Effects of childhood hearing loss on the subcortical and cortical representation of speech

Axelle Calcus[1, 2, 3] & Stuart Rosen[1]

[1] *Department of Speech, Hearing and Phonetic Sciences, UCL, London WC1N 1PF, UK*

[2] *Laboratoire des Systèmes Perceptifs, CNRS UMR 8248, École Normale Supérieure, Paris Sciences et Lettres Research University, Paris, France*

[3] *Center for Research in Cognition & Neurosciences (CRCN), ULB Neuroscience Institute (UNI), Université Libre de Bruxelles (ULB), Brussels, Belgium*

Corresponding author: Axelle Calcus (axelle.calcus@ulb.be)

Abbreviated title: Speech processing in childhood hearing loss

Number of figures: 7; Number of tables: 2 (submit with Tables_v9.docx – supplementary tables and rebuttal_v4)

The authors declare no conflict of interest

Abstract

Little is known about the effects of childhood mild-to-moderate sensorineural hearing loss (MM HL) on the function of the auditory pathway. We aimed to examine the effect of childhood MM HL and the benefit of frequency-specific amplification on both subcortical and cortical auditory processing, and to relate it to speech-perceptual abilities. We recorded subcortical and cortical responses to speech syllables in nineteen children with congenital MM HL (unamplified and amplified), and sixteen children with typical hearing (unamplified sounds only). Speech perception was measured behaviourally. Congenital HL led to smaller subcortical and cortical responses to unamplified speech sounds. There was a significant benefit of amplification on subcortical and early, but not late, cortical responses, with some effects differing across age. No relationship was found between the neural and behavioural measures. Childhood MM HL affects both subcortical and cortical processing of speech. Amplification mostly benefits subcortical processing of speech in younger children. Childhood HL leads to functional changes in the processing of sounds, with amplification differentially affecting subcortical and cortical levels of the auditory pathway.

Introduction

Animal studies have shown biological and physiological alterations in the properties of neurons in the cochlea, subcortex, and cortex following even a temporary, mild-to-moderate (MM) hearing loss (HL)[1,2]. Similar changes have been documented in humans with congenital deafness [3–5], or in older adults with age-related MM HL (for a review see [6]). Nowadays, most individuals with MM HL who seek treatment receive auditory stimuli via hearing aids, which amplify the incoming auditory signal. To date, the bulk of studies investigating the effects of MM HL on the functional integrity of the auditory pathway (and the potential benefit of hearing aid amplification) has focused on older adults. However, congenital hearing loss affects up to 2 live births in 1000[7], milder degrees of loss being more prevalent yet more often overlooked. Here, we evaluate the effect of childhood MM HL and the benefit of amplification on simultaneously recorded subcortical and cortical responses to speech sounds.

In the auditory system, speech sounds travel from the cochleae to the auditory cortices, undergoing increasingly complex levels of processing. At the level of the cochleae, sounds undergo a frequency analysis. The output of each channel can be separated into their relatively slowly fluctuating modulated signals known as the envelope (ENV), imposed upon a rapidly varying carrier waveform, known as the temporal fine structure (TFS). Both ENV and TFS cues are thought to be important in decoding speech. ENV cues support the robust identification of speech in quiet[8], whereas TFS cues are thought to be especially useful in noisy backgrounds[9,10]. Indeed, individuals with typical hearing (TH) typically benefit from the TFS when they glimpse into the "dips" (i.e., local improvements in SNR) of a fluctuating background noise[11,12]. Therefore, speech intelligibility is much better in the presence of fluctuating than steady background noise, a phenomenon termed "masking release" [13,14]. However, adults with HL show reduced ability to use TFS[15], a difficulty that limits the amount of masking release they experience[16,17]. Similarly to adults, children and adolescents with MM HL experience poorer

speech intelligibility in noise[18–20]. They show reduced sensitivity to TFS compared to

children/adolescents with TH[21], which likely limits masking release of speech in the presence

of a fluctuating background noise, even when they wear hearing aids[22].

At the subcortical level of the auditory pathway, the frequency following response

(FFR) reflects the periodicity of speech sounds[23]. Adding the neural responses to speech sounds

presented in alternating polarities is thought to enhance the ENV of the response (henceforth

FFR_{ENV}, here used as an index of fundamental frequency encoding), whereas subtracting them

is considered to enhance the TFS (henceforth FFR_{TFS}, an index of spectral harmonic

encoding[24]). Although the primary generator of the FFR appears to vary according to the

fundamental frequency (F0) of the input stimulus, the current view is that the FFR emerges

from multiple generators, including the cochlear nucleus, inferior colliculus and thalamus[25,26].

Early studies suggested that the auditory brainstem reached maturity within the first 18 months

of life[27]. However, more recent studies indicate that the FFR evoked by speech sounds keeps

maturing well into the childhood years[28,29].

At the cortical level, components of the late auditory event-related potentials (LAERs)

arise which are thought to reflect the initial detection (P1, N1, and P2), classification and

discrimination (P2 and N2) of auditory stimuli, reflecting processing for auditory events

ranging from isolated sounds to complex auditory scenes[30]. LAERs are thought to be generated

in the auditory cortex (for a review, see [31]), with additional contributors from non-auditory (e.g.,

prefrontal, premotor) areas[32]. The mismatch negativity (MMN) is thought to reflect the

discrimination of a deviant sound, and is typically evoked by a comparatively rare deviant in a

stream of repeated standards (i.e. an oddball paradigm; for a review see [33]). The MMN is

thought to have several additional generators, including bilateral temporal cortex, right inferior

frontal gyrus, and bilateral frontal and centro-parietal regions (e.g., [34–36]). In addition to distinct

topographies, both LAERs and MMNs show distinct developmental trajectories. In typical

development, P1 and N2 responses are thought to decrease in amplitude as children grow older,

while N1 and P2 progressively appear in childhood, growing in amplitude until

adolescence[37,38]. MMN amplitude is thought to increase throughout childhood[39], but to be stable

from around 10 years of age to adulthood[40,41].

Both subcortical and cortical levels of the auditory pathway thus appear to follow a

protracted trajectory, maturing until late adolescence. How MM HL affects the development of

subcortical and cortical speech processing remains largely unknown. So far, studies have

focused on *either* subcortical *or* cortical levels of processing, only providing fragmented

information about the physiology of the auditory pathway in children/adolescents with MM

HL. Yet using the right set of recording/analysis parameters, it is now possible to obtain

subcortical and cortical responses during the same recording session [42,43]. Therefore, the first

aim of this project was to evaluate the effect of childhood hearing loss on simultaneously

recorded subcortical and cortical levels of processing. Noteworthily, the existing literature on

this topic can be classified according to the strategy used to manage intensity levels across

listener groups.

Neural processing of speech presented at fixed intensity levels across groups with and without

HL

When studying the effect of hearing loss on auditory processing, researchers often

choose to investigate the unaided performance of individuals with HL. Stimuli are thus

presented at a fixed intensity level for all individuals, with HL or not. Adult studies using this

strategy to evaluate the effect of HL on subcortical processing have led to contradictory findings

(for a review, see [44]). While some authors reported smaller FFR_{ENV} and FFR_{TFS} in adults with

HL than adults with TH[45,46], others did not show such group differences[47,48]. Note that a possible

explanation for this discrepancy lies in the age-matching across groups[47,48], or lack thereof[45,46].

To our knowledge, no study has focused on the effects of childhood MM HL on subcortical processing of speech, using fixed intensity stimuli across groups of children/adolescents.

At the cortical level, studies using fixed intensity levels across groups suggest that childhood MM HL leads to delayed and/or smaller LAERs evoked by speech or pure tones [49,50]. Developmental changes following MM HL were also apparent in the MMN: While 8-11 year old children with MM HL showed age-appropriate MMN responses to sounds, this was no longer the case in 12-16 year old adolescents with MM HL, as shown both cross-sectionally and longitudinally[49]. This suggests that some effects of childhood MM HL might only reveal themselves in adolescence, highlighting the need to take age into account in developmental studies.

Neural processing of speech presented at similar sensation levels across groups

A common limitation of the above-mentioned studies using fixed intensity levels for all participants is that individuals with HL would have perceived the stimuli at reduced sensation levels (SL) relative to controls. Therefore, some researchers have attempted to equate for the effects of peripheral hearing loss by presenting listeners with TH and those with SNHL are presented with sounds at different intensities – more intense for individuals with SNHL (either by simulating the frequency-specific gain provided by individuals' hearing aids, or by recording the neurophysiological response while listeners wear hearing aids). With such adjustments for HL, adults with acquired MM HL show larger FFR_{ENV} to an amplified speech stimulus compared to an unamplified one[51,52] and in one study, than the FFR_{ENV} evoked by unamplified speech presented to TH listeners[47]. A similar benefit of amplification on the FFR_{ENV} evoked by the speech token /suʃi/ was recently reported in 18 children with HL aged 6 to 17 years, compared to the unamplified stimulus[53]. Note that older adults with acquired MM HL did not

show a significant benefit of amplification on FFR$_{TFS}$[47]. To our knowledge, the effect of

amplification and childhood HL on FFR$_{TFS}$ has remained unexplored so far.

At the cortical level, studies looking at the effect of childhood HL on LAERs by

accounting for the effects of peripheral HL have led to contradictory findings. Pilot data from

a small sample of 5 children aged 2 to 6 years show age-appropriate P1 and N2 evoked by

speech stimuli when the children wore their hearing aids[54]. Another study focused on older,

unaided children (9-12 years) with MM HL, to whom speech stimuli were presented on average

18 dB greater than in children with TH[55]. Note that here, the amplification provided to the group

with HL was not frequency-specific – unlike hearing aids. Results indicate age-appropriate P1

and MMN responses in children with MM HL, but smaller (but not later) N2 responses

compared to children with TH. However, 5 to 8 year-old children who wore their hearing aids

(n = 7) showed significantly smaller LAERs evoked by complex tones than age-matched

children with TH[56]. Interestingly, when re-tested three years later, the same children with HL

showed age-appropriate LAERs. At both time-points, MMN amplitudes were age-appropriate

in children with MM HL. To our knowledge, no study has directly investigated the intra-

individual benefit of amplification on the cortical processing of speech in children/adolescents

with MM HL.

