research Aim	Best Method	Pros	Cons
Is there any relationship between microorganisms and metabolites at a global level?	Global associations	MMiRKAT	Robust to data normalization and distance kernels Allow adjustment for covariates	Unable to deal with scenarios with higher number of features than individuals
Are microbiome and metabolome datasets summarizable through a limited number of components?	Data summarization	MOFA2	Robust to data normalization and distance kernels	Running time
Can we identify associations between metabolites and species?	Individual associations	Log-contrast	Compositional and sub-compositional consistent No need to data transformation Allow adjustment for covariates	Limited to few families of generalized linear models
Can we identify core microorganisms and metabolites?	Feature selection (univariate)	CODA-LASSO (compositional covariates)	Compositional and sub-compositional coherent No need to data transformation Allow adjustment for covariates	Limited to few families of generalized linear models
		LASSO (compositional outcomes)	Flexible framework Allow adjustment for covariates	Need a suitable data transformation
	Feature selection (multivariate)	sPLS	Flexible framework Efficiently account for within- between-	Tuning parameters

			correlation	

579 **Table 1: Summary of best methods depending on the research question**

Objective	Methods and Limits	Methodological avenues
Data normalization	- *CLR*: provides still-correlated features in the original space - *ILR*; *Alpha*: are "black-box" transformations providing uncorrelated features in a restricted space	Data normalizations providing uncorrelated features in the original space for facilitating result interpretation
Mechanistic interpretation	- *Log-contrast*; *CODA-LASSO*: unable to provide a mechanistic view of modifications between microbiome and metabolome data	Network-based model to jointly study the modifications of microorganism and metabolite co-occurrence networks [39]
Feature selection	- **CODA-LASSO; Sparse Dirichlet; sPLS; sCCA:** lack of sparse solutions	Extension to knockoff framework [43] or stability selection [44] for improving feature selection performances

580 **Table 2: Overview of avenues for future methodological developments to jointly**

581 **analyze metagenomics and metabolomics data**

583 Methods

584 *Simulation setups*

Microbiome and Metabolome data were simulated using the "*Normal to Anything*" approach

(NORtA), already used for different multi-omics analyses [39, 46, 48]. An appealing feature

of the NORtA algorithm is to provide a framework capable of simulating data from any

marginal distribution while specifying arbitrary correlation structures. Thus, we are able to

generate synthetic microbiome data respecting: (1) *correlation structure*, (2) *zero-inflation*,

and (3) *over-dispersion*, while metabolome was generated similarly removing the zero-

inflation property. This is consistent with real-data characteristics [21]. Moreover, we induced

compositionality for microbiome data by dividing the count of each microorganism by the

sum over all elements in a given individual. Several data transformations for both

microbiome and metabolome were evaluated across our scenarios to account for data

structure (See subsection Data and Distance Kernel Transformation). In order to evaluate

the Type-I error control, we independently generated two datasets under the null hypothesis

of no association between microorganisms and metabolites. Under the alternative

hypothesis, we varied both the number of associations between microorganisms and

metabolites and the strength of associations, mimicking microbiome-metabolome complex

interdependence. Methods were compared under two main scenarios, simulating: (1) 25

microorganisms and 25 metabolites with 100 individuals and (2) 100 microorganisms and

100 metabolites with 500 individuals. Details on the simulation and sensitivity scenarios were

provided in the supplementary. Under all scenarios we simulated 1,000 replicates.

Simulation setup was summarized in Figure 1.

Data and Distance Kernel Transformation

Most methods used in practice need either a normalization step or a distance-based

transformation in order to be applied properly on compositional or over-dispersed data [19].

Thus, we considered in our main analyses three data normalizations for microbiome and one

data transformation for metabolome data. The choice of data normalization depends on the

research objective.

