SEGUID v2: Extending SEGUID checksums for circular, linear, single- and double-stranded biological sequences

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Abstract

Background: Synthetic biology involves combining different DNA fragments, each containing functional biological parts, to address specific problems. Fundamental gene-function research often requires cloning and propagating DNA fragments, such as those from the iGEM Parts Registry or Addgene, typically distributed as circular plasmids. Addgene's repository alone offers over 100,000 plasmids.

To ensure data integrity, cryptographic checksums can be calculated for the sequences. Each sequence has a unique checksum, making

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checksums useful for validation and quick lookups of associated annotations. For example, the SEGUID checksum, uniquely identifies protein sequences with a 27-character string.

Objectives: The original SEGUID, while effective for protein sequences and single-stranded DNA (ssDNA), is not suitable for circular and double-stranded DNA (dsDNA) due to topological differences. Challenges include how to uniquely represent linear dsDNA, circular ssDNA, and circular dsDNA. To meet these needs, we propose SEGUID v2, which extends the original SEGUID to handle additional types of sequences.

Conclusions: SEGUID v2 produces strand and rotation invariant checksums for single-stranded, double-stranded, possibly staggered, linear, and circular DNA and RNA sequences. Customizable alphabets allows for other types of sequences. In contrast to the original SEGUID, which uses Base64, SEGUID v2 uses Base64url to encode the SHA-1 hash. This ensures SEGUID v2 checksums can be used as-is in filenames, regardless of platform, and in URLs, with minimal friction.

Availability: SEGUID v2 is readily available for major programming languages distributed under the MIT license. JavaScript package **seguid** is available on NPM, Python package **seguid** on PyPi, and R package **seguid** on CRAN.

Keywords: checksum, hash, DNA, RNA, protein, SHA-1, Base64url, SEGUID

1 Introduction

The post-genomic era has provided access to more biological sequence data than ever before. These sequence data provide an important resource for both fundamental research and synthetic biology, which aim to understand and harness biology to solve urgent societal problems, such as global warming and food security. Genetic parts contained on DNA fragments are distributed to and from culture collections and repositories, as well as within the research community to study genes and build new synthetic systems. These parts are almost always distributed as plasmids, where the DNA of interest is inserted into a circular double-stranded (ds) DNA plasmid providing replication and selection in a microorganism optimal for producing high-quality DNA, usually *E. coli*.

Each plasmid should have an associated sequence file that is distributed along with the physical clone. Although repositories such as Addgene (Kamens, 2014b,a) and the iGEM Parts collection provide standards, the trans-

mission of biological material and associated information is still very much an ad-hoc procedure.

Data integrity is of great importance since it directly affects the reproducibility of science that relied on the original genetic parts. Checksums are widely used for verifying that information is preserved through space and time, and has also been applied to safe-guard biological sequences. A checksum is a short string of characters that is calculated from, and typically stored with, an arbitrarily sized piece of data. By transmitting the checksum together with the data, a receiver can verify correctness of the data.

```
>AAF20475.1 crc64=44CAAD88706CC153 lsseguid=X5XEaayob1nZLOc7eVT9qyczarY
QSALTQPASVSGSPGQSITISCTGTSSDVGSYNLVSWYQQHPGKAPKLMIYEGSKRPSGVSNRFSGSKSGNTASLTISGLQAEDEADYYCCSYAGSSTWVFGGGTKLTVL
                                                                                                  ⇑       ⇑
>AAF20481.1 crc64=44CAAD88706CC153 lsseguid=BpBeDdcNUYNsdk46JoJdw7Pd3BI
QSALTQPASVSGSPGQSITISCTGTSSDVGSYNLVSWYQQHPGKAPKLMIYEGSKRPSGVSNRFSGSKSGNTASLTISGLQAEDEADYYCSSYAGSSTLVFGGGTKLTVL
                                                                                                  ⇑       ⇑
```

Figure 1: **Two protein sequences with colliding CRC-64 checksums.** The two protein sequences with GenBank accession number *AAF20481.1*$_{\text{GB}}$ and *AAF20475.1*$_{\text{GB}}$ have both 110 amino acids and differ at two positions (91 and 99). Despite this, they share the same UniProt **CRC-64** checksum. The **SHA-1**-based SEGUID v2 checksums differ.

Several existing checksum algorithms are relevant in bioinformatics. One of the first used was the cyclic redundancy check (**CRC**) (W.H. et al., 1993), which is a computationally inexpensive, error-detecting algorithm originally designed for network and storage devices. The UniProt database uses a 64-bit **CRC** (**CRC-64**) checksum[1]. A disadvantage of the **CRC-64** is that collisions have been observed for biological data. For example, the two protein sequences with GenBank accession numbers *AAF20475.1*$_{\text{GB}}$[2] and *AAF20481.1*$_{\text{GB}}$[3], have the same **CRC-64** hexadecimal checksum, despite differing in two of the 110 amino acids (Figure 1).

While collisions are unavoidable for all functions producing finite-length checksums, longer checksums are less likely to generate collisions. However, long checksums are less readable and also more expensive to generate, store, and compare. The SEquence Globally Unique IDentifier (SEGUID) checksum (Babnigg and Giometti, 2006) was introduced in order to provide a stable, unifying key for the same sequence in different databases facilitating linking protein sequences across databases. SEGUID is based on the **SHA-1**

[1]https://www.uniprot.org/help/sequences
[2]UniProt accession number *A0A5C2FXM9*$_{\text{UP}}$
[3]UniParc accession number *UPI0000117F39*$_{\text{UP}}$

algorithm (Dang, 2015), which produces a 160-bit hash, and is therefore less sensitive to collisions compared with the 64-bit **CRC-64** hash. With **SHA-1**, sequence collisions are very unlikely. For example, the above two proteins with clashing **CRC-64** checksums have distinct SEGUID checksums.

More importantly, no current checksum algorithm is directly useful for DNA, since the topology of DNA is most often different from that of proteins. Double-stranded DNA (dsDNA) consists of two complementary, anti-parallel strands. Usually, only one is stored for DNA, as the second strand is the reverse complement of the stored sequence. However, both strands are equally valid representations of the DNA molecule, but give different checksum values, except for the special case where the strands are identical as in palindromic sequences.

Another topology is a circular sequence, e.g. circular single-stranded DNA (ssDNA) and circular dsDNA. For these cases, there are multiple representations for the same circular sequence, and there is no well-defined canonical representation for which the checksum should be calculated.

