Sexual dimorphism in the distribution and density of regulatory interneurons in the striatum

Meghan Van Zandt[a,1], Deirdre Flanagan[b], Christopher Pittenger[a,c,d,e,f,2]

a. Department of Psychiatry, Yale University School of Medicine, New Haven, CT

b. Yale College, New Haven, CT

c. Yale Child Study Center, Yale University School of Medicine, New Haven, CT

d. Department of Psychology, Yale School of Arts and Sciences, New Haven, CT

e. Center for Brain and Mind Health, Yale University School of Medicine, New Haven, CT

f. Wu-Tsai Institute, Yale University, New Haven, CT

Corresponding Authors:

1. Meghan.vanzandt@yale.edu

2. Christopher.pittenger@yale.edu, Phone: (203)-974-7675

34 Park Street, Room W306, New Haven, CT, 06519, USA

Dr. Meghan Van Zandt is handling publication correspondence and is the primary producer of

data, Dr. Christopher Pittenger will handle all future correspondence following publication and

should Dr. Van Zandt depart the Pittenger Lab.

Abstract

Dysfunction of the cortico-basal circuitry – including its primary input nucleus, the striatum – contributes

to neuropsychiatric disorders, including autism and Tourette Syndrome (TS). These conditions show

marked sexual dimorphism, occurring more often in males than in females. Regulatory interneurons,

including cholinergic interneurons (CINs) and parvalbumin-expressing GABAergic fast spiking interneurons

(FSIs), are implicated in human neuropsychiatric disorders such as TS, and ablation of these interneurons

produces relevant behavioral pathology in male mice, but not in females. Here we investigate sexual

dimorphism in the density and distribution of striatal interneurons, using stereological quantification of

CINs, FSIs, and somatostatin-expressing (SOM) GABAergic interneurons in the dorsal and ventral striatum

in male and female mice. Males have a higher density of CINs than females, specifically in the dorsal

striatum; females have equal distribution between dorsal and ventral striatum. FSIs showed similar effects,

with a greater dorsal-ventral density gradient in males than in females. SOM interneurons were denser in

the ventral than in the dorsal striatum, with no sexual dimorphism. These sex differences in the density

and distribution of FSIs and CINs suggest a potential source of the sexual dimorphism seen in TS and autism

spectrum disorder.

Keywords: Sex Differences, Parvalbumin, GABAergic, Cholinergic, Basal Ganglia

Introduction

The cortico-basal circuitry, particularly the striatum (caudate and putamen), is a locus of pathology in

numerous neuropsychiatric disorders, including autism and Tourette's Syndrome (TS) (Swerdlow and Koob,

1987; Macpherson and Hikida, 2019; Chen et al., 2022). The striatum is the primary input nucleus and a

central hub of the basal ganglia circuitry; it is implicated in numerous neural processes, including motion,

effort, habit formation, motivation, and social behavior (Graybiel et al., 1994; Pennartz et al., 2009;

Graybiel and Grafton, 2015; Bamford and Bamford, 2019; Suzuki et al., 2021).

The striatum is composed of GABAergic medium spiny neurons (MSNs, comprising ~95% of the neurons

in the striatum in rodents)(Gerfen and Wilson, 1996) alongside several populations of regulatory

interneurons, including ChAT-expressing cholinergic interneurons (CINs) and multiple subtypes of

GABAergic interneurons, typically identified by their expression of markers such as parvalbumin (PV) and

somatostatin (SOM) (Kawaguchi et al., 1995; Kawaguchi, 1997; Marin et al., 2000; Muñoz-Manchado et

al., 2018). CINs innervate various GABAergic interneurons, including PV-expressing fast-spiking

interneurons (FSIs) (Chang and Kita, 1992; Kocaturk et al., 2022), reciprocally regulate dopamine release

(Yorgason et al., 2017; Howe et al., 2019; Chantranupong et al., 2023), and can regulate excitatory

glutamatergic input to MSNs (Pakhotin and Bracci, 2007; Ian Antón and Jun, 2011). FSIs are important

regulators responsible for fine tuning MSN firing (Bennett and Bolam, 1994; Orduz et al., 2013) and

integrate with other interneurons in coordinating movement bouts (Gritton et al., 2019) and regulating

behavior (Lee et al., 2017). SOM interneurons play a complex role in regulating MSNs, FSIs, and CINs

(Momiyama and Zaborszky, 2006; Cattaneo et al., 2019) and interact with dopaminergic afferents

(Hathway et al., 1999; Gazan et al., 2020). Despite being a small percentage of striatal neurons, these

interneurons critically regulate striatal function and behavioral output.

In alignment with this important functional role, interneuron pathology has been implicated in numerous

neuropsychiatric disorders (Kataoka et al., 2010; Rapanelli et al., 2017a; Poppi et al., 2021; Brady et al.,

2022). Deficits in cholinergic function are implicated in Parkinson's, Huntington's, and Alzheimer's Diseases,

schizophrenia, bipolar disorder, and attention deficit-hyperactivity disorder (Lim et al., 2014). Postmortem

studies in human patients with TS have shown significant reductions in striatal cholinergic (CINs) and

parvalbumin-expressing (PV) GABAergic interneurons (Kalanithi et al., 2005; Kataoka et al., 2010;

Lennington et al., 2016).

