Thymic epithelial organoids mediate T cell development

Tania Hübscher[1], L. Francisco Lorenzo-Martín[1], Thomas Barthlott[2], Lucie Tillard[1], Jakob J. Langer[1], Paul Rouse[3], C. Clare Blackburn[3], Georg Holländer[2,4,5] and Matthias P. Lutolf[1,7,*]

[1] Laboratory of Stem Cell Bioengineering, Institute of Bioengineering, School of Life Sciences and School of Engineering, École Polytechnique Fédérale de Lausanne (EPFL), Lausanne, Switzerland

[2] Department of Biomedicine, University of Basel, Basel, Switzerland

[3] Centre for Regeneration, Institute for Repair and Regeneration, School of Biological Sciences, University of Edinburgh, Edinburgh, UK

[4] Department of Paediatrics and Institute of Developmental and Regenerative Medicine, University of Oxford, Oxford, UK

[5] Pediatric Immunology, Department of Biomedicine, University of Basel and University Children's Hospital Basel, Basel, Switzerland

[6] Department of Biosystems Science and Engineering, Eidgenössische Technische Hochschule Zürich (ETHZ), Basel, Switzerland

[7] Institute of Human Biology (IHB), Pharma Research and Early Development, Roche Innovation Center Basel, Basel, Switzerland

[*]Corresponding author. Email: matthias.lutolf@epfl.ch

Abstract

Although the advent of organoids opened unprecedented perspectives for basic and translational research, immune system-related organoids remain largely underdeveloped. Here we established organoids from the thymus, the lymphoid organ responsible for T cell development. We identified conditions enabling thymic epithelial progenitor cell proliferation and development into organoids with in vivo-like transcriptional profiles and diverse cell populations. Contrary to two-dimensional cultures, thymic epithelial organoids maintained thymus functionality in vitro and mediated physiological T cell development upon reaggregation with T cell progenitors. The reaggregates showed in vivo-like epithelial diversity and ability to attract T cell progenitors. Thymic epithelial organoids provide new opportunities to study TEC biology and T cell development in vitro, pave the way for future thymic regeneration strategies and are the first organoids originating from the stromal compartment of a lymphoid organ.

Summary statement: Establishment of organoids from the epithelial cells of the thymus which resemble their in vivo counterpart and have thymopoietic ability in reaggregate culture.

Keywords: Thymus, Organoids, Thymic epithelial cells, Thymopoiesis

INTRODUCTION

Over the past two decades, organoids have revolutionized the field of stem cell biology. Recapitulating key elements of the architecture, multicellularity, or function of their native organs on a smaller scale (Rossi et al., 2018), organoids have opened up unprecedented opportunities for personalized medicine. These three-dimensional (3D) structures derived from stem or progenitor cells have been established from a wide variety of organs, particularly of the endodermal lineage (Rossi et al., 2018). However, despite the availability of organotypic cultures (e.g. tissue explants (Anderson and Jenkinson, 1998; Owen and Ritter, 1969) and reaggregates (Giger et al., 2022; Jenkinson et al., 1992; Wagar et al., 2021)) or engineering methods (Kim et al., 2019) (e.g. scaffolds (Asnaghi et al., 2021; Bourgine et al., 2018; Campinoti et al., 2020; Fan et al., 2015; Hun et al., 2016; Poznansky et al., 2000; Purwada and Singh, 2017) and organ-on-a-chip (Goyal et al., 2022)), bona fide immune system-related organoids are considerably underdeveloped. Modelling lymphoid organs is indeed particularly challenging, largely due to the intricate crosstalk between immune and stromal cells required for organ development and function (Anderson and Jenkinson, 1998).

One essential organ for adaptive immunity is the thymus as it functions as the site of T cell development. In the thymus, T cell progenitors undergo lineage commitment and various selection processes to ensure the formation of a diverse, functional, and self-tolerant T cell repertoire, essential for effective immune protection. The instruction of the developing T cells (termed thymocytes) is mostly mediated by thymic epithelial cells (TECs). These stromal cells originate from the pharyngeal endoderm and can be subdivided into cortical and medullary lineages, which mediate successive stages of T cell development.

The essential thymopoietic ability of TECs is however mostly lost in vitro, as traditional two-dimensional (2D) cultures fail to maintain their functionality (Anderson and Jenkinson, 1998; Anderson et al., 1998; Mohtashami and Zúñiga-Pflücker, 2006). Alternative approaches employing OP9 or MS5 cell lines have been developed to circumvent this limitation and study T cell development in vitro (Montel-Hagen et al., 2020; Seet et al., 2017), but the absence of TECs still prevents physiological modelling of T cell selection processes. Other efforts focused on obtaining TECs from pluripotent stem cells (Lai and Jin, 2009; Parent et al., 2013; Ramos et al., 2022; Sun et al., 2013) or through direct reprogramming (Bredenkamp et al., 2014), but these cells largely rely on in vivo grafting to reveal thymopoietic functionality. It was also shown that TECs can form colonies in Matrigel, but these cultures still require feeder cells and their functionality was not demonstrated (Lepletier et al., 2019; Meireles et al., 2017; Wong et al., 2014). Thus, currently the only existing way to preserve TEC functionality in vitro is through (reaggregate) thymic organ cultures, which are organotypic 3D cultures containing different cell types.

Here, in light of what has been achieved for other endoderm-derived epithelia, we postulated that TECs could be grown independently of other cell types as 3D organoids in an extracellular matrix-based hydrogel. We identified culture conditions allowing TECs to form organoids mirroring to some extent the native tissue, and proved their functionality through their ability to mediate T cell development upon reaggregation with T cell progenitors. This work establishes the first thymic epithelial organoids with in vitro thymopoietic ability and is generally the first demonstration of organoids originating from the stromal compartment of a lymphoid organ.

RESULTS AND DISCUSSION

Thymic epithelial cells grow and maintain marker expression in defined organoid culture conditions

To establish thymic epithelial organoids, we followed the approach used for other endodermal organs, which included dissociating the tissue, sorting the cells of interest, and seeding them in a basement membrane-rich hydrogel (Matrigel) (Fig. 1A and Fig. S1 A). Since organoids mostly develop from stem or progenitor cells, we focused on the embryonic thymus due to its higher abundance of thymic epithelial progenitor cells compared to the adult organ (Baran-Gale et al., 2020; Kadouri et al., 2020). Although previous attempts to culture TECs often used serum-containing medium (Bonfanti et al., 2010; Campinoti et al., 2020; Wong et al., 2014), we opted for defined organoid basal medium and investigated factors that could promote TEC growth. We hypothesized that mesenchyme-derived factors that have been shown to influence TEC populations both in vivo and in vitro (Alawam et al., 2020; Boehm and Swann, 2013; Chaudhry et al., 2016; James et al., 2021) could also be important for TEC growth in organoid cultures. Among these factors, we found FGF7 of particular interest, as it has recently been shown to sustain the expansion of thymic microenvironments without exhausting the epithelial progenitor pools in vivo (Nusser et al., 2022). Using E16.5 embryonic thymi, we showed that while sorted TECs failed to grow in organoid basal medium, adding FGF7 to the culture supported organoid formation (Fig. 1, B and C). To monitor organoid development, we performed time-lapse imaging from the time of seeding (Fig. 1D and Movie 1) and found that most organoids were derived from single cells with stem/progenitor properties.

Immunostaining confirmed that organoids were generated by thymus-derived EpCAM-positive cells (Fig. 1E). Single cells formed small organoids in which a large majority of cells were positive for Ki67 after 3 days (Fig. 1F), and both proliferating and non-proliferating cells were present 4 days later (Fig. S1B). To investigate whether these cell populations could recapitulate TEC diversity, including cortical and medullary types (cTECs and mTECs), we stained organoids for the cTEC marker Keratin 8 (KRT8), as well as for Keratin 5 (KRT5) and

with UEA1 lectin as mTEC markers. Overall, our TEC culture system demonstrated a canonical feature of organoids in the emergence of different cell types, with varying degrees of KRT5 expression (Fig. 1G) and the presence of both KRT8-positive and UEA1-reactive populations (Fig. S1C). In addition, at least some organoids were positive for MHCII (Fig. 1H), an important marker of TEC functionality required for the development of CD4+ T cells (Kadouri et al., 2020). TEC differentiation, function and maintenance being critically dependent on the transcription factor *Foxn1* (Žuklys et al., 2016), we further sought to detect transcripts for this master regulator using RNAscope. Unlike in standard 2D cultures where it is highly downregulated (Anderson et al., 1998; Mohtashami and Zúñiga-Pflücker, 2006), a clear *Foxn1* expression could be observed in organoids (Fig. 1I).

To benchmark thymic epithelial organoids against standard 2D culture, we performed bulk RNA sequencing. Unsupervised hierarchical clustering showed the higher transcriptional similarity of thymic epithelial organoids to freshly extracted TECs (in vivo) than to 2D-cultured TECs (Fig. 1J, left). Similarly, a differential expression analysis showed that the expression levels of some key TEC genes, including *Foxn1*, *Dll4* and *Psmb11*, were more similar between in vivo TECs and thymic epithelial organoids compared to 2D-cultured TECs (Fig. 1J, right). Conversely, *Il7* and *Cdh1* were maintained in 2D culture as previously reported (Anderson et al., 1998), and *Ly6a* (a marker of specific TEC subpopulations (Klein et al., 2023)) was upregulated. Lastly, gene set enrichment analysis performed on organoids at different time points confirmed the proliferation peak observed with staining (Fig. S1D).

