Title: Changing rounds into squares or combining stripes: Diversity and

formation of checkerboard patterns in Eukaryotes

Authors : Pierre Galipot 1, Julie Zalko, 2

Affiliations:

1. UMR CNRS 6553 ECOBIO, Campus Beaulieu, Rennes, France

2. Université Paris-Saclay, INRAE, CNRS, AgroParisTech, Génétique Quantitative et

Evolution-Le Moulon, IDEEV 12 route 128, 91190 Gif-sur-Yvette, France

Abstract

Important in many human artistic cultures, checkerboard patterns are rare in nature

like many motifs based on squared geometry. Nevertheless, they are expected to be

very detectable by the visual networks due to their periodic geometry, and diverse

plant and animal species bear them, suggesting specific biological functions. Here,

thanks to a biological survey, we first draw the diversity of species bearing

checkerboard patterns. Then, we selected two genera, Sarcophaga *flies and* Fritillaria

flowers to perform simulations and functional studies to decipher the mechanisms

producing these very peculiar patterns. Although they share a similar geometry, these

two genera appear to produce checkered patterns through two very different ways,

showing a convergence of shape but not of mechanism. Together, this shows the

extent of the mechanisms selected during evolution to generate complex forms, and

confirms the importance of describing color patterns through the species diversity

Introduction

Contrary to hexagons and circles, squares patterns are much more uncommon in

nature and many studies have proven the difficulty to produce squares and the instability of

such patterns[1] [2]. Nevertheless, square lattices (to which checkerboard patterns belong) have

been described in some eukaryotic tissues, like the mouse auditory epithelium[3] or the

oviduct epithelium of the Japanese quail[4]. Concerning color patterning, checkerboard

patterns are the combination of two characteristics: a square lattice, and an alternation of

coloration. The latter is very common in color patterning, and many processes are able to

produce them, like the ubiquitous Turing-like mechanism[5]. Two possibilities coexist, either

squares and color alternation are directly produced by the same processus, (in the same

way that in a Turing-like pattern the dots and their hexagonal arrangement are produced by

the same mechanism[6]), or produced by two separated mechanisms, simultaneously or not,

which combine. As they possess a double periodicity, they are expected to be easily

detected by visual systems[7] [8] [9].

If some species are quite famously known for their checkerboard color patterns like the

Fritillaria flower (Liliaceae) which bear them on their tepals[10], few species have been

formerly described for those peculiar shapes, and no work has already presented them

combined in a single study. We then chose to describe as many checkerboard patterns as

possible, using multiple data sources, taking care to cover the maximum of eukaryotic

biodiversity.

After the species survey, a large range of potential species to study was available and we

decided to choose and compare an animal and an angiosperm species (and their genera):

Sarcophaga flies and *Fritillaria* flowers. They presented the patterns closest to a perfect

checkerboard motif, and were accessible to functional studies, although not genetically.

Simulations, microscopy observations and species comparison allowed us to propose two

distinct mechanisms of formations, thanks to a cluster of clues.

Results

Checkerboard color patterns are rare but present in many diverse taxa of Eukaryotes,

notably plant and animal species

To perform a survey of species bearing checkerboard patterns, we first had to define

them properly. We chose to consider two separate characteristics of checkerboard patterns:

square shaped-motifs and an alternation of two colors [11]. Perfect checkerboard patterns are

never reached by real patterns, so we chose to extend the definition and include species

with slightly diverging characteristics, like not perfect squares or partly irregular color

alternation. Since no exhaustive survey was possible, we chose to use keywords to find the

species ('checkerboard', 'checkered', 'square' and '*fritillaria'*, the latin word for 'dice cornet', a

object exhibiting a checkerboard pattern and then used in species names to pinpoint the

presence of this particular pattern, or one close to it[12]).

Thanks to this, we described several species and genera, summarized in Figure 1, and

qualitatively represented in a diagram thanks to the more or less perfectness of their

characteristics. Both angiosperms and animals were found to exhibit checkerboard-like

patterns, but we were not able to find any species belonging to Fungi. Very diverse animal

species (vertebrates, insects, echinoderms, cephalopods) were found, but few plant species

seem to bear such patterns. The two most perfect checkerboard-like patterns were found in

Sarcophaga flies genus, exhibited on their abdomens, and *Fritillaria* genus, with *Fritillaria*

meleagris. We then chose these two groups to explore pattern formation.