TS is diagnosed approximately twice as often in males as in females (Martino et al., 2013; Scahill et al.,

2014). Whether striatal interneuron pathology in TS is similarly sexually dimorphic is unknown, but

preclinical evidence is beginning to suggest that it may be. The striatal circuitry can be impacted by the

estrous cycle (Cao et al., 2018; Zachry et al., 2021); both GABAergic and cholinergic interneurons express

estrogen receptors (Almey et al., 2012; Almey et al., 2016). Furthermore, depletion of CINs during

development (Cadeddu et al, 2023), or conjoint depletion of both CINs and FSIs in adults (Rapanelli et al.,

2017b), produces dysregulated striatal activity and behavioral abnormalities of potential relevance to TS

and related conditions – repetitive behavioral pathology, anxiety, and social deficits – in male mice, but

not in females. This suggests an underlying sexual dimorphism in striatal interneurons, their regulation of

local microcircuits, and their role in the modulation of striatum-dependent behaviors.

Here, using mice as a model system, we provide the first rigorous stereological comparison between males

and females of the density and distribution of three different subtypes of striatal interneurons implicated

in striatal function and in neuropsychiatric disease: PV- and SOM-expressing GABAergic interneurons and

CINs.

Experimental Procedures

Experimental Design

All animal use followed protocols approved by Yale's Institutional Animal Care and Use Committee. Adult

male and female wild-type C57BL/6 mice (10-14 weeks old) were group housed in our animal facility in

self-ventilating cages, maintained on a 12hr light/dark cycle, and provided with food and water *ad libitium*.

For histology, mice were anesthetized (ketamine 100mg/kg and xylazine 10mg/kg, followed by isoflurane

inhalation until no response to noxious stimulus) followed by transcardial perfusion with 4%

paraformaldehyde in phosphate buffered saline (PBS) (Thermo Fisher) and brain extraction. Cryostat

sections were cut at 30 μm and mounted on Diamond White Glass Slides (Globe). For immunostaining for

identification and counting of interneurons, slides were incubated overnight at room temperature with

primary antibodies diluted in 0.1 M PBS containing 10% normal goat serum (Vector Labs) and 1% Triton-X

100 (AmericanBio). Primary antibodies were used as follows: Rabbit recombinant monoclonal anti-

somatostatin (1:250, Abcam, ab111912), mouse monoclonal anti-ChAT (1:250, Thermo Fisher, MA5-

31383), rabbit polyclonal anti-PV (1:250, Abcam, ab11427). Detection was performed with appropriate

secondary antibodies: polyclonal goat anti-rabbit coupled to Alexa Fluor 488 (1:1000, Thermo Fisher, A-

11008), polyclonal goat anti-rabbit coupled to Alexa Fluor 568 (1:1000, Thermo Fisher, A-11011),

recombinant polyclonal goat anti-mouse coupled to Alexa Fluor 488 (Thermo Fisher, A-28175), and

polyclonal goat anti-mouse coupled to Alexa Fluor 568 (1:1000, Thermo Fisher, A-11004). Sections were

then mounted and coverslipped in ProLong Gold with DAPI (Life Technologies).

Immunostained coronal striatal sections (30 μm, bregma 1.1mm-0.26mm) were visualized on a Zeiss

Scope.A1 using a Plan-APOCHROMAT 10x objective and an Axiocam 503 Mono at 100x magnification.

Microscope fields were systematically tiled over the entire dorsal and ventral striatum and stitched

together using the stitching plugin in FIJI (Schindelin et al., 2012) to provide a high-resolution image

throughout the extent of the structure. Only undamaged sections with consistent immunostaining

through the full extent of the striatum were included in analysis.

Whole striatum reconstructions were imported into StereoInvestigator 10 (MBF Biosciences) and regions

of interest were defined. For whole striatal analyses, the entire anatomical area of the striatum contained

in the image, bounded by the internal capsule/corpus callosum white matter and other landmarks

following the atlas of Paxinos (Franklin and Paxinos, 1997), was selected for quantification. For

dorsal/ventral subregion analyses we initially opted for an arbitrary division, since it is not clear *a priori*

what the meaningful subregions for quantification would be. The striatum was divided in half by a

horizontal line placed halfway between the dorsal and ventral extent of the structure (see Figures 1A, 2A,

3A). Neurons were counted using the fractionator tool in StereoInvestigator using a counting box size of

100 x 100 μm, excluding all neurons on the dashed bounding line. Data collected from left and right

striatum were averaged. Following dorsal/ventral analysis, we then analyzed striatal interneuron density

in the Caudate Putamen (CPu) and Nucleus Accumbens (Nacc) subregions, identified by alignment with

the atlas of Paxinos (Franklin and Paxinos, 1997) and then counted using StereoInvestigator as described

above. In total, 22 male and 17 female brains were analyzed, with two randomly selected sections per

animal from within the range specified above.

