Suboptimal refeeding compensates stunting in a mouse model of juvenile malnutrition

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Short title: Suboptimal refeeding after juvenile malnutrition

List of abbreviations:

GTT: Glucose tolerance test

IGF-1: Insulin-like growth factor 1

ITT: Insulin tolerance test

Lp^{WJL}: *Lactiplantibacillus plantarum* WJL

LPD: Low-protein diet

PC: Principal component

PEM: Protein energy malnutrition

MWD: Modified Western Diet

WA: weight-for-age relationship

WD: Total New Western Diet

Abstract:

Background: The highest rate of growth in mammals occurs in early life, particularly after weaning. Growth is conditioned by the nutritional status. Indeed, restriction of dietary protein in early life leads to wasting and/or stunting.

Objective: Here, we developed a mouse model of juvenile protein malnutrition triggering stunting and studied the physiological effects of refeeding using various diets and interventions.

Methods: In a first intervention, we refed the mice with an optimal diet (breeding diet, rich in protein and fiber). We then treated the mice during the refeeding phase with *Lactiplantibacillus plantarum* WJL (Lp^{WJL}), a previously described bacterial strain that has the ability to stimulate growth via the somatotropic axis in early life upon chronic malnutrition. Finally, we established a model of suboptimal refeeding, upon which the mice were given a western diet (33% kcal from fat; 17% kcal from protein) or an isocaloric modified western diet low in proteins (33% kcal from fat; 8% kcal from protein) after juvenile protein malnutrition.

Results: We found that, in females, optimal growth was restored by control diet refeeding. In males, control diet refeeding after a five-week protein restriction was not enough to catch up growth retardation. No supplementary beneficial effect was found associated to the microbial intervention in this context. Surprisingly, our results showed that, in males, suboptimal refeeding with a diet rich in fat but low in protein was sufficient to buffer the deleterious effects of protein restriction on growth. However, this macroscopic benefit was associated to metabolic alteration. While Lp^{WJL} treatment had no effect on growth per se, we found that bacterial treatment further impaired glycemic control upon suboptimal refeeding.

Conclusions: Overall, we describe a novel model of juvenile protein energy malnutrition, where growth can be caught up by suboptimal refeeding.

Keywords: Mice; protein energy malnutrition; double burden of malnutrition; stunting; probiotic intervention

Introduction

Early postnatal malnutrition is a burden in many developing countries, and is considered the most worrying factor of death for young infants under five years old. Consequences of malnutrition include wasting and stunting, which are defined respectively weight-for-height and height-for-age below two standard deviations from the median of the World Health Organization Child Growth Standards. Wasting is indicative of acute growth disturbance from malnutrition, whereas stunting is more symptomatic of chronic malnutrition, with impaired long-term growth (1). In 2020, about 150 million children under five years of age were stunted (2). Stunting is directly caused by an inadequate intake of nutrients, especially dietary protein (3). Besides stunting, severe malnutrition in early life is associated to metabolic disturbances that persist later in life (4,5). Extensive epidemiological studies have demonstrated that malnutrition during early life has detrimental effects on health, which can last throughout the life of the malnourished person (6). This is called nutritional or metabolic programming, a process through which metabolic and physiological adaptations in response to a poor diet persist even after the nutritional stress is removed (7–10). The first 1,000 days of life are crucial for growth because of the high developmental plasticity during that time, after which stunting generally becomes irreversible and leads to growth impairment in adulthood

(11). Recently, three articles of large human cohorts have highlighted the importance of early life interventions to prevent wasting and stunting in more than 30 cohorts of malnourished children (12–14).

In this context, establishing animal models of early life malnutrition seems crucial to understand the mechanisms behind the observed adverse consequences. Previously, we established a model of mouse juvenile malnutrition, upon which mice were fed a low-protein low-fat diet from weaning (post-natal day 21, P21) to eight weeks of age (15,16), which is the rough physiological equivalent of 1 year to 20 years of age in humans (17). We showed that supplementation with the probiotic *Lactiplantibacillus plantarum* WJL (Lp^{WJL}) sustains postnatal growth of malnourished animals by stimulating anabolic metabolism and hormonal changes, namely the somatotropic axis via the secretion of insulin-like growth factor 1 (IGF-1) (15,16). However, knowledge on the long-term consequences of early malnutrition after refeeding is lacking. Indeed, the first intervention to stop malnutrition is resolving the nutritional stress associated to the lack of protein by providing an optimized diet.

Using a mouse model of juvenile protein energy malnutrition, we sought to investigate the consequences on growth catch-up after refeeding with either an optimal or suboptimal diets (richer in fat and poorer in protein). While growth was not affected by a suboptimal diet, we also explored the metabolic alterations associated to this switch.

METHODS

Animals

Male and female specific pathogen-free C57Bl/6J mice, aged 9 to 10 weeks, were purchased from Charles River (L'Arbresle, France). After one week of acclimatization

at the conventional animal room, they were mated in our animal facility and offspring pups were used for experiments. Mice were kept in Innorack IVC Mouse 3.5 disposable cages (Inovive, USA), exposed to 12:12 hours light-dark cycles, supplied with tap water and given free access to food. All experiments were performed in accordance with the European Community Council Directive of September 22, 2010 (2010/63/EU) regarding the protection of animals used for experimental and other scientific purposes. The research project was approved by a local animal care and user committee (C2EA015) and subsequently authorized by the French Ministry of Research (APAFIS#22788-2019091713204925 v8 and APAFIS#32380-2021061809124474 v4).

Experimental design

Specific-pathogen free C57Bl/6J mice were mated. After the delivery, the litter size was reduced to six offspring per dam. From birth to post-natal day 14 (P14), the dams were given free access to breeding diet (control diet; Altromin 1310). At P14, the pups were kept with their mother, but food was changed according to the experimental groups. All diets were irradiated at 40 kGy before use. Composition of the experimental diets is given in Table 1. At P21, mice were weaned on the diet introduced at P14. Body weight and length were controlled once a week. For the measurement of body length, mice were briefly anesthetized with isoflurane and length was measured using a graduated ruler. Body length was defined as the nose-to-anus distance. At the end of the experiments, mice were killed by cervical dislocation. Tissues were sampled, weighed and measured, and snap-frozen in liquid nitrogen.

Experiment 1 (male and female mice)

A control group was given control diet (Altromin 1310) from P14 to P91. Experimental groups were given low-protein diet (LPD, composition in Table 1), which is a modified

isocaloric version of AIN-93G diet. LPD was custom-made by Sciences de l'Animal et de l'Aliment de Jouy unit (SAAJ, INRAE UE1298, Jouy-en-Josas, France). Switch to control diet was operated in 5 different groups from P28 to P56, then mice were kept on control diet until P91. Blood samples were taken from the submandibular vein three times: at diet switch, two weeks after the switch, and at P91. Time-matched samples were taken in the control group.

Experiment 2 (males only)

Three groups were formed: a control group fed control diet until P119; a low-protein group fed LPD from P14 to P119; and a refeeding group, fed LPD from P14 to P56, then control diet from P56 to P119. From P56 to P119, two treatment arms were formed in each group: one receiving a placebo solution (maltodextrin) and one receiving 10^8 CFU *Lactiplantibacillus plantarum* WJL (Lp^{WJL}) as described previously (16). Solutions were given per os 5 times a week, in the morning. At P112, mice were taken to ANIPHY Platform (Lyon) and put in the Minispec LF90 II (Bruker, Wissembourg, France) for non-invasive determination of body composition using NMR. Mice were taken back to our animal facility and given the experimental diet until P119.

