Dorsomedial Striatum CB1R signaling is required for Pavlovian outcome devaluation in male Long Evans rats and reduces inhibitory synaptic transmission in both sexes.

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ABSTRACT

Cannabinoid-1 receptor (CB1R) signaling in the dorsal striatum regulates the shift from flexible to habitual behavior in instrumental outcome devaluation. Based on prior work establishing individual, sex, and experience-dependent differences in Pavlovian behaviors, we predicted a role for dorsomedial striatum CB1R signaling in driving rigid responding in Pavlovian autoshaping and outcome devaluation. We trained male and female Long Evans rats in Pavlovian Lever Autoshaping (PLA). We gave intra-dorsomedial striatum (DMS) infusions of the CB1R inverse agonist, rimonabant, before satiety-induced outcome devaluation test sessions, where we sated rats on training pellets or home cage chow and tested them in brief nonreinforced Pavlovian Lever Autoshaping sessions. Overall, inhibition of DMS CB1R signaling prevented Pavlovian outcome devaluation but did not affect behavior in reinforced PLA sessions. Males were sensitive to devaluation while females were not and DMS CB1R inhibition impaired devaluation sensitivity in males. We then investigated how DMS CB1R signaling impacts local inhibitory synaptic transmission in male and female Long Evans rats. We recorded spontaneous inhibitory postsynaptic currents (sIPSC) from DMS neurons at baseline and before and after application of a CB1R agonist, WIN 55,212-2. We found that male rats showed decreased sIPSC frequency compared to females, and that CB1R activation reduced DMS inhibitory transmission independent of sex. Altogether our results demonstrate that DMS CB1Rs regulate Pavlovian devaluation sensitivity and inhibitory synaptic transmission and suggest that basal sex differences in inhibitory synaptic transmission may underly sex differences in DMS function and behavioral flexibility.

INTRODUCTION

Impairments in behavioral flexibility occur across a range of mental health disorders including Substance Use Disorder, schizophrenia, Obsessive-Compulsive Disorder, and depression (Geramita et al., 2020; Jordan and Andersen, 2017; Kalivas and Volkow, 2005; Listunova et al., 2018; Simmler and Ozawa, 2019; Thoma et al., 2007). Preclinical studies suggest that sex and individual differences influence behavioral control when environmental conditions change from what is expected (Amaya et al., 2020; Bien and Smith, 2023; Keefer et al., 2020; Morrison et al., 2015; Nasser et al., 2015). Understanding the neurobiological underpinnings of individual and sex differences in behavioral flexibility may help to identify novel therapeutic targets for disorders of behavioral control.

Instrumental conditioning procedures in rats identified dorsal striatal regulation of behavioral flexibility, which involves dorsomedial and dorsolateral striatal (DMS, DLS) subdivisions. The shift from goal-directed to habitual behavior that occurs after extended instrumental experience is mediated by a shift from DMS to DLS control, respectively (Amaya and Smith, 2018; Dickinson et al., 1995; Gremel and Costa, 2013; Peak et al., 2019; Yin et al., 2005, 2004). Within the dorsal striatum (DS), multiple cell-types mediate the activity and output of each DS subregion. The majority of neurons in the DS are GABAergic medium spiny neurons (MSNs), the main type of projection neuron arising from the DS (Graveland and Difiglia, 1985). One of the most abundant receptor types in the DS is the Cannabinoid Receptor-1 (CB1R), which is a G-protein coupled receptor that is expressed pre-synaptically on glutamatergic terminals into the DS and locally on terminals of fast-spiking interneurons and MSNs (Gerdeman and Lovinger, 2001; Gerdeman et al., 2002; Lovinger and Mathur, 2012; Brian N Mathur et al., 2013; Wu et al., 2015). An instrumental outcome devaluation study shows that CB1R deletion in the orbitofrontal cortex-DS projection promotes devaluation sensitivity even during schedules of reinforcement that ordinarily drive habitual responding (Gremel et al., 2016), suggesting that that CB1R-mediated inhibition of synaptic inputs to DMS shift behavior towards rigid, devaluation insensitive instrumental actions.

We hypothesize that DMS CB1R signaling also biases behavior towards rigid devaluation insensitive Pavlovian behaviors. The sign-tracking model has uncovered considerable individual, sex and, experience-dependent differences in Pavlovian devaluation sensitivity (Flagel et al., 2009; Keefer et al., 2020; Kochli et al., 2020; Madayag et al., 2017; Pitchers et al., 2015), which suggest differences in endocannabinoid regulation of behavioral flexibility in the DMS. After limited training (<10 sessions) in Pavlovian lever autoshaping (PLA), in which an insertable lever cue predicts a food outcome, goal-tracking (GT) rats show sensitivity to outcome devaluation while sign-tracking (ST) rats do not (Keefer et al., 2020; Morrison et al., 2015; Nasser et al., 2015; Patitucci et al., 2016). After extended training (>10 session), both GT and ST rats show sensitivity to satiety-induced outcome devaluation (Keefer et al., 2020), an effect established in male rats. Female rats show increased levels of sign-tracking, or lever-directed approach during PLA compared to males (Hammerslag and Gulley, 2014; Keefer et al., 2022; King et al., 2020; Kochli et al., 2020; Madayag et al., 2017; Pitchers et al.,

2015), suggesting they may be less sensitive to outcome devaluation even after extended training. In the present study, we use intracranial CB1R inverse agonist, rimonabant, to determine the role of DMS CB1R in mediating Pavlovian devaluation sensitivity in male and female rats.

Opposite to our prediction, we find that DMS CB1R signaling is necessary for flexible behavior in Pavlovian outcome devaluation. Based on this finding, and prior studies establishing that inhibition of DMS promotes rigid responding (Ragozzino et al., 2002; Yin et al., 2005), we hypothesized that CB1Rs located on GABAergic synapses onto MSNs in the DMS act to reduce inhibitory synaptic transmission. To test this, we measured the effect of CB1R activation on spontaneous inhibitory post synaptic currents (sIPSCs) in the DMS. In the slice electrophysiology studies, we include both males and females to investigate potential sex differences in DMS physiology. We found that male rats showed decreased sIPSC frequency compared to females, and that CB1R activation reduced DMS inhibitory transmission independent of sex.

METHODS

Subjects

For behavioral experiments, we used 68 Long Evans rats (33 Male, 35 Female; run as 5 cohorts) in the age range of 7-9 weeks old at the start of training for this study. All rats were double-housed upon arrival and then single-housed 24-48 hours after arrival. We performed all behavioral procedures during the dark phase of the light cycle. All rats had *ad libitum* access to standard laboratory chow and water before we food deprived them to maintain 90% of their baseline weight. We surgerized one cohort prior to any behavioral training and testing and surgerized the remaining cohorts after three days of training. There were no pre- or post-surgery differences in behavior between groups.

For slice electrophysiology experiments, we used 24 Long Evans rats (13 Male, 11 Female) in the age range of 9-15 weeks old at the time of slice electrophysiology recording. All rats were double housed upon arrival. These rats had ad libitum access to standard laboratory chow and water before we food deprived them 24 hours prior to slice electrophysiology recording.

We maintained all rats on a reverse 12hr:12hr light-dark cycle (lights off at 1000). We performed all procedures in accordance with the "Guide for the Care and Use of Laboratory Animals" (8th edition, 2011, US National Research Council) and with approval by [Author University] Institutional Animal Care and use Committee (IACUC).

Apparatus

We conduct behavioral experiments in identical operant chambers (25 X 27 X 30 cm; Med Associates) located in a separate room from the animal colony. An individual sound-attenuating cubicle with a ventilation fan surrounds each chamber. One wall contains a red house light and the opposing wall contains a food cup with photobeam detectors that rests 2 cm above the grid floor. A programmed pellet dispenser attached to the foodcup and dispensed 45 mg food pellets (catalog #1811155; Test Diet Purified rodent Tablet [5TUL]; protein

20.6%, fat 12.7%, carbohydrate 66.7%). We installed one retractable lever at 6cm above the grid floor on either side of the foodcup and we counterbalanced the lever side between subjects.

