Research article

Dimeric ^{R25C}PTH(1-34) Activates the Parathyroid Hormone-1 Receptor *in vitro* and Stimulates Bone Formation in Osteoporotic Female Mice

Minsoo Noh[1, 2, *], Xiangguo Che[3, *], Xian Jin[3], Dong-Kyo Lee[3], Hyun-Ju Kim[3], Doo Ri Park[4], Soo Young Lee[4], Hunsang Lee[2], Thomas Gardella[5], Je-Yong Choi[3, #], Sihoon Lee[1, #]

[1]Department of Internal Medicine and Laboratory of Molecular Endocrinology, Gachon University School of Medicine, Incheon 21565, Republic of Korea

[2]Department of Life Sciences, Korea University, Seoul 02841, Korea

[3]Department of Biochemistry and Cell Biology, Cell and Matrix Research Institute, School of Medicine, Kyungpook National University, Daegu 41944, Republic of Korea.

[4]Department of Life Sciences, Multitasking Macrophage Research Center, Ewha Womans University, Seoul 03760, Korea

[5]Endocrine Unit, Massachusetts General Hospital and Harvard Medical School, Boston, MA 02114, USA

*These two authors are equally contributed to this manuscript.

[#]Correspondence:

Je-Yong Choi D.D.S., Ph.D., Department of Biochemistry and Cell Biology, Cell and Matrix Research Institute, School of Medicine, Kyungpook National University, Daegu 41944, Republic of Korea. Tel.: +82-53-420-4823; Fax: +82-53-422-1466; E-mail: jechoi@knu.ac.kr

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Sihoon Lee, M.D., Ph.D., Department of Internal Medicine and Laboratory of Molecular Endocrinology, Gachon University School of Medicine, Incheon 21565, Republic of Korea. Tel.: +82-32-458-2646; Fax: +82-32-460-2381; E-mail: shleemd@gachon.ac.kr

Abstract

Osteoporosis, characterized by reduced bone density and strength, increases fracture risk, pain, and limits mobility. Established therapies of Parathyroid hormone (PTH) analogs effectively promote bone formation and reduce fractures in severe osteoporosis, their use is limited by potential adverse effects. In the pursuit of safer osteoporosis treatments, we investigated ^{R25C}PTH, a PTH variant wherein the native arginine at position 25 is substituted by cysteine. These studies were prompted by our finding of high bone mineral density in a hypoparathyroidism patient with the R25C homozygous mutation, we explored its effects on PTH type-1 receptor (PTH1R) signaling in cells and bone metabolism in mice. Our findings indicate that ^{R25C}PTH(1-84) forms dimers both intracellularly and extracellularly, and the synthetic dimeric peptide, ^{R25C}PTH(1-34), exhibiting altered activity in PTH1R-mediated cAMP response. Upon a single injection in mice, dimeric ^{R25C}PTH(1-34) induced acute calcemic and phosphaturic responses comparable to PTH(1-34). Furthermore, repeated daily injections increased calvarial bone thickness in intact mice and improved trabecular and cortical bone parameters in ovariectomized (OVX) mice, akin to PTH(1-34). The overall results reveal a surprising capacity of a dimeric PTH peptide ligand to activate the PTH1R *in vitro* and *in vivo*, suggesting a potential new path of therapeutic PTH analog development.

Key words: Parathyroid hormone, PTH type 1 receptor, Bone formation, Osteoporosis

Introduction

Osteoporosis is a prevalent global bone disorder characterized by low bone mineral density (BMD), causing weakened bones prone leading to fragility fractures, particularly in the spine, hip, and wrist. The development of osteoporosis is influenced by factors including gender, being more prevalent in women; hormonal changes, like decreased estrogen levels during menopause; and age, with heightened susceptibility post-menopause in women contributing to bone loss. Recent meta-analysis of previous studies indicates a global osteoporosis prevalence of 23.1 % among women and 11.7 % among men (1, 2). Osteoporosis stands as a noteworthy risk factor that poses challenges to the preservation of autonomous mobility and overall well-being within an aging society. There is thus a pressing need for safe and efficacious therapies for osteoporosis that mitigate fractures, alleviate associated symptoms, and preserve physical functionality.

Anti-resorptive agents (e.g., bisphosphonates, denosumab, and romosozumab) encompass one therapeutic approach that aims to specifically counteract the declines in bone mass by tempering the balance between bone resorption and formation (3-5). It is pertinent to acknowledge, however, that the prolonged use of most of such agents is limited due to potential long-term side effects. Furthermore, most anti-resorptive therapies cannot stimulate new bone formation. In contrast, bone anabolic agents, such as parathyroid hormone (PTH) and its analogs, such as teriparatide (recombinant human PTH(1-34)), increase BMD by stimulating bone formation more than bone resorption (6). PTH has an exceptionally short half-life in the blood of approximately 2-4 minutes (7, 8), which helps in avoiding excessive increases in blood calcium levels that can otherwise limit the utility of PTH-related medications, while yet inducing a desired anabolic effect on bone. It is also worth noting that studies in rodents reveal that long-term administration of a PTH anabolic agent can lead to bone overgrowth, osteosarcoma, as well as hypercalcemia (9, 10). Consequently, a goal of ongoing research is to uncover the underlying molecular mechanisms driving the anabolic and catabolic effects of PTH to thereby secure more effective therapeutic alternatives for osteoporosis (11).

PTH is produced and secreted by the parathyroid glands as a straight-chain monomeric polypeptide of 84 amino acids (12-14) . It plays a vital role in maintaining calcium and phosphate equilibrium by acting on the PTH1R, a class B G protein-coupled receptor (GPCR) family (15-17) that is expressed primarily in cells of bone and kidney (18). The orchestrated downstream effects of PTH

in target cells act to ensure optimal bone health and the maintenance of the ambient blood calcium and phosphate concentrations required for proper nerve conduction, muscle activity, and systemic cellular communication, whereas disturbances in this system can lead to multiple disorders.

Central to the PTH signaling cascade is the activation of intracellular G proteins (19), most prominently Gsα, which in turn activates adenylyl cyclase leading to the synthesis of the second messenger cyclic AMP (cAMP), and the activation of cAMP-dependent protein kinase A (PKA) (20, 21) . PTH can also activate other second messenger cascades, including the Gαq/phospholipase C (PLC) / inositol 1,4,5-trisphosphate (IP3), diacylglycerol (DAG), protein kinase C (PKC) signaling pathway (22, 23), highlighting the diverse biology of PTH and the PTH1R (24). Adding to this, the PTH1R also mediates the actions of parathyroid hormone-related protein (PTHrP), a development protein that acts in the formation of bones and other tissues. PTHrP shares homology with PTH in the first 34 amino acids, which encompass the receptor-binding portions of the two respective ligands. The PTH1R thus has an intrinsic capacity for dual ligand recognition, which opens possibilities for exploring new modes of therapeutic development for diseases such as osteoporosis and hypothyroidism (18, 19, 25-28). Pharmacologically, the PTH1R can adopt at least two distinct ligand-binding conformations, RG and R^0, the selectivity for which can lead to altered modes of signaling *in vitro* and *in vivo* for peptides such as PTH, PTHrP, and various hybrid analogs (29-33).

The current study extends our prior investigation in which we identified in a patient with chronic hypocalcemia and hyperphosphatemia a mutation that changes the arginine at position 25 in the mature PTH(1-84) polypeptide to cysteine (^{R25C}PTH) (34, 35). Antibody assays revealed the ^{R25C}PTH mutant protein to be present in the patient's blood at markedly elevated levels. We now have found that this patient expressing the ^{R25C}PTH variant has higher-than-normal BMD. We further characterize the ^{R25C}PTH protein and find that it can manifest in two distinct molecular forms: as a monomer and as a dimer, and we demonstrate that a dimeric ^{R25C}PTH(1-34) synthetic peptide retains agonistic properties on the PTH1R that are driven by a moderate selectivity for the RG vs. R^0 receptor conformation. Finally, we demonstrate in mice that ^{R25C}PTH(1-34) can induce skeletal responses that are similar to those induced by PTH(1-34), but without triggering an excessive hypercalcemic response. Considering the proven bone-anabolic capacity of several established PTH agonist ligands, and the need for safe, long-term treatments for skeletal disorders, our studies on dimeric ^{R25C}PTH(1-

34) suggest alternative strategies to consider in such drug development programs.

Results

Dimerization of ^{R25C}PTH(1-84)

In our previous studies, we showed that synthetic monomeric peptide, ^{R25C}PTH(1-34), as compared to PTH(1-34), exhibits a moderately diminished PTH1R-binding affinity and decreased cAMP signaling potency *in vitro*, and that with long-term infusion in mice, ^{R25C}PTH(1-34) leads to only minimal calcemic and phosphaturic effects, which corroborates the hypocalcemia seen in the original patient, despite the significantly elevated levels of ^{R25C}PTH in the plasma (34). On subsequent analysis of this patient, we found a particularly high BMD (Supplementary File 1), which prompted us to further characterize the functional properties of ^{R25C}PTH, as described herein. We produced recombinant PTH(1-84) with or without the R25C mutation by expression of the corresponding cDNA in HEK293T cells (Figure 1). We considered the possibility that the introduction of a sole new cysteine within the polypeptide chain of ^{R25C}PTH(1-84) might induce homologous bimolecular dimerization through a disulfide bond involving the thiol functional group in each monomer (36, 37). To specifically investigate this, we designed the cDNA constructs to express either pre-pro-PTH(1-115)-3xFLAG or ^{R56C}pre-pro-PTH(1-115)-3xFLAG, such that after intracellular processing and cleavage of the pre-pro regions, the mature PTH(1-84)-3xFLAG or ^{R25C}PTH(1-84)-3xFLAG peptides would be generated upon transfection in HEK293T cells (Figure 1C) (38-40). We performed western blot analysis of total cell lysates and conditioned culture media collected from the transfected cells to specifically assess the possible presence of a disulfide-bonded dimeric form. Each sample was thus prepared in either reduced or non-reduced form and proteins were detected using an anti-flag antibody. The results demonstrated the presence of both a low molecular weight monomeric, and in the non-reduced samples, a higher molecular weight dimeric form of the ^{R25C}PTH(1-84)-3XFLAG protein in both the cell lysate and extracellular conditioned medium fractions, and the dimer appeared to be at an elevated proportion in the medium relative to the lysate (Figure 1D). To control for potential artifact effects attributed to the 3xFLAG tag, we utilized plasmid constructs pcDNA3.0-(pre-pro-PTH)-IRES and pcDNA3.0-(^{R56C}pre-pro-PTH)-IRES encoding non-tagged PTH variants and an anti-PTH(39-84) antibody for Western blot analysis, which again confirmed the presence of the dimer in total HEK293T cell lysates (Supplementary File 2).

We observed in the above studies that the expression level of ^{R25C}PTH(1-84) was higher than that of wild-type PTH(1-84) in both cell lysate and medium, which we considered might be due to an intrinsic enhancement in protein stability and resistance to protein degradation in the dimeric molecule. To address this, we treated the cells with the proteasome inhibitor MG132, which acts by forming a hemiacetal with the hydroxyl groups of active site threonine residues, and compared the expression levels of wild-type PTH(1-84) and ^{R25C}PTH(1-84) in the treated vs. untreated cells. The results indicated that while the expression level of wild-type PTH was increased by MG132 treatment, it did not reach the level of ^{R25C}PTH, suggesting that the difference in expression is not related to a difference in sensitivity to proteasome-mediated degradation (Supplementary File 3).

