The process that the evolution from C_3 towards C_4 photosynthesis is driven

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Compared to the plants with C_3 photosynthesis, the plants with C_4 photosynthesis have higher
light, water, and nitrogen use efficiencies. Though only 3% of higher plants have C_4
photosynthesis, C_4 photosynthesis contribute about 23% of terrestrial gross primary productivity.
Currently, it remains elusive how the procedure of the evolution from C_3 towards C_4
photosynthesis is driven. Here, *Flaveria* genus, the chief model for C_4 evolutionary researches,
was used to study this driving through a combined analysis of ecological, physiological and
biochemical data. From proto-Kranz to C_4-like, results showed that precipitation was negatively
associated with the activity of C_4 metabolic core enzyme and C_4 flux, respectively. With increased
vein density, an increased temperature and precipitation were observed during the earlier
evolutionary stage. Our study indicated that decreasing precipitation and increasing temperature
accompanied the transition from C_3 to C_2 photosynthesis and further transition from C_2 to
developing C_4 photosynthesis. There was a positive relationship between CO_2 compensation point,
stomatal density and precipitation, supporting the notion that decreasing precipitation enhances
photorespiration through reducing stomatal conductance during the evolution. As the driving force
from proto-Kranz to C_4-like, our study highlights that drought dominated rather than heat during
C_4 evolution. This study suggests that the driving force between certain stages during the
evolution towards C_4 photosynthesis are different and increased water availability should be
needed in the initial phase.

Key words:
Evolution, C_4 photosynthesis, intermediate plants, precipitation, temperature, driving

Introduction
Photosynthesis undergone evolution, which results in different photosynthetic type, and C_4
photosynthesis is one of these photosynthetic types (Sage 2004; Langdale 2011). Compared to C_3
relative, the plant with C_4 photosynthesis has higher water use efficiency, nitrogen use efficiency
and light use efficiency (Vogan and Sage 2011; Taylor et al. 2010). Although only accompanying 3%
plant species, C_4 photosynthesis contributes above 20% of terrestrial gross primary productivity
(Still et al. 2003; Sage et al. 2012). The evolution from C_3 to C_4 photosynthesis go through
multiple stages (C_4 evolution) (Sage et al. 2012; Sage 2004; Schluter and Weber 2016; Brautigam

and Gowik 2016). The first stage of C_4 evolution is preconditioning in which C_3 plants acquire
new cis-elements (Akyildiz et al. 2007).For this stage, duplication of gene occurs (Akyildiz et al.
2007; Monson 2003), vein density increases (Marshall et al. 2007; Mckown and dengler 2007) and
the number of plastids in the bundle sheath increases (Sage et al. 2013). The second stage is
proto-Kranz. In this stage, plants still perform C_3 photosynthesis (Sage et al. 2013) while the
number of plastids continue to increase (Stata et al. 2019; Sage et al. 2012) and chloroplast and
mitochondria localize to the centripetal wall of bundle sheath cells. The third stage is C_2
photosynthesis (Type I) (Schulze et al. 2013; Sage et al. 2012; Edwards and KU 1987), which is
characterize as the change in the location of GLYCINE DECARBOXYLASE P SUBUNIT
(GDCP) and photosynthetic apparatus from mesophyll cell to bundle sheath cell (Schulze et al.
2013; Stata et al. 2019; Sage et al. 2013). The released CO_2 from photorespiration can be
moderately concentrated in the bundle sheath cell (Photorespiratory CO_2 pump) (Keerberg et al.
2014; Sage et al. 2012). The fourth stage is to induce and develop C_4 photosynthesis (Type II)
(Schluter and Weber 2016; Sage et al. 2018; Nakamura et al. 2013; Mallmann et al. 2014). The
establishment of C_2 photosynthesis could cause the imbalance of ammonia between mesophyll cell
and bundle sheath cell, and the induction of C_4 metabolism is proposed as a mechanism to resolve
this trouble (Mallmann et al. 2014). The fifth stage is functionally developed C_4 photosynthesis
(C_4-like) (Stata et al. 2019). For this stage, though there are still some Rubisco in mesophyll
(Taniguchi et al. 2021; Monson et al. 1987; Stata et al. 2019; Sage et al. 2018), various types of
indicators at this stage are close to plants with ultimate C_4 photosynthesis (Vogan and Sage 2011;
Ku et al. 1991; Adachi et al. 2023). The final stage is the mature C_4 photosynthesis (Mature C_4)
(Sage et al. 2012), and mesophyll cell contain few RuBisCO (Stata et al. 2019). These stages in
the evolutionary transition between C_3 and C_4 photosynthesis are called intermediate stage, and
the plants in these stages are intermediate plants (intermediate species).
The evolution from C_3 towards C_4 photosynthesis is considered to be driven by enhancing
photorespiration, which is promoted by low concentration of atmosphere CO_2, drought, salt and
heat (Sage et al. 2012; Brautigam and Gowik 2016; Sage 2004; Edwards and Smith 2010;
Bromham and Bennett 2014). Low concentration of atmosphere CO_2, drought and salt could
decrease the ratio of CO_2 to O_2 concentration, thereby enhancing photorespiration (Sage et al.
2012). Increasing temperature enhances photorespiration by reducing the concentration of CO_2
relative to O_2 in the chloroplast stroma (Ogren 1984) and decreases the specificity of RuBisCO for
CO_2 relative to O_2 (Ogren 1984; Sage et al. 2012). Studies have shown that C_4 evolution are
correlated with the decrease in the concentration of atmosphere CO_2 and the appearance of dry,
salty and hot environment (Christin et al. 2008; Edwards and Smith 2010; Edwards and Still 2008;
Bromham and Bennett 2014), which suggests they are the driving force for C_4 evolution. However,
all these studies assume that the entire process of C_4 evolution is under a single driver. Given C_4
evolution includes multiple phases and the evolution is complicate, it is possible that these phases
of C_4 evolution might be under different driving force rather than the same driving force
persistently along the evolutionary road towards C_4 photosynthesis (Heckmann et al. 2013; Sage et
al. 2012).
Plants obtain fitness in each step of C_4 evolution (Heckmann et al. 2013; Sage et al. 2012). The
increased vein density is an important characteristics in the initial stage of C_4 evolution (Sage et al.
2012; Sedelnikova et al. 2018; Kumpers et al. 2017; Sage et al. 2014) (Mckown and dengler 2007;
Marshall et al. 2007; Williams et al. 2013), which might help supply water to the mesophyll and

then maintain stomatal opening, since evaporation might be enhanced at this time (Brautigam and Gowik 2016; Sage et al. 2018) (Sage et al. 2012). Therefore, available water content in the environment for this stage may be increased to ensure the fitness of plants (Brautigam and Gowik 2016). However, this contradicts with the notion that the origin of C_4 photosynthesis is promoted by dry environment (Sage et al. 2018; Edwards and Smith 2010; Sedelnikova et al. 2018). Since C_2 photosynthesis and C_4 photosynthesis could largely reduce photorespiration (Sage 2004; Schluter and Weber 2016; Sage et al. 2012), the formation of C_2 from C_3 photosynthesis and the transition from C_2 to developing C_4 photosynthesis may be driven by the conditions which could enhance photorespiration (Schluter and Weber 2016; Sage et al. 2018; Sage et al. 2012),but the driving forces during these transitions also remain undefined.

