Spaceflight causes strain-dependent gene expression changes associated with lipid and extracellular matrix dysregulation in the mouse kidney in vivo

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Highlights

- Spaceflight has a significant effect on gene expression in the kidney.
- Responses in the BALB/c indicate milder transcriptomic perturbations than C57BL/6J.
- Lipid metabolism was altered in both strains of mice.
- Extracellular matrix metabolism and TGF-β signalling up in BALB/c down in C57BL/6J.
- Reduced gene expression of hyaluronan synthesis pathway in BALB/c but not in C57BL/6J.

Abstract

To explore new worlds we must ensure humans can survive and thrive in the space environment. Incidence of kidney stones in astronauts is a major risk factor associated with long term missions, caused by increased blood calcium levels due to bone demineralisation triggered by microgravity and space radiation. Transcriptomic changes have been observed in other tissues during spaceflight, including the kidney. We analysed kidney transcriptome patterns in two different strains of mice flown on the International Space Station, C57BL/6J and BALB/c. Here we show a link between spaceflight and transcriptome patterns associated with dysregulation of lipid and extracellular matrix metabolism and altered transforming growth factor-beta signalling. A stronger response was seen in C57BL/6J mice than BALB/c. Genetic differences in hyaluronan metabolism between strains may confer protection against extracellular matrix remodelling through downregulation of epithelial-mesenchymal transition. We intend for our findings to contribute to development of new countermeasures against kidney disease in astronauts and people here on Earth.

Keywords

Transcriptomics
Kidney
Microgravity
Space radiation
Extracellular matrix

1. Introduction

Renal health risks are one of the highest risk factors facing astronauts on long term missions due to increased incidence of kidney stones after missions [1](Afshinnekoo *et al.,* 2020). Current mitigations such as protective shielding and personal medicine are already in place, but are insufficient for extended missions [1](Afshinnekoo *et al.,* 2020).

Ionising radiation in space causes increased levels of reactive oxygen species, which leads to oxidative stress, inflammation, genetic and epigenetic alterations, and mitochondrial dysfunction [2,3](Pavlakou *et al.,* 2018; da Silveira *et al.,* 2020). The kidney is particularly vulnerable to the stresses of spaceflight due to its large number of mitochondria; playing an important part in regulation of oxidative stress in the body and being severely affected by oxidative stress itself. It is affected in a multifactorial way and disruption of its oxidative stress regulation functions act as a positive feedback mechanism for continuous kidney damage [2](Pavlakou *et al.,* 2018).

Previous research has shown downregulated expression of nuclear oxidative phosphorylation genes and upregulated expression of mitochondrial oxidative phosphorylation genes in space in multiple tissues including the kidney; this switch is thought to be caused by reactive oxygen species causing damage to transcripts from nuclear DNA [3](da Silveira *et al.,* 2020). The kidney

is particularly sensitive to oxidative stress due to its large number of mitochondria. Other effects that space has been observed to exert on the kidneys include changes to endothelial cells and the cytoskeleton, vascular senescence, altered fluid distribution and neurohormonal balance [2](Pavlakou *et al.,* 2018).

Animals have been used as models to assess the ability of humans to survive in space for the last seventy years, and the first mouse was launched into space in 1950 [4](Gray, 1998). Previous studies have shown that different strains of mice have different reactions to kidney injury. C57BL/6 mice showed more profound inflammation, intrinsic injury responses and renal architecture disruption than BALB/c mice in response to reversible unilateral ureteral obstruction, a model of renal fibrosis [5](Puri *et al.,* 2010). C57BL/6 mice have also been shown to be more sensitive to the effects of streptozotocin-induced diabetes than BALB/c mice [6](Gurley *et al.,* 2006). Induction of kidney stones experimentally in mice has been found to be significantly more difficult than in rats, leading to the hypothesis that mice have species-specific protective mechanisms against kidney stones. A model of calcium oxalate crystal deposition in the mouse kidney was induced by injection of glyoxylate but began to decrease 12 days after administration [7](Okada *et al.,* 2007). In contrast to the difficulties inducing kidney stones in mice, they are commonly used as subjects in unilateral ureteral obstruction which is a model for chronic kidney disease and renal fibrosis [8](Martínez-Klimova *et al.,* 2019) both of which are diseases correlated with ageing.

Spaceflight is known to promote accelerated ageing, and microgravity in particular has been used as a model for ageing due to the similarities in physiological changes observed in both, including insulin resistance, decreased protein breakdown after meals, decreased immune function and dysregulation of cytokine production [9](Biolo *et al.,* 2003).

One of the changes associated with ageing in the kidney is nephrosclerosis, which consists of glomerulosclerosis, interstitial fibrosis, and arteriosclerosis [10](Denic, Glassock and Rule, 2016). The incidence of chronic kidney disease development is higher in patients with kidney stones and therefore kidney stones are a predictor and risk factor for chronic kidney disease[11,12]. Tissue injury initiates renal fibrosis, and this injury may be caused by kidney stones. The formation of kidney stones in the papilla involves activation of inflammatory cascades, leading to the accumulation of calcium, phosphate, and oxalate ions in the interstitial space, forming Randall's plaque. The plaque can erode into the renal pelvis leading to loss of cells and exposing it to urine supersaturated with calcium phosphate, which both contribute to renal stone formation [13](Chaiyarit and Thongboonkerd, 2020). High levels of reactive oxygen species associated with the formation of kidney stones can also cause lipid peroxidation which can damage cell membranes [13](Chaiyarit and Thongboonkerd, 2020) which is another mechanism by which fibrotic tissue injury may occur.

Fibrosis is a pathological build-up of extracellular matrix caused by trauma or injury, for instance a kidney stone, and is driven by disruption of transforming growth factor-β (TGF-β) signalling. Increased levels of TGF-β and PAI1 cause an increased production and decreased degradation of the extracellular matrix which lead to fibrotic build-up of its components [14](Zhao *et al.,* 2020). Previous studies have shown an association between dysregulation of

lipid metabolism, a decline in kidney function and development of chronic kidney disease [15](Bulbul *et al.,* 2018).

In spaceflight, profibrotic markers have been observed in mouse lung tissue [16](Tian *et al.,* 2010), and lipid dysregulation has been observed in mouse liver tissue [17](Beheshti *et al.,* 2019). TGF-β has also been identified as an important regulator of response to spaceflight [18](Beheshti *et al.,* 2018).

Interestingly, a recent study reported unexpected differences in the intensity of transcriptomic changes in tissues, including the kidney, in mice of the C57BL/6J and BALB/c lineages when exposed to spaceflight. BALB/c presented subtle alterations in gene expression whereas C57BL/6J presented major alterations [3](da Silveira et al., 2020).

In this study, for the first time, we focus on renal transcriptome alterations in space and how they could be impacted by genetic background.

2. Methods

2.1. Subjects

Transcriptomic data related to kidney tissue obtained in the missions Rodent Research-1 (RR-1) version 3 [19], Rodent Research-3 (RR-3) version 4 [20], and Rodent Research-7 (RR-7) version 8 [21] were obtained from NASA's GeneLab Platform [22](https://genelab.nasa.gov/) from dataset identifiers OSDR-102, OSD-163 and OSDR-253, and samples were processed as detailed in [3](da Silveira *et al.,* 2020).

2.2. Differential Expression Analysis

R Studio (version 1.4.1717) [23] and the package DESeq2 (version 1.32.0) [24](Love, Huber and Anders, 2014) were used to perform differential gene expression analysis by fitting a generalised linear model to each gene following a negative binomial distribution. Differentially expressed genes were identified by Flight to Ground Control comparison on C57BL/6J mice in the OSD-102 dataset with an adjusted p value of 0.1. The same method was used on the C57BL/6J and C3H/HeJ mice in the OSD-253 dataset, and separate analyses were carried out for the 25 and 75 day timepoints compared to their ground control counterparts. Using the same method, zero were identified for OSD-163, so the Basal group was used as a common control, and differential expression analysis was carried out between Spaceflight versus Basal, and Ground Control versus Basal groups. Genes differentially expressed in the Ground Control group were excluded to leave only genes differentially expressed in Spaceflight compared to Basal.

