Hill-Robertson interference may bias the inference of fitness effects of new mutations in
highly selfing species

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ABSTRACT

The accurate estimation of the distribution of fitness effects (DFE) of new mutations is critical for population genetic inference but remains a challenging task. While various methods have been developed for DFE inference using the site frequency spectrum of putatively neutral and selected sites, their applicability in species with diverse life history traits and complex demographic scenarios is not well understood. Selfing is common among eukaryotic species and can lead to decreased effective recombination rates, increasing the effects of selection at linked sites, including interference between selected alleles. We employ forward simulations to investigate the limitations of current DFE estimation approaches in the presence of selfing and other model violations, such as linkage, departures from semidominance, population structure, and uneven sampling. We find that distortions of the site frequency spectrum due to Hill-Robertson interference in highly selfing populations lead to mis-inference of the deleterious DFE of new mutations. Specifically, while accounting for the decrease in the effective population size due to linked effects of selection allows an accurate estimation of selection coefficients in moderately selfing populations, this correction is unable to accurately estimate selection coefficients in highly selfing populations when interference between selected alleles is pervasive. In addition, the presence of cryptic population structure with low rates of migration and uneven sampling across subpopulations leads to the false inference of a deleterious DFE skewed towards effectively neutral/mildly deleterious mutations. Finally, the proportion of adaptive substitutions estimated at high rates of selfing is substantially overestimated. Our observations apply broadly to species and genomic regions with little/no recombination and where interference might be pervasive.

INTRODUCTION

The distribution of fitness effects (DFE) of new mutations describes the continuum of selective effects of spontaneous mutations in a population and is critical for understanding the effects of natural selection on genetic variation (Eyre-Walker and Keightley 2007). Because of the considerable impact deleterious mutations can have on allele frequency patterns in a population, knowledge of the DFE is often necessary to produce accurate evolutionary models, which are required for unbiased estimates of both the demographic history of a population and the proportion of adaptive nucleotide substitutions in a lineage (Charlesworth 1994; Eyre-Walker and Keightley 2007; Bank *et al.* 2014; Johri *et al.* 2022a). While the DFE can be estimated with experimental approaches using random mutagenesis or mutation-accumulation lines (*e.g.*, Wloch *et al.* 2001; Estes *et al.* 2004; Sanjuán *et al.* 2004), it can also be inferred from the distribution of allele frequencies from population genetic data (Williamson *et al.* 2005; Eyre-Walker *et al.* 2006; Keightley and Eyre-Walker 2007; Galtier 2016). As many organisms cannot easily be cultured in laboratories, are not genetically manipulatable, and/or have large generation times, a population genetic approach remains an important and the only possible approach to estimating the DFE in some species.

Population genetic methods use putatively neutral sites to fit the demographic history or to account for other potential nuisance parameters, while selected sites are used to infer the DFE (by inferring the parameters of a prespecified probability distribution), conditional on the demographic history obtained from neutral sites (reviewed in Johri *et al.* 2022b). While most two-step population genetic approaches that infer the DFE make a series of assumptions, like random mating and the absence of effects of selection at linked sites, they appear to be robust to certain model violations like mutation rate variability (Tataru *et al.* 2017), linkage (Kim *et al.* 2017; Huang *et al.* 2021), population substructure (Kim *et al.* 2017), and some complex demographic changes (Kousathanas and Keightley 2013). This is because two-step inference methods are usually employed using interdigitated neutral and selected sites and might therefore correct for model violations that skew the neutral and selected SFS similarly. As an aside, because demographic history is unlikely to affect the SFS of neutral and very strongly deleterious mutations equally (Eyre-Walker *et al.* 2006), this might lead to slight inaccuracies in inference using methods that do not explicitly model population size changes. In fact, realistic simulations of chromosomal segments in both *Drosophila* (Huang *et al.* 2021) and humans

(Huber *et al.* 2017) suggest that inference of the DFE of new deleterious mutations remains

accurate, despite the effects of background selection. Similarly, Kim *et al.* (2017) were able to

accurately infer the parameters of the deleterious DFE in a simulated metapopulation that grew

eight times larger and split into eight isolated populations. However, since migration was not

simulated between these populations, the effects of population structure on DFE inference have

not been fully explored.

Despite significant progress, not much work has been done on understanding the effects

of non-random mating, in particular self-fertilization, on the inference of the DFE using

population genetic approaches. Self-fertilization or selfing (*i.e.*, an individual mating with itself)

is common across all eukaryotic species - about 35% of all seed plants and ~50% of all animals

exhibit moderate to high rates (50-100%) of selfing (Goodwillie *et al.* 2005; Jarne and Auld

2006). It is thus important to thoroughly investigate if current methods can be used to obtain

accurate parameters of the DFE in partially selfing organisms. A transition from random mating

to partial selfing has several effects on the population. Firstly, the increase of nonrandom mating

in a partially selfing population decreases the effective population size (N_e), due to a reduction in

the amount of independently sampled gametes (Pollak 1987) . Note that while the amount of drift

experienced by a mutation is larger with a higher rate of selfing, the mean change in allele

frequency at a selected locus is also greater, due to the increased frequency of homozygotes

(Charlesworth and Charlesworth 1987; Caballero and Hill 1992; Charlesworth 1992). There is

no change in the probability of fixation of semidominant mutations, but the probability of

fixation of recessive or partially recessive beneficial mutations is increased by selfing, and the

probability of fixation of recessive or partially recessive deleterious mutations is slightly reduced

(Caballero and Hill 1992; Charlesworth 1992). Thirdly, increased homozygosity in the genomes

of partially selfing organisms results in a higher proportion of recombination events occurring on

nearly identical chromosomes, which decreases the effective rate of recombination (Nordborg

2000). This effect increases the extent of background selection (BGS; Charlesworth *et al.* 1993;

Charlesworth and Charlesworth 1998; Charlesworth 2003) and selective sweeps (Hedrick 1980;

Hartfield and Bataillon 2020) experienced by the population. Finally, as the effective rate of

recombination is lowered drastically in highly selfing populations, there can be a further decrease

in the efficacy of selection due to Hill-Robertson interference (HRI) between selected loci (Hill

and Robertson 1966; McVean and Charlesworth 2000; Comeron and Kreitman 2002; Hartfield

and Glémin 2014, 2016).

Hill-Robertson interference (HRI) describes how the efficacy of selection at an allele at

one locus can be reduced due to the segregation of selected alleles at a second linked locus (Hill

and Robertson 1966; Felsenstein 1974). While BGS and selective sweeps are also a type of HRI,

those models assume that selection is strong and that there is no interference between selected

sites (reviewed in Charlesworth and Jensen 2021). However, because weakly selected mutations

can segregate in the population for longer times, interference between them can become

important especially if they are sufficiently prevalent (McVean and Charlesworth 2000;

Comeron and Kreitman 2002). Such interference between weakly selected alleles can result in

reduced efficacy of selection and has been termed as interference selection (Comeron *et al.* 1999;

Comeron and Kreitman 2002) or weak selection HRI (McVean and Charlesworth 2000; Kaiser

and Charlesworth 2009). Moreover, the effects of such interference have been shown to extend

to more strongly selected mutations in cases where recombination is greatly reduced (Kaiser and

Charlesworth 2009). We therefore refer to the effects of interference selection/weak selection

HRI as simply HRI in this study. Since the entire genomes of populations with high selfing rates

have greatly reduced effective recombination rates (Nordborg 2000) and deleterious mutations

are prevalent across the genome (Bank *et al.* 2014), HRI is likely to play an important role in

determining the dynamics of deleterious mutations in such populations.

Unlike the model of classical BGS, where deleterious alleles are at deterministic

mutation-selection equilibrium, selected sites in populations or genomic regions with pervasive

HRI depart from the standard equilibrium (Comeron and Kreitman 2002; Kaiser and

Charlesworth 2009; Seger *et al.* 2010; O'Fallon *et al.* 2010; Nicolaisen and Desai 2013). The site

frequency spectra (SFS) of neutral and selected sites affected by HRI are skewed towards rare

variants and become more similar to each other as the strength of HRI increases, and these

changes cannot be summarized as a simple change in effective population size (Comeron and

Kreitman 2002; Kaiser and Charlesworth 2009; Seger *et al.* 2010; O'Fallon *et al.* 2010;

Nicolaisen and Desai 2013; Good *et al.* 2014). More recent work assuming equal selection

coefficients across deleterious alleles has shown that when HRI is sufficiently common, the

nucleotide site diversity can be entirely described by the variation of fitness between individuals

in the population. However, populations with different combinations of average selection

coefficients, recombination, and mutation rates can have the same nucleotide diversity in this

regime (Good *et al.* 2014), suggesting difficulties in estimating a distribution of selection

coefficients along with other population genetic parameters in populations with little/no

recombination. While, a recent study suggested that the population-scaled fitness effects of

mutations are mis-inferred due to linked effects of selection when the probability of selfing is

sufficiently high (Gilbert *et al.* 2022), the role of HRI (specifically interference) remains unclear.

Here, we thoroughly characterize how HRI and other linked effects of selection might

bias the inference of both population-scaled as well as direct estimates of selection coefficients in

highly selfing species and in regions of low recombination for a variety of DFE shapes. To

investigate other potential causes of DFE mis-inference in partially selfing natural populations,

we explore the effects of changing the dominance coefficients, coding density, and number of

selected sites, as well as the effects of cryptic population structure and sampling on DFE

inference. Moreover, we characterize the accuracy of inference of both the DFE of new

deleterious mutations as well as the proportion of adaptive substitutions.