Collectively, these findings show that the defined culture conditions identified herein allow TECs (i) to grow independently of other cell types (ii) to form organoids containing diverse cell populations and that are transcriptionally similar to in vivo TECs.

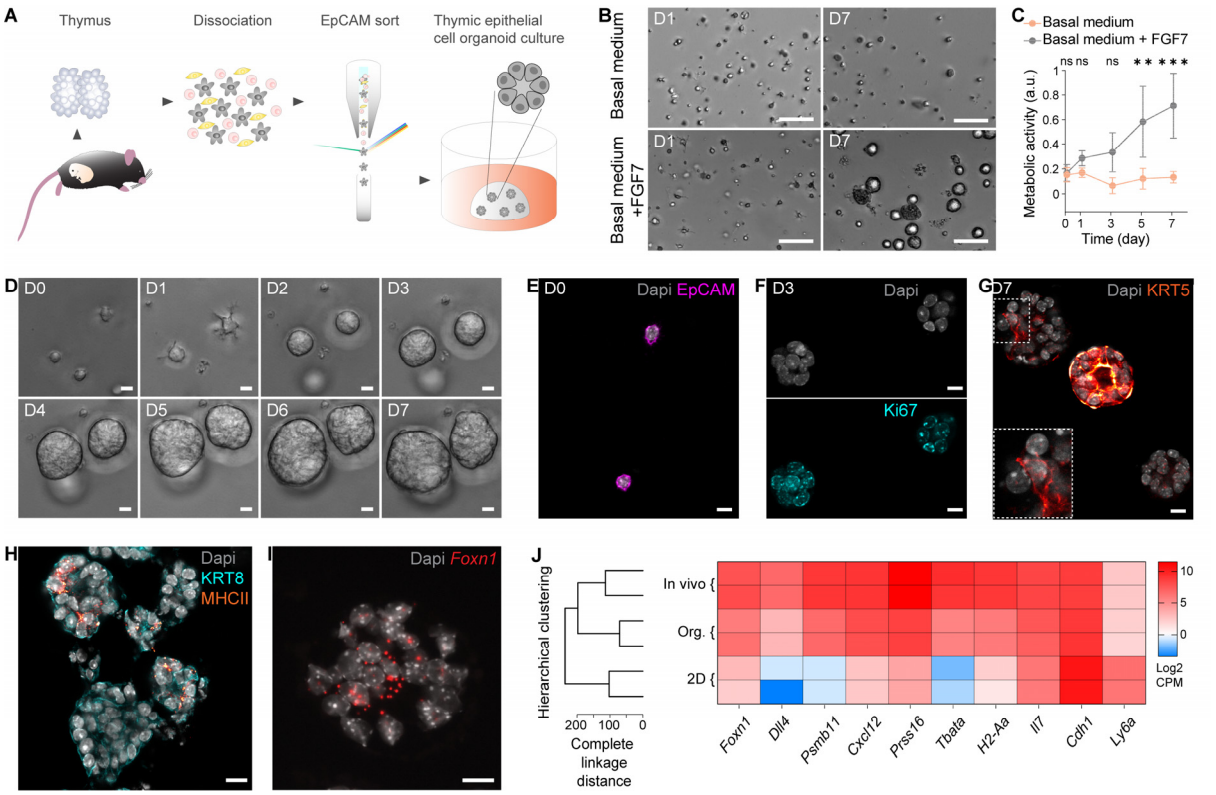

Fig. 1. Thymic epithelial cells grow and maintain marker expression in defined organoid culture conditions. (**A**) Schematic of the experimental workflow to isolate, select and culture thymic epithelial cells (TECs) to obtain organoids. (**B**) Brightfield images of TECs in organoid culture conditions one day (D1) and one week (D7) after seeding, in organoid basal medium and in organoid basal medium supplemented with FGF7. Scale bars, 100µm. (**C**) Metabolic activity (measured using resazurin) of TECs cultured in organoid basal medium and in organoid basal medium with FGF7. **: P = 0.0047, ***: P = 0.0005, ns: P > 0.05 (two-way ANOVA; n = 15 per condition, from 3 mice). Data represent mean ± standard deviation (SD). (**D**) One-week time course showing close up brightfield images of sorted TECs from single cells to multicellular organoids. Scale bars, 10µm. (**E**) Immunofluorescence image of individual EpCAM-positive (magenta) TECs immediately after seeding (D0) with nuclei counterstained using Dapi (grey). Scale bar, 10µm. (**F**) Immunofluorescence image of organoids three days after seeding demonstrating that cells undergo proliferation (Ki67 [cyan], Dapi stains nuclei [grey]). Scale bars, 10µm. (**G**) Immunofluorescence image of organoids showing different cell populations after one week in culture, here with medullary cells (KRT5 [red hot]) present in different patterns. Dapi counterstains nuclei (grey). Scale bar, 10µm. (**H**) Immunofluorescence image highlighting MHCII expression (orange) in D7 organoids also stained with KRT8 (cyan) and Dapi (nuclei, grey). Scale bar, 10µm. (**I**) RNAscope image of organoid at D7 showing *Foxn1* expression (red) with nuclei counterstained using Spectral Dapi (grey). Scale bar, 10µm. (**J**) Gene expression profiling. Left: dendrogram (using hclust) showing clustering of thymic epithelial organoids (Org.) with freshly extracted TECs (In vivo) and not TECs cultured in 2D (2D). Metrics is complete linkage distance. Right: heatmap displaying key TEC genes as well as *Cdh1* and *Ly6a* expression for the same three conditions. n = 2 mice per condition.

TECs cultured as organoids show in vitro functionality when reaggregated with T cell progenitors

To test the functionality of thymic epithelial organoids (i.e. their ability to mediate T cell development), we recapitulated the well-known reaggregate fetal thymic organ culture

167 (RFTOC) approach, wherein selected thymic cell populations are reaggregated together and
168 cultured at the air-liquid interface (Anderson et al., 1993; Jenkinson et al., 1992). To do so, we
169 dissociated TECs cultured as organoids and reaggregated them with an EpCAM-depleted
170 single cell suspension obtained from E13.5 thymi. We performed EpCAM-depletion in order to
171 keep the mesenchymal cells, which have been proven critical for T cell development (Anderson
172 et al., 1993). We used E13.5 embryonic thymi as source of T cell precursors because they
173 contain thymocytes at the earliest stages of development, prior to the expression of CD4 and
174 CD8 (thus referred to as double negative, DN) (Fig. S2A). This allows to easily monitor whether
175 T cell development happens in the reaggregates. To increase cell number and facilitate
176 handling, mouse embryonic fibroblasts (MEFs) were also added, as done previously (Sheridan
177 et al., 2009). We termed the RFTOCs formed with TECs from the organoid cultures organoid
178 RFTOCs (ORFTOCs) (Fig. 2A).

179 After 6 days in culture, ORFTOCs were dissociated and analyzed by flow cytometry (Fig. S2B).
180 At this point, thymocytes expressing both CD4 and CD8 (termed double positive, DP) and
181 constituting a developmental stage following the DN phenotype could be readily detected (Fig.
182 2, B and C), indicating that organoid-derived TECs mediated physiological progression of
183 thymocyte maturation. Notably, the proportion of DPs was similar to that observed in cultured
184 intact thymic lobes (i.e. fetal thymic organ cultures, FTOCs) (Fig. 2, C and D). Conversely,
185 reaggregating only the EpCAM-depleted fraction of E13.5 thymi and MEFs did not yield DP
186 thymocytes (Fig. 2, C and D, and Fig. S2C), demonstrating that organoid-derived TECs are
187 necessary for T cell development in ORFTOCs. Lastly, reaggregates with only organoid-
188 derived TECs and MEFs served as negative control and did not produce immune (CD45+)
189 cells (Fig. 2E), as opposite to the other conditions (Fig. S2B, C, D). To corroborate our findings,
190 we reaggregated organoid-derived TECs with the earliest DN subpopulation (DN1) sorted from
191 adult mice and MEFs, and could also observe thymocyte development (Fig. S2E). The
192 developmental kinetics was however faster in ORFTOCs containing E13.5-derived cells, as
193 expected for first wave early T cell precursors (Rothenberg, 2021).

194 Extending ORFTOC culture period from 6 to 13 days allowed thymocyte maturation to progress
195 further, as an increased proportion of cells expressed the αβ T cell receptor complex (TCR)
196 (Fig. S2F and Fig. 2F), and differentiated into the separate lineages of CD4+ and CD8+ single
197 positive (SP) thymocyte, respectively (Fig. S2F). FTOCs were again used as reference (Fig.
198 S2G) and demonstrated a comparable proportion of mature SP cells (Fig. S2H).

199 As expected for functional TECs, ORFTOCs were positive for *Foxn1* (Fig. 2G).
200 Morphologically, ORFTOCs also presented similarities to FTOCs, here highlighted by KRT8
201 and MHCII staining (Fig. 2H). UEA1 reactivity identified sparse medullary cells throughout

ORFTOCs (Fig 2, I and J), and CD3ε staining confirmed the presence of T cells in between EpCAM-positive epithelial cells (Fig. 2J).

In summary, we demonstrated that thymic epithelial organoids maintain their functionality and, when reaggregated with T cell progenitors, mediate T cell development similarly to intact thymic lobe cultures.

