1 **<u>Eyespots originated multiple times independently across the Lepidoptera</u>**

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Abstract

Eyespot colour patterns function as an effective defence against predators and have evolved

numerous times independently across animals. In Nymphalid butterflies, eyespots have a single

evolutionary origin close to the base of this clade, but eyespots are also present in many other

lepidopteran lineages and may have multiple independent origins. Here we use phylogenetic

comparative methods to investigate the evolution of eyespots across a multi-superfamily phylogeny

of Lepidoptera, and to pinpoint lineages in which eyespots likely originated independently. We find a

total of 30 separate origins of Discal eyespots (in the discal wing region) and 21 separate origins of

Marginal eyespots (in the marginal wing region), a type of serial homologue. In four instances

eyespots were preserved in most extant representatives of a subsequent species radiation. In one

such instance, in the Nymphalidae, we found a Marginal eyespot common ancestor predating the

origin of a Discal eyespot baring ancestor, while in the Saturniidae we observed the opposite. We

conclude that eyespots do not appear to be homologous across the Lepidoptera. However, our

phylogenetic inference provides a roadmap for future developmental and functional studies that can

address whether discal and marginal eyespots share a homologous gene-regulatory network. This

study, therefore, has implications for our understanding of the evolution of serial homologues and of

convergent evolution of visual signals in insects.

25
26 **Introduction**

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Adaptive traits in animals such as protective colour patterns have long been a source of interest and

research (Tylor, 1886; Poulton, 1890; Beddard 1895). Stripes, spots and blotches can all function as

protective colour patterns (Stevens, 2007; Caro et al., 2016; Mizuno et al., 2024), but the ocellus, or

eyespot, which is characterized by the presence of one or more concentric rings of contrasting

colours, is among one of the most researched protective patterns. Eyespots are found across a wide

variety of vertebrate and invertebrate species (Edmunds, 1976; Meadows, 1993; Loyau et al., 2007;

He et al., 2010; Wong & Marek, 2019; Hernández-Palma et al., 2023) and represent a textbook

example of evolutionary convergence in response to selection by visual predators (Stevens et al.,

2008; Oliver et al., 2012; Cunha et al., 2023).

In Lepidoptera (butterflies and moths), eyespots serve a diversity of roles and develop in a diversity of locations in the body of larvae and/or wings of adults. In both larvae and adults, eyespots are used in predator deterrence and in attack deflection, but in adults they also function in courtship and mate selection (Breuker & Brakefield 2002; Robertson and Monteiro 2005; Vallin et al. 2007; Kodandaramaiah, 2011; Merilaita et al., 2011; Prudic et al. 2012; Kodandaramaiah et al., 2013; Hossie et al., 2014; Prudic et al., 2015; Mukherjee & Kodandaramaiah, 2015; Huq et al., 2019; Halali et al., 2019; Crees et al., 2021).

In adult wings, eyespots can be found on different regions and will be named, hereafter, as Marginal and Discal eyespots. Marginal eyespots develop close to the wing margin in between veins (Fig. 1A-B), whereas Discal eyespots are found more centrally located on the wing, either straddling the cross vein in the discal cell region or within the discal cell (Fig. 1 C-D; Otaki et al., 2020). Lepidopterans can have one or both types of eyespots on their wings. Marginal eyespots are often found in more than one unit on the wing margins and are considered serial homologs, whereas Discal eyespots are often found as single units. It is unclear whether Discal eyespots are serial homologs to Marginal eyespots, i.e., it is unclear whether these two eyespot types are using the same gene-regulatory network (Monteiro et al. 2006).

The evolutionary history of Discal and Marginal eyespots across the Lepidoptera is also still largely unknown, as most research on eyespot evolution has focused on Marginal eyespots of brush-footed butterflies. This clade of butterflies (Nymphalidae), which contains ~6,400 described species, accounts for only ~3.3% of lepidopteran diversity (~180,000 total described Lepidoptera; Brakefield, 2003; Heikkilä et al., 2015; Monteiro, 2015; Bhardwaj et al., 2020; Chazot et al., 2021). Phylogenetic comparative methods estimated that Marginal eyespots evolved once in the Nymphalidae, shortly after the origin of the clade (Oliver et al., 2012; Oliver et al., 2014). Thus, all Marginal eyespots in extant nymphalids are thought to be homologous. Nonetheless, Marginal eyespots are found in taxa across the entire order (e.g., Semanturidae, Saturniidae, Cambridae), and thus it is likely that these colour patterns have evolved independently in multiple lineages of this diverse insect clade.

Discal eyespots have also not yet been studied in a phylogenetic context but are widespread in lepidopterans. Because the two wing regions express different genes during development (Banerjee et al., 2023), Marginal and Discal eyespots might have evolved largely independently of each other. If so, we should expect the two eyespot forms to occur primarily in different lineages (Fig. 1E). Alternatively, eyespots might have first evolved on one wing region, and later gained a novel domain

72 of expression on the other region, with the order of these regions potentially varying (Fig. 1F-G).

73 Here, we aim to investigate the evolutionary history of lepidopteran eyespots beyond Nymphalidae,

74 to understand the diversity of paths that resulted in the morphological convergence of this complex

75 colour pattern.

Figure 1. Lepidopteran eyespots have evolved in different wing regions: **A-B** the *Marginal* region, between veins and **C-D** the *Discal* region, straddling the cross vein in the discal cell or found within the cell. **E** These two eyespot types may have evolved independently of each other, and thus be primarily found in different clades. Alternatively, eyespots may first evolve in one region and then acquire a novel expression domain in a different wing region. In the latter scenario, **F** *Discal* eyespots may evolve first followed by *Marginal* eyespots, or **G** Marginal eyespots may enable the subsequent evolution of *Discal* eyespots. Photographs: **A** = *Eupakardia calleta* (Nicky Davis, WildUtah.US.), **B** = *Bicyclus anynana* (Antonia Monteiro, National University of Singapore), **C** = *Limenitis arthemis* (Rawpixel, Rawpixel.com), **D** = *Antheraea polyphemus* (Lacy L. Hyche, Auburn University, Bugwood.org).

We used modern phylogenetic methods to investigate the evolutionary origins of Discal and

Marginal eyespot across Lepidoptera. We first inferred a species-level phylogeny for Lepidoptera,

sampling 5 to 27 molecular sequences per species across 715 taxa. Our phylogeny covers 65% of all

superfamilies in the order Lepidoptera, allowing comparative inferences of eyespot evolution in

moths and butterflies. We then modelled evolutionary origins and losses of eyespots and inferred

ancestral states across the phylogeny. Additionally, for each subtree including multiple (two or

more) eyespot-bearing taxa, we implemented a model comparison approach, based on marginal

likelihood estimation, to quantify support for an ancestral origin of eyespots in the most recent

common ancestor of the subtree. If an ancestral origin of eyespots was supported, we tentatively

concluded that eyespots in all extant taxa of the subtree are likely homologous. These putative

ancestral eyespot origins were then compared with the ancestral state estimates above, to evaluate

the robustness of our results to tree-wide *versus* subtree approaches.

Our results support the hypothesis that eyespots in Lepidoptera have evolved multiple times

independently. Our data also supports the hypothesis that Discal and Marginal eyespots have

evolved in different temporal sequences in the two main clades where they radiated (Nymphalidae

and Saturniidae). We find that both Marginal and Discal eyespots have evolved in a further 10

superfamilies and numerous families across the Lepidopteran tree. Finally, we find that eyespots are

more commonly observed in the Discal wing region (30 independent origins) than the Marginal

region (21 independent origins). Together, our results suggest that eyespot evolution is

commonplace in Lepidoptera. Our study provides a phylogenetic roadmap for the future sampling of

independent lineages to test whether all these eyespots share a common gene-regulatory network.

99
100 **Results**
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102 <u>Tree topology</u>
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While large-scale multi-superfamily phylogenetic trees of Lepidoptera have been inferred in several

recent studies (Heikkilä et al., 2015; Mitter et al., 2017; Kawahara et al., 2019), these studies focused

on resolving deep ancestral nodes within Lepidoptera and sampled taxa accordingly. We aimed,

instead, to maximize taxonomic coverage across the entire clade to enable large-scale comparative

analyses of eyespot evolution, so started by reconstructing a new phylogeny of Lepidoptera. We

focused on Ditrysia, a natural clade that includes 99% of extant species (Heikkilä et al., 2015) and

omitted more basal branching taxa due to inexistent images. We sampled 5 to 27 gene fragments

per species across 715 taxa and conducted maximum likelihood phylogenetic inference using a

generalized time reversible (GTR) substitution model with gamma-distributed site heterogeneity

upon the concatenated alignment. Omission or inclusion of taxa varied across different studies. As

our goal was to include as many superfamilies as possible, our study kept all taxonomically identified

species (Fig. 2), including those which are currently considered to be *incertae sedis* (Uncertain

taxonomic placement) on the lepidopteran tree of life (Heikkilä et al., 2015; Mitter et al., 2017;

Kawahara et al., 2019). Our inferred tree topology is largely congruent with previous phylogenies

(Heikkilä et al., 2015; Mitter et al., 2017; Kawahara et al., 2019).

