# Large gradient of susceptibility to esca disease revealed by long-term monitoring of 46 grapevine cultivars in a common garden vineyard

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## Abstract

Grapevine (*Vitis vinifera* L.) is prone to many fungal diseases, including esca, a severe vascular disease threatening the wine sector for which there is no cost-effective cure. Susceptibility to esca varies between cultivars in different infection conditions. It may therefore be possible to use the genetic diversity of grapevine cultivars to mitigate disease impact. However, the genetic component of esca susceptibility has rarely been investigated in the vineyard, and the specific mechanisms and varietal traits underlying esca susceptibility remain unknown.

In this study, we monitored the incidence and severity of esca foliar symptoms and plant dieback (apoplexy and mortality) at plant level for seven years, on 46 cultivars planted in an experimental common garden, to separate the genetic component of esca susceptibility from the effects of environment and cropping practices. We observed a broad gradient of varietal susceptibility, with a mean incidence of 0 to 26% of vines expressing esca foliar symptoms depending on the variety. This gradient remained similar across years and, unlike the severity of foliar symptoms, the incidence of grapevine dieback was significantly correlated with that of foliar symptoms. We detected a significant but weak and very localised phylogenetic signal for the incidence of esca foliar symptoms in this panel of cultivars.

We then explored the relationships between epidemiological metrics and ecophysiological and phenological traits phenotyped on the same plot. Esca disease incidence was negatively correlated with δ13C across cultivars, suggesting that varieties with higher water use efficiency are less prone to the expression of esca symptoms on leaves. Moreover, the least vigorous cultivars were among the least susceptible, although this relationship was not significant. By contrast, neither phenological stages nor nitrogen status were significantly predictive of cultivar susceptibility to the disease.

36     Together, these results provide new insight into the potential of genetic resources for use in the

37     sustainable management of grapevine trunk diseases and open up new perspectives for studying the

38     pathological and physiological determinants of their incidence.

40     **Keywords**: Ecophysiology, grapevine trunk disease, multi-trait phenotyping, pathogenicity,

41     phenotypic diversity, *Vitis vinifera*

## Introduction

The domesticated grapevine *Vitis vinifera* L. ssp. *vinifera* has a high level of genetic and phenotypic diversity, generated naturally by recombination, hybridisation, and mutation events, as well as through human-assisted selection. More than 5,000 cultivars are currently registered (This *et al.*, 2006), although only a small proportion are widely cultivated worldwide (Anderson and Aryal, 2013). Cultivar diversity is now considered an excellent tool for managing abiotic and biotic pressures in a sustainable manner (Merdinoglu *et al.*, 2018; Wolkovich *et al.*, 2018). There is therefore a need to improve our understanding of the variability between cultivars by phenotyping cultivars for a wide range of traits. Several stress-related traits, including carbon isotope discrimination in berry juice at maturity ($\delta 13C$; Plantevin *et al.*, 2022), cold hardiness (Ferguson *et al.*, 2014), and susceptibility to pests and diseases (Boso *et al.*, 2011; Gaforio *et al.*, 2011; Paňitrur-De La Fuente *et al.*, 2018), have already been studied in a range of cultivars. The susceptibility of the grapevine vascular system to various stresses has also recently been studied. Xylem and hydraulic traits have been phenotyped in a range of *Vitis* genotypes of different origins and levels of drought tolerance (Dayer *et al.*, 2022; Lamarque *et al.*, 2023) or resistance to vascular diseases (Pouzoulet *et al.*, 2020; Fanton and Brodersen, 2021).

Vascular biotic stresses include grapevine trunk diseases, which are responsible for substantial yield losses in vineyards worldwide (Gramaje *et al.*, 2018). One such disease is esca, which is detected in the vineyards through the summer expression of foliar symptoms and yield losses (Lecomte *et al.*, 2012). Esca pathogenesis has been associated with various types of trunk necrosis (Mugnai *et al.*, 1999) probably involving a complex community of fungal pathogens (Bruez *et al.*, 2014). This community includes a few key pathogens from both Ascomycota (including *Phaeomoniella chlamydospora* and *Phaeoacremonium minimum*) and Basidiomycota (e.g. *Fomitiporia mediterranea*). However, no direct role in foliar symptom onset has ever been demonstrated for any

of these pathogens, which are present in both healthy and necrotic wood tissues (Bruez *et al.*, 2016)

and in both symptomatic leaves and asymptomatic plants (Hofsteter *et al.*, 2012; Bruez *et al.*, 2014;

Bortolami *et al.*, 2019). These fungi probably also interact with bacteria (Bruez *et al.*, 2020) and seem

to be restricted to the perennial organs (Bortolami *et al.*, 2019).

Foliar symptoms are associated with xylem hydraulic failure and impaired photosynthesis (Bortolami

*et al.*, 2019, 2021a; Ouadi *et al.*, 2021; Dell'Acqua *et al.*, 2023). The incidence of esca foliar

symptoms is probably affected by multiple factors (as reviewed in Claverie *et al.*, 2020), including

plant age, pedoclimatic conditions, viticulture practices, the plant- and soil-associated microbiota,

and the plant material. The genetic diversity of the grapevine response to esca may be associated with

differences in phenotypic characteristics (ecophysiology, phenology) at cultivar level. Based on what

we now know about plant-to-plant variability within a single cultivar, mineral status, water status and

plant phenology are promising avenues to be explored. The incidence of esca increases following

foliar applications of nutrient sprays, which suggests that a higher nutrient content in the leaves may

be associated with a higher incidence of esca (Calzarano *et al.*, 2009). High levels of grapevine

transpiration may facilitate the translocation of pathogenic toxins and metabolites throughout the vine

(Bortolami *et al.*, 2021b). Differences in phenology can interfere with the co-ordination between

ontogenic susceptibility and favourable climatic periods for disease development (Serra *et al.*, 2018).

The susceptibility of grapevine cultivars to trunk diseases, especially esca, has been assessed with

three methods. The first involves phenotyping the internal necrotic lesions that develop after artificial

inoculation with one or more fungal species, such as *P. chlamydospora* (e.g. Pouzoulet *et al.*, 2017,

2020; Martínez-Diz *et al.*, 2019) and, to a lesser extent, *P. minimum* (Feliciano *et al.*, 2004; Gubler

*et al.*, 2004). However, the results obtained by this method are not readily transferable to field

conditions, particularly as fungal inoculations do not reproduce foliar symptoms (Claverie *et al.*,

2020). The second method is the monitoring of esca incidence (e.g. proportion of plants presenting

foliar symptoms or dieback) in a network of productive vineyards, over an entire region, or even at

the national scale (Bruez *et al.*, 2013). However, one of the major limitations of this approach is the

large number of confounding factors, such as viticulture practices, climatic conditions, soil type, plant

age, and rootstock. These biases can be overcome, making it possible to compare grapevine cultivars

within the same climatic and cultural context, by monitoring a single experimental vineyard planted

with a set of cultivars over multiple years. In the last decade, such experimental setups have been

used to compare numerous cultivars in common environments (e.g. Murolo and Romanazzi, 2014).

However, no experiment to date has been purposely designed to take into account pedoclimatic

microvariability and the origin of the plant material, both of which could account for a significant

amount of the variability in disease susceptibility (Kovács *et al.*, 2017; Gramaje *et al.*, 2018). As a

result, limited information is currently available regarding cultivar differences in terms of the

incidence of esca foliar symptoms and dieback in a common environment. Are the differences

between cultivars consistent between years? What are the ecophysiological drivers of esca

susceptibility at cultivar level?

In this study, we focused on the genetic component of grapevine susceptibility to esca foliar

symptoms and dieback independent of *terroir* effects, and the mechanisms potentially underlying

differences between cultivars. We monitored the incidence and severity of esca foliar symptoms and

plant dieback, and a range of phenological and ecophysiological traits in a common garden vineyard

planted with 46 cultivars, over periods of seven and six years, respectively. This design made it

possible to study cultivar-specific variability without bias due to other factors, such as year, soil, plant

material origin, and viticulture practices. We first investigated the range of cultivar susceptibility to

esca foliar symptoms and grapevine dieback over a period of seven years. We then explored multiple

correlations between different traits and years, to assess the temporal consistency of the ranking of

varieties for susceptibility. Finally, we investigated the relationships between esca foliar symptoms

and dieback on the one hand, and phenological and ecophysiological traits, such as key phenological stages, nitrogen status at flowering, pruning weight, and δ13C, a proxy of water use efficiency, on the other.

## Materials and methods

### 1. Common garden experimental vineyard

The VitAdapt vineyard (as described by Destrac-Irvine and van Leeuwen 2016) was designed as a common garden, and is located at the *Institut National de Recherche pour l'Agriculture, l'alimentation et l'Environnement* (INRAE) research station (Villenave d'Ornon, Nouvelle-Aquitaine, France), at 44°47'23.83 N'', 0°34'39.3' W'. This vineyard, on a sandy-gravel soil, was planted with 52 genotypes (47 *Vitis vinifera* L. cultivars and 5 *Vitis* hybrids). In this study, 45 *V. vinifera* L. cultivars, and one hybrid of several *Vitis* species — Hibernal, an F2 progeny of a Seibel 7053 x Riesling cross (Vitis International Variety Catalogue - VIVC; www.vivc.de) — were monitored. Among them, 28 were red-berried cultivars and 18 were white-berried cultivars (Supplementary Table S1). All plants were grafted onto Selection Oppenheim 4 (SO4) clone 761 rootstocks and planted at a density of 5,555 vines/ha, corresponding to a spacing of 1.8 m between adjacent rows and 1 m between adjacent vines. Eight cultivars were first planted in 2010, all the others being first planted in 2009 (Supplementary Table S1). Each cultivar was tested for major viral diseases before planting, and only non-contaminated material was planted. All vines were pruned according to the double guyot system and grown without irrigation. Pests and diseases were controlled by an integrated management programme, and weeds were controlled by mechanical tillage beneath the row. Cover crops were planted in every other row, alternating from year to year. The VitAdapt vineyard was organized into a five-randomised block design to account for soil variability. Each block

comprised one subplot per cultivar, in which 10 vines were planted in two adjacent rows. In this

study, only four blocks were monitored, resulting in a total of 184 subplots (1,840 vines; 40 per

cultivar, 46 cultivars) subjected to ecophysiological monitoring and esca and dieback monitoring for

six and seven years, respectively.