2.3. Comparison between strains

BALB/c mice possess a distinct set of non-synonymous genetic differences from C57BL/6J mice [25](Timmermans, Van Montagu and Libert, 2017). Pathway enrichment using the WebGestalt biological processes gene ontology database [26](Wang *et al.*, 2017) was used to assess the effect of the genetic differences between the two strains of mice on their responses to spaceflight. Venny [27](Oliveros, 2015) was used to determine genes with non-synonymous mutations between C57BL/6J and BALB/c, compared to differentially expressed genes (adjusted p-value <= 0.1) in the RR-1 and RR-3 datasets. R Studio (version 1.4.1717)[23] and package gplots [28](Warnes *et al.*, 2020) was used to generate supervised heatmaps of the genes identified by these two comparisons.

2.4. Quantification and statistical analysis

RNA-sequence data analysis on murine samples and gene set enrichment analysis using datasets OSD-102, OSD-163 and OSD-253 and carried out as detailed in [3](da Silveira *et al.*, 2020). Additionally, Gene Ontology Hallmarks pathways were run. Gene set enrichment analysis was performed using GSEA (version 4.1.0) [29,30](Subramanian et al., 2005; Mootha et al., 2003) using gene expression independent from the analysis performed using Deseq2. EnrichmentMap (versions 3.3.2 and 3.3.3) and AutoAnnotate (versions 1.3.4 and 1.3.5) applications in Cytoscape (versions 3.8.2 and 3.9.0) were used to visualise enriched pathways using FDR q-value cut off value 0.1 [31,32](Shannon et al., 2003; Merico et al., 2010).

3. Results

3.1. Expression of genes involved in lipid metabolism, extracellular matrix and TGF- β signalling are affected in the kidney by spaceflight

Differential expression analysis of kidney transcriptome data from C57BL/6J (RR-1 mission) and BALB/c mice (RR-3 mission) was carried out to determine the differences in their responses to spaceflight. Differential expression analysis of C57BL/6J was determined by comparison of Spaceflight versus Ground Control groups and found 638 differentially expressed genes (supplementary table 1). The same analysis identified zero differentially expressed genes in RR-3 and an alternative approach to isolate specific genes altered by spaceflight on BALB/c kidney was used, i.e., both Spaceflight and Ground control groups were compared to basal levels of gene expression at the beginning of the experiment and genes that were altered only by spaceflight were selected, a total of 671 genes (supplementary table 2). The gene signature of the differentially expressed genes in C57BL/6J had a stronger and clearer separation between the Spaceflight and Ground Control groups than in BALB/c (Supplementary Figure 3).

Focusing on a subset of these differentially expressed genes containing only those whose expression had doubled or decreased by half after the exposure to spaceflight led to the identification of altered genes in lipid metabolic pathways, extracellular matrix degradation and TGF-β signalling, in the kidneys of both lineages (Fig. 1). Interestingly, the *Ccl28* gene – belonging to the TGF-β signalling pathway - was the most differentially expressed gene in the C57BL/6J samples with a log2 fold change of 2.05 (adjusted p-value <= 0.1) (Fig. 1a, supplementary table 1).

In C57BL/6J mice, the *Wnt11* gene shows downregulation in the spaceflight group compared to the control group, with a log2 fold change of -1.15 (adjusted p-value <= 0.1). This gene is involved in Wnt signalling, a pathway which is involved in crosstalk with TGF-beta signalling.

Fig. 1. Gene expression signature of mouse kidneys exposed to spaceflight shows genes of TGF-β and Lipid Metabolism altered in C57BL/6J and BALB/c mice lineages. Differentially expressed genes in spaceflight in kidney tissue from **a)** the C57BL/6J mice on the RR-1 mission and **b)** the BALB/c mice on the RR-3 mission (adjusted p-value <= 0.1) with a minimum of twice the expected log2 fold change (>1/<-1). FLT = Spaceflight samples, GC = Ground Control Samples.

3.2. Overrepresentation analysis shows spaceflight is associated with positive enrichment of cholesterol metabolic pathways and negative enrichment of ECM pathways in the kidney

The identified upregulated and downregulated genes from each dataset were compared to each other (supplementary figure 2) and pathway analysis was carried out on the differentially expressed genes which were upregulated or downregulated in both datasets (Table 1, supplementary figure 2). Both datasets showed upregulation in genes linked with cholesterol related pathways. Genes connected with extracellular matrix and TGF-β signalling pathways were upregulated in BALB/c (RR-3) and downregulated in C57BL/6J (RR-1). A similar analysis carried out using the GSEA Gene Ontology biological process database, supplementary table 5, showed that pathways related to increased lipid and fat metabolism were enriched in both datasets.

Table 1.
Spaceflight exposed kidney shows cholesterol and protein synthesis pathways positively and negatively enriched, respectively, in both lineages, while extracellular matrix pathways show negative enrichment in C57BL/6J and positive enrichment in BALB/c mice. Biological process pathways enriched by overrepresentation of differentially expressed genes in kidney transcriptomic data from C57BL/6J (RR-1) and BALB/c (RR-3) mice (FDR < 0.05).

	Upregulated in C57BL/6J	Downregulated in C57BL/6J
BALB/c up	Cholesterol Biosynthesis Pathway Cholesterol biosynthetic Cholesterol biosynthesis Superpathway of cholesterol biosynthesis Cholesterol Biosynthesis with Skeletal Dysplasias Cholesterol metabolism (includes both Bloch and Kandutsch-Russell pathways) Steroid biosynthetic Regulation of cholesterol biosynthesis by SREBP (SREBF) Cholesterol biosynthesis, squalene 2,3-epoxide => cholesterol Cholesterol biosynthesis II (via 24,25-dihydrolanosterol)	Genes encoding secreted soluble factors Ensemble of genes encoding ECM-associated proteins including ECM-affiliated proteins, ECM regulators and secreted factors Kisspeptin/Kisspeptin Receptor System in the Ovary E3 ubiquitin ligases ubiquitinate target proteins Ovarian steroidogenesis Protein ubiquitination Ensemble of genes encoding extracellular matrix and extracellular matrix-associated proteins TGF-beta signaling pathway
BALB/c down	N/A	Peptide chain elongation Eukaryotic Translation Elongation metabolism Ribosome Cytoplasmic Ribosomal Proteins Viral mRNA Translation Selenocysteine synthesis Translation Eukaryotic Translation Termination Nonsense Mediated Decay (NMD) independent of the Exon Junction Complex (EJC)

3.3. Diverging patterns of lipid synthesis, protein synthesis and circadian rhythm in response to spaceflight in C57BL/6J and BALB/c mice determined by overrepresentation analysis

Both C57BL/6J (RR-1) and BALB/c (RR-3) hallmarks indicated increased enrichment of pathways associated with epithelial cell remodelling, but different pathways were identified in each dataset supplementary table 3, supplementary table 4.

Comparison of the expression patterns of gene networks in the biological process database for the C57BL/6J (RR-1) and BALB/c (RR-3) datasets identified clusters of gene sets exhibiting overrepresentation, including clusters with opposite patterns between datasets (Fig. 2). Notably, cholesterol biosynthesis and fatty acyl coA biosynthesis were positively overrepresented in C57BL/6J and negatively overrepresented in BALB/c. The opposite pattern was seen in hallmarks related to translation, protein folding and circadian rhythm. Lipid metabolism and storage, cell cycle processes and immune response were positively overrepresented in both C57BL/6J and BALB/c (Supplementary Figure 3).