Relationship between speech processing and perception of speech

A main stake of auditory neurophysiological studies is to relate neural processing to

speech perception in a variety of contexts and clinical populations. Because the FFR reflects

encoding of F0 and lower harmonics of the signal, that are particularly helpful in adverse

listening environments, it is thought to relate to speech perception in noise. Better speech

perception in noise has been associated with subcortical processing of complex sounds in quiet,

as observed in children[57,58], young[59] and older adults with TH[60,61]; but see [62] for a study that did

not find such an association). In fact, FFR_{ENV} evoked by amplitude-modulated tones presented in quiet even seems to predict sentence understanding in noise in adults with TH[63]. Recent evidence suggests that FFR_{ENV} (evoked by speech in quiet) correlates with speech perception in noise in unaided (but not in aided) 4 to 9 year-olds with congenital mild to moderate HL[64]. Whether subcortical processing relates to speech perception in noise in older children/adolescents with congenital HL remains an open question.

Cortical processing of sounds is also thought to relate to behavioural speech perception, mostly in quiet. In 4-month old children with TH or HL, components of the LAER relate to parents' reports of auditory functional performance[65]. In adults, the MMN is thought to relate to perceptual speech discrimination, at least at the group level (for a review, see [66]). However, no clear relationship has emerged between MMN amplitude and speech discrimination in children with TH so far [67–69]. To our knowledge, it remains unclear whether cortical EEG measures relate to speech perception in children/adolescents with congenital HL.

Study objectives

The main goal of this study was twofold: to evaluate (i) the effect of childhood HL and (ii) the benefit of amplification on both subcortical and cortical auditory processing. (i) So far, no study has investigated the effect of childhood MM HL on subcortical processing presented at fixed intensity levels across age-matched groups. Additionally, results from (older) adult studies have led to contradictory results. Therefore, this study will be a first exploration of the effect of MM HL on the processing of unamplified sounds throughout development. At the cortical level, we predict smaller/later LAER and MMN responses in children/adolescents with HL than those with TH, with a possible interaction between age and group on the MMN. (ii) Regarding the second objective, we expect larger FFR_{ENV} in children with HL when sounds are amplified than unamplified. At the cortical level, results from the literature are mixed, but have

never compared LAER/MMN with and without amplification in the same children with HL. We will thus explore the effect of amplification on the cortical responses evoked in children/adolescents with HL.

Given that subcortical and cortical processing have both been related to speech perception, an exploratory objective was (iii) to investigate how these neurophysiological responses relate to speech intelligibility in quiet and in noise.

Methods

Participants

Thirty-five children aged 8 to 16 years were included in the study. Nineteen (8 female) had a diagnosis of mild-to-moderate SNHL (HL group) and sixteen (8 female) were age-matched children with typical hearing (TH group). All participants were monolingual British English natives and obtained a nonverbal IQ (NVIQ) score of at least 85 (Block Design subtest of the Weschler Abbreviated Scale of Intelligence[70]). No child had any known medical, psychological, neurological or developmental disorders other than SNHL. Individual air-conduction audiometric thresholds were measured bilaterally in all children at octave frequencies from 0.125 to 8 kHz using an Interacoustics AC33 audiometer. The study was conducted with the verbal assent of the participants, the written consent of their parents, and was approved by the UCL Research Ethics Committee (2109/004). It was performed in accordance with the Declaration of Helsinki.

Figure 1: Pure-tone air-conduction audiometric thresholds for children with HL (orange) and children with TH (blue and white). Audiometric thresholds are shown across octave frequencies from 0.25 to 8 kHz in the left and right ears. Individual thresholds for the HL group are shown as lines, and the group mean is shown as a bold line. Mean thresholds for the TH group are marked in white, with the shaded blue area representing the range for the TH group.

Participants in the HL group had better-ear pure tone average (BEPTA) thresholds of 21-70 dB HL across 0.25-8 kHz (see Figure 1), with on average 7 dB difference in PTA between the ears. All children in the HL group were spoken language users (i.e., non-signers), and sixteen had prescription hearing aids, but these were not used in this study. Children in the HL group were individually matched in age (± 6 months) to those in the TH group, and mean age did not differ significantly between the two groups (see Table 1). None of the children in the TH group had a known history of hearing loss (including otitis media with effusion), educational difficulties, or speech and language problems (based on parent/guardian report). All had mean PTA thresholds ≤ 20 dB HL across octave frequencies 0.25-8 kHz in both ears and obtained thresholds no higher than 20 dB HL at any given frequency.

Group comparisons are presented in Table 1. Note that some children did not complete all behavioural and electrophysiological tasks. With respect to the behavioural tasks, two children (1 TH and 1 HL) did not perform the speech identification in noise tasks. Due to technical difficulties, two children (1 TH and 1 HL) did not perform the speech identification in a fluctuating noise task. Two children with HL did not perform the speech discrimination in quiet task. With respect to the EEG measurement, one child (HL) was only tested in the amplified, but not the unamplified EEG condition. The sampling frequency of the EEG

recording was accidentally set to 2048 Hz for another child (HL). For this child, we could obtain

the cortical measures, but not the subcortical measures. In all these cases, results were treated

as missing data.

Table 1 : Mean (SD) participant characteristics for the HL and TH groups. Comparisons are
independent-samples t-tests. Effect sizes are Cohen's d. Significant effects are shown in boldface.

Variable	TH (n = 15)	HL (n = 19)	t(df)	p-val	Effect size	95 % CI
Age (years)	12.6 (2.43)	12.4 (2.44)	0.26 (32.1)	.795	0.08	[-1.46, 1.89]
Nonverbal IQ	119.1 (12.6)	102.5 (12.5)	3.83 (31.44)	**<.001**	1.32	[-25.4, -7.7]
Maternal education	20.9 (2.59)	18.3 (2.78)	2.76 (30.99)	**.009**	0.96	[-4.49, -0.67]
BEPTA threshold (dB HL)	3.7 (2.87)	45.0 (13.30)	-13.18 (19.97)	**<.001**	4.12	[-47.86, -34.78]
Age of HL detection (years)	-	3.6 (3.03)				

Data acquisition and pre-processing

Stimuli were digitised /bɑ/ (standard) and /dɑ/ (deviant) syllables (see Figure 2)

originally spoken by a native British female speaker, and identical to those used in[21, 49, 71]. The

intonation contours were made monotone at 220 Hz using Praat[72] and the final stimuli were

RMS equalised with GoldWave[73]. Vowel formant frequencies were approximately 800, 1340

and 2700 Hz for formants F_1, F_2 and F_3 respectively, although there were small differences

between the two syllables as the stimuli were based on natural utterances. Both syllables had

rising F_1s over ~50 ms with a much larger transition for the /dɑ/ (~230 Hz) than the /bɑ/ (~115

Hz). The phonetic contrast was cued primarily by differences in the properties of the initial

release burst (more intense for the /dɑ/, especially at higher frequencies) and on the formant

transitions into the vowel. No F_2 transition was present for the /bɑ/ but the /dɑ/ had a significant

falling transition for F_2 of ~350 Hz.

Each condition consisted of a total of 1320, 175-ms stimuli, presented in a passive

oddball paradigm (deviant probability: 10%). Half of all stimuli were presented in positive

polarity and the other half in negative polarity. The order of presentation of each polarity was

randomized. The inter-stimulus interval was jittered randomly between 275 and 375 ms to limit

neural adaptation[74].

Figure 2: Top row: Spectrograms of the standard and deviant speech syllables in their unamplified form. Bottom
row: Spectra of the vocalic portion (20 to 170 ms) of the standard syllable /bɑ/ in both unamplified and NAL-R
amplified versions. Children with TH were only presented with unamplified versions of the stimuli. Children with
HL were presented with both versions of the stimuli, with the amplification being dependent upon each
individual's audiogram. For visualisation purposes, the amplified stimulus here was generated based on the
average audiogram of the HL group. It can clearly be seen that, as expected, the level of the harmonics below ~2
kHz is unchanged because there is minimal hearing loss at those frequencies, with amplification of the higher
harmonics in the frequency region in which there is substantial HL.

Children with TH were presented with only one condition, consisting of unamplified

stimuli that were presented binaurally at 70 dB SPL through shielded insert earphones (ER-2,

Etymotic Research). Children with HL undertook two conditions - unamplified and amplified

– presented in counter-balanced order. Amplification used the National Acoustic Laboratory-

Revised algorithm (NAL-R[75]) to provide a frequency-specific gain (without compression),

based on each individual child's audiogram. This strategy minimizes the risk of potential

confounds related to sound-field testing as well as potential inter-individual variability in the hearing aid processors of individual children[76]. Stimuli in the amplified condition were limited in intensity such that they did not exceed 85 dB SPL. Each condition lasted approximately 20 minutes, with three short breaks offered every 5 minutes. During the recording, participants were seated comfortably in a shielded, sound-attenuated booth, and watched a silent movie.

EEG was recorded using a BioSemi ActiveTwo system at a sampling rate of 8192 Hz, from 32 scalp electrodes in the standard 10/20 configuration[77]. Additional electrodes were placed on each mastoid and recordings were re-referenced offline to the average of activity at the mastoid electrodes. Electrode offsets were consistently < 50 mV.

