Bacterial defenses and their trade-off with growth are not ubiquitous but depend on ecological contexts

Zhi-Ling Liu[1,2], Jia Liu[1,2], Deng-Ke Niu[1,2*]

[1]MOE Key Laboratory for Biodiversity Science and Ecological Engineering, College of Life Sciences, Beijing Normal University, Beijing, 100875, China

[2]Beijing Key Laboratory of Gene Resource and Molecular Development, College of Life Sciences, Beijing Normal University, Beijing, 100875, China

*Correspondence author:

Deng-Ke Niu, College of Life Sciences, Beijing Normal University, Beijing, 100875, China.

dkniu@bnu.edu.cn, dengkeniu@hotmail.com

ORCID:

Zhi-Ling Liu, https://orcid.org/0000-0001-6579-6642

Jia Liu, Jia Liu, https://orcid.org/0009-0006-5652-5414

Deng-Ke Niu, https://orcid.org/0000-0003-4503-2658

Abstract

Bacteriophages, also known as bacterial viruses, significantly influence microbial ecosystems, driving bacteria to evolve diverse antiviral defense mechanisms. This study explores the intricate relationship between bacterial defenses and growth rates across diverse ecological contexts. Our investigation reveals that bacteria lacking defenses exhibit prolonged doubling times. Integrating phylogenetic eigenvectors into the ecological feature matrix, we employed a phylogenetic random forest model to identify key ecological features influencing defense presence and abundance. Further phylogenetic regressions unveil nuanced dependencies of bacterial defenses on specific environmental factors, challenging assumptions of a universal defense system distribution and underscoring reliance on subtle ecological cues. Notably, symbiotic and endosymbiotic bacteria often exhibit reduced defense systems and negative correlations between defense system abundance and the minimal doubling time. Conversely, free-living and motile bacteria show significant positive correlations between minimal doubling time and defense system abundance. Moreover, we highlight the pivotal role of ecological variables like habitat specificity and nutrient availability in shaping bacterial growth rates and defense mechanisms. Our findings underscore the complexity of microbial interactions and stress the need to consider ecological context in understanding defense strategies. We propose that trade-offs between growth and defense are ubiquitous due to sporadically inefficient optimization of limited resources, particularly in populations with small effective sizes, where both mechanisms may weaken concurrently due to genetic drift. This challenges traditional notions of trade-offs and underscores the impact of ecological context on microbial strategies.

Keywords

Defense system, minimal doubling time, trade-off, symbiosis, endosymbiosis, gut microbiome.

Introduction

Bacterial viruses, also known as phages or bacteriophages, represent the most abundant biological entities on Earth. They significantly outnumber bacterial cells across various samples and microbiomes, with ratios reaching up to ~15-25 fold, as observed in freshwater plankton (1, 2). In environments like seawater, this constant predation results in a substantial daily death rate of bacterioplankton, estimated between 25-40% (3, 4). To combat phage attacks, bacteria have evolved an arsenal of defense tactics targeting every stage of the phage's life cycle. These tactics include diverse mechanistic approaches such as the restriction-modification (RM) system, the Clustered Regularly Interspaced Short Palindromic Repeat and associated protein (CRISPR-Cas) system, and the Abortive Infection (Abi) system (5-7), collectively referred to as antiviral defense systems.

Despite the potential for horizontal gene transfer, bacteria typically only possess a subset of the available anti-phage mechanisms (8). For instance, the CRISPR-Cas systems, which provide adaptive immunity by targeting previously encountered pathogens, are found in approximately 40% of bacteria (9). Similarly, Abi systems are sporadically distributed across bacterial phylogeny (7), while RM systems are widespread but absent in about 4% of bacterial genomes (10). Additionally, the uneven distribution of antiviral defense systems extends to the archaeal lineage (9, 10), reflecting trade-offs between the costs of maintaining such systems and the benefits gained in phage resistance (8, 11-15).

In biological systems, traits often exhibit trade-offs due to constraints on resources such as energy, habitat, or time (16-18). For example, energy allocated for growth cannot simultaneously be used for defense or other energy-intensive processes, leading to necessary compromises among traits. While the concept of growth-defense trade-offs is well-established in plants and animals, its relevance in virus-infected prokaryotes is less recognized (11, 19).

Laboratory research on phage resistance has extensively explored trade-offs using model bacteria and phages such as *Escherichia coli* and *Pseudomonas aeruginosa*. Phage resistance evolution significantly impacts bacterial fitness components, including slowed cell division rates, reduced nutrient uptake, decreased virulence, and impaired cell motility (11, 20-29). Notably, some

phage-resistant strains exhibit trade-ups, where selection improves multiple fitness components

simultaneously, including antibiotic resistance (30, 31).

However, resistance to phages does not consistently correlate negatively with bacterial

growth rates in natural environments. Studies examining marine *Synechococcus* strains and phage

isolates, as well as bacteria and phages from horse chestnut leaves, have shown conflicting results

regarding the growth-defense trade-off (26, 32). Recent research on marine bacterial communities has

observed a negative correlation between growth rates and resistance to protists and viruses,

supporting the growth-defense trade-off hypothesis (33).

To gain a comprehensive understanding of the trade-off between prokaryotic growth and

defense, investigations encompassing diverse environments and species are essential. Recent research

analyzing growth rates and adaptive immunity across bacterial domains revealed a significant positive

correlation, indicating evolutionary trade-offs between growth and adaptive defense against virulent

phages (15). Additionally, besides CRISPR-Cas systems, prokaryotes possess numerous other

antiviral mechanisms, such as the RM and Abi systems, warranting further exploration (5, 6, 34, 35).

Understanding the relationship between growth and antiviral defense across different environments

and prokaryotic lineages holds promise for advancing our knowledge of evolution and informing the

development of antimicrobial strategies.

To grasp the trade-off between prokaryotic growth and defense, it is crucial to take a

comprehensive approach. Despite occasional discrepancies, our objective is to achieve a

comprehensive understanding of whether phage-resistant lineages typically demonstrate slower

growth rates compared to phage-sensitive ones across bacterial domains. These insights could

significantly impact our understanding of evolutionary processes and contribute to the development of

innovative antimicrobial strategies.

Recently, we investigated the relationship between growth rate and adaptive immunity in

4142 bacteria, utilizing predicted minimal doubling times retrieved from the EGGO database and

CRISPR-Cas systems annotated with CRISPRCasFinder (15, 36, 37). Our findings unveiled a

significant positive correlation, indicating evolutionary trade-offs between growth and adaptive

defense against virulent phages within the bacterial domain.

