Biallelic pathogenic variants in *DNAH3* cause male infertility in humans and mice

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Abstract

Axonemal protein complexes, including the outer and inner dynein arms (ODA/IDA), are highly ordered structures of the sperm flagella that drive sperm motility. Deficiencies in several axonemal proteins have been associated with male infertility, which is characterized by asthenozoospermia or asthenoteratozoospermia. Dynein axonemal heavy chain 3 (DNAH3) resides in the IDA and is highly expressed in the testis. However, the relationship between DNAH3 and male infertility is still unclear. Herein, we identified biallelic variants of *DNAH3* in four unrelated Han Chinese infertile men with asthenoteratozoospermia through whole-exome sequencing (WES). These variants contributed to deficient DNAH3 expression in the patients' sperm flagella. Importantly, the patients represented the anomalous sperm flagellar morphology, and the flagellar ultrastructure was severely disrupted. Intriguingly, *Dnah3* knockout (KO) male mice were also infertile, especially showing the severe reduction in sperm movement with the abnormal IDA and mitochondrion structure. Mechanically, nonfunctional DNAH3 expression resulted in decreased expression of IDA-associated proteins in the spermatozoa flagella of patients and KO mice, including DNAH1, DNAH6, and DNALI1, the deletion of which has been involved in disruption of sperm motility. Moreover, the infertility of patients with *DNAH3* variants and *Dnah3* KO mice could be rescued by intracytoplasmic sperm injection (ICSI) treatment. Our findings indicated that *DNAH3* is a novel pathogenic gene for asthenoteratozoospermia and may further contribute to the diagnosis, genetic counseling, and prognosis of male infertility.

Key words: DNAH3, Male infertility, IDA-associated proteins, asthenoteratozoospermia, WES.

Introduction

Infertility is a global public health and social problem that affects approximately one in six couples worldwide. (1) Male infertility, which accounts for half of infertile cases, is a multifactorial disease with common phenotypes, including oligo/azoospermia (poor sperm count or absence of spermatozoa); teratozoospermia (aberrant sperm morphology); asthenozoospermia (weakened sperm motility); and a combination of these phenotypes, such as asthenoteratozoospermia, oligoasthenozoospermia, oligoteratozoospermia and oligoasthenoteratozoospermia. (2, 3)

Asthenoteratozoospermia is one of the most common phenotypes of male infertility, and genetic factors have been established as the predominant cause of asthenoteratozoospermia. Multiple morphological abnormalities of the flagella (MMAF), a subtype of asthenoteratozoospermia, characterized by a mosaic of abnormalities of the flagellar morphology, including absent, short, coiled, bent and/or irregular flagella, is almost always caused by genetic defects. (4, 5) To date, more than 40 genes have been identified as pathogenic genes of MMAF, but these genes can only explain approximately 60% of MMAF-affected cases. (6-9) Therefore, the genetic basis of the remaining cases is still unknown.

The motility of a sperm is driven by its rhythmically beating flagella, and at the center of the flagella lies a conserved axonemal structure, containing the “9 + 2” microtubular arrangement: a ring of nine microtubule doublets (MTDs) surrounding a central pair (CP) of singlet microtubules. Each MTD consists of an A tubule and a B tubule, and the outer (ODA)

and inner (IDA) dynein arms are anchored along the A tubule. (10) The ODA and IDA are ATPase-based protein complexes that drive the movement between the A tubule and the neighboring B tubule of the next doublet, producing the original force for sperm motility. (11, 12) Structural and functional abnormalities of the ODA and IDA have been demonstrated to cause male infertility associated with asthenozoospermia and/or asthenoteratozoospermia. (4, 13, 14)

The dynein axonemal heavy chain (DNAH) family comprises a series of proteins (DNAH1–3, DNAH5–12, and DNAH17) that are precisely assembled with other axonemal dynein motor proteins in the ODAs and IDAs of sperm flagella and motile cilia. (15-17) In humans, DNAH1, DNAH2, DNAH6, DNAH7, DNAH8, DNAH10, DNAH12 and DNAH17 are highly expressed in the testis, and deficiency of these proteins has been demonstrated to cause MMAF-associated asthenoteratozoospermia. (18-25) DNAH3 is an evolutionarily conserved IDA-associated protein and is highly expressed in testes of humans and mice. Deficient DNAH3 has been shown to impair sperm motility in *Drosophila* and cattle. (26, 27) In humans, *DNAH3* has been identified as a novel breast cancer candidate gene. (28) However, the role of DNAH3 in male reproduction in humans and mice remains largely unknown.

In the present study, we identified four biallelic variations in *DNAH3* in four unrelated Han Chinese patients with asthenoteratozoospermia using whole-exome sequencing (WES). The spermatozoa of the patients showed extremely reduced sperm motility and a high proportion of sperm tail defects characterized by the MMAF phenotype. We further generated *Dnah3* knockout (KO) mice, and the male KO mice expectedly showed

aberrations in sperm movement, flagellar IDA, and mitochondrion. Moreover, the absence of DNAH3 led to decreased expression of other IDA-associated proteins, including DNAH1, DNAH6 and DNALI1. Importantly, good outcomes of intracytoplasmic sperm injection (ICSI) treatment were observed in *DNAH3*-deficient patients and *Dnah3* KO mice. This study revealed *DNAH3* as a novel pathogenic gene of asthenoteratozoospermia, and the findings provide valuable suggestions for the clinical diagnosis and treatment of male infertility.

Results

Identification of biallelic pathogenic variants of *DNAH3* in four unrelated infertile men

In the present study, we conducted whole-exome sequencing (WES) to identify potential candidate genes associated with primary asthenoteratozoospermia. Interestingly, after filtering, four biallelic variations in *DNAH3* (NG_052617.1, NM_017539.2, NP_060009.1) were identified in four unrelated patients: a compound heterozygous mutation of c.3590C>T (p.P1197L) and c.3590C>G (p.P1197R) in Patient 1, a homozygous missense mutation of c.4837G>T (p.A1613S) in Patient 2, a compound heterozygous mutation of c.5587del (p.L1863*) and c.10355C>T (p.S3452L) in Patient 3 and a compound heterozygous mutation of c.2314C>T (p.R772W) and c.4045G>A (p.D1349N) in Patient 4 (**Figure 1A**). Importantly, routine semen analysis revealed that all patients showed extremely reduced sperm motility and a high proportion of sperm tail defects (**Table 1**). These variants either were not recorded or had an extremely low frequency in multiple public population databases and were predicted to be potentially deleterious by SIFT, PolyPhen-2 and

Mutation Tester (**Table 2**). Next, Sanger sequencing confirmed these variants in the probands, and their fertile parents carried the heterozygous variants (**Figure 1A**). Moreover, the variant sites are localized in several domains of the DNAH3 protein and are highly conserved across species (**Figure 1B**).

Strikingly, immunofluorescence staining revealed that DNAH3 was exclusively resided in the tail and concentrated in the midpiece of control sperm. However, the fluorescence signal of DNAH3 was hardly detected in the patients' spermatozoa (**Figure 1C**). Additionally, subsequent western blotting analysis yielded consistent results with immunofluorescence staining, indicating that these variants led to disrupted expression of DNAH3 (**Figure 1D**). These results suggested that biallelic variants in *DNAH3* disrupted DNAH3 expression and might be responsible for the infertility of the four patients.

Asthenoteratozoospermia phenotype is observed in patients with *DNAH3* variants

We next investigated the aberrant sperm morphology of the patients using Papanicolaou staining and SEM analysis. Notably, the tails of sperm from the patients exhibited a typical phenotype associated with MMAF, including coiled, short, bent, irregular, and/or absent flagella (**Figure 2A and B**). In addition, a fraction of defects in the sperm head were also present in the patients' sperm (**Figure 2A and B**).

TEM was employed to determine the ultrastructure of the sperm from the patients. Compared to the integrated and well-organized "9 + 2" axonemal arrangement of the sperm flagella from the normal control, spermatozoa from the patients showed absent or disordered CPs, MTDs, and outer dense fibers (ODFs) in different regions of the flagella (**Figure 3A**).

Interestingly, the IDAs of sperm flagella of the patients were hardly captured compared to the control (**Figure 3A**). Additionally, in the midpiece of sperm flagella of the patients, dissolved mitochondrial material was also observed evidently under TEM (**Figure 3A**). We next conducted immunofluorescence staining to label the mitochondria of patients' sperm with TOM20, a subunit of the mitochondrial import receptor. Remarkably, in contrast to the robust TOM20 signals observed in the normal control, the TOM20 signals in the sperm from the patients were considerably diminished, indicating a disrupted mitochondrial function (**Figure 3B**). Together, these data suggested that *DNAH3* may function in sperm flagellar development, and loss-of-function variants were associated with MMAF in humans.

DNAH3 is exclusively expressed in the sperm flagella of humans and mice

To further understand the function of DNAH3 in male reproduction, we explored the expression pattern of DNAH3 in humans and mice. qPCR results revealed that *Dnah3* was predominantly expressed in the mouse testis (**Figure S1A**). Moreover, when observing the expression of *Dnah3* in testes from mice at different postnatal days, we found that *Dnah3* expression was significantly elevated beginning on postnatal Day 22, peaked at postnatal Day 30, and maintained a stable expression level thereafter (**Figure S1B**). In addition, germ cells at different stages were isolated from the testes of humans and mice and were stained with anti-DNAH3 antibody. The results showed that DNAH3 was expressed in the cytoplasm of spermatocytes and spermatogonia and then obviously in the flagellum of early and late spermatids (**Figure S2A and B**). These expression data suggest that DNAH3 may play an important role in sperm flagellar development during spermatogenesis in humans

and mice.

Deletion of *Dnah3* causes male infertility in mice

Considering the absent expression of DNAH3 in the patient sperm, we generated *Dnah3* KO mice using CRISPR–Cas9 technology to further confirm the essential role of DNAH3 in spermatogenesis (**Figure S3A**). PCR, qPCR, and immunofluorescence staining were used to confirm that *Dnah3* was null in KO mice (**Figure S3B-E**). The *Dnah3* KO mice survived without any evident abnormalities in development and behavior. H&E staining further revealed that there were no histological differences in the lung, brain, eye, or oviduct between wild-type (WT) and *Dnah3* KO mice (**Figure S4A**). In addition, no obvious abnormalities in ciliary development were observed in these organs in KO mice compared to WT mice (**Figure S4B**). The *Dnah3* KO female mice were fertile with normal oocyte development (**Figure S5A**). However, the *Dnah3* KO male mice were completely infertile (**Figure 4A**). We next examined the testis and epididymis of *Dnah3* KO male mice to elucidate the etiology of infertility. There was no detectable difference in the testis/body weight ratio of *Dnah3* KO mice when compared to WT mice (**Figure S5B**).