Subcortical response analysis

Subcortical response analyses were performed only on the standard (/bɑ/), to avoid the confound of small spectral differences between the standard and the deviant syllables. 1188 epochs (standard probability: 90%) were obtained by applying a band-pass filter from 90 to 1600 Hz (zero-phase, finite impulse response, -6dB/octave) to the EEG data evoked by the standard syllables (/bɑ/) at Cz, and extracting data 20-170 ms relative to stimulus onset, which covers the duration of the steady portion of the vowel. Trials exceeding ± 100 µV were rejected before averaging. There was no significant difference in the proportion of trials that were rejected across groups [TH-U: 18.5%; HL-U: 19.1%; HL-A: 20.3%; $F(2, 47) = 0.723$, $p = 0.491$, $\eta^2 = 0.03$]. The averaged responses to positive and negative polarities were then added together to enhance the envelope of the response (FFR_{ENV}), and subtracted from each other (i.e. positive minus negative polarity) to enhance the TFS of the response (FFR_{TFS}). Spectral amplitudes were calculated using a fast Fourier transform (FFT), after applying zero-padding to the sampling rate to obtain an FFT with a resolution of 1 Hz. Spectral peaks at F0 (220 Hz) and harmonics (H2: 440 Hz; H3: 660 Hz; H4: 880 Hz; H5: 1120 Hz; and H6: 1320 Hz) were computed as the maximum amplitude within an 8-Hz bin centred on F0 and subsequent harmonics, following

visual inspection of the grand average subcortical response, and in line with previous studies (e.g. [78]). Neural background noise amplitudes were calculated by taking the mean amplitude of the fast Fourier transform across 100-Hz windows surrounding each spectral peak (50 Hz on each side, excluding the ±70 Hz adjacent bins).

Cortical response analysis

Evoked potentials considered to be cortical in origin were obtained by band-pass filtering the EEG recorded at electrode Fz between 0.5 Hz and 35 Hz (zero-phase, finite impulse response, -6dB/octave) and creating epochs from -100 ms to 500 ms relative to each stimulus onset time. 1320 epochs (1188 standards and 132 deviants) were baseline corrected using the mean value from -100 to 0 ms. Trials exceeding ± 100 µV were rejected before averaging. There was no significant difference in the proportion of trials that were rejected across groups [TH-U: 12.7%; HL-U: 13.4%; HL-A: 15.75%; $F(2, 50) = 0.43$, $p = 0.654$, $\eta^2 = 0.01$].

P1 and N2 grand average latency were quantified by means of the half-area latency measure of the standard waveform recorded in the TH group[79], a computation that is less sensitive to high-frequency noise than traditional latency peak measures[80, 81]. The window for computation of the area under the curve of the P1 ranged from 55 to 135 ms and from 250 to 350 ms for the N2, based on visual inspection of the TH grand average. P1 and N2 amplitudes were then computed for each participant and condition as the mean amplitude within a 40-ms window centred around the half-area latency (P1: 96 ms post-stimulus onset; N2: 329 ms post-stimulus onset). P1 and N2 latencies were computed for each participant and condition as the half-area latency for the local peak to appear, within the 40-ms window centred around the TH grand average evoked by standards. Note that N1 and P2 were not analysed because they could not be reliably identified in the younger participants, which is likely due to a protracted maturation of these responses until adolescence[37, 82].

The MMN was computed as the differential wave obtained by subtracting the neural response evoked by the standard to that evoked by the deviant, for each participant and condition. The MMN was then statistically assessed in two ways. First, to determine whether there was evidence for neural discrimination of speech deviants, the presence of an MMN was evaluated for the three combinations of group and amplification conditions. Point-to-point comparison of the differential wave amplitudes was performed to determine the latency period over which the waveforms were significantly smaller than zero, if any. One-sided *t*-tests were computed within the 100-500 ms post-stimulus-onset time window which was identified as the region most likely to contain the MMN by visual inspection of the grand average waveforms. Because adjacent points in the waveform are highly correlated, potentially leading to spurious significant values in short intervals, an MMN was considered present when $p < .01$ (one-tailed) for more than 20 ms at adjacent time-points[83, 84]. Half-area MMN latency was computed on the differential waveform recorded in the TH group. MMN amplitude for each participant and condition were then computed as the mean amplitude in a 100-ms window centred around the grand averaged differential waveform half-area latency peak observed in children with TH (at 426 ms post-stimulus onset[79]. MMN individual latencies were computed as the half-area latency within a 100-ms window centred around the grand average differential waveform of the children with TH.

Speech discrimination in quiet

The speech discrimination in quiet task was the same as that used in previous studies from our laboratory[21, 85]. The two endpoints of the continuum were based on the same digitised /bɑ/ and /dɑ/ used in the electrophysiological recordings. A continuum of 100 stimuli was then constructed (including the endpoints), using the morphing capabilities of the programme STRAIGHT[86]. The task was delivered via a child-friendly computer game, incorporating an adaptive, three-interval, three-alternative, forced-choice paradigm. Stimuli were presented

binaurally via headphones (Sennheiser 25 HD) at a fixed intensity (70 dB SPL), on a tablet, meaning identical levels presented in the two groups. Children were required to detect the odd-one-out, where two of the intervals corresponded to the /bɑ/ end of the continuum, and the third, target stimulus fell somewhere along the rest of the continuum. Participants received visual feedback regarding the accuracy of their responses. A three-down, one-up procedure was used to select, on each trial, the appropriate target sound from a continuum, tracking a performance level of 79.4% correct[87]. This was preceded by an initial one-down, one-up procedure until the first reversal[88]. The initial step size was 15 stimulus places along the continuum, which reduced to five after the first reversal. The threshold was the arithmetic mean of the last four reversals in direction of the adaptive track, expressed as the stimulus number (0-99) of the target stimulus along the continuum. Therefore, smaller numbers indicate better performance. The task was preceded by five practice trials which contained standard-target difference that had previously been deemed suprathreshold for adults with TH[21].

Speech perception in noise

The rationale for choosing the stimuli for this task was to provide the most direct comparison to the neurophysiological stimuli, which were speech syllables. Another advantage of syllables is that they minimise the role of linguistic and cognitive factors on auditory performance, allowing to focus more on lower level auditory properties. Speech reception thresholds (SRTs) were thus measured adaptively for 13 consonants presented in vowel-consonant-vowel (VCV) logatomes, e.g. /imi/, /apa/. Vowels were /i/, /ɑ/, and /u/, and consonants were /p/, /k/, /b/, /d/, /g/, /f/, /m/, /n/, /l/, /v/, /w/, /y/, and /z/. Both the vowels and consonants varied from trial to trial, but only the response for the consonant was scored. Stimuli were recorded by a native British English female speaker in an anechoic chamber and were digitized via a 16-bit analogue-to-digital converter at a 44.1-kHz sampling frequency. The

logatomes were presented together with two types of background noise: steady (i.e. unmodulated, but see [89]) or fluctuating in amplitude sinusoidally at 8 Hz, which corresponds to a syllabic modulation rate[90]. The modulation depth was fixed at 1.0 and the starting phase was randomized between 0° and 360° on each trial. The noise was shaped on-and-off using a raised-cosine with 50-ms rise-fall times.

Participants were seated in a quiet room and the stimuli were presented binaurally over Etymotics ER-2 earphones at an overall level of 70 dB SPL, again meaning identical levels presented in the two groups. They were asked to repeat the logatomes as best as they could. The experimenter scored the participants' responses using a graphical interface which showed the 13 possible consonants. No feedback was provided.

The expected wide variability in intelligibility across consonants at a given SNR would not allow their reliable use in a simple adaptive procedure. Therefore, SRTs for each combination of consonant and vowel were estimated from a prior study of young adults with TH (unpublished), allowing the calculation of adjustment factors in SNR to give all stimuli the same nominal SRT[91]. These factors were then used during the adaptive procedure to adjust the presented SNR. The signal-to-noise ratio (SNR) was varied adaptively by varying the level of the target and fixing the level of the noise at 70 dB SPL. The first logatome was presented at a nominal SNR of -10 dB (taking into account that token's adjustment factor). Depending on the participant's response (correct/incorrect), the SNR of the second logatome was decreased/increased by 10 dB. Again, depending on the participant's response (correct/incorrect), the SNR of the third logatome was decreased/increased by 6 dB. For each subsequent logatome, the SNR increased/decreased by 2 dB for incorrect/correct responses, respectively. Measurement stopped after either 12 reversals or 78 trials, whichever occurred first. The SRT was computed by taking the mean SNR (dB) across all track reversals.

Speech perception in quiet

As a baseline and familiarisation for the speech perception in noise task, we also included a speech perception in quiet task. Percentage correct responses were measured for 78 consonants presented at a fixed intensity of 70 dB SPL, using the same VCV logatomes and procedure for recording the participants' responses as those used in the speech perception in noise task.

Statistical analyses

Regarding the subcortical data, we first ran an outlier detection algorithm. Data points that fell outside the mean ± 3 *SD* (within each combination of group, amplification, FFR component and harmonic) were considered outliers and excluded from further analyses. In total, 2.7% of data points were excluded from the analyses, most of them from the HL group. After excluding outliers, the subcortical data were normally distributed (Kolmogorov-Smirnov test: all ps < .05). Note that for both FFR_{ENV} and FFR_{TFS}, there were significant differences in variance between groups as evidenced by Levene's test for equality of variance (both ps < 0.001).

Second, paired two-tailed *t*-tests with Bonferroni-Holm correction were conducted separately for each group (TH_U, HL_U, HL_A), FFR component (FFR_{ENV} and FFR_{TFS}) and harmonic (F0 to H6) to determine whether the FFR responses were significantly above the neural noise floor. FFR responses are considered present if at least one of the three groups showed a significant difference between target frequency peak and spectral noise floor in each component, at each harmonic. For FFR_{ENV}, peaks were significantly larger than the noise floor only at F0 and H2 (all ps < 0.001). For FFR_{TFS}, peaks were significantly larger than the noise floor at H3, H4, H5 and H6 (all ps < 0.05).

Third, a linear mixed model was used to determine whether amplitude of the spectral noise floor differed across groups (TH_U, HL_U, HL_A), FFR component (FFR_{ENV} and FFR_{TFS}) or

harmonic (F0 to H6). As expected, the only significant predictor was harmonic ($p < 0.001$). In the absence of a significant group effect on the baseline ($p > 0.10$), subsequent analyses were performed on the target frequency peak amplitude.

