The selective cargo loading and release functions of the cellular sterol transporter StarD4 are allosterically controlled by the recognition of specific PIP2 subtypes in the targeted membranes.

Hengyi Xie[1], Harel Weinstein[1,2,*]

[1] Department of Physiology and Biophysics, Weill Cornell Medicine, New York, NY, 10065, USA

[2] Institute for Computational Biomedicine, Weill Cornell Medicine, New York, NY 10065, USA

*Corresponding Author: Harel Weinstein, D.Sc.

Address: Department of Physiology and Biophysics, Weill Cornell Medicine,

1300 York Avenue, New York, NY, 10065

E-mail: haw2002@med.cornell.edu, Phone: 1-212-746-6358

Keywords: Molecular Dynamics (MD) simulations; cholesterol traffic and distribution in cells; conformational changes coupled to modes of cholesterol (CHL) binding; membrane embedding of StarD4; recognition of membranes containing specific PIP2 subtypes; allosteric network coordination between the CHL binding site, the cargo release gates and the binding of specific PIP2 subtypes ; machine learning analysis of MD trajectories; detection of rare events in MD trajectories; non-negative matrix factorization algorithm; N-body Information Theory analysis of MD trajectories; allosteric channels supporting CHL uptake and release on membrane with different composition.

ABSTRACT

Intracellular cholesterol trafficking, vital for cell function, is orchestrated in part by proteins with steroidogenic acute regulator-related lipid transfer (START) domains, such as StarD4 which is a sterol-specific transfer protein that contributes ~ 33% of non-vesicular sterol transport between plasma membrane (PM), the primary cholesterol reservoir, and the endoplasmic reticulum (ER). Comprising a single soluble START domain, StarD4 is known to target both sterol donor-, and acceptor-membranes, through interactions with anionic lipids. Experiments have illuminated the kinetics of this sterol transfer and shown it to be modulated by specific phosphatidylinositol phosphates (PIPs) on the target membrane. The distinct subtype distribution of PIPs in the membranes of cellular organelles serves as a guide directing StarD4 to particular cell components. To discover the molecular mechanism of membrane targeting by the recognition of the PIP2 subtype in the membrane, and how this affects the direction and kinetics of CHL transport (uptake vs release), we used molecular dynamics (MD) to simulate the processes. Analysis of the MD trajectories with machine learning and information theory methods revealed how StarD4 embeds in membrane containing different anionic lipids (PI(4,5)P_2, PI(3,5)P_2, and PS), how it recognizes the anionic lipid content by the different modes in which they bind to the protein, and how the protein responds to this recognition by adopting different orientations on the membrane and undergoing specific conformational changes. We identified the allosteric channels underlying these complex dynamics rearrangements by connecting the various PIP2-subtype-specific conformational states to different CHL binding modes in the pocket, which then relate to the dynamics of the gates that allow either CHL uptake into *apo*-StarD4, or its release from *holo*-StarD4. This reveals the crucial role of PIP2 subtypes in shaping functional StarD4 motifs responsible for organelle selectivity of the cholesterol trafficking, providing fundamental insights into cellular cholesterol regulation.

Introduction

Interest in the process of dynamic trafficking of CHL among diverse locations and organelles in the cell has brought to light the role of cellular proteins containing **st**eroidogenic **a**cute **r**egulator-related lipid transfer (START) domains (StarD), such as StarD4 (1-7). The StarD4 protein specializes in binding and transport sterols (8,9) and contributes 33% of the non-vesicular sterol transport between the primary cholesterol reservoirs (i.e. the plasma membrane (PM) and the endocytic recycling compartment (ERC)) and the endoplasmic reticulum (ER) that governs sterol homeostasis maintenance, and cholesterol concentration sensing and regulation (3,10,11). StarD4 incorporates a single soluble START domain, and targets sterol donor and acceptor membranes by interacting with anionic lipids (9,12). Recent investigations have revealed the modulation of sterol transfer kinetics by StarD4 via specific phosphatidylinositol phosphates (PIPs) subtypes (12), with a preference for StarD4 extracting sterol from donor membranes vesicles resembling plasma membranes that contain phosphatidylinositol 4,5-bisphosphate ($PI(4,5)P_2$), as opposed to membranes vesicles containing phosphatidylinositol 3,5-bisphosphate ($PI(3,5)P_2$) or phosphatidylserine (PS). Given the selective enrichment of distinct PIPs on cellular organelles (13), these negatively charged lipid molecules can serve as organelle-specific fingerprints that guide soluble StarD4 to specific membrane compartments and modulate its sterol transport kinetics. Here we present a mechanistic understanding, at the molecular level, for the recognition of PIP2 subtypes by StarD4 and the resulting modulation of cholesterol transport kinetics by this cholesterol (CHL)-trafficking protein.

Structure determinations have shown the ~210-amino acid START domain to have an α/β helix-grip fold that forms an internal hydrophobic cavity essential for lipid recognition and binding (6,7). However, the structure of a START domain in complex with a sterol remains unresolved. Investigations into sterol binding by the START domains of StarD3 and StarD5, relying primarily on docking and short molecular dynamics (MD) simulations, have proposed a cholesterol binding to residues deep in the hydrophobic pocket (7,14-17). On the other hand, recent structural analyses of LAM proteins in the StARkin superfamily which shares the helix-grip scaffold, have shown that the sterol hydroxyl group forms hydrogen bonds with residues situated in the upper two-thirds of the hydrophobic tunnel (18-20). In our recent study involving extensive MD simulations of StarD4 in aqueous solution (21) we described a dynamic repositioning of CHL in the binding pocket that reconcile the observations from prior studies. Specifically, when cholesterol embeds deep in the hydrophobic pocket, the sterol hydroxyl group interacts with S136 and S147 (termed "the Ser-binding mode"), but in a state of StarD4 that relates to CHL release CHL moves to a new position (termed "Trp-binding mode"), the cholesterol inserts roughly two-thirds into the pocket closer to the gate where its hydroxyl group interacts with W171 in proximity to Y117 and R92 (**Fig. 1A,B**) (21,22). Our simulation trajectories totaling 403 microseconds revealed that spontaneous transitions between the two modes that are stable for long periods, are accompanied by StarD4 conformational changes that involve specific structural motifs identified as membrane-interacting (**Fig. 1C-F**), including the destabilization of the Ω1-H4 front gate (**Fig. 1D**) and the opening of the β8-Ω4 back corridor (**Fig. 1F**) (12). As demonstrated from the analysis of the long trajectories in aqueous solution (21), these changes are dynamically connected to the modes of CHL binding in the pocket by an identified allosteric pathway in the StarD4 structure (21).

Notably, in its transition down the binding pocket from the Trp-binding mode, the CHL hydrocarbon tail becomes exposed to the solvent to a state that is not as stable as the other two (21). We therefore proposed (21) that the Trp-binding mode of the CHL-loaded StarD4 represents a

preparation step for CHL delivery. While the Trp-binding mode of StarD4 was recently reproduced on membrane exposing the ligand cholesterol to the membrane surface (22), the downstream release/uptake processes remained unobserved, prompting further investigation. Here, we are investigating the role of the membrane in this process in the full uptake/release pathway and explore the mechanism by which the recognition and sterol transportation functions are modulated by the PIP2 content as observed experimentally (12).

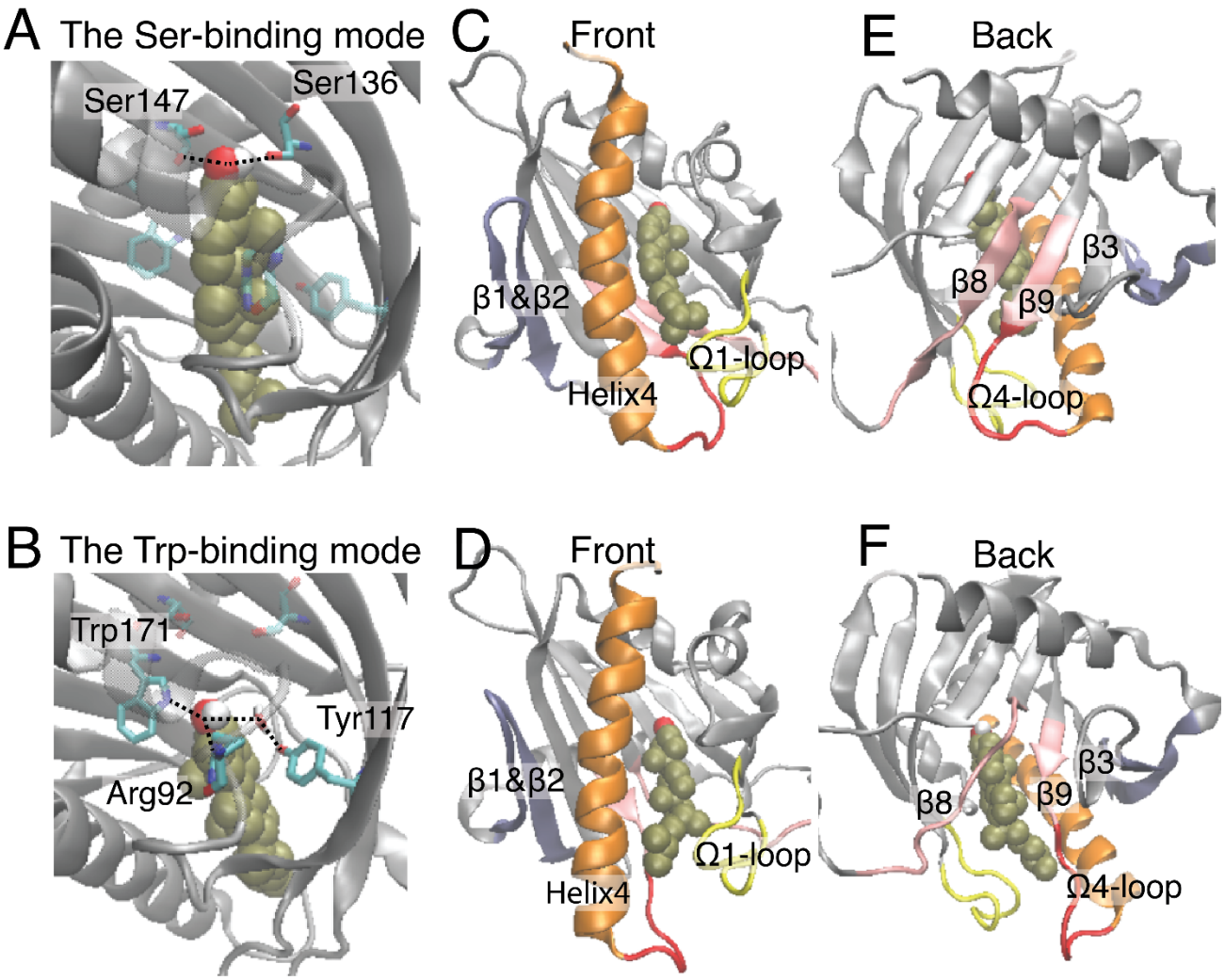

Figure 1: Two modes of cholesterol binding to StarD4 in aqueous solution identified in the long simulation. The two binding modes of CHL in the *holo*-StarD4 complex: **(A)** the "Ser-binding mode"; **(B)** the "Trp-binding mode". StarD4 is rendered in gray. The residues involved in the binding modes (R92, S136, S147,Y117,W171) are rendered in cylinders, with carbons colored in cyan, nitrogen in blue and oxygen in red (hydrogens are not shown). The CHL (in tan) is shown in VDW rendering, with the hydroxyl group in red and white. **(C-F)** The panels depict conformational changes of StarD4 that occur concurrently (see REF (21)) with the translocation of CHL from one binding mode to the other. The structural segments identified in color are involved in the major changes that include the destabilization of the Ω1-H4 front gate and the opening of the back corridor between β8 and β9 Ω4-loop. Helix4 is colored in orange, the Ω1 in yellow, β1&2 in blue, Ω4 in red, and β8&9 in pink. Note that in **(C),** the front gate between Ω1 and H4 is closed, but as the CHL moves towards the gate and H4 moves towards β1, the gate is destabilized and starts towards opening **(D)**. Similarly, the β8-Ω4 back corridor is closed in **(E)** but opened in **(F)** as β9 and Ω4-loop moved towards β3.

To this end, we employed in the present study a battery of extensive molecular dynamics (MD) simulations to collect the data needed to discover and quantify the key stages of the StarD4 cholesterol trafficking between the membranes it targets. As shown in Figs. 2 and 14, the total simulation time in the progression of sampling stages exceeds 2.5 milliseconds. These include the exploration of molecular mechanisms that determine the selectivity and processes of CHL loading from donor membranes and its release to the acceptor membranes. From the analysis of the MD trajectories we identified (i) the dynamic rearrangements underlying the cholesterol uptake and release pathways in StarD4; (ii) the modes of interaction of StarD4 with PI(4,5)P_2 or PI(3,5)P_2 in the corresponding membranes; (iii) how the modes of interaction and the dynamics of StarD4 relate to uptake and release. As discussed here, the detailed definition of the interactions and allosteric channels in the molecular mechanism of PIP2 subtype recognition reveals the organizing elements of cholesterol transfer between the targeted membranes by cellular StarD4.

Specifically, our results show that the *release of CHL* occurs in different conformational states of StarD4 in a PIP2-subtype specific manner. We characterized the preparatory conformational changes leading to cholesterol release from the application of a recently described machine learning method, Rare Event Detection (RED) (23). These preparatory pre-release conformational changes were found to involve dynamic rearrangements of motifs along an allosteric pathway that channels the information from the PIP2-binding motif to the cholesterol-binding site of StarD4. This allosteric route was quantified with the application of N-body Information Theory (NbIT) analysis (24,25). In the separate analysis of the trajectories using a Deep Neural Network (DNN), we found that this allosteric route is a pivotal determinant in distinguishing between StarD4 conformations that interact with PI(3,5)P_2 vs PI(4,5)P_2. Remarkably, the PIP2-subtype specific conformational changes of StarD4 are strongly associated with the functional motifs which are responsible for cholesterol release, revealing the basis for the mechanistic association between the content of the specific PIP2 in the targeted membrane and the selective release of CHL.

The *CHL extraction process* from PI(4,5)P_2-containing membranes into apo-StarD4 was explored with MD simulations using an adaptive sampling protocol involving a sequence of spontaneous interactions of the hydroxyl group of cholesterol with hydrophilic residues lining the hydrophobic pocket. In this process, apo-StarD4 visits a conformational space that differs from the one sampled by the pre-release and CHL release states. In contrast, we found that the same exploration does not produce any cholesterol uptake events in the course of the extensive simulations (see Fig. 14) on PI(3,5)P_2-containing membranes. Indeed, the only events of cholesterol positioning at the pocket entrance occur only when StarD4 adopts a pre-release conformational state. This finding reinforces the observation that when StarD4 interacts with PI(3,5)P_2-containning membrane, it prefers the conformational states that facilitate cholesterol release rather than its extraction from the targeted membrane.

Collectively, these results provide mechanistic insights regarding the molecular basis for organelle selectivity by StarD4 based on PIP2 subtype recognition. The differences in the mode of interaction with the different subtypes lead to conformational preferences that are propagated through an identified allosteric network from the membrane interaction sites to the sterol ligand binding site, to reach the gate of the hydrophobic pocket and the corridor (see Fig. 1). Consequently, the initial PIP2 subtype recognition regulates the kinetics of the cholesterol trafficking function of StarD4.

Results

The process of cholesterol release from *holo*-StarD4 protein.

The dynamics of this process were recorded from MD simulations of StarD4 constructs with CHL in the binding site (see Methods and ref (12)) embedded in 3 different membrane models, each containing only one type of negatively charged lipid: PI(3,5)P_2, or PI(4,5)P_2, or PS. Notably, the simulation design did not aim to replicate the complexities of different organelles; rather, it aimed to faithfully represent the experimental conditions in the research on membrane vesicles where PIP2-subtype-dependent functional dynamics of were observed (12).

Aiming to understand the sensitivity to PIP2 subtype, the simulation process of each of these *holo*-StarD4-membrane systems consisted of three phases, as depicted in **Figure 2**. After equilibration of membrane lipid rearrangement and StarD4 orientation under Mean-field-model simulations (see Methods), spontaneous StarD4 membrane-embedding processes were sampled by MD simulations in Phase 1 (**Fig 2**). In Phase 2 the membrane-embedded CHL-bound StarD4 system was simulated in 24 independent replica runs of StarD4 embedded in each of the three membrane systems (for a total of 344 microseconds, **Fig 2** Phase 2). This was followed in Phase 3 by the simulation of the CHL release process from an identified pre-release state employing an expanded adaptive sampling protocol (see Methods) carried out in 312 replicas (for a total of 725 microseconds, **Fig 2** Phase 3).

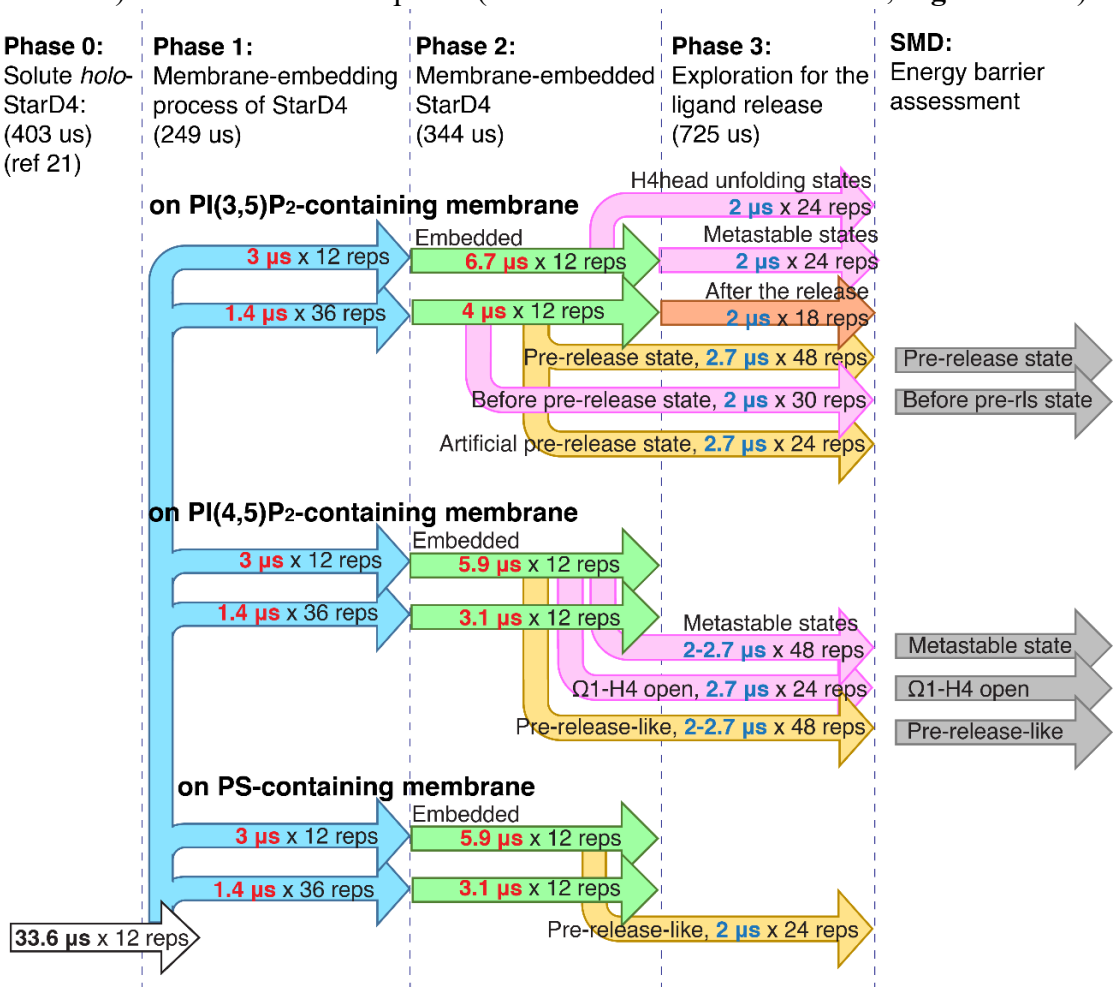

Figure 2. Schematic representation of the atomistic MD simulations of *holo*-StarD4 embedded in the three different membrane models. The labeled components of this schematic plot identify the initial conformations, the simulation time and the number of replicas in each set of simulation contained in Phases 1-3.

The spontaneous releases of the CHL cargo from the StarD4 binding site into the membrane observed in this series of simulations were analyzed to learn the sequence of mechanistic events occurring along the CHL release pathways, as illustrated below for the first complete trajectory sampled in the phase 2 (Release Trajectory 1, RT1).

The starting structure of RT1 of the *holo*-StarD4 embedded in PI(3,5)P_2-containing membranes is from a state in which the CHL had translocated to its Trp-binding-site in the hydrophobic pocket. This CHL binding mode was obtained from the simulations of the protein described in our previous publication (21) and is illustrated in **Figure 1**. In this state the Ω1-H4 front gate (between the β5-β6-loop and the N-terminal of H4) is only slightly open, but the β8-Ω4 back corridor (between the N-terminal of β8 and the β9-H4-loop) is widely open. In the RT1 trajectory we observed a metastable state that appears prior to the release of the CHL cargo in which StarD4 adopted a distinct positional orientation relative to the membrane characterized by an increased tilting angle of the H4 with the protein axis leaning in the direction of the hydrophobic pocket at an orientation angle >40°. These characteristics are depicted in **Suppl. Fig 1** and their evolution is quantified below in **Fig 3A&B** where the occurrence of this pre-release state is identified by the vertical dashed lines marking its start and its end when the CHL release phase of the simulation begins. As shown in **Fig 3C,** in this time frame the CHL tail is surrounded by a hydrophobic environment (in which the water count is reduced) and **Fig 3D** shows that the tail is brought closer to the hydrophobic core of the membrane. In this position, the cholesterol tail engages in direct interaction with the membrane through the opened Ω1-H4 gate. Subsequently, the CHL body (hydrocarbon rings) is positioned closer to the Ω1-H4 gate at the front of StarD4, and has weaker hydrophobic contacts with residues N166, C169, I189, T191 on β8 and β9 at the back of StarD4 (**Fig. 3E**). This allows water molecules to enter the created space and disrupt the interaction between cholesterol and StarD4 in this region (**Fig. 3C**). The rapid distancing of the CHL from its initial binding site at the end of this time interval, seen in **Fig 3F,** indicates that the conditions prevailing in this time interval facilitate the downward repositioning of the CHL to accomplish its release to the membrane.

