Mixed evidence for species diversity affecting ecological forecasts in constant versus

declining light

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Key words: complexity, ecological forecasting, empirical dynamic modeling (EDM),

environmental change, machine learning, microcosm experiment, microorganisms, predictability

Running title: Ecological forecasts in changing environments

Data availability statement

All data and code used in this study will be made **publicly** available on Zenodo upon

publication.

Contributions

Conceptualization: U.D., R.L., O.L.P.; **Data curation**: U.D., R.M.K., R.L., O.L.P.; **Formal**

analysis: U.D., R.L.; **Funding acquisition**: O.L.P.; **Investigation**: Y.C., U.D., A.G., M.J., S.J.,

R.L., S.N., F.P., S.J.v.M, X.Z., D.Z-D.; **Methodology**: U.D., R.M.K., R.L., O.L.P; **Project**

administration: R.L., O.L.P.; **Resources**: O.L.P.; **Software**: U.D., R.M.K.; **Supervision**: R.L.,

O.L.P.; **Validation**: U.D., R.M.K., R.L., F.P., O.L.P.; **Visualization**: U.D.; **Writing – original**

draft: U.D., R.L.; **Writing – review & editing:** all authors.

Abstract

Accurate forecasts of ecological dynamics are critical for ecosystem management and conservation, yet the drivers of forecastability are poorly understood. Environmental change and diversity are considered major challenges to ecological forecasting. This assumption, however, has never been tested experimentally because forecasts have high data requirements. In a long-term microcosm experiment, we manipulated species richness of 30 experimental protist communities and exposed them to constant or gradually decreasing light levels. We collected finely-resolved time series (123 sampling dates over 41 weeks) of species abundances, community biomass, and oxygen concentrations. We then employed data-driven forecasting methods to forecast these variables. We found that species richness and light had a weak interactive effect on forecasts of species abundances: richness tended to reduce forecast accuracy in constant light but tended to increase forecast accuracy in declining light. These effects could partially be explained by differences among time series in variability and autocorrelation. Forecasts of aggregate properties (community biomass, oxygen), however, were unaffected by richness and light, and were not more accurate than those of species abundances. Our forecasts were based on time series that were detrended and standardized. Since real-world forecasting applications require predictions at the original scale of the forecasted variable, it is important to note that the results were qualitatively identical when back-transforming the forecasts to the original scale. Taken together, we found no strong evidence that higher diversity results in lower forecastability. Rather, our results imply that promoting diversity could make populations more predictable when environmental conditions change. From a conservation and management perspective, our findings suggest that diversity conservation might have beneficial effects on

68   decision-taking by increasing the forecastability of species abundances in changing

69   environments.

Introduction

Forecasting the future state of populations, communities, and ecosystems is challenging but necessary to make well-informed decisions in ecosystem conservation and management (Deyle *et al.* 2022; Dietze *et al.* 2024, 2018; Tulloch *et al.* 2020). Forecasts are quantitative predictions of the future (Dietze 2017), i.e. they are based on models fitted to data from the past and present. Forecasts are evaluated by measuring the difference between observed and predicted values, referred to as forecast error, while its inverse is termed forecast accuracy or forecastability. Ecological dynamics are difficult to forecast because organisms are embedded in complex networks of species interactions, respond simultaneously to many different drivers, often in a nonlinear way, and can display stochastic or chaotic dynamics (Beckage *et al.* 2011; Doak *et al.* 2008). However, global change and diversity loss have made ecological forecasting all the more relevant, resulting in rising numbers of published forecasts (Lewis *et al.* 2022). Typically, forecasts are based on observational data of organisms that are relevant in conservation and management (Lewis *et al.* 2022). Examples are forecasts of species that are harvested or hunted (Deyle *et al.* 2013; Henden *et al.* 2020), of pest outbreaks (Piou & Marescot 2023), or of harmful algal blooms (Rousso *et al.* 2020). Likewise, observational data has been used to compare forecast accuracy across different species and ecosystem properties (Lewis *et al.* 2022; Ward *et al.* 2014), in an attempt to gain a more general understanding about the constraints of forecastability.

Only few studies, however, investigated potential drivers of forecastability with experimental approaches (Benincà *et al.* 2008; Daugaard *et al.* 2022; Dumandan *et al.* 2024; Fujita *et al.* 2023). Forecasts of ecological dynamics benefit from long and finely-resolved time series (Dietze 2017), which can be challenging to accomplish with experiments. Yet, controlled

experiments have been a fundamental tool in many fields of ecology (Jessup *et al.* 2004; Jochum

et al. 2020; Petchey *et al.* 1999) and could be useful to advance our understanding about the

potential and the limits of ecological forecasting.

One potential driver of forecastability is the diversity (i.e. species richness) of communities.

This idea, however, has never been tested with an experimental manipulation of diversity.

Previous work focused on comparing forecast accuracy among species within a community

(Daugaard *et al.* 2022), rather than among communities with different properties (e.g. diversity).

The diversity of a community could have both negative and positive effects on the forecastability

of the species embedded in the community, as well as on aggregate ecosystem properties (e.g.

community biomass). First, diversity could reduce forecastability by making the species

interactions network more complex: communities with higher species richness have the potential

for more direct and indirect interactions among species (Polis & Strong 1996; Yodzis 2000),

making the outcome of species interactions less predictable (Montoya *et al.* 2009; Novak *et al.*

2011). Second, diversity could affect forecastability by influencing the temporal variability of

populations and aggregate ecosystem properties. Higher diversity can both increase or reduce the

temporal variability of populations (Xu *et al.* 2021), but usually reduces the temporal variability

of aggregate ecosystem properties (Xu *et al.* 2021). The dampening effect of diversity on the

temporal variability of aggregate properties could be particularly pronounced when

environmental conditions change (Yachi & Loreau 1999), because diversity can buffer aggregate

ecosystem properties against environmental changes (Baert *et al.* 2016; Isbell *et al.* 2015), and

delay abrupt shifts to new ecosystem states (Limberger *et al.* 2023). Provided that lower

variability results in higher forecastability, higher diversity could therefore increase the

forecastability of community biomass (in particular when the environment changes), but reduce

or increase the forecastability of species abundances. In general, aggregate properties are

expected to be easier to forecast than species abundances because they might integrate over

environmental variability (Lewis *et al.* 2023).

When environmental conditions change, ecological forecasting is particularly relevant but

could also be particularly difficult. Of particular importance in a management context are near-

term forecasts (Dietze *et al.* 2024, 2018), that is, forecasts of near-term variability rather than of

long-term trends. Directional environmental change might alter such near-term variability if

different environmental conditions lead to different population dynamics (Fussmann *et al.* 2014).

Previous work showed that random environmental fluctuations can reduce forecastability

(Daugaard *et al.* 2022) but we do not know if the same applies when the environment changes

directionally to novel conditions, as is characteristic of many global change pressures (Tilman *et*

al. 2001). Data-driven forecasting methods (e.g. machine learning) perform better when past

time points hold more information about the future (Pennekamp *et al.* 2019). Population

dynamics can differ among environmental conditions (Fussmann *et al.* 2000, 2014), and changes

to novel environmental conditions could therefore reduce forecastability due to greater

dissimilarity of past and future dynamics. In addition, abiotic change often alters species

interactions (Blois *et al.* 2013; Gilman *et al.* 2010), which could further constrain forecastability

(Dumandan *et al.* 2024). On the other hand, environmental change can reduce stochasticity

(Blasius *et al.* 2020) and increase autocorrelation of population time series (Drake & Griffen

2010; Pace *et al.* 2017). Consequently, future states of such systems depend more on past states,

and should thus be more predictable (Pennekamp *et al.* 2019). Despite the importance of

ecological forecasting in changing environments, we lack empirical studies that test how

directional change to novel environmental conditions influences forecastability.