Regarding the cortical data, we found no outliers ($\pm$ 3 SD from the mean) in the amplitude of either P1 or MMN. The cortical data were normally distributed (Kolmogorov-Smirnov test: all ps < .05). For both P1 and MMN amplitudes, there were no significant differences in variances between groups (Levene's test for equality of variance: both ps > .05).

Linear mixed models (lmerTest package of R^{92}) were used to evaluate group differences in both cortical (P1, N2 and MMN) and subcortical (FFR_{ENV} and FFR_{TFS}) measures. Fixed effect predictors and all their interactions (all orders) were included in the initial model. In all models, the factor listener was used as a random intercept, and random slopes for all the appropriate predictors (harmonic and amplification) that did not lead to convergence issues or inestimable models. Backward stepwise reduction was then used to simplify the models based on F-tests, eliminating insignificant terms, but always keeping lower-order terms that were significant in higher-order interactions. This was done first for the random effects, and then the fixed effects. Only significant results are reported below. Analyses of group differences aimed to answer three main questions.

- First, is there a significant group difference in electrophysiological responses to unamplified speech stimuli? Here, the predictors were group (TH_U vs. HL_U) and age (continuous variable), with harmonic as an additional predictor (FFR_{ENV}: F0 and H2; FFR_{TFS}: H3 to H6) to model the subcortical data.

- Second, for children with HL only, is there a benefit of amplification on the neural encoding of sounds? Here, the predictors were amplification (HL_U vs. HL_A), BEPTA and age, again with harmonic as an additional predictor for the subcortical data.

- Third, does sound amplification in children with HL lead to electrophysiological

responses that are similar to the control group (TH_U vs. HL_A)? Again, the predictors

were group (TH_U vs. HL_U) and age, with harmonic as an additional predictor for the

subcortical data.

With respect to the speech intelligibility measure in quiet, whose outcome variables was

binomial, generalized linear mixed models (logistic regressions) were fitted to the data. Group

(TH, HL) and age, as well as their two-way interaction were included in the model as between-

subjects factors, with both listener and consonant included as random effects. For speech

perception in noise, a linear mixed model was computed with group (TH, HL), noise type

(steady, fluctuating), age, and all their interactions as fixed factors, and listener as a random

effect. Backward stepwise reduction was then applied, as for the EEG measurements.

Finally, the relationship between subcortical and cortical measures, as well as that

between electrophysiological and behavioural measures were investigated using correlations,

conducted separately on the TH and HL groups, and separately for unamplified and amplified

conditions in the latter group.

Table 2: Summary of the main effects of hearing loss (HL_U versus TH_U) and amplification
(HL_A versus HL_U) for subcortical, cortical and behavioural measures. * $p < 0.05$, ** $p < 0.01$,
*** $p < 0.001$.

		Effect of hearing loss (HL_U versus TH_U)	**Benefit of amplification** (HL_U versus HL_A)	**Benefit of amplification** (HL_A versus TH_U)
Subcortical measures				
FFR_{ENV}	Amplitude	$HL_U < TH_U$ ***	$HL_U < HL_A$*	
FFR_{TFS}	Amplitude	$HL_U < TH_U$ *	Amplification × Harmonic ***	Amplification × Harmonic ***
Cortical measures				
P1	Amplitude	$HL_U < TH_U$ **	Amplification × BEPTA *	
	Latency	TH_U earlier than HL_U *	HL_A earlier than HL_U*	
N2	Amplitude	$HL_U < TH_U$ ***		$HL_A < TH_U$ **
	Latency		Amplification × BEPTA *	
MMN	Amplitude	$HL_U < TH_U$ **		$HL_A < TH_U$ **
	Latency			

Results

A summary of the main effects of hearing loss (HL_U versus TH_U) and of amplification (HL_A versus HL_U) is provided in Table 2, for subcortical, cortical and behavioural measures.

1. Subcortical responses

Figure 3 shows the grand-average time waveforms and spectra of the subcortical responses evoked at electrode Cz by unamplified speech for children with TH (left panels), and unamplified and amplified speech for children with HL (middle and right panels, respectively). Grand-averages were derived from the time waveforms and spectra of the individual waveforms. Scatterplots of the amplitude of the various components are shown as a function of age in Figure 4.

First, group differences in the subcortical processing of unamplified speech were examined (TH_U vs. HL_U; see Supplementary Table 1). FFR_{ENV} were significantly smaller (by 0.082 μV) in children with HL_U than in children with TH_U ($p = .001$) for both harmonics, with no effect of age. Similarly, FFR_{TFS} were significantly smaller (by 0.028 μV) in children with HL_U than in children with TH_U ($p = .015$). Note that in neither analysis was there a significant group x harmonic interaction.

Next, the effects of amplification were examined by comparing the subcortical responses of children with HL presented with unamplified versus amplified speech stimuli (HL_U vs. HL_A; see Supplementary Table 2). Overall, FFR_{ENV} were significantly larger (by 0.118 μV) in response to amplified compared to unamplified speech sounds ($p = 0.011$). The amplification × age interaction got excluded during model reduction ($p = 0.051$), yet we observed a marginally significant decreasing benefit of amplification with age on FFR_{ENV} amplitude ($p = 0.019$). With respect to the FFR_{TFS}, we observed a significant amplification × harmonic

interaction. FFR_{TFS} were significantly larger in response to amplified than unamplified speech sounds at H4, H5 and H6, but not at H3.

Last, we compared the amplitude of the responses obtained when amplified sounds were presented to children with HL versus unamplified sounds presented to children with TH (HL_A vs. TH_U; see Supplementary Table 3). The amplitude of the FFR_{ENV} did not significantly differ between HL_A and TH_U ($p = 0.168$). With respect to the FFR_{TFS}, we observed a group × harmonic interaction ($p < 0.001$). FFR_{TFS} were significantly larger in the HL_A than TH_U groups at both H4 and H6, but not at H3 and H5.

Figure 4: Scatterplots of the amplitude (µV) of the FFR$_{ENV}$ (F0 and H2) and FFR$_{TFS}$ (H3 to H6) over age, for each
group (TH$_U$, HL$_U$ and HL$_A$). Regression lines are fit on the basis of each combination of group, FFR component
and harmonic.

2. Cortical responses

Figure 5: Grand average waveforms at Fz, evoked by the standard (lighter line) and deviant (darker line)
presented either unamplified to both children with TH$_U$ (70 dB SPL, left column) and children with HL$_U$ (70
dB SPL, middle column) or amplified to children with HL$_A$ (right column). Voltage maps show the mean
MMN activity during the 100-500 ms post-stimulus time window. Negative values of the MMN are shown in
blue, and positive values in red. The horizontal line represents the duration of a significant MMN in the period
of the 100-500 ms post-stimulus-onset epoch evoked in children with TH. No significant MMNs were observed
in either the unamplified or amplified conditions for the children with HL as a group.

First, we assessed group differences in the cortical processing of unamplified speech (TH$_U$ vs. HL$_U$; see Supplementary Table 4). As shown in Figure 6, P1 was smaller (by 2.05 µV) and later (by 2.79 ms) in the HL$_U$ than the TH$_U$ group (both ps < 0.05). N2 was also smaller (by 3.79 µV; p < 0.001) but not later in the HL$_U$ than the TH$_U$ group. Additionally, children with HL$_U$ had significantly smaller MMN responses (by 3.47 µV) than children with TH$_U$ (p = 0.003).

Next, we sought to determine whether amplification benefited the cortical processing of speech sounds in children with HL (HL$_U$ vs. HL$_A$; see Supplementary Table 5). For P1 amplitude, we found a significant amplification × BEPTA interaction (p = 0.028; see Supplementary Figure 1). Perhaps not surprisingly, the P1 amplitude in children with better BEPTA thresholds was not changed by the relatively small amplification they received, whereas the P1 amplitude for children with poorer BEPTA thresholds (and who received greater degrees of amplification) was increased so as to make P1 amplitude more-or-less independent of BEPTA. P1 latency was longer in the unamplified than the amplified condition (p = 0.027). It was also longer for individuals with higher BEPTA thresholds (p = 0.024) and for younger children (p = 0.005), irrespective of the condition for both. For N2 latency, there was a significant main effect of amplification (p = 0.033). Even though the amplification × BEPTA interaction was also significant (p = 0.030), post-hoc contrasts were not significant (p = 0.842). There was no significant effect of amplification on MMN amplitude or latency (both ps > 0.10). Note that irrespective of the condition, MMN amplitude decreased with age in children with HL (p = 0.040).

Last, we evaluated the potential benefit of amplification for cortical processing of speech in children with HL to that of children with TH (HL$_A$ vs. TH$_U$; see Supplementary Table 6). There was no significant effect of group on P1 amplitude or latency (both ps > 0.10). Yet there remained a significant group difference when looking at N2. Despite amplification, N2

amplitude was smaller (by 2.43 µV; *p* = 0.004), but not later, in the group with HL_A than the

group with TH_U. Similarly, children with HL_A showed significantly smaller MMN than the

group with TH_U (*p* = 0.002). Note that, although the group × age interaction did not come out

significant (*p* = 0.146), post-hoc comparisons indicate a significant decrease of MMN

amplitude with age in HL_A (*p* = 0.038) but not in TH_U (*p* > 0.10).

Supplementary Figure 1: Amplitude (µV) of the P1 for the HL group as a function of BEPTA. Individual (shapes)
and group (lines) data are shown for both unamplified (HL_U, plain lines) and amplified (HL_A, dotted lines) raw.
Shaded lines represent the 95% confidence intervals.

Figure 6: Amplitude (µV; left column) and latency (ms; right column) of the P1, N2 and MMN for each group
and condition. Individual (shapes) and group (colour) data are shown for both unamplified (HL_U, orange) and
amplified (HL_A, red) data. Shaded lines represent the 95% confidence intervals. Latencies (right column) are
illustrated as group boxplots because there was no effect of age.

