Two neuropeptides that promote blood-feeding in *Anopheles stephensi* mosquitoes

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Animals routinely need to make decisions about what to eat and when. These decisions are influenced not only by the availability and quality of food but also by the internal state of the animal, which needs to compute and give weights to these different variables before making a choice. Feeding preferences of female mosquitoes exemplify this behavioural plasticity. Both male and female mosquitoes usually feed on carbohydrate-rich sources of nectar or sap, but the female also feeds on blood, which is essential for egg development. This blood-appetite is modulated across the female's reproductive cycle, yet little is known about the factors that bring it about. We show that mated, but not virgin *Anopheles stephensi* females, a major vector of urban malaria in the Indian sub-continent and West-Africa, suppress blood-feeding between a blood-meal and oviposition. We identify several candidate genes through transcriptomics of blood-deprived and -sated *An. stephensi* central brains that could modulate this behaviour. We show that *short Neuropeptide-F (sNPF)* and *RYamide (RYa)* act together to promote blood-feeding and identify a cluster of cells in the subesophageal zone that expresses *sNPF* transcripts only in the blood-hungry state. Such females also have more *sNPF* transcripts in their midguts. Based on these data, we propose a model where increased *sNPF* levels in the brain and gut promotes a state of blood-hunger, which drives feeding behaviour either by *sNPF's* action in the two tissues independently or via a communication between them.

Anopheles stephensi **| Blood-feeding | Host-seeking | Transcriptomics |** ***sNPF*** **|** ***RYamide***

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Introduction and Results. An animal's dietary choice is an outcome of its internal physiological state and external stimuli. Its changing nutritional demands, reproductive state, and developmental history are integrated with external cues before a food-choice is made. For example, when deprived of a particular nutrient, like proteins, animals show a specific increase in protein-appetite (1–3). Mating influences food choices of female flies by increasing their preference for proteins, sugars, and salt for egg development and oviposition (4–8).

Changes in both sensory perception and central brain circuits have been shown to underlie such behavioural changes. For example, sensory neurons of starved animals become more sensitive to attractive cues and less sensitive to aversive ones, both in vertebrates and invertebrates (9–16). These changes are often brought about by neuromodulators and neuropeptides (9, 10, 17–22). In the central brain, internal nutrient or hormone sensors can monitor the animal's nutritional state. In mice, neurons in the hypothalamus express receptors for hunger and satiety signals like ghrelin and leptin (16, 23, 24). In insects, several neurons in the mushroom body, central complex, and subesophageal zone (SEZ) have been shown to integrate hunger and satiety signals to regulate feeding on sugars or proteins, choose between exploration or exploitation of food sources, or initiate or terminate feeding (25–33).

Female mosquitoes offer a striking model for dissecting how internal state and external cues shape dietary choices. Many female mosquitoes have a dual appetite for carbohydrate-rich nectar or sap – to meet energetic requirements – and protein-rich blood – for egg development. Which meal a female chooses to take is influenced by nutritional demands, reproductive state, and developmental history. For example, mosquitoes with low teneral reserves will take multiple blood meals for egg development (34–37). Nutritional state also influences other internal state outputs on blood-feeding. For example, whether virgin females take blood-meals or not is dependent on prior sugar-feeding in most *Aedes* strains (38) (and references within). However, mating is dispensable in Anophelines even under conditions of nutritional satiety (36, 39–41).

Despite these species- and context-dependent variations, a consistent finding is that female mosquitoes oscillate between states of heightened and suppressed host-seeking across successive reproductive cycles (figure 1A). For example, in *Ae. aegypti* and *An. gambiae*, newly emerged mosquitoes display no or low interest in the host, which increases as they age (34, 42–44) and once fed to repletion, host attraction is suppressed. The strength and timing of this suppression varies between species — being robust and well-characterised in mated *Ae. aegypti* (45–47), and more flexible in Anophelines (36, 48, 49). Sugar-feeding is not modulated in this way, in some cases it is preferred soon after emergence (43, 50).

The modulation of blood-feeding provides a useful framework for understanding how internal and external cues shape meal-choice. What underlying molecular cues represent internal physiological states to influence dietary choice is of interest not only because of its importance in public health, but also because such understanding has implications for other organisms. In *Aedes albopictus* anticipatory expression of the yolk protein precursor gene *vitellogenin 2* in the fat body promotes host-seeking behaviour (51). In *Aedes aegypti*, neuropeptide-Y receptor (NPYLR7), and more recently, short Neuropeptide F (sNPF) and neuropeptide RYamide (RYa) have been shown to suppresses host-

attraction, while Neuropeptide F (NPF) promotes it (47, 52–55). Factors from the host can also both promote (ATP) (56) and terminate feeding (Fibrinopeptide A)(57).

Here we describe the blood- and sugar-feeding patterns of *Anopheles stephensi*, which is endemic to the Indian sub-continent but has expanded its range into West Africa, putting millions more people at risk of malaria (58). Neurotranscriptomics across the different conditions of blood-deprivation and -satiety allowed us to shortlist several candidate molecules that might promote blood-feeding behaviour. Through dsRNA mediated knockdown of nine of them, we identified two neuropeptide genes – *sNPF* and *RYa* - that together promote blood-feeding through its action in the brain and possibly the abdomen. We find that *sNPF* expression pattern reflects this requirement: a cluster of cells in the SEZ expresses the *sNPF* transcripts only in blood-hungry females and such females also have more *sNPF* transcripts in their midguts. *RYa* is not modulated similarly. Both their receptors are expressed in the brain and the *sNPF receptor* is also expressed in the midgut. Together, these data are consistent with a model where increase in levels of *sNPF* in the brain and the midgut promotes a state of blood-hunger in *An. stephensi*.

Blood-, but not sugar-feeding, is modulated across the reproductive cycle. To assess the female *An. stephensi* appetite for blood through the reproductive cycle we designed a behavioural assay (figure 1B, S1A; see methods for details), and tested it on *Ae. aegypti*, a well-studied model. We were able to reproduce the previously reported results that mating enhances blood-feeding in *Aedes* (59–61) and that once fed to repletion, females actively suppress blood-feeding until oviposition (62) (figure 1C).

We used this assay on *An. stephensi*, and employed Generalised Linear Mixed Model (GLMM) to compare appetites across conditions (see supplementary sheet1). We found that these females were uninterested in blood when they emerge but that their appetite increased with age (magenta, D0-D5 in figure 1D, supplementary sheet1: model 1). Once fed to repletion, like *Ae. aegypti*, *An. stephensi* also suppress blood-feeding for at least four days after a blood-meal (magenta, $D1^B$-$D4^B$ in figure 1D) and until oviposition, after which, the suppression was lifted to pre-blood-meal levels (magenta, $D1^O$-$D2^O$ in figure 1D). Unlike blood-feeding, sugar-feeding was not modulated similarly barring a brief suppression post-blood-meal, which was not dependent on oviposition (figure 1E; methods and S1G for details, supplementary sheet1: model 4, 5, 6).

To ascertain whether sugar and blood represent parallel appetites independent of each other, we presented sugar-sated co-housed females a choice between the two. Under these conditions, 70% of the females chose to feed on blood alone and only 21% took both meals (figure 1F). When co-housed females were deprived of sugars for a day prior to testing, the pattern revered: 68% of the females now took both sugar and blood meals and only 24% fed on blood alone (figure 1F). As expected, males fed robustly on sugar in both conditions (grey, figure 1F).

Since females took blood-meals regardless of their prior sugar-feeding status and only sugar-feeding was selectively suppressed by prior sugar access, we infer that these two represent parallel appetites regulated independently. The fact that most females in the starved group took both sugar- and blood-meals, while most females in the sated condition ignored sugar, supports this interpretation. An alternate interpretation of these data are that two represent hierarchical appetites: that some sugar feeding is necessary to stimulate blood-feeding. Determining the order of feeding events in the choice assay would be necessary to comment on this. However, prior reports that females can survive on blood-meals alone (with no access to plant sugars (36)) argue against this model. Together, these data suggest that in newly-emerged females, blood and sugar represent parallel appetites. We note that their interdependence may emerge later in the reproductive cycle ($D1^B$-$D2^B$ in figure 1E).

As mating promotes the first blood-meal in *Ae. aegypti*, we tested if this was true for *An. stephensi*. A post-hoc dissection of each female's spermatheca (where sperm is stored after mating) revealed that many of the blood-fed females in the above assay were unmated (figure 1G). To confirm this, we assayed blood-feeding behaviour in virgin females and found that virgin *An. stephensi*, unlike *Ae. aegypti*, have a robust appetite for blood and that mating enhances it only marginally (cyan, D0-D5, figure 1D, supplementary sheet1: model 1, 2). Interestingly, however, these virgins, unlike mated *An. stephensi* (or *Ae. aegypti)* females, failed to suppress their blood-appetite, even after engorging on blood. Instead, they continued to blood-feed for up to four days after their first blood-meal (cyan, $D1^B$ - $D4^B$ in figure 1D, supplementary sheet1: model 3). A flipped order of the two behaviours - virgins were first given a blood-meal then allowed to mate – also resulted in suppression of the appetite (figure 1H). This suggests that the post-blood-meal suppression of blood-appetite in *An. stephensi*, is dependent upon mating status.

Together, these data describe female *An. stephensi's* feeding behaviours: 1) in newly- emerged females blood and sugar represent parallel appetites, 2) blood-, but not sugar-appetite (barring a brief effect), is suppressed between a blood-meal and oviposition, and 3) mating is not necessary for blood-feeding in newly-emerged females, but it is necessary for its suppression between a blood-meal and oviposition.

Host-seeking is modulated across the reproductive cycle. The modulation of blood-feeding through the reproductive cycle could be due to 1) modulation in peripheral sensitivity to host kairomones, 2) modulation by central brain circuits, or 3) modulation in both peripheral and central circuit elements. Extensive data in other mosquito species suggest that sensitivity to host kairomones change as females age (42, 63–66) and following a blood-meal (49, 67, 68). We tested this in *An. stephensi*.