Overall, we have confirmed that ^{R25C}PTH(1-84) can form a dimeric structure, and the ^{R25C}PTH(1-84) secreted outside the cells predominantly exists in dimeric form. Thus, utilizing dimer ^{R25C}PTH(1-84) in the analysis would be more relevant to understanding the actual function of ^{R25C}PTH. Consequently, we aim to conduct further validation using dimeric ^{R25C}PTH in our subsequent investigations.

Functional characterization of dimeric ^{R25C}PTH(1-34) *in vitro*

To explore the functional properties of dimeric ^{R25C}PTH, we conducted experiments using synthetic peptides of PTH(1-34), ^{R25C}PTH(1-34) (monomeric) and disulfide-bonded dimeric ^{R25C}PTH(1-34). First, we examined the receptor-binding affinity of these ligands by performing competition experiments using membranes prepared from HEK293-derived GP-2.3 cells that stably express the human PTH1R and assay formats designed to assess binding to either the G protein-uncoupled R^0 or G protein-coupled RG receptor conformation. Each experiment was replicated four times (n = 4). The results revealed that monomeric ^{R25C}PTH(1-34) bound to both the R^0 and RG conformations with comparable, albeit slightly weaker affinity as compared to PTH(1-34), while dimeric ^{R25C}PTH(1-34) bound to each conformation with weaker affinity than did the monomeric form while showing an apparent selectivity for higher binding to the RG vs R^0 conformation of PTH1R (Figure 2A).

To investigate the signaling properties of the ligands, we measured the changes increases in intracellular levels of cAMP induced by each ligand in an osteoblastic SaOS-2 derived cell line (SGS-

72 cells) that stably expresses the Glosensor cAMP reporter. Each measurement was replicated four times (n = 4). These assays revealed that each ligand dose-dependently increased the cAMP levels in the cells, detected as an increase in luminescence in an Envision plate reader, and while the potencies were moderately and more substantially reduced for the monomeric and dimeric forms of the ligand, respectively, as compared to PTH(1-34), the maximum response attained by each ligand were comparable (Figure 2B). Dimeric ^{R25C}PTH(1-34) thus retains signaling functionality at the PTH1R that is characterized by a potency approximately commensurate with its affinity for binding to the RG PTH1R conformation.

Effect of single injection of dimeric ^{R25C}PTH(1-34) on calcium and phosphate regulation in mice

To assess whether dimeric ^{R25C}PTH can function *in vivo*, we injected the ligand, and in parallel either vehicle or PTH(1-34) (each peptide at a dose of 50 nmol/kg) into CD1 female mice and measured levels of ionized calcium (Ca^{2+}) in the blood (n = 6 mice/group), inorganic phosphate (Pi) in plasma (n = 12 mice/group), and the excretion of Pi into urine (n = 6 mice/group). Blood Ca^{2+} levels were measured at serial time points of pre-injection, 1, 2, 4, and 6 hours post-injection. Both PTH(1-34) and dimeric ^{R25C}PTH(1-34) induced increases in blood Ca^{2+} levels that were significant, relative to the levels in vehicle-injected mice, at 1 and 2 hours post-injection, and the levels then returned to the baseline levels of vehicle control mice by 4 hours (Figure 3A). Plasma Pi levels were measured in samples obtained pre-injection, at 6 minutes, and 1, 2, and 6 hours post-injection. PTH(1-34) induced a significant decrease in plasma Pi at 1-hour post-injection, and the levels subsequently returned to baseline by 2 hours. Injection of dimeric ^{R25C}PTH(1-34) resulted in a slight decrease in plasma Pi at 2 hours post-injection, but the effect was not significant (Figure 3B). Consistent with this trend, Pi levels in the urine of mice injected with dimeric ^{R25C}PTH(1-34) were increased significantly at 2 hours post-injection and then gradually returned to baseline levels (Figure 3C). These results thus indicate that dimeric ^{R25C}PTH can elicit calcemic and phosphaturic responses *in vivo* that are fully with those expected for an injected PTH1R agonist ligand. We did not detect an increase in plasma cAMP in response to dimeric ^{R25C}PTH(1-34) injected at a dose of either 50 nmol/kg (n = 12) or 100 nmol/kg (n = 2), whereas a significant increase was observed at 15 minutes post-injection of PTH(1-34) at a dose of 50 nmol/kg (Figure 4A and B). This result likely reflects at least in part a reduced cAMP-stimulating

potency of dimeric ^{R25C}PTH(1-34) ligand, relative to PTH(1-34), as revealed by our cell-based Glosensor assays, but other possibilities, such as a reduced stability of the dimeric ligand in the blood, might also account for the weaker cAMP response to the dimeric peptide *in vivo*.

Effect of dimeric ^{R25C}PTH(1-34) on bone calvariae in mice

To initially assess the effects that short-term treatment of dimeric ^{R25C}PTH(1-34) can have on bone, we injected 8-week-old male C57BL/6 mice once a day for six days (days 1-6), with either dimeric ^{R25C}PTH(1-34) (160 µg/kg/day), PTH(1-34) (80 µg/kg/day) or vehicle (N = 6 for group), and after 10 days without treatment (day 16) followed by euthanasia, we isolated the calvariae for histological analysis of new bone formation. Specifically, we examined sections stained with hematoxylin and eosin (H&E) to assess the width of newly formed bone areas along the edge of each sample. These regions exhibited a more vivid coloration compared to the surrounding existing bone tissue, demarcated by a dotted line for clarity (Figure 5A). Measurements were taken below and above the dissection area where new bone had formed, and these measurements were then utilized to calculate the mean values for further analysis. These analyses revealed that both PTH(1-34) and dimeric ^{R25C}PTH(1-34) significantly increased the width of the new bone area by approximately four-fold, as compared to the vehicle group, and the effects of two ligands were not significantly different from each other (Figure 5B). These findings thus support a capacity of dimeric ^{R25C}PTH(1-34) to induce new bone formation *in vivo*, which is of interest given the high BMD observed in the patient with the ^{R25C}PTH mutation, despite the presumably continuous exposure of the bone to the mutant ligand.

Effect of dimeric ^{R25C}PTH on bone mass in osteoporotic mice

To more directly assess the impact of dimeric ^{R25C}PTH(1-34) on bone mass, we administered it to ovariectomized (OVX) mice, which serve as a well-established model for postmenopausal osteoporosis. The OVX mice were injected daily for a duration of 4 weeks with either the dimeric ligand (OVX + dimeric ^{R25C}PTH(1-34), PTH(1-34) (OVX + PTH(1-34)) or vehicle (OVX + vehicle, OVX-controls), and sham-operated (Sham) mice were used as further controls. Mice were euthanized at

the end of the injection period and tissue samples were isolated for analysis. Quantitative micro-computed tomography (μ-CT) analysis of the femurs obtained from each group revealed that, as compared to OVX + vehicle controls, treatment with PTH(1–34) increased femoral trabecular bone volume fraction (Tb.BV/TV) by 121%, cortical bone volume fraction (Ct.BV/TV) by 128%, cortical thickness (Ct.Th) by 115%, and cortical area fraction (Ct.Ar/Tt.Ar) by 118% (Figure 6A). Treatment with dimeric ^{R25C}PTH(1-34) resulted in similar effects on the femoral cortical bone parameters, as it increased Ct.BMD by 104%, Ct.BV/TV by 125%, Ct.Th by 107%, and Ct.Ar/Tt.Ar by 116% (Figure 6B). The increase in cortical bone BMD was significant with dimeric ^{R25C}PTH(1-34) but not with PTH(1-34), whereas an increase in femoral trabecular bone was only observed with PTH(1-34).

The effects of the treatments on bone biomechanical properties were assessed by conducting a three-point bending test on femurs isolated from the mice. The maximum load parameter was significantly decreased in the femurs from OVX-control versus sham mice, and was significantly increased, relative to the OVX-controls, by treatment with the PTH(1-34) but not dimeric ^{R25C}PTH(1-34) (Figure 6C). The slope parameter was significantly decreased in the femurs from OVX-control versus sham mice, and tended to increase versus OVX-controls by treatment with either PTH(1-34) or ^{R25C}PTH(1-34) but the changes were not significant.

We further analyzed the levels of bone metabolism markers in the serum obtained from the mice at the study endpoint (Figure 6D). The levels of serum calcium were within the normal range in all treatment groups, while serum phosphate levels were modestly increased in the OVX mice treated with PTH(1-34) or dimeric ^{R25C}PTH(1-34) as compared to with vehicle, but the effect was significant only with PTH(1-34) . The serum levels of CTX1, a bone resorption marker (41), were elevated in each of the OVX groups versus the sham group, and tended to be lower in the OVX-PTH(1-34) treatment group but the change relative to OVX-vehicle was not significant (Figure 6D). Serum P1NP and alkaline phosphatase levels were each significantly increased in both the PTH(1-34)- and dimeric ^{R25C}PTH(1-34)-treated groups, compared to the OVX-vehicle group, consistent with an enhancing effect of the ligands on bone formation (42, 43). Histological staining of proximal tibial sections for tartrate-resistant acid phosphatase (TRAP) activity, a marker of osteoclast-mediated bone resorption, revealed an apparent increase in this activity in bones of the OVX mice, as compared to those in sham control mice, reflecting a heightened rate of bone turnover, as also suggested by the increased

levels of serum CTX1 in the OVX mice, and the TRAP staining appeared to be reduced in the tibiae of the OVX mice treated with PTH(1-34) (Figure 6D and E). Further histomorphometric analysis confirmed a significant increase in the osteoclast surface area relative to bone surface area (Oc.S/BS) in the proximal tibiae of the OVX-vehicle mice, relative to that in the Sham-control mice, and this parameter was significantly decreased by treatment with PTH(1-34) but not dimeric ^{R25C}PTH(1-34) (Figure 6F).

We then analyzed the bone microstructure in the lumbar vertebrae through von Kossa staining of histological sections and histomorphometric quantification (Figure 7A). The trabecular bone volume fraction (TB; V, %), and trabecular number (Tb N) were significantly reduced in the OVX-vehicle group, as compared to the Sham group, and treatment with either PTH(1-34) or dimeric ^{R25C}PTH(1-34) resulted in a significant increase in each of these parameters, as well as a concomitant reduction in trabecular separation (Tb Sp), as compared to the respective parameters in the OVX-vehicle group (Figure 7B).

Dynamic bone histomorphometry was also performed on the vertebrae to evaluate rates of bone formation (Figure 7C). The trabecular mineral apposition rate (MAR) and cortical MAR were each significantly increased in both the OVX-PTH(1-34) and OVX-dimeric ^{R25C}PTH(1-34) groups, as compared to in the OVX-vehicle group, and although there was a tendency for an increase in the bone formation rate (BFR/BS) in both the trabecular and cortical bone with either PTH(1-34) or dimeric ^{R25C}PTH(1-34) treatment, the differences were not statistically significant, as compared to the OVX control (Figure 7D).

In summary, injection of dimeric ^{R25C}PTH into osteoporotic OVX mice resulted in significant increases in cortical bone in the femurs and trabecular bone in the vertebrae, as well as significant increases in the trabecular MAR and serum markers of the bone formation markers, ALP and PINP, without inducing excessive bone resorption or hypercalcemia.

