Roles of the 5-HT1A receptor in zebrafish responses to potential threat and

in sociality

Loanne Valéria Xavier Bruce de Souza[1,2], Larissa Nunes Oliveira[1,2], Bruna Patrícia Dutra

Costa[1,3], Monica Lima-Maximino[4], Vivianni Veloso[2], Caio Maximino[1,2]

[1] Laboratório de Neurociências e Comportamento "Frederico Guilherme Graeff",

Universidade Federal do Sul e Sudeste do Pará, Marabá/PA, Brazil

[2] Programa de Pós-Graduação em Neurociência e Comportamento, Universidade Federal

Pará, Belém/PA, Brazil

[3] Rede de Biodiversidade e Biotecnologia da Amazônia Legal, Marabá/PA, Brazil

[4] Laboratório de Neurofarmacologia e Biofísica – LaNeF, Universidade do Estado do Pará,

Marabá/PA, Brazil

ABSTRACT

Anxiety is a normal emotion representing a reaction to potential danger, whereas fear can be

defined as a reaction to real, explicit danger. Anxiety-like behavior in animal models has

been associated with differences in the serotonergic system. Treatment of zebrafish cohorts

with 8-OH-DPAT, a full agonist at the 5-HT1A receptor, decreased anxiety-like behavior in

the novel tank test, but increased it in the phototaxis (light-dark preference) assay, both

considered assays for anxiety-like behavior for this species. The same treatment decreased

social approach in both the social investigation and social novelty phases of the social

preference test. Blocking the 5-HT1A receptor with WAY 100,635 shifted the dose-response

curve for the novel tank test rightward. These effects suggest a participation of the 5-HT1A

heteroreceptors in zebrafish anxiety and social preference, increasing anxiety and decreasing

sociality. Thus, the study of this receptor is important for a better understanding of anxiety-

like behavior in zebrafish and its relationship with similar phenomena in vertebrates.

Preprint: https://www.biorxiv.org/content/10.1101/2024.04.17.588464v1

Data: https://github.com/lanec-unifesspa/5HT/tree/main/5HT1A

Keywords: zebrafish; model organism; heteroreceptor; vertebrates; defensive behavior

Lay summary

This study looked at how a certain type of protein, the 5-HT1A receptor, affected anxiety in

zebrafish. It's important to understand that anxiety is a normal feeling we all get when we're

worried or scared about something, while fear is a reaction to an immediate danger. The

researchers wanted to see how a specific drug that activates the 5-HT1A receptor, 8-OH-

DPAT, affected the behavior of the fish. They used different tests to see how the fish reacted

in different situations.

When the fish were given the drug, their anxiety-like behavior decreased in one test where

they were put in a new tank. But in another test where they had to choose between light and

dark areas, their anxiety-like behavior actually increased. It also made the fish less interested

in being social with other fish. When a drug that blocks the 5-HT1A receptor, WAY 100,635,

was give together with 8-OH-DPAT, evidence was found for a receptor reserve.

The researchers found that this drug affected the way the fish behaved in these situations. It

seemed to make them more anxious in some cases and less interested in being social. This

tells us that the receptor has an impact on how the fish are feeling, which can help us learn

more about anxiety in zebrafish and, potentially, in other animals too, including humans.

Significance statement

This research indicates that 5-HT1A heteroreceptors play a key role in increasing anxiety and

decreasing sociality across vertebrates, shedding light on neural mechanisms that could inform

treatments for anxiety-related disorders.

1. Introduction

Under situations of potential threat, humans and non-human animals adjust their

behavior to cautious exploration, risk assessment, and behavioral inhibition, sometimes

described as "anxiety-like behavior" (McNaughton & Corr, 2004). This pattern of defensive

behavior is conceptually related to a fear-like state, but fear usually involves a response to an

actual threat, whether that threat is proximal or distal (Perusini & Fanselow, 2015). While

both are part of a taxonomy of defensive behavior, there is growing evidence to show that

anxiety and fear are distinct emotions in terms of behavioral responses, physiological

settings, and neural bases (Fanselow & Lester, 1988; McNaughton & Corr, 2022;

McNaughton & Zangrossi Jr, 2008; Perusini & Fanselow, 2015).

The relationship between the neurobehavioral responses to threat and the serotonergic

system points to a complex regulation, in which different subpopulations of serotonergic

neurons in the dorsal raphe nucleus and the medial raphe nucleus are uniquely involved in the

pathophysiology of anxiety and affective disorders (Deakin & Graeff, 1991; Maximino,

2012; Paul et al., 2014). This control is thought to be mediated by topographically organized

projections to different areas of the limbic forebrain and midbrain (Abrams et al., 2004, 2005;

Paul & Lowry, 2013). Serotonergic projections to forebrain regions, such as the amygaloid

nuclei and the ventral hippocampus, increase behavioral responses to potential threat when

activated, possibly via 5-HT2 and 5-HT3 receptors (Graeff et al., 1997; Maximino, 2012;

Olivier et al., 2000; Quesseveur et al., 2012). In the midbrain – especially in the

periaqueductal gray area –, activation of 5-HT1A and 5-HT2 receptors lead to an inhibition

of neurobehavioral responses to actual threat (Castilho et al., 2002; Gomes & Nunes-de-

Souza, 2009; Graeff, 2004; Graeff et al., 1997; Lima-Maximino et al., 2020; Paul et al.,

2014). Thus, the Deakin-Graeff theory postulates not only the apparently paradoxical "double

role" of serotonin on fear-like and anxiety-like behaviors in relation to different parts of the

neuraxis, but also the differential participation of serotonin receptors in these effects.

The 5-HT1A receptor (5-HT1AR) is one of the most abundant and widely expressed

receptors in the brain and the main inhibitory serotonergic receptor (Akimova et al., 2009;

Albert & Vahid-Ansari, 2019). This receptor is found in two populations, the presynaptic

autoreceptors (which promote local inhibitory control of neuronal firing rates and 5-HT

synthesis in serotonergic neurons), which are located in the cell body and dendrites of 5-HT

neurons; and postsynaptic heteroreceptors (which modulate the release of other

neurotransmitters in non-serotoninergic neurons), which are located in the dendrites and cell

body of non-serotoninergic neurons in 5-HT projection areas (Albert & Vahid-Ansari, 2019).

There is some evidence for a large receptor reserve for autoreceptors in opposition to

heteroreceptors (Cox et al., 1993; Meller et al., 1990), which could explain why azapirones,

partial agonists of the 5-HT1AR with anxiolytic-like activity in clinical settings (Chessick et

al., 2006), act as antagonists at heteroreceptor sites and full agonists in autoreceptors

(Newman-Tancredi et al., 2019). Low doses of 8-OH-DPAT, a full agonist, and buspirone, a

partial agonist, preferentially activate 5-HT1A autoreceptors, and consequently, the release of

5-HT from serotonergic nerve endings decreases. Therefore, 5-HT synthesis is reduced as a

feedback mechanism. From a systemic point of view, therefore, the effect of buspirone leads

to inhibition of the activity of serotonergic neurons (and consequent decrease in the release of

5-HT) associated with the blockade of 5-HT1A postsynaptic receptors. Full agonists, such as

8-OH-DPAT, appear to act as full agonists on both autoreceptors and heteroreceptors

(Sprouse & Aghajanian, 1988), decreasing the availability of 5-HT in other receptor subtypes

at the same time that post-synaptic 5-HT1ARs are activated. This discrepancy could explain

why both partial agonists and antagonists of the 5-HT1AR decrease defensive behavior to

potential threat (Cao & Rodgers, 1997; Maximino et al., 2013; Rodgers & Cao, 1997). The

putative differential effects of drugs on auto- and heteroreceptors lead to the necessity of

understanding the roles of these different sites in the control of defensive behavior. Studies

using mice lacking 5-HT1A auto- or heteroreceptors (Richardson-Jones et al., 2011), 5-

HT1AR knockouts with expression rescued in pre- or post-synaptic sites (Gross et al., 2002;

Piszczek et al., 2013), intact animals treated with biased agonists that activate pre- or

postsynaptic 5-HT1ARs *in vivo* (Jastrzębska-Więsek et al., 2018), or microinjection of

agonists in the raphe or in postsynaptic sites (File et al., 1996; File & Gonzalez, 1996),

produced conflicting results.

The roles of 5-HT1ARs were also investigated in non-mammalian model organisms,

including zebrafish (*Danio rerio* Hamilton 1822). This model organism has increasingly been

used in neuroscience and behavioral pharmacology to understand the mechanisms of

defensive behavior (Cueto-Escobedo et al., 2022; Gerlai, 2020, 2023; MacRae & Peterson,

2023). While there is considerable conservation of the serotonergic system is in zebrafish,

there are important differences in the degree of neuroanatomical and genomic conservation

(Herculano & Maximino, 2014). For example, zebrafish present extra serotonergic nuclei in

the brain in addition to the raphe (Kaslin & Panula, 2001; Lillesaar et al., 2009), and a

genomic duplication event led to multiple copies of genes involved in serotonin synthesis,

transport, and metabolism, as well as of serotonin receptors (including two copies of the 5-

HT1AR) (Norton et al., 2008; Sourbron et al., 2016). The two copies of the 5-HT1AR-coding

gene, *htr1aa* and *htr1ab,* are expressed in the adult zebrafish brain in a complementary

manner, with high expression of the *htr1aa* isoform as an autoreceptor in the pretectal

diencephalic cluster, superior raphe nucleus, and paraventricular organ, while the *htr1ab*

isoform is expressed as an autoreceptor in the caudal zone of the periventricular

hypothalamus, paraventricular organ, superior raphe, and pretectal cluster (Norton et al.,

2008). As an heteroreceptor, *htr1aa* is found in the ventral nucleus of the ventral

telencephalic area (Vv, homologous to the septal nuclei), preoptic region (PPa),

periventricular gray zone of the optic tectum (PGZ), posterior tuberculum, central posterior

thalamic nucleus, paraventricular organ, and ventral zone of the periventricular hypothalamus

(Hv), while *htr1ab* is found in central gray (homologous to the periaqueductal gray area), Hv,

zona limitans, PGZ, and PPa (Norton et al., 2008). Zebrafish exposed to the phototaxis assay

(light-dark test), a commonly used test for anxiety-like behavior in this species, show

increased *cfos* expression in regions with high density of 5-HT1AR isoforms, such as Vv and

Hv (Lau et al., 2011).

Despite the differences in genes and expression of 5-HT1ARs in zebrafish and

mammals, the role of the serotonergic system appears to be relatively conserved in zebrafish

(Herculano & Maximino, 2014). Some evidence suggests a role for 5-HT1A receptors in

anxiety-like behavior in zebrafish; for example, buspirone produces consistent type-

anxiolytic effects in behavioral assays with zebrafish, regardless of the route of

administration, dose, or test (Abozaid & Gerlai, 2022; Bencan et al., 2009; Connors et al.,

2014; Facchin et al., 2015; Gebauer et al., 2011; Hawkey et al., 2021; Lau et al., 2011;

Maaswinkel et al., 2012, 2013; Maximino et al., 2011, 2014). Maximino et al. (2013)

evaluated behavior characterized by scototaxis (preference for dark environments over light

environments) and geotaxis (the measurement of time at the top or bottom of the apparatus).

