Phylogeography of bar-tailed godwits: pre-LGM structure in Beringia and westward

colonization of post-glacial Europe

Jesse R. Conklin[1,2*], Yvonne I. Verkuil[1,2], Margaux J.M. Lefebvre[3], Phil F. Battley[4], Roeland

A. Bom[2,5], Robert E. Gill, Jr.[6], Chris J. Hassell[7], Job ten Horn[5], Daniel R. Ruthrauff[6], T. Lee

Tibbitts[6], Pavel S. Tomkovich[8], Nils Warnock[9], Theunis Piersma[1,2,5] & Michaël C.

Fontaine[1,3]

[1]Groningen Institute for Evolutionary Life Sciences (GELIFES), University of Groningen,

P.O. Box 11103, 9700 CC Groningen, The Netherlands

[2]BirdEyes, Centre for Global Ecological Change at the Faculties of Science & Engineering

and Campus Fryslân, University of Groningen, Zaailand 110, 8911 BN Leeuwarden, The

Netherlands

[3]MiVEGEC, CNRS, IRD, University of Montpellier, Montpellier, France

[4]Zoology and Ecology Group, Massey University, Palmerston North 4442, New Zealand

[5]Department of Coastal Systems, NIOZ Royal Netherlands Institute for Sea Research, P.O.

Box 59, 1790 AB Den Burg, Texel, The Netherlands

[6]U.S. Geological Survey, Alaska Science Center, 4210 University Drive, Anchorage, AK

99508, USA

[7]Global Flyway Network, PO Box 3089, Broome, WA 6725, Australia

[8]Zoological Museum, Moscow MV Lomonosov State University, Bolshaya Nikitskaya Str. 6,

Moscow 125009, Russia

[9]Audubon Canyon Ranch, Cypress Grove Research Center, Marshall, CA 94940, USA

*Corresponding author: conklin.jesse@gmail.com

ABSTRACT

In migratory birds, high mobility may reduce population structure through increased dispersal

and enable adaptive responses to environmental change, whereas rigid migratory routines

predict low dispersal, increased geographic structure, and limited flexibility to respond to

change. We used nextRAD sequencing of 14,318 single-nucleotide polymorphisms to

explore the population genetics and phylogeographic history of the bar-tailed godwit, *Limosa*

lapponica, a migratory shorebird with six recognized subspecies and known for making the

longest non-stop flights of any landbird. Using scenario-testing in an Approximate Bayesian

Computation framework, we infer that bar-tailed godwits existed in three main lineages at the

Last Glacial Maximum (LGM), when much of their present-day Arctic and sub-Arctic

breeding range persisted in a large, unglaciated Siberian-Beringian refugium. Subsequently,

population structure developed at both longitudinal extremes: in the east, a genetic cline

exists across latitude in the Alaska breeding range of subspecies *L. l. baueri*; in the west, one

lineage diversified into three extant subspecies *L. l. lapponica*, *taymyrensis*, and *yamalensis*,

the former two of which migrate through previously glaciated western Europe. We also

detected unrecognized population structure among bar-tailed godwits wintering in Europe,

wherein a significant proportion of purported *lapponica* individuals were in fact *taymyrensis*,

necessitating a re-assessment of the migrations, ecology, and population estimates for these

subspecies. In the global range of this long-distance migrant, we found evidence of both (1)

fidelity to rigid behavioral routines promoting fine-scale geographic population structure (in

the east), and (2) flexibility to colonize recently available migratory flyways and non-

breeding areas (in the west).

1 | INTRODUCTION

Animal migration is shaped by, and dependent upon, resources and conditions that vary spatially and seasonally (Alerstam et al., 2003), and therefore migratory populations are considered particularly vulnerable to disruptions caused by climate change and anthropogenic habitat modification (Kubelka et al., 2022; Robinson et al., 2009). In birds, migration involves a highly integrated, multi-trait phenotype (including flight capacity, fueling, navigation, molt, etc.; Piersma et al., 2005) and some degree of endogenous behavioral control (Åkesson et al., 2017; Åkesson & Helm, 2020) that may imply limited flexibility to adapt to conditions outside those in which the migration evolved. Conversely, it is clear that some migratory bird populations have persisted through historical large-scale environmental perturbations, such as repeated Pleistocene glaciations (Batchelor et al., 2019; Hewitt, 2000); apparent responses included population sub-division and size changes, loss and gain of migration, and altered migration routes and distances (e.g., Avise & Walker, 1998; Conklin et al., 2022; Milá et al., 2006; Tan et al., 2023; Zink & Gardner, 2017). A phylogeographic approach (Avise, 2009; Knowles & Maddison, 2002) can help us understand what perturbations migratory populations have survived in the past, and therefore their potential resiliance to future changes.

The bar-tailed godwit, *Limosa lapponica*, is a migratory shorebird (Order Charadriiformes, Family Scolopacidae) that breeds on Arctic and sub-Arctic tundra from Scandinavia east to Alaska, and spends the non-breeding season on intertidal mudflats from West Africa to New Zealand (Figure 1). There are up to six recognized subspecies of bar-tailed godwits, with distinct breeding areas and migratory routes (Figure 1), and further distinguished by variation in plumage and body size (Bom et al., 2022; Engelmoer & Roselaar, 1998; Tomkovich,

2010). Based on non-breeding census data, population sizes for these subspecies vary from c.

100,000 to 500,000 individuals (Wetlands International, 2023), except for the subspecies *L. l.*

anadyrensis, which has a poorly described non-breeding range (Chan et al., 2022) but is

believed to number <10,000 individuals. The species is considered Near Threatened by the

IUCN based on widespread population declines, particularly associated with limitations at

migratory stopover sites (Rakhimberdiev et al., 2018; Studds et al., 2017), although the

nominate European-wintering population *L. l. lapponica* is thought to be increasing (BirdLife

International, 2023).

In the western Palearctic, the present-day ranges of three bar-tailed godwit subspecies

(*lapponica*, *taymyrensis*, and *yamalensis*) were strongly affected by the glaciation of Europe

during the last glacial maximum (LGM, c. 20,000–25,000 years before present, ybp; Hewitt,

2000). Accordingly, these three subspecies were indistinguishable at the mtDNA Control

Region (Bom et al., 2022), suggesting a recent postglacial radiation into three migration

routes (Figure 1). In particular, the entire present-day breeding range of *lapponica*, which

makes the shortest migration in the species (one-way distance of c. 2,500–4,500 km), was

unavailable at the LGM, making it presumably the youngest subspecies.

By contrast, the eastern part of the bar-tailed godwit breeding range falls within Beringia, an

area of the Arctic that stretches from the Lena River in northeast Russia to the Mackenzie

River in northwest Canada (Figure 1) and was largely ice-free at the LGM (Ehlers &

Gibbard, 2007; Pielou, 1991) and previous glacial cycles (Batchelor et al., 2019). The much

deeper biogeographical history of this region, which includes the periodic appearance and

disappearance of the Bering Land Bridge, connecting North America with Asia, is reflected

in complex histories of diversification in many Beringian taxa (Winker et al., 2023). Thus,

two eastern subspecies (*menzbieri* and *baueri*) could have persisted near their present-day

ranges for much longer than the western subspecies, and indeed show deeper divergences at

the mitochondrial control region (mtDNA) (Bom et al., 2022). A latitudinal cline in both

morphology and annual-schedule timing across Alaska (Conklin et al., 2010, 2011) suggests

that additional population genetic structure may exist within *baueri*. Conversely, the small

breeding range of *anadyrensis* in Chukotka (Figure 1) was at least partly glaciated at the

LGM (Batchelor et al., 2019), and therefore could represent a recent expansion into newly

available breeding habitat.