Moreover, subsequent computer-assisted sperm analysis (CASA) also showed that sperm isolated from the cauda epididymis were slightly decreased, and nearly all sperm were completely immobile (**Table 3, Movie S1 and Movie S2**). Papanicolaou staining and SEM analysis revealed morphological defects in partial spermatozoa from *Dnah3* KO mice, including coiled, bent, and irregular flagella, as well as aberrant heads and acephalic spermatozoa (**Figure S6A and B**).

TEM was further utilized to evaluate the sperm flagellar ultrastructure of *Dnah3* KO mice. There were no obvious abnormalities of “9 + 2” microtube arrangement in most sperm from the *Dnah3* KO mice when compared to WT mice (**Figure 4B**). However, in contrast to the clear display of an IDA and an ODA on the A-tube of each microtubule doublet in the sperm flagella of WT mice, the sperm flagella of *Dnah3* KO mice exhibited an absence of almost all the IDAs (**Figure 4B**). In addition, the disrupted mitochondria of spermatozoa from *Dnah3* KO mice were also observed under TEM, as manifested by the dilated intermembrane spaces and dissolved mitochondrial material (**Figure 4B and C**). We next performed immunofluorescence staining to label SLC25A4, which is responsible for the exchange of ATP and ADP across the mitochondrial inner membrane. Strikingly, compared to the bright fluorescence signals in the midpiece of WT sperm, the signals *Dnah3* KO were significantly diminished (**Figure 4D**), indicating impaired mitochondrial function. Collectively, DNAH3 is essential for spermatogenesis, and its deficiency seriously damages the sperm motility and IDAs in both humans and mice.

DNAH3 deficiency impairs IDAs related to the reduction of IDA-associated proteins

Considering the disrupted IDAs revealed by TEM analysis in both our patients and *Dnah3* KO mice, we speculated whether the defective IDAs were attributed to the decreased expression of the key IDA-associated proteins. The immunofluorescence data showed that DNAH1/DNAH6 and DNALI1, corresponding to the heavy and light intermediate chains of the IDAs, (16) respectively, were almost invisible along the sperm flagella of the patients when compared to control (**Figure 5A-C**). Consistent results were obtained in our

subsequent western blotting analysis of sperm lysates from the patients (**Figure 5D-F**), indicating that DNAH3 may manipulate the assembly of IDA through regulating the expression of IDA-associated proteins. In contrast, DNAH8/DNAH17 and DNAI1, corresponding to the heavy and intermediate chains of ODAs, (25) were readily detectable in the patients' sperm flagella and were comparable to the control (**Figure S7A-C**), suggesting that DNAH3 may not regulate the expression of ODA-associated proteins. We also performed immunofluorescence staining and western blotting analysis of DNAH1, DNAH6, DNALI1, DNAH8, DNAH17 and DNAI1 on sperm from *Dnah3* KO mice, and the results observed were consistent with those of the patients (**Figure 6A-F, Figure S8A-C**). These findings suggested that other IDA-associated proteins might be downstream effectors of DNAH3, which needs more future research.

ICSI treatment of humans with *DNAH3* variants and *Dnah3* KO mice

ICSI treatment has been reported to be effective in asthenoteratozoospermia-associated infertility. (29, 30) ICSI cycles were attempted for the patients after written informed consent was obtained. The female partners all had normal basal hormone levels and underwent a long gonadotrophin-releasing hormone agonist protocol (**Table 4**). The wife of Patient 1 underwent one ICSI attempt. A total of 21 metaphase II (MII) oocytes were retrieved and microinjected, of which 17 oocytes were successfully fertilized (17/21, 80.95%) and cleaved (17/17, 100%). Thirteen Day 3 (D3) embryos were formed, six of which developed into blastocysts (8/13, 61.54%) after standard embryo culture. Two blastocysts were transferred, one of which was implanted. She eventually achieved clinical

pregnancy, and the pregnancy is ongoing (**Table 4**). The partner of Patient 2 underwent two ICSI attempts. In her first ICSI attempt, six MII oocytes were retrieved, of which three were fertilized (3/6, 50%) and cleaved (3/3, 100%). After standard embryo culture, two D3 embryos were formed and transferred. However, this ICSI failed because no embryos were implanted. In her second ICSI attempt, all five MII oocytes were fertilized and cleaved (5/5, 100%). Five D3 embryos were obtained, of which two were transferred, but no embryos were implanted. The remaining three D3 embryos were cultured continuously, and two available blastocysts were formed and kept to be transferred in the future (**Table 4**). The partner of Patient 3 underwent one ICSI attempt. Of the 20 MII oocytes retrieved, 19 oocytes were fertilized (19/20, 95.0%) and cleaved (19/19, 100%). Fifteen D3 embryos were obtained, and 10 developed into available blastocysts (10/15, 66.7%). One blastocyst was transferred and implanted. She achieved clinical pregnancy, and the pregnancy is ongoing (**Table 4**). The wife of Patient 4 underwent four failed ICSI attempts. In her first two ICSI attempts, 13 and 12 MII oocytes were retrieved, of which five (5/12, 41.67%) and six (6/13, 46.15%), respectively, were fertilized and cleaved. Two available D3 embryos were obtained and transferred in both ICSI attempts, but no embryos were implanted. In her third ICSI attempt, of the eight MII oocytes retrieved, four (4/8, 50%) were fertilized and cleaved (4/4, 100%). However, no available D3 embryos were acquired. In her last ICSI attempt, seven MII oocytes were retrieved, of which three were fertilized (3/7, 42.68%) and two were cleaved (2/3, 66.7%), but no available D3 embryos were formed (**Table 4**). The vivid embryonic development of Patient 1 and Patient 3 after ICSI treatment was shown in **Figure 7A**.

We also carried out ICSI treatment on *Dnah3* KO male mice. Strikingly, favorable outcomes of ICSI were obtained in *Dnah3* KO male mice. After injection of spermatozoa from *Dnah3* KO male mice, pronuclei were observed in most embryos in both the KO and WT groups, indicating a normal fertilization rate (**Figure 7B**). There was no difference in the percentage of 2-cell and blastocyst-stage embryos between the KO and WT groups (**Figure 7B**). Collectively, we observed successful ICSI outcomes in two out of four DNAH3-deficient patients and *Dnah3* KO male mice and therefore suggested ICSI as an optional treatment for infertile men harboring biallelic pathogenic variants in *DNAH3*, and the additional female risk factors for infertility should not be excluded in the failed patients.

Discussion

In the present study, we identified pathogenic variants in *DNAH3* in unrelated infertile men with asthenoteratozoospermia. These variations resulted in the almost absence of DNAH3 and sharply decreased the expression of other IDA-associated proteins, including DNAH1, DNAH6 and DNALI1. Combined with similar findings in *Dnah3* KO mice, we demonstrated that DNAH3 is fundamental for male fertility. Moreover, we suggest that ICSI might be a favorable treatment for male infertility caused by DNAH3 deficiency. Our findings identify a function for DNAH3 in male reproduction in humans and mice and may provide a new view on the clinical practice of male infertility.

Primary ciliary dyskinesia (PCD, MIM: 244400) is a genetic disorder affecting at least one in 7554 individuals. (31) The most common symptoms of PCD are recurrent infections in airways due to malfunction of the motile cilia that are responsible for mucus clearance.

(32) It has been suggested that male infertility associated with sperm defects is highly prevalent (up to 75%) among individuals with PCD. (33) Axonemal defects caused by variants within *DNAH* family members, including *DNAH5*, *DNAH6*, *DNAH7*, *DNAH9* and *DNAH11*, are causative factors for PCD. (34-36) Moreover, deficiency in these PCD-causing *DNAH*s has also been associated with male infertility. (9, 14, 20, 21, 37-39) Additionally, other *DNAH*s, such as *DNAH1*, *DNAH2*, *DNAH8*, *DNAH10*, *DNAH12* and *DNAH17*, are suggested to be pathogenic genes of isolated male infertility. (18, 19, 22, 24, 25) These phenotype–genotype correlations may be attributed to the fact that ciliary and flagellar axonemes have cell type-specific or cell type-enriched DNAHs. (40) DNAH3 resides in the IDA and is expressed in testis and ciliary tissues, including the lung, brain, eye, and oviduct. However, despite its presence in these tissues, the relationship between deficient DNAH3 and disease is unclear to date. Intriguingly, in our study, we observed male infertility associated with abolished sperm movement and IDA structure in both patients carrying biallelic *DNAH3* mutations and *Dnah3* KO mice. However, none of the patients reported suffering from any of the principal PCD symptoms, and the *Dnah3* KO mice showed normal ciliary development in the lung, brain, eye, and oviduct. These findings suggested that DNAH3 may play a more important role in sperm flagellar development than in other motile cilia functions. Given that DNAH3 is expressed in ciliary tissues, its role in these tissues remains intriguing and could be elucidated through sequencing of larger cohorts of individuals with PCD.

In our study, both *Dnah3* KO male mice and *DNAH3*-mutant men are infertile and showed the immotile sperm with disrupted flagellar IDAs and mitochondria. However, there

are still some differences between their phenotypes. The *Dnah3* KO mice displayed moderate sperm morphological defects, and the “9 + 2” microtube arrangement showed no obvious abnormalities. While our patient exhibited a high proportion of sperm tail defects, including coiled, short, bent, and irregular flagella, along with seriously defective “9 + 2” microtube arrangement. Importantly, previous studies have suggested that the phenotypes resulting from depletions of *Dnah*s in mice do not always resemble deficiencies in the human orthologs. Specifically, *Dnah1*$^{-/-}$ mice was infertile due to decreased sperm motility, without any observable structural defects of the axoneme. (41) In contrast, spermatozoa from *DNAH1*-deficient men exhibited significant axonemal disorganization and MMAF phenotype. (18) Combined with our findings, it is conceivable that DNAH3 is crucial for human and mouse male reproduction, but may play distinct roles during flagellar axonemal assembly.