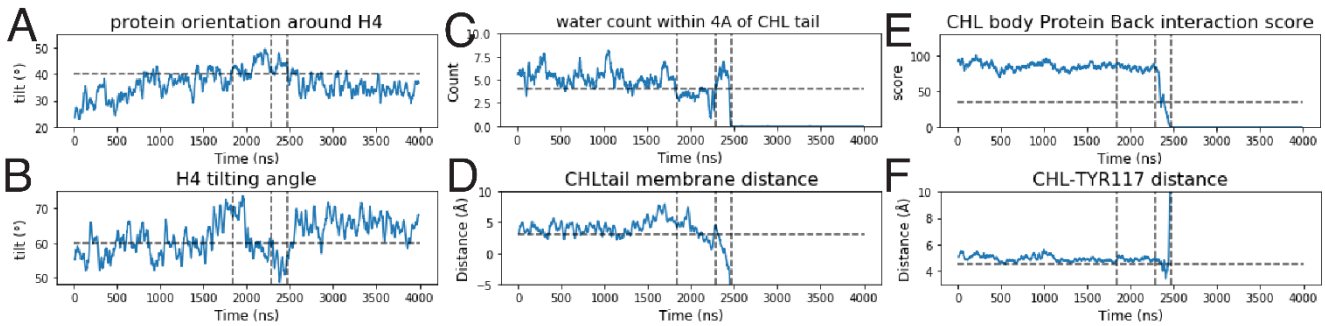

Figure 3. Formation of the pre-release state in the RT1 trajectory of the holo-StarD4 embedded in PI(3,5)P_2-containing membrane, in which StarD4 adopts a specific orientation and a molecular conformation that promotes the release of its cholesterol cargo. Panels (A-F) show the evolution of the structural characteristics of the pre-release state reflected in the measurements of: **(A)** The axial rotation of StarD4 around its H4; **(B)** The tilting angle of H4 in relation to membrane; **(C)** The number of water molecules interacting with the ligand cholesterol; **(D)** The height of cholesterol relative to the membrane; **(E)** The interaction of the CHL hydrocarbon ring with residues of β8&β9; **(F)** The distance between cholesterol and its Y117 binding site. The detailed definitions of the variables are given in the Supplementary Materials, Section 1. The first vertical dashed line marks the entrance into the pre-release; the second vertical dashed line marks the penetration of water; and the third, the rapid release of cholesterol. The auxiliary horizontal dashed lines indicate the values of the characteristic parameters of the pre-release state to emphasize the transitions.

The distinct orientation of StarD4 on the membrane changes immediately after the full release of the CHL as shown in **Fig 3A&E** where the CHL release time point is marked by the third dashed line marked in all the panels of **Fig. 3**. This indicates that the "maximal leaning towards the membrane" conformation of the StarD4 protein is a "release-enabling" configuration. In our study, this particular configuration is not observed at all in the long simulations of *apo*-StarD4, as shown in **Fig 4D-F**, and while it is rare in the simulations of *holo*-StarD4 on PI(3,5)P_2- containing membranes, it is even less frequently observed when the membranes contain only PI(4,5)P_2, and never when PS is the anionic lipid (**Fig 4A-C**).

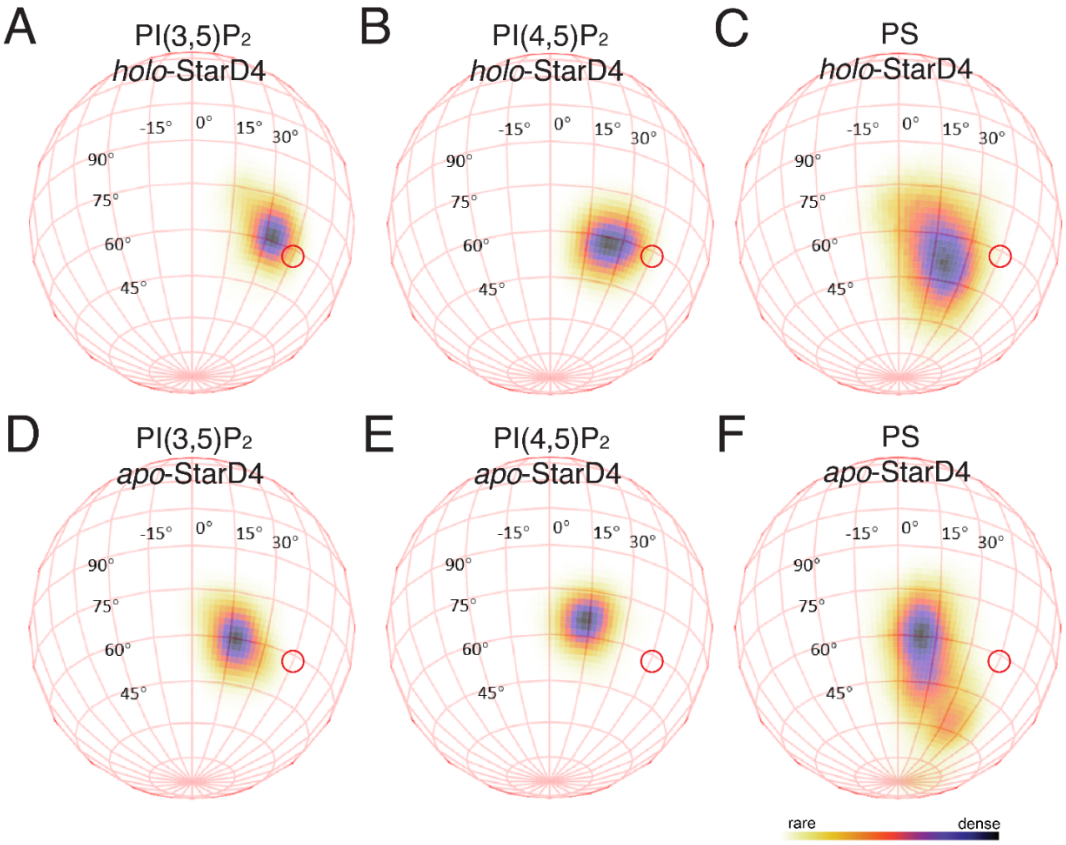

Figure 4. The probability distribution of the of StarD4 orientation in relation to the membrane. In this spherical coordinate system, the vertical axis is the tilting angle of H4, and the horizontal axis measures the axial rotation of protein around H4, as defined in Suppl. Fig. 1. Shown in **A-C** are the orientation distribution of *holo*-StarD4 on PI(3,5)P_2-, PI(4,5)P_2- and PS-containing membrane, respectively. Panels **D-F** show the orientation distribution of *apo*-StarD4 on the three types of membranes. The red circle indicates the region that is shared in the spontaneous release events occurring from the pre-release state.

<u>The temporal sequence of conformational events connecting the initial state of the simulated holo-StarD4 systems to the CHL-release state</u>: We applied the Rare Events Detection method (RED) (21,23) to the RT1 trajectory as described in Methods. Briefly, RED analysis utilizes the data of the temporal evolution of the residue-residue contacts and of the PIP2-residue-pair interaction obtained from each frame in the trajectory. The machine learning (ML) algorithm then identifies collective motions of the structural motifs, and the PIP2-interacting motifs, in each metastable state encoded in the entire trajectory (21,23). **Fig 5** shows the structural transition components identified by RED in the dynamics of *holo*-StarD4 in the RT1 trajectory (see Methods for the definition of these representations

of the structural motifs that dominate the conformation of the membrane embedded StarD4 system during a particular segment of the trajectory). Panel A shows the normalized temporal weight of each component along the RT1 trajectory time. The color code for the components is shown on the right. The weights for all the components are shown at each time point (x-axis) in a representation that stacks their values along the y-axis. For example, at time=2472ns (3rd dashed line), the weight of component 1 (comp 1, in green) is 0.11, comp 3 (in blue) is 0.83, comp 4 (light green) is 0.06, and those of components 0, 2, and 5, are 0.

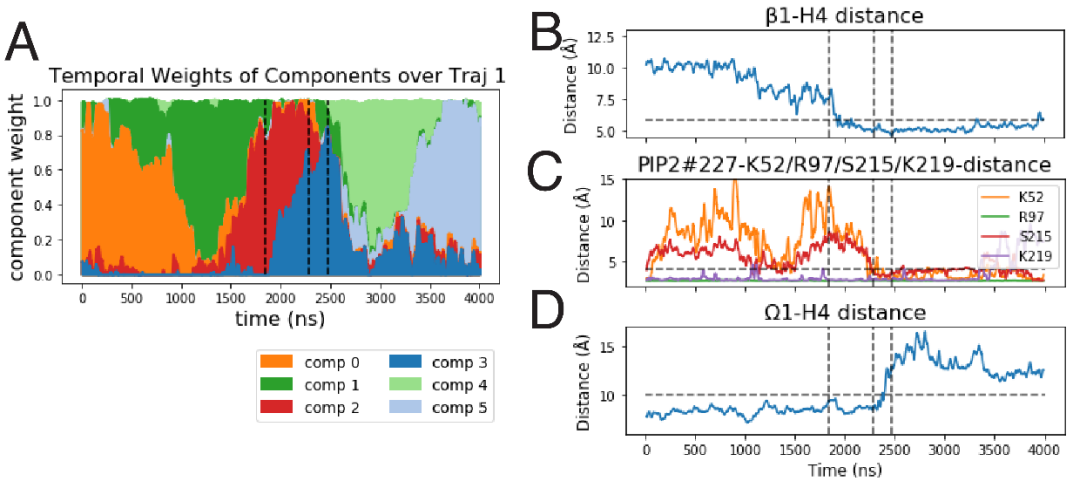

Figure 5. The Rare Event Detection analysis revealed the structural elements that determines the pre-release conformational state. (A) The RED-detected normalized temporal weight of the various components along the RT1 trajectory time (see description of the RED protocol in REF (23) and details in Methods). The color code identifies the RED-components as listed on the bottom. **(B-D)** The time-evolution of the structural elements highlighted by the structure-differentiating contact pairs (SDCPs) obtained from the RED analysis, including: **(B)** The interaction of β1-H4; **(C)** the binding of a PIP_2 simultaneously at K52, R97, S215, K219; **(D)** The destabilization of Ω1-H4 gate. The detailed definitions of the variables are given in the Supplementary Materials, section 1. The vertical dashed lines mark the same events as in Figure 3: the entrance into the pre-release; the penetration of water; and the rapid release of cholesterol.

The structural composition of each of the 6 components reveals the conformational change events occurring when each of them becomes dominant (and then wanes) in a specific time segment of trajectory frames. Fig. 5A reveals that component 3 corresponds to the conformational state that is initiated with the establishment of the hydrophobic environment for the CHL tail described in **Fig. 3C** discussed above and ends after the CHL is released. We note that although the information used by the RED algorithm does not include data about the CHL ligand and relies only on the contact matrices for protein residues and between the residues and PIP2 components of the membrane, the analysis identifies the conformation that is dominant when the CHL is released. This suggests an intrinsic allosteric coupling mechanism between the ligand CHL and the protein, as elaborated further below.

The transition from component 2 to component 3 of the RED analysis (termed event 2→3) is identified to correspond to the transition of the *holo*-StarD4 system into the pre-release state. This event is indicated in **Fig. 5A** by the reduction in the weight of component 2 as that of component 3 concurrently becoming dominant, and is initiated when StarD4 leans towards the membrane to attain the pre-release orientation at the time point indicated by the vertical dashed line at the 1840ns frame. We have shown in Ref. (21) that the specific conformational changes associated with such transitions between components are identified by direct comparison of the normalized spatial arrays (i.e., the collections of residue pairs in-contact during the conformational states represented by each component)

of adjacent components in the RED analysis. As described in the corresponding Method, this comparison identifies **s**tructure-**d**ifferentiating **c**ontact **p**airs (SDCPs) as the contact pairs that contribute significantly to the contact feature in one component, while making minimal contribution in the other component during the transition. These SDCPs are determinant elements for the interpretation of the conformational states represented by the components, as they comprise newly formed contacts in the transition from one component to another, as well as the contacts released during that transition.

The SDCPs in transition event 2→3, summarized in **Suppl. Fig 2A** and **Suppl. Table 1**, reveal the conformational changes of StarD4 and the concurrent PIP2 interactions during the transition into the pre-release state (as quantified in **Fig. 5B-D** and depicted in **Fig. 6**). In this comp2→3 transition, β1 breaks its salt-bridge with the C-terminal of H4 and forms contacts with the N-terminal half of H4 (**Fig 5B, Fig. 6B&C**). During this process, the end of β1 extends downward and initiates an interaction with a PIP2 whose phosphate groups interact simultaneously with K52 on the β1, S215 & K219 on H4, and R97 on Helical loop 3 (H3) (**Fig. 3C, Fig. 6D**). As H4 establishes interactions with β1, it moves farther from the Ω1 loop, thereby increasing the opening of the gate at the lower end of the hydrophobic pocket towards the membrane and facilitating the formation of a hydrophobic pathway for CHL release (**Fig. 3D, Fig. 6E,F**). These events in the CHL translocation process recapitulate our previous observations, in (21), from the dynamics of *holo*-StarD4 in aqueous solution (before the membrane interaction and embedding). There we present the specific coupled dynamics between the downward movement of CHL in the binding site towards the exit gate, and the formation of the β1-H4 contact that causes destabilization of the Ω1-H4 gate and the opening of the β8-Ω4 corridor. Here, with *holo*-StarD4 embedded in the membrane, we also noticed that the water penetration and the disruption of CHL interaction with the residues in β8 and β9 occurred at the same time as the establishment of this PIP2 interaction (second dashed lines, **Fig. 3C&E, Fig 5C**). Thus, nature of these conformational changes suggested to represent the steps preparing for cargo release from StarD4, are captured here by the transition event 2→3 as well as by the events preceding component 2 (**Suppl. Fig 2** and **Suppl. Table 2&3**).

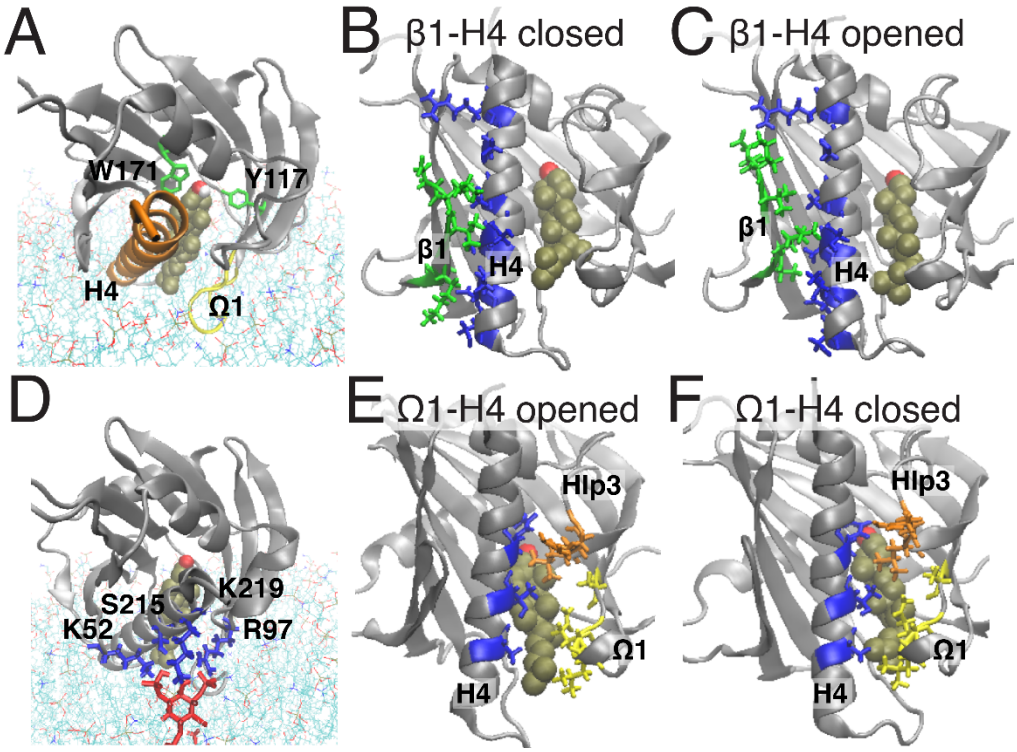

Figure 6. The set of StarD4 conformational changes and PIP_2-StarD4 interactions shown by the RED analysis to lead to the release of cholesterol. Panel (A) shows a representative conformation of the StarD4-CHL-membrane complex in the pre-release state. The StarD4 molecule is rendered in gray cartoon with H4 in orange and Ω1loop in yellow; the CHL is rendered in VDW, and its binding sites W171 and Y117 are in green licorice. Membrane lipids are rendered in lines. **Panels (B,C,E,F)** show conformations of the structural elements identified in the RED analysis: **(B,C)** depict the shift of interactions between β1 and H4; **(E,F)** illustrate the destabilization of the interactions between Ω1 and H4. In **(B,C,E,F)** StarD4 is rendered in gray cartoon and the SDCPs in licorice. The SDCPs residues on β1 are shown in green, on H4 in blue, on Hlp3 in orange, and on the Ω1 loop in yellow. Panel **(D)** shows the PIP_2-binding mode (across H4) that appeared in the pre-release state. The PIP_2 is shown in red while the basic residues it interacts with are shown in blue.

The major SDCPs in the RED analysis results identify the conformational changes of β1β2, the Ω1-H4 gate, and their interaction with PIP2, which brings to light the correlation between the specific modes of binding of PIP2 and the conformational change of StarD4. Indeed, prior experimental results (2,9,12) had shown that (i)-certain basic residues on the membrane-interacting surface of the protein have a critical role in mediating the cholesterol transport function of StarD4, and (ii)-that different PIP2 subtypes exhibit varying preferences for binding to these basic residues (12). We reasoned, therefore, that the characteristic mode of interaction between PIP2 and the basic residues in the pre-release state defined here serves as the basis for PIP2 recognition by StarD4. The subsequent section presents an in-depth analysis of these PIP2-binding modes and their relation to the release of CHL on the different membranes.

The mechanism of cargo release from the *holo*-StarD4 protein is determined by the PIP2 content of the target membrane

<u>The mechanistic role of the pre-release state in the delivery of StarD4 cargo</u>: To reveal the specifics of conformational states visited by the membrane-embedded StarD4 in the CHL release process we analyzed the trajectory data from Phase 2 of the simulations (**Fig. 2**) with the dimensionality reduction algorithm tICA (time-structure based independent component analysis) as described in Methods. In the

projection to the tICA space, the high dimensional data from the simulation trajectories are projected onto a tICA space spanned by collective variables (CVs) that describe the CHL binding modes and changes in structural motifs of StarD4 that determine the functional dynamics of interest. Six such CVs were chosen to describe the conformational changes observed in the β1, H4, Ω1, Ω4, and CHL binding site motifs of StarD4 (**Fig. 7C**). These parameters were used to define the tICA space shown in **Fig. 7A-B**. The first two tICA vectors (**Fig. 7C**) captured 61% of the total dynamics of StarD4 in the membrane-embedded phase defined in **Fig. 2**, phase2.

The tICA representations in **Fig. 7A-B** illustrate the specific ensemble of conformational states adopted by the membrane-embedded *holo*-StarD4 on membranes containing different subtypes of PIP2s. The PS-specific conformational ensemble and the combined ensemble are depicted in **Suppl. Fig. 3A,B**. Superimposing the time-evolution of RT1 onto the tICA space (**Suppl. Fig. 3C**) reveals that RT1 originated from state **2** in **Fig. 7A** and transited to state **1** where the spontaneous cholesterol release event occurs. The structural characteristics of these PIP2-specific states were discerned from the distribution of collective variables (CVs) representing the motif dynamics as detailed in **Fig. 7D**. Notably, it confirms that the pre-release state of StarD4 observed in RT1 as described above is characterized by the interactions between β1-H4 (6th column), the change in protein orientation (8th column), and the wide opening of the β8-Ω4 corridor (3rd column).

In addition to the pre-release state (**Fig. 7A&D**, state **1**), the identified states **2-8** include those with CHL at the Ser-binding site (**Fig. 7**, states **4**&**8**), and conformational states with CHL at the Trp-binding site. The conformational states with CHL at Trp-binding site include some that are found in all membrane-embedded *holo*-StarD4 trajectories (**Fig. 7**, states **2&5**) and others that are specific to one or the other kind of PIP2 subtype composition (**Fig. 7**, states **3,6,7**).