**2.  Monitoring of foliar symptoms of esca and plant dieback**

We monitored 1,840 vines individually by eye, to check for esca foliar symptoms (examples are

shown in Figure 1) and plant dieback (apoplexy and death), between 2017 and 2023. Evaluations

were performed one to three times per year, between early July and mid-September. For each

monitoring campaign, all the plants were inspected during a period of no more than one week. For

each vine, a score was assigned at individual arm level for the observed phenotype (asymptomatic,

presence of esca foliar symptoms/symptomatic, apoplectic, dead, young replanted vine, missing).

When foliar symptoms were observed, severity was scored (light, moderate, severe) by determining,

for each arm, the proportion of symptomatic shoots and the degree of leaf and berry dehydration (as

described in Supplementary Table S2). The score attributed ranged from 1 (light symptoms on a

single arm) to 5 (severe symptoms, i.e. 'tiger-striped' leaves and dried berries, on both arms). In years

with multiple assessments, the maximum severity index was retained.

Three epidemiological metrics were calculated from the data collected at subplot scale for esca foliar

expression and vine dieback. The incidence of esca foliar symptoms was calculated as the proportion

of the total number of productive plants (i.e. excluding totally apoplectic and dead plants, as well as

young replanted vines) displaying typical esca foliar symptoms. The cumulative incidence of foliar

symptoms was also calculated as the proportion of the productive vines expressing esca foliar

symptoms in at least one of the seven years monitored. The mean severity of esca foliar symptoms

was calculated at subplot scale, by averaging the score attributed to each symptomatic vine of the

subplot. Finally, the incidence of plant dieback was calculated as the proportion of mature plants (i.e.

excluding young replanted vines) with at least one unproductive arm (i.e. apoplectic or dead). During

the plant phenotyping process in the field, no *a priori* assumption was made concerning the cause of

plant dieback, which may have multiple abiotic or biotic causes.

**Figure 1. Examples of various foliar phenotypes for esca on different *Vitis vinifera* cultivars grown in a common garden experimental vineyard**

A-C: white-berried cultivars (A) Muscadelle; (B) Chasselas; (C) Viognier. D-F: red-berried cultivars (D) Alvarinho; (E) Pinot Noir; (F) Cabernet Franc. Note that the disease status of a vine is determined by phenotyping the entire shoot, not just individual leaves.

## 3. Phenotyping of ecophysiological and phenological traits

We explored the mechanisms underlying cultivar susceptibility to esca and dieback, using cultivar phenology data and monitoring data for three ecophysiological traits collected at the subplot scale in the VitAdapt vineyard.

### 3.1. Monitoring of phenological stages

The phenological evaluation was performed visually, in the field, on 10 vines of each cultivar in each of the four blocks, as described by Destrac-Irvine *et al.* (2019). Three different stages were monitored: bud break, flowering and veraison. Notations were based on the BBCH scale (Lorenz *et al.*, 1995). The bud-break stage corresponds to the date on which 50% of the buds have reached the BBCH 07 stage; the flowering stage corresponds to the date on which 50% of the flowers have reached the BBCH 65 stage; and the veraison stage corresponds to the date on which 50% of the berries have reached the BBCH 85 stage. Six years of data were included in this study, from 2017 to 2022.

### 3.2. Carbon isotope discrimination ($\delta^{13}C$)

We determined $\delta^{13}C$ levels in berry juice sugars obtained each year, at maturity, from 10 vines of each cultivar in each of the four blocks, as described by Plantevin *et al.* (2022). All samples were analysed in an external laboratory (UMR CNRS/Plateforme GISMO UMR 6282 BIOGEOSCIENCES *Université de Bourgogne*, 21000 Dijon, France). Briefly, the juice was extracted and analysed on a Vario Micro Cube elemental analyser coupled in continuous flow mode to an isotopic ratio mass spectrometer (IsoPrime, Elementar). Results are expressed according to the Vienna Pee Dee Belemnite (VPDB) international reference. Six years of data were included in this study, from 2017 to 2022.

### 3.3. N-tester measurements

N-tester measurements were performed at flowering (as defined above), which is considered to be a

key stage for nitrogen assimilation by grapevines (Celette and Gary, 2013), on 30 leaves of each

cultivar in each of the four blocks, according to the manufacturer's protocol (Yara, Oslo, Østland,

Norway). Briefly, each leaf was clamped with the device, for the measurement of transmittance (at

wavelengths of 650 and 960 nm), which provides a standardised index of chlorophyll content that can

be used as a proxy for vine nitrogen status. As a means of accounting for cultivar-specific nitrogen-

status behaviour under similar conditions, raw values were transformed according to varietal rounded

corrections. The mean value for Sauvignon Blanc was similar to the mean value across all varieties,

so this cultivar was used as the reference variety for this index. For each cultivar, the difference

between the mean value for all available years for the cultivar (2015-2021 for 35 cultivars; 2020-

2021 for the other 11), and the mean value for Sauvignon Blanc was calculated. An Ascendant

Hierarchical Classification (AHC) method was then applied to the N-tester values to group the

cultivars and obtain an average correction for all the cultivars of a single group. The N-tester value

of each cultivar was then corrected with the correction coefficient for its group, as described in

Supplementary Table S3. Five years of data were included in subsequent analyses, from 2017 to 2021.

### 3.4. Pruning weight quantification

Pruning weight was quantified during the winter for all varieties in each of the four blocks. For each

cultivar, two measurements were made for each block, by averaging the value three central vines of

a row (i.e. six vines per subplot). Only the wood from the previous growing season removed from the

plants during pruning was weighed. All measurements were performed during the same period of the

year in each year: between January 15th and 30th. Six years of data were included in this study, from

2017 to 2022.

## 4.  Data analysis

We first evaluated the effect of block on the three epidemiological metrics: esca foliar symptom incidence, esca foliar symptom severity, dieback incidence. Block was found to have a significant impact on the incidences of both foliar symptoms and dieback ($p = 0.03$ and $p = 10^{-5}$, respectively; Supplementary Figure S1). Block was included in subsequent models as a random effect to account for this variability.

We assessed the effects of cultivar and year (entered as fixed effects) on the three epidemiological metrics using three independent mixed modelling procedures, with block as a random effect. The effect of berry skin colour (white/red) on the three epidemiological metrics was independently modelled as a fixed factor, with cultivar and block as random effects.

We investigated the relationships between epidemiological metrics using mean values per cultivar over blocks and years. This made it possible to model the average inter-variety variation of traits independently of temporal and spatial variability. Correlations at cultivar level were assessed in Pearson's correlation tests (i) between epidemiological metrics, (ii) between epidemiological metrics for each single year, and (iii) between years for each single epidemiological metric. Independent modelling procedures were performed for pairs of variables found to be significantly correlated.

The mean values per cultivar over blocks and years were also used to test the relationships between quantitative ecophysiological and phenological traits on the one hand, and the three epidemiological metrics on the other. Correlations between all pairs consisting of one epidemiological and one phenotypic variable were tested in Pearson's correlation tests. Independent modelling was performed for all pairs of variables found to be significantly correlated, with the epidemiological metric modelled as the response variable, and the ecophysiological or phenological traits as fixed effects.

For all these analyses, linear models (LM) or linear mixed models (LMM) were used for response variables following a normal distribution (i.e. foliar symptom severity). For binomial response

variables (i.e. the incidences of foliar symptoms and dieback), generalised linear models (GLM and

GLMM) were fitted, using the binomial family with "cloglog" links (accounting for a non-symmetric

distribution). Each model was graphically validated according to the normality of the residuals (QQ-

plot) and the homogeneity of the residual variance (residuals vs. fitted, residuals vs. predictors). All

analyses involving epidemiological metrics were conducted for the period 2017-2023, whereas

analyses including other phenotypic traits were conducted for the period 2017-2022. Analyses were

performed with R v.4.2.1 software (R Core Team, 2022) and the RStudio interface. Linear modelling

was performed with the "lme4" package (Bates *et al.*, 2015), and model validation was performed

with "DHARMa" (Hartig and Lohse, 2022). Correlation analyses were performed with the "corrplot"

package (Wei, Simko, 2021).

**5. Phylogenetic signal based on epidemiological metrics**

We built a phylogenetic classification of the 46 cultivars included in this study based on grapevine

genotyping data for 20 microsatellite markers (SSR), as described by Laucou *et al.* (2011). We used

the genetic distances between cultivars obtained from different hierarchical clustering analyses: two

classical methods for statistical clustering i.e. Euclidean and Ward) and two methods adapted for

phylogenies (i.e. unweighted pair group method with arithmetic mean (UPGMA) and neighbour

joining (NJ)). For phylogenetic signal analysis, we visually inspected the four phylogenies and

selected the phylogeny best reflecting the known relationships between cultivars. All information on

kinship between the cultivars used in this work was based on the VIVC (www.vivc.de).

