Association of diet and inflammation with the vaginal microbiota of pregnant individuals with or without IBD

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Why was the study conducted?

An altered vaginal microbiota has been implicated in preterm birth. There is no research on the vaginal microbiome and the factors that influence it in pregnant individuals with Inflammatory Bowel Disease (IBD) at a higher risk of preterm delivery.

Key findings

Pregnant individuals with IBD exhibit a comparable vaginal microbiome to healthy pregnant individuals. However, pregnant individuals with IBD present a vaginal immune profile characterized by increased levels of Th17 pro-inflammatory cytokines. High dietary quality, and optimal consumption of vegetables and added sugars were associated with vaginal dominance by the beneficial *L. crispatus*.

What does this add to what is known?

Our results indicate that the vaginal immune environment and not the microbiome might explain poor pregnancy outcomes for individuals with IBD. Moreover, our study supports the importance of diet to favor *L. crispatus,* a bacterium associated with a lower risk of preterm birth.

Abstract

Background and aims: Vaginal dysbiosis has been associated with adverse pregnancy outcomes. Here, we characterized the vaginal microbiota of pregnant individuals with inflammatory bowel disease (IBD) and investigated whether gut or vaginal inflammation and diet influence the vaginal microbiota diversity of these individuals.

Study Design: We recruited 48 individuals in their third trimester of pregnancy (IBD=23 and HC=18). We characterized the vaginal microbiota by *16S rRNA* sequencing and the gut microbiota by shotgun sequencing. We measured fecal calprotectin in stool and pro-inflammatory cytokines in vaginal fluids. We determine dietary quality using validated 24-hour dietary recalls.

Results: Pregnant individuals with IBD exhibit higher levels of fecal calprotectin and increased expression of Th17 pro-inflammatory cytokines (i.e., IL-6, IL-8, IL-17) in the vaginal mucosa compared to healthy pregnant individuals. High fecal calprotectin correlated with high vaginal microbiota diversity. Also, IL-4 (reduced in IBD) was associated with vaginal microbial composition. Regardless of IBD status, pregnant individuals with healthier diets and particularly optimal servings of vegetables and sugars exhibited a vaginal microbiota dominated by *Lactobacillus crispatus*, a species associated with a lower risk of preterm birth and bacterial vaginosis.

Conclusion: Besides gut inflammation, pregnant individuals with IBD also exhibit a Th17 immune tone in the vaginal mucosa. The vaginal microbiota diversity or composition, particularly high in the beneficial *L. crispatus,* is positively associated with healthier diets, regardless of IBD status.

Keywords: Vaginal, microbiota, IBD, inflammation, pregnancy

Introduction

The vaginal microbiota composition of pregnant individuals with IBD is stable throughout pregnancy but has not been directly compared to a healthy cohort (1). Only one study has reported that pregnant individuals with IBD were more likely to have vaginal infections compared to healthy pregnant individuals (2). In general, healthy pregnancies are characterized by vaginal microbiotas with low bacterial diversity and dominant *Lactobacillus* species (3). For instance, pregnant individuals with a vaginal microbiota dominated by *L. crispatus* have a lower risk of preterm birth compared to individuals with low-lactobacilli abundance (4). Moreover, vaginal microbial communities dominated by *L. crispatus* are better protected against infections than those dominated by *L. iners* (5). Therefore, understanding *Lactobacillus* species dominance in individuals with IBD is important to predicting their risk of poor pregnancy outcomes. Our goal is to characterize and compare the vaginal microbiota of pregnant individuals with and without IBD using high-throughput microbiota sequencing.

Moreover, we sought to determine the role of environmental factors, such as diet and local inflammation on the vaginal microbiota composition. To our knowledge, there have been only a few studies evaluating the influence of diet on the vaginal microbiota of pregnant individuals using high throughput microbiota sequencing (6-8), yet none of the studies included all the relevant diet components from validated instruments aiming at measuring dietary intake. Additionally, the immune tone of the vaginal mucosa in pregnant women with IBD remains understudied. Hence, we will test whether diet influences the vaginal microbiota makeup thus diet can be potentially used as a strategy to revert vaginal dysbiosis during pregnancy, and whether an inflammatory environment in the gut and vagina are linked to vaginal microbiota profiles.

Materials and Methods

Recruitment: We conducted a case–control study nested into our ongoing MELODY Trial (9) including participants with 27th–29th weeks of gestation before any dietary intervention. Pregnant women with and without IBD were recruited nationwide under approved IRB protocol (IRB # H00016462) as previously described (10) (see supplementary methods). IBD disease activity was evaluated using validated scoring systems, namely the Harvey Bradshaw index (11) for participants with CD and the 6-point Mayo score (12) for participants with UC.

Sample collection: Vaginal and stool samples were self-collected using the OMNIgeneVAGINAL collection tube (DNA Genotek, Canada) or the ALPCO EasySampler kit (ALPCO, USA) following manufacturer instructions. Samples were received in the lab frozen 30h after sample collection.

Nucleic acid isolation: DNA from both vaginal and stool samples was isolated with Dneasy PowerSoil Pro kit (QIAGEN, Germany) and the RNA from vaginal samples was isolated with PowerMicrobiome kit (QIAGEN, Germany) following the manufacturer's protocol.

Cytokine expression: RNA from vaginal samples was subjected to qRT–PCRs (see supplementary methods). Oligonucleotides used to estimate cytokine expression are listed in **Table S1** (Integrated DNA Technology, USA).

Fecal calprotectin quantification was performed using the CalproLab ELISA ALP (Svar Life Sciences, Norway) according to the manufacturer's instructions. Total protein was quantified using the Pierce BCA Protein Assay kit (Thermo Fisher Scientific, USA). Calprotectin was normalized to initial stool weight (ng calprotectin/mg stool).

Vaginal microbiota sequencing and profiling was performed by *16S rRNA* sequencing of the V3-V4 hypervariable region as previously described (13). Sequencing libraries were sequenced on 600 cycles

using the MiSeq platform (Illumina, CA, USA). QIIME2 was used to process paired-end sequences. The DADA2 (14) algorithm was used for quality control and to obtain representative sequences (Amplicon Sequence Variant or ASV). We used a custom database (15, 16) for taxonomy classification. Only taxa with at least 0.10% abundance and present in a minimum of one sample were used for the analyses, as previously done (17, 18). **Table S2** describes the sequence counts included in the analyses.

Gut microbiota sequencing and profiling were performed using whole genome sequencing. Specifically, library generation and 150bp paired-end sequencing were carried out on the Illumina NextSeq 500 platform. KneadData (dec_v0.1, http://huttenhower.sph.harvard.edu/kneaddata) was used to eliminate human sequences and for quality control. MetaPhlAn4 database (vOct22)(19) was used for taxonomic assignment. **Table S2** describes the sequence counts included in the analyses.

Vaginal and gut microbiota diversity analyses were done in R, particularly the Phyloseq package (20). We performed data imputation for two individuals lacking BMI information (IBD=2), and four individuals (IBD=2, HC=2) without fecal calprotectin measurements using the median values for the IBD or HC group, respectively. All analyses were performed at the ASV level. Microbial alpha diversity was estimated using the Shannon and Simpson's Indexes (1-D) with rarefied sequencing data. Shannon and Simpson's Index were log-transformed to reach 'normality' of the residuals when necessary. To determine associations in alpha diversity we used linear regression models corrected by cofounders as age, body mass index (BMI), antibiotic use, and gestational diabetes. Results for the best-fitted model are reported.

We used Permutational Multivariate Analysis of Variance (PERMANOVA (21)) with adonis2 function to evaluate beta diversity (measured by Aitchison distance) of non-rarefied sequencing data.

We only assessed the associations between the vaginal microbiota and cytokines that were significantly different between IBD and HC. When assessing the association with dietary components, we only included

142 dietary components with no collinearity (Spearman correlation >|0.5|). One participant did not complete
143 dietary recalls and thus was excluded from diet analyses.
144
145 **Discriminant taxa analysis:** Microbial taxa and their association with clinical variables were assessed
146 using MaAsLin2's (22), also including the previously named confounding variables.
147
148 **Community State Types (CST):** Each vaginal sample was classified into CST as described before (23).
149 To compare CST by discrete variables (health status) we use Fisher exact test (24); and by continuous
150 variables (i.e., fecal calprotectin, cytokine expression, or diet score) Kruskal-Wallis test.
151
152 **Dietary assessment:** We conducted 24-hour dietary recalls (24HDRs) around the same time as
153 vaginal/stool collection and estimated the Healthy Eating index-2015 (HEI-2015) as previously described
154 by us (10) (see supplementary methods). Wilcoxon test was used to compare HEI-2015 and its dietary
155 components by health status and CST.
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164 ## Results

165 A total of 48 pregnant individuals were enrolled in the study: 23 with diagnosed IBD (n=18 CD, and n=5,
166 UC) and 25 HC without IBD. Participants' demographics and clinical information are summarized in **Table**
167 **1**. Briefly, participants' mean age was 33.8 years, most had normal BMI (41.7%) or were overweight
168 (37.5%), and most self-identified as White (93.8%). Only a few participants reported gestational diabetes
169 or the use of antibiotics currently or previously during pregnancy. None of the clinical and demographic
170 variables differed by health status (IBD vs. HC, **Table 1**) or IBD diagnosis (CD vs. UC, **Table S3**). More
171 than 50% of the IBD participants were in self-reported remission (**Table 1**).

172

173 ***Vaginal microbiota diversity is associated with gut and vaginal inflammation, regardless of health***
174 ***status.***

175 We first determined whether the vaginal microbiota diversity and composition differed by health status and
176 found no significant differences (**Figure S1**). We also found no significant differences in the vaginal
177 microbiota by IBD medications or when comparing individuals with IBD not on medication with the HC
178 (**Figure S2**).

179

180 Given that most of the study participants were in remission or with mild disease we sought to further
181 determine inflammatory markers that could influence the vaginal microbiota. We observed significantly
182 higher levels of fecal calprotectin in IBD participants compared to their HC counterparts. Concomitantly,
183 fecal calprotectin levels were positively associated with vaginal microbiota Simpson diversity but not with
184 Shannon index (**Figure 1 and S1**) or for the beta diversity (**Figure S1**). Additionally, there were no
185 significant associations between fecal calprotectin and vaginal microbiota diversity by health status (**Figure**
186 **S1**).