3. Behavioural data

3. 1. Speech discrimination in quiet

Figure 7 shows the behavioural performance of the two groups on the speech perception

tasks. For speech discrimination of the speech sounds /bɑ/ vs /dɑ/ in quiet (Figure 7A), there

was no significant effect of group, age, or their interaction on response thresholds (all ps > .10;

see Supplementary Table 7A).

Figure 7: Boxplots of (A) stimulus distance (B) percentage correct consonant identification in quiet and (C) SRTs
for consonant identification in steady (light grey) and fluctuating (dark grey) background noises. Children with
TH are shown in blue, HL children are shown in orange. The boxes show the interquartile range (25th-75th
percentile) of the data with horizontal line indicating the median. The whiskers indicate values that fall within 1.5
times the interquartile range.

596
3. 2. Speech identification in quiet and noise

Figure 7B presents the percentage correct consonant identification in quiet for the two

groups (also see Supplementary Table 7B). Children with HL were significantly poorer than

children with TH ($p < 0.001$). Note that for both groups, performance significantly improved

with age ($p = .003$), although ceiling effects meant only a very small increase in percentage

correct for the TH group.

Finally, Figure 7C presents the SRTs in steady and fluctuating background noise for the

two groups (also see Supplementary Table 7C). The significant main effects of group and noise

type (both ps < .001) need to be considered in light of the significant group × noise type

interaction (p = .006). As expected, both groups experienced masking release when they were

presented with fluctuating noise (both ps < 0.001). However, the magnitude of this effect was

larger in children with TH (8.5 dB SNR) than in children with HL (3.9 dB SNR).

4. Relationship between the neural and behavioural responses

The relationship between electrophysiological activity and behavioural responses was

investigated by means of a set of correlations. Holm-Bonferroni correction was applied to

correct for multiple comparisons where appropriate.

Given the evidence that perception of speech in quiet is dominated by the encoding of

the envelope of the signal, we investigated the relationship between the amplitude of the

FFR_{ENV} at both F0 and H2 and behavioural performance of speech identification in quiet,

separately for children with TH_U and children with HL_U. None of the correlations were

significant (all ps > 0.05).

Adequate perception of speech in noise requires precise spectro-temporal processing of

speech, particularly in the dips of a fluctuating background noise. In fact, the amplitude of

speech-evoked FFR in quiet relates to the perception of speech in noise (see introduction).

Therefore, we investigated the relationship between amplitude of the speech-evoked FFR_{TFS} at

H4 and H6 (in quiet) and behavioural speech perception in steady and fluctuating noise, as well

as masking release, separately for children with TH and children with HL (in both unamplified

and amplified conditions). None of the correlations were significant (all ps > 0.05).

The speech stimuli used for the electrophysiological recordings were the endpoints /bɑ/

and /dɑ/ of the continuum used for the speech discrimination in quiet behavioural task. As such,

we hypothesised that a relationship might be observed between the amplitude of the MMN and the stimulus distance threshold measured in the speech discrimination in quiet behavioural task. However, there was no significant correlation between speech discrimination in quiet and MMN amplitude, as evaluated separately for both groups, and separately for the amplified and unamplified conditions in children with HL (all ps > 0.05).

Last, we investigated the relationship between subcortical and cortical responses, separately for both groups (and separately for HL_A and HL_U). In children with TH, larger FFR_{ENV} at F0 were associated with smaller N2 responses (p = 0.004). There was no other significant correlation between FFR_{ENV} at F0 and P1, N2 or MMN (all ps > 0.05).

Discussion

This study investigated the effect of childhood MM HL and the benefit of amplification on both subcortical and cortical processing of speech sounds. When sounds were presented at the same intensity across groups, both subcortical and cortical responses were smaller in children with HL than those with TH. When sounds were amplified, the subcortical responses increased in children with HL_A compared to HL_U. In fact, they could even be larger in children with HL_A than TH_U (although only significantly so for the FFR_{TFS}). At the cortical level, an amplification benefit was also observed on P1 amplitude but not on the amplitude of later responses (N2 or MMN). In fact, at the group level, children with HL did not show a significant MMN (unlike children with TH), whether the sounds were amplified or not. When looking at individual responses, the amplitude of the MMN tended to decrease with age in children with HL but not in those with TH. Behaviourally, children with HL had poorer speech intelligibility than those with TH both in quiet and in noise, and they showed less benefit from background noise fluctuations. Yet, the relationship between the neurophysiological responses and the behavioural performance remains unclear.

Subcortical processing

When presented with unamplified sounds, children with MM HL showed significantly smaller FFR_{ENV} and FFR_{TFS} than those with TH. This finding replicates and extends previous results from older adults with age-related HL[45,46] – but contrasts with some studies that did not find group differences on unamplified FFR measures [47,48]. The etiology of the HL might thus modulate its effect on subcortical processing of unamplified sounds. When presented with amplified sounds, children with MM HL showed a clear benefit from the frequency-specific gain applied to the sounds. In fact, the amplified FFR_{ENV} of children with MM HL did not significantly differ from that of children with TH_U. Furthermore, the amplified FFR_{TFS} was even larger than that of children with TH_U. Our results thus fit with existing results from the adult and child literature to indicate a clear benefit of amplification at the subcortical level[47,51-53,93,94].

We were initially surprised by the spectacular increase in amplitude of the FFR_{TFS} in the higher harmonics following amplification. Indeed, the amplitude of neural spectral peaks typically *decreases* with increasing frequency (e.g. [47,95,96]). We believe this increase stems from applying the individualized, frequency-specific NAL gains calculated from audiograms with sloping hearing losses, which leads to increasing gains for higher frequency harmonics. To our knowledge, the only other study that used the NAL formula analysed FFR_{TFS} response up to ~700 Hz. In this frequency range, there was no significant difference in the FFR_{TFS} of older adults with HL (in quiet, with or without amplification) and age-matched adults with TH[47] . In our study, FFR_{TFS} was only above noise floor at frequencies at H3 (660 Hz) and above. In fact, our results show significantly larger FFR_{TFS} in HL_A compared to both HL_U and TH_U at H4 and H6, which are close to the first and second formant peaks of our stimulus. In adults with TH, FFR_{TFS} are most clearly detected at harmonics that are close to formant peaks, up to 1500 Hz[24]. Large FFR_{TFS} at H4 and H6 might therefore reflect the interaction between frequency-specific

NAL amplification and the formants of our speech stimuli. Yet a *larger* FFR_{TFS} in children with

HL_A than those with TH_U might indicate hyperactivity at the subcortical level following

childhood HL.

Hyperactivity has been widely reported as a consequence of SNHL (for a review, see

[97]), and is thought to compensate for reduced inputs from damaged peripheral structures to

central brain regions, to maintain sensation[98]. Such compensation fits within the theoretical

framework of homeostatic plasticity, which states that neurons regulate their physiological

properties to compensate for persistent changes in their activity level[99]. Homeostatic

mechanisms are thought to operate from synaptic to network levels of sensory systems. That

FFR amplitude was larger in children with HL than those with TH, but only when sounds were

amplified suggests that compensatory activity is intensity-dependent at the brainstem level. In

mice, changes in auditory cortex responsiveness following conductive HL were only observed

in sounds above a certain threshold, or in certain frequency ranges[100]. Future studies are

warranted to investigate the effect of audibility on homeostatic plasticity of subcortical

processing.

Cortical processing

Despite the medium to large benefit of amplification on subcortical responses, there was

only a small benefit of amplification on cortical responses (P1), or no benefit at all (N2, MMN).

Individualized frequency-specific gain increased P1 amplitude (which reflects improved sound

detectability, [31]), but only for children with poorer BEPTA. Irrespective of BEPTA, P1 was

earlier in HL_A than HL_U. In fact, the P1 amplitude and latency of HL_A did not significantly

differ from that of children with TH_U. This is in line with studies that show age-appropriate P1

when sounds are presented at similar sensation level to children with HL and those with TH [54–

56].

Despite preserved P1, our results indicate that childhood HL leads to smaller (but not later) N2 even when sounds are amplified, in line with [55], but contrary to [54, 56]. Furthermore, we did not observe a significant MMN at the group level on children with HL, even when sounds were amplified. At the individual level, the MMN amplitude of children with HL_A remained smaller than those with TH_U. Note however that there was no group difference in MMN latency. Our finding of absent/smaller MMNs in 8 to 16 year-olds with HL contrast with those of age-appropriate MMNs in 5-12 year-old children[55,56]. Part of the explanation for this discrepancy might lie in the developmental effects of childhood HL on the central auditory pathway (see below for a discussion of the developmental effects observed in this study).

Why does the P1 amplitude benefit from amplification, but not the N2 or MMN? One possible explanation might lie in the different generators of the P1, N2 and MMN. The P1 generator has been reliably located within the secondary auditory cortex[101,102], whereas MMN generators likely include an extended thalamocortical circuitry projecting to primary auditory cortex[103]. Little is known about the generators of the N2, but early intracranial recordings also seem to point towards a subcortical (thalamic) source[104]. Although anatomically close, the primary and secondary auditory cortices receive input from distinct portions of the medial geniculate body. Whereas the lemniscal pathway connects the ventral medial geniculate body to the primary auditory cortex, the nonlemniscal pathway connects the medial/dorsal geniculate body to both primary and secondary auditory cortices[105]. Given that our results indicate smaller N2 and MMN even when children with HL were presented with amplified sounds, these could indicate a specific alteration of the nonlemniscal pathway connecting the medial portion of the geniculate body to the primary auditory cortex. Our cortical findings thus contrast with the subcortical evidence of hyperactivity following childhood HL.