In addition to CRISPR-Cas systems, prokaryotes have evolved various other antiviral

mechanisms, including the RM system, the Abi system, and numerous others that warrant further

exploration (5, 6, 34, 35). Notably, certain prokaryotes, such as the delta proteobacteria *Desulfonema*

limicola, can harbor over 50 defense systems in their genomes (38). Recently, Tesson et al. (38)

developed DefenseFinder, a novel antivirus-system detection program based on the HMM mapping

algorithm, which enables the identification of known antiviral systems across 21,000 fully sequenced

prokaryotic genomes. Leveraging this tool, we explored the relationship between growth and antiviral

defense across bacterial domains, as well as within specific environments.

Materials and Methods

The minimal doubling times of bacteria and archaea were extracted from the EGGO database

(accessed on 2021-11-26), encompassing over 200,000 predictions of estimated growth rates (36).

These values underwent a log transformation using the natural logarithm as the base. Due to its

smaller sample size, we opted not to utilize the dataset of prokaryotic empirical minimal doubling

times provided by Madin et al. (39) in this study. While empirical data typically offer higher accuracy

compared to computationally generated predictions, predictive methodologies have their advantages

(15). Empirical minimal doubling periods may offer superior advantages over computationally

generated predictions due to the inherent limitations of predictive methodologies achieving 100%

precision. However, the wide spectrum of genetic diversity observed among strains of individual

bacterial species necessitates empirical data encompassing various factors, all procured from

equivalent strains, a requirement that is not consistently met. Predicted phenotype values, including

defense systems and growth rate, for a strain originate from the genome sequences of that very strain,

potentially circumventing faults stemming from polymorphism within each species.

The antiviral defense systems of over 21,000 complete microbial genomes were sourced from

Tesson et al. (38). Phylogenetic relationships among the analyzed prokaryotes were obtained from the

Genome Taxonomy Database (GTDB) (40) (accessed 2022-04-08). GTDB offers superior taxonomic

resolution and more precise classification compared to traditional phylogenetic trees by leveraging

genomic data.

The intersecting samples among the three datasets mentioned above consisted of 3532

bacterial genomes and 237 archaeal genomes, some of which belonged to the same species. However,

upon intersecting with subsequent datasets, only a minimal number of archaeal species were acquired,

rendering most analyses impractical. Consequently, this paper focuses solely on the study of bacteria.

Ecological characteristics of prokaryotes were obtained from the ProTraits database (41),

accessed on October 2, 2022, via "ProTraits_precisionScores.txt". This comprehensive database

integrates data from various repositories such as GOLD, NCBI, and KEGG, utilizing automated text

mining and comparative genomic techniques to associate microorganisms with phenotypes. Our study

focuses on ecological features potentially influencing defense system distribution, excluding

physiological traits like glycerol and glutamate. Additionally, features with ambiguous biological

significance, automated physiological features from machine learning, and those with missing P

values exceeding 10% of the overall sample were discarded, retaining 125 prominent ecological

features for analysis. Two columns represent each ecological feature in the database, providing

precision scores for both the presence and absence of the feature, respectively. To facilitate analysis,

these scores were converted into an overall maximal confidence score ranging from 0 to 1, following

the methodology of Weissman et al. (42). Missing values in the ProTraits database were imputed

using phylogenetic information (43). The PVRdecomp function of the R package PVR (Version 0.3)

(44) was employed to extract eigenvectors from the phylogenetic distance matrix. Subsequently, the

missForest function of the R package missForest (Version 1.5) (45) was utilized to estimate missing

values.

Random forest, an integrated algorithm of decision trees, was utilized for regression tasks in

this study. This method optimizes regression tasks, effectively handles high dimensionality, mitigates

overfitting risks, accommodates both categorical and numerical variables, and consistently delivers

reliable outcomes, making it superior to alternative techniques. The eigenvectors extracted from the

phylogenetic distance matrix were incorporated into the ecological feature matrix to control

phylogenetic relationships within the random forest analysis. The randomForest package (version 4.7-

1.1) (46) was implemented in R (version 4.2.0).

Phylogenetic generalized least-squares (PGLS) analysis of correlations was conducted using

the R package phylolm (version 2.6.2) (47). The Pagel's λ model was employed due to its generally

lower AIC values in our dataset. Differences in minimal doubling times between the two groups were

compared using the phylANOVA function (48), while phylogenetic logistic regression analysis with

dichotomous variables as dependent variables was performed using the phyloglm function (49). When

multiple correlation analyses were performed on the same data, *P* values were corrected using the

Benjamini & Hochberg method via the p.adjust function in R (Version 4.0.2).

Results

Phylogenetic Signals in Analyzed Features and Their Implications for Comparative

Analysis

Due to shared ancestry among bacterial samples, feature values may exhibit non-independence,

violating fundamental assumptions of conventional statistical methods. As such, using phylogenetic

comparative methods becomes imperative to mitigate these effects (50, 51). We evaluated the

phylogenetic signals of all analyzed features and observed significant signals in nearly all cases, with

minimal doubling time demonstrating the strongest signal (Supplementary Table S1). Consequently,

we will employ phylogenetic comparative methods whenever applicable.

A recent study by Chen et al. (52) highlighted the potential for conflicting outcomes in PGLS

analysis when interchanging dependent and independent variables. While PGLS cannot establish

causality but rather identifies correlations, Chen et al. utilized simulated data and a gold standard to

establish criteria for selecting the dependent variable. Their findings suggested that choosing the

feature with a stronger phylogenetic signal as the dependent variable led to more accurate correlation

detection than randomly selected variables. In line with this approach, we adopted the feature

exhibiting a stronger phylogenetic signal as the dependent variable in all PGLS analyses conducted in

this study.

Ecological Factors Influencing the Presence of Antiviral Defense Systems in Bacteria

Our analysis stems from a comprehensive program capable of detecting all known antiviral systems in

prokaryotic genomes (38). However, among the 3532 bacterial genomes (Supplementary Table S2)

examined, 7% (243) were found to lack any defense systems. Phylogenetic ANOVA analysis

unveiled that these bacteria exhibited significantly longer minimal doubling times compared to those

encoding one or more defense systems (Mean $\pm$ SD: 5.6 $\pm$ 3.1 h $vs.$ 3.2 $\pm$ 4.4 h. $P = 0.0011$).

Surprisingly, bacteria devoid of defense systems did not demonstrate faster proliferation rates,

contradicting the anticipated trade-off between growth and defense. It is plausible that these bacteria

inhabit specific ecological niches where neither defense mechanisms nor rapid growth are necessary.

To delve into the factors influencing the presence or absence of bacterial defense systems, we

leveraged ecological data from the ProTraits database. Since the database primarily comprises

bacterial species rather than individual strains or genomes, we aggregated some of the 3532 genomes

into species. To capture species-level characteristics, we computed median values of defense system

abundance and minimal doubling time for genomes within the same species, identifying 1044 distinct

bacterial species (Supplementary Table S3).