187

188 We then assessed inflammation in the vaginal mucosae. We observed that pregnant individuals with IBD
189 exhibited higher gene expression of Th17 pro-inflammatory cytokines, specifically IL-6, IL-8, and IL-17 than

190 HC. Conversely, expression of Th1 and Th2 cytokines IFN-γ and IL-4 respectively, was lower in pregnant
191 individuals with IBD compared to HC (**Figure 1**). We observe that the vaginal microbiota composition – not
192 its alpha diversity- correlated with the expression of IL-4 in the vaginal mucosa (**Figures 1 and S3**).
193
194 There were no specific bacteria associated with any of the inflammatory markers studied. We only identify
195 *Dialister* sp. and WAL 1855D (an uncultured bacterium from the *Tissierellaceae* family) increased in
196 pregnant individuals with high BMI (**Figure S4**).
197
198 Finally, we examined the gut microbiota of the pregnant individuals included in the study. There were no
199 significant differences in the gut microbiota diversity and composition by health status or fecal calprotectin
200 levels (**Figure S5**). However, we observed that *Collinsella* was lower in pregnant individuals with IBD
201 compared to HC (**Figure S5**).
202
203 ***Vaginal microbial diversity is associated with the consumption of vegetables and added sugars.***
204 Pregnant individuals in this cohort had a HEI-2015 score of 63.8, comparable to the average of 63.0
205 reported by pregnant individuals in the USA (25, 26). There were no significant differences in the HEI-2015
206 score by health status (**Table S5**). Similarly, there were no significant differences in vaginal microbial
207 diversity or composition by HEI-2015 score (**Figure S6**).
208
209 We further investigated the associations between each dietary component included in the HEI-2015 score
210 and the vaginal microbiota diversity. Consumption of each dietary component was similar between
211 pregnant individuals with IBD and HC (**Table S5**). Whole fruit, fatty acid, and seafood/plant protein were
212 excluded from the analysis due to collinearity (Spearman score >|0.5|). We observed that high scores of
213 total vegetables (high vegetable intake) were predictive of a high vaginal microbiota diversity (**Figure 2**)
214 along with increasing abundance of *L. crispatus* (**Figure 3**). *L. iners* showed an opposite trend, although
215 not significant (**Figures 3 and S7**). We observed that vaginal microbial composition differed by added
216 sugar score, although no bacterial taxon was significantly associated with it (**Figure 2**).

Higher Lactobacillus crispatus *dominance (CST-I) is associated with higher dietary quality and vegetable consumption.*

We determine the vaginal CST for each participant. More than 40% exhibited CST-I (*L. crispatus* dominance), followed by CST-III (*L. iners* dominance*)*, CST-II (*L. gasseri* dominance*)*, CST-IV (non-*Lactobacillus*-dominated), and CST-V (*L. jensenii* dominance. **Table S4**) (27). We did not observe differences in CSTs by health status, IBD diagnosis (CD vs. UC), levels of fecal calprotectin, or any of the vaginal cytokines assessed (**Table S3 and S4**).

Pregnant individuals with CST-I (*L. crispatus*-dominated, N=21) showed a higher HEI-2015 score, reflective of better dietary quality than CST-III individuals (*L. iners*-dominated, N=14. **Figure 3)**. Moreover, scoring for vegetable and added sugar intakes was significantly higher in individuals with CST-I than those with CST-III (**Figure 3**).

Discussion

Principal findings

The vaginal and gut microbiota composition of pregnant individuals with IBD was comparable to HC during the third trimester of pregnancy. Yet pregnant individuals with IBD presented gut inflammation linked to high vaginal microbial diversity and a vaginal pro-inflammatory immune tone. Pregnant individuals with higher dietary quality and optimal consumption of vegetables exhibited a vaginal microbiota profile dominated by the beneficial *L. crispatus*.

Results in the context of what is known.

Reduced levels of IFN-γ and IL-4 and elevated levels of IL-8 and IL-6, as seen in the pregnant individuals with IBD in this study, have been associated with an increased risk of preterm birth (28). To the best of our knowledge, our study is the first one to assess vaginal inflammatory markers in IBD pregnant patients and might explain the increased risk of preterm birth of pregnant individuals with IBD, even when in remission

243 or with mild disease (29, 30). Despite differences in the vaginal immune profile, the vaginal microbiota of
244 pregnant individuals with IBD was comparable to healthy controls. This result is unexpected as
245 inflammation relates to alteration in the microbiota in the vaginal mucosa (31, 32). Here, the vaginal
246 immune profile was assessed by gene expression (by RT-qPCR) and no protein/cytokine levels; thus,
247 further studies are required to accurately evaluate the immune tone in the vaginal mucosa.
248
249 We observe that gut inflammation in IBD individuals is positively associated with increased vaginal
250 microbial diversity. Of note, contrary to the gut microbiota, high microbial diversity in the vagina relates to
251 unhealthy states with increased risk for bacterial vaginosis (reviewed in (33)) and preterm birth (34, 35).
252
253 We then investigated whether diet impacts the vaginal microbiota. Pregnant individuals with higher dietary
254 quality exhibited a vaginal microbiota profile dominated by the beneficial *L. crispatus,* whereas the vaginal
255 microbiota profile of those with lower dietary quality was dominated by *L. iners*. For individual dietary
256 components, our findings showed that high vegetable consumption was associated with greater microbial
257 diversity (linked to vaginal dysbiosis (36)), similar to what previous studies have found for non-pregnant
258 vegetarians compared to non-vegetarians (37). However, our results also showed that, despite the higher
259 diversity, high vegetable intake resulted in a greater abundance of the beneficial *L. crispatus*. This indicates
260 the importance of not only considering a diversity index but identifying members of the vaginal microbiome
261 at the specie level to understand the potential implications of the microbiota in vaginal health.
262
263 Additionally, we found that lower added sugar intake resulted in decreased microbial alpha diversity and
264 increased *L. crispatus*. Concomitantly, high vegetable consumption and low intake of sweetened
265 beverages, has been also positively associated with abundance of *L. crispatus* in White and Black pregnant
266 women (8).
267
268 Importantly, *L. crispatus* is believed to offer the most protective benefits to the host compared to other
269 *Lactobacillus* species, with *L. iners* offering the least protective benefits (reviewed in (5)). *L. crispatus*

270 creates a highly acidic vaginal niche (pH<4.5) inhospitable to non-beneficial microbes, such as bacterial
271 vaginosis-related bacteria (38).
272
273 **Clinical implications**
274 We find that intestinal inflammation correlates with high vaginal microbiota diversity, indicative of unhealthy
275 states with increased risk for bacterial vaginosis and preterm birth. Thus, our results highlight the
276 importance of continuing therapy during pregnancy to reduce IBD-related intestinal inflammation.
277 Moreover, we found that diet can influence the dominance of a beneficial *L. crispatus* associated with
278 decreased risk of pre-term birth and bacterial vaginosis. Hence, emphasizing dietary quality during
279 pregnancy is a must, not only for the sustainment of pregnancy but to fuel a healthy vaginal microbiota.
280
281 **Research implications**
282 Our results highlight the need for a large prospective study that includes pregnant individuals with IBD
283 experiencing different disease activity (i.e., mild, moderate, severe). Future studies including dietary
284 interventions will unveil the role of diet as a strategy to support a healthy vaginal microbiome.
285
286 **Strengths and limitations**
287 Our study is constrained by a few key limitations. The modest sample size for both IBD and HC cohorts
288 curtails the statistical comparisons, particularly when several confounding variables, such as antibiotic use,
289 IBD medications, and gestational diabetes, are considered. This factor reduces the statistical power of the
290 study. Additionally, the IBD samples predominantly represent individuals in remission or with mild disease
291 with CD and only a few participants with UC, which narrows the scope of our conclusions to this specific
292 severity level of IBD and IBD diagnosis. Moreover, the ethnic/racial composition of our study sample, which
293 is mainly White, introduces a limitation since the vaginal microbiome is known to vary with race and
294 ethnicity (39). These limitations suggest the need for studies that include participants with severe disease,
295 an equal representation of IBD diagnosis (UC and CD), as well as individuals with diverse ethnic/racial
296 backgrounds.

297 **Conclusions**

298 Our results demonstrate that although the vaginal and gut microbiota of pregnant individuals with IBD and
299 HC is similar in the third trimester of pregnancy, it varies depending on the immune tone of each mucosa.
300 We show that pregnant individuals with IBD exhibit a pro-inflammatory cytokine profile that has been
301 associated with an increased risk of pre-term birth. Finally, a high-quality diet with optimal intakes of
302 vegetables and added sugars favors *L. crispatus* vaginal dominance.

303

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308

309 **Data availability statement**

310 The raw sequences and their associated metadata will be released at NCBI BioProject ID PRJNA915128
311 upon manuscript publication.

