Title: Long-sperm precedence and other cryptic female choices in *Drosophila melanogaster*

Authors: Brooke Peckenpaugh[1], Joanne Y. Yew[2], and Leonie C. Moyle[1]

[1] Department of Biology, Indiana University, Bloomington, Indiana 47405
[2] Pacific Biosciences Research Center, University of Hawaiʻi at Mānoa, Honolulu, Hawaiʻi 96822

Corresponding author

Leonie C. Moyle, Department of Biology, Indiana University, 1001 E 3rd Street, Bloomington, IN 47405-7000, USA.
Email: lmoyle@iu.edu

Data availability statement

The data and code underlying this article are available in the Dryad Digital Repository, at https://dx.doi.org/[doi].

Author contributions

BP and LCM conceived and designed the experiment. BP performed the experiment and statistical analyses. JYY performed the chemical analysis. BP and LCM wrote the paper. All authors read and provided feedback on the manuscript.

Funding

Indiana University, Bloomington (LCM)
NIH Common Themes in Reproductive Diversity (CTRD) training grant (BP)
National Institute of General Medical Sciences of the National Institutes of Health Grant No. P20GM125508 (JYY), P20GM139753 (JYY)

Conflict of interest statement

The authors have no conflict of interest to declare.

Acknowledgments

We thank Stuart J. Macdonald for providing the DSPR founder lines used in this study, and Brandon S. Cooper for providing the *Drosophila yakuba* line. We thank Sung Nichin for assistance scoring courtship behaviors. We thank Alison Pischedda for advice on the perfuming protocol. All GCMS measurements were performed in the Univ. of Hawaiʻi at Mānoa Microbial Genomics and Analytical Laboratory core. We thank Matthew W. Hahn, Kimberly A. Rosvall, and Michael J. Wade for their helpful comments on the manuscript. The research was supported by Indiana University Department of Biology funding to LCM and BP, as well as the NIH Common Themes in Reproductive Diversity (CTRD) training grant funding to BP.

LONG-SPERM PRECEDENCE AND OTHER CRYPTIC FEMALE CHOICES IN

DROSOPHILA MELANOGASTER

Abstract

Females that mate multiply make postmating choices about which sperm fertilize their

eggs (cryptic female choice); however, the male characteristics they use to make such choices

remain unclear. In this study, we sought to understand female sperm use patterns by evaluating

whether *Drosophila melanogaster* females adjust sperm use (second male paternity) in response

to four main factors: male genotype, male courtship effort, male pheromone alteration, and male

postmating reproductive morphology. Our experiment was replicated across four different *D.*

melanogaster lines, in a full factorial design, including a pheromone manipulation in which

second males were perfumed to resemble heterospecific (*D. yakuba*) males. We found that

females prefer longer sperm—regardless of mating order—in almost all contexts; this observed

pattern of 'long-sperm precedence' is consistent with female postmating choice of high-fitness

male traits. Nonetheless, we also found that this general preference can be plastically altered by

females in response to effects including perfuming treatment; this differential female sperm use

is between otherwise identical males, and therefore solely female-mediated. Furthermore, our

finding that females exercise choice using diverse criteria suggests a possible mechanism for the

maintenance of variation in sexually selected male traits.

Keywords: cryptic female choice, sperm, pheromones, *Drosophila*

     Teaser text

How do females make choices about which sperm will fertilize their eggs? In this study,

we assess four geographically diverse *Drosophila melanogaster* lines to show that females

strongly prefer long sperm—a pattern consistent with female preference for high-fitness males.

In addition, we perfumed males to show that females can adjust this general preference based on

other factors, including pheromones. Because this adjustment occurs between genotypically

identical males, it is entirely female-mediated.

1   Introduction

Reproduction in nature is costly; organisms are therefore expected to use their

reproductive resources judiciously and to adjust them according to reproductive context.

Examples of such behavioral reproductive plasticity span diverse traits and taxa, from palm trees

(e.g., sex allocation in *Attalea speciosa*, Barot et al. 2005) to guppies (e.g., sperm use in *Poecilia*

reticulata, Gasparini & Evans 2018) to flies (e.g., aggression in *Drosophila melanogaster*,

Nandy et al. 2016). One specific strategy for plastically allocating reproductive resources is to

adjust choices about reproductive partners. Females, in particular, are known to do this in

numerous ways prior to mating. For instance, in red jungle fowl, females choose to mate with

males with brighter feathers and longer tails; these traits are more prominent in unparasitized

males and are therefore honest indicators of male health (Zuk et al. 1990). While many such

cases involve choices exercised before mating, some organisms also have the ability to

selectively allocate postmating reproductive resources; for instance, female guppies fertilize

more eggs when mating with more colorful focal males in comparison to stimulus males (Pilastro

et al. 2004). However, because most of these choices are made during and after mating events, it

remains hard to differentiate the relative contributions of each mating partner, the criteria on

which their choices are based, and therefore the degree to which these choices could be adaptive.

94   Interestingly, in *Drosophila*, postmating reproductive plasticity has primarily been

studied in males. Even under optimal conditions, *D. melanogaster* males are known to be

ejaculate-limited, with their progeny output declining after two to five matings in a day (Douglas

et al. 2020, Hopkins et al. 2019, Linklater et al. 2007). Plastic allocation of reproductive

resources could therefore allow them to prioritize sperm and seminal fluid for preferred mates.

Consistent with this, *D. melanogaster* males use premating information, such as female condition

and the relative abundance of potential sexual competitors, to modulate ejaculate composition

(Garbaczewska et al. 2013, Lüpold et al. 2011, Sirot et al. 2011, Sirot 2019, Wigby et al. 2009).

D. melanogaster males also allocate more resources to matings with large females—an

investment that reduces their success at defensive sperm competition in subsequent matings

(Anastasio et al. 2023). Together, these studies broadly indicate that costs to *D. melanogaster*

males drive plasticity in their reproductive allocation during mating, as well as identify specific

criteria used in making these behavioral decisions.

107   Like males, females are also expected to selectively allocate reproductive resources (i.e.,

eggs) at postmating stages, through the action of "cryptic female choice"—the female-mediated

physical and chemical factors that determine which sperm are used to fertilize eggs after mating

(Eberhard 1996, Firman et al. 2017, Thornhill 1983). In *Drosophila,* several proximate

mechanisms of cryptic female choice involve modulating the storage of sperm prior to

fertilization. *Drosophila* females have two types of sperm storage organs and can bias sperm use

through differential sperm storage and/or by preferentially drawing sperm from these alternative

organs (Manier et al. 2013b). Females can also manipulate the identity of sperm available in

storage by adjusting the time until sperm ejection after specific matings, which terminates the

displacement of resident sperm by incoming sperm (Laturney et al. 2018, Lüpold et al. 2013,

Manier et al. 2010, Manier et al. 2013a, Snook & Hosken 2004). Finally, it has been proposed

that the size of sperm storage organs themselves might provide a mechanism by which females

make these choices. For example, females experimentally evolved to have longer seminal

receptacles have been shown to select for males with longer sperm (Miller & Pitnick 2002). Even

when the specific mechanism is unknown, it is evident that *Drosophila* females can change

sperm use patterns depending upon context, including according to male genotype identity

(Chow et al. 2010). What is comparatively poorly understood, however, is the evolutionary

explanation for these apparent female choices, including the criteria used by females to exercise

them.

One reason for this is the challenge of differentiating female choice from primarily male-

mediated factors that can also influence the outcome of postmating processes (Firman et al.

2017). For instance, *Drosophila simulans* males that are more attractive to females also sire more

offspring as second males; however, it is unclear whether this is because females preferentially

use sperm from attractive males, or because attractive males are intrinsically better sperm

competitors (Hosken et al. 2008). While the former explanation would be consistent with an

adaptive function for cryptic female choice, the latter explanation implies passive female

responses to the outcome of male-male sexual selection. One strategy for determining the

specific contribution of female choices to patterns of biased sperm use is to evaluate how these

patterns change when female perception of male identity is manipulated, without influencing

male identity itself. Although female assessment of male identity is likely to be complex, in

Drosophila, female premating preference primarily depends on auditory and chemosensory cues

(Greenspan & Ferveur 2000, Yamamoto & Nakano 1999); manipulating these cues could

therefore modify this assessment. Contact pheromones are one such cue. In insects broadly, and

flies specifically, these pheromones include cuticular hydrocarbons (CHCs)—waxy compounds

on the outer surface (cuticle) of many insects that can be used as sexual pheromones in mate

choice by males (Billeter et al. 2009) and females (Scott 1994), both within and between species

(Blows and Allan 1998). CHC composition can be artificially altered to assess its effect on mate

choice by "perfuming" focal males with the CHCs of other male genotypes (Dyer et. al. 2014,

Wang et al. 2011). For instance, *D. melanogaster* males have divergent pheromone profiles from

the closely related species *Drosophila yakuba,* both in the abundance of CHCs and in the

presence or absence of some CHCs (Figure S1, Khallaf et al. 2021). Perfuming *D. melanogaster*

males with CHCs from *D. yakuba* males, immediately prior to mating, can therefore manipulate

female perception of male identity while keeping male postmating quality the same.