Surgical Procedures

After three days of Pavlovian Lever Autoshaping training, we gave ad libitum access to food before we performed intracranial cannula placement surgery. We anesthetized 8-week-old rats with isoflurane (VetOne, Boise, ID, USA; 5% induction, 2-3% maintenance) then administered the pre-operative analgesic carprofen (5mg/kg, s.c.) and lidocaine (10mg/mL subdermal at incision site). We placed them in a stereotaxic frame (model 900, David Kopf Instruments, Tujunga, CA, USA) over a heating pad to maintain stable body temperature throughout surgery.

We implanted guide cannula (23G; PlasticsOne INC, Roanoke, VA, USA) bilaterally at an 8 degree angle and 1mm above the injection site into the DMS (coordinates from bregma -0.24 mm AP, ± 2.6 mm ML and -4.5 mm DV). We determined distance from bregma using the Paxinos and Watson rat brain atlas (Paxinos and Watson, 2006). Cannula were secured to the skull with jeweler's screws and dental cement. At the end of surgery, we inserted dummy cannula into the guide cannula, which we only removed during infusion habituation and infusion test procedures. We moved rats to a recovery cage over a heating pad, administered carprofen analgesic at 24 hr, 48 hr and 72 hr post-surgery. We gave animals 1 week of recovery before resuming behavioral procedures.

Pavlovian Lever Autoshaping Training

Prior to training, we exposed all rats to the food pellets in their home cage to reduce novelty to the food. Then we trained them in daily Pavlovian lever autoshaping sessions which lasted ~ 26 minutes and included 25 trials of non-contingent lever presentations (conditioned stimulus; CS) and occurred on a VI 60 s schedule (50-70s). At the start of the session, the houselight turned on and remained on for the duration of the session. Each trial consisted of a 10 s lever presentation and retraction of the lever followed immediately by delivery of two 45 mg food pellets into the foodcup. At the end of the session, we returned rats to their cage and colony room. We trained rats in PLA first for 5 days to determine their tracking group, then continued training through 12 days following PLA testing.

Pavlovian Lever Autoshaping Testing

We tested the effects of blocking DMS CB1R during reinforced Pavlovian Lever Autoshaping. We infused rimonabant (SR141716 1 µg/µL or 2 µg/µL; dissolved in 1:1:18 ethanol: emulphor: saline solution) or vehicle bilaterally into DMS at a rate of 0.5 µL/min over the span of 1 minute. We left the infusion cannula in place for an additional minute before slowly removing them and replacing the dummy cannula. We waited 10 min after infusion before placing rats into the behavioral chamber and running the lever autoshaping test. We infused all rats with vehicle, low (1 µg/µL) or high (2 µg/µL) dose of rimonabant across three days and we counterbalanced the dose across days.

Satiety-Induced Outcome Devaluation Testing

After the 12th training session, we gave rats two sessions of satiety-induced outcome devaluation. Rats had one hour of *ad libitum* access to 30 g of either their homecage chow (valued condition) or food pellets used during PLA training (devalued condition) in a ceramic ramekin. Within 15 min of the end of the satiation hour, we performed intra-DMS rimonabant infusions (2 µg/µL) as described in the previous section. We waited 10 min after the infusion before placing rats into the behavioral chamber and running the lever autoshaping test. Tests consisted of 10 non-rewarded lever presentations on VI 60s schedule (50-70s). Immediately after each test, we gave rats a 30 min food choice test in their homecage which included 10 g of homecage chow and 10 g of food pellets in separate ceramic ramekins to confirm satiety was specific to the outcome they had been fed before the test session. We retrained rats on 25 reinforced trials on a separate day between devaluation probe tests.

Brain Slice Preparation for Slice Electrophysiology.

We anesthetized rats with isoflurane then perfused with chilled N-Methyl-D-Glucamine (NMDG)-modified artificial cerebrospinal fluid (NMDG-aCSF; in mM; 92 NMDG, 20 HEPES, 25 Glucose, 30 NaHCO3, 1.3 NaH2PO4, 2.5 KCl, 5 Sodium Abscorbate, 3 Sodium Pyruvate, 2 Thiourea, 10 MgSO4, 0.5 CaCl2) that had been bubbled with carbogen (95% oxygen, 5% carbon dioxide). We collected coronal sections from the DMS (350µM) while the brain was mounted on the cutting stage and submerged in chilled, carbogen-bubbled NMDG-aCSF, using a Leica VT 1200 vibratome. We incubated slices in carbogen-bubbled, 40° NMDG solution for 5-8 minutes then transferred slices to room temperature, carbogen-bubbled HEPES holding solution (in mM; 92 NaCl, 20 HEPES, 25 Glucose, 30 NaHCO3, 1.3 NaH2PO4, 2.5 KCl, 5 Sodium Abscorbate, 3 Sodium Pyruvate, 2 Thiourea, 1 MgSO4, 2 CaCl2). We waited 1 hour before making the first recordings. Sections remained in the holding solution until electrophysiological recordings were performed.

Recordings and Bath Application of Drug.

We visualized cells in the DMS using an Olympus BX50 light microscope. We recorded spontaneous IPSCs (sIPSC) using borosilicate, fire-polished glass pipettes with resistance in the 3-5 MΩ range. We pulled pipettes with a Narshige PC-100 pipette puller and filled them with a CsCl-based internal solution (in mM; 150 CsCl, 10 HEPES, 2 MgCl2*H2O, 0.3 Na-GTP, 3 Mg-ATP, 0.2 BAPTA). We recorded from hemisected slices that were constantly perfused with 37° carbogen-bubbled artificial cerebrospinal fluid (aCSF; in mM; 126 NaCl, 25 NaHCO3, 11 Glucose, 1.2 MgCl2*H2O, 1.4 NaH2PO4, 2.5 KCl, 2.4 CaCl2), containing blockers of AMPA (DNQX, 20µM) and NMDA (AP5, 50µM). We perfused the recording chamber with a basic Longer Pump BT100-2J peristaltic pump. We also recorded from slices submerged in a bath containing DMSO (0.065%) and 2-hydroxypropyl-beta-cyclodextrin (0.006%). We clamped cells at -60 mV using a Molecular Devices Multiclamp 700B amplifier and digitized recordings with a Molecular Devices Axon Digidata 1550B digitizer. We used Molecular Devices Clampex 10.7 software for data acquisition. We excluded recordings when sIPSC baseline was below -200 pA, series resistance was >40 MΩ, or series resistance changed >20% throughout the course of the experiment.

Measurements

For training and devaluation probe tests, we recorded the number and duration of foodcup and lever contacts, the latency to contact, and the probability during the 10 s CS (lever) period. On trials with no contacts, a latency of 10s was recorded. To determine tracking group, we used a Pavlovian Conditioned Approach (PavCA) analysis (Meyer et al., 2012) which quantifies behavior along a continuum where +1.00 indicates behavior is primarily lever directed (sign-tracking) and -1.00 indicates behavior is primarily foodcup directed (goal-tracking). PavCA scores are the average of three separate scores: the preference score (lever contacts minus foodcup contacts divided by the sum of these measures), the latency score (time to contact foodcup minus the time to contact lever divided by 10 s (duration of the cue)) and the probability score (probability to make a lever contact minus the probability to make a foodcup contact across the session). We use the PavCA score from the 5th day of training to determine an individual's tracking group as follows: sign-trackers (ST) have a PavCA sore +0.33 to +1.00, goal-trackers (GT) have a PavCA score -1.00 to -0.33, intermediates (INT) have scores ranging from -0.32 to +0.32. Rats in goal- and intermediate tracking groups were combined into a single GT/INT group as they show similar patterns of outcome devaluation in other studies (Keefer et al., 2020). On day 6, we were unable to record latency data for 6 rats and only retained lever and foodcup contacts for these rats. Preference score was used in place of PavCA for rats on this day.