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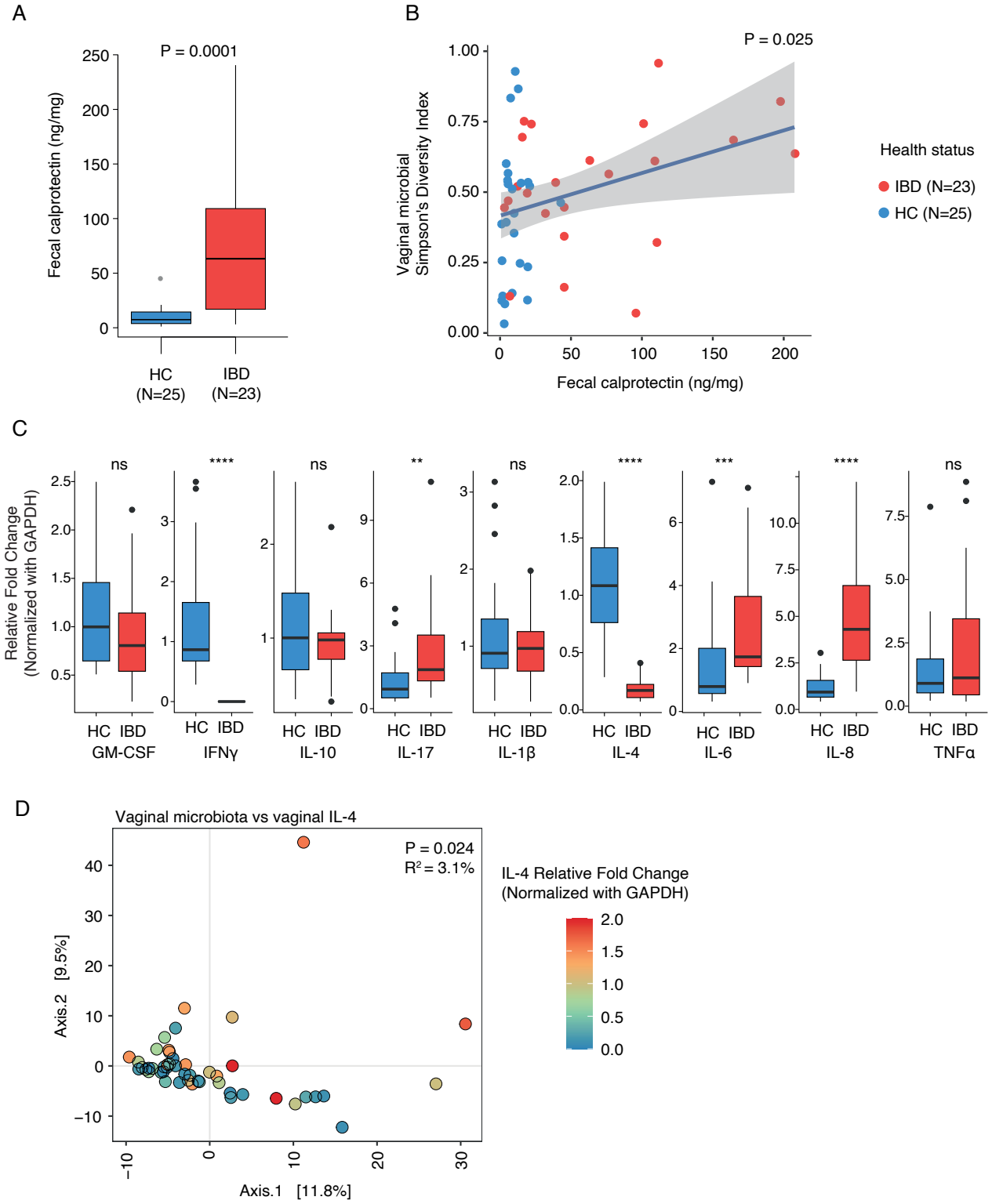

Figure 1. Variation in inflammatory markers and microbial diversity by health status. (A) Fecal calprotectin levels of individuals with Inflammatory Bowel Disease (IBD) or Healthy Controls (HC; Wilcoxon test). (B)

439 Linear regression correlation between vaginal microbial alpha diversity and fecal calprotectin levels. (C)
440 Expression of cytokines on the vaginal mucosa of IBD and HC participants (Wilcoxon test. Asterisks denote
441 ** <5E-2, *** <5E-3, ****<5E-4). (D) Principal Coordinates Analysis (PCoA) of vaginal microbial beta
442 diversity by IL-4 levels (Aitchison distance and PERMANOVA).
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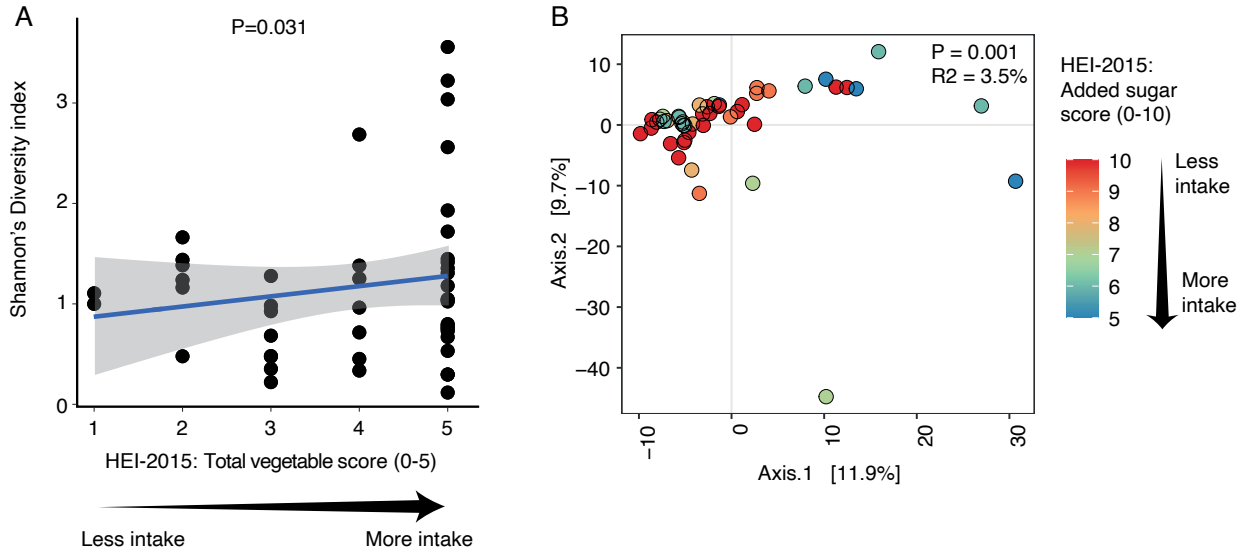

Figure 2. HEI-2015 total vegetable and added sugar components are significantly associated with vaginal microbiota diversity or composition, respectively. (A) Linear regression of microbial alpha diversity and HEI-2015 total vegetable score. Gray shades in the graphs represent the 95% confidence interval. (B) Principal Coordinates Analysis (PCoA) demonstrates significance for vaginal microbial composition by HEI-2015 added sugar score (Aitchison distance and PERMANOVA).

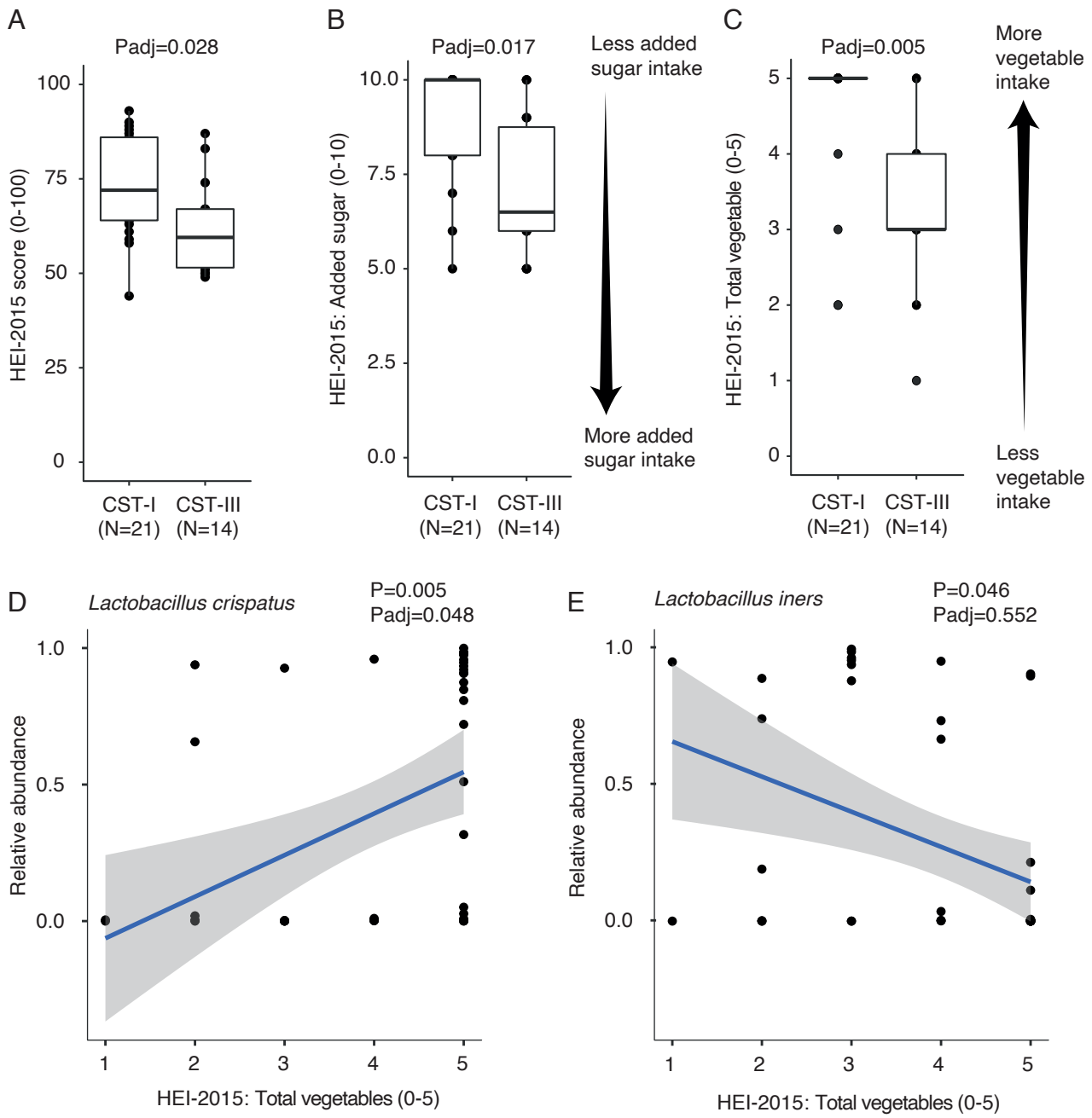

Figure 3. HEI-20215 added sugar and total vegetable correlate with CST classification. Pregnant individuals with CST-I (*L. crispatus*- dominated profile) exhibit higher scores for (A) HEI-2015, (B) added sugar, and (C) total vegetables (Wilcoxon-test, adjusting for multiple comparisons). The arrows indicate the direction of consumption of each dietary component. (D) Total vegetable score positively correlates with the abundance of *L. crispatus* and negatively correlates with the abundance of *L. iners* (E, Spearman Correlation).

470 **Table 1**. Demographic and clinical variables for pregnant individuals with Inflammatory Bowel Disease

471 (IBD) or Healthy Controls (HC) recruited for the study between 2019 and 2022.

