First evidence for the evolution of host manipulation by tumors during the long-term

vertical transmission of tumor cells in *Hydra oligactis*

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Abstract

While host phenotypic manipulation by parasites is a widespread phenomenon, whether

tumors – which can be likened to parasite entities, can also manipulate their hosts is not

known. Theory predicts that this should nevertheless be the case, especially when tumors

(neoplasms) are transmissible. We explored this hypothesis in a cnidarian *Hydra* model

system, in which spontaneous tumors can occur in the lab, and lineages in which such

neoplastic cells are vertically transmitted (through host budding) have been maintained for

over 15 years. Remarkably, the hydras with long-term transmissible tumors show an

unexpected increase in the number of their tentacles, allowing for the possibility that these

neoplastic cells can manipulate the host. By experimentally transplanting healthy as well as

neoplastic tissues derived from both recent and long-term transmissible tumors, we found

that only the long-term transmissible tumors were able to trigger the growth of additional

tentacles. Also, supernumerary tentacles, by permitting higher foraging efficiency for the

host, were associated with an increased budding rate, thereby favoring the vertical

transmission of tumors. To our knowledge, this is the first evidence that, like true parasites,

transmissible tumors can evolve strategies to manipulate the phenotype of their host.

Introduction

It is now widely established that animals (metazoans) are not autonomous entities, but rather holobionts composed of the host cell community, various commensal and mutualistic microorganisms, a diversity of parasitic taxa, and in many cases, tumor cells (Ujvari et al. 2016a, Dheilly et al. 2019). While it has been shown that parasites and mutualistic organisms have frequently evolved the ability to manipulate the host phenotype [e.g., reproduction, behavior, morphology, physiology, body odor (2012, Johnson and Foster 2018, Naundrup et al. 2022)] to favor their transmission and/or survival, sometimes inducing dramatic alterations (Doherty 2020), this phenomenon has been poorly investigated in the context of host-tumor interactions. The only unambiguous evidences of such manipulation is found in the tumoral microenvironment, when, for instance, malignant cells manipulate the surrounding healthy cells to induce the growth of blood vessels that bring oxygen and nutrients to the tumor, a process called neoangiogenesis (Hicklin and Ellis 2005). Evidence regarding the ability of tumor cells to manipulate their host phenotype at other levels is scarce [see (Tissot et al. 2016) for a review] and this probably reflects a real rarity of this phenomenon. Indeed, host manipulation capabilities often rely on sophisticated and complex adaptations that evolve during long coevolutionary processes, whereas tumors (neoplasms) most often evolve for only a few years and then die with their host without being able to transmit any acquired traits (Arnal et al. 2015). However, like "true parasites", transmissible tumor cells [vertically – such as in *Hydra oligactis* (Domazet-Lošo et al. 2014, Boutry et al. 2022b); or horizontally – such as in Tasmanian devils (Jones et al. 2008)] could more easily evolve manipulative abilities (Tissot et al. 2016, Ujvari et al. 2016b). Nevertheless, to date, we lack evidence that transmissible cancers can manipulate the phenotype of their hosts.

Hydra oligactis are cnidarians that frequently develop spontaneous tumors in the lab,

noticeable as individuals with significant increases in body size and visible deformities. These

tumors correspond to abnormal proliferation of germline cells in the hydra's ectoderm

(Boutry et al. 2023). In 2014, Domazet-Lošo and colleagues (Domazet-Lošo et al. 2014)

described a fascinating case of two hydra lineages harboring neoplasms capable of vertical

transmission through buds, during the asexual reproduction of hydras. Thanks to this

transmission, these two tumorous hydra lines have been maintained under laboratory

conditions for more than 15 years now (Lange 2010). Beyond the presence of large tumor

masses, a curious feature of these hydras is their increased number of tentacles. Specifically,

while healthy polyps typically have 6-7 tentacles, tumorous polyps have between 8 and 13

tentacles, even up to 20 (Domazet-Lošo et al. 2014). This phenotype is particularly

impressive given that tentacle numbers in hydras usually vary in a very small range [from 5

to 8 tentacles, depending on temperature and body size (Shostak 2012)]. Recently, Boutry

and colleagues (Boutry et al. 2022a) demonstrated that tumorous hydras featuring these

supernumerary tentacles capture more prey than healthy ones, regardless of whether prey

is rare or abundant in the environment. In contrast, wild specimens of *H. oligactis*

developing spontaneous tumors when placed in laboratory conditions do not seem to

develop additional tentacles (Domazet-Lošo et al. 2014). Since food intake, survival, as well

as budding rate are usually positively correlated in hydras (Tökölyi et al. 2016), the presence

of additional tentacles in certain tumorous hydras is likely to have an adaptive value,

favoring host survival despite the cost of bearing tumors, and this can ultimately enhance

the hydra's reproductive potential and/or the tumor cells' transmission [see also (Boutry et

al. 2022b)].