Topological differences observed between this study and previous ones involve species whose

placement is phylogenetically uncertain (*incertae cedis*), or superfamilies previously shown to

require reclassification (i.e. Tineoidea or the Cossioidea-Sesioidea complex among others; Mutanen

Figure 2. All eyespot occurrences are visually represented in this figure. Species marked with a * are representatives for their specific (Saturniidae or Nymphalidae) taxonomic groups, sharing an eyespot baring common ancestor(s). Colours in the Key represent the corresponding super families in Fig. 3.

significant impact on the main findings of this work, as they are primarily nested within large clades

entirely lacking eyespots. A summary and discussion of novel taxonomic relationships identified in

this work can be found in Supplementary Information (Supplementary information).

<u>Ancestral state reconstructions and evolutionary rates</u>

To investigate the historical evolution of eyespots across Dytrisia, we obtained photographs of as

many species as possible across the phylogeny (only 4 species had missing images) and scored

presence or absence of Discal and/or Marginal eyespots anywhere on the wings of either sex (Fig. 3).

We found that 41 species have Marginal eyespots, 81 species have Discal eyespots, and 12 species

have both eyespot pattern type on their wings across our data set. Eyespots are absent outside the

Nonobectometra (Tineoidea - Epermenoidea) and are also not observed in the Nonobectrometran

Gelechioidea (Fig. 3).

Then, we modelled eyespot evolution as a discrete trait with two alternative states, presence or

absence, regardless of eyespot location on the wings, using RevBayes (v1.21, Höhna et al., 2019). We

monitored joint conditional ancestral states in two independent MCMC simulations and used

stochastic character mapping (Frayman and Höhna, 2019 in Revbayes v1.2.1) to estimate the

number of eyespot gains and losses across the Ditrysia tree. We then repeated these analyses for

Marginal and Discal eyespots separately, to determine if these distinct eyespot types have radiated

in different clades and in different temporal sequences.

Our three ancestral state reconstructions (for the two eyespot types combined, for Marginal, and for

Discal eyespots) indicated that both the most recent common ancestor (MRCA) of all Ditrysia as well

as the MRCA of all Nonobectrometra (including and excluding the Gelenchoidea) did not have

eyespots (Figs. 4, S1, S2). Eyespots, regardless of where they occur, most likely evolved multiple

times independently. We see this through a high number of potential eyespot origins across the

Lepidoptera (Table 1). Eyespots also appear to be evolutionarily short lived, with a higher rate of

losses than origins, regardless of eyespot type (Figs. 4, S1, S2).

Figure 3. Phylogeny of the Lepidoptera compromising 715 species from 28 superfamilies. Superfamily names which are too small to be written on the figure are numbered as follows: 1 = Pterophoridea .2 = Uroidea .3 =Choreutoidea 4. = Immoidea 5. = Galacticoidea .6= Epermenoidea .7 = Hyblaeoidea .8 = Carposinididae .9 = Calliduloidea .10 = Copomorphoidea .11 = Thyridoidea .12 = Hedyloidea .13 = Cimelioidea .14 = Minmalloidea .15 = Drepanoidea .16 = Lasiocampoidea. Species with an eyespot baring common ancestor are highlighted in blue (Marginal) and yellow (Discal). The presence/absence of eyespots of each type for each species is marked at the tips of the tree by a yellow or blue rectangle, as well as a black square indicating the presence or absence of either eyespot type. The origins of each eyespot baring common ancestor, based on a marginal likelihood model comparison approach are shown on ancestral nodes as either red dots (Marginal eyespots) or dark blue dots (Discal eyespots).

Eyespot-bearing species are distributed across 11 superfamilies, but in most cases, eyespots occur in

only one genus or a few genera, within a superfamily. Because taxonomic sampling remains sparse

(22.60% of genera on average across all superfamilies in this study), it is currently difficult to discern

how phylogenetically widespread eyespots are in most of these superfamilies. There are, however,

two major independent occurrences of Marginal and Discal eyespots sharing a single common

ancestor for multiple species in the families Nymphalidae and Saturniidae, where both eyespot types

were retained in multiple lineages after their origin (Figs. 3, 4, S1, S2).

Table 1. Number of independent eyespot origins based on stochastic character maps from discrete-trait models of the evolution of any eyespot type (irrespective of location), Discal eyespots, and Marginal eyespots. For each model we report the median number of origins across the posterior, and the 95% highest posterior density (HPD) interval.

	U95% HPD interval
Discal	[26-35] (Median: 30)
Marginal	[20-23] (Median 21)
Any eyespot type	[33-43] (Median 38)

Figure 4. Eyespot ancestral state reconstruction and transition rates in Lepidoptera for Marginal and Discal eyespots combined. We modelled eyespot evolution as a discrete trait with two alternative states (presence or absence) regardless of marginal or discal location. **A)** Phylogeny of the Lepidoptera with ancestral state reconstructions. Pies indicate the posterior probability of eyespot presence for each internal node. The posterior probability of eyespot presence at the root of Lepidoptera was 0.191. The posterior probability of an eyespot at the base of the Nonobectometra was 0 and finally the posterior probability of an eyespot within the nonobectometra excluding the Gelenchoidea but including all present-day eyespot species is <0.0001. **B)** Posterior distribution of rate estimates for eyespot gain and eyespot loss across Lepidoptera. Higher rate values (i.e. a more rightward distribution) indicate a shorter waiting time for an evolutionary transition (eyespot gain or loss). For any given rate value, a higher posterior density indicates that the rate has high probability, given the model and data. Separate analyses for Marginal and Discal eyespots are shown in the SM (Figs. S1, S2).

 <u>Homology of eyespots with a single common ancestor – common ancestor model vs the multiple</u>
<u>origins model</u>

To validate ancestral state reconstructions pointing to independent eyespot origins, we conducted a

separate test of eyespot homology for each subclade of our Ditrysia tree including at least two

eyespot-bearing taxa (Fig. S3). These tests contrasted the hypothesis of a single ancestral origin of

eyespots in the most recent common ancestor of the subclade (hereafter the 'common ancestor'

model) against the hypothesis of two or more independent eyespot origins within the subclade

(hereafter the 'multiple origins' model). Support for the 'common ancestor' model would be

suggestive of eyespot homology across all extant species in the subclade, especially if congruent

with the results of tree-wide ancestral state reconstructions (Fig. 4), which allowed for eyespot loss

and regain.

The 'common ancestor' and 'multiple origins' models were parametrized as our eyespot evolution

models used for ancestral state reconstructions above (see also Methods), except for one key

difference. We specified alternative priors on the root state frequencies to enforce certainty of an

eyespot presence at the root of each subclade in the 'common ancestor' model, and to enforce

certainty of a common ancestor without eyespots in the 'multiple origins' model. For each subclade,

we then estimated the marginal likelihoods of these two models and compared them using Bayes

factors. Marginal likelihood estimates indicate the likelihood of a model, including all parameters

and priors, given the data. By comparing alternative models that differ only by their prior beliefs on

the character state at the root (presence or absence of eyespots), we thus tested if the assumption

of eyespot homology across extant taxa is more likely, given the phylogeny and character data, than

the assumption of independent eyespot origins.

We found that Discal eyespots are homologous within the Saturniidae and Marginal eyespots are

homologous within the Nymphalidae, according to our SS (Stepping Stone) and PS (Path Sampling)

values (Table 2). The model comparison approach based on all eyespots, Marginal and Discal,

revealed two internal nodes with positive albeit weak support for the 'common ancestor' model (Fig.

3, Table 2). These nodes are positioned close to the base of the MRCA of the Saturniidae and

Nymphalidae families with only the most basal groups excluded in both cases. This result is

consistent with our tree-wide ancestral state reconstructions (Fig. 4). When the circular criteria for

eyespot shape was changed (made more inclusive) to include non-circular shapes (Fig. S4), we found

weak but positive evidence for a more ancestral origin of Discal eyespots in moths, in the common

ancestor of Saturniidae and with the inclusion of some Bombycidae, which traditionally are

considered monophyletic but are paraphyletic in our analysis (BF = 0.144, Fig. S4). We found no

change in the results for Marginal eyespots when making this circular criterion more inclusive, as we

found no new marginal eyespot occurrences.

Table 2. Log 10 of the raw Bayes factors ($\log_{10}$ BF) of comparisons between the 'common ancestor' and the 'multiple-origins' models of eyespot evolution. Values can vary from -1 to +1. For each subclade with two or more eyespot-bearing taxa, we estimated the marginal likelihood of each of these alternative models and compared them via $\log_{10}$ BF. We consider $\log_{10}$ BF > 0.1 as support for the hypothesis of eyespot homology across a subclade. Here, we show subclades with weak but positive evidence for the 'common ancestor' model ($\log_{10}$ BF between 0.1 – 0.5), and Bayes Factors based on marginal likelihood estimates using the stepping stone algorithm. All other subclades examined, either had support for the 'multiple origins' model ($\log_{10}$ BF < -0.1), or showed no positive evidence for either model ($\log_{10}$ BF between -0.1 – 0.1). Results based on marginal likelihood approximation via the path sampling algorithm are qualitatively similar (Table S4). The specific nodes where the 'common ancestor' model was supported for Marginal or Discal eyespots are highlighted in Figs. S1, S2, respectively.