Demographics and clinical variables	IBD (N=23)	HC (N=25)	Overall (N=48)	P value[&]
Age				*0.414*
Mean (SD)	33.3 (4.63)	34.4 (4.97)	33.8 (4.79)	
Median [Min, Max]	33.0 [22.0, 41.0]	36.0 [22.0, 42.0]	34.0 [22.0, 42.0]	
BMI categories [&&]				*0.179*
Underweight	1 (4.3%)	0 (0%)	1 (2.1%)	
Normal	12 (52.2%)	8 (32.0%)	20 (41.7%)	
Overweight	8 (34.8%)	10 (40.0%)	18 (37.5%)	
Obese	2 (8.7%)	7 (28.0%)	9 (18.8%)	
Race				*0.490*
White	23 (100%)	22 (88.0%)	45 (93.8%)	
Asian	0 (0%)	1 (4.0%)	1 (2.1%)	
Other	0 (0%)	2 (8.0%)	2 (4.2%)	
Gestational diabetes				*0.098*
Yes	3 (13.0%)	0 (0%)	3 (6.3%)	
No	16 (69.6%)	21 (84.0%)	37 (77.1%)	
Information unavailable	4 (17.4%)	4 (16.0%)	8 (16.7%)	
Use of antibiotic during all pregnancy				*0.468*
No	19 (82.6%)	22 (88.0%)	41 (85.4%)	
Yes	3 (13.0%)	3 (12.0%)	6 (12.5%)	
Information unavailable	1 (4.3%)	0 (0%)	1 (2.1%)	
IBD diagnosis				-
Crohn's disease	18 (78.3%)	0 (0%)	18 (37.5%)	
Ulcerative colitis	5 (21.7%)	0 (0%)	5 (10.4%)	
Healthy controls	0 (0%)	25 (100%)	25 (52.1%)	
IBD disease activity [&&&]				-
Mild disease	6 (26.1%)	0 (0%)	6 (12.5%)	
Remission	13 (56.5%)	0 (0%)	13 (27.1%)	
Information unavailable	4 (17.4%)	25 (100%)	29 (60.4%)	
Use of IBD medication				*1.70E-06*
No	9 (39.1%)	25 (100%)	34 (70.8%)	
Yes	14 (60.9%)	0 (0%)	14 (29.2%)	
Preterm				*0.106*
No	15 (65.2%)	17 (68.0%)	32 (66.7%)	
Yes	4 (17.4%)	0 (0%)	4 (8.3%)	
Information unavailable	4 (17.4%)	8 (32.0%)	12 (25.0%)	
Infant birth weight (g)				*0.151*
Mean (SD)	3150 (470)	3330 (662)	3240 (579)	
Median [Min, Max]	3230 [1810, 3710]	3290 [1080, 4520]	3230 [1080, 4520]	

[&]Fisher's exact test for categorical variables and Wilcoxon test for continuous variables.

[&&]BMIs categories correspond to the WHO's classifications: Underweight (<18.5), normal weight (18.5–24.9), overweight (≥25.0), and obese (≥30).

[&&&]Disease activity was estimated using the Harvey Bradshaw Index and the Mayo score for individuals with Crohn's Disease or Ulcerative colitis, respectively.

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474 **SUPPLEMENTARY MATERIAL**

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476 **METHODS:**

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478 **Recruitment:** We conducted a case–control study nested into our ongoing MELODY Trial, which is a
479 prospective non-randomized diet intervention trial testing the effects of IBD anti-inflammatory diet during
480 the third trimester of pregnancy on maternal IBD activity and microbiome composition in mothers and their
481 babies(9). Written informed consent was obtained from all eligible participants. Eligible participants were
482 adult (18+ years of age) pregnant individuals with or without IBD, with less than 29 weeks of pregnancy,
483 carrying a singleton, with or without IBD diagnosis. We excluded pregnant individuals with scheduled C-
484 section or induction of vaginal delivery prior to week 37 at the time of enrollment, those with medical
485 conditions that required a special diet, or those unable to speak or understand English.

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487 **Cytokine expression:** RNA from vaginal samples was reverse transcribed using iScript cDNA Synthesis
488 Kit (Bio-Rad, USA), and qRT–PCRs were performed using iTaq Universal SYBR Green-Supermix (Bio-
489 Rad, USA) in an Applied Biosystems ViiA7 Real-Time PCR machine (Thermo-Scientific, USA). Expression
490 of each cytokine was measured in triplicate and the mean was normalized by the expression of a
491 housekeeping gene (GAPDH). The $2^{(-\Delta\Delta Ct)}$ method was used to analyze the relative changes in gene
492 expression.

493

494 **Dietary assessment:** We conducted three, 24-hour dietary recalls (24HDRs) around the same time of
495 vaginal/stool collection. 24HDRs were performed using the USDA Automated Multiple Pass Method(40) in
496 conjunction with the University of Minnesota Nutrition Data for Research (NDSR) software (Current
497 version: NDS-R 2022)(41-44) as we have previously done(45-47). We estimated the Healthy Eating Index
498 2015 (HEI-2015) from the average of the three 24HDRs. The population ratio method(48) was used to
499 compute the individual scores of each of the 13 HEI-2015 components: total fruits, whole fruits, total
500 vegetables, greens and beans, dairy, total protein foods, seafood and plant proteins, fatty acids, refined

501 grains, whole grains, sodium, added sugars and saturated fats. Six food groups have values from 0 to 5;
502 and seven from 0 to 10, for a total maximum score of 100 as previously described(25). Higher values
503 represent better dietary quality. Wilcoxon test was used to compare HEI-2015 and individual HEI-2015'
504 dietary components by health status and CST.
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668 **SUPPLEMENTARY TABLES**

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670 **Table S1.** List of oligos used to determine vaginal cytokine gene expression.

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Serial No	Target gene (qRT primer)	Sequence (5' - 3')
1	IL-1β (F)	CAAGAGTGCTGGAGCGATAA
2	IL-1β (R)	CCGTTACAGCGGAGAGATATAAG
3	TNFα (F)	CACAGAAGACACTCAGGGAAAG
4	TNFα (R)	TCGAGGGAGTCACCCTTAAA
5	IL-6 (F)	GGAGGAGCCAAAGCTCAATAA
6	IL-6 (R)	ACTCCACACCACAAGAAGATG
7	IL-4 (F)	CCTGCTTCATGAGGGAAACT
8	IL-4 (R)	GGTGACAGAACAAGACCCTATC
9	IL-8 (F)	GAAAGGAAGTAGCTGGCAGAG
10	IL-8 (R)	GGGTGGAAAGGTTTGGAGTAT
11	IL-10 (F)	TGGAGTGAGTCCTGGAGAAATA
12	IL-10 (R)	CTCCATGTCCATCACACTTAGG
13	IFN-γ (F)	CCAGCCATCCTCAGAAATGT
14	IFN-γ (R)	CTTGCATCTCCTCACTCTAACC
15	IL-17 (F)	CCATAGTGAAGGCAGGAATCA
16	IL-17 (R)	GAGGTGGATCGGTTGTAGTAATC
17	GM-CSF (F)	CCTCCAGCAGGAATGTCTTAAT
18	GM-CSF (R)	TCTGACTCCTTGGGATGAAATG
19	GAPDH (F)	GTGCTCCCACTCCTGATTT
20	GAPDH (R)	CCTTCTCTAAGTCCCTCCTACA

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680 **Table S2.** Overview number of sequences and amplicon sequence variants (ASVs) obtained from the

681 vaginal samples and stool samples.

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Parameter	Vaginal samples			Stool samples	
	Raw sequences	Post-filtering	Post-filtering/rarefaction*	Raw sequences	Rarefied sequences**
N samples	48	48	48	45	44
Total number of seqs	2,751,605	2,742,919	336,000	652,433,458	236,170,044
Mean seqs/sample	57,325	57,144	7,000	14,498,521	5,367,501
Standard deviation of seqs/sample	24,010	23,945	-	7,468,723	-
Max number of seqs in a sample	123,825	123,720	7,000	30,294,951	5,367,501
Min number of seqs in a sample	12,916	12,872	7,000	715	5,367,501
Total number of different ASVs or Strains	1,425	573	563	1,420	897
Mean ASV or Strains/sample	48	36	33	115	118
Standard deviation of ASV or Strains/sample	38	23	23	46	43
Max number of ASVs or Strains in a sample	208	109	105	249	249
Min number of ASVs or Strains in a sample	6	6	5	1	57

683 * Sequences were rarefied at *7,000 sequences/sample or **5,367,501 sequences/sample, each

684 representing the highest number of sequences that included all vaginal and stool samples, respectively.

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690 **Table S3.** Demographic and clinical variables for pregnant individuals with Crohn's Disease (CD) or

691 Ulcerative Colitis (UC) recruited for this study between 2019 and 2022.

Demographics and clinical variables	CD (N=18)	UC (N=5)	Overall (N=23)	P value[&]
Age				*0.822*
Mean (SD)	33.2 (4.86)	33.4 (4.22)	33.3 (4.63)	
Median [Min, Max]	33.5 [22.0, 41.0]	32.0 [28.0, 39.0]	33.0 [22.0, 41.0]	
BMI categories [&&]				*0.126*
Underweight	1 (5.6%)	0 (0%)	1 (4.3%)	
Normal	10 (55.6%)	2 (40.0%)	12 (52.2%)	
Overweight	6 (33.3%)	2 (40.0%)	8 (34.8%)	
Obese	1 (5.6%)	1 (20.0%)	2 (8.7%)	
Race				-
White	18 (100%)	5 (100%)	23 (100%)	
Asian	0 (0%)	0 (0%)	0 (0%)	
Other	0 (0%)	0 (0%)	0 (0%)	
Gestational diabetes				*0.053*
Yes	1 (5.6%)	2 (40.0%)	3 (13.0%)	
No	14 (77.8%)	2 (40.0%)	16 (69.6%)	
Information unavailable	3 (16.7%)	1 (20.0%)	4 (17.4%)	
Use of antibiotic during pregnancy				*1.000*
No	15 (83.3%)	4 (80.0%)	19 (82.6%)	
Yes	3 (16.7%)	0 (0%)	3 (13.0%)	
Information unavailable	0 (0%)	1 (20.0%)	1 (4.3%)	
Disease activity [&&&]				*0.456*
Mild disease	5 (27.8%)	1 (20.0%)	6 (26.1%)	
Remission	10 (55.6%)	3 (60.0%)	13 (56.5%)	
Information unavailable	3 (16.7%)	1 (20.0%)	4 (17.4%)	
Use of IBD medication				*0.342*
No	6 (33.3%)	3 (60.0%)	9 (39.1%)	
Yes	12 (66.7%)	2 (40.0%)	14 (60.9%)	
Preterm				*0.272*
No	12 (66.7%)	3 (60.0%)	15 (65.2%)	
Yes	2 (11.1%)	2 (40.0%)	4 (17.4%)	
Missing	4 (22.2%)	0 (0%)	4 (17.4%)	
Infant birth weight (g)				*0.411*
Mean (SD)	3100 (515)	3330 (169)	3150 (470)	
Median [Min, Max]	3230 [1810, 3710]	3230 [3180, 3540]	3230 [1810, 3710]	
Fecal calprotectin (ng/mg)				*1.000*
Mean (SD)	69.4 (65.9)	59.5 (42.7)	67.3 (60.9)	
Median [Min, Max]	45.2 [3.11, 208]	45.2 [6.89, 111]	45.2 [3.11, 208]	
Community State Types (CSTs)				*0.209*
I	7 (38.9%)	2 (40.0%)	9 (39.1%)	
II	4 (22.2%)	0 (0%)	4 (17.4%)	
III	2 (11.1%)	3 (60.0%)	5 (21.7%)	