ICSI has been an efficient treatment for male infertility. (42, 43) However, the outcomes of ICSI for male infertility caused by variants in different *DNAH* genes are variable. It has been demonstrated that infertile males with variants in *DNAH1*, *DNAH2*, *DNAH7*, and *DNAH8* have a favorable prognosis, (22, 44-48) while patients with variants in *DNAH17* have poor outcomes after ICSI treatment. (25, 49) Meanwhile, the ICSI outcomes in male infertility caused by *DNAH6* variants may depend on the specific mutation or be controversial. (20, 50, 51) The patients with *DNAH3* mutations in our study experienced different clinical outcomes of ICSI treatment. The partners of Patient 1 and Patient 3 achieved clinical pregnancy. The wives of Patient 2 and Patient 4 obtained favorable fertilization and cleavage rates but experienced no clinical pregnancy due to the

nonimplantation of the transferred embryos. Remarkably, despite the diverse variants within *DNAH3* observed in the four patients, all variants led to a complete absence of DNAH3 expression. Therefore, these different ICSI outcomes might be attributed to additional unexplained factors from the female partners. Considering the successful ICSI outcomes observed in *Dnah3* KO mice, we suggested ICSI as an optimized treatment for infertile men carrying variants in *DNAH3*. More cases are needed to precisely estimate the prevalence of *DNAH3* mutations and determine a prognosis for ICSI treatments.

In conclusion, our study revealed an unexplored role of DNAH3 in male reproduction in humans and mice, suggesting *DNAH3* as a novel causative gene for human asthenoteratozoospermia. Moreover, ICSI is as an optimized treatment for infertile men with *DNAH3* variants. This study expands our knowledge of the relationship between DNAH proteins and disease, facilitating genetic counseling and clinical treatment of male infertility in the future.

Methods

Human subjects

Four unrelated Han Chinese infertile men and their family members were recruited from West China Second University Hospital of Sichuan University and Women and Children's Hospital of Chongqing Medical University. All patients exhibited a normal karyotype (46 XY) without deletion of the azoospermia factor (AZF) region in the Y-chromosome. All of the participants were provided informed consent, and the study was approved by the ethics committee of West China Second University Hospital and The First Affiliated Hospital of

Chongqing Medical University.

Genetic analysis

Peripheral blood samples were obtained from the subjects to extract genomic DNA using a DNA purification kit (TIANGEN, DP304). For WES, 1 μg of genomic DNA was utilized for exon capture using the Agilent SureSelect Human All Exon V6 Kit and sequenced on the Illumina HiSeq X system (150-bp read length). Mutations were annotated and filtered using the ExAC browser, dbSNP, 1000 Genomes Project and HGMD. Then, PolyPhen-2, SIFT, MutationTaster, and CADD were utilized for functional prediction. The candidate pathogenic variants were verified by Sanger sequencing on DNA from the patients' families. The primer pairs used for PCR amplification are listed in **Table S1**.

Electron microscopy

For scanning electron microscopy (SEM), sperm samples were fixed in glutaraldehyde (2.5%, w/v) and dehydrated using an ethanol gradient (30, 50, 75, 85, 95, and 100% ethanol). The samples were dried using a CO_2 critical-point dryer (Eiko HCP-2, Hitachi) and observed under SEM (S-3400, Hitachi).

For transmission electron microscopy (TEM), sperm samples were fixed in glutaraldehyde (3%, w/v) and osmium tetroxide (1%, w/v) and dehydrated with an ethanol gradient. The samples were embedded in Epon 812. Ultrathin sections were stained with uranyl acetate and lead citrate and analyzed under TEM (Tecnai G2 F20).

STA-PUT velocity sedimentation

Single testicular cells from obstructive azoospermia and 8-week-old C57BL male mice were obtained using the STA-PUT velocity sedimentation method as described previously. In brief, total spermatogenic cells were harvested by digesting seminiferous tubules with collagenase (Invitrogen, 17100017), trypsin (Sigma, T4799) and DNase (Promega, M6101) for 15 min each at 37 °C. Cells were diluted in bovine serum albumin (BSA, 3%, w/v) and filtered through an 80 mm mesh to remove fragments. Then, the cells were resuspended in BSA (3%, w/v) and loaded in an STA-PUT velocity sedimentation cell separator (ProScience) to obtain germ cells at different stages.

RNA isolation and quantitative PCR (qPCR)

Total RNA of mouse tissues was extracted using TRIzol reagent (Invitrogen,15596026,) and reverse-transcribed using the 1st Strand cDNA Synthesis Kit (Yeasen, HB210629) according to the manufacturer's instructions. qPCR was carried out on an iCycler RT–PCR Detection System (Bio-Rad Laboratories) using SYBR Green qPCR Master Mix (Bimake, B21202). Primer sequences are listed in **Table S1**.

Immunofluorescence staining

Sperm samples were fixed in paraformaldehyde (4%, w/v), permeabilized with Triton X-100 (0.3% v/v) and blocked with BSA (3%, w/v) at room temperature. Samples were incubated with primary antibodies, including DNAH1 (Cusabio, CSB-PA878961LA01HU, 1:100), DNAH3 (Cusabio, CSB-PA823461LA01HU, 1:100), DNAH6 (Proteintech, 18080-1-AP,

RRID: AB_2878493, 1:50), DNAH8 (Atlas, HPA028447, RRID: AB_10599600, 1:200), DNAH17 (Proteintech, 24488-1-AP, RRID: AB_2879568, 1:50), DNAI1 (Proteintech, 12756-1-AP, RRID: AB_10643244, 1:50), DNALI1 (Proteintech, 17601-1-AP, RRID: AB_2095372, 1:50), TOM20 (Proteintech, 11802-1-AP, RRID: AB_2207530, 1:50), SLC25A4 (Signalway, 32484, RRID: AB_2941094, 1:100), lectin PNA (Invitrogen, L-32460, 1:50) and alpha tubulin (Abcam, ab7291, RRID: AB_2241126, 1:500), overnight at 4 °C. The next day, the samples were washed and incubated with the secondary antibody Alexa Fluor 488 (Invitrogen, A11008, RRID: AB_143165, 1:1000) or Alexa Fluor 594 (Invitrogen, A11005, RRID: AB_141372, 1:1000), and the nuclei were labeled with 4′,6-diamidino-2-phenylindole (DAPI, Sigma–Aldrich, D9542). Image capture was performed by a laser scanning confocal microscope (Olympus, FV3000).

For staining of mouse tissues, samples were first fixed in paraformaldehyde (4%, w/v) and dehydrated with an ethanol gradient. Then, the samples were embedded in paraffin and sliced into 5-μm sections. After deparaffinization and rehydration, sections were processed with 3% hydrogen peroxide and incubated in sodium citrate for antigen repair. Subsequently, sections were blocked with goat serum and incubated with primary antibodies against DNAH3 (Cusabio, CSB-PA823461LA01HU, 1:100) or ac-Tubulin (Abcam, ab24610, RRID: AB_448182, 1:500) at 4 °C overnight. The next day, the sections were incubated with the secondary antibody Alexa Fluor 488, followed by labeling the nuclei with DAPI. Image capture was performed using a fluorescence microscope (Zeiss, Ax10).

Western Blotting

Sperm samples were lysed in RIPA buffer (Beyotime, P0013B) to extract the total protein. For analysis of DNALI1, the protein samples were mixed with SDS loading buffer (P0015, Beyotime, China), boiled at 95 °C for 5 minutes, and separated by 12.5% SDS-PAGE. For analysis of DNAH1, DNAH3, DNAH6, the protein samples were mixed with NuPAGE™ LDS sample buffer (Invitrogen, NP0007), denatured at 70°C for 10 minutes, and separated by 3–8% NuPAGE™ Tris-Acetate gels (EA0375BOX, Invitrogen). Then the resolved proteins were transferred to 0.45 μm PVDF membranes (Merck Millipore, IPVH00010). The membranes were blocked, incubated with primary antibodies, including DNALI1 (Proteintech, 17601-1-AP, RRID: AB_2095372, 1:150), DNAH3 (Cusabio, CSB-PA823461LA01HU, 1:200), DNAH6 (Proteintech, 18080-1-AP, RRID: AB_2878493, 1:150) and alpha tubulin (Proteintech, 11224-1-AP, RRID: AB_2210206, 1:1000) at 4 ℃ overnight. The following day, membranes were washed and incubated with HRP-conjugated secondary antibody (Proteintech, SA00001-2, RRID: AB_2722564, 1:5000). Protein bands were visualized using enhanced chemiluminescence reagents (Millipore, WBKLS0500).

Histology hematoxylin-eosin (H&E) staining

Tissue samples from mice were fixed with 4% paraformaldehyde (w/v) overnight. Following dehydration by ethanol, the samples were embedded in paraffin and sliced into 5-μm sections. The sections were stained with hematoxylin and eosin and observed under a microscope (Zeiss, Axio Imager 2).

Generation of the *Dnah3* KO mouse model

Animal experiments in this study were approved by the Experimental Animal Management and Ethics Committee of West China Second University Hospital, Sichuan University, and complied with the Animal Care and Use Committee of Sichuan University. A *Dnah3* knockout mouse model was generated by the CRISPR–Cas9 system. Briefly, Cas9 and signal-guide RNAs (5'-GTATCAAGTGGATGTAAACC-3') were transcribed using T7 RNA polymerase in vitro and comicroinjected into the cytoplasm of single-cell C57BL/6J mouse embryos to generate frameshift mutations by nonhomologous recombination through introduction of a 1 bp insertion in exon 13. Then, the embryos were cultured and transferred into the oviducts of pseudopregnant female mice at 0.5 days post-coitum. A mutation of *Dnah3* in the founder mouse and their offspring was confirmed using PCR and Sanger sequencing. The primers used for the generation of animal models are listed in **Table S1.**

Intracytoplasmic sperm injection (ICSI)

ICSI was carried out using standard techniques. In brief, one-month-old female KM mice were injected with 5 IU of equine chorionic gonadotropin (eCG) (ProSpec, HOR-272) to induce superovulation. Metaphase II-arrested (MII) oocytes were acquired through another injection of 5 IU human chorionic gonadotropin after 48 hours. MII oocytes were incubated with Chatot-Ziomek-Bavister medium (Easycheck, M2750) at 37.5 °C and 5% CO_2 until use. Mouse cauda epididymal spermatozoa were incubated in human tubal fluid (HTF) medium (Easycheck, M1150) and then frozen and thawed repeatedly to remove sperm tails. For ICSI, a single sperm head was microinjected into an MII oocyte by using a NIKON inverted microscope and a Piezo (PrimeTech, Osaka, Japan) in Whitten's-HEPES medium containing

0.01% polyvinyl alcohol (Gibco,12360-038) and cytochalasin B (3.5 g/ml; Sigma–Aldrich, C-6762). The successfully injected oocytes were transferred into G1-Plus medium (Vitrolife, 10132) and incubated at 37.5 °C and 5% CO_2. The animal experiments were approved by the Experimental Animal Management and Ethics Committee of West China Second University Hospital, Sichuan University.

