Efficient Generation of Protein Pockets with PocketGen

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ABSTRACT

Designing protein-binding proteins plays an important role in drug discovery. However, AI-based design of such proteins is challenging due to complex ligand-protein interactions, flexibility of ligand molecules and amino acid side chains, and sequence-structure dependencies. We introduce PocketGen, a deep generative model that produces both the residue sequence and atom structure of the protein regions where interactions with ligand molecules occur. PocketGen ensures sequence-structure consistency by using a graph transformer for structural encoding and a sequence refinement module based on a protein language model. The bilevel graph transformer captures interactions at multiple granularities across atom, residue, and ligand levels. To enhance sequence refinement, PocketGen integrates a structural adapter with the protein language model, ensuring consistency between structure-based and sequence-based predictions. Results show that PocketGen can generate high-fidelity protein pockets with superior binding affinity and structural validity. It is ten times faster than physics-based methods and achieves a 95% success rate, defined as the percentage of generated pockets with higher binding affinity than reference pockets, along with achieving an amino acid recovery rate exceeding 64%.

Introduction

Modulating protein functions primarily involves the interaction between proteins and small molecule ligands[1–4]. These interactions play a crucial role in biological processes such as enzymatic catalysis, signal transduction, and cellular regulatory mechanisms. Binding small molecules to specific protein sites can induce conformational changes, modulate activity, or inhibit functions. This mechanism is valuable for studying protein functions and designing small molecule-binding proteins with customized properties. Examples include designing enzymes to catalyze reactions without natural catalysts[5–8] and developing biosensors to detect environmental compounds. These biosensors transduce signals useful for environmental monitoring, clinical diagnostics, pathogen detection, drug delivery systems, and food industry applications[9–12]. Often, designs involve modifying existing ligand-binding protein pockets to enable more precise interactions with specific ligands[13–15]. However, the complexity of ligand-protein interactions, the flexibility of ligands and side chains, and sequence-structure relationships pose challenges for the computational generation of high-validity ligand-binding protein pockets[3,15,16].

Methods for pocket design primarily focus on physics modeling or template matching[10,11,13,17,18]. For example, PocketOptimizer[18–20] uses a pipeline that predicts mutations in protein pockets to enhance binding affinity based on physics-inspired energy functions and search algorithms. Starting with a bound protein-ligand complex, PocketOptimizer explores side chain structures and residue types, evaluating these mutants using energy functions and ranking them through integer linear programming techniques. Another approach employs template matching and enumeration methods[11,13,14,17,21]. For instance, Polizzi et al.[13] use a two-step strategy for pocket design. First, they identify and assemble disconnected protein motifs (van der Mer (vdM) structural units) around the target molecule to build protein-ligand hydrogen bonds. They then graft these residues onto the protein scaffold and select the best protein-ligand pairs using scoring functions. This template-matching strategy enabled de novo design of proteins binding the drug apixaban[22]. However, methods based on physics modeling and template matching can be time-consuming, often requiring several hours for a single protein pocket design. Additionally, the focus on specific fold types (e.g., four-helix bundle[13] or NTF2[14]) by these methods limits their broad use.

Recent advances in protein pocket design have been driven by deep learning-based approaches[3,8,16,23–25]. For example, RFDiffusion[26] employs denoising diffusion probabilistic models[27] along with RoseTTAFold[28] for *de novo* protein structure generation. While capable of designing pockets for specific ligands, RFDiffusion's auxiliary guiding potentials lack precision in modeling protein-ligand interactions. RFdiffusion All-Atom (RFdiffusionAA)[16] advances this approach by enabling direct

generation of binding proteins around small molecules through iterative denoising, thanks to architectural modifications that concurrently consider protein structures and ligand molecules. However, both RFDiffusion and RFdiffusionAA derive residue sequences through post-processing with ProteinMPNN[29] or LigandMPNN[30], potentially leading to inconsistencies between sequence and structure modalities.

In contrast, our previous work, FAIR[24], simultaneously designs the complete atom pocket structure and sequence using a two-stage refinement approach. FAIR employs a coarse-to-fine method, initially updating the backbone protein structure before refining the complete atom structure, including side chains. This iterative process continues until convergence is achieved. However, the gap between the two refinement steps can result in instability and limited performance, highlighting the need for an end-to-end generative approach for pocket design. Related research has focused on the co-design of sequence and structure in complementarity determining regions (CDRs) of antibodies[31–35]. While these methods are effective for antibodies, they face challenges when adapting to pocket designs conditioned on target ligand molecules.

Hybrid approaches that combine deep learning models with traditional methods are also being explored[3,8]. For instance, Yeh et al.[8] developed a novel Luciferase by integrating protein hallucination[36], the trRosetta structure prediction neural network[37], hydrogen bonding networks, and RifDock[38], generating a variety of idealized protein structures with diverse pocket shapes for subsequent filtering. Despite its success, this method's applicability is limited to specific protein scaffolds and substrates, lacking a generalized solution. Similarly, Lee et al.[3] combine deep learning with physics-based methods to create proteins with diverse and designable pocket geometries. Their approach involves backbone generation via trRosetta hallucination, sequence design through ProteinMPNN[29] and LigandMPNN[30], and filtering with AlphaFold[39]. Current pocket generation models face several challenges, including achieving sequence-structure consistency and accurately modeling complex protein-ligand interactions.

Here, we introduce PocketGen, a deep generative method for efficient generation of protein pockets. PocketGen employs a co-design scheme (Figure 1a), where the model concurrently predicts the sequence and structure of the protein pocket based on the ligand molecule and the protein scaffold (excluding the pocket). PocketGen consists of two modules: the bilevel graph transformer (Figure 1b) and the sequence refinement module (Figure 1c). PocketGen models the protein-ligand complex as a geometric graph of blocks to handle variable atom counts across different residues and ligands. Initially, the pocket residues are assigned the maximum possible number of atoms (14 atoms) and are later mapped back to specific residue types post-generation.

The graph transformer module uses a bilevel attention mechanism to capture interactions at multiple granularities (atom-level and residue/ligand-level) and aspects (intra-protein and protein-ligand). To account for the influence of the redesigned pocket on the ligand, the ligand structure is updated during refinement to capture potential changes in binding pose. To ensure consistency across protein sequence and structure domains and incorporate evolutionary information encoded in protein language models (pLMs)[40,41], PocketGen integrates a structural adapter into protein sequence updates. This adapter facilitates cross-attention between sequence and structure features, achieving sequence-structure consistency. Only the adapter is fine-tuned during training, while the remaining layers of the protein language models remain unchanged. Results show that PocketGen significantly outperforms current methods in protein pocket generation across two popular benchmarks. PocketGen achieves an average amino acid recovery rate of 63.40% and a Vina score of -9.655 for top-1 generated protein pockets on the CrossDocked dataset. Comprehensive analyses indicate that PocketGen can produce diverse and high-affinity protein pockets for functional molecules, showcasing its efficacy and potential in designing small-molecule binders and enzymes.

Results

Benchmarking generated protein pockets across multiple axes of molecular evaluation

We benchmark PocketGen on two datasets. The **CrossDocked** dataset[42] comprises protein-molecule pairs generated through cross-docking, divided into train, validation, and test sets based on a sequence identity threshold of 30%. The **Binding MOAD** dataset[43] includes experimentally determined protein-ligand complexes, split into train, validation, and test sets according to the proteins' enzyme commission numbers[44]. Given intermolecular distance scales relevant to protein-ligand interactions[45], our default experimental setting considers all residues with atoms within 3.5 Å of any binding ligand atoms as part of the protein pocket, averaging about eight residues per pocket. We also investigate PocketGen's capability to design larger pockets with a radius of 5.5 Å that include more residues (Figure 3c). Methodological details on data processing are provided in Methods.

We use three groups of metrics to evaluate the quality of protein pockets generated by PocketGen. First, we measure the affinity between the generated pocket and the target ligand molecule using the AutoDock **Vina score**[46], **MM-GBSA**[47], and **min-in-place GlideSP score**[48]. Second, we assess the structural validity of the generated pockets using **scRMSD**, **scTM**, and **pLDDT**. We derive the amino acid sequence for the protein pocket structure using ProteinMPNN[29] and predict the pocket structure using ESMFold[49] or AlphaFold2[39]. The scRMSD is obtained by aligning and computing the self-consistency root mean squared deviation between the generated and the predicted structure's backbone atoms. Following an established strategy[50,51], eight sequences are predicted for each generated protein structure and the sequence with the minimum scRMSD is used for reporting the metric values. Similarly, scTM, the self-consistency template modeling score, is obtained by calculating the TM-score[52] between the ESMFold/AlphaFold2-predicted structure and the generated structure, with scores ranging from 0

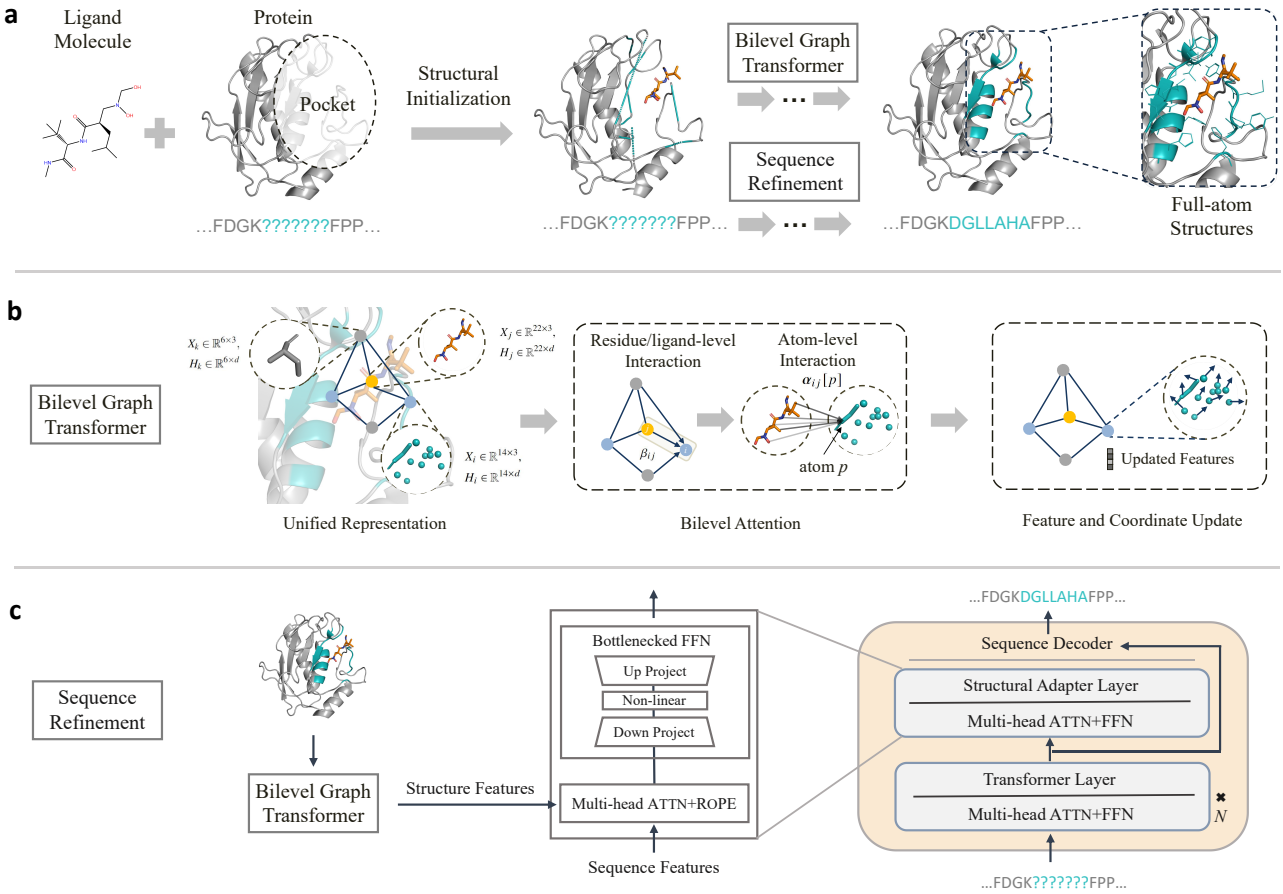

Figure 1. Overview of PocketGen generative model for the design of full-atom ligand-binding protein pockets. a, Conditioned on the binding ligand molecule and the rest part of the protein except the pocket region (i.e., scaffold), PocketGen aims to generate the full atom pocket structure (backbone and sidechain atoms) and the residue type sequence with iterative equivariant refinement. The ligand structure is also adjusted during the protein pocket refinement. **b**, Bilevel graph transformer is leveraged in PocketGen for all-atom structural encoding and update. The bilevel level attention captures both the residue/ligand and atom-level interactions. Both the protein pocket structure and the ligand molecule structure are updated in the refinement. **c**, Sequence refinement module adds lightweight structural adapter layers into pLMs for sequence prediction. Only the adapter's parameters are fine-tuned during training, and the other layers are fixed. In the adapter, the cross-attention between sequence and structure features is performed to achieve sequence-structure consistency.

to 1, where higher values indicate greater designability. We also report the ΔscTM score to evaluate whether the generated pocket improves or degrades the scTM score of the initial protein. The predicted local distance difference test (pLDDT) score[39] reflects the confidence in structural predictions on a scale from 0 to 100, with higher scores indicating greater confidence. We report the average pLDDT score from ESMFold/AlphaFold2 across pocket residues. A generated protein pocket structure is defined as **designable** if the overall structure scRMSD is less than 2 Å and the pocket scRMSD is less than 1 Å[26,53,54]. Table S1 shows the percentage of designable generated pockets. Supplementary Figure S1 describes calculating these metrics. Finally, we report the amino acid recovery (**AAR**) as the percentage of correctly predicted pocket residue types to evaluate the accuracy of the designed sequence. A higher AAR signifies better modeling of sequence-structure dependencies.

We compare PocketGent to six methods, including deep learning methods RFDiffusion[26], RFDiffusionAA (RFAA)[16], FAIR[24], dyMEAN[25], a template-matching method DEPACT[17], and a physics modeling method PocketOpt[18] (refer to Methods for detailed descriptions of baselines). In Figure 2 and Table S1, PocketGen and other methods are tasked to generate 100 sequences and structures for each protein-ligand complex in the CrossDocked and Binding MOAD dataset test set. PocketOpt is not compared because it focuses on mutating existing pockets for optimization and is time-consuming for generating many protein pockets. In Table S1, we show the mean and standard deviation of results over three different runs with random seeds. In Figure 2, we bootstrap the generation results and draw the distributions to show how sensitive the results are to the composition of the data set[55]. Table S1 and Figure 2 show that PocketGen surpasses all the baselines, including state-of-the-art methods

Table 1. The top 1/3/5/10 generated designable protein pocket (ranked by Vina score) on the CrossDocked dataset. The success rate measures the percentage of protein that the model can generate pockets with higher affinity than the reference ones in the datasets. Besides the Vina score, we additionally use MM-GBSA and min-in-space GlideSP score to evaluate the binding affinity. We report the average plDDT of the predicted pocket, the scRMSD of the pocket backbone coordinates, and the change of scTM scores of the whole protein. AF2 means the scores are calculated with AlphaFold2 as the folding tool (ESMFold results in Table S2). co indicates codesign, where codesign methods directly use the designed sequence for consistency calculation. The plDDT, scRMSD, and ΔscTM for PocketOpt are not reported, as PocketOpt keeps protein backbone structures fixed. We use to mark the results of affinity-related metrics, for pocket-structure related metrics, and for whole protein structure metrics. We report the means and standard deviations over three independent runs with random seeds.