The effect of HL on the development of the central auditory pathway

In fact, our cortical findings also contrast with findings of hyperactivity in the auditory cortex as a consequence of age-related HL. Indeed, studies have reported some degree of enhancement of the cortical responses to sounds in older adults with age-related HL compared to age-matched individuals with TH[106-108]. To our knowledge, none of the pediatric studies investigating cortical processing of (amplified or unamplified) sounds in children with HL found such indications of increased cortical responsiveness[49,50,54-56,109-111]. In mice, the same population of neurons showed distinct mechanisms of homeostatic adaptation if visual deprivation occurred in juvenile or adult animals[112]. Such an age-dependent effect of homeostatic plasticity might also be observed in the central auditory pathway following auditory deprivation.

Noteworthily, these age-dependent effects might already be operating between 8 and 16 years of age. Indeed, we observed a significant decrease in the benefit of amplification with age on FFR_{ENV} amplitude. Additionally, the amplitude of the MMN decreased with age in children with HL_A (but not in those with TH_U) – although note that the group × age interaction was not significant. Both results seem to indicate that adolescence could be a period of heightened plasticity for functional changes following auditory deprivation. Future studies are warranted to investigate this claim in a larger sample size, ideally taking puberty into account, as it could act as a biological trigger to those alterations[113-115].

Multiple gain mechanisms within the auditory system

To our knowledge, only three studies so far have combined subcortical and cortical measures from the same individuals to investigate the effects of HL on auditory processing. Speech-evoked subcortical and cortical responses were recorded successively in older adults, with and without hearing aids. When sounds were presented at 65 dB SPL, hearing aid

amplification lead to significantly larger subcortical, but not cortical responses[93]. In gerbils with developmental conductive HL, age-appropriate subcortical, but not cortical responses were recorded to amplitude modulated sounds presented at similar sensation level as in animals with TH[116]. More recently, Hutchison et al., (2023)[117] recorded subcortical and cortical neurophysiological responses in young adults with TH before, throughout and after fitting them with earplugs. The results indicate an increase in the subcortical response, but a reduction of cortical activity ipsilateral to the deprived ear. Together with our observations, the literature suggests that multiple gain mechanisms might underlie homeostatic plasticity throughout the auditory pathway[118]. Neurons from each level of the auditory pathway are thought to display idiosyncratic changes following HL (for a review, see [119]).

Relationship between speech processing & perception

An exploratory objective of our study was to investigate how changes in neurophysiological responses following HL might relate to speech intelligibility in quiet and in noise. To do so, we chose stimuli that were as similar as possible for both neurophysiological and behavioural tasks. However, two methodological choices are worth mentioning here: Unlike the neurophysiological task, stimuli were only presented unamplified in the behavioural tasks. Additionally, syllables were only presented in quiet in the neurophysiological tasks; yet speech perception was investigated in quiet and in noise.

Behaviourally, children/adolescents with HL have higher (i.e. poorer) thresholds in noise than children/adolescents with TH [19,20]. Their thresholds improved in the presence of fluctuating compared to steady background noise, but to a lesser extent than the group with TH. Similarly to adults with HL[15–17], children with MM HL seem less able to listen in the "valleys" of a fluctuating background noise to improve their speech perception. However, we did not find any significant relationship between measures of speech processing and perception. This

contrasts with numerous studies that report small to medium correlations between subcortical responses and speech intelligibility, especially in noise[57-61, 63, 64]. Some correlations have also been observed between cortical responses and speech discrimination[65, 66], although the nature of this relationship remains unclear[67-69]. Again, our sample size might be too small to adequately investigate such possible correlations. Future studies including larger sample sizes are warranted to clarify the relationship between speech processing and perception in the context of congenital HL. These studies would benefit from a direct investigation of the effect of childhood HL on subcortical processing of speech in noise and its relationship with performance.

Conclusions

When presented with unamplified sounds, children with HL show impaired subcortical and cortical responses to speech syllables. Amplification was beneficial for subcortical and early (P1) but not late (N2, MMN) cortical responses. Our results support the existence of multiple gain mechanisms that could underlie homeostatic plasticity throughout the auditory pathway. Responses elicited at different levels of the auditory pathway appear to display idiosyncratic changes following HL, and may be age- and level- dependent. Even though speech perception was poorer in the group with HL than TH (in quiet and in noise), there was no clear relationship between the neurophysiological and behavioural measures. Even mild to moderate congenital HL induces functional alterations to the central auditory pathway, each level showing a specific alteration of its typical response. Future studies are needed to clarify the relationship between these functional changes and the behavioural performance.

Acknowledgments

We are very grateful for the time and efforts of the children who participated in this study and to their parents. We would like to thank Dr Lorna Halliday for her input throughout this project, and for her comments on previous versions of this manuscript. We would also like to thank Gaston Hilkhuysen for his help with aspects of the methods. We thank two anonymous reviewers for their constructive feedback on an earlier version of this manuscript. This work was supported by the FP7 people programme (Marie Curie Actions), grant agreement no FP7-607139 (improving Children's Auditory REhabilitation, iCARE). Axelle Calcus was supported by a MSCA Individual Fellowship (798093, EAR-DNA) and a FNRS MIS (F.4508.22).

Author contributions

A.C. and S.R. conceived the idea, A.C. collected data, A.C. and S.R. analysed the data, A.C. and S.R. interpreted the data, A.C. and S.R. wrote the manuscript.

Data availability

Unidentifiable data, stimuli and statistical analyses scripts are publicly available on: https://github.com/acalcus/MMHL_amplification

Competing interests

The authors declare no competing interests

References

1. Caras, M. L. & Sanes, D. H. Sustained Perceptual Deficits from Transient Sensory Deprivation. *J Neurosci* **35**, 10831–10842 (2015).

2. Mowery, T. M., Kotak, V. C. & Sanes, D. H. Transient Hearing Loss Within a Critical Period Causes Persistent Changes to Cellular Properties in Adult Auditory Cortex. *Cereb Cortex* **25**, 2083–2094 (2015).

3. Daly, D. M., Roeser, R. J. & Moushegian, G. The frequency-following response in subjects with profound unilateral hearing loss. *Electroencephalogr. Clin. Neurophysiol.* **40**, 132–142 (1976).

4. Ponton *et al.* Maturation of the Mismatch Negativity: Effects of Profound Deafness and Cochlear Implant Use. *Audiology Neurotol* **5**, 167–185 (2000).

5. Sharma, A., Kraus, N. & McGee, T. J. Developmental changes in P1 and N1 central auditory responses elicited by consonant-vowel syllables. *Electroencephalogr Clin Neurophysiology Evoked Potentials Sect* **104**, 540–545 (1997).

6. Peelle, J. E. & Wingfield, A. The Neural Consequences of Age-Related Hearing Loss. *Trends Neurosci* 1–12 (2016) doi:10.1016/j.tins.2016.05.001.

7. Lieu, J. E. C., Kenna, M., Anne, S. & Davidson, L. Hearing Loss in Children. *JAMA* **324**, 2195–2205 (2020).

8. Shannon, R. V., Zeng, F.-G., Kamath, V., Wygonski, J. & Ekelid, M. Speech Recognition with Primarily Temporal Cues. *Science* **270**, 303–304 (1995).

9. Qin, M. K. & Oxenham, A. J. Effects of simulated cochlear-implant processing on speech reception in fluctuating maskers. *J Acoust Soc Am* **114**, 446–454 (2003).

10. Zeng, F.-G. *et al.* Speech recognition with amplitude and frequency modulations. *Proc. Natl. Acad. Sci.* **102**, 2293–2298 (2005).

11. Hopkins, K. & Moore, B. C. J. The contribution of temporal fine structure to the intelligibility of speech in steady and modulated noise. *J Acoust Soc Am* **125**, 442–446 (2009).

12. Cooke, M. A glimpsing model of speech perception in noise. *J Acoust Soc Am* **119**, 1562 (2006).

13. Miller, G. A. & Licklider, J. The intelligibility of interrupted speech. *J Acoust Soc Am* **22**, 167–173 (1950).

14. Festen, J. M. & Plomp, R. Effects of fluctuating noise and interfering speech on the speech-reception threshold for impaired and normal hearing. *J Acoust Soc Am* **88**, 1725–1736 (1990).

15. Lorenzi, C., Gilbert, G., Carn, H., Garnier, S. & Moore, B. C. J. Speech perception problems of the hearing impaired reflect inability to use temporal fine structure. *Proc National Acad Sci* **103**, 18866–18869 (2006).

16. Hopkins, K., Moore, B. C. J. & Stone, M. A. Effects of moderate cochlear hearing loss on the ability to benefit from temporal fine structure information in speech. *J Acoust Soc Am* **123**, 1140–1153 (2008).

17. Strelcyk, O. & Dau, T. Relations between frequency selectivity, temporal fine-structure
processing, and speech reception in impaired hearing. *J Acoust Soc Am* **125**, 3328 (2009).

18. Crandell, C. C. Speech Recognition in Noise by Children with Minimal Degrees of Sensorineural
Hearing loss. *Ear Hearing* **14**, 210–216 (1993).

19. Lewis, D. E., Valente, D. L. & Spalding, J. Effect of minimal/mild hearing loss on children's
speech understanding in a simulated classroom. *Ear Hearing* **36**, 136–144 (2015).

20. Goldsworthy, R. L. & Markle, K. L. Pediatric Hearing Loss and Speech Recognition in Quiet and
in Different Types of Background Noise. *J Speech Lang Hear Res* **62**, 758–767 (2019).

21. Halliday, L. F., Rosen, S., Tuomainen, O. & Calcus, A. Impaired frequency selectivity and
sensitivity to temporal fine structure, but not envelope cues, in children with mild-to-moderate
sensorineural hearing loss. *J Acoust Soc Am* **146**, 4299–4314 (2019).

22. Brennan, M. *et al.* Masking Release in Children and Adults With Hearing Loss When Using
Amplification. *J. Speech, Lang., Hear. Res.* **59**, 110–121 (2016).

23. Kraus, N., Anderson, S. & White-Schwoch, T. The Frequency-Following Response: A Window
into Human Communication. in vol. 61 1–15 (Springer Science & Business Media, Cham, 2017).