Utilizing a random forest model, we ranked the importance of 125 ecological features.

Remarkably, factors such as "known habitats = insect endosymbiont," "known habitats =

endosymbiont," "biotic relationship = symbiont," and "host = insects general" emerged as top

influencers, underscoring the significance of symbiotic and endosymbiotic relationships (Fig. 1).

Given that the random forest model cannot discern the directional effect of features on the

presence or absence of antiviral defense systems, we conducted phylogenetic logistic regression

(Table 1). Results indicated a negative correlation between antiviral defense presence and "known

habitats = insect endosymbiont," "biotic relationship = symbiont," and "host = insects general" ($P <$

10^{-8} for all), while "known habitats = endosymbiont" showed no significant correlation ($P = 0.452$).

Ecological Determinants of Defense System Abundance: Insights from Symbiosis and

Environmental Dynamics

The preceding study explored factors influencing the presence or absence of defense systems. Here,

we delve deeper into the variables shaping defense system abundance. Bacteria lacking defense

mechanisms will be recorded as having zero systems, ensuring inclusivity in our analysis.

Among 125 ecological features, "known habitats = insect endosymbiont," "known habitats =

endosymbiont," and "biotic relationship = symbiont" emerged as top influencers, underscoring the

importance of symbiotic associations (Fig. 2). Furthermore, symbiotic relationships with hosts were

supported by additional features within the top rankings, including "ecosystem type = circulatory

system," "ecosystem subtype = blood," "habitat = host associated," "host = insects general,"

"ecosystem category = mammals," "known habitats = host," and "ecosystem = host associated." If

symbiotic bacteria tend to have fewer defense systems for certain reasons, free-living bacteria should

exhibit a comparatively higher abundance of defense systems. Indeed, "habitat = free living" ranks

within the top 19.

Additionally, aquatic environments and energy sources emerged as key factors. "Habitat =

aquatic" and "energy source = photoautotroph" ranked prominently, indicating their influence on

defense system abundance (Fig. 2). Additionally, polyhabitat, filamentous shape, mobility, and

bioremediation also appear to influence defense system abundance significantly.

Further PGLS analysis revealed negative correlations between symbiotic characteristics and

defense system abundance, while free-living bacteria exhibited a positive association (Table 2). While

some correlations may not achieve statistical significance, all other ecological characteristics

positively correlate with the abundance of defense systems (Table 2). Notably, bacteria inhabiting

multiple habitats, exhibiting motility, or being involved in environmental bioremediation tend to

possess a higher abundance of defense systems. Given their exposure to complex or fluctuating

environments, these bacteria are more susceptible to encountering invasive elements such as phages.

Another intriguing finding is that filamentous bacteria harbor significantly more defense systems,

likely due to their filamentous morphology providing increased sites for phage attachment.

Ecological Factors Impacting Bacterial Growth Rates: Insights from Endosymbiosis and

Habitat Dynamics

We investigate whether ecological factors influencing defense system abundance affect bacterial

growth rates. Using PGLS regression, we correlated minimal doubling time with overall maximal

confidence scores for significant ecological features.

Endosymbiosis and symbiosis with insects differ in their effects on bacterial growth rates

compared to symbiosis with mammals (Table 3). "Known habitats = insect endosymbiont," "Known

habitats = endosymbiont," and "Host = insects general" exhibit significantly positive correlations with

minimal doubling time. Conversely, features related to symbiosis with mammals show negative

correlations, albeit not statistically significant. Ambiguous features like "Biotic relationship =

symbiotic" show a significant positive correlation, while others like "Ecosystem = host associated,"

"Known habitats = host," and "Habitat = host associated" exhibit negative correlations.

Aquatic habitat and energy source features positively correlate with minimal doubling time

(Table 3), suggesting slower growth rates in bacteria residing in aquatic environments and utilizing

natural energy sources. Additionally, filamentous morphology correlates with slower growth rates,

while bacteria inhabiting multiple habitats tend to grow faster (Table 3).

Exploring Growth-Defense Relationships Across Ecological Niches

In our analysis utilizing the ProTraits database, each species received an overall maximal confidence

score indicating the likelihood of possessing specific ecological features. We categorized species into

presence or absence groups based on a threshold of 0.5, compiling species counts for the top 30

influential ecological features affecting defense system abundance. Features with fewer than 20

species were filtered out.

To investigate potential growth-defense trade-offs within these ecological niches, we

conducted PGLS regression analysis, correlating bacterial minimal doubling time with defense system

abundance (Table 4). Highly significant correlations ($P < 0.01$) were primarily observed in two niches:

"Habitat = free living" and "Mobility," with extensive sample sizes comprising hundreds of bacterial

species.

Among the features related to symbiosis, significant correlations between growth and defense

were identified in three features, each representing samples of over 400 species: "Ecosystem = host

associated," "Known habitats = host," and "Habitat = host associated." Accounting for phylogenetic

data, closely related lineages with similar characteristics were treated as nearly one effective sample.

Thus, the effective sample size in phylogenetic comparative analysis was substantially lower than the

total number of analyzed lineages. Consequently, the conventional statistical rule of thumb regarding

small and large samples ($n = 30$) should not be applied in phylogenetic comparative studies (53).

Nevertheless, the findings presented in Table 4 suggest the presence of not only a trade-off

but also a trade-up between bacterial growth and defense.

Contextual Dependency in Growth-Defense Correlations Across Bacterial

Environments

In Table 4, diverse correlations emerge between bacterial growth and defense across different

environments. Positive correlations are observed in some bacteria within specific habitats, while

negative correlations are apparent in others; meanwhile, no correlation is discernible in some

instances. Consequently, within a large sample, the prevalence of each bacterial type dictates the

observed correlation pattern: a predominance of bacteria exhibiting positive correlations yields a

positive overall correlation, a balance of the three types results in no discernible correlation, and a

prevalence of bacteria with negative correlations leads to an overall negative correlation. Thus, the

significance of analyzing growth-defense correlations on a global scale diminishes.

Nevertheless, we conducted PGLS regression analysis to explore the growth-defense

correlation in our dataset of 3532 samples, yielding statistically insignificant results ($P = 0.398$).

Despite the lack of statistical significance, our analysis underscores the complex and context-

dependent nature of the relationship between bacterial growth rates and defense mechanisms across

diverse ecological contexts.

Discussion

The analysis of bacterial defense mechanisms, whether assessed by the presence or absence of

defense systems, or their abundance, yielded notable findings: ecological factors associated with

symbiosis and endosymbiosis emerged as the primary influencers of bacterial defenses (Figs. 1, 2).