They observed anxiety-like behavior correlated positively with extracellular levels of 5-HT in

the brain in the novel tank test (NTT), while the correlation was negative in the phototaxis

assay. This suggests that 5-HT may regulate zebrafish behavior differently in the NTT and

phototaxis assay. Nonetheless, buspirone produced an anxiolytic-like effect in both the NTT

and the phototaxis assay. Buspirone also has been shown time spent near a conspecific in a

social investigation test, but not in the social novelty test (Barba-Escobedo & Gould, 2012),

which could either represent an anxiolytic-like effect or increased social motivation.

Puzzlingly, WAY 100635, a full 5-HT1A receptor antagonist, also produced anxiolytic-like

effects in both the NTT and the phototaxis assay (Maximino et al., 2013). Nowicki et al.

(2014), on the other hand, observed an anxiogenic-like effect on the NTT after treatment with

p-MPPF, another 5-HT$_{1A}$ receptor antagonist. On the other hand, treatment of zebrafish with

WAY 100,635 increased responsiveness to an olfactory-evoked fear stimulus (conspecific

alarm substance) in this species (Nathan et al., 2015), suggesting opponent roles of this

receptor in responses to potential vs. actual threat (anxiety vs. fear) in this species.

The similar effects of buspirone, a partial agonist, and WAY 100,635, a full

antagonist, on zebrafish anxiety-like behavior suggest the existence of a receptor reserve in

presynaptic sites. Furthermore, there are no studies with full agonists in this species. Thus,

the aim of the present study is to evaluate the effect of treatment with 8-OH-DPAT, a full

agonist of the 5-HT1A receptor, on behaviors relevant to anxiety and sociability in zebrafish.

8-OH-DPAT is a full agonist with high affinity for the 5-HT1A receptor (pKi for human

receptors between 8.4 and 9.4;

https://www.guidetopharmacology.org/GRAC/LigandDisplayForward?tab=biology&ligandId=7),

although it has been reported to act as a full agonist at 5-HT7 receptors as well (pKi for rat

receptors between 7.3 and 7.5;

https://www.guidetopharmacology.org/GRAC/LigandDisplayForward?tab=biology&ligandId=7).

We hypothesized that treatment with 8-OH-DPAT would be anxiogenic, and predicted that

this treatment would reduce time on white and increase risk assessment in the phototaxis

assay, increase time on bottom and erratic swimming in the NTT, and decrease preference

scores in both the social investigation and social novelty tests. Moreover, given the evidence

for a receptor reserve in mammals (Cox et al., 1993; Meller et al., 1990), we hypothesized

that pre-treatment with the antagonist WAY 100,635 would displace the dose-response curve

rightward.

2. Materials and methods

2.1 Animals and housing

96 adult zebrafish (*Danio rerio*) (>4 months old) from the shortfin phenotype were

used in the experiments. Adult wild-caught zebrafish (4-6 months old) of both sexes from the

strain were obtained from a colony resulting from cross-breeding between animals purchased

in specialized stores (Power Fish Fisheries, Itaguaí/RJ, Brazil). Based on body morphology

(general body shape and presence of genital papilla in females (Parichy et al., 2009), a

male:female ratio of approximately 40%:60% was estimated; sex of samples was ultimately

established after animals were sacrificed, using histological analysis of the gonads (Siegfried

& Steinfeld, 2021). All animals were kept in 50L tanks and acclimatized in the laboratory for

a period of 14 days before the onset of experiments. The tanks were kept constant at a

temperature of 26 ± 2ºC and pH 7.2, oxygenation, 14/10 light-dark cycle provided by

fluorescent lamps, according to standards of care for zebrafish (Lawrence, 2007). Animals

were fed twice a day with commercial flocked feed. Animal density adopted was in

accordance with the protocol adopted in the Brazilian guideline for the care of zebrafish in

the laboratory (Conselho Nacional de Controle de Experimentação Animal - CONCEA,

2017). The project was carried out with the approval of the Animal Use Ethics Committee of

UNIFESSPA (Universidade Federal do Sul e do Sudeste do Pará) protocol number

23479.019576/2022-43.

Sample sizes were calculated based on a model for independent-samples t-tests,

considering an effect size of $d = 1.35$ (based on a metanalysis of the effects of buspirone on

behavioral   tests   in   zebrafish;

https://github.com/lanec-unifesspa/5HT/blob/main/5HT1A/zf_metanalysis_buspirone.pdf), a

power of 0.8, and $\alpha = 0.05$. In these conditions, a minimum sample size of $n = 10$

animals/group was projected; ultimately, 12 animals per group were used in each experiment.

Calculations   were   made   using   the   R   package   pwr   (v.   1.3-0;

https://cran.r-project.org/web/packages/pwr/index.html).

2.2 Drugs and treatments

8-OH-DPAT (8-hydroxy-2-(dipropylamino)tetralin; CAS#87394-87-4) and WAY

100,635 (*N*-[2-[4-(2-Methoxyphenyl)-1-piperazinyl]ethyl]-N-(2-

pyridyl)cyclohexanecarboxamide) were bought from Sigma-Aldrich (St Louis, USA) and

dissolved in Cortland's salt solution (NaCl 124.1 mM, KCl 5.1 mM, Na2HPO4 2.9 Mm,

MgSO4 1.9 mM, CaCl2 1.4 mM, NaHCO3 11.9 mM, 1000 units heparin; Perry et al., 1984).

For all treatments, the solutions were injected intraperitoneally into cold-anesthetized, and

once anesthetized, they were positioned on a sponge-based surgical bed for injection

according to the method described by Kinkel et al. (2010). For both treatments, the injection

was carried out using a microsyringe (Hamilton 701N, 26 gauge needle, 200 conical tip, 10

μL), with a dose proportional to the animal's body weight (0.3 mg/kg) and an injection

volume of 5 μL.

Experimenters were blind to treatment through the use of coded vials; only the

principal investigator had access to the codes before data analysis. Treatment order was

randomized within sessions by using a random number generator

(https://www.randomizer.org/).

2.3. Experiment 1: Novel tank test

In general, this task aims to evaluate zebrafish vertical locomotion/exploration as an

index of defensive responses related to the response to novelty (Bencan et al., 2009; Egan et

al., 2009; Levin et al., 2007). Behavioral assessment of the animals was carried out according

to the methodology previously described in works by the research group

(https://dx.doi.org/10.17504/protocols.io.bp2l6x45klqe/v1), in which parameters related to

locomotion and the vertical position occupied by the animal in the tank (15 cm x 25 cm x 20

cm; width, length, height) were evaluated. Dechlorinated water was used for both the

recovery tank and the test tank. 12 adult zebrafish were used per group; the same 24 animals

were also used in Experiment 2. Immediately after the intraperitoneal injection, the fish were

relocated with a fish net to a recovery tank, where they remained for 30 minutes. Then, using

the fish net, the fish was transferred to the test tank, where it remained for the 6-min

habituation period, during which time the fish's behavior was recorded, generating a spatio-

temporal exploratory profile pattern. The behavioral endpoints were transcribed using The

Real   Fish   Tracker   software   (v.   0.4.0;

https://www.dgp.toronto.edu/~mccrae/projects/FishTracker/). Table 1 summarizes the

endpoints assessed in Experiment 1.

Table 1

Behavioral endpoints assessed in the novel tank test (NTT), phototaxis assay, and social

preference.

		ZBC code		
Test	**Behavior**	(Kalueff et al., 2013)	**Description**	**Operationalization**

NTT				
	Geotaxis (main endpoint)	1.46	Preference for the bottom of the tank, avoidance of the top	Time spent in the bottom third of the tank (s)
	Top-dwelling	1.176	Dwelling of the animal near the surface of a novel tank; can include surfacing (ZBC1.164), with aberrant swimming very close to the water surface	Time spent in the top third of the tank (s)
	Erratic swimming	1.51	Complex behavior in which the swim has no defined direction and the speed changes abruptly	Absolute turn angle (º)

	Hypolocomotion	1.81	Abnormally slow swimming for a sustained period of time, typically related to sedation or neuromotor deficits	Swimming speed (cm/s)
	Hyperlocomotion	1.79	Abnormally fast swimming for a sustained period of time, typically related to psychostimulant action	Swimming speed (cm/s)
Phototaxis assay	Scototaxis (main endpoint)	1.137	Preference for the dark part of the tank	Time spent in the black compartment (s)
	Risk assessment	n/a	Cautious exploration of the environment to investigate possible threats	Partial entries (up to half of body length) or full entries of low duration (< 1 s)

Erratic swimming	1.51	Complex behavior in which the swim has no defined direction and the speed changes abruptly	Absolute turn angle (º)
Freezing	1.68	Complete cessation of movement (except gills and eyes)	Total time swimming at speeds < 0.5 cm/s
Thigmotaxis	1.173	Preference for tank walls	Percent of time, while in the white compartment, spent at a distance of less than 2 cm of a tank wall
Hypolocomotion	1.81	Abnormally slow swimming for a sustained period of time, typically related to sedation or neuromotor deficits	Swimming speed (cm/s)

	Hyperlocomotion	1.79	Abnormally fast swimming for a sustained period of time, typically related to psychostimulant action	Swimming speed (cm/s)
Social preference test	Social approach	1.152	Tendency to spend time close to or inspecting conspecifics	Preference score (time near the conspecific (SI) or "new stranger" (SN) / time near the empty tank + time near the conspecifi)
	Social avoidance	n/a	Tendency to avoid novel conspecifics	Time spent near empty tank (SI) or "old stranger" (SN)
	Erratic swimming	1.51	Complex behavior in which the swim has no defined direction and the speed changes abruptly	Absolute turn angle (º)

Hypolocomotion	1.81	Abnormally slow swimming for a sustained period of time, typically related to sedation or neuromotor deficits	Swimming speed (cm/s)
Hyperlocomotion	1.79	Abnormally fast swimming for a sustained period of time, typically related to psychostimulant action	Swimming speed (cm/s)

2.4. Experiment 2: Phototaxis assay

Light/dark preference in zebrafish, characterized by a preference for either dark or

light environments, has been described and validated as a model for evaluating anxiety-like

behavior for the species (Araujo et al., 2012; Maximino et al., 2010, 2011; Serra et al., 1999).

Preference for either light or dark environments have been described in the literature,

depending on developmental stage (e.g., more preference for light environments in larvae and

juvenile zebrafish; Chen et al., 2015; Steenbergen et al., 2011) and light levels (e.g., more

preference for dark/black environments under bright light; Araujo et al., 2012; Blaser &

Peñalosa, 2011; Facciol et al., 2017, 2019). In both cases, anxiolytic drugs, including

buspirone, have been shown to reduce preference (Maximino et al., 2011; Steenbergen et al.,

2011). 12 zebrafish were used in each group. Since animals were reused in Experiments 1

and 2, the order in which animals were subjected to either the NTT or the phototaxis assay

was randomized. Immediately after the intraperitoneal injection, the fish was relocated with

an fish net to a recovery tank, where it remained for 30 min. Then, using the fish net, the fish

was transferred to an apparatus (15 cm x 10 cm x 45 cm; height, width, length) divided into a

white compartment and a black compartment, with hatches in the middle where the fish is

placed for acclimatization for 3 min. After this, the hatches were removed and the fish was

free to explore the apparatus for 15 min. The protocol established in the research group

(https://dx.doi.org/10.17504/protocols.io.puydnxw) was used. Scototaxis, swimming speed,

erratic swimming, and freezing were extracted using TheRealFishTracker (v. 0.4.0;

https://www.dgp.toronto.edu/~mccrae/projects/FishTracker/), while risk assessment and

thigmotaxis were scored manually with the help of X-Plo-Rat (Tejada et al., 2018;

https://sites.ffclrp.usp.br/scotty/). Table 1 summarizes the endpoints assessed in Experiment

2.