In this article, we propose SEGUID v2, which is an extension to the original SEGUID, that produces checksums for linear, circular, single- and double-stranded sequences of different types, including DNA, RNA, and amino-acid sequences in order to integrate synthetic biology resources and identify redundancies in existing databases.

2 Methods

The problem of defining a checksum for sequences of different kinds, comprise three challenges: (i) identify a unique representation among one or more equivalent representations, and convert it to a linear sequence representation, (ii) choose a hash function to distill the unique representation to a large-enough bit sequence, and (iii) compactly encode the bit sequence to a checksum using a text-friendly encoding alphabet.

Below, we explain how we define each of these steps for the proposed SEGUID v2 checksum, and what objectives we target when there are multiple options available.

2.1 Unique representation of sequence data

2.1.1 Linear single-stranded sequences

A *linear single-stranded sequence* is a single, directional sequence with one or more symbols, where *directional* means it has a unique start and end ("left to right"). The sequence may be amino acids, DNA, RNA, or any

sequence of symbols from a well-defined, case-sensitive alphabet. Examples of DNA sequences are `C`, `AT`, and `GATTACA`. Examples of RNA sequences are `C`, `AU`, and `AGA`. Examples of amino-acid sequences ("proteins") are `A`, `QSA`, and `QSALTQPASV`.

A linear single-stranded sequence is always unique, which makes it natural to use the sequence as-is to calculate the hash. This case was already covered by the original SEGUID algorithm.

2.1.2 Linear double-stranded sequences

A *linear double-stranded sequences* comprise of two sequences that share a region where the two sequences are complementary to each other. For double-stranded DNA, the "upper" strand is often referred to as the "Watson" strand and the "lower" as the "Crick" strand. The Watson and Crick strands are reverse complementary to each other. We use the Watson and Crick strands separated by symbol ↩, where both Watson and Crick are 5′→3′ sequences, to represent a double-stranded sequence. This notation reflects the duality of double-stranded sequences and matches the graphical representation in the figures.

Two types of such sequences must be considered - (i) blunt-ended, and (ii) staggered, linear double-stranded sequences - which are discussed next.

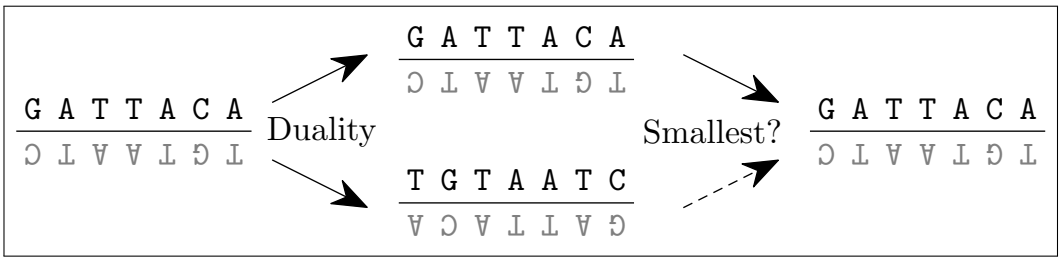


Figure 2: **The SEGUID v2 steps for identifying the canonical representation of a blunt-ended linear, double-stranded DNA sequence.** The input sequence `GATTACA`↩`TGTAATC` (left and top middle) has a dual, reverse-complementary representation `TGTAATC`↩`GATTACA` (bottom middle). The lexicographically smallest of these two is `GATTACA`↩`TGTAATC` (right). It is from this linear representation that the SEGUID v2 checksum is calculated. Legend: Following IUPAC-IUB Commission on Biochemical Nomenclature (1970), we choose to rotate the symbols on the complementary strand in the figure such that the left-to-right, 3′→5′ representation holds when rotating the figure upside-down.

Blunt-ended, linear double-stranded sequences: In its basic form, the two sequences are of the same length and fully overlapping, i.e. the shared region spans their full length. We refer to this type of sequences as *blunt-ended* (short blunt), or non-staggered, double-stranded sequences. For example, `GATTACA`↩`TGTAATC` is a blunt, double-stranded sequence with reverse-complementary sequences `GATTACA` and `TGTAATC`. Importantly, `GATTACA`↩`TGTAATC` and `TGTAATC`↩`GATTACA` are dual representations of the same linear double-stranded DNA sequence (Figure 2). In other words, *without further constraints*, there is no unique representation of it. To address this *uniqueness problem*, we define the unique representation of the two possibilities as the one that is lexicographically smaller[4] than the other. This is possible for all well-defined alphabets. For example, consider the DNA alphabet `A`, `C`, `G`, `T`, ordered such that `A` < `C` < `G` < `T`. This allows us to compare any two sequences lexicographically, e.g. `AT` < `TA`. For example, in the above example, we have that `GATTACA`↩`TGTAATC` < `TGTAATC`↩`GATTACA`, and any checksums will be calculated based on the first of these two representations (Figure 2).

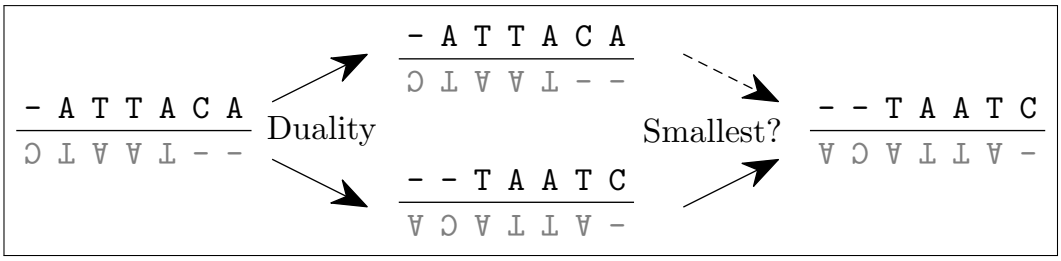


Figure 3: **The SEGUID v2 steps for identifying the canonical representation of a staggered, linear, double-stranded DNA sequence.** The input sequence `-ATTACA`↩`--TAATC` (left and top middle) has a dual, reverse-complementary representation `--TAATC`↩`-ATTACA` (bottom middle). Because the minus symbol (`-`) is lexicographically smaller than the other symbols, the smallest of these two linear representations is `--TTACA`↩`-GTAATC` (right). It is from this linear representation that the SEGUID v2 checksum is calculated.