Fig. 2. Network analysis showing clusters with opposite expression patterns in spaceflight in gene ontology biological process comparison between kidney tissue data from C57BL/6J (RR-1) and BALB/c (RR-3) datasets.

3.4. Genetic background differences between C57BL/6J (RR-1) and BALB/c (RR-3) mice are a potential reason for the difference in expression patterns of genes related to lipid and extracellular matrix metabolism

While mice in both missions were affected by the space environment, the C57BL/6J mice appeared to experience more severe effects than the BALB/c mice. The BALB/c mouse strain used in RR-3 has a distinct set of genes with non-synonymous mutations compared to the C57BL/6J mouse strain used in RR-1. These genetic differences were assessed to provide insight into their different reactions to spaceflight, (Fig. 3, supplementary figure 4, and supplementary table 6). The hyaluronan metabolic pathway had the highest enrichment ratio, and genes in this pathway were differentially expressed by BALB/c mice in spaceflight. Other pathways containing non-synonymous mutations between the two strains which contained genes differentially expressed in BALB/c in spaceflight related to cytoskeleton and the innate immune system. Pathways containing non-synonymous mutations between the two strains which contained genes differentially expressed in C57BL/6J in spaceflight were connected to organic acid metabolism and negative regulation of intracellular signalling.

Fig. 3. Enriched biological process pathways of non-synonymous mutations in BALB/c mice compared to C57BL/6J strain reveal connections with expression of genes involved in pathways related to hyaluronan metabolism in BALB/c and intracellular signalling in C57BL/6J. Top 10 enriched biological process pathways in the genetics of non-synonymous mutations in BALB/c strain of mice compared to C57BL/6J strain[25] (Timmermans, Van Montagu and Libert, 2017) and their connections with enriched pathways in the transcriptomic data of kidneys obtained from C57BL/6J mice (RR-1) (green arrow) and BALB/c (RR-3) (pink arrow).

Comparison between genes with non-synonymous mutations in BALB/c compared to C57BL/6J and differentially expressed genes in C57BL/6J (RR-1) and BALB/c (RR-3) (Fig. 4) showed upregulation of genes related to fatty acid metabolism and downregulation of genes related to cell junction, and expression of genes related to cholesterol metabolism and PPAR signalling were altered in the comparison with differentially expressed genes in C57BL/6J. In comparison with differentially expressed genes in BALB/c, regulation of genes related to lipid transport were identified, as well as altered expression of genes involved in Wnt-protein binding, gap junction and lipid metabolism. In general, a pattern of upregulation of genes involved in lipid processes was seen in C57BL/6J and downregulation of genes involved in lipid processes was seen in BALB/c in response to spaceflight.

Fig. 4. Gene expression signatures of mouse kidneys exposed to spaceflight limited by non-synonymous mutations between the BALB/c and C57BL/6J genes show genes involved in lipid metabolism and cell junction altered in C57BL/6J and BALB/c mouse lineages. Differentially expressed genes in kidney transcriptomic data from **a)** C57BL/6J mice on the RR-1 mission and **b)** BALB/c mice on the RR-3 mission (adjusted p-value <= 0.1). FLT = Spaceflight samples, GC = Ground Control Samples.

3.5. Lack of differences in gene expression patterns in spaceflight and ground control groups of RR-7 mission indicate need to standardise protocols for rodent research on the ISS

Similar analyses were attempted using data from C57BL/6J and C3H/HeJ mice flown on the RR-7 mission, however transcriptomic analysis revealed few differences between the spaceflight and ground control groups. For the C57BL/6J mice in RR-7, zero differentially expressed genes were identified between the 25 and 75 day spaceflight groups each compared with their corresponding ground control groups. The same comparisons carried out C3H/HeJ mouse groups identified one differentially expressed gene at each timepoint. Likewise, functional enrichment analysis of hallmarks resulted in only one significantly enriched gene set (FDR < 0.25) at 25 days and zero at 75 days for C57BL/6J, and 3 gene sets each at 25 days and 75 days for C3H/HeJ.

Comparison of principal component analyses revealed a less clear pattern of separation between groups of both the C57BL/6J and C3H/HeJ strains of mice in the RR-7 dataset compared to C57BL/6J (RR-1) and BALB/c (RR-3), (Fig. 5).

Fig. 5. Principal component analysis (PCA) plots of differential expression analysis performed on experimental and control groups of kidney transcriptomic data from **a)** RR-1 mission involving C57BL/6J mice **b)** RR-3 mission involving BALB/c mice **c)** RR-7 mission involving C57BL/6J mice and **d)** RR-7 mission involving C3H/HeJ mice. FLT = spaceflight group, GC = ground control, BSL = basal control, FLT25 = spaceflight group 25 days in space, FLT75 = spaceflight group 75 days in space, GC = ground control group 25 days, GC75 = ground control group 75 days, VIV25 = vivarium control 25 days, VIV75 = vivarium control 75

4. Discussion

Mice in both the C57BL/6J and BALB/c lineages – RR-1 and RR-3 mission, respectively - showed transcriptomic alterations associated with their exposure to space, however their responses were different. Different sets of genes were differentially expressed in each lineage, however common alterations in pathways including lipid metabolism, extracellular matrix degradation and TGF-β signalling were found. Disruption of lipid metabolism can impair extracellular matrix degradation, via increased levels of TGF-β and PAI1, leading to a build-up of extracellular matrix and promoting fibrosis [14](Zhao *et al.,* 2020). The strong expression of the *Ccl28* gene suggests an inflammatory environment in the C57BL/6J spaceflight exposed kidney as its expression is mediated by proinflammatory cytokines, and causes regulatory T cells to migrate to mucosal surfaces and increase TGF-β production [33](Eksteen *et al.,* 2006).

4.1. Lipids

Alterations in expression of genes and enrichment of pathways connected with lipid metabolism were seen in both C57BL/6J (RR-1) and BALB/c (RR-3) mice when exposed to spaceflight. Lipid storage and fatty acid metabolism were enriched in network analysis of biological processes for both C57BL/6J and BALB/c, and GSEA analysis showed enrichment of pathways related to lipid metabolism and synthesis in genes upregulated in both datasets. This could

indicate increased lipid accumulation which can cause impairment of extracellular matrix degradation [14].

Cholesterol biosynthesis and fatty acyl co-A biosynthesis were positively enriched in C57BL/6J, and negatively enriched in BALB/c, potentially affected by the *Hmgcr* gene which plays a role in cholesterol biosynthesis [34](Goldstein and Brown, 1990) which we have shown is downregulated in spaceflight exposed BALB/c. Accumulation of cholesterol increases TGF-β levels and disrupts extracellular matrix degradation through inhibition of PAI1, which controls normal degradation of the extracellular matrix [35](Proctor *et al.,* 2006). Decreased lipid synthesis may be an adaptive response against contribution to fibrotic damage in the BALB/c mice or may be indicative that they are less affected by oxidative stress.

Genes involved in lipid metabolism were differentially expressed in C57BL/6J. *Slc10a1*, *Npas2* and *Arntl*, were upregulated, while *Hmgcs2* was downregulated with a log2 fold change of -1.68. The *Hmgcs2* gene encodes the protein 3-hydroxy-3-methylglutaryl-CoA (HMG-CoA synthase) which catalyses the conversion of acetyl-CoA and acetoacetyl-CoA to HMG-CoA and CoA, a which is a rate-limiting step in ketogenesis [36](Hegardt, 1999). Impairment of ketogenesis causes hepatic injury and inflammation [37](Cotter *et al.,* 2014) and downregulation of HMG-CoA synthase has been identified as a marker for kidney stone disease in a model of induced urolithiasis in rats [38](Cao *et al.,* 2020); suggesting that the downregulation of HMG-CoA synthase in our data may be a marker for kidney injury.