2.5. Experimental 3: Social preference test

The social preference test was divided in two stages, social interaction and social

novelty tests, and carried out based on the protocol described by Barba-Escobedo and Gould

(2012). These tasks aim to analyze the tendency of zebrafish to approach co-specifics and

investigate novel social situations. 48 adult zebrafish were used as conspecific stimuli, while

24 adult zebrafish were used as focal fish in the test, where 12 animals were injected

intraperitoneally with the vehicle solution (Cortland's saline solution), and 12 animals with 8-

OH-DPAT. Animals from this experiment were not reused from Experiments 1 and 2, due to

the longer extension of the test; exposing animals to three behavioral tests would risk the

drug having been eliminated by the end of the third task. Conspecifics were drawn from

different tanks than the hometank of the focal fish, and therefore focal fish had no prior

contact with them. Immediately after intraperitoneal injection, the focal fish was relocated

using a fish net to a recovery tank, where it remained for 30 min. Then, using the fish net, the

focal fish is transferred to the test tank where it remains for 3 min for acclimatization. For

both the social interaction and social novelty tests, three tanks were used: the main tank (15

cm x 25 cm x 20 cm; width, length, height), for the focal animal, and two other tanks for the

conspecifics (15 cm x 17 cm x 16 cm; width, length, height). Both sides of the main tank

were separated from the lateral tanks with barriers to prevent visual access. Once the fish's

acclimatization period was over, the barriers were removed allowing the focal fish visual

access to the conspecifics. In the social interaction test (SI), the behavior of 1 focal animal

was evaluated for an initial period of 6 min, in which it will have visual access to an

unknown conspecific and an empty tank. After that time interval, barriers were lowered, a

new conspecific was transferred to the empty tank, and the barriers were again removed to

allow the focal fish visual access to the conspecifics. The behavior was observed again for 6

minutes, consisting in the social novelty test (SN). The conspecific that was present in the

former stage was labeled as "old stranger", while the conspecific that was introduced in the

empty tank was labeled as "new stranger". In both stages, the conspecific stimulus was a

female zebrafish, confirmed by body morphology and, later, gonadal histology; female

zebrafish stimuli are reported to produce stronger preferences than male stimuli (Ogi et al.,

2021). In both the social interaction and social novelty stages, behavior was filmed and later

analyzed using TheRealFishTracker software (v. 0.4.0;

https://www.dgp.toronto.edu/~mccrae/projects/FishTracker/); behavioral endpoints are

described in Table 1.

2.6. Experiment 4: Evidence for a receptor reserve

For Experiment 4, animals were first pre-treated with either vehicle or WAY 100,635

(0.001 mg/kg, a dose below the necessary to produce behavioral effects in adult zebrafish;

Maximino et al., 2013; Nathan et al., 2015), and then injected with 8-OH-DPAT at the

following doses: 0.03 mg/kg, 0.3 mg/kg, and 3 mg/kg. 30 min. after injection, animals were

subjected to the novel tank test, as described in Section 2.3, above.

2.7. Statistical analysis

Results obtained from the behavioral tests from Experiment 1-3 were analyzed using

unpaired t-tests or Wilcoxon rank sum test with continuity correction for variables which

violated the assumptions of parametric statistics. No data was removed from the dataset. For

Experiment 4, data were analyzed using two-way (8-OH-DPAT dose X WAY 100,635 dose)

analyses of variance. The data was represented as Gardner-Altman plots (Ho et al., 2019),

with individual datapoints plotted on the upper axes. On the lower axes, each effect size

(Cohen's d; Cohen, 1988) was plotted as a bootstrap sampling distribution. Bootstrap

resamples were taken using Efron's (1987) method; the confidence interval is bias-corrected

and accelerated. Plots were made using R 4.1.2, using the dabestr package (v. 2023.9.12;

https://github.com/ACCLAB/dabestr). Post-hoc power calculations were made using the pwr

package (v. 1.3-0; https://cran.r-project.org/web/packages/pwr/index.html) in R, and the

power of each analysis is reported. While sex was not included as a variable in the analysis,

as the study was not designed to understand sex differences in the effect of 8-OH-DPAT,

exploratory analyses were made on the data, following Joel and Fausto-Sterling (2016).

Potential differences between males and females were explored via interaction effect size

plots. Plots for these exploratory analyses were made using the esci package

(https://github.com/rcalinjageman/esci) in jamovi (v. 2.4; https://www.jamovi.org).

2.7. Open science practices

Hypotheses were not formally preregistered. Data packages and analysis scripts can

be   found   on   our   GitHub   repository

(https://github.com/lanec-unifesspa/5HT/tree/main/5HT1A).

332   **3. Results**

3.1. Novel tank test

8-OH-DPAT decreased geotaxis ($t_{[df = 19.484]}$ = 5.222, p < 0.001, power = 0.999; Figure

1A) and increased top-dwelling ($t_{[df = 16.386]}$ = -4.009, p < 0.001, power = 0.97; Figure 1B). No

effects were observed on erratic swimming ($t_{[df = 21.998]}$ = -0.159, p = 0.875, power = 0.05;

Figure 1C), freezing ($t_{[df = 17.214]}$ = -1.555, p = 0.138, power = 0.31; Figure 1D), or swimming

speed ($t_{[df = 20.107]}$ = 0.971, p = 0.343, power = 0.16; Figure 1E). For all variables, 95%

confidence intervals for the effect sizes for sex X group interactions overlapped zero

(Supplemental File S1); however, plots suggest higher geotaxis and lower top-dwelling in

males, and larger effects of 8-OH-DPAT in females vs. male zebrafish in geotaxis and top-

dwelling.

Figure 1 – 8-OH-DPAT decreases anxiety-like behavior in the zebrafish novel tank test. (A)
Geotaxis. (B) Top-dwelling. (C) Erratic swimming. (D) Freezing. (E) Swimming speed. For a
description of variables, refer to Table 1. Blue dots represent animals treated with vehicle (CTRL),
and orange dots represent animals treated with 8-OH-DPAT (DPAT). The Cohen's *d* between CTRL
and DPAT is shown in Gardner-Altman estimation plot. Both groups are plotted on the left axes; the
effect size (Cohen's *d*) is plotted on floating axes on the right as a bootstrap sampling distribution.
The mean difference is depicted as a dot; the 95% confidence interval is indicated by the ends of the
vertical error bar. 5000 bootstrap samples were taken; the confidence interval is bias-corrected and
accelerated.

3.2. Phototaxis assay

8-OH-DPAT increased scototaxis ($t_{[df = 17.361]}$ = 2.43, p = 0.262, power = 0.64; Figure

2A) and increased risk assessment (W = 33, p = 0.024, power = 0.6; Figure 2B). No effect

were found for erratic swimming ($t_{[df = 20.671]}$ = 0.727, p = 0.475, power = 0.11; Figure 2C),

freezing (W = 71.5, p = 0.99, power = 0.06; Figure 2D), thigmotaxis ($t_{[df = 14.873]}$ = -1.323, p =

0.206, power = 0.24; Figure 2E), or swimming speed ($t_{[df = 21.985]}$ = 1.71, p = 0.101, power =

0.37; Figure 2F). For all variables, 95% confidence intervals for the effect sizes for sex X

group interactions overlapped zero (Supplemental File S2); however, plots suggest higher

scototaxis in females, and larger effects of 8-OH-DPAT in males vs. female zebrafish in this

variable; as well as higher thigmotaxis in males, with a small-sized interaction effect in which

8-OH-DPAT increases thigmotaxis in males but not females.

Figure 2 – 8-OH-DPAT increases anxiey-like behavior in the zebrafish light/dark test.

(A) Scototaxis. (B) Risk assessment. (C) Erratic swimming. (D) Freezing. (E) Thigmotaxis.

(F) Swimming speed. For a description of variables, refer to Table 1. Blue dots represent

animals treated with vehicle (CTRL), and orange dots represent animals treated with 8-OH-

DPAT (DPAT). The Cohen's *d* between CTRL and DPAT is shown in Gardner-Altman

estimation plot. Both groups are plotted on the left axes; the effect size (Cohen's *d*) is plotted

on floating axes on the right as a bootstrap sampling distribution. The mean difference is

depicted as a dot; the 95% confidence interval is indicated by the ends of the vertical error

bar. 5000 bootstrap samples were taken; the confidence interval is bias-corrected and

accelerated.

3.3. Social preference test

During the SI phase, 8-OH-DPAT decreased the preference score ($t_{[df = 13.466]}$ = 2.981, p

= 0.0103, power = 0.81; Figure 3A), with no effects on social avoidance ($t_{[df=21.594]}$ = 0.881, p

= 0.388, power = 0.13; Figure 3B). No effects were observed on erratic swimming ($t_{[df = 21.392]}$

= 0.548, p = 0.589, power = 0.08; Figure 3C), nor on swimming speed ($t_{[df = 19.036]}$ = 1.1803, p

= 0.252, power = 0.2; Figure 3D). For all variables, 95% confidence intervals for the effect

sizes for sex X group interactions overlapped zero (Supplemental File S3); however, plots

suggest no differences in preference score between males and females, and larger effects of

8-OH-DPAT in females vs. male zebrafish in this variable; likewise, no differences are

suggested in social avoidance, but larger effects of 8-OH-DPAT in males than females are

suggested. Plots also suggest that females show higher erratic swimming than males, and 8-

OH-DPAT appears to reduce erratic swimming in males but not females in the SI phase.

Figure 3 – 8-OH-DPAT decreases social approach in the social investigation test. (A)

Preference score. (B) Social avoidance. (C) Erratic swimming. (D) Swimming speed. For a

description of variables, refer to Table 1. Blue dots represent animals treated with vehicle

(CTRL), and orange dots represent animals treated with 8-OH-DPAT (DPAT). The Cohen's *d*

between CTRL and DPAT is shown in Gardner-Altman estimation plot. Both groups are

plotted on the left axes; the effect size (Cohen's *d*) is plotted on floating axes on the right as a

bootstrap sampling distribution. The mean difference is depicted as a dot; the 95% confidence

interval is indicated by the ends of the vertical error bar. 5000 bootstrap samples were taken;

the confidence interval is bias-corrected and accelerated.

During the SN phase 8-OH-DPAT also decreased the preference score ($t_{[df = 21.847]}$ =

2.667, p = 0.014, power = 0.72; Figure 4A), but the time near the "new stranger" was also

decreased ($t_{[df = 21.767]}$ = 2.701, p = 0.013, power = 0.73; Figure 4B). No effect was observed on

erratic swimming ($t_{[df = 21.785]}$ = -0.7101, p = 0.485, power = 0.1; Figure 4C), nor on swimming

speed ($t_{[df = 21.797]}$ = -1.359, p = 0.188, power = 0.25; Figure 4D). For all variables, 95%

confidence intervals for the effect sizes for sex X group interactions overlapped zero

(Supplemental File S4); again, plots suggest no differences in preference score between

males and females, and larger effects of 8-OH-DPAT in males vs. female zebrafish in this

variable; likewise, no differences are suggested in social avoidance, but larger effects of 8-

OH-DPAT in males than females are suggested. Plots also suggest that females show higher

erratic swimming than males, and 8-OH-DPAT appears to reduce erratic swimming in

females but not males in the SN phase.