Clade (*subfamily where applicable)	Eyespot type	$\log_{10}$ BF for common ancestor
Saturniidae	All	0.247
Nymphalidae	All	0.150
Saturniidae	Discal	0.146
Nymphalidae (Nymphalini*)	Discal	0.126
Saturniidae (Attacini*)	Marginal	0.110
Nymphalidae	Marginal	0.131

Marginal eyespots, however, evolved more than once within the Saturniidae and are not

homologous. Separate analyses for Discal and Marginal eyespots suggested that the MRCA of the

Saturniidae first evolved Discal eyespots, followed by a later origin of Marginal eyespots in multiple

species in the Attacini tribe of the subfamily Saturniinae (Fig. 3, Table 2), in some species of the tribe

Saturniini (subfamily Saturniinae), and in the subfamily Oxytenninae (Fig. 3, S6, S7). Our analyses,

however, did not support Marginal eyespot homology across these three groups, (all common

ancestors of Attacini and Saturniinae, and Saturniinae and Oxytenninae have BF = -0.94592)

suggesting that Marginal eyespots evolved independently within the Saturniidae.

In the Nymphalidae, Marginal eyespots are homologous, whereas Discal eyespots appear to have

evolved two times independently (Fig. 3, Fig. S5A, Table 2). Discal eyespots evolved in the ancestors

of Heliconiine and Nymphaline whose Discal eyespots are likely homologous (Table 2), and in the

subfamily Satyrinae (Fig. S5A). The data remained unchanged when we extended Discal and
Marginal eyespot shape criterions to allow for non-circular eyespots. Together, our ancestral state
reconstructs, and model-comparison approach point at two taxonomic groups containing eyespot-
bearing lineages with contrasting evolutionary histories: the Saturniidae, where Discal eyespots
evolved first and were followed by a few origins of Marginal eyespots, and the Nymphalidae
butterflies, where Marginal eyespots evolved first and were followed by two independent origins of
Discal eyespots (Fig. 3, S1, S2).

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225 <u>Correlated evolution of Marginal and Discal eyespots</u>
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Our finding of two subclades, Nymphalidae and Saturniidae, with both Marginal and Discal eyespots
arising in different temporal orders (Fig. 3) led to the hypothesis that eyespots may initially evolve
on a single wing domain (Marginal or Discal), but once the genetic machinery for eyespot
development becomes established, eyespot evolution on a different wing domain is facilitated.
Critically, this novel-domain hypothesis is compatible with either wing region hosting the most
ancestral eyespot, but it requires evidence that the ancestral origin facilitated eyespot evolution in
novel domains, as would be expected if the eyespot gene-regulatory network is redeployed in a
different group of cells as opposed to evolving anew.

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We addressed this hypothesis using a correlated evolution model, wherein the rate of eyespot
evolution on each wing region depends on whether eyespots are already present on the alternative
region. To test for correlated evolution of eyespots between wing regions (Marginal and Discal), we
modelled a reversible jump mixture distribution, sampling the posterior probabilities of correlated vs
independent eyespot evolution, and assuming equal prior probabilities for both scenarios. If
correlated evolution is supported, origin of novel Marginal and Discal eyespots should be
accelerated when eyespots already exist in the alternative wing domain, whereas no such
acceleration should be present if eyespot gains are independent. We tested this hypothesis in the
Papilionoidea (containing Nymphalidae) and Bombycoidea (containing Saturniidae), as both clades
encompass multiple putative origins of both Discal and Marginal eyespots (Figs S1 & S2), and
therefore enable estimation of evolutionary rates on each eyespot domain.

247
Our results demonstrated that Discal and Marginal eyespots are highly unlikely to be evolutionarily
correlated (Figs. S8). Specifically, we see a posterior probability of 36.3% for papilionid discal
eyespots and a posterior probability of 36.4% for papilionid marginal eyespots to evolve in
correlation with each other. In Bombycoidea, we observed a posterior probability of 46.38% for

discal and 56.14% for marginal eyespots to evolve in correlation with each other (Fig. S8C). Both

probabilities are too low (<95%) to ascertain with certainty whether these eyespot domains evolved

independently or not. These results indicate that with the present data, the reversible jump model is

ambiguous between the two evolutionary scenarios. Consistently, the rate of Discal eyespot origins

in Papilionoidea is similar for lineages with and without pre-existing Marginal eyespots, and the rate

of Marginal eyespot origins in Bombycoidea is similar for lineages with and without pre-existing

Discal eyespots (Fig. S8BC). While we found no unequivocal support for correlated evolution

between Discal and Marginal eyespots in either Papilionoidea and Bombycoidea, modelling the joint

evolution of both eyespot types resulted in inferred ancestral origins that were largely consistent

with our previous model comparison approach and previous tree-wide ancestral state

reconstructions of individual eyespot domains (Figs. 3, S9.

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264 <u>Other smaller eyespot observations</u>
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Our stochastic character mapping analyses suggested that both Discal and Marginal eyespots have

evolved multiple times in Dytrisia (Table 1), with only a few of these origins occurring in

Nymphalidae and Saturniidae. The remaining origins of Discal and Marginal eyespots, occurred in

sparsely sampled families (superfamilies), namely : Brahmaeidae (Bombycoidea), Sphingidae

(Bombycoidea), Carthaeidae (Bombycoidea), Cossidae (Cossioidea-Sesioidea), Castniidae

(Cossioidea-Sesioidea), Drepanidae (Drepanoidea), Geometridae (Geometroidea), Sematuridae

(Geometroidea), Hesperiidae (Papilionoidea), Lycaenidae (Papilionoidea), Papilionidae

(Papilionoidea), Pieridae (Papilionoidea), Riodinidae (Papilionoidea), Lasiocampidae

(Lasiocampoidea), Mimallonidae (Mimallonoidea), Erebidae (Noctuoidea), Noctuidae (Noctuoidea),

Crambidae (Pyraloidea) and Limacodidae (Zygaenoidea, Figs 2, 3, S4, Table S5). These families

represent potential subjects for future comparative studies if phylogenies are or become available,

and eyespots are found in more than a handful of species.

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To obtain a preliminary picture of whether eyespots are widespread within these clades, we

searched image databases for close relatives of eyespot-bearing taxa within these families and

recorded presence or absence of eyespots not included in our Ditrysia phylogeny. We looked at

every image available of species within these families and found that 59% of the species with

eyespots represented in Fig. 3 have one or more congeners (species with which they share a genus)

also with eyespots. Of the remaining species that did not have congeners with eyespots, 32% were

part of a subfamily, and 3% part of a family, with eyespot-bearing species, and 6% of species (three

taxa) did not have any congener, subfamily, or family member with eyespots. This could be due to

lack of image data (*Pterodecta felderi* and *Helicomitra pulchra*) or due to these clades being

monotypic to the family level (*Carthaea saturnioides*). A visual summary of the species found on our

tree with eyespots and up to 3 close relatives also supporting eyespots is provided in Fig. S10 and

Table S5.

Discussion

Our results show that eyespot evolution in Lepidoptera is diverse and complex. Most eyespot

research up to now has been conducted in a single clade (Nymphalidae), but here we demonstrate

that eyespots have evolved multiple times independently across the Lepidoptera, including in many

poorly known moth clades (Figs. 2, 3). Our stochastic character mapping analysis demonstrated that

the median number of discal eyespot origins on our tree is 30 and the median number of marginal

eyespots is 21 (Table 1). Most of the clades with eyespots on our phylogeny have low taxonomic

coverage, but we documented eyespots in close relatives of each of these lineages (Fig. S7),

suggesting a less "tippy" distribution than that documented in Fig. 3. Future phylogenetic studies,

with a denser taxonomic sampling, will be required to characterize the evolutionary history of

eyespots in these lesser-known clades. Nonetheless, our study strongly suggests that eyespots are

not homologous across the order Lepidoptera, i.e., the MRCA of all Lepidoptera with eyespots did

not have eyespots. We find that this result is robust to alternative analytical approaches as both

ancestral state reconstructions (from tree-wide eyespot evolution models), and comparisons of

different root-state assumptions within subclades, point at independent eyespot origins in the two

main clades where these traits are known to be widely distributed, the Brush-footed butterflies

(Nymphalidae) and the saturniid moths.

Here we provided the first evidence that Discal eyespots are homologous in the Saturniidae but not

so in the Nymphalidae. Discal eyespots, present in 62% of the represented clades within the family

Saturniidae in this study, are likely to have originated at or near the base of the family (Figs. 3, 4, S6,

S9B). This result stands despite the large diversity of eyespot complexity, colour and size observed

across the Saturniidae (D'Abrera, 1995;1998;2012, Figs. S10). All three analyses (common ancestor

vs multiple origins, ancestral state reconstruction and correlated evolution), support a single Discal

eyespot common ancestor. The evolutionary history of nymphalid Discal eyespots had not been

investigated before and our analyses point to two separate origins within this clade.