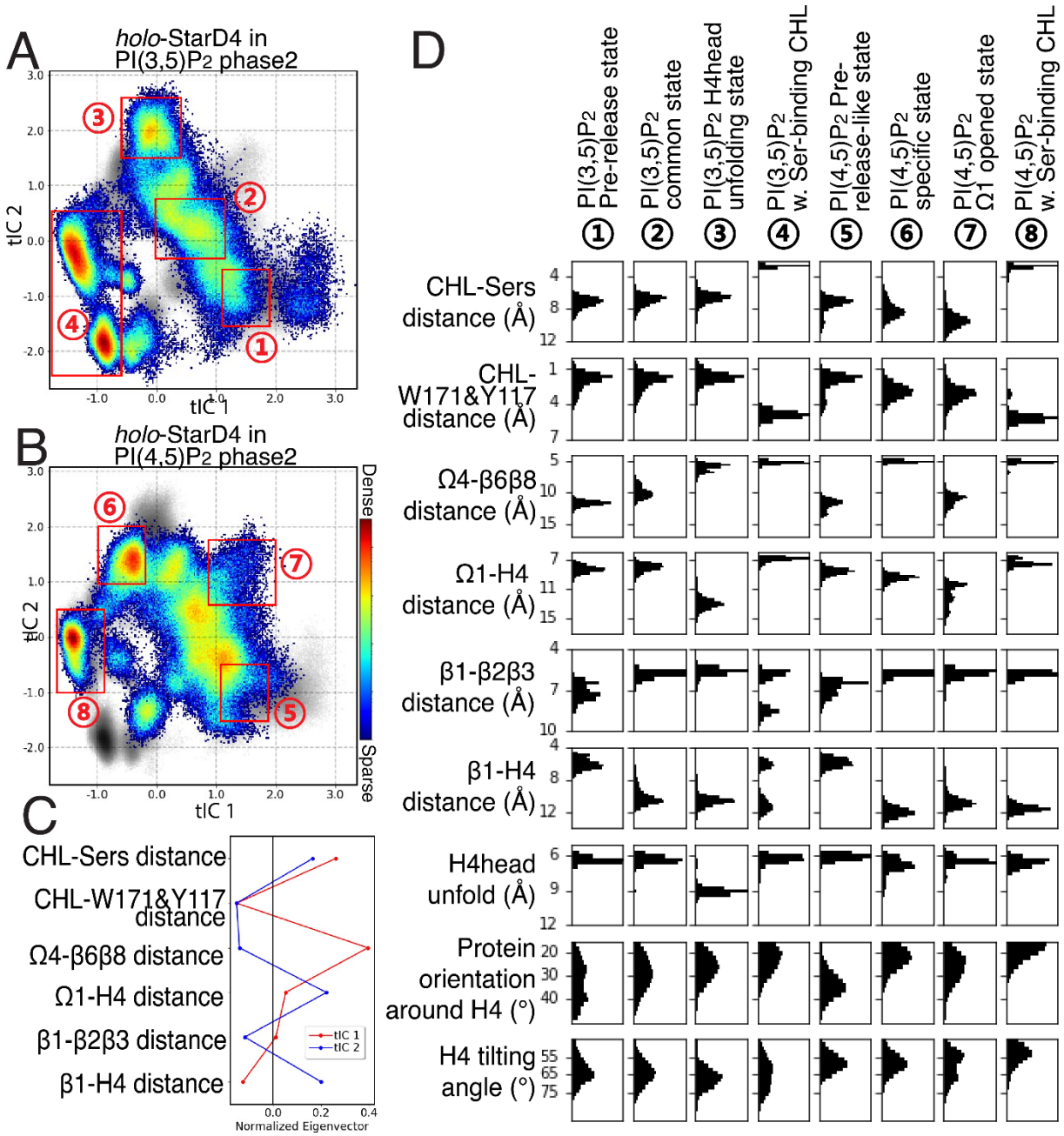

Figure 7. The characteristics of conformational states of membrane-embedded *holo*-StarD4. The high dimensional simulation data are projected onto a 2D tICA space defined on CVs describing the CHL binding modes and the conformational changes of StarD4. **Panels (A,B)** present the population density maps of the conformational space of *holo*-StarD4 embedding on the PI(3,5)P_2-containing membrane **(in A)** and on the PI(4,5)P_2-containing membrane **(B).** The tICA space is spanned by the two vectors (tIC 1, tIC 2) defined in **Panel (C)** by the contributions of individual CVs. The definitions of the CVs are listed in the Supplementary Materials, section 1. The color scale for the representation of population densities is shown on the right. The gray scale in the background represents the combined population density of StarD4s on three types of membranes **(Suppl. Fig. 3B)**. **Panel D** presents the structural characteristics of the pre-release state, the PIP_2-subtype-specific states, and the states common to the protein embedded in membranes regardless of the PIP2 subtype. The states are identified the 8 red boxes in **(A,B)** and their characteristics are represented by probability density histograms of CVs that include the binding of CHL on the Ser-site and on the Trp-site; the interactions of β8-Ω4 corridor, Ω1-H4 gate, H4-β1, β1-β2&3 interactions, H4 partial unraveling, as well as the orientations of StarD4.

The characteristic features of the CHL-release observed in RT1 are reproduced in other Phase 3 simulations: To follow the CHL-release process under various conditions we conducted adaptive samplings simulations from initial seeds taken along the series of conformational changes that lead to the pre-release states of *holo*-StarD4. These pre-release states share similar characteristic features in each of the membrane systems studied – the PI(3,5)P_2-containing membranes (**Fig. 7A**, state 1), the PI(4,5)P_2-containing membranes (**Fig. 7B**, state 5) and PS-containing membranes. As shown in (**Fig. 2**, Phase3, orange set) the cumulative simulation time was 355 microseconds. In this set of simulations, the CHL release process described for **RT1** was successfully reproduced in two additional simulation on PI(3,5)P_2-containing membrane (termed **RT2** and **RT3**), and in one trajectory from the StarD4 embedded in PS-containing membranes (**RT4**) which also shared features of the pre-release state. Additionally, we started a set of 24 trajectories from artificial pre-release states in which the β1-H4 interaction was maintained and the StarD4 was manually positioned in the pre-release strongly leaning orientation (see **Fig 4**) on the PI(3,5)P_2-containing membrane and a PIP2 binding in the characteristic cross-H4 mode. One of these trajectories (**RT5**) also reproduced the CHL release process, albeit partially with cholesterol moving halfway out of the pocket (**Suppl. Fig. 4A**).

The characteristic features observed in RT1 that re-appeared during the cholesterol release process in all these trajectories included the binding of an anionic lipid at K52 and R97 (**Suppl. Fig. 4B**), the β1-H4 interaction and destabilization of the Ω1-H4 gate (**Suppl. Fig. 4C**), the wide opening of the β8-Ω4 corridor (**Suppl. Fig. 4C**), and the orientation of StarD4 relative to the membrane surface (**Suppl. Fig. 4D**). While StarD4 may deviate from this pre-release orientation just before the release process in some of these trajectories, when cholesterol is destabilized from its binding sites in the pocket it consistently rearranges back into the rarely visited orientation on the membrane, leaning towards the hydrophobic pocket. This underscores the pivotal role of StarD4's orientation in cholesterol transportation.

A low probability release mechanism from a high energy initial state: By initiating additional simulations in Phase 3 (**Fig. 2**, phase3, pink set, 334 μs) we explored possible alternatives, other than the canonical pre-release state, as starting points for CHL release pathways. These simulations were initiated from different conformational states of StarD4 with CHL bound at the Trp-binding site (**Fig. 7A,B,** states 2,3,6,7), as indicated by the scatter points on the tICA maps in **Suppl. Fig. 5**. Only two CHL release trajectories were observed (**RT6**, **RT7**), and remarkably, both of them started from a high-energy state (see initial frames situated at the upper right corner of state 7 in **Fig. 7B**) where the population density is very low. The initial structures of the (**RT6**, **RT7**) set feature an Ω1-open configuration on PI(4,5)P_2-containing membranes (**Fig. 7B** state 7), characterized by the wide opening of the Ω1-H4 gate (**Fig. 7D**, 4th column), and they shared with the pre-release state a similar pattern of a widely opened β8-Ω4 corridor (**Fig. 7D**, 3rd column), but without the characteristic β1-H4 interaction (**Fig. 7D**, 6th column). These features are likely related to the membrane lipid tail that had inserted between the Ω1 loop and the CHL in the initial frames (**Suppl. Fig. 6A**), which resulted in a persistent and wide opening of the Ω1-H4 gate (centered at CV "Ω1-H4-dist"=14Å). Notably, this feature is not observed in the pre-release state shared among RT1-RT5 (**Fig. 7D**, 4th column, **Suppl. Fig. 6B,C**).

To assess the likelihood of CHL release from the pre-release state shared by RT1-RT5, compared to a release from RT6&7, we compared the free energy barrier of CHL release in all these trajectories. The release paths were first explored using Steered Molecular Dynamics (SMD) simulations (26,27) as described in Methods. To this end, initial frames were selected from the states of the pre-release conformations on PI(3,5)P_2-containing membrane (**Fig. 7A**, state **1**) and on

PI(4,5)P_2-containing membrane (**Fig. 7A**, state **5**), as well as from the state containing the conformations with opened Ω1 on PI(4,5)P_2-containing membrane (**Fig. 7B**, state **7**). The ligand CHL was steered along the z-axis towards to membrane at an average speed of 0.05 Å/ns for 400 ns in each run, with 24 replica runs per group. The free energy profiles were then evaluated with umbrella sampling protocols (see (28,29) and Methods) along the pathway resulting from the SMD.

The free energy profiles calculated for membrane systems containing either PI(3,5)P_2 or PI(4,5)P_2 are shown in **(Fig. 8)**. The calculated total energy barrier for CHL release from the identified pre-release state was 2.6 kcal/mol (**Fig. 8A**), but it was 4.0 kcal/mol for the release pathway from the Ω1-opened state (**Fig. 8B**). This observation suggests that the kinetics of CHL release from the pre-release state would be approximately 10-fold faster than from the Ω1 opened state. The same free energy profile evaluations were applied to other highly populated StarD4 conformational states with CHL at the Trp-binding site (**Fig. 7**, states **2,5,6**), and these showed higher free energy barriers compared to those starting from the pre-release structures on PI(3,5)P_2-containing membrane (**Suppl. Fig. 7**).

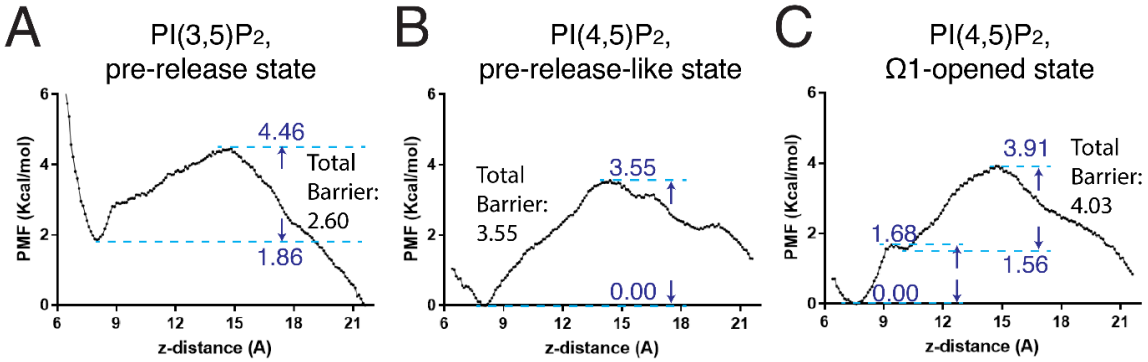

Figure 8. The energy barrier along the CHL release path sampled with Steered MD simulation and evaluated with umbrella sampling. The barrier is represented in terms of the calculated potential of mean force (PMF). The reaction coordination is the z-distance between the CHL and StarD4. Panels **(A-C)** show the PMF values for CHL release initiated from: **(A)** the pre-release state of StarD4 on PI(3,5)P_2-containing membrane; **(B)** the pre-release-like state on PI(4,5)P_2-containing membrane; and **(C)** the low probability Ω1-opened state on PI(4,5)P_2-containing membrane. Local maxima and minima are labeled along the reaction pathways, and the calculated Total Barrier is indicated.

Thus, we find that CHL release from the *holo*-StarD4 systems is energetically preferred from a state exhibiting the features we identified for the pre-release state. The results show that among the conformational states explored in the various long trajectories of the extensive simulations, the pre-release state holding the highest promise for CHL release shares the following features: (1) A position of StarD4 on the membrane that leans maximally in one particular direction (**Fig. 4**); (2) The protein conformation includes an interaction between the N-term of H4 and β1 and the H4 N-terminal; (3) The β8-Ω4 corridor is open; (4) A PIP2 lipid is binding at K52 on β1 and R97 on H3. These features are the results of specific modes of interaction with the PIP2 containing membranes that are communicated in the membrane embedded *holo*-StarD4 through the allosteric mechanisms we described. They determine the dynamics of CHL release as described below.

The modes of PIP2-subtype binding to *holo*-StarD4 are related to the dynamics and repositioning of β1

We observed that one of the key features of the pre-release state is the PIP2 binding at the K52 of β1 and R97 of H3 which positions it across H4. To learn whether the PIP2-StarD4 interaction mode correlates with specific conformational changes of StarD4, we investigated all the binding modes of the PIP2 membrane components to StarD4. We found that the basic residues of StarD4 that interact with membrane anionic lipids are distributed in three groups with specific locations, as illustrated in **Fig 9A**: the *upper-group* (K52, R97, S215, R218, K219, R222), the *left-group* (R46, K49, K50, K52, R58) and the *right-group* (R116, R158, R163, R130, R194).

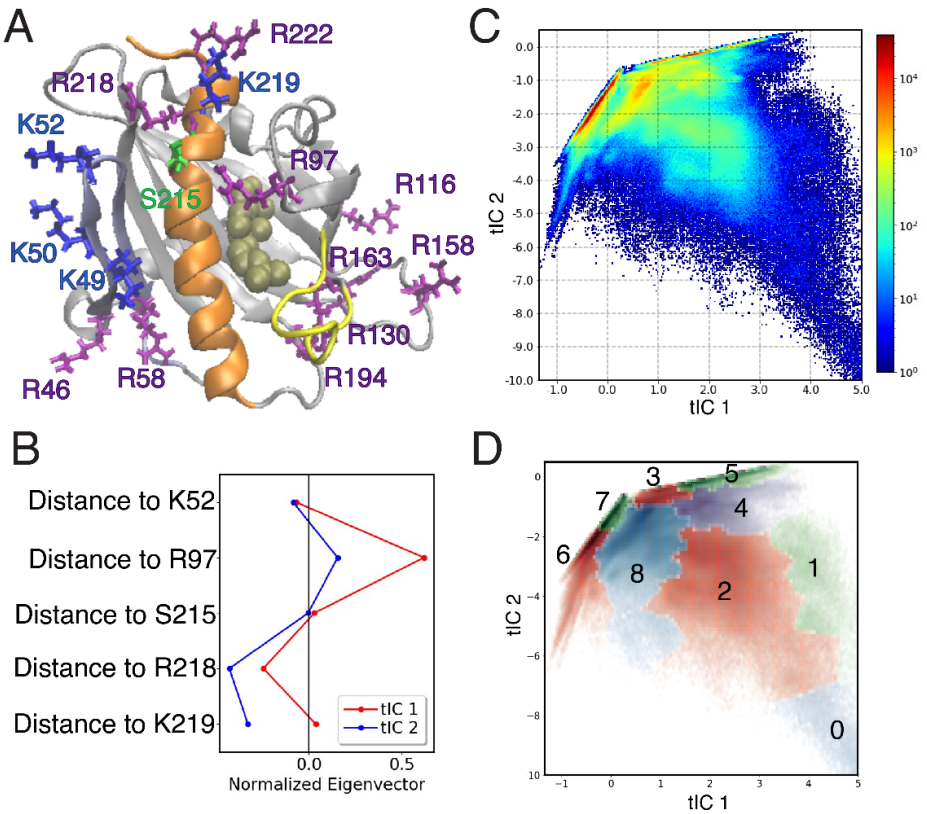

Figure 9. The probability map of PIP2 on putative interaction sites. (A) Basic residues on the StarD4-membrane interface. The StarD4-CHL complex is rendered the same way as figure 1, with the lysine's shown in blue licorice, Arginine's in Purple, and Ser215 in green.
(B) Contributions of individual CVs to the first 2 dimension of the tICA space. The definitions of the CVs shown in this panel are detailed in the supplementary materials section 2. **(C)** The population density map of the 2D tICA space representing the StarD4-interaction modes of PIP2s. The color scale represents the density and is shown on the right. **(D)** The 2D tICA map is color-coded by the lipid binding modes 0-8.

Residues within the same group are located adjacently, leading to a tendency for shared PIP2 interactions among these residues. The binding mode observed in the pre-release state involves a PIP2 traversing the K52, S215, R218 and R97, which is an example of the PIP2-interaction at the upper-group. To facilitate the analysis of the PIP2 subtype interaction mode preferences in this upper-group, we constructed the Interaction Mode Landscape (IML) shown in (**Fig 9B-C)** as described in Methods. This binding mode map is further discretized into 9 macrostates based on their kinetic similarities, named as binding mode 0 to 8 (**Fig 9D**) (detailed in Methods). Each binding mode is characterized by

the basic residues with which it interacts, as depicted in the histograms of PIP2-residue distances (**Suppl. Fig 8**) and are summarized in **Table 1**.

To study the subtype-based interaction mode preference, we separately projected the position of PI(3,5)P_2 and of PI(4,5)P_2 on the 2D IML map. As shown in **Fig. 10A,B** and **Table 1**, PI(4,5)P_2 exhibits a preference for states 3,4 and 5 where its phosphate groups gather in a narrow region. In contrast, PI(3,5)P_2 shows a preference for binding mode 7, where its phosphate groups bind across H4 from β1 to Hlp3 (**Fig. 10C,D**).

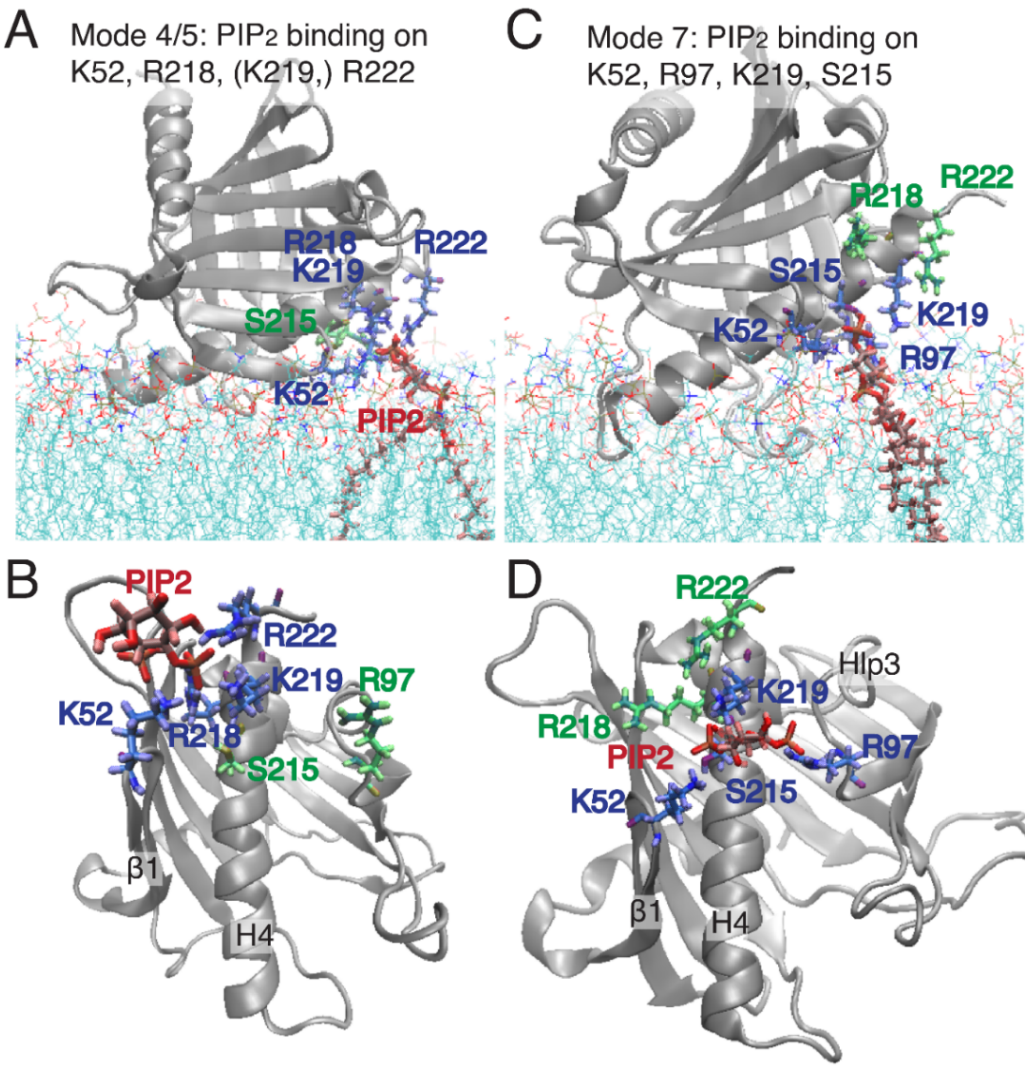

Figure 10. Representative conformations of PIP_2-subtype-specific binding modes. (A,B) The modes of PIP_2 binding to the *upper-group* residues K52 on β1, and R218 K219 R222 on H4, in lateral view **(A)** and upward view **(B)**. **(C,D)** The "cross-H4"-binding mode to the *upper-group* residues in which the PIP_2 is binding with K52 on β1, S215 K219 on H4, and R97 on Hlp3, in lateral view **(C)** and upward view **(D)**. The StarD4 is rendered in Gray cartoon. The *upper-group* basic residues K52 R97 S215 R218 K219 R222 are shown in licorice, with the PIP_2-binding residues in blue and the non-interacting residues in green. The PIP_2 is shown in red licorice. The membrane is shown only in the lateral views.

Notably, binding mode 7 corresponds to the binding mode observed in the pre-release state. The occurrence rates of the binding modes in the PI(3,5)P_2- and PI(4,5)P_2-systems are compared in **Table 1**.

Table 1: The occurrence rates* of the PIP_2 binding modes in the *holo* and *apo* StarD4 systems on membranes with different PIP2 compositions

Mode ID	Characteristics of the Binding Mode	Occurrence rate for *holo*-StarD4		Occurrence rate for *apo*-StarD4	
		$PI(3,5)P_2$	**$PI(4,5)P_2$**	**$PI(3,5)P_2$**	**$PI(4,5)P_2$**
Mode 0	Unbound **	0.6%	0.2%	0.0%	0.0%
Mode 1	Binds on K50. close to K52. **	0.2%	1.0%	0.0%	0.1%
Mode 2	Binds on K50, K52. close to K49.	21.6%	20.7%	21.0%	13.2%
Mode 3	Binds on K52, K219, R222. Close to S215, R218.	12.1%	13.2%	**4.7%**	**27.4%**
Mode 4	Binds on K52, R218, R222.	15.5%	22.3%	**7.0%**	**19.0%**
Mode 5	Binds on K52, R218, K219, R222.	3.3%	3.7%	**0.1%**	**62.1%**
Mode 6	Binds on R97, K219.	42.0%	54.3%	33.1%	15.5%
Mode 7	Binds on K52, R97, K219. Close to S215.	**53.2%**	**32.8%**	64.9%	49.2%
Mode 8	Binds on K52, K219, R222.	65.5%	47.0%	28.6%	15.6%

*Note that the total occurrence %-ages does not sum up to 100% due to simultaneous binding of multiple lipids.

**The occurrence of PIP_2s in states 0&1 are under-estimated, because in these binding modes the PIP_2 lipids don't match the criteria of "PIP_2s binding at the upper basic residue patch", as described in Methods.

As shown above by the results from the analysis with the RED algorithm, the conformational changes that StarD4 undergoes in the transition into the pre-release state (i.e., transition event 2→3), involve a PIP2 binding in mode 7 when the β1 establishes interactions with the N-terminal half of H4 (**Fig. 5A,C**). We therefore investigated the role of this binding mode by comparing the β1 to H4 distance in the presence and absence of a PIP2 binding at mode 7. This brought to light the function enabling correlation between PIP2 binding and the conformational changes in StarD4 (**Fig. 11A**). Thus, the results showed that the correlation between the PIP2 binding mode and the conformational change at the β1-H4 was present not only in the pre-release state but also throughout the entire simulation of membrane-embedded StarD4. This is important mechanistically because we found the β1-H4 interaction to be associated with the opening of the β8-Ω4 back corridor and the destabilization of the Ω1-H4 gates on the pathway of cholesterol release (21), and this correlation is quantified in the following section. Moreover, the binding of $PI(4,5)P_2$ in mode 7 promotes as well another feature of the release state, which is the reorientation relative to the membrane that causes the protein to lean towards the side of hydrophobic pocket (**Fig. 11B**). Collectively, these findings suggest not only that the occurrence of the cross-H4-binding of PIP2 is preferred in the pre-release conformational changes

in StarD4, but also that it underlies the mechanism of PIP2-subtype recognition by StarD4 and its role in the regulation of the cholesterol trafficking function.

