FIRST INSIGHTS OF THE DANUBE STURGEON (*ACIPENSER GUELDENSTAEDTII*) SKIN ADHERENT MICROBIOTA

Răzvan MATACHE[1], György DEÁK[1,2], Abdulhusein JAWDHARI[2], Isabela SADÎCA[1,2], Cristian-Emilian POP[2,3,4*], Sergiu FENDRIHAN[2,3], Nicolae CRĂCIUN[2,4]

This study assesses the microbial composition associated with the skin of Acipenser gueldenstaedtii, emphasizing species that are adherent. Various growth media were employed to cultivate samples collected from a group of 12 sturgeons from the same batch, revealing a diverse and atypical bacterial population. Notably, Ralstonia pickettii is documented here for the first time, as there have been no prior associations reported between this species and either wild or aquaculture sturgeon populations, to the best of our knowledge. The identification of presented adherent and non-adherent species raises questions and concerns regarding the existing microbial communities in sturgeons.

Keywords: Sturgeons; *Acipenser gueldenstaedtii*; skin microbiota; pathogens; bacterial adherence; microbial communities

1. Introduction

Tegument pathogens pose a growing concern in some sectors of the aquaculture industry, a rapidly expanding food sector of considerable economic significance [1, 2]. While the majority of commercially raised fish species intended for human consumption exhibit high water bacteria treatment tolerance and rapid growth, mitigating the severe impact of pathogens on their population, sturgeons however present a distinct scenario. Due to their prolonged maturation process, that is usually taking 8 to 13 years for males and 14 to 23 years for females [3], as well as to their heightened sensitivity to the conventional cooper sulphate bacterial treatment [4, 5], the significance of tegument adherent pathogens is therefore accentuated. For sturgeons, and not only, prolonged

[1] University Politehnica of Bucharest, Romania;
[2] National Institute for Research & Development in Environmental Protection, Bucharest, Romania;
[3] Non-Governmental Research Organization Biologic, Bucharest, Romania;
[4] University of Bucharest, Faculty of Biology, Bucharest, Romania

pathogenic microbiota hosting can notably impact the growth and survival rate [6, 7]. In this study we evaluated *Acipenser gueldenstaedtii* skin adherent microbiota, excluding host specific and general parasites [8–10]. In terms of dermal skeleton, *A. gueldenstaedti* presents five rows of modified diamond-shaped ganoid scales called scutes. These dermal structures are of woven bone type, sharing similarities with the structure of the ewe bone, although the proportion of the apatitic domain seems to be lower for scutes [11]. The outermost layer of the body (covering the scutes as well), is considered tegument, which thus consists of the skin (integument) and its appendices. The skin contains mucosal glands and mucosa skin-associated lymphoid tissues (SALT) [12], which can trigger the innate immune response [13].

The organ is composed of the outer layer epidermis, followed by dermis and hypodermis. It is the epidermis that possesses a variety of cell types that participate in the exudation of surface mucus [14, 15]. These mucus-producing cells are recognized for harboring diverse biologically active large molecules, primarily glycoproteins, which holds significance in pivotal biological processes, particularly those related to immune responses [16]. The fundamental architectural element of the epidermis is comprised of stratified and irregularly shaped epithelial cells that are organized into multiple tiers. The basal layer is composed of columnar basal cells that overlay the basement membrane, these basal cells possess a columnar nucleus situated toward their upper portion.

The intermediate layer encompasses a variety of multilayered, irregular epithelial cells alongside mucous cells. In the intermediate layer, the cell nuclei are typically round or oval, occupying a central position within the cell. Conversely, epithelial cells in proximity to mucous cells exhibit crescent or irregular nuclei. The interface between the epidermis and dermis is demarcated by a relatively substantial basement membrane [14], the dermis is composed of highly oriented, large and parallel collagenous fibers [15]. In sturgeons, like in many other aquatic organisms, the epidermal mucus represents a line of defense in terms of anti-inflammatory and antimicrobial activity [17].

Still, different skin regions are prone to lesions for various reasons [18], and consequently to infections, offering a gateway for pathogenic and opportunistic bacteria.

Known as being common to many fish species and the aquatic environment in general, the microorganisms identified in this work provide the first insights regarding skin adherent microbiota in the particular case of *A. gueldenstaedti*, although the pattern can be valid for other, if not all sturgeon species.

The purpose of the present work was to identify adherent skin microbiota of cultured *A. gueldenstaedti*, as to the best of our knowledge, this is the first such report.