Figure 1. Checkerboard-like patterns of Eukaryotes species. x-axis: distance to perfect squares motifs. y-axis:
pattern alternation regularity. From up to down and left to right: Stomoxys calcitrans, Thamnophix marcianus,
Sarcophaga sp., Synapta maculata, Sepia officinalis, Fritillaria sp., Bulbophyllum fritilariiflorum, Trogonophis
wiegmanni, Huntleya sp., Lutjanus decussatus.

Fritillaria *checkerboard patterns are compatible with a Turing-like mechanism constrained by*

a parallel nervure frame

To extract clues of *Fritillaria* mechanisms producing their checkerboard-like patterns,

we performed microscopy on fresh individuals (Figure 2.A and B) and also from museum

specimens presenting interesting features on their tepals, such as slight deviation from the

classical patterns. We found that depending on the species, colored shaped go from squares

(notably in *F. meleagris*) to sponge shapes for the pigmented areas (i.e. straight on the sides

touching the nervures and with a negative curvature on the other two sides), and round

shapes for the unpigmented areas. In the most distal part of the tepals, sponge colored

shapes tend to break into two parts (Figure 1.B, left panels). In some individuals, especially

in the *Fritillaria meleagris* museum specimens, we observed a very interesting feature in the

places where the nervures are the widest: instead of alternations, several motifs were side

by side, suggesting that nervures act as constraining barriers, but when the patterning

mechanisms has sufficiently space, a spot pattern is produced. Given the periodicity of this

pattern, and the round geometry of the motifs, this strongly suggests a Turing-like

mechanism for its production[13]. Indeed, at that time, it is the only mechanism demonstrated

to produce periodic color dots patterning in angiosperms[14] [15] . Turing pattern relies

classically on reaction and diffusion, and it seems that nervures could act as constraining

semi-permeable barriers, which therefore shape the pattern into square-like shapes. Two

clues suggest that nervures are semi-permeable to the diffusing agent (whatever its nature).

First, no motif overlaps two nervures, indicating an impeded diffusion[16] [17]. But if nervures

were totally impermeable, there is no reason that motifs perfectly alternate from one 'line' to

the next one, which is what we observe in every *Fritillaria* species. The most reasonable

hypothesis is the possibility for a signal to diffuse to a long range, across nervures.

To sum up, to produce a final checkerboard-like patterns, several mechanisms need to

interact: first, the lateral sides of the squares are produced by the straightness and

parallelism of the tepal nervures. Secondly, the two other sides are more or less straight,

depending on the species, and might depend on the Turing parameters of the system.

Finally, the same Turing-like mechanism seems to be involved to produce an alternation of

colored and uncolored zones, characteristics of the checkerboard-like patterns. This is

summarized into a flowchart (Figure 4.A).

Sarcophaga checkerboard patterns are relying on a structural coloration generated by planar polarity of hair, compatible with the interference between two stripe patterns

To explore *Sarcophaga* checkerboard-like patterns, we use several museum specimens of several species in the *Sarcophaga* genus. First, we observe the very peculiarity of these patterns. While it may appear as a black and white squares alternation, the reality is more complex. First, between two consecutive squares, there are tiny black zones, whatever the lighting. Then, under directional lighting, the black squares become light and vice versa depending on the angle of illumination and observation (Figure 3.A). These color changing patterns might then rely at least partially on structural coloration instead of only classical pigments. This was even more strongly suggested when specimens were recovered by a thin layer of platinum for SEM microscopy. Most of the time, this is sufficient

to cover and hide pigment-based patterns. On the contrary, checkerboard-like patterns were

still visible, so the thin layer of platinum that molded the structure underneath seems to have

taken a role of structural coloration[18] [19]. Under the SEM, we observed that the external

tissues of the abdomen were covered by two types of hair, a big and sparse one, and small

and coarse one. Only the latter is compatible with structural coloration, because of its

micrometric scale [20]. By qualitatively describing in a first part the mean direction and length

of the hair (Figure 3.B, third panel), we observe a perfect correlation between the

characteristics of the hair and the colors. The small black gaps between two squares are

made up of tiny hairs, with not strong preferential direction, which give a glimpse of the

structure underneath. On the contrary, color-changing areas correlate with dense hair with a

strong preferential direction. Furthermore, if we compare two areas with opposite coloration

behavior, (i.e. one is light when the other is black under a particular illumination angle), the

hair length is similar, but their direction is symmetrical in relation to the right-left axis of the

abdomen. Together, this strongly suggests that hair is responsible for the color behavior of

the abdomen, through planar polarity[21].