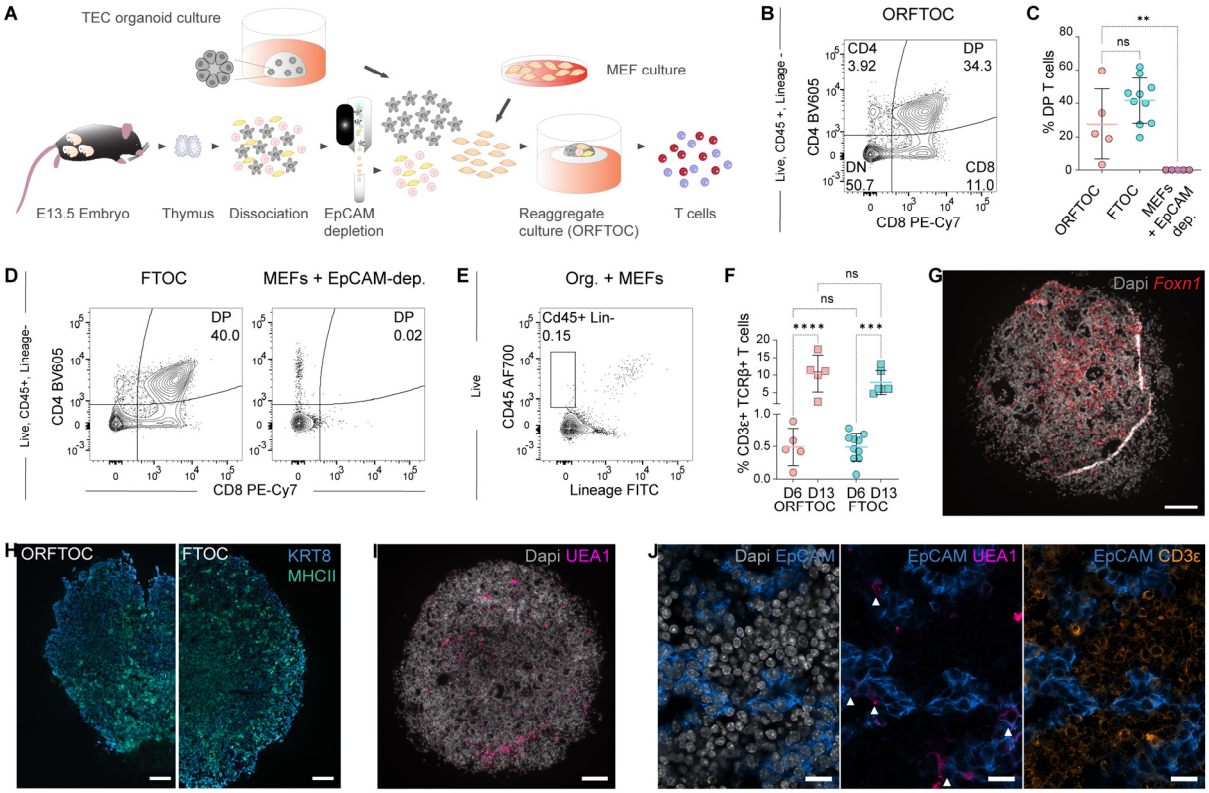

Fig. 2. TECs cultured as organoids show in vitro functionality when reaggregated with T cell progenitors. (**A**) Schematic of the experimental workflow to generate Organoid Reaggregate Fetal Thymic Organ Cultures (ORFTOCs) and analyze T cell development. (**B**) Flow cytometry plot showing T cell development in ORFTOCs after 6 days in culture (D6). Gating strategy is indicated on the left. (**C**) Proportion of double positive (DP) thymocytes at D6 in ORFTOCs and controls (fetal thymic organ cultures [FTOCs] and reaggregates containing only mouse embryonic fibroblasts [MEFs] and the EpCAM-depleted cells from thymic lobes). **: P = 0.0081, ns: P > 0.05 (Mood's median test with P-values adjusted with the false-discovery rate method; n = 5, 10 and 5 for ORFTOC, FTOC and MEFs + EpCAM-depleted cells, respectively, from 5 independent experiments). Graph represents individual datapoints with mean ± SD. (**D**) Flow cytometry plots showing T cell development at D6 in controls (FTOCs [left] and MEFs + EpCAM-depleted cells reaggregates [right]). Gating strategy is indicated on the left. (**E**) Flow cytometry plot showing the absence of a CD45+ Lineage- population in control reaggregates containing only TECs cultured as organoids and MEFs. Gating strategy is indicated on the left. (**F**) Proportion of CD3ε-positive, T cell receptor beta (TCRβ)-positive cells in ORFTOCs and FTOCs at day 6 and 13 (D13). ***: P = 0.0002, ****: P < 0.0001, ns: P > 0.05 (one-way ANOVA with Tukey's multiple comparisons test; n = 5, 5, 10 and 6 for D6 ORFTOCs, D13 ORFTOCs, D6 FTOCs and D13 FTOCs, respectively, from 5 independent experiments). Graph represents individual datapoints with mean ± SD. (**G**) RNAscope image of D13 ORFTOC section highlighting *Foxn1* expression (red) with nuclei counterstained using Spectral dapi (grey). Scale bar, 100µm. (**H**) Immunofluorescence images of D13 ORFTOC (left) and FTOC (right) sections showing KRT8 (blue) and MHCII staining (green). Scale bar, 100µm. (**I**) Immunofluorescence image of D13 ORFTOC section demonstrating the

presence of medullary cells (UEA1-reactivty [magenta]). Dapi counterstains nuclei (grey). Scale bar, 100µm. (**J**) Zoomed immunofluorescence images of D13 ORFTOC section showing epithelial cells (EpCAM [blue]) and nuclei (dapi [grey]) (left), medullary cells (UEA1-reacticity [bright pink]) co-staining with epithelial cells (middle), and T cells (CD3ε [amber]) in-between epithelial cells (right). Scale bars, 100µm.

ORFTOCs recapitulate in vivo-like TEC and T cell populations diversity and physiological T cell development

To further characterize the cell types in ORFTOCs, we profiled them and FTOC controls through single cell RNA sequencing (Fig. 3A). This analysis revealed three main clusters corresponding to the epithelial, immune, and mesenchymal compartments of ORFTOCs (Fig 3B). Unsupervised clustering identified 7 main clusters of epithelial cells (Fig. S3A), which we annotated according to in vivo datasets (Baran-Gale et al., 2020; Bautista et al., 2021; Bornstein et al., 2018; Gao et al., 2022; Nusser et al., 2022; Park et al., 2020): ‘early cTECs’, ‘cTECs’, ‘early mTECs’, ‘pre-Aire mTECs’, ‘Aire and Spink5 mTECs’, ‘tuft-like mTECs’, and ‘adult bipotent progenitor-like’ (Fig. S3B). For the immune cells, clusters covered the main T cell developmental stages defined in vivo (Cordes et al., 2022; Luis et al., 2016; Mingueneau et al., 2013; Park et al., 2020; Rothenberg, 2021; Zhou et al., 2019), spanning from progenitors to mature T cells (Fig. S3, C and D). Both ORFTOCs and FTOCs contributed to all subpopulations (Fig. 3, C and D), suggesting that ORFTOCs faithfully recapitulate the different cell types present in FTOC controls. The biggest differences were observed for clusters representing cTECs and early stages of T cell development: (i) more ‘early cTECs’ were present in FTOCs and (ii) more ‘cTECs’ and ‘thymus-seeding progenitor (TSP) to DN early 1’ and ‘TSP to DN early 2’ cells were present in ORFTOCs. A potential explanation for this is the difference in embryonic age between the epithelial cells used for organoids and FTOCs, as FTOCs were from E13.5 thymi to match T cell progenitors in ORFTOCs, while TECs in ORFTOCs were from E16.5 thymi.

To compare our in vitro populations with the in vivo thymus, we aligned our clusters to the mouse dataset of the reference atlas by Park et al. (Park et al., 2020) (Fig. 3, E and F). We found strong overlap in most epithelial cell types (Fig. 3E), with the cTECs aligning together and most in vitro mTEC clusters matching their in vivo counterparts. However, the ‘adult bipotent progenitor-like’ cluster was smaller in vivo compared to in vitro. Immune clusters from our dataset also matched clusters defined for in vivo populations (Fig. 3F), especially from the ‘DP blast’/‘DP (P)’ stage onwards and, most importantly, for the CD4 and CD8 stages (mature T cells).

Besides gene expression, we also studied in vitro TCR recombination dynamics through V(D)J sequencing, allowing us to map productive T cells bearing both TCR chains on the immune UMAP (Fig. 3G). The quantification of productive chains presenting all V(D)J regions showed that the recombination of the TCRβ (TRB) and -α (TRA) chains were mostly achieved prior to and at the DP stage, respectively (Fig. 3H), similarly to the Park dataset (Park et al., 2020). In addition, thymocytes underwent proliferation (marked by high *Cdk1* expression) in between the recombination stages (marked by high *Rag1* and *Rag2* expression) (Fig. 3I), which also aligns with in vivo data (Park et al., 2020; Rothenberg, 2021).

Taken together, these results show the transcriptional similarity of ORFTOCs to FTOCs and that ORFTOCs preserve in vivo-like TEC diversity and T cell development.