2. Materials and methods

2.1. Specimen preparations and sampling

Nine juvenile specimens of *Acipenser gueldenstaedti* (total length ≈ 13 cm) sourced from a fish farming facility were washed through three successive baths of microbiological pure water, individually rinsed in aseptic conditions and placed in enclosed sterile polycarbonate water tanks (AquaMedic, Germany) with 2.5 L of ultra purified water (Mili-Q, Merck, Germany) each. A group of three separate specimens were kept as control (unwashed). Aeration in aseptic conditions was assured with an air pump through sterile silicon tubbing connected to a microbiological air filter. After 8 hours, each specimen was taken out from the designated water tank and rinsed intensively with ultra purified water in a microbiology class 2 biosafety cabinet, with a focused intensive rinse on the belly region. The sampling was performed by immediately and firmly swabbing a 3 cm^2 linear belly area using a sterile flocked swab, which was then placed in 1 mL of liquid Amies storage media recipient (Copan ESwabs, Italy). No specimens were harmed during this work and all specimens were later released back into the aquaculture facility.

2.1. Cultures and primary identifications

From each sample, 100 µL of Amies media was placed on triptic soy agar (TSA), nutrient, Drigalski and chromogen agar plates (MLT, Romania), and spread until drying using a sterile Drigalski spatula and a Petri dish turntable (Schuett-Biotec, Germany). The plates were incubated at 27° C and colonies were counted using a colony counter (Schuett-Biotec, Germany) after 24 and 48 hours of incubation, prior to strains isolation and primary identification on chromogenic agar. With few exceptions, fish bacteria are inhibited by incubation temperatures above 30° C [19], therefore the 27° C incubation temperature was chosen as an realistic compromise for sturgeon microbiota growth, exceeding this threshold temperature enhances heat shock proteins (hsp) expression in juvenile specimens, especially the protein folding and heat stress marker hsp70 [20].

2.3. Final taxonomic identification

Final taxonomic identification was performed with a Matrix-Assisted Laser Desorption/Ionization Time-of-Flight (MALDI-TOF) Mass Spectrometer Biotyper (MALDI-TOF Microflex LT/SH, BrukerDaltonik, Germany). MSP 96 target polished steel plate (Bruker Daltonics, Germany) spots were loaded with isolated colonies, and α-Cyano-4-hydroxycinnamic acid (Bruker Daltonics, Germany) matrix was added with the stock solution (acetonitrile, trifluoracetic

acid and distilled water). The SR Library Database was used for the identification (Database CD BT BTyp2.0-Sec-Library 1.0) with MBT Compass (v.4.1) software.

3. Results and Discussions

The bacterial load and diversity varied significantly among *A. gueldenstaedti* individuals, although they were belonging to the same batch.

Within this study we isolated and identified a number of 11 species, out of which 3 particularly adherent ones (Table 1), being of high interest for public health monitoring, and even for development of fish skin grafts wound dressings technologies [21] .

Table 1

Colonies and identified isolates

Sample	Media	Total Plate Count		Strains identified
		24 h	48 h	
Control 1	TSA	x	x	*Escherichia coli* *Plesiomonas shigelloides* *Staphylococcus warneri* *Bacillus cereus*
Control 2		x	x	
Control 3		122	x	
S1		X	X	*Bacillus cereus* *Plesiomonas shigelloides*
S2		694	X	
S3		720	X	
S4		10	X	
S5		152	X	
S6		770	X	
S7		170	X	
S8		440	X	
S9		310	X	
Control 1	Nutrient	405	x	*Escherichia coli* *Plesiomonas shigelloides* *Massilia aurea* *Aeromonas hydrophila* *Pseudomonas stutzeri* *Sphingomonas paucimobilis*
Control 2		724	x	
Control 3		508	x	
S1		30	20	*Ralstonia pickettii* *Bacillus cereus*
S2		10	70	

S3		170	230	*Plesiomonas shigelloides*
S4		30	70	
S5		40	110	
S6		20	70	
S7		10	10	
S8		20	20	
S9		120	140	
Control 1	Drigalski	870	x	*Plesiomonas shigelloides* *Aeromonas veronii*
Control 2		120	243	
Control 3		820	x	
S1		0	0	*Plesiomonas shigelloides*
S2		0	35	
S3		0	0	
S4		0	2	
S5		0	3	
S6		0	4	
S7		0	0	
S8		0	3	
S9		0	5	
Control 1	Chromogen	564	x	*Escherichia coli* *Aeromonas veronii* *Aeromonas jandaei* *Plesiomonas shigelloides* *Bacillus cereus*
Control 2		773	x	
Control 3		768	x	
S1		0	2	*Plesiomonas shigelloides* *Bacillus cereus*
S2		0	2	
S3		20	25	
S4		10	14	
S5		70	80	
S6		50	60	
S7		10	13	
S8		20	24	
S9		360	380	

X = plate confluence, uncountable.