For devaluation probe tests, we also report total approach (the sum of food cup and lever contacts during the 10 s CS period) and individual contact measurements. We recorded consumption on the test days and calculated the amount of pellet or chow consumed in grams during the satiety hour and during the 30 min choice test.

We processed sIPSC traces using the template search function in Molecular Devices Clampfit 10.7 software to determine event peak amplitude and event peak start time. We report these measurements in each experiment: Amplitude, calculated as the peak amplitude of an event and averaged across each recording; Frequency, calculated as the number of events per recording divided by the duration of the recording in seconds; Interevent Interval, calculated as the inverse of the time (in seconds) between the peak of an event and the peak of the event prior and represented through a cumulative frequency distribution.

Histology

At the end of behavioral experiments, we deeply anesthetized rats with isoflurane and transcardially perfused 100ml of 0.1M sodium phosphate buffer (PBS), followed by 200ml of 4% paraformaldehyde (PFA) in PBS. We removed brains and post-fixed them in 4% PFA over night before we transferred them to 30% sucrose in dH2O for 48-72 hr at 4 °C. We rapidly froze brains in dry ice before storing them in -20 °C until slicing. We sliced brains with the Leica Microsystems 1850 cryostat to collect 40 μm coronal sections in three series through the cannula placements in the DMS. We mounted sections onto gel-coated slides and then stained with cresyl violet before coverslipping with Permount. We examined placements under a light microscope for confirmation of cannula placement in the DMS (Fig. 2B). We excluded 11 rats due to cannula placements being outside the region of interest.

Experimental Design and Statistical Analysis

We analyzed behavioral data using SPSS 29.0 statistical software (IBM) or Prism (Graphpad software) with

mixed-design repeated measures analysis of variance (ANOVA) or paired t tests, where applicable. Significant

main and interaction effects (p<0.05) were followed by post-hoc repeated-measures ANOVA or Bonferroni

comparisons. Analyses included between subject factors of Tracking (ST, GT/INT) Sex (male, female) and

Treatment (vehicle, rimonabant) and within-subject factors of Session (1-12), Outcome Value (Valued,

Devalued), or Outcome (Nonsated, Sated).

For slice electrophysiology experiments, data are represented as mean ± standard error or presented as

cumulative frequency distribution plots. We performed independent samples student's t-test, two sample

Kolmogorov-Smirnov tests, or Kruskal-Wallis tests with Dunn's post hoc comparisons as appropriate using

either SPSS or Prism. We analyzed mean amplitude and mean frequency data using independent samples t-

tests between males and females. We analyzed the cumulative frequency distribution of interevent interval

between males and females using a Kolmogorov-Smirnov test and reported the effect size using Hedges' *g.*

We analyzed the cumulative frequency of interevent intervals between DMSO and WIN conditions in the bath

and between males and females using the Kruskal-Wallis test with Dunn's post hoc comparisons. Analysis

included within-subject variable of Bath (pre-WIN, post-WIN) and between-subject variable of Sex (Male,

Female). We removed two data points, one from each Sex, based on results from Grubb's Test for Outliers.

RESULTS

Acquisition of Pavlovian Lever Autoshaping differs due to Tracking and Sex

We trained rats for 12 days in Pavlovian Lever Autoshaping in which an insertable lever cue predicts food

pellet delivery. We used the Pavlovian Conditioned Approach Index (PavCA) on the 5th session of training to

determine tracking groups (Fig. 1A). We then used a mixed design repeated measures ANOVA with between

subject factor of Tracking (ST, GT/INT) and within subject factor of Session (1-12). Consistent with group

assignments, ST rats show more lever directed behavior than GT/INT rats (main effect Tracking; $F_{(1,53)}$ =

49.293, p=<0.001). Consistent with prior studies (Villaruel and Chaudhri, 2016; Bacharach et al., 2018; Keefer

et al., 2020) showing that GT and intermediate (GT/INT) rats shift away from food-cup approach and towards

lever approach with extended

training, we observe a main

effect of Session for PavCA

Index, $F_{(11,583)}$=106.292,

p<0.001, and a Session x

Tracking (ST, GT/INT)

interaction, $F_{(11,583)}$=13.909,

p<0.001). Next, we examined

whether there were sex

Figure 1. Acquisition of a Pavlovian Conditioned Approach differs by Tracking and Sex. *A,* PavCA Index mean ± SEM for ST and GT/INT (collapsed on sex) that acquire individual differences in conditioned responding in PLA task. *Main effect of Session. %Significant Session X Tracking interaction. *B,* PavCA Index mean ± SEM for Male and Female rats (collapsed on tracking) that acquire conditioned responding in a PLA task. *Main effect of Session. %Significant Session X Sex interaction.

differences in the acquisition and expression of Pavlovian approach behaviors (Fig. 1B). We used a similar
statistical approach with between-subject factor of Sex (Male, Female) and within subject factor of Session (1-
12) and found a Session x Sex interaction for PavCA Index, F(11,605)=1.823, p=0.047). We analyzed PavCA
indices between males and females using independent samples t-tests for each day. While males and females
show similar PavCA indices during initial acquisition, female rats showed more sign-tracking, via a higher
PavCA Index, than males with extended training (day 8, t(55)=-1.754, p=0.043; day 9,t(55)=-2.007, p=0.025).
However, there were no sex differences in responding on the last day of training (PavCA Index; t(55)=-1.099,
p=0.277), prior to testing in outcome devaluation.

Intra-DMS inhibition of CB1R signaling prevents outcome devaluation but does not affect Pavlovian Approach
during non-sated, reinforced sessions.

We tested rats using a within-subject satiety-induced outcome devaluation procedure in which they were sated
on the training pellet (devalued) or homecage chow (valued) just prior to brief PLA test sessions under
extinction conditions. Prior to test sessions we gave intra-DMS vehicle or CB1R inverse agonist, rimonabant,
injections to determine the effects of inhibiting DMS CB1R signaling on devaluation sensitivity of Pavlovian
approach. To examine how this manipulation generally affects Pavlovian devaluation sensitivity across all rats,
we analyzed total approach which is the sum of lever and foodcup contacts during the 10 s cue presentation. We compared responding during the valued (chow sated) versus devalued (pellet sated) conditions using a mixed design repeated measures ANOVA with between subject factor of Treatment (Vehicle, Rimonabant) and within subject factor of Outcome Value (Valued, Devalued). Figure 2A shows the performance of all rats that received either intra-DMS vehicle or rimonabant infusions during the outcome devaluation probe test. We found a main effect of Outcome Value (F(1,49)=5.558, p=0.022) and an Outcome Value x Treatment interaction (F(1,49)=6.663, p=0.013), indicating that intra-DMS rimonabant impaired Pavlovian

Figure 2. Intra-DMS Rimonabant prevents sensitivity to Pavlovian Outcome Devaluation. Data are represented as within-subject individual data (lines) and group data (bars; mean ± SEM). Rats received intra-DMS injections of either vehicle (left) or rimonabant (right) 10 minutes prior to probe test. **A,** Total behavior (sum of lever and food cup contacts) in outcome devaluation across all rats. We observed a main effect of Outcome Value and a significant Outcome Value X Treatment interaction. **B,** Coronal sections (in mm) depicting the location of DMS injector tips for rimonabant infusion. **p<0.025

devaluation sensitivity across all rats. Under vehicle conditions, rats decreased total approach when sated on
the training pellet (devalued state) compared to when they were sated on the homecage chow (valued state).
In contrast, with intra-DMS rimonabant infusions, rats showed a similar amount of Pavlovian approach in the
valued and devalued states. These results suggest a divergent endocannabinoid mechanism for mediating

Pavlovian outcome devaluation in which DMS CB1R promote flexibility, in contrast to prior studies suggesting that CB1R signaling promotes rigid responding in instrumental settings (Navarro et al., 2001; Hilário et al., 2007; Gremel et al., 2016). Figure 2B shows the approximate location of intra-DMS infusions.