Discussion

In this study, we show the introduction of a cysteine mutation at the 25th amino acid position of mature parathyroid hormone (PTH) facilitates the formation of homodimers comprised of the resulting ^{R25C}PTH peptide. This dimerization surprisingly was compatible with receptor binding affinity, and lead to relatively minor deviations in functional behavior as assessed in our cell-based assays and compared to the standard monomeric control PTH peptide. The homozygous ^{R25C}PTH mutation was identified in patients that presented with hypocalcemia and hyperphosphatemia, despite elevated PTH levels (34, 44), and the mutation was found to impact the bioactive region of PTH (34, 45-48). Our initial research on this ^{R25C}PTH mutant focused on the monomeric state of the peptide, and these studies revealed relatively moderate decreases in PTH1R binding affinity and cAMP stimulating potency *in vitro* and moderately impaired calcemic effects upon infusion in mice. Additional patient observations, however, revealed the patients to have a higher BMD than anticipated for age-matched averages. This elevated bone mass, coupled with the elevated serum PTH levels, prompted our further investigations into the properties of the mutant PTH, as described herein.

Our investigations brought to light the capacity of the cysteine-25 mutation to induce dimer formation in the otherwise monomeric PTH polypeptide as produced in transfected cells. This result was established by comparing western blots of transfected cell lysates analyzed under reducing versus non-reducing conditions of gel electrophoresis. Subsequently, we employed synthetic peptides to further explore the functional properties of dimeric ^{R25C}PTH(1-34). The results of our current studies show some divergence from our previous findings obtained using the monomeric counterpart, ^{R25C}PTH (1-34). Compared to the monomer, dimeric ^{R25C}PTH(1-34) exhibited a more preferential binding affinity for the RG versus R^0 PTH1R conformation, despite a diminished affinity for either conformation. We also observed that the potency of cAMP production in cells was lower for dimeric ^{R25C}PTH as compared to the monomeric ^{R25C}PTH, in accordance with a lower PTH1R-binding affinity.

We further pursued *in vivo* applications in mice. Initially, we assessed the calcemic and phosphatemic responses to a single injection of synthetic peptides of either PTH(1-34) or dimeric ^{R25C}PTH(1-34) in intact mice. We found the dimer could induce increases in plasma calcium levels that were at least as robust and as sustained as those induced by PTH(1-34), and was similarly in phosphaturic (Figure 3). Intriguingly, we did not detect an increase in plasma cAMP levels upon

injection of dimeric ^{R25C}PTH, whereas PTH(1-34) injection resulted in the expected rapid and transient increase in blood cAMP (Figure 4). Determining whether this difference is due to a difference in the capacity of the two ligands to activate the presumably renal pool of PTH receptors that mediate the blood cAMP response to PTH, a difference in the pharmacokinetic properties of the ligands, or some other such effects will require further investigation.

Activation of the canonical Gas-cAMP-PKA signaling pathway is generally thought to underly most of the biological responses induced by PTH1R activation, and our studies in SAOS2-derived osteoblastic cells confirm that dimeric PTH can activate this pathway, albeit not as efficiently as a monomeric PTH peptide. Arg25 resides in the 20-34 (C-terminal region) of PTH(1-34), which plays a significant role in the binding of the ligand to the extracellular domain (ECD) of the PTH1R. In concert with the binding of the 20-24 region of PTH to the ECD, the N-terminal portion of PTH engages the transmembrane domain (TMD) of the PTH1R to then induce the conformational changes involved in G protein coupling and cAMP production (33, 49-52). The precise binding mode used by dimeric ^{R25C}PTH to the PTH1R is unknown, but it may be anticipated that it differs to some extent from that used by the monomeric peptide, due, for example, to the changes in bulk molecular size and display of accessible functional groups. Consequently, the receptor conformational changes and the modes of coupling to downstream effectors may differ for the monomeric versus dimeric ligands, which could potentially lead to altered signaling and biological responses *in vivo*. Whether such changes account for the increased bone density observed in the patient with the homozygous ^{R25C}PTH mutation is unknown but cannot be presently ruled out.

The results of practical assessments of dimeric ^{R25C}PTH(1-34) for effects on calvarial bone after short-term (6-days) injection into normal mice, and for effects on bone mass after long-term (4-weeks) daily injection in osteoporotic OVX mice demonstrate the dimer can mediate significant influences on bone metabolism. Moreover, the results of these studies in OVX mice provided evidence to suggest that the effects of dimeric PTH may favor bone anabolism versus bone catabolism more effectively than a monomeric PTH peptide. While further investigation is necessary, the current experimental data strongly imply that the dimeric form of ^{R25C}PTH can serve as a new peptide with distinct functionalities compared to the wild-type PTH.

Furthermore, the recent identification of a young patient in Denmark displaying homozygous ^{R25C}PTH has opened avenues for observing the direct impacts of ^{R25C}PTH within the human biological system (44). The continual monitoring and observation of patients will contribute to a more profound comprehension of the long-term consequences associated with ^{R25C}PTH exposure. This extensive observation is crucial in delineating the extended effects of this compound on individuals. Consequently, by conducting thorough investigations to confirm the potential bone anabolic effect of ^{R25C}PTH, we hope to develop a novel bone anabolic agent with a targeted focus on the PTH1R.

Materials and Methods

Plasmid construction

The coding sequences (CDS) of pre-pro-PTH and the mutated form, ^{R56C}pre-pro-PTH[^{R25C}PTH], were amplified using primers containing the attB site. These CDS fragments were obtained from pcDNA3.0-(hpre-pro-PTH)-IRES and pcDNA3.0-(h^{R56C}pre-pro-PTH)-IRES, which were used in the previous research conducted by Lee*, et al.* (34). Both CDS were introduced into donor vector pDONR223 with Gateway™ BP Clonase™ II Enzyme mix kit (Invitrogen, USA). Then pre-pro-PTH and ^{R56C}pre-pro-PTH were each cloned into pcDNA3.1-ccdB-3xFLAG-V5 with LR Gateway™ LR Clonase™ II Enzyme mix (Invitrogen, USA) to construct pcDNA3.1-(pre-pro-PTH)-3xFLAG-V5, and pcDNA3.1-(^{R56C}pre-pro-PTH) -3xFLAG-V5. All experimental procedures were done with the manufacturer's instruction. pDONR223 was a gift from Kim Lab (Roswell Park Comprehensive Cancer Center, USA), and pcDNA3.1-ccdB-3xFLAG-V5 was a gift from Taipale Lab (Donnelly Centre, University of Toronto, Canada).

Cell culture

All cell lines were grown at 37°C in 5 % CO_2. HEK293T cells were cultured in Dulbecco's Modified Eagle Medium (Cytiva™, HyClone DMEM/High glucose with L-Glutamine and HEPES, Cat No. SH30243.01, USA) with 10 % FBS (Gibco™, Fetal Bovine Serum, certified, Origin: USA, Cat No. 16000044, Lot No. 2522247RP, USA) and 1 % penicillin-streptomycin (Cytiva™, HyClone Penicillin-Streptomycin 100X solution, Cat No. SV30010, USA). Cell lines obtained from commercial sources were not further authenticated.

Transfection

pcDNA3.1-(pre-pro-PTH)-3xFLAG-V5 and pcDNA3.1-(^{R56C}pre-pro-PTH)-3xFLAG-V5 were each transfected into HEK293T with Lipofectamine™ 3000 Transfection Reagent (Invitrogen™, Cat No. L3000001, USA) according to the manufacturer's instruction. After 48 hours of transfection, a culture medium was collected and used for western blot as a secreted protein sample. The rest of the

cells were lysed by RIPA buffer (Thermo Scientific™, RIPA Lysis and Extraction Buffer, Cat No. 89900, USA) following the manufacturer's instruction and used for western blot as total cell lysate sample.

Western blot

Protein samples were prepared in two types, reduced sample and non-reduced sample. Reduced samples were a mixture of protein (secreted protein or cell lysate), sample buffer (Invitrogen™, NuPAGE™ LDS Sample Buffer (4X), Cat No. NP0007, USA), with reducing agent (Invitrogen™, 10X Bolt™ Sample Reducing Agent, Cat No. B0009, USA) and heated for 5 min at 95℃. Non-reduced samples were a mixture of protein, and sample buffer, without reducing agent, and not heated. The protein samples were loaded onto 4 - 12 % Bis-Tris protein gels (GeneSTAR, StarPAGE Bis-Tris 4-12 %/15well, Cat No. GPG4115), and ran with MOPS/SDS running buffer (GeneSTAR, 20X MOPS / SDS Running Buffer, Cat No. GMB0080). Transfer to the membrane was done by iBlot™ 2 Dry Blotting System (Invitrogen™, iBlot™ 2 Gel Transfer Device, Cat No. IB21001, USA) with PVDF transfer stack (Invitrogen™, iBlot™ 2 Transfer Stacks-PVDF-mini, Cat No. IB24002, USA) according to manufacturer's instruction. The membranes were blocked for 1 hour at room temperature (RT) in 5 % skim-milk solution in Tris-buffered saline (TBS; 20 mM Tris-base, 500 mM NaCl, pH 7.5) and then washed three times for 10 min each with tris-buffered saline with 0.05 % tween-20 (TBST). The membranes were incubated with primary antibody for 1 hour at RT, then washed three times for 10 min each with TBST. If needed, the membranes were incubated with horseradish peroxidase (HRP)-conjugated secondary antibody for 1 hour at RT after primary antibody incubation, then washed three times for 10 min each with TBST. After washing, the membranes were rinsed and soaked in TBS. To develop a blot image, the membranes were treated with a chemiluminescent substrate solution (Merck Millipore, Immobilon ECL Ultra Western HRP Substrate, Cat No. WBULS0500, USA) according to the manufacturer's instruction. The blot images were obtained by LAS 4000 mini (Cytiva, ImageQuant™ LAS 4000 mini, USA). The dilution condition for the anti-FLAG with hHRP conjugated antibody (Sigma-Aldrich, Monoclonal ANTI-FLAG® M2-Peroxidase (HRP) antibody produced in mouse, Cat No. A8592, USA) was 1:2,000. The dilution condition for the anti-HSP90 primary antibody (Santa Cruz Biotechnology Inc., HSP 90 Antibody (AC-16), Cat No. sc-

101494, USA) was 1:5,000. The dilution condition for the anti-mouse secondary antibody was 1:5,000. Each antibody was diluted in TBST with 1 % BSA solution.

Proteasome inhibition assay

HEK293T cells were seeded in culture dishes at approximately 60 % confluence, and they were allowed to grow for about 20 to 24 hours prior to transfection. The transfection of pcDNA3.1-(pre-pro-PTH)-3xFLAG-V5 and pcDNA3.1-(^{R56C}pre-pro-PTH)-3xFLAG-V5 was conducted following the method mentioned earlier. After 24 hours of transfection, MG132, dissolved in DMSO, was added to the cells to achieve a final concentration of 10 μM. For the mock treatment, DMSO alone was added. The cells were then incubated for an additional 24 hours after MG132 treatment. Both the culture medium and cell lysate were prepared for western blot analysis to assess the restored protein levels. The western blot procedure was carried out as described in the previous section.