Plesiomonas shigelloides was the most prevalent species, being found in all groups and grew on all media types that were used in this study, thus showing a particular versatility and adaptability for which it's already known for [22]. This rod-shaped gram-negative pathogen discovered by Ferguson and Henderson in 1947 from clinical fecal samples [23] is native to aquatic organisms and has been isolated from fish, shellfish, crustaceans, marine mammals, amphibians, reptiles, and other aquatic lifeforms [24]. Although initially little attention was given to it as a potential pathogen, *P. shigelloides* now ranks with other established pathogens, like *Aeromonas*, as a potential cause of bacterial gastroenteritis [25–27]. However, unlike *Aeromonas*, how pH, temperature, turbidity, and conductivity affect *P. shigelloides* concentrations in freshwater ecosystems remains poorly understood [28]. The two are known to be often found together, so much so that, traditionally, the genera *Aeromonas* and *Plesiomonas* have been taken together in the past [29, 30], yet afterwards three international workshops have taken into account many different aspects of these two genera and thus the two were distinguished apart [29].

Aeromonas was the second most abundant species, being represented by *A. hydrophila, A. veronii, A. jandaei,* and *A. jandaei*, which all grew in the control groups but none was found in the experimental (rinsed) groups, therefore showing far less adhesivity than *P. shigelloides*. Bacterial adherence is a sign of high pathogenicity [31–33], yet interestingly, *P. shigelloides* infections are less virulent [34], and less frequent than those of *Aeromonas sp.* [22], although *P. shigelloides* was reported to adhere and enter eukaryotic intestinal host cells [35].

Bacillus cereus, another microorganism commonly associated with the aquatic environment [36, 37], was identified as being adherent on the tegument of the studied specimens. There are reports of *Bacillus cereus* colonizing the intestinal tract of fishes [36, 38], and even being used as an biological agent against streptococcosis infections [38], however, to the best of our knowledge, none have reported it as a skin pathogen, especially in sturgeons. The gram negative rod-shaped bacteria is nevertheless a pathogen to humans, being transmitted via consumption of infected fish [39] and its also known for its versatility in terms of temperature, pH and responsible for food-borne outbreaks [40–42].

Pseudomonas stutzeri and *Sphingomonas paucimobilis* were found only on the nutrient media and only on control (non-adherent). The first is a denitrifying bacterium with ubiquitous environmental distribution, has additionally been identified as an opportunistic human pathogen. The species has been suggested as an exemplary model organism for investigations into denitrification processes due to its unique metabolic capabilities. Moreover, numerous strains exhibit inherent natural transformation abilities and some possess the capacity for dinitrogen fixation [43]. The latter, formerly known as *Pseudomonas paucimobilis* and CDC

group IIk-1 [44], is widely distributed in water, including water sources in the hospital environment. *Massilia aurea* was also identified to be non-adherent and grew only on nutrient media. The species is worth taking into account as it was first identified only almost two decades ago in drinking water in Spain [45]. There is little information regarding *Massilia aurea* to date, however *Massilia* strains are extensively distributed and perform diverse ecological roles. Initial investigations indicate that *Massilia* is the predominant species within constructed wetland ecosystems, yet the specifics of its species makeup and distribution in these wetlands remain uncertain [46]. *Massilia* spp. play a constructive role in environmental enhancement, for instance, *M. aromaticvorans* (ML15P13T) possesses the capacity to break down harmful gaseous organic air pollutants such as benzene, toluene, ethylbenzene, and xylene isomers [47].

Still, the most particularly interesting finding in this study was the skin-adherent presence of *Ralstonia pickettii.*

Ralstonia pickettii is an emerging Gram-negative opportunistic pathogen [48]. Formerly called *Pseudomonas pickettii* [49] or *Burkholderia pickettii* [50], *R. pickettii* is a waterborne organism of low virulence and minor clinical importance [49] that, to the best of our knowledge, has not been reported in sturgeons. However, the species was identified in biofilm formation in plastic water piping where it resisted disinfection with numerous potent agents, showing a high resilience [51], and was also found in samples of wild and aquacultured fresh seafood [52]. The identification of this species is of particular importance as it can be considered an extremophile due to it's high oligotrophic and other resistance abilities [53].

4. Conclusions

In this study surprisingly exotic species have been identified, and some found to be adherent on the sturgeon skin. In particular, *Ralstonia pickettii* is hereby reported for the first time, as to the best of our knowledge no links between this species and wild or aquaculture sturgeons has been found. The detected adherent and non-adherent species raise questions and concerns in terms of current sturgeon microbial communities.

Future studies examining the possible presence of microbiota withing the skin layers are required to support the presented preliminary data. Further, bioprospecting microbiota of wildlife sturgeons is recommended as the current climate and ecological changes, as well as the increasingly levels of water contamination with emergent pollutants, can induce not only immune system alterations of fishes, but also significant microbial community swifts that can led to detrimental consequences of unknown extent.

R E F E R E N C E S

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