A coarse-grained model for disordered and multi-domain proteins

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Abstract Many proteins contain more than one folded domain, and such modular multi-domain proteins help expand the functional repertoire of proteins. Because of their larger size and often substantial dynamics, it may be difficult to characterize the conformational ensembles of multi-domain proteins by simulations. Here, we present a coarse-grained model for multi-domain proteins that is both fast and provides an accurate description of the global conformational properties in solution. We show that the accuracy of a one-bead-per-residue coarse-grained model depends on how the interaction sites in the folded domains are represented. Specifically, we find excessive domain-domain interactions if the interaction sites are located at the position of the C_α atoms. We also show that if the interaction sites are located at the centre of mass of the residue, we obtain good agreement between simulations and experiments across a wide range of proteins. We then optimize our previously described CALVADOS model using this centre-of-mass representation, and validate the resulting model using independent data. Finally, we use our revised model to simulate phase separation of both disordered and multi-domain proteins, and to examine how the stability of folded domains may differ between the dilute and dense phases. Our results provide a starting point for understanding interactions between folded and disordered regions in proteins, and how these regions affect the propensity of proteins to self-associate and undergo phase separation.

Introduction

Multi-domain proteins (MDPs) consist of more than one folded domain that are often connected by linkers or longer intrinsically disordered regions (IDRs), and make up a large fraction (around 50%) of the proteomes in eukaryotic and prokaryotic organisms (***Han et al., 2007***; ***Van Der Lee et al., 2014***). Like intrinsically disordered proteins (IDPs), MDPs can display large-amplitude motions that may play prominent roles in biomolecular functions like signalling, catalysis and regulation (***Mackereth and Sattler, 2012***; ***Van Der Lee et al., 2014***; ***Delaforge et al., 2016***; ***Bondos et al., 2021***).

The biological functions of MDPs depend both on the properties of the folded domains and the disordered regions, and so characterizing the conformational ensembles can be key to understanding how these proteins function. In many cases, the folded and disordered regions are studied separately, but the folded domains might affect the conformational properties of the disordered regions (***Mittal et al., 2018***; ***Taneja and Holehouse, 2021***) and the disordered regions may also affect the properties of the folded domains (***Yu and Sukenik, 2023***). For example, there is a complex interplay between the folded and disordered regions in the RNA-binding protein hnRNPA1, that affects its conformational ensemble in solution and its propensity to undergo phase separation (***Martin et al., 2021b***). However, describing the conformational ensembles of MDPs in solution generally requires a combination of biophysical experiments and molecular dynamics (MD) simulations (***Thomasen and Lindorff-Larsen, 2022***).

All-atom MD simulations have been used to generate conformational ensembles of IDPs and MDPs and to study intra- and inter-domain interactions (***Zheng et al., 2020***; ***Sekiyama et al., 2022***). Such simulations, however, are often limited by the large system sizes and long time scales which limit efficient sampling of these dynamic proteins. Coarse-grained (CG) models may increase the sampling efficiency by reducing the number of particles in the simulation systems (***Neri et al., 2005***; ***Monticelli et al., 2008***; ***Bereau and Deserno, 2009***; ***Gopal et al., 2010***). The accuracy, transferability, and efficiency of such models, however, depend on

the degree of coarse-graining and the parameterization strategy (***Heo and Feig, 2024***). One commonly used model is the Martini force field, which uses a four-to-one mapping scheme with explicit solvent (***Souza et al., 2021***). Different versions of Martini have been modified to produce improved ensembles of IDPs and MDPs (***Benayad et al., 2020***; ***Thomasen et al., 2022***, ***2023***). For IDPs, there has in the last years been extensive work using even coarser models where each amino acid residue is represented by a single bead. The interaction sites are generally located at the C_α position and separated by bonds that are 0.38 nm long, and we therefore here term these C_α models. Several related models rely on a similar functional form to the HPS model introduced by ***Dignon et al.*** (***2018***) and may include bonded terms, an Ashbaugh-Hatch potential (***Ashbaugh and Hatch, 2008***) for shorter-range interactions and a Debye-Hückel electrostatic screening potential. Such models have for example been used to study the conformational ensembles and interactions within and between IDPs (***Dignon et al., 2018***; ***Joseph et al., 2021***; ***Regy et al., 2021***; ***Dannenhoffer-Lafage and Best, 2021***; ***Wessén et al., 2022***; ***Tesei and Lindorff-Larsen, 2023***; ***Valdes-Garcia et al., 2023***).

Coarse-grained models developed for IDPs do not represent the stability of folded proteins well, because the finely balanced energy contributions from individual backbone and side-chain interactions are not captured by the reduced representation. As a consequence, additional (often harmonic) restraints are applied to maintain the folded configurations in folded proteins and MDPs (***Souza et al., 2021***; ***Borges-Araújo et al., 2023***). Even when applying such restraints to models developed for IDPs, extra attention needs to be paid to interactions related to folded domains since it is still unclear whether the models are fully transferable to MDPs. In particular, C_α-based one-bead-per-residue mappings do not account for the specific orientations of side chains in folded proteins (***Kolinski and Skolnick, 1998***). For example, hydrophobic residues, whose side chains are 'tucked away' in the hydrophobic core of the protein, may be exposed at the surface of the protein in a C_α based representation. One approach to help overcome this problem is to use a different or scaled set of force field parameters for interactions that involve folded regions (***Kim and Hummer, 2008***; ***Dignon et al., 2018***; ***Krainer et al., 2021***). Another possible solution is the introduction of more terms in the energy function to better describe long-range interactions (***Li et al., 2012***; ***Tan et al., 2023***) or to introduce anisotropic interactions (***Sieradzan et al., 2022***).

As an alternative, other coarse-grained models represent a residue by more than one bead to represent backbone side chain orientations and interactions (***Pappu et al., 1996***; ***Hyeon et al., 2006***; ***Maity et al., 2022***; ***Zhang et al., 2022***; ***Sieradzan et al., 2022***; ***Mugnai et al., 2023***; ***Zhang et al., 2023***; ***Yamada et al., 2023***). In some of these models, one bead is placed at C_α and the other one is at the centre of mass (COM) of side chain atoms. In this way, side chain interactions can be explicitly taken into account, improving the simulated dynamical behaviour of folded protein simulations and model transferability. In previous studies, this strategy has been used to study conformational ensembles of IDPs or unfolding pathways of proteins (***Hyeon et al., 2006***; ***Mugnai et al., 2023***). While effective, using multi-bead-per-residue models increases the time to sample configurations in simulations, and requires the determination of a larger number of force field parameters.

We have previously developed and applied an automated procedure to optimize the 'stickiness' parameters (λ) in a one-bead-per-residue model by improving the agreement with experimental small-angle X-ray scattering (SAXS) and paramagnetic relaxation enhancement (PRE) nuclear magnetic resonance (NMR) data for a large set of IDPs (***Norgaard et al., 2008***; ***Tesei et al., 2021b***; ***Tesei and Lindorff-Larsen, 2023***). The most recent CALVADOS (Coarse-graining Approach to Liquid-liquid phase separation Via an Automated Data-driven Optimisation Scheme) model (CALVADOS 2) was further tuned to describe phase behaviour of multi-chain conformational ensembles of IDPs from simulations by reducing the range of non-ionic interactions (***Tesei and Lindorff-Larsen, 2023***).

Here, we explore the use of the CALVADOS model for simulations of MDPs. We find that when the CALVADOS 2 parameters are used in simulations of MDPs with interaction sites at the C_α positions, the resulting structures in some cases show excessive interactions between the folded domains, leading to compact ensembles that do not agree with SAXS data. To remedy this problem, we describe a strategy where interaction sites in folded regions are located at the COM of the residue, and show that simulations with this model result in substantially improved agreement with experiments. We optimize the parameters in CALVADOS using the COM representation to derive a refined set of CALVADOS parameters (CALVADOS 3). When we combine the COM representation of folded domains with harmonic restraints between residues in the folded domains and the CALVADOS 3 parameters we obtain good agreement with experimental data on single-chain properties of MDPs and IDPs. Finally, we show how this model may be used to study the interactions between folded and disordered regions in proteins that undergo phase separation, and how the stability of folded domains might change during phase separation.

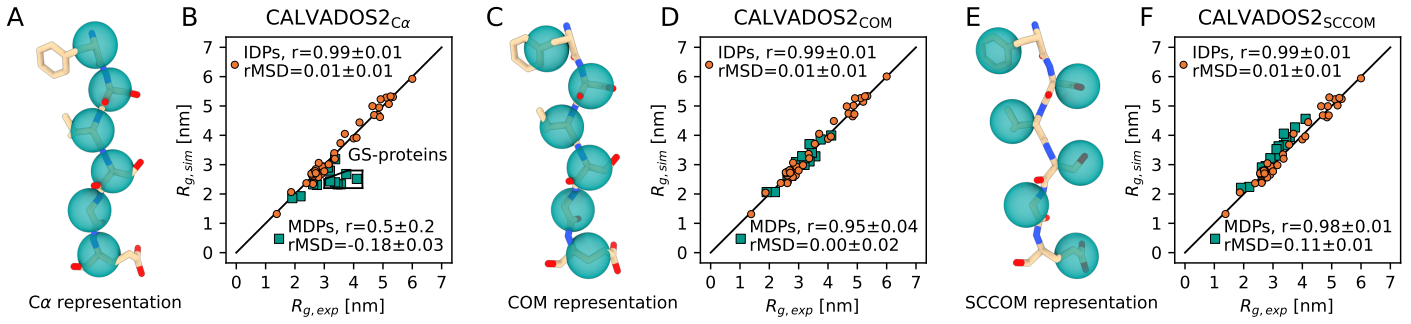

Figure 1. Simulations of MDPs and IDPs using a C_α representation, COM representation or side-chain centre-of-mass (SCCOM) representation. Location of the interaction sites in a β-sheet when using (A) a C_α representation, (C) a COM representation, and (E) a SCCOM representation. Comparison between simulated and experimental R_g values for IDPs (orange) and MDPs (green) using (B) the CALVADOS2$_{C\alpha}$ model (CALVADOS 2 parameters and a C_α representation for both folded and disordered regions), (D) the CALVADOS2$_{COM}$ model (CALVADOS 2 parameters and a COM representation for the interaction sites in the folded regions), and (F) the CALVADOS2$_{SCCOM}$ model (CALVADOS 2 parameters and a SCCOM representation for the interaction sites in the folded regions). The region labelled 'GS-proteins' in panel B contains a number of proteins consisting of pairs of β-sheet-rich fluorescent protein connected by glycine-serine linkers (***Moses et al., 2024***). Pearson correlation coefficients (r) and relative mean signed deviation rMSD $= \langle (R_{g,\mathrm{sim}} - R_{g,\mathrm{exp}})/R_{g,\mathrm{exp}} \rangle$ are reported in the legend, and errors represent standard errors of the mean calculated using bootstrapping. A negative rMSD value indicates that the calculated radii of gyration are systematically lower than the experimental values. The black diagonal lines in panel B, D and F indicate $y = x$.

Results

A modified representation improves accuracy for multi-domain proteins

We first evaluated the accuracy of the original CALVADOS 2 model for simulations of MDPs. We therefore used the CALVADOS 2 parameters (***Tesei and Lindorff-Larsen, 2023***) and a C_α representation to run simulations of 56 IDPs and 14 MDPs (***Table S1***, ***Table S2***, ***Table S3***). In all systems, the interaction sites are located at the C_α positions in both folded and disordered regions; for the MDPs, we applied an additional elastic network model to keep domains intact during simulations (***Figure 1***A, see Methods). We term this combination of the force field parameters (CALVADOS 2) and the C_α representation of the interaction sites in the folded domains as CALVADOS2$_{C\alpha}$. As expected and reported previously (***Tesei and Lindorff-Larsen, 2023***), we found that simulations of IDPs with CALVADOS2$_{C\alpha}$ resulted in good agreement between experimental and calculated values of R_g (***Figure 1***B). In contrast, we found more substantial differences between experimental and calculated values of R_g for several MDPs (***Figure 1***B). In particular, we found that the R_g was underestimated for several MDPs including a series of two fluorescent proteins connected by Gly-Ser linkers of different lengths (here termed GS-proteins; ***Moses et al.*** (***2024***)). This observation was confirmed by calculations of the relative mean signed deviation, rMSD, between experimental and calculated values of R_g that shows that these are on average underestimated by 18% in the MDPs (***Figure 1***B).

As a first attempt at creating a model for both IDPs and MDPs, we used our previously described protocol (***Norgaard et al., 2008***; ***Tesei et al., 2021b***) to optimize the λ stickiness parameters of the CALVADOS model targeting simultaneously SAXS and NMR data on 56 IDPs and 14 MDPs. The resulting λ values were generally smaller than those in CALVADOS 2 (***Figure S1***A) in line with the finding that the MDPs were too compact using CALVADOS 2. Nevertheless, it was also clear that this new parameter set made the agreement worse for disordered proteins (***Figure S1***B-E) and did not result in a satisfactory model to describe both IDPs and MDPs.

We instead hypothesized that the compaction of several MDPs was a result of placing the interaction sites at the C_α positions in the folded domains. In particular for β-sheet-containing proteins, this geometry would mean that residues whose side chains are buried inside the folded domain are represented by interaction sites located closer to the protein surface (***Figure 1***A); thus buried hydrophobic residues might appear as solvent exposed. We therefore constructed a new model where the interaction sites within folded regions were placed at the COM of the residue (***Figure 1***C) and constrained by harmonic restraints; when used with the CALVADOS 2 parameters, we term this model CALVADOS2$_{COM}$. We stress that only the bead locations in the folded domains differ between the CALVADOS2$_{C\alpha}$ and CALVADOS2$_{COM}$ models; residues in disordered regions are represented by one bead centred on the C_α positions in both models. In the absence of folded domains, CALVADOS2$_{COM}$ and CALVADOS2$_{C\alpha}$ are thus identical and simulations with the two models gave comparable results (***Figure 1***B and D). In contrast, simulations of the MDPs with CALVADOS2$_{COM}$ were in substantially better agreement with experiments than simulations with CALVADOS2$_{C\alpha}$ as evidenced e.g. by an increase in Pearson correlation coefficient from 0.5 to 0.95 and an increase in rMSD from -18% to 0% (***Figure 1***B and D). In addition to the COM representation, we also examined whether a side-chain centre-of-mass (SCCOM) representation, shifting bead positions of buried residues further away from the surface, could yield even more accurate R_g

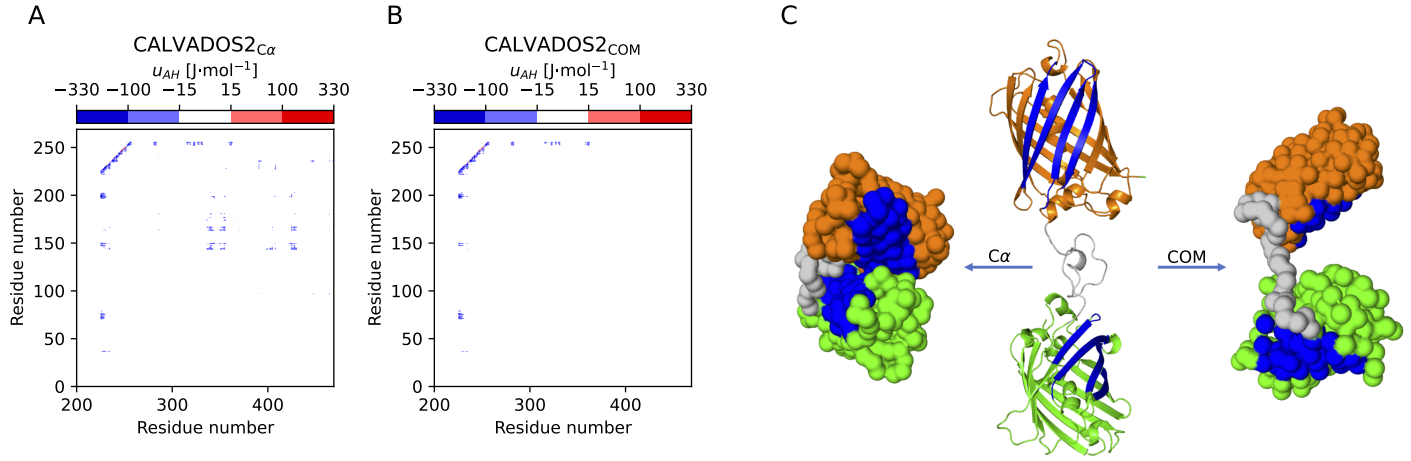

Figure 2. Energy calculations reveal substantial inter-domain interactions. We calculated interaction energy maps (of the Ashbaugh-Hatch term in the force field) from simulations using (A) the CALVADOS2$_{C\alpha}$ model and (B) the CALVADOS2$_{COM}$ model. We show only a subset of the map representing interactions between the first (residues 1–226 on the y-axis) and second (residues 256–470 on the x-axis) folded domains. (C) Examples of structures of GS0 with the same R_g as the average over simulations using CALVADOS2$_{C\alpha}$ (left) and CALVADOS2$_{COM}$ (right). The starting structure of the simulations is shown in the middle, where green and orange parts are the two fluorescent proteins connected by a flexible linker (grey). The regions that interact strongly in the CALVADOS2$_{C\alpha}$ simulations are coloured blue.

predictions than the COM representation (***Figure 1***E). We performed single chain simulations with the CALVADOS 2 parameters and the SCCOM representation (CALVADOS2$_{SCCOM}$) and found that CALVADOS2$_{SCCOM}$ on average resulted in an overestimation of the R_g of MDPs of 11% (***Figure 1***D and F). As an alternative solution to decrease the too strong interactions between folded domains, it has previously been suggested to scale down interactions between pairs of folded domains (by a factor of 0.7) and between folded domains and disordered regions (by a factor of $0.84 = \sqrt{0.7}$) (***Krainer et al., 2021***). While applying this rescaling to CALVADOS 2 (termed CALVADOS2$_{C\alpha}$70%) led to improved agreement with experiments, the improvement was smaller than when using the COM representation, and the simulations had a remaining bias towards underestimating the radii of gyration (***Figure S2***). Therefore, we proceeded by using the COM representation in this study.