Flaveria genus is a widely used system to study C_4 evolution (Sage et al. 2012; Sage 2017; Schluter and Weber 2016; Mallmann et al. 2014; Sage et al. 2013; Vogan and Sage 2011; Nakamura et al. 2013), and the studies on *Flaveria* genus has laid the foundation for the pyramid model of C_4 evolution (Sage et al. 2013; Vogan and Sage 2011; Sage et al. 2012). The photosynthetic types of *Flaveria* species span entire evolutionary stage towards C_4 photosynthesis(Sage et al. 2012; Sage et al. 2014; Schluter and Weber 2016; R.K. Monson 1986). *Flaveria* is also the youngest genus among the known C_4 linage and it's species has the close relationship (Schulze et al. 2013). These make *Flaveria* genus an ideal model to study C_4 evolution (Christin et al. 2011; Sage et al. 2012). The ecology for the habitats of intermediate species and C_4 species could reflect the environmental conditions that drive C_4 evolution (Sage et al. 2012; Sage et al. 2018), therefore, the ecology of *Flaveria* habitats enable us to finely study the driving force in the evolutionary road towards C_4 photosynthesis. In this study, we collected the ecological, biochemical and physiological data for different species of *Flaveria* genus to explore the procedure of driving for the evolutionary trajectory towards C_4 photosynthesis and how these different evolutionary phases help plants gain fitness.

Results

1. Most of *Flaveria* species were distributed inland and a few species were distributed along the coast

The habitats of *Flaveria* species were mapped to the North and South America continents with the longitude and latitude of each *Flaveria* samples downloaded from Global Biodiversity Information Facility (GBIF) (Fig 1A, B). Species were ordered based on the amount of C_4 flux (Fig 1B, C, supplementary fig 1). We found most *Flaveria* species were located inland (Fig 1A, B). A few species (*F.bro*, *F.flo*, *F.lin*) were distributed along the coast (Fig 1B), which was consistent with the previous reports (Powell 1978). During evolution, intermediate species gradually spread to locations with higher latitude (Fig 1B). Compared with the intermediate species, the distribution of C_4 species was more widespread (Fig 1A, B, C), indicating that adaptive capacity of C_4 plants is enhanced (Fig1).

2. The changed pattern for the temperature and precipitation of *Flaveria* growth habitats

The temperature and precipitation of the habitats for each *Flaveria* species were obtained from the worldclim database (Fick and Hijmans 2017). The results from physiological, anatomic, molecular and computational modeling experiments indicate the direction of C_4 evolution could be described by species's order towards C_4 photosynthesis in phylogenetic tree (Figure 2A) (Mckown

and dengler 2007; Nakamura et al. 2013; Gowik et al. 2011; Sage et al. 2012; Sage et al. 2018;
Schluter and Weber 2016). Along the evolutionary pathway towards C_4 photosynthesis (Ordinal
species towards C_4 photosynthesis of the phylogenetic tree in Figure 2A), the annual mean
temperature, mean temperature of warmest quarter, mean temperature of wettest quarter and max
temperature of warmest month first increased and then decreased, though difference in some
species habitats is not significant (Fig 2A, supplementary fig 2, supplementary fig 3,
supplementary fig 4, supplementary file). Mean temperature of warmest quarter, mean
temperature of wettest quarter and max temperature of warmest month increased significantly in
the habitat of *F. rob* (C_3) and *F. son* (Type I) (Fig 2A, B, supplementary fig 2, supplementary fig 3,
supplementary file), compared with those of *F. pri* (C_3). However, these environmental conditions
significantly decreased in the habitat of *F. ang* (Type I), compared with *F. son* (Type I) (Fig 2A,
supplementary fig2, supplementary fig3, supplementary file). For the mean temperature of wettest
quarter, mean temperature of warmest quarter and max temperature of warmest month, compared
with *F. son* (Type I), the habitat of species with C_2 photosynthesis (*F. ang*, *F. chl*, *F. pub*, *F. opp*, *F.
ano*, *F. ram*) had a lower mean temperature, but the results of statistic test for some species are not
significant or in margin (Fig 2A, supplementary fig 3D, supplementary file). The habitats of
species *F. flo* (Type II) and *F. lin* (Type II) had significantly higher mean temperature compared to
most of other intermediate species (Fig 2A, supplementary file). Max temperature of warmest
month for *F. son* (Type II) habitats was extremely high (Fig 2d, supplementary file).
For the precipitation of wettest quarter, precipitation of warmest quarter and precipitation of
wettest month, along C_4 evolutionary pathway, they significantly increased in the *F. rob* (C_3)
habitat, compared with those in *F. pri* habitat (Fig 2A, B, supplementary fig 3, supplementary file).
There was an increased trend from preconditioning to proto-Kranz (Fig 2A, B, supplementary fig
3). Compared with *F. rob* (C_3) habitat, precipitation significantly decreased in the *F. ang* (Type I)
habitat (Fig 2A, supplementary fig 3, supplementary file). Precipitation decreased in *F. son* (Type I)
habitat, and though the difference between *F. rob* and *F. son* is not statistically significant (Fig 2A,
supplementary fig 3, supplementary file). Precipitation of warmest quarter was higher than
precipitation of coldest quarter, and similar to precipitation of wettest quarter (Fig 2A,
supplementary fig 2, supplementary figure 3, supplementary file). Expect *F. flo* (Type II) and *F. lin*
(Type II), precipitation of the habitat of species with C_2 photosynthesis decreased persistently
along the path of C_4 evolution, though difference for the habitats of some species is not significant
(Fig 2A, supplementary fig 3, supplementary file). In the initialization of C_4 evolution, the initial
increase in precipitation and temperature (*F. pri* to *F. rob*) coincided with the initial appearance of
the higher vein density (Fig 2A, B, C, supplementary fig 3, supplementary 4). Although the
precipitation of the Type II species *F. flo* and *F. lin* habitat was high (Fig 2, supplementary fig 3,
supplementary file), they distributed along coast and inhabited the saline land (Table 1), likely
experiencing strong osmotic stress.
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3. The relationship between precipitation (or temperature) and C_4 metabolism related traits during
the transition from proto-Kranz to C_4-like
For the clade A, the activities of C_4 metabolism related enzymes PEPC, PPDK and MDH
showed a negative correlation with precipitation, and there showed a linear relationship between
log enzyme activity and log precipitation (Fig 3 A, B, C, supplementary file). The negative
relationship between C_4 flux and precipitation can be described as a linear function (Fig 3D). For

the clade B, the relationship between log enzyme activity and log precipitation also can be
described as a linear function (Fig 3 I, J, K, supplementary file). However, the linear relation
between log enzyme activity and log precipitation in clade B is weaker than clade A (Fig 3I,J,K,
supplementary file). A weak and negative relationship between C_4 flux and precipitation was
observed in clade B (Fig 3L). The linear relation between log enzyme activity and log temperature
is weaker than that between log enzyme activity and log precipitation, regardless of clade A or
clade B (Fig 3E, F, M, N, G, H, O, P, supplementary file). The linear relationship between C_4 flux
and temperature is weak in both clade A and clade B (Fig 3H, P).

4. The change in temperature and precipitation between the transition of stages during C_4
evolution.
The comparison of *F. rob* and *F. son* suggested that incipiently decreased precipitation along the
evolutionary way towards C_4 photosynthesis concurrently occurred with increased mean
temperature (Fig 4A, F). Precipitation levels of the habitat of *F.ano* and *F.ram* were significantly
lower than that of *F. ang*, and temperature values of the habitat of *F. ram* and *F. ano* were
significantly higher than that of *F. ang* (Fig 4B, G, C, H). Precipitation of *F. pal* habitat was
significantly lower than that of *F. ram*, and precipitation of *F. bro* habitat is significantly lower
than that of *F. flo* (Fig 4D, E). However, temperature for the habitat of *F.pal* and *F. bro* showed the
inconsistent change, temperature for the habitat of *F. pal* was similar to that of *F. ram*, and
temperature of *F. bro* habitat was significantly lower than that of *F. flo* (Fig 4I, J). Furthermore,
compared with their more ancient species, we observed that the decreased precipitation for the
species habitat (*F. son*, *F.ram*, *F.flo*, *F.pal* and *F.bro*) concurrently appeared with the decreased
stomatal density, decreased inhibition of O_2 and CO_2 compensation point, decreased maximum
stomatal conductance (Supplementary Fig 5E, F, G, H). Although the precipitation of *F. flo* and *F.*
lin were increased compared to other species with C_2 photosynthesis (Fig 2A, supplementary fig3,
supplementary fig4), they also had the decreased stomatal density, maximum stomatal
conductance (Supplementary fig 5), which might be related to their salty habitats (Table1).