Furthermore, phylogenetic regression analyses demonstrated a trend wherein symbiotic or

endosymbiotic bacteria tend to exhibit fewer defense systems or lack them altogether (Table 1, 2).

This observation aligns with prior findings noting the scarcity of defense systems among insect

endosymbionts (38, 54-60). Our study unveils statistically significant associations between

symbiosis/endosymbiosis and defense mechanisms, suggesting that such bacteria have reduced the

necessity for robust defensive strategies. Endosymbiotic bacteria, residing within host cells, enjoy

physical insulation from aggressive phages. Similarly, evidence suggests that many symbiotic bacteria

are spatially separated from such phages. Notably, the gut microbiome represents a well-studied

example of symbiotic bacteria, comprising the majority of human symbionts (61, 62). Within the

mammalian small intestine and colon, specialized epithelial cells secrete a mucosal layer (63), which

serves as a formidable barrier against the spread and access of phage populations, providing a refuge

for bacteria (64-66). Consequently, the phage-bacterial ratios in the human gut are significantly lower

compared to other ecosystems, often ranging from 1:1 to 0.1:1 as opposed to 10:1(67). In addition, the

low pH in the stomach, digestive enzymes (mainly proteases), and bile salts make the gastrointestinal

tract hostile to phages (68-70), contributing to a lower virus-to-bacteria ratio in the gut compared to

other environments.

However, the impact of endosymbiosis on bacterial growth rates in insects differs

significantly from symbiosis in mammals (Table 3). The proliferation of endosymbionts is not solely

determined by their inherent ability to replicate but is also influenced by regulatory constraints

imposed by their hosts (71, 72). In contrast, the gut presents a highly complex environment for

bacteria. Nutrient levels fluctuate considerably on a daily basis, and the presence of antimicrobial

molecules and oxygen secreted by the epithelium strongly restricts potential microbial inhabitants

(63), creating a gradient of suitability for bacterial growth from the lumen to the mucosa. Bacteria

residing in isolated microhabitats such as crypts may be tightly controlled by the host epithelium,

while those dispersed into the gut lumen could proliferate in response to nutrient availability. We

hypothesize that the overall impact of gut symbiosis on bacterial growth rate depends on the

distribution of bacteria across different microhabitats.

Due to insufficient sample size, no statistical results have been yielded regarding the

relationship between growth and defense in endosymbiotic bacteria. However, a weak negative

correlation between minimal doubling time and the abundance of defense systems was observed in

symbiotic bacteria (Table 4). This observation suggests that symbiotic bacteria with robust defense

mechanisms tend to exhibit rapid growth. We speculate that, despite potential fluctuations in nutrient

levels, hosts typically provide a eutrophic environment for symbiotic bacteria, where energy and

resources are not limiting factors, thus rendering the trade-off between growth and defense

unnecessary (18). The gut lumen may favor r-selected bacteria, which prioritize rapid growth and

reproduction in unstable environments.

Several ecological features, such as free-living habitat, phototrophic lifestyle, occupancy of

multiple habitats, motility, filamentous morphology, and participation in bioremediation, have been

positively linked to the abundance of defense systems (Table 2). These findings expand upon previous

research on the relationship between defense system abundance and lifestyle, habitat, and geographic

background (38, 54-60). The trade-off between growth and defense has been statistically

demonstrated in free-living and motile bacteria. These findings suggest that certain ecological niches

may favor the allocation of resources towards defense mechanisms, potentially at the expense of

growth rates, in order to cope with environmental challenges such as predation by bacteriophages.

The positive association between defense system abundance and ecological features like free-living

habitat and motility implies that bacteria thriving in diverse and dynamic environments may require

heightened defense strategies to survive and proliferate.

Previous studies have shown a strong association between the abundance of defense systems

and genome size, particularly noting the absence of defense genes in the small genomes of

endosymbionts (38, 56, 60, 73-75). In fact, one study suggests that the number of defense systems

remains largely unaffected by the lifestyle of microbes after controlling for genome size, attributing

the scarcity of defense genes in endosymbionts to their small genomes rather than their lifestyle (60).

In our view, while there is a significant correlation between genome size and the abundance of

defense systems, it does not necessarily mean that studying the quantity of defense systems makes

sense only after controlling for genome size. If, for some reason, the evolutionary driving force

behind an increase in genome size leads to an increase in defense systems, or conversely, if the

evolutionary driving force behind a decrease in genome size leads to a reduction in defense system

genes, then conducting partial correlation analysis controlling for genome size would be necessary.

However, the reduction in genome size in endosymbionts is not a result of natural selection but rather

a consequence of stochastic drift; the genome size decreases because genes are lost. The small

genomes of endosymbionts are the result of many gene losses, including defense systems, rather than

a parallel phenomenon to gene loss. Therefore, controlling for genome size, which effectively

eliminates the gene loss itself, is not only meaningless but also misleading when studying the

evolution of the quantity of defense system genes (both gained and lost).

The concept of trade-offs in evolutionary biology underscores how organisms adapt their life

strategies in response to limited resources (17, 18). While certain environments clearly necessitate a

trade-off between growth and defense strategies, in others, this distinction may become less evident.

Firstly, the resource may occasionally unlimited, the requirement of both growth and defense could be

simultaneously met. Secondly, and more commonly, this ambiguity may arise due to weak selection

efficiency or the prevalence of stochastic drift, especially noticeable in populations with small

effective sizes, which can hinder optimization. In this case, resources saved in one aspect may not be

efficiently reallocated to another. Given this perspective, some endosymbiotic and symbiotic bacteria

might grow slowly despite lacking robust defense systems, challenge the traditional notion of a

positive correlation between growth and defense. Labeling this phenomenon as a trade-up, as

previously done to describe inconsistencies with conventional trade-offs (23, 76-78), may not suffice.

Instead, with smaller effective population size and less efficient natural selection (79), both growth

and defense mechanisms may weaken simultaneously, suggesting that it might be more accurately

termed a trade-down. Expanding on this line of thought, we find an explanation for the dependence of

growth and defense on ecological contexts. In various environments, the majority of bacterial species

may exhibit differing effective population sizes. In environments where most bacterial species boast

large effective population sizes, indicating high natural selection efficiency, a trade-off between

growth and defense, resulting in a negative correlation, is likely to occur. However, in habitats

dominated by species with small populations, where genetic drift prevails, life strategies cannot be

optimized, thereby nullifying the trade-off between growth and defense.

Supplementary Information

Supplementary file 1 in docx format: Table S1. Phylogenetic signals of the bacterial features analyzed

in this study.