Figure 4 – 8-OH-DPAT decreases social preference in the social novelty test. (A)
Preference score. (B) Social avoidance. (C) Erratic swimming. (D) Swimming speed. For a
description of variables, refer to Table 1. Blue dots represent animals treated with vehicle
(CTRL), and orange dots represent animals treated with 8-OH-DPAT (DPAT). The Cohen's *d*
between CTRL and DPAT is shown in Gardner-Altman estimation plot. Both groups are
plotted on the left axes; the effect size (Cohen's *d*) is plotted on floating axes on the right as a
bootstrap sampling distribution. The mean difference is depicted as a dot; the 95% confidence
interval is indicated by the ends of the vertical error bar. 5000 bootstrap samples were taken;
the confidence interval is bias-corrected and accelerated.

3.4. Effects of 8-OH-DPAT after WAY 100,635 pre-treatment

Main effects of 8-OH-DPAT dose ($F_{[3, 72]} = 111.55$, p < 0.001, $\omega^2 = 0.68$, power =
0.99) and WAY 100,635 dose ($F_{[1, 72]} = 32.73$, < 0.0001, $\omega^2 = 0.06$, power = 0.46) were found

for geotaxis. Importantly, an interaction effect was also found ($F_{[3, 72]}$ = 16.71, p < 0.001, ω^2 =

0.1, power = 0.15; Figure 5). Post-hoc tests found that, when animals were not pre-treated

with WAY 100,635, 8-OH-DPAT decreased geotaxis at all doses (p < 0.05, *d* between 1.42

and 5.25); however, in animals that were pre-treated with WAY 100,635, 8-OH-DPAT

produced a significant reduction only at the highest dose (p < 0.001, *d* = 5.17).

Figure 5 – The Hedges' *g* for 7 comparisons against the shared control DPAT0+WAY0 are

shown in a Cumming estimation plot. The raw data is plotted on the upper axes. On the lower

axes, mean differences are plotted as bootstrap sampling distributions. Each mean difference

is depicted as a dot. Each 95% confidence interval is indicated by the ends of the vertical

error bars.

4. Discussion

In the present experiments, differences were observed in anxiety-like behavioral

endpoints in the groups treated with the full agonist of the 5-HT1A receptor, 8-OH-DPAT,

with opposite outcomes in the novel tank test (NTT) and phototaxis assay (light-dark test).

While in the first 8-OH-DPAT decreased anxiety-like behavior, in the latter the drug

increased it. When 8-OH-DPAT was injected in animals pre-treated with WAY 100,635, a

rightward shift in the dose-response curve was found in the NTT. We also observed decreases

in social preference in both the social interaction (SI) and social novelty (SN) tests.

4.1. Effects of 8-OH-DPAT on anxiety tests

One of the most striking findings of this study is the difference in the effects of 8-OH-

DPAT between the phototaxis assay and the NTT. In previous studies with serotonergic drugs

in these two tests, it was shown that increasing 5-HT levels with acute fluoxetine increases

geotaxis (the main endpoint of NTT) and decreases scototaxis (the main endpoint of the

phototaxis assay), while decreasing 5-HT levels with pCPA produces the opposite effects

(Maximino et al., 2013). On the other hand, both buspirone (partial 5-HT1AR agonist) and

WAY 100,635 (full 5-HT1AR antagonist) decreased geotaxis and scototaxis (Maximino et

al., 2013). SB 224,289, an inverse agonist of the 5-HT1B receptor, reduced geotaxis but not

scototaxis, an effect similar to that observed in the present experiment. Unlike buspirone,

which produces consistent reductions in responses to potential threat in both the phototaxis

assay and the NTT (Abozaid & Gerlai, 2022; Araujo et al., 2012; Bencan et al., 2009;

Connors et al., 2014; Facchin et al., 2015; Gebauer et al., 2011; Hawkey et al., 2021; Lau et

al., 2011; Maaswinkel et al., 2012, 2013; Maximino et al., 2011, 2013, 2014; Steenbergen et

al., 2011), 8-OH-DPAT seemed to produce opposite effects in the NTT and phototaxis assay.

Despite being 5-HT1A receptor agonists, there are important differences between buspirone

and 8-OH-DPAT. In a context of large receptor reserves in presynatic neurons, as is the case

with the rodent raphe, buspirone acts as a full agonist at the 5-HT1A autoreceptor, while it

acts as an antagonist at the postsynaptic 5-HT1AR, reducing serotonin avaiability at other

postynapic sites while simultaneously blocking postsynaptic 5-HT1ARs (Newman-Tancredi

et al., 2019). Meanwhile, 8-OH-DPAT is a serotonergic drug that acts as a full agonist at both

sites, reducing serotonin availability for other types of receptor in the postsynaptic sites while

simultaneously activating postsynaptic 5-HT1ARs. It is therefore possible to assume that this

is also the reason why the results differed between the tests.

To confirm the presence of a receptor reserve related to the effects of 8-OH-DPAT,

animals were pre-treated with the antagonist WAY 100,635 and treated with increasing doses

of 8-OH-DPAT before being subjected to the NTT. A rightward shift in the dose-response

curve was observed, with higher doses needed to produce an anxiolytic-like effect in the NTT

when receptors were blocked. When rats are treated with 8-OH-DPAT and the irreversible 5-

HT1A antagonist EEDQ, a rightward shift is observed in the production of 5-HTP, the

serotonin precursor, in the cortex and hippocampus (Meller et al., 1990). Likewise, Cox et al.

(1993) showed that EEDQ induced a rightward shift in the dose-response curve for the

inhibition of dorsal raphe nucleus cell firing by 8-OH-DPAT. Although these studies were the

first to show evidence of a receptor reserve for neurophysiological endpoints of serotonergic

function, other studies found mixed evidence for systemic endpoints. For example, Meller

and Bohmaker (1994) found that EEDQ induces a rightward shift in the attenuation of

plasmatic release of corticosterone and ACTH in rats. However, no evidence for receptor

reserve was found for 5-HT1A receptor-mediated hypothermia in mice (Meller et al., 1992).

Overall, these results are consistent with the hypothesis that, at least for behavior in the NTT,

a receptor reserve is present.

A related question is why 8-OH-DPAT reduced behavioral responses to potential

threat in the NTT and increased it in the LDT. One possibility is that NTT is more sensitive

to pharmacological treatment than LDT. However, in a meta-analysis of the effects of drugs

on the two tests, Kysil et al. (2017) found no differences in the sensitivity of both assays. A

second possibility is that the effect observed on the NTT is not related to a decrease in

responsiveness to potential threat, but represents either serotonin toxicity (Stewart et al.,

2013) or a psychedelic-like effect (Kyzar & Kalueff, 2016). While 8-OH-DPAT increased

top-dwelling in the NTT, which could be interpreted as "surfacing" (an aberrant behavioral

pattern of slow swimming close to the water surface that is typically observed under the

effects of serotonergic hallucinogens; Kalueff et al., 2013; Kyzar & Kalueff, 2016), animals

treated with the drug still spent most of the time in the bottom of the tank, differently from

zebrafish treated with psychedelics. Moreover, psychedelics typically produce an

hypolocomotion phenotype in zebrafish, and changes in the swimming speed were not

observed in any test in the present experiments. A third possibility is that 8-OH-DPAT is

acting on either 5-HT1A and 5-HT7 receptors, producing different behavioral profiles

depending on the receptor. While the affinity of this drug for zebrafish receptors has not been

determined, it could be argued that the opponent effects of 8-OH-DPAT on the NTT and the

phototaxis assay are due to effects on these receptors. The scant amount of behavioral studies

with 8-OH-DPAT investigating these receptors make it difficult to judge that interpretation.

In rodents, 8-OH-DPAT induces hypothermia by acting both on the 5-HT1A and the 5-HT7

receptors (Hedlund et al., 2004). However, 5-HT7R knockout mice showed no differences in

baseline anxiety (Guscott et al., 2005), suggesting that this receptor is involved in other

behavioral functions, at least in mammals. To fully confirm this hypothesis, more

experiments are needed.

An alternative explanation is that the two tests model different defensive processes,

with opposite modulation of the serotonergic system on both. Blaser et al. (2012) analyzed

stimulus control in the two tests, and suggested that geotaxis is mainly controlled by escape

from the top (closer, therefore, to fear), while scototaxis is mainly controlled by approach-

avoidance conflict (closer, therefore, to anxiety). Thus, the effects of 8-OH-DPAT in the

NTT and phototaxis assay could be interpreted as decreasing fear and increasing anxiety,

respectively. In support of this hypothesis, microinjection of 8-OH-DPAT in forebrain

regions such as the hippocampus (Cheeta et al., 2000; File et al., 1996), septum (Cheeta et al.,

2000; Martins De Almeida et al., 1998), and amygdala (Nunes-de-Souza et al., 2000) has

been found to increase anxiety-like behavior in rodents. Conversely, microinjection of 8-OH-

DPAT in the periaqueductal gray (Zangrossi Jr. et al., 2001; Zanoveli et al., 2003, 2010) and

dorsomedial hypothalamus (Nascimento et al., 2014), but not amygdala (Zangrossi et al.,

1999), impair escape in the elevated T-maze, an effect consistent with decreased fear-like

states. In zebrafish, treatment with the 5-HT1A antagonist WAY 100,635 increases

responsiveness to fear-eliciting olfactory stimulus (alarm substance), implicating this receptor

in reducing responses to actual threats (Nathan et al., 2015). Taken together, these results

suggest that the differential effects of 8-OH-DPAT on the NTT and phototaxis assay mediate

different behavioral processes via 5-HT1A heteroreceptors. Experiments using 8-OH-DPAT

to assess its effects on alarm substance-elicited behavioral responses could test this

hypothesis. Considering the participation of the septal homologue Vv and the hypothalamic

Hv in scototaxis (Lau et al., 2011), and considering that both 5-HT1AR isoforms are found in

the latter while only *htr1aa* is found in Vv (Norton et al., 2008), is is likely that these nuclei

are involved in the effects of 8-OH-DPAT.

A last possibility that cannot be discarded is that 8-OH-DPAT acts on receptors

involved with locomotion, either at the hindbrain or spinal cord levels; indeed, *htr1aa* has

been found to be expressed in hindbrain motor circuits (Norton et al., 2008). In zebrafish

larvae, treatment with methysergide, a non-selective agonist of 5-HT1 receptors and

antagonist at 5-HT2 receptors, decreases motor output by prolonging periods of inactivity

(Brustein et al., 2003). Thus – especially in the NTT, in which the main endpoint involves the

ability to swim upwards in the water column –, this motor-inhibitory effect of 5-HT1A

receptor activation could be responsible for the observed effects. While the lack of effects on

swimming speed on all tests weakens this hypothesis, post-hoc power analysis showed that

this variable had lower power than desired across assays, and therefore interpretations on

these effects should be taken with a grain of salt.