However, whether the host or the tumor is responsible for the alteration of the

tentacles number is not clear yet. Furthermore, in one *H. oligactis* lineage, called the St.

Petersburg strain, tumor initiation and maintenance require the presence of a specific

microbiome (Rathje et al. 2020), suggesting that bacteria may also be involved in the

phenotypic alteration of the host to promote their own transmission. Here, through a series

of experiments using a set of healthy and tumor tissue grafts from tumors associated with

different phenotypes, we attempted to determine whether the host or the tumor is

responsible for the growth of additional tentacles in the tumorous hydra.

Material and Methods

Biological material

All hydras were maintained according to standard rearing protocols (Boutry et al.

2022a). Three different main lineages of *Hydra oligactis* were used in our study. The Saint

Petersburg (named thereafter SpB) and the Robusta (named thereafter Rob) strains were

obtained from Thomas Bosch's laboratory in Kiel (Domazet-Lošo et al. 2014). Rob are also

known under the denomination *Pelmatohydra robusta*, as they were described as a distinct

specie from Japan (Ito 1947). Current phylogenetic analyses suggest than they belong to *H.*

oligactis species. (Martínez et al. 2010), and it is currently unknown if populations related to

this strain still exist in nature. For each of these two lines, control lines were also isolated

from healthy polyps collected during the same sampling; these control groups do not harbor

any sign of transmitted tumors and are supposed to be genetically identical to their

respective tumorous lines (Boutry et al. 2022b). A third tumorous lineage has been recently

established in Frédéric Thomas's laboratory in France, and was isolated from a polyp

sampled in Montaud water pond (43.748211°N, 3.989871°E); this lineage, named Montaud

(thereafter Mt), was maintained in the lab since August 2021 (Boutry et al. 2023). To date,

we have been unable to establish a distinct control and vertically transmitted tumorous

lineage for this new strain, due to variable transmission rates of these tumors and the

sporadic occurrence of spontaneous tumors from initially healthy- looking polyps. In

addition, a group named "wild group" was used as recipient of the transplanted tissues; this

group was composed of randomly sampled *H. oligactis* and assumed to be genetically

diverse and distinct from the SpB and Rob lineage. These wild hydras were also sampled in

the Montaud pond, between 15/02/2022 and 15/04/2022.

In this study, two types of tumors were distinguished. "Transmitted tumors"

correspond to those from the tumorous hydra lineages (SpB and Rob) obtained from Thomas

Bosch's laboratory in Kiel; hydras bearing these tumors possess an enhanced number of

tentacles and these tumors have been vertically transmitted through budding for 15 years

(Lange 2010). "Spontaneous tumors" correspond to tumors that sporadically appear in Mt

lineage, but also among healthy polyps of the SpB control lineage (which were isolated from

the common culture as soon as they appeared). Noticeably, tumors were never observed to

appear spontaneously in the Rob control lineage.

Grafting protocol

The transplantation experiment was performed according to (Lenhoff 2013, Dujon et

al. 2022) using healthy and tumorous donors of the SpB, Rob, and Mt lineages. For the

recipients, we used healthy individuals of the SpB lineage and the wild group (see Figure 1

for an illustration of the experimental design). Since transplants on healthy Rob recipients

led to massive mortality, we excluded them from this analysis. The recipients from the "wild

group" (because they may be genetically diverse) were cut in half longitudinally one week

before transplantation and allowed to fully regenerate, in order to obtain two genetically

identical individuals on which to perform the control and tumor transplants. The

manipulations were performed under a binocular magnifying glass (Olympus© model SZ51,

magnification from 0.67X to 4.5X) and the tools were disinfected with 90% alcohol and

rinsed with Volvic© before and after each transplant. Donor and recipient hydras were

starved for 24 hours prior to transplantation to prevent digested food remaining in the

gastric cavity from being released during grafting.