In this study, we sought to understand the nature and relative importance of factors that

females use when exercising cryptic female choice. To do so, we evaluated whether *Drosophila*

melanogaster females adjust their sperm use patterns in response to four main factors: male

genotype, male courtship effort, male pheromone alteration, and male postmating reproductive

morphology (the morphological traits relevant to postmating reproductive success). We

manipulated male pheromones via "perfuming" with heterospecific (*D. yakuba*) males—thereby

changing the perceived, but not actual, identity of these treatment males. We also evaluated

postmating reproductive trait variation in males and females (testis length, sperm length, male

reproductive tract [MRT] mass, sperm storage organ size, female reproductive tract [FRT] mass)

to assess their contribution to sperm preference patterns. We assessed how second-male paternity

(proportion of offspring sired) changed according to each factor, by mating males of each type to

females that had previously mated with a consistent first-male tester genotype. Our experiment

was replicated across four different *D. melanogaster* lines in a full factorial design, to evaluate

whether the effects of these factors on sperm use depend upon female genotype, male genotype,

or their interaction. We find clear evidence that females adjust their use of sperm from the second

male. In particular, females show a general preference for longer sperm, whether that sperm is

from the first or the second male—i.e. 'long-sperm precedence' rather than 'last-male sperm

precedence' (Laturney et al. 2018). Nonetheless, we also find that this general preference can be

overridden in specific contexts by female-mediated changes in response to male genotype,

courtship effort, and perfuming treatment. The presence and direction of these cryptic choices

vary in complex ways, including evidence that females of some genotypes adjust sperm use

based on their perception of male identity, regardless of actual identity. These data allow us to

draw broader inferences about the possible criteria for, as well as the evolutionary causes and

consequences of, such choices.

2   Methods

2.1 Fly stocks

All stocks were reared on standard media prepared by the Bloomington Drosophila Stock

Center and kept at room temperature (~22°C). For this experiment we used four lines from

among the 15 *D. melanogaster* founder lines in the Drosophila Synthetic Population Resource

(DSPR) (King et al. 2012). These fully-sequenced founder lines represent a diverse collection of

genotypes from different locations around the world, and are known to vary in reproductive traits

(i.e., male courtship song, see Pischedda et al. 2014a). Our four DSPR founder lines (A5

[Greece], A7 [Taiwan], B4 [California], and B7 [Malaysia]) were used as focal genotypes for

females and second males in each manipulation. The GFP-labeled *D. melanogaster* line (stock

#32170 from the Bloomington *Drosophila* Stock Center) was used as the first tester male. The *D.*

yakuba line yak_S09_L38 was the heterospecific donor male for the heterospecific-perfumed

treatment (collected by B. Cooper in Sao Tome in 2009). For each conspecific-perfumed

treatment, target males were perfumed (see methods below) with males from their own genotype.

2.2 Experimental design

To assess patterns of preferential sperm use, including after remating with conspecific-

vs. heterospecific-perfumed males, we mated *D. melanogaster* females sequentially to two

males: first, to *D. melanogaster* GFP-labeled tester males, and second, to *D. melanogaster* target

males. Using a GFP-labeled first male allowed us to assess paternity share (first [tester] versus

second [target] males) of offspring from each female) by examining the phenotypes of offspring

generated after the second mating. Any offspring not expressing GFP are sired by the second

focal male. For our CHC manipulation, the second, target males were either perfumed by

genotypically identical males (conspecific-perfumed control) or perfumed by heterospecific *D.*

yakuba males (Figure 1).

We replicated this experimental procedure across all four *D. melanogaster* genotypes, in

a full factorial design. For each line, we set up matings between females and second males from

each of the four DSPR lines, including males from the females' own genotype. This created 16

cross types in total. For each cross-type, we assessed paternity share from crosses with

conspecific-perfumed males and with heterospecific-perfumed males. Each of these cross types

was repeated until successfully replicated at least five times, resulting in over 160 total crosses.

2.3 Perfuming manipulation

Males used in the second mating were perfumed in the morning before setting up crosses.

To do so, we vortexed four target males with 30 donor males, either of the same *D. melanogaster*

genotype (for conspecific-perfumed control crosses) or with 30 heterospecific (*D. yakuba*) donor

males (for heterospecific-perfumed treatment crosses). For both treatments, we first removed

wings from the donor males under CO_2 anesthesia in order to differentiate them from target

males. The flies were vortexed on medium strength for one minute total, in three 20-second

increments with 20-second breaks in between (Pischedda et al. 2014b). The target males were

separated immediately after vortexing and isolated individually in vials. They were allowed to

recover for one hour before setting up crosses.

2.4 Paternity assay

D. melanogaster males were isolated and aged for seven days, while females were

isolated and aged for two days before setting up the first cross with a GFP-labeled tester male.

We allowed mating for 24 hours before removing the males. We allowed the isolated female to

lay eggs for an additional 24 hours and verified successful matings by observing larval activity.

Unmated females (those that failed to produce larvae after the first mating) were removed from

the experiment. Mated females were then moved to a new vial to be paired with the second male.

To determine whether and the degree to which females exhibit premating discrimination against

heterospecific-perfumed males, we observed a subset of crosses for two hours to observe the

number of second matings within this period and recorded the time until remating (latency) and

copulation duration for each (these data are not presented because only 9 of 162 scored pairings

resulted in copulation within two hours). Regardless of whether matings were observed in the

first two hours, seven-day-old second males were housed with females for 24 hours. Males were

then removed and the females were transferred to a new vial. Females continued to be transferred

to new vials every 24 hours for three days. Progeny in these vials were allowed to mature into

adulthood, then counted and assessed for GFP to determine paternity (see details below). All

cross types were repeated until we had five successful replicates (crosses in which there is

evidence for both a first and second mating). Conspecific- and heterospecific-perfumed treatment

crosses for each genotype were completed in parallel and in a randomized order, and the

researcher counting progeny was blind to which cross type or treatment they were measuring.

Upon eclosion, all progeny from the three post-remating vials were anesthetized with

CO_2 and viewed under a Leica M205FA stereo scope equipped with a UV light for visualizing

GFP. The presence or absence of GFP in the ocelli of the eye were used as a marker of paternity;

offspring with GFP ocelli were sired by the first male and were thus counted as P1. Progeny with

wild-type ocelli were sired by the second male and were counted as P2. If no wild-type progeny

were present in the three post-remating vials, we assumed that no remating event occurred, and

these were scored as "unremated".

2.5 Cuticular hydrocarbon extraction and quantification

We confirmed the efficacy of perfuming using gas chromatography–mass spectrometry

(GC/MS) analysis. We extracted cuticular hydrocarbons from males of all four genotypes, for

both conspecific-perfumed and heterospecific-perfumed treatments. Following treatment of

seven-day old virgin males, cuticular hydrocarbons were extracted from pooled samples by

placing five males in a 1.8 mL glass vial (Wheaton 224740 E-C Clear Glass Sample Vials) with

120 μL of hexane (Sigma Aldrich, St Louis, MO, USA) spiked with 10 μg/mL of hexacosane

(Sigma Aldrich). Hexane is a standard non-polar solvent used to extract hydrophobic molecules

from insect cuticles. The alkane hexacosane is not endogenously produced by *Drosophila* and

serves as an internal standard. After 20 min, 100 μL of the solution was removed to a sterilized

1.8 mL glass vial (Wheaton 224740 E-C Clear Glass Sample Vials) and allowed to evaporate

under a fume hood, such that only the extracted cuticular hydrocarbons remained at the bottom

of the tube. Extracts were then stored at −20°C until analysis. The samples were reconstituted in

100 μL of hexane immediately before analysis. Five replicate samples, consisting of five flies per

sample (25 flies total), were prepared for each sample type.