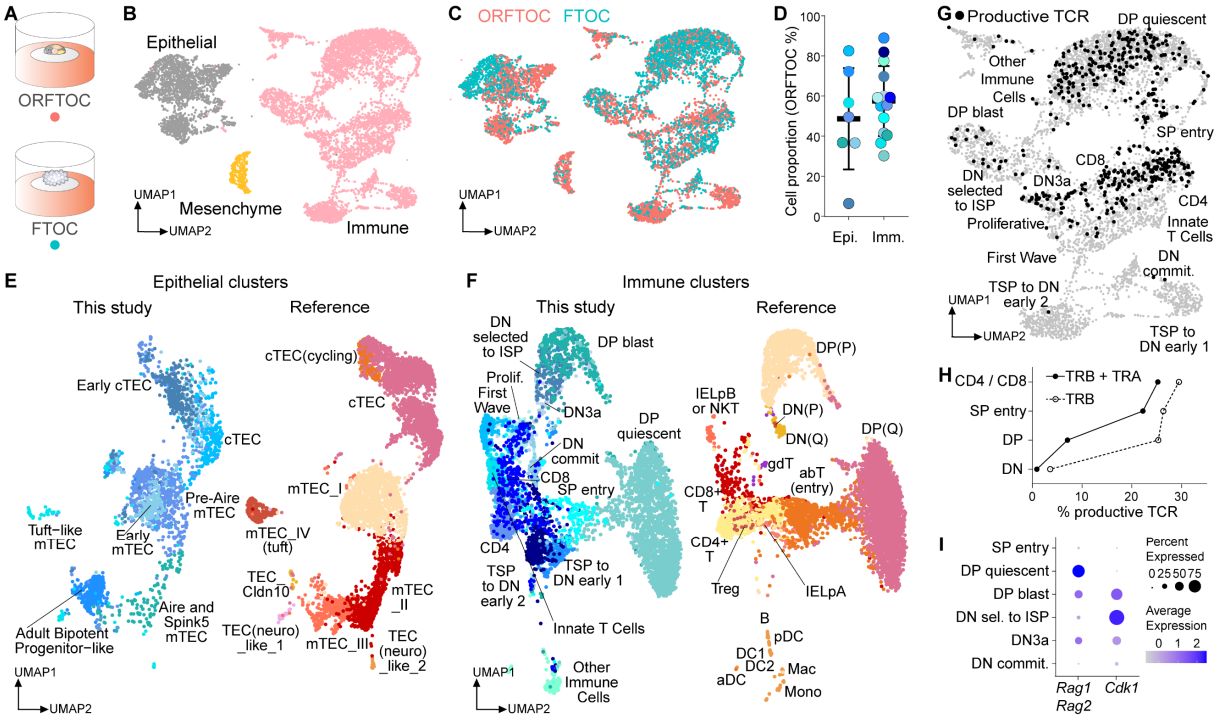

Fig. 3. ORFTOCs recapitulate in vivo-like TEC and T cell populations diversity and physiological T cell development. (**A**) Schematic of the conditions used for ORFTOC and FTOC single-cell RNA sequencing with hashtag antibodies (HTOs) and analyzed after 13 days in culture. (**B**) Uniform Manifold Approximation and Projection (UMAP) showing 3 main clusters corresponding to the main input populations (epithelial, immune and mesenchymal cells). (**C**) UMAP displaying ORFTOC and FTOC cells distribution in the different clusters. (**D**) Dot plot representing ORFTOC proportion for each cluster (dot) within the epithelial or immune main populations, as well as mean ORFTOC proportion and standard deviation. Dot colors are matching clusters colors (Fig. S3, A and C). No outliers within epithelial or immune compartments were identified by Grubbs test. (**E - F**) UMAPs showing the integration of the epithelial (E) and immune (F) clusters identified in this study (left) with the mouse dataset of the reference atlas by Park et al. (Park et al., 2020) (right). (**G**) UMAP of the immune cluster (grey), highlighting cells identified as productive and bearing both TCR chains (black). (**H**) Proportion of productive cells with rearranged TRB or both TRA and TRB chains for the main thymocyte developmental stages. (**I**) Dot plot representing the average expression level and the percentage of cells expressing the recombination enzymes *Rag1* and *Rag2* as well as the cyclin protein *Cdk1* during the recombination and proliferation stages of thymocyte development.

Epi: epithelial, Imm: immune, cTEC: cortical TECs, mTEC: medullary TECs, DN: double negative, ISP: intermediate single positive, Prolif: proliferative, commit: commitment, TSP: thymus-seeding progenitors, DP: double positive, (P): proliferative, (Q): quiescent, IELpB: intestinal intraepithelial lymphocytes precursor B, NKT: natural killer T, IELpA: intestinal intraepithelial lymphocytes precursor A, DC: dendritic cells, pDC: plasmacytoid dendritic cells, aDC; activated dendritic cells, Mac: macrophages Mono: monocytes, sel: selected.

ORFTOCs show thymus-like ability to attract new T cell progenitors and improved epithelial organization upon in vivo grafting

The thymus continuously attracts bone marrow-derived hematopoietic precursors and commits them to the T cell lineage (Lai and Kondo, 2007; Lavaert et al., 2020). To test whether ORFTOCs retain this crucial capacity, we transplanted them under the kidney capsule of syngeneic CD45.1 recipient mice (Fig. 4A). After 5 weeks, all grafts developed into sizeable thymus-like tissues (4/4 ORFTOCs [Fig. 4B], 4/4 FTOC controls). Using flow cytometric analyses (Fig. S4A), we identified all major thymocyte populations (DN, DP, CD4, CD8) in ORFTOC grafts (Fig. 4C), and their proportions were comparable to FTOC control grafts and control thymi (Fig. 4D). This result demonstrated that normal αβ-TCR T cell lineage maturation was supported in ORFTOC grafts. A further detailed analysis (Fig. S4, B and C) detected thymocytes at the DN3 to DN4 transition at the time of ORFTOC graft retrieval, a stage attesting to successful β-selection (Rothenberg, 2021). In addition, DP thymocytes expressing CD69, which indicates positive selection-induced TCR signaling (Steier et al., 2023), were present (Fig. S3 B and D). Together, these results illustrate ORFTOC graft ability to continuously attract and select blood-borne T cell progenitors. Finally, the presence of CD45.1+ mature thymocytes (classified as M1 [CD24+ CD69+] and M2 [CD24+ CD69-], respectively [Fig. 4E and Fig. S4E]) and of regulatory T cells (Fig. 4F and Fig. S4F) showed ORFTOC graft capacity to generate mature CD4 and CD8 T cells, impose their post-selection maturation and select T cells with a regulatory phenotype.

Histological staining showed that ORFTOC grafts, similar to FTOC grafts, displayed the characteristic differences in cellular densities between cortical and medullary areas seen in the native thymus (Gordon and Manley, 2011) (Fig. S4G and Fig. 4G). Immunostaining confirmed the presence of medullary areas (positive for KRT5 and reactive to UEA1) containing Aire-positive cells (Fig. 4G). These medullary areas were larger, better organized and more mature compared to those observed after in vitro culture only (Fig. 2I), likely due to continuous seeding with new T cell progenitors and prolonged crosstalk with immune cells (Irla et al., 2010).

In conclusion, kidney capsule transplants showed that organoid-derived TECs in ORFTOCs have the (i) capacity to mature and reach an organization resembling the native thymus and (ii) long-term ability to attract T cell progenitors and mediate physiological T cell development.

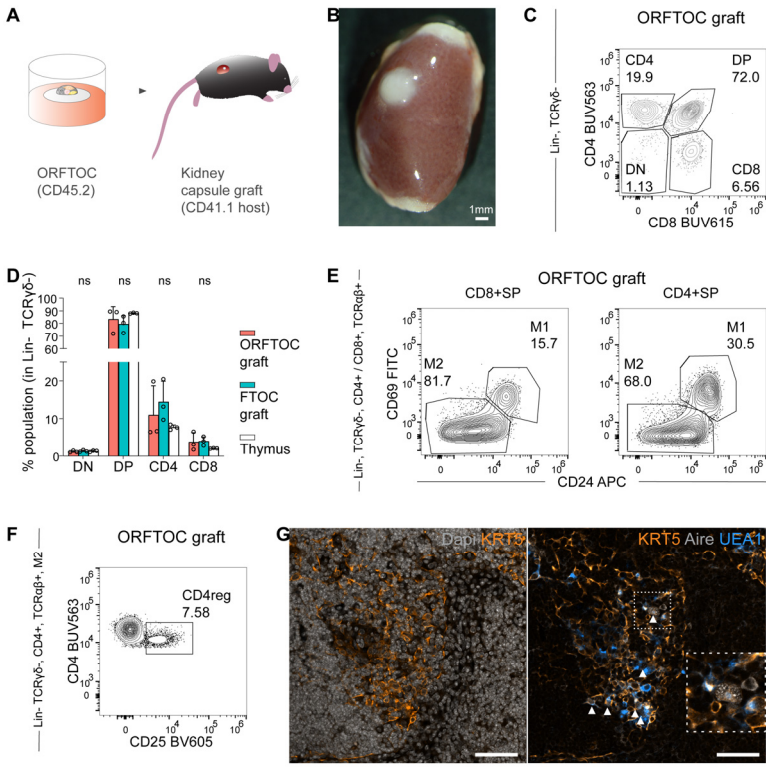

Fig. 4. ORFTOCs show thymus-like ability to attract new T cell progenitors and improved epithelial organization upon in vivo grafting. (**A**) Schematic representing the experimental design for the grafting of ORFTOCs under the kidney capsule. (**B**) Widefield image of an ORFTOC graft retrieved after 5 weeks. Scale bar, 1mm. (**C**) Flow cytometry plot showing host thymocyte development in ORFTOC grafts. Gating strategy is indicated on the left. (**D**) Proportion of the major thymocyte subpopulations in ORFTOC grafts, in control FTOC grafts and thymi. ns: P > 0.05 (one-way ANOVA for each subpopulation between conditions; n = 3 grafts/mice for each condition). Bar graph represents mean ± SD and individual datapoints. (**E**) Flow cytometry plots showing two separate post-selection stages (M1 and M2) within the CD8+ and CD4+ SP populations in ORFTOC grafts. Gating strategy is indicated on the left. (**F**) Flow cytometry plot highlighting the presence of CD4 regulatory T cells (CD4reg) within the M2 population in ORFTOC grafts. Gating strategy is indicated on the left. (**G**) Immunofluorescence images of ORFTOC graft section. Left: medullary cells (KRT5 [amber]) are present in the less dense area (Dapi [grey]). Right: UEA1-reactive (azure) and Aire-positive cells (grey, highlighted with arrowhead) are also present in the medullary region. Scale bars, 100µm.