METHODS

Simulations

All simulations were performed using the forward simulator SLiM (version 4.0.1; Haller and

Messer 2023). The genome architecture and population parameters were modeled to mimic the

genomes of *C. elegans* populations because they have a high rate of selfing, which has been

estimated to be 98%-99.9% using laboratory observations (Hodgkin 1983; Anderson *et al.* 2010),

field work, and population genetic analyses (Barrière and Félix 2005, 2007). A chromosome with

an intron-exon structure was simulated such that each gene consisted of 6 exons (250 bp in

length) and 5 introns (100 bp in length). Genes were interspersed with intergenic regions of

length 3000 bp each. These numbers were close to the median of the length of exons, introns,

and intergenic regions observed in the *C. elegans* genome with a gene density of ~1 gene every

5kb (*C. elegans* Sequencing Consortium 1998). The *C. elegans* genome comprises 30% coding,

25% intronic, and 45% intergenic regions (*C. elegans* Sequencing Consortium 1998). In line

with these genome-wide characteristics, our simulated genome comprised of 30% coding, 10%

intronic, and 60% intergenic regions, and a total of 500 genes, resulting in a full chromosomal

length of 2,503,000 bp. All introns and intergenic regions were assumed to be neutral, while

exons experienced selection.

To take synonymous sites into account, 25% ($= f_{neu}$) of all mutations in exons were

assumed to be fully neutral (with $s=0$) while the rest were under selection. Of the selected

mutations, a fraction f_{pos} were assumed to be beneficial, where f_{pos} was assumed to be either

0.0, 0.01, or 0.001, and the rest ($1 - f_{pos} - f_{neu}$) were deleterious. The DFE of deleterious

mutations was assumed to be a gamma distribution, parametrized by the shape parameter (β) and

the mean population-scaled selection coefficient ($\gamma = 2N_e \overline{s_d}$), where $\overline{s_d}$ is the mean selective

disadvantage of the mutant homozygote relative to wildtype. Three different deleterious DFEs

were modeled, representing fitness effects skewed towards mildly, moderately, and strongly

deleterious classes respectively (Table 1). The DFE of beneficial mutations was assumed to be

exponential with mean $2N_e \overline{s_a} = 200$, where $\overline{s_a}$ is the mean selective advantage of the mutant

homozygote relative to wildtype. All mutations were assumed to be semidominant (where $h =$

0.5) unless otherwise specified.

A Wright-Fisher population under equilibrium was simulated with a population size (N_e)

of 500,000 diploid individuals (following Gilbert *et al.* 2022) and with varying probabilities of

selfing – 0%, 50%, 80%, 90%, 95%, and 99%, where $N_e = N$ for 0% selfing. Mutation (μ) and

recombination (r) rates were assumed to be constant at 3.3×10^{-9} per site/generation (Denver *et*

al. 2004; Konrad *et al.* 2019; Saxena *et al.* 2019) and 3.12×10^{-8} per site/generation (Rockman

and Kruglyak 2009), respectively. Simulations were conducted with a population size of 5000

diploid individuals, with a rescaling factor (Q) of 100, and rates of mutation and recombination

as well as selection coefficients were correspondingly adjusted so that the population-scaled

parameters $N\mu$, Nr, and Ns were equivalent to a population of 500,000. Simulations were run

for a total of $14N$ generations, where $10N$ generations corresponded to the burn-in period and

$4N$ generations were used to calculate the number of substitutions and where N represents the

scaled population size. Five replicates of each evolutionary scenario were simulated and the DFE

was estimated for each replicate separately.

Single-locus simulations with selfing: To investigate the effects of selfing on DFE inference

without linkage, selected, semidominant mutations were simulated at single sites with

probabilities of selfing of 0%, 50%, and 99%. The s_d of deleterious mutations were drawn from

the gamma distributions shown in Table 1. As allele frequencies of neutral mutations are

unaffected by linkage in the absence of selected mutations, 187,500 neutral sites were simulated,

which is equivalent to the number of neutral sites simulated in the simulations of 500 *C. elegans*

genes. 50,000 selected sites were simulated for each scenario (compared to 562,500 selected sites

in previous simulations), then paired with neutral mutations simulated with the appropriate

selfing rate. The simulations of individual selected sites and the neutral sites used rescaled values

of N, μ, and r equal to 5,000 individuals, 3.3×10^{-7} per site/generation and 3.12×10^{-6} per

site/generation respectively, which is equivalent to the previous simulations with linkage. All

simulations were run for $14N$ generations with $10N$ burn-in generations.

Simulations under low recombination rates with no selfing: To clarify if the lower effective

recombination rates in a partially selfing population were responsible for DFE mis-inference, we

conducted simulations with low recombination rates and 0% selfing. As before, these simulations

were conducted assuming a census size of 500,000 diploid individuals and rescaled by a factor of

100 to 5,000 individuals. The *C. elegans* recombination rate of 3.12×10^{-8} was thus rescaled to

3.12×10^{-6}, and multiplied by 0.5, 0.1, 0.05, 0.01, 0.005, or 0.001 to determine a final

simulated recombination rate. Beneficial mutations were not included in these simulations.

Simulations with varying dominance coefficients: All selected mutations shared a single

dominance coefficient of 0.1, 0.25, or 0.75. Populations with selfing probabilities of 0%, 50%,

and 99% were simulated and compared to the original simulations (where $h = 0.5$).

Simulations under varying coding density and chromosome size: Simulations were performed

with varying coding densities by varying the intergenic lengths - 1500 bp and 500 bp. The total

number of genes was not increased in these simulations. Thus, in the simulations with 1500 bp

intergenic regions, the simulated genome was comprised of ~43% coding, ~14% intronic, and

~43% intergenic regions, with a total length of 1,752,100 bp. In the simulations with 500 bp

intergenic regions, the genome was comprised of ~55% coding, ~18% intronic, and ~27%

intergenic regions, with a total length of 1,376,650 bp. These simulations were run with

probabilities of selfing equal to 0%, 50%, and 99%.

To study the effects of a more realistically sized chromosome on DFE inference in
selfers, simulations where the total number of genes was increased from 500 to 3000 were
conducted, corresponding to approximately the number of genes observed on the first
chromosome of the *C. elegans* genome. This increased the size of the simulated chromosome to
15,018,000 bp, which is close to the true size of *C. elegans* chromosome I (15,072,434 bp)
(https://www.ncbi.nlm.nih.gov/datasets/genome/GCF_000002985.6/; last accessed in Dec.
2023).

Simulations with varying rescaling factors: To investigate the impact of rescaling of population
sizes in our forward simulations, simulations were conducted with the rescaling factor $Q = 50$,
20, and 10, where the population-scaled parameters $N\mu$, $Nr,$ and Ns were equivalent to a
population of 500,000. Simulations were run for a total of $14N$ generations, where $10N$
generations corresponded to the burn-in period and $4N$ generations were used to calculate the
number of substitutions and where N represents the scaled population size. Due to the excessive
run time of unscaled simulations, simulations were only run for populations with selfing rates of
95% and 99%. For $Q = 10$, only one replicate of DFE3 at 95% selfing and three replicates each
for DFE2 and DFE3 at 99% selfing reached completion.

Quantifying the effect of BGS in simulated data
To determine the theoretically expected neutral nucleotide diversity (π) in simulations with
partial selfing, we calculated the expected effective population size after correcting for selfing
(N_{self}) as $N_{self} = N/(1 + F)$, where $F = P_{self}/(2 - P_{self})$, and P_{self} represents the proportion
of selfing individuals in a population per generation (Pollak 1987). Without the linked effects of
selection, the neutral π is expected to be approximated by $\pi \sim \theta = 4N_{self}\mu$. In simulations with
deleterious mutations, the extent of BGS was quantified as the observed nucleotide diversity
(π_{obs}) at neutral sites relative to that expected under strict neutrality, *i.e.*,

$$B = \frac{\pi_{obs}}{4N_{self}\mu} \tag{1}$$

Simulations with population structure

To study the effects of population structure on DFE inference in selfers, a finite island model

(Maruyama 1970) with five demes and an equal probability of exchange of migrants between all

demes was considered. Note that in this model the migration rate is independent of the selfing

rate, which applies well to animals but is not necessarily the case in plant species because selfing

can prevent pollen dispersal (reviewed in Sicard and Lenhard 2011). Scenarios with different

migration rates were simulated such that the effective metapopulation size remained constant and

equivalent to the size of the panmictic population simulated above. Genome structure was

identical to that in the previous simulations mimicking *C. elegans* genome architecture. In a

metapopulation with d demes and a deme effective population size N_{deme}, the mean coalescence

time for a pair of alleles sampled from the same deme (T_S) is given by the following equation

(Nei 1973; Slatkin and Voelm 1991; Charlesworth and Charlesworth 2010):

270
$$T_S \sim 2dN_{deme} \tag{2}$$

273 The mean coalescence time for a pair of alleles sampled from different demes (T_B) is given by

274 (Nei 1973; Slatkin and Voelm 1991; Charlesworth and Charlesworth 2010):

276
$$T_B \sim 2dN_{deme} + \frac{(d-1)}{2m} \tag{3}$$

279 where the migration rate m is the probability that an allele in one deme is derived by migration

280 from any other deme in a generation (which means that the migration rate between any two

281 demes is equal to $\frac{m}{d}$). Combining these two equations gives the mean coalescence time (T_T) for a

282 pair of alleles drawn from the entire metapopulation (Nei 1973; Slatkin and Voelm 1991;