Transplantations were realized in a glass Petri dish containing 150 mL of Volvic©.

Using a flat scalpel, the head of the donor hydra was cut and removed from the beaker. Two

cuts were made in the body of the hydra, resulting in a ring-shaped graft (Figure 1). Using

fine forceps, a piece of fishing line (nylon, diameter 0.28 mm) was inserted into the center of

the graft. A small incision was made halfway up the recipient hydra, and the fishing line that

has been previously inserted into the graft was then passed through this incision and exited

through the mouth of the recipient hydra to limit tissue damage. The graft was held to the

recipient by sections of polyethylene tubing (Figure 1. internal diameter 0.30 mm, Sensa©).

The fishing line and the tubes were removed three hours after the graft, and the

experimenter could then check the acceptance of the graft. The hydras whose graft was not

accepted were eliminated and the grafting was repeated [see (Dujon et al. 2022) for a video

of the grafting steps]. Three days later, the protrusion created by the graft was excised using

a scalpel blade, allowing the individual to return to a normal morphology. The period of

three days was assumed to be sufficient time to allow tumor cells to migrate into the tissue

before graft removal (Domazet-Lošo et al. 2014). Freshly transplanted hydras were deprived

of food for a week to avoid water pollution from feeding during regeneration, then returned

to a normal diet on day 8 post-transplant.

Temporal follow-up and statistical analysis

The number of tentacles in donor and recipient individuals was assessed before the

transplantations. The individuals with transplants were then followed weekly for 70 days to

record the number of tentacles, the date of tumor onset, the asexual reproduction through

bud release and the date of eventual death. Detached buds from the grafted individuals

were removed after each feeding and the size of the tumors was qualitatively assessed at

the death of the individual (following the method of Tissot et al. 2023).

Generalized linear mixed-effects models were used to quantify the impact of donor

and recipient characteristics on the different traits observed in grafted individuals. For each

analysis, the most parsimonious model was selected following the method used by Zuur

(Zuur et al. 2009). The use of Akaike's conditional criterion (AICc), therefore, allows us to

exclude some irrelevant effects and to present here only the results of the most relevant

models (see supplementary). All the models, their effects (fixed and random), and their

respective AICc are presented in the appendix. The absence of overdispersion and the

balance of the residuals were also checked using the DHARMa package (Team et al. 2021).

Analyses were performed using R software (version 4.1.3)(R Core Team 2022) and various

packages (see the exhaustive list in Appendix). Because of the unbalanced distribution of

tumor type modalities within the different lines (transmissible tumors in SpB and Rob,

spontaneous tumors in Mt and SpB), we had to separate all analyses into two parts, one

including all lines with transmissible tumors as well as their controls, and one including all

lines with spontaneous tumors and their controls. Random effects of transplantation date

and/or batch were evaluated and included in the models for which they were relevant.

The median number of donor tentacles was compared using Wilcoxon tests between

the tumorous and the control groups. Mortality rates during the experiment were compared

using a random generalized models with a binomial distribution (see the analysis and results

in supplementary section 2). The rate of tumor development was modeled using a random

generalized model with a binomial distribution (see the analysis and results in

supplementary section 3). The different models constructed evaluated the fixed effects of

donor lineage (SpB, Rob, or Mt), donor status (tumor or healthy) and recipient lineage (SpB

or Wild group).

The change in the number of tentacles was then analyzed using a delta variable: the

difference between the maximum number of tentacles developed during the study and the

number of tentacles of the recipient on the day of transplantation (see the analysis and

results in supplementary section 4). This variable was then modeled according to a random

generalized models with a Poisson distribution (we turned all values into positive values by

simply adding 2 to all values). In the models, several fixed effects were tested: tumor status

and lineage of the donor, lineage of the recipient, initial number of tentacles of the recipient

and donor. The impact of tumor development after transplantation on the number of

tentacles was excluded because of a confounding effect between donor status and tumor

development (see in supplementary). The effect of tumor size on the number of

supernumerary tentacles was evaluated using an intra-group linear model (donor lineage,

status, and recipient). This was done to assess how small, medium, and large tumors

influence the variation in tentacle numbers independently of donor and recipient lineage,

for both transmissible and spontaneous tumors (see the analysis and results in

Supplementary Section 4).