Supplementary file 2 in xlsx format: Table S2. A comprehensive dataset of the minimal doubling time

and antiviral defense system features for 3532 bacterial genomes. Table S3. A comprehensive dataset

of ecological characteristics, phylogenetic feature vectors, antiviral defense systems, and other

features for 1044 bacterial species.

Acknowledgments

387   We are grateful to Tian-Hao Zhao for his help in machine learning techniques and Quan-Guo Zhang

388   for hekpful comments. This article has undergone refinement by ChatGPT 3.5 to enhance language

389   and readability (80). Subsequently, the authors thoroughly reviewed the manuscript and made

390   necessary edits, taking full responsibility for the content.

Author contributions

393   Deng-Ke Niu conceived the research hypothesis; Zhi-Ling Liu and Jia Liu analyzed the data. Zhi-

394   Ling Liu and Deng-Ke Niu drafted the manuscript; all authors contributed to the final version and

395   reviewed the manuscript.

Funding

398   This study was funded by the National Natural Science Foundation of China (grant number

399   31671321).

Availability of data and materials

402   The datasets supporting the conclusions of this article are included within the article and its

403 Supplementary files.

Declarations

Ethics approval and consent to participate

407 All authors consent to participate in this study.

Consent for publication

409 All authors give their consent for publication.

Competing interests

411 The authors declare no competing interests.

References

1. López-García P, Gutiérrez-Preciado A, Krupovic M, Ciobanu M, Deschamps P, Jardillier L, López-Pérez
M, Rodríguez-Valera F, Moreira D. 2023. Metagenome-derived virus-microbe ratios across ecosystems.
The ISME Journal 17:1552-1563.

2. Roux S, Brum JR. 2023. Counting dots or counting reads? Complementary approaches to estimate virus-
to-microbe ratios. The ISME Journal 17:1521-1522.

3. Wommack KE, Colwell RR. 2000. Virioplankton: Viruses in aquatic ecosystems. Microbiology and
Molecular Biology Reviews 64:69-114.

4. Suttle CA. 2007. Marine viruses - major players in the global ecosystem. Nature Reviews Microbiology
5:801-812.

5. Koonin EV, Makarova KS, Wolf YI. 2017. Evolutionary genomics of defense systems in archaea and
bacteria. Annual Review of Microbiology 71:233-261.

6. Bernheim A, Sorek R. 2020. The pan-immune system of bacteria: antiviral defence as a community
resource. Nature Reviews Microbiology 18:113-119.

7. Stern A, Sorek R. 2011. The phage-host arms race: Shaping the evolution of microbes. BioEssays 33:43-
51.

8. Rostøl JT, Marraffini L. 2019. (Ph)ighting phages: how bacteria resist their parasites. Cell Host &
Microbe 25:184-194.

9. Makarova KS, Wolf YI, Iranzo J, Shmakov SA, Alkhnbashi OS, Brouns SJJ, Charpentier E, Cheng D,
Haft DH, Horvath P, Moineau S, Mojica FJM, Scott D, Shah SA, Siksnys V, Terns MP, Venclovas C,
White MF, Yakunin AF, Yan W, Zhang F, Garrett RA, Backofen R, van der Oost J, Barrangou R,
Koonin EV. 2020. Evolutionary classification of CRISPR-Cas systems: a burst of class 2 and derived
variants. Nature Reviews Microbiology 18:67-83.

10. Roberts RJ, Vincze T, Posfai J, Macelis D. 2015. REBASE—a database for DNA restriction and
modification: enzymes, genes and genomes. Nucleic Acids Research 43:D298-D299.

11. Ferenci T. 2016. Trade-off mechanisms shaping the diversity of bacteria. TRENDS IN

439    MICROBIOLOGY 24:209-223.

440 12. Thingstad TF. 2022. Competition-defense trade-offs in the microbial world. Proceedings of the National
441    Academy of Sciences 119:e2213092119.

442 13. Winter C, Bouvier T, Weinbauer MG, Thingstad TF. 2010. Trade-offs between competition and defense
443    specialists among unicellular planktonic organisms: the "killing the winner" hypothesis revisited.
444    Microbiology and Molecular Biology Reviews 74:42-57.

445 14. Pančić M, Kiørboe T. 2018. Phytoplankton defence mechanisms: traits and trade-offs. Biological
446    Reviews 93:1269-1303.

447 15. Liu Z-L, Hu E-Z, Niu D-K. 2023. Investigating the relationship between CRISPR-Cas content and
448    growth rate in bacteria. Microbiology Spectrum 11:e03409-22.

449 16. Roff DA, Fairbairn DJ. 2007. The evolution of trade-offs: where are we? Journal of Evolutionary
450    Biology 20:433-447.

451 17. Garland T, Jr. 2014. Trade-offs. Current Biology 24:R60-R61.

452 18. Garland T, Jr., Downs CJ, Ives AR. 2022. Trade-offs (and constraints) in organismal biology.
453    Physiological and Biochemical Zoology 95:82-112.

454 19. Herms DA, Mattson WJ. 1992. The dilemma of plants: to grow or defend. The Quarterly Review of
455    Biology 67:283-335.

456 20. Lenski RE. 1988. Experimental studies of pleiotropy and epistasis in *Escherichia coli*. I. Variation in
457    competitive fitness among mutants resistant to virus T4. Evolution 42:425-432.

458 21. Buckling A, Wei Y, Massey RC, Brockhurst MA, Hochberg ME. 2005. Antagonistic coevolution with
459    parasites increases the cost of host deleterious mutations. Proceedings of the Royal Society B: Biological
460    Sciences 273:45-49.

461 22. Hall AR, De Vos D, Friman V-P, Pirnay J-P, Buckling A. 2012. Effects of sequential and simultaneous
462    applications of bacteriophages on populations of *Pseudomonas aeruginosa* in vitro and in wax moth
463    larvae. Applied and Environmental Microbiology 78:5646-5652.

464 23. Burmeister AR, Turner PE. 2020. Trading-off and trading-up in the world of bacteria-phage evolution.
465    Current Biology 30:R1120-R1124.

466 24. Castledine M, Padfield D, Sierocinski P, Soria Pascual J, Hughes A, Mäkinen L, Friman V-P, Pirnay J-P,
467    Merabishvili M, de Vos D, Buckling A. 2022. Parallel evolution of *Pseudomonas aeruginosa* phage
468    resistance and virulence loss in response to phage treatment in vivo and in vitro. eLife 11:e73679.

469 25. Bohannan BJM, Lenski RE. 2000. Linking genetic change to community evolution: insights from studies
470    of bacteria and bacteriophage. Ecology Letters 3:362-377.