Here we investigated how diversity and environmental change affect the forecastability

of ecological dynamics in a controlled and simplified system. We conducted a microcosm

experiment with a unique combination of high numbers of time points (123), experimental units

(30), and experimental taxa (18). We used aquatic protists as model organisms because they have

short generation times (~ 0.5-3 generations per day under ideal growth conditions (Leary &

Petchey 2009; Tabi *et al.* 2019)), which allows to collect time series of population dynamics

within a relatively short time frame. We assembled communities with three different levels of

species richness and exposed the communities to constantly high light levels or to light that

declined gradually and then remained constantly at low levels. We collected time series of

species abundances and aggregate ecosystem properties (community biomass and oxygen) and

used data-driven forecasting models to forecast these variables. With this dataset we tested four

hypotheses (Supplement Figure S2 a, b): (1) higher diversity leads to higher forecast error of

species abundances (Hypothesis 1, H1), (2) historical environmental change to novel conditions

increases forecast error of species abundances (H2), (3) forecast error of aggregate ecosystem

properties decreases with diversity, in particular in changing environments (H3), and (4) forecast

error of aggregate ecosystem properties is lower than forecast error of species abundances (H4).

Our experimental communities and treatments are simplified versions of real-world

scenarios. Our manipulation of light is meant to represent anthropogenic pressures that are

changing gradually and directionally (Tilman *et al.* 2001). We chose light because it is relevant

in a global change context (Leech *et al.* 2018; Winder & Sommer 2012) but also because we

expected that light would affect the growth of many species in the community, in particular that

of primary producers (algae), but indirectly also that of consumers. However, we did not intend

to mimic the complex characteristics of human-induced light regime changes in natural aquatic

systems: these changes include both increases in light availability (due to prolonged lake

stratification (Winder & Sommer 2012)) and decreases (e.g. due to the browning of lakes (Leech

et al. 2018)), and they can be associated with concurrent changes in other factors (e.g. browning

resulting from increased input of dissolved organic matter). Likewise, our manipulation of

species richness (maximum: 14 species) is a simplified (i.e. shorter) version of diversity

gradients observed in natural systems. To allow high-frequent samplings, the size of our species

pool was limited because we only used species that were distinguishable with automatic

classification. Although our communities are simpler than their natural counterparts, they

represent a model of an important community type, that is, of a food web dominated by

consumer-resource interactions. With this simple and tractable system we aimed to provide first

experimental insights into the effects of diversity and environmental change on ecological

forecasts.

Methods

Experimental design

In a nine-month experiment, we manipulated light (2 levels) and species richness (3 levels) in a

full factorial design. At each of the three richness levels, we assembled five different community

compositions. The composition × light combinations were not replicated, i.e. from each of the 15

compositions we set up two microcosms, one for each of the two light levels, resulting in a total

of 30 microcosms. Light intensity was either constantly high or declined gradually. To

manipulate light, we used eight incubators (IPP260plus, Memmert), four per light level. The

declining light treatment involved three phases: (1) three months of constantly high light (30% of

the maximum incubator light intensity, ~ 22,500 Lux), (2) three months of gradual light decline

(from 30% to 1%, ~ 800 Lux), and (3) three months at the final low light level (Supplement

Figure S3). Throughout the entire experiment, all incubators were set to a light:dark cycle of

16:8 hours and a temperature of 18°C.

We manipulated species diversity by assembling protist communities with different

numbers of species (7, 10, and 14 species, respectively). We used a pool of 18 species of algae

and ciliates, comprising six functional groups: algae which were edible for ciliates (two species),

inedible algae (5), bacterivorous ciliates (4), omnivorous ciliates (4), mixotrophic ciliates (2),

and a predatory ciliate (1) (Supplement Table S1, Figure S1). The three diversity levels were

generated by manipulating diversity within the functional groups of inedible algae, bacterivores,

and omnivores (one, two, or three species per functional group), and in the mixotrophs (one, one,

or two species) (Supplement Table S2). To avoid confounding effects of species richness and

species identity/composition (Huston 1997; Schmid *et al.* 2017), we assembled five different

compositions at each diversity level. The compositions were as dissimilar as possible, and

species occurred with similar frequency both within and across diversity levels (see Supplement

Section S1.3 for computation of community compositions). Each of the 15 compositions

comprised all six functional groups.

Implementation of the experiment

Over a period of four weeks, we inoculated 30 2L-bottles from stock cultures growing at high

densities. Each bottle was filled with 1 L of medium (75% filtered protist pellet medium, i.e.

0.55 g protist pellet in 1L Chalkley's medium, and 25% WC medium), 20 wheat seeds for slow

release of nutrients, and a magnetic stirrer for homogenization during samplings. Diversity was

manipulated in a substitutive design, e.g. the culture volume added of inedible algae was divided

among one to three species depending on the diversity level. During inoculation, we

homogenized bacteria associated with algae and ciliate cultures so that communities did not

differ in their initial bacterial composition. See Supplement Section S1.5 for further details on

inoculation.

After the inoculation phase, we sampled the 30 bottles three times per week (Monday,

Wednesday, Friday) for nine months (41 weeks), resulting in 123 sampling days. On sampling

days, we removed 50 mL from each bottle after homogenization on a magnetic stirrer and then

added 50 mL of fresh medium to the bottle. Samples were taken under sterile conditions using

50-mL glass pipettes.

To allow species to re-colonize bottles if they had gone extinct, we added the species from

the stock cultures every three weeks, mimicking a very low rate of immigration. In protist

microcosms, species often go extinct through competitive or trophic interactions and would have

no chance of re-colonizing the microcosm when conditions become favourable. At these

immigration events, we added a very low number (10 individuals) of each species that was part

of the community of a given bottle. Soon after the start of the experiment, two species went

extinct in the stock cultures (*Stylonychia mytilus* and *Didinium nasutum*) and could therefore not

be re-immigrated. Stock cultures were cultivated in high light conditions (i.e. 30% light).

Quantification of species abundances and ecosystem properties

We measured abiotic ecosystem properties (dissolved oxygen, carbon, and nitrogen), species

abundances of ciliates and algae, and total abundance of bacteria. To quantify dissolved oxygen,

we used a hand-held oxygen meter and two optical oxygen sensors attached to the inner wall of

each bottle. Oxygen was measured in the morning prior to sampling. The light:dark cycle of the

incubators was set such that the oxygen measurements would be made towards the end of the 16-

hour light phase, specifically after 13.5 hours of light. We measured dissolved organic carbon

(DOC) and dissolved nitrogen with a TOC/TN analyzer, bacterial abundance with a

flowcytometer, algae abundances and morphological traits with a FlowCAM, and ciliate

abundances and movement and morphological traits with video microscopy (R-package *bemovi*

(Pennekamp *et al.* 2015)). The predatory ciliate *Didinium* was counted manually under a

dissecting microscope when present. See Supplement Section S2 for details on measurements

and instruments used.

