The hippocampus pre-orders movements for skilled action sequences

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Abstract

Plasticity in the subcortical motor basal ganglia-thalamo-cerebellar network plays a key role in the acquisition and control of long-term memory for new procedural skills, from the formation of population trajectories controlling trained motor skills in the striatum to the adaptation of sensorimotor maps in the cerebellum. However, recent findings demonstrate the involvement of a wider cortical and subcortical brain network in the consolidation and control of well-trained actions, including an area traditionally associated with declarative memory – the hippocampus. Here, we probe which role these subcortical areas play in skilled motor sequence control, from sequence feature selection during planning to their integration during sequence execution. An fMRI dataset collected after participants learnt to produce four finger sequences entirely from memory with high accuracy over several days was examined for both changes in BOLD activity and their informational content in subcortical regions of interest. Although there was a widespread activity increase in effector-related striatal, thalamic and cerebellar regions, the associated activity did not contain information on the motor sequence identity. In contrast, hippocampal activity increased during planning and predicted the order of the upcoming sequence of movements. Our findings show that the hippocampus pre-orders movements for skilled action sequences, thus contributing to the higher-order control of skilled movements. These findings challenge the traditional taxonomy of episodic and procedural memory and carries implications for the rehabilitation of individuals with neurodegenerative disorders.

Introduction

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The neural control of skilled movements is distributed across cortico-striato-cerebellar
circuits linked via interlocking loops in the thalamus[1]. As motor skills are learnt, the dominant
assumption is that there is a transition from associative-cognitive to motor reference frames[2–
4]. This is implemented through a shift towards the involvement of cortical and subcortical
areas with mono- and disynaptic projections to brainstem and spinal control centres which
control the respective effectors, particularly the contralateral primary motor cortex, the
putamen and the ipsilateral anterior lobule of the cerebellum[1,5–7]. This shift is thought to
underlie the formation of a motor "repertoire" enabling automatic rather than slow and
effortful cognitive control of fine-grained kinematic trajectories and sensorimotor
transformations from typing and handwriting to highly skilled musical and athletic
performance[8,9].

Yet, a growing body of evidence from neurophysiology and neuroimaging experiments
in both humans and animal models suggests that the network of brain regions engaged in
later stages of skill learning and control continues to extend beyond the core motor regions[10].
This applies to skills even after prolonged training over weeks or months[11–15] and during
sequence production entirely from memory without visual or other external guidance[16]. The
associated cortical activity patterns have been linked to higher-order control of skilled motor
sequences such as the serial order and timing of motor sequences[17], to enable behavioural
flexibility, a key feature of skilled motor control in humans. In contrast, activity within primary
motor cortical regions shows a lack of tuning to skilled sequences of movements, even
following extensive practice[18–20].

Whilst the cortical control of motor skills has been characterised in detail over the last
three decades, the complementary roles of striato-thalamo-cerebellar loop regions remains
poorly delineated. Specifically, it remains unclear whether the encoding of skilled motor
sequences, e.g. in striatal and cerebellar effector-related areas is linked to high-order control
of sequence chunking and action selection or to lower-order control of movement
kinematics[7,21,22]. Furthermore, the hippocampus has long been overlooked as a contributor
to procedural motor control due to landmark reports of preserved procedural memory in
amnesic patients with medial temporal lobe damage[23,24]. More recently, the original
taxonomy has been questioned by findings pointing towards its contribution to sequence
learning and consolidation, both during sleep and short breaks[25–29]. Although there is now
strong evidence for the hippocampus in procedural motor skill consolidation, its role in the
online motor control of skilled movements remains unclear. It remains uncertain whether this
contribution is related to the retrieval of task-context associations, or directly tied to the
control of specific features of the performed movement sequences.

Here, we set out to tease apart the roles of subcortical areas in skilled movement
sequence planning and execution from memory. Participants produced movement
sequences with particular order and timing entirely from memory in a delayed sequence
production task following two-days of training[16]. Using canonical and multivariate analysis of
fMRI patterns in effector-related contralateral striato-thalamo-cerebellar and bilateral
hippocampal areas, we probed the tuning of these regions to motor sequences and their
ordinal and temporal features during planning and execution. We found that that the
hippocampus increased its activity and carried accurate information on the upcoming
movement sequence – the movement order independently of the planned sequence timing.
In contrast, the activity increase within effector-related striato-thalamo-cerebellar regions

during planning and execution was not sequence-specific. This implies their involvement in
the lower-order control of individual movement components reused across sequences.

Results

Subcortical activity modulation during sequence planning and execution

24 participants were trained to produce four five-finger sequences from memory using
their right hand. Behavioural training progressed from production under visual instruction to
production from memory across two days. On the final third day, participants produced the
sequences entirely from memory in an MRI scanner. The behavioural and cortical results are
reported in detail in a previous paper[16]. To isolate neural patterns relating to movement
production from those relating to movement preparation, we used two independent trial
types: Go trials (Figure 1a), to sample production activity, and No-Go trials (Figure 1b), to
sample preparatory activity. Go trials began by cueing the upcoming sequence with an
abstract fractal image, followed by a fixation cross, then a black hand with a green
background (Go cue) indicated that the sequence should be executed (Figure 1c). A further
fixation cross, then performance feedback, followed. No-Go trials were identical to Go trials
during movement preparation, however, they did not display a Go cue. Rather, a fixation
cross remained on the screen for an extended period. Feedback followed, and participants
were rewarded for not producing any presses (see Methods and materials).

Figure 1. Trial types and target sequences. **a**, Participants were trained to produce four sequences
from memory via visual instruction in 'Go' trials. The target sequence for each trial was indicated by
an abstract fractal cue which preceded a short preparation period. A black hand with a grey

(instructed; a red cue indicated which finger to press with a set temporal structure) or green (from
memory; participants had to produce the sequence without the visual cue) background appeared to
indicate the go cue. A short fixation period followed, after which feedback was provided based on the
accuracy and timing of the sequence. **b**, on 50% of trials, the go cue would be replaced by an
extended fixation cross. Participants were subsequently rewarded for not making a press during these
'No-Go' trials. **c**, all trials for one participant are plotted with the colour indicating which button was
pressed according to the image above the plot. Target sequences consisted of permutations of two
timings and two orders, constituting S1 through S4. Participants performed the sequences using the
right hand on a 10-finger force transducer.

Firstly, we examined whether the subcortical regions of interest with connections to the
active effector (right hand) – the left thalamus, caudate, putamen, and the right CB lobules
IV and V) – as well as the bilateral hippocampi, changed overall BOLD activity during the
delayed sequence production task. We defined all regions of interest (ROI) using each
participant's individual anatomy and extracted the percent signal change during preparation
(No-Go trials) and production (Go trials) relative to rest (see Methods and materials). The
extracted percent signal change values from across all voxels belonging to each ROI were
then averaged for each participant (Figure 2a, b). All tests were Bonferroni corrected twice to
account for phase (preparation and production) in each pre-defined ROI. During preparation,
we observed activity above baseline in the caudate ($t(23) = 3.28$, $p = .006$, $d = 0.67$),
putamen ($t(23) = 2.67$, $p = .028$, $d = 0.55$), and hippocampus on both the left ($t(23) = 6.28$, p
$< .001$, $d = 1.28$) and right ($t(23) = 4.63$, $p < .001$, $d = 0.94$) sides. No significant difference
relative to baseline was found in the thalamus nor in cerebellar lobules IV and V during
preparation ($p > .114$, $d < 0.41$). During production, activity increases were observed in all
subcortical area with connections to the active effector - contralateral thalamus ($t(23) = 7.54$,
$p < .001$, $d = 1.54$), caudate ($t(23) = 2.66$, $p = .028$, $d = 0.54$), putamen ($t(23) = 4.21$, $p <$
$.001$, $d = 0.86$), cerebellar lobule IV ($t(23) = 5.61$, $p < .001$, $d = 1.15$), and CB lobule V ($t(23)$
$= 12.22$, $p < .001$, $d = 2.49$). The hippocampus, however, showed activity decreases during
production on the left ($t(23) = 2.45$, $p = .044$, $d = 0.50$) and right ($t(23) = 3.19$, $p = .008$, $d =$
0.65) sides. In sum, only the striatum and hippocampus were active during sequence
preparation from memory. In contrast, the striato-thalamo-cerebellar regions with direct
relevance to the active effector increased their activity during production, whereas
hippocampal activity decreased.