In this study, we showed that stromal cells of a lymphoid organ, namely epithelial cells of the thymus, can be cultured as organoids similarly to cells from other endoderm-derived organs. We established TEC-specific culture conditions, characterized the organoids, and demonstrated their superiority in maintaining TEC marker expression compared to conventional 2D cultures. Reaggregating TECs from organoid cultures with T cell progenitors proved their functionality and ability to mediate T cell development. TEC and T cell populations in reaggregates resembled the native cell types, and T cell maturation was recapitulated in a physiological manner. Finally, kidney capsule transplants demonstrated the long-term

353 capability of organoid reaggregates to attract new T cell progenitors and mediate their entire
354 development.

355 Overall, this work addressed a long-standing challenge in the thymus field and presents the
356 first method to culture TECs independently of other cell types in a way that maintains their
357 thymopoietic ability. Although thymic epithelial organoids recapitulate many key organoid
358 features such as cell population diversity and possibility to be expanded and passaged,
359 maintaining their functionality in the long term remains challenging. This is probably linked to
360 some niche factors missing in the current relatively minimal culture conditions, which over time
361 either generally prevent functionality to be maintained or enrich for specific subsets that might
362 lack functionality (Gao et al., 2022). Future work including single-cell transcriptomic analysis
363 of the organoids will most likely help identify yet unexplored but necessary niche factors.

364 Thymic epithelial organoids nevertheless open up new opportunities to study T cell
365 development in vitro in a physiological manner and gain new insights into TEC biology. As
366 TECs undergo deterioration during aging and different medical conditions, the development of
367 the current and future culture conditions might also pave the way for novel thymus regeneration
368 strategies. Finally, to the best of our knowledge, the generation of bona fide organoids from
369 the stromal compartment of a lymphoid organ is unprecedented.

370

371

372 ## Materials and Methods

373 ### Mice

374 For all in vitro experiments, C57BL/6J mice were purchased from Charles River France and
375 maintained in the EPFL animal facility until use. For grafting experiments, Ly5.1 and C57BL/6J
376 mice were bred and maintained in the mouse facility of the Department of Biomedicine at the
377 University of Basel. For timed mating, noon of the day of the vaginal plug was considered as
378 day 0.5 of embryonic development (E0.5). Mice were housed in individual cages at 23°C ± 1°C
379 with a 12 h light/dark cycle, and supplied with food and water ad libitum. All animal work was
380 conducted in accordance with Swiss national guidelines, reviewed and approved by the
381 Cantonal Veterinary Offices of Vaud and of Basel-Stadt, license numbers VD3035.1, VD3823
382 and BS2321.
383

384 ### Isolation of thymic epithelial cells (TECs)

385 E16.5 embryonic thymi were dissected and collected in Eppendorf tubes containing FACS
386 buffer (PBS [Gibco Catalog No. 10010-015] + 2% fetal bovine serum [FBS] [Thermo Fisher
387 Scientific, Catalog No. 26140079]). Lobes were rinsed with PBS and digested with 475 µl
388 TrypLE (Gibco Catalog No. 12605-028) for 5 min at 37 °C under agitation (Eppendorf,
389 ThermoMixer C). Lobes were pipetted to promote dissociation, 25 µl DNase (Sigma-Aldrich,
390 Catalog No. 10104159001, from 1 mg/ml stock) was added and the tubes were incubated for
391 another 5 min. Lobes were again pipetted to help dissociation, TrypLE was quenched with 1ml
392 Adv. DMEM/F12 (Thermo Fisher Scientific, Catalog No. 12634028) containing 10 % FBS and
393 the cell suspension was filtered through a 40 µm strainer. The cells were pelleted and
394 resuspended in FACS buffer for staining with the following antibodies for 20 min at 4° C:
395 Ter119-FITC (BioLegend, Catalog No. 116205, 1/100), EpCAM-PE (BioLegend, Catalog No.
396 1198206, 1/80), PDGFR-α-APC (BioLegend, Catalog No. 135907, 1/40), PDGFR-β-ACP
397 (BioLegend, Catalog No. 136007, 1/40), CD31-PE-Cy7 (BioLegend, Catalog No. 102524,
398 1/160), MHCII-APC/Fire750 (BioLegend, Catalog No. 107651, 1/160), CD45-Pacific Blue
399 (BioLegend, Catalog No. 103126, 1/200). Dapi (Tocris, Catalog No. 4748, 0.5 ug/ml) was used
400 to exclude dead cells. After staining, the antibodies were washed and the cells resuspended
401 in FACS buffer for sorting using an Aria Fusion (BD). The sorting strategy for isolating thymic
402 epithelial cells is shown in Fig. S1 A. Sorted cells were collected in TEC medium (see below)
403 containing 2 % FBS and 2.5 µM Thiazovivin (Stemgen, Catalog No. AMS.04-0017).

404

405

406 **Thymic epithelial organoid culture**

407 Sorted thymic epithelial cells were embedded in growth–factor-reduced Matrigel (Corning,
408 Catalog No. 356231) (~1.55 × 10^4 cells per 20 mL drop) and plated in 24-well plates (Corning,
409 Catalog No. 353047, or Ibidi, Catalog No. 82426). After Matrigel polymerization, TEC medium
410 was added. TEC medium consisted of organoid basal medium (Advanced DMEM/F-12
411 supplemented with 1× GlutaMAX [Thermo Fisher Scientific, Catalog No. 35050038], 10 mM
412 HEPES [Thermo Fisher Scientific, Catalog No. 15630056], 100 µg ml^{-1} Penicillin–Streptomycin
413 [Thermo Fisher Scientific, Catalog No. 15140122], 1× B-27 supplement [Thermo Fisher
414 Scientific, Catalog No. 17504001], 1× N2 supplement [Thermo Fisher Scientific, Catalog No.
415 17502001], 1 mM N-Acetylcysteine [Sigma-Aldrich, Catalog No. A9165]) plus 100 ng ml^{-1}
416 FGF7 (Peprotech, Catalog No. 100-19). 2.5µM Thiazovivin was also added to the medium for
417 the first two days. Medium was changed every second day. Organoids were cultured at 37°C
418 with 5% CO_2.

419

420 **Organoid proliferation assays**

421 Sorted thymic epithelial cells were embedded in 10 µl Matrigel drops (~7.5 x 10^3 cells/drop) in
422 a 48-well plate (Corning, Catalog No. 353078). On the day of seeding (day 0), at day 1, 3, 5
423 and 7, 220 µM resazurin (Sigma-Aldrich, Catalog No. R7017) was added to organoid basal
424 medium and incubated with the cells for 4 h at 37 ºC. Afterwards, the resazurin-containing
425 medium was collected and replaced by fresh TEC medium with or without FGF7. Organoid
426 proliferation was estimated by measuring the reduction of resazurin to fluorescent resorufin in
427 the medium using a Tecan Infinite F500 microplate reader (Tecan) with 560 nm excitation and
428 590 nm emission filters. For analysis, data were normalized from minimum to maximum.