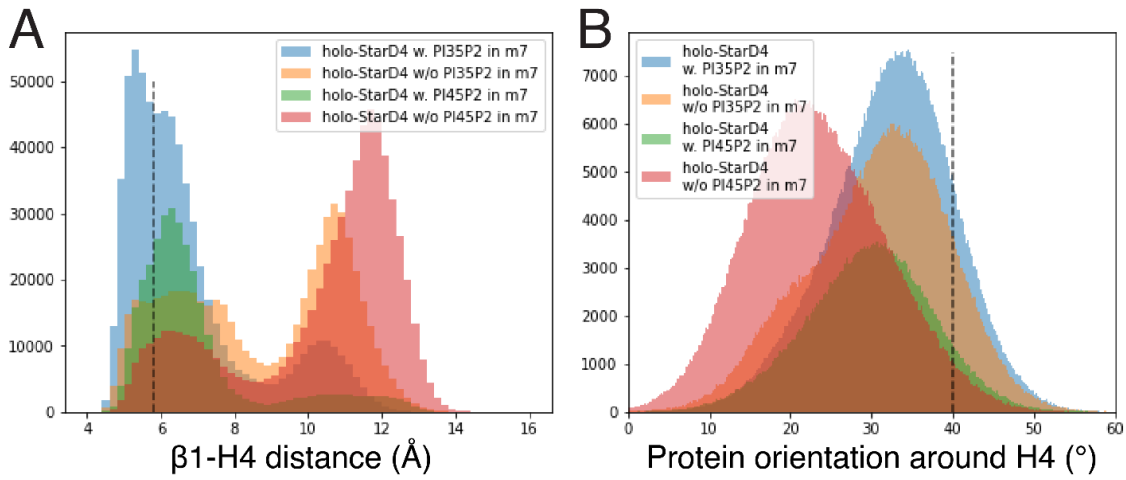

Figure 11. Comparison of the *holo*-StarD4 dynamics with and without PIP_2 binding in the cross-H4 binding mode. (A) The probability density histogram of the β1-H4 distance. **(B)** The probability density histogram of the protein orientation. The vertical dashed lines indicate the values of these characteristic parameters in the pre-release state.
Color code: In blue: the dynamics of StarD4 with a $PI(3,5)P_2$ binding at the cross-H4 binding mode. In orange: the dynamics of StarD4 on $PI(3,5)P_2$-containing membrane without a cross-H4-binding lipid. In green: the dynamics of StarD4 with a $PI(4,5)P_2$ binding at the cross-H4 binding mode. In red: the dynamics of StarD4 on $PI(4,5)P_2$-containing membrane without a cross-H4-binding lipid.
Statistical test: with a time-interval of 2μs per snapshot to eliminate the autocorrelation, the simulations are equivalent to 134, 113, 65, 116 independent samples in the four categories separately. For the comparison of β1-H4 distance, the statistical significance are tested using Mann-Whitney U Test: with & without cross-H4-binding-$PI(3,5)P_2$, $p=3.4e^{-15}$, with & without cross-H4-binding-$PI(4,5)P_2$, $p=1.1e^{-16}$. For the comparison of protein orientation around H4, the statistical significance are tested using T Test: with & without cross-H4-binding-$PI(3,5)P_2$, $p=0.073$, with & without cross-H4-binding-$PI(4,5)P_2$, $p=4.8e^{-11}$.

The allosteric communication between the PIP2-binding modes and the dynamics of the CHL binding pocket.

Having identified the relation between structural motifs of StarD4 involved in the conformational changes to the pre-release state, and the motifs responsible for the differential response to interactions with different PIP2 subtypes, we proceeded to quantify their allosteric communication with the cholesterol binding site. To this end, we applied the N-body Information Theory (NbIT) analysis (24,25) to the trajectories of *holo*-StarD4 on the $PI(3,5)P_2$-containing membrane in the membrane-embedded phase. The relevant component of the NbIT analysis, which can reveal the structural composition of the allosteric network connecting these sites and quantify its strength, is the normalized coordination information (NCI) between motifs. In NbIT this is quantified from the information shared between a specified *receiver* motif and a *transmitter* motif, as described in Methods. The NCI values calculated in this manner between motifs of mechanistic interest are presented in **Table 2** using the residue indexes listed in Methods.

Table 2. Normalized Coordination Information values calculated between various structural motifs in the *holo*-StarD4-membrane complex acting as *transmitters* (columns) and *receivers* (rows).

Transmitters

Receivers	β1	β2&β3	β2β3 loop	H4-Nt	Ω1 loop	Ω4 loop	PIP_2-binding Mode (Pbm)	water influx	CHL sites
β1	11.51	23.5%	9.0%	18.6%	1.3%	10.6%	29.3%	10.6%	7.2%
β2&β3	24.3%	12.62	30.5%	22.8%	3.7%	16.9%	14.4%	23.5%	9.8%
β2β3loop	11.1%	37.5%	7.89	10.9%	1.8%	9.1%	12.9%	14.9%	5.6%
H4 head	20.9%	24.7%	9.3%	17.46	6.3%	30.5%	11.6%	19.1%	10.7%
Ω1 loop	1.6%	4.8%	2.4%	10.1%	8.45	6.8%	3.3%	3.6%	2.9%
Ω4 loop	17.0%	21.1%	9.8%	37.5%	3.7%	5.11	9.1%	31.7%	11.6%
Pbm	33.0%	28.5%	20.5%	28.8%	5.8%	21.5%	0.88	27.5%	30.6%
water influx	18.6%	33.8%	16.3%	26.1%	4.1%	41.8%	13.7%	5.37	23.5%
CHL sites	22.6%	25.9%	15.2%	26.3%	8.1%	25.6%	30.2%	41.9%	1.18

NOTE: Values highlighted on the diagonal represent the Total Correlation for each motif (24,25).

The values in Table 2 quantify the extent of information sharing between the motifs which provides insights into their functional connectivity. Notably, these values are much larger than the NCI values calculated for motifs that are not expected to be involved in allosteric pathways (see **Suppl. Table 4**), indicating the background level of coordination information between unrelated motifs due to the massive dynamics data contained in the MD trajectories. The pattern of communication revealed by the NCI values in Table 2 recapitulates the allosteric network reported recently (21) from simulations of the same system in aqueous solution. For example, β1 remains a strong coordinator to the cholesterol binding site in the membrane-embedded StarD4 although no direct interaction is formed between these motifs. In the embedded system, the importance the of this allosteric connection between β1 and the CHL binding site is illuminated by the finding that the β1 contains the basic residue that respond to the PIP2-subtype and is further coordinated with H4 dynamics. The allosteric network that channels the information from β1 to the CHL binding site in StarD4 involves β1 sharing dynamic information with β2, β3, the H4 N-terminal (H4-Nt). In turn, β2, β3, H4 share dynamic information with β9 & Ω4-loop, and H4 β9 & Ω4 then directly gating the release of CHL (**Fig. 12A**). Remarkably, all the motifs in this allosteric pathway are thus identified as strong coordinators to the cholesterol binding site. With the strong information sharing between the PIP2-subtype-specific binding sites on StarD4 (PIP2-binding mode, Pbm), and the β1, the results confirm and support the role of β1 as the key site for differentiating the PIP2-subtype, and of the Pbm as another strong coordinator of the cholesterol binding site.

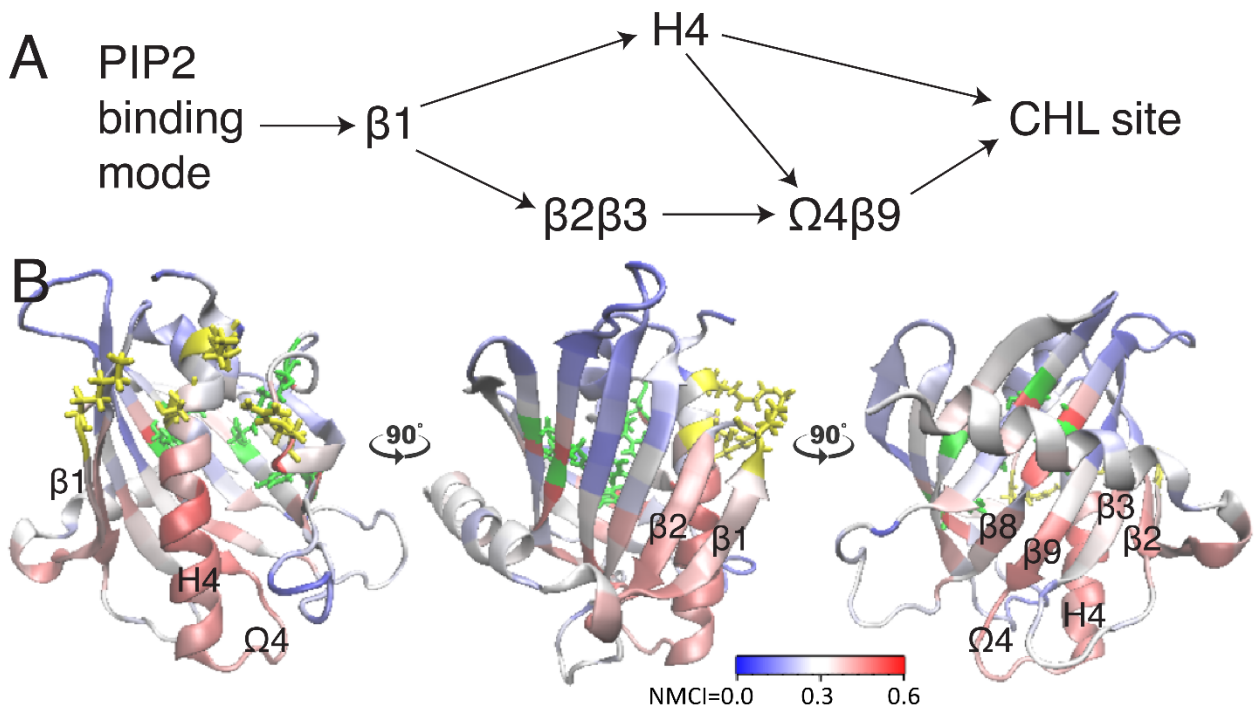

Figure 12. The allosteric coordination channel between the PIP$_2$ binding site and the CHL binding site, which involves Helix4, β1, β2, β3 β9 sheets and Ω4 loop. (A) An allosteric model of information transmission from the PIP$_2$ at cross-H4-binding mode to the cholesterol binding site. The arrows denote the direct information flow identified by the NbIT analysis between interacting motifs. **(B)** The cartoon model of StarD4 is colored according to the value of the normalized mutual coordination information (NCMI) calculated with the NbIT analysis. The color scale that quantifies the Normalized Mutual Coordination Information is at the lower right.

The coordination channel for information transmission from the PIP2 recognition site to the cholesterol binding site is quantitatively identified by calculating the Normalized Mutual Coordination Information (NMCI) term in NbIT (24,25), as described in Methods. Our results are presented in **Fig.12A** in terms of a schematic channel, and the panels in **Fig. 12B** quantify the β1, β2, Ω4 and H4 coordination channel in the structural context of StarD4. Together, the three panels in **Fig. 12B** depict the pattern of NMCI values of the transmission of information from the PIP2 binding site to the cholesterol binding site. The path of the PIP2 subtype-specific information transmitted through the allosteric network comprising the indicated structural motifs determines the preference for pre-release conformational changes that govern CHL release in StarD4.

While the information sharing among β1, β2, Ω4, H4-Nt to the CHL site in the complete trajectory is strong, we note that when the NbIT analysis is used on trajectories starting from the already formed pre-release state of StarD4 on PI(3,5)P$_2$-containning membrane, the calculated information sharing values are weaker (see **Table 3**). This observation confirms that the necessary conformational changes in StarD4 have already occurred in the pre-release state, and the allosteric effect is no longer activated. Importantly, however, the dynamics of the CHL binding site continue to exhibit a strong coupling with the dynamics of residues N166 C169 on β8, and I189 T191 on β9 (referred to as the "water influx" site in **Table 2&3**). Indeed, we had observed that the last event before the release of CHL from the pre-release state of the StarD4 is the destabilization of the interactions of these motifs and the penetration of water to the site.

Table 3. Normalized coordination information between allosteric sites in *holo*-StarD4-membrane complex, analysis in the pre-release state only

Transmitters

Receivers	β1	β2&β3	β2β3 loop	H4-Nt	Ω1 loop	Ω4 loop	PIP_2-binding Mode (Pbm)	water influx	CHL sites
β1	11.93	18.8%	5.6%	14.0%	1.3%	6.3%	24.6%	5.5%	3.9%
β2&β3	20.1%	11.25	27.6%	20.5%	2.7%	16.9%	9.8%	24.6%	7.5%
β2β3loop	6.3%	35.0%	7.00	12.1%	1.7%	12.2%	10.2%	18.0%	4.9%
H4-Nt	16.5%	22.6%	9.3%	16.70	3.4%	24.2%	8.7%	12.7%	6.3%
Ω1 loop	1.6%	2.2%	1.2%	2.7%	6.91	2.3%	2.8%	2.3%	2.2%
Ω4 loop	7.0%	12.3%	7.0%	21.2%	2.1%	5.54	4.0%	17.8%	3.5%
Pbm	23.0%	19.0%	14.3%	20.1%	6.7%	13.9%	0.83	17.8%	25.4%
water influx	8.0%	31.9%	18.5%	13.3%	2.8%	31.5%	7.5%	4.45	13.3%
CHL sites	11.1%	13.8%	9.3%	13.1%	5.4%	9.5%	24.9%	30.8%	1.00

NOTE: Values on highlighted on the diagonal represent the Total Correlation for each motif (24,25).

Deep Neural Networks analysis identifies the structural motifs that respond dynamically to the recognition of specific PIP2-subtypes

As the subtype-dependent PIP2 binding mode is found to be allosterically coupled with the cholesterol binding site through a series of conformational changes along an allosteric network in StarD4, we sought to identify the structural motifs that respond to the specific subtype of PIP2 in the membrane composition. To this end we used Deep Neural Network (DNN) analysis to differentiate first between the PIP2-subtype dependent StarD4 conformations, and then point out the molecular determinants responsible for the PIP2-subtype dependent StarD4 conformational changes.

To prepare the DNN to distinguish between StarD4 modes of interaction with $PI(3,5)P_2$-containing membranes and with $PI(4,5)P_2$-containing membranes in the *holo*-StarD4 trajectories we employed a supervised DNN training pipeline developed in our lab (30). The input datasets consisted of ~46,000 structures defined by the coordinates of protein atoms only, and the DNN was designed to return the PIP2-subtype in the membrane with which the protein structure was interacting. These included data from both the membrane-embedding phase 1 and the membrane-embedded phase 2, sampled at 8ns time-intervals in >180 μs trajectories for each of the systems (**Suppl. Table 6**). The complete dataset was split into sets for training, validation, and test in the ratio of 56:24:20, respectively.

Test set results reached an accuracy of >85% in predicting the subtype of PIP2 (**Table 4**, left). The accuracy achieved for trajectories of the membrane embedding phase of StarD4, in which it had not yet undergone any PIP2-induced conformational changes, is difficult to improve further. However, for the test set based on the membrane-embedded trajectory data, the DNNs achieved significantly higher accuracy, with 95.0% accuracy in predicting and differentiating the interacting PIP2 subtypes based solely on the conformational features of *holo*-StarD4 (**Table 4**, right).

Table 4. Results of the classification by DNNs

		Assessed on the full test set				Assessed on the test set samples with membrane embedded StarD4	
		Prediction				Prediction	
		$PI(3,5)P_2$	$PI(4,5)P_2$			$PI(3,5)P_2$	$PI(4,5)P_2$
Truth	$PI(3,5)P_2$	85.9%	14.1%	Truth	$PI(3,5)P_2$	95.0%	5.0%
	$PI(4,5)P_2$	10.4%	89.6%		$PI(4,5)P_2$	5.1%	94.9%

As the high-accuracy results suggest that the neural networks were successful in recognizing the subtype-specific PIP2-induced conformational changes in the *holo*-StarD4, we proceeded to extract the salient structural features in the PIP2-subtype prediction by the DNNs. To this end we computed *attention heatmaps* featuring the atoms whose conformational changes exerted the greatest influence on the decision of the DNNs (**Suppl. Fig. 9**). The atoms with high attention values are found to be those located mainly in the following motifs: β1, β2, β3, Ω1-loop, β8, β9, and Ω4-loop (**Fig 13**). The salient residues contained those on β1 that interact with anionic lipids (12), and critical components of the allosteric network described above, such as the β8-Ω4 corridor and the Ω1-H4 gate. Notably, the NbIT analysis had shown that these same structural motifs play a crucial role in coupling the CHL binding site with the anionic lipid binding site on β1.

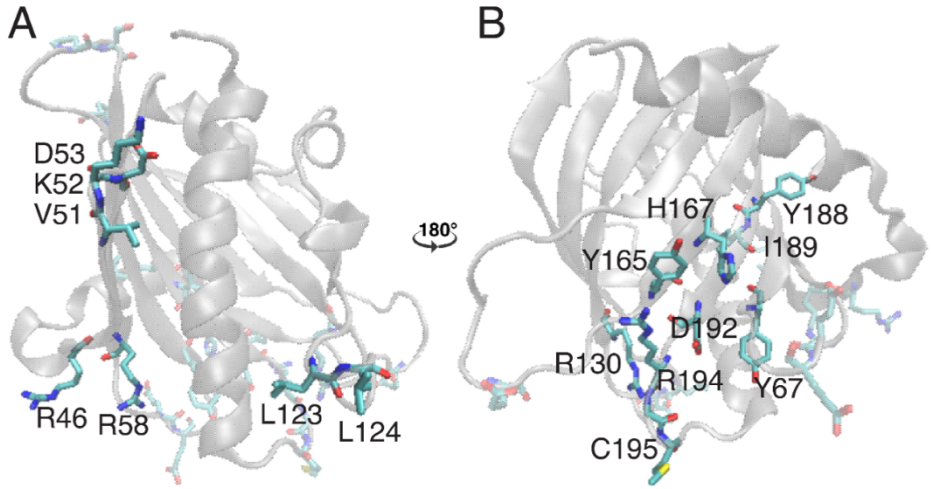

Figure 13. The molecular identified by the DNN to be salient in the differentiation the interaction of StarD4 with $PI(3,5)P_2$-containing membranes vs. $PI(4,5)P_2$-containing membranes. In (A,B) the most important residues for the differentiation are labeled in licorice on the transparent cartoon model of StarD4 viewed from the front **(A),** and from the back **(B)**. The important residues on β1&2 and Ω1 are highlighted non-transparently when viewed from the front, while the important residues on β3 and the β8-Ω4 corridor are highlighted non-transparently when viewed from the back of the protein.

Verification of the contributions of the salient residues to the PIP2-subtype prediction was carried out with a refined DNN based <u>only</u> on the coordinates of the salient residues in the motifs along the allosteric pathway as described above, and trained for the same recognition. Remarkably, this

specialized DNN, focusing exclusively on the salient residues, achieved an accuracy exceeding 96% (**Table 5**). While the information provided to the DNN by the data composed of only the salient residues is a proper subset of the original input, the results in Table 5 showed that the corresponding DNN managed to outperform the one based on the full protein. This suggests that the information derived from the salient residues is sufficient for accurate predictions and is also less affected by noise.

Table 5. Results of the classification by DNNs trained using only the coordinates of the salient residues, assessed on the test set samples from membrane embedded StarD4

		Prediction	
		$PI(3,5)P_2$	$PI(4,5)P_2$
Truth	$PI(3,5)P_2$	95.6%	4.4%
	$PI(4,5)P_2$	2.2%	97.8%

The recognition by the DNNs of these allosterically coupled motifs as pivotal discriminators, substantiates their distinctive dynamics in the response of StarD4 to the difference in PIP2 composition of the membranes. This observation supports our mechanistic conclusion that information about PIP2 subtype is effectively transmitted through the identified allosteric network that regulates the dynamics of the CHL ligand. Thus, the mechanistic inferences from the StarD4 conformation analysis, the PIP2 lipid binding mode analysis, and the analysis of the allosteric pathway come together in a mutually supportive manner. Together, they interpret in full structural detail the mode in which the differential recognition of PIP2 subtypes by the membrane embedded *holo*-StarD4 regulates its preference for release of CHL according to the PIP2-subtype prevalent at the targeted membrane.

The path of Cholesterol uptake by the membrane-embedded *apo*-StarD4

Repositioning of CHL by embedded apo-StarD4: Experimental evidence demonstrated that the $PI(4,5)P_2$-containing membrane is a more efficient cholesterol donor to StarD4 compared to membranes containing only $PI(3,5)P_2$, or PS (12). We investigated the process of cholesterol extraction by StarD4 in the same type of extensive MD simulations of *apo*-StarD4 that produced the mechanistic insights for *holo-StarD4*. For membrane systems containing CHL and one of the negatively charged lipids ($PI(4,5)P_2$, $PI(3,5)P_2$ or PS) the simulations were carried out for 547 microseconds (>166 microseconds per membrane composition) using multi-phase simulations: Phase 1 (70μs) to observe the membrane embedding of StarD4; Phase 2 (48μs) to reveal the conformational changes of membrane-embedded StarD4; and Phase 3 for >48μs to explore the cholesterol extraction process by StarD4 with an open Ω1-H4 gate (see **Fig. 14**, Phases 1-3). The StarD4-membrane interactions examined in phases 2&3 revealed a strong response of the membrane cholesterol to StarD4 that was especially pronounced in $PI(4,5)P_2$-containing membrane.