We identified the recorded algae and ciliate individuals by using support vector machine

(SVM) classifiers (R-package e1071 (Meyer *et al.* 2021)). We trained these classifiers with trait

data collected from monocultures cultivated at different light levels and, if necessary, with

manually annotated data from the experiment. The trait data to train the classifiers was collected

with the same methods used in the experiment, i.e. video microscopy for ciliate traits and

FlowCAM for algae traits. See Supplement Section S2.6 for further details on classifications.

Lastly, we computed community biomass by summing the biomass of all algae, ciliates, and

bacteria in a sample. To this end, we estimated the volume of each particle using its extracted

traits and three-dimensional shape. Further details are given in Supplement Section S4.3.

Processing of recorded time series

After data collection and classification, we carried out several quality controls and made

corrections if needed (for a complete description see Supplement Section S3). Then, we averaged

over technical replicates of density measurements made with flowcytometry and video

microscopy, respectively, and of oxygen concentration measurements made with the oxygen

meter.

The recorded data included some missing data (flowCAM: 0.81%, flowcytometer: 4.84%,

videos: 0.84%, oxygen meter: 1.67%, TOC/TN analyzer: 0.92%, Supplement Table S5). We

used cubic hermite splines to impute the missing data and to interpolate the time series such that

all time points were equidistant (time step of 2.3 days). To make the time series stationary, we

detrended and standardized them, as is commonly done in time series analyses (Benincà *et al.*

2008) by taking the standardized residuals of the regressions of the recorded values against time.

That is, the purpose of our forecasts was to predict the variation around a trend, not the trend

itself. Since forecasts of detrended time series can be back-transformed to the original scale of

the data, detrending does not preclude the use of forecasts when the actual values are needed

(e.g. in conservation and management). To test the robustness of our results, we also evaluated

the forecasts after back-transforming them to the original scale of the forecasted variables

(Supplement Section S7.5).

We detrended and standardized the entire time series, i.e. the test data was included in the

processing of the time series. This approach allows for the best possible time series

transformation and is therefore often used in theoretical studies that compare forecast accuracy

among treatments or species (Benincà *et al.* 2008; Daugaard *et al.* 2022; Munch *et al.* 2023). In

real-world settings, however, the test data is not available prior to making forecasts. We

therefore checked if our results changed when using only the historic part of the time series (i.e.

the 111 time points of the training dataset) to detrend and standardize the time series

(Supplement Section S7.4).

The untransformed and transformed time series are displayed in Supplement Figures S7-S14.

Note that the log-transformation in figures of untransformed time series (Figures S7-S9) is for

better visibility of the time series. However, the data was not log-transformed prior to

detrending.

Forecasting

We forecasted species abundances and two aggregate ecosystem properties (total community

biomass and oxygen concentration). The forecasted species (i.e. forecast targets) included the

ciliate and algae species, bacteria (considered one group), and additional morphotypes

("*Chlamydomonas* clumps", "*Desmodesmus* clumps", "Dividing *Chlamydomonas*" and "Small

cells", i.e. small unidentified cells), 14 targets in total. However, we did not forecast the

abundance of a target if it went extinct during the experiment (Supplement Section S3.2). Across

all bottles, we forecasted 253 taxa time series.

We used different forecast methods to ensure that the results were independent of the chosen

methodology. We used one linear model (Auto-Regressive Integrated Moving Average, i.e.

ARIMA), two nonlinear approaches based on Empirical Dynamic Modeling (Simplex EDM and

Multiview EDM (Sugihara & May 1990; Ye & Sugihara 2016)), and two machine learning

methods (Random Forest RF and Recurrent Neural Networks RNN). Simplex and Multiview

EDM differ insofar as the former allows better parameter optimization, while the latter is

designed for high-dimensional systems. See Supplement Section S5 for a more detailed

description of the forecast models and their parametrization.

We used the first 111 time points of the time series to train the models and then forecasted

the remaining 12 time points (i.e. ~10% of the data, corresponding to the last 4 weeks of

sampling) with one-step-ahead forecasts. Two of the forecast methods (ARIMA, Simplex EDM)

included only the forecast target itself as predictor, whereas the other three methods included

additional predictors. As predictors we used all forecast targets of a given bottle as well as the

dissolved carbon and nitrogen concentrations. The predictor data did not include the forecasted

time step. We quantified the forecast error (i.e. the disagreement between forecasted and

recorded values) as the root mean square error (RMSE). Because we forecasted standardized

time series, an RMSE value below one indicates a better performing model than the average

value of the time series (which has an expected RMSE of one).

Analyzing the effects of species richness and light on forecast error

To analyze the effects of the experimental treatments on forecast error, we computed linear

mixed models (R-package *lme4* (Bates *et al.* 2015)), following common practice in similarly

designed experiments (Pennekamp *et al.* 2018; Steiner *et al.* 2006). In separate models, we used

forecast error of either taxa abundances, community biomass, or oxygen as response variable. As

explanatory variables, we used the experimental treatments, i.e. light condition (binary), species

richness (continuous), and their interaction. We used type III ANOVA to assess the importance

of covariates and interactions. If the interaction was significant or marginally significant (i.e. *p*-

values below 0.1), we kept the interaction in the model and used type III ANOVA. Otherwise,

we removed the interaction from the model and used type II ANOVA to investigate the

importance of the main fixed effects. In the case of a significant interaction, we analyzed the

effect of species richness with a separate linear mixed model at each of the two light levels

(Quinn & Keough 2024), and the effect of light at the lowest and highest richness level with a

post-hoc analysis that estimated the marginal means.

Observed species richness deviated from planned richness because not all species

persisted throughout the experiment (Supplement Section S3.1). Furthermore, in one bottle the

alga *Desmodesmus* was introduced by cross-contamination. We therefore used realized richness

(i.e. the median of the observed richness values) rather than planned richness in our models.

Realized species richness was unaffected by the light conditions and was positively correlated

with planned species richness (for more information and alternative realized species richness

metrics see Supplement Section S4.1). We also evaluated if forecast error was related to further

diversity indices (Supplement Section S9).

In all models, we included random intercepts following the experimental design. That is,

models to investigate treatment effects on the forecast error of taxa abundances included random

intercepts for taxon, bottle, and incubator, whereas models to investigate effects on the forecast

error of aggregate properties included random intercepts for composition and incubator.

We also investigated if treatment effects depended on the level of aggregation of the

response variable by including the mean forecast error of taxa abundances and the forecast error

of community biomass in the response variable. As explanatory variables, we used realized

richness, light condition, a binary variable that specified the level of aggregation (taxa

abundances vs. community biomass), and all possible pairwise interaction terms. Bottle and

incubator were included as random effects. We assessed the importance of model covariates with

type III or type II ANOVA depending on the *p*-value of the interaction, as described above. We

repeated this model for the forecast error of oxygen instead of community biomass.

We repeated the analyses for each of the five forecasting methods. As treatment effects

on forecast error were similar among the five approaches, we chose one method (Simplex EDM)

to describe the treatment effects in the main text and refer to the supplementary material for

results regarding the other four methods.

Explaining the treatment effects on forecast error

To explore possible mechanisms behind observed treatment effects on forecast error, we

investigated how forecast error was related with time series properties. For each time series, we

calculated three metrics: the coefficient of variation, autocorrelation, and permutation entropy.

The coefficient of variation is a measure of temporal variability and was calculated as the ratio of

the standard deviation of the detrended time series and the mean of the original time series.