24. Aiken, S. J. & Picton, T. W. Envelope and spectral frequency-following responses to vowel
sounds. *Hearing Res* **245**, 35–47 (2008).

25. Bidelman, G. M. Subcortical sources dominate the neuroelectric auditory frequency-following
response to speech. *Neuroimage* **175**, 56–69 (2018).

26. Coffey, E. B. J. *et al.* Evolving perspectives on the sources of the frequency-following response.
Nat Commun **10**, 5036 (2019).

27. Hecox, K. & Galambos, R. Brain Stem Auditory Evoked Responses in Human Infants and Adults.
Arch Otolaryngol **99**, 30–33 (1974).

28. Krizman, J. *et al.* Continued maturation of auditory brainstem function during adolescence: A
longitudinal approach. *Clin. Neurophysiol.* **126**, 2348–2355 (2015).

29. Krizman, J., Bonacina, S. & Kraus, N. Sex differences in subcortical auditory processing emerge
across development. *Hear. Res.* **380**, 166–174 (2019).

30. Alain, C. & Tremblay, K. The role of event-related brain potentials in assessing central auditory
processing. *J Am Acad Audiol* **18**, 573–589 (2007).

31. Eggermont, J. J. & Ponton, C. W. The Neurophysiology of Auditory Perception: From Single
Units to Evoked Potentials. *Audiology Neurotol* **7**, 71–99 (2002).

32. Scherg, M., Vajsar, J. & Picton, T. A source analysis of the late human auditory evoked potentials.
Journal of Cognitive Neuroscience (1989).

33. Näätänen, R. *et al.* The mismatch negativity (MMN) - A unique window to disturbed central
auditory processing in ageing and different clinical conditions. *Clin Neurophysiol* **123**, 424–458
(2012).

34. Alho, K. Cerebral Generators of Mismatch Negativity (MMN) and Its Magnetic Counterpart
(MMNm) Elicited by Sound Changes. *Ear Hearing* **16**, 38–51 (1995).

35. Rinne, T., Alho, K., Ilmoniemi, R. J., Virtanen, J. & Näätänen, R. Separate Time Behaviors of the
Temporal and Frontal Mismatch Negativity Sources. *Neuroimage* **12**, 14–19 (2000).

36. Zhang, Y. *et al.* Cortical Areas Associated With Mismatch Negativity: A Connectivity Study
Using Propofol Anesthesia. *Front Hum Neurosci* **12**, 38–10 (2018).

37. Bishop, D. V. M., Hardiman, M., Uwer, R. & von Suchodoletz, W. Maturation of the long-latency
auditory ERP: step function changes at start and end of adolescence. *Developmental Sci* **10**, 565–575
(2007).

38. Mahajan, Y. & McArthur, G. Maturation of auditory event-related potentials across adolescence.
Hearing Res **294**, 82–94 (2012).

39. Wetzel, N., Widmann, A., Berti, S. & Schröger, E. The development of involuntary and voluntary
attention from childhood to adulthood: A combined behavioral and event-related potential study. *Clin.*
Neurophysiol. **117**, 2191–2203 (2006).

40. Mahajan, Y. & McArthur, G. Maturation of mismatch negativity and P3a response across
adolescence. *Neurosci Lett* **587**, 102–106 (2015).

41. Mueller, V., Brehmer, Y., Oertzen, T. von, Li, S.-C. & Lindenberger, U. Electrophysiological
correlates of selective attention: A lifespan comparison. *Bmc Neurosci* **9**, 18–21 (2008).

42. Krishnan, A., Bidelman, G. M. & Smalt, C. J. Relationship between brainstem, cortical and
behavioral measures relevant to pitch salience in humans. *Neuropsychologia* **50**, 2849–2859 (2012).

43. Bidelman, G. M. Towards an optimal paradigm for simultaneously recording cortical and
brainstem auditory evoked potentials. *J Neurosci Meth* **241**, 94–100 (2015).

44. Jacxsens, L. *et al.* Frequency-Following Responses in Sensorineural Hearing Loss: A Systematic
Review. *J. Assoc. Res. Otolaryngol.* **25**, 131–147 (2024).

45. Plyler, P. & Ananthanarayan, A. K. Human Frequency Following Responses: Representation of
second formant transitions in normal-hearing and hearing-impaired listeners. *J A A A* **37**, e91–e103
(2001).

46. Ananthakrishnan, S., Krishnan, A. & Bartlett, E. L. Human frequency following response: neural
representation of envelope and temporal fine structure in listeners with normal hearing and
sensorineural hearing loss. *Ear and hearing* **37**, e91–e103 (2016).

47. Anderson, S. *et al.* Effects of hearing loss on the subcortical representation of speech cues. *J*
Acoust Soc Am **133**, 3030–3038 (2013).

48. Presacco, A., Simon, J. Z., Anderson, S. & Johnson, B. Speech-in-noise representation in the aging
midbrain and cortex: Effects of hearing loss. *Plos One* **14**, e0213899-26 (2019).

49. Calcus, A., Tuomainen, O., Campos, A., Rosen, S. & Halliday, L. F. Functional brain alterations
following mild-to-moderate sensorineural hearing loss in children. *eLife* 1–37 (2019)

50. Ji, H. *et al.* Features of Cognitive Ability and Central Auditory Processing of Preschool Children With Minimal and Mild Hearing Loss. *J Speech Lang Hear Res* **66**, 1867–1888 (2023).

51. BinKhamis, G., Forte, A. E., Reichenbach, T., O'Driscoll, M. & Kluk, K. Speech Auditory Brainstem Responses in Adult Hearing Aid Users: Effects of Aiding and Background Noise, and Prediction of Behavioral Measures. *Trends Hear* **23**, 233121651984829–20 (2019).

52. Easwar, V., Purcell, D. W., Aiken, S. J., and, V. P. E. & 2015. Evaluation of speech-evoked envelope following responses as an objective aided outcome measure: Effect of stimulus level, bandwidth, and amplification in adults with hearing loss. *Ear Hearing* **36**, 635–652 (2015).

53. Easwar, V., Purcell, D. & Wright, T. Predicting Hearing aid Benefit Using Speech-Evoked Envelope Following Responses in Children With Hearing Loss. *Trends Hear.* **27**, 23312165231151468 (2023).

54. Martinez, A., Eisenberg, L. & Boothroyd, A. The Acoustic Change Complex in Young Children with Hearing Loss: A Preliminary Study. *Seminars Hear* **34**, 278–287 (2013).

55. Koravand, A., Jutras, B. & Lassonde, M. Auditory event related potentials in children with peripheral hearing loss. *Clin Neurophysiol* **124**, 1439–1447 (2013).

56. Engström, E. *et al.* Auditory event-related potentials and mismatch negativity in children with hearing loss using hearing aids or cochlear implants – A three-year follow-up study. *Int J Pediatr Otorhi* **140**, 110519 (2021).

57. Anderson, S., Skoe, E., Chandrasekaran, B. & Kraus, N. Neural Timing Is Linked to Speech Perception in Noise. *J Neurosci* **30**, 4922–4926 (2010).

58. Hornickel, J., Skoe, E., Nicol, T., Zecker, S. & Kraus, N. Subcortical differentiation of stop consonants relates to reading and speech-in-noise perception. *Proc. Natl. Acad. Sci. USA* **106**, 13022–13027 (2009).

59. Bidelman, G. M. & Momtaz, S. Subcortical rather than cortical sources of the frequency-following response (FFR) relate to speech-in-noise perception in normal-hearing listeners. *Neurosci Lett* **746**, 135664 (2021).

60. Mai, G., Tuomainen, J. & Howell, P. Relationship between speech-evoked neural responses and perception of speech in noise in older adults. *J Acoust Soc Am* **143**, 1333–1345 (2018).

61. Mai, G. & Howell, P. The possible role of early-stage phase-locked neural activities in speech-in-noise perception in human adults across age and hearing loss. *Hearing Res* **427**, 108647 (2023).

62. Presacco, A., Simon, J. Z. & Anderson, S. Evidence of degraded representation of speech in noise, in the aging midbrain and cortex. *J. Neurophysiol.* **116**, 2346–2355 (2016).

63. Mepani, A. M. *et al.* Envelope following responses predict speech-in-noise performance in normal-hearing listeners. *J Neurophysiol* **125**, 1213–1222 (2021).

64. Hirtum, T. V. *et al.* Neural envelope tracking predicts speech intelligibility and hearing aid benefit in children with hearing loss. *Hear. Res.* **439**, 108893 (2023).

65. Ching, T. Y. C. *et al.* Acoustic change complex for assessing speech discrimination in normal-hearing and hearing-impaired infants. *Clin. Neurophysiol.* **149**, 121–132 (2023).

66. Näätänen, R. The perception of speech sounds by the human brain as reflected by the mismatch
negativity (MMN) and its magnetic equivalent (MMNm). *Psychophysiology* **38**, 1–21 (2001).

67. Sharma, M., Sharma, M., Purdy, S. C., Newall, P. & Wheldall, K. Electrophysiological and
behavioral evidence of auditory processing deficits in children with reading disorder. *Clin*
Neurophysiol **117**, 1130–1144 (2006).

68. Jost, L. B. *et al.* Native and non-native speech sound processing and the neural mismatch
responses: A longitudinal study on classroom-based foreign language learning. *Neuropsychologia* **72**,
94–104 (2015).

69. Shafer, V., Morr, M., Datta, H., Kurtzberg, D. & Schwartz, R. Neurophysiological indexes of
speech processing deficits in children with specific language impairment. *Journal of Cognitive*
Neuroscience **17**, (2005).

70. Wechsler, D. The Wechsler Abbreviated Scale of Intelligence (Psychological Corporation, San
Antonio, TX), (1999).

71. Bishop, D. V. M., Hardiman, M. J. & Barry, J. G. Is auditory discrimination mature by middle
childhood? A study using time-frequency analysis of mismatch responses from 7 years to adulthood.
Developmental Sci **14**, 402–416 (2011).