We searched for a phylogenetic signal in cultivar susceptibility to esca using methods based on

autocorrelations computed both on the global phylogenetic tree and local nodes of the phylogeny

implemented in the "phylosignal" package (Keck *et al.*, 2016). A global Moran's I index was

calculated for each epidemiological metric and tested against the null hypothesis of an absence of

phylogenetic signal. Phylogenetic correlograms were then constructed to visualise the distribution of

this index over a gradient of phylogenetic distances and its confidence envelope based on

bootstrapping (1,000 repetitions). The local Moran's index $I_i$ was also plotted alongside the

phylogeny to locate autocorrelation patterns more precisely through LIPA analysis. The existence of

local phylogenetic signals was assessed in a permutation-based test (999 repetitions).

## Results

**Global incidences of esca and dieback and changes over time**

The mean incidence of esca foliar symptoms, for all cultivars, blocks, and years, was 7.5% (standard

deviation, SD = 13.6%), with considerable variability (relative standard deviation, RSD = 182.2%;

Table 1). The mean cumulative incidence of esca foliar symptoms (i.e. observed in at least one year)

was 30.5%, implying that almost one vine in three expressed esca foliar symptoms in at least one

year. This value is five times higher than the mean annual incidence (Table 1). Foliar symptom

severity had a mean score of 2.5 and was much less variable than the incidence of foliar symptoms

(RSD = 59.4%; Table 1). Plant dieback had a mean incidence of 4.1%, which is lower than that of

foliar symptom incidence by a factor of 1.8, but was much more variable (RSD = 233.0%; Table 1).

An analysis of the declining vines showed that 16% expressed apoplexy on a single arm, 51% had a

single dead arm, and 33% were totally unproductive (i.e. 11% totally apoplectic, 18% totally dead,

and 4% with both phenotypes).

**Table 1. Overall statistics for esca and dieback epidemiological metrics in the VitAdapt common garden experimental vineyard (Villenave d'Ornon, France) between 2017 and 2023**

The statistics given are means, standard deviation, relative standard deviation, minimum and maximum. Values are calculated for 1288 observations (46 cultivars; four blocks; seven years). The severity index ranged from 1 (light esca foliar symptom on a single arm) to 5 (severe esca foliar symptom on both arms) and was scored only for plants with symptoms.

| Trait | Statistical individual | mean | SD | CV | min | max |
|---|---|---|---|---|---|---|
| Foliar symptom incidence | Observation ($n$=1288) | 7.3% | 13.6% | 185.3% | 0.0% | 90% |
| Cumulative incidence over seven years | Cultivar ($n$=46) | 30.5% | 24.0% | 74.4% | 0.0% | 78.5% |
| Foliar symptom severity index | Observation ($n$=858) | 2.5 | 1.5 | 59.4% | 1.0 | 5.0 |
| Dieback incidence | Observation ($n$=1288) | 4.1% | 9.6% | 233.0% | 0.0% | 71.4% |

The incidences of esca and dieback increased significantly between 2017 and 2023, with the ageing of the plot (from 7.8 to 13.8 years old on average). This temporal effect was highly significant for the incidence of foliar symptoms ($p < 10^{-16}$). The incidences of esca and dieback in 2021 and 2022 were significantly higher than those in the previous four years, with mean values almost six times higher than those obtained in 2017. Finally, the highest incidences occurred in 2023, when the mean incidence of foliar symptoms was more than seven times higher than that in 2017 (Figure 2A). A similar pattern was observed for the incidence of dieback ($p < 10^{-16}$). Plant dieback rates were very low during the first four years of monitoring, between 0% and 1%. The proportion of unproductive plants was greatest in 2022 and 2023, when it was significantly higher than in all other years (Figure

2C). The severity of esca foliar symptoms also differed significantly between years ($p = 10^{-11}$).

However, severity did not follow a specific temporal pattern similar to that for incidence (Figure 2B).

**Figure 2. Changes over time in the three variables related to esca foliar symptoms and dieback**

**for all cultivars from 2017 to 2023 (mean plant age increasing from 7.8 years in 2017 to 13.8**

**years in 2023)**

(A) Incidence of foliar symptoms of esca (mean ± SEM); (B) Severity of esca foliar symptoms

(mean ± SEM), i.e. mean severity index (according to the rating scale described in Supplementary

Table S2) calculated on the set of symptomatic plants; (C) Incidence of plant dieback (mean ± SEM).

The letters correspond to the groups of significance according to Tukey tests with an alpha risk of

5%.

**Esca incidence differs considerably between cultivars**

The incidence of esca foliar symptoms differed significantly between the 46 cultivars monitored from

2017 to 2023 ($p < 10^{-16}$). We found a large gradient of susceptibility to esca foliar symptoms, as four

grape varieties (Merlot, Petit Manseng, Tannat and Xinomavro) never expressed symptoms during

the seven years of monitoring whereas eight cultivars (Chenin, Saperavi, Mourvèdre, Cabernet Franc,

Sauvignon, Tempranillo, Castets and Cabernet-Sauvignon) had mean annual incidences above 20%

(Figure 3A).

Excluding the four cultivars with an overall incidence of zero, cumulative incidence ranged from

2.5% to 85%, and was 2.7 to 7 times higher than the mean incidence according to the cultivar (Figure

3A). The cumulative incidence of esca was highly correlated with the incidence of foliar symptoms

($r = 0.95$; $p < 10^{-16}$).

The severity of esca foliar symptoms differed significantly between cultivars ($p = 0.04$) but the

variability was lower than that for incidence (Figure 3B). Each cultivar presenting esca foliar

symptoms could display any degree of disease severity, regardless of its mean incidence of foliar

symptoms.

The incidence of plant dieback also differed significantly between cultivars ($p < 10^{-16}$). Four cultivars

(Muscadelle, Petit Manseng, Prunelard, Petit Verdot) had no apoplectic or dead arms. The maximum

incidence, exceeding 20%, was recorded for Saperavi. The distribution of dieback phenotypes

differed between cultivars. For example, Castets was characterised by a higher incidence of apoplectic

arms than of dead arms, whereas the opposite pattern was observed for Chasselas (Figure 3C).

Berry colour had no significant effect on the incidence of either esca foliar symptoms or plant dieback

($p = 0.95$ and $p = 1$, respectively; Figure 3A and 3C).

**Figure 3. Variability of susceptibility to esca foliar symptoms and dieback among grapevine cultivars for the period 2017 to 2023.**

(A) Mean incidence of esca foliar symptoms for the 46 cultivars monitored. Bars and error bars represent the means ± SEM and the grey triangles indicate the cumulative incidence over the seven years; (B) Range of severity of esca foliar symptoms in symptomatic plants only (mean ± SEM). The severity index, which ranged from 0 to 5, is described in Supplementary Material 2; (C) Range for the incidence of plant dieback. Light grey corresponds to plants with one apoplectic arm (total dehydration of the canopy). Red corresponds to plants with one dead arm. Dark grey corresponds to totally unproductive plants (apoplexy or death on both arms). Grapevine cultivars are ordered according to the mean incidence of foliar symptoms presented in (A). White-berried cultivars are shown in light yellow and red-berried cultivars are shown in dark purple.

Finally, the incidences of esca foliar symptoms and dieback were strongly correlated for each cultivar

(r = 0.72; $p = 10^{-8}$; Supplementary Figure S2A). This relationship was strongest in years with an

incidence of dieback of more than 4% (from 2021 to 2023), whereas it was marginal or non-

significant in the preceding years (Figure 4). Despite this significant correlation, we were able to

identify several cultivars with a greater expression of foliar symptoms than of dieback phenotypes,

including Cabernet-Sauvignon, Sauvignon, Cabernet Franc and Morrastel. Several cultivars,

including Castets, Marselan and Tinto Cao, displayed the opposite behaviour (Figure 3A and C).

By contrast, foliar symptom severity was not significantly correlated with either symptom incidence

(r = 0.22, $p = 0.14$; Supplementary Figure S2B) or dieback incidence (r = 0.26, $p = 0.08$;

Supplementary Figure S2C).

**Figure 4. Relationship between the incidence of esca foliar symptoms and the incidence of dieback**

Each dot corresponds to the value for a cultivar monitored during a single year. The regression line corresponds to a binomial model with a cloglog link fitted for all years (estimate = 6.11; $p < 10^{-16}$). Dots are coloured according to the year.

**The relationships between varietal patterns of susceptibility and genetic distances between cultivars are of borderline significance**

Ward's genetic distances were calculated to test for the existence of a phylogenetic signal in the epidemiological metrics. The clustering based on these distances was highly consistent with known cultivar lineages. For esca foliar symptom incidence, a significant phylogenetic signal was highlighted at global level (I = -0.10; $p = 0.03$). For this trait, phylogenetic autocorrelation values were positive for short phylogenetic distances (Figure 5A). However, this signal was not very robust,

had an extremely wide confidence envelope, and was significant only for very short phylogenetic

distances (Figure 5A). The distribution of the local Moran's index $I_i$ over the phylogeny revealed two

hotspots of phylogenetic positive autocorrelation (i.e. nodes at which the local Moran's index value

$I_i$ was significant) (Figure 5B): (i) a node of six cultivars with low incidences (Prunelard, Cot, Tannat,

Petit Verdot, Petite Arvine, and Cornalin); (ii) a node of cultivars with high incidences, three of which

displayed a significant signal (Sauvignon, Morrastel, Carmenere). By contrast, this method revealed

no phylogenetic autocorrelation for either foliar symptom severity ($I = -0.02$; $p = 0.52$) or the

incidence of dieback ($I = -0.02$, $p = 0.30$).

**Figure 5. Relationship between genetic distance and the incidence of esca foliar symptoms for**

**a panel of 46 grapevine cultivars**

(A) Phylocorrelogram showing the change in Moran's I index (measuring phylogenetic

autocorrelation) along a gradient of phylogenetic distances. Black lines represent Moran's I value.

Dashed lines correspond to a confidence envelope at 95%, estimated from 1,000 bootstrap replicates.