IV-C	3 (16.7%)	0 (0%)	3 (13.0%)
V	2 (11.1%)	0 (0%)	2 (8.7%)

[&]Fisher's exact test for categorical variables and Wilcoxon test for continuous variables.

[&&]BMIs categories correspond to the WHO's classifications: Underweight (<18.5), normal weight (18.5–24.9), overweight (≥25.0), and obese (≥30).

[&&&]Disease activity was estimated using the Harvey Bradshaw Index and the Mayo score for individuals with Crohn's Disease or Ulcerative colitis, respectively.

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713 **Table S4.** Vaginal microbial Community State Types (CSTs) distribution by health status, levels of fecal

714 calprotectin, and expression of vaginal cytokines

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Variables	CST-I (N=21)	CST-II (N=7)	CST-III (N=14)	CST-IV-C (N=4)	CST-V (N=2)	Overall (N=48)	P value[&]
Health status							*0.369*
HC	12 (57.1%)	3 (42.9%)	9 (64.3%)	1 (25.0%)	0 (0%)	25 (52.1%)	
IBD	9 (42.9%)	4 (57.1%)	5 (35.7%)	3 (75.0%)	2 (100%)	23 (47.9%)	
Fecal calprotectin (ng/mg)							*0.373*
Mean (SD)	37.0 (55.8)	36.6 (47.2)	26.3 (26.5)	40.4 (47.8)	120 (124)	37.6 (51.1)	
Median	8.54	12.4	19.4	19.5	120	15.3	
IL-6							*0.114*
Mean (SD)	3.8 (4.1)	4.4 (3.4)	2.9 (2.3)	7.5 (3.9)	8.7 (0.05)	4.1 (3.7)	
Median	2.14	4.05	2.5	7.2	8.76	3.12	
IL-4							*0.474*
Mean (SD)	0.761 (0.658)	0.850 (0.762)	0.855 (0.684)	0.377 (0.301)	0.193 (0.140)	0.746 (0.651)	
Median	0.421	0.877	0.581	0.277	0.193	0.428	
IL-1							*0.086*
Mean (SD)	1.22 (0.453)	1.18 (0.403)	1.04 (0.392)	0.763 (0.519)	2.01 (0.321)	1.16 (0.469)	
Median	1.23	1.06	1.02	0.655	2.01	1.06	
TNF-alpha							*0.237*
Mean (SD)	3.46 (3.14)	4.42 (4.05)	3.00 (2.51)	2.48 (1.90)	11.2 (1.79)	3.71 (3.33)	
Median	1.32	4.74	1.69	1.93	11.2	2.1	
IFN-gamma							*0.431*
Mean (SD)	0.583 (0.671)	0.663 (0.951)	0.776 (0.751)	0.152 (0.303)	0.00340 (0.00157)	0.591 (0.715)	
Median	0.561	0.001	0.712	0.000763	0.0034	0.511	
IL-8							*0.201*
Mean (SD)	3.05 (3.05)	3.96 (4.26)	2.25 (2.12)	4.15 (4.24)	11.5 (2.62)	3.39 (3.48)	
Median	1.57	1.26	1.79	2.68	11.5	1.93	
GM-CSF							*0.076*
Mean (SD)	1.04 (0.453)	1.10 (0.561)	1.15 (0.488)	0.689 (0.565)	4.31 (0.303)	1.19 (0.815)	
Median	1.02	0.917	1.08	0.485	4.31	1.02	
IL-10							*0.176*
Mean (SD)	1.15 (0.625)	1.27 (0.745)	1.20 (0.496)	1.09 (0.500)	5.59 (0.473)	1.36 (1.06)	
Median	1	1.17	1.11	0.922	5.59	1.09	
IL-17							*0.096*
Mean (SD)	1.93 (1.20)	2.60 (1.31)	1.59 (1.08)	1.85 (1.26)	6.03 (1.09)	2.09 (1.44)	
Median	2.07	2.49	1.17	2.08	6.03	1.99	

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717 **Table S5.** HEI-2015 score and individual dietary components scoring distribution by health status.

HEI-2015 dietary components (scoring scale)	IBD (N=22)	HC (N=25)	Overall (N=47)	P value[&]
Healthy Eating Index (0-100)				*0.175*
Mean (SD)	66.6 (13.8)	61.4 (13.2)	63.8 (13.6)	
Median [Min, Max]	67.5 [45.0, 91.0]	64.0 [35.0, 84.0]	65.0 [35.0, 91.0]	
Total vegetable (0-5)				*0.361*
Mean (SD)	3.68 (1.39)	4.20 (1.12)	3.96 (1.27)	
Median [Min, Max]	4.00 [1.00, 5.00]	5.00 [1.00, 5.00]	5.00 [1.00, 5.00]	
Greens and beans (0-5)				*0.179*
Mean (SD)	4.41 (1.47)	3.48 (1.90)	3.91 (1.75)	
Median [Min, Max]	5.00 [0, 5.00]	5.00 [0, 5.00]	5.00 [0, 5.00]	
Total fruit (0-5)				*0.279*
Mean (SD)	3.23 (1.57)	2.80 (1.58)	3.00 (1.57)	
Median [Min, Max]	3.50 [0, 5.00]	3.00 [0, 5.00]	3.00 [0, 5.00]	
Whole fruit (0-5)				*0.281*
Mean (SD)	3.82 (1.76)	3.44 (1.87)	3.62 (1.81)	
Median [Min, Max]	5.00 [0, 5.00]	5.00 [0, 5.00]	5.00 [0, 5.00]	
Whole grains (0-10)				*0.728*
Mean (SD)	5.36 (3.27)	5.52 (3.27)	5.45 (3.24)	
Median [Min, Max]	5.50 [0, 10.0]	6.00 [0, 10.0]	6.00 [0, 10.0]	
Dairy (0-10)				*0.659*
Mean (SD)	6.68 (2.23)	6.76 (2.60)	6.72 (2.41)	
Median [Min, Max]	6.00 [3.00, 10.0]	7.00 [3.00, 10.0]	6.00 [3.00, 10.0]	
Total protein foods (0-5)				*0.508*
Mean (SD)	4.64 (0.727)	4.32 (1.03)	4.47 (0.905)	
Median [Min, Max]	5.00 [3.00, 5.00]	5.00 [1.00, 5.00]	5.00 [1.00, 5.00]	
Seafood and plant protein (0-5)				*0.212*
Mean (SD)	3.86 (1.83)	3.36 (1.78)	3.60 (1.80)	
Median [Min, Max]	5.00 [0, 5.00]	3.00 [0, 5.00]	5.00 [0, 5.00]	
Fatty acid ratio (0-10)				*0.162*
Mean (SD)	5.50 (3.60)	3.04 (2.94)	4.19 (3.46)	
Median [Min, Max]	5.00 [0, 10.0]	2.00 [0, 10.0]	3.00 [0, 10.0]	
Sodium (0-10)				*0.217*
Mean (SD)	5.84 (2.79)	5.82 (3.22)	5.83 (2.97)	
Median [Min, Max]	6.00 [1.00, 10.0]	6.50 [0, 10.0]	6.00 [0, 10.0]	
Refined grains (0-10)				*0.251*
Mean (SD)	6.09 (4.10)	6.76 (2.79)	6.45 (3.44)	
Median [Min, Max]	8.00 [0, 10.0]	7.00 [0, 10.0]	8.00 [0, 10.0]	
Added sugar (0-10)				*0.514*
Mean (SD)	8.18 (1.79)	8.20 (1.91)	8.19 (1.84)	
Median [Min, Max]	9.00 [5.00, 10.0]	9.00 [5.00, 10.0]	9.00 [5.00, 10.0]	
Saturated fats (0-10)				*0.847*
Mean (SD)	5.23 (3.32)	3.48 (3.00)	4.30 (3.24)	
Median [Min, Max]	5.50 [0, 10.0]	3.00 [0, 10.0]	4.00 [0, 10.0]	

& Fisher's exact test for categorical variables and Wilcoxon test for continuous variables

743 **Table S6.** HEI-2015 score and individual dietary components scoring distribution by Community State