Staggered, linear double-stranded sequences: Another form of double-stranded sequences is where the two sequences do not fully overlap. We refer to this type of sequences as *staggered* double-stranded sequences, e.g. `-ATTACA`↩`TGTAATC`, `--TTACA`↩`TGTAATC`, and `-ATTACA`↩`-GTAATC`. Here we use the minus symbol (`-`) to indicate that there is a symbol on the opposite strand but not on the current strand. As for blunt double-stranded

[4]Strictly speaking, "smaller or equal" is sufficent to get a unique representation.

sequences, staggered ones have two possible representation for the same sequence, e.g. `-GTAATC`↩`--TTACA` and `--TTACA`↩`-GTAATC` refer to the same sequence. As before, we define the unique representation to be the lexicographically smallest of the two, where we define the minus symbol (`-`) to be smaller than any other symbol, e.g. with the DNA alphabet we have that `-` < `A` < `C` < `G` < `T`. With this, we can identify `--TTACA`↩`-GTAATC` as the minimal lexicographic representation, because `--TTACA`↩`-GTAATC` < `-GTAATC`↩`--TTACA` (Figure 3). It is from this canonical representation that the hash is calculated.

2.1.3 Circular single-stranded sequences

A *circular single-stranded sequence* is a single, directional sequence with one or more symbols without a beginning and end. The sequence may comprise of symbols from any well-defined alphabet. Any linear single-stranded sequence can be made a circular single-stranded sequences, by combining the two ends, and vice versa. Examples of circular DNA sequences are `C`↻, `AT`↻, `TA`↻, and `GATTACA`↻. Here we use the symbol ↻ to indicate that the two ends are combined. A naturally occurring circular, single-stranded DNA sequences is the 6,407-nucleotide M13 bacteriophage with sequence `AACGCTACTA...GGATGTT`↻ (*NC_003287.2*$_{\text{GB}}$ and *V00604.2*$_{\text{GB}}$). Circular RNA (circRNA) are naturally occurring circular RNA formed from mRNA through a process called back-splicing Jeck et al. (2013), where a downstream splice donor site is joined to an upstream splice acceptor site. The *MW729338*$_{\text{GB}}$ represents an 840-nucleotide circRNA from *Culex pipiens* (northern house mosquito). Examples of circular proteins are `A`↻, `QSA`↻, and `QSALTQPASV`↻. A naturally occurring circular single-stranded amino-acid sequence is the 21 amino-acid long *E. coli* peptide Microcin J25 (`PRO_0000002775` at positions 38–58 in *Q9X2V7*$_{\text{UP}}$) with sequence `GGAGHVPEYFVGIGTPISFYG`↻. This circular peptide is also referred to as `VGIGTPISFYGGGAGHVPEYF`↻, e.g. Blond et al. (1999) and Trabi and Craik (2002).

Because there is no beginning and end of a circular sequence, there is also no unique representation of a circular sequence *without further constraints*. For example, both `AU`↻ and `UA`↻ represents the same circular sequence. Similarly, `GATTACA`↻, `ATTACAG`↻, `TTACAGA`↻, `TACAGAT`↻, `ACAGATT`↻, `CAGATTA`↻, and `AGATTAC`↻ are all the same circular sequence. In general, there are n different representations of the same circular single-stranded sequence of length n.

The non-uniqueness of these representations complicates how to store a circular sequence in databases, how to communicate it electronically, and how to present it in text. Similarly, if we naively would calculate the original

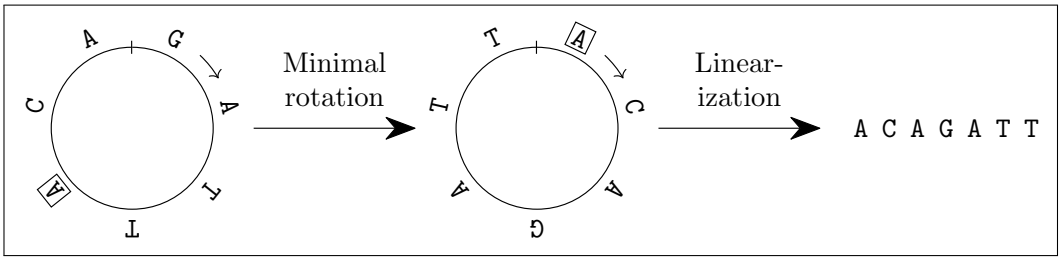


Figure 4: **The SEGUID v2 steps for identifying the canonical representation of a circular, single-stranded DNA sequence.** The input sequence GATTACA↻ (left) is one of several equivalent, non-normalized representations of the same circular sequence. Among all seven possible rotations, there is a minimal rotation, which for this sequence is ACAGATT↻ (middle). The linear representation of the minimal rotation is ACAGATT (right). It is from this linear representation that the SEGUID v2 checksum is calculated.

SEGUID checksum for the above representations, we would get different checksums despite them representing the same circular sequence.

To address the above *uniqueness problem*, we need to identify a single unique representation among all possible representations part of the same *equivalent class*. Analogously to how we addressed the uniqueness problem for double-stranded sequences, a natural candidate is to define the unique representation to be *the sequence with a rotation that has the smallest lexicographical value*. For example, consider the above circular sequence GATTACA↻ with seven equivalent representations. We can order the seven possible rotations as ACAGATT < AGATTAC < ATTACAG < CAGATTA < GATTACA < TACAGAT < TTACAGA. The *minimal lexicographical rotation* of GATTACA↻ is therefore ACAGATT (Figure 4). The minimal rotation can be found in linear time (Pierre Duval, 1983).

2.1.4 Circular double-stranded sequences

A *circular double-stranded sequences* is similar to a *circular single-stranded sequences*, but with a second complementary strand. Examples of circular double-stranded DNA sequences are C↩G↻, AT↩AT↻, and TATGCCAA↩TTGGCATA↻. A real-world example is the 2,686-basepair circular double-stranded DNA pUC19 cloning vector TCGC...CGTC↩GACG...GCGA↻ (*M77789.2*$_{\text{GB}}$ and *L09130.1*$_{\text{GB}}$).