Statistics

All data were imported into JASP Statistical Software (Eric-Jan Wagenmakers, University of Amsterdam) for

statistical analysis. Data was tested for normality using the D'Agostino & Pearson test and outliers were

excluded. For direct comparisons between sexes for whole striatum and to compare ratios of dorsal to

ventral interneurons, a two-tailed unpaired t-test was performed. To compare between sexes for the

dorsal and ventral sub-regions of the striatum, a two-way repeated-measures ANOVA was used, followed

by Fisher's LSD post-hoc test (Table One). Graphs were generated using GraphPad Prism 10 (Graph Pad

Software, LLC). Whiskers on bar and whisker plots indicate the min and max values, with all data points

represented on graphs.

Results

We first quantified CIN density in male and female mice (Figure 1A-B). Male mice had a significantly higher

density of ChAT+ interneurons (Figure 1B; two-tailed t-test, $t[26] = 2.305$, $p=0.031$). Males and females

showed different patterns of CIN density in dorsal and ventral striatum (Figure 1C; 2way ANOVA: subregion

x sex interaction: $F[1.00, 8.00] = 5.84$, $p = 0.042$). The density of ChAT interneurons in the female striatum

was equal between the dorsal and ventral regions (Fisher's LSD: $p=0.43$). In contrast, males showed

significantly higher density of ChAT+ neurons in the dorsal region compared to the ventral region (Fisher's

LSD: $p=0.001$). The higher CIN density in males was restricted to the dorsal striatum (male vs female

Fisher's LSD: $p = 0.03$); there was no difference between males and females in the ventral striatum ($p=0.53$).

Analyzing the ratio of dorsal to ventral ChAT+ interneurons directly also showed higher ratios in males

(1.55 ± 0.13) than in females (1.04 ± 0.022)(two-tailed t-test, $t[29] = 3.670$, $p=0.001$).

Quantification of CIN density in anatomically defined subregions, the CPu and Nacc, showed similar effects

(Figure 1E). There were significant effects of both sex (Figure 1F; 2way ANOVA: $F[1.00, 16.00] = 4.769$, $p =$

0.044) and subregion (F[1.00, 16.00] = 5.991, p=0.026), with no interaction (sex x subregion F[1.00, 8.00]

= 0.614, p=0.456). There was a significantly higher density of ChAT+ interneurons in the CPu compared to

the NAcc in males (Fisher's LSD: p=0.037) but not in females (p=0.236).

Quantification of overall density of PV+ interneurons in whole striatum (Figure 2A-B) did not reveal

significant differences in density between males and females (Figure 2D; t-test, t[36] = 0.595; p=0.56).

However, analyzing across dorsal and ventral subregions uncovered sexually dimorphic effects (Figure 2C;

2-way ANOVA, sub-region x sex: F[1,11] = 6.94, p=0.012). In males, PV+ interneuron density was

significantly higher in the dorsal than in the ventral striatum (Fisher's LSD: p<0.001). In females, there was

a trend in the same direction, but it did not reach statistical significance (Fisher's LSD: p=0.07)(Figure 2D).

Analyzing the ratio of dorsal to ventral PV+ interneurons directly also showed higher ratios in males (1.57

± 0.12) than in females (1.18 ± 0.06)(two-tailed t-test, t[36] = 2.769, p=0.009).

Quantification of PV+ interneurons by anatomical subregion (Figure 2E) also revealed significant sex

differences, with a significant effect of subregion (Figure 2F; Two-way ANOVA: F(1.00, 16.00) = 51.62,

p<0.001) and a significant interaction between sex and subregion (F[1.00, 10.00] = 9.477, p=0.012). Both

sexes showed significantly higher density of PV+ interneurons in the CPu compared to the NAcc (Fisher's

LSD: males p<0.001, females p=0.0013); males showed a significantly higher density of PV+ interneurons

in the CPu compared to females (p=0.012).

SOM+ interneuron densities across the full striatum did not differ between males and females (Figure 3A-

B; t-Test: t[27] = 0.515, p=0.61). Comparing dorsal and ventral striatum revealed higher density of SOM+

cells in the ventral than in the dorsal striatum in both males and females (Figure 3C-D; 2-way ANOVA,

subregion: F[1,15] = 9.96, p = 0.006), but no effect of sex or interaction. Both males and females showed

dorsal/ventral ratios below 1 (0.91 ± 0.05 and 0.90 ± 0.06, respectively; two-tailed t-test, t[27] = 0.08,

p=0.93). Quantification of SOM+ interneurons by anatomical subregion (Figure 3E) showed a significant

effect of subregion (Figure 3F; Two-way ANOVA: F[1.00, 12.00] = 27.45, p<0.001), but no effect of sex or

subregion x sex interaction. Both males (Fisher's LSD: p=0.001) and females (p=0.001) showed significantly

denser SOM+ interneurons in the NAcc compared to the CPu.

Discussion

Interneurons of the striatum are critical to basal ganglia function (Kreitzer, 2009) and have emerged as a

locus of pathology in a number of neuropsychiatric conditions (Rapanelli et al., 2017a; Poppi et al., 2021).