We performed gas chromatography–mass spectrometry (GC/MS) analysis on a 7820A

GC system equipped with a 5975 Mass Selective Detector (Agilent Technologies, Inc., Santa

Clara, CA, USA) and a HP-5ms column ((5%-Phenyl)-methylpolysiloxane, 30 m length, 250 μm

ID, 0.25 μm film thickness; Agilent Technologies, Inc.). Electron ionization (EI) energy was set

at 70 eV. One microliter of the sample was injected in splitless mode and analyzed with helium

flow at 1 mL/min. The column was set at 40°C and held for 3 min, increased to 200°C at a rate

of 35°C/min, then increased to 280°C at a rate of 3°C/min and held for 15 min. Chromatograms

and spectra were analyzed using MSD ChemStation (Agilent Technologies, Inc.). We identified

CHCs based on retention time and electron ionization fragmentation pattern. The abundance of

each compound was then quantified by normalizing the area under each CHC peak to the area of

the hexacosane signal using homebuilt peak selection software (personal correspondence, Dr.

Scott Pletcher, Univ. of Michigan).

2.6 Postmating reproductive trait morphology

We measured several elements of reproductive trait morphology in each of our lines, and

in the tester line, in order to assess their potential influence on patterns of female preferential

sperm use.

2.6.1 Thorax length, testis length, SR length, and ST area

We collected both female and male virgins and aged them to reproductive maturity (N=5

replicates per sex per genotype). We measured thorax length for all flies; this is a standard

measure of body size in *Drosophila* studies because it strongly correlates with the size of other

characters such as wing length (Robertson & Reeve 1952). We then dissected the reproductive

tract into Grace's Insect Medium. For females, we separated the seminal receptacle (SR) and a

randomly chosen spermatheca (ST) from the rest of the reproductive tract and transferred them to

a slide. For males, a randomly chosen testis was separated from the rest of the reproductive tract

and transferred to a slide. In both cases, we then applied a cover slip and took images under an

EVOS FL digital inverted microscope. Using ImageJ, we measured the length of the SR, the

height and width of the ST, and the length of the testis, from these images. Measurements were

replicated across five individuals per genotype.

2.6.2 Sperm length

We dissected sperm out of the seminal vesicles of reproductively mature virgin males into

Grace's insect medium on a slide (N=5 replicates per genotype). Sperm were gently teased apart

with a probe when necessary. The slides were stained with SpermBlue (Microptic), and imaged

under an EVOS FL digital inverted microscope. In ImageJ, we measure sperm length for 5-10

randomly chosen sperm per individual.

2.6.3 Body mass and reproductive tract mass

We collected both female and male virgins and aged them to reproductive maturity (N=5

replicates per sex, per genotype). We measured body and reproductive tract mass according to

Holman et al. (2008). Flies were dissected in distilled water onto a piece of preweighed foil. For

females, the reproductive tract (including the bursa, SR, ST, parovaria, and ovaries) was removed

and transferred to another piece of pre-weighed foil. For males, the reproductive tract (including

testes, seminal vesicles, accessory glands, and the ejaculatory bulb) was removed and transferred

to another piece of pre-weighed foil. For both males and females, the foils holding the body and

reproductive systems of the flies were then placed in a drying oven at 60°C for 18 ± 1 h and

weighed on a Mettler Toledo UMX2 balance to the nearest 0.1 μg.

2.7 Second-male courtship behavior assay

To identify variation in male courtship across genotypes, and to determine whether our

experimental manipulation affects male courtship behavior, we recorded male courtship for each

cross type on video. We recorded videos using a modified FlyPi setup, which combines a

Raspberry Pi (Raspberry Pi Foundation, Cambridge, UK), pi camera, and 3D printed parts (Maia

Chagas et al. 2017). Crosses were set up in a modified cell culture plate consisting of six culture

wells, each with a 3 cm diameter and a small amount of cornmeal media in the bottom, allowing

us to record six crosses simultaneously. Male and female flies were aspirated without

anesthetization into each cell culture well, and the plate was videotaped for two hours. The full

videos were scored for successful copulations, latency to copulation, and copulation duration.

The first 30 minutes of each video was also scored in more detail for the frequency of courtship

song, male tapping, number of male licking attempts, and copulation attempts. Recorded matings

were set up until we had five replicate recordings per cross type.

2.8 Statistical analyses

All statistical analysis and figure construction in this study was performed with R version

4.3.1. All linear models were run using deviation-coded contrasts. This does not change the fit of

the model, but affects the interpretation of regression coefficients such that the intercept

corresponds to the grand mean and main effect estimates represent deviations from the mean.

Model estimates are therefore robust to whichever line is used as the baseline in each

comparison, which is preferable in our experiments where none of our genotypes were *a priori*

designated as the baseline level.

2.8.1 CHC composition

We assessed variation in CHC composition using a principal components analysis (PCA),

which we computed using the prcomp function in R. To test for significant differences among

our principal components, we performed a multivariate analysis of variance (MANOVA) using

the manova function.

2.8.2 Postmating reproductive trait morphology

We assessed variation in female and male reproductive morphology by fitting linear

models using the lm function in R. To test whether male or female genotype predicts the

variation we observed in reproductive morphology, we ran linear models for each morphological

trait separately as the dependent variable. Male or female genotype was always included as the

predictor. The GFP-labelled *D. melanogaster* line was incorporated into each model as a

baseline.

2.8.3 Second-male courtship effort, female second-mating frequency, and second-male paternity

We assessed variation in second-male courtship by fitting a linear model using the lm

function in R. Second male courtship effort was the dependent variable, with female genotype,

male genotype, their interaction, and male perfuming treatment, as predictors.

We assessed variation in female second-mating frequency by fitting a beta-binomial

generalized linear model using the betabin function in R. We coded our response as a binomial

variable, with one column representing the number of successful second-matings, and the other

column representing the number of failed second-matings for each cross type (N=32 total cross

types). The model included female genotype, male genotype, and male perfuming treatment as

predictors (this dataset did not allow us sufficient power to assess interaction terms).

Variation in second male siring success was also assessed with a beta-binomial

generalized linear model. We coded our response as a binomial variable, with one column

representing the number of offspring sired by the second male ("successes"), and the other

column representing the number of offspring sired by the first male ("failures") for each cross

(N=196 total crosses). Our model included male genotype, female genotype, male perfuming

treatment, and all possible interactions (female genotype by male genotype, female genotype by

male perfuming treatment, male genotype by male perfuming treatment, and female genotype by

male genotype by male perfuming treatment) as predictors. California males, California females,

and conspecific-perfumed males were always incorporated into the models as baselines.

2.8.4 Correlations among datasets

Finally, we looked for correlations among our datasets. Across all pairwise genotype

combinations we assessed the strength of association between: second-male courtship effort and

second-male mating success; second male courtship effort and paternity share (P2); female

readiness to remate and second-male paternity share (P2); and, male and female reproductive

trait variation and second-male paternity share (P2). To do this, in each case, we computed

Spearman's rank correlation coefficients using the cor.test function in R. Each correlation was

assessed separately for experiments involving conspecific-perfumed males and heterospecific-

perfumed males.

3   Results

3.1 Males differ in CHC composition based on genotype and perfuming status

We quantified CHC composition across male genotypes and perfuming treatments to

characterize the variation present across lines, as well as to verify that our perfuming treatment

was effective. We found that CHC composition varied substantially across male genotypes

((Figure 2a,b, Table S1, Table S2). For each of the first three PCs—which together account for

92.3% of the variation observed—we detected a significant genotype effect (MANOVA $p <$

0.001; Table S1). CHC composition was also shifted by our perfuming treatment, as shown by

significant treatment effects for PC3 ($p = 0.004$) and PC4 ($p < 0.001$) (MANOVA; Figure 2b,

Table S1). These two PCs account for 8.6% of the variation in CHC composition.

3.2 Postmating reproductive traits differ more among males than among females

We quantified variation in male and female traits, focusing on postmating reproductive

traits, to assess whether these might influence patterns of sperm use. We found that all measured

male traits differed among genotypes, including two overall size traits and three reproductive

size traits (Figure 3, Table S3). Differences among female lines were more modest and only

observed for overall size and one reproductive size trait, RT mass (Figure 3, Table S3).

In terms of non-reproductive traits, both male and female Taiwan individuals consistently

had shorter thoraxes but also the largest dry non-RT body mass. Other differences among lines

were sex-specific and often more modest (for example, Greece males had longer thoraxes but

reduced dry non-RT body mass, and Malaysia females were smaller than average by both

measures) (Table S3).