Statistical analysis

Prism (version 8.4.0, GraphPad, Boston, MA, USA) and SPSS (version 18.0, IBM Corporation, Armonk, NY, USA) were used to perform statistical analyses. All data are presented as the means ± SEMs. Data from two groups were compared using an unpaired, parametric, two-sided Student's *t* test, and a *p* value less than 0.05 was considered statistically significant.

Ethics approval

This study was approved by Ethical Review Board of West China Second University Hospital, Sichuan University. Informed consent was obtained from each participate in this study before taking part.

Data availability

The published article includes all datasets generated or analyzed during this study. The exome-sequencing data supporting the current study have not been deposited in a public repository because of privacy issues but are available from the corresponding author on

request.

Competing interests

The authors declare that they have no conflict of interest.

Author contributions

Xiang Wang: Data curation; formal analysis; investigation; methodology; writing – original draft. **Gan Shen**: Data curation; formal analysis; **Yihong Yang**: Resources; investigation; methodology. **Chuan Jiang**: Data curation; formal analysis. **Tiechao Ruan**: Formal analysis; methodology. **Xue Yang**: Validation; investigation. **Liangchai Zhuo**: Investigation; **Yingteng Zhang**: Investigation, methodology. **Yangdi Ou**: Investigation. **Xinya zhao**: Investigation. **Shunhua Long**: Methodology. **Xiangrong Tang**: Investigation. **Tingting Lin**: Conceptualization; funding acquisition; project administration. **Ying Shen**: Conceptualization; supervision; project administration; writing – review and editing.

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Figure Legends

Figure 1. Identification of biallelic pathogenic variants in *DNAH3* from four unrelated infertile families. (**A**) Pedigrees of four families affected by *DNAH3* variants (M1–M7). Black arrows indicate the probands in these families. (**B**) Location of the variants and conservation of affected amino acids in DNAH3. Black arrows indicate the position of the variants. (**C**) Immunofluorescence staining of DNAH3 in sperm from the patients and normal control. Red, DNAH3; green, α-Tubulin; blue, DAPI; scale bars, 5 μm. (**D**) Western blotting analysis of DNAH3 expressed in spermatozoa from the patients and normal control.

Figure 2. **Defects in sperm morphology of the patients harboring *DNAH3* variants**. (**A**, **B**) Abnormal sperm morphology was observed through Papanicolaou staining (A), and SEM analysis (B) compared to normal control. Scale bars, 5 μm.

Figure 3. Ultrastructural and mitochondrial defects in sperm from infertile men with *DNAH3* variants. (**A**) TEM analysis of sperm obtained from a normal control and patients harboring *DNAH3* variants. Cross-sections of the midpiece, principal piece and endpiece of sperm from normal control showed the typical "9 + 2" microtubule structure, and an IDA and an ODA were displayed on the A-tube of each microtubule doublet. Cross-sections of the midpiece, principal piece and endpiece of sperm from the patients displayed absent or disordered CPs, MTDs and ODFs, as well as an evident missing of the IDAs in different pieces of the flagella. M, mitochondria sheath; ODF, outer dense fiber; MTD, microtubule doublets; CP, central pair; IDA, inner dynein arms; ODA, outer dynein arms. Scale bars, 200

nm. (**B**) Immunofluorescence staining of TOM20 in sperm from the patients and normal control. Red, TOM20; green, α-Tubulin; blue, DAPI; scale bars, 5 μm.

Figure 4. *Dnah3* KO male mice are infertile. (**A**) Fertility of *Dnah3* KO mice. The KO male mice were infertile (n = five biologically independent WT mice or KO mice; Student's *t* test; *, $P < 0.05$; NS, no significance; error bars, s.e.m.). (**B**) TEM analysis of the cross-sections of spermatozoa from *Dnah3* KO mice revealed an obvious absence of IDAs in different pieces of the flagella compared to WT mice. M, mitochondrion sheath; ODF, outer dense fiber; MTD, microtubule doublet; CP, central pair; IDA, inner dynein arm; ODA, outer dynein arm. Scale bars, 200 nm. (**C**) Disrupted mitochondria were observed in spermatozoa tail from *Dnah3* KO mice by TEM analysis. The yellow arrows indicate the normal mitochondria. The red arrowheads indicate the dilated intermembrane spaces and dissolved mitochondrial material. M, mitochondrion sheath. Scale bars, 200 nm. (**D**) Immunofluorescence staining of SLC25A4 indicated impaired mitochondrial formation in *Dnah3* KO mice compared to WT mice. Red, SLC25A4; green, α-Tubulin; blue, DAPI; scale bars, 5 μm.

Figure 5. **Immunofluorescence staining and western blotting analysis of IDA-associated proteins in spermatozoa obtained from normal control and patients with *DNAH3* variants**. (**A** – **C**) Immunofluorescence staining of DNAH1 (A), DNAH6 (B) and DNALI1 (C) in spermatozoa from patients and normal controls. Red, DNAH1 in (A), DNAH6 in (B), DNALI1 in (C); green, α-Tubulin; blue, DAPI; scale bars, 5 μm. (**D** – **F**) Western blotting

analysis of DNAH1(D), DNAH6 (E), DNALI1 (F) in sperm lysates from the patients and normal control.

Figure 6. Immunofluorescence staining and western blotting analysis of IDA-associated proteins in spermatozoa from WT and *Dnah3* KO mice. (**A** – **C**) Immunofluorescence staining of DNAH1 (A), DNAH6 (B) and DNALI1 (C) in spermatozoa from *Dnah3* KO and WT mice. Red, DNAH1 in (A), DNAH6 in (B), DNALI1 in (C); green, α-Tubulin; blue, DAPI; scale bars, 5 μm. (**D** – **F)** Western blotting analysis of DNAH1(D), DNAH6 (E) and DNALI1 (F) in spermatozoa lysates from *Dnah3* KO and WT mice.

Figure 7. ICSI outcomes of *DNAH3*-deficient patients and *Dnah3* KO mice. (**A**) The embryonic development of Patient 1 and Patient 3 after ICSI treatment. MII, metaphase II; PN, pronucleus; scale bars, 40 μm. (**B**) There was no difference in the fertilization rate or 2-cell and blastocyst embryo formation rates between the *Dnah3* KO and WT groups (n = three biologically independent WT mice or KO mice; Student's *t* test; NS, no significance; error bars, s.e.m.).

Tables

Table 1. Semen analysis of the patients in the present study.

		Patient 1	Patient 2	Patient 3	Patient 4	Reference
Semen parameters	Semen volume (ml)	4.3	3.3	0.8	4.6	≥1.5
	Semen concentration (10^6/ml)	2.0	0.5	11.0	21.0	≥15.0
	Motility (%)	6.0	3.0	0	0	≥40.0
	Progressive motility (%)	0	2.3	0	0	≥30.0
Sperm morphology	Normal (%)	1.3	1.1	1.8	1.2	≥4.0
	Head defect (%)	14.2	18.3	10.3	15.7	-
	Tail defect (%)	91.3	87.5	96.5	97.5	-

-, not applicable.

Table 2. Variants analysis of the patients in the present study.

		Patient 1		Patient 2	Patient 3		Patient 4	
Variant	cDNA mutation*	c.3590C>G	c.3590C>T	c.4837G>T	c.5587del	c.10355C>T	c.2314C>T	c.4045G>A
	Protein alteration	p.P1197R	p.P1197L	p.A1613S	p.L1863*	p.S3452L	p.R772W	p.D1349N
	Mutation type	Missense	Missense	Missense	Nonsense	Missense	Missense	Missense
Allele frequency	ExAC Browser	0	0	0.00037135	0	0.0001	0.0005	0.0002
	GnomAD	0	0	0.00026658	0	0.0001	0.0006	0.0002
	1000 Genomes Project	0	0	0.00099840	0	0.000199681	0.00139776	0.000798722
Function prediction	SIFT	Deleterious	Deleterious	Deleterious	/	Tolerated	Deleterious	Deleterious
	Polyphen-2	Probably damaging	Probably damaging	Probably damaging	/	Probably damaging	Probably damaging	Probably damaging
	Mutation Tester	Disease causing	Disease causing	/	/	Disease causing	Disease causing	Disease causing

*, NM_017539.2; /, not applicable.

Table 3. Semen analysis using CASA in the *Dnah3* KO mice.

	WT	KO	P[b] value
Semen parameters			
Sperm concentration (10^6 /ml) [a]	112.32 ± 18.26	105.17 ± 11.15	0.059
Motility (%) [b]	71.56 ± 3.97	4.37 ± 1.15	<0.01
Progressive motility (%) [b]	60.36 ± 4.32	4.37 ± 1.15	<0.01
Sperm locomotion parameters			
Curvilinear velocity (VCL) (μm/s) [b]	67.54 ± 6.79	9.07 ± 1.22	<0.01
Straight-line velocity (VSL) (μm/s) [b]	28.91 ± 4.86	2.68 ± 0.52	<0.01
Average path velocity (VAP) (μm/s) [b]	39.02 ± 5.31	3.85 ± 0.82	<0.01
Amplitude of lateral head displacement (ALH) (μm) [b]	0.71 ± 0.03	0.13 ± 0.04	<0.01
Linearity (LIN) [b]	0.43 ± 0.07	0.30 ± 0.02	0.037
Wobble (WOB, = VAP/VCL) [b]	0.58 ± 0.06	0.42 ± 0.05	0.024
Straightness (STR, = VSL/VAP)	0.74 ± 0.22	0.70 ± 0.13	0.80
Beat-cross frequency (BCF) (Hz) [b]	4.86 ± 0.12	0.73 ± 0.08	<0.01

[a] Epididymides and vas deferens.

[b] A significant difference, two-sided student's t-test. n = 3 biologically independent WT mice or KO mice.