Biological sex modulates basal ganglia structure and function (Giedd et al., 1997; Rijpkema et al., 2012),

and numerous studies show sexual dimorphism in conditions in which basal ganglia dysregulation is

implicated, including TS (Martino et al., 2013; Scahill et al., 2014) and autism (Loomes et al., 2017).

However, no studies to date have systematically examined sexual dimorphism in the structure and function

of striatal interneurons.

Several populations of striatal interneuron have been found to be reduced in number in post-mortem

studies of individuals with persistent adult TS (Kalanithi et al., 2005; Kataoka et al., 2010; Lennington et al.,

2016); these studies included both male and female subjects but were not designed to detect sexually

dimorphic effects. In mice, we have found experimental depletion of cholinergic interneurons (CINs),

parvalbumin-expressing fast-spiking interneurons (FSIs), or both to destabilize the striatum, resulting in

repetitive behavioral pathology, social deficits, and other behavioral abnormalities (Xu et al, 2015; Xu et

al, 2016; Rapanelli et al, 2017; Cadeddu et al, 2023). Interestingly, these effects are sexually dimorphic,

appearing in male but not female mice, despite equivalent levels of interneuron depletion (Rapanelli et al,

2017; Cadeddu et al, 2023). The underlying differences in striatal anatomy or function that lead to this

differential susceptibility remain unclear.

Here, we document intriguing differences between male and female adult mice in striatal interneuron

density and distribution. Differential density and distribution of interneurons suggests differences in

striatal function, and perhaps in resilience and vulnerability to pathophysiological processes.

Cholinergic interneurons (CINs) play a central role in the striatal microcircuitry. CINs regulate the release

of both dopamine (Cachope et al., 2012; Threlfell et al., 2012) and GABA (Nelson et al., 2014) from

dopaminergic afferents from the substantia nigra. They also modulate multiple populations of GABAergic

interneurons, as well as MSNs (Poppi et al., 2021). We find a significant difference between sexes in both

density and distribution of these interneurons: The density of CINs is significantly higher in the dorsal

striatum in males than in females. This matches observations in our previous studies, in which a higher

density of these interneurons was observed in males than in females (Rapanelli et al, 2017; Cadeddu et al,

2023); however, these previous observations were incidental, without the rigorous quantification applied

here. While female mice have relatively even distribution of ChAT+ interneurons between the dorsal and

ventral striatum, males a greater density in the dorsal compared to the ventral area.

The presence of more CINs in the dorsal striatum and CPu of males than of females is perhaps surprising,

as our previous work shows that males are more sensitive to disruption of these cells (Rapanelli et al,

2017; Cadeddu et al, 2023). It might have been expected that a higher basal density of CINs would render

males more resilient to CIN depletion. The current data suggest a different model, that the dorsal
regulatory circuitry in males may be more reliant on CINs than that of females and thus is less able to
accommodate pathological CIN disruption.

PV-expressing FSIs are highly active interneurons innervating a wide variety of targets in the striatum
through both inhibition and disinhibition circuits (Duhne et al., 2021), and are particularly important in
regulating MSNs through feedforward inhibition (Koós and Tepper, 1999; Gittis et al., 2010; Gittis et al.,
2011; Adler et al., 2013; Lee et al., 2017). We find effects of both biological sex and striatal subregion on
FSI density. Male mice again exhibit a significantly higher density in the dorsal striatum than in the ventral
striatum; females also show a difference in density in the same direction, but it is less robust. Because FSIs
are an important source of inhibition in the striatum, this may suggest that the male and female striatum
differ in their excitatory-inhibitory balance. We have previously found depletion of FSIs in the dorsal
striatum to produce both repetitive behavioral pathology and elevated anxiety-like behavior (Xu et al,
2016), though it is not yet clear if these effects are specific to males. When both FSIs and CINs are depleted,
behavioral effects are seen only in males, suggesting that there may be a sex-specific effect (Rapanelli et
al., 2017b).

FSIs and CINs separately regulate MSN activity (Adler et al., 2013; Gritton et al., 2019), and each other
(Chang and Kita, 1992; DeBoer and Westerink, 1994; Faust et al., 2016). Sexual dimorphism in density and
distribution follows the same general pattern for both interneuron types, with higher density in the dorsal
CPu region of the striatum and a greater dorsal-ventral gradient towards the NAcc in males than in females
in both cases. This may suggest a single developmental process underlying both effects.

We found no sex differences in the density or distribution of SOM-expressing GABAergic interneurons.
However, both males and females had more SOM-expressing interneurons in the ventral compared to the
dorsal striatum. This pattern, quite distinct from that seen in CIN and FSI distribution, indicates that the
sexual dimorphism is not uniform across all components of the striatal microcircuitry but rather
differentially affects different cells and processes. This reinforces the idea that differential density,
distribution, and function of interneurons in males and females leads to distinct regional differences in
information processing in the basal ganglia circuitry.