Genes involved in lipid metabolism were also differentially expressed in BALB/c, including *Egr1* which was positively differentially expressed (log2 fold change 1.59), and *Hmgcr* which was negatively differentially expressed (log2 fold change -1.13). *Hmgcr* encodes HMG-CoA reductase which is an enzyme involved in the ketogenesis pathway as is *Hmgcs2* which was negatively differentially expressed in C57BL/6J. HMG-CoA reductase is also involved in the mevalonate pathway which synthesises cholesterol, catalysing the conversion of HMG-CoA to mevalonate [34](Goldstein and Brown, 1990). HMG-CoA reductase inhibitors are used to reduce the risk of death from cardiovascular disease in chronic kidney disease patients, as the inhibition of mevalonate production reduces total cholesterol and low-density lipoprotein cholesterol [39](Campese and Park, 2007). Therefore, downregulation of *Hmgcr* in BALB/c may indicate increased lipid metabolism as shown in a previous study [17](Beheshti *et al.,* 2019), which could act as a protective mechanism against kidney injury during spaceflight. In addition, *Egr1* has an upregulated differential expression in BALB/c. Interestingly, *Egr1* has been shown to play a role in lipid metabolism [40](Magee and Zhang, 2018), but also in an increase of extracellular matrix components and epithelial-mesenchymal transition, both profibrotic processes which can contribute to kidney disease [41](Bhattacharyya *et al.,* 2011). Together these results suggest that BALB/c mice exhibit an adaptive response to lipid dysfunction in spaceflight.

Maintaining homeostasis in lipid metabolism is important to maintain normal kidney function, which is essential for blood pressure and cardiovascular system regulation. Disrupted lipid metabolism can lead to chronic kidney disease and atherosclerosis, which can in turn lead to cardiovascular disease, the leading cause of death in chronic kidney disease [15].

Although physiological examination was not carried out as part of this study to confirm signs of lipid dysfunction in mouse kidneys exposed to spaceflight, previous studies have found both transcriptomic and physiological changes associated with this condition in other organs. Abnormal lipid accumulation was detected in liver tissue from mice exposed to spaceflight by Oil Red O staining, supporting the transcriptomic data on lipid dysfunction pathways in liver [17](Beheshti *et al.,* 2019). In another study, increased numbers of lipid droplets were also observed in the liver of spaceflight mice by CARS microscopy and Oil Red O staining along with upregulation of triglyceride biosynthetic pathways seen in multi-omics analysis. [42]

4.2. Extracellular matrix

Enrichment of pathways related to extracellular matrix metabolism was seen in genes upregulated in BALB/c (RR-3) and downregulated in C57BL/6J (RR-1) in response to spaceflight. In BALB/c, there was an enrichment in several hallmark pathways connected with dysregulated extracellular matrix metabolism, including Myc targets, adipogenesis and epithelial-mesenchymal transition. In the kidney, gene targets of the Myc group of proteins have been linked to an activation of glycolytic metabolism which increases production and deposition of extracellular matrix [43](Lemos *et al.,* 2018) and promotion of TGF-β signalling via transcriptional activation of integrin αv [44](Shen *et al.,* 2017). Adipogenesis and disruption of fatty acid metabolism can impair extracellular matrix degradation via intracellular accumulation of lipids and lipotoxicity [45](Kang *et al.,* 2015). Mesenchymal cells such as fibroblasts produce the components of the extracellular matrix, so increased levels due to epithelial-mesenchymal transition can cause a potentially fibrotic build-up of these components [46](Carew, Wang and Kantharidis, 2012).

Both strains exhibited differential expression in genes involved in extracellular matrix degradation. *Adamts8* was upregulated in C57BL/6J, while expression of *Ncam1* and *Aspn* were altered in BALB/c.

As previously mentioned, lipid accumulation - signs of which were observed in both strains of mice - can also lead to the impairment of normal extracellular matrix degradation. A build up in extracellular matrix components can lead to kidney disease as disrupted wound repair mechanisms are unable to restore kidney function [47] (Wynn, 2010).

4.3. TGF-B

Enrichment of pathways related to TGF-β signalling was seen in genes upregulated in BALB/c (RR-3) and downregulated in C57BL/6J (RR-1) in response to spaceflight. As previously mentioned, both strains showed increased cholesterol biosynthesis, and BALB/c exhibited enrichment in hallmark Myc targets, both of which affect TFG-β signalling. Both strains exhibited differential expression in genes involved in TGF-β signalling. This included *Ccl28,* the most differentially expressed gene in C57BL/6J (log2 fold change 2.05). In BALB/c *Fos* was upregulated (log2 fold change 1.60), while *Ccn2* and *Aspn* were downregulated. TGF-β is a cytokine which increases the production of extracellular matrix components and can lead to kidney dysfunction via glomerulosclerosis and tubulointerstitial fibrosis, leading to renal dysfunction [48](Loeffler and Wolf, 2014).

4.4. Other pathways

In the C57BL/6J mice (RR-1), the *Wnt11* gene shows downregulation in the Spaceflight group compared to the control group, with a log2 fold change of -1.15. Its expression is essential for normal development of the glomeruli, and for uretic epithelial branching in the kidney; and knockout of the gene has been shown to be lethal to mice *in utero* [49](Majumdar *et al.,* 2003). *Wnt11* deficiency in older mice has been found to result in tubular abnormalities, glomerular cysts, and interstitial fibrosis [50](Nagy *et al.,* 2016). *Wnt11* is involved in Wnt/calcium signalling and dysregulation of this pathway has been associated with cellular senescence and diseases related to ageing, including renal fibrosis [51](Hu *et al.,* 2020). Wnt signalling is involved in crosstalk with other profibrotic signalling pathways including the renin-angiotensin system, TGF-β, Notch and Hedgehog [51](Hu *et al.,* 2020). Inhibitors of Wnt signalling have been explored as treatments for fibrosis but have resulted in off-target effects [51](Hu *et al.,* 2020) perhaps in part due to the profibrotic effects of *Wnt11* downregulation.

In C57BL/6J there was enrichment in TNF-alpha signalling, which causes intense temporary inflammation and fibrosis [52](Sime *et al.,* 1998); and mTORC signalling which increases fibroblast activation and interstitial fibrosis [53](Jiang *et al.,* 2013). Interferon alpha response, interferon gamma response and JAK-STAT signalling indicate increase in adaptive immune response, which may be in response to increased inflammation caused by oxidative stress. Also enriched was angiogenesis, the formation of new blood vessels as a part of wound healing [54](Tonnesen, Feng and Clark, 2000), and overactivation of the wound healing process is a profibrotic process which can lead to kidney injury.

Both C57BL/6J and BALB/c showed enrichment of the hallmark E2F targets, which are involved in DNA repair, and BALB/c also shows enrichment of the DNA repair pathway. The DNA damage response has been found to contribute to the progression of fibrosis in systemic sclerosis [55](Vlachogiannis *et al.,* 2020). The JAK-STAT signalling pathway is also enriched in BALB/c, indicating increased inflammation in both lineages.

Clusters of biological processes related to cell cycle and immune response were found to be positively enriched in both datasets, which could represent responses to cellular damage by oxidative stress. Circadian rhythm was affected, negatively enriched in C57BL/6J and positively enriched in BALB/c. Mitochondrial gene expression and function are affected by the circadian clock [56](Aviram, Adamovich and Asher, 2021), which has also been seen to be dysregulated in astronauts [1](Afshinnekoo *et al.,* 2020). Protein re-folding pathways were negatively enriched in C57BL/6J and positively enriched in BALB/c. Oxidative stress can lead to protein unfolding, and if unfolded proteins are not re-folded or destroyed, they can accumulate and cause loss of proteostasis, which is associated with ageing [57](López-Otín *et al.,* 2013). Ribosome activity and translation initiation were also negatively enriched in C57BL/6J and positively enriched in BALB/c, which has been observed previously to be disrupted in spaceflight studies and is linked to loss of proteostasis [3](da Silveira *et al.,* 2020).