To examine in more detail why the CALVADOS2$_{C\alpha}$ model resulted in more compact conformations of MDPs than CALVADOS2$_{COM}$, we calculated the time-averaged non-ionic (Ashbaugh-Hatch) interaction energies between residues of different folded domains. For this analysis we selected GS0, a construct with two fluorescent proteins separated by a 29-residue-long linker (***Moses et al., 2024***), since the R_g value of GS0 deviates substantially from experiments in simulations with CALVADOS2$_{C\alpha}$ (***Figure 1***B). In the energy maps, we see evidence of substantial inter-domain interactions between residue 140–230 of one fluorescent protein and residue 340–440 of the other (***Figure 2***A). In contrast, these domain-domain interactions are not observed when simulating with COM representation (***Figure 2***B). The comparison of the two energy maps thus supports the hypothesis that the too compact conformations of MDPs in simulations with CALVADOS2$_{C\alpha}$ result from inter-domain attractions that are decreased in the COM representation (***Figure 2***C).

Optimizing CALVADOS using a centre-of-mass representation

Having shown that the COM representation gave an improved description of MDPs while preserving the accuracy when simulating IDPs, we proceeded to optimize the CALVADOS model further. We used our iterative Bayesian optimization scheme (***Norgaard et al., 2008***; ***Tesei et al., 2021b***) to optimize the λ stickiness parameters of the CALVADOS model targeting simultaneously SAXS and NMR data on 56 IDPs and 14 MDPs (***Table S1***, ***Table S2***, ***Table S3***). In these simulations we used the COM representation of the folded domains and we thus term the final model CALVADOS3$_{COM}$ to represent both the force field and the COM representation of the folded regions. The resulting λ values in CALVADOS3$_{COM}$ are similar to those in CALVADOS 2 (***Figure 3*** and ***Figure S3***). We found that simulations of IDPs with CALVADOS3$_{COM}$ and CALVADOS2$_{COM}$ gave similar agreement to SAXS experiments. Likewise, we found a similar agreement for the MDPs (***Figure 1***D, ***Figure 3***B and C).

Having optimized λ, we validated the CALVADOS3$_{COM}$ model on 25 IDPs and 9 MDPs (***Table S4***, ***Table S5***) that were not used in training for any of the models (***Figure 4***). For the 25 IDPs we found good agreement for all three models (CALVADOS2$_{C\alpha}$, CALVADOS2$_{COM}$ and CALVADOS3$_{COM}$) (***Figure 4***A–C). We note again that the COM representation is only applied to the folded domain. All IDPs have C_α representations, so CALVADOS2$_{C\alpha}$ and CALVADOS2$_{COM}$ are the same models for IDPs. In contrast, for MDPs we found that CALVADOS3$_{COM}$ and CALVADOS2$_{COM}$ perform substantially better than CALVADOS2$_{C\alpha}$ (***Figure 4***A-C). Our validation results thus show

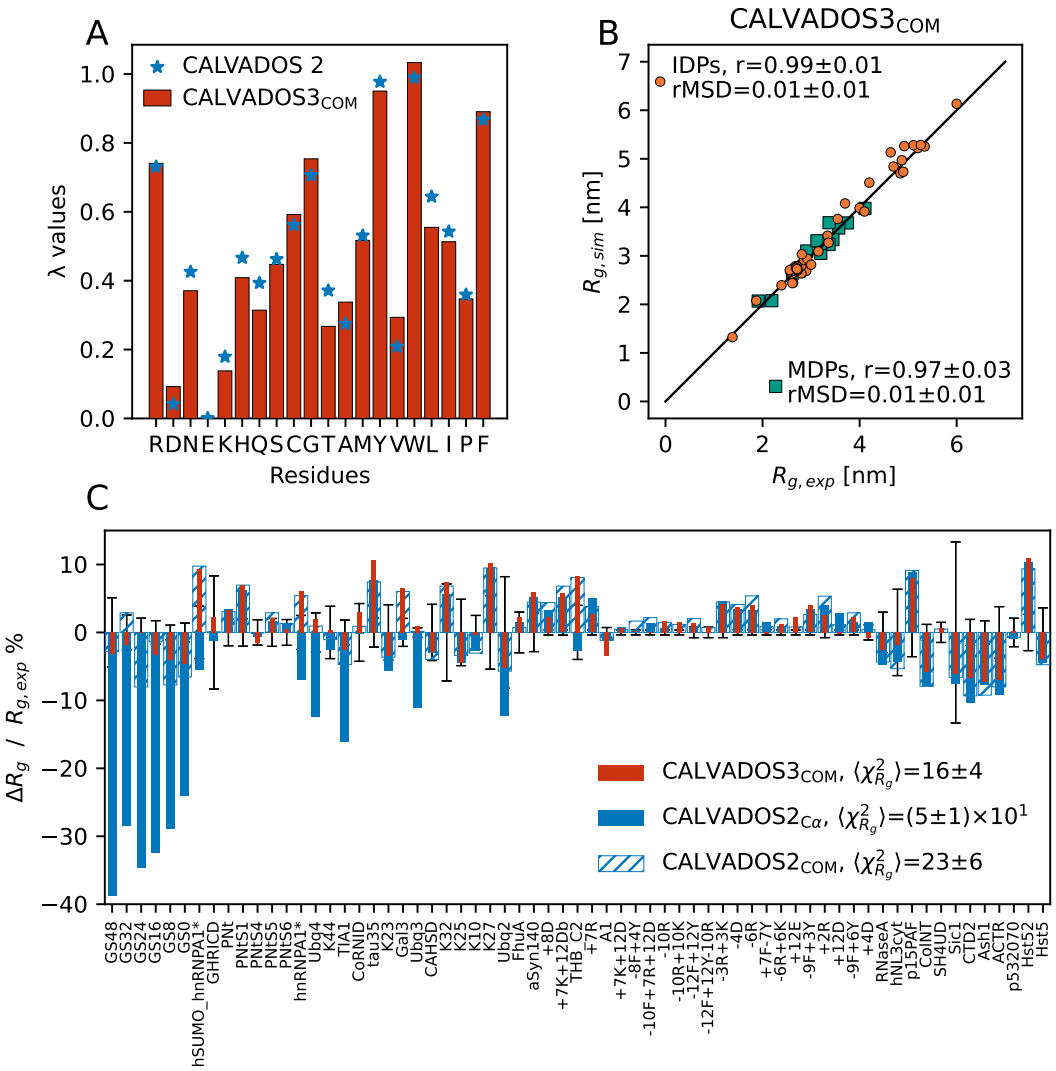

Figure 3. Optimizing the λ parameters using a COM representation for folded domains. (A) Comparison between λ values from CALVADOS 2 (blue) and CALVADOS3$_{\text{COM}}$ (red). (B) Comparison between simulated and experimental R_g values for IDPs (orange) and MDPs (green) using CALVADOS3$_{\text{COM}}$. Pearson correlation coefficients (r) and rMSD are reported in the legend. The black diagonal line indicates $y = x$. (C) Relative difference between experimental and simulated R_g values from CALVADOS3$_{\text{COM}}$ (red), CALVADOS2$_{C\alpha}$ (blue) and CALVADOS2$_{\text{COM}}$ (blue hatched). $\langle \chi^2_{R_g} \rangle$ values across IDPs and MDPs in training set are reported in the legend. Error bars show the experimental error divided by $R_{g,\text{exp}}$.

that the CALVADOS3$_{\text{COM}}$ model gives improved agreement for simulations of MDPs while retaining the accuracy of CALVADOS2$_{C\alpha}$ for simulations of IDPs. Across the 34 independent test proteins we find $\langle \chi^2_{R_g} \rangle$ values of 50, 22 and 15 for CALVADOS2$_{C\alpha}$, CALVADOS2$_{\text{COM}}$ and CALVADOS3$_{\text{COM}}$, respectively (***Figure S4***), and both CALVADOS2$_{\text{COM}}$ and CALVADOS3$_{\text{COM}}$ have essentially no bias (rMSD≈0; ***Figure 4***B and C).

Simulations of phase separation of disordered and multi-domain proteins

We and others have previously used one-bead-per-residue models such as CALVADOS to study the self-association and phase separation of IDPs (***Dignon et al., 2018***; ***Tesei et al., 2021b***; ***Joseph et al., 2021***; ***Regy et al., 2021***; ***Dannenhoffer-Lafage and Best, 2021***; ***Wessén et al., 2022***; ***Tesei and Lindorff-Larsen, 2023***; ***Valdes-Garcia et al., 2023***). In some cases, these models have also been used to study phase separation of proteins that contain a mixture of folded and disordered regions (***Dignon et al., 2018***; ***Conicella et al., 2020***; ***Her et al., 2022***). We therefore examined whether the CALVADOS3$_{\text{COM}}$ model could be used to study phase separation of both IDPs and MDPs. We used multi-chain simulations in a slab geometry (***Dignon et al., 2018***) to simulate the partitioning of proteins between a dilute and dense phase, and calculated the dilute phase concentration (the saturation concentration; c_{sat}) as a sensitive measure of the accuracy of the model. We first simulated 33 IDPs and found that simulations with CALVADOS3$_{\text{COM}}$ gave an agreement with experimental values of c_{sat} that is comparable to that of CALVADOS2$_{C\alpha}$ (***Table S6***, ***Figure S5***, ***Figure S6***, ***Figure S7***).

We then proceeded to use CALVADOS3$_{\text{COM}}$ to study the phase separation of MDPs including hnRNPA1* (where * denotes that residues 259–264 have been deleted from full-length hnRNPA1), full-length FUS (FL_FUS) and other multi-domain proteins with experimental estimates of c_{sat} (***Table S7***; ***Wang et al.*** (***2018***); ***Martin et al.*** (***2021b***)). Simulations of hnRNPA1* with CALVADOS2$_{C\alpha}$, under conditions where the experimental dilute phase concentration is 0.17 mM, resulted in essentially all proteins in the dense phase (c_{sat}=0 mM; ***Figure 5***A). In contrast, simulations using CALVADOS3$_{\text{COM}}$ resulted in a lower propensity to phase separate and a calculated value of c_{sat}=0.14±0.01 mM that is comparable to experiments (***Figure 5***B).

To understand the origin of these differences we calculated interaction energy maps of the proteins in the dense phase. Experiments have shown that the LCD in hnRNPA1* (residues 186–320) plays a central role in driving phase separation (***Molliex et al., 2015***; ***Martin et al., 2021b***), and we indeed found evidence for substantial LCD-LCD interactions in the dense phases in simulations with both CALVADOS2$_{C\alpha}$ (***Figure 5***C) and CALVADOS3$_{\text{COM}}$ (***Figure 5***D). In the simulations with CALVADOS2$_{C\alpha}$ we, however, also observed more substantial interactions between the folded RRM (RNA recognition motif) domains (residues 14–97 and 105–185) and between the RRMs and the LCD. In simulations with CALVADOS3$_{\text{COM}}$ these interactions were much weaker, presumably explaining the increase of c_{sat} in these simulations.

Having demonstrated that CALVADOS3$_{\text{COM}}$ provides a more accurate description of the phase behaviour of hnRNPA1* than CALVADOS2$_{C\alpha}$, we proceeded to perform simulations of several other MDPs for which we found estimates of c_{sat} in the literature (***Figure 6***, ***Figure S8***, ***Figure S9***). As for hnRNPA1*, we found that CALVADOS2$_{C\alpha}$ substantially overestimates the tendency of these proteins to undergo phase separation (i.e. underestimate c_{sat}). The use of the COM representation in CALVADOS3$_{\text{COM}}$ decreases the protein-protein interactions, and thus substantially improves the agreement with experiments, though differences remain.

Examining changes in folding stability in condensates

Experiments have shown that the protein-rich environment of condensates can modulate the stability of folded proteins or nucleic acids (***Nott et al., 2015***; ***Ruff et al., 2022***; ***Chen et al., 2024***; ***Ahmed et al., 2024***). Inspired by these findings, we used the ability to simulate both folded and disordered regions with CALVADOS 3 to examine how partitioning into condensates may shift the folding equilibrium of a folded domain. As it is difficult to sample the folding-unfolding equilibrium by simulations, we studied it indirectly using a thermodynamic cycle that involves differences in partitioning of the folded and unfolded forms into a condensate (***Nott et al., 2015***).

To demonstrate how CALVADOS 3 enables such analyses, we simulated the isolated RRM1 and RRM2 from hnRNPA1* (***Figure 7***A) in the presence of a condensate of the LCD of hnRNPA1* and calculated the free energies of partitioning of the RRM domains in their native, folded state, ΔG^N_{part}. Using the same approach, we performed direct-coexistence simulations without applying harmonic networks to the RRMs to calculate the free energies of partitioning of the RMMs in their unfolded state, ΔG^U_{part}. A comparison of the concentration profiles from our direct-coexistence simulations shows that the unfolded states accumulate in the condensate and are depleted from the dilute phase to a greater extent than the folded states (***Figure 7***B–C); We quantify this via a more negative free energy of partitioning, $\Delta G^U_{\text{part}} < \Delta G^N_{\text{part}}$ (***Figure 7***D). The preference of the unfolded state for the condensate is particularly pronounced for RRM2, for which we estimate a two-fold decrease in the free energy of partitioning ($\Delta G^U_{part} - \Delta G^N_{part} = -0.7$ kcal/mol). From the thermodynamic cycle, this in turn means that the folding stability of RRM2 is 0.7 kcal/mol lower (less stable) in the condensate than in the dilute phase.

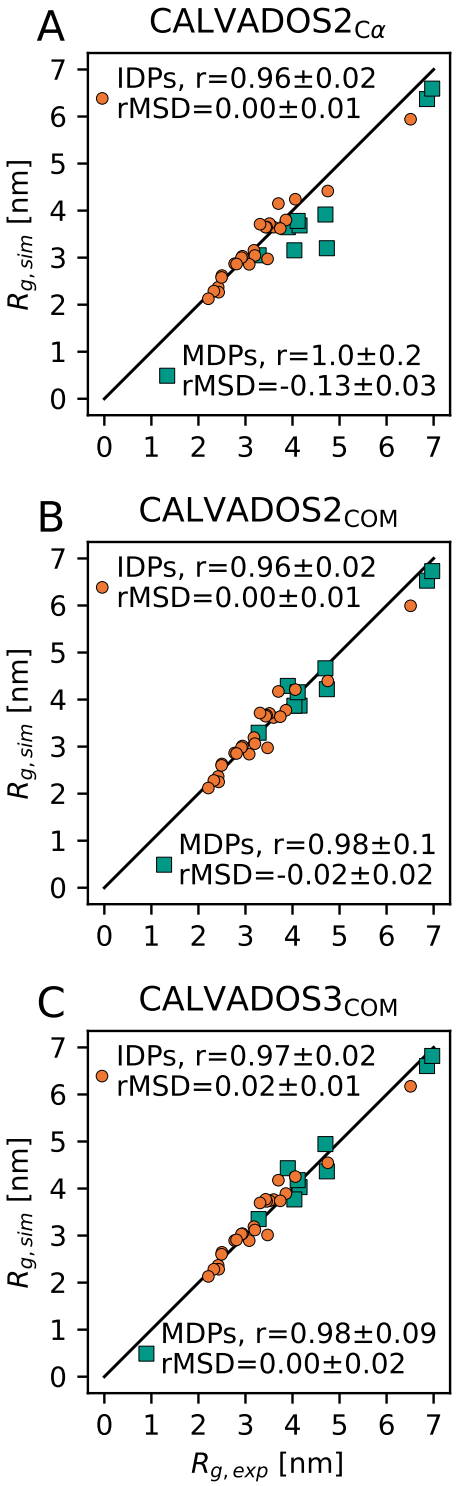

Figure 4. Validation of the CALVADOS3$_{COM}$ model using proteins that were not used during training. Comparison of simulated and experimental R_g values on a validation set using (A) CALVADOS2$_{C\alpha}$, (B) CALVADOS2$_{COM}$ and (C) CALVADOS3$_{COM}$. Pearson correlation coefficients (r) and rMSD are reported in the legend. The black diagonal lines indicate $y = x$.