	PocketOpt	DEPACT	dyMEAN	FAIR	RFDiffusion	RFAA	PocketGen
Top-1 generated protein pocket							
Vina score (↓)	-9.216±0.154	-8.527±0.061	-8.540±0.107	-8.792±0.122	-9.037±0.080	-9.216±0.091	**-9.655**±0.094
MM-GBSA (↓)	-58.754±1.220	-47.130±1.372	48.248±0.816	-51.923±0.588	-54.817±1.091	-59.255±1.260	**-63.542**±0.717
GlideSP (↓)	-8.612±0.127	-7.495±0.053	-7.472±0.088	-7.584±0.094	-8.485±0.069	-8.540±0.065	**-8.916**±0.047
Success Rate (↑)	0.923±0.034	0.750±0.016	0.762±0.029	0.796±0.035	0.891±0.020	0.930±0.027	**0.974**±0.012
pLDDT (AF2) (↑)	-	82.164±0.241	83.053±0.397	83.285±0.240	84.432±0.152	86.571±0.178	**86.830**±0.145
scRMSD (AF2) (↓)	-	0.714±0.025	0.708±0.022	0.693±0.018	0.675±0.015	0.654±0.012	**0.645**±0.009
ΔscTM (AF2) (↑)	-	-0.008±0.003	-0.005±0.002	-0.011±0.005	0.022±0.006	0.020±0.003	**0.028**±0.002
ΔscTM (AF2+co) (↑)	-	-0.012±0.003	-0.025±0.004	-0.032±0.007	-	-	**0.008**±0.002
Top-3 generated protein pockets							
Vina score (↓)	-8.878±0.112	-8.131±0.064	-8.196±0.090	-8.321±0.045	-8.876±0.107	-8.980±0.057	**-9.353**±0.063
MM-GBSA (↓)	-53.372±1.164	-43.790±1.029	-44.151±0.534	-46.050±0.809	-52.423±0.847	-53.593±0.722	**-60.770**±0.589
GlideSP (↓)	-8.360±0.094	-7.377±0.039	-7.325±0.078	-7.348±0.052	-8.219±0.049	-8.233±0.060	**-8.670**±0.056
pLDDT (AF2) (↑)	-	82.049±0.456	82.918±0.237	83.025±0.334	84.260±0.210	**86.289**±0.214	86.280±0.135
scRMSD (AF2) (↓)	-	0.713±0.017	0.722±0.011	0.692±0.016	0.685±0.007	**0.659**±0.014	0.660±0.012
ΔscTM (AF2) (↑)	-	-0.011±0.004	-0.006±0.002	-0.008±0.003	0.021±0.003	0.022±0.002	**0.026**±0.003
ΔscTM (AF2+co) (↑)	-	-0.016±0.005	-0.026±0.004	-0.034±0.003	-	-	**0.005**±0.001
Top-5 generated protein pockets							
Vina score (↓)	-8.702±0.090	-7.786±0.052	-7.974±0.049	-7.943±0.035	-8.510±0.073	-8.689±0.044	**-9.239**±0.076
MM-GBSA (↓)	-52.080±1.071	-35.250±0.823	-37.924±0.340	-37.816±0.402	-46.847±0.700	-51.651±0.809	**-58.083**±0.561
GlideSP (↓)	-8.173±0.089	-7.126±0.035	-7.294±0.042	-7.289±0.041	-8.022±0.030	-8.093±0.048	**-8.417**±0.040
pLDDT (AF2) (↑)	-	82.445±0.307	82.763±0.102	83.748±0.271	84.505±0.288	85.617±0.105	**85.969**±0.080
scRMSD (AF2) (↓)	-	0.716±0.014	0.726±0.011	0.698±0.015	0.680±0.009	0.657±0.006	**0.655**±0.004
ΔscTM (AF2) (↑)	-	-0.009±0.003	-0.007±0.002	-0.012±0.004	0.019±0.003	0.020±0.001	**0.025**±0.001
ΔscTM (AF2+co) (↑)	-	-0.017±0.002	-0.025±0.006	-0.035±0.005	-	-	**0.006**±0.002
Top-10 generated protein pockets							
Vina score (↓)	-8.556±0.104	-7.681±0.040	-7.690±0.054	-7.785±0.028	-8.352±0.061	-8.524±0.038	**-9.065**±0.057
MM-GBSA (↓)	-49.257±0.821	-32.534±0.680	-33.118±0.269	-33.670±0.440	-45.726±0.830	-47.325±0.540	**-54.800**±0.406
GlideSP (↓)	-7.935±0.082	-6.954±0.042	-7.022±0.034	-7.131±0.025	-7.806±0.022	-7.840±0.026	**-8.196**±0.027
pLDDT (AF2) (↑)	-	81.520±0.317	82.467±0.255	83.271±0.228	84.080±0.190	85.442±0.145	**85.945**±0.139
scRMSD (AF2) (↓)	-	0.712±0.013	0.733±0.014	0.706±0.013	0.688±0.009	0.680±0.010	**0.659**±0.007
ΔscTM (AF2) (↑)	-	-0.014±0.002	-0.006±0.001	-0.010±0.003	0.016±0.002	0.019±0.001	**0.023**±0.002
ΔscTM (AF2+co) (↑)	-	-0.018±0.004	-0.030±0.002	-0.033±0.002	-	-	**0.004**±0.002

RFDiffusion and RFDiffusionAA (RFAA) in designability (by 3% and 2% on CrossDocked, respectively) and Vina scores (by 0.199 and 0.123 on CrossDocked, respectively), reflecting its effectiveness in generating structurally valid pockets with high binding affinities. This superior performance can be attributed to PocketGen's capability to capture interactions at multiple granularities (atom-level and residue/ligand-level) and aspects (intra-protein and protein-ligand).

Additionally, PocketGen significantly improves the best-performing alternative method, RFDiffusionAA, by 13.95% on average in AAR, benefiting from incorporating a protein language model that encompasses evolutionary sequence information. In contrast, RFDiffusion and RFDiffusionAA require post-processing to derive amino acid types, which may incur sequence-structure inconsistency and inferior performance in AAR. In protein engineering, the practice often involves mutating several key residues to optimize properties while keeping most residues unchanged to preserve protein folding stability[56,57]. A high AAR achieved by generated protein pockets aligns with this practice.

In Table 1, the top-1, 3, 5, and 10 protein pockets (ranked by Vina score) generated by PocketGen demonstrate the lowest Vina scores, achieving 0.476 lower Vina scores on average than RFDiffusionAA. Besides Vina, two additional affinity metrics, MM-GBSA and GlideSP score further validate that PocketGen is in fact generating higher affinity pockets (4.287 lower MM-GBSA and 0.376 lower GlidsSP score). Moreover, PocketGen also achieves competitive pLDDT, scRMSD, and ΔscTM scores, showcasing PocketGen's ability to produce high-affinity pockets that maintain structural validity and sequence-structure

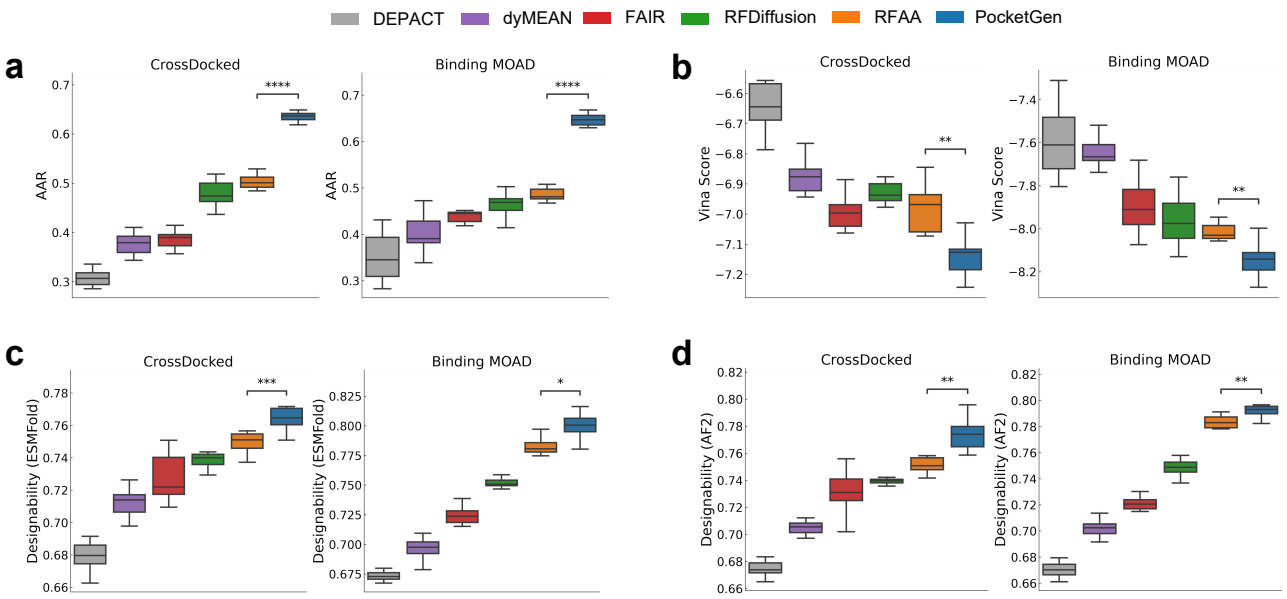

Figure 2. Benchmarking PocketGen on CrossDocked and Binding MOAD datasets. Shown are **a,** amino acid recovery rates (AAR), **b,** Vina score performance, **c,** Designability scores using ESMFold structure prediction method, and **d,** Designability scores using AF2 structure prediction method. Uncertainty is quantified via bootstrapping, two-sided Kolmogorov-Smirnov test is used to compare PocketGen to the best-performing existing model (RFAA). P-value annotation legend: *: $p \in [0,01,0.05]$, **: $p \in [0.001,0.01]$, ***: $p \in [0.0001,0.001]$, ****: $p \leq 0.0001$.

consistency. With a 97% success rate in creating pockets with higher affinity than the reference cases (the success rate of the strongest baseline method is 93% of RFDiffusionAA) on CrossDocked, PocketGen's effectiveness and broad applicability across various ligand molecules are demonstrated.

To investigate the substructure validity and consistency with reference datasets, we conduct a qualitative substructure analysis (Table S4 and Figure S2). This analysis considers three covalent bonds in the residue backbone (C-N, C=O, and C-C), three dihedral angles in the backbone (ϕ, ψ, ω[58]), and four dihedral angles in the side chains ($\chi_1, \chi_2, \chi_3, \chi_4$[59]). Following previous research[60,61], we collect the bond length and angle distributions in the generated pockets and the test dataset and compute the Kullback-Leibler (KL) divergence to quantify the distribution distances. Lower KL divergence scores for PocketGen indicate its effectiveness in replicating the geometric features present in the data.

Probing generative capabilities of PocketGen

Next, we explore PocketGen's generative capabilities. In addition to designing high-quality protein pockets, generative models need to be efficient and maximize the yield of biochemical experiments, meaning that they can rapidly produce high-fidelity pocket candidates and that only a small number of designs are necessary to find a hit. Figure 3a offers a comparative analysis of average **runtime** across different methods. Traditional approaches, such as physics-based modeling (PocketOpt) and template-matching (DEPACT), require over 1,000 seconds to generate 100 pockets. Similarly, the runtime for the advanced protein backbone generation models RFDiffusion and RFDiffusionAA are significant due to their diffusion architecture, requiring 1633.5 and 2210.1 seconds on average to design 100 pockets. In contrast, methods that iteratively refine protein designs, including PocketGen, show a notable decrease in generation time; it takes 44.2 seconds for PocketGen to generate 100 pockets.

While recent methods for pocket generation prioritize maximizing binding affinity with target molecules, this strategy may not always align with practical needs, where pocket diversity also plays a critical role. Examining a batch of designed pockets rather than a single design is beneficial to improve the success rate of pocket design. Thus, we examine the relationship between binding affinity and diversity of generated protein pockets in Figure 3b. **Diversity** is quantified as ($1 -$ average pairwise pocket residue sequence similarity) and can be adjusted by altering the sampling temperature τ (with higher τ resulting in greater diversity). Figure 3b compares PocketGen with the most competitive baseline RFDiffusionAA[16] + LigandMPNN[30], the latest version of ProteinMPNN[29]. Additional results for the RFDiffusion method are in Figure S4. We find that binding affinity and diversity present a trade-off. PocketGen can produce protein pockets with higher affinity than RFDiffusionAA at equivalent levels of diversity.

Figure 3c examines the impact of changes in the redesigned pocket size on PocketGen's performance. The redesign process targets all residues with atoms within 3.5 Å, 4.5 Å, and 5.5 Å of any binding ligand atoms. The average AAR, RMSD, and

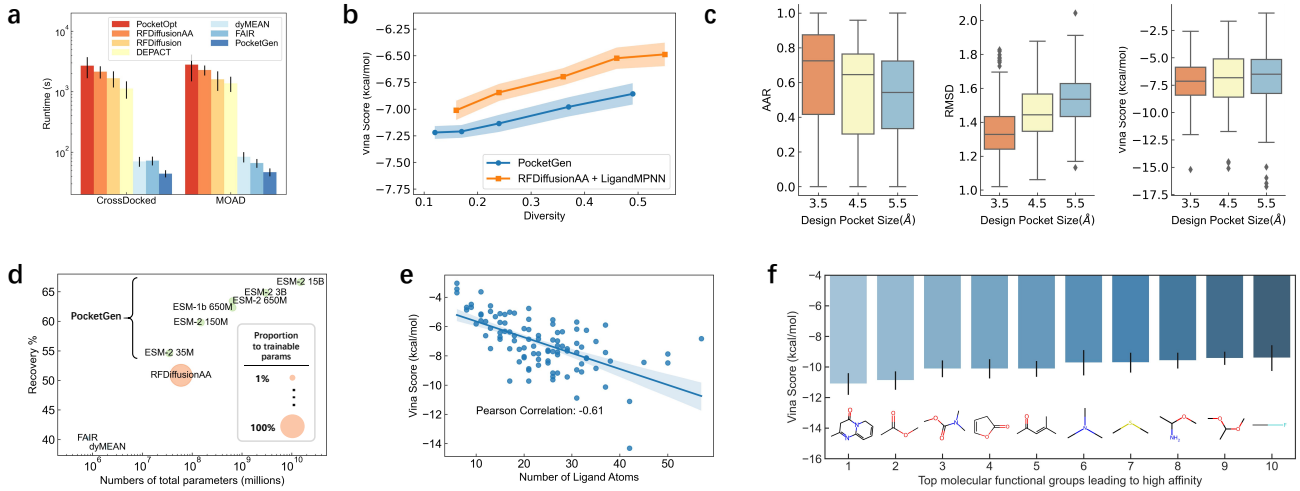

Figure 3. Exploring the capabilities of PocketGen. **a**, The average runtime of different methods for generating 100 protein pockets for a ligand molecule on the two benchmarks. **b**, The trade-off between quality (measured by Vina score) and diversity (1- average pairwise sequence similarity) of PocketGen. We can balance the trade-off by tuning the temperate hyperparameter τ. **c**, The influence of the design pocket size on the metrics. **d**, Performance w.r.t. model scales of pLMs using ESM series on CrossDocked dataset. The green dots represent PocketGen models with different ESMs. The bubble size is proportional to the number of trainable parameters. **e**, PocketGen tends to generate pockets with higher affinity for larger ligand molecules (Pearson Correlation $\rho = -0.61$, bands indicate 95% confidence interval). **f**, The top molecular functional groups leading to high affinity.

Vina scores show a slight decline for larger protein pockets. This trend is attributed to the increased complexity and reduced contextual information associated with expanding the redesigned protein pocket area. Furthermore, an observation of note is that larger pockets tend to enable the exploration of structures with potentially higher affinity, as indicated by the lowest Vina scores, reaching -20 kcal/mol for designs with a 5.5 Å radius. This phenomenon could be due to the enhanced structural complementarity achievable in larger pocket designs. In Extended Data Figure 1ab, we also show PocketGen has the potential to generate the full protein binder for the ligand molecule with two case studies. The generated protein binder demonstrates high scTM scores (0.900 and 0.976, respectively).