Experiment 3 (males only)

Groups were formed as described in Figure 5. A control group was fed control diet from P14 to P119. Experimental diets were control diet (Altromin 1310), LPD, Total New Western Diet (WD; Teklad TD.140203) and a modified isocaloric version of WD (MWD), with reduced protein content. MWD was custom-made for the experiment. WD and MWD were purchased from Envigo (Gannat, France). WD is based on Total Western Diet, as described in (18), with the modifications described by Monsanto et al.

(19). Composition of the diets is given in Table 1. As in Experiment 2, mice were given either a placebo or an Lp^{WJL} solution, from P35 to P119.

Glucose and insulin tolerance tests

In Experiment 3, at P91 and P105, mice underwent respectively a glucose (GTT) or insulin tolerance test (ITT). Mice were food-deprived for 6 hours before the start of the tests. For the GTT, mice were given an intraperitoneal (i.p.) injection of a 20% sterile glucose solution (1 $g.kg^{-1}$ body weight). For the ITT, mice were given an i.p. injection of human insulin (Novorapid, Novo Nordisk) at 0.75 $U.kg^{-1}$ body weight. Glycemia was monitored at the tip of the tail vein at times 0, 15, 30, 45, 60, 90 and 120 minutes post-injection using a glucometer (One Touch Verio Reflect, Life Scan).

IGF-1 measurement in serum

Blood samples were collected from the submandibular vein at the time of diet switch, two weeks after the switch and at the end of the experiment, into a sample tube containing a clotting activator (Micro sample tube serum gel, 1.1 mL, Sarstedt). After centrifugation at 10,000 g (Eppendorf 5424 R) for 5 minutes, sera were collected into sterile 1.5 mL microtubes and kept at -70°C for further use. IGF-1 in serum was measured using Mouse/Rat IGF-1 Quantikine ELISA (R&D Systems) according to manufacturer's instructions.

Data treatment and statistics

Statistical analysis was performed using GraphPad Prism software version 10.0 (GraphPad, San Diego, California, USA). For Experiment 1, we used one-way ANOVA followed by Tukey's multiplicity-adjusted test. Growth catch-up slopes were compared using ANCOVA. Principal Component (PC) Analysis was performed based on the morphological features measured, with PC selection based on percent of total explained

variance. For Experiment 2, we used two-way ANOVA followed by multiplicity-adjusted Šídák's test. For Experiment 3, we used one-way ANOVA followed by Tukey's multiplicity-adjusted test when comparing to control and two-way ANOVA followed by multiplicity-adjusted Šídák's test when comparing treated groups. Unless stated otherwise, results are presented as mean ± SEM. *P*-values are reported after adjustment for multiple comparison. A value of $P < 0.05$ was considered statistically significant.

RESULTS

Optimal refeeding after long-term juvenile malnutrition is insufficient to fully recover growth in males, but not in females

In order to model protein energy malnutrition (PEM), we designed an AIN-93G-based isocaloric low-protein diet (LPD), with 5.1% total kcal coming from protein (Table 1). We first investigated how relevant our model of malnutrition was to human disease. Indeed, although several models have been published (20,21), there is no standard murine model of PEM. We first characterized the impact on weight gain and linear growth in C57Bl/6J mice fed an LPD, starting at P14 and going on after weaning (which took place at P21). We constituted 5 different groups, which corresponded to 5 different times on LPD, going from 2 weeks to 6 weeks. From P28 to P56, mice were refed with control chow diet. To make a parallel with juvenile human malnutrition, we defined stunting and underweight in our model as body size and weight inferior to two standard deviations from the mean of a control group of mice fed chow diet, i.e., optimal diet for mouse growth (dashed lines, Figure 1A-D). As expected, LPD-fed mice had severe growth retardation following weaning, associated to reduced body weight (Figure 1A-D). Since definition of wasting (low weight-for-length) in humans is

complex and requires a Box-Cox power transformation (22), and growth does not occur in the same manner in mice and humans, we plotted length vs. weight in our experimental animals. We found that weight and length are linearly correlated during growth (Figure 1E-H). In LPD-fed mice, the slope was significantly smaller than in age-matched control-fed mice (males and females: ANCOVA: $P < 10^{-4}$), meaning that the mice were wasted (Figure 1E-F). In females, when mice were refed a control chow diet after LPD, growth and weight catch-up occurred within three weeks of diet switch (Figure 1B, D, H). However, in males, the longer the mice stood on LPD, the slower growth and weight catch-up occurred (Figure 1A, C). Indeed, when male mice were fed on LPD from P14 to P56, they were still stunted at P91 (Figure 1C), despite the removal of the nutritional stress. It is noteworthy that, despite the prevalence of stunting in male mice at P91, mice were not wasted after refeeding (Figure 1G-H).

Using the weight-for-age (WA) relationship described by Anstead et al. (20), based on the classification by Gómez et al. (23), we calculated WA in all our groups (Figure 2A-B). After weaning, WA was inferior to 50% in LPD-fed male mice (Figure 2A), showing that the mice suffered from severe malnutrition. In females, however, LPD induced severe malnutrition, but the severity was corrected with time, reaching moderate malnutrition at P56 (Figure 2B). Refeeding with chow diet in male mice only partially corrected the severity of malnutrition, since RF42 mice had a WA score of 85.1 ± 2.1% (5% confidence interval), going as low as 81.6 ± 3.0% for RF56 mice, meaning they still faced mild malnutrition. As was the case previously, refeeding malnourished female mice completely corrected early-life malnutrition (Figure 2B). Overall, our data show a good example of nutritional imprinting of PEM, especially in males. Growth in mice occurs with a linear phase that lasts about two weeks after weaning followed by a

plateau phase (24). We observed that this pattern was similar after refeeding. We thus compared growth catch-up by comparing the slopes following refeeding (or weaning, in the case of the control group) (Figure 2C-D). In males, growth catch-up differed significantly between the groups (P = 0.0205; ANCOVA). After weaning on control diet, mice gained on average 0.11 cm.day^{-1}. This value was similar when mice were refed after only one week on LPD (group RF28), but differed significantly with all other groups, which had a catch-up rate of about 0.08 cm.day^{-1} (Figure 2C). Thus, in males, growth catch-up after refeeding is impaired by long-term exposure to LPD in early life. In females, however, growth catch-up did not differ significantly between groups (P = 0.0504; ANCOVA; Figure 2D), showing a sex-dependent phenotype related to resilience to LPD feeding in early life.

Furthermore, we measured weight of liver, soleus muscle and subcutaneous and perigonadal fat, and length of femur, tibia, small intestine and colon (Figure 3). In males, we observed significantly reduced size of femur and tibia (Figure 3E-F), alongside reductions in intestinal and colon sizes, which were all the more marked as the time on LPD was long (Figure 3G-H). When we plotted the totality of the measured parameters using Principal Component Analysis, we saw that values were mostly spread across PC1, which accounted for 30.44% of the total variance (Figure 3I). While control animals clustered separately from refed mice, the latter formed a cluster, showing a biological difference between control and refed mice. In contrast, in females, we only saw slight variations of fat deposits, but all points clustered together in a Principal Component Analysis (Supplementary Figure 1).