(B) Detection of the phylogenetic signal at a local scale over the phylogeny, by LIPA analysis. The

dendrogram displaying genetic distances between cultivars is based on a set of 20 SSR markers. Bars

correspond to standardised values (i.e. centred and scaled) for the mean incidence of esca foliar

symptoms. Red bars correspond to tips for which the local Moran's index $I_i$ was significant (i.e.

$p < 0.05$ in a test based on permutations).

**Varietal patterns of esca incidence remain constant over time**

We investigated whether the incidence and severity of esca and plant dieback remained constant over

time by performing two-by-two correlation analyses at cultivar level between the three

epidemiological metrics, using the mean values for each of the six years of monitoring (a positive

correlation indicates similar varietal patterns of incidence). For the incidence of foliar symptoms, all

but one of the year-by-year correlations were clearly positive and significant, the exception being

2017 vs. 2022 (Figure 6A). For dieback incidence, all correlations were positive and all but five were

significant (Figure 6C). By contrast, no clear inter-annual correlation pattern was identified for foliar

symptom severity. Only three pairs of years displayed significant correlations, negative in one case

(2018 vs. 2019) and positive in the other two (2017 vs. 2022 and 2021 vs. 2023) (Figure 6B).

**Figure 6. Inter-annual correlations of mean varietal values for the three variables related to esca foliar symptoms and dieback**

(A) Foliar symptom incidence; (B) Severity of esca foliar symptoms, i.e. mean severity index (according to the rating scale described in Supplementary Material 2) calculated for the set of symptomatic plants; (C) Plant dieback incidence. Pearson's correlation coefficients marked with asterisks are significant at the 5% level; *: $p < 0.05$; **: $p < 0.01$; ***: $p < 0.001$.

**A low incidence of esca is associated with high water use efficiency and low vigour at cultivar scale**

The relationships between the three epidemiological metrics and cultivar characteristics were explored by measuring six phenological and ecophysiological traits in the same vineyard: bud burst, flowering and veraison dates, $\delta^{13}C$, N-tester values at flowering, and pruning weight. The variability of these traits is presented in Supplementary Table S4. Globally, the range of variability for these traits was narrow, with RSD values ranging from 3.6% (for veraison date) to 30.2% (for pruning weight). Flowering date was positively correlated with both bud burst and veraison dates (r = 0.6 and r = 0.5, respectively; Supplementary Figure S3); we therefore retained flowering date as the only phenological variable.

Correlations between the mean values of the three epidemiological metrics and these four other

phenotypic traits were tested at cultivar level for the period 2017-2022 (Figure 7). $\delta^{13}C$ value at

harvest was significantly negatively correlated with foliar symptom expression, albeit weakly (r = -

0.33; $p$ = 0.03; Figure 7A). Similarly, this trait was significantly negatively correlated with dieback

incidence (r = -0.32; $p$ = 0.03; Figure 7I). In other words, cultivars with high water- use efficiency

(i.e. a less negative $\delta^{13}C$) were less susceptible to the expression of esca foliar symptoms and dieback

phenotypes. However, when this relationship was considered for each year separately, the correlation

between $\delta^{13}C$ and the incidence of foliar symptoms was significant only for 2022 (Supplementary

Figure S4).

The relationship between the incidence of foliar symptoms and cultivar pruning weight was positive,

but not significant (r = 0.22; $p$ = 0.14; Figure 7B). Despite the weak nature of this relationship, none

of the least vigorous cultivars had a high incidence of foliar symptoms.

There was a moderate negative correlation between these two ecophysiological traits ($\delta^{13}C$ and

pruning weight) at cultivar level (r = -0.49; $p$ = $10^{-5}$; Supplementary Figure S3).

N-tester values at flowering and cultivar phenology (flowering date) were not significantly correlated

with any of the three epidemiological metrics (Figure 7). All 12 correlations presented in Figure 7

were also tested at the subplot scale and similar trends were observed (*data not shown*).

**Figure 7. Relationships between three variables related to esca foliar symptom and dieback and four ecophysiological and phenological traits**

Each dot corresponds to the value for a specific cultivar averaged over all years and blocks for the period 2017-2022. Pearson's correlation coefficients are indicated for each relationship. The regression line is provided for significant correlations only and corresponds to a binomial model with a cloglog link as follows: (A) estimate = -0.61; $p < 10^{-16}$; (E) estimate = -0.63; $p = 10^{-11}$. White-berried cultivars are shown in light yellow and red-berried cultivars are shown in dark purple.

## Discussion

In this study, we focused on esca, a fungal vascular disease of grapevine of growing concern. We used a unique common garden vineyard to assess inter-variety variability in the expression of esca disease. Our results indicate diverse patterns of susceptibility among cultivars, with some cultivars displaying no foliar symptoms in any of the seven years studied and others highly susceptible. We also identified key ecophysiological traits correlated with susceptibility to esca disease, such as cultivar water use efficiency (WUE) and vigour.

The results obtained with VitAdapt indicate that there is a high level of fairly stable cultivar-associated variability in terms of esca expression. The long-term monitoring of the VitAdapt common garden revealed a significant variability of esca foliar symptoms and plant dieback between the 46 grapevine cultivars. Several cultivars only very rarely expressed foliar symptoms or dieback phenotypes, whereas others, including several of high economic importance, were much more strongly affected by esca and dieback. The use of a common garden design and the inclusion of a block effect in statistical models made it possible to prevent possible biases in the calculation of esca incidence due to the variability of soils and viticulture practices. Our results provide new information for consideration in the discussions of cultivar rankings for esca susceptibility obtained in previous multiregional and multicultivar monitoring programmes. For example, Ugni Blanc, for which very high incidences were recorded in the Charentes region (France; Bruez *et al.*, 2013), did not cluster among the most susceptible cultivars in our vineyard, in which all cultivars were subject to the same environment and the same viticulture practices (e.g. double guyot pruning system). Bruez *et al.* (2013) showed that, for a given cultivar, a large proportion of the residual variability is due to the growing area. There are at least two possible explanations for this. First, there can be considerable climatic variation between regions. Second, the incidence of esca disease may be affected by differences in viticulture systems and practices from between regions (Lecomte *et al.*, 2018). This may be the case

for Ugni Blanc, which was analysed here in growth conditions different from those applying to productive contexts in the Charentes region (France), where it is cultivated for the production of Cognac.

Our findings clearly confirmed a number of well-known trends, such as the large differences between elite international cultivars, with Merlot (no foliar symptoms at all from eight to 14 years of age), Syrah and Pinot Noir displaying high levels of resistance to esca, whereas Cabernet-Sauvignon, Sauvignon Blanc, and Tempranillo were highly susceptible (consistent with the findings of Martínez-Diz *et al.*, 2019; Csótó *et al.*, 2023). However, they also provide integrative ecophysiological and esca susceptibility data for lesser-known varieties with promising tolerance to abiotic and biotic stresses. For example, Xinomavro, which never expressed esca in our vineyard, was classified as drought-tolerant by Plantevin *et al.* (2022) and is one of the latest ripening variety in VitAdapt (i.e. it is comparable to Grenache and Carignan, *data not shown*). Conversely, Saperavi was found to be highly prone to the expression of esca and dieback (apoplexy and death), and was also classified as susceptible to drought (Plantevin *et al.*, 2022; Lamarque *et al.*, 2023). More detailed studies are now required to determine whether these preliminary findings are also valid for other commercial clones of these cultivars, and for combinations with different rootstocks, in contrasting productive contexts. More generally, the extent to which esca expression varies between different clones of a single cultivar remains unclear (Murolo and Romanazzi, 2014; Moret *et al.*, 2019). Zooming out to consider the entire pool of *Vitis* spp. might also be promising, to move towards the integration of resistance to trunk diseases as a key trait while selecting and planting both rootstocks (Gramaje *et al.*, 2010; Murolo and Romanazzi, 2014) and hybrid cultivars. Here, the only hybrid variety studied, Hibernal, was found to be of intermediate susceptibility. Comparing 104 interspecific hybrids of diversified pedigrees with 201 *V. vinifera* cultivars, Csótó *et al.* (2023) demonstrated that the interspecific hybrids were globally more tolerant to trunk diseases.

A global increase in esca expression was recorded over the course of the trial, for both foliar

symptoms and plant dieback, probably due to the ageing of the plants (from 8 years old at the start of

monitoring to 14 years old). The incidence of esca disease is known to increase when the plants are

about 10 years of age, generally reaching a maximum in plants between the ages of 15 and 25 years

(Mugnai *et al.*, 1999; Kovács *et al.*, 2017). Moreover, our data indicate that the cumulative incidence

of esca foliar symptoms was much higher than the mean annual incidence, implying that, for a given

subplot, the vines presenting foliar symptoms of esca differ between years. These results clearly

indicate that it is essential to perform monitoring over several years for field trials (Reis *et al.*, 2019).

Even though the incidence of esca increased over time, the relative ranking of the varieties remained

constant between years. As all cultivars were compared in similar environmental conditions, this

suggests that each cultivar has a constitutive level of resistance that is little affected by local variations

of esca disease pressure (which are, in turn, influenced by several traits relating to pathogens, plants

and environment; Claverie *et al.*, 2020). A first genetic locus associated with grapevine susceptibility

to trunk diseases was recently identified in *V. vinifera* cv. 'Gewurztraminer', based on internal

symptoms (proportions of total necrosis and white rot) rather than the monitoring of foliar symptoms

and dieback (Arnold *et al.*, 2023). However, anatomical and physiological traits may contribute to

cultivar susceptibility. A high density of large vessels (>100 µm) has been associated with enhanced

susceptibility following artificial inoculation (Pouzoulet *et al.*, 2020). Based on the results of studies

on very small numbers of cultivars, we can also hypothesise that wood composition (see Rolshausen

*et al.*, 2008 for *Eutypa lata*, another fungal pathogen of wood) and secondary metabolism (see

Lemaitre-Guillier *et al.*, 2020 for *Botryosphaeria* sp. diseases) may have an effect. Another non-

mutually exclusive hypothesis would be the existence of interspecific and intraspecific diversity in

the microbial communities interacting with different cultivars during plant pathogenesis (Laveau *et*

*al.*, 2009; Bekris *et al.*, 2021).