The relationship between the number of buds produced and the number of tentacles

developed was subsequently examined on the complete dataset (transmissible and

spontaneous tumors together) utilizing the same delta variable (refer to the analysis and

results in Supplementary Section 5). The number of buds produced during the 10-week

observation period was then subjected to modeling through random linear models. In these

models, various fixed effects were assessed, including the tumor status of the grafted

individuals, the delta variable representing the number of additional tentacles, and the

group consisting of the lineage and cancerous status of both the donor and recipient. The

latter was treated as random effects to emphasize our focus on intra-group variations in

drawing conclusions. Figures and tables were made with the packages GGplot2 (Wickham

2016), Sjplot (Lüdecke 2023), ggstatsplot (Patil 2021) and the software Keynotes©.

Results

1. Tentacle numbers in hydras bearing transmitted versus spontaneous tumors

217   As expected (see Introduction), hydras from both strains harboring transmissible

tumors had an enhanced number of tentacles, with typically more than 4 additional

tentacles in both the Rob (Figure 2A, control, $\tilde{x}$=6[6-6] (median, first and third quartiles);

transmissible tumors $\tilde{x}$=10[9-12], p < 0.001) and the SpB (Figure 2A, control $\tilde{x}$ =6[5.5-6];

transmissible tumors $\tilde{x}$ =10[8-10], p < 0.001) strains. In contrast, SpB hydras bearing

spontaneous tumors did not show an increased number of tentacles compared to healthy

controls (Figure 2A, control $\tilde{x}$=6[5.5-6]; spontaneous tumors$\tilde{x}$=6[6-6.5], p = 0.371). However,

Mt hydras bearing spontaneous tumors showed an increase in the number of tentacles

compared with controls, but of lower magnitude than for Rob and SpB strains with

transmissible tumors, with typically one additional tentacle (Figure 2A, control $\tilde{x}$ =6[5-6];

spontaneous tumors $\tilde{x}$=7[6-7.5], p < 0.001).

2. Tumor development following tissue transplantation

On average, healthy hydras grafted with tumorous tissue, either from the transmissible or

the spontaneous tumors, have an increased risk to subsequently develop tumors compared

to those grafted with healthy tissue (Figure 3 pie charts). Hydras grafted with transmissible

tumors developed on average 8 times more tumors (IRR (incidence rate ratio) = 8.00 [2.73 -

23.49], p<0.001) than those transplanted with healthy tissue, whereas for spontaneous

tumor transplants the risk only increased 2.61 times (IRR = 2.61 [1.09 - 6.23], p=0.031).

Nevertheless, for the transplantation of transmissible tumors from Rob donors on SpB

healthy recipients, the effect appears weaker (see Figure 3.A pie charts), although it is not

detected as significantly different from the other transplant combinations (see in

supplementary section 3). However, an intriguing result is that hydra grafted with healthy

tissues also developed tumors but at lower, yet non negligible, rate compared to hydra

grafted with tumorous tissues (Figure 3 pie charts; 48.75%, 48.12% and 28.92%, on average,

for healthy tissues from Mt, Spb, and Rob, respectively).

3. Development of new tentacles following tissue transplantation

The number of tentacles (Figure 3A, dot plots) showed a significant increase in individuals

transplanted with tissue from transmitted tumors (even if the effect remains low when the

donor is Rob and the recipient is SpB), whereas there was no significant effect in individuals

transplanted with tissues from spontaneous tumors (Figure 3B&C, dot plots), relative to

individuals transplanted with healthy tissue. On average, an individual transplanted with

transmitted tumor tissues developed 1.45 [1.21 - 1.73] additional tentacles while individuals

transplanted with healthy tissue developed only one additional tentacle (IRR= 1.45 [1.21 -

1.73], p<0.001), with no significant effect of the donor lineage. Furthermore, we also

detected a negative effect of the initial number of tentacles of the recipient (Figure 3, dot

plots): when the recipient already had a relatively high number of tentacles, the increase in

the number of tentacles after transplantation was reduced (IRR=0.82 [0.74 - 0.92], p=0.001),

highlighting a possible threshold effect. In contrast, we do not detect any significant effect of

the number of tentacles of the recipient on the number of tentacles developed after the

graft of tissues from spontaneous tumors (see model selection in supplementary section 4).