In the present experiments, no effects were observed in measures such as erratic

swimming, freezing, and thigmotaxis, which are common secondary endpoints in the NTT

and phototaxis assay. Previously, it has been shown that WAY 100,635 and buspirone

decreased freezing, swimming, and thigmotaxis in the phototaxis assay at higher doses, but

had no effect in these endpoints in the NTT (Maximino et al., 2013). Nowicki et al. (2014),

on the other hand, found that m-PPF, another 5-HT1AR antagonist, increased erratic

swimming at low doses in the NTT. Post-hoc power analyses also showed that these variables

had lower power than desired, suggesting that the lack of effects in the present experiments in

these variables are likely to be false negatives.

4.2. Effects of 8-OH-DPAT on social approach

570   Different phases of the social preference tests used in the present experiment

emphasize the role of social interaction vs. social novelty. Our results with 8-OH-DPAT

showed a decrease in SI and SN, suggesting a broader effect on social approach. Barba-

Escobedo and Gould (2012) showed that buspirone increases social approach in the SI test

(the opposite of what was observed here with 8-OH-DPAT), with no effect on the SN test.

Whether this difference indicates that, as in the case with anxiety-like behavior, the effects of

8-OH-DPAT are likely to be mediated by heteroreceptors is still opened. These results could

be interpreted either as an anxiogenic-like effect on social preference, or a specific effect on

sociality. These hypotheses are difficult to untangle, since, while no effects on erratic

swimming was observed in neither the SI or SN tests, they were also absent in the NTT or

phototaxis assay. While social approach is sensitive to acute stress in adult zebrafish (Cook et

al., 2023), the fact that, differently from lab rodents, zebrafish do not avoid novel

conspecifics, and actively spend time investigating them when possible (Barba-Escobedo &

Gould, 2012; do Nascimento & Maximino, 2023; Ogi et al., 2021), makes it difficult to

understand how threat factors in sociability in this species. The activation of 5-HT2 receptors

has been shown to increase sociability in zebrafish (de Moura et al., 2023; Ponzoni et al.,

2016), and single-nucleotide polymorphisms of the *htr1aa* gene (which code for one isoform

of the 5-HT1AR) mediate populational variability in social exploration and sociability in this

species (Gonçalves et al., 2022). These results further underscore the role of serotonin in

sociability in zebrafish, with opponent modulation by 5-HT1AR and 5-HT2CR agonists.

As was the case with the LDT and NTT, no effects were observed in erratic

swimming. This can suggest that the effects of 8-OH-DPAT are restricted to sociality, and

not an increase in anxiety-like behavior. However, as was the case with the LDT and NTT,

post-hoc power analysis showed lower power than desired, and therefore it is difficult to

understand whether or not the lack of effect in erratic swimming in the SI and SN tests are

false negatives.

4.3. Exploration of potential sex differences

While the current study was not designed to detect sex differences and interactions

between sex and drug effects, some suggestion of such effects were observed. In the NTT, a

small effect sizes of sex were observed in both geotaxis and top-dwelling, with females

displaying less geotaxis and more top-dwelling. While exploratory, these results contradict

what is usually reported in the literature (Genario et al., 2020). Moreover, moderate effect

sizes on the interaction are also suggested, with females showing a possibly higher effect of

8-OH-DPAT than males. In the phototaxis assay, effects appear to be opposite, with higher

anxiety-like behavior in females (small effect size) and a higher effect of 8-OH-DPAT in

males (moderate effect size). Thus, important sex effects could also be responsible for the

opposite directions of 8-OH-DPAT effects in the NTT and phototaxis assay; experiments

explicitly designed to test this hypothesis are needed. In the SI phase of the social preference

test, almost no difference is obserable between males and females, but larger effects of 8-OH-

DPAT in females vs. male zebrafish are suggested in social approach, as are larger effects of

8-OH-DPAT in males than females in social avoidance. Similar effects were observed in the

SN phase, but larger effects of 8-OH-DPAT in males than females are suggested. These

results are consistent with previous work which demonstrated that both males and females

show similar social preferences (Snekser & Diestler, 2023). We emphasize, again, that these

analyses are exploratory, and sample sizes were not planned to detect main or interaction

effects of sex. Thus, further experiments are necessary to confirm these hypotheses of sex

effects.

619 4.4. Limitations

One important limitation of the present study is that, although sample sizes were

higher than those calculated as the minimum to produce an effect size at least as large as that

of buspirone, post-hoc power analyses found that, for important secondary endpoints, power

was lower than desired. Due to IRB restrictions on increasing sample sizes after the study

was conducted, it was unfeasible to re-run the experiments to confirm whether 8-OH-DPAT

did not produce an effect on erratic swimming and freezing, or if the apparent lack of effect is

a false negative due to low power. Further experiments are necessary to untangle this

hypothesis.

Another important limitation relates to how to accurately interpret the results of the

social preference tests in zebrafish. In rats, decreases in social interaction is usually

interpreted as indicating an anxiogenic-like effect (File & Seth, 2003). In zebrafish, some

studies suggest that acute stress decreases social approach (Barcellos et al., 2020; Cook et al.,

2023); nonetheless, while buspirone increases social approach (Barba-Escobedo & Gould,

2012), non-serotonergic anxiolytics such as diazepam appear to decrease it in zebrafish

(Giacomini et al., 2016). Future work should also disentangle the participation of threat (and

therefore of anxiety-like states) and sociability in this test, allowing for a better

comprehension of the roles of the 5-HT1AR in it.

A third limitation of the present study is that it was not explicitly design to take into

account possible sex differences both in the behavioral parameters and in the drug effects.

There is reason to believe that there are differences in the serotonergic system of male and

female zebrafish, as a lower 5-HIAA:5-HT ratio was observed in the forebrain of females

(Dahlbom et al., 2012). Our exploratory analysis suggested that males and females respond

differently to 8-OH-DPAT, and therefore studies explicitly designed to address this

hypothesis are needed.

Finally, our study did not include an explicit dose-response analysis, as only one dose

of 8-OH-DPAT was used. Thus, it is possible that the dose used was not optimal for the

phototaxis assay and the social preference tests. While this could also explain the lack of

effects of 8-OH-DPAT on secondary endpoints, such as freezing and erratic swimming, it is

also important to point that dose-response curves under concomitant treatment with receptor

antagonists are commonly used to assess the presence of a receptor reserve (Cox et al., 1993;

Furchgott & Bursztyn, 1967; Meller et al., 1990). While dose-response curves in behavioral

pharmacology are usually different from the typical sigmoidal curve found elsewhere (e.g.,

lower doses of WAY 100,635 produce larger effects in the NTT and LDT than higher doses;

Maximino et al., 2013), it can be useful to perform such experiments in the future to better

understand whether a receptor reserve is responsible for the different effects of 8-OH-DPAT

and other 5-HT1AR binding drugs in these behavioral tests.

4.5. Translational implications

The present work shed light on a serotonergic mechanism involved in the opponent

regulation of responses to proximal threat in zebrafish. While substantial differences exist

between zebrafish and humans in terms of their serotonergic system (Herculano &

Maximino, 2014), the fact that 8-OH-DPAT appears to produce similar behavioral effects in

both zebrafish and rodents increase the possibility that the function of 5-HT1ARs is

conserved. Indeed, it has been previously proposed that using non-mammalian organisms in

behavioral neuroscience can facilitate a deeper understanding of brain function and

dysfunction at different levels (Gerlai, 2020; Stewart et al., 2014). If similar drug effects are

found in zebrafish, rodents, and humans, it is likely that those effects represent conserved

physiology instead of evolutionary homoplasies (Gerlai, 2020). In the context of molecular

heterogeneity and physiological conservation of the serotonergic system, the present results

point to the importance of interpreting data from behavioral pharmacology in a comparative

framework.

671   **5. Conclusions**

In the present experiments, the full agonist of the 5-HT1A receptor, 8-OH-DPAT,

decreased anxiety-like behavior in the novel tank test (NTT), but increased it in the light-dark

test (LDT), both considered outcomes of anxiety-like behavior for this species. The same

treatment decreased social approach in both the social investigation test and the social

novelty test. In comparison with previous data on the effects of the partial agonist buspirone

and the full antagonist WAY 100,635, and based on the evidence for a receptor reserve for

the anxiolytic-like effect of 8-OH-DPAT in the NTT, we suggest that 8-OH-DPAT increases

anxiety-like behavior mainly by activating heteroreceptors.

Acknowledgments

This work was supported by a Conselho Nacional de Desenvolvimento Científico e

Tecnológico (CNPq/Brazil) productivity grant to CM (#302998/2019-5). LVXBS an LNO

were supported by Fundação Coordenação de Aperfeiçoamento de Pessoal de Nível Superior

(CAPES/Brazil) scholarships. A GPT-3.5-based tool, SciSummary, was used to help create

the lay summary and the significance statement of this article, but no other AI-based tools

were used elsewhere. The graphical abstract for this manuscript was created with

BioRender.com

Conflict of interest statement

The authors declare that they have no conflicts of interest.

Authors' contributions

Loanne Valéria Xavier Bruce de Souza: Conceptualization, Methodology, Formal
Analysis, Investigation, Writing – Original draft, Writing – Review & editing, Visualization;
Larissa Nunes de Oliveira: Investigation, Writing – Original draft, Writing – Review &
editing; **Bruna Patrícia Dutra Costa:** Investigation, Writing – Original draft, Writing –
Review & editing; **Monica Lima-Maximino:** Conceptualization, Methodology, Writing –
Original draft, Writing – Review & editing; **Hellen Vivianni Veloso Corrêa:**
Conceptualization, Methodology, Writing – Original draft, Writing – Review & editing; **Caio**
Maximino: Conceptualization, Methodology, Validation, Formal Analysis, Resources, Data
Curation, Writing – Original draft, Writing – Review & editing, Supervision, Project
administration, Funding acquisition.