Because of the multi-sensory nature of host-seeking, we used a two-choice behavioural assay where multiple host-cues were provided in one arm while the other had none. Mosquitoes had to make a choice between the two arms (fig-

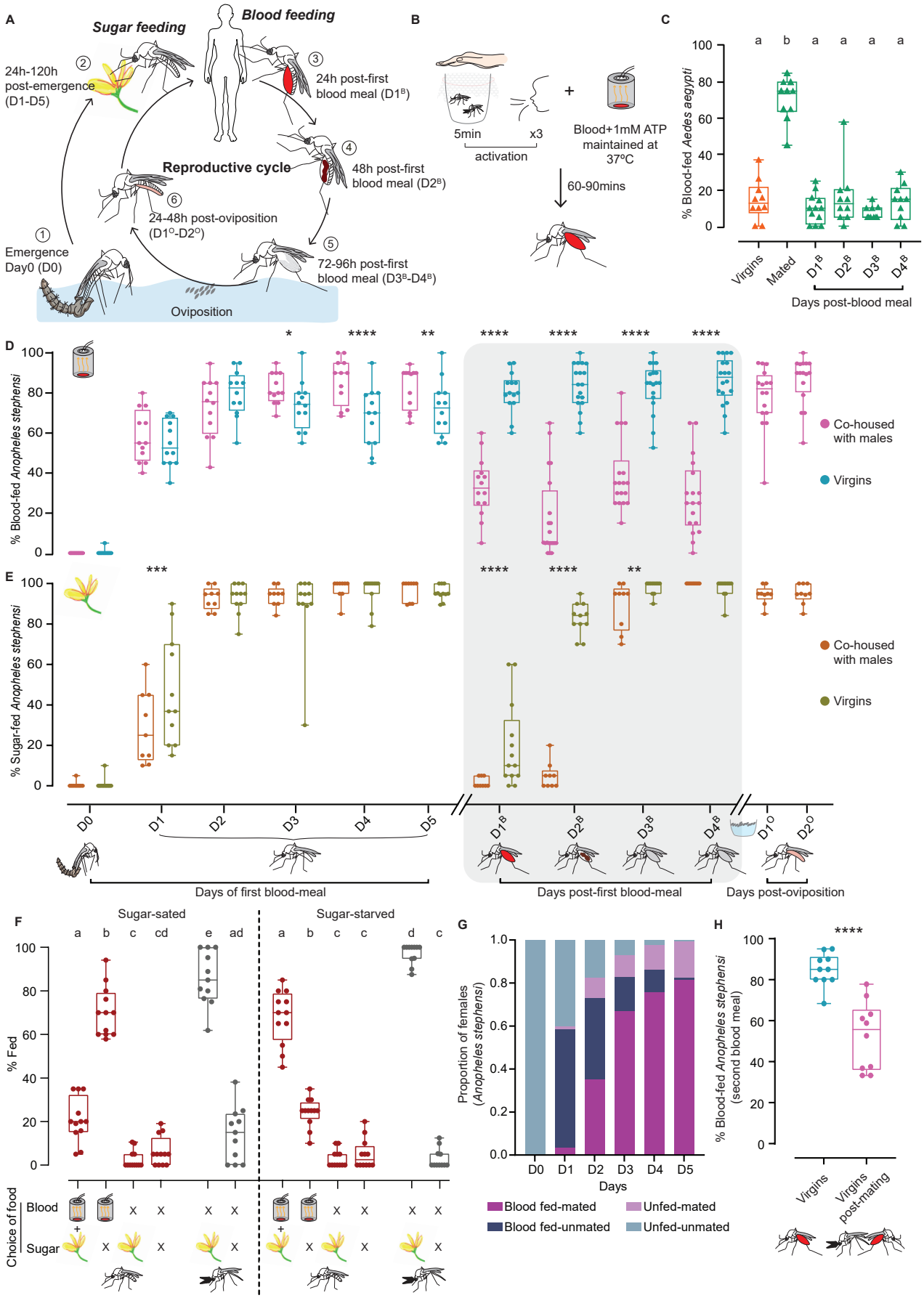
A
Sugar feeding
Blood feeding
24h-120h post-emergence (D1-D5)
24h post-first blood meal (D1B)
48h post-first blood meal (D2B)
Reproductive cycle
24-48h post-oviposition (D1O-D2O)
72-96h post-first blood meal (D3B-D4B)
Emergence Day0 (D0)
Oviposition
B
5min
x3
activation
Blood+1mM ATP maintained at 37ºC
60-90mins
C
% Blood-fed Aedes aegypti
Virgins
Mated
D1B
D2B
D3B
D4B
Days post-blood meal
D
% Blood-fed Anopheles stephensi
Co-housed with males
Virgins
E
% Sugar-fed Anopheles stephensi
Co-housed with males
Virgins
D0
D1
D2
D3
D4
D5
D1B
D2B
D3B
D4B
D1O
D2O
Days of first blood-meal
Days post-first blood-meal
Days post-oviposition
F
Sugar-sated
Sugar-starved
% Fed
Choice of food
Blood
Sugar
G
Proportion of females (Anopheles stephensi)
Days
Blood fed-mated
Blood fed-unmated
Unfed-mated
Unfed-unmated
H
% Blood-fed Anopheles stephensi (second blood meal)
Virgins
Virgins post-mating

Fig. 1. Feeding behaviours of *An. stephensi*. A. Reproductive cycle of *An. stephensi*. Upon emergence [1, D0-D5] females have a dual appetite for protein-rich blood and carbohydrate-rich sources of sugar [2,3]. After a blood-meal [3,4,5, $D1^B$-$D4^B$], the eggs mature and are laid in water [6]. The next reproductive cycle can now begin. B. Blood-feeding assay. 0-7h old mosquitoes were collected in cups and aged appropriately on sugar. Prior to the test, mosquitoes were 'activated' for 5 minutes by presenting a human hand, followed by 3 exhalations. Blood-meals were presented through a Hemotek perfumed with human skin odours. Number of blood-fed mosquitoes were visually scored, irrespective of the quantity fed. C. Blood-feeding behaviour of *Ae. aegypti*. 8-days old virgin and mated females were assessed for their first blood-meal. Fully fed mated females were assessed for second blood-meal ($D1^B$-$D4^B$). Each dot represents a single trial with 18-21 females, n=10-12trials/group. Kruskal-Wallis with Dunn's multiple comparisons test, $p<0.05$. Data labelled with different letters are significantly different. D. Blood-feeding behaviour of *An. stephensi*. Co-housed and virgin females were tested on the day of emergence (D0) and each day for the next 5 days (D1-D5). $D1^B$-$D4^B$ represents their blood-feeding 1-4 days after their first blood-meal and $D1^O$-$D2^O$ represents their blood-feeding 1-2 days after oviposition (see methods for assay details). 18-21 females/trial, n=9-20trials/group. Generalized Linear Mixed Model with post-hoc pairwise comparisons using estimated marginal means and Bonferroni-corrected p-values; *$p<0.05$; **$p<0.01$; ****$p<0.0001$. E. Sugar-feeding behaviour of *An. stephensi*. Females of similar conditions as in (D) were tested for sugar-feeding. 18-21females/trial, n=9-20trials/group. Generalized Linear Mixed Model with post-hoc pairwise comparisons using estimated marginal means and Bonferroni-corrected p-values; **$p<0.01$; ***$p<0.001$; ****$p<0.0001$. F. Choice assay for blood and sugar. Sugar-sated females or sugar-starved females, co-housed with males (indicated by the dashed abdomens) were given a choice between blood and sugar and assessed for their choice of food (bottom): blood and sugar; blood only; sugar only; none. While most sugar-sated females took blood (left of dashed line), most sugar-starved females fed on both (right of dashed line). As an internal control, co-housed males were accessed for sugar-feeding only and most were found to be fed in both conditions. 17-24 mosquitoes/trial; n=11-12 trials/condition. One-way ANOVA with Holm-Šídák multiple comparisons test, $p<0.05$. Data labelled with different letters are significantly different. G. Mating and blood-feeding. The mating status of D0-D5 co-housed females assayed in (D) were determined post-hoc by dissecting their spermathecae. n=232-239 females were analysed for each day. Many blood-fed females were unmated. H. Mating after blood-feeding. Blood-fed virgins were allowed to mate and then offered a second blood-meal. A suppression in blood-appetite was observed. 15-20females/trial, n=10 trials. Unpaired t-test; ****$p<0.0001$.

ure 2A). When no host-cues are provided in either (blank) even blood-hungry females distribute equally between the two arms demonstrating the assay has no bias (black, figure 2B). Using this assay, we found that newly emerged females showed no preference for either the control or the host arm (figure 2B). Five days later, they were strongly attracted to the host-cues and this attraction was suppressed after their blood-meal and until oviposition (magenta, figure 2B). This general trend in attraction to host-cues tracks the trend in blood-feeding. Interestingly, the response of virgin females to host-cues in the Y-maze was highly variable under all conditions tested (cyan, figure 2B). Yet, like in the blood-feeding assay, blood-fed virgins were comparatively more attracted to human host-cues than were the mated blood-fed females (figure 2B).

These data suggest that, like blood-feeding, host-seeking is also modulated through the female's reproductive cycle, and mating influences this modulation by enhancing it initially and suppressing it post-blood-meal. The variability in response of virgin females suggests that mating enhances the female's initial attraction to a host.

Neurotranscriptomics reveals several candidate genes that promote blood-feeding. While sensitivity to host kairomones may modulate blood-feeding, it is also possible that central brain circuitry contributes to it. We reasoned that cues from mating, or signals from other tissues such as midgut could act on brain circuitry to modulate blood-feeding. To determine the molecular underpinnings of this behavioural modulation, we performed RNA sequencing from the central brains of mosquitoes at different stages of their reproductive cycle (consequently different states of blood-deprivation or -satiety). This included females at emergence (D0), 5-days after emergence (D5; sugar-fed, blood-deprived), virgins and mated females one day after a blood-meal ($D1^B$, $D1^B$M), and females one day after oviposition ($D1^O$). Age-matched males were included for the earliest two time points (figure 3A).

For each of our samples, 81-87% of about 20-30million reads aligned to the *An. stephensi* genome (69). The variance between our replicates was low as represented in the Principal Component Analysis (PCA) (figure 3B). In PCA, older female brains, regardless of mating-, blood-feeding-, or oviposition- status clustered together while the youngest female and male brains stood apart on PC1 and PC2 suggesting that age and sex contribute most significantly to the variance in our data (figure 3B). The expression pattern of some hallmark genes showed the expected trends. Genes involved in eclosion behaviours – *eclosion hormone* (70) and *partner of bursicon* (71) – showed the highest expression in the D0 samples and were not expressed later. Male-specific genes, *HMG-B-13-like*, were expressed only in males (72), and genes involved in vitellogenic stages of egg-development – *vitellogenins*, *Ecdysone receptor*, and *E-20-monooxygenase* (73)– were expressed only in post-blood-meal samples (figure 3C).

We next explored this data for insights into the molecular underpinnings of blood-feeding modulation. Reports suggest that anticipatory metabolic changes in female germline cells and a 'vittelogenin wave' in fat bodies can drive specific appetite in *Drosophila* (74) and *Ae. albopictus* (51) respectively. However, we did not find any evidence in the neurotranscriptomes for a similar anticipatory metabolic change that might drive blood-feeding in the mosquito brain (figure S2B). Interestingly, we found that after a blood-meal, glucose is neither spent nor stored, and that the female brain goes into a state of metabolic 'sugar rest', while actively processing proteins (figure S2B, S3). We similarly plotted genes involved in lipid metabolism, neurotransmitters, neuropeptides and their receptors and found no significant patterns (figure S4, S5, S6). In summary, contrary to our expectations, we saw no evidence in the neurotranscriptomes for anticipatory metabolic changes in the brain that might in turn drive blood-feeding behaviour. It is possible, however, that other forms of regulation that do not involve transcriptional changes could still influence metabolism.