Autocorrelation describes the correlation of current and past values and was calculated for a lag

of 5 (i.e. correlation with values 5 time points ago); the choice of lag did not affect the results.

Permutation entropy is a measure of time series complexity, with higher permutation entropy

usually being associated with higher forecast error (Pennekamp *et al.* 2019). We calculated

autocorrelation and permutation entropy based on the transformed time series. See Supplement

Section S4.2 for further details.

We used two approaches to investigate the relationship of the time series metrics with the

experimental treatments and with forecast error of species abundances. First, we computed

separate linear mixed models to test (i) how the experimental treatments (realized richness, light,

and their interaction) affected each of the three time series metrics, and (ii) how each of the three

metrics, light condition, and their interaction affected forecast error. Again, we included random

intercepts for taxon, bottle, and incubator in the models, and assessed the importance of model

covariates with type III or type II ANOVA as described above. As a second approach, we

computed a structural equation model (SEM) to further clarify the relations found with the first

approach, using the R-packages *lavaan* (Rosseel 2012) and *lavaan.survey* (Oberski 2014).

Lavaan.survey allowed us to include random effects according to our experimental design. We

set up the model to estimate the relations between the variables in both light conditions by using

a multiple group analysis, and tested the significance of SEM path coefficients with two-sided t-

tests. All data analyses and forecasts were done in R (version 4.3, package information in

Supplement Section S10).

Results

Effects of species richness and light on forecast error

Richness and light had a weak interactive effect on the forecast error of taxa abundances

(richness x light: p-value= 0.0363; Figure 1a, Table 1, forecast method: Simplex EDM). The

statistical support for this effect was somewhat limited (Figure 1a: overlapping confidence

intervals), potentially due to high amounts of variance in the data. However, the interactive effect

was consistent among forecast method, detrending approach, and scale of forecast error

calculation (see below). The interaction of the two treatments resulted from opposite effects of

species richness in constant versus declining light: forecast error tended to increase with species

richness in constant light but tended to decrease with richness in declining light (Figure 1a).

Specifically, when median richness increased by one species, forecast error increased by 5.6%

(95% CI: -0.8% − 12.1%) in constant light and decreased by 4.6% (95% CI: -11.0% − 1.0%) in

declining light relative to the baseline forecast error of 1 (i.e. the expected forecast error when

the average value of a time series is used as the prediction). Note that the 95% confidence

intervals of the estimates include zero despite the interaction term p-value < 0.05. A discrepancy

such as this can occur when the statistical evidence of an effect is not very strong (e.g., p-value =

0.0363) and when different methods are used to calculate p-values and confidence intervals.

Specifically, approximate p-values were calculated with Wald tests, whereas confidence intervals

were calculated with profile likelihoods due to the greater precision of this approach.

Separate analyses at constant and declining light conditions also indicated opposite

richness effects on taxa abundance forecasts in the two light conditions, as the slopes of the two

relationships had opposite signs and neither of their 95% confidence intervals overlapped with

zero (Supplement Table S7). Furthermore, we investigated the effect of the light decline on the

forecast error of taxa abundances at the low and high end of the diversity gradient. The light

decline reduced forecast error at high species richness, but had no effect at low species richness

(tested with a post-hoc analysis; Supplement Table S9).

The interactive effect of richness and light on the forecast error of taxa abundances

persisted in several robustness analyses (Supplement Section 7). Both the direction and the size

of the treatment effects were similar among the five forecasting methods (Supplement Table S8,

Figure S18, S21). Likewise, treatment effects were similar among richness variables (including

planned richness) and detrending method and were not driven by individual species (Supplement

Figures S21-S24). Furthermore, treatment effects were similar as above when using only the

historical data to detrend and standardize the time series (Supplement Figure S26) and when

evaluating the forecasts after back-transforming the forecasts to the original scale (Supplement

Figure S28).

Forecastability of aggregate ecosystem properties was largely unaffected by the

experimental treatments. Light and richness had no effect on the forecast error of total

community biomass (Table 1, Figure 1b) except when biomass was forecasted with RNN

(Supplement Table S10, Supplement Figure S19). In this case, treatments had similar effects as

on forecast error of taxa abundances. Forecast error of oxygen was unaffected by light and

richness (Supplement Table S11, Supplement Figure S20). For most forecasting methods,

forecast error of aggregate properties (community biomass, oxygen) did not differ from average

forecast error of species abundances (Supplement Tables S12-S14).

Explanations of the treatment effects on forecast error

To investigate possible reasons for differences in forecast error, we analyzed how the forecast

error of species abundances was related with three time series properties: autocorrelation,

coefficient of variation (as a measure of temporal variability), and permutation entropy (as a

measure of time series complexity (Pennekamp *et al.* 2019)). We expected that greater

autocorrelation and lower permutation entropy would be associated with lower forecast error,

while greater variability (i.e. higher coefficient of variation) could be associated with lower or

higher forecast error, depending on whether higher variability is associated with more or less

determinism (Supplement Figure S2 c-e).

Forecast error decreased with increasing autocorrelation and coefficient of variation, and

increased with increasing permutation entropy (Figure 2 a-c, Supplement Table S15). The

negative relationship between forecast error and the coefficient of variation suggests that higher

temporal variability reflected higher deterministic variability (rather than higher stochastic

variability). The experimental treatments affected the three time series metrics, albeit not in an

interactive way: autocorrelation and coefficient of variation were lower in declining light than in

constant light, whereas permutation entropy was unaffected by light but increased with richness

(Figure 2 d-f, Supplement Table S16).

We used a structural equation model (SEM) to disentangle direct and indirect effects of

richness and time series metrics on forecast error of taxa abundances (Figure 3, Supplement

Table S17). A multi-group model provided a good match between model-implied and observed

covariance ($\chi^2 = 3.219$, DF = 4, $p = 0.522$, i.e., no evidence against a good model fit). The

model explained 32.2% of the variation in forecast error in constant light, and 9.7% of the

variation in forecast error in declining light (R^2, Table S18). In line with the bivariate correlation

analyses, the SEM showed in both light conditions that forecast error declined with increasing

autocorrelation, declined with an increasing coefficient of variation, and increased with

increasing permutation entropy (Figure 3; note that not all of these effects were statistically

significant).

Similarly as the linear mixed model analysis, the SEM showed that the effect of richness

on forecast error of taxa abundances depended on the light treatment. Richness had a positive

(albeit not significant) effect on forecast error in constant light but significantly reduced forecast

error in decreasing light (Figure 3, Supplement Table S17). Furthermore, the effect of species

richness on the coefficient of variation and on autocorrelation was reversed between the two light

treatments (Figure 3). Although not significant, these reversed effects of richness on coefficient

of variation and autocorrelation in the two light treatments might explain why increasing

richness had opposite effects on forecast error in the two light treatments.

Discussion

Across the diversity gradient used in this experiment, diversity had only weak effects on the

forecastability of species abundances. Diversity tended to increase forecastability (i.e. reduce

forecast error) of species abundances in declining light but tended to reduce forecastability in

constant light. These effects, although weak, were consistent among different forecast methods,

detrending approaches, and scales of forecast error calculation. In summary, we found only weak

support for the hypothesis that higher diversity makes population dynamics hard to forecast (H1),

and this (weak) negative effect of diversity on forecastability was only observed in constant

light. Our hypothesis that environmental change reduces forecastability (H2) was not supported.