For reproductive traits, California females exhibited the highest RT dry mass and Taiwan

females exhibited the lowest; however, female lines did not differ for either SR length or ST area

(Figure S2). In contrast, male lines differed in RT mass (Taiwan males had lower RT mass, while

Greece males had higher RT mass, than average), as well as testis length and sperm length

(Figure 3, Figure S2). California and Greece males had longer testes than average, while

Malaysia and Taiwan males had shorter testes. Interestingly, California and Taiwan males had

longer sperm than all other lines (1.814 mm and 1.763 mm on average, respectively; Table S3),

and Malaysia and Greece males had significantly shorter sperm (1.662 mm and 1.609 mm on

average, respectively; Table S3). GFP sperm length was intermediate (1.713 mm on average;

Table S3). This indicates that sperm length and testis length are not closely associated among our

D. melanogaster lines: Taiwan males had short testes but also had among the longest sperm,

while the inverse pattern was observed for Greece males.

3.3 Courtship effort varies consistently among male genotypes, but not among female genotypes

being courted or with perfuming

We evaluated courtship across each male-female genotype combination to better

understand if and/or how premating processes influence cryptic female choices, including via our

perfuming treatments. We measured courtship effort as the number of courtship behaviors

exhibited per minute until copulation occurred or until our observation period ended (at two

hours). Note that only 9 of 162 scored pairings resulted in copulation within two hours.

Courtship intensity varied significantly among male genotypes, with Greece and Taiwan males

courting significantly less than the others (Figure 4, Table S4). Male courtship intensity was also

marginally reduced in heterospecific-perfumed males, but this effect was not statistically

significant (Figure 4, Table S4). In addition, although female line identity had only a marginal

influence on courtship, we did find that Taiwan females were consistently courted less than

average across male genotypes (Figure 4, Table S4), possibly as a result of their smaller size

(Turiegano et al. 2012, Table S3). We did not detect any significant female-by-male genotype

interactions.

3.4 Readiness to remate differs among female genotypes and depends on male genotype, but not

on male perfuming status

For each cross type, we assessed whether females chose to mate with second males

versus refused the second-male mating. Remating was evidenced by whether a female produced

offspring from the second male. We found that female genotypes varied in their readiness to

remate following a first mating with the tester male: California females remated more readily

while Greece and Taiwan females mated less readily than average (Figure 5, Table S5). Male

genotype also affected remating: California males were more consistently successful in these

second matings, while Greece males were less successful on average (Figure 5, Table S5). In

contrast, we detected no effect of male perfuming status on second-male premating success

(Figure 5, Table S5).

3.5 Female genotypes vary in their overall use of sperm from the second male, and in whether

they adjust sperm use according to second-male perfuming status

We assessed how paternity of second-males (P2) was affected by genotype and by our

perfuming manipulation. We found strong and significant differences in second-male paternity

depending on both female and second-male genotype, female by male genotype interaction

effects, and female genotype by male genotype by treatment interaction effects. In particular,

Taiwan males gained higher P2 than average, while Greece and Malaysia males had reduced

paternity share (Figure 6, Table 1). In comparison, Greece females exhibited a higher P2 than

average, while Taiwan females exhibited a lower P2 than average, across all genotype pairs

(Figure 6, Table 1). Despite these general effects, second-male paternity share also varied

substantially depending on the particular male-female combination. For instance, Greece females

exhibit higher average P2 when paired with Taiwan second males, while Taiwan females reduce

P2 when paired with Malaysia second males (Figure 6, Table 1).

We also found that females adjust sperm use according to male genotype and male

perfuming treatment, including in female genotype-specific ways. In particular, female

genotypes vary in whether and how sperm use patterns were affected by perfuming. For instance,

Greece females use less sperm from Greece males specifically when those males are

heterospecific-perfumed (Figure 6, Table 1). In contrast, Taiwan females use more sperm from

heterospecific-perfumed Greece males (Figure 6, Table 1). Greece females also use more sperm

from Malaysia males when they are perfumed by heterospecifics (Figure 6, Table 1). In

comparison, California and Malaysia females do not appear to respond to male CHC perfuming

when making sperm use choices.

3.6 Both pre- and postmating choices can be affected by premating behavior, but sperm length

shows the most consistent association with patterns of preferential sperm use

We next assessed correlations between datasets to better understand how these variables

might influence one another. First, we evaluated the relationship between second-male courtship

effort and second-male mating success—i.e., is premating courtship intensity associated with

premating success? We did not see a consistent association for either conspecific-perfumed males

(Figure S4a; Spearman's rank correlation $\rho = 0.308$, $p = 0.246$) or heterospecific-perfumed males

(Figure S4b; Spearman's rank correlation $\rho = 0.174$, $p = 0.519$). Instead, readiness to remate

seems to be more clearly associated with individual female line differences. For instance,

California females tend to remate most readily, and Greece and Taiwan least readily, regardless

of the courtship intensity of the male line they are paired with (Figure S4a,b).

Second, we assessed the relationship between second male courtship effort and paternity

share (P2)—i.e., do females use premating signals to make postmating decisions? We found that

when males are perfumed by conspecifics, male courtship effort is significantly correlated with

siring success (Figure 7a; Spearman's rank correlation $\rho = 0.535$, $p = 0.035$). When males are

perfumed by heterospecifics, the correlation is positive but not significant (Figure 7b;

Spearman's rank correlation $\rho = 0.353$, $p = 0.180$). This weak positive trend between courtship

effort and P2 appears to be largely driven by California males, which are the most active courters

and also received the greatest share of paternity (Figure 7a,b), an association which is unclear in

the other male lines. Thus, we infer that the relationship between these variables is likely

influenced by additional factors not measured here; for instance, Greece males, which have low

siring success, have been shown to exhibit especially long interpulse intervals during courtship

song, which are less preferred by *D. melanogaster* females (Kyriacou & Hall 1982, Pischedda et

al. 2014a).

We next assessed the relationship between female readiness to remate and second-male

paternity share—i.e., is premating choosiness associated with postmating choosiness? We found

no correlation among these factors for conspecific-perfumed males (Figure 7c; Spearman's rank

correlation $\rho = -0.095$, $p = 0.726$), and a slight association for experiments with heterospecific-

perfumed males (Figure 7d; Spearman's rank correlation $\rho = 0.502$, $p = 0.048$). These weak or

absent associations are consistent with heterogenous effects we observe among female lines. For

instance, while Greece females showed relatively high premating choosiness (remating in only

59% of trials) and the highest average P2 of any genotype (consistent with strong second male

preference), Taiwan females exhibit a similar premating choosiness (60% remating frequency)

but significantly lower average P2 (indicating no consistent postmating preference for first or

second male sperm) (Figure 7c,d). This relationship therefore appears to vary substantially

depending upon which of the four genotypes is considered.

Finally, we examined the relationship between second-male paternity share and

reproductive trait variation among lines. Of RT mass (both male and female), SR length, testis

length, sperm length, and SR length minus sperm length, only sperm length and MRT mass were

significantly associated with P2. Of these two, MRT mass was significantly negatively correlated

with P2 among heterospecific-perfumed males (Figure S4d; Spearman's rank correlation $\rho = -$

0.679, $p = 0.004$), but not among conspecific-perfumed males (Figure S4c; Spearman's rank

correlation $\rho = -0.449$, $p = 0.081$). These relationships appear to be substantially influenced by

Taiwan males, which have high P2 but comparatively low RT mass. In comparison, the

relationship between sperm length and P2 was consistently and strongly positive for both

conspecific-perfumed (Figure 7e; Spearman's rank correlation $\rho = 0.631$, $p = 0.009$) and

heterospecific-perfumed groups (Figure 7f; Spearman's rank correlation $\rho = 0.728$, $p = 0.001$).

Second males with longer sperm experience greater siring success. Specifically, when the second

male's sperm is longer than the tester male (as for California or Taiwan second males), P2 is

generally higher; whereas when the second male sperm is shorter than the tester male (as for

Malaysia and Greece), P2 is generally lower. Together, these data indicate that females

preferentially use longer sperm—a generic preference that appears as last male precedence when

the second male's sperm is longer than the first (tester) male's sperm, but can override last male

precedence when the second male's sperm is relatively shorter.