In order to take into account the compositionality of microbiome data while keeping

the original number of features, we considered the centered log-ratio transformation (CLR)

[49] applied on the original count data. This normalization was considered across all the

different considered methods. Basically, the CLR transformation computes the log ratio of

each microbiota count on the geometric mean for a given individual. Formally, the CLR

transformation is given by:

$$CLR(X_j) = \log(\frac{X_j}{g(X)})$$

where $g(X)$ is the geometric mean over all the microorganisms for one sample. This

transformation projects the simplex onto a D compositional subspace under a zero-sum

constraint [24, 50]. By keeping the original number of features the CLR transformation is a

one-one transformation, facilitating result interpretation which is an appealing feature in

practice. We therefore considered the CLR transformation as the reference normalization

when individual associations or feature selection are of interest. However, the CLR

transformation does not ensure independence between features and sub-compositionality

coherence. This latter represents a major limitation for distance-based methods due to

singular covariance matrices. Thus, when distance between features is of interest we

considered the isometric log-ratio (ILR) [25] and alpha transformation [24]. Intuitively, these

two transformations project the original D-dimensional space into an independent D-1 quasi-

orthogonal space, the main difference laying into the transformation used. The ILR

transformation projects the original data onto a Euclidean space. Formally:

$$ILR(X_j) = \sqrt{\left(\frac{j}{j+1}\right)} \log\left(\frac{\prod_{j=1}^{D-1} X_j}{X_j + 1}\right)$$

While the alpha transformation is a Box-Cox type transformation, where the transformed data

follow a multivariate distribution after a suitable alpha-transformation [24].

This facilitates the use of traditional multivariate methods. We therefore considered the ILR

and alpha transformations when evaluating global associations, and data summarization

methods, since the correspondence with the original features does not really matter.

Moreover, since the metabolome data have been shown to be log-normally distributed we

applied a natural log transform on the original count data [51].

Also, we applied different distance kernel transformations before performing some

global association or individual association analyses, highlighting different patterns of

relationships occurring among features. Briefly, we considered Euclidean, Canberra and

Manhattan distances on metabolome matrices of original and log transformed counts, while

considering the Euclidean distance on original and transformed microbiome data.

Interestingly, as presented by [19], the Euclidean distance applied on CLR transformed data

corresponds to the Aitchison distance. This latter has been shown superior to the Bray-Curtis

dissimilarity, representing a true linear relationship, while more stable to data subsetting or

aggregating [52], and will be considered as our reference method here. All data and distance

kernel transformations depending on the method used were summarized in Table S1.

Statistical Analyses

653 Let's assume X and Y, a matrix of microbiome and metabolome, collected on the same set of

654 samples, of size n x p and n x q, where n is the number of samples, p the number of

655 microbiota and q the numbers of metabolites, respectively. X_{ij} represents the jth

656 microorganism in the ith sample, with j = 1,2,.., p, while Y_{ik} is the kth metabolite in the ith

657 sample, where k=1,2,...,q. For the sake of simplicity we considered the case where p=q.

Global Associations

In this paper we refer to global association methods, the statistical approaches providing

global associations between microbiome and metabolome data (Figure 1). We considered

two general methods, the Mantel test [9] and MMiRKAT [10], respectively.

The Mantel test [9] is a statistical framework measuring global correlation between two

datasets measuring on the same set of samples. Traditionally, the Mantel test is applied on

distance or dissimilarity matrices. Here we considered three different distance kernels

applied on the metabolome dataset, Euclidean, Canberra and Manhattan distances. Also, we

applied the Euclidean distance on the original and transformed microbiome matrix, since this

projection leads to more natural interpretations [52] (Table S1). The Mantel test was applied

considering either Pearson's or Spearman's correlation. P-values were obtained empirically

based on permutations using 10,000 replicates. The Mantel test was performed using the

vegan R package.

MMiRKAT is the multivariate extension of MiRKAT providing global association

between a distance-transformed microbiome dataset and a low dimensional continuous

multivariate phenotype [10]. Consistent with distance kernels used in the Mantel test, we

considered Euclidean, Canberra and Manhattan distances applied on the original and

transformed microbiome data, while the entire original or log transformed metabolome matrix

was considered as the outcome (Table S1). MMiRKAT was applied using the *MiRKAT* R

package.

Data Summarization

In this benchmark, we considered 4 distinct data summarization methods, encompassing

CCA, PLS, RDA, and MOFA2. Briefly, all these methods seek to summarize data information

through latent factors.