683 **Table 4. Outcomes of ICSI treatment in the patients with *DNAH3* mutations.**

	Patient	Patient 1	Patient 2		Patient 3	Patient 4			
Subjects	Female age (y)	24	30		30	36			
	Length of primary infertility history (y)	6	1		3	8			
Basal hormones	FSH (IU/L)	7.1	8.5		3.49	4.3			
	LH (IU/L)	4.44	2.5		4.2	2.6			
	E2 (pg/mL)	83	50		43.49	68			
	Prog (ng/mL)	0.2	0.6		0.3	0.3			
ICSI Cycles		Cycle 1	Cycle 1	Cycle 2	Cycle 1	Cycle 1	Cycle 2	Cycle 3	Cycle 4
	E2 level on the trigger day (pg/ml)	3366	1519.6	1582.4	>5000	4440	5000	3152	2206
	No. of follicles ≥ 14 mm on the trigger day	15	6	5	20	13	12	14	11
	No. of follicles ≥ 18 mm on the trigger day	10	3	4	8	11	9	7	5
	No. of oocytes retrieved	24	6	5	25	16	20	21	14
ICSI progress	No. of MII oocytes	21	6	5	20	12	13	8	7
	Fertilization rate (%)	80.95 17/21)	50 (3/6)	100 (5/5)	95 (19/20)	41.67 (5/12)	46.15 (6/13)	50 (4/8)	42.86 (3/7)
	Cleavage Rate (%)	100 (17/17)	100 (2/2)	100 (5/5)	100 (19/19)	100 (5/5)	100 (6/6)	100 (4/4)	66.7 (2/3)
	Available D3 embryos	13	2	5	15	2	2	0	0
	Blastocyst formation rate (%)	61.5 (8/13)	0	66.7 (2/3)	66.7 (10/15)	-	-	-	-
Clinical outcomes	No. of embryos transferred	2 blastocysts	0	2 D3	1 blastocyst	2 D3	2 D3	-	-
	Implantation rate (%)	50 (1/2)	0	0	100 (1/1)	0	0	-	-
	Clinical pregnancy	Yes	No	No	Yes	-	-	-	-
	No. of live birth	ongoing	-	-	ongoing	-	-	-	-

684 -, not applicable.

Supporting information

Figure S1. The expression of DNAH3 in mouse testis. (**A**) qPCR analysis revealed that *Dnah3* was highly expressed in the mouse testis. (**B**) qPCR analysis showed that *Dnah3* expression was significantly elevated beginning on postnatal Day 12, peaked at postnatal Day 30, and maintained a stable expression level thereafter.

Figure S2. DNAH3 is expressed during spermatogenesis in mice and humans. (**A**) Immunofluorescence staining of DNAH3 in isolated mouse germ cells. Pink, PNA; green, DNAH3; blue, DAPI; scale bars, 5 μm. (**B**) Immunofluorescence staining of DNAH3 in isolated human germ cells. Pink, PNA; green, DNAH3; blue, DAPI; scale bars, 5 μm.

Figure S3. Generation of *Dnah3* KO mice. (**A**) Schematic illustration of the strategy for the generation of *Dnah3* KO mice. (**B, C**) PCR sequencing (B) and qPCR (C) were used to confirm the genotype and KO efficiency (n = three biologically independent WT mice or KO mice; Student's *t* test; *, $P<0.05$; error bars, s.e.m.). (**D**) Immunofluorescence staining of DNAH3 in testis of *Dnah3* KO mice and WT mice. Green, DNAH3; blue, DAPI; scale bars, 75 μm. (**E**) Immunofluorescence staining of DNAH3 in spermatozoa isolated from the cauda epididymis of *Dnah3* KO mice and WT mice. Red, DNAH3; green, α-Tubulin; blue, DAPI; scale bars, 5 μm.

Figure S4. Ciliary development of *Dnah3* KO mice. (**A**) H&E staining of lung, brain, eye, and oviduct from *Dnah3* KO mice and WT mice. Scale bars, 100 μm. (**B**) Analysis of ciliary

development in the lung, brain, eye, and oviduct from *Dnah3* KO mice and WT mice by using immunofluorescence staining. Green, Ac-Tubulin; blue, DAPI; scale bars, 20 μm.

Figure S5. Fertility of *Dnah3* KO mice. (**A**) H&E staining of ovary tissue sections from 8-week-old *Dnah3* KO female mice and WT female mice. Scale bars, 75 μm (n = three biologically independent WT mice or KO mice). (**B**) Sizes of the testis and epididymis of the 8-week-old *Dnah3* KO and WT mice (n = three biologically independent WT mice or KO mice; Student's *t* test; NS, no significance; error bars, s.e.m.).

Figure S6. Morphology of sperm isolated from *Dnah3* KO mice. (A, B) Papanicolaou staining (A) and SEM analysis (B) revealed morphological defects in partial spermatozoa from *Dnah3* KO mice compared to WT mice. Scale bars in (A), 5 μm; scale bars in (B), 2.5 μm.

Figure S7. Immunofluorescence staining of ODA-associated proteins in spermatozoa obtained from variants within *DNAH3* patients. (**A** – **C**) The expression of DNAH8 (A), DNAH17 (B) and DNAI1 (C) in spermatozoa of the patients was comparable to that in normal controls. Red, DNAH8 in (A), DNAH17 in (B), DNAI1 in (C); green, α-Tubulin; blue, DAPI; scale bars, 5 μm.

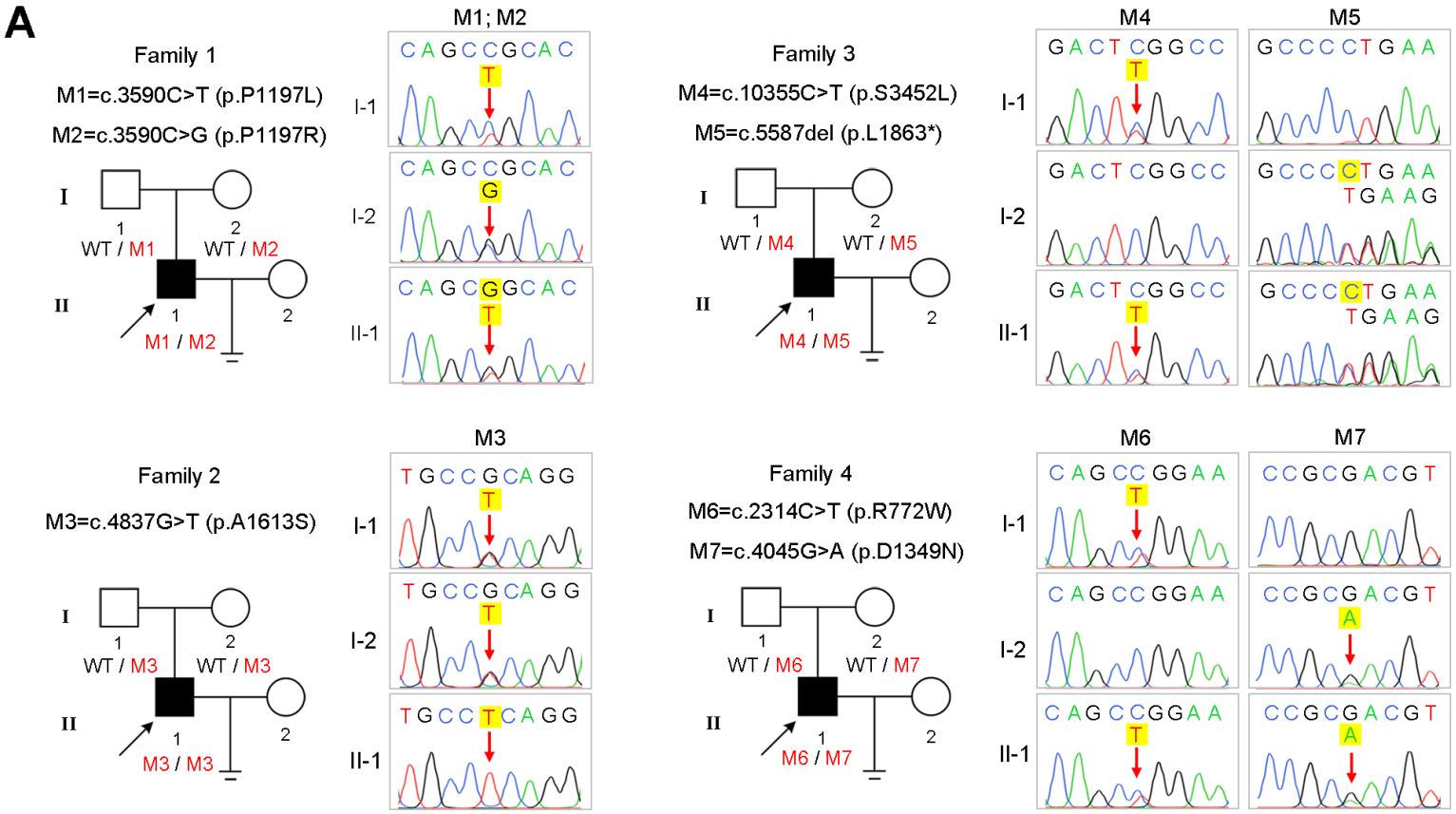
Figure S8. Immunofluorescence staining of ODA-associated proteins in spermatozoa of *Dnah3* KO and WT mice. (**A** – **C**) The expression of DNAH8 (A), DNAH17 (B) and

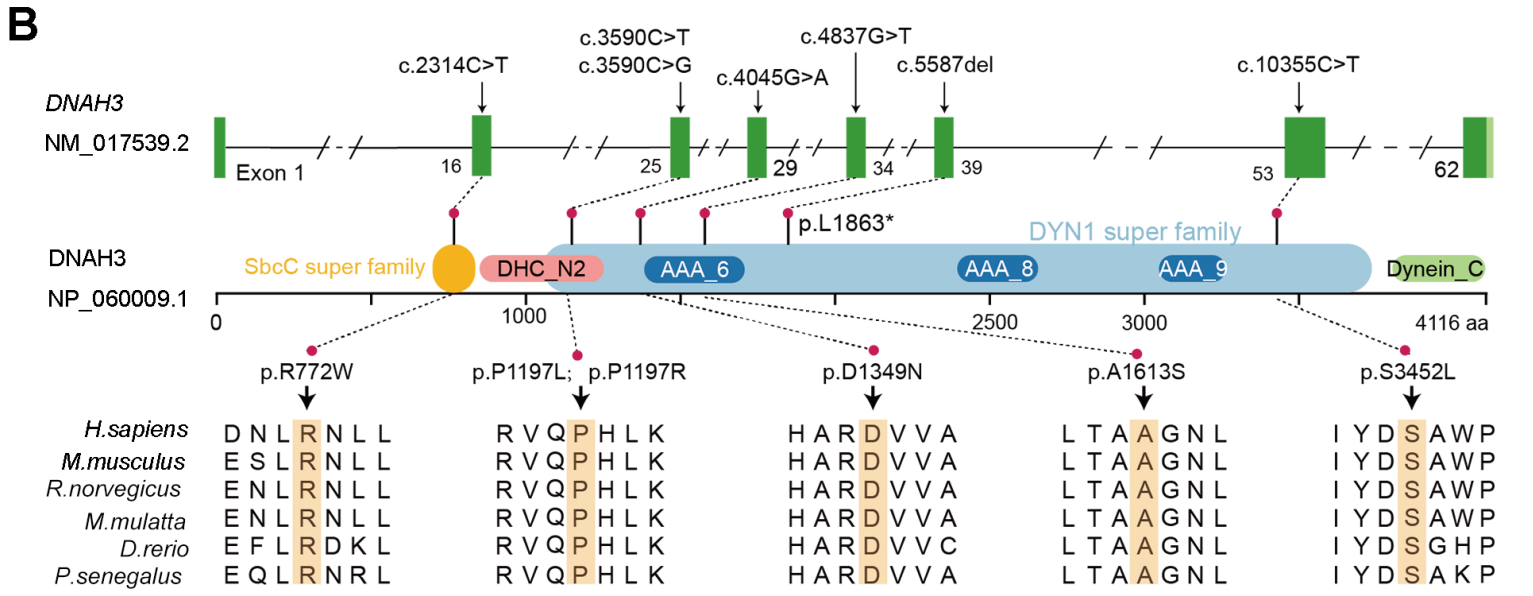
DNAI1 (C) in spermatozoa from *Dnah3* KO mice was comparable to that in spermatozoa from WT mice. Red, DNAH8 in (A), DNAH17 in (B), DNAI1 in (C); green, α-Tubulin; blue, DAPI; scale bars, 5 μm.