Marginal eyespots are homologous within the Nymphalidae but not so in the Saturniidae. The
evolution of Marginal eyespots in the Nymphalidae has been studied before and we report similar
findings to these previous studies. We report a single origin for these eyespots that aligns with both
the 'early' model of eyespot evolution first proposed by Oliver et al. (2012), and the 'late' model
proposed later by Oliver et al. (2014). These two models merely differed in pinpointing the clade
where eyespots first originated, due to a different way of scoring eyespots on the wing. In the 'early'
model, eyespots were scored as being present or absent anywhere on the wing, whereas in the 'late'
model eyespots were scored for specific wing sectors. In our analyses, both results were observed.
The early model was supported by our common ancestor vs multiple origins model comparisons (Fig.
2, Table 2), while the late model was supported by our ancestral state reconstructions (Figs. 3, S2,
S3). The current study, as well as these previous studies, therefore, agree on a single and relatively
early origin of Marginal eyespots within Nymphalidae. Marginal eyespots in the Saturniidae,
however, are not homologous, as we observed four independent origins within this monophyletic
clade. Marginal eyespots evolved and radiated within the Attacini tribe of the subfamily Saturniinae
which includes species of the genus *Samia, Callosamia* and *Hyalophora,* as well as *Epiphora*.
Marginal eyespots evolved separately in species from the genus *Copaxa, Eupakardia* (monotypic)
and *Asthenidia*.
337
Other than what is seen in the Families Nymphalidae and Saturniidae we observe a further 28 Discal
and 19 Marginal eyespot occurrences close to the tips of our tree. Some of these groups are
taxonomically unique e.g., the bombycoid moth *Cathaea saturnoides* (Walker, 1858) which is
monotypic to the level of the family and supports a single large Discal eyespot on each wing surface.
Other species represent more speciose groups such as moths from the Geometrid family
Semanturiidae, or butterflies from the genus *Parnassius* which support interesting and unique
Marginal eyespots.
345
Marginal and Discal eyespots in both the Bombycoidea and the Papilionoidea appear to have
evolved independently from each other. While our taxon sampling for both clades is small, we did
not find support for the hypothesis that the evolution of one eyespot type promoted the evolution
of the other (Figs. S9). Our analyses, however, revealed that eyespots within those clades appeared
in a specific order. Marginal eyespots appeared first in Papilionoidea, followed by Discal eyespots,
whereas the reverse was observed in Bombycoidea.
352

Despite the independent origin of eyespots in the Papilionoidea and Bombycoidea, two

superfamilies currently understood to be ~110 million years apart (Kawahara et al., 2019), it is still

possible that these (and other) Lepidopteran eyespots share the same gene regulatory network

(GRN). Previous research by Murugesan et al. (2022) found that an appendage gene regulatory

network was co-opted to build Marginal eyespots in *Bicyclus anynana*, a nymphalid butterfly. It is

possible that the same GRN was co-opted more than once across the Lepidoptera. To test this

hypothesis, it will be important to molecularly characterize Marginal and Discal eyespot GRN in moth

lineages as well as Discal eyespots in nymphalid butterflies. Early immunochemistry work in two

saturniid species detected the presence of the two eyespot marker proteins previously identified in

nymphalids, Distal-less and Engrailed (Monteiro et al. 2006). Stronger evidence for the use of the

same appendage GRN in these Discal moth eyespots may need to come from CRISPR knockouts of

cis-regulatory elements belonging to common eyespot and appendage genes, showing that both

traits are affected (Murugesan et al. 2022). Demonstrating that all eyespots use a single GRN, or,

alternatively, that different "eyespot" GRNs can evolve using different genes and/or cis-regulatory

elements is a highly interesting question for future research. This line of research would help us

understand molecular mechanisms of convergence of complex phenotypic traits.

From an ecological perspective, the origin of eyespots may be linked to the evolution of body and/or

wing size as eyespots evolved only in the larger members of Dytrisa. This hypothesis is further

strengthened by the occurrence of eyespots in large non Ditrysian primitive Lepidoptera, such as

Zelotypia stacyi (Scott, 1869)*, a* Hepialidae, which was not part of this study. This species has large

Discal eyespots, as well as a large body and wing size (Grehan & Mielke 2018).

The large number of independent origins for eyespots suggests that these traits can evolve readily,

and especially in large wings, perhaps in a quantitative and gradual way. Previous genetic studies

showed that eyespot number in *Bicyclus anynana* hindwings was regulated by a variety of QTLs

around many different genes (Rivera-Colon et al. 2020). This suggests that eyespots differentiate

following some threshold response to quantitative genetic variation. In addition, previous reaction-

diffusion models also showed that eyespot centres could form on the wing by tweaks in a variety of

different parameters (Nihout 1990; Connahs et al. 2019). It is possible that larger wing sector size, as

well as multiple other genetic parameters, facilitate the stable expression of *Distal-less* in the middle

of the wing sector, an essential top regulator of eyespots (Connahs et al. 2019; Matsuoka et al.

2022). Given that the expression of *Distal-less* is linked to the co-option of the appendage GRN

(Murugesan et al. 2022), if *Distal-less* becomes stably expressed in a wing sector, it is possible that

the same appendage GRN gets co-opted every time eyespots independently evolve on the wings of

Lepidoptera. This idea, along with our hypothesis that body size is linked to eyespot development

will require further testing.

Concluding remarks
392
393 While the focus of this work is on eyespot evolution, the phylogeny presented here provides

394 evidence supporting both new and already established relationships among lepidopteran lineages, at

395 the superfamily and potentially family level (see supplementary information). These insights can fuel

396 future systematic research and further comparative work on how wing colour patterns, and other

397 adaptive traits, evolve in Lepidoptera.

398
399 **Materials and methods**
400
401 Molecular data
402
DNA sequence data for 645 species of Lepidoptera (moths and butterflies) and seven species of

Trichoptera (caddisflies, outgroup) were kindly provided by Professor Emeritus Charles Mitter

(University of Maryland, Table S1). Additional sequences for 70 species of Saturniidae (giant silk

moths) were downloaded from NCBI (GenBank). The full dataset was composed of 27 protein-coding

genes (Table S2), with 5-24 fragments available across all species. Our taxonomic sampling includes

~68% (90/133, Van Nieukerken et al., 2011) of all families and ~65% (28/43, Van Nieukerken et al.,

2011) of all superfamilies in the order Lepidoptera. This study focuses on the suborder Ditrysia,

which comprises ~99% of currently described lepidopterans. Of these, ~90% of families (90/100,

Reiger et al., 2009) and ~93-96% (28/29-30) of superfamilies (Van Nieukerken et al., 2011; Heikkilä et

al., 2015; Mitter et al., 2017) are represented. For each superfamily, 1 – 128 species were sampled

(median = 14). The full list of superfamilies as well as the number of species representatives for each

superfamily are outlined in Table S1. We chose the Trichoptera for our outgroup because they are

the closest extant relatives to the Lepidoptera (Mey et al., 2017), but distant enough to provide

ingroup monophyly.

417
418 Sequence alignment and phylogenetic inference
419
DNA sequences were aligned via MAFFT v7.490 (Katoh & Standley, 2013). We used a global

alignment strategy with iterative refinement (G-INS-i) to maximize alignment accuracy based on

weighted sum-of-pairs and a consistency score. The length of the final alignment, consisting of all 27

concatenated gene fragments, was 22,643 bp.

Phylogenetic inference under maximum likelihood was done with IQ-TREE v1.6.12 (Nguyen et al.,

2015, Kalyaanamoorthy et al., 2017; Hoang et al., 2018). All genes were subject to a single

substitution process automatically set by IQ-TREE, using ModelFinder (Kalyaanamoorthy et al.,

2017). Substitutions followed a generalized time reversible (GTR) model, with estimated base

frequencies. Rate variation among sites followed a gamma distribution containing 10 categories.

Node support was estimated using ultra-fast bootstrap in UFBoot (Hoang et al., 2018) for 100,000

iterations. UFBoot is an efficient approximation alternative to the traditional bootstrap method and

it is particularly well-suited for large datasets such as ours. After pruning the outgroup, our

maximum likelihood tree was transformed into an ultrametric tree, with branch lengths scaled to

time, using the *ape* package v5.7-1 (Paradis & Schliep, 2019) in R v4.3.0 (R Core Team 2023). We

assumed an age of ~290 Ma for the most recent common ancestor of Ditrysia (the root node of our

tree), based on the most recent dated phylogeny of Lepidoptera (Kawahara et al., 2019).