Incorporating protein language models (pLMs) distinguishes PocketGen from prior pocket generation models. Beyond employing ESM-2 650M[49] in the default configuration, various versions of ESM with parameter counts ranging from 8M to 15B were also evaluated. As illustrated in Figure 3d, PocketGen's performance improves with the scaling of pLMs. Going from ESM-2 35M to ESM-2 15B protein language models, the performance of PocketGen increases from 54.58% to 66.61%. Specifically, a logarithmic scaling law is observed, aligning with trends noted in large language models[62]. PocketGen achieves efficient training with large pLMs by fine-tuning adapter layers while keeping the majority of pLM layers fixed. PocketGen also has significantly fewer trainable parameters than RFDiffusionAA[16] (7.9M versus 82.9M trainable parameters).

The performance of PocketGen in generating binding pockets is influenced by the characteristics of the ligand molecule. Figure 3e illustrates the relationship between the average Vina score of generated pockets and the number of ligand atoms, indicating that PocketGen tends to create pockets with higher affinity for larger ligand molecules. This trend may be due to the increased surface area for interaction, the presence of additional functional groups, and enhanced flexibility in the conformation of larger molecules[63,64]. Additionally, functional groups in ligand molecules that contribute to high binding affinity were identified using IFG[65]. Figure 3f displays the top 10 molecular functional groups, which include hydrogen bond donors and acceptors (e.g., carbonyl groups), aromatic rings, sulfhydryl groups, and halogens. These groups can form favorable interactions with protein pockets, thereby increasing binding affinity.

As PocketGen also updates ligand structures during pocket generation, we leverage PoseBusters[66] to evaluate the structural validity of the updated ligands. Extended Data Figure 1e shows the detailed validity check, where PocketGen achieves over 95% on all the tests. This is not a surprise because PocketGen only slightly updates the ligand structure during pocket generation and manages to keep ligand structural validity. In Extended Data Figure 1c, we also explore the relationship between affinity and the RMSD to the crystal structure on PDBBind. Generally, lower RMSD leads to higher affinity. Extended Data Fig. 1d shows that PocketGen improves most of the protein-ligand complexes in PDBBind by redesigning the pockets.

We performed ablation studies (Table S5) and hyperparameter analysis (Figure S3) to evaluate the contribution of each module in PocketGen and the impact of loss function hyperparameters on model performance. Among others, we replaced the bilevel graph transformer in PocketGen with other popular encoders in structural biology, such as EGNN[67], GVP[68], and GMN[69]

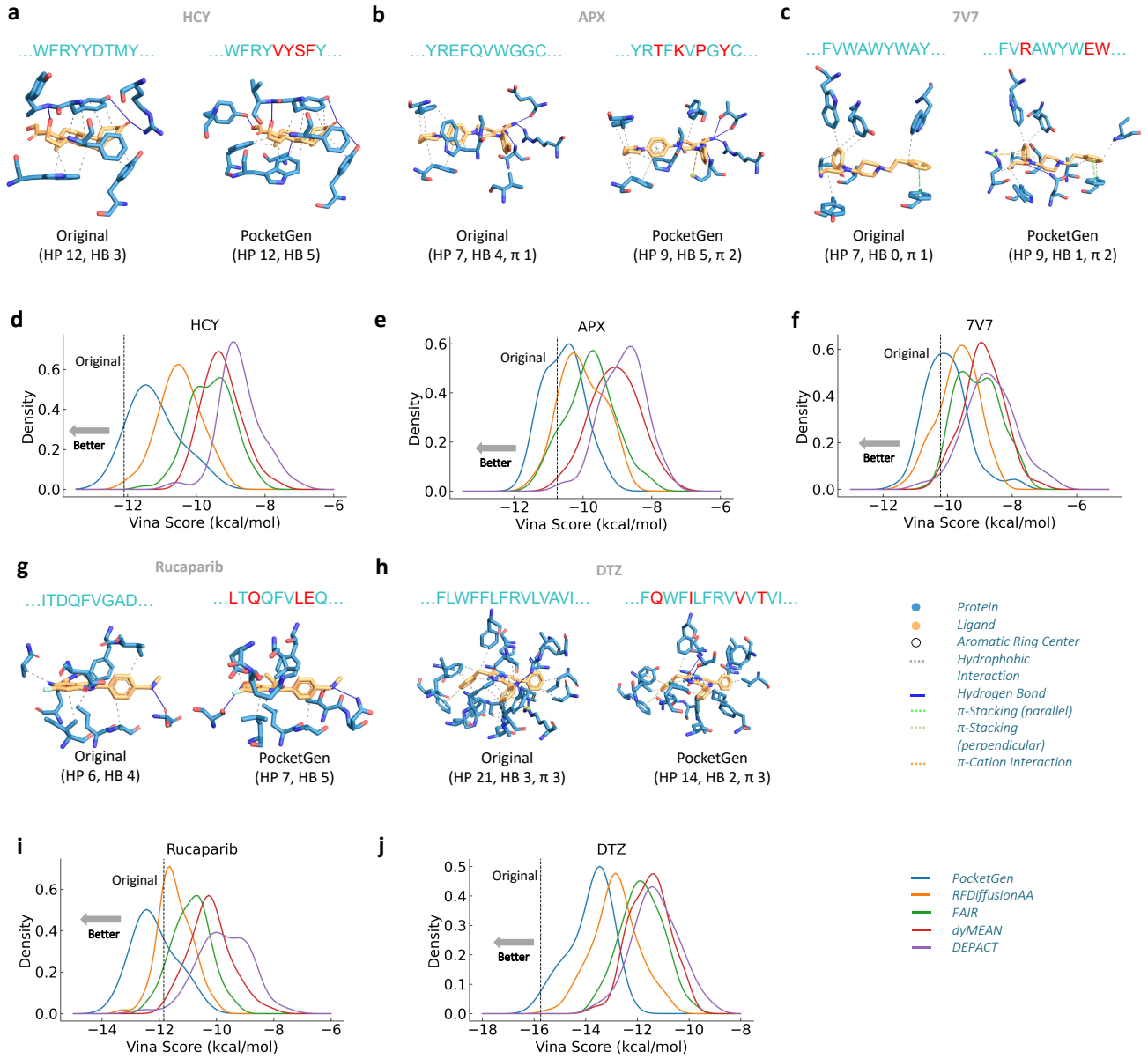

Figure 4. Using PocketGen to design protein pockets for binding with important ligands. a, **b**, **c,** Illustrations of protein-ligand interaction analysis for three target molecules (HCY, APX, and 7V7, respectively). 'PocketGen' refers to the protein pocket designed by PocketGen, and 'Original' denotes the original protein-ligand structure. 'HP' indicates hydrophobic interactions, 'HB' signifies hydrogen bonds, and 'π' denotes the π-stacking/cation interactions. In the residue sequences, red ones denote the designed residues that differ from the original pocket. **d**, **e**, **f,** The pocket binding affinity distributions of PocketGen and baseline methods for three target molecules (HCY, APX, and 7V7, respectively). We mark the Vina Score of the original pocket with the vertical dotted lines. For each method, we sample 100 pockets for each target ligand. The ratio of generated pockets by PocketGen with higher affinity than the corresponding reference pocket are 11%, 40%, and 45%, respectively. **g**, **h,** Protein-ligand interaction analysis for unseen proteins in the training dataset (PiB[21] and luxsit[8]). The target molecules are Rucaparib and DTZ, respectively. **i**, **j,** The pocket binding affinity distributions of PocketGen and baselines for Rucaparib and DTZ.

for comparison. The results show that the bilevel graph transformer and the integration of pLM into PocketGen significantly improve performance. Additionally, PocketGen is robust to hyperparameter variations, consistently producing competitive results.

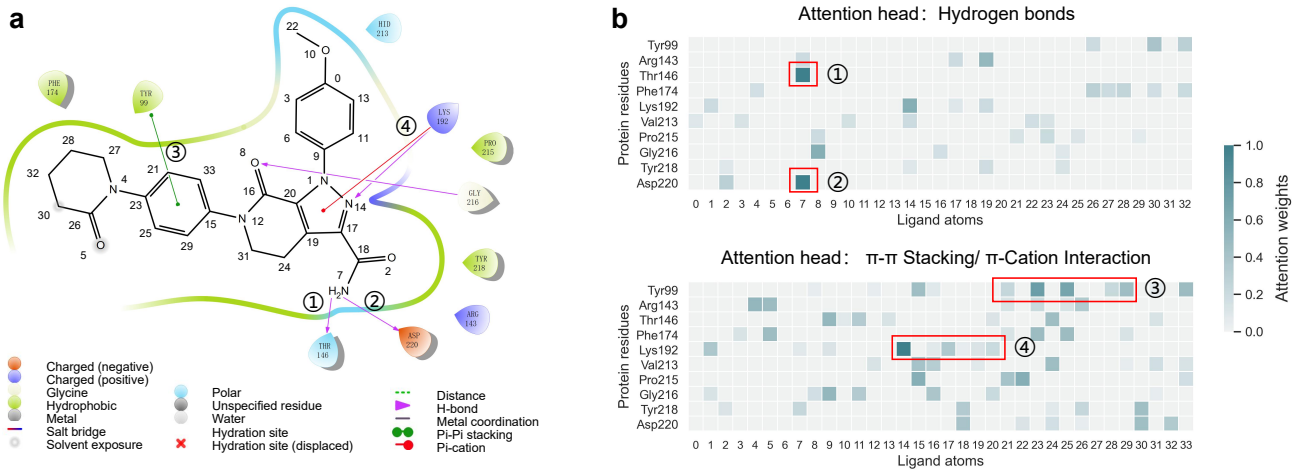

Figure 5. Attention maps in PocketGen capture interactions between atoms in protein and ligand molecules. a, The 2D interaction plot of the designed pocket by PocketGen for APX. **b**, The heatmap of attention matrices between residues and ligand atoms from the last layer of PocketGen. We show two selected attention heads with notable attention patterns marked with red rectangles. We notice that each head emphasizes different interactions. For example, PocketGen recognizes the **hydrogen bond** interaction and assigns a strong attention weight between residue ① THR146, ② ASP220, and ligand atom 7 in the first head. The **π-π stacking** and **π-Cation** interactions of ③ TYR99 and ④ LYS192 are well captured in the second head. The values are normalized by the maximum value ($v_{\max}$) and the minimum value ($v_{\min}$) in each heatmap (i.e., $v' = \frac{v - v_{\min}}{v_{\max} - v_{\min}}$).

Generating protein pockets for therapeutic small molecule ligands

Next, we illustrate the capacity of PocketGen to redesign the pockets of antibodies, enzymes, and biosensors for target ligand molecules, drawing upon previous research[3,10,16]. Specifically, we consider the following molecules. **Cortisol (HCY)**[70] is a primary stress hormone that elevates glucose levels in the bloodstream and serves as a biomarker for stress and other conditions. We redesign the pocket of a cortisol-specific antibody (PDB ID 8cby) to potentially aid in developing immunoassays. **Apixaban (APX)**[71] is an oral anticoagulant approved by the FDA in 2012 for patients with non-valvular atrial fibrillation to reduce the risk of stroke and blood clots[72]. Apixaban targets Factor Xa (fXa) (PDB ID 2p16), a crucial enzyme in blood coagulation that transforms prothrombin into thrombin for clot formation. Redesigning the pocket of fXa could, therefore, have therapeutic significance. **Fentanyl (7V7)**[73] has become a widely abused drug contributing to the opioid crisis. Computational design of fentanyl-binding proteins (biosensors) can facilitate detection and neutralization of the toxin[10]. For example, Baker et al.[10] developed a biosensor (PDB ID 5tzo) for detecting fentanyl in plants. In Figure 4, PLIP[74] describes the interactions between the designed protein pocket and ligands, comparing these predicted interactions to the original patterns.

To generate pockets for the aforementioned small molecules, we pretrained PocketGen on the Binding MOAD dataset and carefully excluded the protein-ligand complexes used in the case studies. The pockets produced by PocketGen replicate most non-bonded interactions from experimentally measured protein-ligand complexes (e.g., achieving a 13/15 match for HCY) and introduce additional physically plausible interaction patterns not present in the original complexes (generated pockets for HCY, APX, and 7V7 molecules form 2, 3, and 4 extra interactions). Specifically, for HCY, PocketGen maintains essential interaction patterns such as hydrophobic interactions (TRP47, PHE50, TYR59, and TYR104) and hydrogen bonds (TYR59) and introduces two new hydrogen bond-mediated interactions with the pocket. For protein pockets designed to bind APX and 7V7 ligands while retaining key interaction patterns, including hydrophobic interactions, hydrogen bonds, and π-π stacking, PocketGen-generated molecules establish additional interactions (e.g., π-Cation interaction with LYS192 for APX and hydrogen bonds with ASN35 for 7V7), thus enhancing binding affinity with the target ligands. In conclusion, PocketGen can establish non-covalent interactions derived from protein-ligand structure data.

With the ability to establish favorable protein-ligand interactions, PocketGen generates high-affinity pockets for these drug ligands. In Figure 4d,e,f, we show the affinity distributions of the generated pockets by PocketGen and baseline methods. The ratio of generated pockets with higher affinity than the reference pocket is 11 %, 40%, and 45%, respectively, for PocketGen. On the contrary, the best runner-up method, RFDiffusionAA, only has 0%, 10 %, and 18%, respectively.

Protein stability is crucial in protein design, ensuring that the designed protein can fold into and maintain its 3D structure[75]. Stability is quantified by the difference in Gibbs free energy ($\Delta\Delta G$) of folded protein between the redesigned protein and the wild-type (original) protein, where $\Delta\Delta G = \Delta G_{\text{orig}} - \Delta G_{\text{redesign}}$. A positive $\Delta\Delta G$ value indicates increased protein stability, while a negative value suggests decreased stability. We used DDMut[76] to predict the change in stability for pockets generated in

Figure 4, with $\Delta\Delta G$ values of 0.09 (HCY), 0.92 (APX), 0.13 (7V7), 0.27 (Rucaparib), and 0.02 (DTZ) respectively. These results indicate that PocketGen can generate protein structures that are likely sufficiently stable to bind ligand molecules.

To demonstrate the generalization capability of PocketGen, we tested it on unseen proteins from the training sets, including PiB[21] and luxsit[8], with binding ligand molecules Rucaparib and DTZ, respectively. Figure 4g and 4h show the interaction analysis, while Figure 4i and 4j display the distribution of Vina scores. PocketGen consistently outperforms baseline methods in generating higher-affinity pockets. Generating pockets with higher affinity than the reference pocket for DTZ is more challenging, as the original pocket was designed using site-saturation mutagenesis[8] to achieve optimal design. In Extended Data Figure 1f, we report case studies with a pair of activity cliff ligand molecules (C19 and C52)[77] to explore PocketGen's adaptability. PocketGen's generated interactions vary across molecular fragments; hydrogen bonds and hydrophobic interactions are generated for one fragment, and halogen bonds are generated for a different fragment, suggesting that PocketGen has learned protein-ligand interaction rules, enabling it to adaptively design high-affinity binding pockets.

Exploring interactions between atoms in protein and ligand molecules produced by PocketGen

We analyze attention maps to understand what PocketGen has learned using a generated pocket for the ligand APX as an example. Figure 5a presents a 2D interaction plot drawn with Schrödinger Maestro software. To assess PocketGen's recognition of meaningful protein-ligand interactions, we plot the heatmap of attention weights produced by the final layer of PocketGen's neural architecture. In Figure 5b, two attention heads are illustrated, with each row and column representing a protein residue or a ligand atom, respectively. The attention heatmaps are sparse, reflecting PocketGen's use of sparse attention (see Methods for more details). The attention heads display diverse patterns, focusing on different aspects. For instance, the first head appears to emphasize hydrogen bonds, assigning high weights between residue THR146, ASP220, and ligand atom 7. The second attention head seems to capture π-π stacking and π-cation interactions, specifically between residue TYR99 and ligand atoms 15, 21, 23, 25, 29, and 33; and residue LYS192 and ligand atoms 1, 14, 17, 19, and 20. These analyses indicate that PocketGen, despite its data-driven nature, has acquired biochemical knowledge to identify intermolecular interactions.