Considering the effect of tumor size on the number of supplementary tentacles

independently of their donor and recipient lineage, large tumors developed after the graft of

transmissible tumors demonstrated a statistically significant increase in the number of

supernumerary tentacles compared to small tumors (Estimate = 1.40, 95% CI [0.14, 2.65], p =

0.03; see figure 5). For spontaneous tumorous donors, the results indicated no significant

effect of the size of the tumors developed after grafting on the number of tentacles (see

Figure 5, model selection and complete results in supplementary section 4).

4. The reproductive costs and benefits of tumors and supernumerary tentacles

Considering only individuals with a stable number of tentacles (-2, +2), we observed that

tumorous individuals produced significantly fewer buds during the ten weeks of the study

(Wilcoxon test, W = 959, p-value = 0.0094, n=171). Regardless of the type of donor and

recipient, the number of buds produced per individual was found to be significantly

correlated with the number of supernumerary tentacles developed by the individual during

the same period (Figure 4). However, the slope of this correlation was notably reduced for

hydras with tumors (Figure 4, n=92, estimate = -2.75 [-5.21 – 0.29], p= 0.029). Consequently,

all other factors being equal, an individual developing a tumor will require more

supplementary tentacles than a healthy individual to achieve the same number of buds

produced. Noticeably, we observed that the impact of the number of tentacles and tumor

development on the number of buds produced became statistically detectable only after the

7th week of the experiment.

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Discussion

Many parasites from different taxa (e.g., viruses, bacteria, protozoa, helminths, fungi,

arthropods) have evolved the ability to manipulate the phenotype of their host to favor their

own transmission and/or survival (2012). A wide variety of host traits can be modified by

parasites, including reproduction, behavior, color, morphology and/or physiology(Barnett

1983, Dawkins 1999, Agnew et al. 2000). Although theory predicts that transmissible

cancers should evolve a true parasitic lifestyle over time (Dujon et al. 2020, Burioli et al.

2021), to date there has been no evidence of host phenotypic manipulation by transmissible

tumor cell lines (Tissot et al. 2016).

In their pioneering work on transmissible tumors in hydra, Domazet-Lošo and

colleagues (Domazet-Lošo et al. 2014) mentioned the existence of an increased number of

tentacles in the tumor-bearing lineages that have been propagated in the laboratory, but the

precise cause of this phenotypic change was not understood. Here, we first highlight that

this is not a general attribute of tumor-developing hydras, since both Mt and SpB polyps

developing spontaneous neoplasms do not show a similar large increase in the number of

tentacles (Figure 2A). These results support the hypothesis that spontaneous tumors, which

have no evolutionary history with their host, do not possess (or just to a small extent) the

ability to influence its phenotype. Conversely, our results showed that transmissible tumors

propagated in the laboratory for more than 10 years can induce the growth of additional

tentacles even when they are transplanted in new hosts (even if this effect seems low with

SpB grafted with tumors from Rob), as they seem to do in their original host lineage. In

addition, while transplantation of spontaneous tumors never promoted an increase in

tentacle numbers, transmissible tumors did, regardless of the origin of the recipient hydra

(although one may suspect a different pattern between recipient strains that may not be

detected due to a lack of power). These results are not consistent with the hypothesis that

the development of additional tentacles could indicate a compensatory response of tumor-

bearing polyps, for example, to better cope with resources redirected to growing tumors.

Similarly, in agreement with previous work showing that the tumor phenotype can be

observed in tumor-bearing hydras that do not consistently have an altered microbiome [this

is the case for SpB, but not Rob (Domazet-Lošo et al. 2014)], our transplantation

experiments do not support the hypothesis that the alteration in tentacles' number could be

caused entirely by bacteria. Indeed, both the SpB and Rob tumors were able to trigger

supernumerary tentacles in their new hosts, even if Rob microbiome is the same between

tumorous and non-tumorous hydras (Domazet-Lošo et al. 2014). Considering a potential size

effect of the donor, individuals with spontaneous tumors grafted also display an enlarged

body similarly to transmissible ones, as the grafted tissues were of similar size compared to

transmissible tumor grafts. Therefore, this is not likely to explain the effect measured here,

which is also consistent with the literature (Ngo et al. 2021).

Thus, although the exact mechanisms promoting the growth of additional tentacles

remain to be identified, this is the first evidence of transmissible tumor cells that have

evolved the ability to manipulate an external phenotypic trait in their host. Interestingly, we

have shown that individuals with large tumors developed after a graft with transmissible

tumors exhibited more supernumerary tentacles, whereas individuals with large tumors

developed after spontaneous tumor grafts did not show the same pattern. This suggests a

proportional effect depending on the number of transmissible tumor cells developing in the

host. Further experiments focusing on transcriptional and proteomic changes (Takahashi et al.