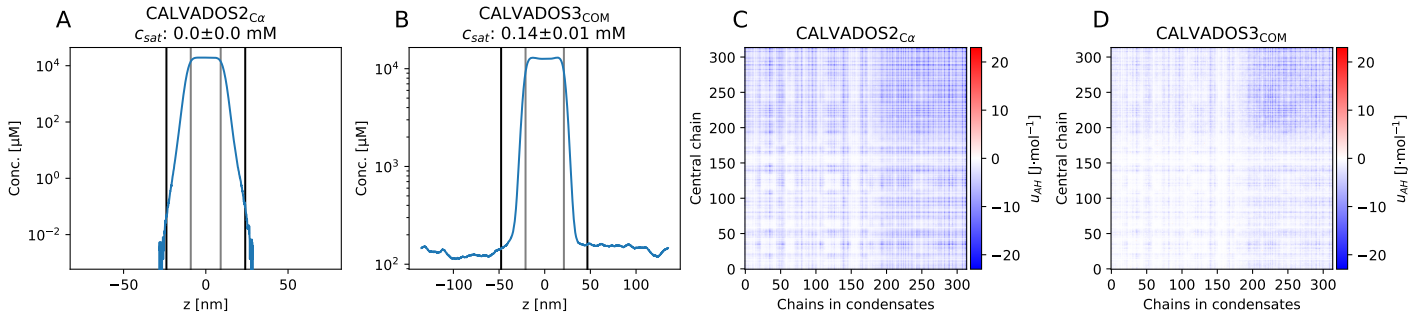

Figure 5. Phase coexistence simulations of hnRNPA1* using (A, C) CALVADOS2$_{C\alpha}$ and (B, D) CALVADOS3$_{COM}$. Simulations were performed at 293 K and an ionic strength of 0.15 M. Equilibrium density profile of hnRNPA1* using (A) CALVADOS2$_{C\alpha}$ and (B) CALVADOS3$_{COM}$. c_{sat} calculated from density profiles are 0 mM and 0.14 mM, respectively. Average residue-residue interaction energies (the Ashbaugh-Hatch term in the force field) between the most central chain and the rest of the condensate for (C) CALVADOS2$_{C\alpha}$ and (D) CALVADOS3$_{COM}$.

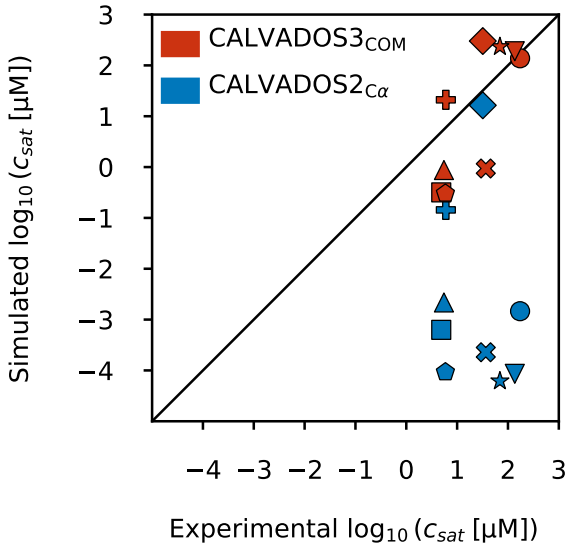

Figure 6. Comparison between simulated and experimental c_{sat} values for MDPs using the CALVADOS3$_{COM}$ model (red) and CALVADOS2$_{C\alpha}$ (blue). The simulated proteins are hnRNPA1* (circle), hSUMO_hnRNPA1* (downward triangle), FL_FUS (upward triangle), GFP_FUS (square), SNAP_FUS (pentagon), SNAP_FUS_PLDY2F_RBDR2K (star), SNAP_FUS_PLDY2F (x symbol), FUS_PLDY2F_RBDR2K (diamond) and hnRNPA3 (plus symbol). The black diagonal line indicates $y = x$.

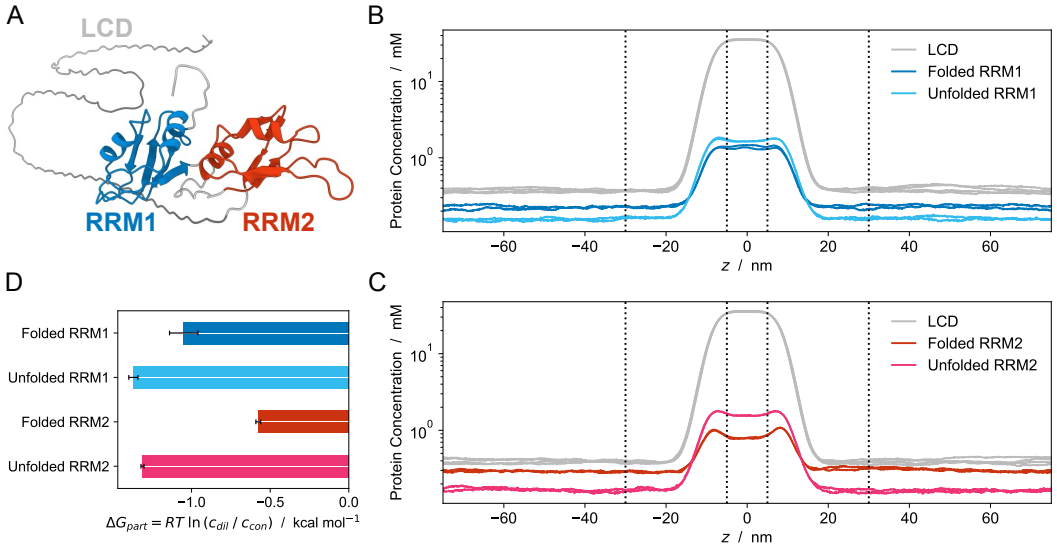

Figure 7. Predicting the effect of the protein-rich environment of a condensate on the stability of folded domains. (A) Structure of hnRNPA1* highlighting the low-complexity domain (grey) and RNA-recognition motifs 1 (blue) and 2 (red). (B) Concentration profiles of the LCD (grey) and RRM1 in the native (blue) and unfolded (cyan) state. (C) Concentration profiles of the LCD (grey) and RRM2 in the native (red) and unfolded (magenta) state. (D) Free energy of partitioning of RRM1 and RRM2 in native and unfolded states into condensates of the LCD. Data estimated from direct-coexistence simulations performed in two independent replicates. Error bars in (D) represent the differences between the replicates.

To put these changes into context, we used a recently developed machine learning approach (***Cagiada et al., 2024***) to predict the absolute protein folding stabilities of the isolated RRMs in the dilute phase, $\Delta G^{\mathrm{dil}}_{N\rightarrow U}$, and obtained 6.6 kcal/mol for RRM1 and 4.4 kcal/mol for RRM2. Using these values and assuming a two-state model, we estimate that the partitioning into the condensate has a negligible effect on the amount of unfolded state for RRM1; in contrast we predict a four-fold increase in the population of the unfolded state of RRM2 from $\exp(-\Delta G^{dil}_{N\rightarrow U}/RT) \approx 1/2000$ to $\exp[-(\Delta G^{dil}_{N\rightarrow U} + \Delta G^{U}_{part} - \Delta G^{N}_{part})/RT] \approx 1/500$. Although substantial additional work is needed to examine the accuracy of CALVADOS 3 for quantifying differences in partitioning of folded and unfolded proteins into condensates, these data show a promising use of our model for predicting unfolding in condensates.

Discussion

In this work, we found that simulations with the CALVADOS2$_{C\alpha}$ model, previously shown to represent single-chain and multi-chain properties of IDPs, underestimated the radii of gyration of MDPs. Changing the CG mapping method from C$_\alpha$ to COM substantially improved the agreement with experimental data. This observation is in line with the finding that reconstruction of all-atom structures from a centre-of-mass representation is more accurate than from a C$_\alpha$ representation (***Heo and Feig, 2024***). We reoptimized the 'stickiness' parameters in the context of a COM-based model based on experimental data for both IDPs and MDPs. The resulting CALVADOS3$_{\mathrm{COM}}$ model provides a good description of both single- and multi-chain simulations of both IDPs and MDPs.

The relatively low c_{sat} value calculated from slab simulations of hnRNPA1* with CALVADOS2$_{C\alpha}$ further supported that interactions between the folded domains are overestimated by C$_\alpha$-based models without any further modifications. Considering that the SCCOM-based model (CALVADOS2$_{\mathrm{SCCOM}}$) overestimated R_g of MDPs, we suggest that the COM-based model (CALVADOS3$_{\mathrm{COM}}$) appears to strike a good balance, leading to improved values of c_{sat} for MDPs. Nevertheless, some systematic differences remain even with this model, which resulted in underestimates of c_{sat} for different constructs of the protein FUS. Together, our results show that the new parameter set and the centre-of-mass representation (CALVADOS3$_{\mathrm{COM}}$) retain the accuracy of CALVADOS 2 for IDPs, but improve the description of proteins with both disordered and folded domains. We therefore term this new model CALVADOS 3, with the implicit notion that this model is used with centre-of-mass representation of residues within folded regions. We note that an earlier version of this preprint (***Cao et al., 2024***) used a slightly different set of parameters, and we suggest to refer to that model as CALVADOS 3beta.

When simulating MDPs with CALVADOS 3 we need to restrain the folded domains using harmonic restraints. In the current work we have manually determined the boundaries for which regions are considered to be folded,

though automated methods will be needed for large-scale applications. Tools for automatic predictions of domain boundaries exist (***Holm and Sander, 1994***; ***Lau et al., 2023***) and might be combined with AlphaFold to set the harmonic restraints (***Jussupow and Kaila, 2023***).

Despite these current limitations, we envision that the CALVADOS 3 model will enable detailed studies of the interactions within and between multi-domain proteins, and pave the way for proteome-wide simulation studies of full-length proteins similar to what has recently been achieved for IDRs (***Tesei et al., 2024***). We also envision that our approach to study changes in protein stability inside condensates can be used together with methods to predict absolute protein stability (***Cagiada et al., 2024***) to learn and expand our knowledge on the rules that underlie phase separation and changes in stability of folded, globular proteins (***Ruff et al., 2022***).

Methods

Description of the model

We modelled each amino acid by one bead. We generated C_α-beads for IDPs and assigned C_α atom coordinates to bead positions for IDRs in multi-domain proteins according to their modelled or experimental structures (see below, Simulations). For structured domains, we used the following rules for the different representations: we placed each bead position at the C_α atom (C_α representation), or the centre of mass calculated for all the atoms in a residue (COM representation), or the centre of mass calculated for only side chain atoms of a residue (SC-COM representation). The CALVADOS 3 energy function consists of bonded interactions, non-bonded interactions and an elastic network model as described below.

Chain connectivity of the beads is described by a harmonic potential,

$$u_{\text{bond}}(r) = k(r - r_0)^2, \tag{1}$$

with force constant $k = 8033$ kJ·mol^{-1}·nm^{-2}. The equilibrium distance r_0 is set to 0.38 nm if two beads are both within IDRs, or the distance between two beads in the initial conformation if at least one bead is within a folded domain.

For non-bonded interactions, we use a truncated and shifted Ashbaugh-Hatch (AH) and Debye-Hückel (DH) potential to model van der Waals and salt-screened electrostatic interactions, respectively. The Ashbaugh-Hatch potential is described by

$$u_{\text{AH}}(r) = \begin{cases} u_{\text{LJ}}(r) - \lambda u_{\text{LJ}}(r_c) + \epsilon(1-\lambda), & r \leq 2^{1/6}\sigma \\ \lambda[u_{\text{LJ}}(r) - u_{\text{LJ}}(r_c)], & 2^{1/6}\sigma < r < r_c \\ 0, & r > r_c \end{cases}, \tag{2}$$

where $u_{\text{LJ}}(r)$ is the Lennard-Jones (LJ) potential,

$$u_{\text{LJ}}(r) = 4\epsilon\left[\left(\frac{\sigma}{r}\right)^{12} - \left(\frac{\sigma}{r}\right)^{6}\right], \tag{3}$$

and where $\epsilon = 0.8368$ kJ·mol^{-1} and $r_c = 2.2$ or 2 nm. Similar to previous work, we use $r_c = 2.2$ nm during the optimization of CALVADOS3$_{\text{COM}}$, and use 2 nm during validation and application (***Tesei and Lindorff-Larsen, 2023***). Both σ and λ are calculated as the arithmetic averages of residue-specific bead size and stickiness, respectively. σ values are van der Waals volumes calculated by Kim and Hummer (***Kim and Hummer, 2008***). λ values are treated as free parameters and optimized iteratively through a Bayesian parameter-learning procedure as described previously (***Tesei et al., 2021b***; ***Tesei and Lindorff-Larsen, 2023***) to minimize the differences in the simulated and experimental R_g and PRE data. In simulations where we scaled down interactions of folded domains (CALVADOS2$_{C\alpha}$70%)), we scaled down ϵ to 0.7ϵ for domain-domain interactions and to $\sqrt{0.7}\epsilon$ for domain-IDR interactions.

The Debye-Hückel potential is described by

$$u_{\text{DH}}(r) = \frac{q_i q_j e^2}{4\pi\epsilon_0\epsilon_r}\frac{\exp(-r/D)}{r}, \tag{4}$$

where q is the average amino acid charge number, e is the elementary charge, $D = \sqrt{1/(8\pi B c_s)}$ is the Debye length of an electrolyte solution of ionic strength c_s, $B(\epsilon_r)$ is the Bjerrum length and ϵ_0 is vacuum permittivity. Electrostatic interactions are truncated and shifted at the cutoff distance $r_c = 4$ nm. The temperature-dependent dielectric constant of the implicit aqueous solution is modelled by the following empirical relationship (***Akerlof and Oshry, 1950***):

$$\epsilon_r(T) = \frac{5321}{T} + 233.76 - 0.9297\times T + 1.417\times10^{-3}\times T^2 - 8.292\times10^{-7}\times T^3. \tag{5}$$

We use the Henderson-Hasselbalch equation to estimate the average charge of the histidine residues, assuming a pK_a value of 6 (***Nagai et al., 2008***).

We use an elastic network model (ENM) with a harmonic potential to restrain non-bonded pairs in the folded domains using

$$u_{\text{ENM}}(r) = k_d(r - r_0)^2. \tag{6}$$

Here, the force constant k_d is 700 kJ·mol^{-1}·nm^{-2}, r is the distance between beads and equilibrium distances r_0 are directly taken from the reference structures. We only apply the ENM to residue pairs with an r_0 below a 0.9 nm cutoff. We determine the predefined boundary of each domain in MDPs by visual inspection of the three-dimensional structures (***Table S8***). Each domain has a starting amino acid and an ending amino acid indicating the range of the domain. Only residue pairs within the same domain are restrained by this harmonic potential except for bonded pairs, which are restrained by the aforementioned bonded potential. All boundaries of MDPs are consistent with definitions in their experimental or simulation articles. In some cases, one domain could be discontinuous because of long loops within the domain so we exclude those regions when defining boundaries. Residues of α-helix, β-sheet and short loops in a structured domain are all restrained equally with the same force constant and cutoff distance. The application of ENM ensures that secondary structures within folded domains would not fluctuate substantially (***Figure S10***). Non-bonded interactions (Ashbaugh-Hatch and Debye-Hückel potential) are excluded for the restrained pairs.