Insulin-like Growth Factor 1 (IGF-1) is a hormone mainly produced by the liver as a downstream effector of growth hormone secretion (25). Circulating levels of IGF-1

directly regulate bone growth during juvenile age (26), and IGF-1 secretion is reduced upon chronic undernutrition (27). We thus measured circulating IGF-1 in males at three timepoints: at diet switch, two weeks after (at the end of the linear growth phase), and at P91 when the mice were killed (Figure 3J). We took time-matched samples in control mice. In control mice, IGF-1 levels did not differ significantly from P28 to P91 (about 350 $ng.mL^{-1}$). In contrast, all mice fed LPD had an IGF-1 serum concentration around 100 $ng.mL^{-1}$ at the time of refeeding, capturing an important signature of stunting in our model of PEM. Two weeks after refeeding, IGF-1 levels were comparable to those of control male mice, and did not significantly vary at P91. Thus, while refeeding with an optimal diet stimulated IGF-1 secretion, it was not enough to compensate for the growth retardation, suggesting irreversible effects on male growth induced by long-term exposure to LPD after weaning.

Lp^{WJL} treatment during the refeeding fails to improve systemic growth

Given the sex-related difference we observed previously, for the reminder of the study we focused on male mice. We chose the most severe stunting phenotype that we observed after refeeding, i.e., mice that were refed at P56. We separated mice in three experimental groups: mice fed control diet from P14 to P119, mice fed LPD during the same period of time, and mice fed LPD from P14 to P56, then control diet from P56 to P119. We decided to keep until day 119, to see if the stunting phenotype observed at day 91 was still present four weeks later.

We previously reported that the Lp^{WJL} strain stimulates growth of juvenile mice upon chronic malnutrition (15,16). In these studies, treatment with Lp^{WJL} was performed in a model where, besides protein, fat was also drastically reduced in the diet. Treatment was performed after weaning, i.e., during the linear phase of growth. We wondered about the

effect of Lp^{WJL} upon PEM and hypothesized that treating our mice during the refeeding phase, which corresponds to a linear phase of growth, would stimulate their growth to a point where mice would totally catch up their growth after a period of protein scarcity. As expected, mice fed LPD all the time showed severe stunting and underweight (Figure 4A-B). Despite a significant increase in body weight and length after refeeding, placebo-treated refed mice (RF56, Figure 4A-B) still presented signs of stunting, since their body length was less than 2 standard deviations from the mean of the control group. Contrary to what we had hypothesized, supplementation with Lp^{WJL} during the refeeding phase did not have an additive effect on growth or weight, with only a slight increase in weight in LPD-fed mice at P91. Indeed, besides fat mass (Figure 4E-F), Lp^{WJL} treatment had no impact on size and weight of several organs (Figure 4C-K). This was evident when running Principal Component Analysis (Figure 4L), where mice clustered mainly along PC1 (59.01% of total variance) according to diet, but not to treatment. Thus, treatment with Lp^{WJL} during the refeeding phase failed to further stimulate growth after prolonged exposition to protein scarcity, highlighting the long-lasting deleterious effects of protein malnutrition during early life.

Suboptimal refeeding with a non-obesogenic Western diet compensates early-life associated stunting, but impairs glucose metabolism

One of the main drivers of malnutrition is suboptimal nutrition, defined as inadequate nutrient intake that does not meet the age-specific recommendations (2). Finally, we sought to characterize the impact on weight gain and growth of LPD feeding followed by refeeding with a suboptimal diet. For this experience, we decided to refeed the animals at P35, i.e., the first time when growth catch-up was impaired upon refeeding with an optimal diet (Figure 1E). Indeed, we sought to establish a novel model of

"double burden of malnutrition", defined as the simultaneous manifestation of both undernutrition and overweight (28). During the first burden (P14 to P35), mice were fed control diet or LPD. During the second burden (P35 to P119), mice were fed two types of suboptimal diet: Total New Western Diet (WD) or a modified version of WD (MWD), with protein accounting for 8.4% of total kcal (instead of 17%). In parallel to these groups, we also fed mice control diet, WD, or MWD from P14 to P112 (Figure 1A). Unlike diet-induced obesity diet traditionally used in nutritional studies, WD has a variety of fat sources and is more similar to the human Western diet. It is noteworthy that this version of WD is not obesogenic in C57Bl/6J mice (19), so the idea was to provide a suboptimal diet without strongly affecting body weight.

During the first burden, mice on LPD gained significantly less weight and were stunted (Figure 5B-C; G3 and G4). However, upon refeeding with WD or MWD, mice quickly gained weight and had a significant growth catch-up (Figure 5B-C; days 35 to 119). Indeed, at P119 all groups of mice had a similar body length (Figure 5D). Feeding with WD after control diet induced a significant gain in body weight compared to control-fed animals (Figure 5E), but this was not the case after LPD. In contrast, MWD did not induce a significant increase in body weight, even when mice were fed MWD from P14 onwards.

To assess whether LPD followed by WD or MWD dysregulates glucose homeostasis, we evaluated glucose and insulin tolerance. At P91 or P119, after 6 hours of food deprivation, no diet had a significant impact on glycemia (Figure 5F-G). It is noteworthy that glycemia did not differ significantly between the two time points (two-way ANOVA: effect of time, $P = 0.81$). When we performed an intraperitoneal glucose tolerance test (GTT), we saw that, unlike WD, MWD feeding after from P14 onwards

(group 6), induced a significant glycemic increase in the early phase of the GTT (the so-called glycemic excursion) (Figure 5H, G6). However, this was not the case when MWD was given after P35 (Figure 5H, G2 and G4). Interestingly, while WD per se, or given after chow feeding, did not modify glucose tolerance (Figure 5H, G1 and G5), priming with LPD during early life induced a transient glucose intolerance (Figure 5H, G3). As a whole, LPD-fed mice were susceptible to develop early-phase glucose intolerance when they were refed with MWD, i.e., a suboptimal diet rich in fat and poor in protein.

To infer if this phenotype was due to a change in insulin sensitivity, we performed an insulin tolerance test (ITT). We saw that insulin tolerance was not modified by any of the different diet combinations (Figure 5I), highlighting that the aforementioned glucose intolerance was not due to impaired insulin sensitivity. Overall, we show that, while it does not affect growth catch-up after protein malnutrition-mediated early life stunting, suboptimal nutrition leads to impaired glucose control later in life.

Supplementation with Lp^{WJL} in suboptimal nutritional conditions further impairs glucose tolerance after early-life protein restriction

In the same line as previously, we investigated the effect of a supplementation with Lp^{WJL} during the second burden of the aforementioned protocol (i.e., from P35 to P119). We found no effect of adding Lp^{WJL} on weight gain and systemic growth (Supplementary Figure 2). Likewise, two-way ANOVA analysis did not reveal any significant effect of Lp^{WJL} on organ size or weight (Supplementary Figure 3). We further assessed glucose metabolism by performing a GTT and an ITT. We previously showed that Lp^{WJL} treatment resulted in improved glucose tolerance during the post-weaning period in chronic malnutrition mice (16). It is noteworthy that, in our experimental

setting, treatment with Lp^{WJL} did not affect glycemia after a 6-hour-long food deprivation (Supplementary Figure 4; effect of Lp^{WJL}, P = 0.0869, two-way ANOVA). Overall, glucose tolerance was not affected in mice fed control diet after weaning (G1 and G2 mice, Figure 6), nor mice that were fed WD or MWD all long (G5 and G6 mice). Surprisingly, mice facing a double burden of malnutrition, particularly those fed WD after LPD (G3 mice), showed significant glucose intolerance when given Lp^{WJL} during the second burden. Furthermore, Lp^{WJL} did not significantly affect insulin tolerance in any of the groups (Figure 6). Overall, these findings indicate that Lp^{WJL} treatment exacerbates glucose intolerance in a mouse model of a double burden of malnutrition manifested by early protein malnutrition followed by western diet nutrition.