Figure 2. Percent signal change in subcortical regions during preparation and production. **a**, volumetric slices of each subcortical region displaying percent signal change for preparation and production, far left and right respectively. Centre shows the level of percent signal change when averaged across each subcortical region. ** $p < 0.01$; * $p < 0.05$; two-sided t test against 0. Error bars represent standard error of the sample mean. **b**, as above, for cerebellum. Black outline indicates significant clusters. Tha, thalamus; Cau, caudate nucleus; Put, putamen; Hip, hippocampus.

We then localised the peak activations within each subcortical area and across the whole cerebellar volume by identifying significant clusters of activation using a random effects analysis (Figure 2a, b). All results were corrected using SPM's small-volume correction and Bonferroni corrected twice to account for phase. During preparation, we found significant clusters of activation in the caudate's ventral anterior head ($t(23) = 6.02$, $pFWE = .008$, extent = 101, MNI coordinates: x=-16, y=22, z=-5), anterior body ($t(23) = 5.56$, $pFWE = $

.020, extent = 83, x -16 y 22 z 7), and dorsal anterior tail (*t*(23) = 5.55, *pFWE* = .008, extent
=106, MNI coordinates: x=-12, y=-6, z=17), posterior putamen (*t*(23) = 5.71, *pFWE* < .001,
extent = 228, MNI coordinates: x=-32, y=-14, z=9), anterior left hippocampus (*t*(23) = 7.97,
pFWE < .001, extent = 766, MNI coordinates: x=-26, y=-22, z=-15), anterior right
hippocampus(*t*(23) = 6.33, *pFWE* < .001, extent = 471, MNI coordinates: x=38, y=-20, z=-
17), and CB right lobule V extending into lobule IV (*t*(23) = 5.09, *pFWE* = .002, extent = 205,
MNI coordinates: x=17, y=-45, z=17). During production, we found significant clusters of
activation widespread in the thalamus which centred middle posterior (*t*(23) = 14.29, *pFWE* <
.001, extent = 851, MNI coordinates: x=-16, y=-24, z=5), the anterior tail of the caudate (*t*(23)
= 5.24, *pFWE* < .001, extent = 208, MNI coordinates: x=-10, y=0, z=9), posterior putamen
(*t*(23) = 7.06, *pFWE* < .001, extent = 783, MNI coordinates: x=-28, y=-18, z=3), and CB
lobules IV-VI (the peak of which is in lobule VI, outside of our pre-defined cerebellar ROIs;
t(23) = 8.46, *pFWE* < .001, extent = 20079, MNI coordinates: x=7, y=-69, z=-16). The
location of the clusters generally shifted or expanded from associative to motor subregions
of the subcortical areas of interest upon execution.

Hippocampal activity during planning predicts the upcoming order of movements

To identify what content is represented in the underlying activity patterns during
planning and production, we trained and tested an LDA classifier to distinguish between the
neural patterns related to sequence identification, specifically of the planned and executed
movement order, movement timing, and their unique combination[16,17]. All decoders were
cross validated across six imaging runs and, for order and timing, across conditions, then
converted into z scores (see Methods and materials).

We first tested whether ROI showed above chance decoding accuracy (Figure 3a, b).
To do so, we obtained accuracy values for each of our three classifiers during preparation
and production when considering all voxels within each ROI. We then assessed accuracy
values in each region using one-tailed one-sample t-tests against a chance level of zero. All
tests were Bonferroni corrected six times to account for classifier (3) and phase (2) in each
pre-defined subcortical ROI. During preparation, we found above chance decoding of the
upcoming sequence order in the left hippocampus (*t*(23) = 3.00, *p* = .018, *d* = 0.61). No
evidence of significantly above-chance decoding accuracy could be found in the other ROIs
(*p* > .264, *d* < 0.36). These results suggest that the left hippocampus is involved in the
planning of the ordinal structure of movement sequences.

Figure 3. Linear discriminant analysis results. **a**, volumetric slices of each subcortical region displaying classification accuracy for each classifier during preparation and production, far left and right respectively. Centre shows classification accuracy when averaged across each subcortical region. * $p < 0.05$; one-sided t test against chance (Bonferroni-corrected for six comparisons within each ROI). Error bars represent standard error of the sample mean. **b**, as above, for cerebellum. Black outline indicates significant cluster. Tha, thalamus; Cau, caudate nucleus; Put, putamen; Hip, hippocampus.

Though considering all present voxels contained within a region is informative as to the general inclination of a region's processing, it does not inform us as to which subregions may be driving decoding accuracy the most. This is especially true when it comes to subcortical regions such as the thalamus, which shows differing functional and anatomical connectivity throughout its various subregions and nuclei[30]. Therefore, we performed

volumetric searchlight analyses using 160 voxel searchlights for each decoder, constrained
to each subcortical ROI (Figure 3a, b). The accuracy value for each given searchlight was
assigned to the centre voxel. We then identified significant clusters using a random effects
analysis on the produced accuracy maps and applied SPM's small-volume correction in a
similar manner to the contrast activity cluster analysis. We Bonferroni corrected significance
values for each cluster six times, to account for classifier (3) and phase (2). During
preparation, we found a significant cluster for the order decoder in the anterior body of the
left hippocampus (t(23) = 5.74, $pFWE$ < .001, extent = 379, MNI coordinates: x=-32, y=-22,
z=-11). No significant clusters were found during production, although prior to six-fold
corrections a cluster for integrated sequence decoding was found in the superior thalamus
during production (t(23) = 4.76, $pFWE$ = .045, extent = 47, MNI coordinates: x=-18, y=-26,
z=15). These findings reinforce that the hippocampus, particularly the anterior body, is
involved in the planning of sequential movement order. Sequence timing and its non-linear
integration with sequence order, however, do not seem to be represented in effector-related
subcortical regions.

Sequence-related activity patterns are not conserved across planning and execution

Given that our target subcortical regions show different activity levels alongside
changes in informational content across preparation and production, we asked whether the
sequence related patterns are maintained prior to and during movement. One possibility is
that the patterns present during preparation undergo linear scaling to exceed a set threshold
and initiate movement, where unintended initiation is thought to be prevented by
suppression in corticospinal circuits[31]. Alternatively, these patterns may alter their
geometries, meaning that they undergo a significant state change. The presence of scaled
cortical neural patterns has been evidenced in single finger movements using fMRI[32],
whereas distinct neural patterns have been shown in multi-unit recordings of cortical
populations[33], however, subcortical pattern dynamics before and during sequence production
are unknown. Accordingly, we assessed the within-sequence, across-phase Euclidean
distance between conditions in our fMRI data across principal components using multi-
dimensional scaling of the representational dissimilarity matrix. Since we saw substantial
activity changes across all regions except for the caudate from preparation to production, the
first principal component is set to represent the different levels of activity as it captures the
dimension with the greatest variance across the conditions. Therefore, we quantified the
Euclidean distance between each sequence's position during preparation and production in
a 2D space with the axes comprising of principal components two and three.

Figure 4. Simulated and empirical cross-phase Euclidean distances using multi-dimensional scaling.
a, multi-dimensional scaling plots of simulated fMRI data along principal components 1, 2, and 3,
showing four simulated sequence conditions across two phases. Red lines connect simulated
sequences with different timings but the same order, whereas blue lines indicate sequences with
different orders but the same timing. Dotted grey lines are drawn between the same sequence during
preparation and production. Preparation and production were either generated using the same

distribution (left panel; no switch) or different distributions (right panel; switch). **b**, multi-dimensional
scaling plots of empirical data from target ROIs showing all four sequences during preparation and
production. **c**, averaged Euclidean distance between preparation and production within sequences
once PC1 is excluded, calculated relative to those same distances from the simulated no switch data
with matched signal to noise ratios. This represents differences for all sequences between
preparation and production which cannot be explained by differences in overall activity. Orange
horizontal lines represent the same distances in the simulations when a switch is induced. * $p < 0.05$;
** $p < 0.001$; one-sided t test against simulated no distance with matched noise, 0 (Bonferroni-
corrected for seven comparisons). Hip, hippocampus; Tha, thalamus; Cau, caudate nucleus; Put,
putamen; Lob IV, lobule IV; Lob V, lobule V.