283 Charlesworth and Charlesworth 2010):

285
$$T_T \sim 2dN_{deme} + \frac{(d-1)^2}{2dm} \tag{4}$$

To keep the effective size of the metapopulation ($0.5T_T$) equivalent to the rescaled size of

5,000 individuals used in previous simulations, N_{deme} and m were varied such that T_T remained

equal to 10,000. To allow a wide range of migration rates, $N_{deme}m$ was varied from 10, 2, 1, 0.5,

and 0.1 in populations with 0%, 50%, and 99% selfing, with separate simulations for DFE1,

DFE2, and DFE3. The exact parameters used in each simulation can be found in Supplementary

Table 7. Parameters were chosen so that the predicted F_{ST} of the entire metapopulation ($F_{ST} =$

$\frac{T_T - T_S}{T_T}$) would cover a range of values (from 0.02 to 0.76) matching previously estimated values

for *C. elegans* populations in the literature (from 0.15 to 0.43; Cutter 2006). Neutral simulations

with identical genome structure and migration parameters were used to verify the joint effects of

population structure and selfing on neutral nucleotide diversity and F_{ST}, and were found to yield

expected results (Supplementary Table 8). The expected neutral nucleotide diversity for partially

selfing populations was calculated assuming that the effective size of each deme will be

equivalent to $N_{deme,self} = \frac{N_{deme}}{1+F}$, which was used to calculate the appropriate T_T and T_S values

for each evolutionary scenario. In SLiM, m represents the proportion of individuals in a deme

that are migrants each generation. Therefore, m still represents the probability that an allele is

derived from migration each generation, and $N_{deme}m$ represents the number of diploid migrants

per deme per generation.

Twenty genomes were sampled from each deme (totaling 100 genomes from the

metapopulation) for DFE inference. To understand the effects of uneven sampling on DFE

inference, two alternative sampling schemes were employed. In the first, 60 genomes were

sampled from one deme, while 10 genomes were drawn from the remaining four demes. In the

second, 35 genomes were drawn from two demes, while 10 genomes were drawn from the

remaining three demes.

Calculating the SFS and the number of adaptive substitutions from simulations

The full output provided by SLiM was used to generate the site frequency spectra of neutral and

selected mutations in exonic regions individually. The fixed class was obtained as the number of

substitutions that occurred after the burn-in period ($= 10N$ generations). The zero class was

calculated by subtracting the number of polymorphic and fixed sites from the total number of

sites for both neutral (= 0.25 × number of exonic sites) and selected (= 0.75 × number of

exonic sites) mutations respectively. Scripts to calculate the SFS are available at

https://github.com/JohriLab/DFESelfing/tree/main/calculateSFS.

The true fraction of beneficial substitutions (α) in simulations was obtained as follows:

$$\alpha = \frac{number\ of\ substitutions\ with\ 2N_e s_a > 5}{number\ of\ substitutions\ at\ selected\ sites} \qquad (5)$$

Only substitutions that occurred after the burn-in with $2N_e s_a > 5$ were included when

calculating the true value of α in order to match the assumptions of GRAPES.

DFE-alpha

Est_dfe (Keightley and Eyre-Walker 2007) was employed to infer the DFE of deleterious

mutations. We fitted a two-epoch model, with starting values of $t2$ (time of change), β, and s_d

equal to 50, 0.5, and -0.01 respectively. $N2$ (current size) and $t2$ were also set to be variable. In

the absence of positive selection, the unfolded SFS was used as input, and the fold setting was

set to 0. In the presence of beneficial mutations, the folded SFS was used instead. DFE-alpha

provides the most likely estimate of the expected value of $\overline{s_d}$, the shape parameter (β) of the

gamma distribution, and a weighted effective population size (N_w). These parameters were used

to obtain the proportion of mutations (f_0, f_1, f_2, f_3) in the four classes- effectively neutral ($0 \leq$

$2N_w s_d < 1$), weakly deleterious ($1 \leq 2N_w s_d < 10$), moderately deleterious ($10 \leq 2N_w s_d <$

100), and strongly deleterious ($100 \leq 2N_w s_d$), respectively.

GRAPES

The program GRAPES (Eyre-Walker and Keightley 2009; Galtier 2016), Version 1.1.1

(https://github.com/BioPP/grapes/releases/tag/v1.1.1) was used to infer the full DFE using

simulations with and without positive selection. GRAPES provides the maximum likelihood

estimate of the shape parameter (β) and the expected population-scaled mean strength of

selection in terms of $2N_e \overline{s_d}$, in order to characterize the gamma distribution representing the

deleterious DFE. GRAPES also infers the parameters of the beneficial DFE, including the

proportion of substitutions that are beneficial mutations (α) and the mean population-scaled

strength of selection favoring new advantageous mutations $2N_e\bar{s_a}$, assuming that advantageous

selection coefficients are exponentially distributed. The unfolded SFS was provided as input for

both scenarios (with and without positive selection). In simulations without positive selection,

the default parameters for the GammaZero model were used (*i.e.*, parameters of the beneficial

DFE were not inferred). In the presence of positive selection, the estimated beneficial DFE was

assumed to follow an exponential distribution (*i.e.*, the GammaExpo model was used), consistent

with our simulations. As GRAPES can use either the folded or the unfolded SFS to provide

estimates of the beneficial DFE, we performed inference using the unfolded and folded SFS

separately to compare the results of both estimation methods. These parameters were accordingly

used to obtain the values f_0, f_1, f_2, f_3, representing the classes of the deleterious DFE as

described above.

Comparison of the expected and estimated DFE when mutations are semidominant

In a partially selfing population, diffusion theory provides the following mean ($M_{\Delta q}$) and

variance ($V_{\Delta q}$) of the change in allele frequency (q) assuming that the change (Δq) in allele

frequency is small (Caballero *et al.* 1991):

$$M_{\Delta q} = q(1-q)s[h + q(1-2h) + F(1-q-h+2qh)] \qquad (6)$$

$$V_{\Delta q} = \frac{q(1-q)}{2N_{self}} = \frac{q(1-q)(1+F)}{2N} \qquad (7)$$

For completely additive mutations ($h = 0.5$), the mean reduces to:

$$M_{\Delta q} = \frac{q(1-q)s(1+F)}{2} \qquad (8)$$

In a panmictic population (of size N) with random mating, the mean and variance of change in

allele frequency (assuming semidominance) is given by:

$$M_{\Delta q} = \frac{q(1-q)s}{2} \tag{9}$$

$$V_{\Delta q} = \frac{q(1-q)}{2N} \tag{10}$$

Thus, by comparing the above set of equations, a selfing population of size N experiences more

genetic drift than a randomly mating population of the same size (Caballero and Hill 1992).

However, the mean change in allele frequency in selfing populations is also larger, by the same

factor, $(1 + F)$, and thus the fixation probability of mutations remains the same in a selfing *vs.* a

randomly mating population (Charlesworth and Charlesworth 1987; Caballero and Hill 1992;

Charlesworth 1992). Thus, for the case of additive mutations and not taking selection at linked

sites into account, the expected DFE would be determined by $2N_{self}s_d(1 + F) = 2\frac{N}{(1+F)}s_d(1 +$

$F) = 2Ns_d$. To account for the relative decrease in population size due to BGS or selective

sweeps, the simulated DFE was also adjusted and presented as $2NBs_d$, where B was calculated

using equation 1.

   In the case of the estimated DFE, because DFE-alpha only infers a relative increase or

decrease in population size ($\frac{N_2}{N_1}$), these estimates of demographic parameters should not be

affected by selfing rates (where the factor $1 + F$ will cancel out). We thus present our inferred

DFE as $2N_w s_d$.

Comparison of the expected and estimated DFE when mutations are not semidominant

When mutations were not additive, our expected DFE was parameterized by $2N_{self}s_d h_{self}$,

where

$$N_{self} = \frac{N}{1 + F} \qquad (11)$$

and

$$h_{self} = h + q(1 - 2h) + F(1 - q - h + 2qh) \qquad (12)$$

Assuming that $q \ll 1$, this equation simplifies to:

$$h_{self} \sim h + F(1 - h) \qquad (13)$$

Note that this approximation is less accurate when selection is weak ($2Ns_d \sim 1$) and thus might not be as accurate for the DFE comprising predominantly weakly deleterious mutations. The estimated DFE (using DFE-alpha and GRAPES) was parameterized by $2N_w s_d h$ where $h = 0.5$ is assumed by both the programs.

Calculation of summary statistics from simulated data

In order to quantify the effects of HRI, the skew in the SFS towards rare alleles was measured using the summary statistic $\Delta\theta_w = 1 - (\frac{\pi}{\theta_w})$, which has been demonstrated to be less sensitive than Tajima's D to the number of SNPs used in the calculation (Becher *et al.* 2020). Here, π is the nucleotide diversity (Nei and Tajima 1981) and θ_w denotes Watterson's θ (Watterson 1975). Both summary statistics were calculated in nonoverlapping 5000 bp windows using *pylibseq* v. 0.2.3 (Thornton 2003). To summarize linkage disequilibrium, D' was calculated for pairs of polymorphisms at frequencies between 1% and 5% in each window. Only low-frequency SNPs were used for D' calculations to correct for the allele frequency dependence of linkage disequilibrium statistics (Lewontin 1988). Here D and D' Lewontin (1964) were computed by *pylibseq* as follows:

$$D = p_{AB} - p_A p_B \qquad (14)$$

and

$$D' = \frac{D}{D_{max}}$$ (15)

where

if $D > 0$

$$D_{max} = \min(p_A p_b, p_a p_B)$$ (16)

if $D < 0$

$$D_{max} = \min(p_A p_B, p_a p_b)$$ (17)

where p_A and p_B represent the minor allele frequency for each polymorphism, p_a and p_b represent the major allele frequency, and p_{AB} represents the frequency of the haplotype AB. All summary statistics were calculated separately for neutral and selected polymorphisms in exonic regions.