Peptide synthesis and quantification

Human PTH(1-34), ^{R25C}PTH(1-34), and dimeric ^{R25C}PTH(1-34) were chemically synthesized by Anygen (Gwangju, Republic of Korea). The purity and mass of each peptide were analyzed by high-performance liquid chromatography (HPLC) and matrix-assisted laser desorption ionization-time of flight mass spectrometry (MALDI-TOF MS) in the manufacturer.

PTH1R competition binding assay

The binding of PTH and its analogs to G protein-uncoupled PTH1R (R^0 conformation) and G protein-coupled PTH1R (RG conformation) was assessed using a competition method with membranes prepared from GP-2.3 cells (HEK-293 cells stably expressing the hPTH1R). For tracer radioligands, we utilized ^{125}I-PTH(1-34) and ^{125}I-MPTH(1-15). The unlabeled ligands tested were PTH(1–34), ^{R25C}PTH(1-34), and dimeric ^{R25C}PTH(1-34). Binding to the R^0 conformation was assessed using ^{125}I-PTH(1-34) as the tracer radioligand, while binding to the RG conformation was assessed

using ^{125}I-MPTH(1-15). The addition of unlabeled ligands PTH(1–34), ^{R25C}PTH(1-34), or dimeric ^{R25C}PTH(1-34) caused dissociation of the tracer radioligand from each receptor if it had affinity to the receptors. Measurement of the dissociated ratio of the radioligand indicated the binding affinity between PTH1R and each unlabeled ligand.

cAMP assay

To measure intracellular cAMP production, SGS-72 cells, derived from SaOS-2 cells and stably expressing the Glosensor cAMP reporter, were utilized to measure intracellular cAMP production. The detection of cAMP-dependent expression was performed using an Envision plate reader (PerkinElmer, Waltham, MA, USA), based on luciferase-based luminescence, as previously described by Maeda*, et al.* (53) .

Animal model used in the study

CD1 female mice were purchased from Charles River Laboratories (Massachusetts, USA), and all animal care and experimental procedures were conducted under the guidelines set by the Institutional Animal Care and Use Committee (IACUC) at Massachusetts General Hospital (MGH). The mice were housed in a specific pathogen-free environment, with 4-5 mice per cage, under a 12-hour light cycle at a temperature of 22±2°C.

C57BL/6N female mice were purchased from KOATECH (Gyeonggi-do, Republic of Korea), and all animal care and experimental procedures were conducted under the guidelines set by the Institutional Animal Care and Use Committees of Kyungpook National University (KNU-2021-0101). The mice were housed in a specific pathogen-free environment, with 4-5 mice per cage, under a 12-hour light cycle at a temperature of 22 ± 2°C. They were provided with standard rodent chow and water ad libitum.

C57BL6 female mice aged 8-10 weeks were used to establish an ovariectomizing (OVX) mouse model of osteoporosis. The mice were divided into the following four groups (n = 6 mice/group) as follows: sham, OVX control group, OVX + PTH (1-34) treated group (40 μg/kg/day), and OVX +

dimeric ^{R25C}PTH treated group (40-80 µg/kg/day). The OVX mice were allowed to recover for 4 weeks. Subsequently, PTH (1-34) and ^{R25C}PTH were subcutaneously administered 5 days per week for 4 weeks. After the treatment period, the mice were sacrificed, and µ-CT and histological analyses were conducted.

Acute injections

The peptides PTH(1-34) and dimeric ^{R25C}PTH(1-34) were diluted in a solution comprising 0.05% Tween 80, 10 mM citric acid, and 150 mM NaCl at a pH of 5.0. Intravenous injections of these peptides were administered at doses ranging from 50-100 µg/kg into 9-week-old CD1 female mice. As a control, mice received only the vehicle. Plasma cAMP levels were assessed before and after peptide or vehicle injection at 15-minute intervals, following the methodology detailed by Maeda*, et al.* (53). Blood-ionized calcium, plasma phosphate, and urine phosphate levels were measured before and after the injection at 1-hour intervals.

Calvarial injection mouse model

C57BL/6 male mice (8-week-old) were divided into the following three groups (n = 6 mice/group): control, PTH (1-34) treated group (80 µg/kg/day), and ^{R25C}PTH treated group (160 µg/kg/day). Subcutaneous injections of the respective drugs were administered once daily for 6 days. On the sixteenth day, the mice were sacrificed, and their bone tissues were harvested and fixed in 10 % formaldehyde at 4°C. The fixed bone tissues were then decalcified in PBS (pH 7.4) containing 0.5 moles of ethylenediaminetetraacetic acid (EDTA). Following decalcification, the tissues were embedded in paraffin, and paraffinized tissues were sectioned to a thickness of 5-7 µm. Histological analysis was performed using the sectioned tissue slides stained with hematoxylin and eosin (H&E). The area of new bone formation, which displays a more intense coloration compared to the existing bone tissue, was examined.

µ-CT analysis

Mouse femurs were fixed in a 4 % paraformaldehyde solution for 24 hours at 4°C. In μ-CT, we used the Quantum FX μ-CT (Perkin Elmer, Waltham, MA, USA). The images were acquired at a 9.7 μm voxel resolution, with settings of 90 kV and 200 μA, a 10 mm field of view, and a 3-minute exposure time. Serial cross-sectional images were reconstructed using the Analyze 12.0 software (Overland Park, KS, USA). To ensure consistent analysis, identical regions of interest (ROIs) were selected for the trabecular and cortical bones. The ROIs were positioned 0.3 mm from the bottom of the growth plate. All bone parameters were evaluated according to the guidelines of the American Society for Bone and Mineral Research (54).

Three-point bending test

The left femur of the mouse was immersed in 0.9 % NaCl solution, wrapped in gauze, and stored at −20°C until ready for a three-point bending test. In this test, we placed the mouse femurs on a suitable mold and set the pressure sensor at a distance that allowed for the maximum allowable pressure without interfering with the test (20.0 mm for the femur). A miniature material testing machine (Instron, MA, USA) was used for this test. The crosshead speed decreased at 1 mm/min. During the test, force-displacement data were collected to determine the maximum load and slope of the bones.

Serum biochemistry analysis

Serum bone resorption and osteogenesis marker levels, specifically the C-terminal telopeptide of type I collagen (CTX) and procollagen type I N-terminal propeptide (P1NP), were assessed in mice from the sham, OVX-control, PTH(1-34), and dimeric ^{R25C}PTH(1-34) groups by using the enzyme-linked immunosorbent assay (ELISA) according to the manufacturer's instructions. Additionally, their concentrations were determined using specific mouse CTX-1 and P1NP ELISA kits (Cloud Clone, Wuhan, China) respectively.

Bone histological analyses

The tibiae were initially fixed in 4% paraformaldehyde at 4°C overnight. The following day, samples were decalcified using 10% ethylenediaminetetraacetic acid (EDTA) solution (pH 7.4) for 4

weeks at 4°C. The decalcified tibiae were then embedded in paraffin and sectioned at 3 μm thick. For TRAP staining, dehydrated paraffin sections were fixed in an acetone/ethanol mixture (1:1) for 1 minute, followed by complete air-drying at RT for 20 minutes. Thereafter, the sections were immersed in TRAP reagent for 30 minutes at 37°C. Osteoclast surface per bone surface (Oc.S/BS) and Osteoclast number per bone surface (Oc.N/BS) analysis followed the ASBMR guidelines (55).

Dynamic bone histomorphometric analysis

To conduct dynamic histomorphometry analysis, we injected the mice with 25 mg/kg body weight of calcein (Sigma-Aldrich) or Alizarin Red S (Sigma-Aldrich) intraperitoneally before sacrifice, with 3- or 10-day intervals between injections. Briefly, femurs or vertebrates were fixed in 4 % paraformaldehyde solution for 24 hours at 4°C. The following day, the samples were washed with phosphate-buffered saline (PBS) solution and then dehydrated using a gradient of ethanol (50 %, 70 %, 85 %, 90 %, and 100 %). Subsequently, we embedded the dehydrated femurs or vertebrates in methyl methacrylate (Sigma) to prepare resin blocks. The resin blocks were sectioned at 6 μm thick by using a Leica SP1600 microtome (Leica Microsystems, Germany). The fluorescence signals of calcein (green) and Alizarin Red S (red) from ROIs were captured using a fluorescence microscope (Leica, Wetzlar, Germany). For vertebral bone analysis, bone mineralization was evaluated by von Kossa staining. The sections were placed in 2-methoxyethyl acetate (Sigma-Aldrich) for 20 minutes, followed by rehydration with serial ethanol solutions (100 %, 90 %, 80 %, 70 %, and 50 %) and distilled water for 2 minutes each. The sections were subsequently dipped in a 1 % $AgNO_3$ solution (Sigma-Aldrich) for 5 minutes under ultraviolet (UV) light photons, washed in distilled water for 5 minutes, and dipped in 5 % sodium thiosulfate solution for 5 minutes to remove nonspecific binding. Finally, we covered the sections with mounting solution and captured images by using a Leica microscope. The parameters of dynamic bone histomorphometry were analyzed using the Bioquant Osteo 2019ME program (Bioquant Osteo, Nashville, TN, USA).

Statistical analysis

Statistical analysis was performed in GraphPad Prism 10.1.2. The data are presented as the mean ± standard error of the means (SEM). Statistically significant differences between the two groups were determined using an unpaired *t*-test. A *p*-value less than 0.05 was considered statistically significant.

Author contributions

Conceptualization, Methodology: M.N., X.C., X.J., D-K.L., H-J.K., D.P., S.Y.L., H.L., T.J.G., J.-Y.C., and S.L.; Validation, Formal Analysis: M.N., and X.C.; Investigation, Resource: M.N., X.C., X.J., D.-K.L., and D.P.; writing - Original Draft Preparation: M.N., and X.C.; Writing – Review & Editing: M.N., X.C., T.J.G., H.L., J.-Y.C., and S.L.; Visualization: M.N., and X.C.; Supervision: H.-J.K., H.L., J.-Y.C., and S.L.; Project Administration: J.-Y.C., and S.L.; Funding Acquisition: S.Y.L., H.L., J.-Y.C., and S.L.

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Figures and Tables

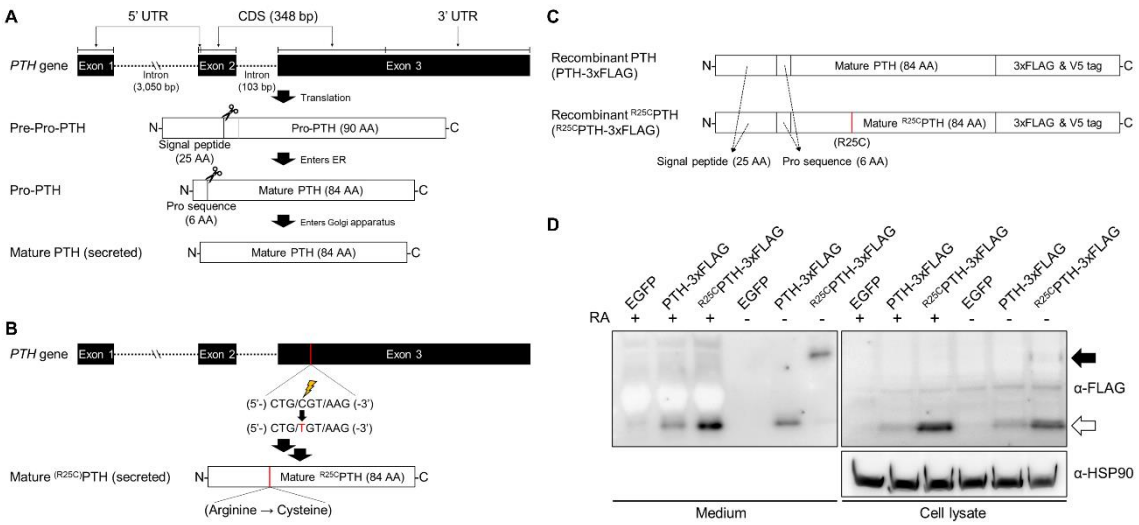


Figure 1. Formation of R25C mutant PTH(1-84) dimer.