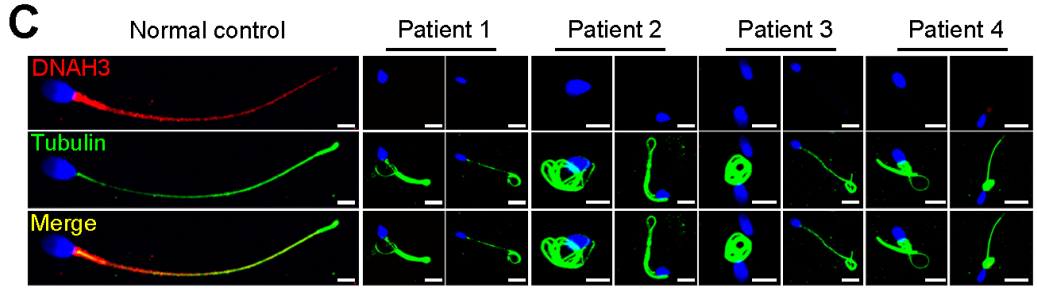
Table S1. Primer pairs used in the present study.

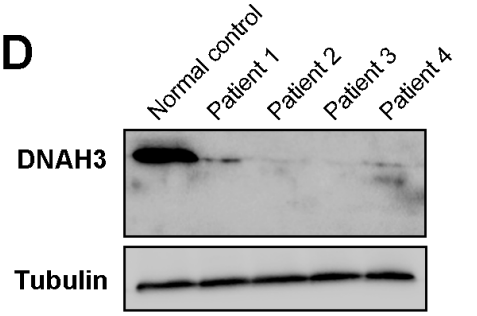
Movie S1. CASA of sperm from WT mice. Sperm from the epididymis of WT mice were collected, incubated, and recorded under a phase-contrast microscope. A normal quantity and motility of sperm were observed in the WT mice (n = three biologically independent WT mice).

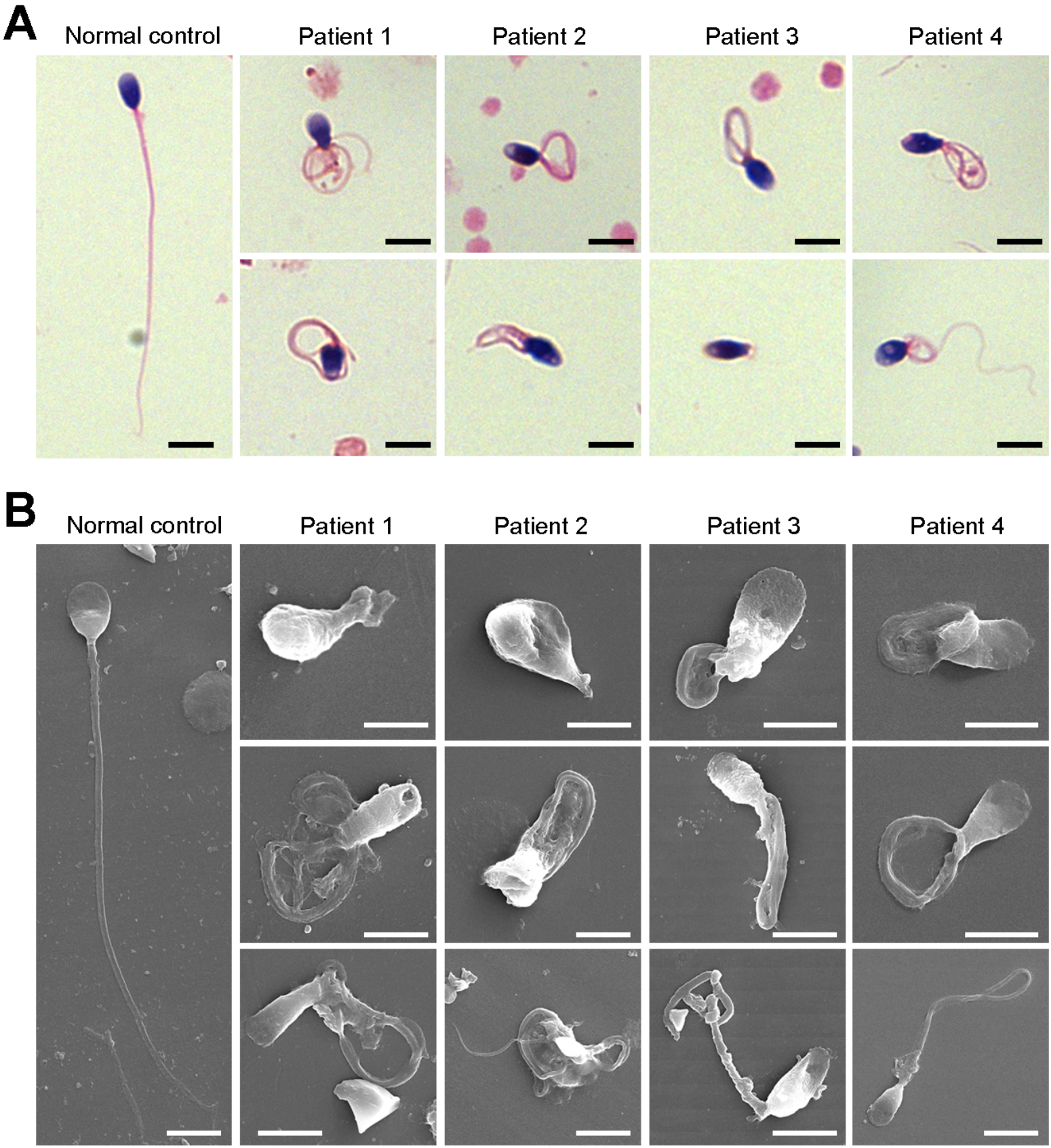
Movie S2. CASA of sperm from *Dnah3* KO mice. Epididymal sperm of *Dnah3* KO mice were collected, incubated in HTF medium at 37 °C for 10 minutes, and recorded under a phase-contrast microscope. The movie showed a significantly reduced motility of sperm from *Dnah3* KO (n = three biologically independent *Dnah3* mice).

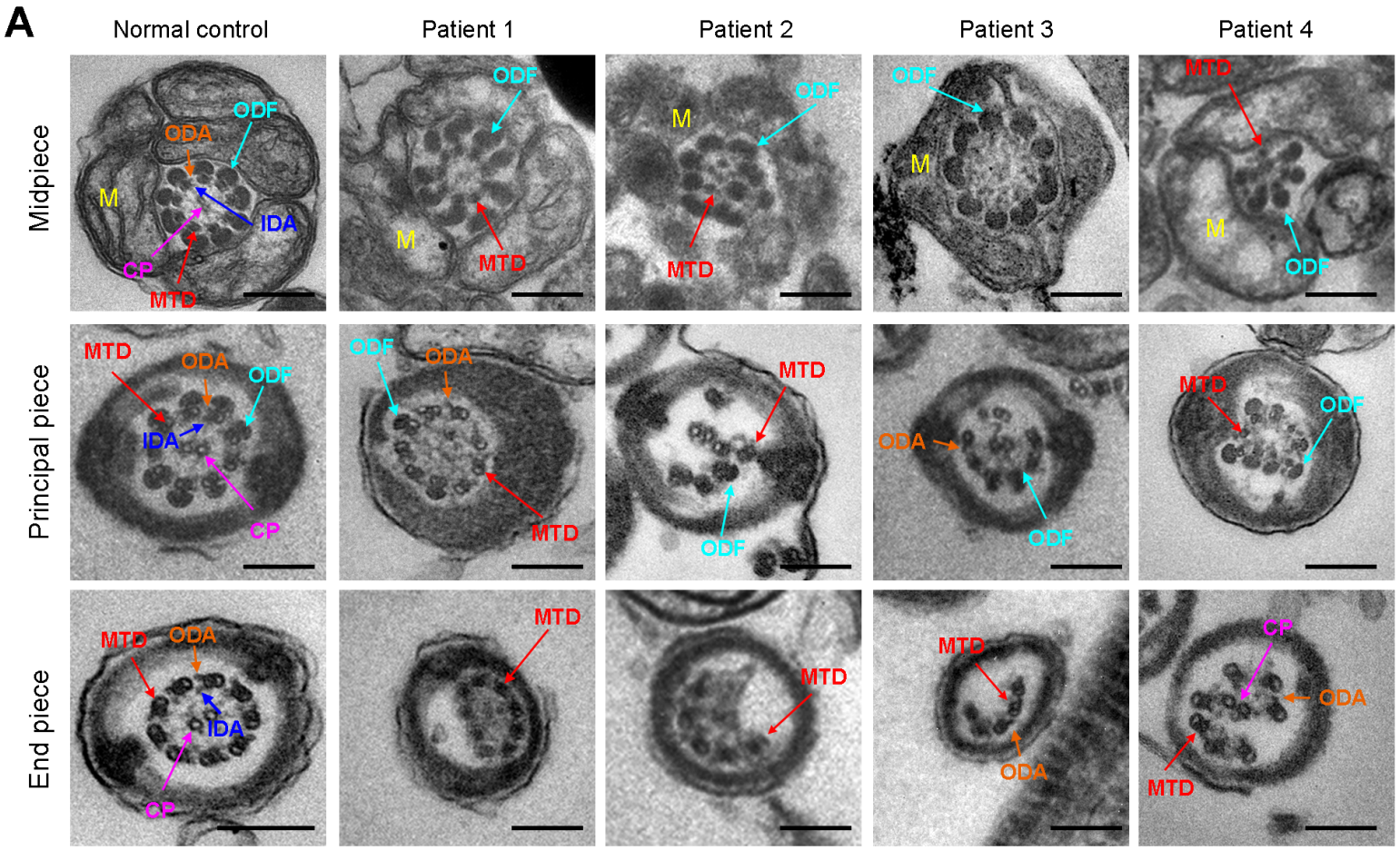

A
Family 1
M1=c.3590C>T (p.P1197L)
M2=c.3590C>G (p.P1197R)
M1; M2
I-1
I-2
II-1
WT / M1
WT / M2
M1 / M2
Family 3
M4=c.10355C>T (p.S3452L)
M5=c.5587del (p.L1863*)
M4
M5
WT / M4
WT / M5
M4 / M5
Family 2
M3=c.4837G>T (p.A1613S)
M3
WT / M3
M3 / M3
Family 4
M6=c.2314C>T (p.R772W)
M7=c.4045G>A (p.D1349N)
M6
M7
WT / M6
WT / M7
M6 / M7