Discussion

Protein-ligand binding is critical in enzyme catalysis, immune recognition, cellular signal transduction, gene expression control, and other biological processes. Recent developments include deep generative models designed to study protein-ligand binding, like Lingo3DMol[78], ResGen[79], and PocketFlow[80] which generate *de novo* drug-like ligand molecules for fixed protein targets; NeuralPLexer[4] can create the structure of protein-ligand complexes given the protein sequence and ligand molecular graph. However, these models do not facilitate the *de novo* generation of protein pockets, the interfaces that bind with the ligand molecule for targeted ligand binding, critical in enzyme and biosensor engineering. We developed PocketGen, a deep generative method capable of generating both the residue sequence and the full atom structures of the protein pocket region for binding with the target ligand molecule. PocketGen includes two main modules: a bilevel graph transformer for structural encoding and updates and a sequence refinement module that uses protein language models (pLMs) for sequence prediction. For structure prediction, the bilevel graph transformer directly updates the all-atom coordinates instead of separately predicting the backbone frame orientation and side-chain torsion angles. To achieve sequence-structure consistency and effectively leverage evolutionary knowledge from pLMs, a structural adapter is integrated into protein language models for sequence updates. This adapter employs cross-attention between sequence and structure features to promote information flow and ensure sequence-structure consistency. Extensive experiments across benchmarks and case studies involving therapeutic ligand molecules illustrate PocketGen's ability to generate high-fidelity pocket structures with high binding affinity and favorable interactions with target ligands. Analysis of PocketGen's performance across various settings reveals its proficiency in balancing diversity and affinity and generalizing across different pocket sizes. Additionally, PocketGen offers computational efficiency, significantly reducing runtime compared to traditional physics-based methods, making it feasible to sample large quantities of pocket candidates. PocketGen surpasses existing methods in efficiently generating high-affinity protein pockets for target ligand molecules, finding important interactions between atoms on protein and ligand molecules, and attaining consistency in sequence and structure domains.

PocketGen creates several fruitful directions for future work. PocketGen could be expanded to design larger areas of the protein beyond the pocket area. While PocketGen has been evaluated on larger pocket designs, modifications will be required to enhance scalability and robustness for generating larger protein areas. Another fruitful future direction involves incorporating additional biochemical priors, such as subpockets[81] and interaction templates[17], to improve generalizability and success rates. For instance, despite overall dissimilarity, two protein pockets might still bind the same fragment if they share similar subpockets[82]. Moreover, conducting wet lab experiments could provide empirical validation of PocketGen's effectiveness. Approaches such as PocketGen have the potential to advance areas of machine learning and bioengineering and help with the design of small molecule binders and enzymes.

Methods

Overview of PocketGen

Unlike previous methods focusing on protein sequence or structure generation, we aim to co-design both residue types (sequences) and 3D structures of the protein pocket that can fit and bind with target ligand molecules. Inspired by previous works on structure-based drug design[79,81] and protein generation[34,35], we formulate pocket generation in PocketGen as a conditional generation problem that generates the sequences and structures of pocket conditioned on the protein scaffold (other parts of the protein except the pocket region) and the binding ligand. To be specific, let $\mathcal{A} = \boldsymbol{a}_1 \cdots \boldsymbol{a}_{N_s}$ denote the whole protein sequence of residues, where N_s is the length of the sequence. The 3D structure of the protein can be described as a point cloud of protein atoms $\{\boldsymbol{a}_{i,j}\}_{1\le i\le N_s, 1\le j\le n_i}$ and let $\boldsymbol{x}(\boldsymbol{a}_{i,j}) \in \mathbb{R}^3$ denote the 3D coordinate of protein atoms. n_i is the number of atoms in a residue determined by the residue types. The first four atoms in any residue correspond to its backbone atoms (C_α, N, C, O), and the rest are the side-chain atoms. The ligand molecule can also be represented as a 3D point cloud $\mathcal{M} = \{\boldsymbol{v}_k\}_{k=1}^{N_l}$ where $\boldsymbol{v}_k$ denotes the atom feature. Let $\boldsymbol{x}(\boldsymbol{v}_k)$ denotes the 3D coordinates of atom $\boldsymbol{v}_k$. Our work defines the protein pocket as a set of residues in the protein closest to the binding ligand molecule: $\mathcal{B} = \boldsymbol{b}_1 \cdots \boldsymbol{b}_m$. The pocket $\mathcal{B}$ can thus be represented as an amino acid subsequence of a protein: $\mathcal{B} = \boldsymbol{a}_{e_1} \cdots \boldsymbol{a}_{e_m}$ where $\boldsymbol{e} = \{e_1, \cdots, e_m\}$ is the index of the pocket residues in the whole protein. The index $\boldsymbol{e}$ can be formally given as: $\boldsymbol{e} = \{i \mid \min_{1\le j\le n_i, 1\le k\le N_l} \|\boldsymbol{x}(\boldsymbol{a}_{i,j}) - \boldsymbol{x}(\boldsymbol{v}_k)\|_2 \le \delta\}$, where $\|\cdot\|_2$ is the L_2 distance norm and δ is the distance threshold. According to the distance range of pocket-ligand interactions[45], we set $\delta = 3.5$ Å in the default setting. With the above-defined notations, PocketGen aims to learn a conditional generative model formally defined as :

$$P(\mathcal{B} | \mathcal{A} \setminus \mathcal{B}, \mathcal{M}), \tag{1}$$

where $\mathcal{A} \setminus \mathcal{B}$ denotes the other parts of the protein except the pocket region. We also adjust the structure ligand molecule $\mathcal{M}$ in PocketGen to encourage protein-ligand interactions and reduce steric clashes.

To effectively generate the structure and the sequence of the protein pocket $\mathcal{B}$, the equivariant bilevel graph transformer and the sequence refinement module with pretrained protein language models and adapters are proposed, which will be discussed in the following paragraphs. The illustrative workflow is depicted in Fig. 1.

Equivariant bilevel graph transformer

It is critical to model the complex interactions in the protein pocket-ligand complexes for pocket generation. However, the multi-granularity (e.g., atom-level and residue-level) and multi-aspect (intra-protein and protein-ligand) nature of interactions brings a lot of challenges. Inspired by recent works on hierarchical graph transformer[81] and generalist equivariant transformer[83], we propose a novel equivariant bilevel graph transformer to well model the multi-granularity and multi-aspect interactions. Each residue or ligand is represented as a block (i.e., a set of atoms) for the conciseness of representation and ease of computation. Then the protein-ligand complex can be abstracted as a geometric graph of sets $\mathcal{G} = (\mathcal{V}, \mathcal{E})$, where $\mathcal{V} = \{\boldsymbol{H}_i, \boldsymbol{X}_i | 1 \le i \le B\}$ denotes the blocks and $\mathcal{E} = \{e_{ij} | 1 \le i, j \le B\}$ include all the edges between blocks. We added self-loops to the edges to capture interactions within the block (e.g., the interactions between ligand atoms). Our model adaptively assigns different numbers of channels to $\boldsymbol{H}_i$ and $\boldsymbol{X}_i$ to accommodate different numbers of atoms in residues and ligands. For example, given a block with n_i atoms, the corresponding block has $\boldsymbol{H}_i \in \mathbb{R}^{n_i \times d_h}$ indicating the atom features (d_h is the feature dimension size) and $\boldsymbol{X}_i \in \mathbb{R}^{n_i \times 3}$ denoting the atom coordinates. Specifically, the p-th row of H_i and $\boldsymbol{X}_i$ corresponds to the p-th atom's trainable feature (i.e., $\boldsymbol{H}_i[p]$) and coordinates (i.e., $\boldsymbol{X}_i[p]$) respectively. The trainable feature $\boldsymbol{H}_i[p]$ is first initialized with the concatenation of atom type embedding, residue/ligand embeddings, and the atom positional embeddings. To build $\mathcal{E}$, we connect the k-nearest neighboring residues according to the pairwise C_α distances. To reflect the interactions between the protein pocket and ligand, we add edges between all the pocket residue and the ligand block. We describe the modules in PocketGen's equivariant bilevel graph transformer, bilevel attention module, and equivariant feed-forward networks.

Bilevel attention module. Our model captures both **atom-level** and **residue/ligand-level** interactions with the bilevel attention module. Firstly, given two block i and j connected by an edge e_{ij}, we obtain the query, the key, and the value matrices with the following transformations:

$$\boldsymbol{Q}_i = \boldsymbol{H}_i \boldsymbol{W}_Q, \qquad \boldsymbol{K}_j = \boldsymbol{H}_j \boldsymbol{W}_K, \qquad \boldsymbol{V}_j = \boldsymbol{H}_j \boldsymbol{W}_V, \tag{2}$$

where $\boldsymbol{W}_Q, \boldsymbol{W}_K, \boldsymbol{W}_V \in \mathbb{R}^{d_h \times d_r}$ are trainable parameters.

To calculate the **atom-level attention** across the i-th and j-th block, we denote $\boldsymbol{X}_{ij} \in \mathbb{R}^{n_i \times n_j \times 3}$ and $\boldsymbol{D}_{ij} \in \mathbb{R}^{n_i \times n_j}$ as the relative coordinates and distances between atom pairs in block i and j, namely, $\boldsymbol{X}_{ij}[p,q] = \boldsymbol{X}_i[p] - \boldsymbol{X}_j[q], \boldsymbol{D}_{ij}[p,q] = \|\boldsymbol{X}_{ij}[p,q]\|_2$.

Then we have:

$$\boldsymbol{R}_{ij} = \frac{1}{\sqrt{d_r}} \left(\boldsymbol{Q}_i \boldsymbol{K}_j^\top \right) + \sigma_D \left(\mathrm{RBF}(\boldsymbol{D}_{ij}) \right), \tag{3}$$

$$\boldsymbol{\alpha}_{ij} = \mathrm{Softmax} \left(\boldsymbol{R}_{ij} \right), \tag{4}$$

where $\sigma_D(\cdot)$ is a Multi-Layer Perceptron (MLP) that adds distance bias to the attention calculation. RBF embeds the distance with radial basis functions. $\boldsymbol{\alpha}_{ij} \in \mathbb{R}^{n_i \times n_j}$ is the atom-level attention matrix obtained by applying row-wise Softmax on $\boldsymbol{R}_{ij} \in \mathbb{R}^{n_i \times n_j}$. To encourage sparsity in the attention matrix, we keep the top-k' elements of each row in $\boldsymbol{\alpha}_{ij}$ and set the others as zeros.

The **residue/ligand-level attention** from the j-th block to the i-th block is calculated as:

$$r_{ij} = \frac{\mathbf{1}^\top \boldsymbol{R}_{ij} \mathbf{1}}{n_i n_j}, \tag{5}$$

$$\beta_{ij} = \frac{\exp(r_{ij})}{\sum_{j \in \mathcal{N}(i)} \exp(r_{ij})}, \tag{6}$$

where $\mathbf{1}$ refers to the column vector with all elements set as ones and $\mathcal{N}(i)$ denotes the neighboring blocks of i. r_{ij} sums up all values in $\boldsymbol{R}_{ij}$ to represent the overall correlation between blocks i and j. Subsequently, β_{ij} denotes the attention across blocks at the block level.

We can update the representations and coordinates using the above atom-level and the residue/ligand-level attentions. PocketGen only updates the coordinates of the residues in the pocket and the ligand molecule. The other protein residues are fixed. Specifically, for the p-th atom in block i:

$$m_{ij,p} = \beta_{ij} \left(\boldsymbol{\alpha}_{ij}[p] \odot \phi_x(\boldsymbol{Q}_i[p] || \boldsymbol{K}_j || \mathrm{RBF}(\boldsymbol{D}_{ij}[p])) \right), \tag{7}$$

$$\boldsymbol{H}'_i[p] = \boldsymbol{H}_i[p] + \sum_{j \in \mathcal{N}(i)} \beta_{ij} \phi_h(\boldsymbol{\alpha}_{ij}[p] \cdot \boldsymbol{V}_j), \tag{8}$$

$$\boldsymbol{X}'_i[p] = \boldsymbol{X}_i[p] + \begin{cases} \sum_{j \in \mathcal{N}(i)} m_{ij,p} \cdot \boldsymbol{X}_{ij}[p], & \text{if } i \text{ belongs to ligand or pocket residues} \\ 0, & \text{if } i \text{ belongs to other protein residues} \end{cases} \tag{9}$$

where ϕ_h and ϕ_x are MLPs with concatenated representations as input (concatenation along the second dimension and $\boldsymbol{Q}_i[p]$ is repeated along rows). $\odot$ computes the element-wise multiplication. $\boldsymbol{H}'_i$ and $\boldsymbol{X}'_i$ denote the updated representation and coordinate matrices, and we can verify that the dimension size of $\boldsymbol{H}'_i$ and $\boldsymbol{X}'_i$ remains the same regardless of the neighboring block size n_j. Furthermore, as the attention coefficients $\boldsymbol{\alpha}_{ij}$ and β_{ij} are invariant under E(3) transformations, the modification of $\boldsymbol{X}'_i$ adheres to E(3)-equivariance. Additionally, the permutation of atoms within each block does not affect this update process.

Equivariant feed-forward network. We adapted the feed-forward network module (FFN) in the transformer model[84] to update $\boldsymbol{H}_i$ and $\boldsymbol{X}_i$. Specifically, the representation and coordinates of atoms are updated to consider the block's feature/geometric centroids (means). The centroids are denoted as:

$$\boldsymbol{h}_c = \mathrm{centroid}(\boldsymbol{H}_i), \qquad \boldsymbol{x}_c = \mathrm{centroid}(\boldsymbol{X}_i), \tag{10}$$

Then we obtain the relative coordinate $\Delta \boldsymbol{x}_p$ and the relative distance representation $\boldsymbol{r}_p$ based on the L2 norm of $\Delta \boldsymbol{x}_p$:

$$\Delta \boldsymbol{x}_p = \boldsymbol{X}_i[p] - \boldsymbol{x}_c, \qquad \boldsymbol{r}_p = \mathrm{RBF}(\|\Delta \boldsymbol{x}_p\|_2), \tag{11}$$

The representation and coordinates of atoms are updated with MLPs σ_h and σ_x. The centroids are integrated to inform of the context of the block:

$$\boldsymbol{H}'[p] = \boldsymbol{H}[p] + \sigma_h(\boldsymbol{H}_i[p], \boldsymbol{h}_c, \boldsymbol{r}_p), \tag{12}$$

$$\boldsymbol{X}'_i[p] = \boldsymbol{X}_i[p] + \Delta \boldsymbol{x}_p \sigma_x(\boldsymbol{H}_i[p], \boldsymbol{h}_c, \boldsymbol{r}_p). \tag{13}$$

To stabilize and accelerate training, layer normalization[85] is appended at each layer of the equivariant bilevel graph transformer to normalize $\boldsymbol{H}$. The equivariant feed-forward network satisfies E(3)-equivariance. Thanks to each module's E(3)-equivariance, the whole proposed bilevel graph transformer has the desirable property of E(3)-equivariance (Theorem 1 in Supplementary Information shows the details). In PocketGen, we use E(3) equivariant model for its simplicity similar to previous works[86,87], which is capable enough to achieve strong performance. We are aware that an SE(3) equivariant model architecture would be better for learning the chirality-related properties of the protein, which we left for future exploration.