5. The relationship between precipitation, stomata and CO_2 compensation point.
Previous work found there was a difference in stomata density between species at different C_4
evolutionary stages in the *Flaveria* genus (Zhao et al. 2022). Furthermore, there was a positive and
linear relationship between stomata density, CO_2 compensation point and precipitation during the
transition between proto-Kranz and C_4-like species in *Flaveria* clade A and clade B (Fig 5A, B, C,
D, E, and F, Supplementary file). The linear relationship between stomata, CO_2 compensation
point and precipitation in *Flaveria* clade B were weaker than that in clade A (Fig 5A, B, C, F, G, H,
Supplementary file). The relation between temperature and stomata density or CO_2 compensation
point was not apparent as the relationships between these two parameters and precipitation (Fig
5D, I, E, J, supplementary file).

Discussion
Evolutionary conditions along the path towards C_4 photosynthesis
Heat and drought have long been thought to be associated with C_4 evolution (Sage et al. 2018;
Taylor et al. 2012). Exploring environmental conditions of the habitats for C_4 and intermediate
species could contribute to elucidate the drivers for C_4 evolution (Sage et al. 2018; Sage et al.

2012; Edwards and Smith 2010). In this work, we examined precipitation and temperature in
diverse *Flaveria* species during different periods (Fig 2A, supplementary fig 2, 3, 4). We noticed
that the quantity and trend of wettest quarter mostly coincided with those of warmest quarter (Fig
2A, supplementary fig 3), mean temperature of wettest quarter was overall higher than mean
temperature of driest quarter (Supplementary fig 3D, Supplementary fig 2E) and precipitation of
warmest quarter was overall higher than precipitation of coldest quarter (Supplementary fig 3C,
supplementary fig 2B). These results suggested that rain and heat were in the same season for the
habitat of *Flaveria*. Sufficient water content was necessary to maintain high photosynthetic
activity for plants, therefore this suggestion was in line with previous work that indicates *Flaveria*
species perform high photosynthesis during summer with heat in Mexico (Powell 1978). Because
there is no apparent change in Rubisco Sc/o (Kubien et al., 2008) and similar Ci/Ca between C_3
and type I intermediate species (Adachi et al., 2023), the higher photosynthetic activity would
indicate also a higher photorespiratory flux in C_3 plants and initial intermediate species. Besides,
under such conditions, photorespiration, long regarded as the driving factor of C_4 evolution, might
further be enhanced by heat (Sage et al. 2012; Sage et al. 2018). Furthermore, any evolutionary
selective pressure on photosynthesis should act in the periods with active photosynthesis (Sage et
al. 2018), i.e. the driest and coldest season should not be considered for the study on the
evolutionary trajectory towards C_4 photosynthesis (Supplementary fig 2). Therefore, the periods
with rain and heat for *Flaveria* genus are valuable to explore the process of the driving of C_4
evolution (Fig 2A, supplementary fig 3).
We found that the initial stage of C_4 evolution was associated with high precipitation and
temperature, and later stage was associated with decreased precipitation (Fig 2A, Fig 4,
supplementary fig 3). Heat is considered to be a major factor that promotes the origin of C_4
photosynthesis (Sage et al. 2018). Here, our results suggested that in addition to increased
temperature, the increase in precipitation was also a vital factor to initiate the evolution towards C_4
photosynthesis (Fig 2, Supplementary fig 3). It is possible that heat occurred with drought to
enhance photorespiration simultaneously and then promoted C_4 evolution (Sage et al. 2012). Here,
data from this study indicated that this process indeed existed, and it was at the transition from C_3
to C_2 (*F. son*) and the transition from C_2 to developing C_4 photosynthesis (*F. ram*, *F. ano*) (Fig 4).
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The evolutionary mechanism during the initial phase of C_4 evolution
The initial stage of C_4 evolution is featured as having increased vein density and increased
number of chloroplasts in bundle sheath (Sage et al. 2012). The increased vein density can help
maintain sufficient water content in leaf and avoid the reduction of stomatal conductance, since
the evaporation might be increased at this stage (Sage et al. 2012; Sage et al. 2018). In agreement
with this, our data showed that, when at the stage of vein density increasing, there was no change
in stomatal development, and temperature was indeed increased (Fig 2A, B, C, D, supplementary
fig 5). The increased temperature could increase vapor pressure between leaf and air and hence
enhance the evaporation (Sage et al. 2018). However, we found that water vaper pressure in
atmosphere was increased in this stage and this could decrease water vapor deficit (Supplementary
fig 6B). No change in the stomatal density and maximum stomatal conductance indicated that
water shortage in leaf did not occur (Supplementary fig 5), since long-term water shortage could
alter stomatal development (Franks et al. 2009; Taylor et al. 2012). Importantly, our data also
showed that precipitation was increased at this stage, which matched with the increased vein

density, and this ensured the sufficient supply of water to the leaf (Fig 2A, B, C). Increased precipitation and keeping stomatal opening could not only maintain intercellular CO_2 concentration, but also could help alleviate the increase of leaf temperature. These changes all benefit the acquirement of carbon. Therefore, the initial stage of C_4 evolution could be promoted by high temperature and precipitation (Fig 2).

The increase in vein density could decrease volume of mesophyll in a leaf (Sedelnikova et al. 2018; Mckown and dengler 2007), which inevitably decreases the photosynthetic capacity of the leaf. In theory, this should create a strong evolutionary pressure to select for the increased photosynthetic capacity of BSC to offset the loss of photosynthetic capacity. In consistent with this, the relative number and size of chloroplast in bundle sheath cell of *F. rob* is slightly increased compared to those of *F.pri* (Sage et al. 2013). Besides, the photosynthetic apparatus located into BSC from MC could benefit the refixation of CO_2, since BSC locates to the inner of leaf and photosynthetic apparatus in BSC locates to the inner cell periphery (Sage et al. 2013). Indeed, the Proportion in half of cell, coverage of inner half of cell area and compensation CO_2 point at high light intensity are also slightly increased (Sage et al. 2013). The light intensity of *F. rob* habitat was higher than that of *F. pri* (Supplementary fig 5A); the size of bundle sheath in *F. rob* decreases compared with *F. pri* (Sage et al. 2013; Mckown and dengler 2007); the size of mesophyll cell decreases compared with *F. pri* (Mckown and dengler 2007); and the thickness of leaf become less (Mckown and dengler 2007); all these enhanced the level of light that bundle sheath experiences, consisted with the notion that selection pressure for bundle sheath with higher photosynthetic capacity might exist at this stage.