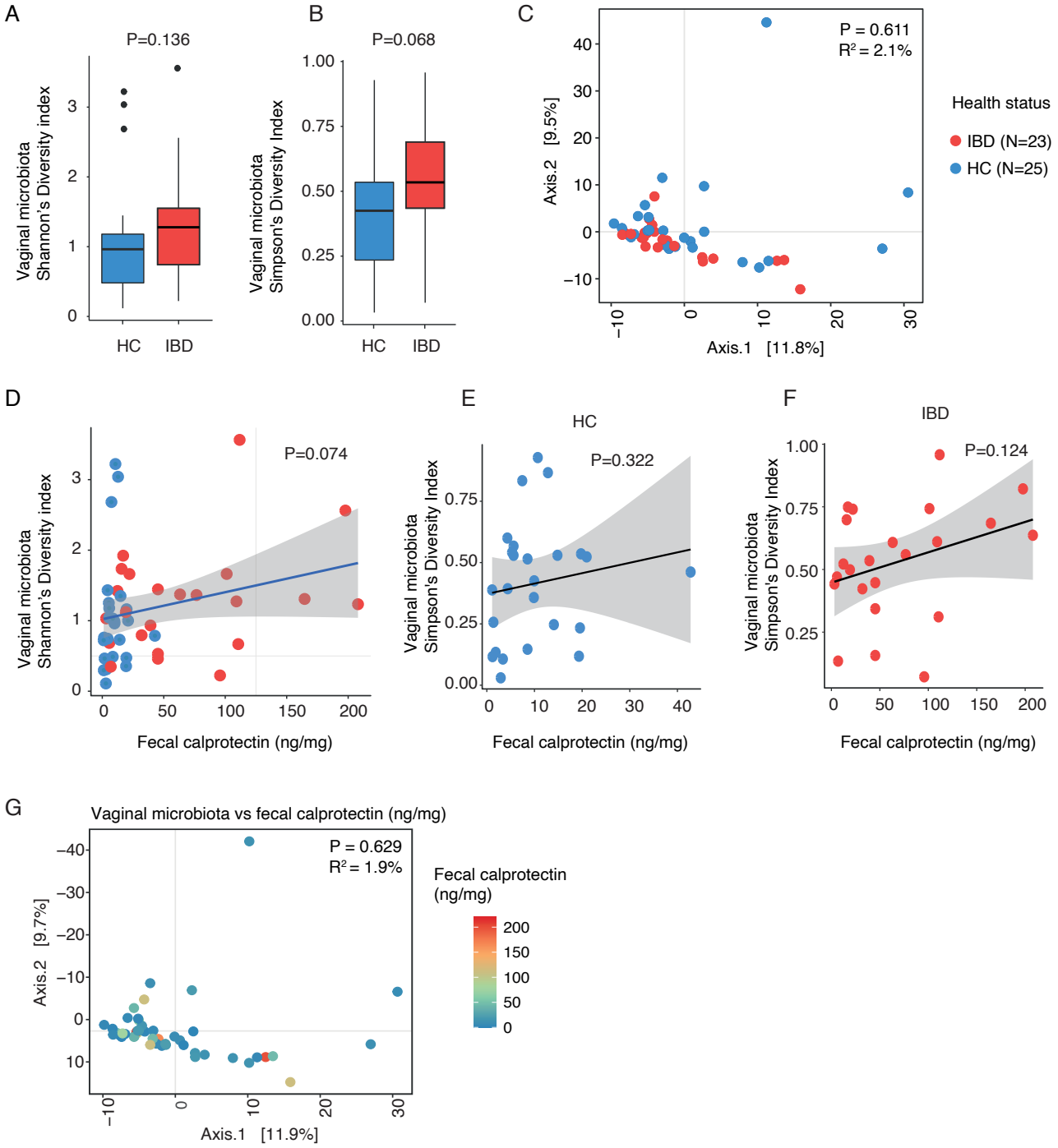
744 Types (CSTs).

	Community State Types						
Diet variables	I (N=21)	II (N=6)	III (N=14)	IV-C (N=4)	V (N=2)	Overall (N=47)	P value[&]
Healthy Eating Index (total score 0-100)							*0.098*
Mean (SD)	**68.4 (11.3)**	61.0 (15.7)	**55.2 (13.7)**	69.0 (9.49)	73.5 (13.4)	63.8 (13.6)	
Median	**67**	65.5	**53**	65.5	73.5	65	
Total vegetables (0-5)							*0.007*
Mean (SD)	**4.57 (0.978)**	3.50 (1.38)	**3.21 (1.12)**	4.25 (1.50)	3.50 (2.12)	3.96 (1.27)	
Median	**5**	4	**3**	5	3.5	5	
Greens and Beans (0-5)							*0.169*
Mean (SD)	4.38 (1.36)	3.50 (2.07)	3.07 (2.20)	4.75 (0.500)	4.50 (0.707)	3.91 (1.75)	
Median	5	4.5	4	5	4.5	5	
Total fruit (0-5)							*0.016*
Mean (SD)	2.86 (1.31)	3.33 (1.86)	2.29 (1.59)	4.75 (0.500)	5.00 (0)	3.00 (1.57)	
Median	3	3.5	2	5	5	3	
Whole fruit (0-5)							*0.271*
Mean (SD)	3.81 (1.72)	3.83 (1.60)	2.79 (2.12)	4.50 (1.00)	5.00 (0)	3.62 (1.81)	
Median	5	4.5	3	5	5	5	
Whole grains (0-10)							*0.947*
Mean (SD)	5.52 (3.33)	5.33 (3.14)	5.14 (3.35)	5.50 (4.43)	7.00 (0)	5.45 (3.24)	
Median	6	5.5	4.5	6	7	6	
Dairy (0-10)							*0.919*
Mean (SD)	6.62 (2.52)	6.00 (2.45)	6.93 (2.53)	7.25 (1.71)	7.50 (3.54)	6.72 (2.41)	
Median	6	6	6.5	7.5	7.5	6	
Total protein foods (0-5)							*0.482*
Mean (SD)	4.33 (1.11)	5.00 (0)	4.50 (0.760)	4.50 (0.577)	4.00 (1.41)	4.47 (0.905)	
Median	5	5	5	4.5	4	5	
Seafood and plant protein (0-5)							*0.112*
Mean (SD)	4.24 (1.37)	3.67 (2.16)	2.79 (1.89)	2.50 (2.38)	4.50 (0.707)	3.60 (1.80)	
Median	5	5	2	2.5	4.5	5	
Fatty acid ratio (0-10)							*0.306*
Mean (SD)	4.95 (3.34)	2.67 (3.27)	3.36 (3.52)	5.25 (3.20)	4.50 (6.36)	4.19 (3.46)	
Median	4	1.5	2	4	4.5	3	
Sodium (0-10)							*0.305*
Mean (SD)	6.57 (2.60)	6.00 (3.35)	4.57 (3.27)	5.00 (2.94)	8.00 (1.41)	5.83 (2.97)	
Median	7	7	3.5	5.5	8	6	
Refined grains (0-10)							*0.627*
Mean (SD)	6.62 (3.80)	6.83 (2.86)	5.36 (3.50)	7.50 (2.52)	9.00 (1.41)	6.45 (3.44)	
Median	8	6.5	5	8	9	8	
Added sugar (0-10)							*0.02*
Mean (SD)	**9.10 (1.51)**	8.00 (1.79)	**7.07 (1.82)**	7.75 (2.06)	8.00 (1.41)	8.19 (1.84)	

Median	**10**	8.5	**6.5**	7.5	8	9	
Saturated fats (0-10)							*0.532*
Mean (SD)	5.10 (3.46)	2.67 (2.50)	3.79 (3.04)	4.75 (3.30)	3.50 (4.95)	4.30 (3.24)	
Median	5	2.5	3.5	6	3.5	4	

& Kruskal-Wallis test followed by Wilcoxon tests for significant variables; in bold are indicated the pairwise analyses yielding significant differences between groups.

767 **SUPPLEMENTARY FIGURES**



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769 **Figure S1.** Comparison of vaginal microbiota diversity by health status and fecal calprotectin levels.
770 Shannon (A) and Simpson's (B) diversity indices to evaluate the microbial diversity. A linear regression
771 model assessed the comparison by health status. A Principal Coordinates Analysis (PCoA) (C) was
772 performed based on the Aitchison distance. The Shannon diversity index (D) by health status, with linear

773 regression modeling for analysis. Simpson's diversity index was employed separately for Healthy Controls
774 (E) and IBD patients (F), indicating no significant correlations; linear regression models were utilized for
775 these assessments. For the PCoA, PERMANOVA was applied to calculate P values and R^2, helping
776 quantify the explained variance.
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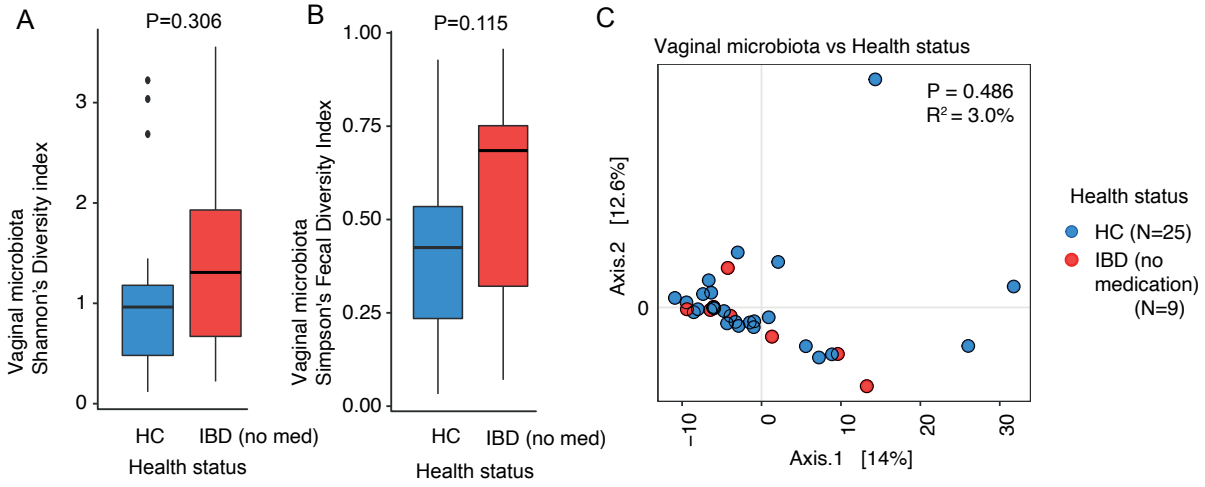


Figure S2. Comparison of vaginal microbial diversity in IBD patients off medication vs Healthy Controls (HC). Shannon's index (A) and Simpson's index (B). The significance levels were computed using a linear regression model. Beta diversity patterns are depicted through Principal Coordinates Analysis (PCoA) based on Aitchison distances (C), statistical significance and explained variance are reported through P values and R^2, using PERMANOVA.