Data accessibility

References

Abozaid, A., & Gerlai, R. (2022). Behavioral Effects of Buspirone in Juvenile Zebrafish of Two
Different Genetic Backgrounds. *Toxics, 10,* 22. https://doi.org/10.3390/toxics10010022

Abrams, J. K., Johnson, P. L., Hollis, J. H., & Lowry, C. A. (2004). Anatomic and functional
topography of the dorsal raphe nucleus. *Annals of the New York Academy of Sciences, 1018,*
46–57. https://doi.org/10.1196/annals.1296.005

Abrams, J. K., Johnson, P. L., Mikkelsen, J. D., Shekhar, A., & Lowry, C. A. (2005). Serotonergic

systems associated with arousal and vigilance behaviors following administration of

anxiogenic drugs. *Neuroscience*, *133*, 983–997.

https://doi.org/10.1016/j.neuroscience.2005.03.025

Akimova, E., Lanzenberger, R., & Kasper, S. (2009). The Serotonin-1A Receptor in Anxiety

Disorders. *Biological Psychiatry*, *66*(7), 627–635.

https://doi.org/10.1016/j.biopsych.2009.03.012

Albert, P. R., & Vahid-Ansari, F. (2019). The 5-HT1A receptor: Signaling to behavior. *Biochimie*,

161, 34–45. https://doi.org/10.1016/j.biochi.2018.10.015

Araujo, J., Maximino, C., Brito, T. M. de, Silva, A. W. B. da, Batista, E. de J. O., Oliveira, K. R. M.,

Morato, S., Herculano, A. M., & Gouveia Jr., A. (2012). Behavioral and pharmacological

aspects of anxiety in the light/dark preference test. Em A. V. Kalueff & A. M. Stewart

(Orgs.), *Zebrafish Protocols for Neurobehavioral Research*. Humana Press.

https://doi.org/10.1007/978-1-61779-597-8_14

Barba-Escobedo, P. A., & Gould, G. G. (2012). Visual social preferences of lone zebrafish in a novel

environment: Strain and anxiolytic effects. *Genes, Brain and Behavior*, *11*, 366–373.

https://doi.org/10.1111/j.1601-183X.2012.00770.x

Barcellos, H. H. de A., Pompermaier, A., Mendonça-Soares, S., Maffi, V. C., Fernandes, M.,

Koakoski, G., Kirsten, K., Baldisserotto, B., & Barcellos, L. J. G. (2020). Aripiprazole

prevents stress-induced anxiety and social impairment, but impairs antipredatory behavior in

zebrafish. *Pharmacology Biochemistry and Behavior*, *189*, 172841.

https://doi.org/10.1016/j.pbb.2019.172841

Bencan, Z., Sledge, D., & Levin, E. D. (2009). Buspirone, chlordiazepoxide and diazepam effects in a

zebrafish model of anxiety. *Pharmacology Biochemistry & Behavior*, *94*(1), 75–80.

https://doi.org/10.1016/j.pbb.2009.07.009

Blaser, R. E., & Peñalosa, Y. M. (2011). Stimuli affecting zebrafish (*Danio rerio*) behavior in the light/dark preference test. *Physiology & Behavior, 104*(5), 831–837. https://doi.org/10.1016/j.physbeh.2011.07.029

Brustein, E., Chong, M., Holmqvist, B., & Drapeau, P. (2003). Serotonin patterns locomotor network activity in the developing zebrafish by modulating quiescent periods. *Journal of Neurobiology, 57*(3), 303–322. https://doi.org/10.1002/neu.10292

Cao, B.-J., & Rodgers, R. J. (1997). Influence of 5-HT1A receptor antagonism on plus-maze behaviour in mice. I. Pindolol enantiomers and pindobind 5-HT1A. *Pharmacology, Biochemistry and Behavior, 58*, 583–591.

Castilho, V., Macedo, C., & Brandão, M. (2002). Role of benzodiazepine and serotonergic mechanisms in conditioned freezing and antinociception using electrical stimulation of the dorsal periaqueductal gray as unconditioned stimulus in rats. *Psychopharmacology, 165*(1), 77–85. https://doi.org/10.1007/s00213-002-1246-4

Cheeta, S., Kenny, P. J., & File, S. E. (2000). Hippocampal and septal injections of nicotine and 8-OH-DPAT distinguish among different animal tests of anxiety. *Progress in Neuro-Psychopharmacology and Biological Psychiatry, 24*, 1053–1067. https://doi.org/10.1016/S0278-5846(00)00129-9

Chen, F., Chen, S., Liu, S., Zhang, C., & Peng, G. (2015). Effects of lorazepam and WAY-200070 in larval zebrafish light/dark choice test. *Neuropharmacology, 95*, 226–233. https://doi.org/10.1016/j.neuropharm.2015.03.022

Chessick, C. A., Allen, M. H., Thase, M. E., Cunha, A. A. B. M. da, Kapczinski, F., Lima, M. S. de, & Souza, J. J. dos S. (2006). Azapirones for generalized anxiety disorder. *Cochrane Database of Systematic Reviews*, CD006115. https://doi.org/10.1002/14651858.CD006115

Cohen, J. (1988). *Statistical power analysis for the behavioral sciences* (2nd ed). Routledge.

Connors, K. A., Valenti, T. W., Lawless, K., Sackerman, J., Onaivi, E. S., Brooks, B. W., & Gould, G. G. (2014). Similar anxiolytic effects of agonists targeting serotonin 5-HT1A or

cannabinoid CB receptors on zebrafish behavior in novel environments. *Aquatic Toxicology*, *151*, 105–113. https://doi.org/10.1016/j.aquatox.2013.12.005

Cook, A., Beckmann, H., Azap, R., & Ryu, S. (2023). Acute Stress Modulates Social Approach and Social Maintenance in Adult Zebrafish. *eNeuro*, *10*(9), ENEURO.0491-22.2023. https://doi.org/10.1523/ENEURO.0491-22.2023

Cox, R. F., Meller, E., & Waszczak, B. L. (1993). Electrophysiological evidence for a large receptor reserve for inhibition of dorsal raphe neuronal firing by 5-HT1A agonists. *Synapse*, *14*, 297–304. https://doi.org/10.1002/syn.890140407

Cueto-Escobedo, J., German-Ponciano, L. J., Guillén-Ruiz, G., Soria-Fregozo, C., & Herrera-Huerta, E. V. (2022). Zebrafish as a Useful Tool in the Research of Natural Products With Potential Anxiolytic Effects. *Frontiers in Behavioral Neuroscience*, *15*, 795285. https://doi.org/10.3389/fnbeh.2021.795285

Dahlbom, S. J., Backström, T., Lundstedt-Enkel, K., & Winberg, S. (2012). Aggression and monoamines: Effects of sex and social rank in zebrafish (Danio rerio). *Behavioural Brain Research*, *228*(2), 333–338. https://doi.org/10.1016/j.bbr.2011.12.011

de Moura, L. A., Pyterson, M. P., Pimentel, A. F. N., Araújo, F., de Souza, L. V. X. B., Mendes, C. H. M., Costa, B. P. D., de Siqueira-Silva, D. H., Lima-Maximino, M., & Maximino, C. (2023). Roles of the 5-HT2C receptor on zebrafish sociality. *Progress in Neuro-Psychopharmacology and Biological Psychiatry*, *125*, 110769. https://doi.org/10.1016/j.pnpbp.2023.110769

Deakin, J. F. W., & Graeff, F. G. (1991). 5-HT and mechanisms of defense. *Journal of Psychopharmacology*, *5*, 305–315.

Diretriz brasileira para o cuidado e a utilização de animais para fins científicos e didáticos - DBCA. Anexo I. Peixes mantidos em instalações de instituições de ensino ou pesquisa científica, Resolução Normativa CONCEA nº 34 218 (2017). https://www.mctic.gov.br/mctic/opencms/legislacao/outros_atos/resolucoes/ Resolucao_Normativa_CONCEA_n_34_de_27072017.html

do Nascimento, B. G., & Maximino, C. (2023). Social investigation and social novelty in zebrafish: Roles of salience and novelty. *Behavioural Processes, 210,* 104903. https://doi.org/10.1016/j.beproc.2023.104903

Efron, B. (1987). Better Bootstrap Confidence Intervals. *Journal of the American Statistical Association, 82*(397), 171–185. https://doi.org/10.1080/01621459.1987.10478410

Egan, R. J., Bergner, C. L., Hart, P. C., Cachat, J. M., Canavello, P. R., Elegante, M. F., Elkhayat, S. I., Bartels, B. K., Tien, A. K., Tien, D. H., Mohnot, S., Beeson, E., Glasgow, E., Amri, H., Zukowska, Z., & Kalueff, A. V. (2009). Understanding behavioral and physiological phenotypes of stress and anxiety in zebrafish. *Behavioural Brain Research, 205*(1), 38–44. https://doi.org/10.1016/j.bbr.2009.06.022

Facchin, L., Duboué, E. R., & Halpern, M. E. (2015). Disruption of Epithalamic Left–Right Asymmetry Increases Anxiety in Zebrafish. *Journal of Neuroscience, 35,* 15847–15859. https://doi.org/10.1523/JNEUROSCI.2593-15.2015

Facciol, A., Iqbal, M., Eada, A., Tran, S., & Gerlai, R. (2019). The light-dark task in zebrafish confuses two distinct factors: Interaction between background shade and illumination level preference. *Pharmacology Biochemistry and Behavior, 179,* 9–21. https://doi.org/10.1016/j.pbb.2019.01.006

Facciol, A., Tran, S., & Gerlai, R. (2017). Re-examining the factors affecting choice in the light–dark preference test in zebrafish. *Behavioural Brain Research, 327,* 21–28. https://doi.org/10.1016/j.bbr.2017.03.040

Fanselow, M. S., & Lester, L. (1988). A functional behavioristic approach to aversively motivated behavior: Predatory imminence as a determinant of the topography of defensive behavior. Em R. C. Bolles & M. D. Beecher (Orgs.), *Evolution and Learning* (p. 185–211). Erlbaum.

File, S. E., & Gonzalez, L. E. (1996). Anxiolytic effects in the plus-maze of 5-HT1A-receptor ligands in dorsal raphe and ventral hippocampus. *Pharmacology, Biochemistry and Behavior, 54,* 123–128.

File, S. E., Gonzalez, L. E., & Andrews, N. (1996). Comparative Study of Pre- and Postsynaptic 5-HT1A Receptor Modulation of Anxiety in Two Ethological Animal Tests. *Journal of Neuroscience, 16*, 4810–4815. https://doi.org/10.1523/JNEUROSCI.16-15-04810.1996

File, S. E., & Seth, P. (2003). A review of 25 years of the social interaction test. *European Journal of Pharmacology, 463*(1–3), 35–53. https://doi.org/10.1016/S0014-2999(03)01273-1

Furchgott, R. F., & Bursztyn, P. (1967). Comparison of Dissociation Constants and of Relative Efficacies of Selected Agonists Acting on Parasympathetic Receptors*. *Annals of the New York Academy of Sciences, 144*, 882–899. https://doi.org/10.1111/j.1749-6632.1967.tb53817.x

Gebauer, D. L., Pagnussat, N., Piato, Â. L., Schaefer, I. C., Bonan, C. D., & Lara, D. R. (2011). Effects of anxiolytics in zebrafish: Similarities and differences between benzodiazepines, buspirone and ethanol. *Pharmacology, Biochemistry & Behavior, 99*, 480–486. https://doi.org/10.1016/j.pbb.2011.04.021

Genario, R., Abreu, M. S. de, Giacomini, A. C. V. V., Demin, K. A., & Kalueff, A. V. (2020). Sex differences in behavior and neuropharmacology of zebrafish. *European Journal of Neuroscience, 52*, 2586–2603. https://doi.org/10.1111/ejn.14438

Gerlai, R. (2020). Evolutionary conservation, translational relevance and cognitive function: The future of zebrafish in behavioral neuroscience. *Neuroscience & Biobehavioral Reviews, 116*, 426–435. https://doi.org/10.1016/j.neubiorev.2020.07.009

Gerlai, R. (2023). Zebrafish (*Danio rerio*): A newcomer with great promise in behavioral neuroscience. *Neuroscience & Biobehavioral Reviews, 144*, 104978. https://doi.org/10.1016/j.neubiorev.2022.104978

Giacomini, A. C. V. V., Abreu, M. S., Giacomini, L. V., Siebel, A. M., Zimerman, F. F., Rambo, C. L., Mocelin, R., Bonan, C. D., Piato, A. L., & Barcellos, L. J. G. (2016). Fluoxetine and diazepam acutely modulate stress induced-behavior. *Behavioural Brain Research, 296*, 301–310. https://doi.org/10.1016/j.bbr.2015.09.027

Gomes, K. S., & Nunes-de-Souza, R. L. (2009). Implication of the 5-HT2A and 5-HT2C (but not 5-HT1A) receptors located within the periaqueductal gray in the elevated plus-maze test-retest paradigm in mice. *Progress in Neuro-Psychopharmacology & Biological Psychiatry*, *33*, 1261–1269. https://doi.org/10.1016/j.pnpbp.2009.07.015

Gonçalves, C., Kareklas, K., Teles, M. C., Varela, S. A. M., Costa, J., Leite, R. B., Paixão, T., & Oliveira, R. F. (2022). Phenotypic architecture of sociality and its associated genetic polymorphisms in zebrafish. *Genes, Brain and Behavior*, *21*(5), e12809. https://doi.org/10.1111/gbb.12809

Graeff, F. G. (2004). Serotonin, the periaqueductal gray and panic. *Neuroscience & Biobehavioral Reviews*, *28*(3), 239–259. https://doi.org/10.1016/j.neubiorev.2003.12.004

Graeff, F. G., Viana, M. B., & Mora, P. O. (1997). Dual Role of 5-HT in Defense and Anxiety. *Science*, *21*(6), 791–799.