In sum, we find consistent sex differences in the density and distribution of striatal interneurons, with a
higher density of both CINs and FSIs in the dorsal striatum, as well as differences in interneuron density
between dorsal and ventral striatum, more prominently in males. The mechanisms underlying these

differential patterns of interneuron density and distribution remain to be elucidated but are likely to

involve sexually dimorphic modulation of developmental processes. Of note, estrogen receptors are

expressed on both cholinergic (Almey et al., 2012) and GABAergic interneurons (Almey et al., 2016),

providing a potential mechanism for sex hormones to regulate interneuron function. These findings

represent a starting point for future work analyzing the impact of these differences on sexual dimorphism

in both normal basal ganglia function and in the pathophysiology of a range of neuropsychiatric conditions,

such as TS and ASD.

Acknowledgements

The authors thank Betsy D'Amico and the staff of the Yale Animal Resource Core and Charles River

Laboratories for animal husbandry.

Funding sources

This work was supported by the grants F32MH123088 (MVZ), K24MH121571 (CP), and R01MH127259

(CP). This work was also supported by the State of Connecticut through its support of the Abraham

Ribicoff Research Facilities at the Connecticut Mental Health Center; the reported work represents the

views of the authors and not of the State of Connecticut.

Conflict of Interest

M. Van Zandt and D. Flanagan declare no competing financial interests. C Pittenger has consulted in the

past year for Biohaven Pharmaceuticals, Freedom Biosciences, Ceruvia Life Sciences, Nobilis

Therapeutics, Transcend Therapeutics, and F-Prime Capital Partners and has received research support

from Biohaven, Freedom, and Transcend; none of these relationships are related to the current work. Dr.

Pittenger holds equity in Alco Therapeutics and has pending patents related to the treatment of OCD, the

actions of psychedelic drugs, and the role of specific antibodies in neuroimmune pathophysiology; again,

none of these relationships are related to the current work.

References

Adler A, Katabi S, Finkes I, Prut Y, Bergman H (2013) Different correlation patterns of cholinergic and

GABAergic interneurons with striatal projection neurons. Frontiers in Systems Neuroscience 7.

Almey A, Milner TA, Brake WG (2016) Estrogen receptor α and G-protein coupled estrogen receptor 1 are

localized to GABAergic neurons in the dorsal striatum. Neuroscience Letters 622:118--123.

Almey A, Filardo EJ, Milner TA, Brake WG (2012) Estrogen receptors are found in glia and at extranuclear
neuronal sites in the dorsal striatum of female rats: evidence for cholinergic but not dopaminergic
colocalization. Endocrinology 153:5373-5383.

Bamford IJ, Bamford NS (2019) The Striatum's Role in Executing Rational and Irrational Economic Behaviors.
The Neuroscientist 25:475-490.

Bennett BD, Bolam JP (1994) Synaptic input and output of parvalbumin-immunoreactive neurons in the
neostriatum of the rat. Neuroscience 62:707-719.

Brady MV, Mariani J, Koca Y, Szekely A, King RA, Bloch MH, Landeros-Weisenberger A, Leckman JF,
Vaccarino FM (2022) Mispatterning and interneuron deficit in Tourette Syndrome basal ganglia
organoids. Molecular Psychiatry 27:5007-5019.

Cachope R, Mateo Y, Mathur BN, Irving J, Wang HL, Morales M, Lovinger DM, Cheer JF (2012) Selective
activation of cholinergic interneurons enhances accumbal phasic dopamine release: setting the
tone for reward processing. Cell Rep 2:33-41.

Cao J, Willett JA, Dorris DM, Meitzen J (2018) Sex Differences in Medium Spiny Neuron Excitability and
Glutamatergic Synaptic Input: Heterogeneity Across Striatal Regions and Evidence for Estradiol-
Dependent Sexual Differentiation. Frontiers in Endocrinology 9:173.

Cattaneo S, Zaghi M, Maddalena R, Bedogni F, Sessa A, Taverna S (2019) Somatostatin-Expressing
Interneurons Co-Release GABA and Glutamate onto Different Postsynaptic Targets in the Striatum.
bioRxiv:566984.

Chang HT, Kita H (1992) Interneurons in the rat striatum: relationships between parvalbumin neurons and
cholinergic neurons. Brain Research 574:307-311.

Chantranupong L, Beron CC, Zimmer JA, Wen MJ, Wang W, Sabatini BL (2023) Dopamine and glutamate
regulate striatal acetylcholine in decision-making. Nature 621:577-585.

Chen J, Tian C, Zhang Q, Xiang H, Wang R, Hu X, Zeng X (2022) Changes in Volume of Subregions Within
Basal Ganglia in Obsessive–Compulsive Disorder: A Study With Atlas-Based and VBM Methods.
Frontiers in Neuroscience 16.

DeBoer P, Westerink BHC (1994) GABAergic Modulation of Striatal Cholinergic Interneurons: An In Vivo
Microdialysis Study. Journal of Neurochemistry 62:70-75.

Duhne M, Lara-González E, Laville A, Padilla-Orozco M, Ávila-Cascajares F, Arias-García M, Galarraga E,
Bargas J (2021) Activation of parvalbumin-expressing neurons reconfigures neuronal ensembles in
murine striatal microcircuits. European Journal of Neuroscience 53:2149-2164.