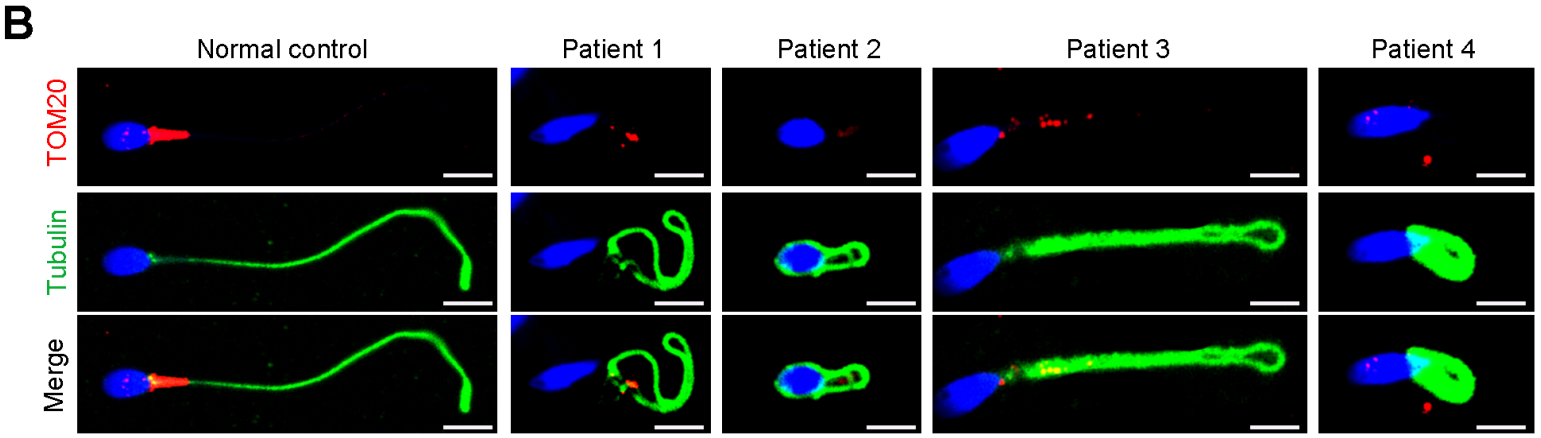

B
DNAH3
NM_017539.2
Exon 1
c.2314C>T
c.3590C>T
c.3590C>G
c.4045G>A
c.4837G>T
c.5587del
c.10355C>T
p.L1863*
DNAH3
NP_060009.1
SbcC super family
DHC_N2
AAA_6
AAA_8
AAA_9
DYN1 super family
Dynein_C
4116 aa
p.R772W
p.P1197L; p.P1197R
p.D1349N
p.A1613S
p.S3452L
H.sapiens
M.musculus
R.norvegicus
M.mulatta
D.rerio
P.senegalus