Of particular interest is the evolution and persistence of the migration of the *baueri*

subspecies, which includes three separate trans-oceanic flights of >5,000 km, including a

southward flight of c. 12,000 km across the Pacific Ocean from Alaska to New Zealand

(Figure 1), the longest non-stop flight recorded in any landbird (Conklin et al., 2017; Gill et

al., 2009). Given this annual routine's apparent reliance on the specific distribution of

habitats and environmental conditions around the Pacific (Conklin et al., 2017; Gill et al.,

2014; Piersma et al., 2022), it is unclear how long godwits have been making this migration,

and whether it will withstand major changes (e.g. in habitat, prevailing winds etc.) in the

future. Unraveling the history of bar-tailed godwits may help reveal whether the trans-Pacific

migratory route, and the physiological capabilities required to use it, arose *in situ*, or were

products of eastward expansion from the Palearctic (Hedenström, 2010; Piersma et al., 2022).

In this study, we describe the history of divergences and degree of neutral genetic

differentiation among global flyway populations of bar-tailed godwits. We used nextera-

tagmented, reductively-amplified DNA (nextRAD) sequencing (Russello et al., 2015) for *de*

novo discovery of genome-wide single-nucleotide polymorphisms (SNPs) for population

genetic analyses, and compared hypothesized evolutionary scenarios in an Approximate

Bayesian Computation (ABC) framework using DIYABC-RF (Collin et al., 2021). By

reconstructing the recent evolutionary history of bar-tailed godwits, we (1) identify

previously unrecognized sub-specific structure, (2) reveal the divergence times and historical

connections among populations, and (3) provide insights regarding the origins and

maintenance of population structure in long-distance migratory birds.

2 | MATERIALS AND METHODS

2.1 | Sampling and DNA extraction

We assembled DNA samples representing all recognized and hypothesized breeding

populations within the global range of bar-tailed godwits (Figure 1, Table S1). Where

possible, we used samples collected in known breeding areas; however, because bar-tailed

godwits breed in low densities in remote regions, for some populations there were few or no

breeding samples available. In these cases, we used samples collected from non-breeding

sites when sampled individuals could be confidently assigned to breeding areas either

because they were remotely tracked to breeding areas using satellite-telemetry or light-level

geolocation, or because long-term research programs had established strong links between

breeding and non-breeding areas (e.g. through mark-recapture/resight programs). There were

no verifiable samples available from the poorly-studied *anadyrensis* population; however,

godwits have been collected from coastal southeast Chukotka, within 300 km of the known

anadyrensis breeding range (Tomkovich, 2010) and far from the expected migration routes of

menzbieri and *baueri* (Figure 1). Therefore, we included eight such samples, in order to

detect genetic variation that might indicate the distinctiveness of *anadyrensis*. All tissue

samples were acquired from museum collections or collected by the authors and colleagues in

the field under all requisite permits appropriate to their respective countries and institutions.

We extracted genomic DNA from samples using two methods. For blood or organ tissue

samples preserved in 95% ethanol, we used the DNeasy Blood and Tissue Kit (Qiagen)

following the manufacturer's instructions for tissue. For blood samples preserved in Queen's

lysis buffer, we used the NucleoSpin Blood QuickPure Kit (Macherey-Nagel). Extract quality

was first assessed on a 1.5% agarose gel to exclude extractions with insufficient yield or

excessively degraded DNA. We then quantified DNA concentrations using a Qubit 3.0

fluorometer (Life Technologies), diluted extracts to achieve relatively even concentrations,

and dried down samples in a SpeedVac concentrator (ThermoFisher Scientific). We delivered

80–162 ng of DNA of 209 individual godwits (Table S1) for SNP discovery and genotyping.

2.2 | SNP genotyping using nextRAD sequencing

Genomic DNA was converted into nextRAD genomic fragment libraries (SNPsaurus, LLC,

USA) following the method described by Russello et al. (2015). Genomic DNA was first

fragmented with Nextera reagent (Illumina, Inc.), which also ligates short adapter sequences

to the ends of the fragments. The Nextera reaction was scaled for fragmenting 10 ng

of genomic DNA, although 20–30 ng of genomic DNA was used for input to compensate for

degraded DNA in the samples and to increase fragment sizes. Fragmented DNA was then

amplified, with one of the primers matching the adapter and extending 10 nucleotides into

the genomic DNA with the selective sequence 'GTGTAGAGCC'. Thus, only fragments

starting with a sequence that can be hybridized by the selective sequence of the primer were

efficiently amplified. PCR amplification was done at 74°C for 27 cycles. The nextRAD

libraries were sequenced on an Illumina HiSeq-4000 at the Genomics Core Facility,

University of Oregon, USA.

For SNP-calling, reads were first trimmed using custom scripts (SNPsaurus, LLC) in bbduk

(BBMap tools; Bushnell, 2016). Next, a *de novo* reference was created from abundant reads

(after removal of low-quality (phred-scale quality <20) and very high-abundance reads) and

reads that aligned to these. All 205,854,244 reads were mapped to the reference with an

alignment identity threshold of 95% using bbmap (BBMap tools). Genotype calling was

performed using SAMtools and BCFtools (samtools mpileup -gu -Q 10 -t DP, DPR -f

ref.fasta -b samples.txt | bcftools call -cv - > genotypes.vcf), applying a minimum read depth

filter of 7x. The genotype table was then filtered using VCFtools v.01.14 (Danecek et al.,

2011) to remove SNPs called in <80% of samples and putative alleles with a population

frequency <3%, to exclude artefactual variants. The resulting VCF file included 6,424 unique

loci (150-bp sequences) containing 21,031 SNPs.

Additional cleaning was performed using VCFtools to remove indels ($n = 478$ SNPs), low-

quality SNPs (phred-scale quality [QUAL] <999; $n = 15$ SNPs), and samples that failed to

sequence ($n = 1$ *menzbieri* individual) or were of unknown breeding population ($n = 9$

individuals from New Zealand that could not be confidently assigned to a breeding region

within Alaska based on tracking data). Next, SNPs deviating from Hardy-Weinberg (H-W)

proportions in all seven hypothesized populations (excluding *anadyrensis*) were identified

and removed using VCFtools ($p < 0.05$; $n = 48$ SNPs); the small sample size in *anadyrensis*

made it statistically insensitive to H-W deviations. To ensure independence of loci, we used

PLINK v.1.9 (Chang et al., 2015) to identify and remove all SNPs in linkage disequilibrium

(LD; r^2 >0.20; n = 6,188 SNPs). The LD-pruned data set included 199 individuals and 14,318

unlinked SNPs on 6,140 loci.