Considering the established individual differences in devaluation sensitivity in Pavlovian autoshaping (Keefer et al., 2020; Morrison et al., 2015; Nasser et al., 2015; Patitucci et al., 2016; Smedley and Smith, 2018), we added Tracking and Sex as between-subject factors in this analysis. We observed an Outcome Value x Treatment x Sex x Tracking interaction (F(1,49)=4.545, p=0.038)) which points to differences in the effects of treatment on devaluation sensitivity that differ by Sex and/or Tracking. In male rats we observed a main effect of Outcome Value and an Outcome Value x Treatment interaction (Fig. 3A, Value: F(1,25)=6.084, p=0.021; Value X Treatment: F(1,25)=6.440, p=0.018). Bonferroni post hoc comparisons confirm that under vehicle conditions, male rats were sensitive to outcome devaluation (t(53)=3.905, p<0.007) responding more to cues in valued than in devalued conditions. We observed that intra-DMS rimonabant impaired devaluation sensitivity in male rats, as they responded similarly between valued and devalued conditions (t(53)=0.0534, p>0.999). In female rats, we did not observe any significant main effects or interactions (Fig. 3B; *Fs*<0.890, *ps*>0.353), indicating they were not sensitive to Pavlovian outcome devaluation; thus, we could not evaluate treatment

Figure 3. Male, but not female, rats are sensitive to Pavlovian Outcome Devaluation, and this sensitivity is blocked by intra-DMS Rimonabant regardless of Tracking type. Data are represented as within-subject individual data (lines) and group data (bars; mean ± SEM). Rats received intra-DMS injections of either vehicle (left) or rimonabant (right) 10 minutes prior to probe test. **A,** In Male rats, we observed a significant main effect of Outcome Value and a significant Outcome Value X Treatment interaction. **B,** In Female rats, we did not observe any significant main effects or interactions. **C, D,** In ST rats, we observe a significant Outcome Value X Treatment X Sex interaction on total behavior. **C,** In Male ST rats we observed a significant Outcome Value X Treatment interaction. **D,** In ST Female rats there were no main effects or interactions. We then performed a parallel analysis in our GT/INT rats. **E, F** In GT/INT rats, we observe an Outcome Value X Treatment interaction, but no interaction with Sex. #p=0.067*p<0.05 **p<0.025

effects on this behavior.

In a prior study using male rats, it was established that initially devaluation insensitive ST rats become devaluation sensitive after extended training (Keefer, 2020). The present study replicates this finding and

shows that under vehicle conditions, male ST rats are sensitive to outcome devaluation (Fig. 3C, Bonferroni post-hoc; t(13)=2.679, p=0.037). Here we use both sexes and identify an Outcome Value x Treatment x Sex interaction in ST rats (F(1,24)=6.210, p=0.020), suggesting potential sex differences in devaluation sensitivity and/or effects of CB1R signaling inhibition. First, we confirmed the Outcome Value x Treatment interaction that was observed overall (Fig. 2A) is also observed in male ST rats (Fig. 3C, F(1,12)=5.063, p=0.044). Post-hoc analyses confirmed that intra-DMS rimonabant injections impaired devaluation sensitivity in male rats with similar levels of Pavlovian approach for valued and devalued conditions (t(13)=0.9205, p=0.7482). We found similar trends for male ST rats in lever contacts (the dominant response of ST rats) during outcome devaluation (Fig. 3-1A), in which there was a significant Outcome Value X Treatment interaction (F(1,13)=4.810, p=0.047) but post hoc tests did not reach significance even for the vehicle condition (t<2.484, p>0.0548). As expected, we observed no significant effects when analyzing male ST foodcup contacts (Fig. 3-2A). In contrast to males, female ST rats showed similar levels of responding in all probe tests and intra-DMS rimonabant had no effects (Total Behavior, Fig. 3D, *Fs*<1.236, *ps*>0.288; Lever, Fig. 3-1B; Foodcup, Fig. 3-2B).

Consistent with prior studies, male GT/INT rats were sensitive to outcome devaluation after extended training (main effect of Outcome Value (Fig. 3E; F(1,11)=5.203, p=0.043). In contrast to the ST group, we observed no main effects or interactions with Sex in GT/INT group. Despite this, we performed parallel analyses and found a marginal devaluation effect under vehicle condition in male GT/INT rats (t(11)=2.425, p=0.0675). For GT/INT the dominant response is food cup contacts, and for this measure there was a significant Outcome Value X Treatment interaction (Fig. 3-2C; F(1,11)=7.279, p=0.0207) and post hoc analysis revealed that under vehicle conditions, male GT/INT rats were sensitive to outcome devaluation (t(11)=2.872, p=0.0304) which was not the case with intra-DMS rimonabant (t(11)=0.8692, p=0.8066). We observed no significant differences when analyzing lever contacts alone (Fig. 3-1C). Female GT/INT rats showed a significant Outcome Value X Treatment interaction for total behavior (Fig. 3F; F(1,14)=5.100, p=0.040) that was driven by opposite patterns of behavior for the two treatments, however differences between value conditions did not reach significance for either treatment (vehicle, valued vs. devalued, t(14)=1.907, p=0.1545, rimonabant, valued vs. devalued (t(14)=1.329, p=0.410). We found a similar Outcome Value X Treatment interaction when looking at female GT/INT lever contacts alone (Fig. 3-1D; F(1,14)=4.953, p=0.043) but no significant interactions for food cup contacts (Fig. 3-2D); however, none of the post hoc analysis for these measures reached significance in female GT/INT rats.

Altogether, these results point to sex differences in Pavlovian outcome devaluation sensitivity and to treatment effects on Pavlovian devaluation sensitivity in male rats. Male rats are sensitive to devaluation after extended training, while female rats are not. The effects of intra-DMS CB1R blockade on devaluation sensitivity in male rats are consistent across tracking groups but are response specific. In male ST rats this sensitivity is driven by lever contacts, while in male GT/INTs, this sensitivity is driven by food cup contacts.

These effects of DMS CB1R signaling inhibition were specific to the satiety-specific outcome devaluation test. We found no difference in responding between intra-DMS vehicle and rimonabant groups during a non-sated,

non-reinforced Pavlovian lever autoshaping test of the same duration (10 trials, Fig. 2-1A; Sex x Treatment x

Response (lever, foodcup), $Fs<0.479$, $ps>0.493$). This suggests that intra-DMS rimonabant treatment effects

on Pavlovian approach emerge only after outcome-specific satiety. The observed effects are also not due to

differences in consumption between male and female rats during the 1-hour satiation period. To account for

body weight differences between male and female rats of the same strain and age, we normalized the amount

(g) of food consumed (either for the satiation period or post-probe choice test) to each rat's average body

weight across both days of outcome devaluation tests (Council, 1995; Lenglos et al., 2013). We found no

difference in the amount of food consumed during the satiation period prior to the probe test (g/bw chow Mean:

Male, 0.032, SEM ±0.002; Female, 0.031, SEM ±0.002; g/bw pellet Mean: Male, 0.039, SEM ±0.002; Female,

0.036, SEM ±0.002; $Fs<1.153$, $ps>0.288$). To confirm devaluation of the sated food, we gave rats a post-probe

choice test between the chow and training pellets (Fig. 2-1B) immediately after the end of the outcome

devaluation probe test. Rats consumed less of the food when they were sated on and more of the alternative

food, verified by a main effect of Outcome ($F(1,45)=8.134$, $p<0.007$) and this did not differ by sex or treatment

($Fs<1.790$, $ps>0.187$).