429

430 **Bulk transcriptome profiling**

431 Sorted thymic epithelial cells were culture as indicated above. As controls, sorted thymic
432 epithelial cells from E16.5 embryos were either directly lysed in RLT buffer (QIAGEN, Catalog
433 No. 74004) containing 40 mM DTT (ITW Reagents, Catalog No. A2948) or cultured in 2D on
434 plates coated with 6 µg/ml laminin (R&D Systems, Catalog No. 3446-005-01). Cultures were
435 done in TEC medium. Organoids were collected in cold PBS to dissolve Matrigel and then
436 lysed in RLT buffer with DTT. They were collected after 3 and 7 days. Cells cultured in 2D were
437 directly collected in RLT buffer with DTT. They were collected once a confluent monolayer
438 formed, after 3 days, as prolonged culture in these conditions lead to cell detachment and
439 death. RNA was extracted using QIAGEN RNeasy Micro Kit (QIAGEN, Catalog No. 74004)
440 according to manufacturer's instructions. Purified RNA was quality checked using a

441 TapeStation 4200 (Agilent), and 88 ng were used for QuantSeq 3′ mRNA-Seq library
442 construction according to manufacturer's instructions (Lexogen, Catalog No. 015.96). Libraries
443 were quality checked using a Fragment Analyzer (Agilent) and were sequenced in a NextSeq
444 500 (Illumina) using NextSeq v2.5 chemistry with Illumina protocol #15048776. Reads were
445 aligned to the mouse genome (GRCm39) using star (version 2.7.0e). R (version 4.1.2) was
446 used to perform differential expression analyses. Count values were imported and processed
447 using edgeR (Robinson et al., 2010). Expression values were normalized using the trimmed
448 mean of M values (TMM) method and lowly-expressed genes (< 1 counts per million) and
449 genes present in less than three samples were filtered out. Differentially expressed genes were
450 identified using linear models (Limma-Voom) (Smyth et al., 2018), and P-values were adjusted
451 for multiple comparisons by applying the Benjamini-Hochberg correction method (Reiner et al.,
452 2003). Voom expression values were used for hierarchical clustering using the function hclust
453 (Murtagh, 1987) with default parameters, and for heatmap generation. Single sample gene set
454 enrichment analysis (GSEA) (Subramanian et al., 2005) was used to score the E2F targets
455 hallmark proliferation gene set (Howe et al., 2018; Liberzon et al., 2015) between samples.

456

457 **Whole-mount immunofluorescence staining**

458 Organoid samples were fixed in 4% paraformaldehyde (Thermo Fisher Scientific, Catalog No.
459 15434389) in PBS for 30 min at room temperature and subsequently washed with PBS.
460 Samples were permeabilized in 0.2 % Triton X-100 (Sigma-Aldrich, Catalog No. T8787), 0.3
461 M glycine (Invitrogen, Catalog No. 15527-013) in PBS for 30 min at room temperature and
462 blocked in 10 % serum (goat [Thermo Fisher Scientific, Catalog No. 16210064] or donkey
463 [Abcam, Catalog No. ab7475]), 0.01% Triton X100 and 0.3M glycine in PBS for 4h at room
464 temperature. Samples were then incubated with primary antibodies overnight at 4 °C, washed
465 with PBS, incubated with secondary antibodies overnight at 4 °C, and washed with PBS.
466 Mounting was done with Fluoromount-G (SouthernBiotech, Catalog No. 0100-01). The
467 following primary and secondary antibodies were used: MHCII-Biotin (BioLegend, Catalog No.
468 107603, 1/200), UEA1 (Vector Laboratories Catalog No. B-1065, 1/500), Keratin 5 (BioLegend,
469 Catalog No. 905501, 1/200), Keratin 8 (Abcam Catalog. No. ab53280, 1/200), Ki67 (BD
470 Pharmingen, Catalog No. 550609, 1/200), EpCAM-APC (Invitrogen, Catalog No. 17-5791-82,
471 1/200), Streptavidin Alexa 488 (Thermo Fisher Scientific, Catalog No. S-11223, 1/500),
472 Streptavidin Alexa 647 (Thermo Fisher Scientific, Catalog No. S-21374, 1/500), Goat anti-Rat
473 Alexa 647 (Thermo Fisher Scientific, Catalog No. A-21247, 1/500), Donkey anti-Mouse Alexa
474 568 (Thermo Fisher Scientific, Catalog No. A-10037, 1/500), Donkey anti-Rabbit Alexa 488
475 (Thermo Fisher Scientific, Catalog No. A-21206, 1/500), Donkey anti-Rabbit Alexa 568

476 (Thermo Fisher Scientific, Catalog No. A-11077, 1/500). Dapi (Tocris, Catalog No. 47481
477 mg/ml) was used to stain nuclei.

478 **Reaggregate culture**

479 E13.5 embryonic thymi were dissected and collected in Eppendorf tubes containing FACS
480 buffer. Lobes were rinsed with PBS and digested with 475 ul TrypLE and 25 µl DNase (from 1
481 mg/ml stock) for 5 min under agitation. Lobes were pipetted to help dissociation and TrypLE
482 was quenched with 1ml Adv. DMEM/F12 containing 10 % FBS. The cells were pelleted and
483 resuspended in FACS buffer for immunomagnetic cell separation with EpCAM-conjugated
484 beads (Miltenyi Biotec, Catalog No. 130-105-958, 1/4). After 20 min incubation at 4°C, the
485 unbound complexes were washed and the cells processed through magnetic columns (Miltenyi
486 Biotec, Catalog No. 130-042-401) following manufacturer instruction. The EpCAM-depleted
487 fraction was collected and used to prepare reaggregates with dissociated thymic epithelial
488 organoids and mouse embryonic fibroblasts (MEFs).

489 Thymic epithelial organoids at day 7 of culture were collected in cold Advanced DMEM/F-12
490 supplemented with 1× GlutaMAX, 10 mM HEPES and 100 µg ml^{-1} Penicillin–Streptomycin.
491 Organoids were pelleted and digested with 950 µl TrypLE and 50 µl DNase (from 1 mg/ml) for
492 5min at 37 °C. Organoids were pipetted to improve dissociation. In case digestion was
493 insufficient, organoids were further digested for 5 min with Trypsin + 0.25% EDTA (Gibco,
494 Catalog No. 25200-072) at 37 °C and pipetted until the obtention of a single cell suspension.
495 Dissociation was quenched with Adv. DMEM/F12 containing 10 % FBS and the cells pelleted.

496 Wild-type MEFs were a kind gift from the Blackburn laboratory. MEFs were cultured in
497 Advanced DMEM/F-12 supplemented with 1× GlutaMAX, 1x Non-Essential Amino Acids
498 (Gibco, Catalog No. 11140035), 100 µg ml^{-1} Penicillin–Streptomycin and 10 % FBS on gelatin-
499 coated dishes (0.1% gelatin in H_2O) (Sigma-Aldrich, Catalog No. G1890). MEFs were
500 harvested using Trypsin EDTA 0.25% for 2 min at 37 °C. Dissociation was quenched with Adv.
501 DMEM/F12 containing 10 % FBS and the cells pelleted.

502 For reaggregates using adult double negative 1 (DN1) thymocytes as input population, adult
503 thymi were dissected from 4 weeks old female C57BL/6J mice. Thymi were cut in small pieces
504 with a scalpel to liberate thymocytes, which were filtered to a single cell suspension with a 40
505 µm strainer. Cells were incubated with APC anti-mouse CD8a Antibody (BioLegend, Catalog
506 No. 100711, 1/50) for 20 min at 4°C in FACS buffer and washed. Cells were then incubated
507 with anti-APC magnetic beads (Miltenyi Biotec, Catalog No. 130-090-855, 1/4) for 20 min at
508 4°C. The unbound beads were washed away and the cells processed through magnetic
509 columns following manufacturer instruction. The APC depleted fraction was collected and used
510 for staining with the following antibodies: Ter119-FITC (BioLegend, Catalog No. 116205,

511 1/800), Cd45R-FITC (BioLegend, Catalog No. 103205, 1/800), CD11b-FITC (Thermo Fisher
512 Scientific, Catalog No. 11-0112-82, 1/800), Ly-6G-FITC (BioLegend, Catalog No. 108405,
513 1/800), Cd11C-FITC (BioLegend, Catalog No. 117306, 1/800), NK-1.1-FITC (BioLegend,
514 Catalog No. 108705, 1/800), CD3-FITC (BioLegend, Catalog No. 100306), CD4-FITC
515 (BioLegend, Catalog No. 100510), CD45-Pacific Blue (BioLegend, Catalog No. 103126, 1/200)
516 or CD45-AF700 (BioLegend, Catalog No. 103127, 1/160), CD44-PE (BioLegend, Catalog No.
517 103008, 1/160), CD25-BV711 (BioLegend, Catalog No. 102049, 1/160) and Dapi (Tocris,
518 Catalog No. 4748, 0.5 ug/ml). After staining, the antibodies were washed and the cells
519 resuspended in FACS buffer for sorting using an Aria Fusion (BD). The sorting strategy for
520 isolating DN1 thymocytes was gating on cells, single cells, live cells, CD45+ cells, CD44+
521 CD25- cells. DN1 thymocytes were collected in ORFTOC medium (see below).

522 Organoids reaggregate fetal thymic organ culture (ORFTOCs) were prepared as previously
523 described (Sheridan et al., 2009). Briefly, the cell suspension for each ORFTOC typically
524 contained 10^5 EpCAM-depleted cells, 10^5 thymic epithelial organoid cells, and 10^5 MEFs (or
525 10^5 thymic epithelial organoid cells, $4x10^4$ DN1 thymocytes and 10^5 MEFs). These cells were
526 transferred to an Eppendorf tube and pelleted. The pellet was resuspended in 60 µl of the
527 medium used for culture, and transferred to a tip sealed with parafilm inside a 15 ml Falcon
528 tube. Cells were pelleted inside the tip for 5 min at 470 rcf. The pellet was then gently extruded
529 on top of a filter membrane (Merck, Catalog No. ATTP01300) floating on culture medium in
530 24well plate. ORFTOC culture medium consisted of advanced DMEM/F-12 supplemented with
531 1× GlutaMAX, 1x Non-Essential Amino Acids, 100 µg ml^{-1} Penicillin–Streptomycin, 2 % FBS
532 and 100 ng/ml FGF7. 2.5µM Thiazovivin was added for the first two days of culture and half of
533 the medium volume was changed every second day.