Faust TW, Assous M, Tepper JM, Koós T (2016) Neostriatal GABAergic Interneurons Mediate Cholinergic
Inhibition of Spiny Projection Neurons. The Journal of Neuroscience 36:9505-9511.

Franklin KBJ, Paxinos G (1997) The mouse brain in stereotaxic coordinates. San Diego: Academic Press San
Diego.

Gazan A, Rial D, Schiffmann SN (2020) Ablation of striatal somatostatin interneurons affects MSN
morphology and electrophysiological properties, and increases cocaine-induced hyperlocomotion
in mice. Eur J Neurosci 51:1388-1402.

Gerfen CR, Wilson CJ (1996) Chapter II The basal ganglia. In: Handbook of chemical neuroanatomy, pp 371-
468: Elsevier.

Giedd JN, Castellanos FX, Rajapakse JC, Vaituzis AC, Rapoport JL (1997) Sexual dimorphism of the
developing human brain. Prog Neuropsychopharmacol Biol Psychiatry 21:1185-1201.

Gittis AH, Nelson AB, Thwin MT, Palop JJ, Kreitzer AC (2010) Distinct roles of GABAergic interneurons in the
regulation of striatal output pathways. J Neurosci 30:2223-2234.

Gittis AH, Leventhal DK, Fensterheim BA, Pettibone JR, Berke JD, Kreitzer AC (2011) Selective inhibition of
striatal fast-spiking interneurons causes dyskinesias. J Neurosci 31:15727-15731.

Graybiel AM, Grafton ST (2015) The striatum: where skills and habits meet. Cold Spring Harb Perspect Biol
7:a021691.

Graybiel AM, Aosaki T, Flaherty AW, Kimura M (1994) The Basal Ganglia and Adaptive Motor Control.
Science 265:1826-1831.

Gritton HJ, Howe WM, Romano MF, DiFeliceantonio AG, Kramer MA, Saligrama V, Bucklin ME, Zemel D,
Han X (2019) Unique contributions of parvalbumin and cholinergic interneurons in organizing
striatal networks during movement. Nature Neuroscience 22:586-597.

Hathway GJ, Humphrey PP, Kendrick KM (1999) Evidence that somatostatin sst2 receptors mediate striatal
dopamine release. Br J Pharmacol 128:1346-1352.

Howe M, Ridouh I, Allegra Mascaro AL, Larios A, Azcorra M, Dombeck DA (2019) Coordination of rapid
cholinergic and dopaminergic signaling in striatum during spontaneous movement. eLife
8:e44903 , citation = eLife 42019;44908 e44903.

Ian Antón O, Jun BD (2011) Cholinergic modulation of synaptic integration and dendritic excitability in the
striatum. Current Opinion in Neurobiology 21:425-432.

Kalanithi PS, Zheng W, Kataoka Y, DiFiglia M, Grantz H, Saper CB, Schwartz ML, Leckman JF, Vaccarino FM
(2005) Altered parvalbumin-positive neuron distribution in basal ganglia of individuals with
Tourette syndrome. Proc Natl Acad Sci U S A 102:13307-13312.

Kataoka Y, Kalanithi PS, Grantz H, Schwartz ML, Saper C, Leckman JF, Vaccarino FM (2010) Decreased number of parvalbumin and cholinergic interneurons in the striatum of individuals with Tourette syndrome. J Comp Neurol 518:277-291.

Kawaguchi Y (1997) Neostriatal cell subtypes and their functional roles. Neurosci Res 27:1-8.

Kawaguchi Y, Wilson CJ, Augood SJ, Emson PC (1995) Striatal interneurones: chemical, physiological and morphological characterization. Trends Neurosci 18:527-535.

Kocaturk S, Guven EB, Shah F, Tepper JM, Assous M (2022) Cholinergic control of striatal GABAergic microcircuits. Cell Rep 41:111531.

Koós T, Tepper JM (1999) Inhibitory control of neostriatal projection neurons by GABAergic interneurons. Nat Neurosci 2:467-472.

Kreitzer AC (2009) Physiology and pharmacology of striatal neurons. Annu Rev Neurosci 32:127-147.

Lee K, Holley SM, Shobe JL, Chong NC, Cepeda C, Levine MS, Masmanidis SC (2017) Parvalbumin Interneurons Modulate Striatal Output and Enhance Performance during Associative Learning. Neuron 93:1451-1463 e1454.

Lennington JB, Coppola G, Kataoka-Sasaki Y, Fernandez TV, Palejev D, Li Y, Huttner A, Pletikos M, Sestan N, Leckman JF, Vaccarino FM (2016) Transcriptome Analysis of the Human Striatum in Tourette Syndrome. Biol Psychiatry 79:372-382.

Lim SAO, Kang UJ, McGehee DS (2014) Striatal cholinergic interneuron regulation and circuit effects. Frontiers in Synaptic Neuroscience 6.

Loomes R, Hull L, Mandy WPL (2017) What Is the Male-to-Female Ratio in Autism Spectrum Disorder? A Systematic Review and Meta-Analysis. J Am Acad Child Adolesc Psychiatry 56:466-474.

Macpherson T, Hikida T (2019) Role of basal ganglia neurocircuitry in the pathology of psychiatric disorders. Psychiatry and Clinical Neurosciences 73:289-301.