4   Discussion

In this study, we sought to understand whether and how females make plastic choices

about sperm use. We found that sperm use patterns vary based on male genotype, female

genotype, their interaction, as well as based on male pheromone profile (even when male identity

is the same). Among these effects, male sperm length, specifically the relative difference in

sperm length between the first and second male, appears to play a particularly important role in

determining female sperm use patterns. While sperm length has been recognized as an important

component of male-male competition (e.g., Immler et al. 2011, Lüpold & Pitnick 2018, Snook

2005), our analysis enables us to infer that the preference for longer sperm is also female-

mediated, context-dependent, and controlled by factors other than the size of the female sperm

storage organs, as we expand upon below.

4.1 Female sperm use choices vary among lines in their direction and magnitude, and in the

criteria used to evaluate males

Consistent with previous work in *D. melanogaster*, we observe clear evidence for sperm

choice mediated by females (Chow et al. 2010, Clark & Begun 1998, Clark et al. 1999, Miller &

Pitnick 2002). Female lines show patterns of sperm use in control crosses (with their own

unmanipulated second males) that significantly shift in response to changing either second-male

genotype or perfuming treatment or both (depending on the female line). In addition, both first

and second males have the potential to shape some paternity outcomes via plastic allocation

during the transfer of sperm, including adjusting ejaculate composition based on the identity of

the female. In some cases, it is difficult to differentiate between these male or female plastic

effects; for instance, our male by female genotype interaction effects on paternity share could be

shaped by both cryptic male and cryptic female choice. However, our design specifically aimed

to include effects where the influence of male plasticity should be minimized. In particular, to

attribute perfuming effects on paternity to cryptic male choice, males would have to plastically

adjust their ejaculate based on one minute of exposure to conspecific vs. heterospecific males,

but only when mating with certain female genotypes. A more parsimonious and biologically

plausible inference is that females are adjusting paternity—exercising choice—in response to

male pheromone differences. Thus, in addition to the effects of male reproductive plasticity, our

results underscore the potential for cryptic female choice to influence genotypic representation

among the resulting offspring, and to generate sexual selection on males (Pitnick et al. 2001).

We found substantial and consistent variation among our four female lines in the

direction and magnitude of preferential sperm use patterns. *D. melanogaster* is conventionally

described as having second (or last) male sperm precedence (Lefevre & Jonsson 1962, Pischedda

& Rice 2012). However, depending on the specific male-female combination, our estimated

preference for the sperm of second-males ranged from 0.2 to 0.95. Values below 0.5 (e.g.,

Taiwan females with Malaysia males) indicate consistent first-male precedence in these specific

male-female combinations. Moreover, female lines differ in how strongly they differentiate

among sperm donors. For example, with the exception of California males (which they strongly

prefer; average P2 = 0.79), California females do not consistently differentiate between the first

and second males' sperm when paired with the other three second male lines in our experiment

(i.e., mean P2 is approximately 0.5, with a broad range of values among individual CA females;

Figure 6, Table S6).

We also clearly observed among-line variation in the characteristics that females use to

evaluate males as reproductive donors. In particular, Greece and Taiwan females modulate their

use of second male sperm in response to perfuming status, albeit in complex ways. For example,

Greece females decrease their use of Greece sperm when these males are heterospecific-

perfumed, but Taiwan females increase their use of Greece sperm after the same treatment

(Figure 6, Table S7). In comparison, male perfuming appears to have no influence on cryptic

choice by California or Malaysia females. This diversity in sperm choice patterns depending on

intrinsic (female genotype) and extrinsic (perfumed status) factors points to a much greater

heterogeneity of sperm use decision-making among *D. melanogaster* females than simple last

male precedence (see also Laturney et al. 2018). Given the broad geographic origin of these

lines, differences in the magnitude and direction of female choices could reflect reproductive or

other differences among lines that operated prior to their collection and lab establishment.

4.2 Cryptic female choice is strongly shaped by male traits, especially sperm length

Within the diversity of individual sperm use criteria, however, we did observe several

broad patterns. In particular, every female line differentiated among male genotypes in at least

one postmating experimental context, indicating that one or more aspects of male genotype are

important in making paternity allocation decisions. Our analysis also identifies specific male

traits that likely mediate these male genotype effects. As noted above, some female lines adjust

sperm use in response to perfuming treatment, indicating that male cuticular hydrocarbon profile

affects their postmating choices. Among conspecific-perfumed second males, paternity share is

also associated with courtship effort, suggesting that females may prefer the sperm of male

genotypes who court more, although this effect is shaped largely by California males. Most

strikingly, however—and more consistently than either chemosensory or courtship traits—we

find that sperm length is positively associated with preferential sperm use across many contexts

and conditions. The majority of female lines preferentially used the second-male's sperm when

that sperm was longer than the first-male's sperm (i.e., when California or Taiwan was the

second male). This result is consistent with previous studies demonstrating the increased

competitive ability of long sperm (e.g., Miller & Pitnick 2002). However, if this process were

solely mediated by sperm length, we would expect paternity outcomes to be consistent across

female genotypes. Instead, we see significant variation in male siring success among female lines

and perfuming treatments; for instance, Taiwan males show increased siring success when

mating with Greece females compared to California females (Figure 6, Table 1), indicating that

females also play a role in shaping these outcomes. Moreover, in several cases (for example,

when Taiwan females mate with Malaysia males—see Figure 6) female lines subverted the

expected precedence for last males' sperm specifically when the first (GFP tester) male had

longer sperm than the second male (i.e. Greece and Malaysia males). That is, our results indicate

that 'long-sperm precedence', in addition to 'last-male sperm precedence' (Lefevre & Jonsson

1962), jointly contribute to female sperm use patterns in *D. melanogaster*.

Finally, although our data indicate diverse criteria for female sperm choice overlaid on a

generic preference for longer sperm, it also indicates that these patterns are not explained by at

least one studied mechanism of female sperm choice—SR length (Miller & Pitnick 2002). SR

length has been proposed as a primary mechanism via which females discriminate between

sperm of different lengths (Pitnick et al. 1999); in particular, females with longer SRs are thought

to be most selective because they can better discriminate between the sperm length of the first

and second male. However, while we observe clear variation in female discrimination of sperm

based on length—e.g., Greece females exhibit generally high P2, regardless of length, whereas

California females only exhibit high P2 for their own (long-spermed) males (Figure 6, Table 1)—

we do not observe significant variation in SR length among our female lines (Figure 3, Table S3,

Table S2). Thus, SR length is not the mechanism by which these diverse *D. melanogaster* female

lines discriminate against short sperm in our experiments.

4.3 Evolutionary implications of the observed variation in female sperm choice

Consistent patterns in choice criteria among female genotypes provide an opportunity to

evaluate the nature of past drivers of female choice in our system, including hypotheses where

cryptic female choice is not beneficial to females versus where it has specific fitness benefits.

For example, one proposal is that female sperm use patterns result primarily from manipulation

by males; that is, cryptic female choice is the product of sexual conflict rather than providing an

adaptive benefit to females. In this case, female sperm use patterns are expected to be random

with respect to actual measures of male quality. Our data do not convincingly support this

hypothesis, given the general postmating female preferences we observe for males who have the

longest sperm and, under some conditions, for males that court frequently.

Alternatively, Birkhead (1998) proposed that female sperm choice might function to bias

sperm use against the sperm of closely related males. However, our findings are not consistent

with inbreeding avoidance as a primary function of cryptic female choice. If that were the case,

second-male success (against the unrelated tester first male) should consistently be lower in

crosses with males from the same line as females, compared to males from different lines.

However, female lines differed in their choice of own male sperm, ranging from consistent high

preference to low preference.

Apart from inbreeding avoidance, other hypothesized functions of sperm choice include

female benefits from sire discrimination (e.g., choice of high-quality males ['good genes'

hypothesis; Eberhard 1996], or choice of attractive males ['sexy sons' hypothesis; Weatherhead

& Robertson 1979]). We find that females have a generic preference for longer sperm (Figure

7e,f); in this case, choosing longer-spermed males increases the frequency of longer-spermed

(i.e. 'sexier') sons, which themselves could benefit from the same generic preference. As studies

have shown that long sperm have a competitive advantage in displacing shorter sperm (Lüpold et

al. 2012, Lüpold et al. 2016), this preference likely also aligns specifically with higher

reproductive fitness in males. Thus, our data support the hypothesis that females could gain

indirect fitness benefits from exercising cryptic choice based on specific male traits like sperm

length.

The differences we observe among female lines in the criteria they used for sperm choice

also have implications for the expected future drivers of postmating reproductive evolution.