If one were to assume that activity patterns during production are simply upscaled
versions of those during preparation, the hypothetical distance between patterns after
removal of the first principal component should be zero. However, given a non-zero level of
noise in our data, these distances are unlikely to equal zero which would bias our results
away from indicating a maintained distribution. To control for this, we simulated fMRI data
sets with the same conditions as our empirical data and generated preparation and
production from either the same distribution (no switch) or different distributions (switch),
whilst matching their signal to noise ratios to our regions of interest (Figure 4a). Additionally,
we scaled this cross-phase distance by the average of distances within preparation and
production (Figure 4b). We then performed one-tailed, one-sample *t*-tests against zero to
identify significantly elevated Euclidean distance in each of our subcortical regions,
Bonferroni corrected seven times for region (Figure 4c). We found significant elevation in the
thalamus ($t(23) = 6.06$, $p < .001$, $d = 1.24$), caudate ($t(23) = 3.20$, $p = .014$, $d = 0.65$),
putamen ($t(23) = 3.12$, $p = .017$, $d = 0.64$), hippocampus in the left ($t(23) = 3.05$, $p = .020$, $d
= 0.62$) and right ($t(23) = 3.93$, $p = .002$, $d = 0.80$) hemispheres. No significant differences
were found in CB lobule IV ($t(23) = 0.37$, $p = 1.00$, $d = 0.08$) and CB lobule V ($t(23) = 2.05$, $p
= .182$, $d = 0.42$). These results suggest that all subcortical regions apart from CB show
significantly higher cross-phase distances than a matched simulation which does not predict
a change in distributions across phase, providing evidence for a substantial state change
which alters sequence-specific neural patterns form prior to and during movement.

260   **Discussion**
Acquiring a repertoire of skilled motor sequences has been linked to plasticity in
subcortical motor-related brain areas – basal ganglia, the cerebellum, and, more recently,
the hippocampus[9,27,34]. However, the contribution of these areas to higher-order action
selection and lower-order movement control in skilled movements is debated[35–38]. Here, we
examined the contribution of effector-related subcortical regions and the hippocampus to
motor sequence control in the peri-movement phase (Figure 5). Further, we assessed
whether the neural patterns during planning in those subcortical areas were simply a
subthreshold versions of those during execution[31,39], or qualitatively distinct neural states[33].
We found that both hippocampal and effector-related striatal regions showed increased
activity during sequence planning. However, only the hippocampal activity was related to
sequence-specific information, specifically predicting the sequence order shortly before each
execution. In contrast, effector-related striatal, thalamic, and cerebellar regions increased
activity during execution, yet lacked sequence-specific tuning, suggesting their role may be
related to the control of lower-level kinematics of individual movement elements, rather than
of a whole movement sequence. All regions, apart from the cerebellum, showed
fundamental shifts in the sequence-related activity patterns from planning to execution,
indicating that movement onset is associated with a fundamental state shift in subcortical

areas rather than a scaling up of subthreshold patterns, similar to the previous results in the
cortex[16,33].

The hippocampus pre-orders upcoming movements

The hippocampus has traditionally been associated with episodic recall in the non-
motor domain and is key to the preservation of event order[40,41]. Here we show that the
hippocampus is part of online motor control of a procedural skill which was trained over
several days and produced without external guidance. Specifically, it is key to setting up the
serial order of movements in an upcoming sequence from long-term memory (Figure 5). This
is a high-order control function, as the same hippocampal patterns are retrieved regardless
of the upcoming temporal structure (timing) of the sequence. These findings show that the
hippocampal involvement goes beyond its contribution to the acquisition of motor skills,
which has been reported in the context of sleep-related or wakeful consolidation during short
rest periods interspersed with sequence learning[26,27]. Instead, it is participating in the rapid
planning of a well-trained skill.

The hippocampus may retrieve the order of the upcoming motor sequence via
competitive queueing of the upcoming movements[11,42,43]. This computational framework
suggests that upcoming movements are pre-ordered in parallel prior to their serial execution
during production in the parallel planning layer[44] which has been confirmed
experimentally[42,45]. Alternatively, the hippocampus may engage in serial order preplay[46],
which is temporally compressed approximately 20-fold relative to the acquired skill and is
associated with skill consolidation[27]. How competitive queueing and preplay are related to
each other is unclear. It is possible that they either measure the same underlying
mechanism, e.g. pre-ordered sequence elements being related to incomplete replay sweeps,
or constitute co-existing modes of serial order control across domains.

Our previous findings in the cortex suggest that the superior parietal lobule (SPL) also
plays a role in the definition of movement order during planning[16] and represents effector
non-specific movement intentions [47,48]. The SPL and hippocampus share extensive
connections[49] and interact closely during navigation tasks to convert world-centred into body-
centred coordinates[50,51]. As such, interaction between the hippocampus and the SPL may
serve to map the world-centred order retrieved by the hippocampus[42] onto intrinsic reference
frames[52] prior to execution.

Figure 5. Schematic representation of sequence feature control during planning and execution.
Higher level sequence features, order (blue) and timing (red), are defined by cortex and hippocampus
during planning. During execution, the activity patterns in hippocampus, basal ganglia, and thalamus
shift while the cortex integrates order and timing into low-level sequence specific trajectories (green),
while maintaining higher-level independent movement timing. The green gradient fill in thalamus,
basal ganglia, and cerebellum indicates regions that implement individual sequential elements. Tha,
thalamus; BG, basal ganglia; CB, cerebellum; HC, hippocampus.

Effector-relevant striatal, thalamic, and cerebellar activity lacks sequence-specific tuning

Despite robust activity increases, in particular during sequence execution, we found no
evidence that effector-relevant contralateral striatal, thalamic, or ipsilateral cerebellar activity
contained any sequence-specific information. These findings speak against both the
hypothesized role of the striatum in higher-order action selection [36,53–55] or the integration of
sequence features for kinematic control of the whole sequence as one motor program[56–58].
Human neuroimaging studies report movement concatenation-related activity in the
striatum[59], however our data suggests that these control signals are not sequence-specific[60],
but might instead be related to the process of concatenation itself. Further, multi-unit
recordings in the dorsomedial striatum (caudate nucleus in humans) showed ramping activity
which scales with temporal intervals between movements[61,62], and is sensitive to the
temporal structure of sequential movements[63]. However, we find no evidence that the
striatum encodes differences in upcoming or ongoing temporal interval structure of
sequences that are matched in structural complexity – neither during planning, nor during
execution.
Recent results in rodent models showed that optogenetic lesions lead to kinematic
deficits in overtrained sequence production[38]. Interestingly, this mapping breaks down when
highly trained sequences share elements with other sequences the animal has been

exposed to, e.g. lever presses appearing in a different order, with the skill becoming once-
more cortically-dependent[64]. This is typical for many motor skills in humans, such as typing,
handwriting and music production where elements are reused in different permutations,
including highly overtrained sequences such as signing with ones name or writing "Best
wishes". It is reflected in the factorial design of our study, which requires a well-trained
recombination of movements into new sequences and shows no evidence of striatal tuning
to these sequences. Thus, kinematic control in the striatum is likely associated with fused
movements that are not broken up for flexible recombination – here, individual muscular
commands associated with the different finger presses on the force transducer.

Cerebellar motor regions have traditionally been associated with low-level sensorimotor
learning and online motor control, as evidenced by cerebellar ataxia[65,66], adaptation to
sensory perturbations[67] and eye blink conditioning[35]. However, recent findings demonstrated
the cerebellar involvement beyond short-timescales[68,69]. Transient perturbations of the deep
cerebellar nuclei during planning resulted in mice making fewer correct directional licks and
impacted subsequent movement direction whilst kinematics remained stable[37,68,69]. The
interconnected ventrolateral thalamus, which constitutes an interface between striatum,
cerebellum and cortex, has been shown to help maintain preparatory activity[70] and control
cortical dynamics from preparation to production for rapid and precise motor behaviour[71].
This extends the notion of the cerebellum as an online motor controller and the thalamus as
a sensorimotor relay[72,73]. However, we did not observe any activity increase or sequence
related patterns in ipsilateral effector-relevant cerebellar regions during motor sequence
planning. This supports the classical findings that cerebellar motor regions contribute to the
online control of individual movement elements during movement production, rather than the
planning and execution of a whole sequence, e.g. planning "on the fly" once the production
of a sequence has started[74,75]. Notably, sequence-related patterns could be found in
cerebellar regions known to be interconnected with prefrontal areas (Crus I). This was only
observed during motor execution, again emphasizing the role of the cerebellum in the online
control of movements beyond effector-related regions.