On the contrary, light decline increased forecastability when richness was high but had no

significant effect when richness was low. Forecastability of aggregate properties (community

biomass, oxygen) was unaffected by our experimental treatments (H3) and was not higher than

that of species abundances (H4). Taken together, we found no strong evidence that higher

diversity leads to low forecastability, which could be partly due to the comparatively short

diversity gradient. However, our results suggest preliminary support for higher diversity

increasing the forecastability of species abundances in changing environments. Diversity effects

on forecastability could be stronger across broader diversity gradients, which calls for further

research in more complex systems with larger diversity gradients to explore the generality of our

findings.

Light decline increased the forecastability of species abundances at the upper end of the

diversity gradient. This positive effect of light decline on forecastability potentially resulted from

higher autocorrelation in changing conditions (Figure 2d). Higher correlation between successive

time points means that past states have higher information content, resulting in higher accuracy

of forecasts (Ward *et al.* 2014). Accordingly, higher autocorrelation was associated with lower

forecast error (Figure 2a, Figure 3). Higher autocorrelation in changing conditions was due to

both direct and indirect effects of the environmental driver. Decreasing light put some species on

deterministic trajectories of population decline, which indirectly led to abundance increases of

inferior competitors and prey organisms (Supplement Figures S29, S30). Such indirect effects of

environmental change via altered species interactions are widespread (Amundrud & Srivastava

2016; Bell *et al.* 2019; Martin & Maron 2012), and may be more frequent than direct effects

(Ockendon *et al.* 2014). As such, autocorrelation can spread through the community even if only

few species are directly affected by environmental change. Taken together, our results illustrate

that forecastability of species abundances can be high in changing environments if direct and

indirect effects of environmental change result in higher autocorrelation of time series, and

therefore in greater information content of past states.

In the declining light treatment, forecastability of species abundances tended to decrease as

diversity decreased. We can only speculate about a possible mechanism behind this pattern.

Possibly, high forecast error at the low end of the diversity gradient might have resulted from

declining light leading to previously unobserved dynamics. Specifically, in the two communities

with lowest diversity, an entire functional group (consumers of algae, i.e. omnivores +

mixotrophs) was represented by only one species, which declined or suddenly increased in

response to the environmental driver, resulting in pronounced changes in prey abundances

(Supplement Section S8.1). In more diverse communities, however, the consumer functional

group thrived across the entire light gradient and did not change suddenly as conditions changed.

For example, the decline of the dominant consumer in the decreasing light treatment was

compensated by increases in other consumer species (Supplement Section S8.1). This process is

known as the insurance effect of diversity (Loreau *et al.* 2021; Yachi & Loreau 1999), i.e. the

loss of species due to environmental change is compensated by other species that perform similar

functions but tolerate and potentially thrive in the new environmental conditions (Bestion *et al.*

2020; Hong *et al.* 2022; Isbell *et al.* 2015). This mechanism might have contributed to the

positive effect of diversity on forecastability in the changing light treatment by maintaining

functional groups in the community, thereby preventing pronounced changes in population

dynamics.

The statistical support for the treatment effects was somewhat limited, possibly reflecting

high variance in the data. Having more replicates of each richness × composition × light

combination would be useful in future research to gain stronger statistical support for treatment

effects. In the case of the light treatment, high variance might have resulted from counteracting

effects of light on forecastability. As discussed above, it seems that environmental change had

contrasting effects on forecastability, that is, light decline increased forecastability by increasing

autocorrelation but reduced forecastability by leading to previously unobserved dynamics. The

first of these effects (higher autocorrelation) dominated when diversity was high, whereas the

second effect (change in dynamics) dominated when diversity was low. Still, however, both

effects likely occurred along the entire diversity gradient, with the importance of these effects

likely differing among species and compositions. These contrasting effects of environmental

change on forecastability might have contributed to the high variation in the data.

The limited statistical support for the diversity effect on taxa abundance forecasts could be

partly due to the rather short diversity gradient. To enable high-frequent samplings, we only used

species that were distinguishable by automatic classification, which limited the size of the

species pool. This limitation is typical for studies on ecological forecasting: the high number of

required time points constrains studies to focusing either on comparatively few species

(Daugaard *et al.* 2022; Dumandan *et al.* 2024) or on few experimental units (Benincà *et al.*

2008). Despite this limitation, our diversity gradient was in the range of previous studies that

used experimental protist communities to draw important conclusions about the effect of

diversity on the stability of communities (Pennekamp et al. 2018; Steiner et al. 2005). In contrast

to stability, however, forecastability is associated with more sources of error resulting from the

training of forecast models. Consequently, it is to be expected that variance in the data is larger

for forecastability than for response variables such as stability or species abundances.

Although diversity explained only a part of the variation in forecastability of taxa

abundances (Supplement Section S6.1), our results nevertheless indicate that diversity tends to

increase forecastability in changing environments. From an ecosystem management perspective,

our results therefore suggest that diversity conservation could have a positive effect on the

accuracy of population forecasts in changing environments. More-diverse communities might be

less prone to ecological surprises as environmental conditions change because diversity can

promote the resistance of functional groups (Baert *et al.* 2016; Isbell *et al.* 2015). As implied by

our findings (see discussion above and Supplement Section S8.1), the loss or rise of functional

groups can have large impacts if it concerns species with strong ecosystem-effects. For example,

the decline of keystone predators or consumers can have cascading effects on other groups

(Amundrud & Srivastava 2016; Estes *et al.* 2011), potentially leading to novel dynamics of many

species in the community, and therefore to overall low forecastability. Such species with strong

ecosystem-effects might thus be of particular concern in conservation and management, not only

because of their effects on ecosystem functioning but potentially also on population forecasts. To

confirm that our findings are transferable to a management context, we conducted robustness

analyses which showed that treatment effects on forecastability were similar when we back-

transformed the forecasts to the original scale of the time series (Supplement Section 7.5). Taken

together, diversity maintenance might reduce ecological surprises in changing environments,

thereby increasing the forecastability of species abundances.

In contrast to our hypothesis, the forecastability of aggregate properties (i.e. community

biomass, oxygen) was largely unaffected by our experimental treatments. Trends of treatment

effects went in the same rather than opposite direction as effects on forecastability of species

abundances. We had expected that contrasting effects of diversity on the temporal variability of

aggregate and non-aggregate properties (Tilman 1996; Xu *et al.* 2021) may translate into

contrasting effects on forecastability. However, in our experiment, species richness affected

neither the temporal variability of population abundances (Figure 2e) nor that of community

biomass or oxygen (Supplement Figure S31). In general, the experimental treatments had similar

effects on time series properties irrespective of the level of aggregation of the response variable,

possibly explaining the similarity in treatment effects on forecastability. Overall, our results

suggest that diversity and environmental change are less influential in affecting the

forecastability of aggregate ecosystem properties.

Also in contrast to our expectations, forecastability of community biomass did not differ

from average forecastability of species abundances (Supplement Table S12). Forecastability is

expected to increase with the level of aggregation because aggregate properties might integrate

over environmental variability (Lewis *et al.* 2023). Indeed, community biomass had lower

temporal variability than the biomass of individual species (Supplement Figure S32). However,

our results suggests that lower temporal variability does not necessarily translate into higher

forecastability. Rather, we observed the opposite pattern: time series with higher variability were

forecasted better, indicating that higher temporal variability did not reflect more stochastic but

more deterministic variation (e.g. abundance changes in response to the light decline or to

changes in competitor abundances). At the level of the community, compensatory dynamics

among species might have led to reduced variability of community biomass without increasing

its forecastability because biomass integrated over primarily deterministic (not stochastic)

variation. In conclusion, our results do not support the prospect that forecastability increases by

aggregating individual ecosystem components.