471 26. Lennon JT, Khatana SAM, Marston MF, Martiny JBH. 2007. Is there a cost of virus resistance in marine
472    cyanobacteria? The ISME Journal 1:300-312.

473 27. Bohannan BJM, Kerr B, Jessup CM, Hughes JB, Sandvik G. 2002. Trade-offs and coexistence in
474    microbial microcosms. Antonie Van Leeuwenhoek International Journal of General and Molecular
475    Microbiology 81:107-115.

476 28. Jessup CM, Bohannan BJM. 2008. The shape of an ecological trade-off varies with environment.
477    Ecology Letters 11:947-959.

478 29. Quance MA, Travisano M. 2009. Effects of temperature on the fitness cost of resistance to bacteriophage
479    T4 in *Escherichia coli*. Evolution 63:1406-1416.

480   30.   Burmeister AR, Fortier A, Roush C, Lessing AJ, Bender RG, Barahman R, Grant R, Chan BK, Turner
481   PE. 2020. Pleiotropy complicates a trade-off between phage resistance and antibiotic resistance.
482   Proceedings of the National Academy of Sciences 117:11207-11216.

483   31.   Moulton-Brown CE, Friman V-P. 2018. Rapid evolution of generalized resistance mechanisms can
484   constrain the efficacy of phage–antibiotic treatments. Evolutionary Applications 11:1630-1641.

485   32.   Koskella B, Taylor TB, Bates J, Buckling A. 2011. Using experimental evolution to explore natural
486   patterns between bacterial motility and resistance to bacteriophages. The ISME Journal 5:1809-1817.

487   33.   Yang JW, Chang FH, Yeh YC, Tsai AY, Chiang KP, Shiah FK, Gong GC, Hsieh CH. 2023. Trade-offs
488   between competitive ability and resistance to top-down control in marine microbes. MSYSTEMS
489   8:e01017-22.

490   34.   Georjon H, Bernheim A. 2023. The highly diverse antiphage defence systems of bacteria. Nature
491   Reviews Microbiology 21:686-700.

492   35.   Tesson F, Bernheim A. 2023. Synergy and regulation of antiphage systems: toward the existence of a
493   bacterial immune system? Current Opinion in Microbiology 71:102238.

494   36.   Weissman JL, Hou S, Fuhrman JA. 2021. Estimating maximal microbial growth rates from cultures,
495   metagenomes, and single cells via codon usage patterns. Proceedings of the National Academy of
496   Sciences 118:e2016810118.

497   37.   Couvin D, Bernheim A, Toffano-Nioche C, Touchon M, Michalik J, Néron B, Rocha EPC, Vergnaud G,
498   Gautheret D, Pourcel C. 2018. CRISPRCasFinder, an update of CRISRFinder, includes a portable
499   version, enhanced performance and integrates search for Cas proteins. Nucleic Acids Research 46:W246-
500   W251.

501   38.   Tesson F, Hervé A, Mordret E, Touchon M, d'Humières C, Cury J, Bernheim A. 2022. Systematic and
502   quantitative view of the antiviral arsenal of prokaryotes. Nature Communications 13:2561.

503   39.   Madin JS, Nielsen DA, Brbic M, Corkrey R, Danko D, Edwards K, Engqvist MKM, Fierer N,
504   Geoghegan JL, Gillings M, Kyrpides NC, Litchman E, Mason CE, Moore L, Nielsen SL, Paulsen IT,
505   Price ND, Reddy TBK, Richards MA, Rocha EPC, Schmidt TM, Shaaban H, Shukla M, Supek F, Tetu
506   SG, Vieira-Silva S, Wattam AR, Westfall DA, Westoby M. 2020. A synthesis of bacterial and archaeal
507   phenotypic trait data. Scientific Data 7:170.

508   40.   Parks DH, Chuvochina M, Rinke C, Mussig AJ, Chaumeil P-A, Hugenholtz P. 2022. GTDB: an ongoing
509   census of bacterial and archaeal diversity through a phylogenetically consistent, rank normalized and
510   complete genome-based taxonomy. Nucleic Acids Research 50:D785-D794.

511   41.   Brbić M, Piškorec M, Vidulin V, Criško A, Šmuc T, Supek F. 2016. The landscape of microbial
512   phenotypic traits and associated genes. Nucleic Acids Research 44:10074-10090.

513   42.   Weissman JL, Laljani RMR, Fagan WF, Johnson PLF. 2019. Visualization and prediction of CRISPR
514   incidence in microbial trait-space to identify drivers of antiviral immune strategy. The ISME Journal
515   13:2589-2602.

516   43.   Debastiani VJ, Bastazini VAG, Pillar VD. 2021. Using phylogenetic information to impute missing
517   functional trait values in ecological databases. Ecological Informatics 63:101315.

518   44.   Santos T. 2018. Phylogenetic eigenvectors regression and phylogentic signal-representation curve, v0.3.
519   https://cran.r-project.org/package=PVR.

520   45.   Stekhoven DJ, Bühlmann P. 2011. MissForest—non-parametric missing value imputation for mixed-type
521   data. Bioinformatics 28:112-118.

46.   Liaw A, Wiener M. 2002. Classification and regression by randomForest. R News 2:18-22.

47. Ho LST, Ane C. 2014. A linear-time algorithm for Gaussian and non-Gaussian trait evolution models.
Systematic Biology 63:397-408.

48. Garland T, Jr., Dickerman AW, Janis CM, Jones JA. 1993. Phylogenetic analysis of covariance by
computer simulation. Systematic Biology 42:265-292.

49. Ives AR, Garland T, Jr. 2010. Phylogenetic logistic regression for binary dependent variables. Systematic
Biology 59:9-26.

50. Felsenstein J. 1985. Phylogenies and the comparative method. The American Naturalist 125:1-15.

51. Symonds MRE, Blomberg SP. 2014. A primer on phylogenetic generalised least squares, p 105-130. *In*
Garamszegi LZ (ed), Modern Phylogenetic Comparative Methods and Their Application in Evolutionary
Biology: Concepts and Practice doi:10.1007/978-3-662-43550-2_5. Springer Berlin Heidelberg, Berlin,
Heidelberg.

52. Chen Z-L, Guo H-J, Niu D-K. 2023. Dependent variable selection in phylogenetic generalized least
squares regression analysis under Pagel's lambda model. bioRxiv
doi:10.1101/2023.05.21.541623:2023.05.21.541623.

53. Hu E-Z, Lan X-R, Liu Z-L, Gao J, Niu D-K. 2022. A positive correlation between GC content and
growth temperature in prokaryotes. BMC Genomics 23:110.

54. Burstein D, Sun CL, Brown CT, Sharon I, Anantharaman K, Probst AJ, Thomas BC, Banfield JF. 2016.
Major bacterial lineages are essentially devoid of CRISPR-Cas viral defence systems. Nature
Communications 7:10613.