We then used VCFtools to calculate proportion of missing SNP calls per individual (range 1–

77%) and removed 15 individuals with >30% missing data. Because inclusion of related

individuals can bias population genetic analyses (Anderson & Dunham, 2008; Rodríguez-

Ramilo & Wang, 2012), we estimated individual pairwise relatedness in PLINK using the

Identity-by-Descent estimator PI HAT; we found no cases of relatedness involving half-

siblings or closer (PI HAT >0.20). The final data set included 184 individuals and 14,318

SNPs. We used VCFtools, PLINK, and PGDspider v.2.1.0.3 (Lischer & Excoffier, 2012) to

convert data to different formats required for further analysis.

2.3 | Inference of population structure and diversity

To assess major axes of genetic variation and structure, we performed a principal component

analysis (PCA; Patterson et al. 2006) using the R packages *gdsfmt* v.1.14.1 and *SNPRelate*

v.1.12.2 (Zheng et al., 2012). We determined the number of ancestral populations or clusters

(K) and ancestry proportions of these for each individual using ADMIXTURE v.1.3.0

(Alexander et al., 2009). We conducted 20 replicate runs (with random seeds) for each

putative number of clusters (K) ranging from 1 to 8. We used cross-validation (CV)

(Alexander & Lange, 2011) to assess how the CV error rate varied with increasing K values

and which value provided the lowest CV error. Then, we used the CLUMPAK (Cluster

Markov Packager Across K; Kopelman et al., 2015) web server (http://clumpak.tau.ac.il/)

with default settings to summarize estimates of individual ancestry proportions to each

cluster across replicate runs, inspect potential distinct solutions identified, and visualize the

most likely ancestry proportions at each value of K.

Globally and for each putative population, we characterized genetic diversity by calculating

nucleotide diversity (π), heterozygosity, and inbreeding coefficient (F_{IS}) using VCFtools. We

evaluated evidence for deviations from mutation-drift equilibrium, indicative of departures

from a null hypothesis of constant population size, by estimating the per-locus Tajima's D

values for each population using VCFtools. We estimated the degree of genetic

differentiation among populations by calculating pairwise F_{ST} (Weir & Cockerham, 1984)

using the *diffCalc* function in the R package *diversity* v.1.9.90 (Keenan et al., 2013), with

95% confidence intervals derived from 500 bootstraps. We calculated *p*-values for F_{ST}

estimates using the *pairwiseTest* function (1,000 permutations) in the R package *strataG*

v.2.4.905 (Archer et al., 2017).

2.4 | Population evolutionary relationships and demographic history

To visualize evolutionary relationships among populations, we first constructed a midpoint-

rooted neighbor-joining tree based on Nei's genetic distance (Nei, 1972) using the R

packages *poppr* v.2.8.5 (Kamvar et al., 2014) and *ape* v.5.3 (Paradis & Schliep, 2019), with

missing data replaced by mean allele counts, and node support calculated from 1,000

bootstraps.

To identify likely potential sources of admixture in the tree, we first conducted the three-

population test for admixture (Reich et al., 2009) in R using ADMIXTOOLS 2.0.0 (Maier et

al., 2023) for all possible population triads. Briefly, the *f*₃ statistic (Patterson et al., 2012)

assesses shared genetic drift to test the null hypothesis that a target population (A) is related

to two other populations (B, C) in a simple tree-like manner. We tested significance of the *f*₃

test using a blocked-jackknife resampling of SNPs, considering blocks of 100 SNPs. A

significantly negative value of f_3 (A;(B,C)) indicates that the target population must have

arisen from an admixture of the other two populations, or populations closely related to them.

To further investigate the genetic relationships among godwit populations, we estimated

population networks using the approach implemented in ADMIXTOOLS 2.0.0 (Maier et al.,

2023), which uses the shared and private genetic ancestry components of genome-wide allele

frequency data to infer population branching while accounting for potential historical

migration and admixture events among populations. To find the best network topology, we

used the function *find_graphs* for 0 to 5 admixture events (*m*), each with 100 replicates and a

maximum of 300 generations. For each replicate, we calculated the out-of-sample score to

account for the difference in likelihood score caused by an increasing number of admixture

events, which also increase the degrees of freedom. We used the *baueri* South population as a

root to fold the networks; even if that population is not an actual outgroup, it allowed

visualizing population relationships in a consistent way. The actual population branching

order was later assessed statistically using the Approximate Bayesian Computation (ABC)

(Beaumont et al., 2002) Random Forest (RF) statistical framework (Pudlo et al., 2016;

Raynal et al., 2019). For the best-supported replicates (i.e., those displaying the likelihood

score closest to zero), we assessed the goodness-of-fit with the R package *admixture-graph*

v.1.0.2 (Leppälä et al., 2017). This approach allows comparing the observed and expected

values of f_4 statistics among the different alternatives and identifying the graph that best fits

the data (i.e., the one properly predicting the observed f_4 values for all possible four-

population combinations).

Finally, we evaluated the support for different possible scenarios of population divergence

and admixture using the ABC-RF statistical framework (Pudlo et al., 2016; Raynal et al.,

2019) in the R package DIYABC-RF v.1.2.1 (Collin et al., 2021). ABC-RF can estimate

posterior probabilities of historical scenarios, based on massive coalescent simulations of

genetic data. Simulations are compared to observed SNP data using summary statistics to

identify the best-fitting model by calculating the number of RF votes and to approximate the

posterior probability for the best model (Pudlo et al., 2016). The best-fitting posterior

parameter distribution values for the best model can be estimated using a RF procedure

applied in a regression setting (Raynal et al., 2019). Estimated parameters include the

effective size (N_e) for each population, admixture events and their rates (r), and split and

admixture times (t). We evaluated potential evolutionary scenarios in a stepwise manner, as

follows.

As our primary question arising from the above analyses concerned the nature and relative

timing of the admixed origin of *menzbieri* (see *Results*), we sought to first reduce the number

of potential scenarios by fixing the topology of the relationships among the three *baueri*

populations. So, in Step 1, we tested whether the apparently clinal variation among these

populations was best represented by one of two possible simple branching topologies or with

baueri Central as an admixture of *baueri* North and South (Figure S4, Table S2).

In Step 2, we created four possible scenarios (Figure S5c–f) for an admixed origin of

menzbieri from neighboring populations to the west and east, while allowing populations on

the two main branches to diverge at different times relative to each other. Essentially, the

admixed origin of *menzbieri* could occur: 1) before either branch began to diverge (Figure

S5c); 2) after populations on one branch diverged but before the other branch began to

diverge (Figure S5d,e); or 3) after populations on both branches had diverged (Figure S5f).

To compete with these scenarios, we created two "null-hypothesis" scenarios in which

menzbieri was not admixed, but simply diverged from one of the two main branches (Figure

S5a,b).

The scenario parameters were considered as random variables drawn from prior distributions

(Tables S2 and S3). We used DIYABC-RF to simulate 20,000 genetic data sets per scenario,

with the same properties as the observed data set (number of loci and proportion of missing

data). Simulated and observed data sets were summarized using the whole set of summary

statistics proposed by DIYABC-RF for SNP markers (Table S4), describing population

genetic variation (e.g. heterozygosity, proportion of monomorphic loci), differentiation (e.g.