Our study illustrates that experiments with microbial organisms are a useful tool to

investigate potential drivers of forecastability. Data-driven forecasting methods require long time

series (Dietze 2017), which can be collected within a comparatively short time frame for

microbial organisms because of their short generation times. Our experimental system is a model

of a food web dominated by consumer-resource interactions. Investigating how diversity and

environmental change influence forecastability using other systems (e.g. mutualistic networks)

and other environmental factors would be a promising avenue of future research to come to a

more general understanding about the drivers of forecastability. Likewise, manipulation of

different aspects of community structure could shed further light on the potential and limits of

ecological forecasting. We manipulated species richness and attempted to keep the number of

functional groups constant across bottles. However, since not all species persisted, as is common

in such experiments (Steiner *et al.* 2006), our communities differed in the number of functional

groups (Supplement Section S9). While the number of trophic levels was the same in all but one

bottle, the number of functional groups was positively correlated with median realized richness

(Supplement Figure S33) and had similar effects on forecast error as realized richness

(Supplement Figure S34). Our experiment therefore does not allow to disentangle whether the

diversity effects were due to differences in the number of species or in the number of functional

groups. Teasing apart such effects could be a useful next step in future experiments.

Our findings give novel insights into the drivers of forecastability. Previous work has

focused on comparing forecastability among different species and ecosystem properties (Lewis

et al. 2022; Ward *et al.* 2014). For example, observational research showed that small, fast-

growing species are less forecastable than large, slow-growing species (Ward *et al.* 2014), and

experimental research found that species with fewer (and stronger) interactions are less

forecastable than species with more (and weaker) interactions (Daugaard *et al.* 2022). Time

series properties have been identified as a further driver of forecastability, with higher time series

complexity and stronger non-linearity leading to lower forecastability (Pennekamp *et al.* 2019).

Previous work has also explored how transferrable forecasting models are to novel biotic

contexts and showed that models are less transferrable for a species that is more impacted by a

change in biotic context than for a species that is less impacted (Dumandan *et al.* 2024). Our

findings advance our understanding about ecological forecasting by focusing on how properties

of the community (i.e. diversity) and of the environment (constant versus changing) affect the

forecastability of the species embedded in these communities. We found only weak support for

the expectation that higher diversity reduces the forecastability of populations. Whether a longer

diversity gradient would alter this finding is an open question. If species in very diverse

communities have more (but weaker) interactions, a longer diversity gradient might not

necessarily result in lower forecastability, given that more (and weaker) interactions can lead to

higher forecastability (Daugaard *et al.* 2022). Future work on forecastability could also explore

the effect of other community properties (e.g. number of trophic levels), of other global change

drivers, and of the number of drivers (single versus multiple drivers). While our study gave first

experimental insights, more work is needed to further improve our understanding about the

drivers of forecastability.

In conclusion, our results suggest that diversity does not necessarily reduce the

forecastability of ecological systems. Rather, when environmental conditions change, diversity

might promote forecastability by buffering against ecological surprises that can result from a

decline or rise of functional groups. In contrast, low diversity in combination with environmental

change could make ecosystems more unpredictable. Our results therefore caution against

assuming high forecastability of ecosystems with low diversity. Forecasting efforts for

management or conservation purposes might require particularly finely-resolved time series in

ecosystems of low diversity, so as not to miss unexpected changes in ecological dynamics as

environmental conditions change. Considering anthropogenic pressures on diversity, reduced

forecastability of less-diverse systems may pose an additional challenge to conserve populations,

communities, and ecosystems in changing environments.

Acknowledgements

631 We thank Fabian Avesani for help in the lab, Bernhard Schmid for discussions on the

632 experimental design, and the Predictive Ecology Group of the University of Zurich for feedback

633 on the manuscript. We thank Frederik Hammes for input on flowcytometry settings and for

634 access to a backup flowcytometer, with help from Jürg Sigrist. This project was part of SNF

635 Project 310030_188431.

References

Amundrud, S.L. & Srivastava, D.S. (2016). Trophic interactions determine the effects of drought

on an aquatic ecosystem. *Ecology*, 97, 1475–1483.

Baert, J.M., Laender, F.D., Sabbe, K. & Janssen, C.R. (2016). Biodiversity increases functional

and compositional resistance, but decreases resilience in phytoplankton communities.

Ecology, 97, 3433–3440.

Bates, D., Mächler, M., Bolker, B. & Walker, S. (2015). Fitting linear mixed-effects models

using lme4. *J. Stat. Softw.*, 67, 1–48.

Beckage, B., Gross, L.J. & Kauffman, S. (2011). The limits to prediction in ecological systems.

Ecosphere, 2, art125.

Bell, G., Fugère, V., Barrett, R., Beisner, B., Cristescu, M., Fussmann, G., *et al.* (2019). Trophic

structure modulates community rescue following acidification. *Proc. R. Soc. B Biol. Sci.*,

286, 20190856.

Benincà, E., Huisman, J., Heerkloss, R., Jöhnk, K.D., Branco, P., Van Nes, E.H., *et al.* (2008).

Chaos in a long-term experiment with a plankton community. *Nature*, 451, 822–825.

Bestion, E., Barton, S., García, F.C., Warfield, R. & Yvon-Durocher, G. (2020). Abrupt declines

in marine phytoplankton production driven by warming and biodiversity loss in a

microcosm experiment. *Ecol. Lett.*, 23, 457–466.

Blasius, B., Rudolf, L., Weithoff, G., Gaedke, U. & Fussmann, G.F. (2020). Long-term cyclic

persistence in an experimental predator–prey system. *Nature*, 577, 226–230.

Blois, J.L., Zarnetske, P.L., Fitzpatrick, M.C. & Finnegan, S. (2013). Climate change and the

past, present, and future of biotic interactions. *Science*, 341, 499–504.

Daugaard, U., Munch, S.B., Inauen, D., Pennekamp, F. & Petchey, O.L. (2022). Forecasting in

the face of ecological complexity: number and strength of species interactions determine

forecast skill in ecological communities. *Ecol. Lett.*, 25, 1974–1985.

Deyle, E.R., Bouffard, D., Frossard, V., Schwefel, R., Melack, J. & Sugihara, G. (2022). A

hybrid empirical and parametric approach for managing ecosystem complexity: water

quality in Lake Geneva under nonstationary futures. *Proc. Natl. Acad. Sci.*, 119,

e2102466119.

Deyle, E.R., Fogarty, M., Hsieh, C., Kaufman, L., MacCall, A.D., Munch, S.B., *et al.* (2013).

Predicting climate effects on Pacific sardine. *Proc. Natl. Acad. Sci.*, 110, 6430–6435.

Dietze, M., White, E.P., Abeyta, A., Boettiger, C., Bueno Watts, N., Carey, C.C., *et al.* (2024).

Near-term ecological forecasting for climate change action. *Nat. Clim. Change*, 1–9.

Dietze, M.C. (2017). *Ecological forecasting*. Princeton University Press, Princeton.

Dietze, M.C., Fox, A., Beck-Johnson, L.M., Betancourt, J.L., Hooten, M.B., Jarnevich, C.S., *et*

al. (2018). Iterative near-term ecological forecasting: needs, opportunities, and

challenges. *Proc. Natl. Acad. Sci.*, 115, 1424–1432.