2005) within grafted individuals, with a quantitative assessment of tumors size, will certainly

improve our knowledge of these phenotypic manipulations.

While it seems clear that the growth of supernumerary tentacles is induced only by

the presence of transmissible tumor cells, to argue for host manipulation in the strict sense

[as observed in the case of manipulative parasites, see (Ewald 1980, Poulin 2010, 2012)], it is

necessary to demonstrate that this phenotypic change is indeed associated with an increase

in tumor transmission. One possibility is that the supernumerary growth of tentacles, by

increasing the rate of prey capture, merely compensates for the pathological costs

associated with the presence of the tumor, the growth of which requires additional

resources. Under this hypothesis, tumorous hydra with supernumerary tentacles would not

necessarily exhibit an increased budding rate, as it would be more indicative of a host

manipulation aimed at better tolerating the tumor [i.e., compensating for vital resources

diverted toward the tumor (Thomas et al. 2020)]. It is also possible than the growth of

supernumerary tentacles, through a higher feeding rate, enhances the host's budding rate

and, consequently, the vertical transmission of the tumor. Our results do support the

hypothesis that tumors do indeed impose a cost on the reproductive potential of the host in

terms of the weaker budding of tumoral individuals, and this cost can be offset by the

growth of supernumerary tentacles (see Figure 4). Beyond this first conclusion, our data also

support the hypothesis that the supernumerary growth of tentacles can sometimes not only

compensate for the cost of the tumor, but also leads for some cancerous individuals to a

significant increase in the number of buds, beyond the values presented by healthy

individuals with a normal number of tentacles (c.f., overcompensation area in Figure 4).

Given that the transmission rate of tumors remains constant regardless of the budding rate

(Boutry et al. 2022b), all our results thus suggest that tumor-induced supernumerary

tentacle growth has likely evolved as a tumoral adaptation that reduces the cost for the host

in order to maintain and enhance tumor transmission. Further research should focus on

underlying the bio-physiological pathways and energetical costs of this manipulated

phenotype (Takahashi et al. 2005, Sebens et al. 2017).

An intriguing finding from the present study is that in *H. oligactis*, new tumors can

develop following injury and/or transplantation of healthy tissue alone. Additional

experiments are required to formally describe these outgrowths as tumors [although there

are strong phenotypic similarities with spontaneous tumors; see (Domazet-Lošo et al. 2014)]

and to understand the cause and mechanisms underlying their development. We can

speculate that the act of grafting is likely to weaken the hydra's ability to control abnormal

cell proliferation and/or make them more susceptible to dysbiosis known to promote

tumorigenesis (Rathje et al. 2020). In any case, despite the appearance of spontaneous

tumors even after transplantation of healthy tissue, it is striking that only transmissible

tumors are able to consistently alter and increase the number of tentacles in the

transplanted individuals, which ultimately reinforces our conclusions.

An alternative explanation for our results would be that the increase in tentacle

number following transplantation is a response of the hydra when non-self-tissues are

introduced into the polyps. This would imply that the tumors, owing to their own evolution

over years of vertical transmission, have diverged so significantly that they are perceived as

non-self by the recipient hydras. However, we do not favor this hypothesis since when *Hydra*

vulgaris are grafted with tissue from *Hydra oligactis*, we simply observe a destruction of the

non-self-tissue (Bosch and David 1986). On the opposite, there is evidence of a lack of intra-

specific non-self-recognition in *H. vulgaris* (Kuznetsov and Bosch 2003), without any

reporting of growth of additional tentacles. Also, since healthy and spontaneously tumorous

hydras have been shown to sometimes grow a few additional tentacles, it would necessary

to explore in the future the extent to which the responsible environmental factors are

similar to those potentially produced/exploited by transmissible tumors [see, for instance

(Lefèvre et al. 2008)]. Further work, including experimental evolution, would be needed to

assess the extent to which tentacle growth promotion is a capability that could be acquired

over time from spontaneous tumor cells, at least for those that also acquired the capacity to

be transmitted.

Transmissible tumors in hydras have significant implications for their ecological (Boutry et al.

2022a) and reproductive dynamics (Boutry et al. 2022b). For instance, these tumors have

been associated with changes in life history traits, notably increased early budding rates and

reduced sexual reproduction. Under experimental conditions, transmissible tumors have

also been shown to influence biotic interactions with other species (Boutry et al. 2021).