F_{ST} and Nei's genetic distances), or admixture (e.g. admixture coefficient, f_3 and f_4 statistics;

Patterson et al., 2012); see the full list and details in Table S4. Linear discriminant analysis

(LDA) components were also used as additional summary statistics (Estoup et al., 2012). The

total number of summary statistics was 50 and 1,708 for Step 1 and 2, respectively.

We used the random forest (RF) classification procedure (Pudlo et al., 2016) to compare the

likelihood of the competing scenarios at each step with DIYABC-RF. RF is a machine-

learning algorithm that uses hundreds of bootstrapped decision trees to perform classification,

using the summary statistics as a set of predictor variables. Some simulations are not used in

decision tree building at each bootstrap (i.e., the out-of-bag simulations), and are used to

compute the "prior error rate," which provides a direct method for estimating the cross-

validation error rate. At each step, we built a training set of 20,000 simulated data sets per

scenario, with the same number of loci and individuals as the observed data set, and then

grew classification forests of increasing size (100, 500, and 2,000 trees) to test convergence.

The RF computation provides a classification vote for each scenario (i.e., the number of times

a model is selected from the decision trees). We selected the scenario with the highest

classification vote as the most likely scenario, and we estimated its posterior probability

following the recommendation of Pudlo et al. (2016). We assessed the global performance of

our chosen ABC-RF scenario, by calculating the prior error rate based on the available out-

of-bag simulations and we repeated the RF analysis (3 times in Step 1; 10 times in Step 2) to

ensure that the results converged.

For the best-supported model in Step 2, we estimated posterior distribution values of all

parameters using a regression by RF methodology (Raynal et al., 2019) in DIYABC-RF. We

used a classification forest of 500 decision trees, based on a training set of 100,000

simulations, to conduct the parameter estimations. We converted divergence time estimates to

years assuming a generation time of 8 years (delayed maturity with adult annual survival c.

0.86; Méndez et al., 2018) and the genome-wide mutation rate calculated by Zhang et al.

(2014) for Order Charadriiformes: 1.5×10^{-9} substitutions per site per year. All of the steps

involved in the ABC-RF analysis (simulations, computation of summary statistics, model

checking, scenario comparisons, and estimations of parameter posterior distributions) were

performed with DIYABC-RF.

3 | RESULTS

3.1 | Summary of nextRAD SNP data set

The final data set comprised 184 unrelated bar-tailed godwits genotyped at 14,318 unlinked

high-quality SNPs; this included 15–31 individuals in each of seven hypothesized

populations, but only three purported *anadyrensis* individuals (Table S1). On average,

individuals were genotyped at 93.7% (range: 72–99%) of SNPs and with a mean read depth

of 50.4 (range: 15–116). Each SNP was genotyped in an average of 172.5 (range: 149–184,

or 81–100%) individuals. Globally, nucleotide diversity (π) was 0.245, and heterozygosity

was 20.6%.

3.2 | Population structure and diversity

The first three axes (PC1–3) of the principal component analysis explained 4.0% of the total

genetic variation (Figure 2a). On PC1, individuals were essentially distributed on an axis of

longitude in three clusters: a western Palearctic group containing all of *lapponica,*

taymyrensis and *yamalensis*; an Alaska group containing all of *baueri*; and an intermediate

group of *menzbieri*, suggesting an admixed origin (Figure 2a). PC2 separated 12 *lapponica*

individuals from the main western Palearctic cluster. PC3 stretched *baueri* individuals along

a latitudinal cline, partly separating the North, South and Central populations. The three

purported *anadyrensis* individuals did not form a coherent cluster, one clustering with

menzbieri and two with *baueri*. Higher-order axes of variation (PC4–6) explained an

additional 2.2% of variation but revealed negligible further sub-structure (Figure S1).

Genetic ancestry analyses with ADMIXTURE for $K = 2$ ancestral populations provided the

lowest CV error criterion (Figure S2) and 100% support for the major clusters across 20

replicate runs. At $K = 2$, populations were clearly divided between a western Palearctic

cluster and an Alaska cluster, with *menzbieri* reflecting admixture between the two (Figure

2b). Additional structure identified in the PCA was apparent at $K = 3$ and 4; i.e. the clear

structure within purported *lapponica* and the clinal variation within *baueri*, respectively. At K

= 5, the identification of a predominantly *menzbieri* cluster may suggest significant genetic

drift specific to that population, as opposed to an origin of simple and recent admixture.

Higher values of K revealed no further important geographic structure (Figure S2). Again, the

purported *anadyrensis* individuals did not form a coherent cluster, but appears to include one

menzbieri and two *baueri* individuals (Figure 2b).

In both PCA and ADMIXTURE, only 6 of 27 purported *lapponica* individuals sampled on

wintering grounds in The Netherlands clearly grouped with the four individuals sampled at

known *lapponica* breeding areas in western Russia (see Figure 1, Table S1). Of the remaining

21, 19 clearly grouped with *taymyrensis* (sampled at breeding areas in central Russia and a

non-breeding area in Mauritania), and two appear to reflect admixture between *lapponica* and

taymyrensis (Figure 2a,b). Therefore, for further analysis, we considered a new population of

'*taymyrensis* Europe' ($n = 19$), and a reduced *lapponica* population ($n = 10$ 'true' *lapponica*

plus the 2 admixed individuals). For clarity, we hereafter refer to the original *taymyrensis*

samples as '*taymyrensis* Africa' ($n = 29$). As PCA and ADMIXTURE results do not support

an identifiable *anadyrensis* population, we excluded those samples ($n = 3$) from further

analysis. Therefore, we proceeded with 181 individuals in eight populations.

Globally, nucleotide diversity was 0.245, and this was similar for all populations (Table S5).

Mean heterozygosity was also generally uniform (range of means: 0.198–0.215), with no

evidence of inbreeding in any population (all mean $F_{IS} < 0.20$; Table S5). Values of Tajima's

D were low and similar among populations (range of means: 0.214–0.339; Table S5),

suggesting no evidence of dramatic population-size changes or additional hidden structure.

Pairwise F_{ST} among the eight hypothesized populations ranged 0.001–0.048 (Table 1), and

were greatest between the westernmost and easternmost breeding populations (*lapponica* vs.

baueri; F_{ST} values ranging from 0.046 to 0.048 for the three *baueri* sub-populations). The

two groups that winter in Europe, all previously considered *lapponica* (*lapponica* and

taymyrensis Europe), were moderately differentiated ($F_{ST} = 0.016$). Differentiation among the

three sub-populations of *baueri* ($F_{ST} = 0.006$–0.009) was lower than for most recognized

subspecies pairs, with 95% CIs that included zero. The lowest values were among the three

groups breeding in central Russia (*taymyrensis* Europe, *taymyrensis* Africa, and *yamalensis*;

$F_{ST} = 0.001$–0.003).

3.3 | Population evolutionary relationships and demographic history

The neighbor-joining (NJ) tree based on Nei's distance (Figure 3a) again identified two main

branches (western Palearctic and Alaska groups), with low node support within each group,

and *menzbieri* occupying an intermediate position between them.

The three-population test (based on the Patterson's f_3 statistic of the form (A;(B,C)); Table

S6) identified *menzbieri* as a likely product of admixture between the two main branches

identified by the NJ tree (see Figure 3a). Of 168 possible population triads, 12 produced a

significantly negative f_3 statistic; all of these included *menzbieri* as the target population (A)

and one member of each of the two main branches (*lapponica/taymyrensis/yamalensis* or the

baueri group, respectively) as the source populations (B, C) (Table S6).