Negative enrichment in photoreceptor development and light perception was present in GSEA analysis of both datasets; many genetic diseases affect both the kidney and the retina due to the

presence of common developmental pathways [58](Savige, Ratnaike and Colville, 2011) so these enrichment patterns likely represent expression of genes common to both the retina and the kidney with different functions in each.

4.5. Strain mutations

To seek explanations for the difference in responses to spaceflight from C57BL/6J and BALB/c mice, we determined enriched pathways for the non-synonymous mutations between these lineages. Hyaluronan metabolism was the pathway found to be most enriched.

No genes involved in the metabolism of hyaluronan were differentially expressed in C57BL/6J, but one gene involved in hyaluronan metabolism, *Egf* was downregulated in BALB/c. EGF treatment in rat mesothelial tissue was shown to result in the increase of hyaluronan synthesis and epithelial-mesenchymal transition [59](Koistinen *et al.,* 2017). The hyaluronan synthesis process is linked with morphological changes such as mitosis and anchorage-independent growth seen in epithelial-mesenchymal transition, by inducing de-adhesion and budding of extracellular vesicles [59](Koistinen *et al.,* 2017). Increased EGF protein in the urine has been identified as a hallmark of renal interstitial fibrosis, and is correlated with increased transcription of the *Egf* gene [60](Ju *et al.,* 2015). Downregulation of *Egf* in the BALB/c mice could potentially confer resistance to extracellular matrix remodelling via reduced hyaluronan synthesis.

Previous studies have shown similar, milder effects on BALB/c mice compared to C57BL/6J in response to injury and stress. A study on retinal ischemia/reperfusion injury, a model for diabetic retinopathy, caused increased inflammation and damage due by oxidative stress in C57BL/6J compared to BALB/c mice [61](Shi, Ebrahim and Berger, 2018). BALB/c has also been shown to have milder reactions to UVB exposure than C57BL/6J [62](Sharma, Werth and Werth, 2011), despite their lack of protective pigmentation. In this study, higher hyaluronan levels and lower collagen levels were measured in C57BL/6J mice. Hyaluronan is a component of the extracellular matrix, plays an important role in the wound repair process, and has been shown to contribute to the development of fibrosis [63](Albeiroti, Soroosh and de la Motte, C. A., 2015). Increased metabolism of hyaluronan showed the highest enrichment in a functional enrichment analysis of the genetic differences in BALB/c compared to C57BL/6J mice.

Other genes with non-synonymous mutations between the strains were also differentially expressed in BALB/c including genes with links to lipid transport which were downregulated, and genes linked to lipid metabolism, gap junction and Wnt-protein binding were also affected. Many more genes concerning lipids were identified in the comparison with BALB/c, the majority of which were downregulated, including *Mogat1*, which is a lipid precursor which usually has high levels of expression in the kidney [64](Sankella, Garg and Agarwal, 2016).

Genes with non-synonymous mutations between the strains which were differentially expressed in C57BL/6J including alterations in expression of genes involved in cholesterol metabolism and PPAR signalling, upregulation of genes related to fatty acid metabolism, and downregulation of genes related to cell junction. *Fads* and *Elovl5* are both involved in fatty acid synthesis, potentially contributing to impairment of extracellular matrix via increasing build-up of lipids and lipotoxicity [45](Kang *et al.,* 2015). These genes are not differentially expressed in BALB/c

(RR-3), so the differences in these genes between the two mouse strains may confer some resistance to extracellular matrix remodelling. Variation in the *Hmgcs2* gene in BALB/c compared to C57BL/6J may also confer some fibrotic resistance to BALB/c as its downregulation as seen in C57BL/6J (RR-1) promotes proinflammatory ketogenesis [36](Hegardt, 1999; Cotter *et al.,* 2014). Genes related to cell junction were downregulated, potentially contributing to de-adhesion which is a part of epithelial-mesenchymal transition.

Genes which were not found to be differentially expressed in the datasets but do not possess non-synonymous mutations between the lineages may also have an effect as their function could be altered without their expression being altered. This could suggest the presence of synonymous mutations in the upstream regulators of these genes, i.e., transcription factors and non-coding RNAs and will be an area of future study.

4.6. Standardising procedures

Vastly different results were observed in RR-7 compared to RR-1 and RR-3, especially as the same strain, C57BL/6J was used in RR-1 and RR-7. We cannot currently draw any conclusions about the response of the C3H/HeJ strain to spaceflight due to the dramatic difference between the response of the C57BL/6J strain in the RR-1 and RR-7 missions.

Need to standardise kidney dissection for omics so data from different omics analyses and missions are comparable and more useful information can be obtained.

4.7. Kidney stones/countermeasures

While the C57BL/6J (RR-1) and BALB/c (RR-3) mice showed pro-fibrotic hallmarks they did not develop kidney stones, as seen in astronauts. Mice on Earth have previously proven to be poor experimental models of kidney stones [7](Okada *et al.,* 2007) but good models of renal fibrosis [8](Martínez-Klimova *et al.,* 2019) and this research indicates that the same may be true in space. Kidney stones and fibrosis can both result from metabolic disturbances, which have been seen both in this data and previous studies. Better understanding of the metabolic effects of spaceflight could lead to the development of new treatments and protective measures, and to inform risk assessments for astronauts.

Countermeasures against the increased risk of kidney stones were first suggested by [65](Cockett, Beehler and Roberts, 1962) including a physical exercise regimen and increased fluid intake for astronauts. A current countermeasure against bone loss on the ISS since 2008 is the advanced resistive exercise device, which has been shown to prevent bone mineral density loss, while increasing lean mass and decreasing fat mass. Vitamin D supplementation is necessary due to the shielding of the ISS from radiation, as this vitamin is essential for bone health and calcium metabolism [66](Smith *et al.,* 2014).

The current mitigations in place are not sufficient to bring the risk of kidney issues on long term space missions to within acceptable levels [1] (Afshinnekoo *et al.,* 2020). The long-term effects of bone loss due to extended and repeated exposure to microgravity remains an unresolved issue, and recommendations from the 2010 NASA Bone Summit included improving pre-flight risk

assessment and post-flight monitoring of bone health, continuing implementation of current countermeasures and development of pharmacological interventions [67](Orwoll *et al.,* 2013). Better understanding of lipid dysregulation in renal disease and renal fibrosis could lead to better diagnosis and treatment for patients. Drugs and treatments developed for use in astronauts could also be of benefit to people on Earth.

4.8. Conclusion

In summary, gene signatures from kidney tissue of both C57BL/6J (RR-1) and BALB/c (RR-3) mice showed differential expression consistent with remodelling of the extracellular matrix and development of fibrosis. C57BL/6J mice on the RR-1 mission showed a stronger and clearer reaction to spaceflight, while the reaction of BALB/c mice in RR-3 was more subtle.

Currently, kidney stones are considered the primary risk to the renal health of astronauts, but because of the link between the development of kidney stones and renal fibrosis, and observations of profibrotic markers in this and other studies, fibrosis should be considered as an additional serious potential risk on long term space missions. Biomarkers for risk factors for fibrosis may be used to inform pre-flight risk assessments for astronauts. Currently available antifibrotic drugs used for protection against chronic kidney disease, renin-angiotensin system blockers, mineralocorticoid receptor antagonist and sodium/glucose cotransporter 2 inhibitors [68] may also help protect astronauts. Further study into the specific mechanisms which protect BALB/c mice from gene expression changes in space, such as differences in hyaluronan metabolism, could also be a starting point to develop new countermeasures.