Simulations

We generated initial conformations of all IDPs as Archimedes' spirals with a distance of 0.38 nm between bonded beads. Atomistic structures of all MDPs used in optimization procedures, single-chain validation and slab simulations either came from our recent work (***Thomasen et al., 2023***) or were modelled by superposing experimental domain structures (if available) on AlphaFold predictions (***Jumper et al., 2021***; ***Varadi et al., 2022***). We then mapped all of these MDPs to CG structures based on different CG representations (C_α, COM, SCCOM).

We conducted Langevin dynamics simulations using OpenMM 7.6.0 (***Eastman et al., 2017***) in the NVT ensemble with an integration time step of 10 fs and friction coefficient of 0.01 ps^{-1}. Single chains of N residues were simulated in a cubic box with a $(N-1)\times 0.38 + 4$ nm box edge length under periodic boundary conditions. Each chain was simulated in 20 replicas for $6.3\sim 77.7$ ns depending on the sequence length of the disordered regions (***Tesei and Lindorff-Larsen, 2023***; ***Tesei et al., 2024***). Final trajectories had 4000 frames for each protein, excluding the initial 10 frames in each replica.

We performed direct-coexistence simulations in a cuboidal box using $[L_x, L_y, L_z] = [17, 17, 300]$ and $[15, 15, 150]$ nm to simulate multi-chains of Ddx4WT and the other IDPs, respectively. For MDPs, box sizes are shown in ***Table S7***. To keep the condensates thick enough and reduce finite-size surface effects, we chose 150 chains for hnRNPA1* and 100 chains for all the other IDPs and MDPs (see also below). We generated each IDP chain as an Archimedes spiral with a distance of 0.38 nm between bonded beads in the xy-plane. Each spiral was placed along the z-axis with a 1.47 nm interval. To avoid steric clashes of densely packed MDP input structures, we chose the most compact conformation sampled by single-chain simulations with CALVADOS 2 parameters and corresponding CG representation as the initial conformation for each MDP chain. Before production simulations, we performed equilibrium runs where we used an external force to push each chain towards the centre of the box so that a condensate could be formed. We then continued to perform production simulations, saving frames every 0.125 ns and discarded the first 150 ns before analysis. The slab in each frame was centred in the box and the equilibrium density profile $\rho(z)$ was calculated by taking the averaged densities over the trajectories as previously described (***Tesei and Lindorff-Larsen, 2023***).

To examine finite-size effects of the direct-coexistence simulations we performed additional simulations of hnRNPA1* varying both the box dimensions (L_x, L_y, L_z) and the number of chains. We calculated both dense and dilute phase concentrations from each simulation and find that unless we use a very small patch ($L_x = L_y = 11$ nm), the results are consistent (***Figure S11***, ***Figure S12***, ***Table S9***), in line with previous analyses of such finite-size effects (***Dignon et al., 2018***; ***Joseph et al., 2021***). Convergence of the IDP simulations was assessed as previously described (***Tesei et al., 2021b***).

To indicate the computational performance of single- and multi-chain CALVADOS simulations, we show the performance for systems of different sizes run either on an Intel Xeon Gold 6130 CPU (for single-chain simulations) on an NVIDIA Tesla V100 GPU (for multi-chain simulations) (***Figure S13***).

To estimate the free energy of partitioning of RRM1 (residues 11–89) and RRM2 (residues 105–179) into condensates of hnRNPA1* LCD (GS followed by residues 186–314), we performed direct-coexistence simulations at 298 K, pH 7.5, and 150 mM ionic strength, in a cuboidal box with sidelengths $[L_x, L_y, L_z] = [15, 15, 150]$ nm. The structures of the native states of RRM1 and RRM2 were based on the crystal structure (***Shamoo et al., 1997***) as previously described (***Martin et al., 2021b***). We performed two independent simulations, each 21 µs long, for each system and, after centering the LCD condensate in the middle of the box, calculated concentration

profiles along the z-axis using the last 20 μs of each trajectory. We estimated the free energies of partitioning as $\Delta G_{\text{part}} = RT \ \ln(c_{\text{dil}} \ / \ c_{\text{con}})$ where R is the gas constant and c_{dil} and c_{con} are the average concentrations of the RRMs in the dilute phase and in the LCD condensate, respectively. The error on ΔG_{part} was estimated as the difference between the values from the two independent simulation replicas. Absolute folding stabilities of RRM1 and RRM2 were calculated using the Google Colab implementation of a recently described model for predicting absolute protein stability (***Cagiada et al., 2024***).

Parameter optimization

Our Bayesian Parameter-Learning Procedure (***Tesei and Lindorff-Larsen, 2023***) of the 'stickiness' parameters, λ, aimed to minimize the following cost function:

$$\mathcal{L}(\lambda) = \langle \chi^2_{R_g} \rangle + \eta \langle \chi^2_{\text{PRE}} \rangle - \theta \ln(P(\lambda)). \tag{7}$$

$\chi^2_{R_g}$ and χ^2_{PRE} denoting R_g and PRE differences between experiments and simulations are estimated as

$$\chi^2_{Rg} = \left(\frac{R_g^{\text{exp}} - R_g^{\text{calc}}}{\sigma^{\text{exp}}} \right)^2 \tag{8}$$

and

$$\chi^2_{\text{PRE}} = \frac{1}{N_{\text{labels}} N_{\text{res}}} \sum_j^{N_{\text{labels}}} \sum_i^{N_{\text{res}}} \left(\frac{Y_{ij}^{\text{exp}} - Y_{ij}^{\text{calc}}}{\sigma_{ij}^{\text{exp}}} \right)^2. \tag{9}$$

Here $P(\lambda)$ is a statistical prior of λ (***Tesei et al., 2021b***; ***Tesei and Lindorff-Larsen, 2023***), σ^{exp} is the error on the experimental values, Y is PRE data, either $I_{\text{para}}/I_{\text{dia}}$ or Γ_2 is calculated using the rotamer library approach implemented in DEER-PREdict (***Tesei et al., 2021a***), N_{labels} is the number of spin-labeled mutants, and N_{res} is the number of measured residues. The prior loss, $\theta \ln(P(\lambda))$, quantifies the difference between prior distribution $P(\lambda)$ and current λ values (with min-max normalization at each step) to avoid overfitting. The coefficients are set to η = 0.1 and θ = 0.08. λ is not allowed to be negative but can be greater than 1.0 during optimization.

We used a training set consisting of 56 IDPs and 14 MDPs to perform the optimization. All of those proteins were from our previous studies (***Tesei and Lindorff-Larsen, 2023***; ***Thomasen et al., 2023***). A summary of the training data and other properties of different CALVADOS models is shown in the supporting material (***Table S10***). 51 IDPs and 14 MDPs in this training set were used for fitting against experimental SAXS R_g data and 5 IDPs were used for fitting against experimental PRE data (***Table S1***, ***Table S2***, ***Table S3***). We then used a validation set to validate the performances of our new optimized models on reproducing experimental R_g. This validation set was composed of 25 IDPs and 9 MDPs. 12 IDPs in this validation set were from our previous work and the rest (13 IDPs and 9 MDPs) were newly collected experimental R_g data in this work (***Table S4***, ***Table S5***). We also collected nine MDPs with measured values of c_{sat} to examine the accuracy of the phase behaviour simulated with the models presented in this work (***Table S7***).

The optimization procedure went through several cycles until convergence of the final total cost ($|\Delta\mathcal{L}| < 1$, $\Delta\mathcal{L}$ is the difference of final total cost between the current and previous cycle, ***Equation 7***). Within each cycle, we use the optimized λ values from the previous cycle to perform new single-chain simulations (initial λ values for the first cycle are CALVADOS 2 parameters, (***Tesei and Lindorff-Larsen, 2023***)), calculate R_g and PRE for each frame and then nudge values in the λ set iteratively to minimize the cost function (five residues are randomly subjected to small perturbations sampled from a Gaussian distribution with $\mu = 0, \sigma = 0.05$). This trial λ set (λ_k) is used to calculate the Boltzmann weights of each frame by $w_i = \exp(-[U(r_i, \lambda_k) - U(r_i, \lambda_0)]/k_{\text{B}}T)$, where U is the AH potential, r_i are coordinates of a conformation, k_{B} is the Boltzmann constant and T is temperature. The resulting weights are then used to calculate the effective fraction of frames by $\phi_{\text{eff}} = \exp[-\sum_i^{N_{\text{frames}}} w_i \log(w_i \times N_{\text{frames}})]$; if $\phi_{\text{eff}} \geq 0.6$, trial λ_k acceptance probability is determined by the Metropolis criterion, $\min\{1, \exp\left(\frac{\mathcal{L}(\lambda_{k-1}) - \mathcal{L}(\lambda_k)}{\xi_k}\right)\}$, where ξ_k is a unitless control parameter, its initial value is set to 0.1 and scaled down by 1% at each iteration until $\xi < 10^{-8}$, which means a micro-cycle is complete. Within a cycle, a total of 10 micro-cycle are performed. In this work, the optimization procedure converged within three cycles. Therefore, we used the resulting λ values from the third cycle as the final parameter set. We ran one additional optimization cycle to confirm the convergence of the training.

Data and software availability

Scripts and data to reproduce the work are available via https://github.com/KULL-Centre/_2024_Cao_CALVADOSCOM.

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Table S1. Experimental solution conditions and radii of gyration of IDPs included in the training set for the Bayesian parameter-learning procedure.

Protein	N	R_g ± Err [nm]	T [K]	c_s [M]	pH	Ref.
Hst5	24	1.38 ± 0.05	293.00	0.150	7.5	(***Jephthah et al., 2019***)
Hst52	48	1.87 ± 0.05	298.00	0.150	7.0	(***Fagerberg et al., 2020***)
p532070	62	2.39 ± 0.05	277.00	0.100	7.0	(***Zhao et al., 2021***)
ACTR	71	2.63 ± 0.1	278.00	0.200	7.4	(***Kjaergaard et al., 2010***)
Ash1	81	2.9 ± 0.05	293.00	0.150	7.5	(***Martin et al., 2016***; ***Jin and Gräter, 2021***)
CTD2	83	2.614 ± 0.05	293.00	0.120	7.5	(***Jin and Gräter, 2021***)(***Gibbs et al., 2017***)]
Sic1	92	3.0 ± 0.4	293.00	0.200	7.5	(***Gomes et al., 2020***)
SH4UD	95	2.71 ± 0.04	293.15	0.216	8.0	(***Shrestha et al., 2019***)
ColNT	98	2.8 ± 0.033	277.00	0.433	7.6	(***Johnson et al., 2017***)
p15PAF	111	2.81 ± 0.1	298.00	0.150	7.0	(***De Biasio et al., 2014***)
hNL3cyt	119	3.15 ± 0.2	293.00	0.300	8.5	(***Paz et al., 2008***)
RNaseA	124	3.36 ± 0.1	298.00	0.150	7.5	(***Riback et al., 2017***)
+4D	137	2.72 ± 0.03	298.00	0.150	7.0	(***Bremer et al., 2022***)
-3R+3K	137	2.63 ± 0.02	298.00	0.150	7.0	(***Bremer et al., 2022***)
-6R+6K	137	2.79 ± 0.01	298.00	0.150	7.0	(***Bremer et al., 2022***)
-10R+10K	137	2.85 ± 0.01	298.00	0.150	7.0	(***Bremer et al., 2022***)
-4D	137	2.64 ± 0.01	298.00	0.150	7.0	(***Bremer et al., 2022***)
+2R	137	2.62 ± 0.02	298.00	0.150	7.0	(***Bremer et al., 2022***)
+12D	137	2.8 ± 0.01	298.00	0.150	7.0	(***Bremer et al., 2022***)
+12E	137	2.85 ± 0.01	298.00	0.150	7.0	(***Bremer et al., 2022***)
+7K+12D	137	2.92 ± 0.01	298.00	0.150	7.0	(***Bremer et al., 2022***)
+7R	137	2.71 ± 0.01	298.00	0.150	7.0	(***Bremer et al., 2022***)
-12F+12Y-10R	137	2.61 ± 0.02	298.00	0.150	7.0	(***Bremer et al., 2022***)
-10F+7R+12D	137	2.86 ± 0.01	298.00	0.150	7.0	(***Bremer et al., 2022***)
+8D	137	2.69 ± 0.01	298.00	0.150	7.0	(***Bremer et al., 2022***)
+7K+12Db	137	2.56 ± 0.01	298.00	0.150	7.0	(***Bremer et al., 2022***)
-9F+6Y	137	2.66 ± 0.01	298.00	0.150	7.0	(***Bremer et al., 2022***)
-10R	137	2.67 ± 0.01	298.00	0.150	7.0	(***Bremer et al., 2022***)
-9F+3Y	137	2.68 ± 0.01	298.00	0.150	7.0	(***Bremer et al., 2022***)
-8F+4Y	137	2.71 ± 0.01	298.00	0.150	7.0	(***Bremer et al., 2022***)
+7F-7Y	137	2.72 ± 0.01	298.00	0.150	7.0	(***Bremer et al., 2022***)
-12F+12Y	137	2.6 ± 0.02	298.00	0.150	7.0	(***Bremer et al., 2022***)
A1	137	2.76 ± 0.02	298.00	0.150	7.0	(***Bremer et al., 2022***)
-6R	137	2.57 ± 0.01	298.00	0.150	7.0	(***Bremer et al., 2022***)
aSyn140	140	3.55 ± 0.1	293.00	0.200	7.4	(***Ahmed et al., 2021***)
FhuA	144	3.34 ± 0.1	298.00	0.150	7.5	(***Riback et al., 2017***)
K27	167	3.7 ± 0.2	288.00	0.150	7.4	(***Mylonas et al., 2008***)
K10	168	4.0 ± 0.1	288.00	0.150	7.4	(***Mylonas et al., 2008***)
K25	185	4.1 ± 0.2	288.00	0.150	7.4	(***Mylonas et al., 2008***)
K32	198	4.2 ± 0.3	288.00	0.150	7.4	(***Mylonas et al., 2008***)
CAHSD	227	4.84 ± 0.2	293.00	0.070	7.0	(***Hesgrove et al., 2021***)
K23	254	4.9 ± 0.2	288.00	0.150	7.4	(***Mylonas et al., 2008***)
tau35	255	4.64 ± 0.1	293.20	0.150	7.4	(***Lyu et al., 2021***)
CoRNID	271	4.7 ± 0.2	293.15	0.192	7.5	(***Cordeiro et al., 2019***)
K44	283	5.2 ± 0.2	288.00	0.150	7.4	(***Mylonas et al., 2008***)
PNt	334	5.11 ± 0.1	298.00	0.150	7.5	(***Riback et al., 2017***; ***Bowman et al., 2020***)
PNtS1	334	4.92 ± 0.1	298.00	0.150	7.5	(***Bowman et al., 2020***)
PNtS4	334	5.34 ± 0.1	298.00	0.150	7.5	(***Bowman et al., 2020***)
PNtS5	334	4.87 ± 0.1	298.00	0.150	7.5	(***Bowman et al., 2020***)
PNtS6	334	5.26 ± 0.1	298.00	0.150	7.5	(***Bowman et al., 2020***)
GHRICD	351	6.0 ± 0.5	298.00	0.350	7.3	(***Seiffert et al., 2020***; ***Pesce et al., 2023***)

Table S2. Experimental solution conditions and PRE data included in the training set for the Bayesian parameter-learning procedure.

Proteins	N	N_{labels}	$\omega_I/2\pi[MHz]$	T [K]	c_s [M]	pH	Ref.
A2	155	2	850	298	0.005	5.5	(***Ryan et al., 2018***)
aSyn	140	5	700	283	0.200	7.4	(***Dedmon et al., 2005***)
OPN	220	10	800	298	0.150	6.5	(***Kurzbach et al., 2016***)
FUS	163	3	850	298	0.150	5.5	(***Monahan et al., 2017***)
FUS12E	164	3	850	298	0.150	5.5	(***Monahan et al., 2017***)

Table S3. Experimental solution conditions and radii of gyration of MDPs included in the training set for the Bayesian parameter-learning procedure.