Sequence refinement with pretrained protein language models and adapters

Protein language models (pLMs), such as the ESM family of models[40,41], have learned extensive evolutionary knowledge from the vast array of natural protein sequences, demonstrating a strong ability to design protein sequences. In PocketGen, we propose to leverage pLMs to help refine the designed protein pocket sequences. To infuse the pLMs with structural information, we implant lightweight structural adapters inspired by previous works[88,89]. Different from LM-Design[89] which focuses on protein sequence design given fixed backbone structure, PocketGen codesigns both the amino acid sequence as well as the full atom structure of the protein pocket. In our default setting, only one structural adapter was placed after the last layer of pLM. Only the adapter layers are fine-tuned during training, and the other layers of PLMs are frozen to save computation costs. The structural adapter mainly has the following two parts.

Structure-sequence cross attention. The structural representation of the i-th residue $\boldsymbol{h}_i^{\text{struct}}$ is obtained by mean pooling of $\boldsymbol{H}_i$ from the bilevel graph transformer. In the input to the pLMs, the pocket residue types to be designed are assigned with the mask, and we denote the i-th residue representation from pLMs as $\boldsymbol{h}_i^{\text{seq}}$. In the structural adapter, we perform cross-attention between the structural representations $\boldsymbol{H}^{\text{struct}} = \{\boldsymbol{h}_1^{\text{struct}}, \boldsymbol{h}_2^{\text{struct}}, \cdots, \boldsymbol{h}_{N_s}^{\text{struct}}\}$ and sequence representations $\boldsymbol{H}^{\text{seq}} = \{\boldsymbol{h}_1^{\text{seq}}, \boldsymbol{h}_2^{\text{seq}}, \cdots, \boldsymbol{h}_{N_s}^{\text{seq}}\}$. The query, key, and value matrices are obtained as follows:

$$\boldsymbol{Q} = \boldsymbol{H}^{\text{seq}}\boldsymbol{W}_Q, \qquad \boldsymbol{K} = \boldsymbol{H}^{\text{struct}}\boldsymbol{W}_K, \qquad \boldsymbol{V} = \boldsymbol{H}^{\text{struct}}\boldsymbol{W}_V, \tag{14}$$

where $\boldsymbol{W}_Q, \boldsymbol{W}_K, \boldsymbol{W}_V \in \mathbb{R}^{d_h \times d_r}$ are trainable weight matrices. Rotary positional encoding[90] is applied to the representations, and we omit it in the equations for simplicity. The output of the cross attention is obtained as:

$$\text{CrossAttention}(\boldsymbol{Q}, \boldsymbol{K}, \boldsymbol{V}) = \text{Softmax}\left(\frac{\boldsymbol{Q}\boldsymbol{K}^\top}{\sqrt{d_r}}\right)\boldsymbol{V}. \tag{15}$$

Bottleneck feed-forward network. A bottleneck feed-forward network (FFN) is appended after the cross-attention to impose non-linearity and abstract representations, inspired by previous works such as Houlsby et al.[88]. The intermediate dimension of the bottleneck FFN is set to be half of the default representation dimension. Finally, the predicted pocket residue type $\boldsymbol{p}_i$ is obtained using an MLP on the output residue representation.

Training protocol

Inspired by AlphaFold2[39], we use a recycling strategy for model training. Recycling facilitates the training of deeper networks without incurring extra memory costs by executing multiple forward passes and computing gradients solely for the final pass. The training loss of PocketGen is the weighted sum of the following three losses:

$$\mathcal{L}_{\text{seq}} = \frac{1}{T}\sum_t \sum_i l_{\text{ce}}(\hat{\boldsymbol{p}}_i, \boldsymbol{p}_i^t); \tag{16}$$

$$\mathcal{L}_{\text{coord}} = \frac{1}{T}\sum_t \left[\sum_i l_{\text{huber}}(\hat{\boldsymbol{X}}_i, \boldsymbol{X}_i^t) + \sum_j l_{\text{huber}}(\hat{\boldsymbol{x}}(\boldsymbol{v}_j), \boldsymbol{x}^t(\boldsymbol{v}_j))\right]; \tag{17}$$

$$\mathcal{L}_{\text{struct}} = \frac{1}{T}\sum_t \left[\sum_{b\in\mathcal{B}} l_{\text{huber}}(\hat{b}, b^t) + \sum_{\theta\in\Theta} l_{\text{huber}}(\cos\hat{\theta}, \cos\theta^t)\right]; \tag{18}$$

$$\mathcal{L} = \mathcal{L}_{\text{seq}} + \lambda_{\text{coord}}\mathcal{L}_{\text{coord}} + \lambda_{\text{struct}}\mathcal{L}_{\text{struct}}, \tag{19}$$

where T is the total refinement rounds. $\hat{\boldsymbol{p}}_i, \hat{\boldsymbol{X}}_i, \hat{\boldsymbol{x}}(\boldsymbol{v}_j), \hat{b}$, and $\cos\hat{\theta}$ are the ground-truth residue types, residue coordinates, and ligand coordinates, bond lengths, and bond/dihedral angles; $\boldsymbol{p}_i^t, \boldsymbol{X}_i^t, \boldsymbol{x}^t(\boldsymbol{v}_j), b^t$, and $\cos\theta^t$ are the predicted ones at the t-th round by PocketGen. The sequence loss $\mathcal{L}_{\text{seq}}$ is the cross-entropy loss for pocket residue type prediction; the coordinate loss $\mathcal{L}_{\text{coord}}$ uses huber loss[91] for the training stability; the structure loss $\mathcal{L}_{\text{struct}}$ is added to supervised bond lengths and bond/dihedral angles for realistic local geometry. $\mathcal{B}$ and Θ denote all the bonds and angles in the protein pocket (including side chains). λ_{coord}, and λ_{struct} are hyperparameters balancing the three losses. We perform a grid search over $\{0.5, 1.0, 2.0, 3.0\}$ and choose these hyperparameters based on the validation performance to select the specific parameter values. In the default setting, we set λ_{coord} to 1.0 and λ_{struct} to 2.0.

Generation protocol

In the generation procedure, PocketGen initializes the sequence with uniform distributions over 20 amino acid types and the coordinates based on linear interpolations and extrapolations. Specifically, we initialize the residue coordinates with linear interpolations and extrapolations based on the nearest residues with known structures in the protein. Denote the

sequence of residues as $\mathcal{A} = \boldsymbol{a}_1 \cdots \boldsymbol{a}_{N_s}$, where N_s is the length of the sequence. Let $\boldsymbol{x}(\boldsymbol{a}_{i,1}) \in \mathbb{R}^3$ denote the C_α coordinate of the i-th residue. We take the following strategies to determine the C_α coordinate of the i-th residue: (1) We use linear interpolation if there are residues with known coordinates at both sides of the i-th residue. Specifically, assume p and q are the indexes of the nearest residues with known coordinates at each side of the i-th residue ($p < i < q$), we have: $\boldsymbol{x}(\boldsymbol{a}_{i,1}) = \frac{1}{q-p}[(i-p)\boldsymbol{x}(\boldsymbol{a}_{q,1}) + (q-i)\boldsymbol{x}(\boldsymbol{a}_{p,1})]$. (2) We conduct linear extrapolation if the i-th residue is at the ends of the chain, i.e., no residues with known structures at one side of the i-th residue. Specifically, let p and q denote the index of the nearest and the second nearest residue with known coordinates. The position of the i-th residue can be initialized as $\boldsymbol{x}(\boldsymbol{a}_{i,1}) = \boldsymbol{x}(\boldsymbol{a}_{p,1}) + \frac{i-p}{p-q}(\boldsymbol{x}(\boldsymbol{a}_{p,1}) - \boldsymbol{x}(\boldsymbol{a}_{q,1}))$. Inspired by previous works[33,34], we initialize the other backbone atom coordinates according to their ideal local coordinates relative to the C_α coordinates. We initialize the side-chain atoms' coordinates with the coordinate of their corresponding C_α, added with Gaussian noise. We initialize the ligand molecular structure with the reference ligand structure from the dataset. The ligand structure is updated during pocket generation and the updated ligand is used for Vina score calculation.

Since the number of pocket residue types and the number of side chain atoms are unknown at the beginning of generation, each pocket residue is assigned 14 atoms, the maximum number of atoms for residues. After rounds of refinement by PocketGen, the pocket residue types are predicted, and the full atom coordinates are determined by mapping the coordinates to the predicted residue types (taking the first n coordinates according to residue type). In PocketGen, we directly predict the absolute atom coordinates, which reduces the model complexity and flexibly captures atom interactions. We also notice PocketGen aligns with the recent trend of directly predicting full atom coordinates. For example, the recent AlphaFold3[92] directly predicts the full atom coordinates, replacing the AlphaFold2 structure module that operated on amino-acid-specific frames and side-chain torsion angles, and achieves better performance on protein structure prediction. For generation efficiency, we set the number of refinement rounds to 3.

Experimental setting

Datasets. We consider two widely used datasets for benchmark evaluation: **CrossDocked** dataset[42] contains 22.5 million protein-molecule pairs generated through cross-docking. Following previous works[24,60,93], we filter out data points with binding pose RMSD greater than 1 Å, leading to a refined subset with around 180k data points. For data splitting, we use mmseqs2[94] to cluster data at 30% sequence identity, and randomly draw 100k protein-ligand structure pairs for training and 100 pairs from the remaining clusters for testing and validation, respectively; **Binding MOAD** dataset[43] contains around 41k experimentally determined protein-ligand complexes. Following previous work[95], we keep pockets with valid and moderately 'drug-like' ligands with QED score ≥ 0.3. We further filter the dataset to discard molecules containing atom types $\notin \{C, N, O, S, B, Br, Cl, P, I, F\}$ as well as binding pockets with non-standard amino acids. Then, we randomly sample and split the filtered dataset based on the Enzyme Commission Number (EC Number)[44] to ensure different sets do not contain proteins from the same EC Number main class. Finally, we have 40k protein-ligand pairs for training, 100 pairs for validation, and 100 pairs for testing. For all the benchmark tasks in this paper, PocketGen and all the other baseline methods are trained with the same data split for a fair comparison. In real-world pocket generation and optimization case studies, the protein structures were downloaded from PDB[96].

Implementation. Our PocketGen model is trained with Adam[97] optimizer for 5k iterations, where the learning rate is 0.0001, and the batch size is 64. We report the results corresponding to the checkpoint with the best validation loss. It takes around 48 hours to finish training on 1 Tesla A100 GPU from scratch. In PocketGen, the number of attention heads is set as 4; the hidden dimension d is set as 128; k is set to 8 to connect the k-nearest neighboring residues to build $\mathcal{E}$; k' is set as 3 to encourage sparsity in the attention matrix. For all the benchmark tasks of pocket generation and optimization, PocketGen and all the other baseline methods are trained with the same data split for a fair comparison. We follow the implementation codes provided by the authors to obtain the results of baseline methods. Algorithm 1 and 2 in the supplementary show the pseudo-codes of the training and generation process of PocketGen.

Baseline methods. PocketGen is compared with five state-of-the-art representative baseline methods. **PocketOptimizer**[18] is a physics-based method that optimizes energies such as packing and binding-related energies for ligand-binding protein design. Following the suggestion of the paper, we fixed the backbone structures. **DEPACT**[17] is a template-matching method that follows a two-step strategy[98] for pocket design. It first searches the protein-ligand complexes in the database with similar ligand fragments. It then grafts the associated residues into the protein scaffold to output the complete protein structure with PACMatch[17]. Both the backbone and the sidechain structures are changed in DEPACT. **RFDiffusion**[26], **RFDiffusionAA**[16], **FAIR**[24], and **dyMEAN**[25] are deep-learning-based models that for protein generation. RFDiffusion does not explicitly model protein-ligand interactions and is not directly applicable to small molecule-binding protein generation. Following the suggestions in RFDiffusion[26] and RFDiffusionAA[16], we use a heuristic attractive-repulsive potential to encourage the formation of pockets with shape complementarity to a target molecule. The residue sequence for the generated protein by RFDiffusion is derived with ProteinMPNN, and the side-chain conformation is decided with Rosetta[99] side-chain packing. RFDiffusionAA is the latest

version of RFDiffusion, which can directly generate protein structures surrounding small molecules by combining residue-based representation of amino acids with atomic representation of small molecules. For RFDiffusion and RFDiffusionAA, we let them in paint the pocket area to obtain a consistent setting with other methods for comparison. We also note that RFDiffusion and RFDiffusionAA do not provide the training/finetuning scripts, so we use the provided pre-trained checkpoints for all the related experiments in our paper. FAIR[24] was specially designed for full-atom protein pocket design via iterative refinement. dyMEAN[25] was originally proposed for full atom antibody design, and we adapted it to our pocket design task with proper modifications. Detailed information on baselines is included in Supplementary Notes. The setting of the key hyperparameters is summarized in Table. S6. All the baselines are run on the same Telsa A100 GPU for a fair comparison with our PocketGen.

Data availability

This study's training and test data are available at Zenodo (https://zenodo.org/records/10125312). The project website for PocketGen is at https://zitniklab.hms.harvard.edu/projects/PocketGen.

Code availability

The source code of this study is freely available at GitHub (https://github.com/zaixizhang/PocketGen) to allow for replication of the results of this study.

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Author contributions statement

Z.X.Z., Q.L., and M.Z. designed the research, Z.X.Z. conducted the experiments, Z.X.Z., Q.L., and M.Z. analyzed the results. Z.X.Z., W.X.S., Q.L., and M.Z. wrote the manuscript. All authors reviewed the manuscript.

Competing interests

The authors declare no competing interests.

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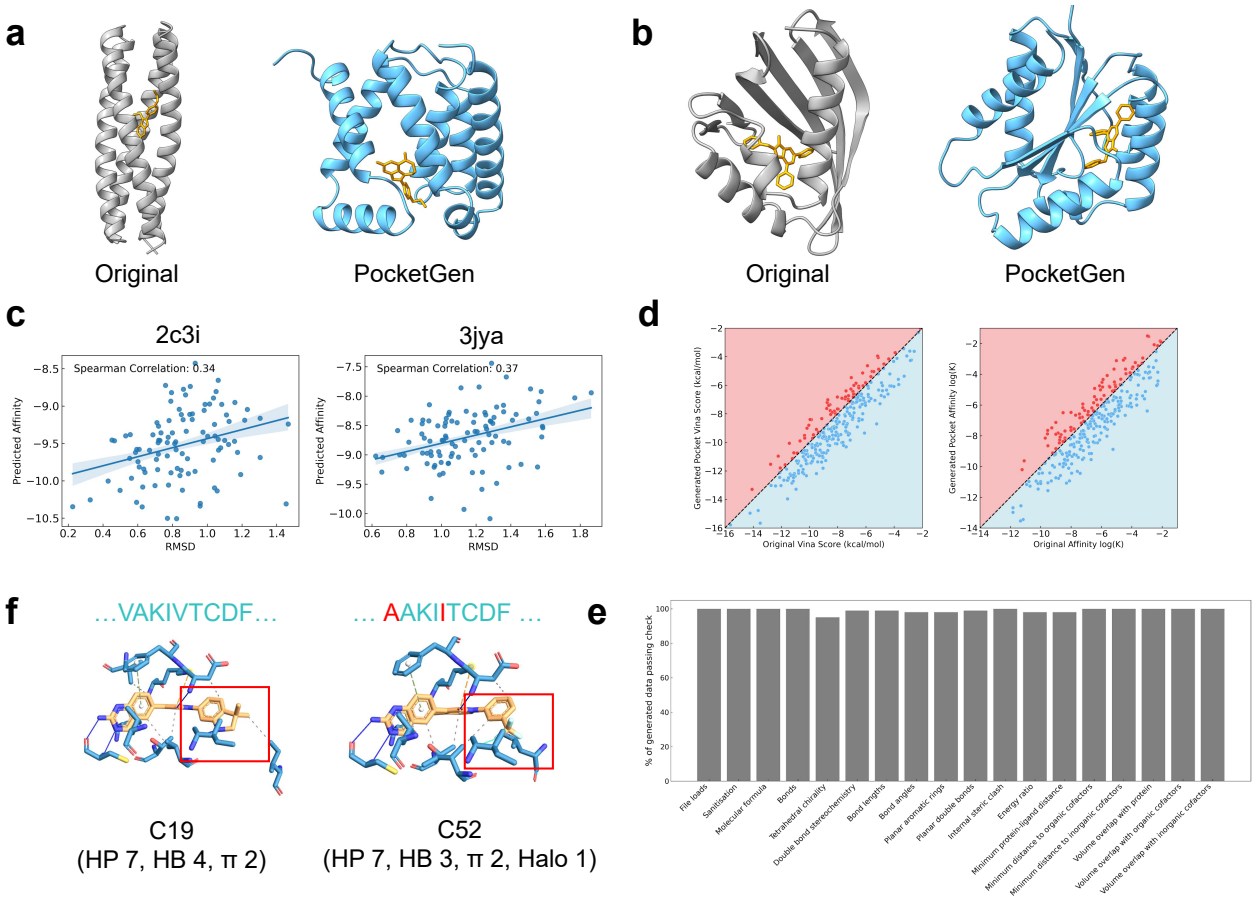