Then, to explain the organizing into squares, we involved two observations: first, if we look at

the thorax of the *Sarcophaga* flies, it is characterized by a striped pattern, parallel to the left-

right axis[22]. If we hypothesized that this mechanism of stripe patterning is also present in the

abdomen, it could explain alternations of the checkerboard-like pattern parallel to the antero-

posterior axis. In the perpendicular direction, we observed that there are exactly two

alternations per abdomen segments. Many insect species including many flies possess a

bicolor coloration in each abdomen segment [23] [24], one color on the anterior side, and the

other on the posterior side. If we assume that this is also the case for Sarcophaga flies, this

could explain the alternations perpendicular to the antero-posterior axis. Finally, as

coloration seems to be coded by the preferential direction of the hair, the combination of two

perpendicular signals may explain this checkerboard-pattern. This is summarized into a

flowchart (Figure 4.B).

Figure 3. Checkerboard-like patterns of Sarcophaga sp. A: detail of Sarcophaga sp. abdomen color patterns depending on the incident light angle. Square color changes from dark to light depending on the light angle. B: upper panels: details of the abdomen under light microscopy (left) and SEM (right). Fro, up to down panels: detail of the most anterior segment, hair phenotype details, preferential direction and size vectors, color correspondence.

Figure 4. Morphogenesis flowchart of Fritillaria sp. (A) and Sarcophaga sp. (B) checkerboard-like patterns. Yellow rectangles: shape, pink trapezoid: generative mechanism, blue shape: Modifying mechanism.

Material and methods

Species survey and phylogenetic tree

The sources of photography were multiple, scientific literature was preferred, then online encyclopedias, Wikipedia (https://en.wikipedia.org/) and Wikimedia Commons (https://commons.wikimedia.org), eventually alternative sources (through search engines) with keywords such as "checkerboard", "checkered", "squares", "fritillaria".

Phylogenetic relationships between species were built with phyloT (V2, https://phylot.biobyte.de/) based on NCBI taxonomy. Phylogenetic trees were plotted and annotated in iTOL (Interactive Tree Of Life, version 6.8.1, https://itol.embl.de/) [25].

Scanning Electron Microscopy

A total of three abdomens were sampled from *Sarcophaga* sp. museum specimens (National Museum of Natural History, Paris, France). Samples were mounted on the aluminum stubs with colloidal graphite, sputter-coated with platinum using an EM ACE600 fine coater (Leica, Wetzlar, Germany), and observed using a SU3500 scanning electron microscope (SEM, Hitachi, Tokyo, Japan).

Light Microscopy

A total of 20 tepals were sampled from 8 specimens of *Fritillaria thunbergii*, harvested in the botanical gardens of the University of Tokyo (Koishikawa Botanical Gardens, Tokyo, Japan). Tepals gas were extracted by a vacuum pump to make the tissues more transparent and suitable for observation. Abaxial and adaxial sides were mounted in water and observed under light microscopy.

A total of 8 tepals were sampled from 3 specimens of *Fritillaria meleagris*, from museum specimens (National Museum of Natural History, Paris, France), after a survey of hundreds of museum specimens. They were rehydrated and observed under light microscopy.

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References

1. Yang, L., Dolnik, M., Zhabotinsky, A. M. & Epstein, I. R. Turing patterns beyond hexagons and stripes. *Chaos: An Interdisciplinary Journal of Nonlinear Science* **16**, 037114 (2006).

2. Li, J., Wang, H. & Ouyang, Q. Square Turing patterns in reaction-diffusion systems with coupled layers. *Chaos: An Interdisciplinary Journal of Nonlinear Science* **24**, 023115 (2014).