Protein	N	R_g ± Err [nm]	T [K]	c_s [M]	pH	Ref.
THB_C2	137	1.91 ± 0.076	295.15	0.15	6.5	(***Michie et al., 2016***)
Ubq2	162	2.19 ± 0.18	293.00	0.33	8.0	(***Jussupow et al., 2020***)
Ubq3	228	2.62 ± 0.018	293.00	0.33	8.0	(***Jussupow et al., 2020***)
Gal3	250	2.91 ± 0.06	303.00	0.04	7.0	(***Lin et al., 2017***)
TIA1	275	2.75 ± 0.05	293.15	0.10	6.0	(***Sonntag et al., 2017***)
Ubq4	304	3.19 ± 0.092	293.00	0.33	8.0	(***Jussupow et al., 2020***)
hnRNPA1*	314	3.12 ± 0.078	293.15	0.15	7.5	(***Martin et al., 2021b***)
hSUMO_hnRNPA1*	433	3.37 ± 0.13	293.15	0.10	7.5	(***Martin et al., 2021b***)
GS0	470	3.2 ± 0.044	293.15	0.15	7.4	(***Moses et al., 2024***)
GS8	486	3.37 ± 0.036	293.15	0.15	7.4	(***Moses et al., 2024***)
GS16	502	3.45 ± 0.06	293.15	0.15	7.4	(***Moses et al., 2024***)
GS24	518	3.57 ± 0.075	293.15	0.15	7.4	(***Moses et al., 2024***)
GS32	534	3.75 ± 0.097	293.15	0.15	7.4	(***Moses et al., 2024***)
GS48	566	4.11 ± 0.21	293.15	0.15	7.4	(***Moses et al., 2024***)

Table S4. Experimental solution conditions and radii of gyration of IDPs included in the validation set.

Protein	N	R_g ± Err [nm]	T [K]	c_s [M]	pH	Ref.
ChiZ164	67	2.42 ± 0.01	293.00	0.0650	7.0	(***Hicks et al., 2020***)
DomainV	67	2.43 ± 0.024	288.15	0.1985	7.0	(***Chan-Yao-Chong et al., 2019***)
DSS1	71	2.5 ± 0.1	288.00	0.1700	7.4	(***Pesce et al., 2023***)
BMAL1P624A	98	2.77 ± 0.09	283.25	0.1540	7.2	(***Garg et al., 2019***)
VWF	103	3.08 ± 0.03	293.00	0.1530	7.4	(***del Amo-Maestro et al., 2021***)
p27Cv56	107	2.328 ± 0.1	293.00	0.0950	7.2	(***Das et al., 2016***)
p27Cv14	107	2.936 ± 0.13	293.00	0.0950	7.2	(***Das et al., 2016***)
p27Cv78	107	2.211 ± 0.03	293.00	0.0950	7.2	(***Das et al., 2016***)
p27Cv31	107	2.81 ± 0.18	293.00	0.0950	7.2	(***Das et al., 2016***)
p27Cv44	107	2.492 ± 0.13	293.00	0.0950	7.2	(***Das et al., 2016***)
p27Cv15	107	2.915 ± 0.1	293.00	0.0950	7.2	(***Das et al., 2016***)
PTMA	111	3.7 ± 0.2	288.00	0.1600	7.4	(***Pesce et al., 2023***)
GON7	114	3.18 ± 0.04	283.00	0.2110	6.5	(***Arrondel et al., 2019***)
NHE6cmdd	116	3.2 ± 0.2	288.00	0.1700	7.4	(***Pesce et al., 2023***)
hKISS1	120	3.47 ± 0.05	283.15	0.1590	7.0	(***Ibáñez de Opakua et al., 2017***)
TtASR1	141	3.31 ± 0.08	293.15	0.1500	7.3	(***Hamdi et al., 2017***)
HvASR1	143	3.51 ± 0.09	293.15	0.1500	7.3	(***Hamdi et al., 2017***)
TIF2NRID	150	3.74 ± 0.092	283.15	0.1750	6.8	(***Senicourt et al., 2021***)
ED4	163	4.06 ± 0.11	293.15	0.1530	7.4	(***Gondelaud et al., 2021***)
ANAC046	167	3.6 ± 0.3	298.00	0.1400	7.0	(***Pesce et al., 2023***)
PARCL	180	3.43 ± 0.065	293.15	0.1700	7.5	(***Ostendorp et al., 2022***)
N_FATZ1	191	3.45 ± 0.062	293.15	0.1920	7.5	(***Sponga et al., 2021***)
D91_FATZ1	209	4.0 ± 0.1	293.00	0.1800	7.5	(***Sponga et al., 2021***)
cDAXX	246	4.75 ± 0.05	293.00	0.1300	8.0	(***Schmit et al., 2019***)
ED3	373	6.51 ± 0.15	293.15	0.1530	7.4	(***Gondelaud et al., 2021***)

Table S5. Experimental solution conditions and radii of gyration of MDPs included in the validation set.

Protein	N	R_g ± Err [nm]	T [K]	c_s [M]	pH	Ref.
SH4UD_SH3_SH2	264	3.28 ± 0.06	293.15	0.216	8.0	(***Gurumoorthy et al., 2023***)
H46	381	4.15 ± 0.05	283.00	0.163	6.5	(***Elena-Real et al., 2023***)
TDP43W2A	415	4.11 ± 0.04	293.15	0.312	8.0	(***Wright et al., 2020***)
PCPE	424	4.04 ± 0.11	293.15	0.506	7.4	(***Bernocco et al., 2003***)
NiV_V	457	6.97 ± 0.02	293.15	0.232	8.0	(***Salladini et al., 2017***)
HeV_V	458	6.86 ± 0.03	293.15	0.232	8.0	(***Salladini et al., 2017***)
D14	483	3.9 ± 0.17	283.15	0.156	7.5	(***Hajizadeh et al., 2018***)
S4FL	552	4.7 ± 0.1	283.15	0.169	7.2	(***Gomes et al., 2021***)
ChiAM	682	4.73 ± 0.077	293.15	0.282	8.0	(***Mazurkewich et al., 2020***)

Table S6. IDPs and experimental conditions used for slab simulations in this work.

Proteins	N	c_s [M]	pH	T [K]	$c_{\text{sat,exp}}$ [μM]	Ref.
LAF1	176	0.15	7.5	293	44.0	(***Schuster et al., 2020***)
LAF1D2130	166	0.15	7.5	293	275.0	(***Schuster et al., 2020***)
LAF1shuf	176	0.15	7.5	293	6.0	(***Schuster et al., 2020***)
A1S150	131	0.15	7.0	293	218.1	(***Martin et al., 2021a***)
A1S200	131	0.20	7.0	293	159.8	(***Martin et al., 2021a***)
A1S300	131	0.30	7.0	293	93.4	(***Martin et al., 2021a***)
A1S500	131	0.50	7.0	293	66.5	(***Martin et al., 2021a***)
-12F+12Y	137	0.15	7.0	293	60.3	(***Bremer et al., 2022***)
+4D	137	0.15	7.0	277	4.5	(***Bremer et al., 2022***)
-6R	137	0.15	7.0	277	7.1	(***Bremer et al., 2022***)
A1	137	0.15	7.0	293	102.2	(***Bremer et al., 2022***)
+2R	137	0.15	7.0	277	18.0	(***Bremer et al., 2022***)
+8D	137	0.15	7.0	277	18.7	(***Bremer et al., 2022***)
-14N+14Q	137	0.15	7.0	293	171.6	(***Bremer et al., 2022***)
-10G+10S	137	0.15	7.0	293	268.1	(***Bremer et al., 2022***)
+7F-7Y	137	0.15	7.0	293	209.0	(***Bremer et al., 2022***)
-20G+20S	137	0.15	7.0	293	469.4	(***Bremer et al., 2022***)
-23S+23T	137	0.15	7.0	293	342.2	(***Bremer et al., 2022***)
-8F+4Y	137	0.15	7.0	277	63.2	(***Bremer et al., 2022***)
-3R+3K	137	0.15	7.0	277	83.1	(***Bremer et al., 2022***)
-4D	137	0.15	7.0	277	88.8	(***Bremer et al., 2022***)
-9F+3Y	137	0.15	7.0	277	115.0	(***Bremer et al., 2022***)
+23G-23S	137	0.15	7.0	293	46.1	(***Bremer et al., 2022***)
+23G-23S+7F-7Y	137	0.15	7.0	293	194.0	(***Bremer et al., 2022***)
+23G-23S-12F+12Y	137	0.15	7.0	293	6.5	(***Bremer et al., 2022***)
-30G+30S	137	0.15	7.0	293	841.8	(***Bremer et al., 2022***)
FUS	163	0.15	5.5	297	105.0	(***Murthy et al., 2019***)
A2	155	0.01	5.5	297	15.0	(***Ryan et al., 2021***)
Ddx4WT	236	0.13	6.5	297	230.0	(***Brady et al., 2017***)
allF	137	0.15	7.0	293	250.0	(***Alshareedah et al., 2023***)
allY	137	0.15	7.0	293	85.0	(***Alshareedah et al., 2023***)
allW	137	0.15	7.0	293	1.0	(***Alshareedah et al., 2023***)
FUS_long	216	0.15	7.0	285	46	(***Farag et al., 2023***)

Table S7. Multi-domain proteins and experimental conditions used for slab simulations in this work.

Proteins	N	c_s [M]	pH	T [K]	$c_{\text{sat,exp}}$ [μM]	Box [nm]	Ref.
hnRNPA1*	314	0.15	7.5	293	173.0	[20, 20, 270]	(***Martin et al., 2021b***)
hnRNPA3	381	0.116	7.4	298	6	[25, 25, 190]	(***Kar et al., 2022***)
hSUMO_hnRNPA1*	433	0.15	7.5	293	136.2	[25, 25, 190]	(***Martin et al., 2021b***)
FL_FUS	526	0.15	7.4	293	5.5	[20, 20, 270]	(***Wang et al., 2018***)
GFP_FUS	764	0.15	7.4	293	4.9	[25, 25, 300]	(***Wang et al., 2018***)
SNAP_FUS	708	0.15	7.4	293	5.9	[25, 25, 300]	(***Wang et al., 2018***)
SNAP_FUS_PLDY2F_RBDR2K	710	0.15	7.4	293	69.4	[29, 29, 340]	(***Wang et al., 2018***)
SNAP_FUS_PLDY2F	710	0.15	7.4	293	36.6	[25, 25, 300]	(***Wang et al., 2018***)
FUS_PLDY2F_RBDR2K	528	0.15	7.4	293	32.0	[25, 25, 340]	(***Wang et al., 2018***)

Table S8. Domain boundaries of MDPs used in this study for the Bayesian parameter-learning procedure and validation. Brackets indicate the first and last residue of the domain, respectively. Nested brackets indicate subdomains (restrained) separated by long linkers (unrestrained).

Protein	N	Domain 1	Domain 2	Domain 3	Domain 4	Domain 5	Domain 6	Domain 7
THB_C2	137	[6, 42]	[50, 137]					
Ubq2	162	[11, 82]	[87, 158]					
Ubq3	228	[1, 72]	[77, 148]	[153, 224]				
Gal3	250	[117, 250]						
SH4UD_SH3_SH2	264	[94, 150]	[166, 258]					
TIA1	275	[6, 82]	[95, 172]	[190, 275]				
Ubq4	304	[1, 72]	[77, 148]	[153, 224]	[229, 300]			
hnRNPA1*	314	[11, 89]	[105, 179]					
H46	381	[140, 355]						
TDP43W2A	415	[5, 77]	[107, 177]	[193, 260]	[321, 329]			
PCPE	424	[12, 125]	[134, 249]	[293, 412]				
hSUMO_hnRNPA1*	433	[44, 114]	[132, 209]	[224, 298]				
NiV_V	457	[406, 457]						
HeV_V	458	[404, 456]						
GS0	470	[1, 226]	[256, 470]					
D14	483	[31, 121]	[157, 246]	[265, 354]	[400, 479]			
GS8	486	[1, 226]	[272, 486]					
hSUMO_TIA1PrLD	492	[32, 102]	[114, 186]	[212, 287]	[321, 389]			
GS16	502	[1, 226]	[288, 502]					
GS24	518	[1, 226]	[304, 518]					
FL_FUS	526	[286, 368]	[423, 451]					
FUS_PLDY2F_RBDR2K	528	[288, 370]	[425, 453]					
GS32	534	[1, 226]	[320, 534]					
S4FL	552	[15, 138]	[[287, 294], [323, 466], [492, 542]]					
GS48	566	[1, 226]	[352, 566]					
ChiAM	682	[8, 89]	[92, 172]	[178, 257]	[266, 356]	[359, 462]	[471, 567]	[578, 668]
SNAP_FUS	708	[286, 368]	[423, 451]	[[537, 564], [586, 701]]				
SNAP_FUS_PLDY2F_RBDR2K	710	[288, 370]	[425, 453]	[[539, 566], [588, 703]]				
SNAP_FUS_PLDY2F	710	[288, 370]	[425, 453]	[[539, 566], [588, 703]]				
GFP_FUS	764	[286, 368]	[423, 451]	[529, 755]				

Table S9. Analysis of the system size effects on slab simulation of hnRNPA1*. The protein concentration is fixed throughout all simulation configurations and is above the experimental saturation concentration. ND: In simulations with 150 chains and box size [11.0, 11.0, 900] nm we did not observe a stable condensed phase.

	Number of chains & Box size [nm]	Simulation length [μs]	Dilute phase conc. [mM]	Dense phase conc. [mM]
Varying (L_x, L_y)	45 chains & [11.0, 11.0, 270]	10	0.3±0.1	12.0±0.2
	75 chains & [14.1, 14.1, 270]	10	0.19±0.02	12.62±0.02
	300 chains & [28.3, 28.3, 270]	10	0.18±0.01	12.37±0.02
	450 chains & [34.6, 34.6, 270]	5	0.104±0.008	12.32±0.02
Varying L_z	45 chains & [20.0, 20.0, 81]	10	0.16±0.02	11.62±0.05
	75 chains & [20.0, 20.0, 135]	10	0.15±0.01	12.34±0.03
	300 chains & [20.0, 20.0, 540]	10	0.10±0.01	12.51±0.02
	450 chains & [20.0, 20.0, 810]	5	0.08±0.01	12.48±0.03
Varying (L_x, L_y, L_z)	150 chains & [11.0, 11.0, 900]	10	ND	ND
	150 chains & [14.1, 14.1, 540]	10	0.17±0.03	12.5±0.1
	150 chains & [28.3, 28.3, 135]	10	0.16±0.01	12.11±0.03
	150 chains & [34.6, 34.6, 90]	10	0.17±0.01	10.79±0.06

Table S10. Summary of CALVADOS models. The number of IDPs and MDPs, and cutoff distance for the AH potential used during optimization, cutoff distance of AH potential for validation (production) simulations, and references are shown.

models	IDPs	MDPs	cutoff_optimization [nm]	cutoff_production [nm]	Ref.
CALVADOS 1	48	0	4.0	4.0	(***Tesei et al., 2021b***)
CALVADOS 2	56	0	2.4	2.0	(***Tesei and Lindorff-Larsen, 2023***)
CALVADOS 3 ($CALVADOS3_{COM}$)	56	14	2.2	2.0	this study