55. Siozios S, Jimenez PN, Azagi T, Sprong H, Frost CL, Parratt SR, Taylor G, Brettell L, Liew KC, Croft L,
King KC, Brockhurst MA, Hypša V, Novakova E, Darby AC, Hurst GD. 2023. Genome dynamics across
the evolutionary transition to endosymbiosis. bioRxiv
doi:10.1101/2023.05.02.539033:2023.05.02.539033.

56. Beavogui A, Lacroix A, Wiart N, Poulain J, Delmont TO, Paoli L, Wincker P, Oliveira PH. 2024. The
defensome of complex bacterial communities. Nature Communications 15:2146.

57. Leplae R, Geeraerts D, Hallez R, Guglielmini J, Drèze P, Van Melderen L. 2011. Diversity of bacterial
type II toxin–antitoxin systems: a comprehensive search and functional analysis of novel families.
Nucleic Acids Research 39:5513-5525.

58. Makarova KS, Wolf YI, Koonin EV. 2009. Comprehensive comparative-genomic analysis of Type 2
toxin-antitoxin systems and related mobile stress response systems in prokaryotes. Biology Direct 4:19.

59. Pandey DP, Gerdes K. 2005. Toxin-antitoxin loci are highly abundant in free-living but lost from host-
associated prokaryotes. Nucleic Acids Res 33:966-76.

60. Puigbò P, Makarova KS, Kristensen DM, Wolf YI, Koonin EV. 2017. Reconstruction of the evolution of
microbial defense systems. BMC Evolutionary Biology 17:94.

61. Sender R, Fuchs S, Milo R. 2016. Are we really vastly outnumbered? Revisiting the ratio of bacterial to
host cells in humans. Cell 164:337-340.

62. Sender R, Fuchs S, Milo R. 2016. Revised estimates for the number of human and bacteria cells in the
body. PLOS Biology 14:e1002533.

63. Donaldson GP, Lee SM, Mazmanian SK. 2016. Gut biogeography of the bacterial microbiota. Nature
Reviews Microbiology 14:20-32.

64. Lourenço M, Chaffringeon L, Lamy-Besnier Q, Pédron T, Campagne P, Eberl C, Bérard M, Stecher B,
Debarbieux L, De Sordi L. 2020. The spatial heterogeneity of the gut limits predation and fosters

coexistence of bacteria and bacteriophages. Cell Host & Microbe 28:390-401.e5.

65.   Guerin E, Hill C. 2020. Shining light on human gut bacteriophages. Frontiers in Cellular and Infection
Microbiology 10:481.

66.   Shkoporov AN, Turkington CJ, Hill C. 2022. Mutualistic interplay between bacteriophages and bacteria
in the human gut. Nature Reviews Microbiology 20:737-749.

67.   Shkoporov AN, Hill C. 2019. Bacteriophages of the human gut: the "known unknown" of the mcrobiome.
Cell Host & Microbe 25:195-209.

68.   Jończyk-Matysiak E, Łodej N, Kula D, Owczarek B, Orwat F, Międzybrodzki R, Neuberg J, Bagińska N,
Weber-Dąbrowska B, Górski A. 2019. Factors determining phage stability/activity: challenges in
practical phage application. Expert Review of Anti-infective Therapy 17:583-606.

69.   Dlamini SB, Gigante AM, Hooton SPT, Atterbury RJ. 2023. Efficacy of Different Encapsulation
Techniques on the Viability and Stability of Diverse Phage under Simulated Gastric Conditions.
Microorganisms 11:2389.

70.   Yang Y, Du H, Zou G, Song Z, Zhou Y, Li H, Tan C, Chen H, Fischetti VA, Li J. 2023. Encapsulation
and delivery of phage as a novel method for gut flora manipulation in situ: A review. Journal of
Controlled Release 353:634-649.

71.   Whittle M, Barreaux AMG, Bonsall MB, Ponton F, English S. 2021. Insect-host control of obligate,
intracellular symbiont density. Proceedings of the Royal Society B: Biological Sciences 288:20211993.

72.   Whittle M, Bonsall MB, Barreaux AMG, Ponton F, English S. 2023. A theoretical model for host-
controlled regulation of symbiont density. Journal of Evolutionary Biology 36:1731-1744.

73.   Makarova KS, Wolf YI, Snir S, Koonin EV. 2011. Defense islands in bacterial and archaeal genomes
and prediction of novel defense systems. Journal of Bacteriology 193:6039-6056.

74.   Oliveira PH, Touchon M, Rocha EPC. 2014. The interplay of restriction-modification systems with
mobile genetic elements and their prokaryotic hosts. Nucleic Acids Research 42:10618-10631.

75.   Bernheim A, Bikard D, Touchon M, Rocha EPC. 2019. Atypical organizations and epistatic interactions
of CRISPRs and cas clusters in genomes and their mobile genetic elements. Nucleic Acids Research
48:748-760.

76.   Behringer MG, Ho W-C, Miller SF, Worthan SB, Cen Z, Stikeleather R, Lynch M. 2024. Trade-offs,
trade-ups, and high mutational parallelism underlie microbial adaptation during extreme cycles of feast
and famine. Current Biology 34:1403-1413.e5.

77.   Reding-Roman C, Hewlett M, Duxbury S, Gori F, Gudelj I, Beardmore R. 2017. The unconstrained
evolution of fast and efficient antibiotic-resistant bacterial genomes. Nature Ecology & Evolution 1:0050.

78.   Reznick D, King K. 2017. Antibiotic resistance: Evolution without trade-offs. Nature Ecology &
Evolution 1:0066.

79.   Moran NA. 1996. Accelerated evolution and Muller's rachet in endosymbiotic bacteria. Proceedings of
the National Academy of Sciences 93:2873-2878.

80.   OpenAI. 2024. ChatGPT (4) [Large language model]. https://chat.openai.com.

Table 1. Phylogenetic logistic regression of defense system presence/absence on bacterial habitats

Dependent variable ~ independent variable	Slope	P	$P_{adjusted}$
Presence/absence of defense ~ Known habitats = insect endosymbiont	−16.108	5.6×10^{-16}	2.3×10^{-15}
Presence/absence of defense ~ Known habitats = endosymbiont	0.215	0.543	0.543
Presence/absence of defense ~ Biotic relationship = symbiotic	−7.387	6.6×10^{-10}	1.3×10^{-9}
Presence/absence of defense ~ Host = insects general	−8.558	3.4×10^{-9}	4.5×10^{-9}

The default parameter logistic MPLE was used in the phylogenetic logistic regression of the

relationships. In the regression analysis, the presence of defense systems was assigned as 1 and the

absence was assigned as 0. The habitats are measured by the overall maximal confidence score

described in Materials and Methods section. The ratio of samples with defense systems to those

without related systems was 965 vs 79. The $P_{adjusted}$ value represents the corrected P-value computed

using the Benjamini & Hochberg method.