Despite participants' ability to fluently produce the target movement sequences from
memory after two days of training, the movement kinematics for the upcoming sequence are
not pre-planned. Rather, the order and timing of sequences is recombined trial-by-trial likely
through communication with the cortex, which has shown a mechanism whereby the order
and timing are planned and subsequently integrated during execution in the cortex[16]. This
maintenance of separation up until the go cue likely affords the individual the ability to adjust
movement plans should task demands suddenly change.

Shift in sequence pattern activity across planning and execution

Motor planning in the cortex has been proposed to be a scaled down version of
execution, held at bay by suppression in corticospinal circuits [31,39]. More recently, however,
planning activity in the motor cortex has been proposed to exist in an output-null dimension
to execution, where increases and decreases in output from neurons projecting to the same
source cause no net change in activity, but prepare the system to shift into the output-potent
dimension [33,76], including for sequences of movements [16]. Our current results suggest that
this shift also takes place in multiple subcortical areas: striatum, thalamus, and
hippocampus. Here, sequence-related patterns across planning and execution were
significantly distinct across the peri-movement phases. This could not be explained by a
simple upscaling through changes in activity and matched by our no-change simulations of

the same dimensions that considered empirical noise levels in those areas. This could be driven by separate populations becoming active at different times around movement execution or, alternatively, changes of the patterns of the same neuronal pool.

Implications for clinical disorders

Here we show that the hippocampus, but not the basal ganglia-cerebellar-thalamic loop is carrying sequence-related information during the planning of skilled movements produced from memory. Thus, breakdowns in movement sequence control in Parkinson's Disease[77] and Cerebellar Ataxia[78] cannot be explained by deficits in the planning or execution of entire sequences. Instead we hypothesize that they might be a result of the reduced capacity to concatenate movement elements or execute movement elements from the motor repertoire. In contrast, our results predict that Alzheimer's Disease which is associated with a degeneration of the hippocampal formation and the medial temporal lobe, affects everyday movement sequence production such as handwriting and ideomotor action organization[79] because of the inability to retrieve the correct order of the upcoming movement elements[43]. We hypothesize that this applies particularly to contexts where movement sequences need to be retrieved from memory flexibly in difference permutations. In humans, this applies to everyday skilled actions which utilise the same elements from the motor repertoire across different ordinal sequences, including overtrained sequences of movements, e.g. in typing, handwriting and tool use.

In summary, our results question traditional classifications of motor skill memory and control, along with their associated subcortical substrates. Research on skilled movement planning in diverse clinical populations is needed to inform appropriate rehabilitation interventions and support, e.g. by strategically boosting access to compensatory sequence planning mechanisms to enhance performance.

Methods and materials

Participants

24 participants (14 female and 10 male; mean age = 21.00 years, SD = 1.64 years) completed the three-day experiment, having met all behavioural and imaging requirements. All participants had no history of neurological disorders. 23 participants were right-handed with a mean Edinburgh Handedness Inventory (https://www.brainmapping.org/shared/Edinburgh.php; adapted from [80]) score of 75.22 (SD = 20.97, range: 25-100), one was left-handed with a score of -70. Although our preregistration (www.osf.io/g64hv) stated that we would exclude left-handed individuals, this participant's data did not differ qualitatively from the rest of the sample so were included. Data collected from an additional 17 participants were excluded for the following reasons: one due to unforeseen technical difficulties with the apparatus, one because of a corrupted functional scan, and 15 further did not reach target performance after two days of training. Target performance required an error rate < 20% (group mean = 6.54%, SD = 6.03%) and statistically distinct sequence timing structures that were maintained across different sequence orders; the full details regarding the exclusion criteria have been described elsewhere[16]. Participants were recruited either through a participation panel at Bangor University and awarded module credits, or through social media and given monetary reward at a standard rate. Individuals with professional musical qualifications were excluded from recruitment. All participants provided informed consent, including consent to data analysis

and publication, through an online questionnaire hosted by Qualtrics. The Bangor University
School of Psychology Ethics Committee approved this experiment and its procedures (ethics
approval number 2019-16478).

Apparatus

Two custom-built force transducer keyboards captured presses from all 10 fingers on
the right and left hands of participants as they produced movement sequences. Each key
had a small groove where participants placed their finger and could be adjusted for comfort
according to the size of the participant's hand. Below each key, a force transducer sampled
force at 1000 Hz (Honeywell FS Series, with a range of up to 15N). The keys were not
depressible. Force acquisition occurred in each trial from 500ms before sequence cue onset
to the end of the production period in production trials, and the end of the false production
period in No-Go trials. Traces from the right hand were baseline-corrected trial-by-trial using
the first 500ms of acquisition (500ms before the sequence cue) and smoothed to a Gaussian
window of 100ms. Button presses were recognised when a channel's force exceeded a fixed
threshold relative to baseline (2.5N for the first 8 participants and 1N for the subsequent 16
of 24 participants). The further apparatus used during the experiment have been described
in a previous paper[16].

Behavioural task

Participants were trained to produce four five-finger sequences with defined timing
structures, defined by inter-press intervals (IPIs), from memory in a delayed sequence
production paradigm. Go trials began with an abstract fractal image (Sequence cue), which
was associated with a sequence. Following the Sequence cue, a fixation cross was shown to
allow participants to prepare the upcoming sequence. A black hand with a green background
(Go cue) then appeared to cue sequence production from memory. Succeeding the Go cue,
another fixation cross was presented. Feedback was then displayed to participants based on
their performance during the preceding production period, finally followed by an inter-trial
interval where a further fixation cross was displayed. During training, participants learned the
sequences through repeated exposure to visually guided (Instructed) Go trials. These
Instructed Go trials were functionally identical to From Memory Go trials, but featured a Go
cue with a grey background and a red dot on the tip of each finger on the hand image, which
moved from finger to finger in the target production order and in-pace with the target timing
structure. No-Go trials had the same structure as Go trials, but the go cue did not appear
following the preparatory fixation cross. Instead, the fixation cross continued to show for an
extended period. This was succeeded by a further fixation cross, feedback, and ITI, as in Go
trials.

The four five-finger target sequences consisted of permutations of two finger orders
(Order 1 and 2) and two IPI orders (Timing 1 and 2) matched in finger occurrence and
sequence duration. Sequence orders were generated randomly for each participant, making
sure to avoid ascending and descending press triplets and any identical sequences.
Furthermore, each participant's trained sequences began with the same finger press to
avoid differences in the first press driving the decoding of sequence identity during
preparation [81]. Timing structures were the same across participants, which consisted of four
target IPI sequences as follows: 1200ms - 810ms - 350ms - 650ms (Timing 1), and 350ms -
1200ms - 650ms - 810ms (Timing 2).

The trial-by-trial feedback used a points-based scale, ranging from 0 to 10. Points
were awarded based on initiation reaction time and temporal deviation from target timing. 0

points were awarded if the executed press order was incorrect. If the executed press order
was correct, participants were awarded their earned timing points. In No-Go trials, 5 points
were awarded if no press was made as instructed. Participants were presented with a
feedback screen after each trial showing the number of points achieved in the current trial,
as well as visual feedback on whether they pressed the correct finger at the correct time. A
horizontal line was drawn across the centre of the screen, with four symbols displayed
equidistantly along the line representing each of the five finger presses. Correct presses
were indicated by an "X" symbol and incorrect presses were represented by a "–" symbol, for
each respective sequence position. The vertical position of these symbols above ("too late")
or below ("too early") the horizontal line was proportional to the participant's timing of the
respective press relative to target IPI. Using these cues, participants were able to adjust their
performance online to ensure maximum accuracy of sequence production. During the first
two days of training, auditory feedback in the form of successive rising tones corresponding
to the number of points (0 - 10) was played alongside the visual feedback. Auditory feedback
was absent during the fMRI session, to prevent any auditory processing driving decoding
accuracy. All aspects of the behavioural task for this experiment are described in greater
detail in a previous study[16].