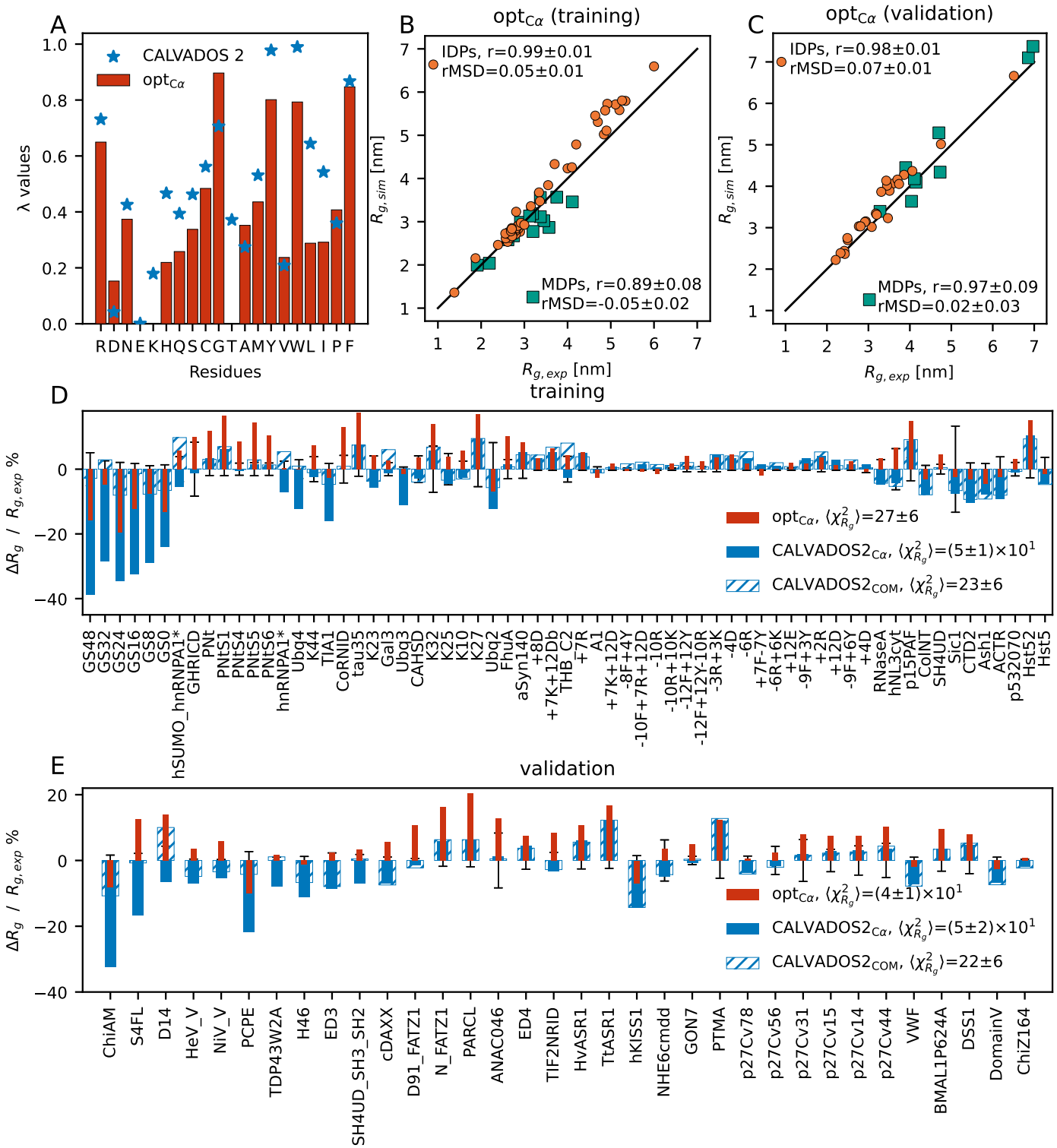

Figure S1. Optimizing the λ parameters using a Cα representation for folded domains. (A) Comparison between λ values from CALVADOS 2 (blue) and opt$_{C\alpha}$ (red). Comparison between simulated and experimental R_g values for IDPs (orange) and MDPs (green) using opt$_{C\alpha}$ in (B) the training set and (C) the validation set. Pearson correlation coefficients (r) and rMSD are reported in the legend. The black diagonal lines indicate $y = x$. Relative difference between experimental and simulated R_g values from opt$_{C\alpha}$ (red), CALVADOS2$_{C\alpha}$ (blue) and CALVADOS2$_{COM}$ (blue hatched) in (D) the training set and (E) the validation set. $\langle\chi^2_{R_g}\rangle$ values across IDPs and MDPs are reported in the legend. Error bars show the experimental error divided by $R_{g,exp}$. Results from CALVADOS2$_{C\alpha}$ and CALVADOS2$_{COM}$ are presented as the same data in ***Figure S4*** and ***Figure 3***.

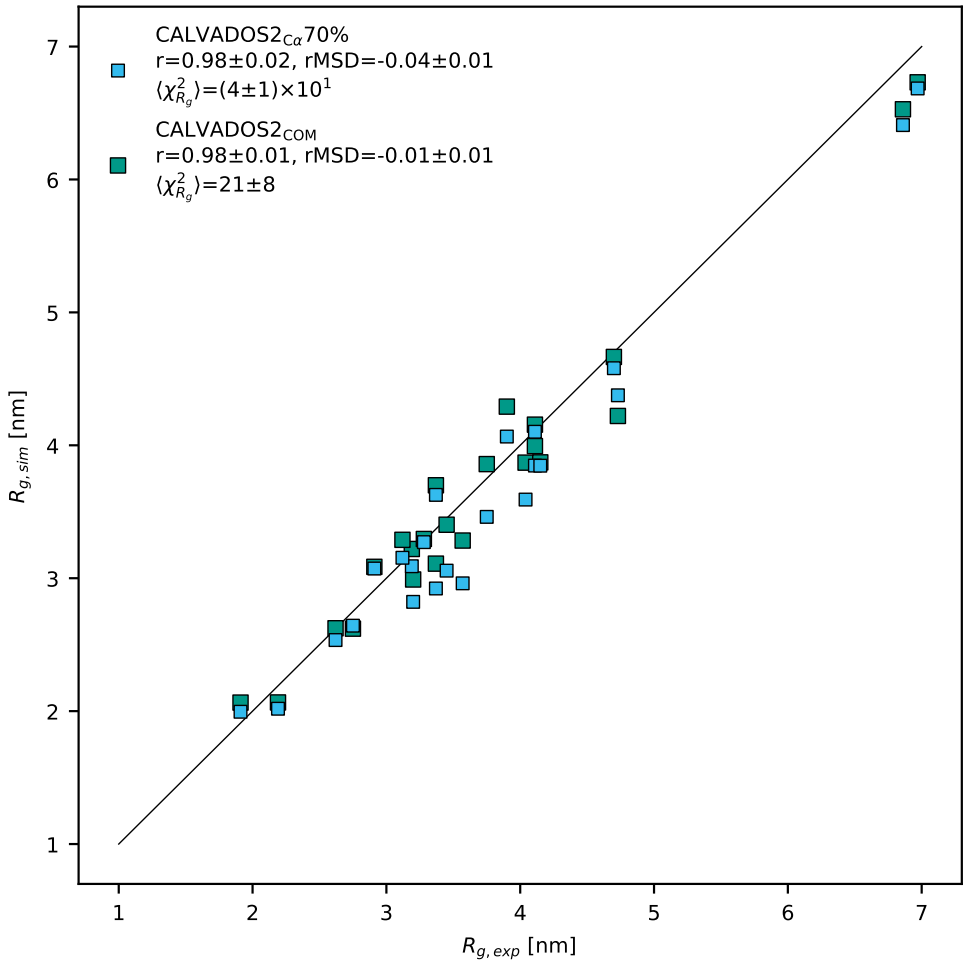

Figure S2. Comparison between simulated and experimental R_g values for all MDPs in the training and validation set using CALVADOS2$_{C\alpha}$70% (cyan) and CALVADOS2$_{COM}$ (green). Pearson correlation coefficients (r), rMSD and $\langle\chi^2_{R_g}\rangle$ values are reported in the legend. The black diagonal line indicates $y = x$.

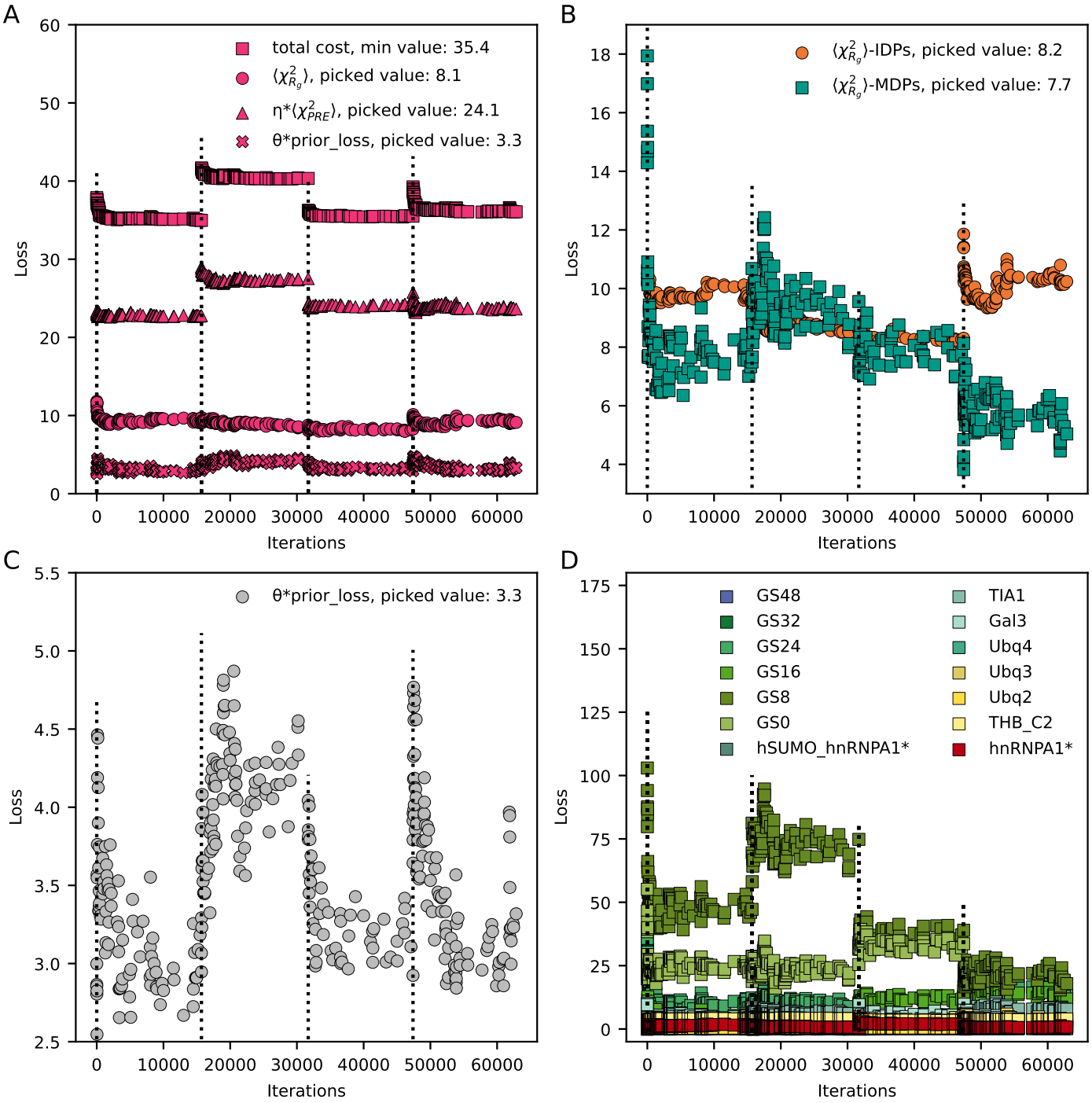

Figure S3. Optimization based on COM representation. The dotted lines indicate the starting points for each cycle. The minimal total cost was used to determine the final λ set and the corresponding values of each term are shown in the legends. (A) Evolution of the total cost (square), $\langle\chi^2_{R_g}\rangle$ (circle), $\eta\langle\chi^2_{\mathrm{PRE}}\rangle$ (triangle) and prior loss (cross) in the optimization, with $\theta = 0.08$ and $\eta = 0.1$. (B) Evolution of $\langle\chi^2_{R_g}\rangle$ contributed from IDPs (orange) and MDPs (green), respectively. (C) Evolution of prior loss. (D) Evolution of $\chi^2_{R_g}$ for each MDP during optimization.

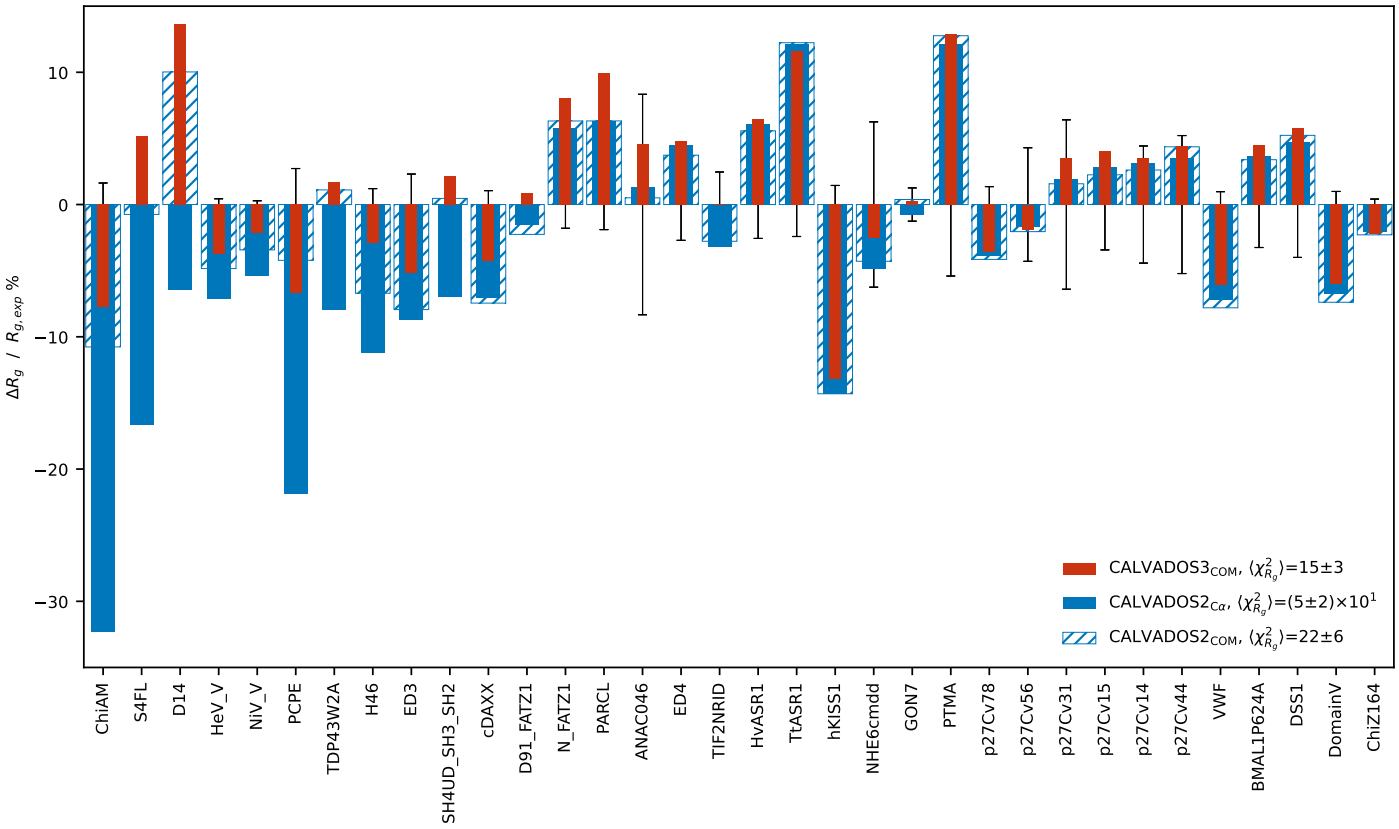

Figure S4. Relative difference between experimental and simulated R_g values from CALVADOS3$_{\text{COM}}$ (red), CALVADOS2$_{C\alpha}$ (blue) and CALVADOS2$_{\text{COM}}$ (blue hatched). $\langle \chi^2_{R_g} \rangle$ values across IDPs and MDPs in validation set are reported in the legend. Error bars show the experimental error divided by $R_{\text{g,exp}}$.

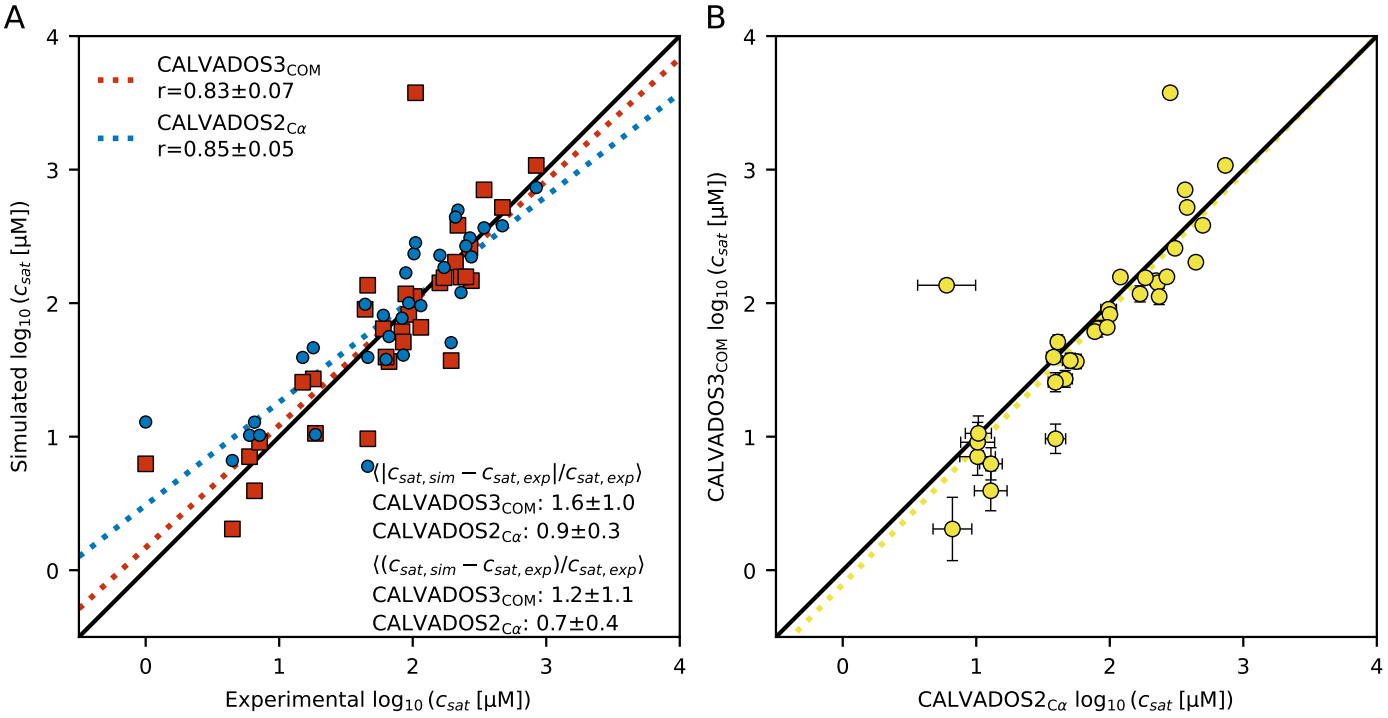

Figure S5. (A) Correlation between c_{sat} from simulations yielded by CALVADOS3$_{\text{COM}}$ (red) or CALVADOS2$_{C\alpha}$ (blue) and experiments for IDPs. (B) Correlation of simulated c_{sat} between CALVADOS3$_{\text{COM}}$ and CALVADOS2$_{C\alpha}$ for the same set of IDPs. Error bars show the error propagation with logarithm from simulated c_{sat}. The black diagonal lines indicate $y = x$. The dotted lines indicate the corresponding linear regression.