(A) Schematic representation of PTH gene structure and expression. (B) Schematic representation of ^{R56C}pre-pro-PTH(1-115) (In mature form, ^{R25C}PTH(1-84)) gene structure and expression (C) Schematic representation of recombinant PTH proteins (D) *In vitro* dimerization of ^{R25C}PTH. Recombinant protein constructs were transfected into HEK293T cells, and expression of PTH-3xFLAG and ^{R25C}PTH3xFLAG in culture medium or cell lysate was demonstrated by western blot. The result confirms the presence of dimeric R25CPTH. (*bp: base pairs; *AA: amino acids; *RA: reducing agent)

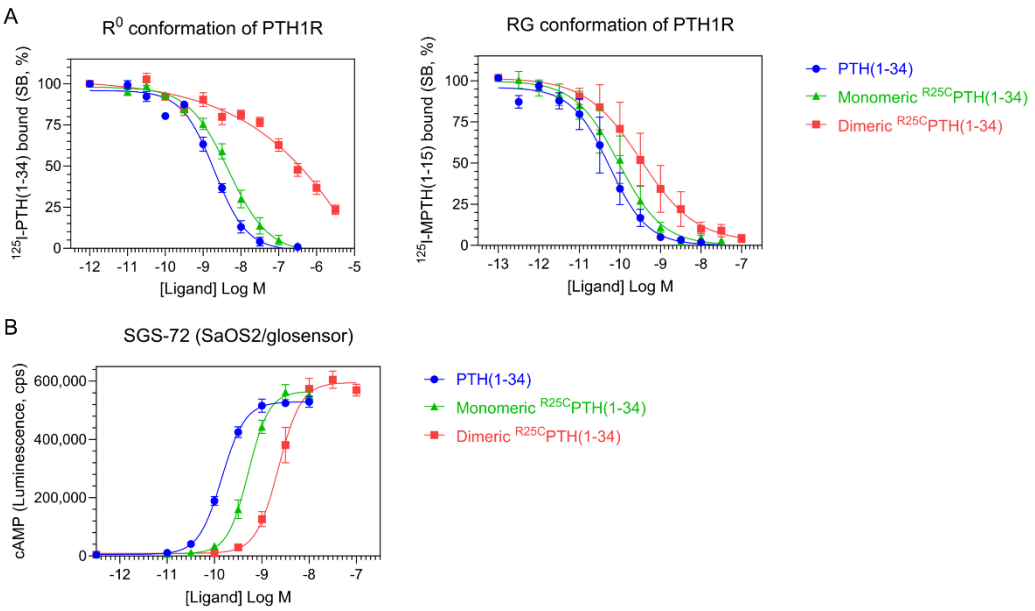


Figure 2. Effect of PTH, monomeric ^{R25C}PTH, and dimeric ^{R25C}PTH to the PTH1R *in vitro*.

(A) The binding of PTH(1-34), monomeric ^{R25C}PTH(1-34), and dimeric ^{R25C}PTH(1-34) to the PTH1R in R^0 conformation of RG conformation was assessed by competition methods using ^{125}I-PTH(1-34) and ^{125}I-MPTH(1-15) as radioligand. (B) Ligand potency for cAMP signaling was assessed in SGS-72 cells, which were derived from SaOS2 cells modified to express Glosensor cAMP reporter. The cells were preloaded with luciferin and treated with varying concentrations of PTH(1-34), monomeric ^{R25C}PTH(1-34), and dimeric ^{R25C}PTH(1-34).

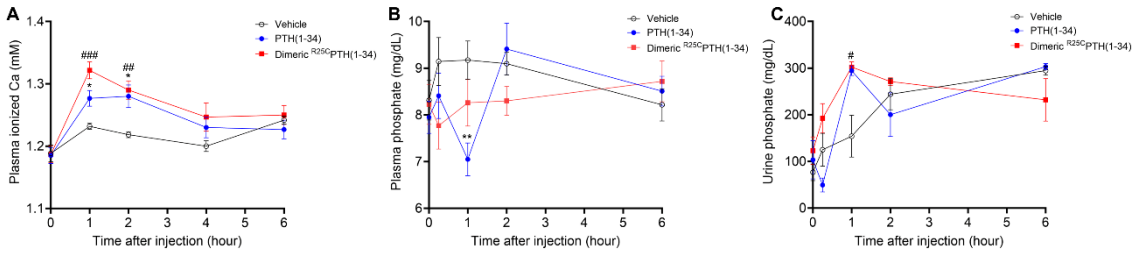


Figure 3. Calcemic and phosphatemic responses by PTH injection in CD1 female mice.

(A) Plasma Calcemic Response after Injection (n = 6). Both PTH(1-34) and dimeric ^{R25C}PTH(1-34) significantly elevate ionized calcium levels in plasma at 1 to 2 hours post-injection. After 2 hours post-injection, plasma ionized calcium level gradually restored to baseline levels similar to those of the vehicle group. (B) Plasma Phosphatemic Response after Injection (n = 12). Following PTH(1-34) injection, plasma phosphate levels significantly decrease at 1-hour post-injection, subsequently returning to baseline akin to those of the vehicle group. Conversely, dimeric ^{R25C}PTH(1-34) injection shows no significant alteration in phosphatemic response but demonstrates a tendency towards a slight decrease in phosphate levels, gradually restoring to baseline levels akin to those of the vehicle group. (C) Urine Phosphatemic Response after Injection (n = 6). The urine phosphate levels markedly increased at 1 hour post-injection for both PTH(1-34) and dimeric ^{R25C}PTH(1-34), followed by a return to baseline levels akin to those of the vehicle group. This analysis was conducted using 9-week-old female CD1 mice. The mice were administered PTH(1-34) and dimeric ^{R25C}PTH(1-34) at a concentration of 50 nmol/kg for each compound. Error bars represent mean ± standard error. *p*-values were determined using the *t*-test. * denotes *p*-value < 0.05 for PTH(1-34) compared to vehicle, ** denotes *p*-value < 0.01 for PTH(1-34) compared to vehicle, # denotes *p*-value < 0.05 for dimeric ^{R25C}PTH(1-34) compared to vehicle, ## denotes *p*-value < 0.01 for dimeric ^{R25C}PTH(1-34) compared to vehicle, ### denotes *p*-value < 0.001 for dimeric ^{R25C}PTH(1-34) compared to vehicle.

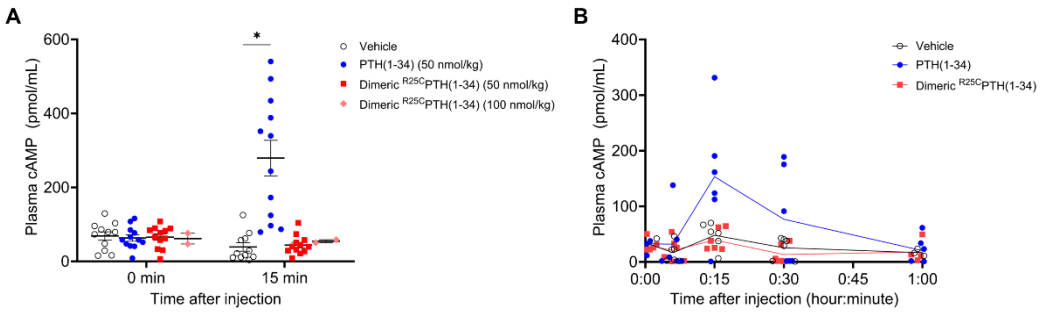


Figure 4. Effect of cAMP production by PTH injection in CD1 female mice.

(A) Comparison of cAMP levels induced following the injection of PTH(1-34) (50 nmol/kg) and two concentrations (50, 100 nmol/kg) of dimeric ^{R25C}PTH(1-34). cAMP levels were assessed both before and 15 minutes after injection, confirming induced cAMP production exclusively by PTH(1-34). (B) To compare the amount of cAMP generated after injecting PTH(1-34) and two concentrations of dimeric ^{R25C}PTH(1-34) more precisely, we measured cAMP levels in a group of 6 mice (n = 6) before injection and at 6, 15, 30, and 60 minutes after injection. We observed that cAMP production was induced only by PTH(1-34), and at the 15-minute time point, the cAMP levels reached their peak and gradually decreased thereafter. Female CD1 mice at 9 weeks old were used for each analysis. The error bars indicate mean ± standard error. *p*-values were obtained using the *t*-test. * indicates *p*-value < 0.05 against vehicle.

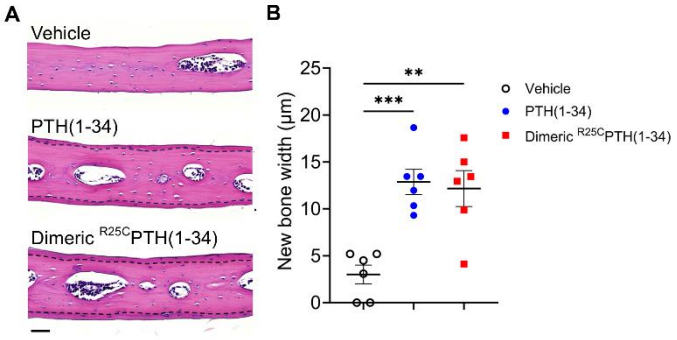


Figure 5. Effect of dimeric ^{R25C}PTH(1-34) in calvarial injection model

(A) Dissections of the calvarial bones. Calvarial injections were performed on eight-week-old male C57BL/6 mice (N = 6 per group) that received daily administrations of vehicle, PTH(1-34), or dimeric ^{R25C}PTH(1-34) for six days. Following a 10-day treatment period, histological sections of calvariae, stained with hematoxylin (pink; representing bone matrix) and eosin (blue-purple; indicating cell nuclei), were obtained. The area of new bone formation, with more intense staining compared to the existing bone tissue, is denoted by the dotted line. The bar indicates 50 µm. (B) Quantification of new bone width in calvarial injection model. The result showed a significant increase in new bone width following injections of both PTH(1-34) and dimeric ^{R25C}PTH(1-34) compared to the vehicle group. ** indicates *p*-value <0.01 against vehicle, *** indicates *p*-value <0.001 against vehicle.