CCA initially proposed by [11] summarizes the relationship between two datasets by

finding linear combinations of the two matrices maximizing the correlation. CCA was

performed using the *CCA* R package.

Unlike CCA, PLS seeks for linear combinations maximizing the covariance between

the two datasets [12]. Also, in PLS directionality of effect of one matrix on the other can be

taken into account, leading to two general forms of PLS, regression and canonical,

respectively [13]. Thus, canonical PLS and regression PLS were applied with the *mixOmics*

R package.

Moreover, RDA is a two-step procedure, combining multivariate linear regression and

PCA [13]. In the first step, a multivariate linear regression is fitted between each element of

the matrix of responses and the matrix of predictors. Then a PCA is applied on the matrix of

predicted values. RDA was performed using the *vegan* R package.

Finally, MOFA2 is an unsupervised multi-omics framework able to untangle sources

of variability shared by different omics [14]. MOFA2 is a Bayesian probabilistic model able to

find latent factors linking two omics by putting priors on model parameters. We applied

MOFA2 using the related R package *MOFA2* with default parameters.

Except for MOFA2 where the best number of latent factors were chosen by the

model, we kept all the components corresponding to the minimal number of features

observed in one dataset.

Individual associations

When individual relationships are of interest, we consider different regression models taking

into account the compositionality induced by microbiome data as predictors.

Indeed, for microbiota that are explanatory variables, we fitted 3 different models, a

log-linear regression on the CLR transformed microbiome, a log-contrast model [22] and

MiRKAT [10].

Formally the log-linear model of the CLR transformed microbiome (referred to as clr-

lm in the Result section) is given by:

716
$$E(Y_{ik}^* | X_{ij}^*, \beta_j) = \beta_0 + X_{ij}^* \beta_j + \epsilon_i, \forall (j, k)$$

where Y^* is the log transformed metabolome matrix and X^* the CLR transformed

microbiome data. Although the compositionality in the microbiome data is taken into account

using the CLR transformation, the previous model is not robust to the subset of

microorganisms, not preserving the sub-compositionality feature of microbiome data. Thus,

the log-contrast model by imposing a zero-sum constraint on regression coefficient

preserves the scale invariance property needed to ensure the sub-compositionality

characteristic of microbiome data [22]. Formally, the model is given by:

$$E(Y_{ik}^* | X_{i.}, \beta) = X_{i.} \beta + \epsilon_i, \sum_{j=1}^{p} \beta_j = 0$$

Under the log-contrast framework, following [22] we applied the global significance F-test in

order to determine whether there is an association between at least one microorganism and

a given metabolite. The log-contrast model was performed using the *Compositional* R

package. Aligned with the idea of global association, MiRKAT is a statistical framework

exploiting semi-parametric kernel machine regression framework in order to summarize

microbiome relationships [10]. One major feature of MiRKAT compared to other approaches

is permitting the use of several distance kernels at the same time. This is particularly

appealing since it is often unclear in practice which kernel is the more suitable. In our

context, we considered Euclidean, Canberra and Manhattan distances either on original or

transformed microbiome data, while considering the original or log transformed metabolome

as outcome. MiRKAT was applied with the *MiRKAT* R package.

Feature Selection: Univariate

Adapted from [23] we considered two different models accounting for compositional

predictors, when fitting models with metabolites as outcomes. Firstly, we considered the

CLR-LASSO, performing the CLR transformation on microbiome data before fitting a

univariate or multivariate LASSO log-linear regression [16]. We referred to as LASSO and

MLASSO in the Results section. Formally for a metabolite k, the LASSO log-linear model is

given by:

$$\sum_{i=1}^{n}(Y_{ik}^{*} - \sum_{j=1}^{p} X_{ij}\beta_j)^2 + \lambda \sum_{j=1}^{p} |\beta_j|$$

with Y^* is the log transformed metabolome matrix and X^* the CLR transformed

microbiome data. Best penalty parameters λ were chosen using a 10-fold cross-validation

through a 10 step grid-search from 0.01 to 1. LASSO or MLASSO models were fitted using

the *glmnet* R package.