534 Controls where one of the cell population is absent were made the same way. For FTOC
535 controls, E13.5 dissected lobes were directly placed on top of a filter membrane and also
536 cultured in ORFTOC medium.

537 All cultures were done at 37°C with 5% CO_2.

538

539 **Flow cytometry analysis of ORFTOCs and FTOCs**

540 After 6 and 13 days in culture, ORFTOCs, FTOCs, reaggregates with DN1 thymocytes and
541 controls reaggregates were gently detached from the filter membrane by pipetting and
542 transferred to Eppendorf tubes, together with the culture medium to collect recently emigrated
543 T cells. Samples were pelleted, rinsed with PBS and digested with 200 µl TrypLE for 10 min at
544 37° C with agitation on an Eppendorf shaker (800 rpm). Dissociation was quenched with 1ml
545 Adv. DMEM/F12 containing 10 % FBS and the cells pelleted. Cells were resuspended in FACS

546 buffer for staining. The cells were incubated for 20 min with the following antibodies: Ter119-
547 FITC (BioLegend, Catalog No. 116205, 1/800), Cd45R-FITC (BioLegend, Catalog No. 103205,
548 1/800), CD11b-FITC (Thermo Fisher Scientific, Catalog No. 11-0112-82, 1/800), Ly-6G-FITC
549 (BioLegend, Catalog No. 108405, 1/800), Cd11C-FITC (BioLegend, Catalog No. 117306,
550 1/800), NK-1.1-FITC (BioLegend, Catalog No. 108705, 1/800) together referred as Lineage,
551 CD44-PE (BioLegend, Catalog No. 103008, 1/160), CD69-APC (BioLegend, Catalog No.
552 104513, 1/160), CD4-BV605 (BioLegend, Catalog No. 100548, 1/40), CD3-PerCP/Cy5.5
553 (BioLegend, Catalog No. 100327, 1/160), CD8-PE/Cy7 (BioLegend, Catalog No. 100722,
554 1/160), CD25-BV711 (BioLegend, Catalog No. 102049, 1/160), CD45-AF700 (BioLegend,
555 Catalog No. 103127, 1/160), TCRβ-BV421 (BioLegend, Catalog No. 109230, 1/80) and Dapi
556 (Tocris, Catalog No. 4748, 0.5 ug/ml). After staining, the antibodies were washed and the cells
557 resuspended in FACS buffer for analyzing using a LSR Fortessa Cytometer (BD). The gating
558 strategy for analysis is shown in Fig. S2 B. Beads (UltraComp, Thermo Fisher Scientific
559 Catalog No. 01-3333-42) were used for single color staining for compensation. Gates were
560 based on T cells extracted from a young adult. Flow cytometry data were analyzed using
561 FlowJo (BD, version 10.9.0).

562

563 **Single-cell transcriptome profiling**

564 After 13 days in culture, ORFTOC and FTOC samples were collected and dissociated as
565 described for flow cytometry analysis. After dissociation, two ORFTOC samples and two FTOC
566 samples were pooled, respectively. For each pool, 500 000 cells were incubated with 1ul
567 TotalSeq Antibody (HTO) (BioLegend, Catalog No. 155863 and 155861) in 50 µl FACS buffer
568 for 30 min on ice. Antibodies were washed two times with FACS buffer and the single cell
569 suspensions filtered through a 40 µm strainer. After cell count, samples were mixed in a 1:1
570 ratio and processed using Chromium Next GEM Single Cell 5' Reagent Kits v2 (Dual Index)
571 with Feature Barcode technology for Cell Surface Protein & Immune Receptor Mapping
572 reagents (10X Genomics, Catalog No. PN-1000265, PN-1000256, PN-1000190, PN-1000287,
573 PN-1000215 and PN-100025) following manufacturer's instruction. Single Cell Mouse TCR
574 amplification Kit (10X Genomics Catalog No. 1000254) was used to prepare TCR libraries.
575 Sequencing was done using NovaSeq v1.5 STD (Illumina protocol #1000000106351 v03) for
576 around 100,000 reads per cell. The reads were aligned using Cell Ranger v6.1.2 to the mouse
577 genome (mm10). Raw count matrices were imported into R and analyzed using Seurat v4.2.0
578 (Hao et al., 2021). HTO with less than 100 features and less than 1 count were discarded.
579 Cells with less than 600 features, less than 0.4 or more than 10 percent mitochondrial genes
580 were discarded. Demultiplexing was performed using HTODemux with standard parameters.
581 Doublets were removed using recoverDoublets from scDblFinder package (Germain et al.,

582 2022) and based on doublets identified from HTOs. Data were normalized using SCTransform
583 and with cell cycle score as variable to regress. The three clusters representing the main cell
584 types were obtained using PCA and UMAP with 18 dimensions and a resolution of 0.005. Each
585 cell type was then subset and thresholded based on *EpCAM*, *Ptprc* and *Pdgfra* expression.
586 Epithelial clusters were identified using 18 dimensions and a resolution of 0.4, leading to 7
587 clusters that were named based on markers from previous datasets (Baran-Gale et al., 2020;
588 Bautista et al., 2021; Bornstein et al., 2018; Gao et al., 2022; Kernfeld et al., 2018; Park et al.,
589 2020). Immune clusters were identified using 18 dimensions and a resolution of 3. Immune
590 clusters were further merged to obtain 14 clusters representing main T cell developmental
591 stages based on markers from previous datasets (Cordes et al., 2022; Mingueneau et al.,
592 2013; Park et al., 2020; Rothenberg, 2021). The number of cells per clusters in both FTOC
593 and RFTOC samples were calculated to show HTO repartition between both samples. TCR
594 analysis was conducted using scRepertoire (Borcherding et al., 2020). Filtered contig output
595 from Cell Ranger was used as input and added to immune cells metadata. Productive cells
596 with both TRA and TRB chains were plotted on the UMAP, and percentage of productive cell
597 (either at least TRB chain with no NA and no double chain, or both TRA and TRB chains with
598 no NA and double chain accepted only for TRA) per cluster calculated. The mouse samples
599 from the dataset from Park et al. (Park et al., 2020) were used for alignment. H5ad files were
600 converted to Seurat object, TECs were subset from the stromal dataset and 4 weeks-old T
601 cells from the mouse total dataset. Alignment was performed using SCTransform and
602 canonical correlation analysis (CCA) with the Park dataset labeled as reference and otherwise
603 default parameters.

604

605 **Kidney capsule grafting and analysis**

606 ORFTOCs were grafted in CD45.1 host mice and FTOC controls in CD45.2 host mice. Mice
607 were treated with the analgesic Carprofen (10 mg/kg in drinking water) 12-24 h prior to
608 transplantation. Mice were anesthetized with Ketalar/Rompun (100 mg/kg Ketamin and 20
609 mg/kg Xylazin, intraperitoneal). Lacrinorm eye gel (Bausch & Lomb) was administered to avoid
610 dehydration of the cornea during the procedure. Anesthetized mice were shaved laterally and
611 disinfected using Betadine. The surgery was performed on a heating pad in order to minimize
612 body temperature drop. A small incision of approximately 1 cm was done first on the skin and
613 then in the peritoneum. By pulling at the posterior fat of the kidney with forceps, the kidney was
614 exposed outside of the peritoneum and kept wet with PBS. Under the microscope, an incision
615 and a channel were done with watchmaker-forceps on the kidney capsule's membrane and
616 one ORFTOC or FTOC was placed under the membrane. After positioning the kidney back
617 into the peritoneum, the wound was closed with two stitches (resorbable suture material 5/0;

618 Polyactin 910; RB-1 plus; Johnson&Johnson). The skin opening was closed with staples,
619 which were removed 7-10 days later. An analgesic (Temgesic, Buprenorphine 0.1 mg/kg,
620 subcutaneous) was administered at the end of the procedure followed by continuous treatment
621 of transplanted mice by Carprofen (10 mg/kg in drinking water) for 3 days. After the transplants,
622 mice were monitored daily and weighed every second day to confirm their wellbeing. Grafts
623 were analyzed 5 weeks after transplantation.
624 At the time of analysis, mice were sacrificed with CO_2 and kidneys retrieved. Grafts were
625 separated from the kidney under the microscope. To collect T cells, grafts were mechanically
626 dissociated by pipetting in FACS buffer. Single cell suspensions were then stained with Zombie
627 NIR (BioLegend, Catalog No. 423105, 1/1000) for 30 min at 4° C. Samples were then washed
628 with FACS buffer and incubated with the following Lineage antibodies for 30 min at 4°C: CD11b
629 Biotin (BioLegend, Catalog No. 101204, 1/1000), CD11c Biotin (BioLegend, Catalog No.
630 117304, 1/1000), CD19 Biotin (BioLegend, Catalog No. 101504, 1/1000), DX5 Biotin
631 (BioLegend, Catalog No. 108904, 1/1000), MHCII Biotin (BioLegend, Catalog No. 116404,
632 1/1000), GR1 Biotin (BioLegend, Catalog No. 108404, 1/1000), F4/80 Biotin (BioLegend,
633 Catalog No. 123100, 1/1000), Ter119 Biotin (BioLegend, Catalog No. 116204, 1/1000) and
634 NK-1.1 Biotin (BioLegend, Catalog No. 108704, 1/1000). After washes, samples were
635 incubated with the following antibodies for 30 min at 4°C : CD45.1-PerCP-Cy5.5 (BioLegend,
636 Catalog No. 110728, 1/500), CD45.2-BV650 (BioLegend, Catalog No. 109836, 1/200), CD4-
637 BUV563 (Thermo Fisher Scientific, Catalog No. 365-0042-82, 1/1000), CD8-BUV615 (Thermo
638 Fisher Scientific, Catalog No. 366-0081-82, 1/500), TCRαβ-PE-Dazzle594 (BioLegend,
639 Catalog No. 109220, 1/500), TCRγδ-PE (BioLegend, Catalog No. 118108, 1/500), CD69-FITC
640 (BioLegend, Catalog No. 104506, 1/500), CD24-APC (BioLegend, Catalog No. 101814,
641 1/1000), CD44-BV785 (BioLegend, Catalog No. 103059, 1/500), Ckit-BUV737 (Thermo Fisher
642 Scientific, Catalog No. 367-1171-82, 1/200), CD71-PE-Cy7 (BioLegend, Catalog No. 113812,
643 1/200), Sca1-BUV395 (Thermo Fisher Scientific, Catalog No. 363-5981-82, 1/500), CD25-
644 BV605 (BioLegend, Catalog No. 102036, 1/500), CD5-APC-eF780 (Thermo Fischer Scientific,
645 Catalog No. 47-0015-82, 1/500) and Streptavidin-BV510 (Biolegend, Catalog No. 405234,
646 1/500). After final washes, samples were resuspended in FACS buffer and analyzed on an
647 Aurora flow cytometer (Cytek Biosciences). Flow cytometry data were then analyzed using
648 FlowJo (version 10.9.0).