Procedure

Training duration was consistent across participants and occurred across the first two
days of the experiment over three distinct training stages. In the first training stage, 80% of
all trials were instructed Go trials, and the remaining 20% were No-Go trials. During the
second training stage, 40% of trials were instructed Go trials, 40% were from-memory Go
trials, and 20% were No-Go trials. In the third and final stage of training, 80% of trials were
from-memory Go trials, and 20% were No-Go trials. The third day took place inside of the
MRI scanner and consisted of a short refresher stage, made up of the same proportion of
trials as the second stage of training, during which T1 images were collected. Next, there
was an fMRI stage consisting of six imaging runs, featuring 50% from-memory Go trials and
50% No-Go trials. In addition, before and after the last training stage, participants completed
a synchronization task which has been described elsewhere[16].

MRI acquisition

Data were acquired using a 32-channel head coil in a Philips Ingenia Elition X 3T MRI
scanner. T1 anatomical scans at a 0.937 x 0.937 x 1 resolution were acquired using
MPRAGE, encoded in the anterior-posterior dimension with an FOV of 240 x 240 x 175 (A-P,
R-L, F-H). For the functional data, T2*-weighted scans were collected across six runs of 230
volumes at a 2 mm isotropic resolution, with 60 slices at a thickness of 2 mm. The functional
images were acquired at a TR of 2s, a TE of 35 ms, and a flip angle of 90°. These were
obtained at a multiband factor of 2, in an interleaved odd-even EPI acquisition. To allow the
stabilization of the magnetic field, four images were discarded at the beginning of each run.
The whole brain of most participants was covered, except for the central prefrontal cortex,
the anterior temporal lobe, and ventral parts of the cerebellum. Jitters were used within each
trial during preparation periods, post-production fixation crosses, and ITIs, to give us a more
accurate estimate of the Hemodynamic Response Function (HRF) by varying which part of
the trial is sampled by each TR[82].

Pre-processing and first-level analysis

All fMRI pre-processing steps were completed using SPM12 (revision 7219) in MATLAB (The MathWorks). We applied slice timing correction using the first slice as a reference to interpolate all other slices to, ensuring analysis occurred on slices which represent the same time point. Realignment and unwarping were conducted using a weighted least-squares method correcting for head movements using a 6-parameter motion algorithm. A mean EPI was produced using SPM's Imcalc function, wherein data acquired across all six runs were combined into a mean EPI image to be co-registered to the anatomical image. Mean EPIs were co-registered to anatomical images using SPM's coreg function, and their alignment was checked and adjusted by hand to improve the alignment, if necessary. All EPI runs were then co-registered to the mean EPI image. For the GLM, regressors were defined for each sequence separately for both preparation and production, resulting in eight regressors of interest per run. Preparation- and production-related BOLD responses were independently modelled from No-Go and Go trials, respectively, to tease out activity from these brief trial phases despite the haemodynamic response lag[83]. The preparation regressor consisted of boxcar function starting at the onset of the Sequence cue in No-Go trials and lasting for the duration of the maximum possible preparation phase (2500ms). The production regressor consisted of a boxcar function starting at the onset of the first press with a fixed duration of 0 (constant impulse), to capture activity related to sequence initiation and extract sequence production-related activity from the first finger press that was matched across sequences within each participant. We aimed at capturing BOLD responses related to neuronal populations that become differentially active for different sequences[84], for which a single estimate of sequence production has been used to successfully identify sequence representations in several previous fMRI studies[12,17,52,81,85]. Additionally, we included several regressors of no interest: (1) error trials (incorrect or premature presses during Go trials and presses during No-Go trials), which were modelled from sequence cue onset to the end of the ITI; (2) the preparation period in Go trials (1000-2500 ms from Sequence cue); and (3) the temporal derivate of each regressor. The boxcar model was then convolved with the standard HRF. To remove the influence of movement-related artifacts, we used a weighted least-squares approach[86]. This resulted in us obtaining beta weight images for each of the eight conditions per scanning run, for all six runs. We further calculated the percent signal change during preparation and production relative to rest and compared each to a baseline of zero using two-tailed one-sample t-tests (Figure 2a). These tests were Bonferroni-corrected two times, to account for phase (2) within each pre-defined ROI. Additionally, we identified significant clusters of activity constrained to each ROI using a random effects analysis with an uncorrected threshold of $t(23) > 3.48$, $p < 0.001$ and a cluster-wise p value for the cluster of that size[87].

Subcortical and cerebellar regions of interest

We used Freesurfer's automatic segmentation[88] to segment subcortical regions of interest consisting of the thalamus, caudate nucleus, putamen, and hippocampus, from each participant's T1 anatomical image. We then resliced each region's mask into the same resolution as the functional images (2x2mm isotropic) and further masked them using functional activity. Only beta weights from voxels within these subcortical functional masks were extracted, constraining analyses to voxels belonging to subcortical regions of interest. Further, to assess the spatial organisation of multivariate patterns, we defined 160-voxel volumetric searchlights with a maximum radius of 6 mm in native space within each subcortical region of interest. LDA accuracy and RSA dissimilarity values (see Linear

discriminant analysis and Representational similarity analysis sections) obtained were
assigned to the centre voxel of the active searchlight.

For the CB, we used the SUIT cerebellar toolbox[89] to segment the whole structure
based on individual anatomical T1 images. We then resliced the lobular probabilistic
cerebellar atlas[90] into each participant's native space to acquire masks for CB lobules IV and
V. We then resliced these masks into functional resolution and further masked them using
functional activity. Beta weights were extracted from these regions using these masks to
constrain analyses to voxels of interest. Additionally, we defined a 160-voxel volumetric
searchlight across the whole cerebellum using each participant's individual anatomy. LDA
accuracy and RSA dissimilarity values were assigned to the centre voxel of the active
searchlight in a similar manner to the subcortical analysis. Classification accuracy and
distance maps were subsequently resliced into SUIT space to display group results on a
cerebellar surface flatmap[91].

Linear discriminant analysis

LDA was used to detect sequence-specific representations[16,17,42], programmed in a
custom-written MATLAB (The MathWorks) code. We extracted mean patterns and common
voxel-by-voxel covariance matrices for each class from the training dataset (five of the six
imaging runs), and then a gaussian linear discriminant classifier was used to distinguish
between the same classes in the test dataset (the remaining imaging run). The factorized
classification of finger order, timing, and integrated order and timing followed the previous
approach[16,17] and was performed on betas estimated from the sequence preparation and
production periods independently. For the decoding of sequence order, the classifier was
trained to distinguish between two sequences with different orders but matching timing
across five runs and was then tested on two sequences with the same orders paired with a
different timing in the remaining run. This classification was then cross-validated across runs
and across training/test sequences, for a total of 12 cross-validation folds. For the decoding
of sequence timing, the classifier was trained to distinguish between two sequences with
differing timings paired with the same order and tested on two sequences with the same two
timings paired with a different order and underwent the same cross-validation procedure.
This method of training and testing the linear discriminant classifier allowed for identification
of sequence feature representations that were transferrable across conditions they are
paired with and therefore independent. The integrated classifier was trained to distinguish
between sequences on five runs and then tested on the remaining run. Here, the mean
activity for each timing (collapsed across two orders) and finger order (collapsed across two
timings) condition within each run was subtracted from the overall activity for each run,
separately[16,17]. This allowed for the measurement of residual activity patterns that were not
explained by a linear combination of timing and order, rather spatiotemporal idiosyncrasies
that would result in the generation of unique kinematics for the respective sequence that
were not transferrable to others. We then trained our integrated classifier to distinguish
between all four sequences based on these residual activity patterns across five imaging
runs and then tested its accuracy on the remaining imaging run, cross-validated across runs.