RESULTS

Inference of the DFE of new deleterious mutations in self-fertilizing populations

We simulated a single *C. elegans*-like population with varying rates of selfing – 0, 50, 80, 90, 95, and 99% and three different DFEs (following a gamma distribution; parameters specified in Table 1) skewed towards mildly (DFE1), moderately (DFE2), and strongly deleterious (DFE3) classes respectively. Here we assumed that all new mutations were semidominant. We used DFE-alpha and GRAPES to infer the shape of the deleterious DFE, specified to be gamma-distributed. As expected, when there is no selfing, inference was accurate (Figure1, Supplementary Figure 1, Supplementary Table 1, Supplementary Table 2). As the selfing rate increased ($\geq$ 90%), the estimated DFE was skewed strongly towards the class of mildly deleterious mutations (Figure 1), when compared to the simulated DFE, with a particularly strong skew in the most highly selfing population, consistent with Gilbert *et al.* (2022). This mis-inference corresponded with large decreases in the mean values of population-scaled selection coefficients of the inferred gamma distributions (Supplementary Figure 1). The β parameter of the gamma distribution was also mis-inferred as the selfing rate increased, with the inferred β becoming 2-3× smaller than the simulated value at 99% selfing (Supplementary Table 2). While

the overall shape and skew of the DFE were largely recovered when there was some chance of

outcrossing (*e.g.*, with $P_{self} \sim 90\%$; Figure 1), the mean population-scaled selection coefficients

remained slightly biased even when selfing rates were as low as 80% (*e.g.* for DFE2;

Supplementary Figure 1, Supplementary Table 2).

Because BGS will lead to a decrease in the effective population size, we hypothesized

that the true population-scaled selection coefficients in our simulated populations are likely to be

smaller than those simulated (*i.e.*, $2Ns_d$). To account for the reduction in the efficacy of

purifying selection due to BGS, we corrected the simulated DFE with the relative rate of

coalescence (B) with and without BGS observed in the simulated population. Note that this

adjusted DFE accounts for the relative decrease in population size due to linked effects of

selection only, not due to selfing (see details in Methods). While the inferred DFE matches the

corrected expected DFE substantially well for moderate levels of selfing (Figure 1), the DFE

accounting for BGS is somewhat inaccurate at high selfing levels, especially for the DFEs with

more weakly and moderately deleterious mutations. That is, the inferred DFE tends to have both

more strongly deleterious mutations and nearly neutral mutations than the DFE accounting for

linked effects of selection and will thus result in the inference of a distribution of "*s*" that is more

platykurtic than the simulated distribution (Supplementary Figure 2). This suggests that a simple

rescaling of the effective population size can correct for the misinference when selfing rates are

low/moderate (when the population is likely experiencing classical BGS) but not when rates are

high (when the population is likely in the interference selection regime). As expected, this effect

is especially strong when the DFE consists of many weakly and strongly moderately mutations

(Supplementary Figure 2). We, therefore, conclude that Hill-Robertson interference (HRI)

among deleterious alleles is likely causing the mis-inference of the DFE of new mutations in

highly selfing populations.

As expected, the simulated DFEs were accurately estimated from single-locus simulations with

no linkage even in highly selfing populations (Supplementary Figure 3), despite a significant

reduction in the total number of segregating sites at high selfing (Supplementary Table 3).

Concordantly, the observed SFSs at selected and neutral sites were highly similar across different

selfing levels, confirming that selfing does not drastically influence the proportions of allele

frequencies in a population without linkage (Supplementary Figure 4), assuming that deleterious

mutations are additive. Importantly, as high rates of selfing increase the levels of homozygosity,

the site frequency spectrum (SFS) obtained from a random sample of diploid *individuals*

deviated starkly from one obtained by a random sampling of *genomes* (an example shown in

Supplementary Figure 5A). The inferred DFEs were very similar when we sampled individuals

vs. genomes (Supplementary Figure 5B) and thus all results that follow were obtained by

sampling genomes.

The effects of linkage on DFE inference in regions of low recombination

HRI is more likely in genomic regions of low recombination and may similarly bias DFE

inference. With decreasing rates of recombination (0.5, 0.1, 0.05, 0.01, 0.005, and 0.001×), the

extent of mis-inference of the DFE increased (Figure 2). As before, the estimated DFE was

strongly skewed towards mildly deleterious mutations compared to the simulated DFE,

consistent with a decrease in the efficacy of selection due to linked effects of selection.

The relative decrease in nucleotide diversity ($B = \frac{\pi}{\pi_0}$) due to background selection,

measured in simulations of regions with low recombination ($B \sim 0.05 - 0.92$) captures the range

of B values measured in the simulations with selfing and linkage ($B \sim 0.08 - 0.95$). To account

for the decrease in the effective population size due to BGS, the inferred DFE was compared to

the DFE given by the distribution of $2NBs_d$. Again, while the DFE accounting for BGS

explained the estimated DFEs quite well for higher recombination regions, at very low

recombination rates, it did not match the inferred DFEs. As with high selfing, the estimated

distribution of selection coefficients at low recombination was observed to be more platykurtic

than the DFE accounting for BGS, and mis-inference was much stronger at the lowest

recombination rates, even for the most strongly deleterious DFE (Supplementary Figure 6). This

reflects the fact that a simple rescaling effect of BGS (*i.e.*, the decrease in the rate of

coalescence) is unlikely to explain mis-inference observed in regions of very low recombination.

When recombination rates are extremely low, the probability of interference between selected

alleles at multiple loci increases, leading to a departure of the SFS from equilibrium. Overall,

increased HRI caused by severely reduced recombination rates, which are of particular

importance in selfing organisms, might lead to the observed DFE mis-inference.

The quantification of HRI using summary statistics

Much theoretical and computational work on HRI has suggested that the decrease in the efficacy of selection results in a decrease in the nucleotide diversity at selected and linked neutral sites as well as an increase in divergence at selected sites due to the increased fixation of deleterious mutations (Comeron *et al.* 1999; McVean and Charlesworth 2000). Moreover, the site frequency spectrum for selected and neutral sites is expected to be skewed towards low-frequency variants as the strength of BGS and HRI increases (mimicking the genealogy of an expanding population; Zeng and Charlesworth 2011; Walczak *et al.* 2012; Nicolaisen and Desai 2013; Good *et al.* 2014), and the nucleotide diversity and site frequency spectrum of selected mutations are expected to become more similar to those of neutral mutations (Comeron and Kreitman 2002; Kaiser and Charlesworth 2009; Seger *et al.* 2010; O'Fallon *et al.* 2010; Nicolaisen and Desai 2013). Because interference results in a gene genealogy with longer external branches than expected under a standard neutral coalescent (Walczak *et al.* 2012; Nicolaisen and Desai 2013), and consistent with a skew towards low-frequency alleles, new mutations are more likely to occur on separate branches of the genealogy, generating negative LD. Thus signed measures of linkage disequilibrium between selected mutations and nearby neutral sequences become more negative as HRI becomes stronger (Hill and Robertson 1966; McVean and Charlesworth 2000; Comeron and Kreitman 2002; Garcia and Lohmueller 2021). In addition, in populations with pervasive HRI, the SFS has been observed to show non-monotonic behavior at high frequencies, resulting in a U-shape, which deviates from the predictions of the classical BGS theory (Seger *et al.* 2010; Neher and Hallatschek 2013; Good *et al.* 2014; Zeng and Corcoran 2015; Cvijović *et al.* 2018).

In our simulations, the SFSs of neutral and selected sites shifted towards lower frequency mutations and became more similar at 90% selfing and above (Figure 3A, Supplementary Figures 7A & 8A), which is consistent with the extent of DFE mis-inference and with previously simulated results in highly selfing populations (Charlesworth *et al.* 1993). In addition, the SFS at high frequency alleles showed non-monotonic (U-shaped) behavior (Figure 3B, Supplementary Figures 7B & 8B), which has been previously observed in populations with pervasive HRI (Seger *et al.* 2010; Neher and Hallatschek 2013; Good *et al.* 2014) or very strong background selection (Cvijović *et al.* 2018). Note that the SFS of neutral sites at 0% selfing was very similar to the unlinked neutral SFS (Supplementary Figure 4), consistent with high B values. Trends in

$\Delta\theta_w$, a measure of the skewness of the SFS (with positive values indicating a relative increase in

rare alleles), were concordant with the patterns in the SFSs, with $\Delta\theta_w$ values for neutral and

selected sites sharply increasing and converging towards each other at 90% selfing and above

(Figure 4A). Values of $\Delta\theta_w$ at neutral sites nearly converge with values at selected sites at 99%

selfing, reflecting the reduced efficacy of selection and almost complete linkage.

For selfing rates between 80% and 95%, D' tended to become closer to 0 at both selected

and neutral sites (Figure 4B). This is similar to an effect observed in simulations by Kamran-

Disfani and Agarwal (2014), where partially selfing populations with intermediate selfing rates

showed increases in linkage disequilibria. This phenomenon is expected at higher selfing levels

if the efficacy of selection is not overwhelmed by HRI, because of correlations in homozygosity

at different loci across the genome (Roze and Lenormand 2005). At 99% selfing, D' values

became considerably more negative for selected and neutral sites, consistent with a population

experiencing pervasive HRI.