<u>Image data collection</u>

We scored images of each of the 715 species in our phylogeny for presence or absence of eyespot

patterns (in all wing surfaces and in both sexes). Colour patterns were identified as eyespots if they

met the following criterion: Patterns contained a circular or oblong spot encircled by one or more

concentric rings of a distinct colour that also contrasted with the overall background colour of the

wing (Fig. 1). To explore the sensitivity of our results to this conservative criterion, we repeated our

main comparative analyses under a more inclusive definition of eyespots, where non-circular/oblong

shapes (e.g., triangles, rectangles, teardrops, Fig. S10) encircled by another ring of Colour were also

considered. Eyespots were further classified into Marginal or Discal, based on their locations on the

wing surface (Figs. 1).

We first queried a wide range of online databases for images of the type specimen of each of the

sampled species (Table S3, Supplementary data). If not available, images of other (non-type)

specimens were collected from the same databases with a preference for museum specimens over

other specimens. In cases where sequence data was assigned to a particular subspecies, we

prioritized images of the same taxonomic rank when available. 96 species included in our dataset

lacked publicly available and/or reliably identified images of any form. We photographed 81 of these

species at the Mcguire Center for Lepidoptera (MGCL, Florida Museum of Natural History, 3215 Hull

Rd, Gainesville, Fl 32611), using a Cannon D50 DSLR camera. We were unable to obtain images for 4

species (0.56%), which are treated as missing data in all comparative analyses.

<u>Modelling the origin and loss of eyespots in Lepidoptera</u>

461
To model eyespot evolution across Lepidoptera, we used RevBayes v1.2.1 (Höhna et al., 2016).

RevBayes is an open-source software designed for Bayesian phylogenetic inference. It allows users

to build probabilistic graphical models using an interactive model-specification language. Eyespots

were modelled as a discrete trait with two states (presence/absence). Eyespot states had unequal

transition rates, drawn from identical exponential priors. The rate parameters of these priors were

set to reflect an expectation of 10 events (10 eyespot gains and 10 eyespot losses) along the tree.

Root state frequencies were in turn drawn from a Dirichlet prior, assuming equal probabilities of

presence or absence of eyespots at the origin of the tree. Eyespot evolution was modelled

separately using three datasets: eyespot presence/absence irrespective of eyespot location on the

wing; presence/absence of Marginal eyespots; and presence/absence of Discal eyespots. For all

three datasets, eyespots could be located on any wing surface (dorsal/ventral surfaces of fore or

hindwings) and in either sex.

Each model was run for 100,000 iterations after an initial burn-in of 10,000, and tuning parameter

proposals every 1,000 iterations on two independent chains. We evaluated the convergence and

stationarity of the MCMC chains for each model using the R package *Convenience* v1.0.0 (Fanreti et

al., 2013). Joint conditional ancestral states were sampled every 100 iterations and plotted using

RevGadgets v1.1.1 (Tribble et al., 2023), *ggplot2* v3.4.3 (Wickham et al., 2016) and *ggtree* v3.9.0 (Yu

et al., 2017 & 2018, Yu, 2020 & 2022; Xu et al., 2022). Posterior probability data for specific nodes

was obtained using the R package Treeio (Wang et al., 2020). in R. Stochastic character histories (in

Revbayes 1.2.1, Frayman and Höhna, 2019) were sampled every 100 iterations and summarized by

computing the median number of transitions (eyespots gains and losses) and corresponding 95%

highest posterior density (HPD) intervals. ITOL was used to enhance the aesthetics of phylogenetic

figures (ITOL, 2023).

486
487 <u>Testing eyespot homology in selected clades</u>
488
We next implemented a model testing approach to investigate eyespot homology among extant

species of Lepidoptera. We identified all subclades including two or more taxa bearing eyespots,

regardless of eyespot location. If the most recent common ancestor (MRCA) of these clades also

displayed eyespots, it is likely that eyespots in extant taxa are homologous — i.e. inherited from

their common ancestor. To test this hypothesis, we extracted all the above-mentioned subclades

(Fig. 5 for schematic example) from the complete ultrametric phylogeny (Fig. 2), and for each

subclade applied two alternative versions of the discrete-trait model described previously. In the

first version (hereafter the 'multiple origins' model), we constrained the root state to eyespot

absence, by assigning a fully informative prior to the root state parameter. In the second version

(the common ancestor model), we instead enforced a MRCA with eyespots by setting the root

frequency of eyespot presence to one.

We subsequently computed the marginal likelihood of the two alternative models for each subclade

and for our entire phylogeny. A marginal likelihood indicates the fit of a model, averaged across its

entire parameter space, to a data set. Marginal likelihood estimation is routinely used in

phylogenetic analysis, for instance, in the selection of appropriate nucleotide substitution models

and for species delimitation applications (Oaks et. Al 2019). Unlike other model comparison

approaches, marginal likelihoods are weighted by the prior, as prior beliefs are bound to influence

model fit (Xie et al. 2011). Here, the two models being compared (the 'multiple origins' model and

the 'common ancestor' model) differ only by the prior controlling the character state at the root.

Their direct comparison thus serves as a statistical test for the presence of eyespots in the most

recent common ancestor (MRCA) of each subclade with eyespots. Marginal likelihood was estimated

via two sampling methods, steppingstone sampling (SS) and path sampling (PS) (Lartillot and

Philippe, 2006, Fan et al., 2011; Xie et al., 2011). For both methods the power posterior analysis was

split into 50 intervals between the prior and posterior and was run for 5,000 iterations with a burn-in

of 5,000 generations. We repeated all marginal likelihood approximations and confirmed that both

PS and SS estimates were stable (i.e. differing by no more than 0.5 between independent runs).

Marginal likelihood estimates were then used to calculate the log 10 of the raw Bayes Factor ($\log_{10}$

BF) between the 'multiple origins' and 'common ancestor' models. Positive $\log_{10}$BF values support

the 'common ancestor' model and were interpreted as follows: < 0.1 = weak evidence not worth

mentioning, 0.1 – 0.5 weak evidence but worth mentioning, 0.5 – 1.0 strong evidence, and > 1.0 very

strong evidence. Negative values were in turn interpreted as support for the multiple origins model.

The main results of these analyses were summarized on the tree using ITOL (2023) after initial

visualization in R using *phytools* v1.9-16 (Revell, 2012), *ggplot2* v3.4.3 (Wickham et al., 2016) and

ggtree v3.9.0 (Yu et al., 2017 & 2018, Yu, 2020 & 2022; Xu et al., 2022).

<u>Correlated evolution of Marginal and Discal eyespots</u>

We then focused on two clades (Papilionoidea and Bombycoidea), in which both Marginal and Discal

eyespots have evolved in distinctly ordered sequences (see Results). These findings suggested that

an ancestral eyespot origin on one wing region (Marginal or Discal) could facilitate the subsequent

evolution of eyespots on the alternative region, by redeploying the ancestral eyespot gene-

regulatory network on a novel tissue domain. We tested this hypothesis using a correlated evolution

analysis for each clade, where the probability of gaining Marginal or Discal eyespots can depend on

whether the alternative eyespot type is already present. To do this, we modelled a reversible jump

mixture distribution, which samples the posterior probabilities of parameter spaces for correlated

and independent eyespot evolution models. Support for the correlated evolution model in the

posterior distribution of the reversible jump parameter would indicate that the evolution of one

eyespot types depends on the presence of the other and could then be followed up with inspection

of transition rates estimates to determine if ancestral eyespot expression facilitates the evolution of

new eyespot domains. Alternatively, support for the independent model would suggest that Discal

and Marginal are similarly likely to evolve whether the alternative eyespot type is present or absent.

As in our previous analyses, we modelled all transition rates (eyespots gains and losses on each wing

region) as drawn from identical exponential priors, Revbayes v1.2.4 (Höhna et al., 2016) was used to

run the comparative evolution analysis. The rate parameter of the exponential priors corresponded

to a mean of 10 transitions across the tree. Root frequencies were drawn from a flat Dirichlet

distribution. The models were run for 10,000,000 iterations, with a pre-burn-in of 25% and tuning

parameter proposals every 5,000 iterations on two independent chains. Joint conditional ancestral

states were sampled every 10,000 iterations and plotted using *RevGadgets* v1.1.1 (Tribble et al.,

2023) in R. We evaluated the convergence and stationarity of the MCMC chains for each model using

the R package *Convenience* V1.0.0 (Fanreti et al., 2013).

551
552 Identifying potential eyespot pattern common ancestors
553
Our initial image data collection included only species with available sequence data, resulting in

several taxa (19 families in 11 superfamilies) with a single eyespot (Discal and/or Marginal) bearing

taxon. To determine if these taxa belong to larger a single common ancestor of eyespot-bearing

species, warranting future research, we investigated the phenotype of all species currently classed in

the same family as one of the species on our tree (via images available online) which were part of

the 28 discal and/or 19 other marginal eyespot occurrences identified by our stochastic mapping

analysis. Whether species closely related to taxa represented in our tree (Fig. 2) have a Discal and or

Marginal eyespot pattern. Due to the large number of species and the variation in number of species

with eyespots across the 19 different families (11 superfamilies) we identified as having eyespots,

we summarize our findings in both table and figure form with up to 3 species per family (Fig. S10,

Table S5).