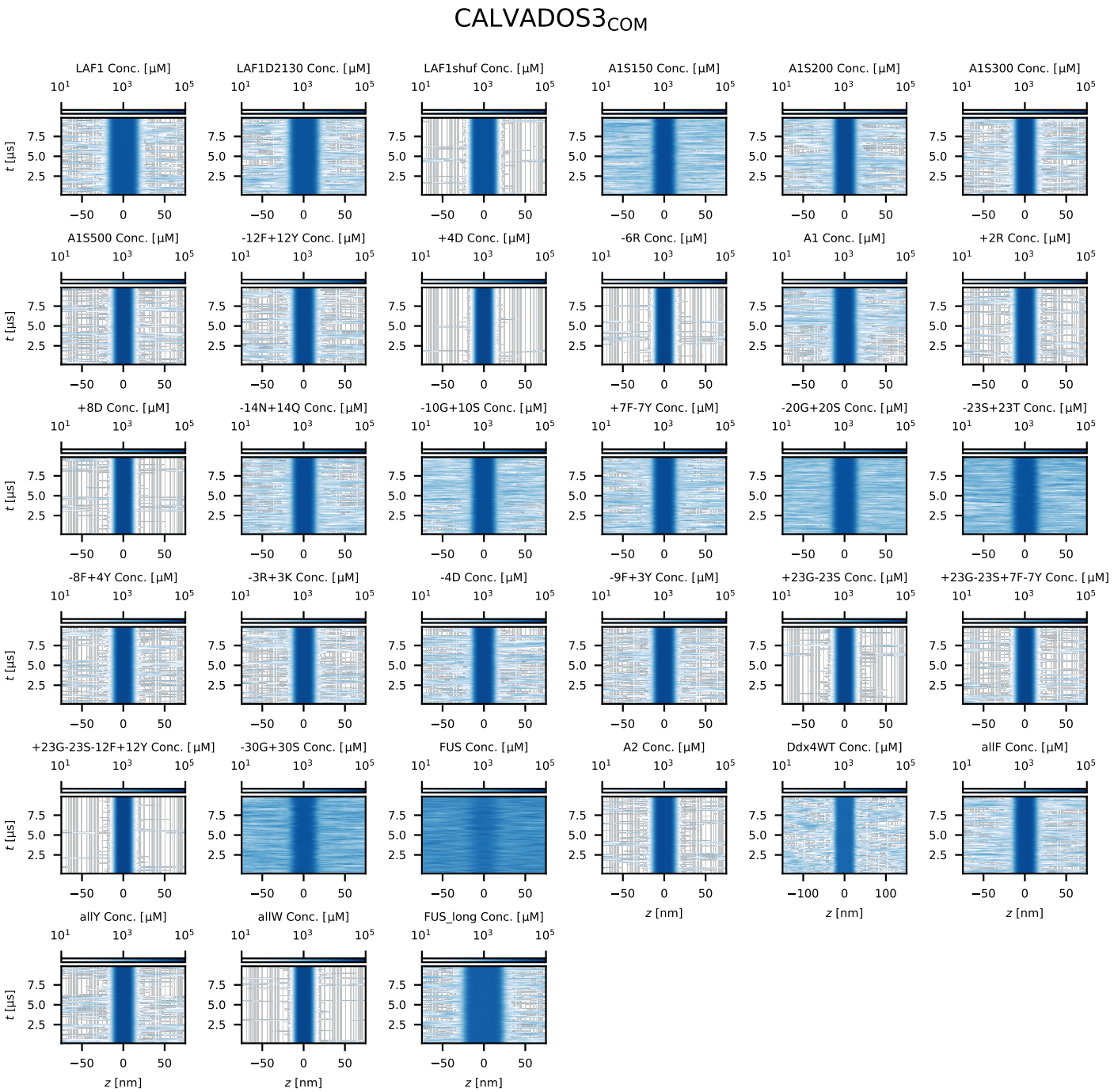

Figure S6. Time evolution of the protein concentration profiles from slab simulations of 33 IDPs using CALVADOS3$_{COM}$ parameters. A more intense colour intensity indicates higher protein concentration.

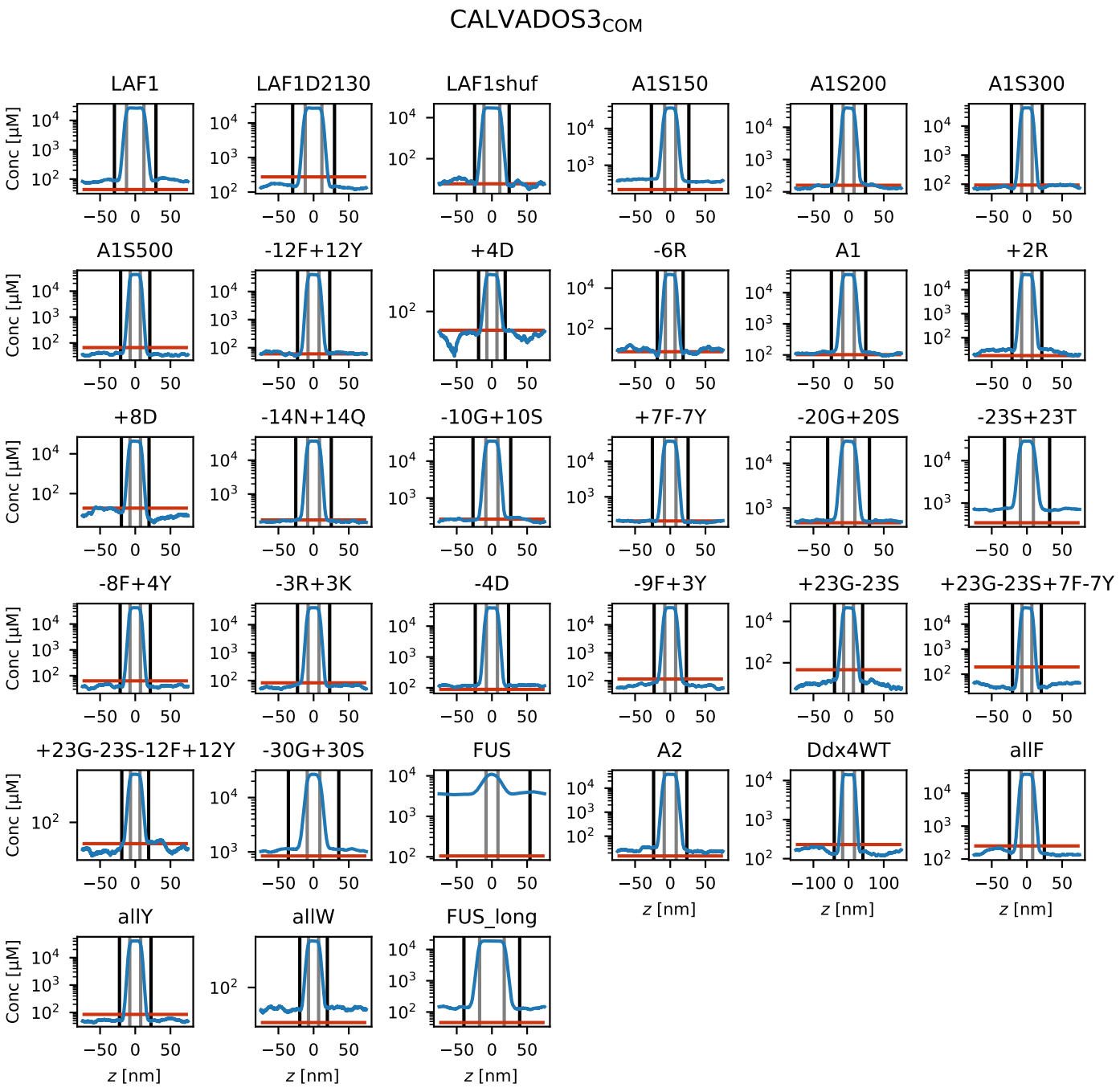

Figure S7. Equilibrium density profiles of slab simulations of 33 IDPs using CALVADOS3$_{COM}$. The red horizontal lines indicate experimental saturation concentrations.

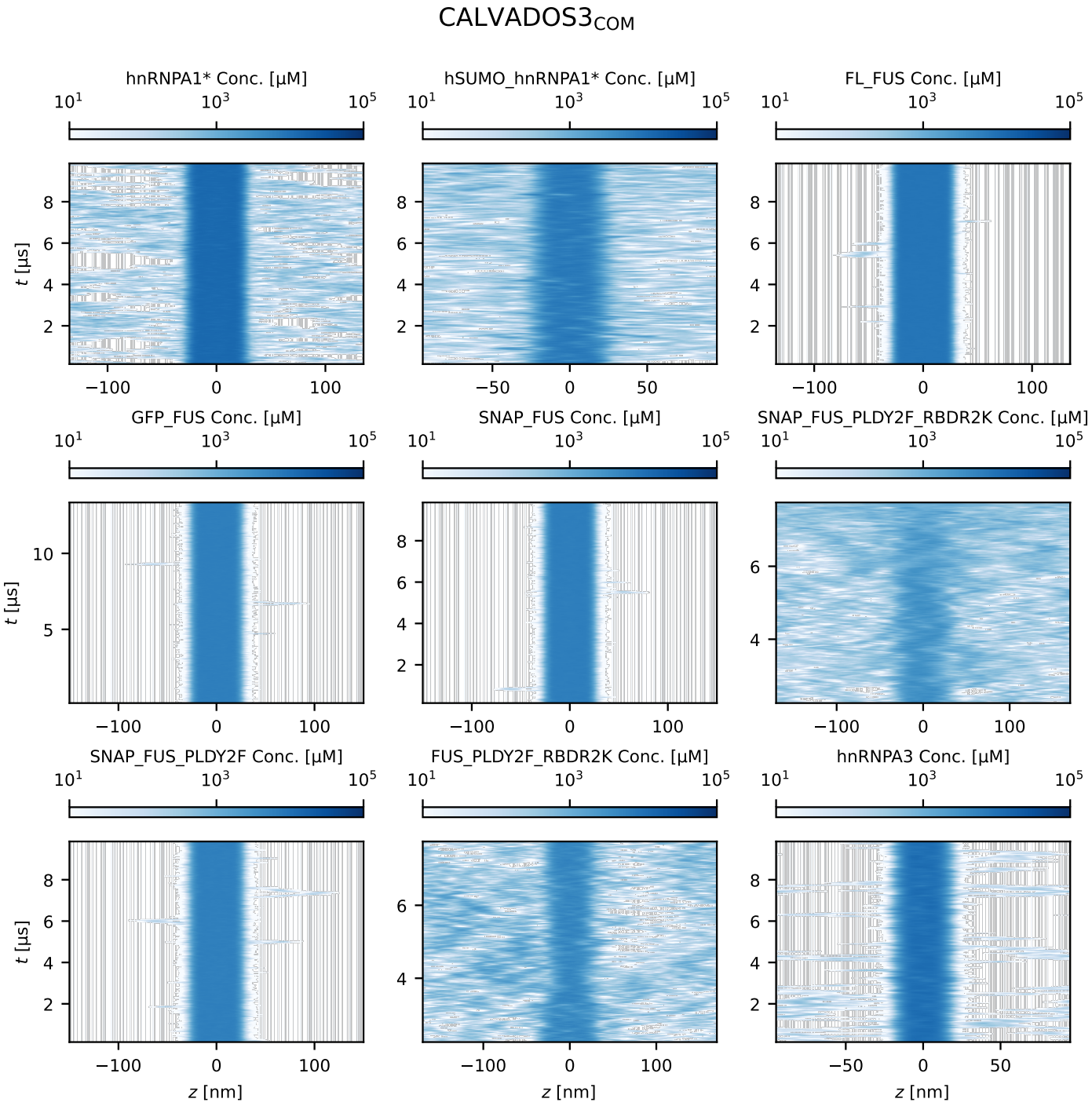

Figure S8. Time evolution of the protein concentration profiles from slab simulations of 9 MDPs using CALVADOS3$_{COM}$ parameters. A more intense colour intensity indicates higher protein concentration.

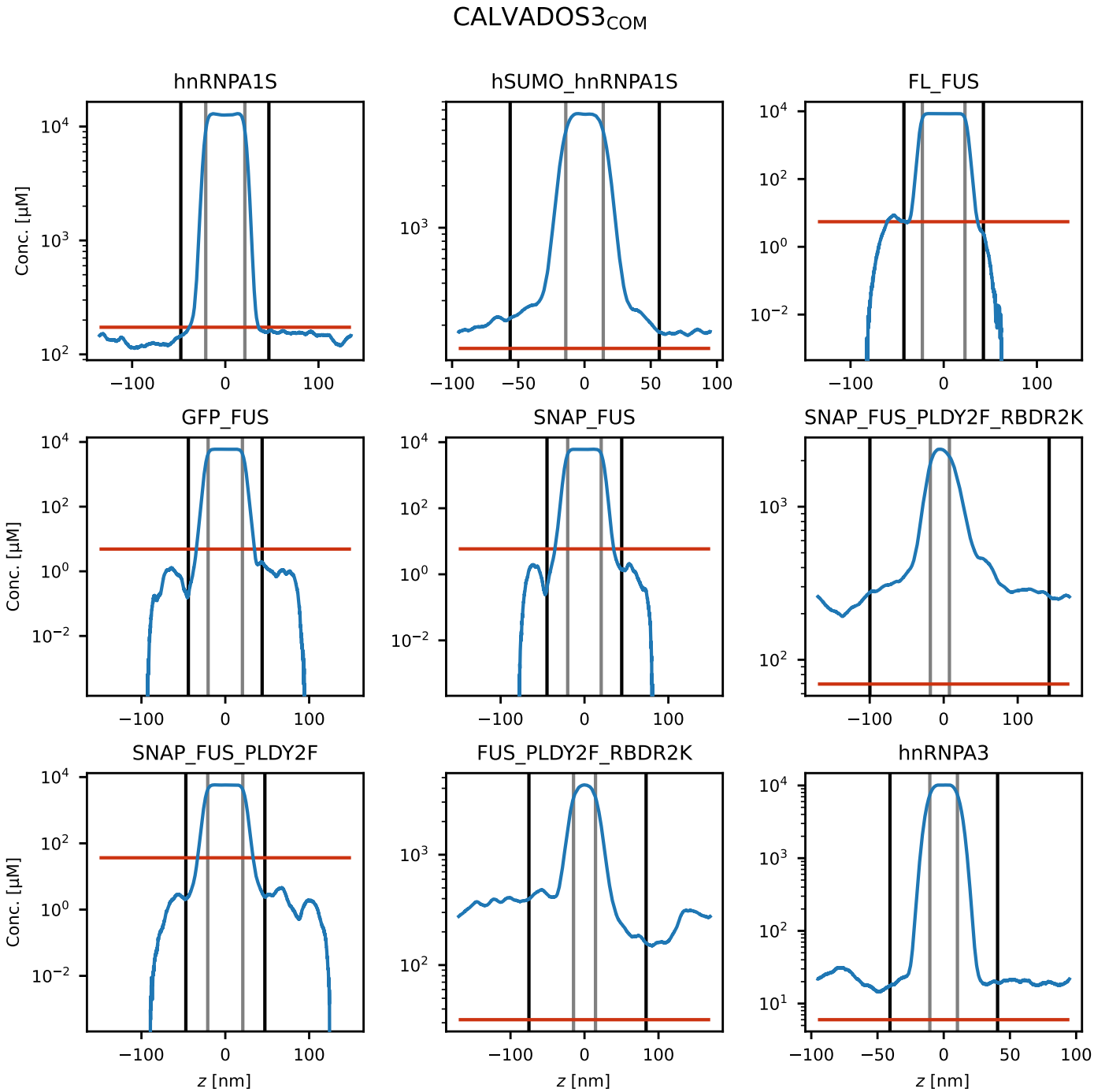

Figure S9. Equilibrium density profiles of slab simulations of nine MDPs using CALVADOS3$_{COM}$. The red horizontal lines indicate experimental saturation concentrations.