Extended Data Fig. 1: More case studies and evaluations of PocketGen. **a**, The originally designed protein binder for Rucaparib[21] (left panel) and the generated protein binder by PocketGen (right panel). **b**, The originally designed protein binder for DTZ[8] (left panel) and the generated protein binder by PocketGen (right panel). Note that PocketGen generates the whole protein instead of the pocket region in **a**&**b**. The generated protein binder has high scTM scores (0.900 and 0.976). **c**, The predicted affinity (log K) by GIGN[100] of the generated pockets by PocketGen with respect to RMSD. We randomly select two protein-ligand complexes from PDBBind (PDB id 2c3i and 3jya). **d**, The Vina score/binding affinity (log K) of the generated pockets by PocketGen and the original pockets from PDBBind. The black region/dots indicate the generated pockets have higher affinities than the original pockets while the red region/dots indicate lower affinities. **f**, The generated interactions by PocketGen with respect to a pair of activity cliff ligand molecules, i.e., C19 and C52[77]. As marked with red rectangles, PocketGen adaptively generates different interactions for different molecular fragments (hydrogen bonds+hydrophobic interactions and halogen bonds respectively). 'HP' indicates hydrophobic interactions, 'HB' signifies hydrogen bonds, 'π' denotes the π-stacking/cation interactions, and 'Halo' indicates the Halogen bonds. **e**, Detailed validity check with PoseBusters on CrossDocked and Binding MOAD.

Supplementary Information

Supplementary Notes

Training and Generation Algorithms

Algorithm 1 Training Algorithm of PocketGen

Input: protein sequences $\mathcal{A} = \boldsymbol{a}_1 \cdots \boldsymbol{a}_{N_s}$ and structures $\{\boldsymbol{x}(\boldsymbol{a}_i)\}_{i=1}^{N_s}$, ligand molecule $\mathcal{M}$, total iteration rounds T.
Initial: Initialize the coordinates of pocket residues $\{\boldsymbol{x}(\boldsymbol{b}_i)\}_{i=1}^{m}$. Initialize the sequences of pocket residues with uniform distributions over 20 residue categories. Initialize the residue/ligand representations $\{\boldsymbol{H}_i\}$.

while Model training is not converged **do**
  # Recycling training
  for $t = 1, \cdots T$ **do**
    Obtain the residue representations $\{\boldsymbol{H}_i\}$ and updated coordinates $\{\boldsymbol{X}_i\}$ with the equivariant bilevel graph transformer.
    Predict residue types $\boldsymbol{p}_i^t$ with structure refinement modules.
    if $t == T$ **then**
      $\mathcal{L}_{\text{seq}} = \sum_i l_{\text{ce}}(\hat{\boldsymbol{p}}_i, \boldsymbol{p}_i^t)$.
      $\mathcal{L}_{\text{coord}} = \sum_i l_{\text{huber}}(\hat{\boldsymbol{X}}_i, \boldsymbol{X}_i^t) + \sum_j l_{\text{huber}}(\hat{\boldsymbol{x}}(\boldsymbol{v}_j), \boldsymbol{x}^t(\boldsymbol{v}_j))$.
      $\mathcal{L}_{\text{struct}} = \sum_{b \in \mathcal{B}} l_{\text{huber}}(\hat{b}, b^t) + \sum_{\theta \in \Theta} l_{\text{huber}}(\cos\hat{\theta}, \cos\theta^t)$
    end if
  end for
  Minimize $\lambda_{\text{seq}}\mathcal{L}_{\text{seq}} + \lambda_{\text{coord}}\mathcal{L}_{\text{coord}} + \lambda_{\text{struct}}\mathcal{L}_{\text{struct}}$.
end while

Algorithm 2 Generation Algorithm of PocketGen

Input: protein sequences $\mathcal{A} = \boldsymbol{a}_1 \cdots \boldsymbol{a}_{N_s}$ and structures $\{\boldsymbol{x}(\boldsymbol{a}_i)\}_{i=1}^{N_s}$, ligand molecule $\mathcal{M}$, total iteration rounds T.
Initial: Initialize the coordinates of pocket residues $\{\boldsymbol{x}(\boldsymbol{b}_i)\}_{i=1}^{m}$. Initialize the sequences of pocket residues with uniform distributions over 20 residue categories. Initialize the residue/ligand representations $\{\boldsymbol{H}_i\}$.

for $t = 1, ..., T$ **do**
  Obtain the residue representations $\{\boldsymbol{H}_i\}$ and updated coordinates $\{\boldsymbol{X}_i\}$ with the equivariant bilevel graph transformer.
  Predict residue types $\boldsymbol{p}_i^t$ with structure refinement modules.
end for
Map $\{\boldsymbol{X}_i\}$ to the predicted residue types.
Output designed pockets.

Loss Functions

In Equation 19, we use Huber loss[91] in $\mathcal{L}_{\text{coord}}$ and $\mathcal{L}_{\text{struct}}$ for the stability of optimization, which are defined as follows:

$$l_{huber}(x, y) = \begin{cases} 0.5\ (x-y)^2,\ if\ |x-y| < \epsilon, \\ \epsilon \cdot (|x-y| - 0.5 \cdot \epsilon),\ else, \end{cases} \tag{20}$$

where x and y represent the predicted and ground-truth coordinates/ bond length/ angles. The Huber loss has the following property: if the L1 norm of $|x-y|$ is smaller than ϵ, it is MSE loss, otherwise it is L1 loss. At the beginning of the model training, the deviation between the predicted and ground-truth coordinates is large, and the L1 term makes the loss less sensitive to outliers than MSE loss. The deviation is small when the training is almost complete, and the MSE loss is applied for further finetuning. In practice, we find that directly using MSE loss sometimes leads to instability at the beginning of the training (e.g., very large gradient norm), while Huber loss makes the training procedure more stable. Following the suggestion of previous works[32,34], we set $\epsilon = 1$ in all our experiments.

Implementation Details of Baseline Methods and Hyperparameter Settings

PocketOptimizer[18] [1] is a physic-based computational protein design method that predicts mutations in the binding pockets of proteins to increase affinity for a specific ligand. We use the latest version, i.e., PocketOptimizer 2.0[18]. There are generally four main steps in PocketOptimizer: structure preparation, flexibility sampling, energy calculations, and computation of design solutions. Specifically for the energy calculations, both packing-related energies and binding-related energies are considered. Following the suggestions in the original paper, we use AMBER ff14S force field[101] for energy computation and the Dunbrack rotamer library[102] for rotamer sampling for PocketOptimizer in our implementation. We fixed the backbone structures following the suggestions of the original paper. As for the output design solutions, we select the top 100 designs identified by PocketOptimizer based on integer linear programming for downstream metric calculations.

DEPACT[17] [2] is a template-matching method that follows a two-step strategy for pocket design. Firstly, it searches the protein-ligand complexes in the template database with similar ligand fragments and constructs a cluster model (a set of pocket residues). The template databases are constructed based on the corresponding training datasets for fair comparisons. Secondly, it grafts the cluster model into the protein pocket with PACMatch. It works by placing residues from the cluster model on protein scaffolds by matching the atoms of residues with atoms of the protein scaffold. The backbone coordinates of the pocket residues are also modified in the process. The qualities of the generated pockets are evaluated and ranked based on a statistical scoring function. We take the top 100 designed pockets for evaluation. The output of DEPACT+PACMatch is complete protein structures with redesigned pockets. In the paper, we only use DEPACT to represent the whole method of DEPACT+PACMatch for conciseness.

RFDiffusion[26] [3] is one of the state-of-the-art method for *de novo* protein backbone generation. It combines the RoseTTAFold structure prediction network with the diffusion probabilistic models (DDPMs) framework. To model the influence of ligand molecules, we use a heuristic attractive-repulsive potential to encourage the formation of pockets with shape complementarity to a target molecule following the suggestions of RFDiffusion[26] and RFDiffusionAA[16]. The residue sequence is further decided with ProteinMPNN[29], and the side-chain conformation is added with Rosetta[99] side-chain packing.

RFDiffusionAA[16] [4] is the latest version of RFDiffusion which combines a residue-based representation of amino acids and atomic representations of all other groups to model protein-small molecules/metals/nucleic acids/covalent modification complexes. Starting from random distributions of amino acid residues surrounding target small molecules, RFDiffusionAA can directly generate the small molecule binding protein backbone. Furthermore, with LigandMPNN[30], the latest version of ProteinMPNN[29], we can assign residue types and predict sidechain conformations considering the protein-ligand interactions. Experiments in RFDiffusionAA[16] show that the generated protein by RFDiffusionAA has better binding affinity than those obtained by RFDiffusion with auxiliary potential.

dyMEAN[25] [5] is an end-to-end full-atom model for E(3)-equivariant antibody design given the epitope and the incomplete sequence of the antibody. Its previous version, MEAN[34], only considers the backbone atoms, while dyMEAN considers the complete atom structure and performs better on downstream tasks. Generally, dyMEAN co-designs antibody sequence and structure via a multi-round progressive full-shot refinement manner, which is more efficient than auto-regressive or diffusion-based approaches. An adaptive multi-channel equivariant encoder is used in dyMEAN, which can process protein residues of variable sizes when considering full atoms. To adapt dyMEAN to our pocket design task, we replace the antigen with the target ligand molecule to provide the context information for pocket generation.

FAIR[24] [6] is our previous method for full atom pocket sequence-structure co-design. FAIR operates in two steps, proceeding in a coarse-to-fine manner (backbone refinement to full atoms refinement, including side chains) for full-atom generation. In FAIR, residue types and atom coordinates are updated using a hierarchical graph transformer composed of a residue-level and atom-level encoder.

[1] https://github.com/Hoecker-Lab/pocketoptimizer
[2] https://github.com/chenyaoxi/DEPACT_PACMatch
[3] https://github.com/RosettaCommons/RFdiffusion
[4] https://github.com/baker-laboratory/rf_diffusion_all_atom
[5] https://github.com/THUNLP-MT/dyMEAN
[6] https://github.com/zaixizhang/FAIR

Proof of E(3)-Equivariance of PocketGen

The E(3)-transformation on the euclidean coordinate $\boldsymbol{x} \in \mathbb{R}^3$ can be represented as: $T_g \cdot \boldsymbol{x} = \boldsymbol{O}\boldsymbol{x} + \boldsymbol{t}$, where $\boldsymbol{O} \in \mathbb{R}^3$ is the orthogonal transformation matrix, $\boldsymbol{t} \in \mathbb{R}^3$ is the translation vector. Implementing T_g on a coordinate matrix $\boldsymbol{X} \in \mathbb{R}^{n\times 3}$ means transforming each coordinate (i.e., each row) with T_g. PocketGen has the desirable property of **E(3)-equivariance** as follow:

Theorem 1. *Denote the E(3)-transformation as T_g and the generative process of PocketGen as $\{(\boldsymbol{b}_i, \boldsymbol{x}(\boldsymbol{b}_i))\}_{i=1}^m = p_\theta(\mathcal{A}\setminus\mathcal{B}, \mathcal{M})$, where $\{(\boldsymbol{b}_i, \boldsymbol{x}(\boldsymbol{b}_i))\}_{i=1}^m$ indicates the designed protein pocket seqeuce and structure. We have $\{(\boldsymbol{b}_i, T_g \cdot \boldsymbol{x}(\boldsymbol{b}_i))\}_{i=1}^m = p_\theta(T_g \cdot (\mathcal{A}\setminus\mathcal{B}), T_g \cdot \mathcal{M})$. Here, $T_g \cdot (\mathcal{A}\setminus\mathcal{B})$ and $T_g \cdot \mathcal{M}$ denote applying E(3)-transformation on the protein structures except for the pocket region and the molecular structure respectively.*

Then we prove the E(3)-equivariance of each module in PocketGen as follows.

Lemma 1. *Denote the bilevel attention module as $\{\boldsymbol{H}'_i, \boldsymbol{X}'_i\} = \mathrm{Att}(\{\boldsymbol{H}_i, \boldsymbol{X}_i\})$, then it is E(3)-equivariant. Namely, for any E(3)-transformation T_g, we have $\{\boldsymbol{H}'_i, T_g \cdot \boldsymbol{X}'_i\} = \mathrm{Att}(\{\boldsymbol{H}_i, T_g \cdot \boldsymbol{X}_i\})$.*

Proof. The key to the proof of Lemma 1 is to prove that the propagation in Eq. 2-9 is E(3)-invariant on $\boldsymbol{H}_i$ and E(3)-equivariant on $\boldsymbol{X}_i$. The correlation $\boldsymbol{R}_{ij}$ between block i and block j in Eq. 3 is E(3)-invariant because all the inputs, including the query $\boldsymbol{Q}_i$, the key $\boldsymbol{K}_j$, and the distance matrices $\boldsymbol{D}_{ij}$, are not influenced by the geometric transformation T_g. Therefore, we can immediately derive that the atom-level attention $\boldsymbol{\alpha}_{ij}$ in Eq. 4 is E(3)-invariant. Similarly, the residue/ligand-level attention β_{ij} in Eq. 6 is E(3)-invariant because it only operates on r_{ij} in Eq. 5 which aggregates $\boldsymbol{\alpha}_{ij}$. Finally, we can derive the E(3)-invariance on $\boldsymbol{H}$ and the E(3)-equivariance on $\boldsymbol{X}$ (ligand or pocket residues) as below:

$$
\begin{aligned}
\boldsymbol{H}'_i[p] &= \boldsymbol{H}_i[p] + \sum_{j\in\mathcal{N}(i)} \beta_{ij}\phi_h(\boldsymbol{\alpha}_{ij}[p]\cdot\boldsymbol{V}_j),\\
T_g\cdot\boldsymbol{X}'_i[p] &= T_g\cdot(\boldsymbol{X}_i[p] + \sum_{j\in\mathcal{N}(i)} \beta_{ij}\left(\boldsymbol{\alpha}_{ij}[p]\odot\phi_x(\boldsymbol{Q}_i[p]||\boldsymbol{K}_j||\mathrm{RBF}(\boldsymbol{D}_{ij}[p]))\right)\cdot\boldsymbol{X}_{ij}[p])\\
&= \boldsymbol{O}(\boldsymbol{X}_i[p] + \sum_{j\in\mathcal{N}(i)} \beta_{ij}\left(\boldsymbol{\alpha}_{ij}[p]\odot\phi_x(\boldsymbol{Q}_i[p]||\boldsymbol{K}_j||\mathrm{RBF}(\boldsymbol{D}_{ij}[p]))\right)\cdot\boldsymbol{X}_{ij}[p]) + \boldsymbol{t}\\
&= (\boldsymbol{O}\boldsymbol{X}_i[p] + \boldsymbol{t}) + \sum_{j\in\mathcal{N}(i)} \beta_{ij}\left(\boldsymbol{\alpha}_{ij}[p]\odot\phi_x(\boldsymbol{Q}_i[p]||\boldsymbol{K}_j||\mathrm{RBF}(\boldsymbol{D}_{ij}[p]))\right)\cdot\begin{bmatrix}\boldsymbol{O}(\boldsymbol{X}_i[p]-\boldsymbol{X}_j[1])\\ \vdots\\ \boldsymbol{O}(\boldsymbol{X}_i[p]-\boldsymbol{X}_j[n_j])\end{bmatrix}\\
&= (\boldsymbol{O}\boldsymbol{X}_i[p] + \boldsymbol{t}) + \sum_{j\in\mathcal{N}(i)} \beta_{ij}\left(\boldsymbol{\alpha}_{ij}[p]\odot\phi_x(\boldsymbol{Q}_i[p]||\boldsymbol{K}_j||\mathrm{RBF}(\boldsymbol{D}_{ij}[p]))\right)\cdot\begin{bmatrix}\boldsymbol{O}\boldsymbol{X}_i[p]+\boldsymbol{t}-(\boldsymbol{O}\boldsymbol{X}_j[1]+\boldsymbol{t})\\ \vdots\\ \boldsymbol{O}\boldsymbol{X}_i[p]+\boldsymbol{t}-(\boldsymbol{O}\boldsymbol{X}_j[n_j]+\boldsymbol{t})\end{bmatrix}\\
&= T_g\cdot\boldsymbol{X}_i[p] + \sum_{j\in\mathcal{N}(i)} \beta_{ij}\left(\boldsymbol{\alpha}_{ij}[p]\odot\phi_x(\boldsymbol{Q}_i[p]||\boldsymbol{K}_j||\mathrm{RBF}(\boldsymbol{D}_{ij}[p]))\right)\cdot\begin{bmatrix}T_g\cdot\boldsymbol{X}_i[p]-T_g\cdot\boldsymbol{X}_j[1]\\ \vdots\\ T_g\cdot\boldsymbol{X}_i[p]-T_g\cdot\boldsymbol{X}_j[n_j]\end{bmatrix},
\end{aligned}
$$

which concludes the proof of Lemma 1. □

Lemma 2. *Denote the equivariant feed-forward network as as $\{\boldsymbol{H}'_i, \boldsymbol{X}'_i\} = \mathrm{FFN}(\{\boldsymbol{H}_i, \boldsymbol{X}_i\})$, then it is E(3)-equivariant. Namely, for any E(3)-transformation T_g, we have $\{\boldsymbol{H}'_i, T_g\cdot\boldsymbol{X}'_i\} = \mathrm{FFN}(\{\boldsymbol{H}_i, T_g\cdot\boldsymbol{X}_i\})$.*

Proof. The proof of Lemma 2 focuses on the single-atom updates in Eq. 10-13. First, it is easy to obtain the E(3)-equivariance of the centroid in Eq. 10:

$$T_g\cdot\boldsymbol{x}_c = T_g\cdot\mathrm{centroid}(\boldsymbol{X}_i) = \mathrm{centroid}(T_g\cdot\boldsymbol{X}_i).$$

Then we have the following equation on the relative coordinate $\Delta\boldsymbol{x}$ in Eq. 11:

$$\boldsymbol{O}\Delta\boldsymbol{x}_p = (\boldsymbol{O}\boldsymbol{X}_i[p]+\boldsymbol{t}) - (\boldsymbol{O}\boldsymbol{x}_c+\boldsymbol{t}) = T_g\cdot\boldsymbol{X}_i[p] - T_g\cdot\boldsymbol{x}_c.$$

Then we can obtain the E(3)-invariance of $\boldsymbol{r}_p$ in Eq. 11:

$$\boldsymbol{r}_p = \mathrm{RBF}(\|\Delta\boldsymbol{x}_p\|_2) = \mathrm{RBF}(\|\boldsymbol{X}_i[p]-\boldsymbol{x}_c\|_2) = \mathrm{RBF}(\|(\boldsymbol{O}\boldsymbol{X}_i[p]+\boldsymbol{t}) - (\boldsymbol{O}\boldsymbol{x}_c+\boldsymbol{t})\|_2) = \mathrm{RBF}(\|T_g\cdot\boldsymbol{X}_i[p]-T_g\cdot\boldsymbol{x}_c\|_2).$$

Finally we can derive the E(3)-invariance on $\boldsymbol{H}'[p]$ and the E(3)-equivariance on $\boldsymbol{X}_i'[p]$:

$$\begin{aligned}
\boldsymbol{H}'[p] &= \boldsymbol{H}[p] + \sigma_h(\boldsymbol{H}_i[p], \boldsymbol{h}_c, \boldsymbol{r}_p), \\
T_g \cdot \boldsymbol{X}_i'[p] &= T_g \cdot (\boldsymbol{X}_i[p] + \Delta \boldsymbol{x}_p \sigma_x(\boldsymbol{H}_i[p], \boldsymbol{h}_c, \boldsymbol{r}_p)) \\
&= \boldsymbol{O}(\boldsymbol{X}_i[p] + \Delta \boldsymbol{x}_p \sigma_x(\boldsymbol{H}_i[p], \boldsymbol{h}_c, \boldsymbol{r}_p)) + \boldsymbol{t} \\
&= \boldsymbol{O}\boldsymbol{X}_i[p] + \boldsymbol{t} + \boldsymbol{O}\Delta \boldsymbol{x}_p \sigma_x(\boldsymbol{H}_i[p], \boldsymbol{h}_c, \boldsymbol{r}_p) \\
&= T_g \cdot \boldsymbol{X}_i[p] + (T_g \cdot \boldsymbol{X}_i[p] - T_g \cdot \boldsymbol{x}_c)\sigma_x(\boldsymbol{H}_i[p], \boldsymbol{h}_c, \boldsymbol{r}_p) \\
&= T_g \cdot \boldsymbol{X}_i[p] + (T_g \cdot \boldsymbol{X}_i[p] - \text{centroid}(T_g \cdot \boldsymbol{X}_i))\sigma_x(\boldsymbol{H}_i[p], \boldsymbol{h}_c, \boldsymbol{r}_p),
\end{aligned}$$

which concludes the proof of Lemma 2 □

The sequence refinement module with the pretrained protein language models and adapters operates on the scaler residue representations unaffected by the geometric transformation T_g. To sum up, with Lemma 1-2 at hand, it is obvious to deduce the E(3)-equivariance of the PocketGen.

Supplementary Figures

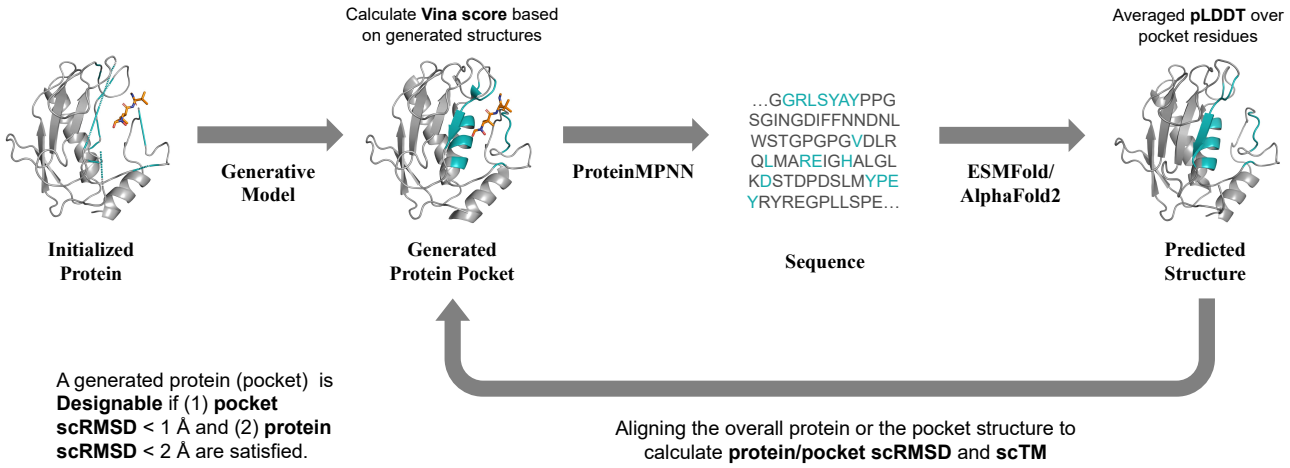


Figure S1. Schematic of computing metrics with respect to the generated pocket structures. black parts refer to the pocket, while the others are gray. The generative model refers to PocketGen or baseline methods leveraged to generate the protein pocket. The Vina score is calculated based on the generated protein-ligand complex. To calculate self-consistency scores, we first use ProteinMPNN to derive the residue sequence and then use ESMFold/AlphaFold2 to predict the structure. By aligning the predicted structure with the generated structure, we can obtain the protein/pocket scRMSD. Similarly, we can obtain the scTM score of the protein structure. We also report the averaged pLDDT of the pocket residues from ESMFold.

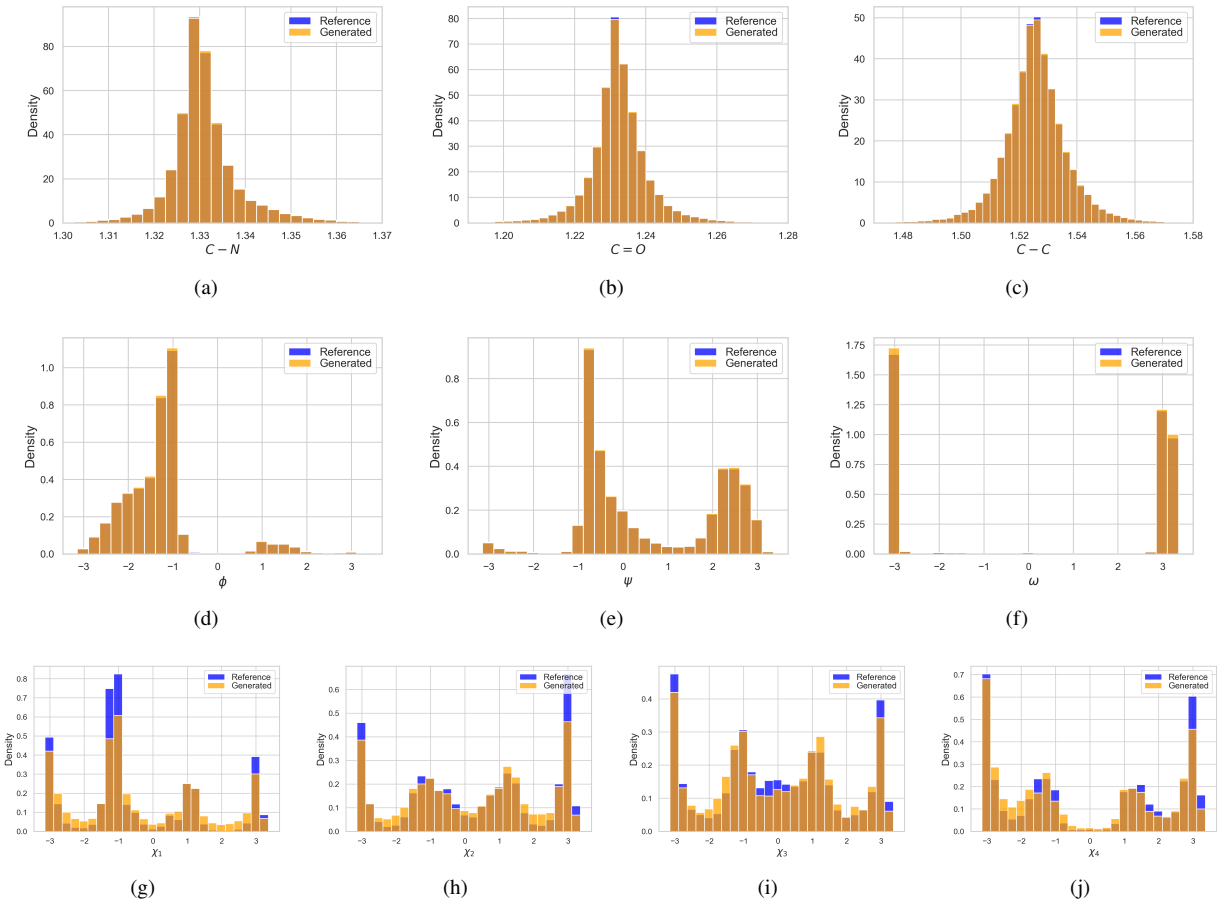
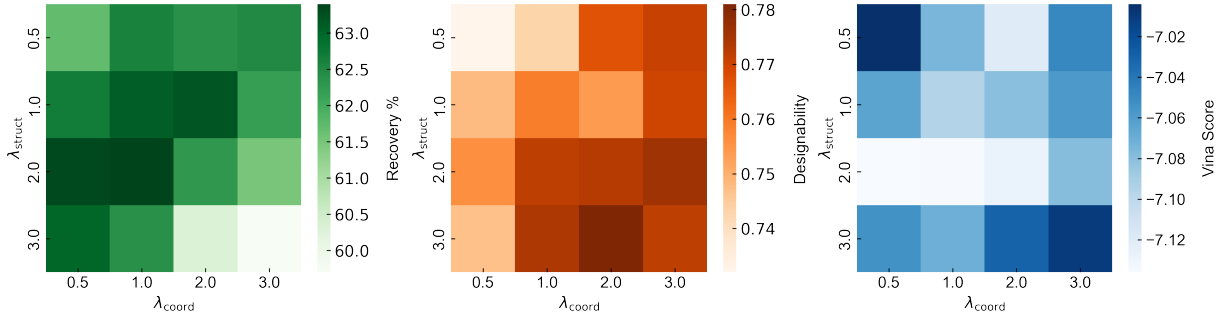


Figure S2. The bond length and dihedral angle distributions of the generated pockets and the reference (training set of CrossDocked dataset).



(a) AAR (b) Designability (c) Vina Score

Figure S3. Hyperparameter Analysis. Influence of hyperparameters (loss weights $\mathcal{L}_{\text{coord}}$ and $\mathcal{L}_{\text{struct}}$) on PocketGen's performance (AAR, Designability, and Vina Score) on the CrossDocked dataset. We perform a grid search over $\{0.5, 1.0, 2.0, 3.0\}$ and choose these hyperparameters based on the validation performance to select the specific parameter values. In the default setting, we set λ_{coord} to 1.0 and λ_{struct} to 2.0.

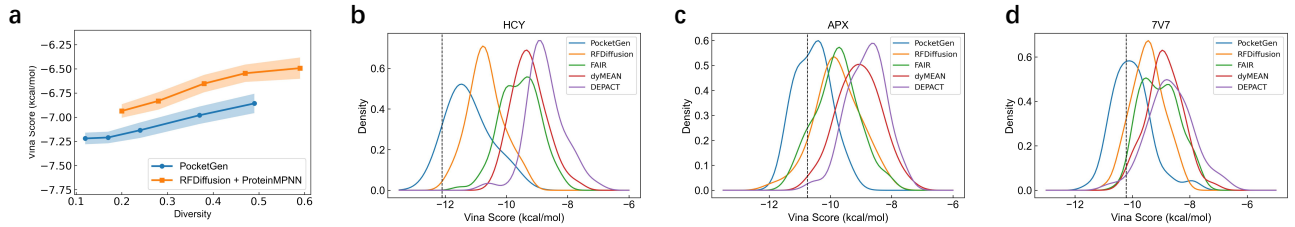


Figure S4. Additional results with RFDiffusion. a, The trade-off between quality (measured by Vina score) and diversity (1-average pairwise sequence similarity) of PocketGen. **b, c, d,** The pocket binding affinity distributions of PocketGen and baseline methods for three target molecules (HCY, APX, and 7V7, respectively).