Similarly to the single-stranded cases, there are multiple representations for circular double-stranded sequences. For example, the eight rotational variants TATGCCAA↩TTGGCATA↻, ATGCCAAT↩ATTGGCAT↻, TGCCAATA↩TATTGGCA↻,

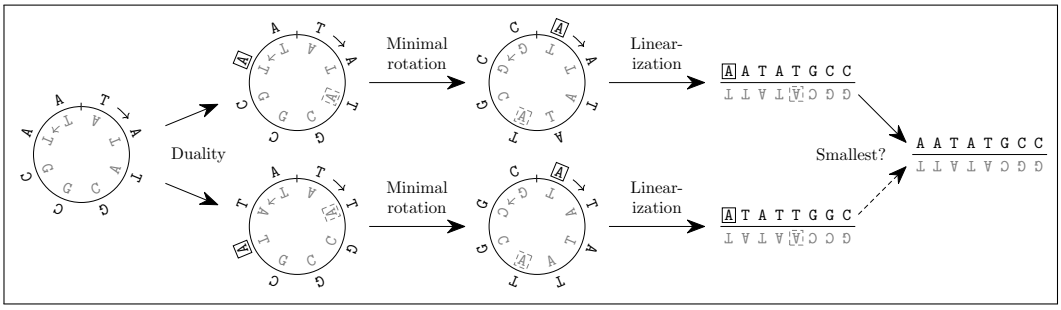


Figure 5: **The SEGUID v2 steps for identifying the canonical representation of a circular, double-stranded DNA sequence.** The input sequence TATGCCAA↩TTGGCATA↻ (left, and top left) is one of several equivalent, non-normalized representations of the same circular sequence. This double-stranded sequence has the dual reverse-complementary representation TTGGCATA↩TATGCCAA↻ (bottom left). The minimal rotations for these dual representations are AATATGCC↩GGCATATT↻ (top middle) and ATATTGGC↩GCCAATAT↻. The smallest of the two linear minimal-rotated representations (top right and top bottom), is AATATGCC↩GGCATATT (right). It is from this linear representation that the SEGUID v2 checksum is calculated.

..., ATATGCCA↩TGGCATAT↻ represent the same eight-basepair circular dsDNA, but so does the eight reverse-complementary counterparts TTGGCATA↩TATGCCAA↻, ATTGGCAT↩ATGCCAAT↻, TATTGGCA↩TGCCAATA↻, ..., TGGCATAT↩ATATGCCA↻. In general, there are $2 \cdot n$ different representations of the same circular double-stranded sequence of length n. We define the unique representation as *the smallest of the minimal lexicographical rotations of the two strands*. For instance, AATATGCC↩GGCATATT↻ is the minimal for one of original representation, and ATATTGGC↩GCCAATAT↻ for the reverse-complementary duality. Between these two, AATATGCC↩GGCATATT↻ is the lexicographically smallest, because AATATGCC↩GGCATATT↻ < ATATTGGC↩GCCAATAT↻. Thus, the canonical representation of the circular double-stranded sequence TATGCCAA↩TTGGCATA↻ is AATATGCC↩GGCATATT↻ (Figure 5). Note, it is sufficient to compare only AATATGCC < ATATTGGC to identify AATATGCC↩GGCATATT↻ to be the minimal lexicographically representation. This is true also for non-bijective complementary alphabets.

2.2 Hash function (SHA-1)

SEGUID v2 supports linear and circular, single- and double-stranded sequences. As shown above, circular sequences are transformed to unique,

linear representations based on minimal rotations and lexicographic ordering. Thus, regardless of input sequence, the hash is always calculated on a linear representation. To apply the hash function to these representations, we need to coerce them to single strings[5]. There are two types of linear representations we have to consider; the single-stranded one and the double-stranded one. A linear *single*-stranded sequence can be coerced to a string as-is, e.g. the sequence `GATTACA` becomes the string `"GATTACA"`. For a linear *double*-stranded sequence, we chose to coerce it to a string by concatenating the $5' \rightarrow 3'$ Watson sequence and the $5' \rightarrow 3'$ Crick sequences with a semicolon (`;`). For example, the blunt, for linear dsDNA sequence `GATTACA`$\hookleftarrow$`TGTAATC`, the hashed string is `"GATTACA;TGTAATC"`. Similarly, for the staggered, linear dsDNA sequence `--TTACA`$\hookleftarrow$`-GTAATC` the hashed string is `"--TTACA;-GTAATC"`.

The original SEGUID method uses **SHA-1** (Secure Hash Algorithm 1) to generate a 160-bit hash from the input sequence. As argued in Babnigg and Giometti (2006), **SHA-1** is less sensitive to collisions compared to **CRC-64** and sequence checksum collisions are very unlikely when using **SHA-1**. Another, popular hash function is the Message Digest Algorithm 5 (**MD5**) (Rivest, 1992), which produces a 128-bit hash. Although we are not aware of any collisions for biological data, it has been shown(Wang and Yu, 2005) that **MD5** collisions can be created artificially. Although we have not yet observed a **SHA-1** collision, one could argue that we should "upgrade" to **SHA-256** to further lower the risk for collisions. On the other hand, that comes with a cost of producing longer checksums. For example, the popular and widely available **SHA-256** algorithm, part of **SHA-2**, produces a 256-bit long hash, which is 60% longer than a **SHA-1** hash, which in turn would lengthen the final checksum the same amount. There are downsides to having too long checksums. For instance, they may be too long to be used for metadata headers in some sequence-based file formats, they risk being line-wrapped in communications such as email threads, and they add to the storage size for very-large lookup tables. Moreover, some file systems have a limit on how long a canonical pathname may be, e.g. on MS Windows the default pathname-length limit is 256 characters. A very long checksums in a directory or file name, in an already deeply nested directory structure, may result in this limit being exceeded.

Stevens et al. (2017) and Leurent and Peyrin (2020) demonstrated that it is feasible to artificially construct **SHA-1** clashes on modern computers. Because of this, **SHA-1** is no longer considered safe from a cryptography point

[5]Technically, hash functions operates on raw bytes. To calculate a hash for a string, we ensure to use ASCII strings such that each characters uniquely maps to an integer byte.

of view. For instance, the U.S. National Institute of Standards and Technology (NIST) "*recommends that federal agencies transition away from SHA-1 for all applications as soon as possible. Federal agencies should use SHA-2 or SHA-3 as an alternative to SHA-1*" (Computer Security Resource Center (CSRC), 2022). The **SHA-2** and **SHA-3** family of hash functions produce longer hashes, making it harder to construct collisions. That said, sequence checksums are used for efficient storage, communication, and lookup of sequences - not to secure them cryptographically. Arguing that the checksum of a full sequence can guarantee that the full sequence has not be compromised is problematic and risky. To achieve that, cryptographical protocols based on, for instance, public-and-private keys should be used, which is a problem beyond sequence checksums. We find no reason to incorporate such features in SEGUID v2, because there already exist well-established, readily available protocols for encrypted communication and storage.