As the genetic proximity between plant species and populations is often associated with common

susceptibilities to pathogens and pests (Gilbert *et al.*, 2015), we tested the hypothesis of closely

related cultivars having similar patterns of susceptibility to esca. We found a significant but weak

phylogenetic signal for esca susceptibility based on the incidence of foliar symptoms. This signal was

associated exclusively with local nodes of closely related cultivars and was similar for all four

clustering methods tested *(data not shown)*. This signal was not as robust as that likely to arise at

broader levels of grouping and may be strongly affected by the choice of genetic markers. Based on

a local-scale analysis, the main hotspots of phylogenetic signal corresponded to a group of weakly

susceptible cultivars. Pedigrees were available for only two of these six cultivars, Cot being an

offspring of Prunelard in the VIVC. The relationships between the remaining varieties were less clear,

and they even came from contrasting locations (Bacilieri *et al.*, 2013). We can hypothesise that traits

reducing susceptibility to esca in grapevine cultivars are converging within this group of interest.

Susceptible cultivars were overrepresented at another node of the phylogeny: Sauvignon, Chenin,

Marselan, Mourvedre, Morrastel, Cabernet-Sauvignon, Castets, Cabernet Franc, Carmenere. All

these cultivars originate from Western and Central Europe, with six originating from South-Western

France (Bacilieri *et al.*, 2013). Most belong to the Savagnin and Cabernet Franc families.

Nevertheless, the signal on these branches appeared unstable (i.e. significant for only three cultivars).

Moreover, phylogenetic correlation did not apply to all members of a group or lineage, as represented

in the vineyard studied. Thus, despite promising findings, it is difficult to conclude that esca

susceptibility is clearly driven by phylogenetic patterns. Here, the genetic panel studied consisted of

cultivars belonging from a single species, *V. vinifera* L. (with the exception of the *Vitis* sp. hybrid

'Hibernal') and was not truly representative of the overall diversity of grapevine. The search for

genetic markers of trunk disease incidence would probably benefit from studies of specifically

designed diversity panels (e.g. Nicolas *et al.*, 2016).

Interestingly, the incidences of foliar symptoms and dieback were strongly and consistently correlated

between cultivars and years. This suggests that the cultivars with a high incidence of esca were also

those with a high proportion of apoplectic or dead plants. This finding echoes the work of Guérin-

Dubrana *et al.* (2013) in the Bordeaux region, which showed that, in Cabernet-Sauvignon, mortality

was higher in plants that had expressed esca symptoms in previous years. Nevertheless, other studies

have shown that this correlation is not always valid at plant level in individual cultivars. For instance,

Andreini *et al.* (2014) noted a discordance between esca symptom expression and plant mortality,

especially for Cabernet-Sauvignon. Dewasme *et al.* (2022) suggested that the impact of esca on the

mortality of this cultivar might be overestimated. Investigations of the mechanisms underlying foliar

symptoms and plant decline, especially at vascular level, would be a promising approach to clarifying

this issue.

Conversely, a lack of correlation was noted between the metrics reflecting disease incidence and those

reflecting the severity of foliar symptoms. There was also no clear varietal or temporal pattern for

foliar symptom severity. We hypothesise that a number of different mechanisms and factors underlie

these differences in severity levels. The link between incidence and severity has long been called into

question in plant science. These two traits are known to be positively correlated for a large number

of airborne fungal diseases (Seem, 1984). Nevertheless, no such relationship would be expected for

wilt diseases with symptoms occurring in organs (here, in leaves) distal to the infection area (here,

woody organs; Seem, 1984). Few studies have specifically tested this hypothesis for trunk disease

pathosystems, although several previous studies concluded that there was a positive correlation

between incidence and severity in a single (Calzarano *et al.*, 2018) or multiple cultivars (Romanazzi

*et al.*, 2009).

There have been many studies of the impact of esca disease on grapevine physiology, including plant

growth (Gramaje *et al.*, 2010; Dell'Acqua *et al.*, 2023), photosynthesis, water relations (Magnin-

Robert *et al.*, 2011; Bortolami *et al.*, 2021a,b), and phenology (Andreini *et al.*, 2013), but very little

effort has, as yet, been devoted to the opposite issue: the ecophysiological traits predicting esca

incidence or severity. Here, we found that a low incidence of esca appeared to be associated with high

water use efficiency and low vigour at the cultivar scale. At this same scale, $\delta^{13}C$ levels were

significantly negatively related to the incidences of both foliar symptoms and dieback. This indicator,

averaged over several years of contrasting water availability, is used as a proxy for varietal WUE

(Bchir *et al.*, 2016). Cultivars with higher WUE, associated with lower levels of stomatal opening,

were found to be less prone to the expression of esca symptoms. The underlying mechanism may be

at least partly related to that proposed by Bortolami *et al.* (2021b) to elucidate the antagonistic effects

of drought and esca. Genotypes with a high WUE control their transpiration much more strongly,

thereby slowing the translocation of toxins from the trunk to the leaves via the vascular apparatus. Di

Marco and Osti (2008) drew similar conclusions for wood decay in kiwifruit (*Actinidia deliciosa* var.

deliciosa). However, in our case, this relationship was found to be unstable over time. The strongest

correlation was observed in 2022, a year combining high esca intensity and low water availability.

Indeed, it is known that the differences in WUE between varieties can be exacerbated by drought

conditions (Plantevin *et al.*, 2022), particularly when different plant genotypes are planted in the same

vineyard.

Water availability is considered a driver of esca pathogenicity. High water levels are considered to

activate esca expression (Bortolami *et al.*, 2021b; Monod *et al.*, 2023). In this study, the overall

increase in esca incidence over time (with the ageing of the vines) made it difficult to decipher

precisely the role of the water conditions in each year. Our findings suggest that, in our experimental

context, plant age makes a much greater contribution to esca incidence than plant water use.

The least vigorous cultivars appear to be grouped among the cultivars least susceptible to esca. This

relationship has already been suggested for trunk diseases, but never clearly demonstrated. It may

reflect the narrower xylem vessels of less vigorous vines, increasing compartmentalisation efficiency

and decreasing susceptibility to esca (Pouzoulet *et al.*, 2020). We found that $\delta^{13}$C levels and pruning

weight were negatively correlated. In particular, a set of weakly susceptible cultivars of low vigour

were found to have less negative values for $\delta^{13}$C (e.g. Petit Manseng, Tannat, Mavrud, Colombard).

We suggest that this tandem action of these two traits in the prediction of esca incidence is not a

coincidence. As discussed in previous studies, these cultivars are more able to decrease their hydraulic

conductivity markedly and rapidly in response to non-optimal water conditions, thus ensuring larger

hydraulic safety margins but constraining plant growth and productivity (Torrez-Ruiz *et al.*, 2024).

Nevertheless, this relationship is ambiguous, and depends on water availability. Our findings are

consistent with those of Tambussi *et al.* (2007), who found that, for common cereals, plants with a

lower WUE have a competitive advantage for growth and productivity in climates with rainfall

occurring during the growing season (here, a temperate oceanic climate).

Esca disease expression is also considered to be positively correlated with leaf nitrogen status

(Calzarano *et al.*, 2009; Li, 2015) through a roughly twofold mechanism that has already been

discussed for plants (e.g. Mur *et al.*, 2017; Sun *et al.*, 2020) involving modifications to plant

metabolism (e.g. enhanced vigour) and a favouring of fungal development and pathogenicity. In our

experimental vineyard, normalised nitrogen index was not a determinant of varietal susceptibility to

esca. As nitrogen status and vigour were only weakly correlated in this dataset (Supplementary

Material 7), it seems likely that the variability of nitrogen status was limited in this trial, potentially

accounting for the lack of correlation between vine nitrogen status and esca symptoms. This result

and those of Monod *et al.* (2023), who demonstrated a negative correlation between nitrogen status

and esca severity in a single cultivar growing in contrasting conditions in different vineyards, argue

in favour of a reconsideration, across a wide range of genotypes and growing conditions, of the link

between vine mineral status and the incidence of trunk disease.

Finally, we found no significant effect of phenology on the variability of esca expression between

varieties. There have been few studies of the links between trunk diseases and phenology. Serra *et al.*

(2018) assumed that phenology affects esca expression in relation to climatic variables. As foliar

symptoms emerge only between fruit set and veraison and accumulate through the summer season in

a systematic progressive sigmoidal pattern (Lecomte *et al.*, 2024), we can hypothesise that varietal

shifts in phenological stage may influence cultivar-specific patterns of esca expression, particularly

when climatic conditions fluctuate sharply. Monitoring over a time scale finer than one year would

improve our understanding of the interaction between grapevine phenology and disease dynamics.

## Conclusion

This study, making use of a unique experimental facility, adds to our knowledge of the contribution

of genetic diversity to the sustainable management of grapevine dieback. Our findings confirm the

utility of performing pluriannual monitoring in natural conditions, within common garden

experimental vineyards, for accurate comparisons of susceptibility to trunk diseases between

cultivars. The ranking of varieties for susceptibility, based on the incidence of esca foliar symptoms,

was consistent with the findings of a previous field trial conducted in Italy (Murolo and Romanazzi,

2014; Figure 8A). A weaker correlation was also identified with the ranking obtained in a network of

French vineyards (Figure 8B). By contrast, no correlation was found with the susceptibility gradient

obtained by artificially inoculating cuttings with *P. chlamydospora* (Pouzoulet *et al.*, 2020; Figure

8C). This confirms that the susceptibility of grapevine genotypes to trunk diseases cannot be assessed

solely on the basis of inoculations under control conditions, as already discussed by several authors

(e.g. Sosnowski *et al.*, 2007; Reis *et al.*, 2019). Furthermore, susceptibility to trunk diseases is

potentially plastic in response to the environment. It therefore seems necessary to replicate such

monitoring under contrasting conditions, to facilitate the selection of the most suitable cultivars for

each production context. Within this framework, a complementary strategy would involve monitoring

a single cultivar in multiple environments (see Monod *et al.*, 2023).