For admixture graphs estimated by ADMIXTOOLS 2, the greatest increase in likelihood

occurred at $m = 2$ admixture edges in the population graph (Figure S3a), but out-of-sample

scores indicated the highest likelihood at $m = 3$ (Figure S3b). Overall, the five best-supported

replicates (i.e., those with likelihood scores closest to zero) included two distinct network

topologies for $m = 3$ and one for $m = 2$. The best-supported topologies at $m = 2$ and 3 (Figure

S3c,d) each provided a perfect fit with the data as estimated by *Admixture-graph* (i.e., no

outlying observed f_4 statistic values compared to those theoretically expected from these

graphs; Figure S3e,f). Therefore, according to parsimony we present the model at $m = 2$

(Figure 3b). In this graph, *menzbieri* arose from the admixture of a population ancestral to

present-day western Palearctic populations (86% contribution) and a population closely

related to *baueri* North (14%). Additionally, *baueri* North has received 4% ancestry from a

population closely related to *taymyrensis* Europe.

To resolve the nature and relative timing of the admixed origin of *menzbieri* suggested by all

previous analyses, we compared the likelihood of alternative scenarios using the approximate

Bayesian computation – random forest approach (ABC-RF). In Step 1, we tested whether the

apparently clinal variation among the three *baueri* populations was best represented by a

simple branching topology or by historical admixture (Figure S4). The best-fitting scenario,

representing *baueri* Central as an admixture of *baueri* North and South, received the greatest

proportion of RF votes (44.5%), with a posterior probability of 51.0% (Figure S4 and Table

S7). This ABC-RF analysis had good power to discriminate among the three scenarios, as

suggested by the low prior error rate of 11.4% and the confusion matrix (Table S8).

Therefore, we fixed the topology of the main Alaskan branch accordingly in the global

scenarios evaluated in Step 2.

In Step 2 (Figure S5), we compared six possible scenarios for the origin of *menzbieri*: two

"null-hypothesis" scenarios in which *menzbieri* simply diverged from one of the two main

branches (*a,b*), and four in which an admixed *menzbieri* arose at different time points relative

to the diversification within each branch (*c–f*). The best-supported scenario was *c*, in which

the historical admixture occurred prior to the main diversifications within both the western

Palearctic and Alaska groups (Figure 4c); this scenario received (mean ± SD) 26.4 ± 0.9% of

votes, with a posterior probability of 46.1 ± 2.6% (Figure S5, Table S9). The global prior

error rate of 22.0 ± 0.0% indicated that discriminating power among the six scenarios was

good, but with some difficulties in discriminating between scenarios *c* and *e* (Table S10).

Scenario *e* was the second best-supported scenario, receiving 25.3 ± 1.0% of votes. This

scenario was most similar to *c*, but with the admixture occurring before diversification among

baueri populations and after *lapponica* diverged from the other western Palearctic

populations (Figure 4e).

Each of the 20 demographic parameters of the best-supported scenario was estimated within

the ABC-RF framework (Table S11). Timing parameter estimates support the existence of

three main lineages at the LGM, with the divergence of the western Palearctic and Alaskan

branches occurring c. 38.2K ybp (modal estimate; 5th–95th quantiles: 22.0–93.1K), followed

by an admixture giving rise to *menzbieri* c. 22.0K ybp (13.4–52.5K; Figure 5, Table S11).

The post-LGM warming period featured *lapponica* diverging from the western Palearctic

lineage (c. 16.4K ybp; 9.1–28.9K) and the diversification of *baueri* populations (c. 11.6–

12.3K ybp); although we modeled the *baueri* diversification as a divergence followed by an

admixture (see Step 1), the two time estimates were essentially indistinguishable, indicating a

relatively contemporaneous radiation of three populations, rather than contact after a

prolonged isolation (Figure 5). Most recently, the remaining western Palearctic lineage

diversified into *taymyrensis* (Europe and Africa) and *yamalensis* populations (c. 4.5–5.2K

ybp); these two divergence times were also indistinguishable, rather than clearly sequential

(Figure 5).

4 | DISCUSSION

4.1 | Population structure

Previous research showed that the three western Palearctic bar-tailed godwit populations (*lapponica*, *taymyrensis*, and *yamalensis*) were indistinguishable at the mtDNA control region (Bom et al., 2022), consistent with a very recent post-glacial expansion into three breeding areas and migration routes. However, in genome-wide SNPs, we found that *lapponica* was strongly differentiated from *taymyrensis* and *yamalensis*, which were weakly but significantly differentiated from each other (Table 1). This suggests a more complicated phylogeographic history, including a post-LGM colonization by *lapponica* of recently-deglaciated Europe, followed later by emerging structure along two flyways in the source population breeding in the central Russian Arctic. These differences from patterns found in mtDNA illustrate the greater resolution of genome-wide nuclear markers to detect subtle and recent population structure (Edwards & Bensch, 2009).

We also found unexpected structure among samples collected in the Dutch Wadden Sea during winter, purported to belong to *lapponica*: of these 27 samples, only 6 grouped with the 4 individuals sampled in the breeding area of *lapponica*, whereas 19 clearly clustered with *taymyrensis*, and another 2 appeared to represent admixture between the two subspecies (Figure 2). Due to long-term mark-resighting programs on the flyway, we were able to restrict our purported *lapponica* samples to marked individuals that had been observed in multiple years in mid-winter (December–January) in the Wadden Sea and had never been reported in west Africa; this should have provided the best chance to exclude *taymyrensis* individuals from that sample, as *taymyrensis* is expected to pass through western Europe on migration between Russian breeding grounds and non-breeding grounds in west Africa but to

be absent in the winter (Figure 1; Drent & Piersma, 1990; Duijns et al., 2012). Our results

strongly indicate that both *taymyrensis* and *lapponica* are present in Europe in the winter.

The currently recognized *lapponica* and *taymyrensis* subspecies were once considered

differentially migrating populations of the nominate subspecies *L. l. lapponica*, until the

recognition of plumage and morphometric differences between the Fennoscandian and

central Russian breeding areas led to the full subspecies status of *taymyrensis* (Engelmoer &

Roselaar, 1998); this taxon was itself later split with the recognition of *yamalensis* (Bom et

al., 2022). *Lapponica* and *taymyrensis* are generally considered 'leap-frog' migrants, with the

entirely of *taymyrensis* wintering in west Africa, south of European-wintering *lapponica*

(Figure 1), consistent with historical mark-resight and tracking evidence (Duijns et al., 2012).

By contrast, based on morphometric data, Engelmoer (2008) predicted that approximately

20% of bar-tailed godwits wintering in the Dutch Wadden Sea were *taymyrensis*, and thus the

migration along the flyway was in fact a 'partial leap-frog system'. Despite this uncertainty,

the current population estimate for *lapponica* (150,000–180,000 and increasing; Wetlands

International, 2023) is based entirely on winter census data from Europe, assuming no mixing

of subspecies. If our results from the Wadden Sea (i.e. c. 70% *taymyrensis*) reflect the

subspecies proportions of the entire European wintering population, then the currently

accepted population estimate for *lapponica* might overstate the actual population size by a

factor of three, and any perceived increase could reflect unknown dynamics in two conflated

populations. This should be tested through genetic assignment of a larger and geographically

broader sample of the European wintering population, followed by a re-evaluation of the

population sizes and conservation status of both *lapponica* and *taymyrensis*.