Based on the above, we choose to use **SHA-1** also for SEGUID v2.

2.3 Encoding (Base64url)

A hash, by itself, is sufficient to uniquely identify a sequence. However, hashes, such as **SHA-1**, are very large integers, larger than can easily be represented in common programming languages, which makes them complicated to coerce as-is to and from textual representations. Encoding is a technique to turn an internal binary representation to a string, preferably in a compact way. Different encoding algorithms have different "compression" ratios (Table 1).

Description	Value	Length
SHA-1 (binary)	$10011010110110\ldots0010100100_2$	160 bits
SHA-1+Base16	`9adaef6edbb0afafcca01c6fb66e01129bfe8ca4`	40 symbols
SHA-1+Base64	`mtrvbtuwr6/MoBxvtm4BEpv+jKQ=`	28 symbols
SHA-1+Base64url	`mtrvbtuwr6_MoBxvtm4BEpv-jKQ=`	28 symbols
SEGUID v1	`mtrvbtuwr6/MoBxvtm4BEpv+jKQ`	27 symbols
SEGUID v2	`mtrvbtuwr6_MoBxvtm4BEpv-jKQ`	27 symbols

Table 1: **Examples of different encodings of a SHA-1 hash**. The **SHA-1** hash was generated based on the sequence string `ACAGATT`. The **Base64** specification requires that the output comprise a multiple of four symbols, and if not, the output should be padded with equal signs (`=`) at the end. Because of this, and a **Base64** encoding of a **SHA-1** hash will always have a single, non-informative padding symbol at the end. This padding symbol is excluded in the SEGUID checksums.

For instance, the **SHA-1** hash, which is a 160-bit integer, or equivalently

a 20-byte integer, could be encoded to a string of length 160 comprising symbols zero (`0`) and one (`1`). Assuming an 8-bit ASCII string, this encoding is 800% larger than the internal 20 bytes. That would be an inefficient way to represent the hash. A more compact, and commonly used encoding is the hexadecimal representation, which comprise an 16-character alphabet (`0-9` and `a-f`). In this representation, a **SHA-1** hash is of length 40 characters; one for every four ($\log_2 16$) bits, or two for each byte. To achieve a more compact checksum, a larger set of symbols is needed. The **Base64** encoding (Josefsson, 2006) uses a 64-character alphabet (`a-z`, `A-Z`, `0-9`, `-`, and `/`). With this, each character can encode six ($\log_2 64$) bits, i.e. a 160 bits can be represented by $\lceil 160/6 \rceil = 27$ characters.[6]

The original SEGUID method uses **Base64**, modulo the mandatory, non-informative padding symbol, to encode the **SHA-1** hash, resulting in a 27-character checksum. This was chosen to get a compact representation of the hash, while still being text-friendly. One problem with a **Base64**-based checksum is that it cannot be used as part of a filename, because of the forward slash (`/`), which is used to represent directories on most file systems. For similar reason, it cannot be used as-is as part of a Uniform Resource Locator (URL), where symbols `/` and `+` have special meaning. In order to use a SEGUID checksum in a URL, these symbols need to be re-encoded. To address these needs, a filename- and URL-safe variant of **Base64** is the **Base64url** encoding (Josefsson, 2006). It substitutes forward slashes (`/`) and plus symbols (`+`) with underscores (`_`) and minus symbols (`-`). The **Base64url** algorithm is linear ($O(n)$) in the number of bits, which is always 160 bits when using **SHA-1**.

In contrast to the original SEGUID, for SEGUID v2, we choose to use the **Base64url** encoding. For example, the linear single-stranded DNA sequence `ACAGATT` yields the SEGUID v1 checksum `mtrvbtuwr6/MoBxvtm4BEpv+jKQ`, while the SEGUID v2 checksum is `mtrvbtuwr6_MoBxvtm4BEpv-jKQ` (Table 1).

2.4 SEGUID v2 checksums have a prefix

In order to preserve what type of information is encoded in the checksum, SEGUID v2 checksums are by default prefixed with either `lsseguid`, `csseguid`, `ldseguid`, or `cdseguid`, followed by an equal sign (`=`). For example, `lsseguid=IQiZThf2zKn_I1KtqStlEdsHYDQ` indicates a SEGUID v2 checksum for a linear single-strand sequence.

[6]Technically, **Base64** encoding requires a multiple of four symbols. To meet this requirement, the 27-character string representing a **SHA-1** hash should be padded with a special symbol (`=`) at the very end.

2.5 SEGUID Short ID

For practical purposes, SEGUID v2 defines a *short form* of its checksum. Specifically, a *SEGUID Short ID* comprise the first six (6) characters of the checksum without the prefix. For example, the Short ID of the SEGUID v2 checksum `lsseguid=IQiZThf2zKn_I1KtqStlEdsHYDQ` is `IQiZTh`.

While we designed the SEGUID v2 checksum to be text-friendly, we designed the Short ID to be human-friendly. The *SEGUID Short ID* provides a compact way to reference a specific sequence. The short length makes it easy to communicate in both written and spoken form, and they allow for quick manual comparisons. It is also feasible to memorize and recognize, especially in projects that have a few sequences that are commonly used.

The risk for collisions among Short ID is still relatively small, especially for any given project working with a limited set of sequences. Theoretically, a six-character Short ID can encode $2^{36} = 1,073,741,824$ different sequences[7].

2.6 Supported sequence types and alphabets

The SEGUID v2 algorithm is designed to support DNA, RNA, and proteins, but also sequences with a custom alphabet (Table 2). Formally, the input sequence must not include symbols outside of the declared *case-sensitive* alphabet. By requiring case sensitivity, we allow for a richer set of custom alphabets, but we also avoid the risk of introducing non-deterministic behavior from, for instance, automatically coercing lower-case letters to upper-case ones. In addition, we limit alphabets to digits, and upper-, and lower-case letters. This is to assure a well-defined lexicographic ordering of symbols (per the 7-bit ASCII character set). Importantly, there are *reserved symbols* that must never be part of the alphabet. Specifically, the dash symbol (`-`), the semicolon (`;`), and the newline (`\n`) are reserved symbols.