In an alternate approach to identifying genes that might modulate blood-feeding, we took the intersection of genes that were enriched in the three blood-hungry conditions (and absent in males). These comparisons were made against the two conditions that were not motivated to blood-feed: females at emergence and mated-blood-fed females (figure

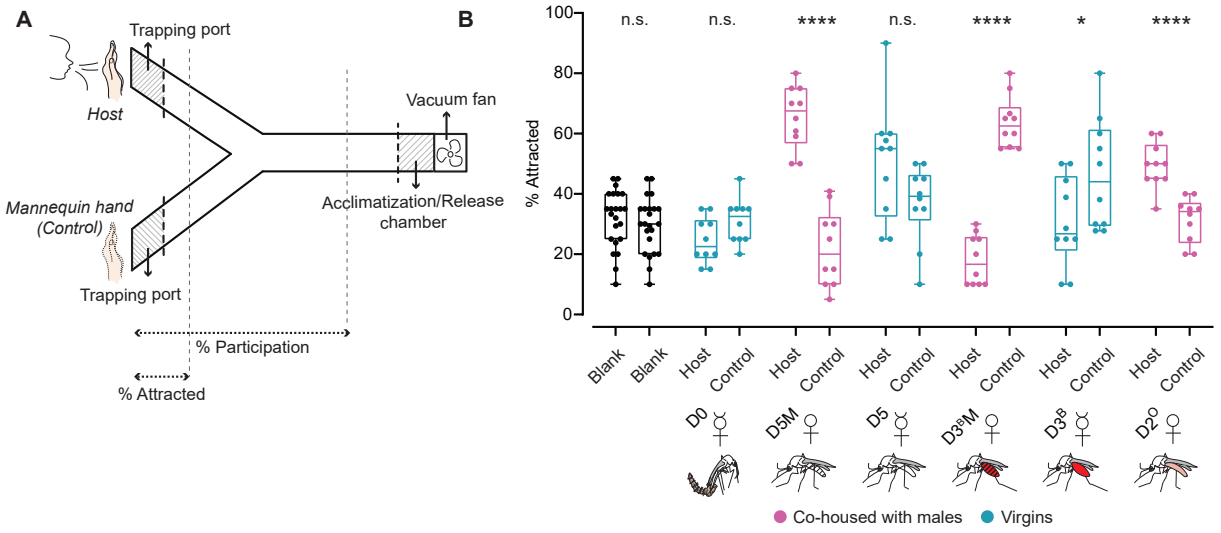

Fig. 2. Host-seeking is modulated like blood-feeding is in *An. stephensi*. A. Schematic of the Y-maze used to assess host-seeking behaviour of female *An. stephensi*. Females were allowed to acclimatise in the chamber for 5 minutes while being exposed to host kairomones that were presented in a test arm. A fan sucked the air from both host and control arms at 0.3-0.6m/s. Mosquitoes were released after acclimatisation and allowed to choose between the two arms. B. Percent females attracted to either host-cues or control arm at the indicated time points in the Y-maze. 17-21 females/trial, n=10 trials/group. Unpaired t-test; n.s.: not significant, p>0.05.; * p<0.05; ****p<0.0001. (D0: day of emergence ; D5M: 5days post-emergence, mated; D5: 5days post-emergence, virgin; $D3^{B}M$: 3days post-blood-meal, mated; $D3^{B}$: 3days post-blood-meal, virgin; $D2^{O}$: 2days post-oviposition)

3D). These shortlisted 38 and 34 genes respectively, gave us 68 unique candidates (excluding common genes and isoforms) that might promote blood-feeding in *An. stephensi* (figure S7). Notably, we did not find any genes to be differentially expressed between the two blood-fed conditions (virgins and mated), despite their contrasting behaviours (figure S2A). The strongest signatures we observed in these females were those related to metabolism (figure S2B). It is possible that the differences lie in small populations of cells, which get obscured in the averaged transcriptomes of the entire brain.

Neuropeptides *sNPF* and RYamide promote blood--feeding in *An. stephensi*. We employed multiple, independent criteria while further short-listing candidates from the above analysis: 1) genes that showed the expected expression pattern: low in newly-emerged males and females; high in blood-hungry females; low in blood-sated females, 2) all neuropeptides, since these are known to regulate feeding behaviours, and 3) all genes related to juvenile hormone (JH) and 20-hydroxyecdysone (20E) signalling pathway, since they regulate development and reproduction in mosquitoes. This gave us 9 candidates (figure 3E): Two sodium-dependent amino acid transporters *118504075 and 118504077*. Transporters can sense nutrients and regulate feeding in *Drosophila* (29, 75). The neuropeptides included *Orcokinin (Ork)*, *Prothoracicostatic peptides (Prp)*, *RYamide (RYa)* and *short Neuropeptide F (sNPF)*, and a metabolic hormone *Eiger*, which responds to low nutrient conditions in *Drosophila* (76). *Protein Hairy* and *Juvenile hormone epoxide hydrolase 1-like (JHEH)* are involved in the JH pathway (77, 78)(figure 3E).

Using dsRNA, we knocked each of these genes down and tested their blood-feeding behaviour. In each case, we ensured efficiency of gene knockdown post-hoc (figure 4A). We first confirmed that the injection itself did not cause any behavioural changes (figure 4B) and in all following experiments either *dsGFP*-injected or uninjected females were included as controls.

We saw no change in behaviour upon loss of *Eiger*, *JHEH*, and *Ork* function (figure S8A-C, S8A'-C'). We were able to downregulate *Hairy* and *Prp* transcript levels by only 30-35%, hence cannot comment on their role in modulating blood-feeding (figure S8D, S8E, S8D', S8E'). Downregulation of the two transporters was efficient and resulted in a marginal reduction in blood-feeding (figure S8F, S8G, S8F', S8G'). However, their combined knockdowns, though effective, did not show any significant change in behaviour (figure S8H, S8H').

Two neuropeptides - *sNPF* and *RYa* - showed about 25% and 40% reduced mRNA levels in the heads and these animals showed no change in blood-feeding (figure S9B, S9C, S9E, S9F). We noticed, however, that many of these animals did not engorge and instead took smaller blood-meals (figure S9D, S9G). Given this, and since neuropeptides are known to act in concert (79), we tested the possibility that *sNPF* and *RYa* might act together to regulate blood-feeding behaviour. Indeed, their simultaneous dsRNA injections resulted in about 40% of the animals not taking blood-meals (figure 4C). Since RNA interference (RNAi) can be variable, and each female is an independent experiment, we pooled the females that took blood-meals and those that didn't separately before quantifying mRNA from them. We noted that knockdown efficiency was higher in the heads of females that did not take blood-meals suggesting that downregulating these genes caused the suppression of blood-feeding (figure 4D). Importantly, since these females were offered both blood and sugar, we noted that their sugar-feeding was unaffected (figure 4C).

Since RNAi results in systemic knockdowns and neuropep-

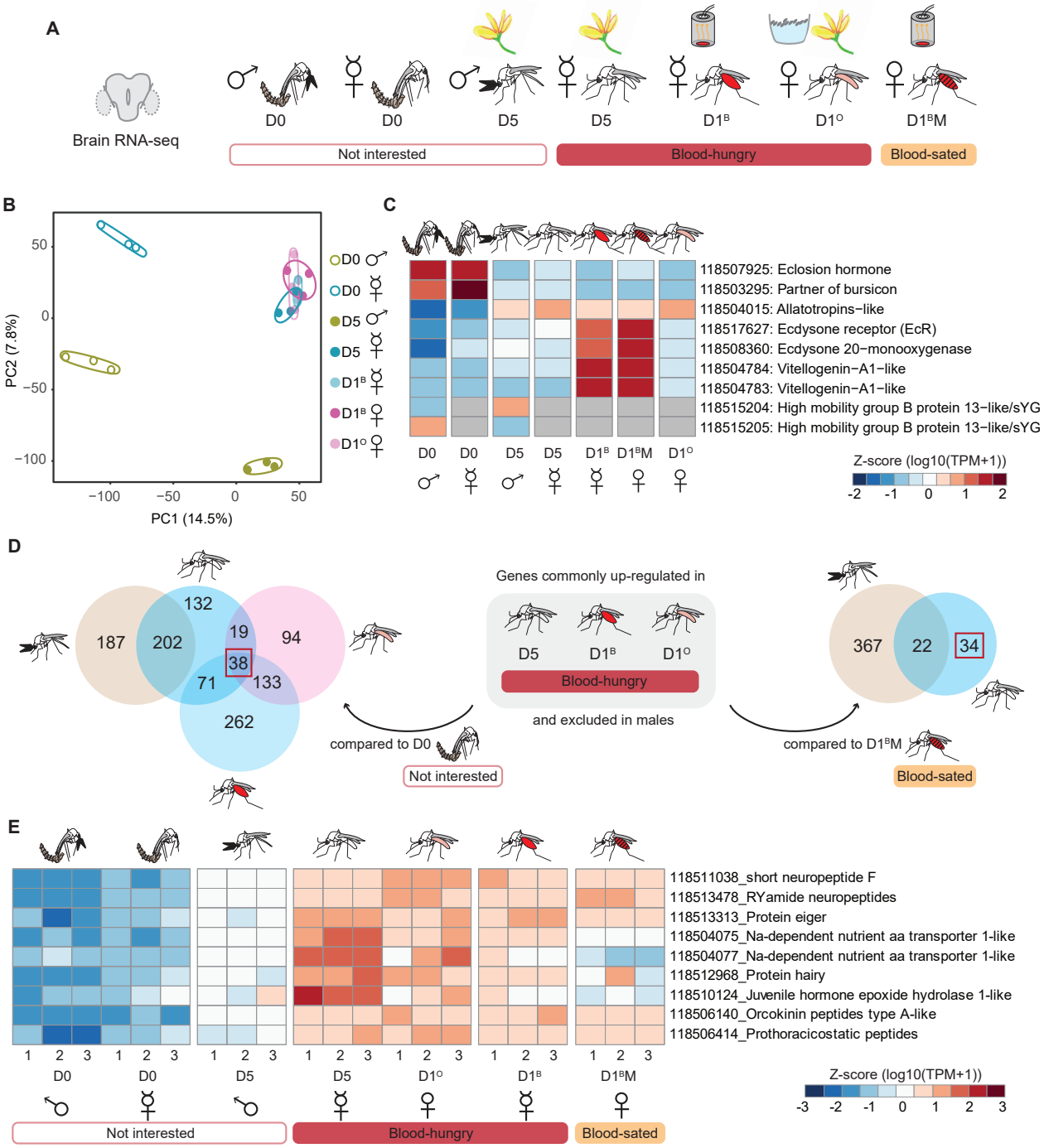

Fig. 3. Neurotranscriptome of *An. stephensi*. A. Central brains were sampled from males and females at different ages and feeding preferences for bulk RNA-seq. Uninterested in blood: D0 males, D0 females, D5 males. Blood-hungry: D5, sugar-fed virgin females, $D1^B$ virgin females, and $D1^O$ females. Blood-sated: $D1^B$ mated females. n=100 central brains per replicate; 3 biological replicates per sample, 7 samples. B. Principal Component Analysis (PCA) of the normalised counts-per-million from different brain RNA-seq samples. Replicates cluster closely, and age and sex contribute most to the differences between the samples. C. Genes (listed on the right) with known expression patterns plotted across different samples. These known expression patterns are reproduced in our data. Average of the triplicate data are represented as z-score normalised log10 (TPM+1) values. Grey cells represent no data. D. Identification of candidates potentially involved in promoting blood-feeding behaviour. Two types of comparisons were made: (left) genes commonly up-regulated in the three blood-hungry conditions (see figure 3A) when compared against D0 females and excluded in males, and (right) genes up-regulated in D5 sugar-fed virgin females when compared against $D1^B$ mated females (blood-sated), and excluded in males. Number of candidate genes identified from two different sets of analyses are boxed in red. E. Nine candidate genes (listed on the right), shortlisted for further validation. RNA-seq data from central brain samples in triplicates are represented as z-score normalised log10 (TPM+1) values. (D0: day of emergence; D5: 5days post-emergence, virgin; $D1^B$: 1day post-blood-meal, virgin; $D1^BM$: 1day post-blood-meal, mated; $D1^O$: 1day post-oviposition)