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References:

Aarvik & Karisch in Aarvik L, Karisch T, plazi (2009). Revision of Multiquaestia, Karisch (Lepidoptera:
Tortricidae: Grapholitini). Plazi.org taxonomic treatments database. Checklist dataset
https://doi.org/10.5281/zenodo.274748 accessed via GBIF.org on 2023-02-22.

Banerjee, T. D., Murugesan, S. N., Connahs, H., & Monteiro, A. (2023). Spatial and temporal
regulation of Wnt signaling pathway members in the development of butterfly wing patterns.
Science Advances, 9(30), eadg3877.

Bazinet, A. L., Cummings, M. P., Mitter, K. T., & Mitter, C. W. (2013). Can RNA-Seq resolve the rapid
radiation of advanced moths and butterflies (Hexapoda: Lepidoptera: Apoditrysia)? An exploratory
study. PloS one, 8(12), e82615.

Beddard, F. E. (1895). Animal coloration: an account of the principal facts and theories relating to the
colours and markings of animals: S. Sonnenschein.

Bhardwaj, S., Jolander, L. S.-H., Wenk, M. R., Oliver, J. C., Nijhout, H. F., & Monteiro, A. (2020). Origin
of the mechanism of phenotypic plasticity in satyrid butterfly eyespots. Elife, 9, e49544.
Blest, A. D. (1957). The function of eyespot patterns in the Lepidoptera. Behaviour, 209-256.

Brakefield, P. M. (2003). The power of evo-devo to explore evolutionary constraints: experiments
with butterfly eyespots. Zoology, 106(4), 283-290.

Breuker, C. J., & Brakefield, P. M. (2002). Female choice depends on size but not symmetry of dorsal
eyespots in the butterfly Bicyclus anynana. Proceedings of the Royal Society of London. Series B:
Biological Sciences, 269(1497), 1233-1239.

Caro, T., Sherratt, T. N., & Stevens, M. (2016). The ecology of multiple colour defences. Evolutionary
ecology, 30, 797-809.

Carter, D. J., & Hargreaves, B. (1986). A field guide to caterpillars of butterflies and moths in Britain
and Europe: Collins.

Chazot, N., Condamine, F. L., Dudas, G., Peña, C., Kodandaramaiah, U., Matos-Maraví, P., ... &
Wahlberg, N. (2021). Conserved ancestral tropical niche but different continental histories explain
the latitudinal diversity gradient in brush-footed butterflies. Nature Communications, 12(1), 5717.

Crees, L. D., DeVries, P., & Penz, C. M. (2021). Do hind wing eyespots of Caligo butterflies function in
both mating behavior and antipredator defense?(Lepidoptera, Nymphalidae). Annals of the
Entomological Society of America, 114(3), 329-337.

Cunha, C., Narotamo, H., Monteiro, A., & Silveira, M. (2023). Detection and measurement of
butterfly eyespot and spot patterns using convolutional neural networks. Plos one, 18(2), e0280998.
620
621   D'Abrera, B. (1995). Saturniidae Mundi, Saturniid moths of the world part I. Keltern, Germany:
622   Automeris press in association with Victoria Australia Melborne & London: Hill house.
623
624   D'Abrera, B. (2012). Saturniidae Mundi, Saturniid moths of the world part II. Victoria Australia,
625   Melborne & London: Hill house.
626
627   D'Abrera, B. (1998). Saturniidae Mundi, Saturniid moths of the world part III. Keltern, Germany:
628   Goecke & Evers in association with Victoria Australia Melborne & London: Hill house.
629
630   De Prins, J., & Kawahara, A. Y. (2012). Systematics, revisionary taxonomy, and biodiversity of
631   Afrotropical Lithocolletinae (Lepidoptera: Gracillariidae). Zootaxa, 3594(1), 1–283-281–283.
632
633   De Prins W. & Steeman C. 2003–2023. *Catalogue of the Lepidoptera of Belgium*. Last accessed on
634   the 22/02/2023. Available online at https://projects.biodiversity.be/lepidoptera
635
636   Dilão, R., & Sainhas, J. (2004). Modelling butterfly wing eyespot patterns. Proceedings of the Royal
637   Society of London. Series B: Biological Sciences, 271(1548), 1565-1569.
638
639   Edmunds, M. (1976). The defensive behaviour of Ghanaian praying mantids with a discussion of
640   territoriality. Zoological Journal of the Linnean Society, 58(1), 1-37.
641
642   Epstein, M. E. (1996). Revision and phylogeny of the limacodid-group families, with evolutionary
643   studies on slug caterpillars (Lepidoptera: Zygaenoidea).
644
645   Fabreti, L. Hoehna, S & Magee, A. (2023). _convenience: Convergence assessment for phylogenetic
646   inference_. R package version 1.0.0.
647
648   Fan, Y., Wu, R., Chen, M.-H., Kuo, L., & Lewis, P. O. (2011). Choosing among partition models in
649   Bayesian phylogenetics. Molecular biology and evolution, 28(1), 523-532.
650
651   Freyman, W. A., & Höhna, S. (2019). Stochastic character mapping of state-dependent diversification
652   reveals the tempo of evolutionary decline in self-compatible Onagraceae lineages. Systematic
653   Biology, 68(3), 505-519.
654
655   Benson, D. A., Cavanaugh, M., Clark, K., Karsch-Mizrachi, I., Ostell, J., Pruitt, K. D., & Sayers, E. W.
656   (2018). GenBank. *Nucleic acids research*, *46*(D1), D41-D47.
657
658   Grehan, J. R., & Mielke, C. G. (2018). Evolutionary biogeography and tectonic history of the ghost
659   moth families Hepialidae, Mnesarchaeidae, and Palaeosetidae in the Southwest Pacific (Lepidoptera:
660   Exoporia). Zootaxa, 4415(2), 243-275.
661
662   Halali, D., Krishna, A., Kodandaramaiah, U., & Molleman, F. (2019). Lizards as predators of
663   butterflies: shape of wing damage and effects of eyespots. The Journal of the Lepidopterists' Society,
664   73(2), 78-86.
665
666   He, B., Liu, Y., Shi, H., Zhang, J., Hu, M., Ma, Y., Fu, L., Hong, M., Wang, J., Fong, J.J., Parham, J.F.
667   (2010). Captive breeding of the Four-eyed Turtle (Sacalia quadriocellata). Asian Herpetological
668   Research, 1(2), 111-117.

669
670   Heikkilä, M., Kaila, L., Mutanen, M., Peña, C., & Wahlberg, N. (2012). Cretaceous origin and repeated
671   tertiary diversification of the redefined butterflies. Proceedings of the Royal Society B: Biological
672   Sciences, 279(1731), 1093-1099.
673
674   Heikkilä, M., Mutanen, M., Kekkonen, M., & Kaila, L. (2014). Morphology reinforces proposed
675   molecular phylogenetic affinities: a revised classification for Gelechioidea (Lepidoptera). Cladistics,
676   30(6), 563-589.
677
678   Heikkilä, M., Mutanen, M., Wahlberg, N., Sihvonen, P., & Kaila, L. (2015). Elusive ditrysian phylogeny:
679   an account of combining systematized morphology with molecular data (Lepidoptera). BMC
680   Evolutionary Biology, 15(1), 1-27.
681
682   Hernández-Palma, T. L., Rueda-Solano, L. A., Valkonen, J. K., & Rojas, B. (2023). Predator response to
683   the coloured eyespots and defensive posture of Colombian four-eyed frogs. Journal of Evolutionary
684   Biology, 36(7), 1040-1049.
685
686   Hoang, D, T. Chernomor, O. Von Haeseler, A. Minh, B, Q & Vinh, L, S. (2018). UFBoot2: Improving the
687   ultrafast bootstrap approximation. Mol. Biol. Evol., 35:518–522.
688
689   Höhna, Landis, Heath, Boussau, Lartillot, Moore, Huelsenbeck, Ronquist. (2016). RevBayes: Bayesian
690   phylogenetic inference using graphical models and an interactive model-specification language.
691   Systematic Biology, 65:726-736.
692
693   Hombría, J. C.-G. (2011). Butterfly eyespot serial homology: enter the Hox genes. BMC biology, 9(1),
694   1-3.
695
696   Hossie, T. J., Skelhorn, J., Breinholt, J. W., Kawahara, A. Y., & Sherratt, T. N. (2015). Body size affects
697   the evolution of eyespots in caterpillars. Proceedings of the National Academy of Sciences, 112(21),
698   6664-6669.
699
700   Hsu Y-F, Powell JA (2005) Phylogenetic relationships within Heliodinidae and systematics of moths
701   formerly assigned to Heliodines Stainton (Lepidoptera: Yponomeutoidea). University California
702   Publications Entomology 124: 1–158.
703
704   Huq, M., Bhardwaj, S., & Monteiro, A. (2019). Male *Bicyclus anynana* butterflies choose females on
705   the basis of their ventral UV-reflective eyespot centers. Journal of insect science, 19(1), 25.
706   Jeffreys, H. (1998). The theory of probability: OuP Oxford.
707
708   Kalyaanamoorthy, S. Minh, B, Q. Wong, T, K, F. Von Haeseler, A. & Jermiin, L, S. (2017). ModelFinder:
709   Fast Model Selection for Accurate Phylogenetic Estimates, Nature Methods, 14:587–589.
710   Katoh, K. & Standley, D. M. (2013). MAFFT Multiple Sequence Alignment Software Version 7:
711   Improvements in Performance and Usability. Molecular Biology and Evolution, 30(4), 772–780.
712
713   Kawahara, A., Ohshima, I., Kawakita, A., Regier, J., Mitter, C., Cummings, M., . . . Lopez-Vaamonde, C.
714   (2011). Increased gene sampling provides stronger support for higher-level groups within gracillariid
715   leaf mining moths and relatives (Lepidoptera: Gracillariidae). BMC Evol. Biol, 11, 182.
716
717   Kawahara, A. Y., & Rubinoff, D. (2012). Three new species of Fancy Case caterpillars from threatened
718   forests of Hawaii (Lepidoptera, Cosmopterigidae, Hyposmocoma). ZooKeys(170), 1.
719