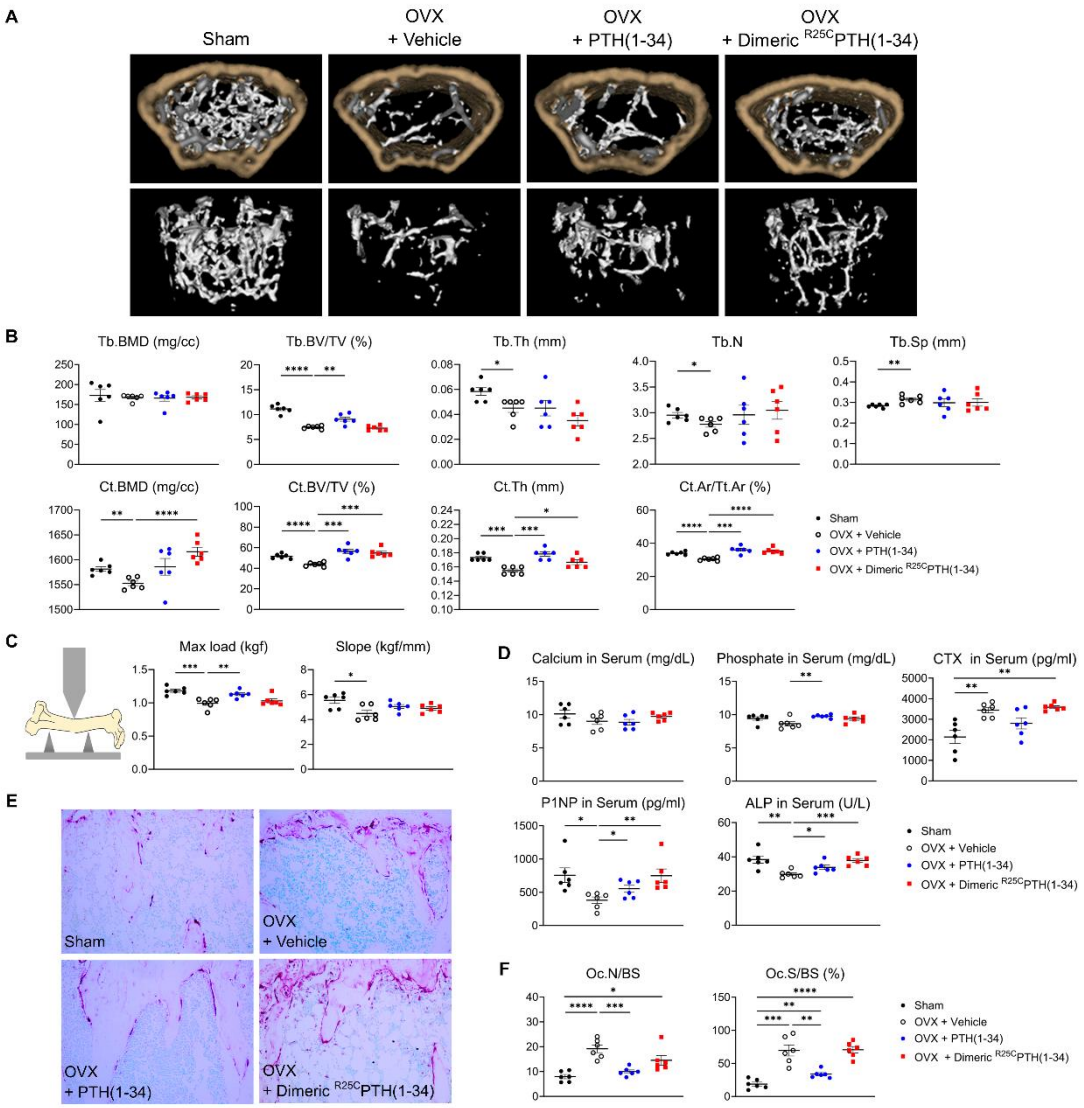


Figure 6. Impact of ^{R25C}PTH(1-34) on Bone Turnover.

The effects of Sham, OVX-Control (OVX + vehicle), OVX treated with PTH(1-34) (OVX + PTH(1-34)), and OVX treated with dimeric ^{R25C}PTH(1-34) (OVX + dimeric ^{R25C}PTH(1-34)) on bone turnover in mice. (A) Femurs obtained from mice in each group were subjected to µCT analyses for the assessment of bone mass. (B) Several parameters of (A) were quantified using µCT measurements, including trabecular bone mineral density (Tb.BMD), trabecular bone volume to tissue volume (Tb.BV/TV), trabecular bone thickness (Tb.Th), trabecular number (Tb.N), trabecular separation (Tb.Sp), cortical bone mineral density (Ct.BMD), cortical bone volume to tissue volume (Ct.BV/TV), cortical thickness (Ct.Th), and cortical area to total tissue area (Ct.Ar/Tt.Ar). (C) A 3D-point bending test was conducted with femurs obtained from mice in each group. The left panel describes a schematic model of a 3D-

point bending test. The middle and right panels each indicate the maximum bending load (kgf) and slope (kgf/mm). (D) Serum levels of calcium, phosphorus, CTX, P1NP, and ALP were measured for each group using an ELISA assay. (E) TRAP staining of histological sections of proximal tibias was carried out to visualize osteoclast activity. The scale bars represent 100 μm (F) Quantification of osteoclast number per bone surface (Oc.N/BS), and osteoclast surface per bone surface (Oc.S/BS) was performed. Each group consisted of six samples (n = 6). The error bars indicate mean ± standard error. *p*-values were obtained using the *t*-test to compare the mean of each column with the mean of a control column. * indicates p-value < 0.05, ** indicates *p*-value < 0.01, *** indicates *p*-value < 0.001, **** indicates *p*-value < 0.0001.

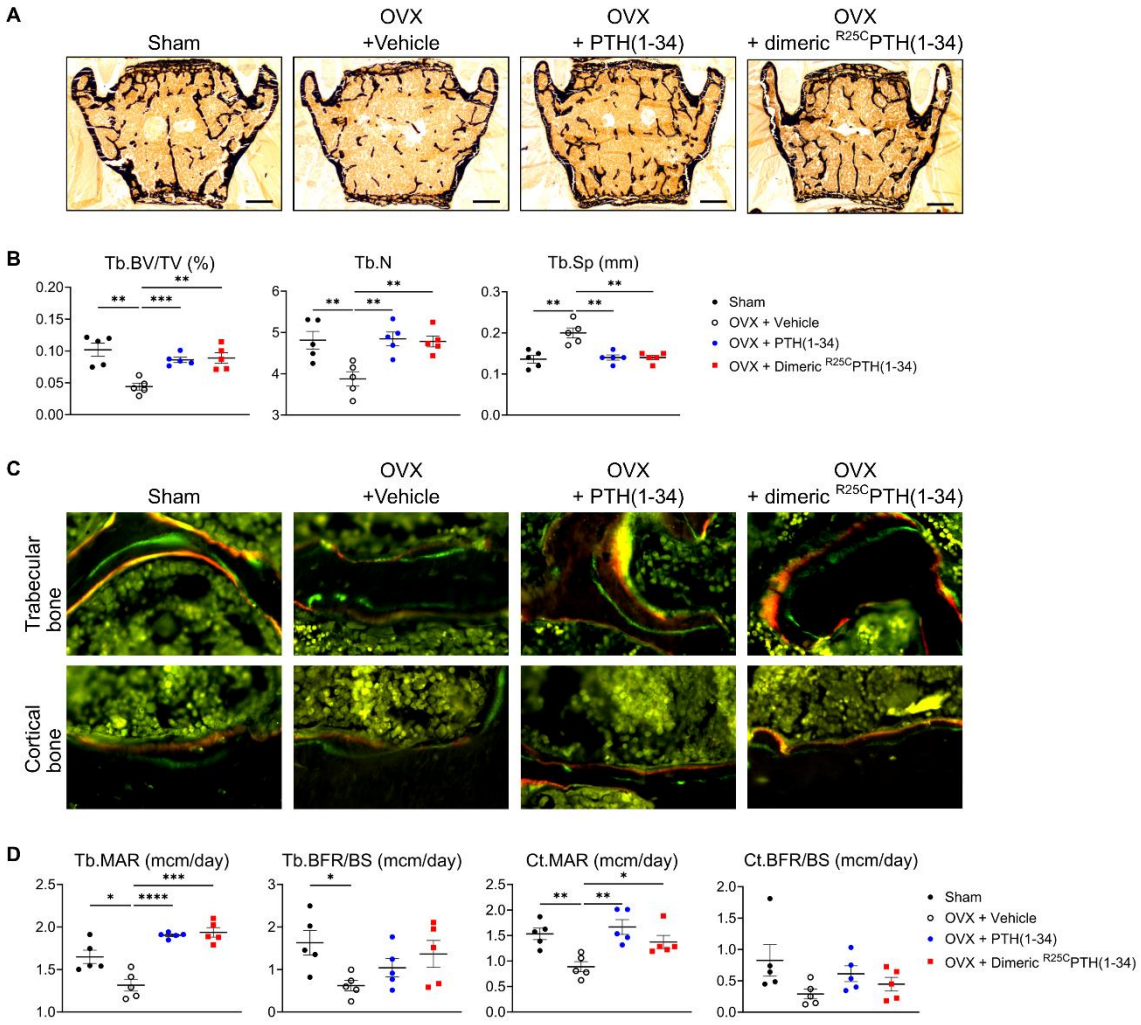
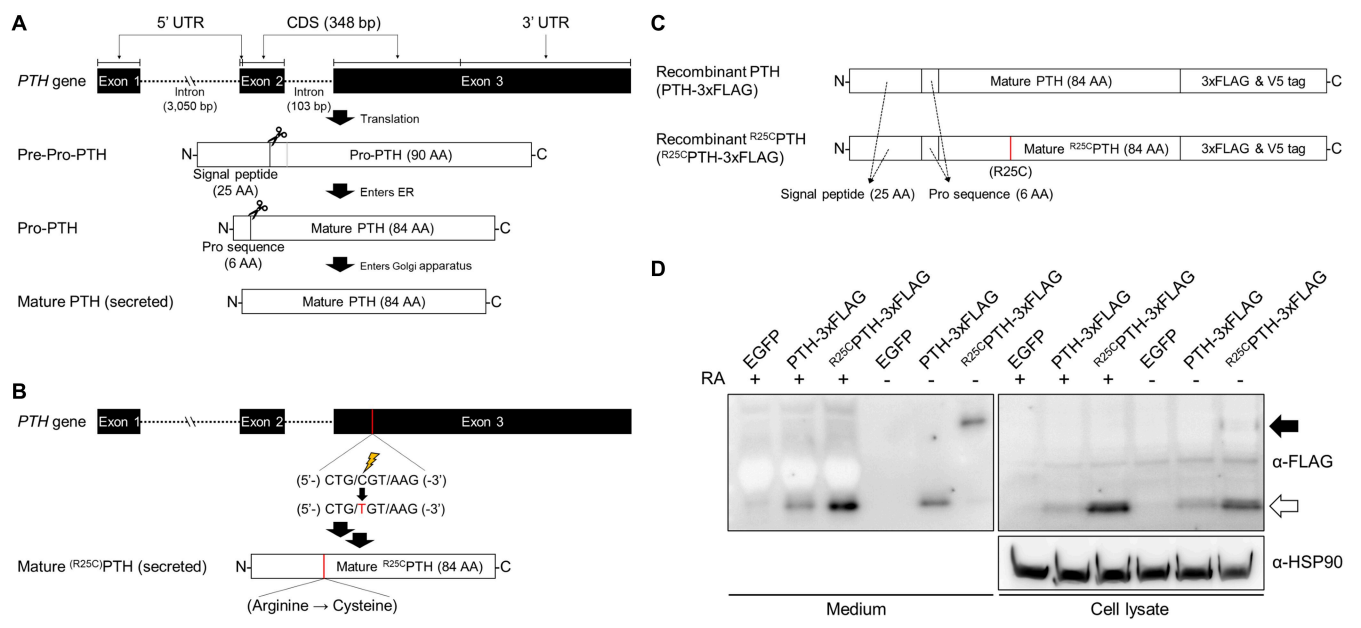
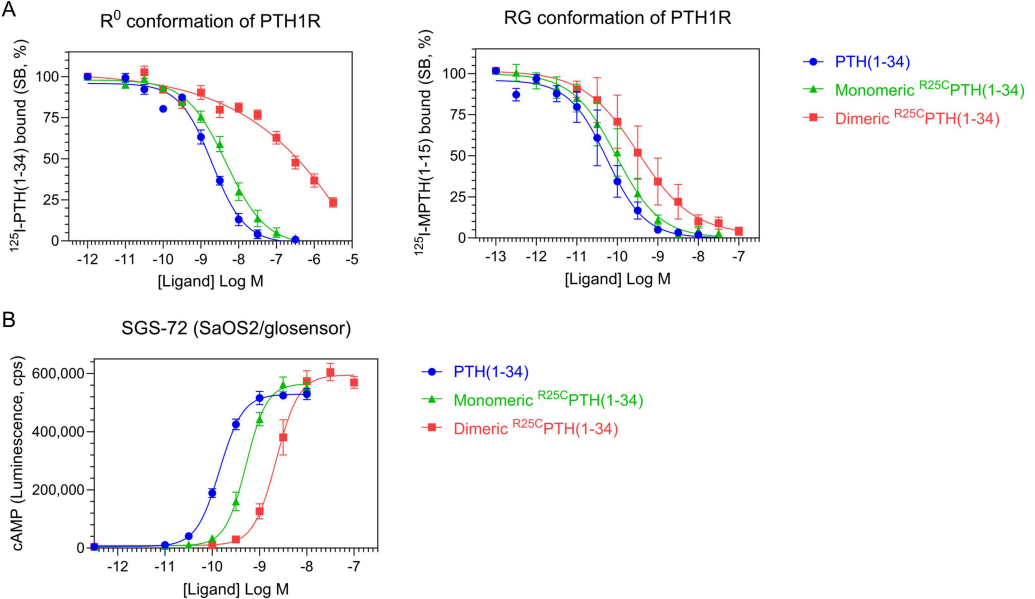