Marin O, Anderson SA, Rubenstein JL (2000) Origin and molecular specification of striatal interneurons. J Neurosci 20:6063-6076.

Martino D, Macerollo A, Leckman JF (2013) Neuroendocrine aspects of Tourette syndrome. Int Rev Neurobiol 112:239---279.

Momiyama T, Zaborszky L (2006) Somatostatin Presynaptically Inhibits Both GABA and Glutamate Release Onto Rat Basal Forebrain Cholinergic Neurons. Journal of Neurophysiology 96:686-694.

Muñoz-Manchado AB, Gonzales CB, Zeisel A, Munguba H, Bekkouche B, Skene NG, Lönnerberg P, Ryge J, Harris KD, Linnarsson S, Hjerling-Leffler J (2018) Diversity of Interneurons in the Dorsal Striatum Revealed by Single-Cell RNA Sequencing and PatchSeq. Cell Reports 24:2179--2190.e2177.

Nelson AB, Hammack N, Yang CF, Shah NM, Seal RP, Kreitzer AC (2014) Striatal cholinergic interneurons
Drive GABA release from dopamine terminals. Neuron 82:63-70.

Orduz D, Bischop DP, Schwaller B, Schiffmann SN, Gall D (2013) Parvalbumin tunes spike-timing and
efferent short-term plasticity in striatal fast spiking interneurons. The Journal of Physiology
591:3215-3232.

Pakhotin P, Bracci E (2007) Cholinergic Interneurons Control the Excitatory Input to the Striatum. The
Journal of Neuroscience 27:391-400.

Pennartz CMA, Berke JD, Graybiel AM, Ito R, Lansink CS, Meer Mvd, Redish AD, Smith KS, Voorn P (2009)
Corticostriatal Interactions during Learning, Memory Processing, and Decision Making. The
Journal of Neuroscience 29:12831-12838.

Poppi LA, Ho-Nguyen KT, Shi A, Daut CT, Tischfield MA (2021) Recurrent Implication of Striatal Cholinergic
Interneurons in a Range of Neurodevelopmental, Neurodegenerative, and Neuropsychiatric
Disorders. Cells 10.

Rapanelli M, Frick LR, Pittenger C (2017a) The Role of Interneurons in Autism and Tourette Syndrome.
Trends Neurosci 40:397-407.

Rapanelli M, Frick LR, Xu M, Groman SM, Jindachomthong K, Tamamaki N, Tanahira C, Taylor JR, Pittenger
C (2017b) Targeted Interneuron Depletion in the Dorsal Striatum Produces Autism-like Behavioral
Abnormalities in Male but Not Female Mice. Biol Psychiatry 82:194-203.

Rijpkema M, Everaerd D, van der Pol C, Franke B, Tendolkar I, Fernández G (2012) Normal sexual
dimorphism in the human basal ganglia. Hum Brain Mapp 33:1246-1252.

Scahill L, Specht M, Page C (2014) The Prevalence of Tic Disorders and Clinical Characteristics in Children.
J Obsessive Compuls Relat Disord 3:394-400.

Schindelin J, Arganda-Carreras I, Frise E, Kaynig V, Longair M, Pietzsch T, Preibisch S, Rueden C, Saalfeld S,
Schmid B, Tinevez J-Y, White DJ, Hartenstein V, Eliceiri K, Tomancak P, Cardona A (2012) Fiji: an
open-source platform for biological-image analysis. Nature Methods 9:676-682.

Suzuki S, Lawlor VM, Cooper JA, Arulpragasam AR, Treadway MT (2021) Distinct regions of the striatum
underlying effort, movement initiation and effort discounting. Nature Human Behaviour 5:378-
388.

Swerdlow NR, Koob GF (1987) Dopamine, schizophrenia, mania, and depression: Toward a unified
hypothesis of cortico-striatopallido-thalamic function. Behavioral and Brain Sciences 10:197--208.

Threlfell S, Lalic T, Platt NJ, Jennings KA, Deisseroth K, Cragg SJ (2012) Striatal dopamine release is triggered
by synchronized activity in cholinergic interneurons. Neuron 75:58-64.

Yorgason JT, Zeppenfeld DM, Williams JT (2017) Cholinergic Interneurons Underlie Spontaneous Dopamine
Release in Nucleus Accumbens. J Neurosci 37:2086-2096.
Zachry JE, Nolan SO, Brady LJ, Kelly SJ, Siciliano CA, Calipari ES (2021) Sex differences in dopamine release
regulation in the striatum. Neuropsychopharmacology 46:491-499.