Some of our lines altered sperm use patterns based on contact pheromones (Greece and Taiwan),

indicating that pheromone bouquet can be a criterion for evaluating preferred sires. Other lines

(California and Malaysia) were insensitive to pheromone manipulation in their sperm use

choices. Variation in choice criteria makes it more challenging for males to tailor specific traits to

satisfy, or manipulate (Ryan 1990), a broad range of female biases. Moreover, when the specific

preferences of females differ according to male trait values and male trait identity, there is no

single most preferred male genotype, and variation should be maintained at male traits. This

expectation is consistent with our observation, and with previous analyses (e.g., Clark et al.

1999, Chow et al. 2010, Miller & Pitnick 2002), showing that the outcomes of cryptic female

choice are demonstrably influenced by male by female interactions.

That these criteria for female sperm choice can also vary behaviorally is particularly

evident in the case of Greece females. When sequentially mated with males from other lines,

Greece females uniformly favor whichever genotype (tester versus second male) has the longer

sperm. However, when mated with their own conspecific-perfumed second males, Greece

females preferentially use this sperm (Figure 6), even though it is significantly shorter than the

first (tester) male (Figure 3). In this case, the female preference for local males appears to

override their generic preference for longer sperm. We know that this decision is both female-

mediated and based on chemosensory assessment, because it is reversed when Greece females

are paired with heterospecific-perfumed Greece males. Greece females instead favor the longer

(tester male) sperm in these pairings (Figure 6), even though these Greece males are identical in

every sense to conspecific-perfumed males—except for their CHC profile. Flexible changes in

preference, such as this case where females specifically prefer own males despite generic

preferences for longer sperm, is another potential mechanism by which sexually selected

variation could be maintained in males.

5   Conclusion

The direction and magnitude of female-mediated sperm choice, as well as the male traits

influencing these choices, varies significantly among four female lines of *D. melanogaster*. This

diversity in female sperm choices is overlaid on a generic preference for longer sperm,

suggesting that paternity decisions may ultimately benefit female fitness indirectly through the

production of higher-quality offspring or more attractive offspring. Interestingly, at least one

proposed proximate mechanism of female sperm choice—SR length—does not account for

variation in observed sperm use. Diversity in cryptic female choices—both in the specific

modalities used to assess and choose among males, and in the plasticity of these choices—may

explain how variation in male postmating sexually selected traits is maintained over time.

References

Anastasio, O. E., Sinclair, C. S., & Pischedda, A. (2023). Cryptic male mate choice for high-quality females reduces male postcopulatory success in future matings. *Evolution*, 77(6), 1396-1407. doi:10.1093/evolut/qpad064

Barot, S., Mitja, D., Miranda, I., Meija, G. D., & Grimaldi, M. (2005). Reproductive plasticity in an Amazonian palm. *Evolutionary Ecology Research*, *7*(7), 1051-1065.

Billeter, J. C., Atallah, J., Krupp, J. J., Millar, J. G., & Levine, J. D. (2009). Specialized cells tag sexual and species identity in Drosophila melanogaster. *Nature*, 461(7266), 987-991. doi:10.1038/nature08495

Birkhead, T. R. (1998). Cryptic Female Choice: Criteria for Establishing Female Sperm Choice. *Evolution*, 52(4), 1212-1218. doi:10.1111/j.1558-5646.1998.tb01848.x

Blows, M. W., & Allan, R. A. (1998). Levels of mate recognition within and between two Drosophila species and their hybrids. *Am Nat*, 152(6), 826-837. doi:10.1086/286211

Chow, C. Y., Wolfner, M. F., & Clark, A. G. (2010). The genetic basis for male x female interactions underlying variation in reproductive phenotypes of Drosophila. *Genetics,* 186(4), 1355-1365. doi:10.1534/genetics.110.123174

Clark, A. G., & Begun, D. J. (1998). Female genotypes affect sperm displacement in Drosophila. *Genetics*, 149(3), 1487-1493. doi:10.1093/genetics/149.3.1487

Clark, A. G., Begun, D. J., & Prout, T. (1999). Female x male interactions in Drosophila sperm competition. *Science*, 283(5399), 217-220. doi:10.1126/science.283.5399.217

Douglas, T., Anderson, R., & Saltz, J. B. (2020). Limits to male reproductive potential across mating bouts in Drosophila melanogaster. *Animal Behaviour*, 160, 25-33. doi:10.1016/j.anbehav.2019.11.009

Dyer, K. A., White, B. E., Sztepanacz, J. L., Bewick, E. R., & Rundle, H. D. (2014). Reproductive character displacement of epicuticular compounds and their contribution to mate choice in Drosophila subquinaria and Drosophila recens. *Evolution*, 68(4), 1163-1175. doi:10.1111/evo.12335

Eberhard, W. (1996). Female control: sexual selection by cryptic female choice (Vol. 69): Princeton University Press.

Firman, R. C., Gasparini, C., Manier, M. K., & Pizzari, T. (2017). Postmating female control: 20 years of cryptic female choice. *Trends in Ecology & Evolution*, *32*(5), 368-382. doi:10.1016/j.tree.2017.02.010

Garbaczewska, M., Billeter, J. C., & Levine, J. D. (2013). Drosophila melanogaster males increase the number of sperm in their ejaculate when perceiving rival males. *J Insect Physiol,* 59(3), 306-310. doi:10.1016/j.jinsphys.2012.08.016

Gasparini, C., & Evans, J. P. (2018). Female control over multiple matings increases the opportunity for postcopulatory sexual selection. *Proceedings of the Royal Society B*, *285*(1888), 20181505. doi:10.1098/rspb.2018.1505

Greenspan, R. J., & Ferveur, J. F. (2000). Courtship in Drosophila. *Annu Rev Genet*, 34, 205-232. doi:10.1146/annurev.genet.34.1.205. doi:10.1146/annurev.genet.34.1.205

Holman, L., Freckleton, R. P., & Snook, R. R. (2008). What use is an infertile sperm? A comparative study of sperm-heteromorphic Drosophila. *Evolution*, *62*(2), 374-385. doi: 10.1111/j.1558-5646.2007.00280.x

Hopkins, B. R., Sepil, I., Thézénas, M. L., Craig, J. F., Miller, T., Charles, P. D., ... & Wigby, S. (2019). Divergent allocation of sperm and the seminal proteome along a competition gradient in Drosophila melanogaster. *Proceedings of the National Academy of Sciences*, *116*(36), 17925-17933. doi:10.1073/pnas.1906149116

Hosken, D. J., Taylor, M. L., Hoyle, K., Higgins, S., & Wedell, N. (2008). Attractive males have greater success in sperm competition. *Curr Biol*, 18(13), R553-554. doi:10.1016/j.cub.2008.04.028

Immler, S., Pitnick, S., Parker, G. A., Durrant, K. L., Lupold, S., Calhim, S., & Birkhead, T. R. (2011). Resolving variation in the reproductive tradeoff between sperm size and number. *Proc Natl Acad Sci U S A*, 108(13), 5325-5330. doi:10.1073/pnas.1009059108

Khallaf, M. A., Cui, R., Weißflog, J., Erdogmus, M., Svatoš, A., Dweck, H. K., ... & Knaden, M. (2021). Large-scale characterization of sex pheromone communication systems in Drosophila. *Nature communications*, *12*(1), 4165. doi:10.1038/s41467-021-24395-z

King, E. G., Merkes, C. M., McNeil, C. L., Hoofer, S. R., Sen, S., Broman, K. W., . . . Macdonald, S. J. (2012). Genetic dissection of a model complex trait using the Drosophila Synthetic Population Resource. *Genome Res*, 22(8), 1558-1566. doi:10.1101/gr.134031.111

Kyriacou, C. P., & Hall, J. C. (1982). The function of courtship song rhythms in Drosophila. *Animal Behaviour*, *30*(3), 794-801. doi:10.1016/S0003-3472(82)80152-8

Laturney, M., van Eijk, R., & Billeter, J. C. (2018). Last male sperm precedence is modulated by female remating rate in Drosophila melanogaster. *Evolution letters*, *2*(3), 180-189. doi:10.1002/evl3.50