Drought as a more important factor than heat for driving C_4 evolution

Some studies have proposed that drought is a primary driving force for C_4 evolution (Sage 2004; Zhou et al. 2018; Sage et al. 2018; Schluter and Weber 2016; Edwards and Smith 2010), and other studies suggest that heat is the critical environment factor driving C_4 evolution (Sage et al. 2018; Sage et al. 2012). In this study, we proposed that drought played a more important role than heat during C_4 evolution (Fig 3). From proto-Kranz to C_4-like, precipitation declined step by step, however, there was no clear trend of change in temperature (Fig 2). Regardless of clade A or Clade B, we found a negative association between core C_4 enzyme activity and precipitation (Fig 3, supplementary file) and positive association between CO_2 compensation point and precipitation (Fig 5, supplementary file). In particular, for clade A, there was a strong negative relationship between precipitation and the activities of core C_4 enzymes (Fig3, supplementary file). However, the relationship wasn't apparent for temperature (Fig 3, supplementary file). These results indicated that progressing C_2 photosynthesis and developing C_4 photosynthesis mainly associated to decreasing precipitation (Fig 2A, B, C, D, E, F, Fig 3). Therefore, drought rather than heat, was suggested to be the major driving force for the evolution from proto-Kranz to C_4-like. Flaveria genus has long been thought as the chief model in C_4 evolutionary study and it represents C_4 evolution of all evolutionary route to a large extent. Therefore, drought might dominate this transitional in all C_4 evolutionary route. C_4 evolution repeat independently above 60 times (Sage et al. 2012). It is possible that heat dominates this transitional process in the evolution of other C_4 lineages. In future, the drivers for this transitional process in other C_4 evolutionary lineages still need to be checked.

Different evolutionary drivers at the transition of the stages in C_4 evolution

The environmental drivers for the evolution towards C_4 photosynthesis have been intensively
studied. For example, decreased concentration of atmosphere CO_2 has been thought as a driving
force of C_4 evolution (Christin et al. 2008; Concentrations and Paleogene). Drought, salt and heat
were also thought to be conditions that promote C_4 evolution (Sage et al. 2018; Sage 2004;
Bromham and Bennett 2014) (Edwards and Still 2008; Edwards and Smith 2010; Pearcy and
Ehleringer 2006; Sage et al. 2012). These studies assume that entire C_4 evolutionary process is
promoted by a single ecological condition. However, here, our results suggested that
environmental conditions promoted C_4 evolution in an orderly and stepwise manner (Fig 6), and
different selection pressure drove the transition of certain stages during the evolutionary way
towards C_4 photosynthesis (Fig 6).

Besides the initialization of C_4 evolution, the evolution towards C_2 photosynthesis is also
considered to be a crucial step during C_4 evolution (Sage et al. 2012; Schluter and Weber 2016).
Previous studies have reported that *Flaveria* species with C_2 photosynthesis can represent a
transition between C_3 and C_4 photosynthesis during the evolution (Heckmann et al. 2013;
Mallmann et al. 2014; Schluter and Weber 2016; Sage et al. 2012), and *F. ram* and *F. ano* have
enhanced C_4 cycle flux, C_4 cycle related enzyme activity and gene expression levels (Adachi et al.
2023; Mallmann et al. 2014; Vogan and Sage 2011; R.K. Monson 1986). According to the
phylogenetic tree, *F. ang* is closer to *F.ram* as compared with *F.son* (Fig 2A), therefore, among all
Flaveria species, *F.son* most likely represents the emergence of C_2 photosynthesis in *Flaveria*.
Similarly, *F. ram* and *F. ano* most likely represent the emergence of developing C_4 photosynthesis
in clade A and clade B, respectively, and *F. pal* and *F. bro* most likely represent the emergence of a
fully functional C_4 cycle for clade A and clade B, respectively (Fig 2A). Therefore, the increased
temperature and decreased precipitation simultaneously promoted the emergence of C_2
photosynthesis (Fig 4A, F). Increasing temperature and decreasing precipitation could enhance
photorespiration and thus promote the formation of photorespiratory CO_2 pump which improves
the fitness of these intermediate plants. The photorespiratory CO_2 pump largely decreases the
photorespiration (Sage et al. 2018; Sage et al. 2012; Schluter and Weber 2016; Ku et al. 1991),
which is indicated by the large decrease in CO_2 compensation point and O_2 inhibition (Sage et al.
2013; Ku et al. 1991). Study proposes photorespiration promotes C_4 evolution by decreasing
stomatal conductance under drought (Sage et al. 2012). As the support, we provided the evidence
that there was a positive relationship between stomatal density, CO_2 compensation point and
precipitation (Fig 5). These also suggested that the concurrent evolution for decreased stomata
density and the emergence of C_2 and C_4 photosynthesis during C_4 evolution (Fig 5). The
enhancement of C_2 photosynthesis has long been considered to cause the imbalance of nitrogen
between mesophyll cell and bundle sheath cell, which induces C_4 cycle (Mallmann et al. 2014). In
case of the evolutionary process from C_2 photosynthesis to developing C_4 photosynthesis,
precipitation decreased and temperature increased (Fig 4B, C, G, H), and this evolutionary process
could be driven. The formation of functionally developed C_4 photosynthesis was driven by
inconsistent conditions for Clade A and Clade B (Fig 4I, J), and precipitation have played a
dominated role in this step. There was no significant difference and a decrease in temperature of
the habitats (Figure 4 I, J), which suggested that the temperature was not a deciding factor
controlling this transition.

In summary, this study analyzed the data of ecology, physiology and biochemistry for different
species of *Flaveria* genus to reveal the process that the evolution from C_3 towards C_4
photosynthesis is driven, and results indicated plants suffered carbon limitation in the early phase
of C_4 evolution and the primary driving force for this phase is carbon limitation (Fig 6). Later,
plants suffered water limitation leading to decease in acquirement of carbon, therefore, the
primary driving force for these phases of C_4 evolution is water limitation (Fig 6). For a long time,
most of studies proposed that environmental condition causing the carbon restriction through
enhancing photorespiration (drought, heat, salty and low concentration of atmosphere CO_2) drove
the evolution towards C_4 photosynthesis(Christin et al. 2008; Edwards and Smith 2010; Schluter
and Weber 2016; Sage 2004; Sage et al. 2012; Sage et al. 2018). Our study proposed these studies
were not subtlety-oriented and suggested that certain steps of the evolution from C_3 to C_4
photosynthesis were driven by different environmental factors which sequentially promoted C_4
evolution, in contrast to the current notion that the entire process of C_4 evolution is driven by same
environmental factor (Fig 6). The initial evolution was promoted by high precipitation, while later
decreased precipitation positively dominated the transition among C_3, C_2 and C_4 photosynthesis
(Fig 6). More important, the increase in vein density could represent a crucial characteristics for
the starting point of the evolution from C_3 towards C_4 photosynthesis (Christin et al. 2012;
Mckown and dengler 2007; Sage et al. 2012), therefore, increased humidity and temperature
together may be the most important condition that facilitated the initial evolution towards C_4.
Therefore, the origin of C_4 evolution might be in the habitat with raised humidity and temperature,
rather than drought, heat, salt and low concentration of atmosphere CO_2 (Bromham and Bennett
2014; Christin et al. 2008; Sage et al. 2018). Our study could benefit the prediction of the
occurrence of C_4 evolution. C_4 photosynthesis is crucial for the ecology and environment.
Therefore, this study may help us handle the climate and environment changes currently and in the
future. Additionally, the understanding provided in this study for the driving of C_4 evolution might
benefit to identify the regulatory innovation that facilitated C_4 evolution, which might help us to
perform C_4 engineering.
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Methods
1. Obtain the ecological data of *Flaveria* species.
First, the geographical distribution of *Flaveria* was acquired from Global Biodiversity
Information Facility (GBIF) database using the rgibf software package. Using ArcMAP software,
we acquired from the Worldclim database the elevation data of the respective locations and 19
Bioclimatic factors, including Mean Diurnal Range, annual mean Temperature, Isothermality,
Temperature Seasonality, Max Temperature of Warmest Month, Temperature Annual Range, Mean
Temperature of Wettest Quarter, Mean Temperature of Warmest Quarter, Annual Precipitation,
Precipitation of Wettest Month, Precipitation of Warmest Quarter, Precipitation of Wettest Quarter,
Precipitation of Coldest Quarter, Min Temperature of Coldest Month, Precipitation of Driest
Quarter, Mean Temperature of Coldest Quarter, Precipitation of Driest Month, Precipitation
Seasonality (Coefficient of Variation), Precipitation of Coldest Quarter based on the geographical
distribution information of the *Flaveria*. Mean Diurnal Range, Isothermality, Temperature
Seasonality, Temperature Annual Range, Precipitation Seasonality (Coefficient of Variation) were
not used in this study. Specifically, we downloaded the data from the worldclim at first. Secondly,
we prepared the file for the information of sampling location. Thirdly, we imported the layers file