Doak, D.F., Estes, J.A., Halpern, B.S., Jacob, U., Lindberg, D.R., Lovvorn, J., *et al.* (2008).

Understanding and predicting ecological dynamics: are major surprises inevitable?

Ecology, 89, 952–961.

Drake, J.M. & Griffen, B.D. (2010). Early warning signals of extinction in deteriorating

environments. *Nature*, 467, 456–459.

Dumandan, P.K.T., Simonis, J.L., Yenni, G.M., Ernest, S.K.M. & White, E.P. (2024).

Transferability of ecological forecasting models to novel biotic conditions in a long-term

experimental study. *Ecology*, 105, e4406.

Estes, J.A., Terborgh, J., Brashares, J.S., Power, M.E., Berger, J., Bond, W.J., *et al.* (2011).

Trophic downgrading of planet Earth. *Science*, 333, 301–306.

Fujita, H., Ushio, M., Suzuki, K., Abe, M.S., Yamamichi, M., Iwayama, K., *et al.* (2023).

Alternative stable states, nonlinear behavior, and predictability of microbiome dynamics.

Microbiome, 11, 63.

Fussmann, G.F., Ellner, S.P., Shertzer, K.W. & Jr, N.G.H. (2000). Crossing the Hopf bifurcation

in a live predator-prey system. *Science*, 290, 1358–1360.

Fussmann, K.E., Schwarzmüller, F., Brose, U., Jousset, A. & Rall, B.C. (2014). Ecological

stability in response to warming. *Nat. Clim. Change*, 4, 206–210.

Gilman, S.E., Urban, M.C., Tewksbury, J., Gilchrist, G.W. & Holt, R.D. (2010). A framework

for community interactions under climate change. *Trends Ecol. Evol.*, 25, 325–331.

Henden, J.-A., Ims, R.A., Yoccoz, N.G., Asbjørnsen, E.J., Stien, A., Mellard, J.P., *et al.* (2020).

End-user involvement to improve predictions and management of populations with

complex dynamics and multiple drivers. *Ecol. Appl.*, 30, e02120.

Hong, P., Schmid, B., De Laender, F., Eisenhauer, N., Zhang, X., Chen, H., *et al.* (2022).

Biodiversity promotes ecosystem functioning despite environmental change. *Ecol. Lett.*,

25, 555–569.

Huston, M.A. (1997). Hidden treatments in ecological experiments: re-evaluating the ecosystem

function of biodiversity. *Oecologia*, 110, 449–460.

Isbell, F., Craven, D., Connolly, J., Loreau, M., Schmid, B., Beierkuhnlein, C., *et al.* (2015).

Biodiversity increases the resistance of ecosystem productivity to climate extremes.

Nature, 526, 574–577.

Jessup, C.M., Kassen, R., Forde, S.E., Kerr, B., Buckling, A., Rainey, P.B., *et al.* (2004). Big

questions, small worlds: microbial model systems in ecology. *Trends Ecol. Evol.*, 19,

189–197.

Jochum, M., Fischer, M., Isbell, F., Roscher, C., van der Plas, F., Boch, S., *et al.* (2020). The

results of biodiversity–ecosystem functioning experiments are realistic. *Nat. Ecol. Evol.*,

4, 1485–1494.

Leary, D.J. & Petchey, O.L. (2009). Testing a biological mechanism of the insurance hypothesis

in experimental aquatic communities. *J. Anim. Ecol.*, 78, 1143–1151.

Leech, D.M., Pollard, A.I., Labou, S.G. & Hampton, S.E. (2018). Fewer blue lakes and more

murky lakes across the continental U.S.: Implications for planktonic food webs. *Limnol.*

Oceanogr., 63, 2661–2680.

Lewis, A.S.L., Rollinson, C.R., Allyn, A.J., Ashander, J., Brodie, S., Brookson, C.B., *et al.*

(2023). The power of forecasts to advance ecological theory. *Methods Ecol. Evol.*, 14,

746–756.

Lewis, A.S.L., Woelmer, W.M., Wander, H.L., Howard, D.W., Smith, J.W., McClure, R.P., *et*

al. (2022). Increased adoption of best practices in ecological forecasting enables

comparisons of forecastability. *Ecol. Appl.*, 32, e2500.

Limberger, R., Daugaard, U., Gupta, A., Krug, R.M., Lemmen, K.D., van Moorsel, S.J., *et al.*

(2023). Functional diversity can facilitate the collapse of an undesirable ecosystem state.

Ecol. Lett., 26, 883–895.

Loreau, M., Barbier, M., Filotas, E., Gravel, D., Isbell, F., Miller, S.J., *et al.* (2021). Biodiversity

as insurance: from concept to measurement and application. *Biol. Rev.*, 96, 2333–2354.

Martin, T.E. & Maron, J.L. (2012). Climate impacts on bird and plant communities from altered

animal–plant interactions. *Nat. Clim. Change*, 2, 195–200.

Meyer, D., Dimitriadou, E., Hornik, K., Weingessel, A. & Leisch, F. (2021). *e1071: Misc*

Functions of the Department of Statistics, Probability Theory Group (Formerly: E1071),

TU Wien.

Montoya, J.M., Woodward, G., Emmerson, M.C. & Solé, R.V. (2009). Press perturbations and

indirect effects in real food webs. *Ecology*, 90, 2426–2433.

Munch, S.B., Rogers, T.L., Symons, C.C., Anderson, D. & Pennekamp, F. (2023). Constraining

nonlinear time series modeling with the metabolic theory of ecology. *Proc. Natl. Acad.*

Sci., 120, e2211758120.

Novak, M., Wootton, J.T., Doak, D.F., Emmerson, M., Estes, J.A. & Tinker, M.T. (2011).

Predicting community responses to perturbations in the face of imperfect knowledge and

network complexity. *Ecology*, 92, 836–846.

Oberski, D. (2014). lavaan.survey: an R package for complex survey analysis of structural

equation models. *J. Stat. Softw.*, 57, 1–27.

Ockendon, N., Baker, D.J., Carr, J.A., White, E.C., Almond, R.E.A., Amano, T., *et al.* (2014).

Mechanisms underpinning climatic impacts on natural populations: altered species

interactions are more important than direct effects. *Glob. Change Biol.*, 20, 2221–2229.

Pace, M.L., Batt, R.D., Buelo, C.D., Carpenter, S.R., Cole, J.J., Kurtzweil, J.T., *et al.* (2017).

Reversal of a cyanobacterial bloom in response to early warnings. *Proc. Natl. Acad. Sci.*,

114, 352–357.

Pennekamp, F., Iles, A.C., Garland, J., Brennan, G., Brose, U., Gaedke, U., *et al.* (2019). The

intrinsic predictability of ecological time series and its potential to guide forecasting.

Ecol. Monogr., 89, e01359.

Pennekamp, F., Pontarp, M., Tabi, A., Altermatt, F., Alther, R., Choffat, Y., *et al.* (2018).

Biodiversity increases and decreases ecosystem stability. *Nature*, 563, 109–112.

Pennekamp, F., Schtickzelle, N. & Petchey, O.L. (2015). BEMOVI, software for extracting

behavior and morphology from videos, illustrated with analyses of microbes. *Ecol. Evol.*,

5, 2584–2595.