We were unable to completely resolve structure among the three Beringian subspecies due to

the small number and high sequencing failure rate of purported *anadyrensis* samples; we thus

can conclude little regarding the status or history of that subspecies, which has a distinct

migration route through eastern Russia but an incompletely described non-breeding range

(Chan et al., 2022; Tomkovich, 2010). However, the three samples in our final data set did

not cluster together, grouping with either *menzbieri* ($n = 1$) or *baueri* ($n = 2$; Figure 2). This

suggests that these individuals, sampled in southeast Chukotka, were most likely passage

migrants from the neighboring subspecies to the west and east, rather than from the closest

breeding population, but leaves unanswered the question of the potential genetic

distinctiveness of *anadyrensis*. Therefore, genetic evaluation of the status of *anadyrensis*

awaits analysis of confirmed breeding members of the subspecies.

Within *baueri*, we confirm that behavioral and morphological differences observed along a

latitudinal cline in the Alaska breeding range (Conklin et al., 2010, 2011; Conklin & Battley,

2011) are mirrored by a similar genetic cline (Figure 2). Bar-tailed godwits from the entire

Alaska breeding range regularly mix at non-breeding sites in New Zealand (Battley et al.,

2020; Conklin et al., 2011) and Australia (Wilson et al., 2007) and at migratory staging sites

in the Yellow Sea and southwest Alaska (Battley et al., 2012). In the Yellow Sea, many

baueri individuals also mix freely in staging flocks of *menzbieri* on northward migration (see

Figure 1; Battley et al., 2012; Choi et al., 2015). The clear genetic structure maintained

among (sub-) populations that regularly meet in the non-breeding season implies that

behavioral rigidity, in terms of strict migration timing and fidelity to a breeding region, is

sufficient to preclude panmixia among and within subspecies. Within *baueri*, previous

research found no clear link between migration timing, which generally follows the

latitudinal genetic cline we detected, and variation at the gene Clock, thought to be associated

annual-cycle scheduling in birds (Parody-Merino et al., 2019). Therefore, it remains unclear

whether the phenotypic and genetic cline within *baueri* is primarily driven by neutral genetic

drift or involves selection, and whether it is emerging or stably maintained over a long time

period.

4.2 | Phylogeographic history

Our demographic reconstruction in DIYABC-RF provides the first estimates for divergence

times in bar-tailed godwits, and new insights regarding the historical origins of present-day

flyway populations. Our divergence time estimate of c. 38K ybp for the two main lineages

suggests the ancestors of the western Palearctic and Alaskan populations existed in Siberia

and Beringia well before the LGM (Figure 5). This corresponds to a period (c. 40K ybp)

when the Bering Sea separated Asia and North America (Farmer et al., 2023) and much of

the Chukotka Peninsula was glaciated (Batchelor et al., 2019); either or both of these factors

could have caused or perpetuated isolation of the two lineages. We inferred a subsequent

admixture of the two lineages c. 22K ybp, giving rise to present-day *menzbieri* (Figure 5).

During this period, just prior to and including the LGM, Beringia was largely unglaciated

(Batchelor et al., 2019) and lower sea levels created a connection between the continents via

the Bering Land Bridge (c. 36–11K ybp; Farmer et al., 2023), a situation that could have

created new breeding habitat available for colonization from both the west and the east, or

conceivably re-connected breeding populations in a continuous swath of tundra habitat

stretching from Chukotka to Alaska. Indeed, reconstructed vegetation histories indicate that

at least part of the Bering Land bridge hosted graminoid and herbaceous tundra, the preferred

breeding habitat of bar-tailed godwits, during the LGM (Wang et al., 2017). Presumably,

subsequent re-flooding of the Bering Strait (by c. 11K ybp) and incremental reduction and

isolation of tundra habitat associated with the warming period leading to the mid-Holocene

Climatic Optimum (c. 8K ybp; Kraaijeveld & Nieboer, 2000; Stewart & Dalén, 2008;

Wauchope et al., 2017) served to isolate these three extant lineages into their present-day

breeding ranges (Figure 1); similar structure across Beringia is found in numerous avian taxa

(Winker et al., 2023)

Because significant portions of Alaska have remained unglaciated for hundreds of thousands

of years (Batchelor et al., 2019; Winker et al., 2023) and our results suggest no temporal limit

to *baueri*'s occupation of its present-day breeding range, we can infer no theoretical

maximum age of that population's trans-Pacific migration (Gill et al., 2009). Therefore, we

cannot help discern between scenarios of *in situ* evolution of these extremely long non-stop

flights in the Pacific versus incremental up-scaling of flight distances through eastward

expansion of godwits from Asia (Hedenström, 2010; Piersma et al., 2022). We can only infer

that any such eastward expansion did not appear to occur recently (e.g. post-LGM), as *baueri*

has apparently existed in Alaska for sufficient time for genetic and morphological structure to

develop along a latitudinal cline (Conklin et al., 2011). However, our inference of a radiation

into this cline c. 12K ybp is potentially consistent with expansion northward or southward

within Alaska, or with emerging isolation-by-distance across a stably occupied area.

Paleoclimatic modeling, to assess change in both atmospheric circulation and habitats in the

Pacific Basin across time, may reveal more about how long *baueri* could theoretically have

been migrating across the Pacific in the current manner (Piersma et al., 2022).

If bar-tailed godwits existed in a Siberian-Beringian refugium (stretching from the West

Siberian Plain to Alaska) at the LGM and then expanded westward with the de-glaciation of

Europe (c. 14K ybp), then we would expect *lapponica* to have diverged very recently from

other western Palearctic lineages, particularly given the expected delay between geographic

separation and genetic divergence. In this light, our inferred divergence of *lapponica* c. 16K

ybp is unexpected, as their present-day breeding range in Fennoscandia was not available

until at least 3,000 years after that (Stroeven et al., 2016). This divergence-time estimate is

not easily explained by a simple westward expansion, and favors a scenario of isolation

followed by expansion from tundra breeding areas that were available south or southeast of

glaciated Europe during or soon after the LGM. In fact, such refugia, including in

central/southern Europe, have been inferred for tundra-breeding shorebirds (Arcones et al.,

2021) and other taxa (Hewitt, 2004; Sommer & Nadachowski, 2006; Taberlet et al., 1998). If

the breeding grounds of *lapponica* gradually moved toward Fennoscandia from central

Europe as glaciers retreated north, then its present-day migration route would re-trace that

northward expansion, as has been hypothesized for numerous migratory birds that either

originally evolved from sedentary populations (Rappole & Jones, 2002; Salewski &

Bruderer, 2007) or oscillated between short- and long-distance migration as northern areas

were impacted by cyclic glaciations (Bruderer & Salewski, 2008; Zink & Gardner, 2017).