Figure One. Cholinergic Interneurons are denser and show a dorsally skewed distribution in the striatum

of male versus female mice. A. Representative whole striatum reconstruction of high-resolution

microscope images of ChAT-expressing interneurons from a male mouse striatum. Dashed lines indicate

counting regions for dorsal and ventral striatum. **B.** Stereological quantification of the density of ChAT+

interneurons per mm^2 over the whole mouse striatum showing a higher density of ChAT+ neurons in males

compared to females (*Two-tailed t-test, t[26] = 2.305, p=0.029). **C.** Quantification of the density of ChAT+

interneurons divided over the dorsal and ventral sub-regions of the striatum reveals a significant effect of

subregion (f[1.00, 14.00] = 10.42, p=0.006), a trend towards significant effect of sex (f[1,14] = 3.79, p=0.07),

and a significant interaction of subregion and sex (2way ANOVA, f[1,8] = 5.55, p=0.046). ChAT+

interneurons in the male dorsal striatum were significantly denser than both female dorsal striatum

(*Fisher's LSD, p=0.032) and male ventral striatum (**p=0.001). **D.** Alternative representation of the data

from C, showing distribution gradients of ChAT+ interneurons by individual animal. **E.** Representative

whole striatum reconstruction of high-resolution microscope images of ChAT-expressing interneurons,

with dashed lines indicating counting regions for CPu and NAcc. **F.** Quantification of the density of ChAT+

interneurons divided over the CPu and NAcc sub-regions of the striatum reveals a significant effect of

subregion (2way ANOVA, f[1,16] = 5.991, p=0.026) and sex (f[1,16] = 4.769, p=0.044); without interaction

(f[1,8] = 0.614, p=0.456). Males showed significantly higher ChAT+ interneuron density in the CPu

compared to the NAcc (*Fisher's LSD: p=0.037) while females did not (p=0.237) and there was a trend

towards higher density of ChAT in the male CPu compared to the female CPu (p=0.07).

Figure Two. Parvalbumin-expressing interneurons are more concentrated in the dorsal versus the

ventral striatum in male, but not female mice. A. Representative whole striatum reconstruction of high-

resolution microscope images of PV-expressing interneurons from a male mouse striatum. Dashed lines

indicate counting regions for dorsal and ventral striatum. **B.** Quantification of the density of PV+

interneurons per mm^2 over the whole mouse striatum using stereological methods shows similar densities

of PV+ neurons between sexes (Two-tailed t-test, t[36] = 0.595; p=0.56). **C.** Quantification of the density

of PV+ interneurons divided over the dorsal and ventral sub-regions of the striatum reveals a significant

effect of subregion (f[1,20] = 25.32), p<0.001) and a significant interaction of subregion x sex (2way ANOVA,

f[1,13] = 8.39, p=0.01). Males show a different distribution of PV+ interneurons than females, in that they

are significantly denser in the dorsal striatum (****Fisher's LSD, p<0.001), while females had a more equal

distribution between the dorsal and ventral regions (p=0.10). **D.** Alternative representation of the data

from C, showing distribution gradients of PV+ interneurons by individual animal. **E.** Representative whole

striatum reconstruction of high-resolution microscope images of PV-expressing interneurons, with dashed

lines indicating counting regions for CPu and NAcc. **F.** Quantification of the density of PV+ interneurons

divided over the CPu and NAcc sub-regions of the striatum reveals a significant effect of subregion (2way

ANOVA, f[1,16] = 51.62, p<0.001) and a significant interaction of sex and subregion (f[1,10] = 9, p=0.044).

Males showed significantly higher PV+ interneuron density in the CPu compared to the NAcc (****Fisher's

LSD: p<0.001) while females showed a similar, but less robust trend (***p=0.001). PV+ interneurons in the

CPu were significantly denser in males than in females (*p=0.012), but there were no differences in density

in the NAcc (p=0.96).

Figure Three. Somatostatin-expressing interneurons are denser in the ventral striatum in both male and

female mice. A. Representative whole striatum reconstruction of high-resolution microscope images of

SOM-expressing interneurons from a male mouse striatum. **B.** Quantification of the density of SOM+

interneurons per mm^2 over the whole mouse striatum using stereological methods, showing similar

densities of SOM+ neurons between sexes (Two-tailed t-test, t[27] = 0.515, p=0.61). **E.** Quantification of

the density of SOM+ interneurons divided over the dorsal and ventral sub-regions of the striatum reveals

an effect of subregion (f[1,14] = 8.981, p=0.009) but no interaction with sex (f[1,10] = 0.20, p=0.677). Both

males (*Fisher's LSD, p=0.04) and females (*p=0.03) show slight but significantly higher densities of SOM

interneurons in the ventral compared to the dorsal striatum, but no differences between sexes in either

region. **D.** Alternative representation of the data from C, showing distribution gradients of SOM+

interneurons by individual animal. **E.** Representative whole striatum reconstruction of high-resolution

microscope images of SOM-expressing interneurons, with dashed lines indicating counting regions for CPu

and NAcc. **F.** Quantification of the density of SOM+ interneurons divided over the CPu and NAcc sub-

regions of the striatum reveals a significant effect of subregion (2way ANOVA, f[1,12] = 27.45, p<0.001)

and no effect of sex (f[1,12] = 1.308, p=0.275) or interaction (f[1,8] = 0.019, p=0.894). Both males and

females showed significantly higher SOM+ interneuron density in the NAcc compared to the CPu

(**Fisher's LSD: p=0.001 for both) with no sex differences in density for either the CPu (p=0.369) or the

NAcc (p=0.302).

447