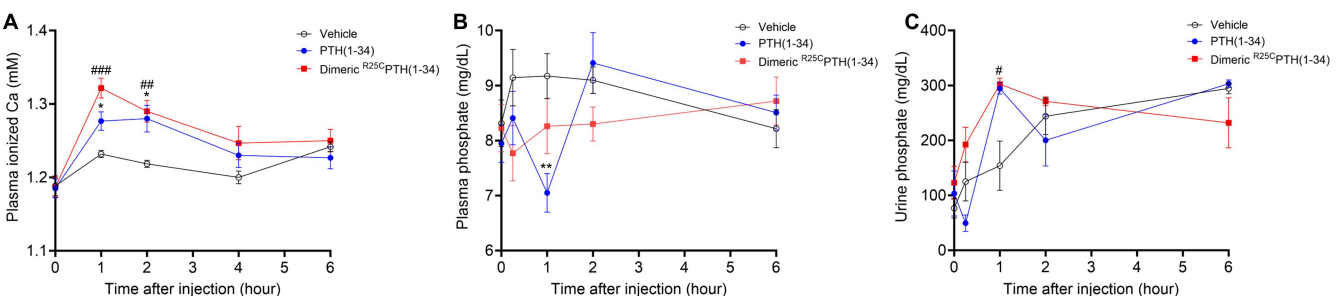
Figure 7. Impact of ^{R25C}PTH(1-34) on Osteoblast Function in Vertebrae

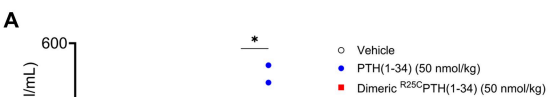
The effects of Sham, OVX-Control (OVX + vehicle), OVX treated with PTH(1-34) (OVX + PTH(1-34)), and OVX treated with dimeric ^{R25C}PTH(1-34) (OVX + dimeric ^{R25C}PTH(1-34)) on osteoblast function in mice. (A) Mineralization of vertebrae obtained from each group was assessed through Von Kossa staining. (B) Quantification of trabecular bone parameters including trabecular bone volume to tissue volume (Tb.BV/TV), trabecular number (Tb.N), and trabecular separation (Tb.Sp) was performed using the Bioquant Osteo 2019 v19.9.60 program. Each bar represents 500 µm. (C) Fluorescent microscopic observations of trabecular and cortical bone sections from each group demonstrate the apposition of xylenol (red) and calcein (green) labels. (D) Quantification of trabecular bone parameters such as trabecular bone mineral apposition rate (Tb.MAR), trabecular bone formation rate to bone surface (Tb.BFR/BS), cortical bone MAR (Ct.MAR), and trabecular MAR (Tb.MAR) was carried out using the Bioquant Osteo 2019 v19.9.60 program. Each group consisted of five samples

(n = 5). The error bars indicate mean ± standard error. *p*-values were obtained using the *t*-test compare the mean of each column with the mean of a control column. * indicates *p*-value < 0.05, ** indicates *p*-value < 0.01, *** indicates *p*-value < 0.001, **** indicates *p*-value < 0.0001.

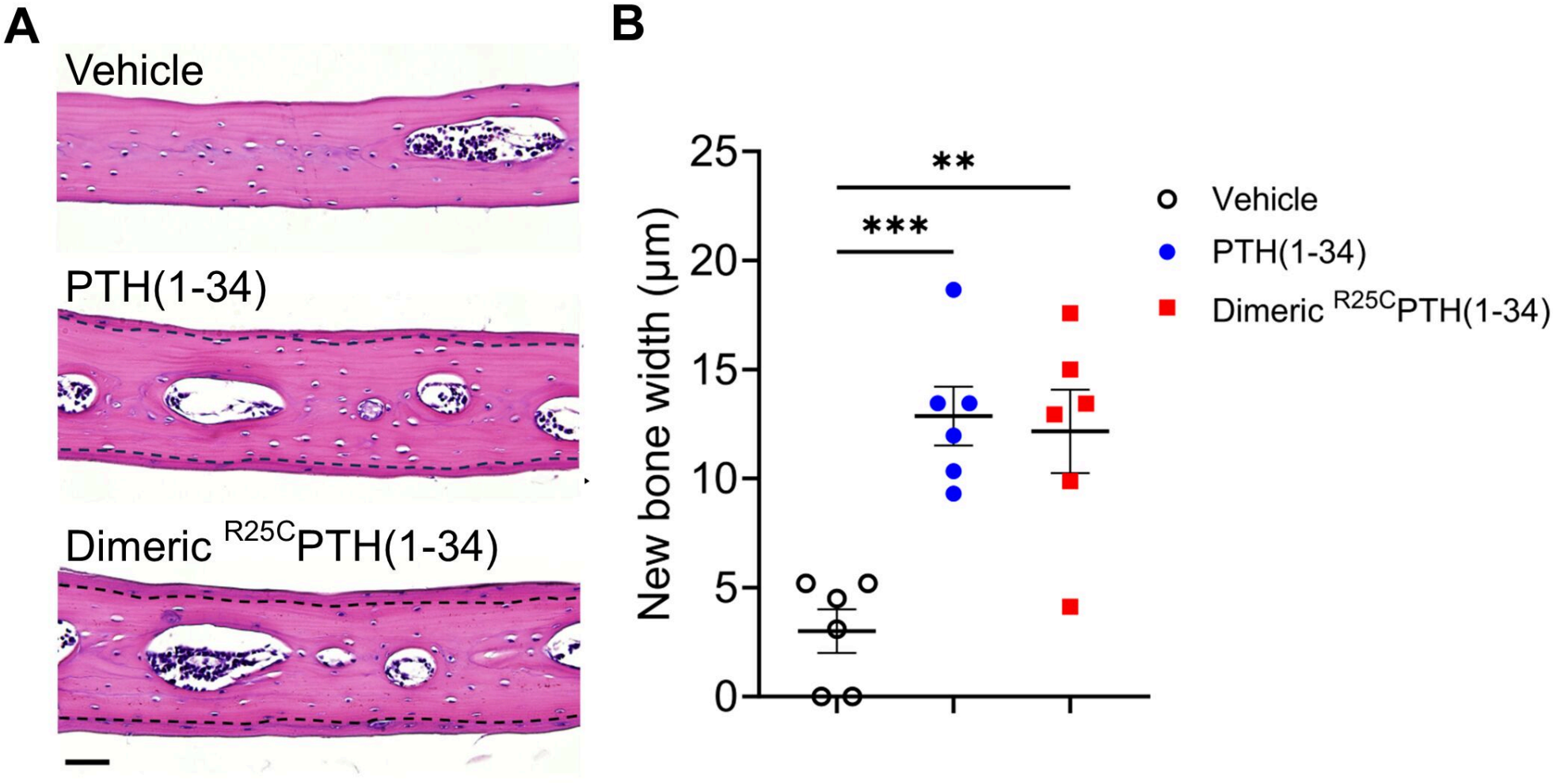

A
5' UTR
CDS (348 bp)
3' UTR
PTH gene
Exon 1
Intron (3,050 bp)
Exon 2
Intron (103 bp)
Exon 3
Translation
Pre-Pro-PTH
N-
Pro-PTH (90 AA)
-C
Signal peptide (25 AA)
Enters ER
Pro-PTH
N-
Mature PTH (84 AA)
-C
Pro sequence (6 AA)
Enters Golgi apparatus
Mature PTH (secreted)
N-
Mature PTH (84 AA)
-C
B
PTH gene
Exon 1
Exon 2
Exon 3
(5'-) CTG/CGT/AAG (-3')
(5'-) CTG/TGT/AAG (-3')
Mature (R25C)PTH (secreted)
N-
Mature R25CPTH (84 AA)
-C
(Arginine → Cysteine)
C
Recombinant PTH (PTH-3xFLAG)
N-
Mature PTH (84 AA)
3xFLAG & V5 tag
-C
Recombinant R25CPTH (R25CPTH-3xFLAG)
N-
Mature R25CPTH (84 AA)
3xFLAG & V5 tag
-C
(R25C)
Signal peptide (25 AA)
Pro sequence (6 AA)
D
EGFP
PTH-3xFLAG
R25CPTH-3xFLAG
EGFP
PTH-3xFLAG
R25CPTH-3xFLAG
EGFP
PTH-3xFLAG
R25CPTH-3xFLAG
EGFP
PTH-3xFLAG
R25CPTH-3xFLAG
RA
+ + + - - -
+ + + - - -
α-FLAG
α-HSP90
Medium
Cell lysate