of climate factor. Fourthly, we imported the information of sampling location. Fifth, generating the
new layers. Lastly, we exported the file that including sample and the data of geographical factors.
All bioclimatic variables were averaged from 1970 to 2000. All data from the worldclim database
have a maximum spatial resolution of 30 arc-seconds. The water vaper pressure and solar radiation
were also extracted from the World clim database(Fick and Hijmans 2017), based on the
geographical distribution information of *Flaveria*. The water vaper pressure and solar radiation in
June, July and August were extracted. The water vaper pressure and solar radiation were also from
1970-2000.
405
2. Obtain the biochemical, anatomical and physiological data of *Flaveria* species.
The biochemical, anatomical and physiological data for *Flaveria* was acquired from previous
research. The data of vein density and distance between vein was from (Mckown and dengler
2007). The data of vein density and distance between vein was digitized by Origin 2018.
Specifically, first, importing the figure with the picture format; Second, compiling the coordinate
axis with the Image digitizing tool in Origin 2018; Third, grabbing and obtaining the data point
with the Image digitizing tool in Origin 2018; Fourth, exporting the data. The data of CO_2
compensation point and O_2 inhibition was from (Ku et al. 1991), and the means of the data of CO_2
compensation point and O_2 inhibition were used in this study. The data related to stomata was
from our work (Zhao et al. 2022), and the means were used in this study. The data of C_4 metabolic
enzyme activity was from (Supplementary table S7) in (Adachi et al. 2023). The growth habitat
was acquired from (Powell 1978; Kocacinar et al. 2008).
418
3. Map the *Flaveria* species
We mapped and marked the locations for the habitats of different *Flaveria* species to a world
map using the software google earth Pro (version 7.3) based on the Latitude and longitude from
Global Biodiversity Information Facility (GBIF). Coordinates were plotted as placemarks, with
customized icons to distinguish between different species to facilitate the visualization and
analysis of distribution patterns and geographical clusters. Google Earth Pro, version 7.3. (2020).
[Software]. Google LLC. Available at: https://www.google.com/earth/.
426
4. Statistical analysis, regression analysis and Graph plotting.
The graph was plotted and the regression analysis was performed by using GraphPad prism 8.
The statistic test was performed by GraphPad prism 8, WPS office or Microsoft office. The
statistic test for the difference in precipitation or temperature between all *Flaveria* species was
performed by one-way ANOVA, and the difference between each group was conducted using
Tukey's multiple test (Figure 2A, supplementary Figure 2,3,4, supplementary file). The rest of
other statistic test (Figure 2B, C, Figure 4) was conducted by Student's t test, and the detail in the
statistic test was described in each figure legends. The linear regression in Figure 3 was conducted
by using the $\log_2$ value of the precipitation of wettest quarter and $\log_2$ value of precipitation of C_4
metabolic enzyme activities, and linear regression between them were performed with GraphPad
prism 8 (Supplementary file). Specifically, conducting the option of analyze; Then, selecting linear
regression; Exporting the result. The parameters in GraphPad prism 8 for the linear regression
were the defaults. The results of the linear regression were in supplementary file. The
phylogenetic tree for *Flaveria* was drawn by integrating the studies in (Lyu et al. 2015; Mckown

2005).

446   Table 1 The habitat of *Flaveria* species

Species (photosynthetic type)	Growth habitat	Geographic location
Flaveria pringlei (C_3)	Rather arid and gypseous habitats, scattered localities, waste places, cultivated fields, roadsides	Southern Mexico
Flaveria cronquistii (C_3)	Rocky limestone (possibly gypseous) areas	Southern Mexico
Flaveria robusta (C_3)	Gypsum and slate, sandy clay, or open rocky slopes, weedy at roadsides	Western Mexico
Flaveria sonorensis (Type I)	Close to warm mineral springs	North-western Mexico
Flaveria angustifolia (Type I)	Sandy or loamy soils in open fields	Southern Mexico
Flaveria pubescens (Type II)	Scattered localities, gypsum soils	Central Mexico
Flaveria oppositifolia (Type I)	high gypsum plains or other areas with dilute gypsum, saline, or rarely loamy soil, usually in moist areas or in disturbed habitats at roadside	Central Mexico
Flaveria linearis (Type II)	Near the coast and on offshore islands	Western Florida, USA
Flaveria floridana (Type II)	Restricted to saline, white sand near the beaches of brackish marshes	Western Florida, USA
Flaveria ramosissima (Type II)	Sandy-clay bottoms, along dry washes, fields and roadsides, possibly in gypseous soils,	Southern Mexico
Flaveria anomala (Type II)	High gypsum plains, often in disturbed sites	Central Mexico
Flaveria palmeri (C_4-like)	Gypsum flats and knolls, or clay-loam soils probably with dilute gypsum, bolsons near lagunas, distributed areas near roads and fields	Northern Mexico
Flaveria brownie (C_4-like)	Saline, sandy, and marshy areas of the coastal flats and islands of the lower Texas Gulf Coast	North-eastern Mexico
Flaveria vaginata (C_4-like)	Arid and gypseous soils, scrub vegetation, cultivated fields	South-central Mexico
Flaveria bidentis (C_4)	A weed in moist places on waste ground, near rivers, streams, and barrancas, in fields, pastures, disturbed ground, near streets and sidewalks in towns, claygravel, or sand	Africa, Caribbean, South America, southern USA

Flaveria trinervia (C$_4$)	a variety of habitats, usually saline, gypseous, and disturbed, usually near ephemeral or permanent water sources	Africa, Caribbean, Central/South America, Mexico, USA
Flaveria chloraefolia (Type I)	Permanent or ephemeral saline marshy soils, springs, streams, roadsides	North Mexico

The information of habitat of *Flaveria* species was from (Powell 1978; Kocacinar et al. 2008)

Figure legends

Fig 1 Distribution of the *Flaveria* species

(A) The location of *Flaveria* species was mapped to the world map. The main habitat of *Flaveria*

is North and South America, which was displayed. (B) The location of *Flaveria* species except C$_4$

species was mapped to the world map. (C) and (D) The latitude and longitude of different *Flaveria*

species. The latitude and longitude were acquired from GBIF. Different colors indicate different

photosynthetic types. *F.rob* (n=39), *F.cro* (n=48), *F. pri* (n=140), *F.son* (n=6), *F.ang* (n=50), *F.ano*

(n=145), *F.opp* (n=135), *F.pub* (n=25), *F.ram* (n=65), *F.chl* (n=96), *F.flo* (n=56), *F.lin* (n=799),

F.cam (n=95), *F.bro* (n=19), *F.bid* (n=525), *F.pal* (n=44), *F.tri* (n=1652), *F.vag* (n=8).