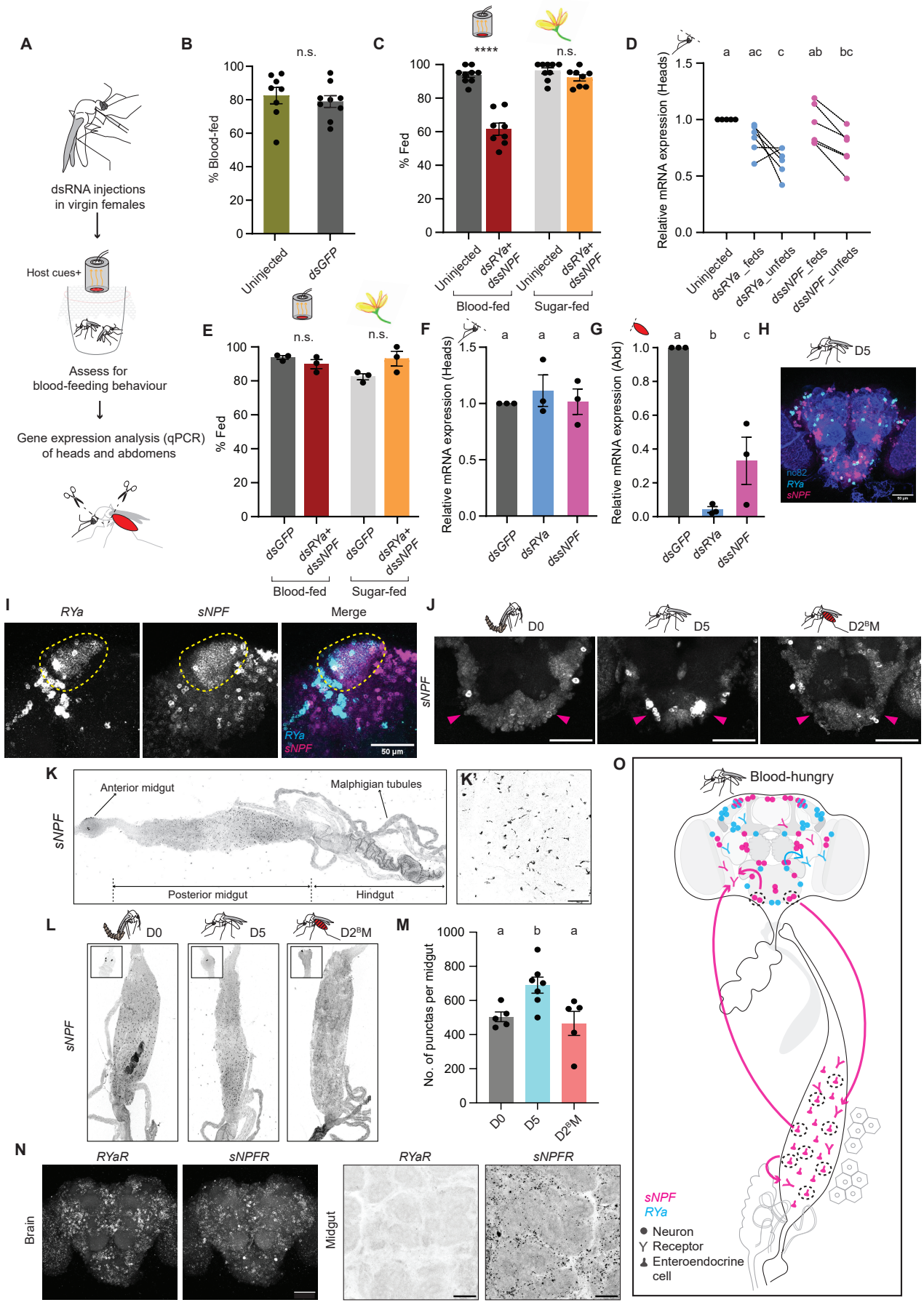
A
dsRNA injections
in virgin females
Host cues+
Assess for
blood-feeding behaviour
Gene expression analysis (qPCR)
of heads and abdomens
B
n.s.
% Blood-fed
Uninjected
dsGFP
C
****
n.s.
% Fed
Uninjected
dsRYa+
dssNPF
Blood-fed
Sugar-fed
D
a
ac
c
ab
bc
Relative mRNA expression (Heads)
Uninjected
dsRYa_feds
dsRYa_unfeds
dssNPF_feds
dssNPF_unfeds
E
n.s.
n.s.
% Fed
dsGFP
dsRYa+
dssNPF
Blood-fed
Sugar-fed
F
a
a
a
Relative mRNA expression (Heads)
dsGFP
dsRYa
dssNPF
G
a
b
c
Relative mRNA expression (Abd)
dsGFP
dsRYa
dssNPF
H
D5
nc82
RYa
sNPF
50 µm
I
RYa
sNPF
Merge
RYa
sNPF
50 µm
J
D0
D5
D2BM
sNPF
K
Anterior midgut
Malphigian tubules
sNPF
Posterior midgut
Hindgut
K'
L
D0
D5
D2BM
sNPF
M
a
b
a
No. of punctas per midgut
D0
D5
D2BM
N
RYaR
sNPFR
Brain
RYaR
sNPFR
Midgut
O
Blood-hungry
sNPF
RYa
Neuron
Receptor
Enteroendocrine
cell

Fig. 4. ***short Neuropeptide F (sNPF)* and *RYamide (RYa)* together promote blood-feeding in *An. stephensi*.** A. Schematic representation of the pipeline used for functional validation. dsRNAs against candidate genes were injected in adult virgins for targeted RNAi. After recovery these were tested for blood-feeding. Knockdown efficiency was determined via qPCR in whole heads and abdomens of fed and/or unfed females after the behaviour. B. Comparison of uninjected females and *dsGFP*-injected females for blood-feeding behaviour revealed no difference between the two, indicating that the animals recovered from the injection procedure. 19-25 females/ replicate, n= 8-9 replicates/group. Unpaired t-test; n.s.: not significant, p>0.05. C,D. Blood-feeding and sugar-feeding behaviour of females when both *RYa* and *sNPF* are knocked down in the head (C). Relative mRNA expressions of *RYa* and *sNPF* in the heads of *dsRYa+dssNPF*- injected blood-fed and unfed females, as compared to that in uninjected females, analysed via qPCR (D). Simultaneous knockdown of both neuropeptides suppressed the blood-feeding behaviour of females, without affecting the sugar-feeding behaviour. Note the more efficient knockdown in the heads of unfed females, as compared to blood-fed females. 14-25 females/replicate, n=8-9 replicates/group. Unpaired t-test; n.s.: not significant, p>0.05; ****p<0.0001 (C). One-way ANOVA with Holm-Šídák multiple comparisons test, p<0.05 (D). Data labelled with different letters are significantly different. E,F,G: Blood-feeding and sugar-feeding behaviour of females when both *RYa* and *sNPF* are knocked down only in the abdomen (E). Relative mRNA expressions of *RYa* and *sNPF* in the heads (F) and abdomens (G) of *dsRYa+dssNPF*- injected fed females, as compared to that in dsGFP-injected females, analysed via qPCR. No change in the blood-feeding or sugar-feeding behaviour was observed, despite the strong knockdown of both the genes in the abdomens, while the levels in the heads remain unaltered. 17-21 females/replicate, n=3 replicates/group.Unpaired t-test; n.s.: not significant, p>0.05. (E). One-way ANOVA with Holm-Šídák multiple comparisons test, p<0.05 (F,G). Data labelled with different letters are significantly different. H, I. Whole mount mRNA *in situ* hybridisation of central brain of 5days old sugar-fed virgin female, with *RYa* and *sNPF* probes (H). Both neuropeptides are expressed in multiple clusters (cyan: *RYa*, magenta: *sNPF*, blue: nc82). Co-expression was observed only in the Kenyon cells of the mushroom body (I). Maximum intensity projections of the confocal stacks are shown. Scale bar, 50µm. J. *sNPF* expression in the SEZ region of the central brains of females uninterested in blood (D0), blood-hungry females (D5) and blood-sated females ($D2^BM$). Note the appearance of a novel cluster only in the blood-hungry (D5) condition. Maximum intensity projections of selected confocal stacks are shown. Scale bar, 50µm. K. Whole mount mRNA *in situ* hybridisation of gut of 5days old sugar-fed virgin female with *sNPF* probes. *sNPF* expression was observed in the enteroendocrine cells (EECs) in the posterior midgut, shown in higher magnification in K'. Scale bar, 50µm. L,M. *sNPF* expression in the posterior midguts of females uninterested in blood (D0), blood-hungry females (D5) and blood-sated females ($D2^BM$). In addition to the EECs in the posterior midgut, two cells in the anterior midgut also show strong *sNPF* expression (L, inset). Higher number of cells express *sNPF* in the blood-hungry condition (M). n=5-7 guts per condition. One-way ANOVA with Holm-Šídák multiple comparisons test, p<0.05. N. Whole mount mRNA *in situ* hybridisation of central brain (left) and midgut (right) of 5days old sugar-fed virgin female, with *sNPF receptor (sNPFR)* and *RYa receptor(RYaR)* probes. While both receptors are expressed in the brain, only *sNPFR* is expressed in the gut. Scale bar, 50µm. O. Summary schematic of *sNPF* and *RYa* function in modulating blood-appetite in *An. stephensi*. *RYa* and *RYaR* (cyan) are expressed only in the brain, while *sNPF*and *sNPFR* (magenta) are expressed both in the brain and the gut. Increased *sNPF* levels in both the tissues (dashed circles) promote a state of blood-hunger, which may drive feeding behaviour either by *sNPF's* action in the two tissues independently or via a communication between them. This happens in the context of *RYa* signalling in the brain. (D0: day of emergence; D5: 5days post-emergence, virgin; $D2^BM$: 2days post-blood-meal, mated)

tides are known to influence behaviour from their action in non-neuronal tissues such as the gut (55, 79, 80), we modified our dsRNA injection protocol to knock *sNPF* and *RYa* down in the abdomen while leaving their levels in the head unaltered (figure 4F, 4G, S9J, S9L) (see methods for details). Abdomen-specific knockdown had no effect on blood-feeding (figure 4E, S9I, S9K), indicating that abdominal *sNPF* and *RYa* alone cannot influence blood-feeding and their action in the brain is necessary. In contrast, since every head knockdown of these neuropeptides also abolished their abdominal expression (figure S9C, S9F), we cannot rule out the possibility that both brain- and gut- derived neuropeptides modulate blood-feeding. Indeed, since such neuropeptide mediated gut-brain control of feeding is commonly seen (79), we propose a model, consistent with our data, where brain-derived *sNPF/RYamide* is essential to trigger blood-feeding, while gut-derived peptide acts as a modulatory feedback signal. This remains to be tested with cell-specific tools that are currently not available for this species.