DISCUSSION

PEM early in life has deadly consequences, being one of the most important causes of infant death in the developing world (2). Maintenance of adequate nutrition and rehabilitation are the most effective interventions to reduce the deleterious effects of PEM (29). In this study, we aimed at establishing a mouse model of PEM, followed by a nutritional rehabilitation, in order to explore the long-lasting consequences of early-life PEM.

Early-life PEM results in stunting (30). In our protein malnutrition model, slow growth and weight gain rate occurred after weaning, with over 50% reduction after 5 weeks on LPD. In humans, the two main clinical syndromes of the extreme forms of children with PEM are marasmus and kwashiorkor (31). While marasmus results from adaptation to severe depletion of calories and nutrients, kwashiorkor (“sickness of the weaning” in Ga

language) is the consequence of a diet with inadequate protein but reasonably normal caloric intake. Thus, our mouse model seems closer to kwashiorkor than marasmus. However, it does not exhibit classic traits of kwashiorkor such as edema and hepatomegaly (31), probably due to the fact that our mice are not in contact with any enteric pathogens and thus are not susceptible to infection and inflammation. Indeed, several studies have shown that malnourished mice are more susceptible to infection, thus mimicking one of the hallmarks of human malnutrition (20,21). Furthermore, nutritional rehabilitation after kwashiorkor results in increased IGF-1 levels, but the values are lower than in healthy children (32). In our mouse model, IGF-1 levels are fully restored by nutritional rehabilitation, suggesting that mice are more resilient than humans to early-life nutritional stress. This is confirmed by the fact that, in our model, mice are kept under severe protein malnutrition for several weeks after weaning. While this results in a severe phenotype regarding growth and weight, our mice did not show any particular sign of torpor, reduced activity, or change in appetite during the nutritional challenge. Moreover, unlike Ferreira-Paes et al. (21), who describe a model of marasmus using weanling BALB/c mice, nutritional rehabilitation in male mice did not fully resolve the severity of protein malnutrition. Indeed, in our experimental setting, male mice were stunted after nutritional rehabilitation, although early-life wasting was corrected. It is noteworthy that the earlier the rehabilitation took place, the better the response was. Nevertheless, even after post-natal day 56 (i.e., young adult mice), growth occurred at a rate of 0.08 $cm.day^{-1}$ after refeeding on an optimal diet, showing the remarkable capacity of mice to adapt to deleterious nutritional environments.

Recent advances in the knowledge of malnutrition have shown that the gut microbiota is a contributor to the deleterious stunting and wasting phenotype (33). Indeed, colonization of germ-free mice with kwashiorkor-associated microbiota in combination with a Malawian diet leads to severe weight loss (34). Our previous work has shown that modulating the microbiota of chronically malnourished weanling mice with Lp^{WJL} stimulates growth via activation of the somatotropic axis (15,16). We hypothesized that Lp^{WJL} would be able to stimulate growth during the refeeding phase in our PEM model, thus compensating the stunting phenotype we had previously observed. In our experimental setting, we saw no improvement of growth associated to Lp^{WJL} during the refeeding phase. Thus, it is likely that the effects of the bacterium are associated to an overall dietary challenge, with reduced protein and fat, rather than low-protein feeding per se. In line with that, the model we previously described exhibited a marked decrease of weight after weaning,

Overall, our results confirm the Developmental Origins of Health and Disease (DOHaD) paradigm, which stipulates that developmental and early postnatal insults, such as malnutrition, can have deleterious impacts in adult life (35). In line with that, the DOHaD paradigm is particularly appropriate in understanding the nutritional transitions in countries that face the double burden of malnutrition. Therefore, we sought to establish a model with suboptimal refeeding after a moderately long period of low-protein diet. It is well known that severe acute malnutrition affects glucose homeostasis in infants (4,36), a feature that persists later in life (37,38). Furthermore, studies in rodents have shown that post-weaning PEM induces hepatic alterations, such as steatosis and impaired hepatic mitochondrial turnover (39–41). Interestingly, two independent studies using a high-fat diet (58% kcal from fat) after LPD have shown that

PEM after weaning does not potentiate fat accumulation, hepatic steatosis and insulin resistance in adult young mice (39,42). In their mouse model of double burden of malnutrition, Dalvi et al. described that LPD feeding for 4 weeks post-weaning induces transient glucose intolerance after a recovery on chow feeding (39), showing that LPD priming induces deleterious effects later in life associated to poor metabolic health. Indeed, in our model, while we saw no differences in growth with mice on MWD after LPD, LPD priming also induces deficiencies in glucose control later in life. Although results on mouse models are still inconsistent and fail to fully recapitulate the metabolic alterations observed in malnourished humans, they provide key mechanistic insights into the different pathologies associated to stunting in early life.

Indeed, cohorts of malnourished mice and humans have shown that growth and metabolic functions are impaired, associated to alterations in the gut microbiota, and that this impaired phenotype could be transferred (33). Similarly, extensive research has shown that the overnourished microbiota is associated to loss of richness of taxa and enrichment of bacteria associated to inflammation (43,44). While most studies have been correlative on this issue, they have given an insightful look into the alterations of microbial ecology in cases of malnutrition. Little is known on humans facing the double burden of malnutrition. A 2018 study on a Mexican cohort of 36 patients (undernourished, obese or controls) showed similarities in the composition of the microbiota between the obese and undernourished group. In line with these findings, we hypothesized that treatment with Lp^{WJL} could have a beneficial impact in mice facing a double burden of malnutrition. Not only did we not see any beneficial effect of Lp^{WJL} in this context, but the treatment induced further impairment of glucose tolerance in mice

fed a suboptimal Western diet. Thus, caution should be used when developing probiotic strategies for PEM when infants also face a double burden of malnutrition.

Overall, we provide a new model of refeeding after early-life PEM, where we show sex-associated differences in the long-term response to refeeding. We provide data showing that suboptimal refeeding with a moderately low-protein diet, which is also rich in fat, is sufficient to induce growth in malnourished mice, although this is associated to metabolic perturbations. Further investigations into the specific mechanisms driving these features will help uncover mechanisms associated to PEM and the double burden of malnutrition.

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JLT, FL, and FDV designed research; JLT, AC and DS conducted research; JLT and FDV analyzed data; and JLT, FL and FDV wrote the paper. FDV had primary responsibility for final content. All authors read and approved the final manuscript.

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FIGURE LEGENDS

Figure 1: Chronically malnourished juvenile male and female mice differ in their dynamics of growth.

Body length (**A, B**) and weight (**C, D**) of male and female control diet- or low-protein diet-fed mice refed with control diet respectively at post-natal day 28 (RF28), 35 (RF35), 42 (RF42), 49 (RF49), or 56 (RF56). Arrows correspond to the day of refeeding. **(E, F)** Length-versus-weight of control- and low-protein diet- (LPD) fed animals. (**G, H**) Length-versus-weight of control and refed mice, after the time of refeeding.

SD: standard deviation of control group.

Data are mean ± SEM.

Figure 2: Optimal refeeding after long-term juvenile malnutrition is insufficient to fully recover systemic growth in males, but not in females.

(A, B) Weight-for-age relationship of of male and female control diet- or low-protein diet-fed mice refed with control diet respectively at post-natal day 28 (RF28), 35 (RF35), 42 (RF42), 49 (RF49), or 56 (RF56). Arrows correspond to the day of refeeding. **(C, D)** Body length evolution during the two weeks following refeeding or from day 21 to day 35 in control mice.

Data are mean ± SEM.

Figure 3: Optimal refeeding after early-life malnutrition in males is insufficient to fully recover organ growth.