Finally, we recommend tackling the highly multifactorial nature of plant dieback through integrative

approaches taking multiple factors, such as other pests and diseases, climatic hazards, soil fertility

and technical management, into account.

**Figure 8. Scatterplots showing the relationships between cultivar ranks for susceptibility to esca as assessed in this work and reported in previous publications.**

Each dot corresponds to the ranking of a cultivar in a given study, for the range of cultivars common to the two studies compared. (A) Relationship between the cultivar rankings obtained in this study and those obtained by Murolo and Romanazzi (2014) in a one-year (2008) study monitoring of foliar symptom expression in an experimental vineyard in Italy (14 cultivars common to the two studies). The regression line corresponds to a linear model, and the standard error is provided; (B) Relationship between the cultivar rankings obtained in this study and those obtained by Etienne *et al.* (unpublished data), in a study over six years (2003-2022) monitoring foliar symptom expression in a network of 2084 vineyards located in French grape-producing areas (19 cultivars common to the two studies); (C) Relationship between the cultivar rankings obtained in this study and those obtained by Pouzoulet *et al.* (2020) by phenotyping the length of stem necrosis induced by *Phaeomoniella chlamydospora* after 12 weeks of incubation (11 cultivars common to the two studies). Rank-based Kendall's correlation coefficients are indicated for each relationship.

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## Competing Interests statement

The authors have no competing interests to declare.

## References

**References**

Anderson, K., & Aryal, N. R. (2013). *Which Winegrape Varieties are Grown Where? : A global empirical picture.*

University of Adelaide Press. https://www.jstor.org/stable/10.20851/j.ctt1t304tz

Andreini, L., Cardelli, R., Bartolini, S., Scalabrelli, G., & Viti, R. (2014). Esca symptoms appearance in *Vitis vinifera* L. :

Influence of climate, pedo-climatic conditions and rootstock/cultivar combination. *Vitis*, *53*(1), 33-38.

Andreini, L., Guarino, L., Viti, R., & Scalabrelli, G. (2013). Evaluation of the effect of esca disease on bud break in *Vitis*

*vinifera* L. : Possible relationship between cultivars and rootstocks. VITIS - Journal of Grapevine Research, 52(1), 33-40.

https://doi.org/10.5073/vitis.2013.52.33-40

Arnold, G., Prado, E., Dumas, V., Butterlin, G., Duchêne, E., Avia, K., & Merdinoglu, D. (2023). *Discovery of a locus*

*associated with susceptibility to esca dieback in grapevine* (p. 2023.10.20.563213). bioRxiv.

https://doi.org/10.1101/2023.10.20.563213

Bacilieri, R., Lacombe, T., Le Cunff, L., Di Vecchi-Staraz, M., Laucou, V., Genna, B., Péros, J.-P., This, P., & Boursiquot,

J.-M. (2013). Genetic structure in cultivated grapevines is linked to geography and human selection. *BMC Plant Biology*,

*13*(1), 25. https://doi.org/10.1186/1471-2229-13-25

Bates, D., Mächler, M., Bolker, B., & Walker, S. (2015). Fitting Linear Mixed-Effects Models Using lme4. *Journal of*

*Statistical Software*, *67*, 1-48. https://doi.org/10.18637/jss.v067.i01

Bchir, A., Escalona, J. M., Gallé, A., Hernández-Montes, E., Tortosa, I., Braham, M., & Medrano, H. (2016). Carbon

isotope discrimination ($\delta13C$) as an indicator of vine water status and water use efficiency (WUE) : Looking for the most

representative sample and sampling time. *Agricultural Water Management*, *167*, 11-20.

https://doi.org/10.1016/j.agwat.2015.12.018

Bekris, F., Vasileiadis, S., Papadopoulou, E., Samaras, A., Testempasis, S., Gkizi, D., Tavlaki, G., Tzima, A., Paplomatas,

E., Markakis, E., Karaoglanidis, G., Papadopoulou, K. K., & Karpouzas, D. G. (2021). Grapevine wood microbiome

analysis identifies key fungal pathogens and potential interactions with the bacterial community implicated in grapevine

trunk disease appearance. *Environmental Microbiome*, *16*(1), 23. https://doi.org/10.1186/s40793-021-00390-1

Bortolami, G., Farolfi, E., Badel, E., Burlett, R., Cochard, H., Ferrer, N., King, A., Lamarque, L. J., Lecomte, P.,

Marchesseau-Marchal, M., Pouzoulet, J., Torres-Ruiz, J. M., Trueba, S., Delzon, S., Gambetta, G. A., & Delmas, C. E.

L. (2021a). Seasonal and long-term consequences of esca grapevine disease on stem xylem integrity. *Journal of*

*Experimental Botany*, *72*(10), 3914-3928. https://doi.org/10.1093/jxb/erab117

Bortolami, G., Gambetta, G. A., Cassan, C., Dayer, S., Farolfi, E., Ferrer, N., Gibon, Y., Jolivet, J., Lecomte, P., &

Delmas, C. E. L. (2021b). Grapevines under drought do not express esca leaf symptoms. *Proceedings of the National*

*Academy of Sciences*, *118*(43), e2112825118. https://doi.org/10.1073/pnas.2112825118

Bortolami, G., Gambetta, G. A., Delzon, S., Lamarque, L. J., Pouzoulet, J., Badel, E., Burlett, R., Charrier, G., Cochard,

H., Dayer, S., Jansen, S., King, A., Lecomte, P., Lens, F., Torres-Ruiz, J. M., & Delmas, C. E. L. (2019). Exploring the

Hydraulic Failure Hypothesis of Esca Leaf Symptom Formation. *Plant Physiology*, *181*(3), 1163-1174.

https://doi.org/10.1104/pp.19.00591

Boso, S., Alonso-Villaverde, V., Gago, P., Santiago, J. l., & Martínez, M. c. (2011). Susceptibility of 44 grapevine (*Vitis*

*vinifera* L.) varieties to downy mildew in the field. *Australian Journal of Grape and Wine Research*, *17*(3), 394-400.

https://doi.org/10.1111/j.1755-0238.2011.00157.x

Bruez, E., Baumgartner, K., Bastien, S., Travadon, R., Guérin-Dubrana, L., & Rey, P. (2016). Various fungal communities

colonise the functional wood tissues of old grapevines externally free from grapevine trunk disease symptoms. *Australian*

*Journal of Grape and Wine Research*, *22*(2), 288-295. https://doi.org/10.1111/ajgw.12209

Bruez, E., Lecomte, P., Grosman, J., Doublet, B., Bertsch, C., Fontaine, F., Ugaglia, A., Teissedre, P.-L., Costa, J.-P. D.,

Guerin-Dubrana, L., & Rey, P. (2013). Overview of grapevine trunk diseases in France in the 2000s. *Phytopathologia*

*Mediterranea*, *52*(2), 262-275. https://doi.org/10.14601/Phytopathol_Mediterr-11578

Bruez, E., Vallance, J., Gautier, A., Laval, V., Compant, S., Maurer, W., Sessitsch, A., Lebrun, M.-H., & Rey, P. (2020).

Major changes in grapevine wood microbiota are associated with the onset of esca, a devastating trunk disease.

*Environmental Microbiology*, *22*(12), 5189-5206. https://doi.org/10.1111/1462-2920.15180

Bruez, E., Vallance, J., Gerbore, J., Lecomte, P., Costa, J.-P. D., Guerin-Dubrana, L., & Rey, P. (2014). Analyses of the

Temporal Dynamics of Fungal Communities Colonizing the Healthy Wood Tissues of Esca Leaf-Symptomatic and

Asymptomatic Vines. *PLOS ONE*, *9*(5), e95928. https://doi.org/10.1371/journal.pone.0095928

Calzarano, F., Amalfitano, C., Seghetti, L., & Cozzolino, V. (2009). Nutritional status of vines affected with esca proper.

Phytopathologia Mediterranea, 48(1), 20-31. https://doi.org/10.14601/Phytopathol_Mediterr-2872

Calzarano, F., Osti, F., Baránek, M., & Di Marco, S. (2018). Rainfall and temperature influence expression of foliar

symptoms of grapevine leaf stripe disease (esca complex) in vineyards. *Phytopathologia Mediterranea*, *47*, 488-505.

https://doi.org/10.14601/Phytopathol_Mediterr-23787

Celette, F., & Gary, C. (2013). Dynamics of water and nitrogen stress along the grapevine cycle as affected by cover

cropping. *European Journal of Agronomy*, *45*, 142-152. https://doi.org/10.1016/j.eja.2012.10.001

Claverie, M., Notaro, M., Fontaine, F., & Wery, J. (2020). Current knowledge on Grapevine Trunk Diseases with complex

etiology : A systemic approach. *Phytopathologia Mediterranea*, *59*(1), 29-53. https://doi.org/10.36253/phyto-11150

Csótó, A., Nagy, A., Laurinyecz, N., Nagy, Z. A., Németh, C., Németh, E. K., Csikász-Krizsics, A., Rakonczás, N.,

Fontaine, F., Fekete, E., Flipphi, M., Karaffa, L., & Sándor, E. (2023). Hybrid Vitis Cultivars with American or Asian

Ancestries Show Higher Tolerance towards Grapevine Trunk Diseases. *Plants*, *12*(12), 2328.

https://doi.org/10.3390/plants12122328

Dayer, S., Lamarque, L. J., Burlett, R., Bortolami, G., Delzon, S., Herrera, J. C., Cochard, H., & Gambetta, G. A. (2022).