At the LGM, the lineage that recently gave rise to *taymyrensis* and *yamalensis* could have

existed east or south of the glaciers that extended from Europe to the Taimyr Peninsula of

central Russia (Arcones et al., 2021; Batchelor et al., 2019). We may then expect that the

present-day breeding range and migration of *yamalensis* (Figure 1) would most closely

resemble the state of the western Palearctic lineage at the LGM. Consistent with this, there is

evidence of tundra habitats existing south of the ice-dammed West Siberian Lake at the LGM

(Binney et al., 2017; Mangerud et al., 2004), close to the present-day breeding area of

yamalensis in the West Siberian Plain. Currently, *taymyrensis* and *yamalensis* have some

degree of isolation in breeding areas along a latitudinal and longitudinal gradient (Figure 1;

Bom et al., 2022). This may suggest that a northeastward expansion of breeding range toward

the Taimyr Peninsula was associated with discovery of the migration route of *taymyrensis*

through western Europe (Figure 1) after glaciers retreated.

We have presented the first unequivocal evidence that *taymyrensis* can be found in Europe in

winter, and it is therefore unclear whether this is a new, perhaps emerging, development in

modern times (i.e. tens or hundreds of years) or a stable situation that went previously

undetected. However, the detection of two apparent F1 *lapponica*/*taymyrensis* hybrids (see

Figure 2) in our relatively small sample from the Wadden Sea appears more indicative of a

brief period of significant contact between the subspecies. Also, the negligible differentiation

between European- and Africa-wintering *taymyrensis* ($F_{ST} = 0.001$) provides no evidence of

further structure within that subspecies. Taken together, our results suggest that the existence

of *taymyrensis* in Europe, or at least conditions promoting successful cross-population

recruitment, is a recent or increasing phenomenon.

4.3 | Population structure in long-distance migrants

Highly mobile organisms, such as migratory birds, are interesting cases for studying the

effects of spatial isolation on population structure. On one hand, highly mobile species are

expected to have high dispersal ability, which should decrease the strength of physical or

ecological barriers to gene flow, resulting in shallow structure, or even panmixia, across large

geographic areas. Accordingly, a negative relationship between dispersal ability and

population structure has been demonstrated in various taxa (Bohonak, 1999; Medina et al.,

2018). In birds, both migratory tendency and flight capacity *per se* have been linked with

increased dispersal and decreased population structure and speciation (Arguedas & Parker,

2000; Claramunt et al., 2012). By this reasoning, we should expect unimpeded gene flow in

an extreme long-distance migrant like the bar-tailed godwit, which can fly distances >11,000

km across formidable ecological barriers without resting, eating, or drinking (Gill et al.,

2009; Piersma et al., 2022), predicting negligible sub-specific population genetic structure.

Paradoxically, migration has also been linked with increased rates of speciation and

diversification in birds (Rolland et al., 2014; Winger et al., 2014). Long-distance migration is

associated with high site-fidelity and rigid behavioral and physiological traits that enable

timely exploitation of predictable seasonal resources across hemispheres (Åkesson et al.,

2017; Conklin et al., 2013; Winger et al., 2019). These traits predict low effective dispersal

among neighboring populations, either through strict spatial or temporal isolation, or by

preventing successful recruitment by immigrants (Nosil et al., 2005). In this light, we might

expect bar-tailed godwits to exhibit strong structure among flyway populations.

Within this species, we see evidence for both ends of the continuum: in the west, negligible

structure between two populations with adjacent breeding areas but separate ranges in the

non-breeding season (*taymyrensis* and *yamalensis*), and in the east, clinal structure within a

single subspecies, where individuals freely mix throughout the non-breeding season (*baueri*).

The latter situation, involving birds making the longest non-stop migratory flights on earth,

demonstrates the weak link between dispersal *ability* (in terms of flight capacity) and actual

dispersal. In fact, bar-tailed godwits are known for extremely rigid site-fidelity at non-

breeding and migratory staging sites (Conklin et al., 2013; Rakhimberdiev et al., 2018), and

are presumed to show similar fidelity to breeding sites as adults. The very fine-scale structure

we demonstrate within the Alaska breeding range suggests that natal dispersal is also quite

restricted in *baueri* (i.e. relative to distances of c. 400–800 km between our sampling sites;

Figure 1). As we have no reason to expect weaker site-fidelity in other bar-tailed godwit

populations, we presume the weak differentiation between *taymyrensis* and *yamalensis*,

which may meet at pre- and post-breeding areas and breed <500 km from each other (Bom et

al., 2022), is a result of their recent divergence (c. 6,000 ybp) rather than high levels of gene

flow.

While not clearly related to dispersal and gene flow, flight capacity may be more indicative

of a species' propensity for exploration and colonization of new migratory flyways. As

illustrated in other long-distance migratory shorebirds, such as dunlin, *Calidris alpina*

(Wenink et al., 1996), and red knot, *C. canutus* (Conklin et al., 2022), such species appear to

respond relatively quickly to major climatic changes, such as retreat of glacial ice sheets,

with establishment of new flyway populations (as we have demonstrated with the *lapponica*

subspecies). As migration of adult birds tends to be more conservative, such 'innovations' in

a population may result largely from inexperienced juveniles conducting their first migration

(Cresswell, 2014; Verhoeven et al., 2022). Intriguingly, bar-tailed godwits and other long-

distance migrants also seem flexible (at least in evolutionary timescales) with regard to

migration distance. For example, *lapponica* and *yamalensis*, among the youngest of bar-tailed

godwit subspecies, migrate as little as 2,500–5,000 km between breeding and non-breeding

grounds, whereas the other four subspecies travel one-way distances of 10,000–18,000 km

(Figure 1). This shows that behavioral and physiological 'upscaling' to accommodate the

world's longest flights (Hedenström, 2010) is not a one-way journey; it may be reversed,

with regression toward short-distance migration, if the ecological opportunity arises.

ACKNOWLEDGEMENTS

We thank Per Palsbøll and Ritsert Jansen for guidance and help to develop the funding proposal, and to members of the Palsbøll and Fontaine laboratories at the University of Groningen and Montpellier for helpful discussions during project conception and analysis, respectively. We thank Anneke Bol at NIOZ Royal Netherlands Institute for Sea Research for DNA extractions of Mauritania samples; Marco van der Velde for laboratory assistance in Groningen; Eric Johnson and Paul Etter (SNPsaurus) for sequencing, bioinformatics, and technical expertise; and the Centre for Information Technology of the University of Groningen (particularly Bob Dröge and Cristian Marocico) for technical support and access to the Peregrine high-performance computing cluster. For providing samples, we thank Massey University, Moscow Lomonosov State University Zoological Museum, NIOZ, Royal Ontario Museum (Oliver Haddrath and Mark Peck), U.S. Geological Survey Alaska Science Center, University of Groningen, and University of Washington Burke Museum. For godwit capture and field sampling, we thank Luke DeCicco, Jimmy de Fouw, Maks Dementyev, Nick Hajdukovich, Jim Johnson, Murray Potter, Adrian Riegen, David Melville, Dan Mulcahy, Adrian Riegen, Rob Schuckard, Craig Steed, and Cleland Wallace, plus numerous local volunteers. In Australia, we thank Broome Bird Observatory and the Australasian Wader Studies Group for logistical and field work support, and we acknowledge the Yawuru People via the offices of Nyamba Buru Yawuru Limited for permission to capture godwits on the shores of Roebuck Bay, traditional lands of the Yawuru people. Sampling was supported by the David and Lucile Packard Foundation (Alaska, Australia, and New Zealand), BirdLife Netherlands and WWF-Netherlands (Australia), and a Royal Society of New Zealand Marsden Fund grant to P.F.B. and Andrew Fidler (New Zealand). The Prins Bernhard Cultuurfonds grant to T.P. started up the long-term efforts in Mauritania, supported now by

operational grants from NIOZ; we thank the authorities of the Parc National du Banc

d'Arguin for access and support. Work in the Wadden Sea was supported by Waddenfonds