3 Results

SEGUID v2 is designed to be robust against mistakes in order to prevent incorrect checksums from propagating in the scientific process. For instance, part of the requirements is that the implementation should validate that correctness of the input sequence, e.g. all symbols are part of the alphabet and double-stranded sequences are correctly complementary. At the same time as being strict, SEGUID v2 is designed to be flexible such that it can support

[7] The Short ID holds $6\,\text{symbols} \cdot \log_2 64\,\text{bits/symbol} = 36\,\text{bits}$ (22.5%) of the original 160 bits of the **SHA-1** hash.

Alphabet	#Symbols	Complementary	Definition
DNA	4	Bijective	Nucleotides A↔T, C↔G.
Extended DNA	15	Bijective	DNA + K↔M, R↔Y, S↔S, W↔W, B↔V, D↔H, N↔N.
RNA	4	Bijective	Nucleotides A↔U, C↔G.
Extended RNA	15	Bijective	RNA + K↔M, R↔Y, S↔S, W↔W, B↔V, D↔H, N↔N.
Protein	20	No	Amino acids A, C, D, E, F, G, H, I, K, L, M, N, P, Q, R, S, T, V, W, Y, plus U, O.
Extended protein	26	No	Protein + B, J, Z, X.
Custom	$1 - 60$	Optional	Digits (0-9), upper-case letters (A-Z), lower-case letters (a-z).

Table 2: **Alphabets supported by SEGUID v2**. SEGUID v2 has built-in support for DNA, RNA, and proteins. These alphabets comprise upper-case letters only. IUPAC is a standard to represent DNA, RNA, and amino-acid sequences with an "extended" set of symbols (IUPAC-IUB Commission on Biochemical Nomenclature, 1970, 1984). Custom alphabets, complementary or not, are also supported. The concept of complementarity does not apply to proteins. Proteins are effectively single-stranded sequences, meaning only the single-stranded SEGUID v2 methods apply to them. Amino-acid symbols J, O, and U are not part of the IUPAC standard.

existing and future sequences not already anticipated. For example, in the recent Viner et al. (2024), the authors define an expanded epigenetic DNA alphabet (Table 3) that encodes also known modifications to cytosine (`C`). Using this alphabet, methylation motifs can be represented as, for instance, `TTGmGCAA`, `AT2hAAAT`, and `TGCm1m1`. Their complementary counterparts are `TTGC1CAA`, `ATTT2hAT`, and `m1m1GCA`. They also define an "extended" version of this alphabet, with additional complementary symbol pairs. Both of these alphabets can be used with SEGUID v2 by defining them as custom alphabets.

Alphabet	#Symbols	Complementary	Definition
Expanded epigenetic alphabet	12	Bijective	DNA + `m`↔`1`, `h`↔`2`, `f`↔`3`, `c`↔`4`.
Ambiguous expanded epigenetic alphabet	20	Bijective	Expanded epigenetic alphabet + `w`↔`6`, `x`↔`7`, `y`↔`8`, `z`↔`9`.
DNA with Uracil	5	Non-bijective	`A`↔`T`, `A`↔`U`, `C`↔`G`.

Table 3: **Custom alphabets with SEGUID v2**. (i) The expanded epigenetic alphabet, defined in Table 1 of Viner et al. (2024), includes known modifications to cytosine (`C`) and symbols for each guanine (`G`) complementary to a modified nucleobase. (ii) The "ambiguous" version of this alphabet, defined in Table 2 of Viner et al. (2024), addresses the case when the knowledge on some of nucleotides is incomplete. (iii) Uracil (`U`) may occur naturally in DNA, which in case adenine (`A`) is complementary to either thymine (`T`) or uracil (`U`). This can be represented by a non-bijective complementary alphabet.

Another example arise from the fact that uracil (`U`) may occur naturally in DNA, e.g. phage DNA from dUTPase, uracil-DNA glycosylase (*dut ung*) mutant hosts (Wang and Mosbaugh, 1988), as an apoptosis inducer during *Holometabola* development (Muha et al., 2012), and as a mean for somatic hypermutation resulting from U:G mismatch repair during vertebrate antibody development (Muramatsu et al., 2000). See Figure 6 for an illustration.

This also occurs in the USER™ Uracil excision cloning protocol (Geu-Flores et al., 2007). In these cases, adenine (`A`) may bind to either thymine (`T`) or uracil (`U`), which can be described by a *non-bijective* complementary alphabet (Table 3).

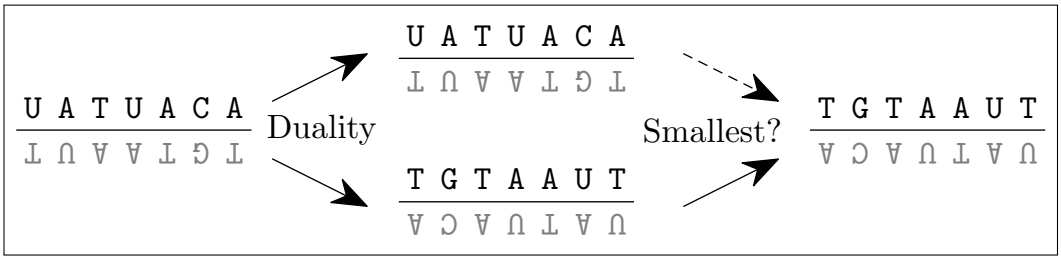


Figure 6: **The SEGUID v2 algorithm on double-stranded sequence of DNA with uracil.** The SEGUID v2 algorithm supports non-bijective complementary alphabets such as DNA with uracil (`U`). The input sequence `UATUACA`↔`TGTAAUT` (left and top middle) comprise basepairs `A`↔`T` and `A`↔`U`. It has a dual, reverse-complementary representation `TGTAAUT`↔`UATUACA` (bottom middle), which is also the lexicographically smallest (right).