We normalised classification accuracy by transforming to z scores, assuming a
binomial distribution of the number of correct guesses[17]. We then tested these z scores
against zero (chance level) in subcortical and cerebellar ROIs across participants for
statistical analysis. Z-score transformed classification accuracy maps were subjected to a
random effects analysis, similar to the RSA distance maps, with an uncorrected threshold of

$t(23) > 3.48$, $p < .001$ and a cluster-wise p value for a cluster of that size[87]. Analyses that
considered all voxels within respective regions of interest were also subjected to one-tailed
one-sample t tests relative to zero, Bonferroni-corrected 6 times to account for phase (2) X
classifier (3), to identify decoding accuracy above chance level. We also ran repeated
measures ANOVAs on regional distance measures with factors of phase, classifier, and
region to assess interaction effects and ran post-hoc pairwise comparisons to investigate
significant interaction effects.

Multi-dimensional scaling

We firstly multivariately pre-whitened beta weights prior to multi-dimensional scaling to
increase the reliability of our distance measurements, using a regularised estimate of the
overall noise-covariance matrix[92]. We then identified the first three principal components in
our pre-whitened beta weights using classical multi-dimensional scaling of the variance-
covariance matrix. This method plots all eight conditions in 3D space according to the first
three eigenvectors (principal components) associated with the largest eigenvalues. Given
that activity levels varied within regions across preparation and production (see Results), we
measured the Euclidean distance between preparation and production within each sequence
upon exclusion of the first principal component, since the largest eigenvalues are likely
associated with differences in activity levels across phase. Hence, we calculated the
Euclidean distance between points plotted in the 2D space represented by principal
components one and two. However, given a non-zero level of noise in the data, the
Euclidean distance between two points will be biased away from zero. We therefore
simulated fMRI activity patterns with a matched number of voxels, sample size, and signal to
noise ratio relative to the mean across participants of each ROI collected in the empirical
data. We then calculated the cross-phase within-sequence Euclidean distances to provide
baselines for the one-sample t-tests carried out in each region (Bonferroni corrected seven
times for region). Additionally, these distances were scaled and normalised according to the
average of the average distances between conditions during preparation and production
respectively.

References

1. Bostan, A.C., and Strick, P.L. (2018). The basal ganglia and the cerebellum: Nodes in an integrated network. Nat. Rev. Neurosci. *19*, 338–350. 10.1038/s41583-018-0002-7.

2. Dayan, E., and Cohen, L.G. (2011). Neuroplasticity subserving motor skill learning. Neuron *72*, 443–454. 10.1016/j.neuron.2011.10.008.

3. Hikosaka, O., Nakamura, K., Sakai, K., and Nakahara, H. (2002). Central mechanisms of motor skill learning. Curr. Opin. Neurobiol. *12*, 217–222. 10.1016/S0959-4388(02)00307-0.

4. Verwey, W.B. (2023). C-SMB 2.0: Integrating over 25 years of motor sequencing research with the Discrete Sequence Production task. Psychon. Bull. Rev. 10.3758/s13423-023-02377-0.

5. Bostan, A.C., Dum, R.P., and Strick, P.L. (2013). Cerebellar networks with the cerebral cortex and basal ganglia. Trends Cogn. Sci. *17*, 241–254. 10.1016/j.tics.2013.03.003.

6. Kelly, R.M., and Strick, P.L. (2003). Cerebellar loops with motor cortex and prefrontal cortex of a nonhuman primate. J. Neurosci. Off. J. Soc. Neurosci. *23*, 8432–8444.

7. Graybiel, A.M., and Grafton, S.T. (2015). The Striatum: Where Skills and Habits Meet. Cold Spring Harb. Perspect. Biol. *7*, a021691. 10.1101/cshperspect.a021691.

8. Wong, A.L., and Krakauer, J.W. (2019). Why Are Sequence Representations in Primary Motor Cortex So Elusive? Neuron. 10.1016/j.neuron.2019.09.011.

9. Harpaz, N.K., Hardcastle, K., and Ölveczky, B.P. (2022). Learning-induced changes in the neural circuits underlying motor sequence execution. Curr. Opin. Neurobiol. *76*, 102624.

10. Diedrichsen, J., and Kornysheva, K. (2015). Motor skill learning between selection and execution. Trends Cogn. Sci. *19*. 10.1016/j.tics.2015.02.003.

11. Averbeck, B.B., Chafee, M.V., Crowe, D.A., and Georgopoulos, A.P. (2003). Neural activity in prefrontal cortex during copying geometrical shapes. I. Single cells encode shape, sequence, and metric parameters. Exp. Brain Res. *150*, 127–141. 10.1007/s00221-003-1416-6.

12. Berlot, E., Popp, N.J., and Diedrichsen, J. (2020). A critical re-evaluation of fmri signatures of motor sequence learning. eLife *9*, 1–24. 10.7554/eLife.55241.

13. Berlot, E., Popp, N.J., Grafton, S.T., and Diedrichsen, J. (2021). Combining Repetition Suppression and Pattern Analysis Provides New Insights into the Role of M1 and Parietal Areas in Skilled Sequential Actions. J. Neurosci. *41*, 7649–7661. 10.1523/jneurosci.0863-21.2021.

14. Mushiake, H., Saito, N., Sakamoto, K., Itoyama, Y., and Tanji, J. (2006). Activity in the Lateral Prefrontal Cortex Reflects Multiple Steps of Future Events in Action Plans. Neuron *50*, 631–641. 10.1016/j.neuron.2006.03.045.

15. Shima, K., and Tanji, J. (1998). Both supplementary and presupplementary motor areas are crucial for the temporal organization of multiple movements. J. Neurophysiol. *80*, 3247–3260.

16. Yewbrey, R., Mantziara, M., and Kornysheva, K. (2023). Cortical Patterns Shift from Sequence Feature Separation during Planning to Integration during Motor Execution. J. Neurosci. *43*, 1742–1756. 10.1523/JNEUROSCI.1628-22.2023.

17. Kornysheva, K., and Diedrichsen, J. (2014). Human premotor areas parse sequences into their spatial and temporal features. eLife *3*, e03043. 10.7554/eLife.03043.

18. Shima, K., and Tanji, J. (2000). Neuronal activity in the supplementary and presupplementary motor areas for temporal organization of multiple movements. J. Neurophysiol. *84*, 2148–2160.

19. Yokoi, A., and Diedrichsen, J. (2019). Neural Organization of Hierarchical Motor Sequence Representations in the Human Neocortex. Neuron *103*, 1178-1190.e7. 10.1016/j.neuron.2019.06.017.

20. Zimnik, A.J., and Churchland, M.M. (2021). Independent generation of sequence elements by motor cortex. Nat. Neurosci. *24*, 412–424. 10.1038/s41593-021-00798-5.

21. Wolff, S.B.E., Ko, R., and Ölveczky, B.P. (2022). Distinct roles for motor cortical and thalamic inputs to striatum during motor skill learning and execution. Sci. Adv. *8*. 10.1126/sciadv.abk0231.

22. Dhawale, A.K., Wolff, S.B.E., Ko, R., and Ölveczky, B.P. (2021). The basal ganglia control the detailed kinematics of learned motor skills. Nat. Neurosci. *24*, 1256–1269. 10.1038/s41593-021-00889-3.

23. Brooks, D.N., and Baddeley, A.D. (1976). What can amnesic patients learn? Neuropsychologia *14*, 111–122. 10.1016/0028-3932(76)90012-9.

24. Cohen, N.J., and Squire, L.R. (1980). Preserved Learning and Retention of Pattern-Analyzing Skill in Amnesia: Dissociation of Knowing How and Knowing That. Science *210*, 207–210. 10.1126/science.7414331.

25. Albouy, G., Sterpenich, V., Balteau, E., Vandewalle, G., Desseilles, M., Dang-Vu, T., Darsaud, A., Ruby, P., Luppi, P.-H., Degueldre, C., et al. (2008). Both the hippocampus and striatum are involved in consolidation of motor sequence memory. Neuron *58*, 261–272. 10.1016/j.neuron.2008.02.008.

26. Albouy, G., King, B.R., Maquet, P., and Doyon, J. (2013). Hippocampus and striatum: dynamics and interaction during acquisition and sleep-related motor sequence memory consolidation. Hippocampus *23*, 985–1004. 10.1002/hipo.22183.

27. Buch, E.R., Claudino, L., Quentin, R., Bönstrup, M., and Cohen, L.G. (2021). Consolidation of human skill linked to waking hippocampo-neocortical replay. Cell Rep. *35*, 109193. 10.1016/j.celrep.2021.109193.