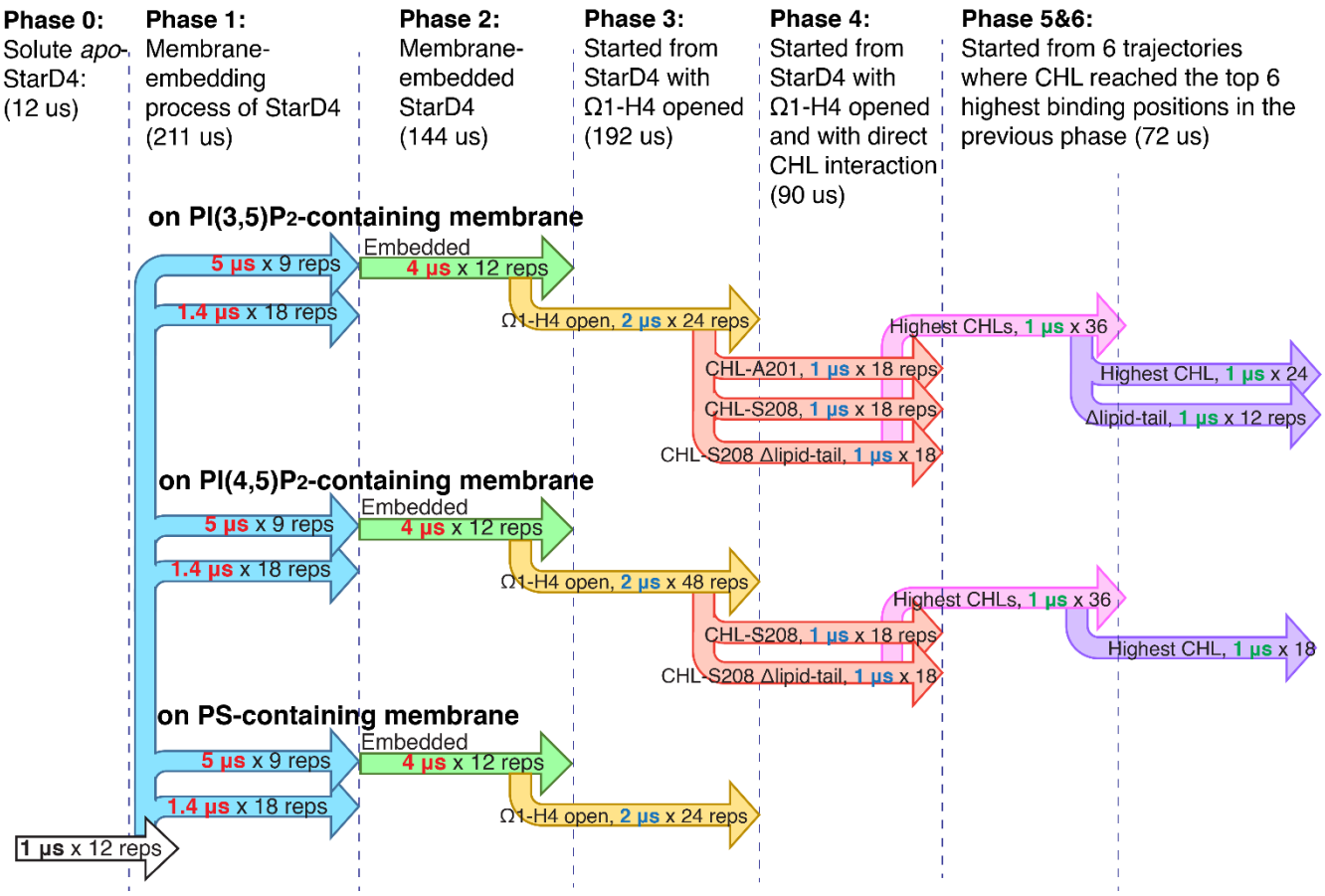

Figure 14. Schematic representation of the atomistic MD simulations of the *apo*-StarD4 membrane complex. The criteria for the choice of initial conformations, the simulation times, and the number of replicas for each set of simulation shown in the labels of the schematic plot.

At the concentration used, membrane-residing cholesterols are located predominantly beneath the membrane surface, with their oxygen closest to the lipid phosphate groups, and tend to have an inclination of ~20° from the upright position (**Fig. 15G-I**). We found that with the StarD4 embedded in the membrane, a subset of membrane cholesterols neighboring either the Ω1 or H4 motifs is repositioned to reach above the membrane or tilt beyond 40° (**Fig. 15D-F**). This repositioning is more pronounced for cholesterols located right at the Ω1-H4 gate with interactions involving both motifs (**Fig. 15A-C**). Moreover, this repositioning, and especially the rising CHL, is more prevalent within PI(4,5)P_2-containing membrane. It is less prominent in PI(3,5)P_2-containing membrane, and weakest in PS-containing membrane. Consistently, the dwell time of cholesterol at the Ω1-H4 gate follows the same trend (**Suppl. Fig. 10**). These observations align with experimental findings that the kinetics of cholesterol extraction by StarD4 is faster in PI(4,5)P_2-containing membrane compared to those containing PI(3,5)P_2 and PS (12), which suggests that this repositioning of CHL is a preparatory step in its process of extraction by StarD4.

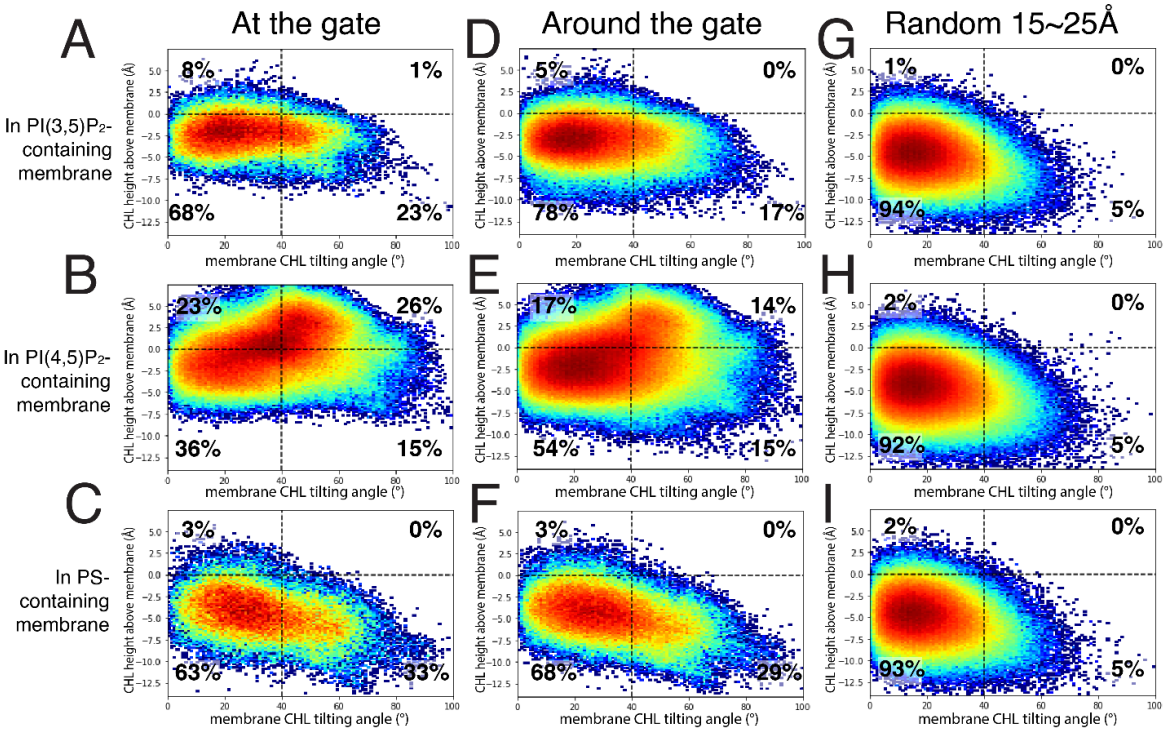

Figure 15. The probability distribution of the tilt and height positions of membrane-CHLs in relation to the membrane. X-axis: the tilting angle of the membrane-cholesterol; **Y-axis:** the height above the membrane surface. The positions are compared at three categories of positions relative to the embedded StarD4: when the membrane-CHL is at the H4-Ω1 gate **(A-C)**; when it is around the H4-Ω1 gate **(D-F)**; when it is far from the H4-Ω1 gate **(G-I)**. The negative lipid compositions of the membranes are shown in rows: **(A,D,G)** PI(3,5)P_2-containing membranes; **(B,E,H)** PI(4,5)P_2-containing membranes; **(C,F,I)** PS-containing membranes. The horizontal and vertical dashed lines are auxiliary lines that facilitate the comparison of the probability distributions by splitting the map to four regions, and the populations in each quadrant are indicated at the four corners of each figure. The figures are color-coded with densest regions in red and sparsest regions in blue. The detailed definitions of the variables and the criteria of the three categories are listed in the supplementary materials section 3.

<u>The modes of interaction of the membrane-residing CHL with the embedded StarD4:</u> The analysis of the simulation trajectories of *apo*-StarD4 in the three membrane systems showed three different sites of major interaction. These are indicated in **Fig. 16A-C** by the high-density regions in the 2D-space constructed for each membrane composition. The 2D-space is spanned by the distance of the CHL oxygen to StarD4 residues A201 (on the Y-axis), and to S208 (on the X-axis). The sites are indicated in the **Fig. 16A-C** panels, and the structural definition of the sites is illustrated in **Fig. 16D-F**. Site **1** represents A201-binding, Site **2** represents S208-binding, while Site **0** represents membrane-CHL molecules that are distant from both A201 and S208, and have no direct interaction with StarD4. Comparison of the distributions in **Figs. 16A, 16B, and 16C** shows that in membranes containing PI(3,5)P_2 and PS, CHL favors Sites 0&1 (**Fig. 16A,C**), while Site 2 is prominent in the PI(4,5)P_2-containing membrane but not in the PI(3,5)P_2-containing membrane (**Fig. 16A,B**). Notably, the CHL molecule is reaching above the membrane surface only at Site 2 (S208-binding). This leads us to propose that the S208-binding site likely serves as a crucial intermediate state in preparing for the subsequent uptake of the CHL into StarD4. In this scenario, the low occurrence of the S208-binding mode of cholesterol in the PI(3,5)P_2-containing membrane, and even more so in the PS-containing membrane, relates directly to the observed slower kinetics of CHL uptake in these membranes.

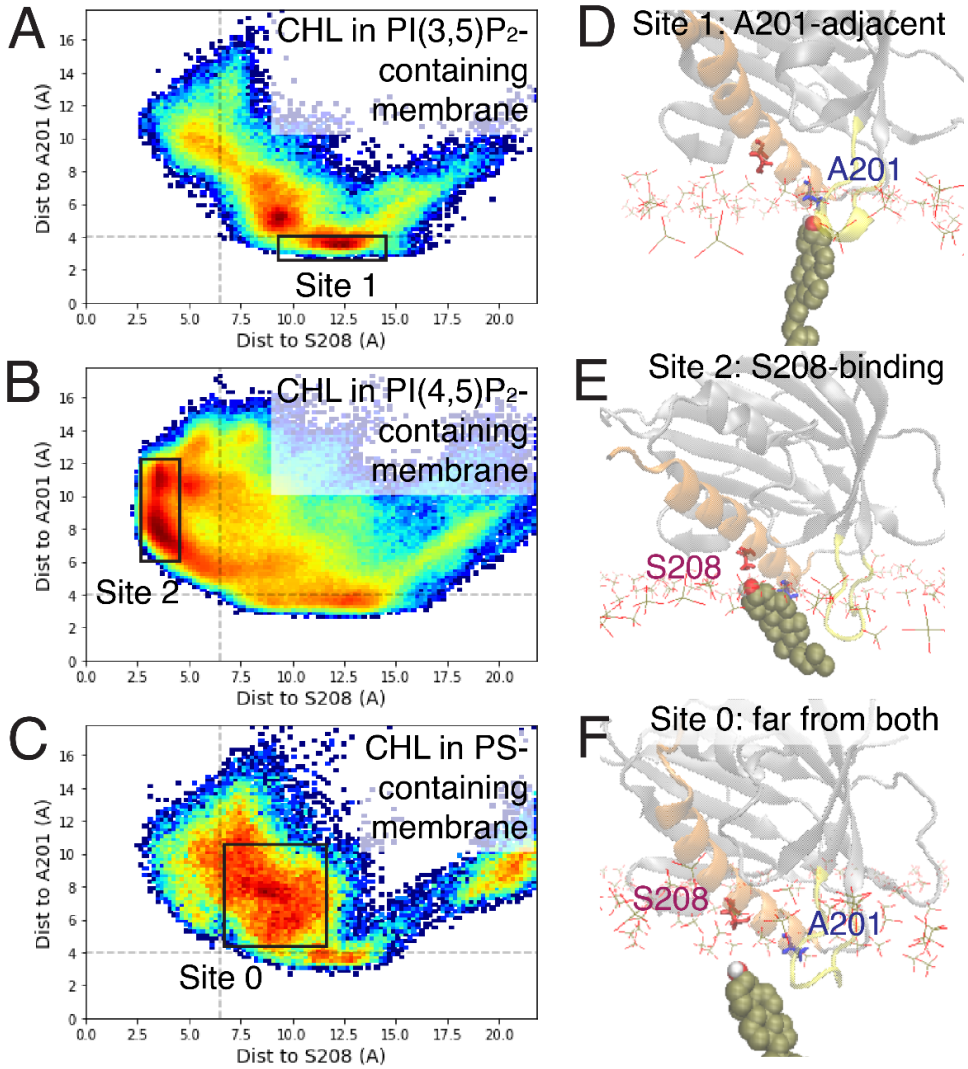

Figure 16. The interactions of membrane-residing cholesterols with membrane-embedded StarD4. (A-C) The probability distribution of the position of membrane-CHLs in relation to the StarD4 (X-axis: the distance to S208; Y-axis: the distance to A201), compared between the three types of membranes: **(A)** in PI(3,5)P_2-containing membrane; **(B)** in PI(4,5)P_2-containing membrane; **(C)** in PS-containing membrane. The figures are color-coded with dense regions in red and sparse regions in blue. The rectangles mark the metastable sites of CHL binding. **(D-F)** The representative conformations of *apo*-StarD4 complexes with membrane-CHL: Site 1, adjacent to A201 **(D)**; Site 2, binding to S208 **(E)**; Site 0, far from both A201 and S208 **(F)**. The protein is rendered in transparent cartoon in gray, with H4 in orange and Ω1 loop in yellow. The membrane cholesterol adjacent to StarD4 is shown in VDW. The cholesterol binding site residues are shown in licorice: S208 in red, A201 in blue. The membrane surface is denoted by the lipid phosphate headgroups shown in lines rendering.

<u>The pathway of Cholesterol uptake:</u> The uptake of CHL by *apo*-StarD4 from PI(4,5)P_2-containing and PI(3,5)P_2-containing membranes was sampled in Phase 4 MD simulations (**Fig. 14)** with an adaptive sampling protocol starting from the protein bound to a membrane cholesterol molecule at Site 2 (the S208). With an open Ω1-H4 gate, adaptive sampling simulations were initiated from both the unaltered adaptive sampling seeds and the modified ones with "pocket-penetrating" lipid tails removed (**Suppl. Fig. 12**). Additionally, the *apo*-StarD4 with cholesterol at the A201-binding on PI(3,5)P_2-containing membrane was also included as a set of adaptive sampling seeds. As shown in (**Fig. 14**, phase 4), this simulation setup results in a total of 90 replica runs, each of 1 microsecond in length. In the next adaptive step, 6 trajectories that had reached the highest position within the Ω1-H4 gate during the

previous phase were selected from each membrane composition and used as initial points for new, 72 replica runs of 1 microsecond each. This iterative process was repeated across two phases to expand the sampling of CHL along its extraction pathway (**Fig. 14**, phases 5,6). As the cholesterol molecule moved up progressively from the gate to the W171, Y117 binding site, we represent these dynamics by mapping the interaction mode of CHL along its uptake pathway with its location described by the CVs listed in **Fig. 17A** (see further details in Supplementary Methods, section 3).

The CVs describing the upward movement of the CHL were used as parameters for in the tICA dimensionality reduction transformation of the CHL dynamics data as described above (e.g., see Fig.7) and detailed in Methods. The first 2 tICA vectors describe 92% of the total dynamics of the cholesterol along the uptake pathway. The 2D map of the tICA space spanned by these two vectors (**Fig. 17B**) displays multiple metastable states. These include, in addition to Site **1** (A201-binding) and Site **2** (S208-binding) shown in **Fig. 17C**, three stepwise binding states inside the hydrophobic pocket: Site **3**, which represents T119-binding (**Fig. 17D,E**); Site **4** representing T191-binding (**Fig. 17D,F**); and Site 5 representing W171,Y117-binding (**Fig. 17D,G**). Site **5** marks the end of the uptake pathway as the W171,Y117-binding mode is commonly found in *holo*-StarD4. The identification of these sites as metastable states of the CHL dynamics is confirmed by their kinetics similarity, and is characterized by their CV distributions detailed in **Suppl. Fig. 13**.

The key comparative finding from this analysis is that the binding sites inside the hydrophobic pocket were observed exclusively for the PI(4,5)P_2-containing membrane system (**Fig. 17D**), while the *apo*-StarD4 embedded on the PI(3,5)P_2-containing membrane managed only rarely to extract a membrane CHL even up to the lowest binding site (Site 3 or 4) inside the gate (**Fig. 17C**). This difference agrees well with the experimental findings (12).

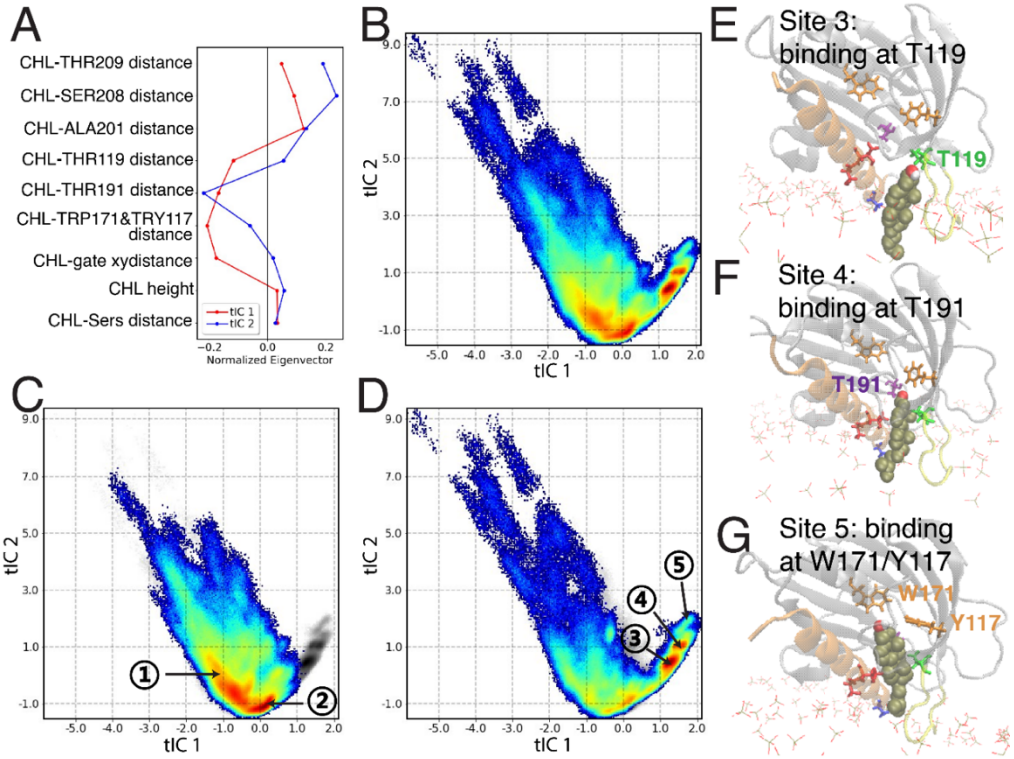

Figure 17. The CHL extraction pathway. (A-D) Extraction dynamics mapped on a 2D tICA space. **(A)** Contributions of individual CVs to the first 2 dimension of the tICA space they span. **(B)** Population density map of the position of the extracted CHL in StarD4. **(C)** Same, on the PI(3,5)P_2-containing membrane; **(D)** On the PI(4,5)P_2-containing membrane. Dense regions are in red and sparse regions in blue. Numbers mark the metastable CHL binding sites in **(C)** and **(D)**. In **(E-G)** the representative conformations of metastable CHL binding sites in the StarD4 hydrophobic pocket are rendered as in **Fig. 16**. **(E)** Site 3: T119-binding; **(F)** Site 4: T191-binding; **(G)** Site 5: W171/Y117-binding. The CHL binding sites are shown in licorice: S208 T209 in red, A201 in blue, T119 in green, T191 in purple, W171 Y117 in orange.

Comparing the membrane interactions and conformations of the *apo*- and *holo*-StarD4 in their respective functional mechanisms

As we has shown here that the preference for CHL delivery by *holo*-StarD4 to a PI(3,5)P_2-containing membrane was associated with a distinct orientation of the protein with respect to the membrane, we reasoned that the extraction of CHL from the membrane may also be associated with a preferred orientation of the *apo* protein. As the membrane embedded *apo*-StarD4 is more likely to pick up the CHL from a PI(4,5)P_2-containing membrane than from a PI(3,5)P_2-containing membrane, we investigated the conformation differences of *apo*-StarD4 that underlie this preference in the corresponding MD simulation trajectories. We found that in contrast to the CHL release process where a large protein orientation of >40° towards the right side (the side of hydrophobic pocket) is favored, the CHL uptake process on PI(4,5)P_2-containing membrane involves a distribution of orientation centered at ~10°, with only minor leaning towards the right side (**Fig. 4E**). This orientation is less visited by *apo*-StarD4 on PI(3,5)P_2-containing membrane, or by *holo*-StarD4 (**Fig. 4**).

As shown in the comparison of the *apo*-StarD4 dynamics on membranes with different PIP_2 subtype compositions and different modes of membrane-CHL binding presented in **Fig. 18A** the interaction with the N-terminal of H4 (H4-Nt) is absent in *apo*-StarD4 on PI(4,5)P_2-containing membrane (CV "β1-H4 interaction" >10Å). This is likely due to the observation that PI(4,5)P_2 phospholipid binding to *apo*-StarD4 displays distinct features, with a strong preference for binding mode 5 in PI(4,5)P_2-*apo*-StarD4 interactions. This is shown by the comparative occurrence rates of binding mode probabilities of PIP2 phospholipids to StarD4 listed in **Table 1,** above. Thus, in mode 5, the phosphate groups of the lipid are inserted into the upper-left corner on the basic residue patch (**Fig. 10A,B**). This arrangement is likely to hinder β1 from shifting downwards and forming the interaction with the N-terminal of H4 (H4-Nt). In contrast, the *apo*-StarD4 on PI(3,5)P_2-containing membrane, with no PIP2 binding in mode 5, exhibits a growing portion of closer β1-H4 interaction as the simulations progress (**Fig. 18A**). Thus, the effect of PIP2 binding mode 5 which appears to prevent the β1-H4 interaction, contrasts with the impact of binding mode 7 of PIP2, which is associated with the β1-H4 interaction. We had shown (21) that the movement of H4 towards β1 is correlated with the movement of Ω4-loop towards β2&3 away from β8 & Ω1-loop, which leads to the opening of the β8-Ω4 back corridor. In PI(4,5)P_2-membrane-embedded *apo*-StarD4, the absence of β1-H4 interaction also eliminates the opening of β8-Ω4 back corridor (**Fig. 18B**). With the back corridor β8-Ω4 closed on PI(4,5)P_2-containing membrane, we reasoned that the conformation provides a hydrophobic environment that stabilizes the incoming CHL that is being extracted from the PI(4,5)P_2-containing membrane.