Limitations of Study

Mice were part of different, non-identical missions. In each mission, the studies consisted of only female mice. Analysis was not conducted on actual kidney tissue to determine if physiological changes had occurred.

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Declaration of Interests

The authors declare no competing interests.

References

[1] E. Afshinnekoo, R.T. Scott, M.J. MacKay, E. Pariset, E. Cekanaviciute, R. Barker, S. Gilroy, D. Hassane, S.M. Smith, S.R. Zwart, M. Nelman-Gonzalez, B.E. Crucian, S.A. Ponomarev, O.I. Orlov, D. Shiba, M. Muratani, M. Yamamoto, S.E. Richards, P.A. Vaishampayan, C. Meydan, J. Foox, J. Myrrhe, E. Istasse, N. Singh, K. Venkateswaran, J.A. Keune, H.E. Ray, M. Basner, J. Miller, M.H. Vitaterna, D.M. Taylor, D. Wallace, K. Rubins, S.M. Bailey, P. Grabham, S.V. Costes, C.E. Mason, A. Beheshti, Fundamental Biological Features of Spaceflight: Advancing the Field to Enable Deep-Space Exploration, Cell 183 (2020) 1162–1184. https://doi.org/10.1016/j.cell.2020.10.050.

[2] P. Pavlakou, E. Dounousi, S. Roumeliotis, T. Eleftheriadis, V. Liakopoulos, Oxidative Stress and the Kidney in the Space Environment, Int J Mol Sci 19 (2018) 3176. https://doi.org/10.3390/ijms19103176.

[3] W.A. Da Silveira, H. Fazelinia, S.B. Rosenthal, E.C. Laiakis, M.S. Kim, C. Meydan, Y. Kidane, K.S. Rathi, S.M. Smith, B. Stear, Y. Ying, Y. Zhang, J. Foox, S. Zanello, B. Crucian, D. Wang, A. Nugent, H.A. Costa, S.R. Zwart, S. Schrepfer, R.A.L. Elworth, N. Sapoval, T. Treangen, M. MacKay, N.S. Gokhale, S.M. Horner, L.N. Singh, D.C. Wallace, J.S. Willey, J.C. Schisler, R. Meller, J.T. McDonald, K.M. Fisch, G. Hardiman, D. Taylor, C.E. Mason, S.V. Costes, A. Beheshti, Comprehensive Multi-omics Analysis Reveals Mitochondrial Stress as a Central Biological Hub for Spaceflight Impact, Cell 183 (2020) 1185-1201.e20. https://doi.org/10.1016/j.cell.2020.11.002.

[4] T. Gray, A Brief History of Animals in Space, NASA Communications (1998). https://history.nasa.gov/animals.html (accessed February 10, 2022).

[5] T.S. Puri, M.I. Shakaib, A. Chang, L. Mathew, O. Olayinka, A.W.M. Minto, M. Sarav, B.K. Hack, R.J. Quigg, Chronic kidney disease induced in mice by reversible unilateral ureteral obstruction is dependent on genetic background, Am J Physiol Renal Physiol 298 (2010) F1024-1032. https://doi.org/10.1152/ajprenal.00384.2009.

[6] S.B. Gurley, S.E. Clare, K.P. Snow, A. Hu, T.W. Meyer, T.M. Coffman, Impact of genetic background on nephropathy in diabetic mice, Am J Physiol Renal Physiol 290 (2006) F214-222. https://doi.org/10.1152/ajprenal.00204.2005.

[7] A. Okada, S. Nomura, Y. Higashibata, M. Hirose, B. Gao, M. Yoshimura, Y. Itoh, T. Yasui, K. Tozawa, K. Kohri, Successful formation of calcium oxalate crystal deposition in mouse kidney by intraabdominal glyoxylate injection, Urol Res 35 (2007) 89–99. https://doi.org/10.1007/s00240-007-0082-8.

[8] E. Martínez-Klimova, O.E. Aparicio-Trejo, E. Tapia, J. Pedraza-Chaverri, Unilateral Ureteral Obstruction as a Model to Investigate Fibrosis-Attenuating Treatments, Biomolecules 9 (2019) 141. https://doi.org/10.3390/biom9040141.

[9] G. Biolo, M. Heer, M. Narici, F. Strollo, Microgravity as a model of ageing, Curr Opin Clin Nutr Metab Care 6 (2003) 31–40. https://doi.org/10.1097/00075197-200301000-00006.

[10] A. Denic, R.J. Glassock, A.D. Rule, Structural and Functional Changes With the Aging

Kidney, Adv Chronic Kidney Dis 23 (2016) 19–28.
https://doi.org/10.1053/j.ackd.2015.08.004.

[11] T.-F. Chuang, H.-C. Hung, S.-F. Li, M.-W. Lee, J.-Y. Pai, C.-T. Hung, Risk of chronic kidney disease in patients with kidney stones—a nationwide cohort study, BMC Nephrol 21 (2020) 292. https://doi.org/10.1186/s12882-020-01950-2.

[12] A.D. Rule, E.J. Bergstralh, L.J. Melton, X. Li, A.L. Weaver, J.C. Lieske, Kidney Stones and the Risk for Chronic Kidney Disease, Clinical Journal of the American Society of Nephrology 4 (2009) 804–811. https://doi.org/10.2215/CJN.05811108.

[13] S. Chaiyarit, V. Thongboonkerd, Mitochondrial Dysfunction and Kidney Stone Disease, Front Physiol 11 (2020) 566506. https://doi.org/10.3389/fphys.2020.566506.

[14] X. Zhao, J.Y.Y. Kwan, K. Yip, P.P. Liu, F.-F. Liu, Targeting metabolic dysregulation for fibrosis therapy, Nat Rev Drug Discov 19 (2020) 57–75. https://doi.org/10.1038/s41573-019-0040-5.

[15] M.C. Bulbul, T. Dagel, B. Afsar, N.N. Ulusu, M. Kuwabara, A. Covic, M. Kanbay, Disorders of Lipid Metabolism in Chronic Kidney Disease, Blood Purif 46 (2018) 144–152. https://doi.org/10.1159/000488816.

[16] J. Tian, M.J. Pecaut, J.M. Slater, D.S. Gridley, Spaceflight modulates expression of extracellular matrix, adhesion, and profibrotic molecules in mouse lung, J Appl Physiol (1985) 108 (2010) 162–171. https://doi.org/10.1152/japplphysiol.00730.2009.

[17] A. Beheshti, K. Chakravarty, H. Fogle, H. Fazelinia, W.A. da Silveira, V. Boyko, S.-H.L. Polo, A.M. Saravia-Butler, G. Hardiman, D. Taylor, J.M. Galazka, S.V. Costes, Multi-omics analysis of multiple missions to space reveal a theme of lipid dysregulation in mouse liver, Sci Rep 9 (2019) 19195. https://doi.org/10.1038/s41598-019-55869-2.

[18] A. Beheshti, S. Ray, H. Fogle, D. Berrios, S.V. Costes, A microRNA signature and TGF-β1 response were identified as the key master regulators for spaceflight response, PLoS One 13 (2018) e0199621. https://doi.org/10.1371/journal.pone.0199621.

[19] J. Galazka, R. Globus, Rodent Research-1 (RR1) NASA Validation Flight: Mouse kidney transcriptomic, proteomic, and epigenomic data, (2017). https://doi.org/10.26030/YN9M-2D19.

[20] R. Globus, J. Galazka, R. Smith, M. Cramer, Rodent Research-3-CASIS: Mouse kidney transcriptomic, proteomic, and epigenomic data, (2018). https://doi.org/10.26030/Q8VT-7P92.