72. Boersma, P., and Weenink, D. "Praat: A system for doing phonetics by computer," *Glot. Int.* **5**,
341–347. (2001).

73. Craig, C. GoldWave (Version 5.23) [Sound Editing Program] St Johns, Newfoundland: GoldWave
Inc. (2008).

74. Malmierca, M. S., Sanchez-Vives, M. V., Escera, C. & Bendixen, A. Neuronal adaptation, novelty
detection and regularity encoding in audition. *Frontiers Syst Neurosci* **8**, 111 (2014).

75. Byrne, D. & Dillon, H. The National Acoustic Laboratories' (NAL) New Procedure for Selecting
the Gain and Frequency Response of a Hearing Aid. *Ear Hear.* **7**, 257–265 (1986).

76. Billings, C. Uses and Limitations of Electrophysiology with Hearing Aids. *Seminars Hear* **34**,
257–269 (2013).

77. Jasper, H. The ten-twenty system of the International Federation. *Electroencephalogr Clin*
Neurophysiol, Suppl **52**: 3-6. (1958).

78. Schoof, T. & Rosen, S. The Role of Age-Related Declines in Subcortical Auditory Processing in
Speech Perception in Noise. *J Assoc Res Otolaryngology* **17**, 1–20 (2016).

79. Zhang, G. & Luck, S. J. Variations in ERP data quality across paradigms, participants, and scoring
procedures. *Psychophysiology* **60**, e14264 (2023).

80. Finke, M., Büchner, A., Ruigendijk, E., Meyer, M. & Sandmann, P. On the relationship between
auditory cognition and speech intelligibility in cochlear implant users: An ERP study.
Neuropsychologia **87**, 169–181 (2016).

81: Luck, S. J. An introduction to the event-related potential technique (2nd ed.). MIT Press. (2014).

82. Sussman, E., Steinschneider, M., Gumenyuk, V., Grushko, J. & Lawson, K. The maturation of
human evoked brain potentials to sounds presented at different stimulus rates. *Hearing Res* **236**, 61–79
(2008).

83. Kraus, N., McGee, T., Micco, A., Sharma, A. & Carrell, T. Mismatch negativity in school-age
children to speech stimuli that are just perceptibly different. *Electroencephalogr Clin Neurophysiology*
Evoked Potentials Sect **88**, 123–130 (1993).

84. McGee, T., Kraus, N. & Nicol, T. Is it really a mismatch negativity? An assessment of methods for
determining response validity in individual subjects. *Electroencephalogr Clin Neurophysiology*
Evoked Potentials Sect **104**, 359–368 (1997).

85. Halliday, L. F., Tuomainen, O. & Rosen, S. Auditory processing deficits are sometimes necessary
and sometimes sufficient for language difficulties in children: Evidence from mild to moderate
sensorineural hearing loss. *Cognition* **166**, 139–151 (2017).

86. Kawahara, H., Masuda-Katsuse, I. & de Cheveigné, A. D. Restructuring speech representations
using a pitch-adaptive time–frequency smoothing and an instantaneous-frequency-based F0 extraction:
Possible role of a repetitive structure in sounds. *Speech Commun* **27**, 187–207 (1999).

87. Levitt, H. Transformed up-down methods in psychoacoustics. *J Acoust Soc Am* **49**, 467 (1971).

88. Baker, R. J. & Rosen, S. Evaluation of maximum-likelihood threshold estimation with tone-in-
noise masking. *British Journal of Audiology* (2001).

89. Stone, M. A. & Moore, B. C. J. On the near non-existence of "pure" energetic masking release for
speech. *J Acoust Soc Am* **135**, 1967–1977 (2014).

90. Greenberg, S., Carvey, H., Hitchcock, L. & Chang, S. Temporal properties of spontaneous
speech—a syllable-centric perspective. *J. Phon.* **31**, 465–485 (2003).

91. Plomp, R. & Mimpen, A. M. Speech-reception threshold for sentences as a function of age and
noise level. *J. Acoust. Soc. Am.* **66**, 1333–1342 (1979).

92. Kuznetsova, A., Brockhoff, P. B. & Christensen, R. H. B. lmerTest Package: Tests in Linear
Mixed Effects Models. *J. Stat. Softw.* **82**, (2017).

93. Jenkins, K. A., Fodor, C., Presacco, A. & Anderson, S. Effects of amplification on neural phase
locking, amplitude, and latency to a speech syllable. *Ear and hearing* **39**, 810–824 (2017).

94. Easwar, V., Peng, Z. E., Boothalingam, S. & Seeto, M. Neural Envelope Processing at Low
Frequencies Predicts Speech Understanding of Children With Hearing Loss in Noise and
Reverberation. *Ear Hear.* (2024)

95. Krishnan, A. Human frequency-following responses: representation of steady-state synthetic
vowels. *Hear. Res.* **166**, 192–201 (2002).

96. Russo, N., Nicol, T., Musacchia, G. & Kraus, N. Brainstem responses to speech syllables. *Clin
Neurophysiol* **115**, 2021–2030 (2004).

97. Herrmann, B. & Butler, B. E. Hearing loss and brain plasticity: the hyperactivity phenomenon.
Brain Struct Funct **226**, 2019–2039 (2021).

98. Chambers, A. R. *et al.* Central Gain Restores Auditory Processing following Near-Complete
Cochlear Denervation. *Neuron* **89**, 867–879 (2016).

99. Turrigiano, G. G. & Nelson, S. B. Homeostatic plasticity in the developing nervous system. *Nat.*
Rev. Neurosci. **5**, 97–107 (2004).

100. Teichert, M., Liebmann, L., Hübner, C. A. & Bolz, J. Homeostatic plasticity and synaptic scaling
in the adult mouse auditory cortex. *Sci. Rep.* **7**, 17423 (2017).

101. Godey, B., Schwartz, D., Graaf, J. B. de, Chauvel, P. & Liégeois-Chauvel, C. Neuromagnetic
source localization of auditory evoked fields and intracerebral evoked potentials: a comparison of data
in the same patients. *Clin Neurophysiol* **112**, 1850–1859 (2001).

102. Liegeois-Chauvel, C., Musolino, A., Badier, J. M., Marquis, P. & Chauvel, P. Evoked potentials
recorded from the auditory cortex in man: evaluation and topography of the middle latency
components. *Electroencephalogr Clin Neurophysiology Evoked Potentials Sect* **92**, 204–214 (1994).

103. Lakatos, P. *et al.* The Thalamocortical Circuit of Auditory Mismatch Negativity. *Biol. Psychiatry*
87, 770–780 (2020).

104. Velasco, M., Velasco, F. & Velasco, A. L. Intracranial studies on potential generators of some
vertex auditory evoked potentials in man. *Stereotact Funct Neurosurg* (1989).

105. Webster, D. B. The Mammalian Auditory Pathway: Neuroanatomy. *Springer Handb. Audit. Res.*
1–22 (1992) doi:10.1007/978-1-4612-4416-5_1.

106. Tremblay, K. L., Piskosz, M. & Souza, P. Effects of age and age-related hearing loss on the
neural representation of speech cues. *Clin. Neurophysiol.* **114**, 1332–1343 (2003).

107. Alain, C., Roye, A. & Salloum, C. Effects of age-related hearing loss and background noise on
neuromagnetic activity from auditory cortex. 1–12 (2014) doi:10.3389/fnsys.2014.00008/abstract.

108. Millman, R. E., Mattys, S. L., Gouws, A. D. & Prendergast, G. Magnified Neural Envelope
Coding Predicts Deficits in Speech Perception in Noise. *J Neurosci* **37**, 7727–7736 (2017).

109. Rance, G., Cone-Wesson, B., Wunderlich, J. & Dowell, R. Speech Perception and Cortical Event
Related Potentials in Children with Auditory Neuropathy. *Ear Hearing* **23**, 239–253 (2002).

110. Glista, D., Easwar, V., Purcell, D. W. & Scollie, S. A Pilot Study on Cortical Auditory Evoked
Potentials in Children: Aided CAEPs Reflect Improved High-Frequency Audibility with Frequency
Compression Hearing Aid Technology. *Int J Otolaryngology* **2012**, 1–12 (2012).

111. Uhler, K., Tollin, D. J. & Gilley, P. M. EEG Alpha Band Responses Reveal Amplification
Benefits in Infants with Hearing Loss. *Children* **10**, 600 (2023).

112. Wen, W. & Turrigiano, G. G. Developmental Regulation of Homeostatic Plasticity in Mouse
Primary Visual Cortex. *J. Neurosci.* **41**, 9891–9905 (2021).

113. Gracia-Tabuenca, Z., Moreno, M. B., Barrios, F. A. & Alcauter, S. Development of the brain
functional connectome follows puberty-dependent nonlinear trajectories. *Neuroimage* **229**, 117769
(2021).

114. Laube, C., Bos, W. van den & Fandakova, Y. The relationship between pubertal hormones and
brain plasticity: Implications for cognitive training in adolescence. *Dev Cogn Neuros-neth* **42**, 100753
(2020).

115. Vijayakumar, N. *et al.* A longitudinal analysis of puberty-related cortical development.
Neuroimage **228**, 117684 (2021).

116. Yao, J. D. & Sanes, D. H. Developmental deprivation-induced perceptual and cortical processing
deficits in awake-behaving animals. *eLife* (2018).

117. Hutchison, P. *et al.* Acoustic deprivation modulates central gain in human auditory brainstem and
cortex. *Hear. Res.* **428**, 108683 (2023).

118. Munro, K. J., Turtle, C. & Schaette, R. Plasticity and modified loudness following short-term
unilateral deprivation: Evidence of multiple gain mechanisms within the auditory system. *J Acoust Soc*
Am **135**, 315–322 (2014).

119. Sanes, D. Synaptic and cellular consequences of hearing loss. In: Deafness, pp 129-149, Ed: Kral,
Popper & Fay (2013).