Weight of liver **(A)**, subcutaneous fat **(B)**, perigonadal fat **(C)**, and soleus muscle **(D)**; length of femur **(E)**, tibia **(F)**, small intestine **(G),** and colon **(H)** of male control diet- or low-protein diet-fed mice refed with control diet respectively at post-natal day 28 (RF28), 35 (RF35), 42 (RF42), 49 (RF49), or 56 (RF56). **(I)** Principal Component Analysis of parameters measured in **A** to **H**. **(J)** Serum concentration of IGF-1 measured at the switch of diet, 2 weeks after switch and at the end of the experiment in aforementioned mice.

Data are mean ± SEM.

* $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$; *** $P < 10^{-4}$, one-way ANOVA followed by Tukey's post-hoc test.

Figure 4: Lp^{WJL} treatment during the refeeding phase fails to improve systemic growth.

Body length **(A)** and weight **(B)**; weight of liver **(C)**; fat content **(D)** ; weight of subcutaneous fat **(E)**, perigonadal fat **(F)**, and soleus muscle **(G)**; length of femur **(H)**, tibia **(I)**, small intestine **(J)**, and colon **(K)** of male mice refed at post-natal day 56 (RF56) or maintained on low-protein diet (LPD) and treated with *Lactiplantibacillus plantarum* WJL *(*Lp^{WJL}) or a placebo. **(L)** Principal Component Analysis of parameters measured in **C** and **E** to **K**.

Data are mean ± SEM.

* $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$; *** $P < 10^{-4}$, two-way ANOVA followed by Šídák's post-hoc test.

Figure 5: Suboptimal refeeding with a non-obesogenic Western diet compensates early life-associated stunting, but impairs glucose metabolism.

(A) Experimental procedure . Body length **(B)** and weight **(C)** of mice described in panel **A**. Length **(D)** and weight **(E)** at post-natal day 119. Plasma glucose after 6 hours of food deprivation at post-natal day 91 **(F)** and 112 **(G)** of of mice described in panel **A**. Intraperitoneal glucose tolerance test **(H)** and insulin tolerance test **(I)** performed at post-natal day 91 and 112 respectively on mice described in panel **A**. Inset: area under the curve (AUC) of the respective tests.

Data are mean ± SEM.

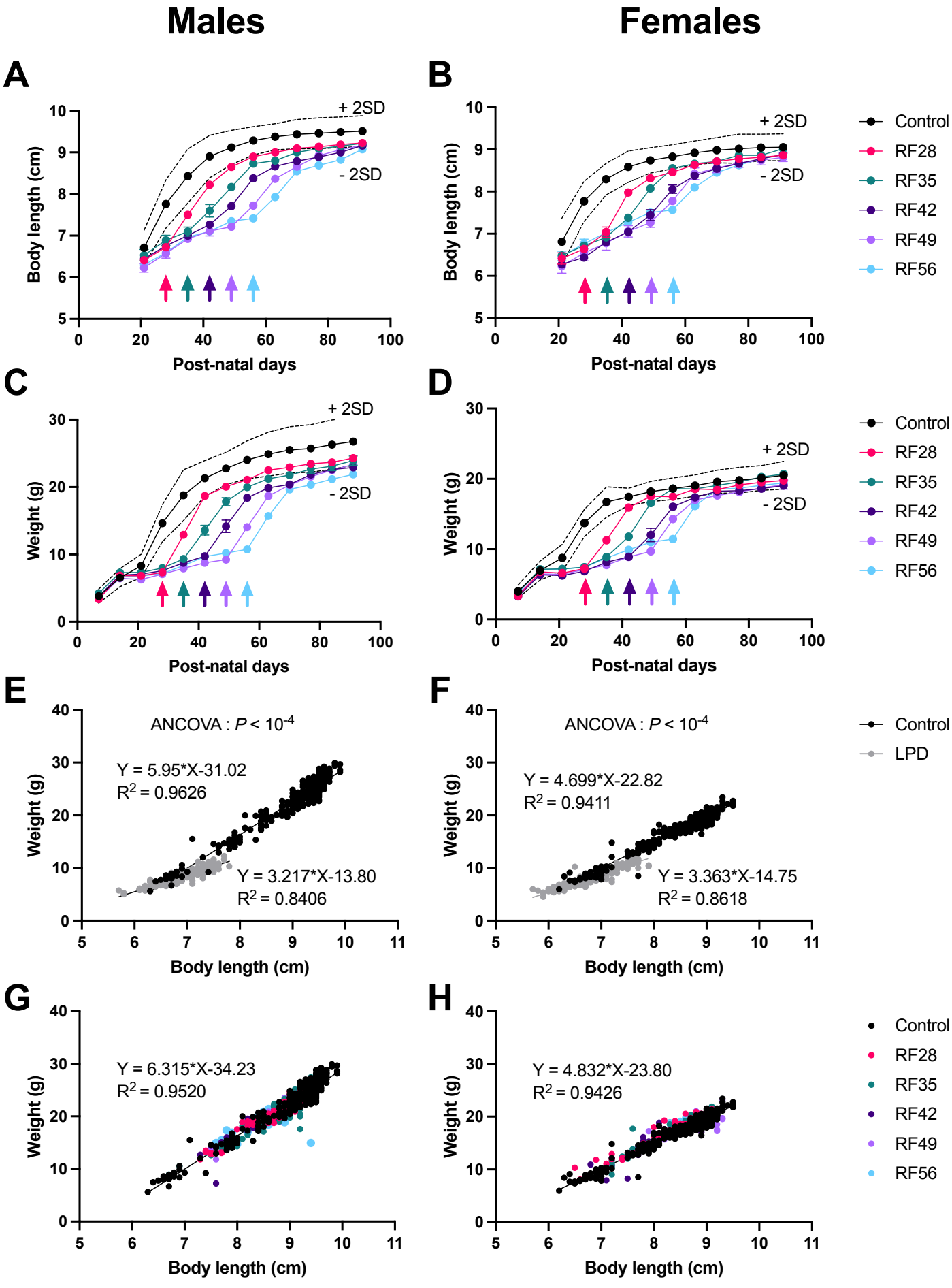
* $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$; *** $P < 10^{-4}$, one-way ANOVA followed by Tukey's post-hoc test (when not indicated, value vs. control).