[21] J.M. Galazka, S.-H. Lai Polo, A.M. Saravia-Butler, H.W. Fogle, N. Bense, M.T. Dinh, V. Boyko, S. Narang, Y.-C. Chen, S.V. Costes, S.G. Gebre, Transcriptional analysis of kidneys from mice flown on the RR-7 mission, (2019). https://doi.org/10.26030/4MX6-5X80.

[22] NASA GeneLab: Open Science for Life in Space, (2021). https://genelab.nasa.gov/ (accessed August 6, 2021).

[23] R: The R Project for Statistical Computing, (n.d.). https://www.r-project.org/ (accessed June 28, 2021).

[24] M.I. Love, W. Huber, S. Anders, Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2, Genome Biol 15 (2014) 550. https://doi.org/10.1186/s13059-014-0550-8.

[25] S. Timmermans, M. Van Montagu, C. Libert, Complete overview of protein-inactivating sequence variations in 36 sequenced mouse inbred strains, Proc Natl Acad Sci U S A 114

(2017) 9158–9163. https://doi.org/10.1073/pnas.1706168114.

[26] J. Wang, S. Vasaikar, Z. Shi, M. Greer, B. Zhang, WebGestalt 2017: a more comprehensive, powerful, flexible and interactive gene set enrichment analysis toolkit, Nucleic Acids Res 45 (2017) W130–W137. https://doi.org/10.1093/nar/gkx356.

[27] J.C. Oliveros, Venny. An interactive tool for comparing lists with Venn's diagrams, (2015). https://bioinfogp.cnb.csic.es/tools/venny/index.html (accessed October 18, 2021).

[28] G.R. Warnes, B. Bolker, L. Bonebakker, R. Gentleman, W. Huber, A. Liaw, T. Lumley, M. Maechler, A. Magnusson, S. Moeller, M. Schwartz, B. Venables, T. Galili, gplots: Various R Programming Tools for Plotting Data, (2020). https://CRAN.R-project.org/package=gplots (accessed November 1, 2021).

[29] A. Subramanian, P. Tamayo, V.K. Mootha, S. Mukherjee, B.L. Ebert, M.A. Gillette, A. Paulovich, S.L. Pomeroy, T.R. Golub, E.S. Lander, J.P. Mesirov, Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles, Proc Natl Acad Sci U S A 102 (2005) 15545–15550. https://doi.org/10.1073/pnas.0506580102.

[30] V.K. Mootha, C.M. Lindgren, K.-F. Eriksson, A. Subramanian, S. Sihag, J. Lehar, P. Puigserver, E. Carlsson, M. Ridderstråle, E. Laurila, N. Houstis, M.J. Daly, N. Patterson, J.P. Mesirov, T.R. Golub, P. Tamayo, B. Spiegelman, E.S. Lander, J.N. Hirschhorn, D. Altshuler, L.C. Groop, PGC-1alpha-responsive genes involved in oxidative phosphorylation are coordinately downregulated in human diabetes, Nat Genet 34 (2003) 267–273. https://doi.org/10.1038/ng1180.

[31] P. Shannon, A. Markiel, O. Ozier, N.S. Baliga, J.T. Wang, D. Ramage, N. Amin, B. Schwikowski, T. Ideker, Cytoscape: a software environment for integrated models of biomolecular interaction networks, Genome Res 13 (2003) 2498–2504. https://doi.org/10.1101/gr.1239303.

[32] D. Merico, R. Isserlin, O. Stueker, A. Emili, G.D. Bader, Enrichment map: a network-based method for gene-set enrichment visualization and interpretation, PLoS One 5 (2010) e13984. https://doi.org/10.1371/journal.pone.0013984.

[33] B. Eksteen, A. Miles, S.M. Curbishley, C. Tselepis, A.J. Grant, L.S.K. Walker, D.H. Adams, Epithelial Inflammation Is Associated with CCL28 Production and the Recruitment of Regulatory T Cells Expressing CCR10, The Journal of Immunology 177 (2006) 593–603. https://doi.org/10.4049/jimmunol.177.1.593.

[34] J.L. Goldstein, M.S. Brown, Regulation of the mevalonate pathway, Nature 343 (1990) 425–430. https://doi.org/10.1038/343425a0.

[35] G. Proctor, T. Jiang, M. Iwahashi, Z. Wang, J. Li, M. Levi, Regulation of renal fatty acid and cholesterol metabolism, inflammation, and fibrosis in Akita and OVE26 mice with type 1 diabetes, Diabetes 55 (2006) 2502–2509. https://doi.org/10.2337/db05-0603.

[36] F.G. Hegardt, Mitochondrial 3-hydroxy-3-methylglutaryl-CoA synthase: a control enzyme in ketogenesis, Biochem J 338 (Pt 3) (1999) 569–582.

[37] D.G. Cotter, B. Ercal, X. Huang, J.M. Leid, D.A. d'Avignon, M.J. Graham, D.J. Dietzen, E.M. Brunt, G.J. Patti, P.A. Crawford, Ketogenesis prevents diet-induced fatty liver injury and hyperglycemia, J. Clin. Invest. 124 (2014) 5175–5190. https://doi.org/10.1172/JCI76388.

[38] Y. Cao, B. Duan, X. Gao, E. Wang, Z. Dong, iTRAQ-Based Comparative Proteomics Analysis of Urolithiasis Rats Induced by Ethylene Glycol, Biomed Res Int 2020 (2020) 6137947.

https://doi.org/10.1155/2020/6137947.

[39] V.M. Campese, J. Park, HMG-CoA reductase inhibitors and the kidney, Kidney Int 71 (2007) 1215–1222. https://doi.org/10.1038/sj.ki.5002174.

[40] N.S. Magee, Y. Zhang, Hepatocyte Early Growth Response 1 (EGR1) Regulates Lipid Metabolism in Nonalcoholic Fatty Liver Disease, The FASEB Journal 32 (2018). https://doi.org/10.1096/fasebj.2018.32.1_supplement.670.56.

[41] S. Bhattacharyya, M. Wu, F. Fang, W. Tourtellotte, C. Feghali-Bostwick, J. Varga, Early growth response transcription factors: key mediators of fibrosis and novel targets for anti-fibrotic therapy, Matrix Biol 30 (2011) 235–242. https://doi.org/10.1016/j.matbio.2011.03.005.

[42] K.R. Jonscher, A. Alfonso-Garcia, J.L. Suhalim, D.J. Orlicky, E.O. Potma, V.L. Ferguson, M.L. Bouxsein, T.A. Bateman, L.S. Stodieck, M. Levi, J.E. Friedman, D.S. Gridley, M.J. Pecaut, Spaceflight Activates Lipotoxic Pathways in Mouse Liver, PLoS One 11 (2016) e0152877. https://doi.org/10.1371/journal.pone.0152877.

[43] D.R. Lemos, M. McMurdo, G. Karaca, J. Wilflingseder, I.A. Leaf, N. Gupta, T. Miyoshi, K. Susa, B.G. Johnson, K. Soliman, G. Wang, R. Morizane, J.V. Bonventre, J.S. Duffield, Interleukin-1β Activates a MYC-Dependent Metabolic Switch in Kidney Stromal Cells Necessary for Progressive Tubulointerstitial Fibrosis, JASN 29 (2018) 1690–1705. https://doi.org/10.1681/ASN.2017121283.

[44] Y. Shen, N. Miao, B. Wang, J. Xu, X. Gan, D. Xu, L. Zhou, H. Xue, W. Zhang, L. Yang, L. Lu, c-Myc promotes renal fibrosis by inducing integrin αv-mediated transforming growth factor-β signaling, Kidney Int 92 (2017) 888–899. https://doi.org/10.1016/j.kint.2017.03.006.

[45] H.M. Kang, S.H. Ahn, P. Choi, Y.-A. Ko, S.H. Han, F. Chinga, A.S.D. Park, J. Tao, K. Sharma, J. Pullman, E.P. Bottinger, I.J. Goldberg, K. Susztak, Defective fatty acid oxidation in renal tubular epithelial cells has a key role in kidney fibrosis development, Nat Med 21 (2015) 37–46. https://doi.org/10.1038/nm.3762.