The observed effects of inhibiting CB1R signaling were also not evident during non-sated, reinforced Pavlovian

lever autoshaping sessions. We tested the effect of intra-DMS rimonabant infusion on a subset of rats (N=12)

during non-sated, reinforced PLA sessions and found no significant difference between vehicle, low (1µg/µl) or

high dose (2µg/µl) of rimonabant on lever presses (Fig. 2-2A; $Fs<1.972$, $ps>0.198$) or on foodcup contacts

(Fig. 2-2B; $Fs<1.078$, $ps>0.329$) across sex or tracking. We did observe a significant main effect of Sex for

lever contacts ($F(1,10)=5.395$, $p=0.043$), which is in line with acquisition data, during which females showed

more sign-tracking. Overall, rimonabant inhibition of DMS CB1R signaling did not impact conditioned approach

under reinforced conditions.

Baseline spontaneous IPSC recordings in DMS neurons differ between male and female Long Evans rats.

Based on the sex differences in behavioral flexibility,

we predicted that there may be differences in

inhibitory synaptic transmission in the DMS, in which

male rats may show reduced inhibitory synaptic

transmission. We recorded spontaneous IPSCs from

cells in the DMS in males and females (Fig. 4A). We

examined the mean amplitude (absolute value), the

mean frequency, or total events across the duration of

the recording, and the cumulative frequency

distribution for interevent interval, or the time between

event peaks, during 5-min recordings. We found that

there is no difference in the mean amplitude of DMS

sIPSCs between males and females when slices are

Figure 4: sIPSCs in DMS cells show reduced frequency and larger inter-event intervals in males as compared to females. *A,* Representative sIPSC recordings from DMS cells in aCSF bath of male (blue, n=8) and female (purple, n=9) Long Evans Rats. Scale bars: 20 pA and 1 sec. Data are presented as mean ± SEM. *B,* Mean Amplitude *C,* Mean Frequency *D,* Cumulative Frequency Plot of Interevent Interval. **p<0.025

perfused with an aCSF bath (Fig. 4B, t= -1.226, p=0.239). However, we did see a difference in both the

frequency and interevent interval. Male rats show a lower frequency as compared to females (Fig. 4C, t= -

2.561, p=0.022) and a larger inter-event interval (Fig. 4D, Kolmogorov-Smirnov test, D=0.2498, p<0.0001,

Hedge's *g* = 0.426). This difference in frequency and inter-event interval of sIPSCs suggests that male rats

show less inhibitory synaptic transmission onto recorded DMS neurons than females.

WIN 55, 21-2 bath application changes sIPSC measures in both males and females relative to DMSO bath

application.

We hypothesized that activation of DMS CB1R would reduce inhibitory synaptic transmission in male rats and

included females to investigate if there are sex differences in the effect of CB1R manipulation on sIPSCs in the

DMS. We recorded sIPSCs from DMS cells for 5 mins at baseline and following a 10-minute bath application of

a CB1R agonist, WIN 55,212-2 (WIN; 10μM, Fig. 5A). We found that there were no differences in the mean amplitude of sIPSCs due to WIN or Sex (Fig. 5B, *Fs*<1.182, *ps*>0.290). However, we did see differences in frequency and inter-event interval (Fig. 5C,D). We found a main effect of WIN for frequency (F(1,19)=6.306, p=0.021) but no main effect or interaction with Sex (*Fs*<0.825, p>0.375). We found that application of WIN shifted the interevent interval cumulative distribution curves to the right (Kruskal Wallis, H=1359, p<0.001) and post hoc comparisons confirmed that this occurred for both male and female rats (DMSO vs. WIN; Dunn's comparisons; male, p<0.0001, Hedges' *g* = 0.2085; female, p<0.0001, Hedges' *g* = 0.2291). This rightward shift suggests that WIN increases the interevent interval in both sexes. Application of WIN in the bath caused a

Figure 5: Activation of CB1R by WIN reduces sIPSC frequency and increases sIPSC interevent interval, regardless of Sex. *A,* Representative sIPSC recordings from DMS cells pre- (blue) and post-WIN (light blue) bath application for Long Evans Rats (n=21). Scale bars: 20 pA and 1 sec. Data are presented as mean ± SEM. *B,* Mean Amplitude with individual data for males (blue) and females (purple). *C,* Mean Frequency with individual data for males and females *D,* Cumulative Frequency Plot of Interevent Interval of sIPSCs. **p<0.025

reduction in the frequency of inhibitory events and an increase in the inter-event interval across all rats,

suggesting that CB1R located on presynaptic inhibitory inputs suppresses release of GABA in the DMS.

DISCUSSION

In the current studies we investigated the role of DMS CB1R signaling in Pavlovian outcome devaluation and

regulation of inhibitory synaptic transmission. We found that after extended training in PLA, males are sensitive

to outcome devaluation, while females are not and that DMS CB1Rs were necessary for the devaluation

sensitivity in males. Slice electrophysiology studies revealed that male rats showed a reduced frequency of

inhibitory events in the DMS as compared to females but that activating DMS CB1Rs reduced the probability of

GABA release similarly in both sexes.

The current results align with prior research that established significant individual-, experience-, and sex-dependent differences in Pavlovian devaluation. Consistent with previous studies (Pitchers et al., 2015; Keefer et al., 2022; Kochli et al., 2020), we found that female rats showed more lever-directed behaviors than males during extended training, but this difference diminished before testing in outcome devaluation (Fig. 1B). Under vehicle conditions, we replicated prior findings that male rats show devaluation sensitivity after extended training in PLA (Keefer, 2020). We extended this work to include females, for which we do not observe devaluation sensitivity after extended training (Fig. 2). These results echo the findings of other studies that indicate females are less sensitive to instrumental and Pavlovian devaluation (Bien and Smith, 2023; Quinn et al., 2007; Schoenberg et al., 2019; Sood and Richard, 2023). Additionally, further analyses into either lever or foodcup contacts revealed that male rats were sensitive to devaluation for their preferred response of their tracking group. ST male rats reduced lever contacts when the outcome is devalued (Fig. 3-1A), while GT/INT male rats reduce foodcup contacts (Fig. 3-2C) as has been shown previously in studies that examine Pavlovian outcome devaluation after extended training (Keefer, 2020; Keefer et al., 2022).

At first, we predicted that dorsal striatal CB1R signaling would promote rigid, or habitual, behaviors as has been shown for instrumental outcome devaluation (Gremel et al., 2016). However, our study suggests that CB1Rs in DMS promote behavioral flexibility in male rats, running counter to this established understanding. There are several factors that contribute to the divergence of results including species differences, the use of Pavlovian versus instrumental devaluation procedures and the subregion-specific effects of experimental manipulations. This prior study trained CB1R flox mice in both random-ratio (RR) and random interval (RI) schedules of instrumental reinforcement and generated an OFC-DS specific CB1R knockout. Competing action-outcome and stimulus-response associations mediate instrumental devaluation, and studies show that goal-directed behaviors shift to habit with extended training or with RR schedules of reinforcement (Adams, 1982; Adams and Dickinson, n.d.; Gremel et al., 2016). This is not the case with Pavlovian behaviors that are sensitive to devaluation even after extended training (Holland, 1998; Keefer et al., 2020), suggesting stimulus-outcome associations support adaptive reward seeking despite overtraining. Thus, differences in Pavlovian and instrumental processes may, in part, underline divergent findings between studies. Another possibility is methodological differences in the way CB1R were manipulated between studies. CB1R deletion in the OFC-DS projection promoted "goal-directed" devaluation sensitivity even during schedules of reinforcement that ordinarily drive "habitual" devaluation insensitivity (Gremel et al., 2016). Our current behavioral study inhibits CB1R signaling indiscriminately- likely affecting both inhibitory and excitatory synaptic transmission- rather than specifically on glutamatergic OFC afferents to the dorsal striatum, as in the prior study. Never-the-less, prior work has shown that systemic activation of CB1Rs promotes rigid responding (Hilário et al., 2007; Nazzaro et al., 2012) and while both DLS and DMS express CB1Rs (Hohmann and Herkenham, 2000; Fusco et al., 2004; Van Waes et al., 2012), more of the CB1R work within subregions of the DS has focused on the DLS. The DLS does express CB1R more densely than DMS, thus, it is possible that off-target effects impacted DLS function, an area with high CB1R density (Hohmann and Herkenham, 2000; Fusco et al., 2004; Van Waes et al., 2012) and this could confound our results. We think this is unlikely given the volume of rimonabant