768 Lefevre, G., Jr., & Jonsson, U. B. (1962). Sperm transfer, storage, displacement, and utilization
769   in Drosophila melanogaster. *Genetics*, 47(12), 1719-1736.
770   doi:10.1093/genetics/47.12.1719
771
772 Linklater, J. R., Wertheim, B., Wigby, S., & Chapman, T. (2007). Ejaculate depletion patterns
773   evolve in response to experimental manipulation of sex ratio in Drosophila
774   melanogaster. *Evolution*, *61*(8), 2027-2034. doi:10.1111/j.1558-5646.2007.00157.x
775
776 Lüpold, S., Manier, M. K., Ala-Honkola, O., Belote, J. M., & Pitnick, S. (2011). Male Drosophila
777   melanogaster adjust ejaculate size based on female mating status, fecundity, and
778   age. *Behavioral Ecology*, *22*(1), 184-191. doi:10.1093/beheco/arq193
779
780 Lüpold, S., Manier, M. K., Berben, K. S., Smith, K. J., Daley, B. D., Buckley, S. H., . . . Pitnick,
781   S. (2012). How multivariate ejaculate traits determine competitive fertilization success in
782   Drosophila melanogaster. *Curr Biol*, 22(18), 1667-1672. doi:10.1016/j.cub.2012.06.059
783
784 Lüpold, S., Pitnick, S., Berben, K. S., Blengini, C. S., Belote, J. M., & Manier, M. K. (2013).
785   Female mediation of competitive fertilization success in Drosophila
786   melanogaster. *Proceedings of the National Academy of Sciences*, *110*(26), 10693-10698.
787   doi:10.1073/pnas.1300954110
788
789 Lüpold, S., Manier, M. K., Puniamoorthy, N., Schoff, C., Starmer, W. T., Luepold, S. H. B., ... &
790   Pitnick, S. (2016). How sexual selection can drive the evolution of costly sperm
791   ornamentation. *Nature*, *533*(7604), 535-538. doi:10.1038/nature18005
792
793 Lüpold, S., & Pitnick, S. (2018). Sperm form and function: what do we know about the role of
794   sexual selection? *Reproduction*, 155(5), R229-R243. doi:10.1530/REP-17-0536
795
796 Maia Chagas, A., Prieto-Godino, L. L., Arrenberg, A. B., & Baden, T. (2017). The€ 100 lab: A
797   3D-printable open-source platform for fluorescence microscopy, optogenetics, and
798   accurate temperature control during behaviour of zebrafish, Drosophila, and
799   Caenorhabditis elegans. *PLoS biology*, *15*(7), e2002702.
800   doi:10.1371/journal.pbio.2002702
801
802 Manier, M. K., Belote, J. M., Berben, K. S., Novikov, D., Stuart, W. T., & Pitnick, S. (2010).
803   Resolving mechanisms of competitive fertilization success in Drosophila
804   melanogaster. *Science*, *328*(5976), 354-357. doi:10.1126/science.1187096
805
806 Manier, M. K., Lupold, S., Pitnick, S., & Starmer, W. T. (2013). An analytical framework for
807   estimating fertilization bias and the fertilization set from multiple sperm-storage organs.
808   *Am Nat*, 182(4), 552-561. doi:10.1086/671782
809
810 Manier, M. K., Lupold, S., Belote, J. M., Starmer, W. T., Berben, K. S., Ala-Honkola, O., . . .
811   Pitnick, S. (2013). Postcopulatory sexual selection generates speciation phenotypes in
812   Drosophila. *Curr Biol*, 23(19), 1853-1862. doi:10.1016/j.cub.2013.07.086
813

814 Miller, G. T., & Pitnick, S. (2002). Sperm-female coevolution in Drosophila. *Science*, 298(5596),
815   1230-1233. doi:10.1126/science.1076968
816

817 Nandy, B., Dasgupta, P., Halder, S., & Verma, T. (2016). Plasticity in aggression and the
818   correlated changes in the cost of reproduction in male Drosophila melanogaster. *Animal*
819   *Behaviour*, *114*, 3-9. doi:10.1016/j.anbehav.2016.01.019
820

821 Pilastro, A., Simonato, M., Bisazza, A., & Evans, J. P. (2004). Cryptic female preference for
822   colorful males in guppies. *Evolution*, *58*(3), 665-669. doi: 10.1111/j.0014-
823   3820.2004.tb01690.x
824

825 Pischedda, A., & Rice, W. R. (2012). Partitioning sexual selection into its mating success and
826   fertilization success components. *Proc Natl Acad Sci U S A*, 109(6), 2049-2053.
827   doi:10.1073/pnas.1110841109
828

829 Pischedda, A., Cochrane, V. A., Cochrane, W. G., & Turner, T. (2014). Genetic dissection of
830   courtship song variation using the Drosophila Synthetic Population Resource. *bioRxiv*,
831   006643. doi:10.1101/006643
832

833 Pischedda, A., Shahandeh, M. P., Cochrane, W. G., Cochrane, V. A., & Turner, T. L. (2014).
834   Natural variation in the strength and direction of male mating preferences for female
835   pheromones in Drosophila melanogaster. *PLoS One*, 9(1), e87509.
836   doi:10.1371/journal.pone.0087509
837

838 Pitnick, S., Marrow, T., & Spicer, G. S. (1999). Evolution of Multiple Kinds of Female Sperm-
839   Storage Organs in Drosophila. *Evolution*, 53(6), 1804-1822. doi:10.1111/j.1558-
840   5646.1999.tb04564.x
841

842 Pitnick, S., Brown, W. D., & Miller, G. T. (2001). Evolution of female remating behaviour
843   following experimental removal of sexual selection. *Proc Biol Sci*, 268(1467), 557-563.
844   doi:10.1098/rspb.2000.1400
845

846 Robertson, F. W., & Reeve, E. C. (1952). Heterozygosity, environmental variation and heterosis.
847   *Nature*, 170(4320), 286. doi:10.1038/170286a0
848

849 Ryan, M. J. (1990). Sexual selection, sensory systems and sensory exploitation. *Oxford surveys*
850   *in evolutionary biology*, *7*, 157-195.
851

852 Scott, D. (1994). Genetic Variation for Female Mate Discrimination in Drosophila Melanogaster.
853   *Evolution*, 48(1), 112-121. doi:10.1111/j.1558-5646.1994.tb01298.x
854

855 Sirot, L. K., Wolfner, M. F., & Wigby, S. (2011). Protein-specific manipulation of ejaculate
856   composition in response to female mating status in Drosophila melanogaster. *Proc Natl*
857   *Acad Sci U S A*, 108(24), 9922-9926. doi:10.1073/pnas.1100905108
858

859 Sirot, L. K. (2019). Modulation of seminal fluid molecules by males and females. *Curr Opin*
860   *Insect Sci*, 35, 109-116. doi:10.1016/j.cois.2019.07.009
861
862 Snook, R. R., & Hosken, D. J. (2004). Sperm death and dumping in Drosophila. *Nature*,
863   428(6986), 939-941. doi:10.1038/nature02455
864
865 Snook, R. R. (2005). Sperm in competition: not playing by the numbers. *Trends Ecol Evol*, 20(1),
866   46-53. doi:10.1016/j.tree.2004.10.011
867
868 Thornhill, R. (1983). Cryptic female choice and its implications in the scorpionfly Harpobittacus
869   nigriceps. *The American Naturalist*, *122*(6), 765-788. doi:10.1086/284170
870
871 Turiegano, E., Monedero, I., Pita, M., Torroja, L., & Canal, I. (2013). Effect of Drosophila
872   melanogaster female size on male mating success. *Journal of insect behavior*, *26*, 89-100.
873   doi:10.1007/s10905-012-9342-8
874
875 Wang, L., Han, X., Mehren, J., Hiroi, M., Billeter, J.-C., Miyamoto, T., . . . Anderson, D. J.
876   (2011). Hierarchical chemosensory regulation of male-male social interactions in
877   Drosophila. *Nature neuroscience*, 14(6), 757. doi:10.1038/nn.2800
878
879 Weatherhead, P. J., & Robertson, R. J. (1979). Offspring quality and the polygyny threshold:" the
880   sexy son hypothesis". *The American Naturalist*, *113*(2), 201-208. doi:10.1086/283379
881
882 Wigby, S., Sirot, L. K., Linklater, J. R., Buehner, N., Calboli, F. C., Bretman, A., . . . Chapman,
883   T. (2009). Seminal fluid protein allocation and male reproductive success. *Curr Biol*,
884   19(9), 751-757. doi:10.1016/j.cub.2009.03.036
885
886 Yamamoto, D., & Nakano, Y. (1999). Sexual behavior mutants revisited: molecular and cellular
887   basis of Drosophila mating. *Cell Mol Life Sci*, 56(7-8), 634-646.
888   doi:10.1007/s000180050458
889
890 Zuk, M., Johnson, K., Thornhill, R., & Ligon, J. D. (1990). Mechanisms of Female Choice in
891   Red Jungle Fowl. *Evolution*, 44(3), 477-485. doi:10.1111/j.1558-5646.1990.tb05933.x
892

Table 1. Specific genotype, treatment, and interaction effects on second male paternity share (P2). Shown is the beta-binomial generalized linear model of variation in second male paternity share (P2), depending on female line, second male line, treatment group, and all possible interactions. California females, California males, and conspecific-perfumed males are incorporated into the model as a baseline.