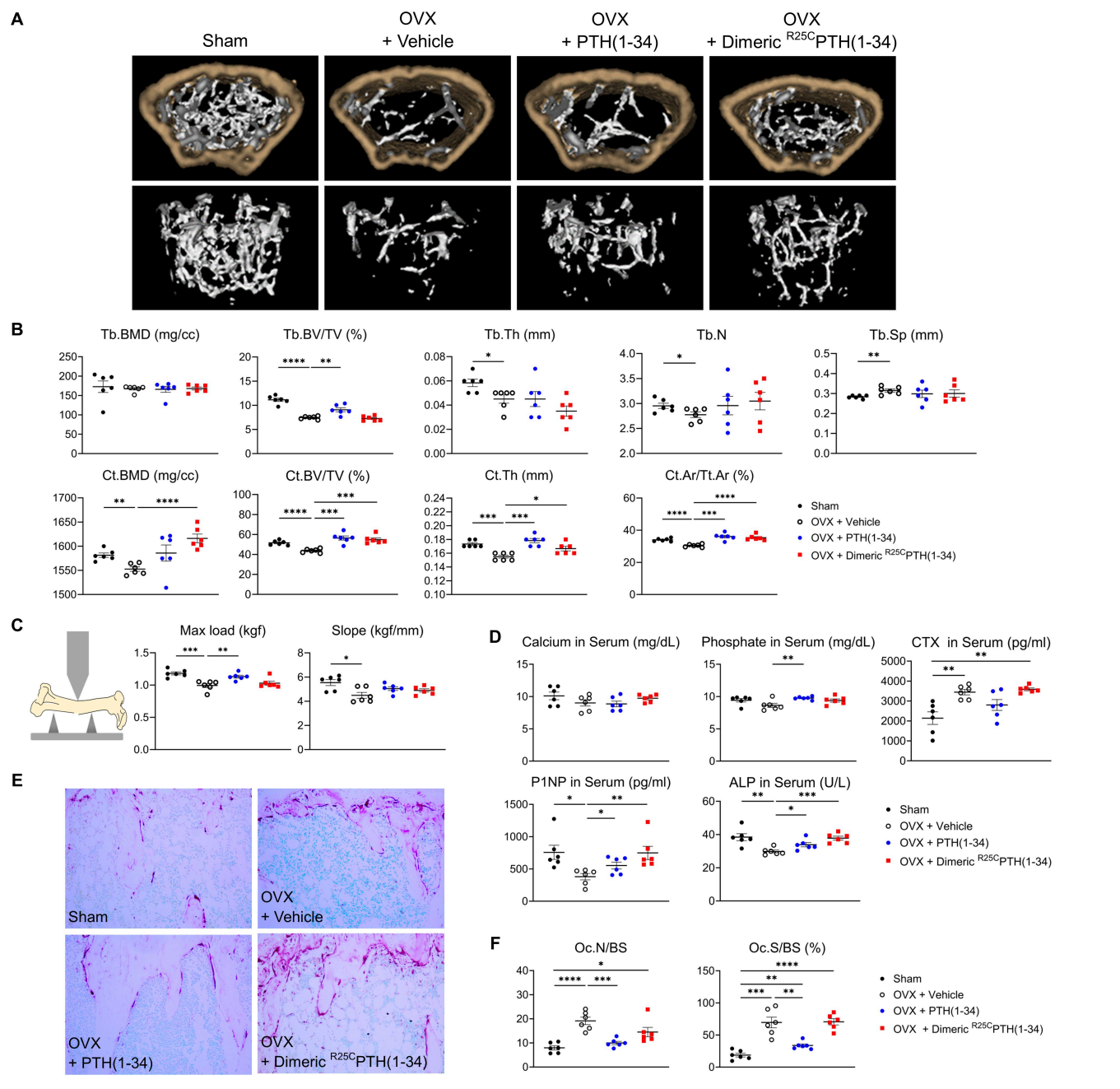

A
R0 conformation of PTH1R
125I-PTH(1-34) bound (SB, %)
100
75
50
25
0
-12
-11
-10
-9
-8
-7
-6
-5
[Ligand] Log M
RG conformation of PTH1R
125I-MPTH(1-15) bound (SB, %)
100
75
50
25
0
-13
-12
-11
-10
-9
-8
-7
[Ligand] Log M
PTH(1-34)
Monomeric R25CPTH(1-34)
Dimeric R25CPTH(1-34)
B
SGS-72 (SaOS2/glosensor)
cAMP (Luminescence, cps)
600,000
400,000
200,000
0
-12
-11
-10
-9
-8
-7
[Ligand] Log M
PTH(1-34)
Monomeric R25CPTH(1-34)
Dimeric R25CPTH(1-34)

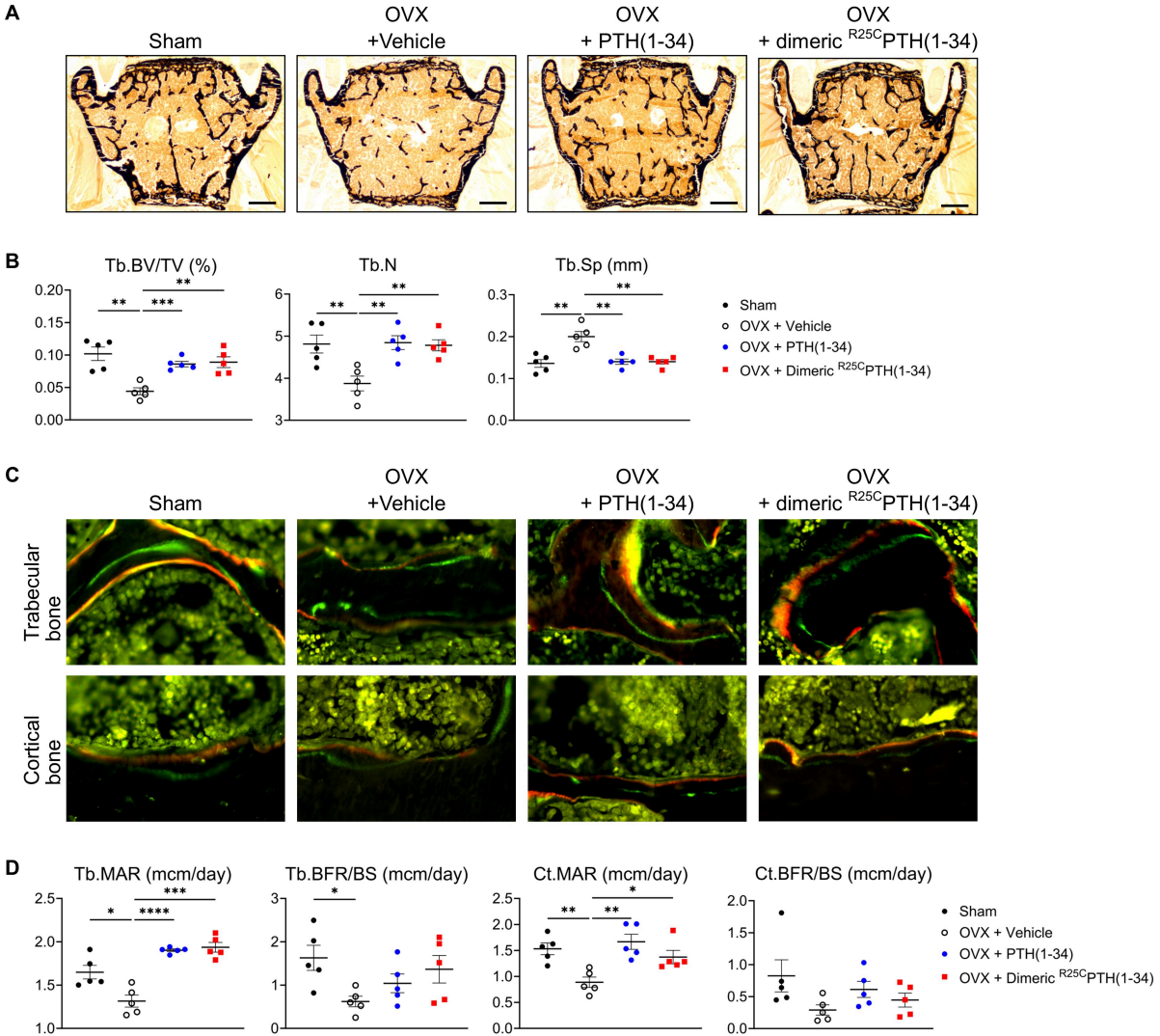

A
Plasma ionized Ca (mM)
1.4
1.3
1.2
1.1
###
##
*
*
0
1
2
3
4
5
6
Time after injection (hour)
Vehicle
PTH(1-34)
Dimeric R25C PTH(1-34)
B
Plasma phosphate (mg/dL)
10
9
8
7
6
**
Time after injection (hour)
Vehicle
PTH(1-34)
Dimeric R25C PTH(1-34)
C
Urine phosphate (mg/dL)
400
300
200
100
0
#
Time after injection (hour)
Vehicle
PTH(1-34)
Dimeric R25C PTH(1-34)


A
600
400
200
0
Plasma cAMP (pmol/mL)
*
0 min
15 min
Time after injection
Vehicle
PTH(1-34) (50 nmol/kg)
Dimeric R25CPTH(1-34) (50 nmol/kg)
Dimeric R25CPTH(1-34) (100 nmol/kg)

B
400
300
200
100
0
Plasma cAMP (pmol/mL)
0:00
0:15
0:30
0:45
1:00
Time after injection (hour:minute)
Vehicle
PTH(1-34)
Dimeric R25CPTH(1-34)


A
Vehicle
PTH(1-34)
Dimeric R25CPTH(1-34)
B
New bone width (µm)
25
20
15
10
5
0
**
***
Vehicle
PTH(1-34)
Dimeric R25CPTH(1-34)


A
Sham
OVX + Vehicle
OVX + PTH(1-34)
OVX + Dimeric R25CPTH(1-34)
B
Tb.BMD (mg/cc)
Tb.BV/TV (%)
Tb.Th (mm)
Tb.N
Tb.Sp (mm)
Ct.BMD (mg/cc)
Ct.BV/TV (%)
Ct.Th (mm)
Ct.Ar/Tt.Ar (%)
Sham
OVX + Vehicle
OVX + PTH(1-34)
OVX + Dimeric R25CPTH(1-34)
C
Max load (kgf)
Slope (kgf/mm)
D
Calcium in Serum (mg/dL)
Phosphate in Serum (mg/dL)
CTX in Serum (pg/ml)
P1NP in Serum (pg/ml)
ALP in Serum (U/L)
Sham
OVX + Vehicle
OVX + PTH(1-34)
OVX + Dimeric R25CPTH(1-34)
E
Sham
OVX + Vehicle
OVX + PTH(1-34)
OVX + Dimeric R25CPTH(1-34)
F
Oc.N/BS
Oc.S/BS (%)
Sham
OVX + Vehicle
OVX + PTH(1-34)
OVX + Dimeric R25CPTH(1-34)


A
Sham
OVX +Vehicle
OVX + PTH(1-34)
OVX + dimeric R25CPTH(1-34)
B
Tb.BV/TV (%)
Tb.N
Tb.Sp (mm)
Sham
OVX + Vehicle
OVX + PTH(1-34)
OVX + Dimeric R25CPTH(1-34)
C
Sham
OVX +Vehicle
OVX + PTH(1-34)
OVX + dimeric R25CPTH(1-34)
Trabecular bone
Cortical bone
D
Tb.MAR (mcm/day)
Tb.BFR/BS (mcm/day)
Ct.MAR (mcm/day)
Ct.BFR/BS (mcm/day)
Sham
OVX + Vehicle
OVX + PTH(1-34)
OVX + Dimeric R25CPTH(1-34)