[46] R.M. Carew, B. Wang, P. Kantharidis, The role of EMT in renal fibrosis, Cell Tissue Res 347 (2012) 103–116. https://doi.org/10.1007/s00441-011-1227-1.

[47] T.A. Wynn, Fibrosis under arrest, Nat Med 16 (2010) 523–525. https://doi.org/10.1038/nm0510-523.

[48] I. Loeffler, G. Wolf, Transforming growth factor-β and the progression of renal disease, Nephrol Dial Transplant 29 Suppl 1 (2014) i37–i45. https://doi.org/10.1093/ndt/gft267.

[49] A. Majumdar, S. Vainio, A. Kispert, J. McMahon, A.P. McMahon, Wnt11 and Ret/Gdnf pathways cooperate in regulating ureteric branching during metanephric kidney development, Development 130 (2003) 3175–3185. https://doi.org/10.1242/dev.00520.

[50] I.I. Nagy, Q. Xu, F. Naillat, N. Ali, I. Miinalainen, A. Samoylenko, S.J. Vainio, Impairment of Wnt11 function leads to kidney tubular abnormalities and secondary glomerular cystogenesis, BMC Dev Biol 16 (2016) 30. https://doi.org/10.1186/s12861-016-0131-z.

[51] H.-H. Hu, G. Cao, X.-Q. Wu, N.D. Vaziri, Y.-Y. Zhao, Wnt signaling pathway in aging-related tissue fibrosis and therapies, Ageing Res Rev 60 (2020) 101063. https://doi.org/10.1016/j.arr.2020.101063.

[52] P.J. Sime, R.A. Marr, D. Gauldie, Z. Xing, B.R. Hewlett, F.L. Graham, J. Gauldie, Transfer of tumor necrosis factor-alpha to rat lung induces severe pulmonary inflammation and patchy interstitial fibrogenesis with induction of transforming growth factor-beta1 and

myofibroblasts, Am J Pathol 153 (1998) 825–832. https://doi.org/10.1016/s0002-9440(10)65624-6.

[53] L. Jiang, L. Xu, J. Mao, J. Li, L. Fang, Y. Zhou, W. Liu, W. He, A.Z. Zhao, J. Yang, C. Dai, Rheb/mTORC1 Signaling Promotes Kidney Fibroblast Activation and Fibrosis, Journal of the American Society of Nephrology 24 (2013) 1114–1126. https://doi.org/10.1681/ASN.2012050476.

[54] M.G. Tonnesen, X. Feng, R.A. Clark, Angiogenesis in wound healing, J Investig Dermatol Symp Proc 5 (2000) 40–46. https://doi.org/10.1046/j.1087-0024.2000.00014.x.

[55] N.I. Vlachogiannis, M. Pappa, P.A. Ntouros, A. Nezos, C.P. Mavragani, V.L. Souliotis, P.P. Sfikakis, Association Between DNA Damage Response, Fibrosis and Type I Interferon Signature in Systemic Sclerosis, Front Immunol 11 (2020) 582401. https://doi.org/10.3389/fimmu.2020.582401.

[56] R. Aviram, Y. Adamovich, G. Asher, Circadian Organelles: Rhythms at All Scales, Cells 10 (2021) 2447. https://doi.org/10.3390/cells10092447.

[57] C. López-Otín, M.A. Blasco, L. Partridge, M. Serrano, G. Kroemer, The Hallmarks of Aging, Cell 153 (2013) 1194–1217. https://doi.org/10.1016/j.cell.2013.05.039.

[58] J. Savige, S. Ratnaike, D. Colville, Retinal abnormalities characteristic of inherited renal disease, J Am Soc Nephrol 22 (2011) 1403–1415. https://doi.org/10.1681/ASN.2010090965.

[59] V. Koistinen, K. Härkönen, R. Kärnä, U.T. Arasu, S. Oikari, K. Rilla, EMT induced by EGF and wounding activates hyaluronan synthesis machinery and EV shedding in rat primary mesothelial cells, Matrix Biol 63 (2017) 38–54. https://doi.org/10.1016/j.matbio.2016.12.007.

[60] W. Ju, V. Nair, S. Smith, L. Zhu, K. Shedden, P.X.K. Song, L.H. Mariani, F.H. Eichinger, C.C. Berthier, A. Randolph, J.Y.-C. Lai, Y. Zhou, J.J. Hawkins, M. Bitzer, M.G. Sampson, M. Thier, C. Solier, G.C. Duran-Pacheco, G. Duchateau-Nguyen, L. Essioux, B. Schott, I. Formentini, M.C. Magnone, M. Bobadilla, C.D. Cohen, S.M. Bagnasco, L. Barisoni, J. Lv, H. Zhang, H.-Y. Wang, F.C. Brosius, C.A. Gadegbeku, M. Kretzler, ERCB, C-PROBE, NEPTUNE, and PKU-IgAN Consortium, Tissue transcriptome-driven identification of epidermal growth factor as a chronic kidney disease biomarker, Sci Transl Med 7 (2015) 316ra193. https://doi.org/10.1126/scitranslmed.aac7071.

[61] H. Shi, A.S. Ebrahim, E.A. Berger, A Contrast in Pathogenic Responses between C57BL/6J and BALB/cJ Mice Using a Model of Retinal Injury, Am J Pathol 188 (2018) 2717–2728. https://doi.org/10.1016/j.ajpath.2018.08.010.

[62] M.R. Sharma, B. Werth, V.P. Werth, Animal models of acute photodamage: comparisons of anatomic, cellular and molecular responses in C57BL/6J, SKH1 and Balb/c mice, Photochem Photobiol 87 (2011) 690–698. https://doi.org/10.1111/j.1751-1097.2011.00911.x.

[63] S. Albeiroti, A. Soroosh, C.A. de la Motte, Hyaluronan's Role in Fibrosis: A Pathogenic Factor or a Passive Player?, Biomed Res Int 2015 (2015) 790203. https://doi.org/10.1155/2015/790203.

[64] S. Sankella, A. Garg, A.K. Agarwal, Characterization of the Mouse and Human Monoacylglycerol O-Acyltransferase 1 (Mogat1) Promoter in Human Kidney Proximal Tubule and Rat Liver Cells, PLoS One 11 (2016) e0162504.

https://doi.org/10.1371/journal.pone.0162504.

[65] A.T.K. Cockett, C.C. Beehler, J.E. Roberts, Astronautic Urolithiasis: A Potential Hazard During Prolonged Weightlessness in Space Travel, Journal of Urology 88 (1962) 542–544. https://doi.org/10.1016/S0022-5347(17)64841-9.

[66] S.M. Smith, S.R. Zwart, M. Heer, E.K. Hudson, L. Shackelford, J.L. Morgan, Men and women in space: bone loss and kidney stone risk after long-duration spaceflight, J Bone Miner Res 29 (2014) 1639–1645. https://doi.org/10.1002/jbmr.2185.

[67] E.S. Orwoll, R.A. Adler, S. Amin, N. Binkley, E.M. Lewiecki, S.M. Petak, S.A. Shapses, M. Sinaki, N.B. Watts, J.D. Sibonga, Skeletal health in long-duration astronauts: nature, assessment, and management recommendations from the NASA Bone Summit, J Bone Miner Res 28 (2013) 1243–1255. https://doi.org/10.1002/jbmr.1948.

[68] M. Ruiz-Ortega, S. Lamas, A. Ortiz, Antifibrotic Agents for the Management of CKD: A Review, Am J Kidney Dis 80 (2022) 251–263. https://doi.org/10.1053/j.ajkd.2021.11.010.