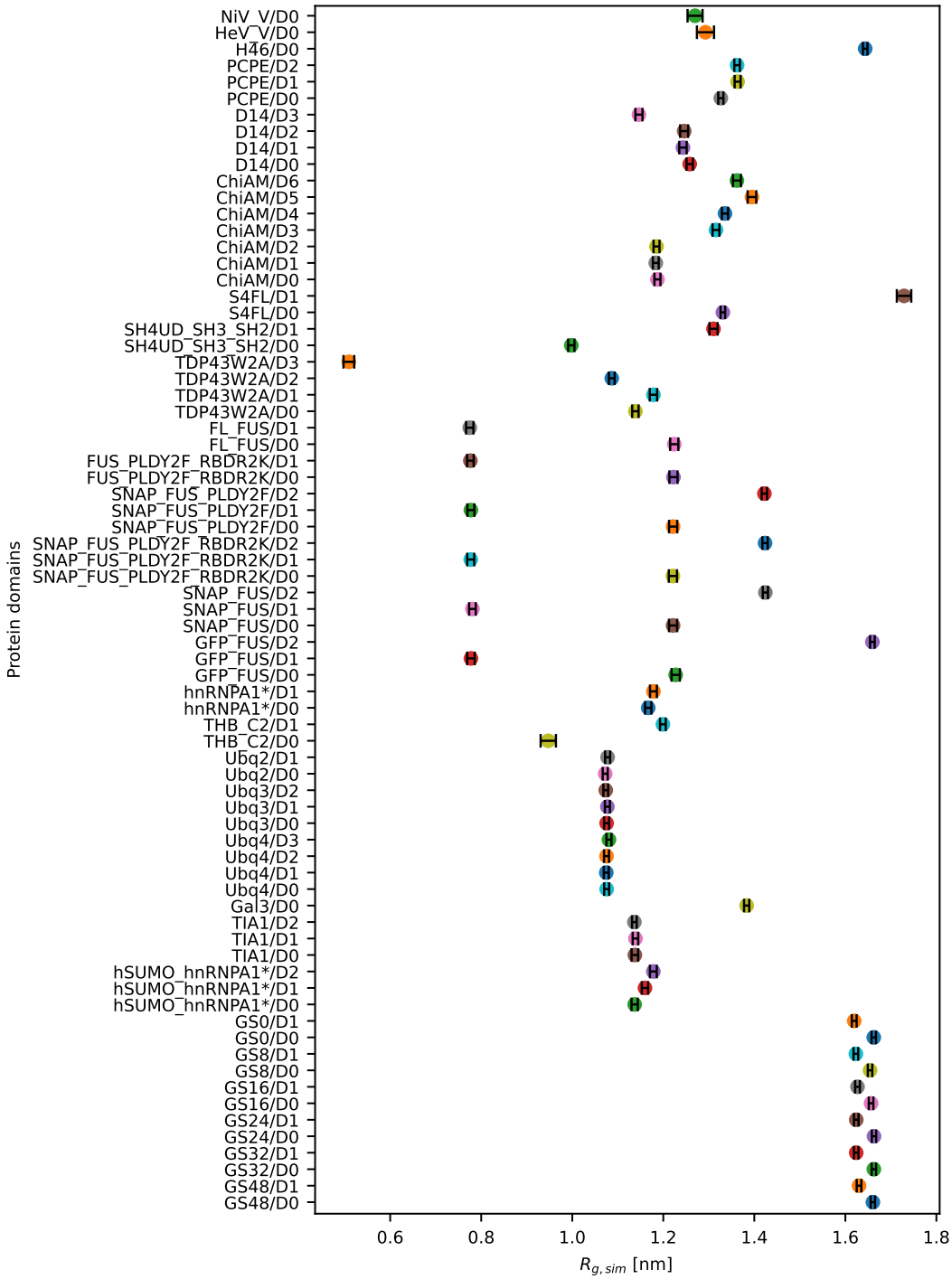

Figure S10. Simulated R_g of domains restrained by elastic network model. Domains in a protein are indicated by D0, D1, D2, etc. Multi-domain proteins in the training set, validation set and slab simulations set are shown.

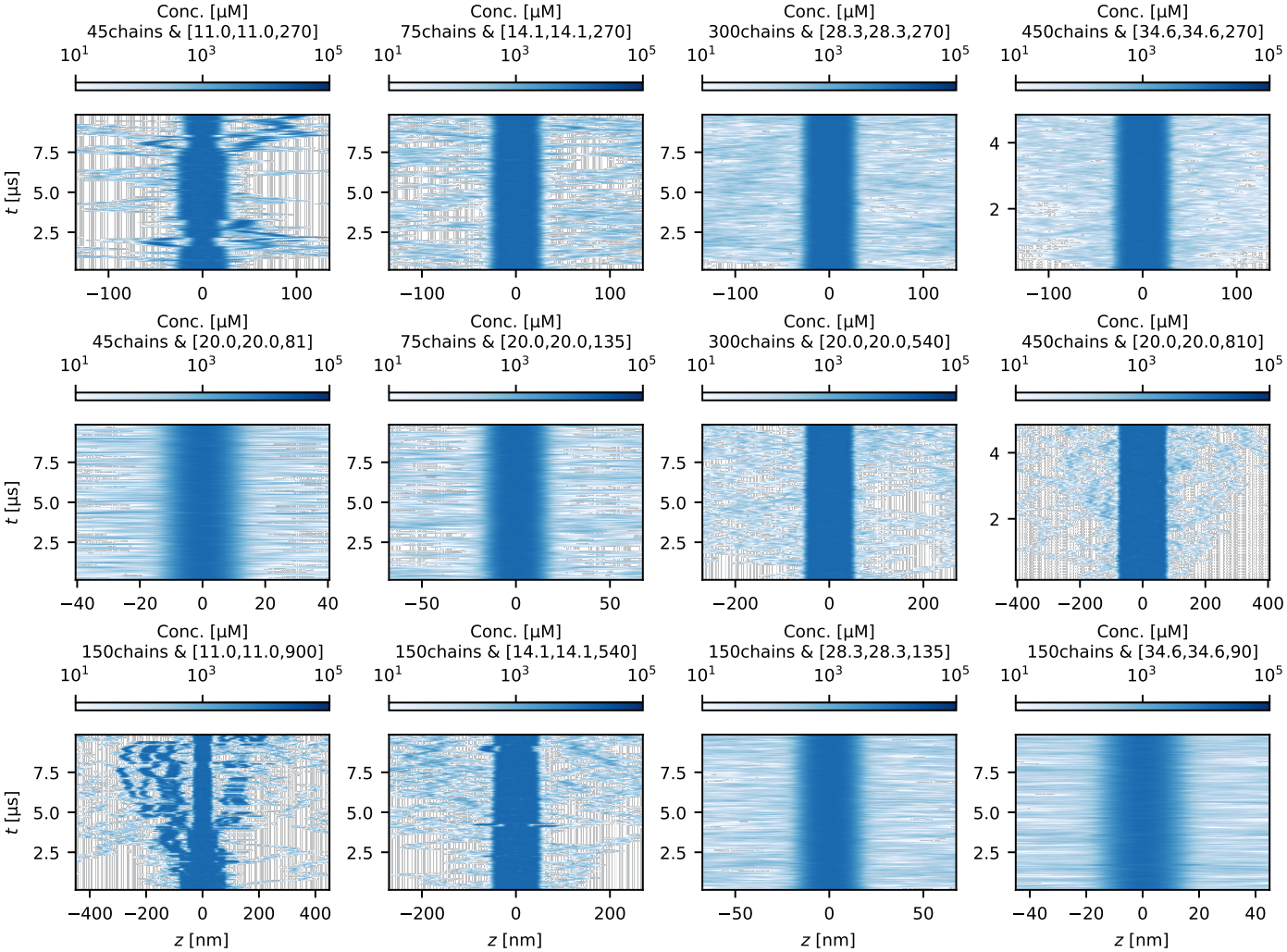

Figure S11. Time evolution of the protein concentration profiles from slab simulations of hnRNPA1* using CALVADOS3$_{COM}$ parameters for analysis of finite-size effects. A more intense colour intensity indicates higher protein concentration. The units of the box sizes are nm.

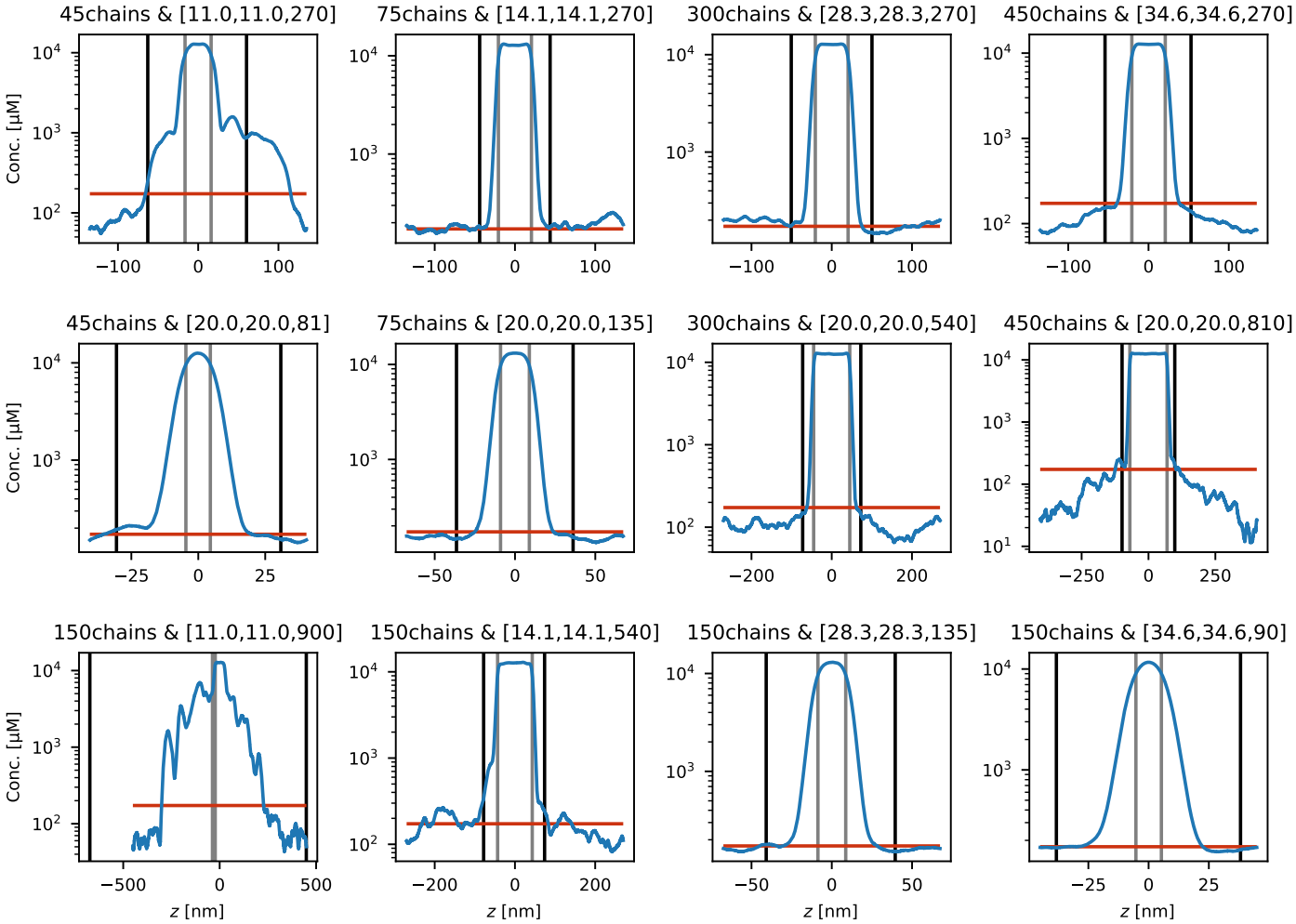

Figure S12. Equilibrium density profiles of slab simulations of hnRNPA1* using CALVADOS3$_{COM}$ for analysis of finite-size effects. The red horizontal lines indicate experimental saturation concentrations. The units of the box sizes are nm.

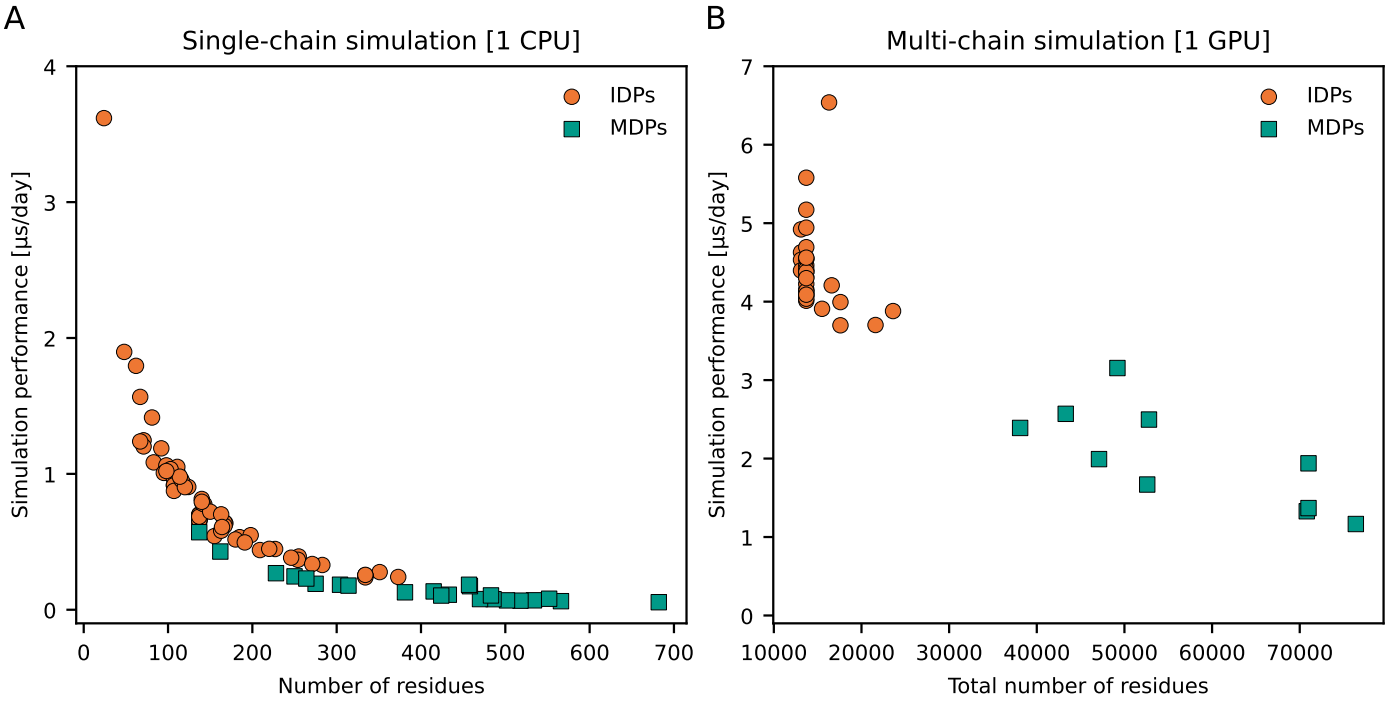

Figure S13. Simulation performance of CALVADOS 3 model on IDPs (orange) and MDPs (green) for (A) single-chain simulations on an Intel Xeon Gold 6130 CPU and (B) multi-chain simulations on an NVIDIA Tesla V100 GPU.