Simulation rescaling alters the effects of HRI

Due to computational time constraints, it was necessary to rescale the parameters $N\mu$, Nr, and

Ns for most simulations in this study by lowering N and increasing μ, r, and s by the rescaling

factor $Q = 100$. While equivalent equilibria are expected according to diffusion theory if the

population-scaled parameters are held constant (Hill and Robertson 1966; Crow and Kimura, M.

1970; Ewens 1979; Li 1987), it has previously been observed that high levels of rescaling can

increase the effects of HRI (Comeron and Kreitman 2002). To understand the effects of rescaling

at high selfing rates, we once again simulated *C. elegans*-like populations with 500 genes and

selfing rates of 95% and 99% and the three deleterious DFEs, with $Q = 50$, 20, and 10, though

at $Q = 10$ only a limited number of simulations were able to be completed within a reasonable

timeframe. We find that as Q decreases from the original value of 100, the effects of background

selection and HRI decrease, with the B values decreasing in a linear fashion with Q

(Supplementary Figure 9). While the extent of DFE mis-inference decreases as Q is lowered, it is

important to note that the DFEs estimated at low Q values and 99% selfing were still mis-

inferred, especially the DFEs with more weakly and moderately deleterious mutations. As

expected based on the inferred DFEs, the SFS at higher selfing rates and low Q was still skewed

towards lower frequency mutations and U-shaped, though less than at high Q (Supplementary

Figures 10-11). Thus, it is likely that fully unscaled simulations ($Q = 1$) would have even less

HRI effects, especially for the strongly deleterious DFE, though it is not unrealistic to expect

strong BGS and some interference between selected mutations based on the trends observed

here. For instance, we observed much stronger effects of BGS and substantially more mis-

inference of the DFE when larger genomic regions were simulated (Supplementary Figure 12),

suggesting full length chromosomes simulated at $Q = 1$ will likely have more pervasive linked

effects of selection. Indeed, Gilbert *et al.* (2022) conducted unscaled simulations of the entire

genome, including all full-length chromosomes and a reduced recombination rate in chromosome

centers, with *C. elegans*-like parameters and found a strong skew in their inferred DFEs relative

to their simulated DFEs at intermediate levels of selfing (see figure 4 of Gilbert *et al.*), so

exploration of our scaled simulations with pervasive HRI is still relevant to highly selfing

populations.

Extent of mis-inference does not depend on coding density

The effects of HRI are expected to increase when there is a higher density of selected sites, as

deleterious mutations arising in the population will be more closely linked on average (Comeron

and Kreitman 2002). Gene density varies along the *C. elegans* chromosomes, with higher gene

densities found in the chromosome centers than in the chromosome arms (*C. elegans* Sequencing

Consortium 1998). To understand if increases in coding density change the extent of DFE mis-

inference, we decreased the length of intergenic regions in our simulations, while keeping the

number and length of genes constant, producing simulations with coding densities 2× and 6×

higher than our original coding density. We observed no obvious changes in DFE inference in

these scenarios (Supplementary Figure 13), suggesting that DFE mis-inference will not increase

in more coding-dense regions of the genome, although local gene densities could increase more

than we simulated. In addition, the rate of recombination in *C. elegans* is higher in the gene-poor

chromosome arms than in the gene-rich cores (Rockman and Kruglyak 2009), which could

worsen DFE mis-inference in regions with high gene density due to low recombination rates.

The impact of dominance on DFE inference

DFE-alpha and GRAPES assume that all new mutations are semidominant, but it is known that

dominance coefficients of new deleterious mutations vary considerably and are partially

recessive on average, with strongly deleterious mutations tending to be highly recessive

(Simmons and Crow 1977; Agrawal and Whitlock 2011; Manna *et al.* 2011). The effect of

dominance on DFE inference is not fully understood. Most DFE inference methods assume that

new mutations are semidominant and thus provide estimates of the joint distribution of the

selection and dominance coefficients, "$s_d h$". A recent study by Wade *et al.* (2023) suggests that,

if there are recessive lethal mutations, current DFE inference approaches can lead to

underestimating the true proportion of strongly deleterious mutations, as more strongly

deleterious mutations are likely to persist in the population if they are recessive. Furthermore,

since selfing increases the levels of homozygosity in the genome, the efficacy of selection

against large effect and highly recessive deleterious mutations is expected to be greater

(Charlesworth and Charlesworth 1998), leading to more efficient purging of deleterious

mutations. Thus, with higher rates of selfing one might instead expect to observe a decrease in

the allele frequencies of mildly and moderately deleterious mutations, leading to the false

inference of the presence of more strongly deleterious mutations in the population.

To investigate the joint effects of dominance and selfing on DFE inference, we performed

simulations where all selected mutations had fixed dominance coefficients equal to 0.1, 0.25, or

0.75. Note that the outputs of DFE-alpha and GRAPES were adjusted to represent $2N_{self} s_d h_{self}$

rather than $2N s_d$, and the expected values from the simulations were adjusted to account for the

dominance and homozygosity of selfing genomes (Caballero and Hill 1992), as described in

Methods. At 0% selfing, as new deleterious mutations became more recessive (especially when

$h = 0.1$), the distribution of the product of the selection and dominance coefficients was

somewhat mis-inferred. When mutations were predominantly moderately and strongly

deleterious, there was an underestimation of the proportion of strongly deleterious mutations

(Figure 5, Supplementary Table 5), consistent with Wade *et al.* (2023). Additionally, the

frequency of the nearly neutral class of mutations was underestimated in all three DFEs, an effect

that can also be observed to a smaller degree in Wade *et al.* (2023). However, when new

mutations were comprised largely of mildly deleterious mutations (*i.e.*, DFE1), there was an

overestimation of the fraction of strongly deleterious mutations. In contrast, when more

dominant mutations were simulated, DFE inference resulted in mis-inference towards both

weakly and strongly deleterious mutations, though this effect was fairly small. As expected, mis-

inference was stronger when the true DFE was skewed towards mildly deleterious mutations, as

these mutations are more likely to be present as homozygotes in the population.

As selfing increases homozygosity, the inferred DFEs in partially selfing populations at

different dominance levels are expected to become more similar to each other, making the

dominance of mutations a less important factor for the efficacy of selection (Caballero and Hill

1992; Charlesworth 1992). As expected, at 50% selfing, the DFE was inferred more accurately

(Supplementary Figure 14, Supplementary Table 5). While the dominance coefficient played a

negligible role in DFE inference when the selfing rate was 99%, the estimated DFEs were

strongly skewed towards mildly deleterious mutations (Figure 5). Thus, just as expected, at high

rates of selfing, dominance coefficients play a relatively minor role, while HRI and BGS are

primarily responsible for DFE mis-inference.

Increased purging of recessive deleterious mutations has been hypothesized to reduce

their average allele frequencies, leading to DFE mis-inference (Arunkumar *et al.* 2015; Gilbert *et*

al. 2022). However, observable effects of purging are likely to occur only when mutations are

highly recessive (Nei 1968) and strongly deleterious (Glémin 2003). Concordantly, we observed

that the distribution of allele frequencies of partially recessive ($h = 0.25$) mutations at

intermediate selfing levels was the same as those of semidominant mutations, suggesting that

increased purging of recessive deleterious alleles did not have a strong effect on the average

allele frequencies in a population sample at equilibrium (see the SFS in Supplementary Figure

15).

Inference of the full DFE of new deleterious and beneficial mutations

Both BGS and HRI lead to a decrease in the efficacy of selection at linked selected sites, thus

reducing the probability of fixation of beneficial mutations while increasing the probability of

fixation of deleterious mutations (Charlesworth 1992; Kamran-Disfani and Agrawal 2014). It is

therefore likely that the proportion of fixed mutations at selected sites that are beneficial (α) is

mis-estimated when effective recombination rates are low. We conducted simulations with 0%,

50%, 80%, 90%, 95%, and 99% selfing, where 0.1% and 1% of new mutations in exons were

beneficial, following an exponential distribution with $2N\bar{s}_a = 200$. When 0.1% of new exonic

mutations were beneficial, the inference of the deleterious DFE by GRAPES was fairly accurate

at 0% and 50% selfing (Figure 6A). However, the deleterious DFE was mis-inferred at a much

lower selfing rate than in scenarios without beneficial mutations and mis-inference was

particularly strong when the deleterious DFE was highly skewed towards weakly deleterious

mutations. When 1% of new mutations were beneficial, the deleterious DFE estimated by

GRAPES was strongly skewed towards nearly neutral mutations even at 0% selfing for the

weakly deleterious and moderately deleterious DFEs (Supplementary Figure 16). Due to

recurrent selective sweeps in this scenario as well as BGS, there is likely to be a substantial

decrease in effective population size experienced by selected alleles (and thus a reduced $2N_e s_d$).

We thus accounted for this decrease again by comparing the inferred DFE to the distribution of

$2N(\pi/\pi_0)s_d$, where π/π_0 represents the neutral nucleotide site diversity with and without

selection. Again, the DFE accounting for the decrease in the effective size did not explain the

inferred DFE at high selfing rates, including 95% selfing, suggesting that HRI effects might be

responsible for the mis-inference, even at moderate levels of selfing (Figure 6A). The selected

SFSs resembled the neutral SFSs in simulations with DFE mis-inference, reflecting the observed

results of DFE inference (Supplementary Figure 17). With a high rate of beneficial mutations

entering the population, hitchhiking can reduce the efficacy of selection on weakly deleterious

alleles (Campos and Charlesworth 2019), leading to the mis-inference of the DFE for the more

weakly deleterious DFEs. In addition, the failure of DFE inference programs to fully correct for

the presence of mildly beneficial alleles in the SFS could also have contributed to the observed

mis-inference (Tataru *et al.* 2017).