In contrast, the opened back corridor apo-StarD4 on PI(3,5)P_2-containing membrane hinders the uptake. Indeed, we observed an incipient cholesterol uptake near THR191 only in 4% of cases where the structural characteristics include a strong β1-H4 interaction (**Fig. 18C**) and a widened back corridor between Ω1 and Ω4 (**Fig. 18D**). Notably, the conformational state of StarD4 in these few occurrences is akin to the pre-release state of *holo*-StarD4. This is shown in **Fig. 18E** by the fact that the probability distribution of the conformation of the incipient CHL-uptake conformation of *apo*-StarD4 on PI(3,5)P_2-containing membranes projected on the tICA space defined for the *holo*-StarD4, lands close to the pre-release state (see **Fig. 7** for details of the tICA map). In contrast, the full CHL uptake process from PI(4,5)P_2-containing membrane is seen to land in a highly populated PI(4,5)P_2-specific state (**Fig. 18F**). This shows that the *apo*-StarD4 on PI(3,5)P_2 adopted a conformation favoring

CHL release, not uptake, whereas the *apo*-StarD4 on PI(4,5)P_2 adopted a conformation favoring CHL entrance and binding in the canonical site. These findings are in agreement with experimentally observed difference of CHL uptake kinetics from PI(3,5)P_2- containing vs PI(4,5)P_2-containing membrane, and provide a mechanistic explanation.

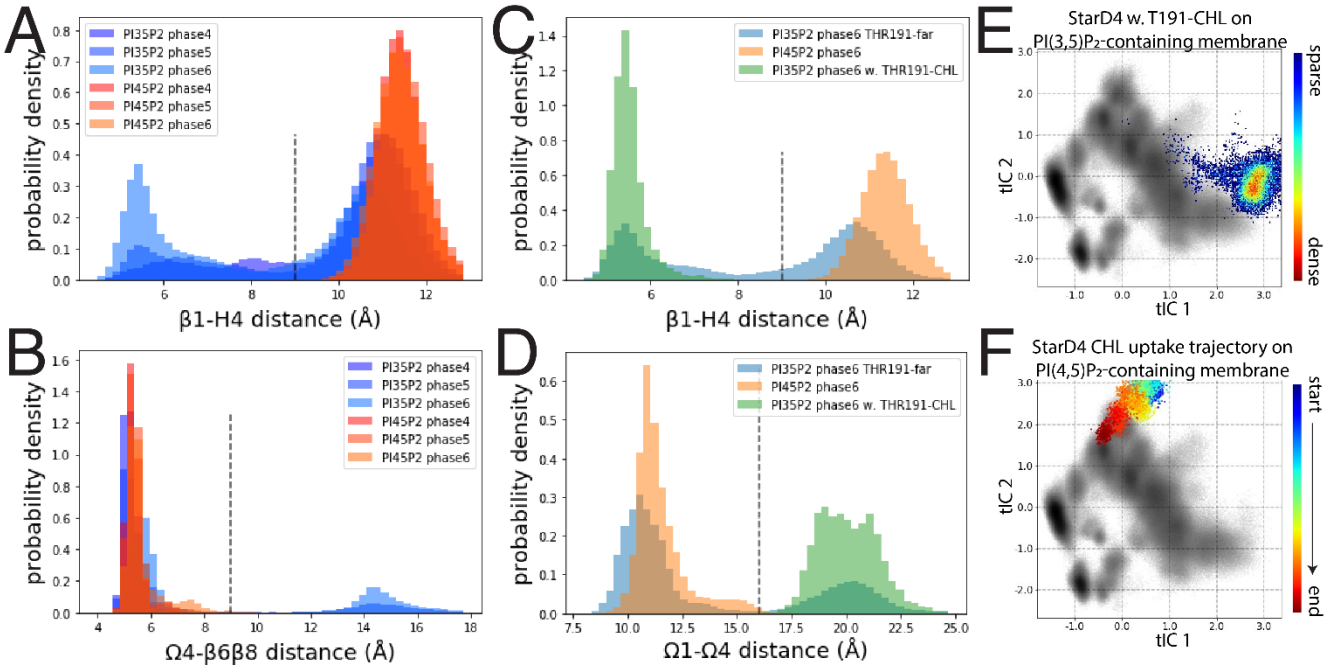

Figure 18. Comparison of the *apo*-StarD4 dynamics on membranes with different PIP_2 subtype compositions and different modes of membrane-CHL binding. (A,B) The probability density histogram compared between the *apo*-StarD4 on PI(3,5)P_2-containing membrane (red-orange scheme) and on the PI(4,5)P_2-containing membrane (blue-purple scheme) during different phases of the adaptive sampling: **(A)** The interaction between β1 & H4; **(B)** The opening of the β8-Ω4 back corridor. The dashed line in Panels (A-D) indicate the state transition. **(C,D)** Color coded probability density histograms comparing StarD4 on PI(3,5)P_2-containing membrane with a CHL close to the T191 binding site (green) and without any CHL close to the T191 binding site (blue). StarD4 on PI(4,5)P_2-containing membranes is in orange. **(C)** The β1-H4 distance; **(D)** The Ω1-Ω4 distance compared. **(E)** The probability distribution of the conformation of the CHL-T191-adjacent *apo*-StarD4 obtained from the simulations on PI(3,5)P_2-containing membranes, projected on the tICA space defined for the *holo*-StarD4, with the background represents the combined population density of *holo*-StarD4 in gray scale, as in Fig 7. The population density of *apo*-StarD4 is color-coded with dense regions in red and sparse regions in blue. **(F)** The time-evolution of the trajectory that successfully extracted a CHL from PI(4,5)P_2-containing membrane, projected on the tICA space defined for the *holo*-StarD4. The evolution of the conformation encoded in the trajectory is color-coded from the starting point (in blue) to the end point (in red).

In sum, we showed that: ***1.*** Different subtypes of PIP2 membrane components prefer different binding modes to StarD4; ***2.*** StarD4 adopts different conformation in response to different PIP2 binding modes; ***3.*** The preparation for the release of CHL from *holo*-StarD4 and the stability of the extraction of membrane CHL by *apo*-StarD4 prefer specific and different conformations of the protein. Collectively, these results and the quantitative findings from the NbIT and DNN analyses provide compelling evidence for an allosteric mechanism in which StarD4 senses the subtype of PIP2 through its interaction modes at the upper basic residue patch (*upper-group*), transmit the information through an allosteric network involving β1, β2, β9, Ω4 and H4 that enhances or discourages the conformational

changes leading to the pre-release state, and thereby establish the preferred direction of cargo translocation within StarD4.

Discussion

Experimental evidence has demonstrated that sterol traffic by StarD4 – a cytosolic protein in the family of STARD proteins involved in non-vesicular sterol transport between organelles – is modulated by its interaction with anionic lipids in the target membrane, particularly PI(4,5)P_2 (9,12,31). As such traffic is essential for subcellular cholesterol homeostasis (1-7), we considered it essential to discover the nature of the molecular the mechanisms of enabling recognition of such anionic phospholipids by StarD4, and how it guides the functional mechanism. We therefore carried out extensive MD simulations mirroring experimental conditions to explore the processes of CHL uptake and release by the StarD4 protein interacting with membranes containing each of the anionic lipids examined in these experiments.

Many studies of non-vesicular sterol transport and sterol transfer proteins (14-16) have identified a sterol binding pocket in START domains and considered the orientation of the ligand to place the hydroxyl group at the base of the hydrophobic pocket (in StarD4, interacting with S136 and S147, **Fig. 1A**), but no structures of any sterol-bound START domain have been resolved. Recent structural analyses (17) suggested that the size of the hydrophobic pocket of StarD4 is marginally insufficient to accommodate a cholesterol molecule, underscoring the necessity for a conformational change. Indeed, our simulation found a series of conformational changes of StarD4 in the complex with cholesterol, and showed (**Fig. 7D)** that the opening of Ω1-H4 front gate enlarges the hydrophobic pocket and facilitates the translocation of CHL from the Ser-binding site to the Trp-binding site. It is noteworthy that this Trp-binding-cholesterol-StarD4 complex we described recently (12,21,22) closely resembles the reported structures of cholesterol-binding StARkin domains of LAM proteins (**Suppl. Fig. 14**) (18-20). This similarity is especially relevant because previous *in silico* studies of the Lam4S2 protein have demonstrated the release of a sterol ligand from a similar complex (32). Similarly, we found from our MD simulations of the *holo*-StarD4 in an aqueous environment – before it embeds in the membrane (21) – that CHL can continue to move further down from the Trp-binding site to the farthest end of the hydrophobic cavity to position itself close to the entrance of the pocket, and we suggested that this represents a preparatory step before its release (21). The release mechanism from the *holo*-StarD4 embedded in the membrane described in the present work supports this prediction of a preparatory state.

The release mechanism from such a preparatory state is described here for *holo*-StarD4 embedded in a variety of membrane systems mimicking those investigated experimentally (9,12). The mechanism is driven by the allosteric communication that connects the modes of interaction of StarD4 with these membranes to the conformational changes that enable the translocation of the CHL in a manner dependent on the nature of the anionic lipids. These findings agree with the experimental results that show clear differences between the functional behaviors of StarD4 when embedded in the different PIP2 and in PS phospholipids. Because the modes of interaction with different anionic lipids are different, the activation of allosteric channels (**Fig. 12A**) leads to different conformational changes, as described in **Fig. 11**.

From our parallel studies of *holo*- and *apo*-StarD4, we identified the molecular mechanisms that determine the preferred actions of this protein (CHL uptake vs its release) in agreement with the experimentally observed CHL traffic differences in the different systems (12). Thus, we identified the structural motifs and interactions that determine the conformations and dynamics that lead to a kinetically preferred CHL release into membranes that contain PI(3,5)P_2 lipids, and those favoring CHL uptake from membranes containing PI(4,5)P_2. Quantitative analysis of allosteric communication among the key structural motifs illuminated the allosteric pathways connecting the modes of StarD4-membrane interactions with the dynamics of CHL and its binding sites. From such structure-based mechanistic details of the CHL release from *holo*-StarD4 embedded in PI(3,5)P_2-containing membranes emerged the definition of a pre-release state of StarD4 where the Ω1-H4 gate and the β8- Ω4 corridor are open so that the ligand in the hydrophobic pocket is exposed to the membrane surface and destabilized by water molecules (**Fig. 6**). The CHL release process on the PI(3,5)P_2 containing membranes starts from this pre-release state which is not visited in the uptake process that is preferred on PI(4,5)P_2 containing membranes. The formation of the pre-release conformational state is characterized by the destabilization of the Ω1-H4 gate, accompanied by a movement of the β9/Ω4/H4 motif towards the β1-β3 motif which, in turn produces a shift in the β1-H4 interaction and the distancing of Ω4 from β6&β8 (**Fig. 7D** State 1). The β1, Ω4 and H4 are membrane interacting motifs whose dynamics are shown in Fig. 11&18 to be related to the interaction with anionic lipids. Their conformational changes induced water penetration and decrease the hydrophobic interaction between CHL and the hydrophobic pocket of StarD4, thereby creating conditions conducive to cargo release.

Notably, we observed CHL release from an additional state (**Fig. 7D** state 7) in which both the β8-Ω4 back corridor and the Ω1-H4 gate are open, but the gate is held open by a penetrating lipid tail instead of the interaction between β1 and H4. While this confirms the essential structural determinants for CHL release regarding the openings, we found that in this alternative state energy barrier of the process of release from state 7 is much higher because the Ω1-H4 gate is not freely opened (**Fig. 8**). The energy profile reaches its peak at z-dist=15Å where the bulky hydrocarbon rings are passing through the gate, and the energy barrier suggests a 10x slower kinetics compared to the release from state 1.

These comparisons of the structural characteristics of the CHL release mechanisms underscore the importance of water penetration through the β8-Ω4 back corridor as a prerequisite for CHL release, while emphasizing the facilitating role of the β1-H4 interaction in this process. This underscores the mechanistic contrast to the process of CHL uptake which is observed exclusively for a StarD4 conformation lacking the shifted β1-H4 interaction and the opening of the β8-Ω4 back corridor (**Fig. 18**). Together, the findings substantiate the critical role of conformational changes of the membrane binding motifs of StarD4 in regulating CHL release and uptake, which rely on different interactions with the kinds of anionic lipids in the membrane. Our detailed analysis revealed how these different modes of interaction lead to the difference in the preferred direction of cholesterol translocation by StarD4.

Thus, our analysis of the MD trajectories revealed an allosteric network that links the modes of interaction of the membrane binding motifs to the dynamics of CHL release from the cholesterol binding pocket in StarD4. The difference in the modes of interaction of StarD4 with different types of anionic lipids relates to the larger distance between the phosphate groups in PI(3,5)P_2 than in PI(4,5)P_2. Specifically, the larger distance in PI(3,5)P_2 favors the "cross-H4" binding mode that spans β1, H4 and

Hlp3. In contrast, when the membrane contains the PI(4,5)P_2 whose phosphates groups are adjacent the "corner" binding mode with K52, R218, K219, R222 is preferred for StarD4-membrane interactions (**Fig. 10, Table 1**). Previous research has highlighted subtype preferences in the binding of PIP2 lipids to R46 and R58 at the N-terminal of β1 and the C-terminal of β2 (12). The different modes of β1 recognition by the PIP2 subtypes changes the dynamics in the allosteric network that regulates the actions of StarD4 in CHL traffic.

This conclusion emerged from mutually consistent findings resulting from the application of the various methods of trajectory analysis: (1) examination of the dynamics by comparing the distribution of collective variables in the presence and absence of lipid at key binding modes (**Fig. 11,18**); (2) quantifying the *mutual information* between the allosteric network and the anionic lipid binding site using NbIT (**Fig. 12 and Table 2**); (3) identifying the key motifs of the allosteric network that contribute in the PIP2-dependent conformational changes using DNN (**Fig. 13**). Together, these findings affirm the crucial role of basic residues of β1 in the binding and subtype recognition of membrane anionic lipids (9,31).

Regarding the findings from experiments with membrane in which the anionic lipid is phosphatidylserine (PS) (12), we also found that its smaller negative charge leads to a weaker adsorption energy of StarD4. Using the Mean-Field Model we calculated the adsorption energy when StarD4 approached a 10% PS-containing membrane the adsorption energy, and found that it is only -1.1 kBT compared to the -3.8 kBT energy observed when approaching a 10% PIP2-containing membrane. Correspondingly, the average number of PS lipids binding simultaneously at each of the three basic residue groups (**Suppl. Table 5**) is significantly lower than for the PIP2 lipids, the calculated anionic lipid dwell time is shorter for PS than for the PIP2 lipids (**Suppl. Fig. 11**), and the orientation of StarD4 on PS-containing membranes is less stable than on the PIP2-containing membranes (**Fig. 4**). The broader range of orientations of StarD4 on PS-containing membranes relates to a weaker repositioning effect of the membrane cholesterol (**Fig. 15,16**). We note that the differences we report here and in previous publications (**Fig. 4** and ref (12)) between the orientations of StarD4 on PIP2- and PS-containing membranes, is reproduced in recently published MD simulations (22) where the StarD4 on the PS-containing membrane was also shown to adopt a wider range of orientations in different trajectories and to have a more superficial anchoring on the membrane (12,22).

Our mechanistic analysis has brought to light the central role of the β1 strand in sensing the type of anionic lipid enriched in the membrane, and in transmitting the information to the CHL binding site through its dynamic interactions with H4, Ω4, and β9. Notably, both the β1 and β2 motifs are absent in the StARkin domain of LAM proteins despite the general conservation of various structural motifs among START and StARkin domains (14-16). This difference is, however, consistent with the difference in the functional mechanisms of the protein between StarD4 and the LAM StARkins which are constrained to specific membrane sites via their membrane anchors. The structural difference represents a functional adaptation that provides the soluble StarD4 family proteins with a membrane-target sensing mechanism that regulates their sterol trafficking directions.

We note that the comparative experiments investigating the sterol trafficking properties of StarD4 utilized the mouse StarD4 (mStarD4) protein (9,12,33) and thus our simulations were done with this same system. The membrane-interaction interface is highly conserved between mStarD4 and the human hStarD4, including the basic residues involved in the interactions with anionic phospholipids. Thus, most residues are identical, except for R97 and S215 in mStarD4 which correspond to S82 and G199 in the human, and R222 in mStarD4 which extends beyond the human

StarD4 sequence (**Fig. 9A**). The S215A mutation has been reported to lower the kinetics of mStarD4 on PI(3,5)P_2-containing membrane to a level that is significantly below than on PS, while the acceleration of the kinetics by the PI(4,5)P_2-containing membrane relative to PS persists in the S215A mutant (12). Our findings are aligned with the experimental results, as we found the S215A mutation to block the favorite binding mode of PI(3,5)P_2, which underscores the functional importance of the "cross-H4" binding mode we described and its subtype preference. On the other hand, the "corner" binding mode we found to be favored by PI(4,5)P_2 is centered on the interaction at R218. In agreement with this finding, the R218A mutation has been reported (12) to slow the kinetics of StarD4 on PI(4,5)P_2-containing membranes to the level measured on PS-containing membrane. Similarly congruent are the findings that mutations of K52 and K219 – which we showed to have essential roles in both the PI(3,5)P_2 "cross-H4" binding mode and the "corner" binding mode favored by PI(4,5)P_2 – impede StarD4 cholesterol transportation on anionic membranes (9). Together, these findings substantiate the insights we obtained about the mechanistic role of the specific binding motifs for each PIP2 subtype and their components in modulating the cholesterol transport dynamics of StarD4.

Naturally occurring mutations of StarD4 have been documented in the context of dysfunctions associated with various diseases. While expression rates of StarD4 have been related to cancer (34,35), it remained unclear how mutations of the basic residues may contribute. For example, the naturally occurring R202Q mutation in human StarD4 (corresponding to a mutation of R218 in the mouse StarD4 discussed here (**Fig. 10A,B**)) has been associated with 13 malignant melanoma and breast carcinoma samples in five independent studies (36-41). Similarly, the G199C mutation (corresponding to S215 in mouse StarD4 (**Fig. 10C,D**)) has also been reported in a case of skin melanoma (42). As our results underscore the mechanistic role of subtype specific PIP2 binding modes in the modulation of cholesterol transport function of StarD4, the dysregulation caused by mutations in the preferred PIP2 binding sites can explain a role in disease.

In conclusion, the PIP2-subtype-dependent regulatory mechanism of the StarD4 CHL uptake and delivery functions that emerged in this study, leverages differences in the dynamic response of the protein (both the *apo* and *holo* forms) to interactions with different anionic lipids. As described, these differences are propagated through the allosteric channels to the CHL binding site to modulate the stability of the cholesterol cargo, thereby regulating the conformations and the orientations of the protein on the target membranes that determine the steps and directions of the cholesterol traffic in cellular processes.

MATERIALS AND METHODS

Modeling the initial structures

The cholesterol-StarD4 complex: The structure of *apo*-StarD4 was derived from the X-ray structure (PDB ID: 1jss) where residues 24 to 222 are resolved (8). To complete the structure, the missing residues K223 and A224 are added using Modeler 9.23 software, resulting in the conformation of StarD4 segment spanning residues 22-224, with acetylation on the N-terminus and carboxylation on the C-terminus (43). The initial structure of cholesterol bound (*holo*) StarD4 was obtained by docking the cholesterol into the hydrophobic pocket of *apo*-StarD4 using the Schrodinger Induced Fit Docking protocol (44-46). Both the *apo*- and *holo*-StarD4 constructs were then solvated in 0.15M K^+Cl^- ionic aqueous solution and equilibrated using the NAMD simulation platform version 2.12 (47).

Assembly of the initial lipid membrane: Explicit all-atom lipid membrane models were constructed with 400 lipids in symmetric bilayers composed of a 44:23:23:10 mixture of POPC, POPE, cholesterol, and anionic lipids, (either POPS, POPI(4,5)P_2, or POPI(3,5)P_2), which is consistent with the experimental conditions that revealed the PIP2-subtype-dependent StarD4 kinetics modulations (21). The initial structure of membrane bilayers was assembled using the CHARMM-GUI webserver (48). The full equilibration of the membrane patch was done with StarD4 restrained at a 2Å distance adjacent to the membrane as described in the section below.

Construction of StarD4-membrane complexes.

A three-stage simulation protocol was employed to obtain the StarD4-membrane complexes:

Stage 1. *Equilibration of StarD4 in aqueous solution.* The conformational space of the StarD4 constructs prior to its interaction with membrane was explored for *apo*- and *holo*-StarD4 in aqueous environments in all atom MD simulations with OpenMM software (49). The protein was solvated in a 70×70×70 $Å^3$ water box ionized with 0.15M K^+Cl^-, resulting in a system containing approximately 32,500 atoms. The OpenMM simulations were conducted in NPT ensemble (T=310K, P=1 atm) using a 4fs integration time-step and a Monte Carlo Barostat. As described recently (21), 12 statistically independent replicas were generated after equilibration for each system and simulated for a duration of 1μs/each for the *apo*-StarD4 system, and for 33.6 μs/each for the *holo*-StarD4 system. The analyses and results were previously reported (21). Following the simulations, clustering analysis was performed to identify distinct conformations, resulting in 3 representative conformations for *apo*-StarD4, and 9 snapshots in 3 representative conformation states for *holo*-StarD4. These were used to build the atomistic models for the simulations of StarD4-membrane complexes.

Stage 2. *Distribution of the membrane anionic lipids in the electrostatic field of approaching StarD4 molecules.* To speed up the convergence of the membrane reorganization in the electrostatic field of the approaching StarD4, we used the Mean-field model (MFM) approach developed in the lab to assess the impact of long-range electrostatic interactions between StarD4 and membrane (12,50-53). MFM sampling accelerates the exploration under important degrees of freedom, particularly electrostatics and lipid mixing, which play crucial roles in the long-range interaction to establish an optimized state for the StarD4-membrane combination. This allowed us to investigate the effect of the mutual interaction on the orientation of StarD4 as it approaches the negatively charged membranes, as well as the corresponding rearrangement of anionic lipids in the membrane in response to the presence of StarD4.