720   Kawahara, A. Y., & Breinholt, J. W. (2014). Phylogenomics provides strong evidence for relationships
721   of butterflies and moths. Proceedings of the Royal Society B: Biological Sciences, 281(1788),
722   20140970.
723

724   Kawahara, A. Y., Plotkin, D., Ohshima, I., LOPEZ-VAAMONDE, C., Houlihan, P. R., Breinholt, J. W., . . .
725   Davis, D. R. (2017). A molecular phylogeny and revised higher-level classification for the leaf-mining
726   moth family Gracillariidae and its implications for larval host-use evolution. Systematic Entomology,
727   42(1), 60-81.
728

729   Kawahara, A. Y., Plotkin, D., Espeland, M., Meusemann, K., Toussaint, E. F., Donath, A., . . . Dos Reis,
730   M. (2019). Phylogenomics reveals the evolutionary timing and pattern of butterflies and moths.
731   Proceedings of the National Academy of Sciences, 116(45), 22657-22663.
732

733   Kawahara, A. Y., Storer, C., Carvalho, A. P. S., Plotkin, D. M., Condamine, F. L., Braga, M. P., . . .
734   Lohman, D. J. (2023). A global phylogeny of butterflies reveals their evolutionary history, ancestral
735   hosts and biogeographic origins. Nature Ecology & Evolution. doi:10.1038/s41559-023-02041-9
736   Kristensen, N. P., & Schmidt-Rhaesa, A. (1998). Volume 1: Evolution, Systematics, and Biogeography
737   (Vol. 1): Walter de Gruyter.
738

739   Kodandaramaiah, U. (2011). The evolutionary significance of butterfly eyespots. Behavioral Ecology,
740   22(6), 1264-1271.
741

742   Kodandaramaiah, U., Lindenfors, P., & Tullberg, B. S. (2013). Deflective and intimidating eyespots: a
743   comparative study of eyespot size and position in Junonia butterflies. Ecology and evolution, 3(13),
744   4518-4524.
745

746   Kyaw, K. M. M., Yagi, S., Oku, J., Sakamaki, Y., & Hirowatari, T. (2019). Taxonomic study of Thiotricha
747   Meyrick (Lepidoptera, Gelechiidae) in Japan, with the description of two new species. *ZooKeys*, *897*,
748   67.
749

750   Labandeira, C. C., Yang, Q., Santiago-Blay, J. A., Hotton, C. L., Monteiro, A., Wang, Y.-J., . . . Rose, T. R.
751   (2016). The evolutionary convergence of mid-Mesozoic lacewings and Cenozoic butterflies.
752   Proceedings of the Royal Society B: Biological Sciences, 283(1824), 20152893.
753

754   Lartillot, N., & Philippe, H. (2006). Computing Bayes factors using thermodynamic integration.
755   Systematic biology, 55(2), 195-207.Miller, J. Y. (2008). Studies in the Castniidae. V. Description of a
756   New Species of Zegara: Florida Museum of Natural History, University of Florida.
757

758   Loyau, A., Gomez, D., Moureau, B., Théry, M., Hart, N. S., Jalme, M. S., . . . Sorci, G. (2007). Iridescent
759   structurally based coloration of eyespots correlates with mating success in the peacock. Behavioral
760   Ecology, 18(6), 1123-1131.
761

762   Meadows, D. W. (1993). Morphological variation in eyespots of the foureye butterflyfish (Chaetodon
763   capistratus): implications for eyespot function. Copeia, 1993(1), 235-240.
764

765   Merilaita, S., Vallin, A., Kodandaramaiah, U., Dimitrova, M., Ruuskanen, S., & Laaksonen, T. (2011).
766   Number of eyespots and their intimidating effect on naive predators in the peacock butterfly.
767   Behavioral Ecology, 22(6), 1326-1331.
768

Mey, W., Wichard, W., Müller, P., & Wang, B. (2017). The blueprint of the Amphiesmenoptera-
Tarachoptera, a new order of insects from Burmese amber (Insecta, Amphiesmenoptera). Fossil
Record, 20(2), 129-145.

Miller, J. Y. (2008). *Studies in the Castniidae. V. Description of a New Species of Zegara*. Florida
Museum of Natural History, University of Florida.

Mitter, C., Davis, D. R., & Cummings, M. P. (2017). Phylogeny and evolution of Lepidoptera. Annual
review of entomology.

Mizuno, A., Lagisz, M., Pollo, P., Yang, Y., Soma, M., & Nakagawa, S. (2024). A systematic review and
meta-analysis of anti-predator mechanisms of eyespots: conspicuous pattern vs eye mimicry.

Monteiro, A., Glaser, G., Stockslager, S., Glansdorp, N., & Ramos, D. (2006). Comparative insights
into questions of Lepidopteran wing pattern homology. BMC Developmental Biology, 6(1), 1-13.

Monteiro, A., Chen, B., Scott, L. C., Vedder, L., Prijs, H. J., Belicha-Villanueva, A., & Brakefield, P. M.
(2007). The combined effect of two mutations that alter serially homologous color pattern elements
on the fore and hindwings of a butterfly. BMC genetics, 8, 1-10.

Monteiro, A. (2015). Origin, development, and evolution of butterfly eyespots. Annual review of
entomology, 60, 253-271.

Muff, S., Nilsen, E. B., O'Hara, R. B., & Nater, C. R. (2022). Rewriting results sections in the language
of evidence. Trends in ecology & evolution, 37(3), 203-210.

Mukherjee, R., & Kodandaramaiah, U. (2015). What makes eyespots intimidating–the importance of
pairedness. BMC Evolutionary Biology, 15(1), 1-10.

Murugesan, S. N., Connahs, H., Matsuoka, Y., Das Gupta, M., Tiong, G. J., Huq, M., . . . Werner, T.
(2022). Butterfly eyespots evolved via cooption of an ancestral gene-regulatory network that also
patterns antennae, legs, and wings. Proceedings of the National Academy of Sciences, 119(8),
e2108661119.

Mutanen, M., Wahlberg, N., & Kaila, L. (2010). Comprehensive gene and taxon coverage elucidates
radiation patterns in moths and butterflies. Proceedings of the Royal Society B: Biological Sciences,
277(1695), 2839-2848.

Nguyen, L, T. Schmidt, Heiko A. Haeseler, A & Minh, B, Q. (2015). IQ-TREE: A fast and effective
stochastic algorithm for estimating maximum likelihood phylogenies.

National Center for Biotechnology Information (NCBI)[Internet]. Bethesda (MD): National Library of
Medicine (US), National Center for Biotechnology Information; [1988] – [cited 2022 Sept 06].
Available from: https://www.ncbi.nlm.nih.gov/

Oliver, J. C., Tong, X.-L., Gall, L. F., Piel, W. H., & Monteiro, A. (2012). A single origin for nymphalid
butterfly eyespots followed by widespread loss of associated gene expression.

Oliver, J. C., Beaulieu, J. M., Gall, L. F., Piel, W. H., & Monteiro, A. (2014). Nymphalid eyespot serial
homologues originate as a few individualized modules. Proceedings of the Royal Society B: Biological
Sciences, 281(1787), 20133262.

Otaki, J. M. (2020). Morphological and spatial diversity of the discal spot on the hindwings of
nymphalid butterflies: Revision of the nymphalid groundplan. Insects, 11(10), 654.

Paradis, E. & Schliep, K. (2019). ape 5.0: an environment for modern phylogenetics and evolutionary
analyses in R. Bioinformatics 35: 526-528.

Phillips, E., & Solís, M. A. (1996). Neurophyseta (Lepidoptera: Crambidae) de Costa Rica. *Revista de*
Biología Tropical, *44*(2B), 693-717.