4 Implementation and availability

4.1 Minimal, core software implementations

The SEGUID v2 algorithm, together with original SEGUID algorithm, is implemented and distributed as libre, open-source software under the MIT license. We provide small, self-contained implementations in JavaScript (ECMA International, 2021), Python (van Rossum and de Boer, 1991), and R (R Core Team, 2023) that have a minimal set of dependencies. For JavaScript, Python and R, we provide packages named **seguid**, which are available on npm, PyPi, and CRAN, respectively. These packages implement a standardized application protocol interface (API), e.g. `csseguid("GATTACA")`. They also provide a command-line interface (CLI), e.g. `npx seguid --type=lsseguid <<< 'GATTACA'`, `python -m seguid --type=csseguid <<< 'GATTACA'`, and `Rscript -e seguid::seguid --type=cdseguid <<< 'TATGCCAA;TTGGCATA'`. Support for additional programming languages are planned, specifically C++ and Rust, and will be made available via the SEGUID project website (`https://www.seguid.org`). Documentation and examples can be found via the SEGUID website.

4.2 Validation and testing

It is essential that any implementation of the SEGUID v2 checksum algorithm is correct and comply with the SEGUID v2 specifications as described here. For instance, we do not want to incorrectly produced checksums to be used

in production, appear in publications, or entering shared databases. We provide a rich set of unit and redundancy tests (see SEGUID website) to lower the risk for this to ever occur. These tests assert correctness, and identical behavior and results regardless of which programming language was used for the implementation. All new implementations must adhere to these tests in order to be considered SEGUID v2 compliant. These tests run on all releases and on a nightly basis. This test framework is designed to make it as simple as possible for a third-party contributor to run these tests. We use continuous-integration testing to validate all known implementations.

4.3 Higher-level implementations

The main purpose of the above **seguid** packages is to provide a minimal, language-agnostic API of the SEGUID v2 algorithm that can be imported in higher-level implementations. The scope of these packages will be kept at a minimum to avoid feature creep of both of the API and the SEGUID v2 algorithm. This way, the packages also serve as reference implementations and help lower the threshold for implementing the API in other languages.

For these reasons, we consider tasks like coercing a DNA sequence in lower-case letters to upper-case letters, trimming strings, and converting a sequence to its reverse-complementary counterpart to be out of scope. Such features have to be implemented by other tools and packages, e.g. Python packages **pydna** (Pereira et al., 2015) and **Biopython** (Cock et al., 2009; Bassi and Gonzalez, 2007). For example, **pydna** implements the SEGUID v2 methods for the `Dseqrecord` class, which holds double-stranded DNA. It also provides a rich set of utility functions for operating on DNA and RNA sequences. **Biopython** already implements the original SEGUID method and we anticipate it will also provide SEGUID v2 functionalities in a future release.

4.4 Graphical user interfaces (GUIs)

Some users are not comfortable with the provided command-line interfaces, or unexperienced with in one of the supported programming languages. In order to make SEGUID v2 accessible to these users it is important the method is also available through a Graphical User Interface (GUI). A readily available GUI will boost the adoption of SEGUID v2 to a broader audience as it also caters to users of varying expertise levels. Below are two GUIs that can be used for calculating SEGUID checksums.

The **SEGUID CALCULATOR** (Figure 7) is a browser-based GUI for the SEGUID v2 algorithm. It is implemented in JavaScript and allows for

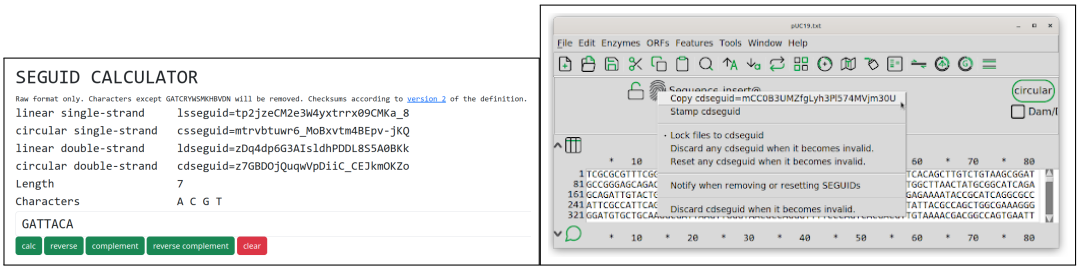


Figure 7: **GUI implementations of SEGUID checksums.** Two screenshots of GUI tools for calculating SEGUID v2 checksums. The **SEGUID CALCULATOR** tool (left) calculates the different types of SEGUID v2 checksums based on the Watson sequence as it is being entered. This a JavaScript-based implementation that fully runs within the web browser. The A plasmid Editor (**ApE**) tool (right) is specifically designed for double-stranded DNA. Because of this, it is sufficient to enter the Watson sequence, from which the Crick sequence is automatically inferred.

computations such that entered data never leaves the web browser or the local computer. This type of instant interactive feedback enhances understanding of how the checksum works.

The popular, freely available **ApE** (A plasmid Editor) (Davis and Jorgensen, 2022) allows calculating and stamping dsDNA sequences with SEGUID v2 checksums (Figure 7). **ApE** is available from the Jorgensen Lab website[8].

5 Applications

SEGUID checksums can be used to map sequence metadata both within and across biological databases. For example, by indexing different types of identifiers based on SEGUID checksums, the SEGUID checksum can serve as the common denominator across heterogeneous databases. For example, at the Argonne National Laboratory, this technique was used to build a real-time, interactive search engine for protein annotation data collected from multiple resources. For motivations similar to those of the Short ID, the search engine was designed to list matching candidates as soon as the user had typed three letters (Figure 8).

Analogously to how SEGUID checksums can be used for quick lookup of sequence data via database tools, they can use for navigating and lookin up sequence data hosted directly on the file system. For example, sequence-

[8] `https://jorgensen.biology.utah.edu/wayned/ape`

annotation data files of different kinds can be grouped in folders named as the corresponding SEGUID v2 checksums, or the individual files can comprise the checksum.