Then, consistently with the log-contrast model, we applied the coda-LASSO

considering a log-linear response of the metabolome level. Briefly, the coda-LASSO is a

penalized log-contrast model, permitting to select only the most contributive features, with a

zero-sum constraint on regression coefficients, ensuring scale invariance, a property needed

for compositional data. The model considered in the coda-LASSO framework is a direct

extension of the model initially proposed by [53]. This latter fits a two-stage model on all

possible log-ratios between each pair of microbiota, leading to sparse solutions. The R

package *coda4microbiome* with the default parameters were used when applying coda-

LASSO.

Then, following the same rationale, when fitting models with microorganisms as

outcomes, we considered two different approaches, adjusting a univariate or multivariate

LASSO linear model on the CLR transformed microorganisms or taking advantage of the

sparse Dirichlet regression framework [27]. For the former, the model for the jth

microorganism is given by:

$$\sum_{i=1}^{n}(X_{ij}^{*} - \sum_{k=1}^{p}Y_{ik}\beta_j)^2 + \lambda\sum_{j=1}^{p}|\beta_j|$$

where X^* is the CLR transformed microbiome data. Here we considered the original and

the log transformed metabolome signal as explanatory variables. In the sparse Dirichlet

regression we used a multinomial dirichlet distribution. These models are direct extensions of

the original LASSO model assuming X following a Dirichlet distribution [27]. Consistently with

the methodology used in LASSO, best penalty parameters were chosen from a 0.1 step grid-

search between 0.01 and 1 using a 10-fold cross-validation. Sparse Dirichlet regression

framework was applied using the *MGLM* R package.

Feature Selection: Multivariate

Sparse Canonical Correlation Analysis (sCCA) [17] and sparse Partial Least Squares (sPLS)

[18] are two penalized extensions of CCA and PLS permitting to summarize data information

through latent factors while proceeding to feature selection.

For sCCA we used L1 penalty on the two datasets, only keeping features contributing

on the two first components. Best penalty parameters were found using 25 permutation-

based samples considering a 0.1-step grid search from 0.01 to 1. sCCA were performed

using the *PMA* R package.

Consistently, canonical and regression sPLS were tuned using a 10-fold cross

validation, considering a 5 step grid search ranging from 5 to 25 in our low dimensional

setting and from 10 to 50 in our high dimensional scenario. We maximally kept two

components in order to select the most contributive features. sPLS were applied using the

mixOmics R package. For both sCCA and sPLS, features on the two first components with

non-null loadings were considered as informative variables hence were kept to compute the

performance metrics.

P-value combinations

In order to provide fair comparisons across our individual association methods with

compositional predictors, we combined p-values using the Aggregated Cauchy-based test

(ACAT) [47] when CLR-lm was considered. Indeed, for a large number of microbiota and

metabolites, applying univariate methods can lead to p x q possible correlations, limiting the

statistical power due to multiplicity. Similarly to the log-contrast model or MiRKAT, in practice

one can be interested in having the global association between one metabolite and several

microorganisms. Thus, in order to provide a powerful method controlling the Type-I error rate

well, we combined p-values for all microorganisms in a given metabolite using ACAT [47],

resulting from p p-values. We argue that this approach may result in more detected signals,

since the multiplicity burden is drastically reduced. Briefly, ACAT is a method combining p-

values through a Cauchy distribution.

Formally for one metabolite, the aggregated p-values across the p microbiota can be

approximated by:

$$0.5 - \frac{\arctan(\frac{T}{w})}{\pi}$$

where $T = \sum_{j=1}^{p} w_j \tan(\{0.5 - p_j\}\pi)$.

One important feature of ACAT compared to other aggregation methods, such as Fisher's

method, is that the method can efficiently control the Type-I error rate even in presence of

correlated p-values, while maintaining good power [47]. Also, the method does not require

any resampling step, facilitating its application to large datasets.

812 *Performance Metrics*

Since all the methods considered in this benchmark exploit different statistical concepts, the

outputs cannot be directly compared. Consequently, we opted for several performance

metrics depending on the research question.