Gross, C., Zhuang, X., Stark, K., Ramboz, S., Oosting, R., Kirby, L., Santarelli, L., Beck, S., & Hen, R. (2002). Serotonin1A receptor acts during development to establish normal anxiety-like behaviour in the adult. *Nature*, *416*, 396–400.

Guscott, M., Bristow, L. J., Hadingham, K., Rosahl, T. W., Beer, M. S., Stanton, J. A., Bromidge, F., Owens, A. P., Huscroft, I., Myers, J., Rupniak, N. M., Patel, S., Whiting, P. J., Hutson, P. H., Fone, K. C., Biello, S. M., Kulagowski, J. J., & McAllister, G. (2005). Genetic knockout and pharmacological blockade studies of the 5-HT7 receptor suggest therapeutic potential in depression. *Neuropharmacology*, *48*(4), 492–502. https://doi.org/10.1016/j.neuropharm.2004.11.015

Hawkey, A. B., Hoeng, J., Peitsch, M. C., Levin, E. D., & Koshibu, K. (2021). Subchronic effects of plant alkaloids on anxiety-like behavior in zebrafish. *Pharmacology Biochemistry and Behavior*, *207*, 173223. https://doi.org/10.1016/j.pbb.2021.173223

Hedlund, P. B., Kelly, L., Mazur, C., Lovenberg, T., Sutcliffe, J. G., & Bonaventure, P. (2004). 8-OH-DPAT acts on both 5-HT1A and 5-HT7 receptors to induce hypothermia in rodents.

European Journal of Pharmacology, *487*(1), 125–132.

https://doi.org/10.1016/j.ejphar.2004.01.031

Herculano, A. M., & Maximino, C. (2014). Serotonergic modulation of zebrafish behavior: Towards a

paradox. *Progress in Neuro-Psychopharmacology and Biological Psychiatry*, *55*, 50–66.

https://doi.org/10.1016/j.pnpbp.2014.03.008

Ho, J., Tumkaya, T., Aryal, S., Choi, H., & Claridge-Chang, A. (2019). Moving beyond P values:

Data analysis with estimation graphics. *Nature Methods*, *16*(7), 565–566.

https://doi.org/10.1038/s41592-019-0470-3

Jastrzębska-Więsek, M., Partyka, A., Rychtyk, J., Śniecikowska, J., Kołaczkowski, M., Wesołowska,

A., Varney, M. A., & Newman-Tancredi, A. (2018). Activity of Serotonin 5-HT1A Receptor

Biased Agonists in Rat: Anxiolytic and Antidepressant-like properties. *ACS Chemical

Neuroscience*, *9*(5), 1040–1050. https://doi.org/10.1021/acschemneuro.7b00443

Joel, D., & Fausto-Sterling, A. (2016). Beyond sex differences: New approaches for thinking about

variation in brain structure and function. *Philosophical Transactions of the Royal Society B:

Biological Sciences*, *371*(1688), 20150451. https://doi.org/10.1098/rstb.2015.0451

Kalueff, A. V., Gebhardt, M., Stewart, A. M., Cachat, J. M., Brimmer, M., Chawla, J. S., Craddock,

C., Kyzar, E. J., Roth, A., Landsman, S., Gaikwad, S., Robinson, K., Baatrup, E., Tierney, K.,

Shamchuk, A., Norton, W., Miller, N., Nicolson, T., Braubach, O., … Zebrafish Neuroscience

Research Consortium. (2013). Towards a comprehensive catalog of zebrafish behavior 1.0

and beyond. *Zebrafish*, *10*, 70–86. https://doi.org/10.1089/zeb.2012.0861

Kaslin, J., & Panula, P. (2001). Comparative anatomy of the histaminergic and other aminergic

systems in zebrafish (Danio rerio). *Journal of Comparative Neurology*, *440*, 342–377.

https://doi.org/10.1002/cne.1390

Kinkel, M. D., Eames, S. C., Philipson, L. H., & Prince, V. E. (2010). Intraperitoneal injection into

adult zebrafish. *Journal of Visualized Experiments*, *42*, 2126. https://doi.org/10.3791/2126

Kysil, E. V., Meshalkina, D. A., Frick, E. E., Echevarria, D. J., Rosemberg, D. B., Maximino, C., Lima, M. G., Abreu, M. S., Giacomini, A. C., Barcellos, L. J. G., Song, C., & Kalueff, A. V. (2017). Comparative Analyses of Zebrafish Anxiety-Like Behavior Using Conflict-Based Novelty Tests. *Zebrafish, 14*(3), 197–208. https://doi.org/10.1089/zeb.2016.1415

Kyzar, E. J., & Kalueff, A. V. (2016). Exploring Hallucinogen Pharmacology and Psychedelic Medicine with Zebrafish Models. *Zebrafish, 13*(5), 379–390. https://doi.org/10.1089/zeb.2016.1251

Lau, B. Y. B., Mathur, P., Gould, G. G., & Guo, S. (2011). Identification of a brain center whose activity discriminates a choice behavior in zebrafish. *Proceedings of the National Academy of Sciences, 108*, 2581–2586. https://doi.org/10.1073/pnas.1018275108

Lawrence, C. (2007). The husbandry of zebrafish (*Danio rerio*): A review. *Aquaculture, 269*, 1–20. https://doi.org/10.1016/j.aquaculture.2007.04.077

Levin, E. D., Bencan, Z., & Cerutti, D. T. (2007). Anxiolytic effects of nicotine in zebrafish. *Physiology & Behavior, 90*, 54–58. https://doi.org/10.1016/j.physbeh.2006.08.026

Lillesaar, C., Stigloher, C., Tannhäuser, B., Wullimann, M. F., & Bally-Cuif, L. (2009). Axonal projections originating from raphe serotonergic neurons in the developing and adult zebrafish, Danio rerio, using transgenics to visualize raphe-specific pet1 expression. *Journal of Comparative Neurology, 512*(2), 158–182. https://doi.org/10.1002/cne.21887

Lima-Maximino, M., Pyterson, M. P., Silva, R. X. do C., Gomes, G. C. V., Rocha, S. P., Herculano, A. M., Rosemberg, D. B., & Maximino, C. (2020). Phasic and tonic serotonin modulate alarm reactions and post-exposure behavior in zebrafish. *Journal of Neurochemistry, 153*(4), 495–509. https://doi.org/10.1111/jnc.14978

Maaswinkel, H., Le, X., He, L., Zhu, L., & Weng, W. (2013). Dissociating the effects of habituation, black walls, buspirone and ethanol on anxiety-like behavioral responses in shoaling zebrafish. A 3D approach to social behavior. *Pharmacology, Biochemistry & Behavior, 108*, 16–27. https://doi.org/10.1016/j.pbb.2013.04.009

Maaswinkel, H., Zhu, L., & Weng, W. (2012). The immediate and the delayed effects of buspirone on zebrafish (*Danio rerio*) in an open field test: A 3-D approach. *Behavioural Brain Research*, *234*, 365–374. https://doi.org/10.1016/j.bbr.2012.07.014

MacRae, C. A., & Peterson, R. T. (2023). Zebrafish as a Mainstream Model for In Vivo Systems Pharmacology and Toxicology. *Annual Review of Pharmacology and Toxicology*, *63*(Volume 63, 2023), 43–64. https://doi.org/10.1146/annurev-pharmtox-051421-105617

Martins De Almeida, R. M., Giovenardi, M., Charchat, H., & Bolten Lucion, A. (1998). 8-OH-DPAT in the median raphe nucleus decreases while in the medial septal area it may increase anxiety in female rats. *Neuroscience & Biobehavioral Reviews*, *23*, 259–264. https://doi.org/10.1016/S0149-7634(98)00026-8

Maximino, C. (2012). *Serotonin and Anxiety: Neuroanatomical, Pharmacological, and Functional Aspects*. Springer.

Maximino, C., da Silva, A. W. B., Gouveia, A., & Herculano, A. M. (2011). Pharmacological analysis of zebrafish (*Danio rerio*) scototaxis. *Progress in Neuro-Psychopharmacology and Biological Psychiatry*, *35*, 624–631. https://doi.org/10.1016/j.pnpbp.2011.01.006

Maximino, C., Marques de Brito, T., Dias, C. A. G. de M., Gouveia, A., & Morato, S. (2010). Scototaxis as anxiety-like behavior in fish. *Nature Protocols*, *5*(2), Artigo 2. https://doi.org/10.1038/nprot.2009.225

Maximino, C., Oliveira, D. L. de, Rosemberg, D. B., Batista, E. de J. O., Herculano, A. M., Oliveira, K. R. M., Benzecry, R., & Blaser, R. (2012). A comparison of the light/dark and novel tank tests in zebrafish. *Behaviour*, *149*(10–12), 1099–1123. https://doi.org/10.1163/1568539X-00003029

Maximino, C., Puty, B., Benzecry, R., Araújo, J., Lima, M. G., de Jesus Oliveira Batista, E., Renata de Matos Oliveira, K., Crespo-Lopez, M. E., & Herculano, A. M. (2013). Role of serotonin in zebrafish (*Danio rerio*) anxiety: Relationship with serotonin levels and effect of buspirone, WAY 100635, SB 224289, fluoxetine and para-chlorophenylalanine (pCPA) in two

behavioral models. *Neuropharmacology, 71*, 83–97.

https://doi.org/10.1016/j.neuropharm.2013.03.006

Maximino, C., Silva, A. W. B. da, Araújo, J., Lima, M. G., Miranda, V., Puty, B., Benzecry, R.,

Picanço-Diniz, D. L. W., Jr, A. G., Oliveira, K. R. M., & Herculano, A. M. (2014).