injected (0.5 µL per hemisphere) and our ex vivo confirmation of reduced inhibitory synaptic transmission with CB1R activation in the DMS. The current targeting of DMS, as compared to DLS, may in part explain why our results diverge from observations that dorsal striatal CB1Rs support rigid responding via inhibition of glutamatergic inputs and our findings fit within the context of the DMS' role of biasing behavior towards "goal-directed" responding (Yin et al., 2005; Corbit and Janak, 2010; Gremel and Costa, 2013; Li et al., 2022).

These prior studies established that the DMS supports flexible, goal-directed instrumental conditioned responding. Reducing the activity of the DMS through lesion or pharmacological inhibition impairs flexible responding in a variety of tasks. To be interpreted in this conceptual framework, our behavioral pharmacology results suggest that CB1R signaling disinhibits the DMS, and thus, reducing CB1R signaling has a net inhibitory effect on DMS, resulting in impaired "goal-directed" Pavlovian devaluation sensitivity. Based on this interpretation, we hypothesized that DMS CB1R signaling at GABAergic inputs to DMS medium spiny neurons reduces inhibitory transmission in the area, allowing DMS activation to promote flexible responding in Pavlovian devaluation.

Our slice electrophysiology studies focused on inhibitory synaptic currents to investigate this hypothesis. At baseline, we found that males showed reduced inhibitory events as compared to females (Fig. 4). Within the above framework of striatal contributions to goal-directed and habitual control of behavior, lower levels of DMS inhibitory transmission (as seen in males) would promote flexibility and higher levels of inhibitory transmission (as seen in females) would prevent the expression of outcome devaluation, consistent with our devaluation findings in male and female rats, respectively. While we did not confirm the identity of the cells we recorded from, approximately 90% of cells across the dorsal striatum are medium spiny neurons (MSNs), the main type of projection neurons arising from the striatum (Graveland & Difiglia, 1985). Due to their abundance, we are likely to be recording from MSNs in the DMS. Multiple studies have shown that intact female rats and males treated with estradiol have increased striatal MSN excitability (Tansey et al., 2008; Dorris et al., 2015; Cao et al., 2018; Proaño et al., 2018) and estradiol decreases GABA release (Schultz et al., 2009). However, these studies are not specific to the DMS. Additionally, some studies have shown lower numbers of GABAergic neurons in males compared to females (Ovtscharoff et al., 1992), which may explain reduced inhibitory synaptic transmission in males. However, there are many types of GABAergic cells in the DMS. GABAergic Medium Spiny Neurons (MSNs) are the main projection neuron of the DMS, and they also project locally to other MSNs (Wilson and Groves, 1980; Somogyi et al., 1981; Graveland and Difiglia, 1985; Tunstall et al., 2002; Czubayko and Plenz, 2002; Burke et al., 2017). There are also multiple GABAergic interneuron types, predominately Parvalbumin positive fast-spiking interneurons (FSIs) and somatostatin interneurons (SOM). In fact, a study focusing on sex differences in the number of interneurons shows that some GABAergic interneurons are more dense in males than females (FSIs) while other interneurons are less dense in males than females (Van Zandt et al., 2024). Thus, further work must be done to isolate inhibitory synaptic transmission from these different sources and better understand sex differences in the DMS with cell-type specificity.

We show that CB1R activation reduces the frequency of inhibitory events regardless of sex. This should be interpreted with caution, as we only tested a single dose of the CB1R agonist. We applied WIN 55,212-2 at a concentration of 10 µM, which is a high concentration for bath application. Other studies use much lower doses and have seen sex differences in other brain regions (Tabatadze et al., 2015; Ferraro et al., 2020). Both males and females express CB1R in the dorsal striatum and males express CB1R more densely in the striatum and other brain regions than females (Laurikainen et al., 2019; Liu et al., 2020). Thus, it is possible that application of WIN at a lower dose may reveal more sensitivity to CB1R manipulation in males due to this higher concentration of receptors. Another caveat of these electrophysiological findings is that rats we recorded from did not have any behavioral training. It is possible that behavioral experience alters DMS inhibitory tone or changes DMS activity, as has been shown in other studies examining DMS activity after extended training or under different schedules of reinforcement (Fanelli et al., 2013; Gremel and Costa, 2013; Vandaele et al., 2019).

CB1Rs are located on multiple cell types in the dorsal striatum so further work must been done to identify the cell-type that supports Pavlovian flexibility in male rats. One notable possibility is the parvalbumin positive FSIs. CB1Rs are expressed on striatal PV-FSIs and mediate a form of inhibitory LTD that disinhibits MSNs, a mechanism that is associated with striatal regulation of behavioral flexibility (DePoy et al., 2013; Brian N. Mathur et al., 2013). CB1Rs are also expressed on cortical inputs that target MSNs and MSNs themselves (Gerdeman and Lovinger, 2001; Gerdeman et al., 2002; Wu et al., 2015; Lovinger and Mathur, 2012; Lovinger et al., 2022), but it has not yet been established whether cortical projections targeting PV-FSIs also contain CB1Rs. CB1R signaling at cortical-striatal FSI synapses would be expected to reduce inhibitory tone and increase DMS MSN activation, a similar result to CB1R signaling at FSI-MSN synapses. Direct manipulation of DLS PV-FSIs shows that their activity is critical to supporting habitual responding (O'Hare et al., 2017; Patton et al., 2020) but much less is known about DMS PV-FSIs and their contribution to habitual or goal-directed responding. Thus, these two hypotheses must be tested further to discover the mechanism of DMS CB1R regulation of Pavlovian devaluation sensitivity.

Overall, the current studies show that males are sensitive to Pavlovian outcome devaluation, a result that may be explained by reduced inhibitory synaptic transmission in the DMS. We find that the devaluation sensitivity of male rats requires DMS CB1R, but more work is needed to identify the cell-type specific population of CB1Rs that support flexible responding. Additionally, it is possible that DMS CB1Rs would be necessary for the devaluation sensitivity of females in cases where they respond flexibly at baseline. Thus, future studies should manipulate DMS endocannabinoids under conditions in which males and females respond similarly to determine if CB1Rs play a sex-specific role in mediating behavioral flexibility.

REFERENCES

Adams CD (1982) VARIATIONS IN THE SENSITIVITY OF INSTRUMENTAL RESPONDING TO REINFORCER DEVALUATION.
Adams CD, Dickinson A (n.d.) INSTRUMENTAL RESPONDING FOLLOWING REINFORCER DEVALUATION.

Amaya KA, Smith KS (2018) Neurobiology of habit formation. Curr Opin Behav Sci 20:145–152.

Amaya KA, Stott JJ, Smith KS (2020) Sign-tracking behavior is sensitive to outcome devaluation in a devaluation context-dependent manner: implications for analyzing habitual behavior. Learn Mem 27:136–149.