(project Metawad, WF209925) and operational funding from NIOZ. Work in Oman was

financially supported by the Research Council (TRC) of the Sultanate of Oman

(ORG/EBR/12/002). The core of this project was supported by a Dutch Research Council

(NWO) grant to T.P. (ALW-Open Programme grant, 'Ecological drivers of global flyway

evolution' 824.01.001). Any use of trade, product or firm names is for descriptive purposes

only and does not imply endorsement by the U.S. Government.

AUTHOR CONTRIBUTIONS

J.R.C., Y.I.V., M.C.F. and T.P. conceived and designed the study. P.F.B., R.A.B., R.E.G.,

C.J.H., J.t.H., D.R.R., T.L.T., N.W., and P.S.T. organized and performed field sampling

and/or maintained individual resight-history databases. J.R.C. and Y.I.V. curated samples and

conducted the labwork. J.R.C., M.J.M.L. and M.C.F. analysed and interpreted the data, with

assistance from Y.I.V. J.R.C. and M.C.F. wrote the manuscript, with major contributions

from Y.I.V. and T.P. All authors provided feedback and helped edit the manuscript.

DATA AVAILABILITY STATEMENT

Demultiplexed nextRAD short-read data were deposited in NCBI's SRA archives under

BioProject ID xxx (TBA). Associated files (VCF and metadata) were deposited in DRYAD

(doi: TBA).

BENEFIT-SHARING STATEMENT

All genetic samples were collected with collaboration and/or consultation with local

communities, and all principal collaborators who provided samples are included as co-

authors. The research addresses an international priority concern, which is the conservation

of the study organism. Benefits from this research accrue from the sharing of our data and

results with the scientific and stakeholder communities, both directly and on public databases

as described above.

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Figure 1. Global distribution and sampling of bar-tailed godwits. For each of six recognized
subspecies (indicated by colour), arrows indicate general migration routes between northern
breeding areas and southern non-breeding (i.e. boreal winter) areas (coloured areas).
Numbers indicate total individuals sampled in each area (black circles); three shades of blue
indicate sampling for *baueri* North, Central, and South. Letters refer to sampling sites (in
black; see Table S1 for details) and other locations mentioned in the text (in gray): A,
Murmansk, Russia; B, Wadden Sea, The Netherlands; C, Banc d'Arguin, Mauritania; D,
Taimyr Peninsula, Russia; E, Barr al Hikman, Oman; F, Roebuck Bay, Australia; G, SE
Chukotka, Russia; H, Colville River, Alaska; I, Seward Peninsula, Alaska; J, Yukon-
Kuskokwim Delta, Alaska; K, North Island, New Zealand; L, South Island, New Zealand; M,
West Siberian Lake; N, West Siberian Plain; O, Lena River Delta; P, Bering Land Bridge; Q,
Mackenzie River Delta. Dashed box indicates approximate extent of the Beringia region.

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Figure 2. (a) Population structure estimated by principal component analysis. The individual
scores (*n* = 184 individuals) for the first three principal components (PC1–3) are shown. The
scree plot (inset) indicates the proportion of explained genetic variance by each PC, with
PC1–3 explaining 4.0% of total variation. (b) Individual genetic ancestries assigned to major
clusters for *K* = 2–5 estimated using ADMIXTURE. At each value of *K*, the ancestry
proportions for the 184 individuals for the dominant solution were determined by
CLUMPAK summary of 20 replicate runs. Numbers in parenthesis indicate proportion of
replicate runs contributing to the dominant solution (see Figure S2 for minor clusters for *K* =
3–5). Names below plot indicate sampling locations (see Figure 1 and Table S1); colors
indicate *a priori* purported breeding populations.

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Figure 3. (a) Unrooted neighbour-joining tree, based on Nei's minimum distance. Numbers
indicate bootstrapped node confidence (%). (b) AdmixtureGraph showing best graph with
two migration reticulations (*m* = 2), arbitrarily rooted with *baueri* South population (see
Methods). Numbers indicate relative genetic distances (unlabeled branches = 0 distance).
Dotted lines represent admixtures, with percentages indicating admixture proportions, and
arrows indicating direction of admixture.

Figure 4. Scenarios tested in Step 2 of DIYABC-RF analysis: six possible scenarios for the origin of *menzbieri*, by either simple branching (a,b) or by admixture of the two main branches at relatively ancient (c), intermediate (d,e) or recent (f) time periods. Extant (sampled) populations are indicated by colours; inferred historical populations are shown in shades of grey. The best-supported scenario was (c), followed by (e), as inferred by the proportion of Random Forest classification votes; see Figure S5 and Tables S9 and S10 for details.

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Figure 5. Best-supported Scenario in Step 2 of DIYABC-RF analysis (see Figure 4), scaled
to relative time-parameter estimates (converted to years assuming a generation time of 8
years) for five divergence events (branches) and two admixture events (horizontal bars). On
left, posterior distributions for each time parameter estimate, indicating mode, mean, median,
and interval of 5th–95th quantiles (vertical line). For reference, estimated timing of Last
Glacial Maximum, Holocene Climatic Optimum, and most recent presence of Bering Land
Bridge and glaciation of Europe are shown.

Table 1. Mean population pairwise F_{ST} and 95% confidence intervals of estimates. All comparisons are significantly different from zero (1,000
permutations; $*p = 0.038$, $**p < 0.001$).
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	lapponica	*taymyrensis* **Europe**	*taymyrensis* **Africa**	*yamalensis*	*menzbieri*	*baueri* **North**	*baueri* **Central**
taymyrensis **Europe**	0.016** [0–0.042]						
taymyrensis **Africa**	0.014** [0–0.040]	0.001* [0–0.014]					
yamalensis	0.015** [0.0003–0.040]	0.003** [0–0.016]	0.002** [0–0.010]				
menzbieri	0.026** [0.001–0.048]	0.011** [0.002–0.024]	0.010** [0.004–0.020]	0.013** [0.006–0.022]			
baueri **North**	0.048** [0.031–0.073]	0.032** [0.021–0.045]	0.031** [0.023–0.042]	0.034** [0.026–0.045]	0.016** [0.007–0.026]		
baueri **Central**	0.047** [0.028–0.071]	0.032** [0.018–0.053]	0.031** [0.019–0.045]	0.033** [0.022–0.048]	0.015** [0.003–0.031]	0.007** [0–0.027]	
baueri **South**	0.046** [0.030–0.070]	0.031** [0.021–0.044]	0.030** [0.024–0.040]	0.033** [0.026–0.040]	0.014** [0.007–0.024]	0.009** [0–0.020]	0.006** [0–0.023]

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