Fingerprinting of Psychoactive Drugs in Zebrafish Anxiety-Like Behaviors. *PLOS ONE, 9*(7),

e103943. https://doi.org/10.1371/journal.pone.0103943

McNaughton, N., & Corr, P. J. (2004). A two-dimensional neuropsychology of defense: Fear/anxiety

and defensive distance. *Neuroscience & Biobehavioral Reviews, 28*(3), 285–305.

https://doi.org/10.1016/j.neubiorev.2004.03.005

McNaughton, N., & Corr, P. J. (2022). The non-human perspective on the neurobiology of

temperament, personality, and psychopathology: What's next? *Current Opinion in Behavioral*

Sciences, 43, 255–262. https://doi.org/10.1016/j.cobeha.2021.11.005

McNaughton, N., & Zangrossi Jr, H. (2008). Theoretical approaches to the modeling of anxiety in

animals. Em R. J. Blanchard, D. C. Blanchard, G. Griebel, & D. J. Nutt (Orgs.), *Handbook of*

Anxiety and Fear (p. 11–27). Elsevier B. V.

Meller, E., Bohmaker, K. (1994). Differential receptor reserve for 5-HT1A receptor-mediated

regulation of plasma neuroendocrine hormones. *Journal of Pharmacology and Experimental*

Therapeutics, 271, 1246-1252.

Meller, E., Goldstein, M., & Bohmaker, K. (1990). Receptor reserve for 5-hydroxytryptamine1A-

mediated inhibition of serotonin synthesis: Possible relationship to anxiolytic properties of 5-

hydroxytryptamine1A agonists. *Molecular Pharmacology, 37*, 231–237.

Meller, E., Chalfin, M., & Bohmaker, K. (1992). Serotonin 5-HT1a receptor-mediated hypothermia in

mice: Absence of spare receptors and rapid induction of tolerance. *Pharmacology,*

Biochemistry & Behavior, 43, 405-411. https://doi.org/10.1016/0091-3057(92)90169-G

Nascimento, J. O. G., Kikuchi, L. S., de Bortoli, V. C., Zangrossi Jr., H., & Viana, M. B. (2014).

Dorsomedial hypothalamus serotonin 1A receptors mediate a panic-related response in the

elevated T-maze. *Brain Research Bulletin*, *109*, 39–45.

https://doi.org/10.1016/j.brainresbull.2014.09.011

Nathan, F. M., Ogawa, S., & Parhar, I. S. (2015). Kisspeptin1 modulates odorant-evoked fear

response via two serotonin receptor subtypes (5-HT1A and 5-HT2) in zebrafish. *Journal of*
Neurochemistry, *133*(6), 870–878. https://doi.org/10.1111/jnc.13105

Newman-Tancredi, A., Cussac, D., Ormière, A.-M., Lestienne, F., Varney, M. A., & Martel, J.-C.

(2019). Bell-shaped agonist activation of 5-HT1A receptor-coupled Gαi3 G-proteins:

Receptor density-dependent switch in receptor signaling. *Cellular Signalling*, *63*, 109383.

https://doi.org/10.1016/j.cellsig.2019.109383

Norton, W. H. J., Folchert, A., & Bally-Cuif, L. (2008). Comparative analysis of serotonin receptor

(HTR1A/HTR1B families) and transporter (slc6a4a/b) gene expression in the zebrafish brain.

Journal of Comparative Neurology, *511*(4), 521–542. https://doi.org/10.1002/cne.21831

Nowicki, M., Tran, S., Muraleetharan, A., Markovic, S., & Gerlai, R. (2014). Serotonin antagonists

induce anxiolytic and anxiogenic-like behavior in zebrafish in a receptor-subtype dependent

manner. *Pharmacology Biochemistry and Behavior*, *126*, 170–180.

https://doi.org/10.1016/j.pbb.2014.09.022

Nunes-de-Souza, R. L., Canto-de-Souza, A., Da-Costa, M., Fornari, R. V., Graeff, F. G., & Pelá, I. R.

(2000). Anxiety-induced antinociception in mice: Effects of systemic and intra-amygdala

administration of 8-OH-DPAT and midazolam. *Psychopharmacology*, *150*, 300–310.

Ogi, A., Licitra, R., Naef, V., Marchese, M., Fronte, B., Gazzano, A., & Santorelli, F. M. (2021).

Social Preference Tests in Zebrafish: A Systematic Review. *Frontiers in Veterinary Science*,
7, 590057.

Olivier, B., van Wijngaarden, I., & Soudijn, W. (2000). 5-HT3 receptor antagonists and anxiety; a

preclinical and clinical review. *European Neuropsychopharmacology*, *10*, 77–95.

https://doi.org/10.1016/S0924-977X(99)00065-6

Parichy, D. M., Elizondo, M. R., Mills, M. G., Gordon, T. N., & Engeszer, R. E. (2009). Normal table of postembryonic zebrafish development: Staging by externally visible anatomy of the living fish. *Developmental Dynamics*, *238*, 2975–3015. https://doi.org/10.1002/dvdy.22113

Paul, E. D., Johnson, P. L., Shekhar, A., & Lowry, C. A. (2014). The Deakin/Graeff hypothesis: Focus on serotonergic inhibition of panic. *Neuroscience & Biobehavioral Reviews*, *46*, 379–396. https://doi.org/10.1016/j.neubiorev.2014.03.010

Paul, E. D., & Lowry, C. A. (2013). Functional topography of serotonergic systems supports the Deakin/Graeff hypothesis of anxiety and affective disorders. *Journal of Psychopharmacology*, *27*, 1090–1106. https://doi.org/10.1177/0269881113490328

Perry, S. F., Davie, P. S., Daxboeck, C., Ellis, A. G., & Smith, D. G. (1984). Perfusion methods for the study of gill physiology. Em W. S. Hoar & D. J. Randall (Orgs.), *Fish Physiology, Vol. 10. Gills: Part B. Ion and Water Transfer*. Academic Press.

Perusini, J. N., & Fanselow, M. S. (2015). Neurobehavioral perspectives on the distinction between fear and anxiety. *Learning & Memory, 22*(9), 417–425. https://doi.org/10.1101/lm.039180.115

Piszczek, L., Schlax, K., Wyrzykowska, A., Piszczek, A., Audero, E., & Gross, C. (2013). Serotonin 1A auto-receptors are not sufficient to modulate anxiety in mice. *European Journal of Neuroscience*. https://doi.org/10.1111/ejn.12260

Ponzoni, L., Sala, M., & Braida, D. (2016). Ritanserin-sensitive receptors modulate the prosocial and the anxiolytic effect of MDMA derivatives, DOB and PMA, in zebrafish. *Behavioural Brain Research*, *314*, 181–189. https://doi.org/10.1016/j.bbr.2016.08.009

Quesseveur, G., Nguyen, H. T., Gardier, A. M., & Guiard, B. P. (2012). 5-HT2 ligands in the treatment of anxiety and depression. *Expert Opinion on Investigational Drugs, 21*, 1701–1725. https://doi.org/10.1517/13543784.2012.719872

Richardson-Jones, J. W., Craige, C. P., Nguyen, T. H., Kung, H. F., Gardier, A. M., Dranovsky, A., David, D. J., Guiard, B. P., Beck, S. G., Leonardo, E. D., & Hen, R. (2011). Serotonin-1A

autoreceptors are necessary and sufficient for the normal formation of circuits underlying innate anxiety. *Journal of Neuroscience, 31*, 6008–6018. https://doi.org/10.1523/JNEUROSCI.5836-10.2011

Rodgers, R. J., & Cao, B.-J. (1997). Anxiolytic-like profile of p-MPPI, a novel 5HT1A receptor antagonist, in the murine elevated plus-maze. *Psychopharmacology, 129*, 365–371.

Serra, E. L., Medalha, C. C., & Mattioli, R. (1999). Natural preference of zebrafish (*Danio rerio*) for a dark environment. *Brazilian Journal Of Medical And Biological Research*, *32*, 1551–1553.

Siegfried, K. R., & Steinfeld, J. S. (2021). Histological Analysis of Gonads in Zebrafish. *Methods in Molecular Biology (Clifton, N.J.), 2218*, 253–263. https://doi.org/10.1007/978-1-0716-0970-5_20

Snekser, J. L., & Diestler, E. (2023). Sex differences in the expression of aggressive behavior and influences on social choice in zebrafish (*Danio rerio*). *Behavioural Processes, 208*, 104871. https://doi.org/10.1016/j.beproc.2023.104871

Sourbron, J., Schneider, H., Kecskés, A., Liu, Y., Buening, E. M., Lagae, L., Smolders, I., & de Witte, P. (2016). Serotonergic Modulation as Effective Treatment for Dravet Syndrome in a Zebrafish Mutant Model. *ACS Chemical Neuroscience*, *7*(5), 588–598. https://doi.org/10.1021/acschemneuro.5b00342

Sprouse, J. S., & Aghajanian, G. K. (1988). Responses of hippocampal pyramidal cells to putative serotonin 5-HT1A and 5-HT1B agonists: A comparative study with dorsal raphe neurons. *Neuropharmacology*, *27*(7), 707–715. https://doi.org/10.1016/0028-3908(88)90079-2

Steenbergen, P. J., Richardson, M. K., & Champagne, D. L. (2011). Patterns of avoidance behaviours in the light / dark preference test in young juvenile zebrafish: A pharmacological study. *Behavioural Brain Research, 222*, 15–25. https://doi.org/10.1016/j.bbr.2011.03.025

Stewart, A. M., Braubach, O., Spitsbergen, J., Gerlai, R., & Kalueff, A. V. (2014). Zebrafish models for translational neuroscience research: From tank to bedside. *Trends in Neurosciences*, *37*(5), 264–278. https://doi.org/10.1016/j.tins.2014.02.011

Stewart, A. M., Cachat, J., Gaikwad, S., Robinson, K. S. L., Gebhardt, M., & Kalueff, A. V. (2013). Perspectives on experimental models of serotonin syndrome in zebrafish. *Neurochemistry International*, *62*, 893–902. https://doi.org/10.1016/j.neuint.2013.02.018

Tejada, J., Chaim, K. T., & Morato, S. (2018). X-PloRat: A Software for Scoring Animal Behavior in Enclosed Spaces. *Psicologia: Teoria e Pesquisa*, *33*. https://doi.org/10.1590/0102.3772e3322

Zangrossi, H., Viana, M. B., & Graeff, F. G. (1999). Anxiolytic effect of intra-amygdala injection of midazolam and 8-hydroxy-2-(di-*n*-propylamino)tetralin in the elevated T-maze. *European Journal of Pharmacology, 369*(3), 267–270. https://doi.org/10.1016/S0014-2999(99)00075-8

Zangrossi Jr., H., Viana, M. B., Zanoveli, J., Bueno, C., Nogueira, R. L., & Graeff, F. G. (2001). Serotonergic regulation of inhibitory avoidance and one-way escape in the rat elevated T-maze. *Neuroscience & Biobehavioral Reviews, 25*, 637–645. https://doi.org/10.1016/S0149-7634(01)00047-1

Zanoveli, J. M., Nogueira, R. L., & Zangrossi Jr., H. (2003). Serotonin in the dorsal periaqueductal gray modulates inhibitory avoidance and one-way escape behaviors in the elevated T-maze. *European Journal of Pharmacology, 473*, 153–161. https://doi.org/10.1016/S0014-2999(03)01970-8

Zanoveli, J. M., Pobbe, R. L. H., de Bortoli, V., Carvalho, M. C., Brandão, M. L., & Zangrossi Jr, H. (2010). Facilitation of 5-HT1A-mediated neurotransmission in dorsal periaqueductal grey matter accounts for the panicolytic-like effect of chronic fluoxetine. *International Journal of Neuropsychopharmacology, 13*, 1079–1088.