To determine in which cells *sNPF* and *RYa* act, we used hybridisation chain reaction (HCR) (81, 82) *in situ*. We found that their transcripts are expressed in several non-overlapping neuronal clusters distributed across many regions in the brain (figure 4H, S10A) and only in the Kenyon cells of the mushroom body were they co-expressed (figure 4I, S10B). This is reminiscent of their expression in *Drosophila* and *Bombyx* (83–85). Importantly, we found a cluster of cells in the SEZ that expresses the *sNPF* transcript only in the blood hungry-state (figure 4J, supplementary movie). Its transcripts were either absent or diminished from this cluster in newly-emerged or sated females. Since the RNAi experiments suggested the abdominal involvement of these neuropeptides, we also looked for their expression in the midgut where we could detect only *sNPF's* transcripts, not *RYa's* (figure 4K,K', S11A, S11B, S11E). Interestingly, *sNPF's* transcript expression was also modulated in the midgut enteroendocrine cells (EECs) - hormone-producing cells of the gut. In these cells, *sNPF* transcripts were specifically increased in blood-hungry females when compared to newly-emerged or sated females (figure 4L,M).

Neuropeptides work by acting on their receptors to trigger downstream signalling cascades in the cells that express the receptors. We identified putative receptors for *sNPF* and *RYa* (see methods) and found that both were expressed in several overlapping cell-clusters in the brain (figure 4N, S10C, S10D). In the midgut, however, we could only detect *sNPFR* and not *RYaR* (figure 4N, S11C, S11D, S11F). Since these receptors were identified based on homology, their functional validation will be necessary to implicate them in blood-feeding behaviour. However, neuropeptide receptors are often ligand-promiscuous and confirming their specificity would require pharmacological characterisation beyond the scope of this study.

Together, these data are consistent with a model where increased *sNPF* expression in both the brain and midgut might drive the blood-feeding behaviour, but that this occurs in the context of *RYa's* action in the brain (figure 4O).

Conclusions

Here, we report the blood- and sugar-feeding behaviours of *An. stephensi*. While sharing key features with other mosquito species, *An. stephensi* exhibits distinct regulatory patterns. Unlike *Ae. aegypti*, it does not require mating to initiate blood-feeding, though the presence of males is necessary to sustain the enhanced state. In both species, blood-feeding is suppressed after a blood-meal and until oviposition, but this suppression is mating-dependent in *An. stephensi*. Since unmated *Ae. aegypti* females did not blood-feed in our assays, we cannot comment on whether this suppression also requires mating.

Mosquito feeding behaviour is highly context-dependent, shaped by physiological state, environment, and experimental design (36). Our comparisons between *Ae. aegypti* and *An. stephensi* were conducted under standardised conditions. While direct comparisons across studies must be made cautiously, patterns observed under particular conditions can offer meaningful insights. For example, virgins of other *Anopheles* species readily blood-feed (36, 39–41), suggesting that this is typical of the genus, whereas *Ae. aegypti* virgins do so only under specific conditions and these responses vary among strains (38) (and references within). Whether blood-fed *Aedes* virgins suppress subsequent feeding is likely also context dependent with reports suggesting both that they do (86, 87) and do not (45). Post-blood-meal suppression of host-seeking - well characterised and robust in mated *Aedes* - appears weaker in mated *Anopheles* and can be lifted prior to oviposition (88). Indeed, we too observed a stronger post-blood-meal suppression of feeding in *Ae. aegypti* than in *An. stephensi* (compare Fig. 1C to Fig. 1D). Few studies have investigated this suppression in *Anopheline* virgins, but when they do, they report a weak suppression (89). Taken together, *An. stephensi* displays characteristics of both *Aedes* and *Anopheline* feeding patterns - its virgin feeding patterns align more closely with other Anophelines, yet, like mated *Ae. aegypti*, it still exhibits clear suppression of blood-feeding until oviposition.

What mechanisms might underlie such mating-induced modulation of blood-feeding? In *Drosophila melanogaster*, mating induces a suite of behavioural changes - including increased protein and sugar intake - that are mediated by male-derived sex-peptide that acts on central brain circuits in the female brain (4–7, 90–92). These changes are anticipatory in nature rather than a homeostatic control of feeding behaviour to meet reproductive demands (7, 8). In mosquitoes, the molecular mechanisms driving post-mating behavioural changes are not likely to be the same (93). Instead, mating-derived modulators such as HP1 in *Aedes* (62, 94) and male-deposited steroid hormone (20-hydroxyecdysone) in *Anopheles* (95, 96) have been shown to induce sexual refractoriness. Whether they also shape feeding decisions remains unknown.

We have identified two neuropeptides - *RYa* and *sNPF* - that act synergistically to promote blood-feeding in *An. stephensi*. Our data are consistent with a model where an increase in *sNPF* levels in the brain - likely released by a cluster of neurons in the SEZ - as well as in the midgut, promotes a state of blood-hunger. Whether this reflects gut-to-brain communication or parallel, tissue-local computations to modulate behaviours remains unclear, especially since we detected the transcript of both *sNPF* and its receptor in both tissues. Inter-organ signalling is well established in feeding regulation (79, 97). For example, gut-derived NPF in flies can influence brain circuits to increase protein intake (80). More broadly, feeding behaviour is likely regulated by a distributed network of hormonal and neuronal cues across multiple organs. Signals from the ovary, fatbody, and other nutrient-sensing tissues may also contribute, acting through both central and peripheral circuits. Many of these inputs remain to be identified.

It is possible that *sNPF* promotes blood-feeding independent of *RYa*. However, we were unable to knock *sNPF* down completely to test this. Its synergistic requirement with *RYa*, despite no apparent modulation of *RYa* or its receptor, could result from a general state of hunger that *RYa* promotes, in the context of which, we were able to uncover the blood-feeding role of the diminished *sNPF*. Since many neurons co-express these receptors, it is possible that neurons downstream of *sNPF* or *RYa* expressing neurons integrate the two inputs to modulate behaviour.

We note that our findings differ from those reported in *Aedes*. In *Aedes sNPF* and *RYamide* act as satiety brakes—peptide or receptor activation curbs host-seeking (52, 53, 55) —whereas in *Anopheles*, our knockdowns show that the same neuropeptides act as a hunger signal, stimulating blood-feeding. While these differences are interesting, in the context of their evolutionary histories, and the known differences in how neuropeptides can act differently across species, they are not entirely unexpected. *Aedes* and *Anopheles* lineages diverged ≈150-200 million years ago (98), and the two genera have since diverged in almost every facet of biology—diel activity, host preference, feeding and oviposition sites, egg-desiccation tolerance, genome size, and chemosensory repertoires among other traits. Neuropeptide pathways are known to drive opposing behaviours in different species: *sNPF* is known to be orexic in *D. melanogaster*, *A. mellifera*, *L. decemlineata*, *B. mori*, and *P. americana*, while it has anorexic effects in *S. gregaria* (99). Against this backdrop, it is unsurprising that the same neuropeptide pathway can drive opposite behaviours in *Aedes* and *Anopheles*. These comparative studies underscore a key lesson for vector control strategies in general. A strategy that is effective in one mosquito species may inadvertently boost vectorial capacity in another. Behavioural interventions, chemical modulators, and even gene-drives therefore require rigorous cross-species validation because results from one mosquito species cannot be assumed to hold true for another.

Methods

Mosquito culture. *Anopheles stephensi* (Indian strain; Bangalore strain, TIGS-1) and *Aedes aegypti* (Bangalore collection) were reared in BugDorm cages and maintained at 26-28°C, 70-80% relative humidity with a photo-period of 12h light: 12h dark. All the behavioural assays were performed at these conditions of temperature and humidity. Larvae were fed on a mixture of yeast and dog biscuit (1:3 ratio) and cultured in trays in appropriate densities to prevent overcrowding. Adult mosquitoes were provided *ad libitum* access to 10% sugar solution (8% sucrose +2% glucose +0.05% methyl paraben +5% vitamin syrup (Polybion SF Complete, Merck Ltd.)) at all times, unless specified otherwise. Females were provided O+ve human blood, secured periodically from blood banks, both for general maintenance and feeding assays.

Regulatory permissions. This project was approved by the Institutional Human Ethics Committee (Ref: inStem/IEC-22/02) and the Institutional Biosafety Committee (TIGS/M-7/8/2020-1).

Blood-feeding assay: first blood-meal, 0-120h post-emergence (D0-D5). All *An. stephensi* feeding experiments were performed from ZT12-ZT15 and *Ae. aegypti* experiments from ZT10-ZT12 (Zeitgeber time 0 is defined as the time of lights ON).

0-7h old mosquitoes were cold anaesthetised and collected during the day in 1100ml paper cups covered with a net and secured with a rubber band. These were maintained at the conditions of temperature and humidity described above. To collect virgins, males were separated at the time of collection. 20 females with males (1:1 ratio) or without males were collected for each trial and were aged appropriately with *ad libitum* access to 10% sugar solution (supplementary figure S1A).

Females were assayed for their blood-feeding behaviour from the day of emergence (D0, 0-7h old) to 5 days of age (D5), every 24h. They were provided constant access to sugar, except when they were offered blood. A detailed schematic has been shown in figure S1A. On the day of the test, appropriately aged females were offered a blood-meal via a Hemotek feeder (Hemotek Ltd), which was maintained at 37°C and perfumed with human skin odours. It has been reported that female mosquitoes need to be 'activated' to take blood-meals. This involves the presentation of CO_2 (and other host-cues) to the females, likely because CO_2 has been shown to gate temperature and visual cues (100, 101). So, prior to offering the blood-meal, the mosquitoes were 'activated' by the presentation of a hand placed above the cup for 5 minutes, followed by 3 exhalations from the experimenter. Additionally, blood was supplemented with 1mM ATP (Sigma A6419) before loading the feeder (figure 1B). Females were allowed to feed for 60-90mins in dark and abdomens were scored visually for presence of blood, irrespective of the amount ingested (inset, figure S1A). For D0, no sugar was provided at the time of collection. Females were allowed to acclimatise for at least 30 mins in the cups and then offered a choice of both sugar and blood simultaneously, to assess their preference for blood soon after emergence (figure S1A).

In the case of co-housed females, males and females were housed together from the time of collection until the day of the experiment. Males were separated post-hoc and spermathecae of the females were dissected to score for the presence or absence of sperm (figure 1G). All the assays with virgin and co-housed females were performed in parallel by a single experimenter. Mosquitoes from at least 4 independent batches were tested for each day of the experimental timeline.