As the folded SFS might provide better estimates of the deleterious DFE when beneficial

mutations are segregating at high frequencies (Keightley and Eyre-Walker 2007; Eyre-Walker

and Keightley 2009), we also tested the inference using the folded SFS. Overall, the deleterious

DFE inferred by GRAPES was comparable to the unfolded results when the folded SFS was

used (Supplementary Figures 18A & 19A). However, the DFEs estimated by DFE-alpha were

biased towards weakly deleterious mutations even with no selfing, with much larger uncertainty

in the estimated DFE at high selfing levels, especially when 1% of all new selected mutations

were beneficial. This indicates that folding the SFS is inadequate to correct for the mis-inference

of the deleterious DFE when the rate of new beneficial mutations is extremely high.

As expected, the true fraction of beneficial substitutions (α) decreased as the selfing rate

increased (Figure 6B, Supplementary Figure 16B). Changes in this parameter were driven by

both increases in the number of deleterious substitutions ($\sim 2 - 4 \times$ higher at 99% selfing

relative to 0% selfing) as well as the decreases in the number of beneficial substitutions ($\sim 3 \times$

lower at 99% selfing relative to 0% selfing; Supplementary Table 6), consistent with previous

observations in Kamran-Disfani and Agrawal (2014). In simulations with both 0.1% beneficial

mutations and 1% beneficial mutations, α tended to be overestimated for the weakly deleterious

DFE at all selfing levels, with mis-inference increasing steadily as the selfing rate increased. For

the moderately and strongly deleterious DFEs, α was only overestimated at high selfing levels,

with the strongly deleterious DFE corresponding with the most accurate inference of α. This is

consistent with the expectation that a decrease in the efficacy of selection will increase the

fixation of weakly deleterious alleles, which will be misinterpreted as beneficial fixations by

GRAPES, leading to an overestimation of α at high selfing levels. Very similar trends were

observed when the folded SFS was used to estimate α (Supplementary Figures 18B & 19B).

Overall, at high selfing levels, α values were substantially overestimated by GRAPES.

Effects of population structure and sampling on the accuracy of inference

A final factor that could bias DFE inference in partially selfing populations is population

structure because the inference procedure of DFE-alpha and GRAPES is not designed to account

for the effects of population structure on the SFS. Moreover, selfing populations tend to have

highly structured populations, and DFE inference in the context of population structure has been

largely unexplored, though the effects of cryptic structure (without migration) were tested in Kim

et al. (2017). Here, we simulated an island model, with five demes that exchanged migrants at a

total rate m each generation. Multiple values of m were used to vary the degree of isolation

between demes in this model, and the number of individuals in each deme (N_{deme}) was varied so

that the effective size of the metapopulation (N_{meta}) was equivalent to 500,000 individuals

(Supplementary Table 7). Under strict neutrality and while selfing was taken into account (see

Methods), the neutral nucleotide diversity of the entire metapopulation ($2T_T\mu$), of individual

subpopulations ($2T_S\mu$), and F_{ST} was equal to theoretical expectations (equations 2-4,

Supplementary Table 8). We performed simulations at $N_{deme}m = 0.1, 0.5, 1,$ and 2, which

corresponded to $F_{ST} = 0.76, 0.38, 0.24,$ and 0.14 respectively at 99% selfing. These values

were chosen to mimic F_{ST} values observed empirically amongst *C. elegans* populations, which

range from 0.44 to 0.83 (Barrière and Félix 2005; Richaud *et al.* 2018).

Population structure by itself did not result in mis-inference of the DFE when the rate of
migration was sufficiently high (*i.e.*, $N_{deme}m = 2, 1$, and 0.5). High selfing rates with
population structure again led to inferring a DFE skewed almost entirely towards mildly
deleterious mutations, thus producing a very similar extent of mis-inference as that observed in a
panmictic population (Supplementary Figure 20 A, B, and C, respectively). However, at low
rates of migration between demes ($N_{deme}m$ of 0.1), the DFE was mis-inferred even at 0%
selfing, with a bias towards inferring a lower efficacy of selection than the simulated DFE
(Figure 7; Supplementary Figure 20). This was largely because the relative increase in
intermediate frequency alleles due to population structure as opposed to panmixia
(Supplementary Figures 21-22) was falsely inferred as a bottleneck (mean fold change = 0.45-
0.50; similar to Chikhi *et al.* 2010) by DFE-alpha, leading to incorrect estimates of the DFE. As
expected, these distortions of intermediate frequency alleles in the SFS caused by population
structure were observed to increase at higher selfing levels and at lower migration rates, with
particularly strong deviations from panmixia observed at $N_{deme}m = 0.1$.
Empirical studies that draw from worldwide population samples may knowingly or
unknowingly sample unevenly from structured populations when performing DFE inference. To
investigate if this has unintended consequences on DFE inference, we repeated our experiments
with population structure, altering our sampling strategy to primarily draw genomes from one
deme (60%) or two demes (35% from each), with the remaining demes contributing only 10% of
the sampled genomes each. As expected, the site frequency spectra from both unevenly sampled
populations were observed to be even more distorted at intermediate frequencies than the evenly
sampled populations with low migration rates (Supplementary Figures 23-26). For the sampling
strategy primarily drawing from one deme, some DFE mis-inference towards nearly neutral and
strongly deleterious mutations was observed at 0% and 50% selfing even for $N_{deme}m = 1$ and
more strongly at $N_{deme}m = 0.5$, and this effect was stronger at the DFE with predominantly
weakly deleterious mutations (Supplementary Figure 27). This effect was also observed for the
sampling strategy primarily drawing from two demes, but to a lesser extent (Supplementary
Figure 28). At the lowest migration rate, the inferred DFEs for both uneven sampling cases were
substantially less deleterious at 0% and 50% selfing than for even sampling, particularly for the
DFE with predominantly weakly deleterious mutations (Figure 7). At higher selfing levels, the
inferred DFEs for all sampling schemes falsely inferred a larger proportion of weakly deleterious

mutations than predicted by the DFE corrected for BGS (Figure 7), likely due to a combination

of HRI and uneven sampling. The BGS-corrected DFE was unable to correct for the mis-

inference of the DFE for all selfing levels at the lowest migration rate, reflecting the fact that

DFE mis-inference is likely driven by changes in the SFS caused by population structure rather

than changes in the level of BGS.

778   Overall, these results indicate both DFE inference approaches are fairly robust to

population structure in an island model, even with uneven sampling, until the populations

become extremely isolated from each other, or individuals are mostly drawn from one deme,

especially when the true DFE has a significant proportion of mildly deleterious mutations.

Additionally, both low rates of migration and highly uneven sampling result in the inference of

an even more weakly deleterious DFE than simulated. Testing DFE inference with more models

of population structure and additional sampling strategies might be necessary to interpret

empirically estimated DFEs from natural populations.

DISCUSSION

Inference of the distribution of fitness effects (DFE) of new mutations is a critical step in

understanding the forces that shape genetic variation in a population. Population genetic methods

to infer the DFE that account for nonequilibrium demography usually assume independent

segregation of putatively neutral and selected sites (Eyre-Walker and Keightley 2007; Keightley

and Eyre-Walker 2007; Galtier 2016; Tataru *et al.* 2017). However, organisms that have little/ no

recombination, for instance, those that undergo high rates of selfing or asexual reproduction, are

likely to violate this assumption. A major consequence of strong linkage is the increase in BGS,

selective sweeps, and Hill-Robertson effects that lower the efficacy of selection. Particularly in

the regime where HRI is pervasive, many linked selected polymorphisms segregate in a

population, reducing the population's variance in fitness and thus the efficacy of selection

(Comeron and Kreitman 2002; Kaiser and Charlesworth 2009; Seger *et al.* 2010; O'Fallon *et al.*

2010; Nicolaisen and Desai 2013). In this study, we demonstrate that when HRI is pervasive, the

DFE of new deleterious mutations is mis-inferred. When the regime of classical BGS and sweeps

is predominant, population-scaled selection coefficients remain biased (as expected), but

accounting for the relative decrease in the rate of coalescence can provide reasonably accurate

estimates of the selection coefficients. The latter is applicable to moderately recombining

populations/regions. We also find that while purging of partially recessive deleterious mutations

does not lead to mis-inference, the presence of cryptic population structure where there is a low

rate of migration between subpopulations and/or sampling from predominantly one

subpopulation leads to inferring a DFE skewed towards effectively neutral/mildly deleterious

class of mutations. Moreover, the proportion of beneficial substitutions (α) is substantially

overestimated in highly selfing species, suggesting caution when performing inference in species

or genomic regions with little/no recombination.

Why is the DFE estimated from *C. elegans* populations not skewed towards mildly

deleterious mutations?