Plummer, M. Best, N. Cowles, K. & Vines, K. (2006). CODA: Convergence Diagnosis and Output
Analysis for MCMC, R News, vol 6, 7-11.

Poulton, E. B. (1890). The colours of animals: their meaning and use, especially considered in the
case of insects (Vol. 67): D. Appleton.

R Core Team (2023). R: A language and environment for statistical computing. R Foundation for
Statistical Computing, Vienna, Austria. URL https://www.R-project.org/.

Regier, J. C., Zwick, A., Cummings, M. P., Kawahara, A. Y., Cho, S., Weller, S., . . . Parr, C. (2009).
Toward reconstructing the evolution of advanced moths and butterflies (Lepidoptera: Ditrysia): an
initial molecular study. BMC Evolutionary Biology, 9(1), 1-21.

Regier, J. C., Brown, J. W., Mitter, C., Baixeras, J., Cho, S., Cummings, M. P., & Zwick, A. (2012). A
molecular phylogeny for the leaf-roller moths (Lepidoptera: Tortricidae) and its implications for
classification and life history evolution. PLoS ONE, 7(4), e35574. (2012A)

Regier, J. C., Mitter, C., Mitter, K., Cummings, M. P., Bazinet, A. L., Hallwachs, W., . . . Zwick, A.
(2017). Further progress on the phylogeny of Noctuoidea (Insecta: Lepidoptera) using an expanded
gene sample. Systematic Entomology, 42(1), 82-93.

Regier, J. C., Mitter, C., Solis, M. A., Hayden, J. E., Landry, B., Nuss, M., . . . Cummings, M. P. (2012). A
molecular phylogeny for the pyraloid moths (Lepidoptera: Pyraloidea) and its implications for higher-
level classification. Systematic Entomology, 37(4), 635-656. (2012B)

Regier, J. C., Mitter, C., Zwick, A., Bazinet, A. L., Cummings, M. P., Kawahara, A. Y., . . . Davis, D. R.
(2013). A large-scale, higher-level, molecular phylogenetic study of the insect order Lepidoptera
(moths and butterflies). PLoS ONE, 8(3), e58568.

Regier, J. C., Mitter, C., Davis, D. R., Harrison, T. L., SOHN, J. C., Cummings, M. P., . . . Mitter, K. T.
(2015). A molecular phylogeny and revised classification for the oldest ditrysian moth lineages (L
epidoptera: T ineoidea), with implications for ancestral feeding habits of the mega-diverse D itrysia.
Systematic Entomology, 40(2), 409-432.

Regier, J. C., Mitter, C., Solis, M. A., Hayden, J. E., Landry, B., Nuss, M., . . . Cummings, M. P. (2012). A
molecular phylogeny for the pyraloid moths (Lepidoptera: Pyraloidea) and its implications for higher-
level classification. *Systematic Entomology, 37*(4), 635-656.

Revell, L. J. (2012) phytools: An R package for phylogenetic comparative biology (and other things).
Methods in Ecology and Evolution, 3, 217-223.

Rivera-Colón, A. G., Westerman, E. L., Van Belleghem, S. M., Monteiro, A., & Papa, R. (2020).
Multiple loci control eyespot number variation on the hindwings of Bicyclus anynana butterflies.
Genetics, 214(4), 1059-1078.

Schachat, S. R., Oliver, J. C., & Monteiro, A. (2015). Nymphalid eyespots are co-opted to novel wing
locations following a similar pattern in independent lineages. BMC evolutionary biology, 15(1), 1-9.

Sohn, J.-C., Regier, J. C., Mitter, C., Davis, D., Landry, J.-F., Zwick, A., & Cummings, M. P. (2013). A
molecular phylogeny for Yponomeutoidea (Insecta, Lepidoptera, Ditrysia) and its implications for
classification, biogeography and the evolution of host plant use. PLoS ONE, 8(1), e55066.

Sohn, J. C., Regier, J. C., Mitter, C., Adamski, D., LANDRY, J. F., Heikkilä, M., . . . Zwick, A. (2016).
Phylogeny and feeding trait evolution of the mega-diverse Gelechioidea (Lepidoptera:
Obtectomera): new insight from 19 nuclear genes. Systematic Entomology, 41(1), 112-132.

Sohn, J.-C., & Choi, S.-W. (2016). First report of Saridoscelinae (Lepidoptera, Yponomeutidae) in
Korea with new records of Saridoscelis kodamai Moriuti from Korea and China. Korean journal of
applied entomology, 55(4), 347-350.

Sondhi, S., Karmakar, T., Sondhi, Y., & Kunte, K. (2021). Moths of Tale Wildlife Sanctuary, Arunachal
Pradesh, India with seventeen additions to the moth fauna of India (Lepidoptera: Heterocera).
Tropical Lepidoptera Research, 1-53.

Stevens, M. (2005). The role of eyespots as anti-predator mechanisms, principally demonstrated in
the Lepidoptera. Biological Reviews, 80(4), 573-588.

Stevens, M. (2007). Predator perception and the interrelation between different forms of protective
coloration. Proceedings of the Royal Society B: Biological Sciences, 274(1617), 1457-1464.

Stevens, M., Hardman, C. J., & Stubbins, C. L. (2008). Conspicuousness, not eye mimicry, makes
"eyespots" effective antipredator signals. Behavioral Ecology, 19(3), 525-531.

Tylor, A. (1886). Colouration in animals and plants: Alabaster, Passmore and Sons.

Tribble, C. May, M. Freyman, W. Landis, M. Ying, L. Barido-Sottani, J. Magee, A. Kopperud, B &
Hohna, S. (2023). _RevGadgets: Visualization and Post-Processing of 'RevBayes' Analyses

Van Nieukerken, E., Kaila, L., Kitching, I., Kristensen, N., Lees, D., Minet, J., . . . Simonsen, T. (2011).
Animal Biodiversity: an Outline of Higher-level Classification and Survey of Taxonomic Richness.
Zootaxa (3148).

Wang, L.-G., Lam, T. T.-Y., Xu, S., Dai, Z., Zhou, L., Feng, T., Guo, P., Dunn, C. W., Jones, B. R., Bradley,
T., Zhu, H., Guan, Y., Jiang, Y., & Yu, G. (2020). treeio: an R package for phylogenetic tree input and
output with richly annotated and associated data. Molecular Biology and Evolution, 37(12), 3443-
3445. https://doi.org/10.1093/molbev/msaa137

Weber, M. G., & Agrawal, A. A. (2012). Phylogeny, ecology, and the coupling of comparative and
experimental approaches. Trends in ecology & evolution, 27(7), 394-403.

Wickham, H. (2016). ggplot2: Elegant Graphics for Data Analysis. Springer-Verlag New York.
921

922 Wickham H, François R, Henry L, Müller K, Vaughan D (2023). dplyr: A Grammar of Data
923 Manipulation. R package version 1.1.41(https://github.com/tidyverse/dplyr)
924
925 Wong, V. L., & Marek, P. E. (2019). Super black eyespots of the Eyed elater (2167-9843).
926
927 Xie, W., Lewis, P. O., Fan, Y., Kuo, L., & Chen, M.-H. (2011). Improving marginal likelihood estimation
928 for Bayesian phylogenetic model selection. Systematic biology, 60(2), 150-160.
929
930 Xu, S. Lin, L. Luo, X. Chen, M. Tang, W. Zhan, L. Dai, Z. Lam, T. Guan, Y. & Yu, G. (2022). Ggtree: A
931 serialized data object for visualization of a phylogenetic tree and annotation data. iMeta 2022,
932 4(1):e56. doi:10.1002/imt2.56
933
934 Yu, G. (2020). Using ggtree to visualize data on tree-like structures. Current Protocols in
935 Bioinformatics, 2020, 69:e96. doi: 10.1002/cpbi.96
936
937 Yu, G. (2022). Data Integration, Manipulation and Visualization of Phylogenetic Trees (1st edition).
938 Chapman and Hall/CRC. doi:10.1201/9781003279242
939
940 Yu, G. Lam, T. Zhu, H. & Guan, Y. (2018). Two methods for mapping and visualizing associated data
941 on phylogeny using ggtree. Molecular Biology and Evolution 2018, 35(2):3041-3043. doi:
942 10.1093/molbev/msy194
943
944 Yu, G. Smith, D. Zhu, H. Guan, & Y. Lam, T. (2017). ggtree: an R package for visualization and
945 annotation of phylogenetic trees with their covariates and other associated data. Methods in
946 Ecology and Evolution 2017, 8(1):28-36. doi:10.1111/2041-210X.12628
947
948 Zahiri, R., Kitching, I. J., Lafontaine, J. D., Mutanen, M., Kaila, L., Holloway, J. D., & Wahlberg, N.
949 (2011). A new molecular phylogeny offers hope for a stable family level classification of the
950 Noctuoidea (Lepidoptera). Zoologica Scripta, 40(2), 158-173.
951
952 Zhu, H. (2024). kableExtra: Construct Complex Table with 'kable' and Pipe Syntax. R package version
953 1.4.0. DOI: 10.32614/CRAN.package.kableExtra