Alt text: A table containing the output of our beta-binomial generalized linear model. Our model includes significant terms for at least some components of female genotype, second male genotype, female genotype by second male genotype, and female genotype by second male genotype by perfuming treatment.

Model component	Estimate	Std. error	z	p
Intercept	0.734	0.082	8.903	<0.001 ***
Female line (Greece)	0.951	0.163	5.844	<0.001 ***
Female line (Malaysia)	-0.035	0.127	-0.279	0.780
Female line (Taiwan)	-0.383	0.134	-2.853	0.004 **
Second male line (Greece)	-0.661	0.137	-4.820	<0.001 ***
Second male line (Malaysia)	-0.870	0.133	-6.633	<0.001 ***
Second male line (Taiwan)	0.467	0.143	3.271	0.001 ***
Treatment (het-perfumed)	0.032	0.080	0.399	0.690
Female line (Greece) x second male line (Greece)	-0.241	0.270	-0.893	0.372
Female line (Malaysia) x second male line (Greece)	-0.042	0.214	-0.196	0.845
Female line (Taiwan) x second male line (Greece)	0.128	0.233	0.550	0.582
Female line (Greece) x second male line (Malaysia)	-0.090	0.237	-0.379	0.705
Female line (Malaysia) x second male line (Malaysia)	0.038	0.212	0.180	0.857
Female line (Taiwan) x second male line (Malaysia)	-0.452	0.221	-2.040	0.041 *
Female line (Greece) x second male line (Taiwan)	0.782	0.308	2.537	0.011 *
Female line (Malaysia) x second male line (Taiwan)	-0.003	0.217	-0.015	0.988
Female line (Taiwan) x second male line (Taiwan)	-0.009	0.224	-0.038	0.969
Female line (Greece) x treatment (het-perfumed)	-0.126	0.162	-0.777	0.437
Female line (Malaysia) x treatment (het-perfumed)	0.113	0.127	0.893	0.372
Female line (Taiwan) x treatment (het-perfumed)	-0.011	0.134	-0.085	0.932
Second male line (Greece) x	-0.090	0.136	-0.661	0.508

treatment (het-perfumed)				
Second male line (Malaysia) x treatment (het-perfumed)	0.155	0.129	1.197	0.231
Second male line (Taiwan) x treatment (het-perfumed)	0.120	0.142	0.843	0.400
Female line (Greece) x second male line (Greece) x treatment (het-perfumed)	-0.737	0.270	-2.732	0.006 **
Female line (Malaysia) x second male line (Greece) x treatment (het-perfumed)	-0.007	0.214	-0.031	0.976
Female line (Taiwan) x second male line (Greece) x treatment (het-perfumed)	0.685	0.234	2.933	0.003 **
Female line (Greece) x second male line (Malaysia) x treatment (het-perfumed)	0.519	0.237	2.189	0.029 *
Female line (Malaysia) x second male line (Malaysia) x treatment (het-perfumed)	-0.302	0.021	-1.429	0.153
Female line (Taiwan) x second male line (Malaysia) x treatment (het-perfumed)	-0.231	0.221	-1.045	0.296
Female line (Greece) x second male line (Taiwan) x treatment (het-perfumed)	0.309	0.308	1.005	0.315
Female line (Malaysia) x second male line (Taiwan) x treatment (het-perfumed)	0.115	0.217	0.531	0.596
Female line (Taiwan) x second male line (Taiwan) x treatment (het-perfumed)	-0.396	0.224	-1.770	0.077

Figure 1. Experimental procedure for assessing paternity share of second (focal) males.
Females were first mated with a GFP tester male. After 48 hours, these females were paired for second matings with focal males, which were either perfumed by conspecifics (left, control) or by heterospecifics (right, treatment). Unmated females were removed from the experiment. In remated females, we assessed paternity for first and second males based on the presence/absence of the GFP marker in resulting offspring.

Alt text: A diagram of our experimental design. Females are mated to two males before their offspring are assessed, such that we can compare patterns of paternity between male genotypes and perfuming treatments.

Figure 2. CHC composition varied based on genotype (G, Greece; T, Taiwan; C, California; and M, Malaysia) and perfuming status (conspecific-perfumed males shown in grey, heterospecific-perfumed males shown in purple) (Table S1). Both panels show standardized principal component (PC) values generated from multivariate CHC composition data for replicate males of each genotype and treatment. Left panel: PCs 1 and 2 largely reflect differences between male genotypes. Right panel: PCs 3 and 4 reflect additional shifts in CHC composition associated with heterospecific perfuming treatment.

Alt text: A principal component analysis of CHC composition. On the left, we see CHC composition clustering according to male genotype. On the right, CHC composition separates out according to perfuming treatment. Points are labeled using the first letter of their genotype.

Figure 3. Variation in (a) thorax length in mm and (b) reproductive trait length in mm in females and males across *D. melanogaster* lines. We observed significant variation among lines for thorax length (both males and females), testis length, and sperm length, but SR length did not significantly differ between lines (Table S3). Box plots show the median (central line) and first and third quartiles; whiskers extend to the smallest and largest values (not including outliers >1.5x the inter-quartile range).

Alt text: A graphical representation of variation in morphology across male and female *D. melanogaster* lines. Data are represented using box plots and individual data points.

Figure 4. Courtship effort (courtship behaviors per minute) consistently differed between second male genotypes but did not consistently vary by the female genotype being courted or by male perfuming status (Table S4). Conspecific-perfumed males are shown in grey, and heterospecific-perfumed males are shown in purple. The dashed line indicates the average courtship effort for each male line. Box plots show the median (central line) and first and third quartiles; whiskers extend to the smallest and largest values (not including outliers >1.5x the inter-quartile range). The y-axis has been truncated to make variation in the data easier to visualize; the non-truncated data (as used in statistical analyses) is shown in Figure S3.

Alt text: A matrix of graphs representing second male courtship effort across all possible male by female combinations, divided according to conspecific- or heterospecific-perfumed treatments. Data are represented using box plots and individual data points.

Figure 5. Counts of mating successes (dark shading) and mating failures (light shading) differs across male (column) and female (row) genotypes (Table S5). Results are shown separately for conspecific-perfumed males (grey) and heterospecific-perfumed males (purple) within each male x female panel.

Alt text: A matrix of graphs representing mating successes vs. failures across all possible male by female combinations, divided according to conspecific- or heterospecific-perfumed treatments. Data are represented with stacked bars, where the lower bar represents the count of mating failures and the upper bar represents mating successes.

Figure 6. Female use of second male sperm (P2) varies across female genotypes, male genotypes, and female by male genotype combinations (Table 1). P2 also varies depending on male perfuming status, as shown in grey (conspecific-perfumed males) and purple (heterospecific-perfumed males). The dashed line indicates the average P2 for each male line. Box plots show the median (central line) and first and third quartiles; whiskers extend to the smallest and largest values (not including outliers >1.5x the inter-quartile range).

Alt text: A matrix of graphs representing second male siring success across all possible male by female combinations, divided according to conspecific- or heterospecific-perfumed treatments. Data are represented using box plots and individual data points.

Figure 7. Second male courtship effort is significantly correlated with siring success when males are perfumed by conspecifics (a),but is not significant when perfumed by heterospecifics (b). Female remating rate is not correlated with second male siring success when males are conspecific-perfumed (c) but is correlated when males are heterospecific-perfumed (d). Sperm length is significantly correlated with siring success for both conspecific-perfumed (e) and heterospecific-perfumed males (f). Crosses are labeled as female x male genotype (G, Greece; T, Taiwan; C, California; and M, Malaysia).

Alt text: Scatterplots representing the correlation between datasets. Points are represented using the first letter of each genotype in that cross (for example, C x G is California female by Greece male). Data is divided between conspecific-perfumed and heterospecific-perfumed males. Sperm length and second male siring success show the clearest pattern of correlation.