28. Doyon, J., Bellec, P., Amsel, R., Penhune, V., Monchi, O., Carrier, J., Lehéricy, S., and Benali, H. (2009). Contributions of the basal ganglia and functionally related brain structures to motor learning. Behav. Brain Res. *199*, 61–75. 10.1016/j.bbr.2008.11.012.

29. Lungu, O., Monchi, O., Albouy, G., Jubault, T., Ballarin, E., Burnod, Y., and Doyon, J. (2014). Striatal and Hippocampal Involvement in Motor Sequence Chunking Depends on the Learning Strategy. PLoS ONE *9*, e103885. 10.1371/journal.pone.0103885.

30. Kumar, V.J., van Oort, E., Scheffler, K., Beckmann, C.F., and Grodd, W. (2017).
Functional anatomy of the human thalamus at rest. NeuroImage *147*, 678–691.
10.1016/j.neuroimage.2016.12.071.

31. Duque, J., and Ivry, R.B. (2009). Role of corticospinal suppression during motor
preparation. Cereb. Cortex *19*, 2013–2024. 10.1093/cercor/bhn230.

32. Ariani, G., Pruszynski, J.A., and Diedrichsen, J. (2022). Motor planning brings human
primary somatosensory cortex into action-specific preparatory states. eLife *11*.
10.7554/elife.69517.

33. Kaufman, M.T., Churchland, M.M., Ryu, S.I., and Shenoy, K.V. (2014). Cortical activity in
the null space: Permitting preparation without movement. Nat. Neurosci. *17*, 440–448.
10.1038/nn.3643.

34. Khilkevich, A., Zambrano, J., Richards, M.-M., and Mauk, M.D. (2018). Cerebellar
implementation of movement sequences through feedback. eLife *7*, e37443.
10.7554/eLife.37443.

35. Christian, K.M., and Thompson, R.F. (2003). Neural Substrates of Eyeblink Conditioning:
Acquisition and Retention. Learn. Mem. *10*, 427–455. 10.1101/lm.59603.

36. Frank, M.J. (2011). Computational models of motivated action selection in corticostriatal
circuits. Curr. Opin. Neurobiol. *21*, 381–386. 10.1016/j.conb.2011.02.013.

37. Gao, Z., Davis, C., Thomas, A.M., Economo, M.N., Abrego, A.M., Svoboda, K., De
Zeeuw, C.I., and Li, N. (2018). A cortico-cerebellar loop for motor planning. Nature *563*,
113–116. 10.1038/s41586-018-0633-x.

38. Mizes, K.G.C., Lindsey, J., Escola, G.S., and Ölveczky, B.P. (2023). Dissociating the
contributions of sensorimotor striatum to automatic and visually guided motor
sequences. Nat. Neurosci. *26*, 1791–1804. 10.1038/s41593-023-01431-3.

39. Cisek, P., and Kalaska, J.F. (2005). Neural correlates of reaching decisions in dorsal
premotor cortex: Specification of multiple direction choices and final selection of action.
Neuron *45*, 801–814. 10.1016/j.neuron.2005.01.027.

40. Davachi, L., and DuBrow, S. (2015). How the hippocampus preserves order: the role of
prediction and context. Trends Cogn. Sci. *19*, 92–99. 10.1016/j.tics.2014.12.004.

41. Jafarpour, A., Lin, J.J., Knight, R.T., and Buffalo, E.A. (2023). Multiple memory systems
for efficient temporal order memory. Hippocampus *33*, 1154–1157. 10.1002/hipo.23550.

42. Kornysheva, K., Bush, D., Meyer, S.S., Sadnicka, A., Barnes, G., and Burgess, N.
(2019). Neural Competitive Queuing of Ordinal Structure Underlies Skilled Sequential
Action. Neuron *101*, 1166-1180.e3. 10.1016/j.neuron.2019.01.018.

43. Rhodes, E., Lamar, M., Libon, D.J., and Giovannetti, T. (2019). Memory for Serial Order
in Alzheimer's Disease and Vascular Dementia: A Competitive Queuing Analysis. Arch.
Clin. Neuropsychol. *34*, 2–13. 10.1093/arclin/acy013.

44. Houghton, G., and Hartley, T. (1995). Parallel models of serial behavior: Lashley
revisited. Psyche (Stuttg.) *2*, 2–25.

45. Averbeck, B.B., Chafee, M.V., Crowe, D.A., and Georgopoulos, A.P. (2002). Parallel
processing of serial movements in prefrontal cortex. Proc. Natl. Acad. Sci. U. S. A. *99*,
13172–13177. 10.1073/pnas.162485599.

46. Dragoi, G., and Tonegawa, S. (2011). Preplay of future place cell sequences by
hippocampal cellular assemblies. Nature. 10.1038/nature09633.

47. Henderson, M.M., Rademaker, R.L., and Serences, J.T. (2022). Flexible utilization of
spatial- and motor-based codes for the storage of visuo-spatial information. eLife *11*,
e75688. 10.7554/eLife.75688.

48. Shushruth, S., Zylberberg, A., and Shadlen, M.N. (2022). Sequential sampling from
memory underlies action selection during abstract decision-making. Curr. Biol. *32*, 1949-
1960.e5. 10.1016/j.cub.2022.03.014.

49. Huang, C.-C., Rolls, E.T., Hsu, C.-C.H., Feng, J., and Lin, C.-P. (2021). Extensive
Cortical Connectivity of the Human Hippocampal Memory System: Beyond the "What"
and "Where" Dual Stream Model. Cereb. Cortex N. Y. NY *31*, 4652–4669.
10.1093/cercor/bhab113.

50. Rolls, E.T. (2020). Spatial coordinate transforms linking the allocentric hippocampal and
egocentric parietal primate brain systems for memory, action in space, and navigation.
Hippocampus *30*, 332–353. 10.1002/hipo.23171.

51. Whitlock, J.R., Sutherland, R.J., Witter, M.P., Moser, M.-B., and Moser, E.I. (2008).
Navigating from hippocampus to parietal cortex. Proc. Natl. Acad. Sci. *105*, 14755–
14762. 10.1073/pnas.0804216105.

52. Wiestler, T., and Diedrichsen, J. (2013). Skill learning strengthens cortical
representations of motor sequences. eLife *2013*, 801. 10.7554/eLife.00801.

53. Gurney, K., Prescott, T.J., and Redgrave, P. (2001). A computational model of action
selection in the basal ganglia. I. A new functional anatomy. Biol. Cybern. *84*, 401–410.
10.1007/PL00007984.

54. Mink, J.W. (1996). THE BASAL GANGLIA: FOCUSED SELECTION AND INHIBITION
OF COMPETING MOTOR PROGRAMS. Prog. Neurobiol. *50*, 381–425. 10.1016/S0301-
0082(96)00042-1.

55. Redgrave, P., Prescott, T.J., and Gurney, K. (1999). The basal ganglia: a vertebrate
solution to the selection problem? Neuroscience *89*, 1009–1023.

56. Friend, D.M., and Kravitz, A.V. (2014). Working together: basal ganglia pathways in
action selection. Trends Neurosci. *37*, 301–303. 10.1016/j.tins.2014.04.004.

57. Graybiel, A.M. (1998). The basal ganglia and chunking of action repertoires. In
Neurobiology of Learning and Memory (Neurobiol Learn Mem), pp. 119–136.
10.1006/nlme.1998.3843.

58. Wymbs, N.F., Bassett, D.S., Mucha, P.J., Porter, M.A., and Grafton, S.T. (2012).
Differential Recruitment of the Sensorimotor Putamen and Frontoparietal Cortex during
Motor Chunking in Humans. Neuron *74*, 936–946. 10.1016/j.neuron.2012.03.038.

59. Wymbs, N.F., and Grafton, S.T. (2015). The human motor system supports sequence-
specific representations over multiple training-dependent timescales. Cereb. Cortex *25*,
4213–4225. 10.1093/cercor/bhu144.

60. Bednark, J.G., Campbell, M.E.J., and Cunnington, R. (2015). Basal ganglia and cortical
networks for sequential ordering and rhythm of complex movements. Front. Hum.
Neurosci. *9*, 421. 10.3389/fnhum.2015.00421.