Fig 2 The environment conditions of the habitats for *Flaveria* species

(A) The average temperatures of the warmest quarter for different *Flaveria* species (above panel).

The precipitation of the wettest quarter for different *Flaveria* species (below panel). (B) The

average temperatures of the wettest quarter for *F. rob* and *F. pri*. (C) The precipitation of wettest

quarter for *F.rob* and *F.pri*. (D) The changes in vein density of C$_3$ and intermediate species.

Different colors indicate different photosynthetic types. Orange color represents C$_3$, grey color

represents intermediate species, green color represents C$_4$-like, blue color represents C$_4$ species.

Black lines represent median values. These environmental data are from 30 years. *F.rob* (n=39),

F.cro (n=48), *F. pri* (n=140), *F.son* (n=6), *F.ang* (n=50), *F.ano* (n=145), *F.opp* (n=135), *F.pub*

(n=25), *F.ram* (n=65), *F.chl* (n=96), *F.flo* (n=56), *F.lin* (n=799), *F.cam* (n=95), *F.bro* (n=19), *F.bid*

(n=525), *F.pal* (n=44), *F.tri* (n=1652), *F.vag* (n=8). For (A), the statistic test for the difference in

the environmental conditions was performed with One-way ANOVA, and the difference between

each group was performed by multiple Tukey test. For (B, C), the statistic test for the difference

between *F. rob* and *F. pri* was performed with Student t-test (Unpaired, Two-tails).

Fig 3 The association between precipitation and temperature with C$_4$ enzyme activities or C$_4$ flux

Here the precipitation denotes the precipitation of the wettest quarter. The temperature denotes the

average temperature of the wettest quarter. A-H, clade A. (A) The relationship between Log PEPC

activity per CHL and Log precipitation. (B) The relationship between Log PPDK activity per CHL

and Log precipitation. (C) The relationship between Log MDH activity per CHL and Log

precipitation. (D) The relationship between C$_4$ flux and precipitation. (E) The relationship between

Log PEPC activity per CHL and Log temperature. (F) The relationship between Log PPDK

activity per CHL and Log temperature. (G) The relationship between Log MDH activity per CHL

and Log temperature. (H) The relationship between C$_4$ flux and Log temperature. I-P, clade B. (I)

The relationship between Log PEPC activity per CHL and log precipitation. (J) The relationship

between Log PPDK activity per CHL and log precipitation. (K). The relationship between Log
MDH activity per CHL and log precipitation. (L). The relationship between C_4 flux and
precipitation. (M) The relationship between log PEPC activity per CHL and log temperature. (N)
The relationship between log PPDK activity per CHL and log temperature. (O) The relationship
between log MDH activity per CHL and log temperature. (P) The relationship between C_4 flux and
temperature. The results of linear regression analysis are in supplementary file.
491
Fig 4 Precipitation and temperature during the transition from proto-Kranz to C_4-like
(A) Change in precipitation of wettest quarter during the transition from proto-Kranz (*F. rob*) to
C_2 (*F. son*). (B) Change in precipitation of wettest quarter during the transition from C_2 (*F. son*) to
developing C_4 (*F. ano*) in clade B. (C) Change in precipitation of wettest quarter during the
transition from proto-Kranz (*F.son*) to developing C_4 (*F.ram*) in clade A. (D) Change in
precipitation of wettest quarter during the transition from developing C_4 (*F.ram*) to functionally
developed C_4 photosynthesis (*F.pal*) in clade A. (E) Change in precipitation of wettest quarter
during the transition from developing C_4 (*F.flo*)to functionally developed C_4 photosynthesis (*F.bro*)
in clade B. (F) Change in mean temperature of wettest quarter during the transition from
proto-Kranz (*F.rob*) to C_2 (*F.son*). (G) Change in mean temperature of wettest quarter during the
transition from C_2 (*F. son*) to developing C_4 (*F. ano*) in clade B. (H) Change in mean temperature
of wettest quarter during the transition from C_2 (*F. son*) to developing C_4 (*F. ram*) in clade A. (I)
Change in mean temperature of wettest quarter during the transition from developing C_4 (*F.ram*)
to functionally developed C_4 photosynthesis (*F. pal*) in clade A. (J) Change in mean temperature
of the wettest quarter during the transition from developing C_4 (*F. ano*) to functionally developed
C_4 photosynthesis (*F. bro*) in clade B. The transition from C_3 to C_2 and the emergence of C_4 flux
was promoted by increased temperature and decreased precipitation, indicated by the direction of
arrow. That the arrow become broad indicates the enhancement of C_4 flux. The statistic test for the
difference between each group was performed with Student t-test (Unpaired, Two-tails).
511
Fig 5 The association for stomatal density, CO_2 compensation point, precipitation and temperature
Here the precipitation denotes the precipitation of the wettest quarter. The temperature denotes the
average temperature of the wettest quarter. A-E, clade A. (A) The relationship between stomatal
density and precipitation. (B) The relationship between CO_2 compensation point and precipitation.
(C) The relation between stomatal density and CO_2 compensation point. (D) The relationship
between stomatal density and temperature. (E) The relationship between CO_2 compensation point
and temperature. F-J, clade B. (F) The relationship between stomatal density and precipitation. (G)
The relationship between CO_2 compensation point and precipitation. (H) The relationship between
stomatal density and CO_2 compensation point. (I) The relationship between stomatal density and
temperature. (J) The relationship between CO_2 compensation point and temperature. The data of
stomatal density was from (Zhao et al. 2022), and the data of CO_2 compensation point was from
(Ku et al. 1991). The average of stomatal density and the average of CO_2 compensation point were
used in this study. The results of linear regression analysis are in supplementary file.
525
Fig 6 Schematic diagram for the driving of C_4 evolution.
The schematic diagram for the conditions driving the evolutionary process for C_4 photosynthesis.
The raindrop number denotes the precipitation. The thermometer denotes the temperature. The