Bacharach SZ, Nasser HM, Zlebnik NE, Dantrassy HM, Kochli DE, Gyawali U, Cheer JF, Calu DJ (2018) Cannabinoid receptor-1 signaling contributions to sign-tracking and conditioned reinforcement in rats. Psychopharmacology (Berl) 235:3031–3043.

Bien E, Smith K (2023) The role of sex on sign-tracking acquisition and outcome devaluation sensitivity in Long Evans rats. Behav Brain Res 455:114656.

Burke DA, Rotstein HG, Alvarez VA (2017) Striatal local circuitry: a new framework for lateral inhibition. Neuron 96:267–284.

Cao J, Willett JA, Dorris DM, Meitzen J (2018) Sex differences in medium spiny neuron excitability and glutamatergic synaptic input: Heterogeneity across striatal regions and evidence for estradiol-dependent sexual differentiation. Front Endocrinol 9:173.

Corbit LH, Janak PH (2010) Posterior dorsomedial striatum is critical for both selective instrumental and Pavlovian reward learning. Eur J Neurosci 31:1312–1321.

Council NR (1995) Nutrient Requirements of Laboratory Animals. National Academies Press (US).

Czubayko U, Plenz D (2002) Fast synaptic transmission between striatal spiny projection neurons. Proc Natl Acad Sci 99:15764–15769.

DePoy L, Daut R, Brigman JL, MacPherson K, Crowley N, Gunduz-Cinar O, Pickens CL, Cinar R, Saksida LM, Kunos G, Lovinger DM, Bussey TJ, Camp MC, Holmes A (2013) Chronic alcohol produces neuroadaptations to prime dorsal striatal learning. Proc Natl Acad Sci U S A 110:14783–14788.

Dickinson A, Balleine B, Watt A (1995) Motivational control after extended instrumental training. Anim Learn Behav 23:197–206.

Dorris DM, Cao J, Willett JA, Hauser CA, Meitzen J (2015) Intrinsic excitability varies by sex in prepubertal striatal medium spiny neurons. J Neurophysiol 113:720–729.

Fanelli RR, Klein JT, Reese RM, Robinson DL (2013) Dorsomedial and dorsolateral striatum exhibit distinct phasic neuronal activity during alcohol self-administration in rats. Eur J Neurosci 38:2637–2648.

Ferraro A, Wig P, Boscarino J, Reich CG (2020) Sex differences in endocannabinoid modulation of rat CA1 dendritic neurotransmission. Neurobiol Stress 13:100283.

Flagel SB, Akil H, Robinson TE (2009) Individual differences in the attribution of incentive salience to reward-related cues: Implications for addiction. Neuropharmacology 56 Suppl 1:139–48.

Fusco F r., Martorana A, Giampà C, De March Z, Farini D, D'Angelo V, Sancesario G, Bernardi G (2004) Immunolocalization of CB1 receptor in rat striatal neurons: A confocal microscopy study. Synapse 53:159–167.

Geramita MA, Yttri EA, Ahmari SE (2020) The two-step task, avoidance, and OCD. J Neurosci Res 98:1007–1019.

Gerdeman G, Lovinger DM (2001) CB1 Cannabinoid Receptor Inhibits Synaptic Release of Glutamate in Rat Dorsolateral Striatum. J Neurophysiol 85:468–471.

Gerdeman GL, Ronesi J, Lovinger DM (2002) Postsynaptic endocannabinoid release is critical to long-term depression in the striatum. Nat Neurosci 5:446–451.

Graveland GA, Difiglia M (1985) The frequency and distribution of medium-sized neurons with indented nuclei in the primate and rodent neostriatum. Brain Res 327:307–311.

Gremel CM, Chancey JH, Atwood BK, Luo G, Neve R, Ramakrishnan C, Deisseroth K, Lovinger DM, Costa RM (2016) Endocannabinoid Modulation of Orbitostriatal Circuits Gates Habit Formation. Neuron 90:1312–1324.

Gremel CM, Costa RM (2013) Orbitofrontal and striatal circuits dynamically encode the shift between goal-directed and habitual actions. Nat Commun 4:2264.

Hammerslag LR, Gulley JM (2014) Age and sex differences in reward behavior in adolescent and adult rats. Dev Psychobiol 56:611–21.

Hilário MRF, Clouse E, Yin HH, Costa RM (2007) Endocannabinoid Signaling is Critical for Habit Formation. Front Integr Neurosci 1:6.

Hohmann AG, Herkenham M (2000) Localization of cannabinoid CB1 receptor mRNA in neuronal subpopulations of rat striatum: A double-label in situ hybridization study. Synapse 37:71–80.

Holland P (1998) Amount of training affects associatively-activated event representation, pp461–469. Pergamon.

Jordan CJ, Andersen SL (2017) Sensitive periods of substance abuse: Early risk for the transition to dependence. Dev Cogn Neurosci 25:29–44.

Kalivas PW, Volkow ND (2005) The Neural Basis of Addiction: A Pathology of Motivation and Choice. Am J Psychiatry 162:1403–1413.

Keefer SE, Bacharach SZ, Kochli DE, Chabot JM, Calu DJ (2020) Effects of Limited and Extended Pavlovian Training on Devaluation Sensitivity of Sign- and Goal-Tracking Rats. Front Behav Neurosci 14.

Keefer SE, Kochli DE, Calu DJ (2022) Inactivation of the Basolateral Amygdala to Insular Cortex Pathway Makes Sign-Tracking Sensitive to Outcome Devaluation. eNeuro 9:ENEURO.0156-22.2022.

King CP, Tripi JA, Hughson AR, Horvath AP, Lamparelli AC, Holl KL, Chitre A, Polesskaya O, Richards JB, Woods LCS, Palmer AA, Robinson TE, Flagel SB, Meyer PJ (2020) Sensitivity to food and cocaine cues are independent traits in a large sample of heterogeneous stock rats. bioRxiv 2020.05.13.066944.

Kochli DE, Keefer SE, Gyawali U, Calu DJ (2020) Basolateral Amygdala to Nucleus Accumbens Communication Differentially Mediates Devaluation Sensitivity of Sign- and Goal-Tracking Rats. Front Behav Neurosci 14.

Laurikainen H, Tuominen L, Tikka M, Merisaari H, Armio R-L, Sormunen E, Borgan F, Veronese M, Howes O, Haaparanta-Solin M, Solin O, Hietala J (2019) Sex difference in brain CB1 receptor availability in man. NeuroImage 184:834–842.

Lenglos C, Mitra A, Guèvremont G, Timofeeva E (2013) Sex differences in the effects of chronic stress and food restriction on body weight gain and brain expression of CRF and relaxin-3 in rats. Genes Brain Behav 12:370–387.

Li DC, Dighe NM, Barbee BR, Pitts EG, Kochoian B, Blumenthal SA, Figueroa J, Leong T, Gourley SL (2022) A molecularly integrated amygdalo-fronto-striatal network coordinates flexible learning and memory. Nat Neurosci 25:1213–1224.

Listunova L, Roth C, Bartolovic M, Kienzle J, Bach C, Weisbrod M, Roesch-Ely D (2018) Cognitive Impairment Along the Course of Depression: Non-Pharmacological Treatment Options. Psychopathology 51:295–305.

Liu X, Li X, Zhao G, Wang F, Wang L (2020) Sexual dimorphic distribution of cannabinoid 1 receptor mRNA in adult C57BL/6J mice. J Comp Neurol 528:1986–1999.

Lovinger DM, Mateo Y, Johnson KA, Engi SA, Antonazzo M, Cheer JF (2022) Local modulation by presynaptic receptors controls neuronal communication and behaviour. Nat Rev Neurosci 23:191–203.

Lovinger DM, Mathur BN (2012) Endocannabinoids in striatal plasticity. Parkinsonism Relat Disord 18:S132–S134.

Madayag AC, Stringfield SJ, Reissner KJ, Boettiger CA, Robinson DL (2017) Sex and Adolescent Ethanol Exposure Influence Pavlovian Conditioned Approach. Alcohol Clin Exp Res 41:846–856.