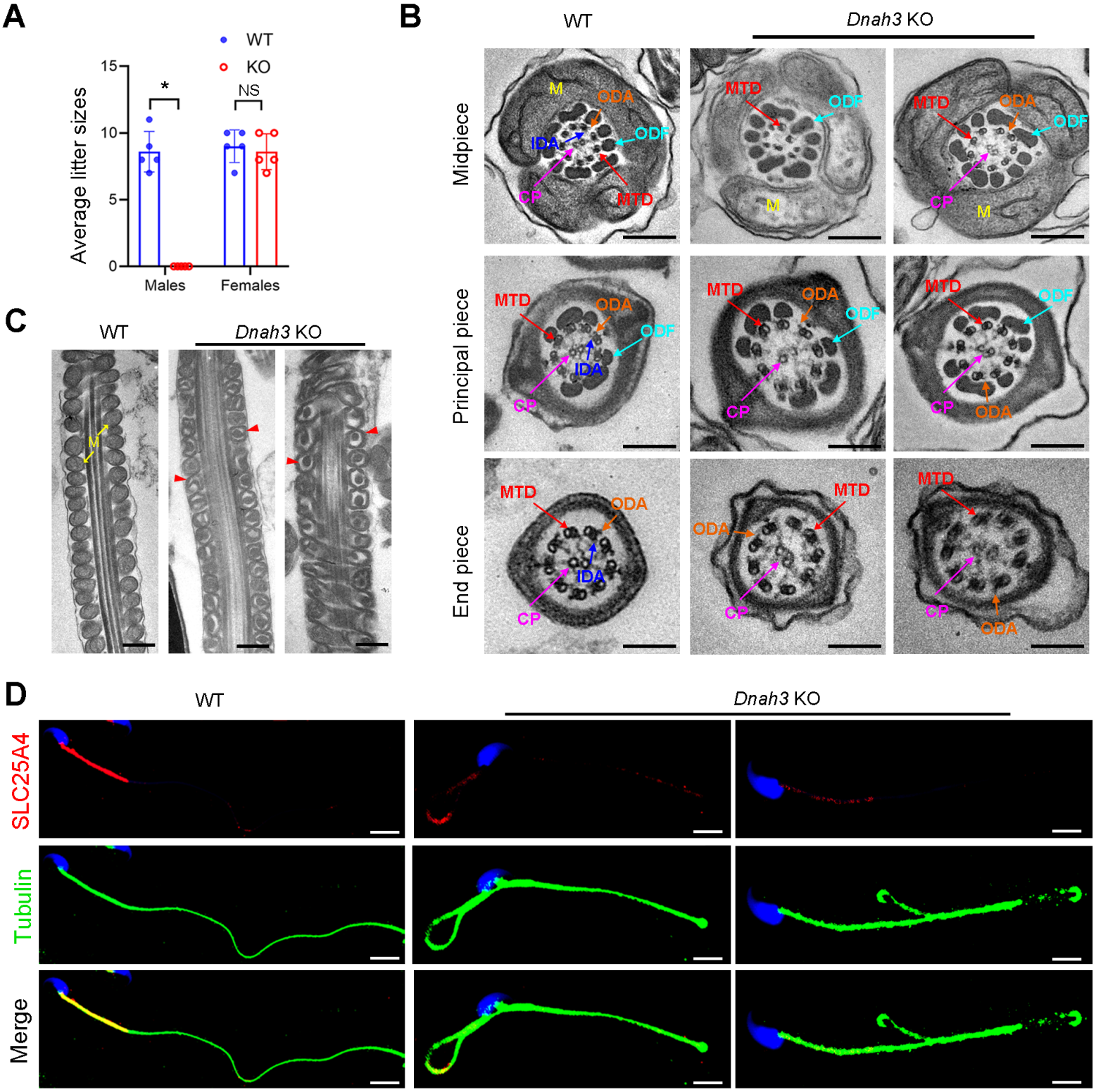

C
Normal control
Patient 1
Patient 2
Patient 3
Patient 4
DNAH3
Tubulin
Merge

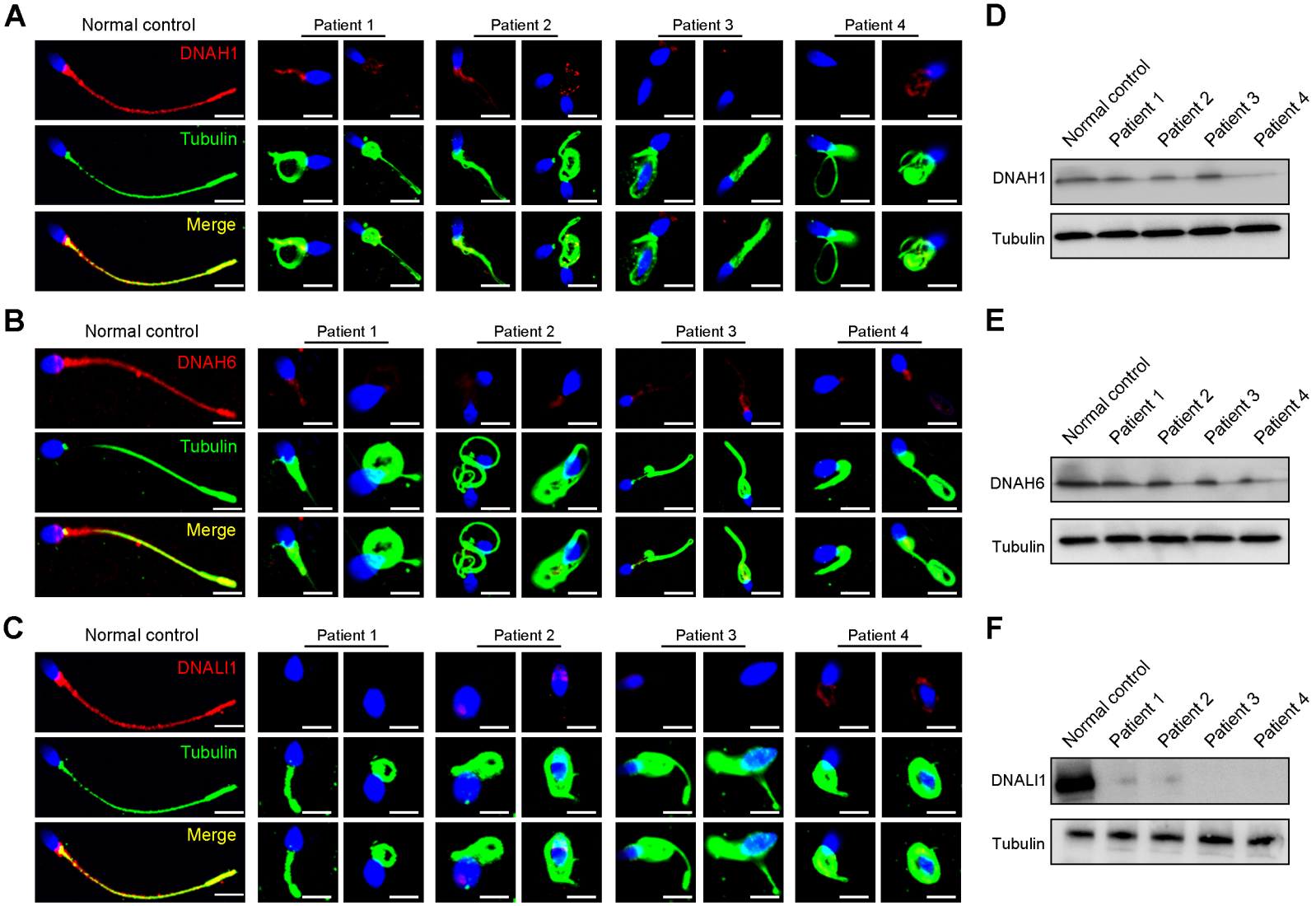

D
Normal control
Patient 1
Patient 2
Patient 3
Patient 4
DNAH3
Tubulin

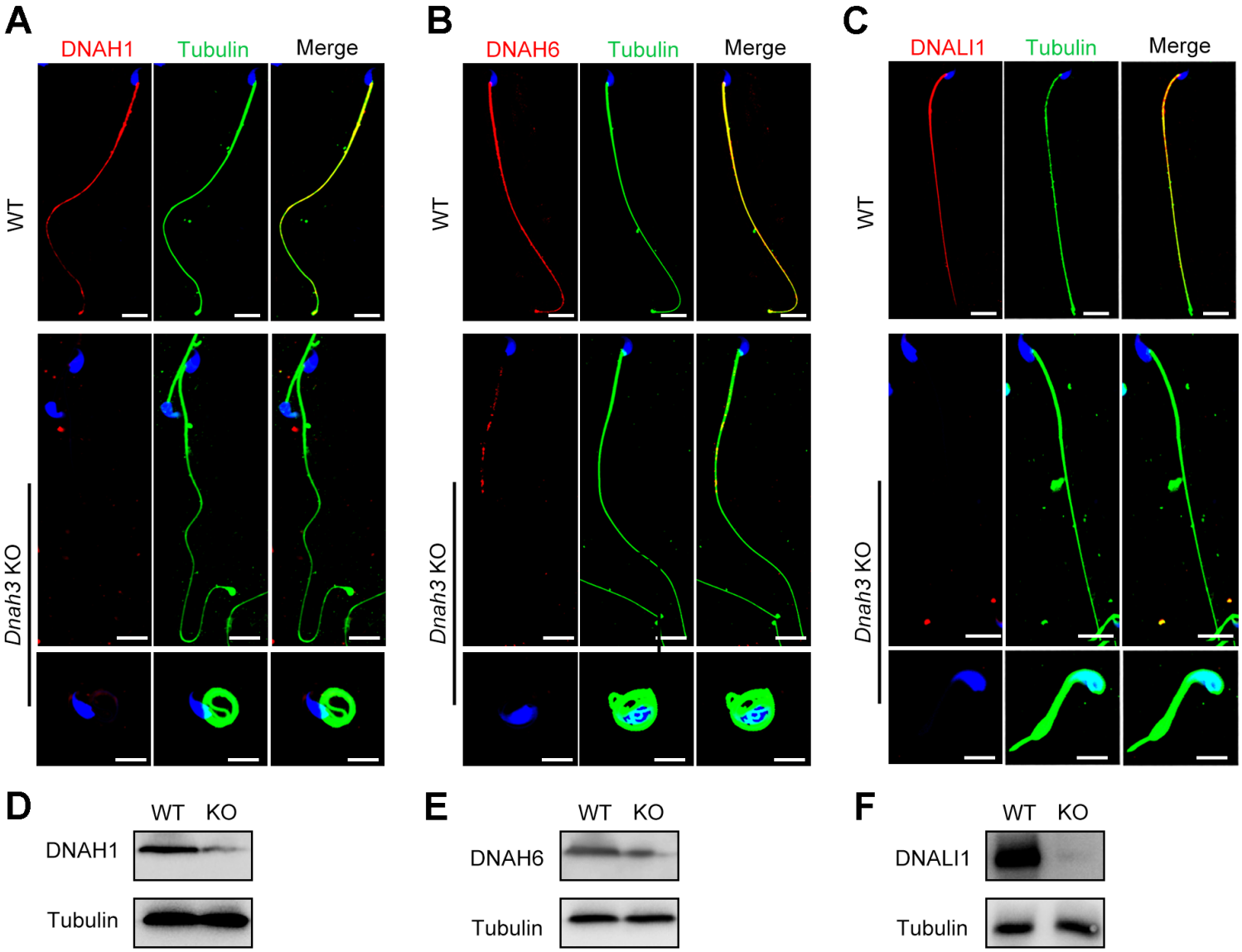

A
Normal control
Patient 1
Patient 2
Patient 3
Patient 4
B
Normal control
Patient 1
Patient 2
Patient 3
Patient 4

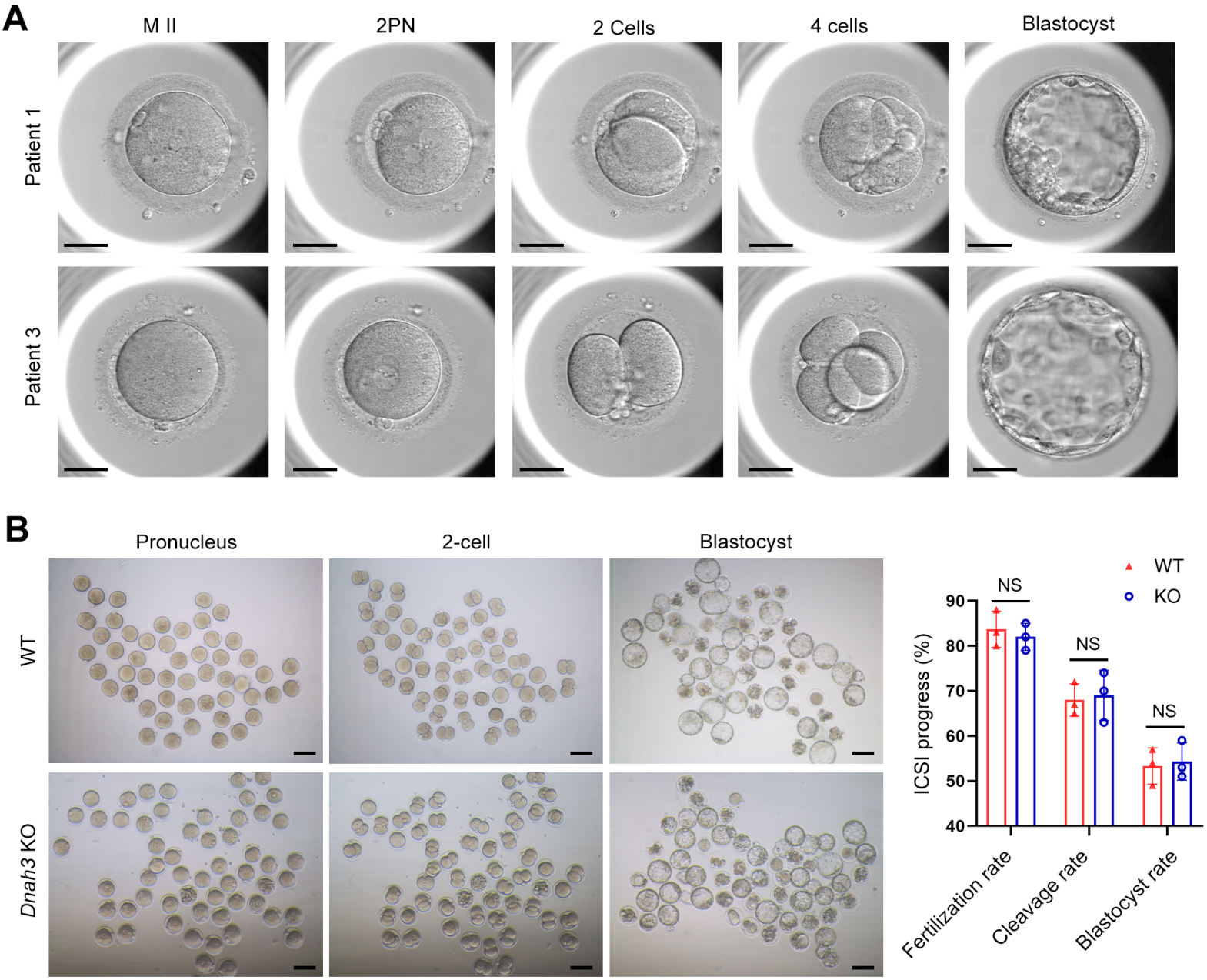

A
Normal control
Patient 1
Patient 2
Patient 3
Patient 4
Midpiece
Principal piece
End piece
ODF
ODA
M
IDA
CP
MTD


B
Normal control
Patient 1
Patient 2
Patient 3
Patient 4
TOM20
Tubulin
Merge


A
WT
KO
Average litter sizes
15
10
5
0
*
NS
Males
Females
B
WT
Dnah3 KO
Midpiece
Principal piece
End piece
M
ODA
ODF
IDA
CP
MTD
C
WT
Dnah3 KO
M
D
WT
Dnah3 KO
SLC25A4
Tubulin
Merge


A
Normal control
Patient 1
Patient 2
Patient 3
Patient 4
DNAH1
Tubulin
Merge
B
Normal control
Patient 1
Patient 2
Patient 3
Patient 4
DNAH6
Tubulin
Merge
C
Normal control
Patient 1
Patient 2
Patient 3
Patient 4
DNALI1
Tubulin
Merge
D
Normal control
Patient 1
Patient 2
Patient 3
Patient 4
DNAH1
Tubulin
E
Normal control
Patient 1
Patient 2
Patient 3
Patient 4
DNAH6
Tubulin
F
Normal control
Patient 1
Patient 2
Patient 3
Patient 4
DNALI1
Tubulin


A
DNAH1
Tubulin
Merge
WT
Dnah3 KO
B
DNAH6
Tubulin
Merge
WT
Dnah3 KO
C
DNALI1
Tubulin
Merge
WT
Dnah3 KO
D
WT
KO
DNAH1
Tubulin
E
WT
KO
DNAH6
Tubulin
F
WT
KO
DNALI1
Tubulin


A
M II
2PN
2 Cells
4 cells
Blastocyst
Patient 1
Patient 3
B
Pronucleus
2-cell
Blastocyst
WT
Dnah3 KO
WT
KO
NS
NS
NS
ICSI progress (%)
90
80
70
60
50
40
Fertilization rate
Cleavage rate
Blastocyst rate