3. Togashi, H. *et al.* Nectins Establish a Checkerboard-Like Cellular Pattern in the Auditory Epithelium. *Science* **333**, 1144–1147 (2011).

4. Yamanaka, H. I. & Honda, H. A checkerboard pattern manifested by the oviduct epithelium of the Japanese quail. *Int J Dev Biol* **34**, 377–383 (1990).

5. Kondo, S. The present and future of Turing models in developmental biology. *Development* **149**, dev200974 (2022).

6. Chen, Y. & Buceta, J. A non-linear analysis of Turing pattern formation. *PLOS ONE* **14**, e0220994 (2019).

7. De Valois, K. K., De Valois, R. L. & Yund, E. W. Responses of striate cortex cells to grating and checkerboard patterns. *The Journal of Physiology* **291**, 483–505 (1979).

8. Issa, N. P., Trepel, C. & Stryker, M. P. Spatial Frequency Maps in Cat Visual Cortex. *J. Neurosci.* **20**, 8504–8514 (2000).

9. Broderick, W. F., Simoncelli, E. P. & Winawer, J. Mapping spatial frequency preferences across human primary visual cortex. *Journal of Vision* **22**, 3 (2022).

10. Ramawat, K. G., Mérillon, J.-M. & Shivanna, K. R. *Reproductive Biology of Plants*. (CRC Press, 2014).

11. Peng, C.-H., Jiang, C., Wonka, P. & Pottmann, H. Checkerboard patterns with black rectangles. (2019).

12. Day, P. D. Studies in the genus Fritillaria L. (Liliaceae). (Queen Mary University of London, 2018).

13. Kondo, S. An updated kernel-based Turing model for studying the mechanisms of biological pattern formation. *Journal of Theoretical Biology* **414**, 120–127 (2017).

14. Galipot, P., Damerval, C. & Jabbour, F. The seven ways eukaryotes produce repeated colour motifs on external tissues. *Biological Reviews* **96**, 1676–1693 (2021).

15. Ding, B. *et al.* Two MYB Proteins in a Self-Organizing Activator-Inhibitor System Produce Spotted Pigmentation Patterns. *Current Biology* **30**, 802-814.e8 (2020).

16. Manukyan, L., Montandon, S. A., Fofonjka, A., Smirnov, S. & Milinkovitch, M. C. A living mesoscopic cellular automaton made of skin scales. *Nature* **544**, 173–179 (2017).

17. Fofonjka, A. & Milinkovitch, M. C. Reaction-diffusion in a growing 3D domain of skin scales generates a discrete cellular automaton. *Nat Commun* **12**, 2433 (2021).

18. Minh, N. H., Kim, K., Kang, D. H., Yoo, Y.-E. & Yoon, J. S. Fabrication of robust and reusable mold with nanostructures and its application to anti-counterfeiting surfaces based on structural colors. *Nanotechnology* **32**, 495302 (2021).

19. Saison, T. *et al.* Replication of butterfly wing and natural lotus leaf structures by nanoimprint on silica sol–gel films. *Bioinspir. Biomim.* **3**, 046004 (2008).

20. Iridescence | SpringerLink. https://link.springer.com/chapter/10.1007/978-0-387-34120-0_1.

21. Maung, S. M. T. W. & Jenny, A. Planar cell polarity in Drosophila. *Organogenesis* **7**, 165–179 (2011).

22. Sinha, S. K. & Mahato, S. Intra-puparial development of flesh fly Sarcophaga dux (Thomson) (Diptera, Sarcophagidae). *Current Science* **111**, 1063–1070 (2016).

23. KonDo, Y. *et al.* Toll ligand Spätzle3 controls melanization in the stripe pattern formation in caterpillars. *Proc Natl Acad Sci U S A* **114**, 8336–8341 (2017).

24. Raja, K. K. B. *et al.* The regulation of a pigmentation gene in the formation of complex color patterns in Drosophila abdomens. *PLOS ONE* **17**, e0279061 (2022).

25. Letunic, I. & Bork, P. Interactive Tree Of Life (iTOL) v4: recent updates and new developments. *Nucleic Acids Research* **47**, W256–W259 (2019).