Model-assisted ideotyping reveals trait syndromes to adapt viticulture to a drier climate. *Plant Physiology*, *190*(3),

1673-1686. https://doi.org/10.1093/plphys/kiac361

Dell'Acqua, N., Gambetta, G. A., Delzon, S., Ferrer, N., Lamarque, L. J., Saurin, N., Theodore, P., & Delmas, C. E. L.

(2023). Mechanisms of grapevine resilience to a vascular disease : Investigating stem radial growth, xylem development,

and physiological acclimation. *Annals of Botany*, 23, 1-15. https://doi.org/10.1093/aob/mcad188

Destrac-Irvine A., van Leeuwen C. (2016). VitAdapt: an experimental program to study the behavior of a wide range of

Vitis vinifera varieties in a context of climate change in the Bordeaux vineyards.Climwine, sustainable grape and wine

production in the context of climate change, 11-13 April 2016, Bordeaux. Full text proceedings paper, 165-171.

Destrac-Irvine, A., Barbeau, G., Resseguier, L. de, Dufourcq, T., Dumas, V., Cortazar-Atauri, I. G. de, Ojeda, H., Saurin,

N., Leeuwen, C. van, & Duchêne, É. (2019). Measuring the phenology to more effectively manage the vineyard. *IVES*

*Technical Reviews, Vine and Wine*. https://doi.org/10.20870/IVES-TR.2019.2586

Dewasme, C., Mary, S., Darrieutort, G., Roby, J.-P., & Gambetta, G. A. (2022). Long-Term Esca Monitoring Reveals

Disease Impacts on Fruit Yield and Wine Quality. *Plant Disease*, *106*(12), 3076-3082. https://doi.org/10.1094/PDIS-11-

21-2454-RE

Di Marco, S., & Osti, F. (2008). Foliar Symptom Expression of Wood Decay in *Actinidia deliciosa* in Relation to

Environmental Factors. *Plant Disease*, *92*(8), 1150-1157. https://doi.org/10.1094/PDIS-92-8-1150

Fanton, A. C., & Brodersen, C. (2021). Hydraulic consequences of enzymatic breakdown of grapevine pit membranes.

*Plant Physiology*, *186*(4), 1919-1931. https://doi.org/10.1093/plphys/kiab191

Feliciano, A., Eskalen, A., & Gubler, W. (2004). Differential susceptibility of three grapevine cultivars to

*Phaeoacremonium aleophilum* and *Phaeomoniella chlamydospora* in California. *Phytopathologia Mediterranea*, *43*,

66-69. https://doi.org/10.14601/Phytopathol_Mediterr-1727

Ferguson, J. C., Moyer, M. M., Mills, L. J., Hoogenboom, G., & Keller, M. (2014). Modeling Dormant Bud Cold

Hardiness and Budbreak in Twenty-Three Vitis Genotypes Reveals Variation by Region of Origin. *American Journal of*

*Enology and Viticulture*, *65*(1), 59-71. https://doi.org/10.5344/ajev.2013.13098

Gaforio, L., García-Muñoz, S., Cabello, F., & Muñoz-Organero, G. (2011). Evaluation of susceptibility to powdery

mildew (*Erysiphe necator*) in *Vitis vinifera* varieties. VITIS - Journal of Grapevine Research, 50(3), 123-126.

https://doi.org/10.5073/vitis.2011.50.123-126

Gilbert, G. S., Briggs, H. M., & Magarey, R. (2015). The Impact of Plant Enemies Shows a Phylogenetic Signal. *PLOS*

*ONE*, *10*(4), e0123758. https://doi.org/10.1371/journal.pone.0123758

Gramaje, D., García-Jiménez, J., & Armengol, J. (2010). Field Evaluation of Grapevine Rootstocks Inoculated with Fungi

Associated with Petri Disease and Esca. *American Journal of Enology and Viticulture*, *61*(4), 512-520.

https://doi.org/10.5344/ajev.2010.10021

Gramaje, D., Úrbez-Torres, J. R., & Sosnowski, M. R. (2018). Managing Grapevine Trunk Diseases With Respect to

Etiology and Epidemiology : Current Strategies and Future Prospects. *Plant Disease*, *102*(1), 12-39.

https://doi.org/10.1094/PDIS-04-17-0512-FE

Gubler, W., Thind, T., Feliciano, A., & Eskalen, A. (2004). Pathogenicity of *Phaeoacremonium aleophilum* and

*Phaeomoniella chlamydospora* on Grape Berries in California. *Phytopathologia Mediterranea*, *43*, 70-74.

https://doi.org/10.14601/Phytopathol_Mediterr-1735

Guerin-Dubrana, L., Labenne, A., Labrousse, J. C., Bastien, S., Rey, P., & Gégout-Petit, A. (2013). Statistical analysis of

grapevine mortality associated with esca or Eutypa dieback foliar expression. Phytopathologia Mediterranea, 52(2), 276-

288. https://doi.org/10.14601/Phytopathol_Mediterr-11602

Hartig, F., & Lohse, L. (2022). *DHARMa : Residual Diagnostics for Hierarchical (Multi-Level / Mixed) Regression*

*Models* (0.4.6) [Logiciel]. https://cran.r-project.org/web/packages/DHARMa/index.html

Hofstetter, V., Buyck, B., Croll, D., Viret, O., Couloux, A., & Gindro, K. (2012). What if esca disease of grapevine were

not a fungal disease? *Fungal Diversity*, *54*(1), 51-67. https://doi.org/10.1007/s13225-012-0171-z

Keck, F., Rimet, F., Bouchez, A., & Franc, A. (2016). phylosignal : An R package to measure, test, and explore the

phylogenetic signal. *Ecology and Evolution*, *6*(9), 2774-2780. https://doi.org/10.1002/ece3.2051

Kovács, C., Balling, P., Bihari, Z., Nagy, A., & Sándor, E. (2017). Incidence of grapevine trunk diseases is influenced by

soil, topology and vineyard age, but not by *Diplodia seriata* infection rate in the Tokaj Wine Region, Hungary.

*Phytoparasitica*, *45*(1), 21-32. https://doi.org/10.1007/s12600-017-0570-5

Lamarque, L. J., Delmas, C. E. L., Charrier, G., Burlett, R., Dell'Acqua, N., Pouzoulet, J., Gambetta, G. A., & Delzon,

S. (2023). Quantifying the grapevine xylem embolism resistance spectrum to identify varieties and regions at risk in a

future dry climate. *Scientific Reports*, *13*(1), 7724. https://doi.org/10.1038/s41598-023-34224-6

Laucou, V., Lacombe, T., Dechesne, F., Siret, R., Bruno, J.-P., Dessup, M., Dessup, T., Ortigosa, P., Parra, P., Roux, C.,

Santoni, S., Varès, D., Péros, J.-P., Boursiquot, J.-M., & This, P. (2011). High throughput analysis of grape genetic

diversity as a tool for germplasm collection management. *Theoretical and Applied Genetics*, *122*(6), 1233-1245.

https://doi.org/10.1007/s00122-010-1527-y

Laveau, C., Letouze, A., Louvet, G., Bastien, S., & Guerin-Dubrana, L. (2009). Differential aggressiveness of fungi

implicated in esca and associated diseases of grapevine in France. Phytopathologia Mediterranea, 48(1), 32-46.

https://doi.org/10.14601/Phytopathol_Mediterr-2873

Lecomte, P., Bénétreau, C., Diarra, B., Meziani, Y., Delmas, C., & Fermaud, M. (2024). Logistic modeling of summer

expression of esca symptoms in tolerant and susceptible cultivars in Bordeaux vineyards. *OENO One*, *58*(1).

https://doi.org/10.20870/oeno-one.2024.58.1.7571

Lecomte, P., Darrieutort, G., Liminana, J.-M., Comont, G., Muruamendiaraz, A., Legorburu, F.-J., Choueiri, E., Jreijiri,

F., El Amil, R., & Fermaud, M. (2012). New Insights into Esca of Grapevine : The Development of Foliar Symptoms and

Their Association with Xylem Discoloration. *Plant Disease*, *96*(7), 924-934. https://doi.org/10.1094/PDIS-09-11-0776-

RE

Lecomte, P., Diarra, B., Carbonneau, A., Rey, P., & Chevrier, C. (2018). Esca of grapevine and training practices in

France : Results of a 10-year survey. *Phytopathologia Mediterranea*, *57*(3), 472-487.

https://doi.org/10.14601/Phytopathol_Mediterr-22025

Lemaitre-Guillier, C., Fontaine, F., Roullier-Gall, C., Harir, M., Magnin-Robert, M., Clément, C., Trouvelot, S., Gougeon,

R. D., Schmitt-Kopplin, P., & Adrian, M. (2020). Cultivar- and Wood Area-Dependent Metabolomic Fingerprints of

Grapevine Infected by Botryosphaeria Dieback. *Phytopathology®*, *110*(11), 1821-1837. https://doi.org/10.1094/PHYTO-

02-20-0055-R

Li, S. (2015). *Modélisation spatio-temporelle pour l'esca de la vigne à l'échelle de la parcelle* [Phdthesis, Université de

Bordeaux]. https://theses.hal.science/tel-01793300

Lorenz, D. h., Eichhorn, K. w., Bleiholder, H., Klose, R., Meier, U., & Weber, E. (1995). Growth Stages of the Grapevine :

Phenological growth stages of the grapevine (*Vitis vinifera* L. ssp. *vinifera*)—Codes and descriptions according to the

extended BBCH scale†. *Australian Journal of Grape and Wine Research*, *1*(2), 100-103. https://doi.org/10.1111/j.1755-

0238.1995.tb00085.x

Magnin-Robert, M., Letousey, P., Spagnolo, A., Rabenoelina, F., Jacquens, L., Mercier, L., Clément, C., Fontaine, F.,

Magnin-Robert, M., Letousey, P., Spagnolo, A., Rabenoelina, F., Jacquens, L., Mercier, L., Clément, C., & Fontaine, F.