Supplementary Tables

Table S1. Benchmarking PocketGen and other approaches for pocket generation on two datasets. Reported are average and standard deviation values across three independent runs (random seeds on the same dataset). The best results are bolded.

Model	CrossDocked			Binding MOAD		
	AAR (↑)	Designability (↑)	Vina (↓)	AAR (↑)	Designability (↑)	Vina (↓)
Test set	-	0.77	-7.016	-	0.79	-8.076
DEPACT	31.52±3.26%	0.68±0.04	-6.632±0.18	35.30±2.19%	0.67±0.06	-7.571±0.15
dyMEAN	38.71±2.16%	0.71±0.03	-6.855±0.06	41.22±1.40%	0.70±0.03	-7.675±0.09
FAIR	40.16±1.17%	0.73±0.02	-7.015±0.12	43.68±0.92%	0.72±0.05	-7.930±0.15
RFDiffusion	46.57±2.07%	0.74±0.01	-6.936±0.07	45.31±2.73%	0.75±0.05	-7.942±0.14
RFDiffusionAA	50.85±1.85%	0.75±0.03	-7.012±0.09	49.09±2.49%	0.78±0.03	-8.020±0.11
PocketGen	**63.40±1.64%**	**0.77±0.02**	**-7.135±0.08**	**64.43±2.35%**	**0.80±0.04**	**-8.112±0.14**

Table S2. The top 1/3/5/10 generated protein pocket (ranked by Vina score) with designability on the CrossDocked dataset. The success rate measures the percentage of protein that the model can generate pockets with higher affinity than the reference ones in the datasets. We report the average plDDT of the predicted pocket, the scRMSD of the pocket backbone coordinates, and the change of the scTM score of the whole protein. ESMFold means the scores are calculated with AlphaFold2 as the folding tool. The plDDT, scRMSD, and ΔscTM for PocketOpt are not reported, as PocketOpt keeps protein backbone structures fixed. We use to mark the results of affinity-related metrics, for pocket-structure related metrics, and for whole protein structure metrics. We report the means and standard deviations over three independent runs with random seeds.

	DEPACT	dyMEAN	FAIR	RFDiffusion	RFAA	PocketGen
Top-1 generated protein pocket						
pLDDT (ESMFold) (↑)	82.130±0.187	83.327±0.206	83.254±0.412	84.559±0.230	86.346±0.238	**87.065**±0.180
scRMSD (ESMFold) (↓)	0.705±0.023	0.703±0.024	0.680±0.019	0.676±0.017	0.654±0.011	**0.642**±0.008
ΔscTM (ESMFold) (↑)	-0.009±0.002	-0.004±0.003	-0.011±0.004	0.014±0.005	0.021±0.003	**0.030**±0.004
ΔscTM (ESMFold+co) (↑)	-0.015±0.005	-0.016±0.003	-0.023±0.005	-	-	**0.014**±0.003
Top-3 generated protein pockets						
pLDDT (ESMFold) (↑)	81.991±0.303	82.825±0.218	83.145±0.297	84.636±0.186	86.224±0.190	**86.450**±0.075
scRMSD (ESMFold) (↓)	0.706±0.025	0.724±0.024	0.685±0.017	0.679±0.014	**0.653**±0.012	0.655±0.007
ΔscTM (ESMFold) (↑)	-0.014±0.003	-0.010±0.004	-0.013±0.005	0.019 ±0.004	0.020±0.003	**0.024** ±0.001
ΔscTM (ESMFold+co) (↑)	-0.016±0.003	-0.022±0.002	-0.026±0.003	-	-	**0.012**±0.002
Top-5 generated protein pockets						
pLDDT (ESMFold) (↑)	82.070±0.276	82.910±0.231	83.168±0.208	84.320±0.219	85.735±0.087	**86.414**±0.110
scRMSD (ESMFold) (↓)	0.717±0.019	0.725±0.014	0.690±0.012	0.680±0.012	0.656±0.006	**0.654**±0.010
ΔscTM (ESMFold) (↑)	-0.018±0.004	-0.007 ±0.002	-0.024±0.002	0.016±0.001	0.019±0.001	**0.027**±0.001
ΔscTM (ESMFold+co) (↑)	-0.014±0.004	-0.023±0.003	-0.027±0.002	-	-	**0.015**±0.002
Top-10 generated protein pockets						
pLDDT (ESMFold) (↑)	81.580±0.232	82.771±0.203	83.048±0.165	84.234±0.195	85.377±0.142	**86.090**±0.124
scRMSD (ESMFold) (↓)	0.710±0.014	0.734±0.013	0.705±0.013	0.684±0.012	0.672±0.007	**0.657**±0.005
ΔscTM (ESMFold) (↑)	-0.025±0.002	-0.014±0.001	-0.026±0.002	0.014±0.003	0.019±0.000	**0.022**±0.001
ΔscTM (ESMFold+co) (↑)	-0.015±0.003	-0.022±0.002	-0.026±0.003	-	-	**0.013**±0.001

Table S3. The top 1/3/5/10 generated protein pocket (ranked by Vina score) with designability on the Binding MOAD dataset. Besides Vina score, we also calculate MM-GBSA to evaluate the binding affinity. The success rate measures the percentage of protein that the model can generate pockets with higher affinity than the reference ones in the datasets. We report the average plDDT of the predicted pocket, the scRMSD of the pocket backbone coordinates, and the change of the scTM score of the whole protein. co indicates codesign, where codesign methods directly use the designed sequence for consistency calculation. The plDDT, scRMSD, and ΔscTM for PocketOpt are not reported, as PocketOpt keeps protein backbone structures fixed. We use to mark the results of affinity-related metrics, for pocket-structure related metrics, and for whole protein structure metrics. We report the means and standard deviations over three independent runs with random seeds.

Methods:	PocketOpt	DEPACT	dyMEAN	FAIR	RFDiffusion	RFAA	PocketGen
Top-1 generated protein pocket							
Vina score (↓)	-9.828±0.120	-9.216±0.071	-9.352±0.082	-9.530±0.064	-9.901±0.055	-10.120±0.063	**-10.322**±0.057
MM-GBSA (↓)	-62.293±0.819	-55.180±0.906	-55.504±0.628	-59.851±0.723	-63.440±0.836	-64.305±1.04	**-67.862**±0.618
Success Rate (↑)	0.891±0.024	0.784±0.045	0.779±0.02	0.841±0.033	0.905±0.030	0.923±0.027	**0.952**±0.016
pLDDT (ESMFold) (↑)	-	83.145±0.426	83.280±0.247	83.536±0.217	85.068±0.335	**87.232**±0.314	87.106±0.224
scRMSD (ESMFold) (↓)	-	0.642±0.024	0.622±0.017	0.620±0.025	0.581±0.018	**0.572**±0.016	0.575±0.013
ΔscTM (ESMFold) (↑)	-	-0.004±0.001	-0.001±0.002	0.008±0.003	0.025±0.005	**0.031**±0.005	0.030±0.004
ΔscTM (ESMFold+co) (↑)	-	-0.025±0.003	-0.019±0.004	-0.008±0.005	-	-	0.004±0.003
Top-3 generated protein pocket							
Vina score (↓)	-9.403±0.102	-8.876±0.064	-8.971±0.076	-9.234±0.050	-9.589±0.048	-9.634±0.045	**-10.135**±0.039
MM-GBSA (↓)	-58.204±0.739	-51.255±0.660	-52.371±0.540	-54.946±0.632	-57.235±0.803	-60.801±0.829	**-64.108**±0.421
pLDDT (↑)	-	82.769±0.370	82.131±0.263	82.826±0.207	84.912±0.328	85.920±0.301	**86.535**±0.220
scRMSD (ESMFold)(↓)	-	0.645±0.023	0.620±0.015	0.596±0.014	0.580±0.016	0.574±0.015	**0.573**±0.011
ΔscTM (ESMFold) (↑)	-	-0.009±0.002	-0.004±0.003	0.004±0.002	0.020±0.005	0.025±0.004	**0.029**±0.003
ΔscTM (ESMFold+co) (↑)	-	-0.027±0.002	-0.020±0.003	-0.007±0.002	-	-	**0.005**±0.002
Top-5 generated protein pocket							
Vina score (↓)	-9.260±0.091	-8.759±0.050	-8.842±0.050	-9.195±0.043	-9.478±0.045	-9.569±0.035	**-9.950**±0.030
MM-GBSA (↓)	-55.728±0.536	-49.204±0.745	-50.289±0.431	-52.152±0.577	-55.490±0.830	-56.306±0.687	**-62.970**±0.412
pLDDT (ESMFold) (↑)	-	81.939±0.261	81.915±0.163	82.346±0.219	85.150±0.322	85.830±0.205	**86.438**±0.160
scRMSD (ESMFold) (↓)	-	0.639±0.015	0.628±0.014	0.612±0.015	0.587±0.010	0.583±0.011	**0.581**±0.008
ΔscTM (ESMFold) (↑)	-	-0.007±0.001	-0.002±0.002	0.003±0.002	0.024±0.003	0.024±0.003	**0.026**±0.001
ΔscTM (ESMFold+co) (↑)	-	-0.027±0.004	-0.023±0.001	-0.005±0.002	-	-	**0.005**±0.002
Top-10 generated protein pocket							
Vina score (↓)	-8.981±0.087	-8.632±0.056	-8.690±0.051	-8.827±0.063	-9.268±0.042	-9.320±0.054	**-9.630**±0.037
MM-GBSA (↓)	-51.337±0.505	-45.480±0.519	-47.804±0.420	-48.203±0.625	-53.660±0.522	-55.763±0.643	**-60.106**±0.284
pLDDT (ESMFold) (↑)	-	81.632±0.270	81.774±0.264	82.739±0.340	84.582±0.191	85.337±0.229	**86.456**±0.079
scRMSD (ESMFold) (↓)	-	0.644±0.012	0.626±0.014	0.592±0.013	0.589±0.014	0.586±0.007	**0.584**±0.005
ΔscTM (ESMFold) (↑)	-	-0.003±0.002	-0.006±0.001	0.006±0.003	0.012±0.002	0.018±0.002	**0.027**±0.001
ΔscTM (ESMFold+co) (↑)	-	-0.029±0.002	-0.021±0.001	-0.008±0.002	-	-	**0.006**±0.003

Table S4. Substructure analysis of the generated molecules. We consider three covalent bonds in the backbone (C-N, C=O, and C-C), three conventional dihedral angles in the backbone (ϕ, ψ, ω[58]), and four dihedral angles in the side chains ($\chi_1, \chi_2, \chi_3, \chi_4$[59]). The KL divergence of the bond lengths and dihedral angles between the training set (CrossDocked) and the generated pockets are calculated following previous works[60,61]. Since PocketOpt sets the pocket backbone fixed, the corresponding backbone metrics are not calculated.

Methods	C-N	C=O	C-C	ϕ	ψ	ω	χ_1	χ_2	χ_3	χ_4
PocketOpt	-	-	-	-	-	-	**0.140**	0.098	0.071	0.104
DEPACT	0.010	0.011	0.009	0.033	0.032	0.097	0.190	0.126	0.109	0.148
dyMEAN	0.012	0.012	0.022	0.036	0.017	0.078	0.224	0.187	0.130	0.125
FAIR	0.014	0.011	0.020	0.025	0.024	0.080	0.235	0.067	0.110	0.167
RFDiffusion	0.009	0.006	0.006	0.027	0.016	0.066	0.160	0.092	0.073	**0.060**
RFDiffusionAA	0.008	0.007	**0.005**	**0.025**	0.014	**0.062**	0.159	0.087	0.071	0.064
PocketGen	**0.006**	**0.004**	**0.005**	0.029	**0.013**	0.077	0.158	**0.053**	**0.065**	0.097

Table S5. Ablations studies of PocketGen. **PocketGen w/o ligand update** indicates the ligand structure is fixed during the generation of pockets by PocketGen. **PocketGen w/o residue-level attention** denotes only the basic atom-level attention performed in the bilevel attention module. **PocketGen w/o pLM** means no pretrained protein language model is leveraged for sequence refinement. Moreover, **PocketGen w/ EGNN/GVP/GMN** uses EGNN[67], GVP[68], and GMN[69] respectively to replace the bilevel graph transformer in PocketGen as the structural encoder for comparison. We report the means and standard deviations over three different runs (%). The best results are bolded.

Model	CrossDocked			Binding MOAD		
	AAR (↑)	Designability (↑)	Vina (↓)	AAR (↑)	Designability (↑)	Vina (↓)
PocketGen w/o ligand update	59.20±1.56%	0.75±0.03	-6.838±0.10	59.61±2.13%	0.77±0.05	-7.865±0.12
PocketGen w/o residue-level attention	58.19±1.78%	0.66±0.04	-6.924±0.08	57.30±2.28%	0.67±0.04	-7.805±0.10
PocketGen w/o pLM	42.70±1.45%	0.71±0.05	-6.894±0.07	44.23±1.31%	0.69±0.06	-7.787±0.12
PocketGen w/ EGNN encoder	58.85±1.21%	0.71±0.02	-6.916±0.03	58.64±1.14%	0.72±0.06	-7.804±0.11
PocketGen w/ GVP encoder	61.10±0.90%	0.72±0.04	-6.930±0.08	59.97±1.40%	0.75±0.05	-7.883±0.10
PocketGen w/ GMN encoder	60.49±1.33%	0.68±0.03	-6.947±0.05	59.82±1.67%	0.74±0.04	-7.869±0.08
PocketGen	**63.40±1.64%**	**0.77±0.02**	**-7.135±0.08**	**64.43±2.35%**	**0.80±0.04**	**-8.112±0.14**

Table S6. Hyperparameters for the baselines and our PocketGen.

hyperparameter	value	description
PocketOptimizer		
n_confs	50	The number of sampled ligand conformers.
n_poses	10000	The max number of ligand poses to sample.
ligand_scaling	1.0	The ligand scaling factor.
vdw_filter_thresh	100	The energy threshold for filtering rotamers (kcal/mol).
DEPACT		
interaction_threshold	-1.0	The threshold of ligand-subpocket interaction score for filtering.
mathcing_threshold	1.5 Å	The RMSD cutoff for finding geometrically consistent ways of placing the seeding residues.
num_CCSS	1000	The number of common chemical substructure (CCSS).
weight_seed	100	The weight of seeding residues in the scoring function.
FAIR		
hidden_size	128	Size of the hidden states in its hierarchical message passing network (MPN).
num_heads	4	Number of attention heads.
n_atom_layers	6	Number of atom-level layers in the MPN.
n_residue_layers	2	Number of residue-level layers in the MPN.
k_atom_neighbors	24	Number of neighbors for each node in the atom KNN graph.
k_residue_neighbors	8	Number of neighbors for each node in the residue KNN graph.
n_backbone_iter	5	Number of iterations in backbone refinement.
n_fullatom_iter	10	Number of iterations in full atom refinement.
RFDiffusion		
potential_scale	1.0	Scale of the auxiliary substrate contact potential.
temp_ProteinMPNN	0.1	The temperature in ProteinMPNN for sequence inference.
n_steps	50	Number of the diffusion steps.
RFDiffusionAA		
temp_ProteinMPNN	0.1	The temperature in LigandMPNN for sequence inference.
n_steps	50	Number of the diffusion steps.
dyMEAN		
embed_size	64	Size of the residue type embedding and the position number embedding.
hidden_size	128	Size of the hidden states in the MPN
n_layers	3	Number of layers in the MPN
n_iter	3	Number of iterations in the progressive full-shot decoding.
k_neighbors	9	Number of neighbors for each node in the KNN graph.
d	16	Size of the attribute vector of each channel (equal to the size of the atom type
PocketGen		
hidden_size	128	Size of the hidden states in the MPN
n_layers	4	Number of layers in the MPN
num_heads	4	Number of attention heads.
n_iter	3	Number of refinement rounds.
k_sparse	3	Number of elements to keep in the sparse attention.
k_neighbors	8	Number of neighbors for each node in the KNN graph.