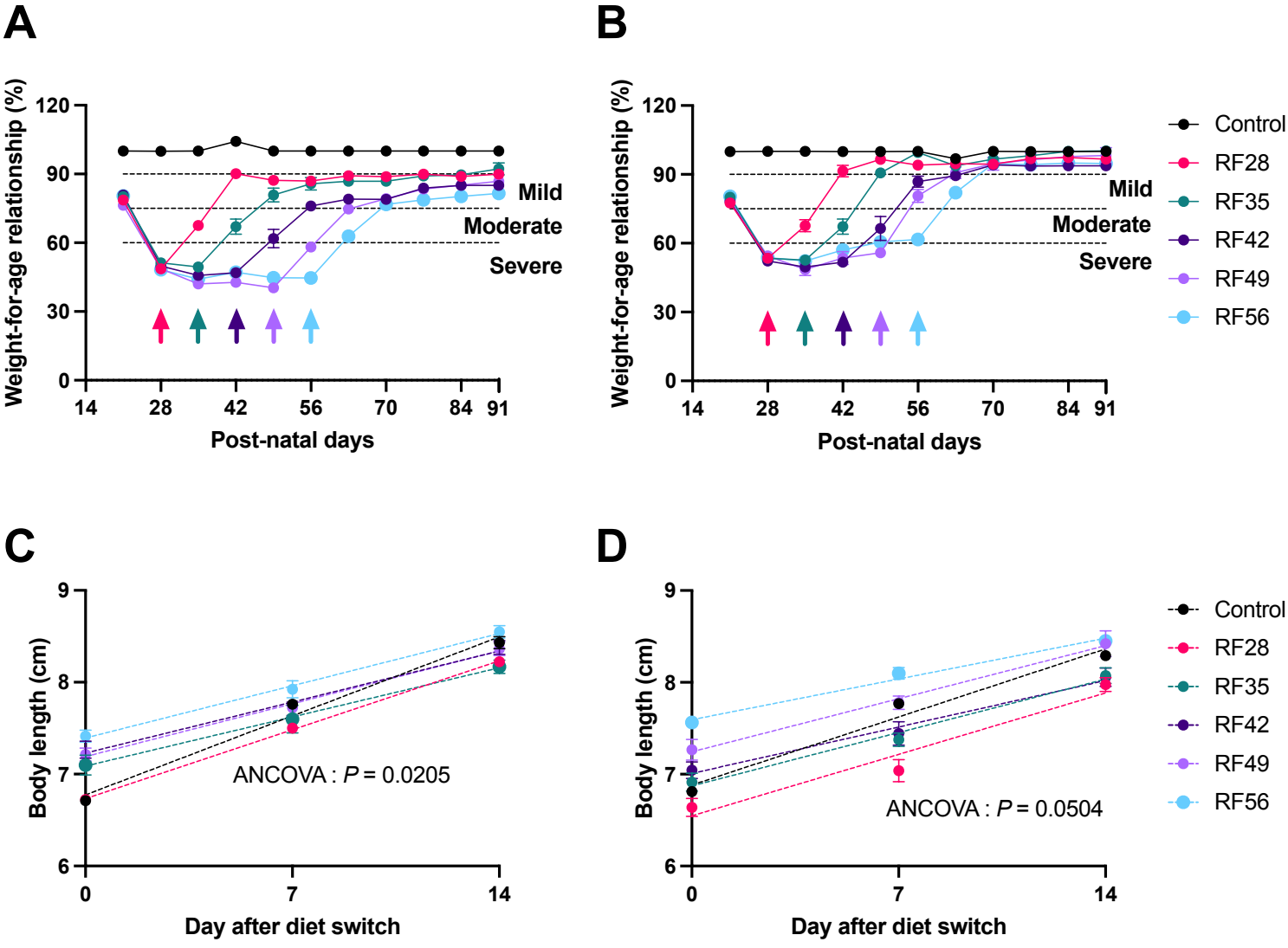
Figure 6: Supplementation with Lp^{WJL} in suboptimal conditions further impairs glucose tolerance after early-life protein restriction.

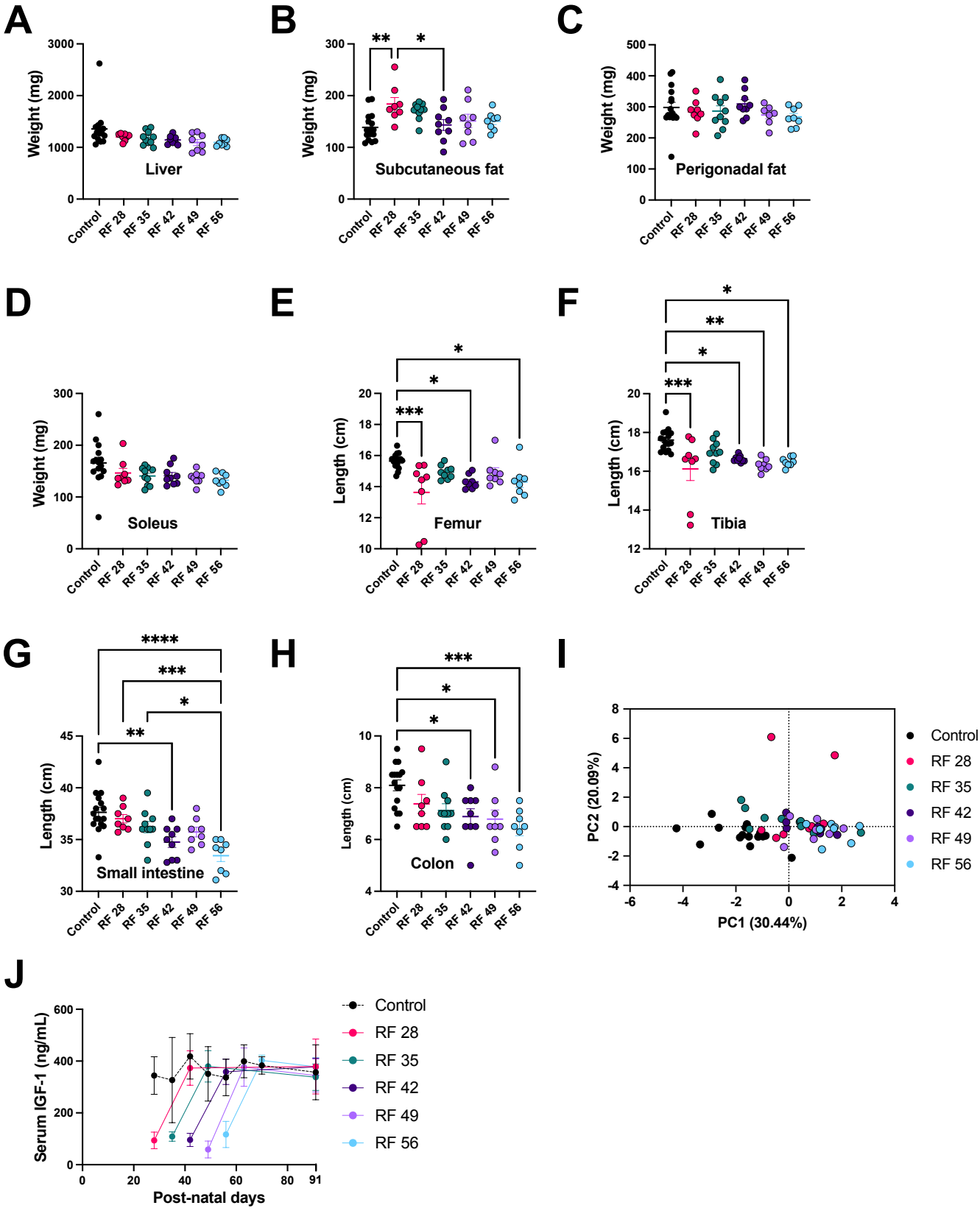
Intraperitoneal glucose tolerance test (GTT) and insulin tolerance test (ITT) performed respectively at P91 and P112 on male mice treated with *Lactiplantibacillus plantarum* WJL *(*Lp^{WJL}) or a placebo from P35 to P119. Experimental groups are described in Figure 5**A**.