In the MFM approach, StarD4 is represented at the detailed 3D atomistic level, while the membrane is simplified as a two-dimensional smooth charged surface that represents the behavior of lipid polar headgroups. The positioning of StarD4 is systematically explored in various orientations while ensuring that all atoms are at least 2 Å away from the membrane surface. As described (50,53), self-consistent minimization is employed to sample the rearrangement of membrane lipid position corresponding to the StarD4s by optimizing the governing mean-field-based free energy function (F), which comprises the *electrostatic component* (F_{el}), the *translational entropy* of mobile ions (F_{IM}) that depends on the local lipid component densities $\varphi_{(x,y)}$, and the *lipid mixing entropy* (F_{lip}) that depends on the mobile ion concentrations c^- and c^+. The minimization of this free energy function is carried

out under the influence of the electrostatic field generated by the StarD4 protein, which yields a correspondingly optimized lipid rearrangement that is quantified from the local lipid density. The absorption energy of StarD4 into the membrane is determined by comparing the free energy of the StarD4-complex to the sum of the free energies of StarD4 and of the membrane considered separately. The best orientation of the membrane-approaching StarD4 is determined as the position that yields the strongest absorption energy among the geometries sampled.

Here, the calculations were carried out in a $(256\ Å)^3$ cubic space containing a 0.15M ionic solution of monovalent counterions, corresponding to λ=8.09Å Debye length, under the temperature of 310K.

Stage 3. *Equilibration of the mutual StarD4/membrane interaction at 2Å above the membrane*. For the equilibration of the mutually adapted system from Stage 2, the system was solvated in 0.15M K^+Cl^- ionic solution, resulting in a size of ~138,000 atoms. The equilibration was carried out in 3 steps with NAMD version 2.12 at temperature of 310K in NPT ensemble using Monte Carlo Membrane Barostat, with isotropic XY ratio as established previously (12). The 3 consecutive steps start with restraining forces of 1 kcal/mol·$Å^2$ applied on the protein backbone atoms, the heavy atoms of the docked cholesterol, and the heavy atoms of the membrane lipids heads. After the first 0.5 ns simulations the restraints are decreased gradually, first to 0.5 kcal/mol·$Å^2$, and then to 0.1 kcal/mol·$Å^2$ in the subsequent runs of 0.5ns/each.

Following these 3 equilibration steps, the subsequent MD simulations were carried out with the OpenMM software (49), using 4 fs integration time-step, in NPT ensemble (T=310K, P=1 atm) under Monte Carlo Membrane Barostat. These started with a 500 ns equilibration phase in which the relative height of StarD4 (z-distance to the membrane surface) and the orientation of StarD4 (the z-distance differences between the left-side-half and right-side-half of StarD4, and between the top-side-half and the bottom-side-half the StarD4) are strongly restrained with a force constant of 20 kcal/mol·$Å^2$. This constraint ensured proximity and consistent orientation of StarD4 with the membrane, facilitating the equilibration of membrane lipid rearrangement without direct contact. After the initial 500 nanoseconds, all these constrains are released, allowing StarD4 to establish direct contact with the membrane, and initiating the production run of the StarD4-membrane systems.

Production run simulations of the membrane embedded *holo*-StarD4

Production runs of the membrane embedded *holo*-StarD4 were carried out with the OpenMM software with the same parameter and conditions as above (4 fs integration time-step, NPT ensemble (T=310K, P=1 atm), Monte Carlo Membrane Barostat). The 3-phase design of the entire unbiased simulation protocol of the membrane embedded *holo*-StarD4 is shown in **Fig. 2**, with the following aims:

Phase 1 of 0.25 milliseconds: To establish an equilibrated system of the StarD4 embedded in the membrane. Nine conformations from three representative states in phase 0 (*holo*-StarD4 in aqueous solution) were brought to the vicinity of the membranes using the *preparation protocol* described above. Parallel simulations were conducted on three different types of membranes, utilizing 141 statistically independent replicas, amounting to a total simulation time of 249 microseconds. StarD4 was considered embedded in membrane when the Ω1-H4 gate establishes substantial contact with the membrane: during the phase 1 simulation, the relative position of the Ω1-loop to the membrane surface converged to 2 stable states that contain >95% of the samples, either with no substantial contact with membrane (contact area <70$Å^2$), or with multiple hydrophobic residues inserted into the membrane

and the contact surface area >300Å^2. Thus, we considered the Ω1-loop embedded in membrane when its membrane contact surface area is >200 Å^2, and similarly, the β9-H4 motif is considered embedded when its membrane contact surface area is >150 Å^2. StarD4 was considered embedded in membrane when both the Ω1 loop and the β9-H4 motif are membrane embedded simultaneously.

Phase 2 of 0.34 milliseconds: To explore StarD4 conformational changes and CHL translocation after membrane embedding. For each membrane type, 4 trajectories of embedded StarD4 from Phase 1 were replicated ×6 and continued in this Phase to expedite exploration of the cholesterol release pathway. These involved 72 independent runs (24/each for each membrane type), totaling 344 microseconds of cumulative simulation time.

Phase 3 of 0.73 milliseconds: To explore the cholesterol release pathways. A total of 48 conformations listed in the Table below were selected from different states described in the text, which included the pre-release state, highly populated states, and states of PIP2-subtype-specfic binding (as illustrated in **Fig. 2** and **Fig. 7A,B**):

#initial seeds	From the pre-release state	From populated states	From PIP2-subtype-specfic states
w. PI(3,5)P_2	8	5	11
w. PI(4,5)P_2	8	8	4
w. PS	4	-	-

In addition, 4 conformations referred to in the text as "artificial pre-release state", were manually constructed to mimic the pre-release state on the PI(3,5)P_2-containing membrane. To this end, StarD4 in the defined pre-release state protein conformation was repositioned on the membrane to adopt the pre-release orientation. After an equilibration as described in the preparatory steps, adaptive sampling runs were carried out for a total of 312 replicas (6 replicas from each initial point), resulting in a total simulation time of 725 microseconds.

Steered MD simulation and the calculation of potential mean force using umbrella sampling

The Steered MD simulation was undertaken for accelerated sampling of the release paths of CHL from StarD4 in different conformational states. The collective variable guiding this process was the z-distance between the center of mass of cholesterol and the StarD4 α/β helix-grip structure (residues 70-76, 79-86, 103-108, 113-118, 131-140, 146-150, 170-175, 184-189, 211-220). Using a spring constant of 5 kcal/mol·Å^2, the pulling phase of target was carried out with a very low pulling velocity of 0.0002Å/ps for 2ns, followed by an equilibration phase without target movement for 8ns. This 10ns cycle was continued within the first 200 ns. Subsequently, the velocity was increased to 0.0003Å/ps in the pulling phase, and the cycle is continued for another 200 ns. The total pulling lengths of 20Å was covered in 400ns, which moved the CHL from the binding site to the membrane. The 24 replicas of SMD generated from each state of interest (states 1,2,5,6,7 in **Fig. 7A,B**), were run according to this protocol, amounting to a total of 48 microseconds of SMD exploration.

The potential of mean force calculations used the same collective variable – i.e. the z-distance between the cholesterol and the α/β helix-grip structure – as the reaction coordinate. Fifteen umbrella sampling windows were defined along the path, covering z=7 to 21Å with a spacing of 1Å. Each window utilized a harmonic potential function with a force constant of 1 kcal/mol·Å^2. For each window, 15 independent runs of 40 ns were performed. The initial 10 ns of each run were considered

equilibration and were excluded from subsequent analysis. The potential of mean force energy profiles were constructed using the Weighted Histogram Analysis Method (WHAM) algorithms (54).

The production run simulations of the membrane embedded *apo*-StarD4

The production run simulations of the membrane embedded *apo*-StarD4 were carried out on the same computational platform as the simulations of *holo*-StarD4. The simulation design is outlined in **Fig. 14.**

Phases 1&2 of 0.21, 0.14 milliseconds: The initial two phases mirror the design employed for *holo*-StarD4 simulations. In Phase 1 we establish an equilibrated membrane embedded StarD4-membrane, utilizing 81 statistically independent replicas amounting to 211 microseconds. This system is explored in Phase 2 to reveal the conformational changes after membrane embedding of StarD4 in the presence of membrane cholesterol, utilizing 36 statistically independent replicas amounting to 144 microseconds.

Phase 3 of 0.19 milliseconds: Probes the movement of membrane cholesterols near StarD4 with its Ω1-H4 gate open. The starting points were conformations resulting from the simulation of membrane embedded apo-StarD4 in phase 2 with a Ω1-H4-distance (defined as described in Supplementary Methods section 1) of >9Å. These conformations were replicated and continued. The protocol of Phase 3 includes 24 independent runs for the PI(3,5)P_2- or PS-containing membrane, and 48 independent runs for the PI(4,5)P_2-containing membrane, totaling 192 microseconds of cumulative simulation time.

Phases 4-6 of 90, 72, 72 microseconds: The protocols of the Phase 4-6 simulations explore the CHL uptake pathways from the membrane to the hydrophobic pocket. As shown in Figure 14, the initial configurations for Phase 4 originated in Phase 3 and were selected if their CHL molecule had raised above the membrane surface, interacting with StarD4 at S208 or A201. Subsequently, the 6 trajectories in which cholesterol had reached highest above the membrane surface in Phase 4 simulations, were replicated as shown in **Figure 14** and continued in Phase 5. This process was repeated in going from Phase 5 to Phase 6. In the groups termed " Δlipid-tail", a specific alteration was made: the membrane lipid tails that occasionally also extended above the membrane surface were manually removed. This adjustment aimed to speed uptake convergence by preventing the cycles of competition for the hydrophobic pocket with the extracted CHL. The modification involved substituting the lipid with a conformation where both lipid tails resided within the membrane, while maintaining the interaction of the protein with the lipid head (**Suppl. Fig 12**). The initial seeds that underwent this manual modification were re-equilibrated before the production run, with constrains applied on the protein backbone and the target cholesterol (following the preparation protocol described for the of StarD4-membrane complex).

Rare Event Detection analysis

As described in the main text, we used the Rare Event Detection (RED) protocol detailed in our publications (21,23) to find the structural motifs that move together to establish new conformations and PIP2-protein interaction modes that dominate during specific time segments of the trajectory. Briefly, the protocol implements the Non-negative Matrix Factorization (NMF) module in the Scikit-Learn Python package (55) to identify rare structural rearrangement events in the molecular dynamics

(MD) trajectory data by decomposing a trajectory into structural motifs and detect those undergoing simultaneous changes that dominate the conformation of the protein for specific periods along the trajectory. The input to the NMF algorithm is a contact matrix constructed from the entire trajectory, which therefore contains the information about the evolving protein structure. In the NMF algorithm this matrix is further decomposed into a spatial array (W) and a temporal array (H) as described in (23), with the former defining "components" (which are sets of conformational features that evolve concurrently in the trajectory), and the latter depicting the time-ordered contribution of each component along the trajectory (21,23).

Here, the contact matrix contains residue-residue interactions and residue-pair/PIP2 interactions calculated in each of the 10,000 frames sampled along the 4-microseconds simulation of RT1. For residue-residue contacts matrix, a contact is marked as 1 if any atom from one residue is within 3.5Å of that of another residue, resulting in a $201^2 \times 10000$ matrix. In the residue-pair-PIP2 contact matrix, a diagonal entity is marked as 1 if any atom from one residue is within 3.5Å of a PIP2, while an off-diagonal entity is marked as 1 if both residues are within 3.5Å of a shared PIP2, resulting in another $201^2 \times 10000$ matrix for 10000 frames. These two matrices of size 40401×10000 are concatenated and result in an 80802×10000 matrix representing the residue-pair/PIP2 interactions for every frame. To capture dynamic information, contacts that remain unchanged by these criteria throughout the simulation are excluded. The resulting contact map contains 2171 columns of pairs of residues that alter their contact states during the simulation, and has 10,000 rows, recorded at intervals of 0.4ns. Smoothing (see (21,23)) is applied across the time dimension using a sliding window of 20 nanoseconds to refine the array.

The NMF algorithm then decomposes this contact matrix into the W and H components that place structural states in spatial and temporal matrices. In this study, the choice of six components (see (21,23)) reveals the transition of conformational states in RT1. The spatial arrays are normalized, allowing direct comparison between components, and the differences between components help identify structure-differentiating contact pairs (SDCPs). These SDCPs, highlighted in the transition from one component to another as described in (21), denote pairs that contribute significantly to structural variations, which yields a comprehensive identification of rare events within the trajectory data.

tICA - the time-structure based independent component analysis used for dimensionality reduction and identification of metastable states

The dimensionality reduction approach we used for the analysis of the conformational space as described in the main text was carried out with the well-known tICA method (56-60). The tICA is constructed by projecting the simulation data onto low dimensionality (usually 2D) spaces (see **Figs. 7C, 9B, 17A**) defined by several collective variables (CVs) that quantify the dynamics of interest. The CVs we used are described in detail in Supplementary Methods. The tICA transformation of the simulation data identifies the slowest reaction coordinates in the conformational space by solving for the generalized eigenvectors and eigenvalues of the time-lagged covariance matrix:

$$C^x(\tau)\mathrm{V}_i = \lambda_i C^x(0)\mathrm{V}_i$$

where $C^x(\tau)$ is the time-lagged covariance matrix ($C^x(\tau) = \langle X(t)X^T(t+\tau)\rangle_t$) between collective variables, $C^x(\tau)$ is the regular covariance matrix ($C^x(0) = \langle X(t)X^T(t)\rangle_t$), $X(t)$ is the data vector at

time t, τ is the lag-time of the covariance matrix, V_i is the ith tICA eigenvector, and λ_i is the ith tICA eigenvalue of the covariance matrix. In this study, the trajectory was sampled at a stride of 0.4 ns/frame, and the lag-time τ for constructing the covariance matrix was set to 20 ns (50 frames). This choice allowed the exclusion of fluctuations faster than 20 ns, which were considered not relevant to the slow processes in the system. The first 200 ns in every independent run were discarded to reduce equilibration artifacts before analysis. This dimensionality reduction technique facilitated a focused exploration of the slow processes and relevant reaction coordinates in the simulated dynamics as described below.

Mapping the cholesterol uptake pathway and assignment of macrostates in the tICA space: This analysis focused on the Phases 4-6 of the simulation of *apo*-StarD4 membrane complex (**Fig. 14**) in which adaptive sampling was used to simulate the progress of membrane cholesterol into the hydrophobic pocket of StarD4. Key collective variables were recorded to assess the position of the cholesterol (as defined supplementary methods) in the 234 simulation trajectories conducted in these three phases. The CHL position data included the height reached above the membrane surface, the distance to the Ω1-H4 gate, and its distance to the hydrophilic residues that served as a ladder into the hydrophobic pocket of StarD4. These data were used as CVs to define the tICA transformation. To identify metastable states in the cholesterol uptake process, the 2D tICA space was discretized into 100 microstates using K-means clustering. Subsequently, these microstates were further clustered into 10 macrostates based on their kinetic similarities, determined from the transitional rate between microstates. This clustering process was achieved using the Robust Perron Cluster Analysis (PCCA+) algorithm (61).

Construction of the PIP2 Interaction Mode Landscape (IML) and assignment of PIP2 binding modes.

The IML for PIP2 binding modes to the StarD4 (**Fig. 9C,D**) represents the positions the PIP2 molecules that had established stable interactions with the basic residues. The criterion for considering a PIP2 molecule to be binding to the upper basic residue patch is that it remains for at least 10 ns within a distance of <4.5Å from at least two of residues K49, K50, K52, R97, S215, R218, K219, R222 ("upper basic group"). The position of such a PIP2 molecule was recorded from the first frame where it formed an interaction with upper-group residue until the last frame of its binding in the same trajectory. If unbinding and later rebinding occur in the same trajectory for the same PIP2, its position is continuously recorded.

This analysis was performed on the trajectories of Phases 2&3 (**Figs. 2&14)** of both *apo*- and *holo*-StarD4 embedded in either PI(3,5)P_2- or PI(4,5)P_2-containing membranes. A total of 288 trajectories of StarD4-membrane complexes were included in this analysis. From this set of simulations, a total of 872 PIP2s that bind to upper group residues were recorded. (Note that multiple PIP2s can bind simultaneously to each StarD4). The positions of bound PIP2 were defined using 5 collective variables (CVs), each measuring the distance between the PIP2 anionic atoms and the closest basic atoms in the K52/R97/S215/R218/K219 patch, resulting in 872 position-time matrices representing the evolution of the PIP2-StarD4 interactions over time. These CVs served in the construction of the tICA dimensionality reduction analysis as described above.

The first two tICA vectors composed of the 5 CVs were found to describe 85% of the total dynamics of the PIP2 interacting with the upper basic patch. To facilitate the comparison of population

densities and preferences between PI(3,5)P_2 and PI(4,5)P_2 binding modes, the 2D tICA space was discretized into 100 microstates using K-means clustering, and segregated into 9 macrostates based on their kinetics similarities (see above). The assignment of macrostates provides a valuable mapping tool that facilitates direct comparison between PI(3,5)P_2 and PI(4,5)P_2 population densities in the states they represent. These macrostates are herein designated as the definition of the binding modes 0 to 8 of PIP2. Every timestep in the 872 position-time matrices of PIP2 is assigned to its macrostate, representing the binding mode of a certain lipid at the given time. Then, these 872 trajectories of PIP2 are mapped back to the original 288 trajectories of StarD4-membrane system where multiple PIP2 may binding simultaneously on one StarD4, resulting a "matrix of binding mode over time" which quantifies the number of lipid(s) in each lipid binding mode on a StarD4 at any given time. This matrix provides a summary of the occurrence and distribution of different binding modes on StarD4 throughout the simulation. We note, however, that because the kinetics similarities of the microstates that served in the definition of the 9 macrostates were derived from a mixture of PI(3,5)P_2 and PI(4,5)P_2 trajectories, the macrostate assignment from this map may contain microstates from both, and cannot be used to study the dynamics of the protein in the individual environments.

The Deep Neural Network

Input data structure: The DNNs described in the main text were trained to study the difference between the StarD4 interacting with membranes containing PI(3,5)P_2 vs membranes containing PI(4,5)P_2. The input dataset for the DNNs consisted of the ensemble of trajectories containing ~46,000 StarD4 protein structures, without the information regarding the ligand CHL or the membrane lipids. The protein structures were sampled from the simulations of StarD4-membrane systems including both the membrane-embedding phase and the membrane-embedded phase, at 8ns time-intervals. The first 160ns of every trajectory were considered equilibration time and excluded from the data set. The simulations used for the DNN dataset sampling are listed in **Suppl. Table 6**.

The trajectory data were subjected first to a *debiasing protocol* that consists of random scrambling of the position and orientation of the protein to eliminate their influence on the decision-making process of the DNNs. With the debiased data, the DNNs rely solely on intramolecular protein conformational changes to predict the labeled lipid type. To further augment the data, each protein structure is scrambled into three different random orientations, resulting in approximately 14,000 samples. This dataset was divided into training, validation, and test sets in a ratio of 56:24:20, respectively, keeping track of the three samples derived from the orientation scrambling of the same structure.

The analysis of the molecular dynamics trajectories employed image-recognition convolutional neural networks in a pipeline developed in our lab (30) in which the coordinates of the 3D protein structures are transformed into 2D pixel representations that retain their 3D information. Briefly, to convert a structure into a picture, the (X,Y,Z) coordinates of each atom are normalized and converted into a set of (R,G,B) values of a color point. These color points are then arranged in sequential order according to the protein sequence from top left to bottom right in a 2D input picture readable by a convolutional neural network. In the case of DNN based on the full StarD4 protein, the first 3192 atoms of the residues in the 24 to 223 range are converted into a picture representation with a size of (56,57,3), where the 56 rows and 57 columns collectively contain 3192 pixels representing each atom, and the third dimension contains 3 (R,G,B) colors representing the X,Y,Z of the atom. In the case of

DNN based on the salient residues, the first 400 atoms in the residues 46 51 52 53 58 62 63 67 123 124 130 160 165 167 175 180 181 188 189 192 194 195 196 are converted into a picture representation with a size of (20,20,3). This method allows for the efficient application of image-recognition convolutional neural networks to the analysis of molecular dynamics trajectories.

Building the DNN: The recognition of the protein structures in their image representation is carried out with custom Dense Convolutional Networks in Keras with a Tensorflow based on a pre-existing implementation (62). The DNNs utilize Dense Blocks, where feature-maps from all the preceding layers are used as inputs for the subsequent layer. In the case of the DNN based on full protein atoms, the DNNs for StarD4 take images of size (56,57,3) as inputs, starting with a convolutional layer with a stride of 2, followed by 3 dense blocks containing 6, 12, 24 convolutional layers respectively. When only the residues identified as the salient contributing features by the DNN (see above) were used as input to another DNN analysis run as described in the main text, the input was constructed as images of size (20,20,3), starting with a convolutional layer with a stride of 1, followed by 3 dense blocks containing 6, 12, 24 convolutional layers. The DNNs utilize a growth rate of 48 filters per layer, 96 initial filters, a reduction ratio of 0.5 in the transition between Dense Blocks. A dropout rate of 0.2 is applied in every convolutional layer during training.

Visualization of the salient contributing features in DNN: The saliency analyses are conducted among the top 1000 samples, where the DNN reported the highest confidences in correctly identifying the PIP2 subtype. The contribution of each atom to the decision-making by DNN was evaluated using the visual saliency package provided by keras-vis (63) to calculate the gradient of the output label over the input features with guided back-propagation. The contributions of each atom are averaged over the 1000 samples, resulting in a saliency map of the same shape as the input sample size.

Quantifying the allosteric mechanism using N-body information theory (NbIT)

We used the information theory-based NbIT analysis methodology (24,25) to calculate the terms that quantify the information sharing between structural motifs involved in the allosteric communication and pathways. The structural motifs involved in the analysis are defined below, at the end of this section.

A key term defined in detail (24,25) and used as described in recent publications (64-67) is the *Coordination Information* (CI) between motifs that is calculated from the entropy in each motif $H(X)$ as described in detail in (24) where

$H(X) = \frac{1}{2}\ln(|2\pi e C(X)|)$, where $C(X)$ is the covariance matrix of the position array X.