Indeed, for global and individual association methods, we systematically evaluated

model performance through Type-I error control and power, since the considered methods

are frequentist frameworks. Briefly, Type-I error control assesses whether a method provides

a good control of false positives at a given significance threshold. In other words, under the

null hypothesis of no association, at a significance threshold equals to 0.05 we maximally

expect 5% of false positives for a method that performs well. Type-I error control was

evaluated using the quantile-quantile plot of the -log10 of p-values. Similarly, the power is the

capability of a method to detect a significant signal (at a given significance threshold) when

we know that there is an association. In practice, researchers want methods maximizing the

power while accurately controlling the Type-I error.

Data summarization methods were compared based on the proportion of the

explained variance. We refer to explained variance, the amount of data variability kept by

latent factors built by methods.

Moreover, inspired from [42] when univariate *and* multivariate feature *selection*

methods were evaluated, we considered sparsity and reliability as primary performance

metrics. For univariate *methods* sparsity corresponds to the total number of relevant

associations found by the method (here with coefficients different from zero), while reliability

is the capability of a method to accurately discriminate true from false associations between

two features. However, we adapted both sparsity and reliability calculation when considering

multivariate feature selection methods. Indeed, sparsity was computed by the total number of

nonzero coefficients on the total number of features while reliability was adapted to capture

the model performance to keep true contributive variables within the two datasets. Reliability

was evaluated using the F1-Score (harmonic mean of the precision and recall). In practice,

researchers are looking for sparse methods with high F1-Score. Performance metrics

depending on the considered method were summarized in Figure 1. Technical details on the

performance metric calculation and adaptations were provided in the supplementary.

Methods.

Konzo data analysis workflow

Stool samples collected from individuals from study populations in Masi-Manimba (n = 65)

and Kahemba (n = 106) regions of the Democratic Republic of the Congo were used for

metagenomics and metabolomics assessment, where a proportion of the cohort is affected

with Konzo. Shotgun metagenomics sequencing was performed on DNA extracted from

~250mg of stool with the goal of generating ~50 million reads per sample. Data was

analyzed following similar methodology as described previously using Kracken2 and

Bracken for taxonomic classifications [29] . Additionally, stool was analyzed by the company

Metabolon, harnessing their large in-house repository of rigorously tested and validated

metabolites that are used as reference, to detect metabolites present in the samples.

Analysis was performed on the 1,098 microorganisms and 1,340 metabolites across the 171

individuals unconditionally of the disease status. Microbiome data at the genus level were

normalized using the CLR transformation while metabolome data were log transformed. The

workflow was as follows 1) global association, 2) data summarization 3) univariate and

multivariate feature selection and 4) individual associations. Moreover, we considered

microorganisms as explanatory variables and the microorganisms as outcomes. For global

associations, since the number of features exceeds the number of individuals, we performed

the Mantel test instead of MMiRKAT. We further discussed this aspect in the Discussion

section. Then, we applied MOFA2 in order to detect the most contributing microorganisms

and metabolites on the first component. Following the same methodology as presented in

the Method section, we extracted the core microorganisms and metabolites using the

regression sPLS, keeping only the features with nonzero loadings on the two first

components. We finally applied the log-contrast and CODA-LASSO in order to highlight

contributions of microorganisms on metabolites. We summarized the workflow in Figure S26.

Ethics approval and consent to participate

869 Not applicable

Consent for publication

871 Not applicable

Availability of data and materials

873 Codes to reproduce the analyses are available at:

874 https://github.com/lmangnier/Benchmark_Integration_Metagenomics_Metabolomics. The

875 simulated data are produced using the simulate_data.R script available in the same Github

876 repository. R 4.2.2 is required to reproduce results from the paper. The metagenomics and

877 metabolomics data for Konzo disease are available upon request from Matthew S. Bramble.

Competing interests

879   The authors declare no competing interests.

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881   Not applicable

Authors' contributions

884   LM designed, conducted, performed the data analysis, and wrote the manuscript. MM

885   performed the data analysis. AM and NV wrote the manuscript. AB, MPSB, MSB, and AD

886   revised the manuscript. All authors read and approved the final version of the manuscript.

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