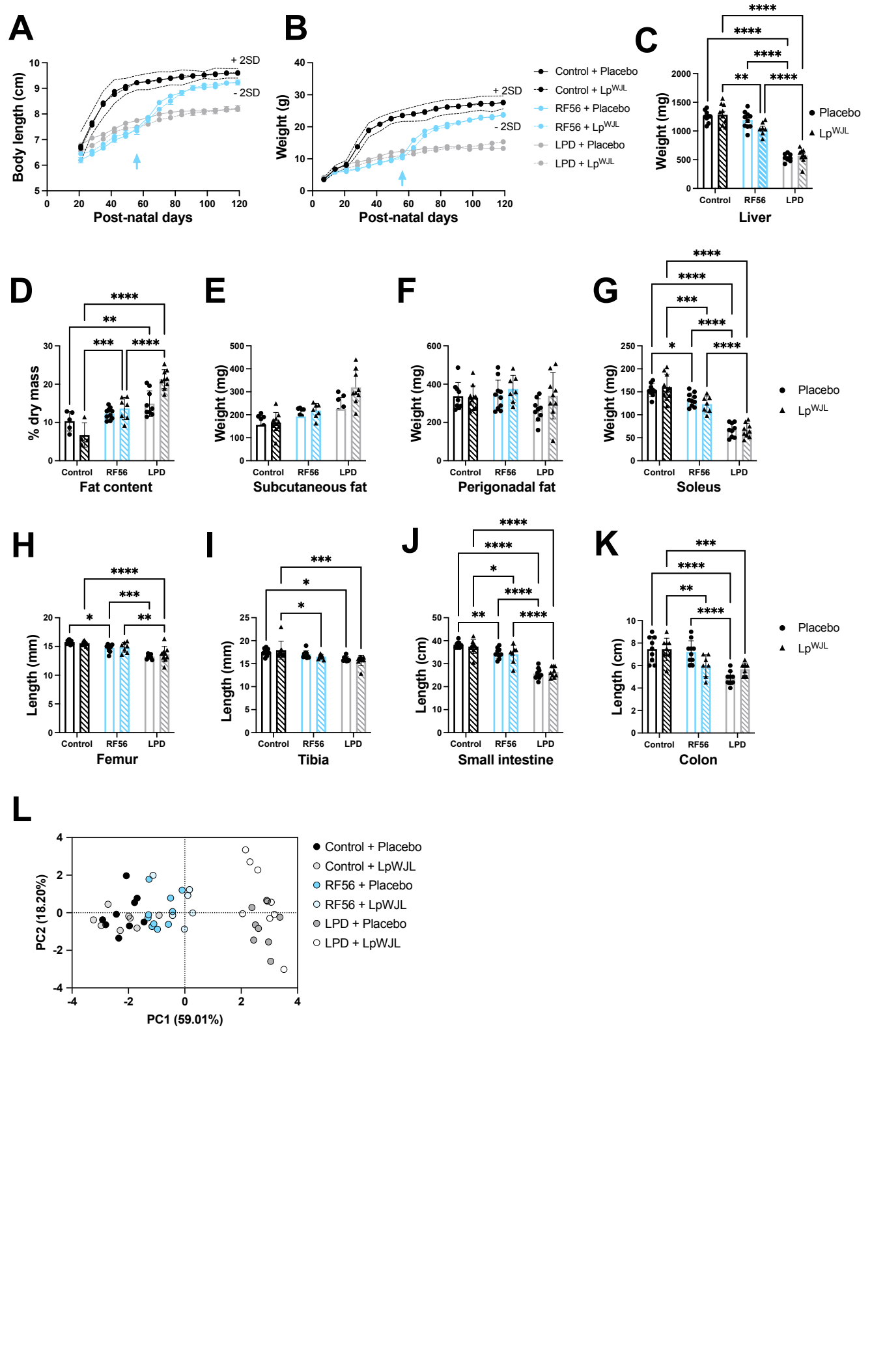
Data are mean ± SEM.

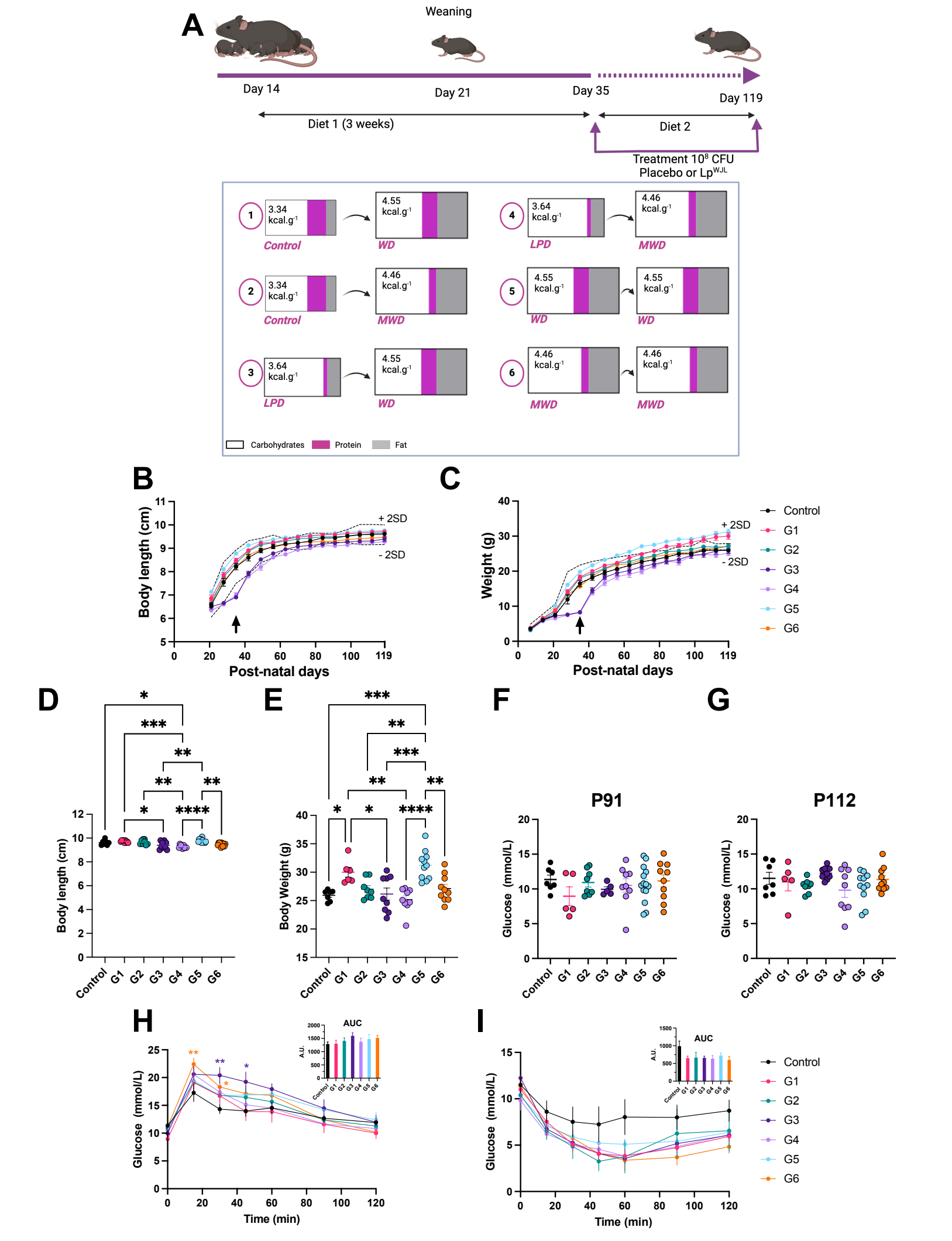
* $P < 0.05$; ** $P < 0.01$, Student's *t*-test with Welch's correction.