Briefly summarized, the *Coordination Information calculation* starts from the total information shared by structural motifs within a set, termed *Total Correlation* (TC), that is calculated from differential entropy terms as:

$$TC(X_1, \dots, X_N) = \sum_i^N H(X_i) - H(X_1, \dots, X_N).$$

A *Conditional Total Correlation*, indicating the remaining information within a set of motifs $\{X_1, \dots, X_N\}$ given the dynamics of another motif X_m, is then obtained as:

$$TC(X_1, \dots, X_N \mid X_m) = \sum_i^N (H(X_i, X_m) - H(X_m)) - \big(H(X_1, \dots, X_N, X_m) - H(X_m)\big).$$

By relating the *Total Correlation* and the *Conditional Total Correlation*, we obtain the *Coordination Information* (CI) that quantifies the portion of information shared in the structural motifs described in the main text as *receptors* $\{X_1, \dots, X_N\}$, that is also shared with X_m motif that functions as the *transmitter*:

$$\overline{CI}(\{X_1, \dots, X_N\}, X_m) = \frac{TC(X_1,\dots,X_N) - TC(X_1,\dots,X_N \mid X_m)}{TC(X_1,\dots,X_N)} * 100\%.$$

In **Tables 2&3** and the **Suppl. Table 4**, the *Total Correlation* within each motif is displayed along the diagonal. The off-diagonal elements quantify the *Normalized Coordination Information*. In these Tables, the structural motifs shown at the top of each column are acting as the *Transmitter* and those on the left side of each row indicate the coordinated *Receiver*.

The same information is used to identify the allosteric channels that connect distant motifs, identified by calculating a term named *Normalized Mutual Coordination Information* ($NMCI$) (see (24)), which measures the portion of coordination information shared between the *receiver* $\{X_1, \dots, X_N\}$ and the *transmitter* motifs X_m that is also shared with another structural element X_n:

$$NMCI(\{X_1, \dots, X_N\}, X_m, X_n) = \frac{CI(\{X_1, \dots, X_N\}, X_m) + CI(\{X_1, \dots, X_N\}, X_n) - CI(\{X_1, \dots, X_N\}, X_m X_n)}{CI(\{X_1, \dots, X_N\}, X_m)}$$

Definition of the structural motifs included in the NbIT analysis: **CHL sites**: S136 S147 W171 R92 Y117; **β1**: R46 V47 A48 K49 K50 V51; **β2&β3**: R58 K59 P60 Y67 L68 Y69; **β2β3 loop**: E62 E63 F64 N65; **H4-N-terminal (H4-Nt)**: Q199 S200 A201 D203 T204 A207; **Ω1 loop**: L124 N125 I126; **Ω4 loop**: D192 R194 G195; **PIP2 binding mode (Pbm)**: K52, R97, S215, K219; **Water influx**: N166, C169, I189, T191; **β8β9 Loop**: K177 D178 S179 P180; **β4-Nt**: H107 F108 E109; **H4-Ct**: G220 L221 K223 A224.

ACKNOWLEDGEMENTS

We gratefully acknowledge helpful discussions with Drs. Ambrose Plante, and Derek M. Shore, and with Prof. George Khelashvili. HX gratefully acknowledges their helpful guidance regarding the principles, applicability, implementation, and interpretation of the RED, NbIT and TPT analysis. We are grateful for very helpful discussions with Prof. Frederick R. Maxfield and experimental data from his lab members. Support from the 1923 Foundation for the project "*How Needed Molecular Precision is Achieved for Addressing, Pickup, and Delivery in the Trafficking of Cholesterol Among Cell Membranes*" is gratefully acknowledged. The computational resources and technical help at the Center for Computational Innovations (CCI) at the Rensselaer Polytechnic Institute (RPI), and the efficient and sustained access to the AiMOS supercomputer at CCI generously awarded through the COVID-19 High Performance Computing Consortium, are gratefully acknowledged. We are grateful for the computational resources under Project BIP109 at the Oak Ridge Leadership Computing Facility, which is a DOE Office of Science User Facility supported under Contract DE-AC05-00OR22725, and for the in-house computational resources of the David A. Cofrin Center for Biomedical Information in the Institute for Computational Biomedicine at Weill Cornell Medical College.

References

1. Hao, M., Lin, S. X., Karylowski, O. J., Wustner, D., McGraw, T. E., and Maxfield, F. R. (2002) Vesicular and non-vesicular sterol transport in living cells. The endocytic recycling compartment is a major sterol storage organelle. *J Biol Chem* **277**, 609-617
2. Iaea, D. B., Mao, S., Lund, F. W., and Maxfield, F. R. (2017) Role of STARD4 in sterol transport between the endocytic recycling compartment and the plasma membrane. *Mol Biol Cell* **28**, 1111-1122
3. Iaea, D. B., and Maxfield, F. R. (2015) Cholesterol trafficking and distribution. *Essays Biochem* **57**, 43-55
4. Prinz, W. A. (2007) Non-vesicular sterol transport in cells. *Prog Lipid Res* **46**, 297-314
5. Wong, L. H., Gatta, A. T., and Levine, T. P. (2019) Lipid transfer proteins: the lipid commute via shuttles, bridges and tubes. *Nat Rev Mol Cell Biol* **20**, 85-101
6. Letourneau, D., Lefebvre, A., Lavigne, P., and LeHoux, J. G. (2015) The binding site specificity of STARD4 subfamily: Breaking the cholesterol paradigm. *Mol Cell Endocrinol* **408**, 53-61
7. Mathieu, A. P., Fleury, A., Ducharme, L., Lavigne, P., and LeHoux, J. G. (2002) Insights into steroidogenic acute regulatory protein (StAR)-dependent cholesterol transfer in mitochondria: evidence from molecular modeling and structure-based thermodynamics supporting the existence of partially unfolded states of StAR. *J Mol Endocrinol* **29**, 327-345
8. Romanowski, M. J., Soccio, R. E., Breslow, J. L., and Burley, S. K. (2002) Crystal structure of the Mus musculus cholesterol-regulated START protein 4 (StarD4) containing a StAR-related lipid transfer domain. *Proc Natl Acad Sci U S A* **99**, 6949-6954
9. Iaea, D. B., Dikiy, I., Kiburu, I., Eliezer, D., and Maxfield, F. R. (2015) STARD4 Membrane Interactions and Sterol Binding. *Biochemistry* **54**, 4623-4636
10. Radhakrishnan, A., Goldstein, J. L., McDonald, J. G., and Brown, M. S. (2008) Switch-like control of SREBP-2 transport triggered by small changes in ER cholesterol: a delicate balance. *Cell Metab* **8**, 512-521
11. Liscum, L., and Munn, N. J. (1999) Intracellular cholesterol transport. *Biochim Biophys Acta* **1438**, 19-37
12. Zhang, X., Xie, H., Iaea, D., Khelashvili, G., Weinstein, H., and Maxfield, F. R. (2022) Phosphatidylinositol phosphates modulate interactions between the StarD4 sterol trafficking protein and lipid membranes. *J Biol Chem* **298**, 102058
13. Maxfield, F., and Hao, M. (2013) Lipid Trafficking in Cells. in *Encyclopedia of Biophysics* (Roberts, G. K. ed.), Springer Berlin Heidelberg. pp 1289-1296
14. Tsujishita, Y., and Hurley, J. H. (2000) Structure and lipid transport mechanism of a StAR-related domain. *Nat Struct Biol* **7**, 408-414
15. Murcia, M., Faraldo-Gomez, J. D., Maxfield, F. R., and Roux, B. (2006) Modeling the structure of the StART domains of MLN64 and StAR proteins in complex with cholesterol. *J Lipid Res* **47**, 2614-2630
16. Thorsell, A. G., Lee, W. H., Persson, C., Siponen, M. I., Nilsson, M., Busam, R. D., Kotenyova, T., Schuler, H., and Lehtio, L. (2011) Comparative structural analysis of lipid binding START domains. *PLoS One* **6**, e19521
17. Tan, L., Tong, J., Chun, C., and Im, Y. J. (2019) Structural analysis of human sterol transfer protein STARD4. *Biochem Biophys Res Commun* **520**, 466-472
18. Horenkamp, F. A., Valverde, D. P., Nunnari, J., and Reinisch, K. M. (2018) Molecular basis for sterol transport by StART-like lipid transfer domains. *EMBO J* **37**
19. Tong, J., Manik, M. K., and Im, Y. J. (2018) Structural basis of sterol recognition and nonvesicular transport by lipid transfer proteins anchored at membrane contact sites. *Proc Natl Acad Sci U S A* **115**, E856-E865
20. Jentsch, J. A., Kiburu, I., Pandey, K., Timme, M., Ramlall, T., Levkau, B., Wu, J., Eliezer, D., Boudker, O., and Menon, A. K. (2018) Structural basis of sterol binding and transport by a yeast StARkin domain. *J Biol Chem* **293**, 5522-5531
21. Xie, H., and Weinstein, H. (2023) Allosterically coupled conformational dynamics in solution prepare the sterol transfer protein StarD4 to release its cargo upon interaction with target membranes. *Frontiers in Molecular Biosciences* **10**

22. Reza, T., Larissa van, E., Charlotte, G., Dandan, X., Mahmoud, M., Anne-Claude, G., and Nathalie, R. (2023) Membrane specificity of the human cholesterol transfer protein STARD4. *bioRxiv*, 2023.2012.2004.569895
23. Plante, A., and Weinstein, H. (2021) Ligand-Dependent Conformational Transitions in Molecular Dynamics Trajectories of GPCRs Revealed by a New Machine Learning Rare Event Detection Protocol. *Molecules* **26**
24. LeVine, M. V., and Weinstein, H. (2014) NbIT--a new information theory-based analysis of allosteric mechanisms reveals residues that underlie function in the leucine transporter LeuT. *PLoS Comput Biol* **10**, e1003603
25. LeVine, M., Perez-Aguilar, J. M., and Weinstein, H. (2014) *N-body Information Theory (NbIT) Analysis of Rigid- Body Dynamics in Intracellular Loop 2 of the 5-HT 2A Receptor*,
26. Sotomayor, M., and Schulten, K. (2007) Single-molecule experiments in vitro and in silico. *Science* **316**, 1144-1148
27. Izrailev, S., Stepaniants, S., Balsera, M., Oono, Y., and Schulten, K. (1997) Molecular dynamics study of unbinding of the avidin-biotin complex. *Biophys J* **72**, 1568-1581
28. Kumar, S., Rosenberg, J. M., Bouzida, D., Swendsen, R. H., and Kollman, P. A. (1995) Multidimensional free-energy calculations using the weighted histogram analysis method. *Journal of Computational Chemistry* **16**, 1339-1350
29. Roux, B. (1995) The calculation of the potential of mean force using computer simulations. *Computer Physics Communications* **91**, 275-282
30. Plante, A., Shore, D. M., Morra, G., Khelashvili, G., and Weinstein, H. (2019) A Machine Learning Approach for the Discovery of Ligand-Specific Functional Mechanisms of GPCRs. *Molecules* **24**
31. Mesmin, B., Pipalia, N. H., Lund, F. W., Ramlall, T. F., Sokolov, A., Eliezer, D., and Maxfield, F. R. (2011) STARD4 abundance regulates sterol transport and sensing. *Mol Biol Cell* **22**, 4004-4015
32. George Khelashvili, N. C., Kalpana Pandey, David Eliezer, Anant K Menon. (2019) Exchange of water for sterol underlies sterol egress from a StARkin domain. *eLife* **8:e53444**
33. Iaea, D. B., Spahr, Z. R., Singh, R. K., Chan, R. B., Zhou, B., Bareja, R., Elemento, O., Di Paolo, G., Zhang, X., and Maxfield, F. R. (2020) Stable reduction of STARD4 alters cholesterol regulation and lipid homeostasis. *Biochim Biophys Acta Mol Cell Biol Lipids* **1865**, 158609
34. Huang, T., Shan, R., Zhang, M., Li, L., Huang, J., Liu, B., and Zhou, W. (2021) [Steroidogenic acute regulatory protein-related lipid transfer 4 (StarD4) promotes breast cancer cell proliferation and its mechanism]. *Sheng Wu Yi Xue Gong Cheng Xue Za Zhi* **38**, 1118-1125
35. Yue, X., Kong, Y., Zhang, Y., Sun, M., Liu, S., Wu, Z., Gao, L., Liang, X., and Ma, C. (2023) SREBF2-STARD4 axis confers sorafenib resistance in hepatocellular carcinoma by regulating mitochondrial cholesterol homeostasis. *Cancer Sci* **114**, 477-489
36. Ferrari, A., Vincent-Salomon, A., Pivot, X., Sertier, A. S., Thomas, E., Tonon, L., Boyault, S., Mulugeta, E., Treilleux, I., MacGrogan, G., Arnould, L., Kielbassa, J., Le Texier, V., Blanche, H., Deleuze, J. F., Jacquemier, J., Mathieu, M. C., Penault-Llorca, F., Bibeau, F., Mariani, O., Mannina, C., Pierga, J. Y., Tredan, O., Bachelot, T., Bonnefoi, H., Romieu, G., Fumoleau, P., Delaloge, S., Rios, M., Ferrero, J. M., Tarpin, C., Bouteille, C., Calvo, F., Gut, I. G., Gut, M., Martin, S., Nik-Zainal, S., Stratton, M. R., Pauporte, I., Saintigny, P., Birnbaum, D., Viari, A., and Thomas, G. (2016) A whole-genome sequence and transcriptome perspective on HER2-positive breast cancers. *Nat Commun* **7**, 12222
37. Dutton-Regester, K., Kakavand, H., Aoude, L. G., Stark, M. S., Gartside, M. G., Johansson, P., O'Connor, L., Lanagan, C., Tembe, V., Pupo, G. M., Haydu, L. E., Schmidt, C. W., Mann, G. J., Thompson, J. F., Scolyer, R. A., and Hayward, N. K. (2013) Melanomas of unknown primary have a mutation profile consistent with cutaneous sun-exposed melanoma. *Pigment Cell Melanoma Res* **26**, 852-860
38. Van Allen, E. M., Wagle, N., Sucker, A., Treacy, D. J., Johannessen, C. M., Goetz, E. M., Place, C. S., Taylor-Weiner, A., Whittaker, S., Kryukov, G. V., Hodis, E., Rosenberg, M., McKenna, A., Cibulskis, K., Farlow, D., Zimmer, L., Hillen, U., Gutzmer, R., Goldinger, S. M., Ugurel, S., Gogas, H. J., Egberts, F., Berking, C., Trefzer, U., Loquai, C., Weide, B., Hassel, J. C., Gabriel, S. B., Carter, S. L., Getz, G., Garraway, L. A., Schadendorf, D., and Dermatologic Cooperative Oncology Group of, G. (2014) The

genetic landscape of clinical resistance to RAF inhibition in metastatic melanoma. *Cancer Discov* **4**, 94-109
39. Sanborn, J. Z., Chung, J., Purdom, E., Wang, N. J., Kakavand, H., Wilmott, J. S., Butler, T., Thompson, J. F., Mann, G. J., Haydu, L. E., Saw, R. P., Busam, K. J., Lo, R. S., Collisson, E. A., Hur, J. S., Spellman, P. T., Cleaver, J. E., Gray, J. W., Huh, N., Murali, R., Scolyer, R. A., Bastian, B. C., and Cho, R. J. (2015) Phylogenetic analyses of melanoma reveal complex patterns of metastatic dissemination. *Proc Natl Acad Sci U S A* **112**, 10995-11000
40. Hayward, N. K., Wilmott, J. S., Waddell, N., Johansson, P. A., Field, M. A., Nones, K., Patch, A. M., Kakavand, H., Alexandrov, L. B., Burke, H., Jakrot, V., Kazakoff, S., Holmes, O., Leonard, C., Sabarinathan, R., Mularoni, L., Wood, S., Xu, Q., Waddell, N., Tembe, V., Pupo, G. M., De Paoli-Iseppi, R., Vilain, R. E., Shang, P., Lau, L. M. S., Dagg, R. A., Schramm, S. J., Pritchard, A., Dutton-Regester, K., Newell, F., Fitzgerald, A., Shang, C. A., Grimmond, S. M., Pickett, H. A., Yang, J. Y., Stretch, J. R., Behren, A., Kefford, R. F., Hersey, P., Long, G. V., Cebon, J., Shackleton, M., Spillane, A. J., Saw, R. P. M., Lopez-Bigas, N., Pearson, J. V., Thompson, J. F., Scolyer, R. A., and Mann, G. J. (2017) Whole-genome landscapes of major melanoma subtypes. *Nature* **545**, 175-180
41. ICGC(BRCA-FR) : Breast Cancer - FR.
42. ICGC(SKCM-US) : Skin Cutaneous melanoma - TCGA, US.
43. Sali, A. (1995) Comparative protein modeling by satisfaction of spatial restraints. *Mol Med Today* **1**, 270-277
44. Sherman, W., Day, T., Jacobson, M. P., Friesner, R. A., and Farid, R. (2006) Novel procedure for modeling ligand/receptor induced fit effects. *J Med Chem* **49**, 534-553
45. Sherman, W., Beard, H. S., and Farid, R. (2006) Use of an Induced Fit Receptor Structure in Virtual Screening. *Chemical Biology & Drug Design* **67**, 83-84
46. Miller, E. B., Murphy, R. B., Sindhikara, D., Borrelli, K. W., Grisewood, M. J., Ranalli, F., Dixon, S. L., Jerome, S., Boyles, N. A., Day, T., Ghanakota, P., Mondal, S., Rafi, S. B., Troast, D. M., Abel, R., and Friesner, R. A. (2021) Reliable and Accurate Solution to the Induced Fit Docking Problem for Protein-Ligand Binding. *J Chem Theory Comput* **17**, 2630-2639
47. Phillips, J. C., Braun, R., Wang, W., Gumbart, J., Tajkhorshid, E., Villa, E., Chipot, C., Skeel, R. D., Kale, L., and Schulten, K. (2005) Scalable molecular dynamics with NAMD. *J Comput Chem* **26**, 1781-1802
48. Jo, S., Kim, T., Iyer, V. G., and Im, W. (2008) CHARMM-GUI: a web-based graphical user interface for CHARMM. *J Comput Chem* **29**, 1859-1865
49. Eastman, P., Swails, J., Chodera, J. D., McGibbon, R. T., Zhao, Y., Beauchamp, K. A., Wang, L. P., Simmonett, A. C., Harrigan, M. P., Stern, C. D., Wiewiora, R. P., Brooks, B. R., and Pande, V. S. (2017) OpenMM 7: Rapid development of high performance algorithms for molecular dynamics. *PLoS Comput Biol* **13**, e1005659
50. Khelashvili, G., Weinstein, H., and Harries, D. (2008) Protein diffusion on charged membranes: a dynamic mean-field model describes time evolution and lipid reorganization. *Biophys J* **94**, 2580-2597
51. Mondal, S., Khelashvili, G., Shan, J., Andersen, O. S., and Weinstein, H. (2011) Quantitative modeling of membrane deformations by multihelical membrane proteins: application to G-protein coupled receptors. *Biophys J* **101**, 2092-2101
52. Khelashvili, G., and Harries, D. (2010) Modeling Signaling Processes across Cellular Membranes Using a Mesoscopic Approach. in *Annual Reports in Computational Chemistry* (Wheeler, R. A. ed.), Elsevier. pp 236-261
53. Khelashvili, G., Galli, A., and Weinstein, H. (2012) Phosphatidylinositol 4,5-biphosphate (PIP(2)) lipids regulate the phosphorylation of syntaxin N-terminus by modulating both its position and local structure. *Biochemistry* **51**, 7685-7698
54. Kumar, S., Rosenberg, J. M., Bouzida, D., Swendsen, R. H., and Kollman, P. A. (1992) THE weighted histogram analysis method for free-energy calculations on biomolecules. I. The method. *Journal of Computational Chemistry* **13**, 1011-1021
55. Fabian Pedregosa, G. V., Alexandre Gramfort, Vincent Michel, Bertrand Thirion, Olivier Grisel, Mathieu Blondel, Peter Prettenhofer, Ron Weiss, Vincent Dubourg, Jake Vanderplas, Alexandre Passos, David

Cournapeau, Matthieu Brucher, Matthieu Perrot, Édouard Duchesnay. (2011) Scikit-learn: Machine Learning in Python. *JMLR* **12**, 2825-2830
56. Molgedey, L., and Schuster, H. G. (1994) Separation of a mixture of independent signals using time delayed correlations. *Physical Review Letters* **72**, 3634-3637
57. Naritomi, Y., and Fuchigami, S. (2011) Slow dynamics in protein fluctuations revealed by time-structure based independent component analysis: the case of domain motions. *J Chem Phys* **134**, 065101
58. Perez-Hernandez, G., Paul, F., Giorgino, T., De Fabritiis, G., and Noe, F. (2013) Identification of slow molecular order parameters for Markov model construction. *J Chem Phys* **139**, 015102
59. Schwantes, C. R., and Pande, V. S. (2013) Improvements in Markov State Model Construction Reveal Many Non-Native Interactions in the Folding of NTL9. *J Chem Theory Comput* **9**, 2000-2009
60. Razavi, A. M., Khelashvili, G., and Weinstein, H. (2017) A Markov State-based Quantitative Kinetic Model of Sodium Release from the Dopamine Transporter. *Sci Rep* **7**, 40076
61. Deuflhard, P., and Weber, M. (2005) Robust Perron cluster analysis in conformation dynamics. *Linear Algebra and its Applications* **398**, 161-184
62. Gao Huang, Z. L., Laurens van der Maaten, Kilian Q. Weinberger. (2018) Densely Connected Convolutional Networks. *arXiv*, 1608.06993
63. Kotikalapudi, R., & Contributors. (2017) keras-vis. . *Retrieved from https://github.com/raghakot/keras-vis*
64. Khelashvili, G., Kots, E., Cheng, X., Levine, M. V., and Weinstein, H. (2022) The allosteric mechanism leading to an open-groove lipid conductive state of the TMEM16F scramblase. *Commun Biol* **5**, 990
65. Osei-Owusu, J., Kots, E., Ruan, Z., Mihaljevic, L., Chen, K. H., Tamhaney, A., Ye, X., Lu, W., Weinstein, H., and Qiu, Z. (2022) Molecular determinants of pH sensing in the proton-activated chloride channel. *Proc Natl Acad Sci U S A* **119**, e2200727119
66. LeVine, M. V., Cuendet, M. A., Khelashvili, G., and Weinstein, H. (2016) Allosteric Mechanisms of Molecular Machines at the Membrane: Transport by Sodium-Coupled Symporters. *Chem Rev* **116**, 6552-6587
67. Stolzenberg, S., Michino, M., LeVine, M. V., Weinstein, H., and Shi, L. (2016) Computational approaches to detect allosteric pathways in transmembrane molecular machines. *Biochim Biophys Acta* **1858**, 1652-1662