For *Ae. aegypti*, both virgin and mated females (mating status was confirmed via post-hoc dissection of spermathecae and all co-housed females were found to be mated) were tested for first blood-meal at only 8days post-emergence (D8, figure 1C). Behavioural experiments were performed the same way as described for *An. stephensi*.

Blood-feeding assay: second blood-meal, 24-96h post-first blood-meal ($D1^B$-$D4^B$). To ensure that the second blood-meal is not driven due to an incomplete first blood-meal, we selected only fully-engorged females to test for subsequent feedings (figure S1A). 5days old *An. stephensi* (both virgins and co-hosued with males) and 7-9days old *Ae. aegypti* (mated, as previously determined) were fed on blood and fully fed females were separated and collected in paper cups as described above. As a female takes about 2 days to digest blood, the second blood- meal ingested 24h post-first blood-meal ($D1^B$) was detected by the presence of Rhodamine B (Sigma R6626), which fluoresces in the red range (figure S1A). Spiking the second blood-meal with Rhodamine B (0.04μg/μL of blood) allowed visualisation of the freshly ingested blood, without affecting the feeding efficiency (supplementary figure S1B-D). To assess feeding at subsequent days ($D2^B$-$D4^B$), abdomens were dissected to differentiate between the bright red freshly ingested blood from the dark and clotted first blood-meal. In the case of co-housed *An. stephensi* females, spermathecae were not dissected to confirm mating as females were presumed to be mated when collected for this assay (5days old and co-housed with males for the entire duration from the time of emergence), as previously determined in figure 1G.

To confirm whether mating suppresses subsequent feeding (figure 1H), fully fed virgins were collected and allowed to mate for 3-4 days with aged-match males. No males were introduced in the virgin-only controls. Blood-feeding assay was performed as described above. Spermathecae were dissected post-hoc to determine the mating status and unmated females were not considered for the final analysis. Datasets with less than 80% total mating were discarded.

Blood-feeding assay: post-oviposition, 24-48h post-egg laying ($D1^O$-$D2^O$). For blood-feeding behaviour post-oviposition, fully fed females were separated and a water cup lined with moist paper (ovipositor) was provided for egg laying 2days post-blood-meal (determined in a separate assay described below, supplementary figure S1E; figure S1A). Females were allowed to lay eggs overnight. The following morning, non-gravid females were visually identified by their lean bellies and separated in cups. Blood-feeding assay was performed the same evening for 1day post-oviposition ($D1^O$) and the following day ($D2^O$), as described above. Females were dissected post-hoc and scored for absence of mature ovaries, as a sign of successful oviposition. Gravid females were not considered for the analysis.

Determination of oviposition timing for *An. stephensi*. To determine the day of maximum oviposition, 5-6days old females co-housed with males were fed on blood, as described above. Fully fed females were separated and collected in BugDorm cages in groups of 10 females/cage. An ovipositor was provided the same day and females were allowed to lay eggs overnight. Eggs laid were counted the next day and a fresh ovipositor was provided. This cycle was repeated for the next 5 days. Total eggs laid each day were di-

vided by the number of females present at the time of counting (supplementary figure S1E).

Sugar-feeding assay for *An. stephensi*. As mosquitoes feed on sugar in small bouts, as opposed to a replete blood-meal, we assessed sugar-feeding behaviour of females in windows of 24h for each test day. 0-7h old mosquitoes were collected during the day, separated in cups and aged appropriately. They were given access to 10% sugar solution at all times, as described above, except for the day of emergence (D0) when no sugar was provided at the time of collection (similar to the blood-feeding assay performed at D0).
To assess their sugar-feeding behaviour on the day of emergence (D0), 0-7h old females were acclimatised in the cups for 1h and sugar spiked with Rhodamine B (1mg/25ml of sugar; Sigma R6626) was presented for 2.5-3h. The fluorescence of Rhodamine B in the red channel allowed visualisation of even the tiniest amounts of sugar ingested by a female (figure S1G). For D1 feeding, 3-10h old females (with access to normal sugar so far) were then provided with Rhodamine B-sugar and allowed to feed on it for 24h. For D2 feeding, 27-34h old females (with access to normal sugar so far) were fed on Rhodamine B-sugar for the next 24h and scored as D2. This cycle was repeated until D5. In the case of co-housed females, males were separated post-hoc and spermathecae of the females were dissected to score for the presence or absence of sperm (figure S1H).
To assess sugar-feeding post-blood-meal, females fully fed on blood were collected and separated in cups, as described above. They were allowed to feed on Rhodamine B-sugar for 24h following a blood-meal and scored as $D1^B$. The next set of females (with access to normal sugar so far) were then provided with Rhodamine B-sugar from 24h-48h post-blood-meal and scored as $D2^B$. This cycle was repeated until 4 days post-blood-meal ($D4^B$, figure 1E).
For sugar-feeding post-oviposition, blood-fed females were provided an ovipositor 3 days later and were forced to oviposit during daytime by artificially creating dark conditions. Oviposited females (non-gravid) were then visually identified, separated and presented with Rhodamine B-sugar for the following 24h and scored as $D1^O$. The next set (with access to normal sugar so far) was then provided with Rhodamine B-sugar, 24-48h post-oviposition and scored as $D2^O$(figure 1E). For each day tested, females were dissected post-hoc and scored for absence of mature ovaries, as a sign of successful oviposition. Gravid females were not considered for the analysis.

Dual choice assay of blood and sugar. Mosquitoes were collected as described above. Males and females were co-housed for 4-6days with *ad libitum* access to sugar. To assess the choice between sugar and blood, mosquitoes were either given continuous access to normal sugar (sugar-sated) or starved with water 24h prior to the assay (sugar-starved). At the time of the assay, post-activation (as described above), blood and Rhodamine B-spiked sugar (1mg/25ml of sugar, Sigma R6626) were provided in parallel. After 60mins, abdomens were scored visually for ingestion of blood under white light, and under fluorescent microscope (red channel) for ingestion of sugar (figure S1F). Co-housed males served as an internal control for sugar-feeding. As males are unable to feed on blood, they were only scored for presence or absence of Rhodamine B-spiked sugar. Results are presented in figure 1F.

Statistical analysis of feeding behaviours. Statistical analysis was performed using R (version 4.4.2) and GraphPad Prism (v10.2.0). Blood-feeding and sugar-feeding behaviours of virgins and co-housed females in figure 1D, E, were analysed using Generalized Linear Mixed-effects Models (GLMMs) with a binomial distribution and logit link function. The models were fitted using the "glmer" function from the "lme4" package in R. The probabilities of blood- or sugar-feeding, both pre- and post-blood-meal were modeled as a function of Day (age) and Group (virgin vs. co-housed with males), including their interaction term to test whether group differences varied across days. Replicate identities (BioID) and larval batches (only for first blood-meal in sugar-fed females) were included as random factors to account for replicate-level and batch-level variations respectively. Post-hoc analyses were conducted using estimated marginal means (emmeans package) for age-effects within groups (p-value adjusted for multiple comparisons using Tukey's method) and pairwise comparison between groups at each day (p-value adjusted using Bonferroni method). To test whether mating success altered the sugar- and blood-feeding propensity of females co-housed with males, the probabilities of feeding were modeled only the co-housed data, as a function of Day (age) and Mating (yes or no), including their interaction term and accounting for variation due to replicate IDs (BioID). Statistical significance was set at $\alpha = 0.05$ for all analyses. Complete model outputs are provided in supplementary sheet1. Statistical analyses for data in Figures 1C, 1F, and 1H were performed using GraphPad Prism (v10.2.0) with appropriate tests as indicated in figure legends.

Y-maze olfactory assay for host-seeking behaviour in *An. stephensi*. A 30inch Y-maze was constructed as per the WHO guidelines (`https://www.who.int/publications/i/item/9789241505024`). Each arm of the Y-maze had a trapping port, while the stem was attached to a holding port. Each port had a mesh screen at one end, while the other was fitted with a sliding door to trap or release mosquitoes. The holding port was attached to a fan which pulled air from the two arms, over the mosquitoes, and out at a velocity of 0.3-0.6m/s. Optic flow information (alternate black and white stripes) was placed underneath the Y-maze to act as visual guide for navigation.
Following conditions were tested: D0 virgins, D5 sugar-fed virgins, D5 sugar-fed mated females (D5M), 3days post-blood-meal virgins ($D3^B$), 3days post-blood-meal mated females ($D3^BM$), and 2days post-oviposition females ($D2^O$). For mated conditions, females were co-housed with males for 5days post-emergence and assumed to be mated based on our previous findings (figure 1G). For each condition, mosquitoes were collected and provided *ad libitum* access to sugar as de-

scribed above. Females were transferred to holding ports 7-8h prior to the assay and were starved during this time with only access to water. For each run, mosquitoes were acclimatised for 5 minutes: 2 min with clean air and 3 minutes with an experimenter presenting a hand at one of the arms and simultaneously exhaling at resting rates. To account for disruption in airflow, the hand of a mannequin was placed at the other arm (control). Post-acclimatisation, the door of the holding port was opened to release the mosquitoes and they were allowed to make a choice for 5 minutes in the presence of continued host-cues. The ports were then closed and mosquitoes in each arm counted. All the mosquitoes trapped in a port, sitting on the door or resting on the half-length of the Y-arm were considered as "attracted" to that stimuli (host or control, figure 2A). Participation percentage was also calculated as number of mosquitoes that passed halfway through the Y-stem/total number of mosquitoes released (figure S1I).

Several controls were included to avoid experimental artefacts. Each experimental day started with a blank trial (where no stimuli were presented on either arm of the Y-maze) and the side of the host-cues presentation was alternated with each trial to eliminate side biases. Trials for blood-fed virgins and mated females were done both in parallel and on multiple days to eliminate the influence of external factors which might be present on a given day. When testing blood-fed females, aged-matched sugar-fed females (blood-hungry) were included as positive controls where ever possible, with satisfactory results. Lastly, we made sure that at least 50% of the mosquitoes had made either choice in all runs, except for D0 (newly emerged females) and blank runs. In the rare cases that this was not true, we did not consider those for the analysis. Statistical analysis was performed using GraphPad Prism (v10.2.0). To compare the attraction of females to host-cues and control in figure 2B, each time-point was treated as an independent experiment and p-values were computed for each pair-wise comparison.

Bulk RNA sequencing of central brains and data analysis. For bulk RNA sequencing, central brains were dissected from the following conditions: D0 and D5 sugar-fed males; D0 and D5 sugar-fed virgin females; 1day post-blood-meal virgin ($D1^B$) and mated females ($D1^BM$) and 1day post-oviposition females ($D1^O$). For post-blood-meal conditions, 5days old females were fed on blood and fully fed females were separated for sample preparation. Females co-housed with males for 5 days were assumed to be mated based on our previous findings (figure 1G). Three replicates from independent batches were prepared for each condition.