While our study suggests that DFE inference in highly selfing species should yield DFEs skewed

towards weakly/moderately deleterious mutations, surprisingly, the previous empirical estimate

of the *C. elegans* DFE using DFE-alpha is heavily skewed towards the class of strongly

deleterious mutations ($2N_e\overline{s_d} > 10,000$, Gilbert *et al.* 2022). One explanation of this

discrepancy is that the mean $2N_e\overline{s_d}$ of the population is even greater than 10,000. Interestingly,

direct measurements of fitness-related traits in mutation accumulation lines suggest that *C.*

elegans has a highly deleterious DFE, with estimates of the mean deleterious selection

coefficient ranging from -0.09 to -0.46 (Vassilieva *et al.* 2000; Baer *et al.* 2005; Katju *et al.*

2015). Thus, using previously calculated *C. elegans* N_e ranging from 5,000 to 80,000 (Table 4 in

Teterina *et al.* 2023), the expected $2NB\overline{s_d}$ would range from 900 to 72,000, consistent with this

explanation. However, there are a few major caveats to these observations. (a) The experimental

estimates of selection coefficients are likely biased towards more deleterious selection

coefficients due to the difficulty of measuring small fitness changes in experimental lines

(Davies *et al.* 1999; Eyre-Walker and Keightley 2007), and might not reflect fitness effects in the

wild. (b) It is unclear whether previous estimates of the population sizes of *C. elegans*

populations using neutral nucleotide diversity (Barrière and Félix 2005; Cutter 2006; Rockman

and Kruglyak 2009; Teterina *et al.* 2023) are accurate due to the joint effects of selfing, HRI, and

demography (c) Empirical estimates of the deleterious DFE in *C. elegans* (Gilbert *et al.* 2022)

were obtained by sampling worldwide rather than from local populations, and thus other factors

such as complex demographic history and population structure could play a role as well. For

instance, selfing populations are likely to follow extinction-recolonization dynamics

(Charlesworth and Wright 2001; Ingvarsson 2002; Wright *et al.* 2013) and such models remain

understudied.

A second explanation for the discrepancy between the expected and observed DFE in *C.*

elegans is that the true selfing rate in wild *C. elegans* might be lower than 99%, thus reducing

linked effects of selection (Figure 1). Though selfing rate estimates from unique populations

range from 80% to ~100% using various methods (Barrière and Félix 2005; Sivasundar and Hey

2005; Teterina *et al.* 2023), in general, the strong LD in *C. elegans* populations (haplotype

blocks of 2.5 Mb on average; Cutter 2006) along with low frequency of males in wild specimens

(Barrière and Félix 2005; Richaud *et al.* 2018) suggest the outcrossing rate is very low in most

populations. A recent study using whole-genome sequencing data focusing on Hawaiian *C.*

elegans estimated a relatively lower selfing rate of 93%, suggesting that selfing rate can vary

across populations (Teterina *et al.* 2023). Importantly, as the samples used in Gilbert *et al.*

(2022) were unevenly sampled worldwide (with many samples from Hawaiian populations),

variation in selfing rate across populations or through time could additionally influence DFE

inference results.

A third possibility relates to the difficulty in obtaining the SFS using highly inbred

strains. The SNP-calling pipeline used to obtain *C. elegans* population-genetic data converted

heterozygous sites into homozygous reference or alternate alleles depending on if the mutant was

above or below 50% (Cook *et al.* 2017). This method of SNP processing could lead to the

inference of a more deleterious DFE if the heterozygote frequency is higher at neutral sites than

selected sites by skewing the neutral SFS towards higher frequencies and the selected SFS

towards lower frequencies, though the low frequency of heterozygotes in highly selfing

populations makes this unlikely to be a large issue. Future studies working with inbred strains

should evaluate the effect of employing such SNP processing approaches on population genomic

inference.

Finally, we have only examined a population under equilibrium in the current study,

whereas a recent population expansion/ contraction could decrease the observed effects of

interference between selected alleles at multiple loci, allowing for a correct inference of selection

coefficients. HRI effects in non-equilibrium populations remain to be studied more thoroughly.

Unfortunately, as linked effects of selection will also lead to falsely inferring recent population

growth (Supplementary Table 1; Ewing and Jensen 2016; Schrider *et al.* 2016; Johri *et al.* 2021),

there is no straightforward solution to accounting for the effects of the true demography (Johri *et*

al. 2022b). Thus performing a joint inference of demography and the DFE as employed by Johri

et al. (2020) is likely to be the most promising approach to circumvent this challenge.

How broadly do our results apply?

Previous studies (*e.g.*, Messer and Petrov 2013; Gilbert *et al.* 2022) that have observed similar

biases in inference using DFE-alpha due to linked effects of selection suggested that this might

be because DFE-alpha cannot account for the false signal of extreme recent growth generated by

hitchhiking effects. Our study shows that DFE mis-inference was similarly observed when using

the method GRAPES, which employs a reweighting scheme of the selected relative to the neutral

SFS and does not explicitly model a specific demographic history. We, therefore, conclude that

the observed mis-inference is a general phenomenon (likely due to a combination of BGS,

sweeps, and HRI effects) and not due to errors unique to a particular method like DFE-alpha.

Our results have implications for the population genetics of other highly selfing

organisms, many of which could fall into a parameter space where linked effects of selection are

pervasive. For instance, highly selfing species with large N_e such as *Arabidopsis thaliana*,

Capsella rubella, *Capsella bursa-pastoris*, and *Capsella orientalis* have all been shown to have

more weakly deleterious DFEs than comparable outcrossing species (*A. lyrata* and *C.*

grandiflora; Douglas *et al.* 2015; Chen *et al.* 2020)*,* consistent with the effects of selection at

linked sites potentially playing an important role in determining their genomic diversity. In

addition, classical BGS has been predicted to play a more important role in shaping diversity in

C. rubella, A. thaliana, and *M. truncatula*, than in *C. elegans* populations (Buffalo 2021).

A potential limitation of the applicability of our study is that it is not entirely clear what

the extent of HRI in natural populations is expected to be. A key factor that determines the extent

of HRI is the number (U_d) of new deleterious mutations per genome per generation, with higher

values of NU_d relative to NR causing more interference. Thus, the extent of HRI in across

species would likely differ due to differences in outcrossing rates, population sizes, mutation and

recombination rates, and genome architecture (McVean and Charlesworth 2000; Comeron and

Kreitman 2002; Kaiser and Charlesworth 2009; Good *et al.* 2014). For instance, the rate of LD

decay is significantly smaller in *C. elegans* where a complete decay of LD occurs over several

Mb (Cutter 2006; Rockman and Kruglyak 2009) compared to *A. thaliana* (10-50 kb; Ross-Ibarra

et al. 2008; Shimizu and Tsuchimatsu 2015), suggesting a higher possibility of HRI affecting

DFE inference in *C. elegans*. Interestingly, *C. elegans* has been observed to have a U-shaped

SFS in many samples (Freund *et al.* 2023), consistent with HRI, though in empirical data a

variety of explanations such as ancestral allele misorientation, positive selection, and various

demographic effects could explain a U-shaped SFS (Freund *et al.* 2023). However, some highly

selfing species have more coding-dense genomes and are thus predicted to have higher numbers

of deleterious point mutations per genome per generation (U_d) relative to *C. elegans* ($U_d =$

0.04), which could increase the chances of interference between selected mutations; these

include *Oryza sativa* ((99.3% selfing; $U_d = 0.72$; Lynch *et al.* 2016; Bah *et al.* 2017) and *A.*

thaliana (97% selfing; $U_d = 0.29$; Lynch *et al.* 2016). In addition, pathogenic species like

Plasmodium ($U_d = 0.25$; Lynch *et al.* 2016) that have variable rates of sexual reproduction

between genetically identical clones (and thus experience reduced rates of recombination), seem

to have nearly identical neutral and selected site frequency spectra (Figure 3 in Parobek *et al.*

2016), suggesting that HRI effects might play an important role in shaping variation across their

genomes. We therefore also expect DFE inference to be biased due to HRI in asexual organisms,

mitochondrial genomes, and the Y chromosome, all of which have low effective recombination

rates (Barton and Charlesworth 1998; Weinreich and Rand 2000; Charlesworth and

Charlesworth 2000). Importantly, as structural variants are a significant source of deleterious

mutations (Adrion *et al.* 2017; Abel *et al.* 2020) accounting for the input of other mutation types,

including small insertion-deletions is likely to increase the estimated value of U_d, making HRI

more likely in populations with little/no recombination.

An interesting exception is the previous study performed in *Eichhornia paniculata*

populations (Arunkumar *et al.* 2015), where effects of selection at linked sites did not confound

inference of population-scaled selection coefficients. Their simulated populations were

comparable to ours – a high selfing rate (98%), a mutation to recombination rate ratio of 0.14 (*vs.*

0.11 in this study), a deleterious DFE (with mean $N\overline{s_d} \sim 50 - 100$), and 15,000,000 selected sites

(*vs.* 562,500 in this study). However, the expected values of NU_d in the full haploid genomes of

E. paniculata vs. C. elegans are ~ 25 and ~ 37,125 respectively, assuming 20% and 30% coding

density respectively. Higher NU_d will lead to stronger effects of BGS and has been shown to

cause populations with moderate selection coefficients that would otherwise be in the classical

BGS regime to have pervasive HRI (Good *et al.* 2014), so it is plausible that differences in NU_d

explain the differences between our studies.