(2011). Leaf stripe form of esca induces alteration of photosynthesis and defence reactions in presymptomatic leaves.

*Functional Plant Biology*, *38*(11), 856-866. https://doi.org/10.1071/FP11083

Martínez-Diz, M. del P., Díaz-Losada, E., Barajas, E., Ruano-Rosa, D., Andrés-Sodupe, M., & Gramaje, D. (2019).

Screening of Spanish *Vitis vinifera* germplasm for resistance to *Phaeomoniella chlamydospora*. *Scientia Horticulturae*,

*246*, 104-109. https://doi.org/10.1016/j.scienta.2018.10.049

Merdinoglu, D., Schneider, C., Prado, E., Wiedemann-Merdinoglu, S., & Mestre, P. (2018). Breeding for durable

resistance to downy and powdery mildew in grapevine. *OENO One*, *52*(3), 203-209. https://doi.org/10.20870/oeno-

one.2018.52.3.2116

Monod, V., Zufferey, V., Wilhelm, M., Viret, O., Gindro, K., Croll, D., & Hofstetter, V. (2023). *A systemic approach*

*allows to identify the pedoclimatic conditions most critical in the susceptibility of a grapevine cultivar to*

*esca/Botryosphaeria dieback* (p. 2023.05.23.541976). bioRxiv. https://doi.org/10.1101/2023.05.23.541976

Moret, F., Lemaître-Guillier, C., Grosjean, C., Clément, G., Coelho, C., Negrel, J., Jacquens, L., Morvan, G., Mouille,

G., Trouvelot, S., Fontaine, F., & Adrian, M. (2019). Clone-Dependent Expression of Esca Disease Revealed by Leaf

Metabolite Analysis. *Frontiers in Plant Science*, *9*. https://www.frontiersin.org/article/10.3389/fpls.2018.01960

Mugnai, L., Graniti, A., & Surico, G. (1999). Esca (Black Measles) and Brown Wood-Streaking : Two Old and Elusive

Diseases of Grapevines. *Plant Disease*, *83*(5), 404-418. https://doi.org/10.1094/PDIS.1999.83.5.404

Mur, L. A. J., Simpson, C., Kumari, A., Gupta, A. K., & Gupta, K. J. (2017). Moving nitrogen to the centre of plant

defence against pathogens. *Annals of Botany*, *119*(5), 703-709. https://doi.org/10.1093/aob/mcw179

Murolo, S., & Romanazzi, G. (2014). Effects of grapevine cultivar, rootstock and clone on esca disease. *Australasian*

*Plant Pathology*, *43*(2), 215-221. https://doi.org/10.1007/s13313-014-0276-9

Nicolas, S. D., Péros, J.-P., Lacombe, T., Launay, A., Le Paslier, M.-C., Bérard, A., Mangin, B., Valière, S., Martins, F.,

Le Cunff, L., Laucou, V., Bacilieri, R., Dereeper, A., Chatelet, P., This, P., & Doligez, A. (2016). Genetic diversity,

linkage disequilibrium and power of a large grapevine (*Vitis vinifera* L) diversity panel newly designed for association

studies. *BMC Plant Biology*, *16*, 74. https://doi.org/10.1186/s12870-016-0754-z

Ouadi, L., Bruez, E., Bastien, S., Yacoub, A., Coppin, C., Guérin-Dubrana, L., Fontaine, F., Domec, J.-C., & Rey, P.

(2021). Sap Flow Disruption in Grapevine Is the Early Signal Predicting the Structural, Functional, and Genetic Responses

to Esca Disease. *Frontiers in Plant Science*, *12*. https://www.frontiersin.org/articles/10.3389/fpls.2021.695846

Paňitrur-De La Fuente, C., Valdés-Gómez, H., Roudet, J., Acevedo-Opazo, C., Verdugo-Vásquez, N., Araya-Alman, M.,

Lolas, M., Moreno, Y., & Fermaud, M. (2018). Classification of winegrape cultivars in Chile and France according to

their susceptibility to *Botrytis cinerea* related to fruit maturity. *Australian Journal of Grape and Wine Research*, *24*(2),

145-157. https://doi.org/10.1111/ajgw.12315

Plantevin, M., Gowdy, M., Destrac-Irvine, A., Marguerit, E., Gambetta, G. A., & Leeuwen, C. van. (2022). Using δ13C

and hydroscapes for discriminating cultivar specific drought responses. *OENO One*, *56*(2). https://doi.org/10.20870/oeno-

one.2022.56.2.5434

Pouzoulet, J., Rolshausen, P. E., Charbois, R., Chen, J., Guillaumie, S., Ollat, N., Gambetta, G. A., & Delmas, C. E. L.

(2020). Behind the curtain of the compartmentalization process : Exploring how xylem vessel diameter impacts vascular

pathogen resistance. *Plant, Cell & Environment*, *43*(11), 2782-2796. https://doi.org/10.1111/pce.13848

Pouzoulet, J., Scudiero, E., Schiavon, M., & Rolshausen, P. E. (2017). Xylem Vessel Diameter Affects the Compartmentalization of the Vascular Pathogen *Phaeomoniella chlamydospora* in Grapevine. *Frontiers in Plant Science*, *8*. https://www.frontiersin.org/article/10.3389/fpls.2017.01442

R Core Team. (2022). *R: A Language and Environment for Statistical Computing* [Logiciel]. R Foundation for Statistical Computing. https://www.R-project.org/

Reis, P., Pierron, R., Larignon, P., Lecomte, P., Abou-Mansour, E., Farine, S., Bertsch, C., Jacques, A., Trotel-Aziz, P., Rego, C., & Fontaine, F. (2019). Vitis Methods to Understand and Develop Strategies for Diagnosis and Sustainable Control of Grapevine Trunk Diseases. *Phytopathology®*, *109*(6), 916-931. https://doi.org/10.1094/PHYTO-09-18-0349-RVW

Rolshausen, P. E., Greve, L. C., Labavitch, J. M., Mahoney, N. E., Molyneux, R. J., & Gubler, W. D. (2008). Pathogenesis of Eutypa lata in grapevine : Identification of virulence factors and biochemical characterization of cordon dieback. *Phytopathology*, *98*(2), 222-229. https://doi.org/10.1094/PHYTO-98-2-0222

Romanazzi, G., Murolo, S., Pizzichini, L., & Nardi, S. (2009). Esca in young and mature vineyards, and molecular diagnosis of the associated fungi. *European Journal of Plant Pathology*, *125*(2), 277-290. https://doi.org/10.1007/s10658-009-9481-8

Seem, R. C. (1984). Disease Incidence and Severity Relationships. *Annual Review of Phytopathology*, *22*(1), 133-150. https://doi.org/10.1146/annurev.py.22.090184.001025

Serra, S., Ligios, V., Schianchi, N., Prota, V., & Scanu, B. (2018). Expression of grapevine leaf stripe disease foliar symptoms in four cultivars in relation to grapevine phenology and climatic conditions. Phytopathologia Mediterranea, 57(3), 337-568. https://doi.org/10.14601/Phytopathol_Mediterr-24088

Sosnowski, M. R., Lardner, R., Wicks, T. J., & Scott, E. S. (2007). The Influence of Grapevine Cultivar and Isolate of Eutypa lata on Wood and Foliar Symptoms. *Plant Disease*, *91*(8), 924-931. https://doi.org/10.1094/PDIS-91-8-0924

Sun, Y., Wang, M., Mur, L. A. J., Shen, Q., & Guo, S. (2020). Unravelling the Roles of Nitrogen Nutrition in Plant Disease Defences. *International Journal of Molecular Sciences*, *21*(2), 572. https://doi.org/10.3390/ijms21020572

Tambussi, E. a., Bort, J., & Araus, J. l. (2007). Water use efficiency in C3 cereals under Mediterranean conditions : A review of physiological aspects. *Annals of Applied Biology*, *150*(3), 307-321. https://doi.org/10.1111/j.1744-7348.2007.00143.x

This, P., Lacombe, T., & Thomas, M. R. (2006). Historical origins and genetic diversity of wine grapes. *Trends in*
*Genetics*, *22*(9), 511-519. https://doi.org/10.1016/j.tig.2006.07.008

Torres-Ruiz, J. M., Cochard, H., Delzon, S., Boivin, T., Burlett, R., Cailleret, M., Corso, D., Delmas, C. E. L., De Caceres,
M., Diaz-Espejo, A., Fernández-Conradi, P., Guillemot, J., Lamarque, L. J., Limousin, J.-M., Mantova, M., Mencuccini,
M., Morin, X., Pimont, F., De Dios, V. R., … Martin-StPaul, N. K. (2024). Plant hydraulics at the heart of plant, crops
and ecosystem functions in the face of climate change. *New Phytologist*, *241*(3), 984-999.
https://doi.org/10.1111/nph.19463

Wolkovich, E. M., García de Cortázar-Atauri, I., Morales-Castilla, I., Nicholas, K. A., & Lacombe, T. (2018). From Pinot
to Xinomavro in the world's future wine-growing regions. *Nature Climate Change*, *8*(1), 29-37.
https://doi.org/10.1038/s41558-017-0016-6

Wei T, Simko V (2021). *R package 'corrplot': Visualization of a Correlation Matrix*. (Version 0.92),
https://github.com/taiyun/corrplot.