Mosquitoes were cold anaesthetised before dissections. Central brains (ie. brains without optic lobes) were dissected in RNase-free 1X PBS and immediately placed into ACME solution (13:3:2:2 of water, methanol, acetic acid, and glycerol) on ice, which mildly fixes the tissues (102). For each condition, 100 brains were dissected per replicate. The mildly fixed brains were washed with cold 1X PBS and centrifuged at 13,000 rpm for 5 min at 4°C to remove ACME. Brains were then homogenised in TRIzol (Invitrogen), flash-frozen, and stored at -80°C until RNA extraction. RNA was extracted using Direct-zol RNA MicroPrep (Zymo Research) columns. RNA quantity and quality were assessed using the ThermoFisher Qubit 4 and RNA 6000 Nano kit Bioanalyser, respectively.

The cDNA library was prepared for each sample using the NEBNext Ultra II Directional RNA Library Prep kit (Catalog no-E7765L), with an average fragment size of 250 bp. Some of the library samples were subjected to size and quality checks using a bioanalyser. Paired-end sequencing was carried out on the Illumina Hiseq 2500 sequencing platform, generating 2×100 bp reads. The library preparation and sequencing were performed at the Next Generation Genomics Facility (NGGF) at Bangalore LifeScience Cluster.

Post-sequencing, samples were demultiplexed and delivered as raw Fastq files. After passing the quality check using the FastQC tool, the reads were mapped onto the reference *An. stephensi* transcriptome (NCBI RefSeq assembly GCF013141755.1) (69) using Kallisto v0.43.1 (103) with default parameters. 81-87% of reads aligned to the mosquito transcriptome.

Transcript-level abundance was quantified using Kallisto and gene-level summarisation was performed using the Tximport function in R, with transcript isoform information retained (supplementary sheet 2). Using the EgdeR package, the raw counts were converted to Counts Per Million (CPM) and log2 transformed before data filtering (genes with less than 1 CPM in at least 3 or more samples were filtered out) and normalisation, using the trimmed mean of M-values (TMM) method. Principal component analysis (PCA) was performed using the prcomp function in R and visualised using the ggplot function. Data was checked for batch effects using gPCA (104) in R and the test statistic "delta" was found to be 6.957686e-05, with a p-value of <0.001, indicating no batch-effects.

Differential gene expression analysis

Differential expression analysis was performed using the Limma package in R. A design matrix was created to specify the sample conditions, and the Voom function was used to estimate the mean-variance relationship across genes. A linear model was then fitted to the data using the mean-variance relationship. Bayesian statistics were calculated for the data by creating a contrast matrix to extract the linear model fit. To identify differentially expressed genes (DEGs), the “decideTests” function was used by applying a fold change threshold of 2 (log2fc=1) and a FDR (false discovery rate) adjusted p-value cut-off of 0.05. The number of genes identified to be upregulated and downregulated in each pair-wise comparison are shown in figure S2A and the details are provided in supplementary sheet 3.

To plot Transcripts Per Million (TPM) values as heatmaps, the Fastq files for the central brain samples were mapped onto the reference *An. stephensi* transcriptome and abundance was quantified using Kallisto the same way as described above. Gene-level summarisation was done using Tximport as described above, except all the isoforms of a given transcript were mapped to a single gene (supplementary sheet

2). The resultant TPM values were first filtered for TPM>0.1 in each sample. Next, the genes that were not quantified in at least 2 of the 3 replicates of each sample group were removed. The filtered dataset were then log10 transformed and heatmaps were plotted using the pheatmap function in R. To construct heatmaps of genes involved in carbohydrate and lipid metabolism (supplementary figure S3, S4), key enzymes of the respective canonical pathways were manually curated based on the existing literature. Gene annotations for neuropeptides and neuropeptide receptors were curated from published sources (105–107). Fat body, midgut and ovary data (SRX620223, SRX618937 and SRX620224; bioproject SRP043489) for 2-4days old, sugar-fed *An. stephensi* females were taken from Prasad et al.(108). Fastq files were downloaded from the SRA explorer and processed similarly (supplementary sheet 2).

Gene set enrichment analysis

eggNOG mapper tool 2.1.9 (109) (parameters: 70% identity and minimum 70% query coverage) was used to find functionally annotated orthologs of the identified DEGs in *An. gambiae*. *An. gambiae*-specific orthologs were fed into the GOSt tool from gProfiler2 (110) toolset for functional enrichment analysis. A custom gene list comprising of all the genes identified in our RNA-seq experiment was provided as a background instead of the whole genome. Using this background, an ordered query was run with an FDR adjusted p-value threshold of 0.05 to determine which GO terms (Biological Processes) were over-represented. This was done for all sets of DEGs and the top 10 terms of each set were plotted using ggplot function in R (figure S2B).

dsRNA-mediated gene knockdown. To synthesise the double-stranded RNA (dsRNA), coding regions (500-600bp) of candidate genes were amplified from cDNA of *An. stephensi* females using forward and reverse primers with T7 promoter sequence at 5' ends. The gel purified fragments were then used as a template for *in vitro* transcription reactions using Megascript RNAi kit (Invitrogen) or High-Yield T7 RNAi Kit (Jena Biosciences) as per the manufacturers' protocols. The dsRNAs were purified using the phenol-chloroform method and pellet dissolved in nuclease-free water. The dsRNA for green fluorescent protein (GFP) was synthesised from plasmid pAC5.1B-EGFP (Addgene 21181) to be used as a negative control for off-target effects of dsRNA-mediated knockdown.

Virgins were collected for injections as described above. The day and concentration of dsRNA to be injected was determined for each target gene individually. *dsEiger*: injected in 24h old virgins at 5-6μg/μL; *dsHairy*: injected in 24h old virgins at 4-5μg/μL; *dsJHEH*: injected in 72h old virgins at 5μg/μL; *dsPrp*: injected in 72h old virgins at 3-5μg/μL; *dsOrk*: injected in 96h old virgins at 5μg/μL; *ds118504077* and *ds118504075*: 96h old virgins at 6μg/μL. To achieve tissue-specific knockdown of *sNPF* and *RYa*, injections were performed at different days: in 0-10h old virgins (4μg/μL or 7μg/μL) to achieve abdomen-specific knockdowns and in 96h old virgin females (7μg/μL) to achieve knockdowns in both head and abdominal tissues. For dual injections, purified dsRNAs were mixed in equal amounts to a total of 7μg/μL and injected on the day previously determined.

Injections were performed on a FemtoJet 4i system (Eppendorf) and each female was injected with 0.5-0.6μL of the respective concentrated dsRNAs in the upper thorax. In some cases, dsRNAs were also injected in the abdomens to ensure that the site of injection does not alter the behaviour. Uninjected or *dsGFP*-injected (same concentration as that of the candidate gene) females were included as a control each time. 15-25 females were injected for each trial and at least 2 trials from independent batches of mosquitoes were performed for each target gene.

Injected females were allowed to recover in paper cups with *ad libitum* access to sugar and assessed for blood-feeding behaviour at 3 days post-injection, as previously described. Fed and unfed females were visually scored and whole heads and abdomens were collected separately for analysis. Total RNA was extracted using Direct-zol RNA MicroPrep (Zymo Research) columns, as per the manufacturer's protocol. RNA was quantified using Qubit Broad range assay (Invitrogen). For each sample, 200-220ng RNA was used to prepare cDNA using the SuperScript™ IV Reverse Transcriptase (Invitrogen 18090050) or Maxima H Minus Reverse transcriptase (Thermo Scientific EP0752). Quantitative PCR (qPCR) was performed using PowerUp™ SYBR™ Green Master Mix (Applied Biosystems) to determine the knockdown efficiency. Relative mRNA abundances were calculated using the delta-delta Ct method (111), normalised to RpS7. Statistical analyses were performed using GraphPad Prism (v10.2.0). All the primers used in the study are listed in supplementary table 1.

To assess the sugar-feeding behaviour of dsRNA injected females (*dssNPF*, *dsRYa* and *dsRYa+sNPF*), injections were performed as described above. Injected females were allowed to recover with *ad libitum* access to normal sugar for the first two days, which was then replaced with Rhodamine B-spiked sugar (1mg/25ml of sugar, Sigma R6626) for the next 24h. Control females were treated similarly. The blood-feeding assay was performed 3 days post- injection as described above, with both blood and sugar provided in parallel. Abdomens were scored visually for ingestion of blood under white light, and under fluorescent microscope (red channel) for ingestion of sugar (figure S9A).

Hybridisation chain reaction (HCR). Previously published protocol (112) was modified to perform HCR *in situ* hybridisations on brain and gut tissues collected from females under three different conditions: D0 (not interested in blood), D5 virgins (blood-hungry) and $D2^B$, mated (blood-sated). All the reagents, including the probes, amplifiers and buffers were procured from Molecular Instruments Inc. A set of custom-designed 20 probe pairs were used per target gene: *sNPF (118511038)*, *RYa (118513478)* and their receptors. To find *sNPF* and *RYa* receptors in *An. stephensi*, putative protein sequences were blasted against orthologs in *Drosophila melanogaster*, *Ae. aegypti* and *An. gambiae*. Based on high identities, consistent in all three organisms, *118517381* and

118507167 (NPY receptor type 2) were identified as *RYa receptor (RYaR)* and *sNPF receptor (sNPFR)* respectively.

5-7 brains for each target, per condition were dissected in ice cold PBS and fixed with 4% PFA (in PBS) for 20 minutes at room temperature. Post-washing steps with PTw (0.1% Tween-20 in PBS), 4 times 10 mins each, tissues were permeabilised using detergent solution (50mM Tris-Cl, pH7.5, 150mM NaCl, 1mM EDTA, pH8, 0.5% Tween-20, 1% SDS) for 30 mins at room temperature. Brains were then pre-hybridised in probe hybridisation buffer for 30 mins at 37°C, followed by incubation with neuropeptide probes (0.8pmol each) at 37°C for 48h in a humid chamber. Samples were washed with pre-warmed wash buffer, 4 times 15 mins each, at 37°C to remove unbound probes. This was followed by 2 washes, 5 mins each with 5X SSCT (0.1% Tween-20 in 5X SSC) at room temperature. Samples were then pre-amplified using amplification buffer for 30 mins at room temperature. For each sample, 2μL (3μM stock) of each hairpins were mixed and prepared by heating at 95°C for 90secs for annealing. They were allowed to cool to room temperature for 30mins in a dark chamber and 100μL amplification buffer was added to the mix. Brains were incubated in the hairpin solution for 12-16h at 37°C in dark and excess hairpins were washed off with multiple 5X SSCT washes: twice for 5 mins, twice for 30mins, once for 5 mins. For nc82 staining, all the steps were performed in 0.5% PTw buffer (0.5% Tween-20 in PBS). Brains were equilibrated in 0.5% PTw and incubated with anti-mouse nc82 (1:200; DSHB, RRID: AB2314866) for three overnights at 4°C. Post-washing, goat anti-mouse Alexa-fluor405 (1:400, Invitrogen) was added and samples were left in dark, for one-two overnights at 4°C. After washing off the unbound secondary antibody, samples were equilibrated for at least an hour in Vectashield and mounted in Vectashield for imaging. *In situs* against neuropeptide receptors were performed the same way, except the washing steps to remove the unbound probes were performed at 42°C and samples were incubated with the hairpin solution at room temperature (to minimise background). As controls for background signal, samples were prepared the same way, except no probes were added in the hybridisation buffer (figure S10E). Samples for all three conditions were dissected, processed and imaged the same day.