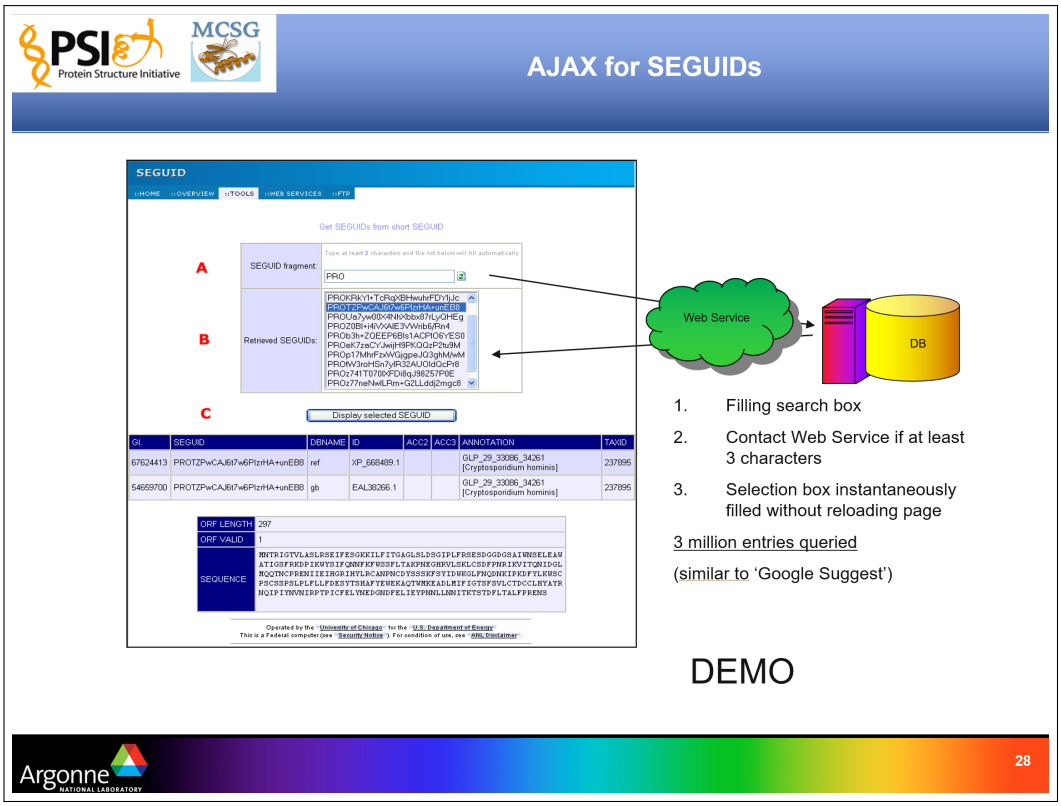


Figure 8: **Type-ahead database search based on partial SEGUID checksums.** Illustration of a how partial SEGUID checksums can be used to autocomplete searches as you type. In this example, the user enters `PRO` and the Ajax-based web interface instantaneously returns all SEGUID checksums starting with those characters. Sequence data is pulled from multiple databases concurrently for entries matching the selected SEGUID checksum.

The **ApE** tool and the **pydna** package can stamp and save sequence files in GenBank format[9] where the checksum is saved in the `COMMENT` section of the GenBank file (Figure 9). An existing checksum can be verified by each tool.

[9] `https://www.ncbi.nlm.nih.gov/genbank/samplerecord/`

ApE

```
LOCUS       New_DNA                    7 bp    DNA     circular     11-MAY-2023
DEFINITION  .
ACCESSION
VERSION
SOURCE      .
  ORGANISM  .
COMMENT     ApEinfo:methylated:0
COMMENT     ApEinfo:csseguid=mtrvbtuwr6_MoBxvtm4BEpv-jKQ
ORIGIN
        1 gattaca
//
```

pydna

```
LOCUS       name                       7 bp    DNA     circular UNK 01-JAN-1980
DEFINITION  description.
ACCESSION   id
VERSION     id
KEYWORDS    .
SOURCE      .
  ORGANISM  .
COMMENT     pydna csseguid=mtrvbtuwr6_MoBxvtm4BEpv-jKQ 2023-10-13T10:11:54
FEATURES    Location/Qualifiers
ORIGIN
        1 gattaca
//
```

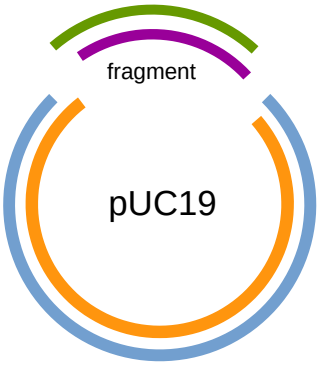
Figure 9: **SEGUID checksums added to GenBank sequence files.** The files were stamped with SEGUID checksum using the **ApE** tool (upper panel) or the **pydna** Python package (lower panel). In both cases, the checksum is added in the `COMMENT` field of the GenBank record. In addition, **pydna** appends an ISO-8601 timestamp after the checksum.

Cloning exercise

The plasmid vector pUC19 (2,686 bp, `cdseguid=zhw8Yrxfo3FO5DDccx4PamBVPCQ`) is a widely used cloning vector. The sequence is available from GenBank under the accession number `L09137.2`.

Simulate the non-directional cloning of the below dsDNA fragment using the unique *Bam*HI site in pUC19. Enter the SEGUID checksums for the two resulting vectors (pUC19_frag_A and pUC19_frag_B) in the **shared spreadsheet** next to your name.

```
gatccATGCAATAAg
    gTACGTTATTcctag
```



Sequences	Short ID
pUC19_frag_A	**`CzM-IG`**
pUC19_frag_B	**`S4WZki`**

Figure 10: **Cloning exercise.** Students, in undergraduate courses at the Department of Biology of the University of Minho, are given a cloning simulation problem (top) that will result in two vectors. They are given the Short IDs of the correct vectors for them to self-correct before filing the results.

A common use of the SEGUID checksum is in undergraduate teaching of practical molecular biology in the Department of Biology at the University of Minho. An example problem is depicted in Figure 10. The students are given a task, typically to simulate a sub-cloning problem on their computers. This kind of problem has a unique correct answer, making it possible to calculate checksums for the correct solution beforehand. The problem of assessing the students is simplified by the students access to a local or web-based SEGUID-capable software, e.g. **SEGUID CALCULATOR**. Giving access to a partial checksum for the correct answer can help the students to rapidly determine whether their procedures are incorrect. We have found this approach practical to teach in-silico cloning to over 60 students at a time. Overall, it lowers the number of mistakes filed and the efforts on reviewing homework assignments.

6 Discussions

The SEGUID v2 checksum is capable of providing the scientific community with stable keys for the many thousands of plasmids created in the process of designing new recombinant organisms or synthetic biological constructs. Important plasmid sequences can be stored in a distributed fashion, without the need for expensive centralized infrastructure. The integrity of sequences distributed among members of the scientific community can easily be verified. Using SEGUID v2 makes it trivial to confirm that members of a research groups are all using the same sequence for a particular DNA fragment containing a genetic part. The SEGUID v2 checksum would also enable a global reference service for circular DNA, such as the registry provided by EMBL-EBI at identifiers.org.

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