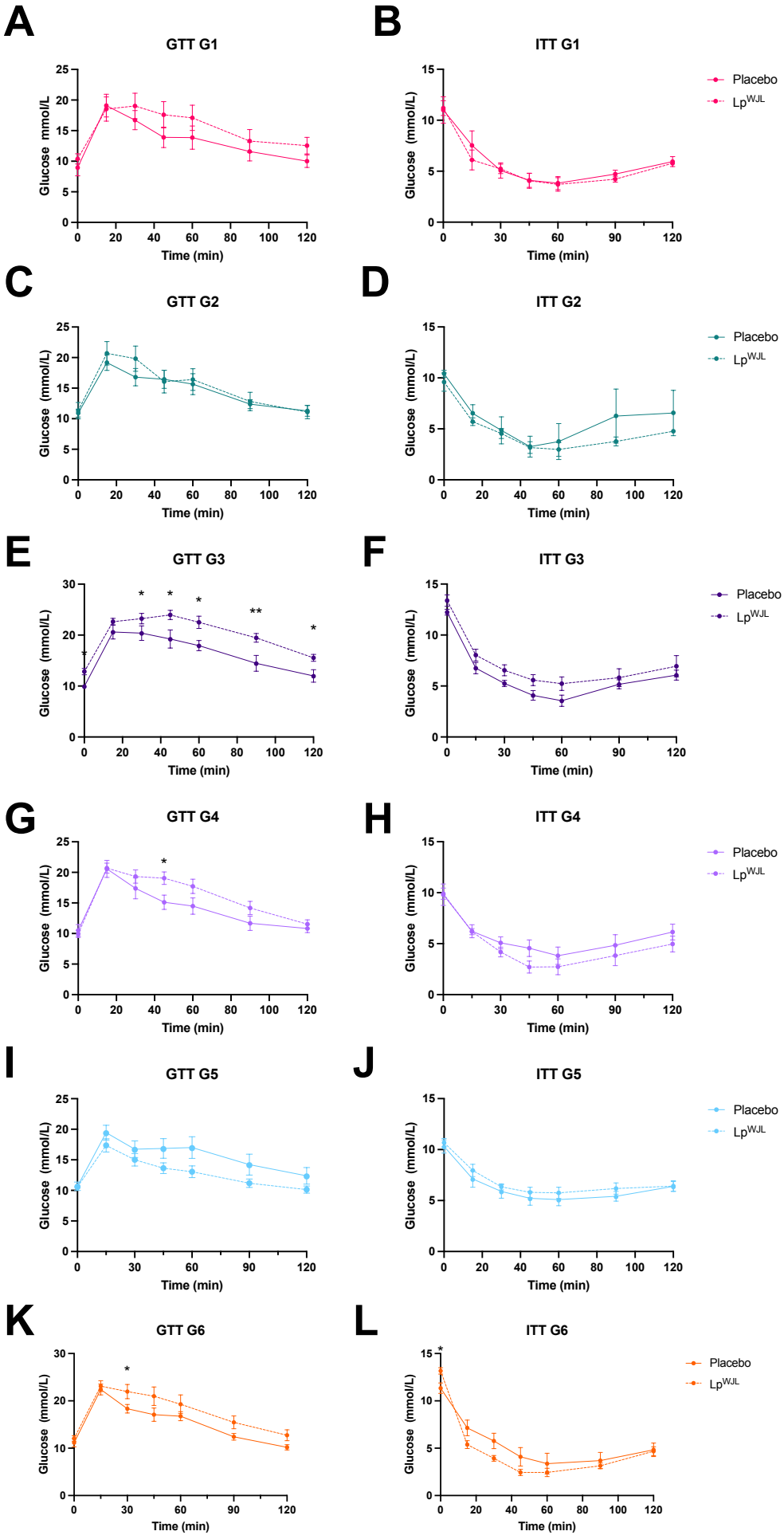

Males
Females
A
B
C
D
E
F
G
H
+ 2SD
- 2SD
Body length (cm)
Weight (g)
Post-natal days
Control
RF28
RF35
RF42
RF49
RF56
LPD
ANCOVA : P < 10-4
Y = 5.95*X-31.02
R2 = 0.9626
Y = 3.217*X-13.80
R2 = 0.8406
Y = 4.699*X-22.82
R2 = 0.9411
Y = 3.363*X-14.75
R2 = 0.8618
Y = 6.315*X-34.23
R2 = 0.9520
Y = 4.832*X-23.80
R2 = 0.9426
Body length (cm)


A
B
C
D
Weight-for-age relationship (%)
Post-natal days
Mild
Moderate
Severe
Control
RF28
RF35
RF42
RF49
RF56
Body length (cm)
Day after diet switch
ANCOVA : P = 0.0205
ANCOVA : P = 0.0504


A
Weight (mg)
Liver
B
Weight (mg)
Subcutaneous fat
C
Weight (mg)
Perigonadal fat
D
Weight (mg)
Soleus
E
Length (cm)
Femur
F
Length (cm)
Tibia
G
Length (cm)
Small intestine
H
Length (cm)
Colon
I
PC2 (20.09%)
PC1 (30.44%)
J
Serum IGF-1 (ng/mL)
Post-natal days
Control
RF 28
RF 35
RF 42
RF 49
RF 56


A
Body length (cm)
Post-natal days
+ 2SD
- 2SD
B
Weight (g)
Post-natal days
+ 2SD
- 2SD
Control + Placebo
Control + Lp^WJL
RF56 + Placebo
RF56 + Lp^WJL
LPD + Placebo
LPD + Lp^WJL
C
Weight (mg)
Liver
Control
RF56
LPD
Placebo
Lp^WJL
D
% dry mass
Fat content
E
Weight (mg)
Subcutaneous fat
F
Weight (mg)
Perigonadal fat
G
Weight (mg)
Soleus
H
Length (mm)
Femur
I
Length (mm)
Tibia
J
Length (cm)
Small intestine
K
Length (cm)
Colon
L
PC2 (18.20%)
PC1 (59.01%)
Control + Placebo
Control + LpWJL
RF56 + Placebo
RF56 + LpWJL
LPD + Placebo
LPD + LpWJL


A
Weaning
Day 14
Day 21
Day 35
Day 119
Diet 1 (3 weeks)
Diet 2
Treatment 10⁸ CFU
Placebo or Lp^WJL
1
3.34 kcal.g⁻¹
Control
4.55 kcal.g⁻¹
WD
2
3.34 kcal.g⁻¹
Control
4.46 kcal.g⁻¹
MWD
3
3.64 kcal.g⁻¹
LPD
4.55 kcal.g⁻¹
WD
4
3.64 kcal.g⁻¹
LPD
4.46 kcal.g⁻¹
MWD
5
4.55 kcal.g⁻¹
WD
4.55 kcal.g⁻¹
WD
6
4.46 kcal.g⁻¹
MWD
4.46 kcal.g⁻¹
MWD
Carbohydrates
Protein
Fat
B
Body length (cm)
+ 2SD
- 2SD
Post-natal days
C
Weight (g)
+ 2SD
- 2SD
Post-natal days
Control
G1
G2
G3
G4
G5
G6
D
Body length (cm)
E
Body Weight (g)
F
P91
Glucose (mmol/L)
G
P112
Glucose (mmol/L)
H
AUC
A.U.
Glucose (mmol/L)
Time (min)
I
AUC
A.U.
Glucose (mmol/L)
Time (min)


A
GTT G1
Glucose mmol/L
Time (min)
B
ITT G1
Glucose (mmol/L)
Time (min)
Placebo
Lp^WJL
C
GTT G2
Glucose (mmol/L)
Time (min)
D
ITT G2
Glucose (mmol/L)
Time (min)
Placebo
Lp^WJL
E
GTT G3
Glucose (mmol/L)
Time (min)
F
ITT G3
Glucose (mmol/L)
Time (min)
Placebo
Lp^WJL
G
GTT G4
Glucose (mmol/L)
Time (min)
H
ITT G4
Glucose (mmol/L)
Time (min)
Placebo
Lp^WJL
I
GTT G5
Glucose (mmol/L)
Time (min)
J
ITT G5
Glucose (mmol/L)
Time (min)
Placebo
Lp^WJL
K
GTT G6
Glucose (mmol/L)
Time (min)
L
ITT G6
Glucose (mmol/L)
Time (min)
Placebo
Lp^WJL