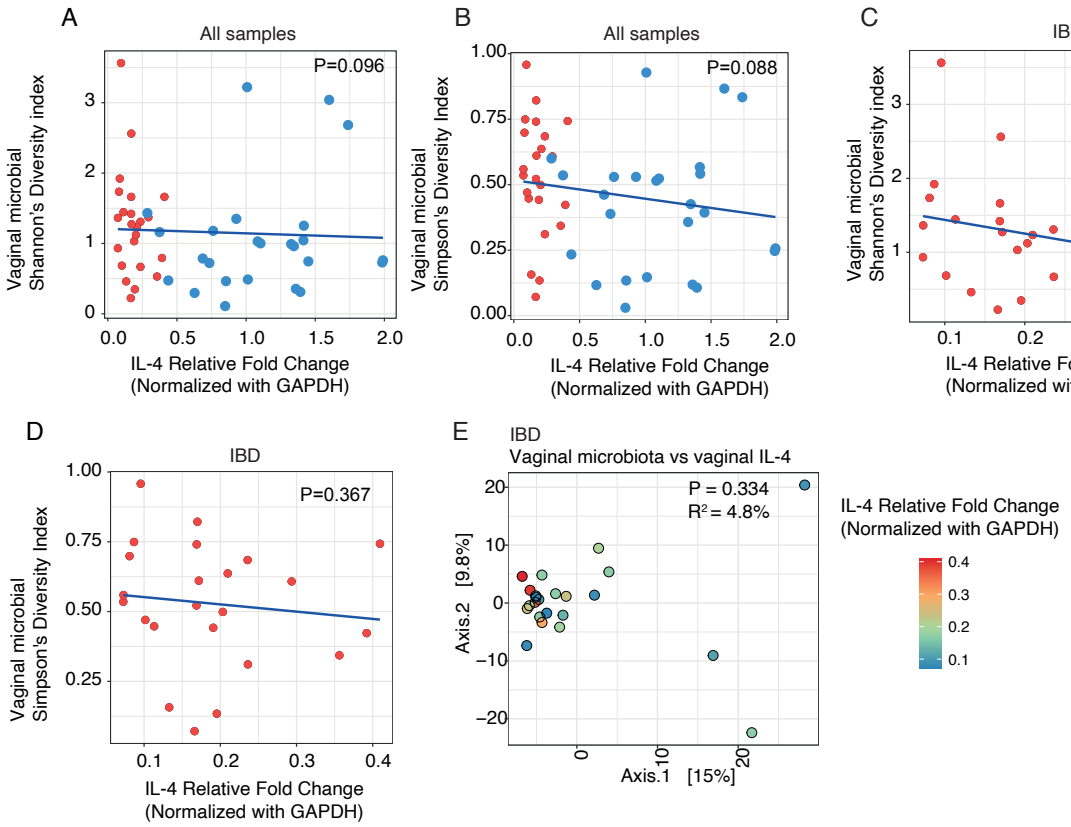


Figure S3. Vaginal microbial diversity and its of association with vaginal IL-4. Vaginal microbial diversity using Shannon (A) and Simpson's (B) diversity index by vaginal IL-4 relative fold change among all samples, reveals no significant associations, with linear regression models. Same analysis, although only among IBD patients using Shannon (C) and Simpson's (D) diversity index by vaginal IL-4 relative fold change, was not significant either. Principal Coordinates Analysis (PCoA), based on Aitchison distance, demonstrates no significance for vaginal microbial composition by IL-4 including only IBD patients (E). For PCoA PERMANOVA was used to obtain the P valued and to quantify the variance explained, as shown by P values and R^2.

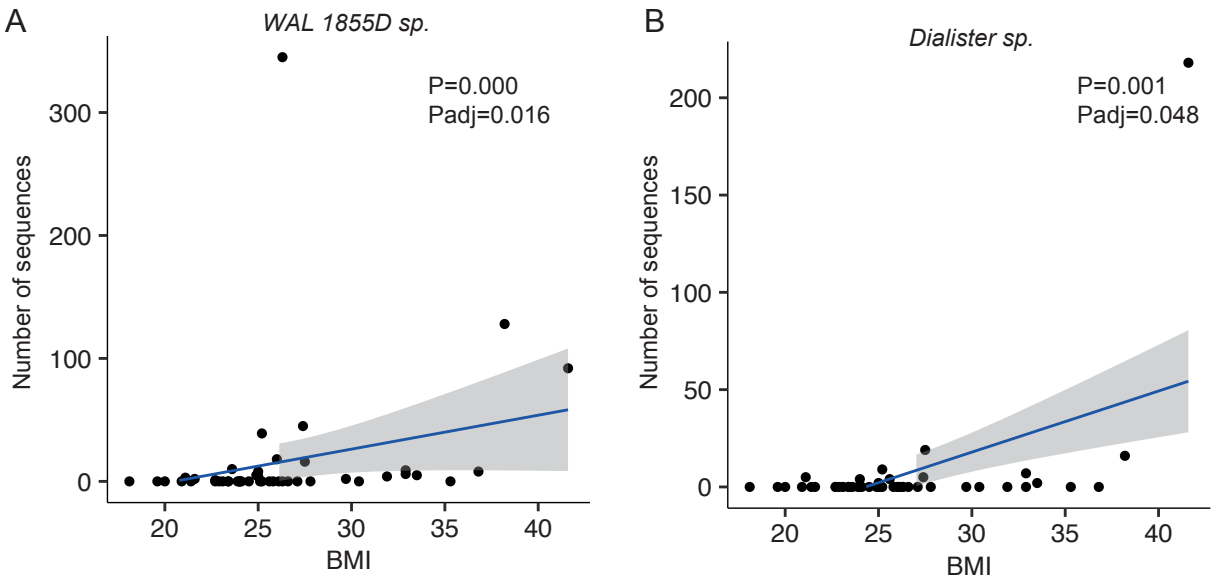


Figure S4. Vaginal species positively correlated with women's BMI. Number sequences by BMI for WAL 1855D sp (A) and *Dialister* sp (B). Analysis was performed with MaAsLin2 algorithm. P values were adjusted for multiple comparisons.

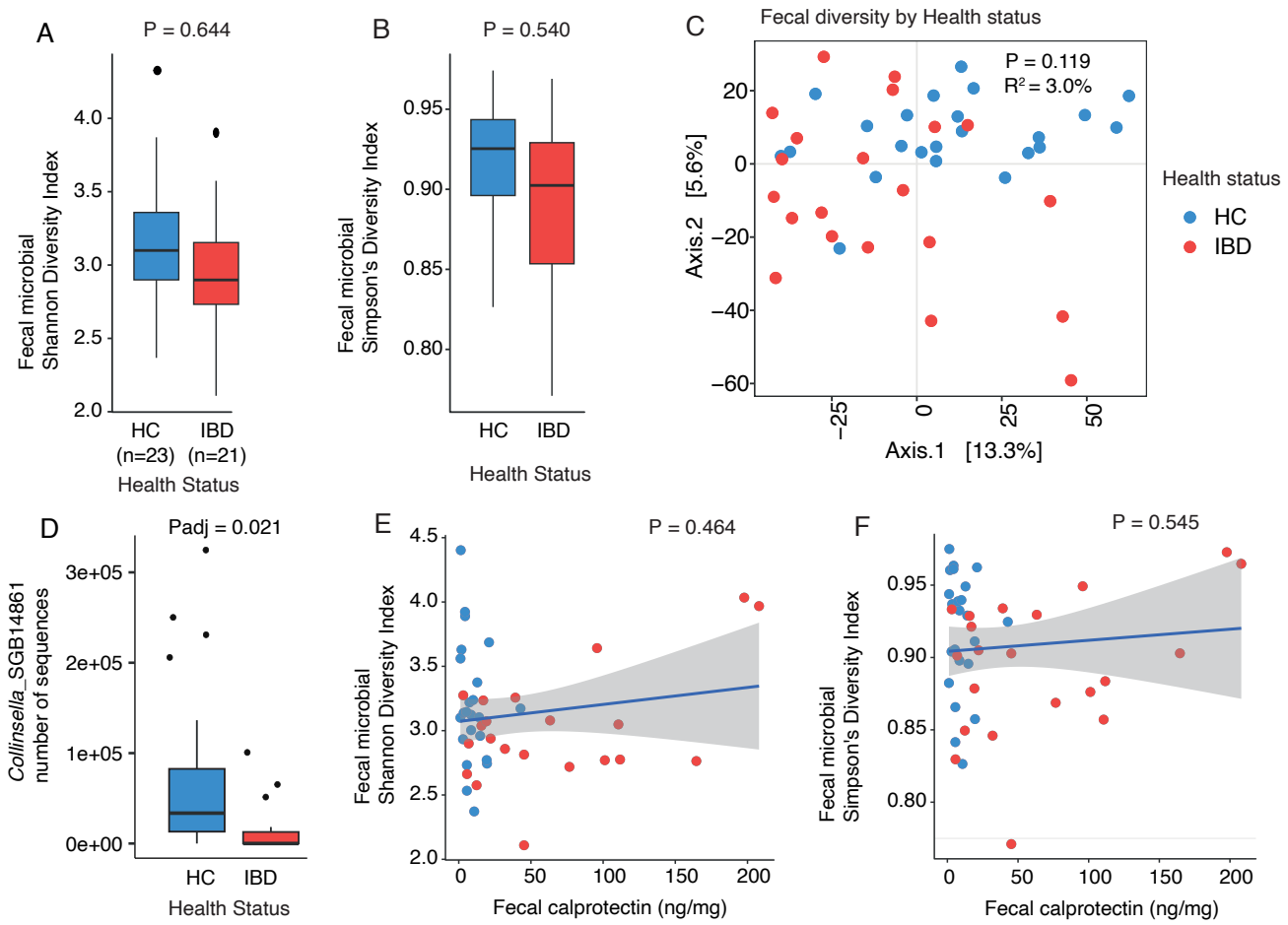


Figure S5. Comparison of fecal microbiota diversity by health status and fecal calprotectin levels. Shannon (A) and Simpson's (B) diversity indices to evaluate the microbial diversity. A linear regression model assessed the comparison by health status. A Principal Coordinates Analysis (PCoA) was performed based on the Aitchison distance to compare health status (C). Taxa significantly different by health status based on the MaAsLin2 algorithm (D). Shannon (E) and Simpson's (F) diversity indexes by fecal calprotectin levels, with linear regression modeling for analysis. For the PCoA, PERMANOVA was applied to calculate P values and R^2, helping quantify the explained variance