649

650 **Sectioning, immunofluorescence staining and RNA scope on sections**

651 Organoids, ORFTOCs, FTOCs and grafts were fixed in 4% paraformaldehyde in PBS for 30
652 min at room temperature (organoids) to overnight at 4 °C (ORFTOCs, FTOCs, grafts). Samples
653 were then washed with PBS and either processed for cryosectioning or for paraffin embedding.

654 For cryosectioning, samples were incubated in 30% (W/V) sucrose (Sigma-Aldrich, Catalog
655 No. S1888) in PBS until the sample sank. Subsequently, samples were incubated for 12 h in
656 a mixture of Cryomatrix (Epredia, Catalog No. 6769006) and 30% sucrose (Sigma-Aldrich,
657 Catalog No. 84097) (mixing ratio 50/50), followed by a 12 h incubation in pure Cryomatrix. The
658 samples were then embedded in a tissue mold, frozen on dry ice or in isopentane cooled by
659 surrounding liquid nitrogen. 10 µm-thick sections were cut at -20°C using a CM3050S cryostat
660 (Leica).
661 For paraffin embedding, organoid, ORFTOC and FTOC samples were
662 embedded in HistoGel (Thermo Fisher Scientific, Catalog No. HG-4000-012) before being
663 placed into histology cassettes. Cassettes were then processed with a Tissue-Tek VIP 6 AI
664 Vacuum Infiltration Processor (Sakura) and embedded in paraffin. 4 µm paraffin sections were
665 obtained with a Leica RM2265 microtome. Slides were processed through de-waxing and
666 antigen retrieval in citrate buffer at pH 6.0 using a heat-induced epitope retrieval PT module
667 (Thermo Fisher Scientific) before proceeding with immunostaining.
668 Sections were then blocked and permeabilized for 30 min in 1% BSA (Thermo Fischer
669 Scientific, Catalog No. 15260-037), 0.2% Triton X100 in PBS and blocked for 30 min in 10%
670 goat or donkey serum in PBS at room temperature. Primary antibodies were incubated O/N at
671 4 °C in PBS, 1.5% donkey or goat serum. On the following day, slices were washed twice in
672 1% BSA, 0.2% Triton X-100 in PBS and incubated with secondary antibodies at room
673 temperature for 45 min. Finally, slices were washed twice in 0.2% Triton X-100 in PBS and
674 mounted with Fluoromount-G. The following primary and secondary antibodies were used:
675 UEA1 (Vector Laboratories, Catalog No. B-1065, 1/500), Keratin 5 (BioLegend, Catalog No.
676 905501, 1/200), Keratin 8 (Abcam, Catalog. No. ab53280, 1/200), CD3ε (Thermo Fisher
677 Scientific, Catalog No. MA5-14524, 1/200), MHCII-Biotin (BioLegend, Catalog No. 107603,
678 1/200) EpCAM-PE (BioLegend, Catalog No. 118206, 1/200), Aire (Thermo Fischer Scientific,
679 Catalog No. 14-5934-82, 1/50), Streptavidin Alexa 488 (Thermo Fisher Scientific, Catalog No.
680 S-11223, 1/500), Goat anti-Rat Alexa 568 (Thermo Fisher Scientific, Catalog No. A-11077,
681 1/500), and Donkey anti-Rabbit Alexa 647 (Thermo Fisher Scientific, Catalog No. A-31573,
682 1/500). Nuclei were again stained with Dapi (Tocris, Catalog No. 4748, 1 ug/ml).
683 RNAscope Multiplex Fluorescent V2 assay (Bio-Techne, catalog no. 323110) was performed
684 according to the manufacturer's protocol. Paraffin sections were hybridized with the probes
685 Mm-Foxn1 (Bio-Techne, catalog no. 482021). Mm-3Plex probes (Bio-Techne, catalog no.
686 320881) and 3Plex Dapb probes (Bio-Techne, catalog no. 320871) were used as positive and
687 negative controls, respectively. Probes were incubated at 40°C for 2 hours, and the different
688 channels were revealed with TSA Opal570 (Akoya Biosciences, catalog no. FP1488001KT).
689 Tissues were counterstained with Dapi and mounted with ProLong Gold Antifade Mountant

690 (Thermo Fisher Scientific, P36930). Hematoxylin and eosin staining was performed using a
691 Ventana Discovery Ultra automated slide preparation system (Roche).

692

693 **Microscopy and image analysis**

694 Live brightfield imaging was performed using a Nikon Eclipse Ti2 inverted microscope with
695 4×/0.13 NA, 10×/0.30 NA, and 40×/0.3 NA air objectives and a DS-Qi2 camera (Nikon
696 Corporation). Time lapse was imaged with a Nikon Eclipse Ti inverted microscope system
697 equipped with a 20×/0.45 NA air objective and a DS-Qi2 camera (Nikon Corporation). Both
698 microscopes were controlled using the NIS-Elements AR software (Nikon Corporation).
699 Extended depth of field (EDF) of brightfield images was calculated using a built-in NIS-
700 Elements function. Fluorescent confocal imaging of fixed whole-mount and sections was done
701 on a Leica SP8 microscope system, equipped with a 20×/0.75 NA air and a 40x/1.25 glycerol
702 objectives, 405 nm, 488 nm, 552nm and 638 nm solid state lasers, DAPI, FITC, RHOD and Y5
703 filter cubes, a DFC 7000 GT (Black/White) camera and a CCD grayscale chip. Sections were
704 also imaged on a Leica DM5500 upright microscope equipped with a 20x/0.7 NA air and a
705 40x/1 NA oil objectives, a DFC 3000 (Black/White) or a DMC 2900 (Color) cameras and a CCD
706 grayscale or a CMOS color chip, respectively. Both Leica microscopes were controlled by the
707 Leica LAS-X software (Leica microsystems). For image processing, only standard contrast-
708 and intensity-level adjustments were performed, using Fiji/ImageJ (NIH) (version 2.1.0/1.53c).

709

710 **Statistics**

711 The number of replicates (*n*), the number of independent experiments or animals, the type of
712 statistical tests performed, and the statistical significance are indicated for each graph in the
713 figure legend. Statistical significance was analyzed using one- or two-way ANOVA, Brown-
714 Forsythe ANOVA in case of heteroscedasticity or Mood's median test in the absence of normal
715 distribution. For multiple comparisons, one-way ANOVA were followed by Tukey's test, Brown-
716 Forsythe ANOVA by Dunnet's T3 test, and Mood's test results adjusted for false-discovery
717 rate. Data normality and equality of variances were previously tested with Shapiro-Wilk and
718 Brown-Forsythe test, respectively. Grubbs test was used to determine the presence of outliers
719 across scRNAseq subpopulations. In all cases, values were considered significant when $P \leq$
720 0.05. Graphs show individual datapoints with mean ± standard deviation (SD). Tests were
721 performed using Prism (GraphPad, version 9.4.0), except Grubbs test which was performed
722 using GraphPad website (https://www.graphpad.com/quickcalcs/grubbs1/) and Mood's test
723 which was performed using the package rcompanion (Mangiafico, 2016) in R (version 4.1.2).
724 Graphs were made using Prism.

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Author contributions

TH conceived the study, designed and carried experiments, analyzed results, prepared artwork, and wrote the manuscript. LFLM helped with experimental design, experimental and analysis work and provided feedback on the manuscript. TB performed grafting experiment and analysis and provided feedback. LT and JJL helped with experiments. PR taught methods, provided feedback and shared reagents. CCB and GH provided feedback on the work and edited the manuscript. MPL conceived the work, designed experiments, and carried out the final editing of the manuscript.

Competing interests

The authors declare no competing interests.

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Data availability

Sequencing data reported in this paper have been deposited in the Gene Expression Omnibus (GEO) public repository under the accession number GSE240698. The SubSeries GSE240696 and GSE240697 corresponds to bulk and single-cell RNA-seq, respectively. Code used for analysis is available upon request and will be published on Github.

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