arrows denote the direction of C_4 evolution. The cycle composed by white and black denotes the
primary driving element that promote the corresponding stage of C_4 evolution. The primary
driving reason in C_3 and proto-Kranz was carbon limitation, and the primary driving reason in the
latter stage was water limitation. The black become narrow indicating the carbon limitation was
increasing gradually, and the white become narrow indicating the water limitation was increasing
gradually.
535
536 Supplementary fig 1. The C_4 flux for *Flaveria*.
537 The data of C_4 flux was from table 1 in (Vogan and Sage 2011)
538
Supplementary fig 2. Temperature and precipitation of habitat for *Flaveria* species in driest and
coldest period.
(A) Precipitation of the driest quarter. (B) Precipitation of the coldest quarter. (C) Precipitation of
the driest month. (D) Mean temperature of the driest quarter. (E) Mean temperature of coldest
quarter. (F) Min temperature of coldest month. Orange, C_3; Purple, Type I; Yellow, Type II; Green,
C_4-like; Blue, C_4. Black lines indicate median. The plants abbreviate genus written. *rob* (n=39),
cro (n=48), *pri* (n=140), *son* (n=6), *ang* (n=50), *ano* (n=145), *opp* (n=135), *pub* (n=25), *ram*
(n=65), *chl* (n=96), *flo* (n=56), *lin* (n=799), *cam* (n=95), *bro* (n=19), *bid* (n=525), *pal* (n=44), *tri*
(n=1652), *vag* (n=8).
548
Supplementary fig 3. Temperature and precipitation of habitat for *Flaveria* species in wettest and
warmest period.
(A) Max temperature of warmest month. (B) Precipitation of westest month. (C) Precipitation of
warmest quarter. (D) Mean temperature of wettest quarter. Orange indicates C_3, purple indicates
Type I, yellow indicates Type II, green indicates C_4-like, blue indicates C_4. Black lines indicate
median. The plants abbreviate genus written. *rob* (n=39), *cro* (n=48), *pri* (n=140), *son* (n=6), *ang*
(n=50), *ano* (n=145), *opp* (n=135), *pub* (n=25), *ram* (n=65), *chl* (n=96), *flo* (n=56), *lin* (n=799),
cam (n=95), *bro* (n=19), *bid* (n=525), *pal* (n=44), *tri* (n=1652), *vag* (n=8).
557
Supplementary fig 4. Annual temperature and precipitation of habitats for *Flaveria* species
(A) Annual mean temperature. (B) Annual precipitation. Orange, C_3; Purple, Type I; Yellow, Type
II; Green, C_4-like; Blue, C_4. Black lines indicate median. The plants abbreviate genus written. *rob*
(n=39), *cro* (n=48), *pri* (n=140), *son* (n=6), *ang* (n=50), *ano* (n=145), *opp* (n=135), *pub* (n=25),
ram (n=65), *chl* (n=96), *flo* (n=56), *lin* (n=799), *cam* (n=95), *bro* (n=19), *bid* (n=525), *pal* (n=44),
tri (n=1652), *vag* (n=8).
564
565 Supplementary fig 5. The trend in features for the evolution towards C_4 photosynthesis.
566 (A) Precipitation of wettest quarter. (B) Mean temperature of wettest quarter. (C) Vein density. (D)
567 Distance between vein. (E) O_2 inhibition. (F) CO_2 compensation point. (G) Maximum stomatal
568 conductance (G_{smax}) (H) Stomatal density (SD). The plants abbreviate genus written.
569
Supplementary fig 6. The difference in solar radiation and water vaper pressure between *F. pri* and
F. rob.
A, The difference in solar radiation between *F.pri* and *F.rob*. B, The difference in water vaper

pressure between *F.pri* and *F.rob*. *F. rob* (n=39). *F. pri* (n=140).

Table 1 The habitat for *Flaveria* species

Supplementary file 1: The statistics test and regression analysis. The plants abbreviate genus

written.

Acknowledgements

Thanks for the research group, and the postdoc support in shanghai. National Research and

Development Program of Ministry of Science and Technology of China (2020YFA0907600)

Author contribution

YYZ and ZGZ designed the study. YXZ and YYZ extracted the climate data and the latitude

and longitude data, and YYZ analyzed these climate data. XQY and YYZ mapped and marked the

coordinates of *Flaveria* genus location and do the visualization. YYZ extracted the physiological

data from the previous works. YYZ analyzed all these data, and did the statistic test and regression

analysis. YYZ plotted the graphs. YYZ and XGZ wrote the paper.

Competing interests

The authors declare no competing financial interests.

References

Adachi S, Stata M, Martin DG, Cheng S, Liu H, Zhu X-G, Sage RF (2023) The Evolution of C4

Photosynthesis in Flaveria (Asteraceae): Insights from the Flaveria linearis Complex. Plant

physiology 191 (1):233-251. doi:10.1093/plphys/kiac467

Akyildiz M, Gowik U, Engelmann S, Koczor M, Streubel M, Westhoff P (2007) Evolution and

function of a cis-regulatory module for mesophyll-specific gene expression in the C_4 dicot

Flaveria trinervia. Plant Cell 19 (11):3391-3402. doi:10.1105/tpc.107.053322

Brautigam A, Gowik U (2016) Photorespiration connects C3 and C4 photosynthesis. J Exp

Bot 67 (10):2953-2962. doi:10.1093/jxb/erw056

Bromham L, Bennett TH (2014) Salt tolerance evolves more frequently in C4 grass lineages.

Journal of Evolutionary Biology 27 (3):653-659. doi:10.1111/jeb.12320

Christin P-A, Osborne CP, Chatelet DS, Columbus JT, Besnard G, Hodkinson TR, Garrison LM,

Vorontsova MS, Edwards EJ (2012) Anatomical enablers and the evolution of C4

photosynthesis in grasses. Proceedings of the National Academy of Sciences 110

(4):1381-1386. doi:10.1073/pnas.1216777110

Christin PA, Besnard G, Samaritani E, Duvall MR, Hodkinson TR, Savolainen V, Salamin N

(2008) Oligocene CO2 decline promoted C4 photosynthesis in grasses. Curr Biol 18 (1):37-43.

doi:10.1016/j.cub.2007.11.058

Christin PA, Osborne CP, Sage RF, Arakaki M, Edwards EJ (2011) C(4) eudicots are not

younger than C(4) monocots. J Exp Bot 62 (9):3171-3181. doi:10.1093/jxb/err041

Concentrations MDiACD, Paleogene Dt <science.1110063.pdf>.

Edwards EJ, Smith SA (2010) Phylogenetic analyses reveal the shady history of C4 grasses.

Proc Natl Acad Sci U S A 107 (6):2532-2537. doi:10.1073/pnas.0909672107

Edwards EJ, Still CJ (2008) Climate, phylogeny and the ecological distribution of C4 grasses. Ecol Lett 11 (3):266-276. doi:10.1111/j.1461-0248.2007.01144.x

Edwards Ge, KU Msb (1987) Biochemistry of C3-C4 Intermediates. The Biochemistry of Plants, 10

Fick SE, Hijmans RJ (2017) WorldClim 2: new 1-km spatial resolution climate surfaces for global land areas. International Journal of Climatology 37 (12):4302-4315. doi:10.1002/joc.5086

Franks PJ, Drake PL, Beerling DJ (2009) Plasticity in maximum stomatal conductance constrained by negative correlation between stomatal size and density: an analysis using Eucalyptus globulus. Plant, cell & environment 32 (12):1737-1748. doi:10.1111/j.1365-3040.2009.002031.x

Gowik U, Bräutigam A, Weber KL, Weber APM, Westhoff P (2011) Evolution of C_4 Photosynthesis in the Genus Flaveria: How Many and Which Genes Does It Take to Make C_4? The Plant Cell 23 (6):2087-2105. doi:10.1105/tpc.111.086264

Heckmann D, Schulze S, Denton A, Gowik U, Westhoff P, Weber AP, Lercher MJ (2013) Predicting C4 photosynthesis evolution: modular, individually adaptive steps on a Mount Fuji fitness landscape. Cell 153 (7):1579-1588. doi:10.1016/j.cell.2013.04.058

Keerberg O, Parnik T, Ivanova H, Bassuner B, Bauwe H (2014) C_2 photosynthesis generates about 3-fold elevated leaf CO_2 levels in the C_3-C_4 intermediate species Flaveria pubescens. J Exp Bot 65 (13):3649-3656. doi:10.1093/jxb/eru239

Kocacinar F, McKown AD, Sage TL, Sage RF (2008) Photosynthetic pathway influences xylem structure and function in Flaveria (Asteraceae). Plant, cell & environment 31 (10):1363-1376. doi:10.1111/j.1365-3040.2008.01847.x

Ku MS, Wu J, Dai Z, Scott RA, Chu C, Edwards GE (1991) Photosynthetic and photorespiratory characteristics of flaveria species. Plant physiology 96 (2):518-528. doi:10.1104/pp.96.2.518

Kumpers BM, Burgess SJ, Reyna-Llorens I, Smith-Unna R, Boursnell C, Hibberd JM (2017) Shared characteristics underpinning C4 leaf maturation derived from analysis of multiple C3 and C4 species of Flaveria. J Exp Bot 68 (2):177-189. doi:10.1093/jxb/erw488