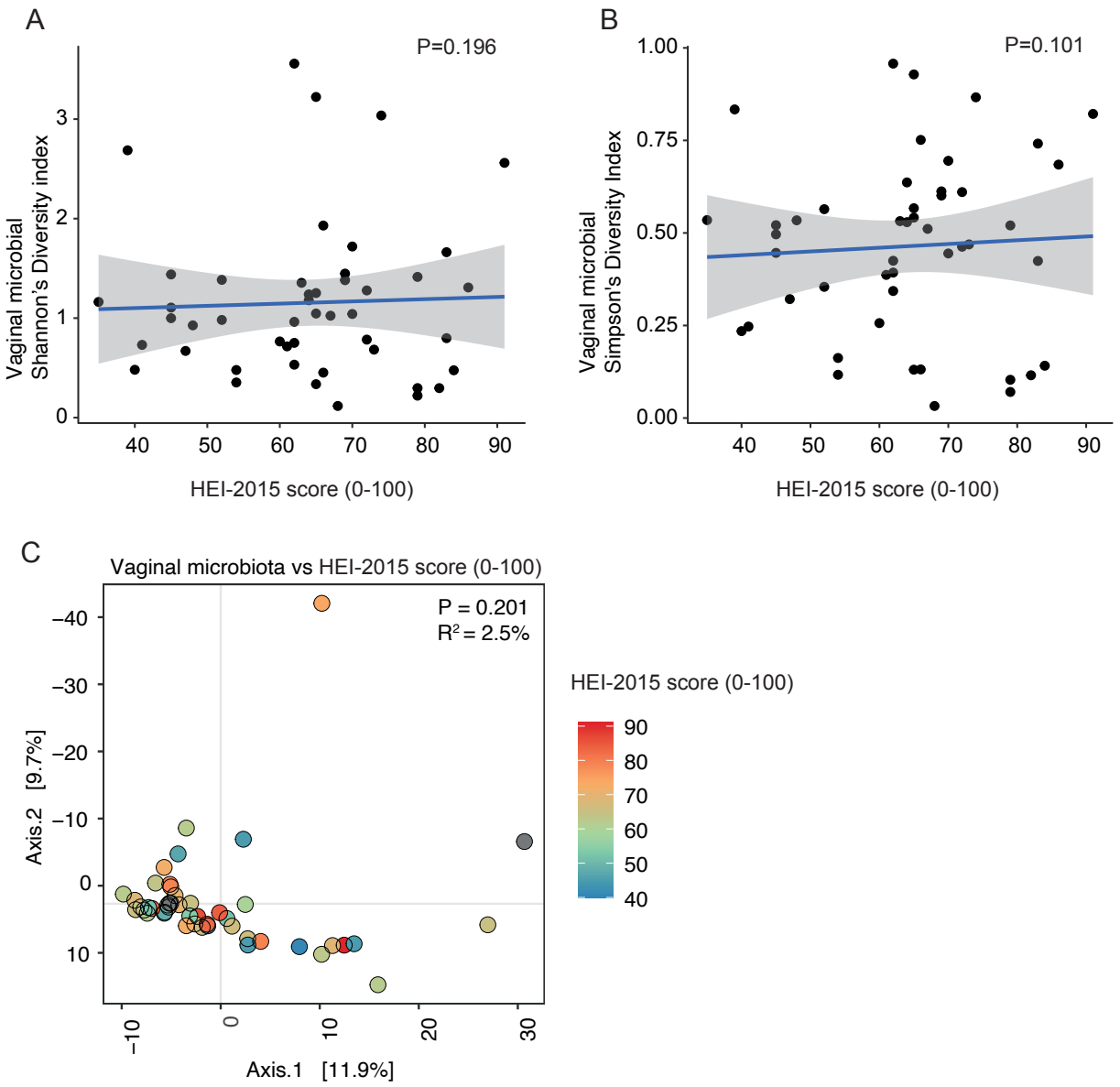


Figure S6. Diet quality is not associated with vaginal microbial alpha or beta diversity. HEI-2015 score for alpha diversity measured with Shannon's (A) and Simpson's (B) diversity indexes. P values were calculated with a linear regression model. Principal Coordinates Analysis (PCoA), based on Aitchison distance, for vaginal microbial composition by HEI-2015 score (B). PERMANOVA was used to obtain the P valued and to quantify the variance explained, as shown by P values and R2.

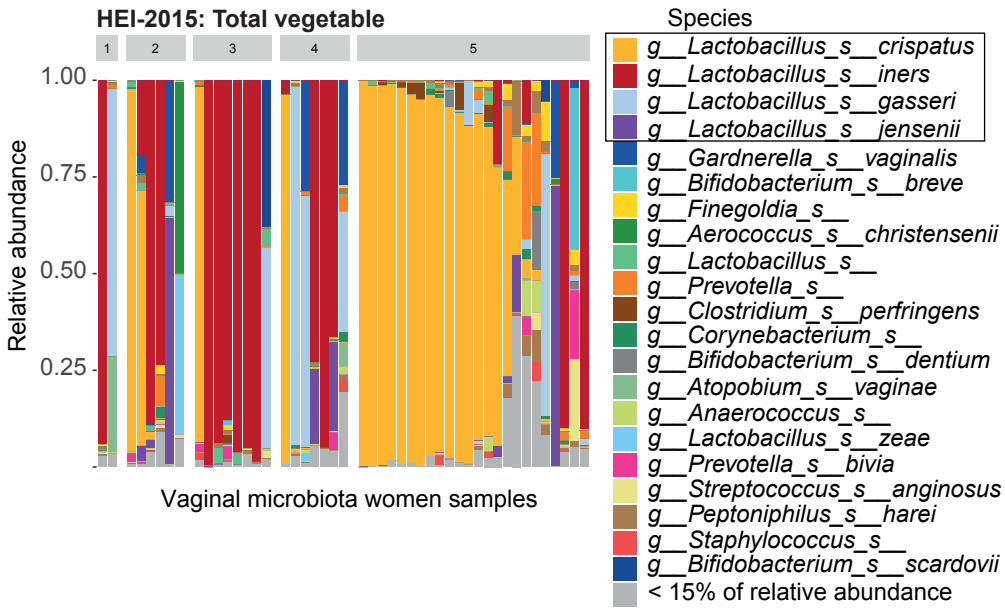
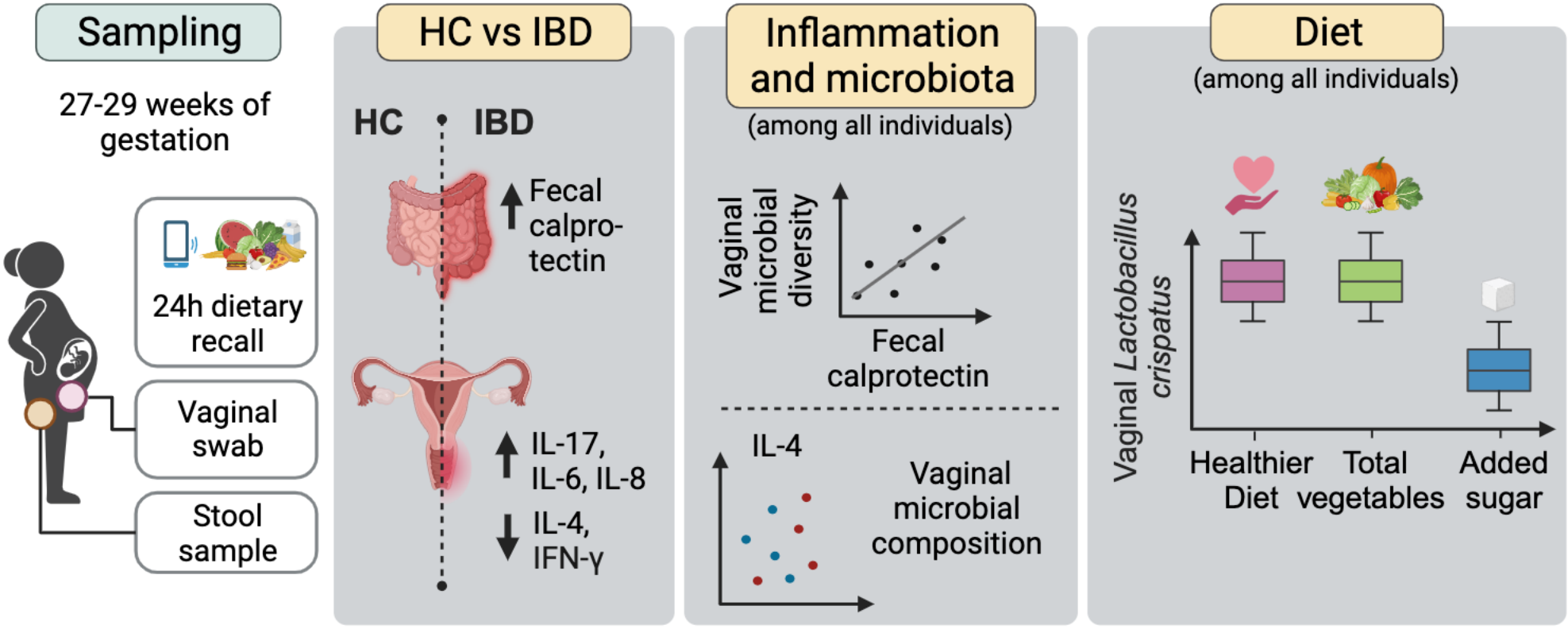


Figure S7. Microbial profile of pregnant individuals included in the study grouped by HEI-2015 total vegetable score and sorted by *L. crispatus* relative abundance. Each bar represents a unique sample.


Sampling
27-29 weeks of gestation
24h dietary recall
Vaginal swab
Stool sample
HC vs IBD
HC
IBD
Fecal calpro-tectin
IL-17, IL-6, IL-8
IL-4, IFN-γ
Inflammation and microbiota
(among all individuals)
Vaginal microbial diversity
Fecal calprotectin
IL-4
Vaginal microbial composition
Diet
(among all individuals)
Vaginal Lactobacillus crispatus
Healthier Diet
Total vegetables
Added sugar