755 Petchey, O.L., McPhearson, P.T., Casey, T.M. & Morin, P.J. (1999). Environmental warming

756   alters food-web structure and ecosystem function. *Nature*, 402, 69–72.

757 Piou, C. & Marescot, L. (2023). Spatiotemporal risk forecasting to improve locust management.

758   *Curr. Opin. Insect Sci.*, 56, 101024.

759 Polis, G.A. & Strong, D.R. (1996). Food web complexity and community dynamics. *Am. Nat.*,

760   147, 813–846.

761 Quinn, G.P. & Keough, M.J. (2024). *Experimental design and data analysis for biologists*. 2nd

762   edn. Cambridge University Press, UK.

763 Rosseel, Y. (2012). lavaan: an R package for structural equation modeling. *J. Stat. Softw.*, 48, 1–

764   36.

765 Rousso, B.Z., Bertone, E., Stewart, R. & Hamilton, D.P. (2020). A systematic literature review

766   of forecasting and predictive models for cyanobacteria blooms in freshwater lakes. *Water*

767   *Res.*, 182, 115959.

768 Schmid, B., Baruffol, M., Wang, Z. & Niklaus, P.A. (2017). A guide to analyzing biodiversity

769   experiments. *J. Plant Ecol.*, 10, 91–110.

770 Steiner, C.F., Long, Z.T., Krumins, J.A. & Morin, P.J. (2005). Temporal stability of aquatic food

771   webs: partitioning the effects of species diversity, species composition and enrichment.

772   *Ecol. Lett.*, 8, 819–828.

773 Steiner, C.F., Long, Z.T., Krumins, J.A. & Morin, P.J. (2006). Population and Community

774   Resilience in Multitrophic Communities. *Ecology*, 87, 996–1007.

775 Sugihara, G. & May, R.M. (1990). Nonlinear forecasting as a way of distinguishing chaos from

776   measurement error in time series. *Nature*, 344, 734–741.

Tabi, A., Petchey, O.L. & Pennekamp, F. (2019). Warming reduces the effects of enrichment on

stability and functioning across levels of organisation in an aquatic microbial ecosystem.

Ecol. Lett., 22, 1061–1071.

Tilman, D. (1996). Biodiversity: population versus ecosystem stability. *Ecology*, 77, 350–363.

Tilman, D., Fargione, J., Wolff, B., D'Antonio, C., Dobson, A., Howarth, R., *et al.* (2001).

Forecasting agriculturally driven global environmental change. *Science*, 292, 281–284.

Tulloch, A.I.T., Hagger, V. & Greenville, A.C. (2020). Ecological forecasts to inform near-term

management of threats to biodiversity. *Glob. Change Biol.*, 26, 5816–5828.

Ward, E.J., Holmes, E.E., Thorson, J.T. & Collen, B. (2014). Complexity is costly: a meta-

analysis of parametric and non-parametric methods for short-term population forecasting.

Oikos, 123, 652–661.

Winder, M. & Sommer, U. (2012). Phytoplankton response to a changing climate.

Hydrobiologia, 698, 5–16.

Xu, Q., Yang, X., Yan, Y., Wang, S., Loreau, M. & Jiang, L. (2021). Consistently positive effect

of species diversity on ecosystem, but not population, temporal stability. *Ecol. Lett.*, 24,

2256–2266.

Yachi, S. & Loreau, M. (1999). Biodiversity and ecosystem productivity in a fluctuating

environment: the insurance hypothesis. *Proc. Natl. Acad. Sci.*, 96, 1463–1468.

Ye, H. & Sugihara, G. (2016). Information leverage in interconnected ecosystems: overcoming

the curse of dimensionality. *Science*, 353, 922–925.

Yodzis, P. (2000). Diffuse effects in food webs. *Ecology*, 81, 261–266.

Table 1: Results of two linear mixed models investigating the effects of species richness and

light on the forecast error of taxa abundances and community biomass, respectively. We

forecasted 253 taxa time series (across all bottles) and community biomass in 30 bottles. In both

models, median centered richness and light treatment were fixed effects. The first model

(response: forecast error of taxa abundances) included taxon, bottle, and incubator as random

intercepts. The second model (response: forecast error of community biomass) included

composition and incubator as random intercepts. Forecast error is based on forecasts computed

with Simplex Empirical Dynamic Modeling (EDM). In the second model, the interaction was not

significant (p-value > 0.1) and was therefore removed from the model. The last four columns

report the ANOVA results. P-values < 0.05 in bold. See Supplement Table S6 for further details.

Response	Covariate	Estimate	95% CI	SS	DF	*F*-value	*p*-value
Forecast error of taxa abundances	Intercept	0.685	0.503 – 0.865				
	Richness	0.056	-0.008 – 0.121	0.008	1, 21.1	0.046	0.8328
	Light (decreasing)	-0.095	-0.265 – 0.081	0.225	1, 6.1	1.230	0.3092
	Richness:Light	-0.102	-0.193 – -0.014	0.922	1, 19.7	5.046	**0.0363**
Forecast error of community biomass	Intercept	0.657	0.447 – 0.866				
	Richness	0.015	-0.107 – 0.134	0.004	1, 19.8	0.058	0.8118
	Light (decreasing)	-0.017	-0.230 – 0.186	0.002	1, 14.1	0.028	0.8698

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Figure 1: Effects of species richness and light on forecast error. The two treatments had

interactive effects on forecast error of taxa abundances (a) but no effect on the forecast error of

community biomass (b). Forecasts were computed with Simplex Empirical Dynamic Modeling

(EDM), and forecast error was calculated as root mean square error (RMSE). RMSE is

standardized (unitless). Observed richness deviated from planned richness because not all species

persisted throughout the experiment. We therefore used realized richness (i.e. the median of the

observed richness values) as explanatory variable. Median richness was centered; a centered

median richness of zero corresponds to a median richness of 5.5 (minimum median richness: 3;

maximum median richness: 8). Lines display the fit of the linear mixed model; shaded areas

denote the 95% confidence intervals. In (a), we forecasted 253 taxa time series (across all

bottles); each point represents a forecasted taxon in a bottle. Taxon, bottle, and incubator were

included as random intercepts in the model. Inserts at the top indicate the number of bottles for

the respective richness level (yellow: constant light, blue: decreasing light). In (b), we forecasted

community biomass in 30 bottles. Composition and incubator were included as random

intercepts in the model.

Figure 2: Relationship of time series metrics with forecast error (a-c), and with experimental

treatments (d-f). Time series metrics are (a, d) autocorrelation at lag 5, (b, e) coefficient of

variation as a measure of temporal variability, and (c, f) permutation entropy as a measure of

time series complexity. All variables are unitless. Time series metrics were calculated for 253

taxa time series (across all bottles). Lines display the fit of the corresponding linear mixed

model, shaded areas denote the 95% confidence intervals. Taxon, bottle, and incubator were

included as random effects in the model. Forecast error is based on forecasts computed with

Simplex EDM. Note that in (b) and (e) the coefficient of variation was $\log_{10}$-transformed so that

linear model assumptions were met.

Figure 3: Structural equation model (SEM) showing the effects of species richness and time

series metrics on forecast error of species abundances in (a) constant light, and (b) decreasing

light. Solid and dashed arrows indicate significant and non-significant effects, respectively (see

Supplement Table S17). Values next to arrows are the path coefficients. The color matches the

sign of the coefficients (blue=positive, red=negative). Bidirectional arrows are covariances

between the error terms of the involved variables.