While our study focused on the modification of tentacle number, it is also promising to

investigate behavioral traits and the life cycle of grafted individuals in future research. This

would provide a more comprehensive understanding of the phenotypic changes in hydras

bearing transmissible tumors.

Supplementary material: This repository contains complete dataset, details of the

performed analysis and supplementary analysis at

https://justineboutry.github.io/Supplementary.io/

Funding: This work was supported by the Hoffmann Family and by the ANR EVOSEXCAN

(ANR-23-CE13-0007) to FT. JB salary was funded by the university of Montpellier.

Acknowledgment: We are grateful to Thomas Bosch's laboratory, and in particular to

Alexander Klimovich, for providing the first hydras, methods and ideas on which the authors

first met and built their research. We would like to thank the two referees who reviewed the

previous version of this manuscript. Their comments have significantly improved the

manuscript.

Author contributions: Conceptualization: JB, FT; Methodology and data collection: JB, OR,

LG, JM, ST; Analysis and visualization: JB, OR, LG; Supervision: JB and FT; Writing of the

original draft: JB, FT, OR, Review & editing: JB, JT, AMN, ST, NS, BU, RH, AD

Competing interests: All other authors declare they have no competing interests.

Figures

Figure 1. Grafting protocol. Images represent each step of the transplantation process,

starting with the recipient polyp (left); the donor is then cut into a ring-shaped tissue graft

that is inserted on a needle, put in contact with the recipient tissue and blocked by stopping

tubes. At the end (right) a transplanted individual can be seen 3 days after the transplant.

Figure 2. Donor and recipient tentacle phenotype. (A) Boxplots represent the number of tentacles in each donor line; the average number of tentacles in each group is written above in the boxes, and the rows represent the comparison between the control and the tumor medians in each line, with the p-values of the corresponding Wilcoxon-Mann-Whitney tests. (B) Boxplots represent the number of tentacles in different groups of recipients.

Figure 3. Tumor appearance and changes in tentacle number after the grafts.

Three sets of matrices showing (i) the proportion of individuals (pie charts) that have

developed tumors (in red) or lacked tumors (in green) and (ii) the variation in tentacle

numbers (graphs) among non-tumorous individuals (green dots) and tumorous individuals

(red dots), following the grafting of healthy tissue (green bars) or tumorous tissue (from

spontaneous – orange bars, and transmitted – red bars, tumors) from Robusta, St.

Petersburg or Montaud donor lineages (left panel) onto healthy recipients from the St.

Petersburg or wild groups (top panels). For the donor lineages, the arrows indicate that the

individuals with spontaneous tumors were isolated from the same cultures as the healthy

individuals, whereas the individuals with transmitted tumors represent a distinct lineage; the

median number of supernumerary tentacles in the tumorous donor individuals are also

shown (in red circles).

Figure 4. Number of buds produced depending on the development of supernumerary tentacles. On the left, the box plot exhibits the bud production of healthy (n=47, green) and tumorous individuals (n=28, red) with a constant number of tentacles (-2, +2) during the 10-week monitoring post-grafting (Wilcoxon test, W=959, p=0.009, $n_{healthy}$=79, $n_{tumorous}$ = 92). The arrow signifies the reduction in bud production attributed to tumor costs. On the right, each point represents bud production for an individual (healthy in green, n=83, and tumorous in red, n=116) based on the number of tentacles developed in the 10 weeks following the transplantation. The orange zone represents tentacle-induced compensation for tumor costs compared to healthy individuals with a stable number of tentacles, while the blue zone indicates overcompensation beyond expected values for a healthy individual. The interaction between tentacle number and tumor occurrence significantly explains bud production variation (Random linear mode, estimate = -2.75, CI: -5.21 — -0.29, p=0.029).

Figure 5. Effect of tumor size on the number of supernumerary tentacles developed after grafting of transmissible tumors (left panel, n=53) versus spontaneous tumors (right panel, n=39). For transmissible tumors, there was a significant increase in supernumerary tentacles with large tumors (linear model, estimate=1.40 [0.14-2.65], p=0.03) compared to small ones, while medium tumors show no significant difference (N.S). In contrast, spontaneous tumors do not show a significant effect on the number of supernumerary tentacles across all tumor sizes (N.S).

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