Langdale JA (2011) C4 cycles: past, present, and future research on C4 photosynthesis. Plant Cell 23 (11):3879-3892. doi:10.1105/tpc.111.092098

Lyu MJ, Gowik U, Kelly S, Covshoff S, Mallmann J, Westhoff P, Hibberd JM, Stata M, Sage RF, Lu H, Wei X, Wong GK, Zhu XG (2015) RNA-Seq based phylogeny recapitulates previous phylogeny of the genus Flaveria (Asteraceae) with some modifications. BMC Evol Biol 15:116. doi:10.1186/s12862-015-0399-9

Mallmann J, Heckmann D, Brautigam A, Lercher MJ, Weber AP, Westhoff P, Gowik U (2014) The role of photorespiration during the evolution of C_4 photosynthesis in the genus Flaveria. Elife 3:e02478. doi:10.7554/eLife.02478

Marshall DM, Muhaidat R, Brown NJ, Liu Z, Stanley S, Griffiths H, Sage RF, Hibberd JM (2007) Cleome, a genus closely related to Arabidopsis, contains species spanning a developmental progression from C(3) to C(4) photosynthesis. Plant J 51 (5):886-896. doi:10.1111/j.1365-313X.2007.03188.x

Mckown A, dengler N (2007) Key innovations in the evolution of Kranz anatomy and C4
vein pattern in Flaveria (Asteraceae). American Journal of Botany 94:382–399

Mckown AD (2005) Phylogeny of flaveria (Asteraceae) and inference of C4 photosynthesis
evolution. American Journal of Botany 92:1911–1928

Monson, William S. Schuster, Ku MSB (1987) Photosynthesis in Flaveria brownil A.M.
Powell. Plant physiology 85:1063-1067

Monson RK (2003) Gene duplication, neofunctionalization, and the evolution ofC4
photosynthesis. Int J Plant Sci

Nakamura N, Iwano M, Havaux M, Yokota A, Munekage YN (2013) Promotion of cyclic
electron transport around photosystem I during the evolution of NADP-malic enzyme-type C4
photosynthesis in the genus Flaveria. New Phytol 199 (3):832-842. doi:10.1111/nph.12296

Ogren DBJaWL (1984) The $CO_2\_O_2$ specificity of ribulose 1,5-bisphosphate carboxylase
oxygenase_ dependence on ribulosebisphosphate concentration, pH and temperature. planta
161:308-313

Pearcy RW, Ehleringer J (2006) Comparative ecophysiology of C3 and C4 plants. Plant, cell
& environment 7 (1):1-13. doi:10.1111/j.1365-3040.1984.tb01194.x

Powell AM (1978) Systematics of Flaveria (Flaveriinae--Asteraceae). Annals of the Missouri
Botanical Garden 65:590-636

R.K. Monson BdM, M.S.B. Ku and G.E. Edwards (1986) Co-function of C3- and
C4-photosynthetic pathways in C3, C4 and C3-C4 intermediate Flaveria species. Planta

Sage RF (2004) The evolution of C_4 photosynthesis. New Phytologist 161:341-270.
doi:10.1046/j.1469-8137.2004.00974.x

Sage RF (2017) A portrait of the C4 photosynthetic family on the 50th anniversary of its
discovery: species number, evolutionary lineages, and Hall of Fame. J Exp Bot 68
(2):4039-4056. doi:10.1093/jxb/erx005

Sage RF, Khoshravesh R, Sage TL (2014) From proto-Kranz to C4 Kranz: building the bridge
to C4 photosynthesis. J Exp Bot 65 (13):3341-3356. doi:10.1093/jxb/eru180

Sage RF, Monson RK, Ehleringer JR, Adachi S, Pearcy RW (2018) Some like it hot: the
physiological ecology of C4 plant evolution. Oecologia 187 (4):941-966.
doi:10.1007/s00442-018-4191-6

Sage RF, Sage TL, Kocacinar F (2012) Photorespiration and the evolution of C_4
photosynthesis. Annu Rev Plant Biol 63:19-47. doi:10.1146/annurev-arplant-042811-105511

Sage TL, Busch FA, Johnson DC, Friesen PC, Stinson CR, Stata M, Sultmanis S, Rahman BA,
Rawsthorne S, Sage RF (2013) Initial events during the evolution of C4 photosynthesis in C3
species of Flaveria. Plant physiology 163 (3):1266-1276. doi:10.1104/pp.113.221119

Schluter U, Weber AP (2016) The Road to C4 Photosynthesis: Evolution of a Complex Trait
via Intermediary States. Plant Cell Physiol 57 (5):881-889. doi:10.1093/pcp/pcw009

Schulze S, Mallmann J, Burscheidt J, Koczor M, Streubel M, Bauwe H, Gowik U, Westhoff P
(2013) Evolution of C4 photosynthesis in the genus flaveria: establishment of a
photorespiratory CO2 pump. Plant Cell 25 (7):2522-2535. doi:10.1105/tpc.113.114520

Sedelnikova OV, Hughes TE, Langdale JA (2018) Understanding the Genetic Basis of C_4
Kranz Anatomy with a View to Engineering C_3 Crops. Annu Rev Genet 52:249-270.
doi:10.1146/annurev-genet-120417-031217

Stata M, Sage TL, Sage RF (2019) Mind the gap: the evolutionary engagement of the C4

metabolic cycle in support of net carbon assimilation. Curr Opin Plant Biol 49:27-34.
doi:10.1016/j.pbi.2019.04.008

Still CJ, Berry JA, Collatz GJ, DeFries RS (2003) Global distribution of C3and C4vegetation:
Carbon cycle implications. Global Biogeochemical Cycles 17 (1):6-1-6-14.
doi:10.1029/2001gb001807

Taniguchi YY, Gowik U, Kinoshita Y, Kishizaki R, Ono N, Yokota A, Westhoff P, Munekage YN
(2021) Dynamic changes of genome sizes and gradual gain of cell-specific distribution of C4
enzymes during C4 evolution in genus Flaveria. Plant Genome:e20095.
doi:10.1002/tpg2.20095

Taylor SH, Franks PJ, Hulme SP, Spriggs E, Christin PA, Edwards EJ, Woodward FI, Osborne
CP (2012) Photosynthetic pathway and ecological adaptation explain stomatal trait diversity
amongst grasses. New Phytol 193 (2):387-396. doi:10.1111/j.1469-8137.2011.03935.x

Taylor SH, Hulme SP, Rees M, Ripley BS, Woodward FI, Osborne CP (2010) Ecophysiological
traits in C_3 and C_4 grasses: a phylogenetically controlled screening experiment. New Phytol
185 (3):780-791. doi:10.1111/j.1469-8137.2009.03102.x

Vogan PJ, Sage RF (2011) Water-use efficiency and nitrogen-use efficiency of C_3-C_4
intermediate species of Flaveria Juss. (Asteraceae). Plant, cell & environment 34
(9):1415-1430. doi:10.1111/j.1365-3040.2011.02340.x

Williams BP, Johnston IG, Covshoff S, Hibberd JM (2013) Phenotypic landscape inference
reveals multiple evolutionary paths to C_4 photosynthesis. elife 2:e00961.
doi:10.7554/eLife.00961

Zhao YY, Lyu MA, Miao F, Chen G, Zhu XG (2022) The evolution of stomatal traits along the
trajectory toward C4 photosynthesis. Plant physiology 190 (1):441-458.
doi:10.1093/plphys/kiac252

Zhou H, Helliker BR, Huber M, Dicks A, Akcay E (2018) C_4 photosynthesis and climate
through the lens of optimality. Proc Natl Acad Sci U S A 115 (47):12057-12062.
doi:10.1073/pnas.1718988115

Clade A

Clade B