Mathur Brian N, Tanahira C, Tamamaki N, Lovinger DM (2013) Voltage drives diverse endocannabinoid signals to mediate striatal microcircuit-specific plasticity. Nat Neurosci 16:1275–1283.

Meyer PJ, Lovic V, Saunders BT, Yager LM, Flagel SB, Morrow JD, Robinson TE (2012) Quantifying Individual Variation in the Propensity to Attribute Incentive Salience to Reward Cues. PLOS ONE 7:e38987.

Morrison SE, Bamkole MA, Nicola SM (2015) Sign Tracking, but Not Goal Tracking, is Resistant to Outcome Devaluation. Front Neurosci 9:468.

Nasser HM, Chen YW, Fiscella K, Calu DJ (2015) Individual variability in behavioral flexibility predicts sign-tracking tendency. Front Behav Neurosci 9:289.

Navarro M, Carrera MR, Fratta W, Valverde O, Cossu G, Fattore L, Chowen JA, Gomez R, del Arco I, Villanua MA, Maldonado R, Koob GF, Rodriguez de Fonseca F (2001) Functional interaction between opioid and cannabinoid receptors in drug self-administration. J Neurosci 21:5344–50.

Nazzaro C, Greco B, Cerovic M, Baxter P, Rubino T, Trusel M, Parolaro D, Tkatch T, Benfenati F, Pedarzani P, Tonini R (2012) SK channel modulation rescues striatal plasticity and control over habit in cannabinoid tolerance. Nat Neurosci 15:284–293.

O'Hare JK, Li H, Kim N, Gaidis E, Ade K, Beck J, Yin H, Calakos N (2017) Striatal fast-spiking interneurons selectively modulate circuit output and are required for habitual behavior. eLife 6:e26231.

Ovtscharoff W, Eusterschulte B, Zienecker R, Reisert I, Pilgrim C (1992) Sex differences in densities of dopaminergic fibers and GABAergic neurons in the prenatal rat striatum. J Comp Neurol 323:299–304.

Patitucci E, Nelson AJD, Dwyer DM, Honey RC (2016) The Origins of Individual Differences in How Learning Is Expressed in Rats: A General-Process Perspective. J Exp Psychol Anim Learn Cogn 42:313–324.

Patton MS, Heckman M, Kim C, Mu C, Mathur BN (2020) Compulsive alcohol consumption is regulated by dorsal striatum fast-spiking interneurons. Neuropsychopharmacol 2020 462 46:351–359.

Paxinos G, Watson C (2006) The Rat Brain in Stereotaxic Coordinates - 6th Edition.

Peak J, Hart G, Balleine BW (2019) From learning to action: the integration of dorsal striatal input and output pathways in instrumental conditioning. Eur J Neurosci 49:658–671.

Pitchers KK, Flagel SB, O'Donnell EG, Woods LCS, Sarter M, Robinson TE (2015) Individual variation in the propensity to attribute incentive salience to a food cue: influence of sex. Behav Brain Res 278:462–469.

Proaño SB, Morris HJ, Kunz LM, Dorris DM, Meitzen J (2018) Estrous cycle-induced sex differences in medium spiny neuron excitatory synaptic transmission and intrinsic excitability in adult rat nucleus accumbens core. J Neurophysiol 120:1356–1373.

Quinn JJ, Hitchcott PK, Umeda EA, Arnold AP, Taylor JR (2007) Sex chromosome complement regulates habit formation. Nat Neurosci 10:1398–1400.

Ragozzino ME, Ragozzino KE, Mizumori SJY, Kesner RP (2002) Role of the Dorsomedial Striatum in Behavioral Flexibility for Response and Visual Cue Discrimination Learning. Behav Neurosci 116:105–115.

Schoenberg HL, Sola EX, Seyller E, Kelberman M, Toufexis DJ (2019) Female rats express habitual behavior earlier in operant training than males. Behav Neurosci 133:110–120.

Schultz KN, von Esenwein SA, Hu M, Bennett AL, Kennedy RT, Musatov S, Toran-Allerand CD, Kaplitt MG, Young LJ, Becker JB (2009) Viral vector-mediated overexpression of estrogen receptor-alpha in striatum enhances the estradiol-induced motor activity in female rats and estradiol-modulated GABA release. J Neurosci Off J Soc Neurosci 29:1897–1903.

Simmler LD, Ozawa T (2019) Neural circuits in goal-directed and habitual behavior: Implications for circuit dysfunction in obsessive-compulsive disorder. Neurochem Int 129:104464.

Smedley EB, Smith KS (2018) Evidence of structure and persistence in motivational attraction to serial Pavlovian cues. Learn Mem 25:78–89.

Somogyi P, Bolam JP, Smith AD (1981) Monosynaptic cortical input and local axon collaterals of identified striatonigral neurons. A light and electron microscopic study using the golgi-peroxidase transport-degeneration procedure. J Comp Neurol 195:567–584.

Sood A, Richard JM (2023) Sex-biased effects of outcome devaluation by sensory-specific satiety on Pavlovian-conditioned behavior. Front Behav Neurosci 17.

Tabatadze N, Huang G, May RM, Jain A, Woolley CS (2015) Sex Differences in Molecular Signaling at Inhibitory Synapses in the Hippocampus. J Neurosci 35:11252–11265.

Tansey EM, Arbuthnott GW, Fink G, Whale D (2008) Oestradiol-17β Increases the Firing Rate of Antidromically Identified Neurones of the Rat Neostriatum. Neuroendocrinology 37:106–110.

Thoma P, Wiebel B, Daum I (2007) Response inhibition and cognitive flexibility in schizophrenia with and without comorbid substance use disorder. Schizophr Res 92:168–180.

Tunstall MJ, Oorschot DE, Kean A, Wickens JR (2002) Inhibitory Interactions Between Spiny Projection Neurons in the Rat Striatum. J Neurophysiol 88:1263–1269.

Van Waes V, Beverley JA, Siman H, Tseng KY, Steiner H (2012) CB1 cannabinoid receptor expression in the striatum: Association with corticostriatal circuits and developmental regulation. Front Pharmacol 3 MAR:21.

Van Zandt M, Flanagan D, Pittenger C (2024) Sex differences in the distribution and density of regulatory interneurons in the striatum. Front Cell Neurosci 18.

Vandaele Y, Mahajan NR, Ottenheimer DJ, Richard JM, Mysore SP, Janak PH (2019) Distinct recruitment of dorsomedial and dorsolateral striatum erodes with extended training. eLife 8.

Villaruel FR, Chaudhri N (2016) Individual Differences in the Attribution of Incentive Salience to a Pavlovian Alcohol Cue. Front Behav Neurosci 10:238.

Wilson CJ, Groves PM (1980) Fine structure and synaptic connections of the common spiny neuron of the rat neostriatum: A study employing intracellular injection of horseradish peroxidase. J Comp Neurol 194:599–615.

Wu YW, Kim JI, Tawfik VL, Lalchandani RR, Scherrer G, Ding JB (2015) Input- and cell-type-specific endocannabinoid-dependent LTD in the striatum. Cell Rep 10:75–87.

Yin HH, Knowlton BJ, Balleine BW (2004) Lesions of dorsolateral striatum preserve outcome expectancy but disrupt habit formation in instrumental learning. Eur J Neurosci 19:181–189.

Yin HH, Ostlund SB, Knowlton BJ, Balleine BW (2005) The role of the dorsomedial striatum in instrumental conditioning. Eur J Neurosci 22:513–523.

87
88
89
90

A

Total Behavior (Lever + Foodcup Contacts)

☐ Valued
▨ Devalued

**

Vehicle   Rimonabant

B

+1.44
+0.96
+0.72
+0.48
+0.12