Considerations for inferring the DFE in highly selfing organisms

Given our observations, we here provide a few points to consider when inferring the DFE of new

mutations in organisms with little/no recombination, where effects of BGS, selective sweeps, or

HRI might be pervasive:

(1) The DFE of new deleterious mutations estimated using the two-step population-

genetic methods is most closely explained by the distribution of $2NBs_d$, which is equivalent to

$2N_{emp}(1 + F)s_d$ in partially selfing populations. Here N_{emp} represents the effective population

size estimated using neutral nucleotide diversity (not accounting for selfing), which includes the

effects of BGS and is what is usually estimated using demographic inference approaches that do

not account for linked effects of selection. Thus, if there exist accurate experimentally validated

estimates of selfing rates, it is possible to accurately infer the distribution of selection

coefficients in populations with moderate levels of selfing. However, in many organisms selfing

rates are inferred using allele frequencies and those estimates are likely to be biased by

population history and effects of selection at linked sites. Moreover, this raises the question of

how one can accurately simulate an estimated DFE (in terms of population-scaled selection

coefficients) when multiple linked selected sites are to be simulated, which will be especially

important to determine the extent of HRI effects in populations with little/no recombination.

(2) Different models of population structure and the effects of demographic changes need

to be considered to fully understand the robustness of DFE inference. For instance, it is known

that local populations of selfing organisms are prone to recurrent extinction and recolonization

events (Pannell and Charlesworth 1999; Ingvarsson 2002) which may bias DFE inference

further. Indeed, recent field studies of *C. elegans* populations suggest that 3-10 individuals

colonize a food source and quickly reproduce to census sizes of up to 10,000 individuals before

dispersing in search of new food sources (Barrière and Félix 2005; Richaud *et al.* 2018). When

sampling across many demes, we suggest also inferring the DFE in isolated populations in order

to minimize biases due to cryptic population structure or uneven sampling and to downsample

the number of genomes such that sampling is as even as possible across demes. Sampling

strategies such as a scattered sampling scheme may be useful in removing such bias (Wakeley

1999; Städler *et al.* 2009), though the reduction in sample size may reduce the ability to

accurately infer the moderate and deleterious portions of the DFE (Kim *et al.* 2017).

(3) If possible, it could be helpful to infer the DFE in closely related non-selfing species

or in different populations of the same species with known differences in selfing rates. For

instance, *C. elegans* and *A. thaliana* populations vary in the estimated rates of selfing by ~10%,

and the mostly outcrossing *A. lyrata* has several populations that have evolved partial selfing

(Mable *et al.* 2005; Bomblies *et al.* 2010; Teterina *et al.* 2023). Though differences such as

population structure and demography need to be carefully accounted for, such comparisons

across populations/species could also be used to approximate the extent of HRI in a selfing

species.

(4) Finally, in the cases where HRI is determined to be extensive, joint inference of

demography and the DFE as employed by Johri *et al.* in *D. melanogaster* (2020) and human

(2022) populations could be especially useful for DFE inference. They employed an approximate

Bayesian computation (Beaumont *et al.* 2002) framework combined with forward simulations to

jointly infer parameters of the DFE of new deleterious mutations and an arbitrary size change,

while accounting for the effects of selection at linked sites. While highly successful, such an

approach has been limited to a relatively simple demographic model of a single size change in

one population due to the large number of parameters required to be jointly estimated. In

addition, our work here indicates that simulation rescaling may have to be limited to accurately

capture the dynamics of Hill-Robertson interference, which will further increase the

computational burden of simulations. Thus, if computationally feasible, such an approach could

yield the most likely distribution of $2Ns_d$. More importantly, the distribution of $2Ns_d$ describing

the fitness effects of new mutations is likely not the distribution of $2Ns_d$ describing the fixation

probabilities of new mutations, which will be relatively smaller. Furthermore, the distortions of

the SFS from a neutral model caused by interference cannot be explained by any constant N_e

(see discussion in Neher 2013; Good *et al.* 2014; Melissa *et al.* 2022). Thus, until theoretical

models can accurately estimate the effects of HRI in organisms with little/no recombination with

complex DFEs, simulation-based approaches will be the most helpful in inferring the DFE of

new mutations in such populations, as well as obtaining the fixation probability of new

mutations, which will be altered by HRI.

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DATA AVAILABILITY

The following data are publicly available on https://github.com/JohriLab/DFESelfing: (1) Data

and scripts to reproduce figures; (2) Config files used for DFE inference; (3) Simulation outputs

in SLiM format; (4) templates and example scripts for launching SLiM simulations.

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Figure 1: Effects of selfing on the inference of the DFE of new deleterious mutations using DFE-alpha (red bars) and GRAPES (purple bars). Results are shown in terms of the proportion of mutations in effectively neutral (f_0), weakly deleterious (f_1), moderately deleterious (f_2) and strongly deleterious (f_3) classes. Selfing rates were varied between 0 and 99 %. The nucleotide site diversity with background selection (B) relative to its expectation under strict neutrality is shown in each panel. The simulated (black bars) and adjusted (grey bars) DFE represents the distribution of $2Ns_d$ and $2NBs_d$ respectively. The error bars denote the standard deviation of proportions estimated from 5 independent replicates.

Figure 2: Effects of low recombination rate on the inference of the DFE of new deleterious mutations using DFE-alpha (red bars) and GRAPES (purple bars). Results are shown in terms of the proportion of mutations in effectively neutral (f_0), weakly deleterious (f_1), moderately deleterious (f_2) and strongly deleterious (f_3) classes. Recombination rates were varied between $0.5 \times$ mean and $0.001 \times$ mean, where the mean recombination rate was 3.12×10^{-8} per site/generations. The nucleotide site diversity with background selection (B) relative to its expectation under strict neutrality is shown in each panel. The simulated DFE (shown in black) and adjusted DFE (shown in grey) represents the distribution of $2Ns_d$ and $2NBs_d$ respectively. The error bars denote the standard deviation of proportions estimated from 5 independent replicates.

Figure 3: Site frequency spectra of neutral and selected alleles from 100 genomes sampled from simulations where all selected mutations were deleterious, for the moderately deleterious DFE (DFE2). For **(A)** the y-axis represents the proportion of segregating polymorphisms that fall into the given derived allele count. The last class (11+) refers to the derived allele counts 11-99. The error bars denote the standard deviation of proportions estimated from 5 independent replicates. For **(B)** the y-axis is plotted on a log scale, with all derived allele counts displayed. Lines were smoothed with LOESS.

Figure 4: Effects of selfing on the SFS and linkage disequilibrium (LD) at neutral (red lines) and selected (blue lines) sites. The skew in the SFS was quantified using $\Delta\theta$ for DFE1 (**A**), DFE2 (**C**), and DFE3 (**E**), calculated using alleles at all frequencies, while LD was summarized by the statistic D' for DFE1 (**B**), DFE2 (**D**), and DFE3 (**F**), calculated for alleles at frequencies of 1%-5%. The error bars denote the standard deviation of the genome-wide average of each summary statistic estimated from 5 independent replicates.

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Figure 5: Effects of selfing on DFE inference of new deleterious mutations with varying
dominance coefficients using DFE-alpha (red bars) and GRAPES (purple bars). The dominance
of selected mutations is noted at the top of each column. Results are shown for populations with
(**A**) 0% selfing and (**B**) 99% selfing. Here the effectively neutral (f_0), weakly deleterious (f_1),
moderately deleterious (f_2) and strongly deleterious (f_3) classes represent $0 < 2N_{self}s_dh_{self} <$
1, $1 < 2N_{self}s_dh_{self} < 10$, $10 < 2N_{self}s_dh_{self} < 100$, and $100 < 2N_{self}s_dh_{self} < 2N_{self}$,
respectively. The error bars denote the standard deviation of proportions estimated from 5
independent replicates.

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Figure 6. Effects of selfing on the inference of (A) the DFE of new deleterious mutations and
(B) the proportion of beneficial fixations (α) using GRAPES (shown in purple bars). (A) Results
are shown in terms of the proportion of mutations in effectively neutral (f_0), weakly deleterious
(f_1), moderately deleterious (f_2) and strongly deleterious (f_3) classes. The adjusted DFE
represents the distribution of $2N(\pi_{obs}/\pi_{exp})s_d$, where π_{obs} is the neutral nucleotide site
diversity observed in each scenario and π_{exp} is the neutral nucleotide site diversity expected in a
population of size N at strict neutrality. For both (A) and (B), 0.1% of new exonic mutations
were beneficial and exponentially distributed with a mean $2Ns_a = 200$. The error bars denote
the standard deviation of proportions estimated from 5 independent replicates.

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Figure 7 Effects of uneven sampling, population structure, and selfing on the inference of the DFE of new deleterious mutations using DFE-alpha (red bars) and GRAPES (purple bars). Results are shown in terms of the proportion of mutations in effectively neutral (f_0), weakly deleterious (f_1), moderately deleterious (f_2) and strongly deleterious (f_3) classes. Populations were simulated under an island model with five demes such that the metapopulation effective size was equal to 5000 at 0% selfing and $N_{deme}m$ was 0.1. Results are shown for the most weakly deleterious DFE (*i.e.*, DFE1). The error bars denote the standard deviation of proportions estimated from 5 independent replicates. Rows represent the sampling scheme for genomes, where sampling was even, sampling was primarily from one deme, or sampling was primarily from two demes.

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1374 **Table 1**: Parameters of the simulated DFEs of deleterious mutations in terms of the proportion of
1375 mutations in effectively neutral (f_0), weakly deleterious (f_1), moderately deleterious (f_2) and
1376 strongly deleterious (f_3) classes. DFEs consisted of predominantly mildly deleterious (DFE1),
1377 moderately deleterious (DFE2), or strongly deleterious (DFE3) mutations.

	β	$2Ns_d$	f_0	f_1	f_2	f_3
DFE1	0.9	5	0.204	0.655	0.140	0.000
DFE2	0.5	50	0.113	0.233	0.497	0.157
DFE3	0.3	1000	0.098	0.097	0.192	0.614

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