61. Emmons, E.B., De Corte, B.J., Kim, Y., Parker, K.L., Matell, M.S., and Narayanan, N.S.
(2017). Rodent Medial Frontal Control of Temporal Processing in the Dorsomedial
Striatum. J. Neurosci. Off. J. Soc. Neurosci. *37*, 8718–8733.
10.1523/JNEUROSCI.1376-17.2017.

62. Wang, J., Narain, D., Hosseini, E.A., and Jazayeri, M. (2018). Flexible timing by
temporal scaling of cortical responses. Nat. Neurosci. *21*, 102–112. 10.1038/s41593-
017-0028-6.

63. Grahn, J. a, and Brett, M. (2007). Rhythm and beat perception in motor areas of the
brain. J. Cogn. Neurosci. *19*, 893–906. 10.1162/jocn.2007.19.5.893.

64. Mizes, K.G.C., Lindsey, J., Escola, G.S., and Ölveczky, B.P. (2023). Motor cortex is
required for flexible but not automatic motor sequences. Preprint at bioRxiv,
10.1101/2023.09.05.556348 10.1101/2023.09.05.556348.

65. Diener, H.-C., and Dichgans, J. (1992). Pathophysiology of cerebellar ataxia. Mov.
Disord. *7*, 95–109. 10.1002/mds.870070202.

66. Miall, R.C., Christensen, L.O.D., Cain, O., and Stanley, J. (2007). Disruption of State
Estimation in the Human Lateral Cerebellum. PLoS Biol. *5*, e316.
10.1371/journal.pbio.0050316.

67. Donchin, O., Rabe, K., Diedrichsen, J., Lally, N., Schoch, B., Gizewski, E.R., and
Timmann, D. (2012). Cerebellar regions involved in adaptation to force field and
visuomotor perturbation. J. Neurophysiol. *107*, 134–147. 10.1152/jn.00007.2011.

68. Chabrol, F.P., Blot, A., and Mrsic-Flogel, T.D. (2019). Cerebellar Contribution to
Preparatory Activity in Motor Neocortex. Neuron *103*, 506-519.e4.
10.1016/j.neuron.2019.05.022.

69. Li, N., and Mrsic-Flogel, T.D. (2020). Cortico-cerebellar interactions during goal-directed
behavior. Curr. Opin. Neurobiol. *65*, 27–37. 10.1016/j.conb.2020.08.010.

70. Guo, Z.V., Inagaki, H.K., Daie, K., Druckmann, S., Gerfen, C.R., and Svoboda, K.
(2017). Maintenance of persistent activity in a frontal thalamocortical loop. Nature *545*,
181–186. 10.1038/nature22324.

71. Inagaki, H.K., Chen, S., Ridder, M.C., Sah, P., Li, N., Yang, Z., Hasanbegovic, H., Gao,
Z., Gerfen, C.R., and Svoboda, K. (2022). A midbrain-thalamus-cortex circuit
reorganizes cortical dynamics to initiate movement. Cell *185*, 1065-1081.e23.
10.1016/j.cell.2022.02.006.

72. Sherman, S.M., and Guillery, R.W. (2002). The role of the thalamus in the flow of
information to the cortex. Philos. Trans. R. Soc. B Biol. Sci. *357*, 1695–1708.
10.1098/rstb.2002.1161.

73. Sherman, S.M. (2007). The thalamus is more than just a relay. Curr. Opin. Neurobiol.
17, 417–422. 10.1016/j.conb.2007.07.003.

74. Ariani, G., Kordjazi, N., Pruszynski, J.A., and Diedrichsen, J. (2021). The Planning
Horizon for Movement Sequences. eNeuro *8*, ENEURO.0085-21.2021.
10.1523/ENEURO.0085-21.2021.

75. Ariani, G., and Diedrichsen, J. (2019). Sequence learning is driven by improvements in
motor planning. J. Neurophysiol. *121*, 2088–2100. 10.1152/jn.00041.2019.

76. Kaufman, M.T., Seely, J.S., Sussillo, D., Ryu, S.I., Shenoy, K.V., and Churchland, M.M.
(2016). The largest response component in the motor cortex reflects movement timing
but not movement type. eNeuro *3*. 10.1523/ENEURO.0085-16.2016.

77. Wilkinson, L., Khan, Z., and Jahanshahi, M. (2009). The role of the basal ganglia and its
cortical connections in sequence learning: Evidence from implicit and explicit sequence
learning in Parkinson's disease. Neuropsychologia *47*, 2564–2573.
10.1016/j.neuropsychologia.2009.05.003.

78. Doyon, J., Gaudreau, D., Jr., R.L., Castonguay, M., Bédard, P.J., Bédard, F., and
Bouchard, J.-P. (1997). Role of the Striatum, Cerebellum, and Frontal Lobes in the
Learning of a Visuomotor Sequence. Brain Cogn. *34*, 218–245.
10.1006/brcg.1997.0899.

79. Förstl, H., and Kurz, A. (1999). Clinical features of Alzheimer's disease. Eur. Arch.
Psychiatry Clin. Neurosci. *249*, 288–290. 10.1007/s004060050101.

80. Oldfield, R.C. (1971). The assessment and analysis of handedness: The Edinburgh
inventory. Neuropsychologia *9*, 97–113. 10.1016/0028-3932(71)90067-4.

81. Yokoi, A., Arbuckle, S.A., and Diedrichsen, J. (2018). The role of human primary motor
cortex in the production of skilled finger sequences. J. Neurosci. *38*, 1430–1442.
10.1523/JNEUROSCI.2798-17.2017.

82. Serences, J.T. (2004). A comparison of methods for characterizing the event-related
BOLD timeseries in rapid fMRI. NeuroImage *21*, 1690–1700.
10.1016/j.neuroimage.2003.12.021.

83. Logothetis, N.K. (2003). The underpinnings of the BOLD functional magnetic resonance
imaging signal. J. Neurosci. *23*, 3963–3971. 10.1523/jneurosci.23-10-03963.2003.

84. Tanji, J., and Shima, K. (1994). Role for supplementary motor area cells in planning
several movements ahead. Nature *371*, 413–416. 10.1038/371413a0.

85. Nambu, I., Hagura, N., Hirose, S., Wada, Y., Kawato, M., and Naito, E. (2015). Decoding
sequential finger movements from preparatory activity in higher-order motor regions: A
functional magnetic resonance imaging multi-voxel pattern analysis. Eur. J. Neurosci.
42, 2851–2859. 10.1111/ejn.13063.

86. Diedrichsen, J., and Shadmehr, R. (2005). Detecting and adjusting for artifacts in fMRI
time series data. NeuroImage *27*, 624–634. 10.1016/j.neuroimage.2005.04.039.

87. Worsley, K.J., Marrett, S., Neelin, P., Vandal, A.C., Friston, K.J., and Evans, A.C.
(1996). A unified statistical approach for determining significant signals in images of

cerebral activation. Hum. Brain Mapp. *4*, 58–73. 10.1002/(SICI)1097-
0193(1996)4:1<58::AID-HBM4>3.0.CO;2-O.

88. Dale, A.M., Fischl, B., and Sereno, M.I. (1999). Cortical surface-based analysis: I.
Segmentation and surface reconstruction. NeuroImage *9*, 179–194.
10.1006/nimg.1998.0395.

89. Diedrichsen, J. (2006). A spatially unbiased atlas template of the human cerebellum.
NeuroImage *33*, 127–138. 10.1016/j.neuroimage.2006.05.056.

90. Diedrichsen, J., Balsters, J.H., Flavell, J., Cussans, E., and Ramnani, N. (2009). A
probabilistic MR atlas of the human cerebellum. NeuroImage *46*, 39–46.
10.1016/j.neuroimage.2009.01.045.

91. Diedrichsen, J., and Zotow, E. (2015). Surface-based display of volume-averaged
cerebellar imaging data. PLoS ONE *10*. 10.1371/journal.pone.0133402.

92. Walther, A., Nili, H., Ejaz, N., Alink, A., Kriegeskorte, N., and Diedrichsen, J. (2016).
Reliability of dissimilarity measures for multi-voxel pattern analysis. NeuroImage *137*,
188–200. 10.1016/j.neuroimage.2015.12.012.

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