Table 2. PGLS regressions of ecological features on the defense system abundance in bacteria

	Slope	P	$P_{adjusted}$
Known habitats = insect endosymbiont	−0.001	0.001	0.004
Known habitats = endosymbiont	−0.001	4.4×10^{-4}	0.002
Biotic relationship = symbiotic	−0.002	0.022	0.031
Ecosystem type = circulatory system	−0.001	0.019	0.029
Ecosystem subtype = blood	−0.001	0.068	0.084
Habitat = host associated	−0.002	0.002	0.005
Host = insects general	−0.002	0.002	0.005
Ecosystem category = mammals	−0.002	0.015	0.024
Known habitats = host	−0.002	0.008	0.017
Ecosystem = host associated	−0.002	0.011	0.019

Habitat = free living	0.002	0.007	0.016
Habitat = aquatic	0.001	0.054	0.071
Known habitats = freshwater	0.001	0.190	0.210
Ecosystem category = aquatic	0.001	0.284	0.284
Energy source = photoautotroph	0.001	0.251	0.264
Energy source = phototroph	0.003	8.5×10^{-6}	9.0×10^{-5}
Energy source = chemolithotroph	0.001	0.105	0.123
Habitat = multiple	0.002	0.001	0.004
Shape = filamentous	0.002	3.8×10^{-6}	8.0×10^{-5}
Mobility	0.002	0.010	0.019
Ecosystem category = bioremediation	0.002	3.9×10^{-5}	2.7×10^{-4}

Due to the weaker phylogenetic signal of defense system abundance in bacteria compared to

ecological features (Table S1), it was utilized as the independent variable in the PGLS regression

analysis. Ecological features were measured using the overall maximal confidence score described in

the Materials and Methods section. A total of 1044 bacterial species were included in these

regressions. The P_{adjusted} value represents the corrected P-value computed using the Benjamini &

Hochberg method. PGLS, phylogenetic generalized least-squares.

Table 3. PGLS regressions of bacterial minimal doubling time on ecological features

	Slope	P	P_{adjusted}
Known habitats = insect endosymbiont	1.809	1.0×10^{-5}	3.0×10^{-5}

Known habitats = endosymbiont	0.799	0.004	0.006
Host = insects general	1.07	3.9×10^{-9}	4.1×10^{-8}
Biotic relationship = symbiotic	0.609	3.3×10^{-5}	8.7×10^{-5}
Ecosystem = host associated	−0.859	8.4×10^{-8}	5.9×10^{-7}
Known habitats = host	−0.625	9.2×10^{-5}	1.9×10^{-4}
Habitat = host associated	−0.478	0.004	0.006
Ecosystem type = circulatory system	−0.197	0.325	0.341
Ecosystem subtype = blood	−0.27	0.097	0.113
Ecosystem category = mammals	−0.111	0.503	0.503
Habitat = free living	0.253	0.089	0.110
Habitat = aquatic	0.907	9.7×10^{-7}	3.4×10^{-6}
Known habitats = freshwater	0.693	6.7×10^{-5}	1.6×10^{-4}
Ecosystem category = aquatic	0.92	7.7×10^{-7}	3.2×10^{-6}
Energy source = photoautotroph	1.235	7.5×10^{-7}	3.2×10^{-6}
Energy source = phototroph	0.422	0.035	0.049
Energy source = chemolithotroph	1.518	2.5×10^{-12}	5.3×10^{-11}
Habitat = multiple	−0.656	2.2×10^{-4}	4.2×10^{-4}
Shape = filamentous	0.845	0.002	0.004
Mobility	−0.305	0.081	0.106
Ecosystem category = bioremediation	0.293	0.311	0.341

Since the minimal doubling time of bacteria exhibits a stronger phylogenetic signal than the

ecological features (Table S1), we employed it as the dependent variable in the PGLS regression

analysis. Ecological features were measured using the overall maximal confidence score described in

the Materials and Methods section. A total of 1044 bacterial species were included in these

regressions. The P_{adjusted} value represents the corrected P-value computed using the Benjamini &

Hochberg method. PGLS, phylogenetic generalized least-squares.

Table 4. PGLS regressions of bacterial minimal doubling time on the abundance of defense systems

	n	Slope	P
Host = insects general	30	0.092	0.478
Biotic relationship = symbiotic	98	−0.009	0.210
Ecosystem = host associated	499	−0.015	0.028
Known habitats = host	518	−0.016	0.021
Habitat = host associated	434	−0.017	0.020
Ecosystem type = circulatory system	52	−0.012	0.194
Ecosystem subtype = blood	92	−0.018	0.104
Ecosystem category = mammals	81	−0.019	0.129
Habitat = free living	605	0.013	0.008
Habitat = aquatic	194	0.007	0.265
Known habitats = freshwater	147	0.001	0.901
Ecosystem category = aquatic	302	0.007	0.243
Energy source = photoautotroph	20	0.028	0.023
Energy source = chemolithotroph	22	0.01	0.524
Mobility	410	0.02	0.002
Habitat = multiple	206	0.01	0.264

Shape = filamentous	22	−0.001	0.837

Since the minimal doubling time of bacteria exhibits a stronger phylogenetic signal than the

abundance of defense systems (Table S1), we used it as the dependent variable in the PGLS

regression analysis. We discarded the ecological features where less than 20 bacterial samples were

obtained. We did adjust the *P*-values in this table using the Benjamini & Hochberg method because

the multiple regressions were not performed on the same dataset. PGLS, phylogenetic generalized

least-squares.

Figure 1. Importance ranking of ecological features for defense system presence/absence in bacteria.

The vertical axis represents the names of ecological features, with higher ranks indicating a greater

impact on the presence or absence of the defense system. MeanDecreaseGini, displayed on the

horizontal axis, quantifies feature importance, with higher values indicating greater significance. The

figure displays the top thirty features in the model. Features starting with the letter "c" represent

eigenvectors representing phylogenetic relationships.

Figure 2. Importance ranking of ecological features for defense system abundance in bacteria. The

vertical axis represents the names of ecological features, with higher ranks indicating a greater impact

on the abundance of the defense system. IncNodePurity, displayed on the horizontal axis, quantifies

feature importance, with higher values indicating greater significance. The figure displays the top

thirty features in the model. Features starting with the letter "c" represent eigenvectors representing

phylogenetic relationships.