Protocol for gut *in situs* was adapted from Slaidina et al. (113). 5-8 gut samples for each target, per condition were dissected in ice-cold PBS and fixed with 4% PFA (in PTw) for 20 minutes at room temperature. Post-washing steps with PTw, tissues were dehydrated on ice with graded methanol washes: 25% methanol (in PTw), 50% methanol (in PTw), 75% methanol (in PTw) and 100% methanol for 10 mins each. Samples were stored in 100% methanol for up to a month, at -20°C. For rehydration, samples were washed sequentially with graded methanol: 75% methanol (in PTw), 50% methanol (in PTw), 25% methanol (in PTw), followed by a wash in permeabilisation solution (1% Triton-X in PBS). Samples were permeabilised for 2h at room temperature and fixed again with 4% PFA (in PTw) for 20 minutes. Following 10mins washes: washed once with PTw, twice with 50% PTw/50% 5X SSCT and twice with 5X SSCT, samples were pre-hybridised in probe hybridisation buffer for 30 mins at 37°C, followed by incubation with probes (0.8pmol each) at 37°C for 48h in a humid chamber. Post washing steps with wash buffer at 37°C and 5X SSCT at room temperature, samples were pre-amplified in amplification buffer for 30 mins at room temperature. Hairpins were prepared the same way as described above and samples were incubated in hairpin solutions for 12-16h in dark at room temperature. Following 2 washes 5 mins each with 5X SSCT, samples were incubated with DAPI (1:1000 in 5X SSCT) for one hour at room temperature. Samples were washed with 5X SSCT, 4 times, 10mins each and equilibrated in Vectashield for at least an hour before mounting. As controls for background signal, samples were prepared the same way, except no probes were added in the hybridisation buffer (figure S11G, H). Samples for different conditions, prepared on different days and stored at -20°C, were processed and imaged together. Statistical analyses were performed using GraphPad Prism (v10.2.0).
All samples were imaged at Central Imaging and Flow Cytometry Facility (CIFF) at NCBS. Images were acquired with a 40X/1.3NA or 10X/0.4NA (whole guts) objectives at a resolution of 512X512 pixels, on an Olympus FV3000 system. Images were viewed and manually analysed using ImageJ/FIJI (114). Maximum intensity projections of all z-stacks in an image are shown, except in figures 4H, 4I, S10B, S10D, where stacks were manually selected to capture relevant brain structures.

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Author contributions

PB conceptualised the project, performed all experiments except RNA-seq and host-seeking assay, performed all analysis, including RNA-seq, wrote and reviewed the manuscript. RP performed the RNA-seq experiments and analysis, standardised HCR, and reviewed the manuscript. PDB performed the host-seeking assay, provided technical assistance during HCR standardisations and determination of mating status, and reviewed the manuscript. SQS conceptualised and supervised the project and wrote and reviewed the manuscript.

Declaration of interests

None of the authors have any competing interests to declare.

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Supplementary data

Supplementary files (as attachments)

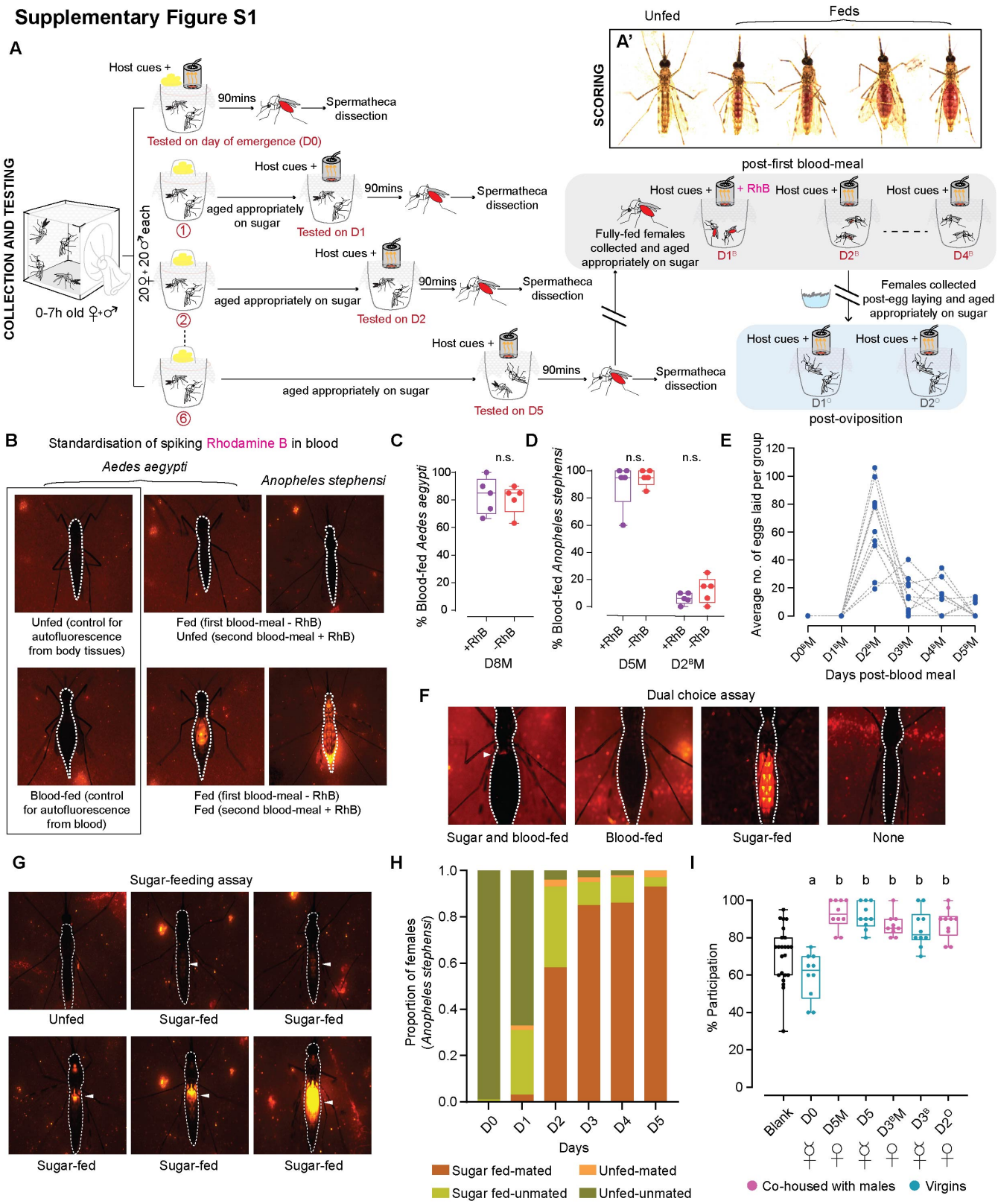
Supplementary sheet 1: Summary of statistical analysis of feeding behaviours using Generalised Linear Mixed Models (GLMM).

Supplementary sheet 2: Table of transcript abundance (in TPM values) for genes identified in the central brain RNAseq data across the gonotrophic cycle.

Supplementary sheet 3: Table of differentially expressed genes (DEGs) identified across states of blood-hunger and -satiety.

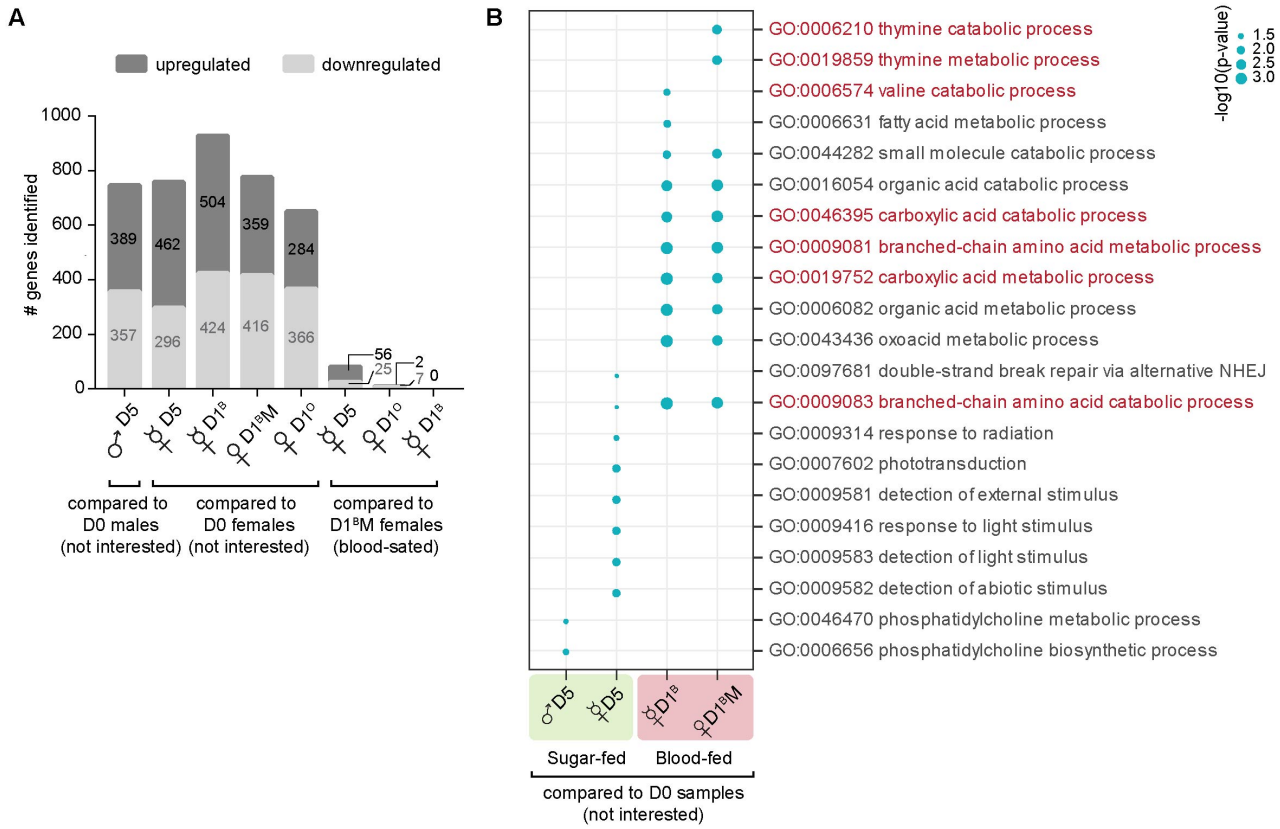
Supplementary movie: *sNPF* expression in the central brain of females uninterested in blood (D0), blood-hungry females (D5) and blood-sated females ($D2^{B}M$). Confocal stacks of whole brains probed for *sNPF* mRNA expression via HCR *in situ* hybridization, at three different conditions of blood-hunger and -satiety, are shown as movies. Note the appearance of novel clusters expressing *sNPF* transcripts (indicated by magenta arrows) in the subesophageal ganglionic region of the brain, only in the blood-hungry (D5) condition.

Supplementary Figure S1



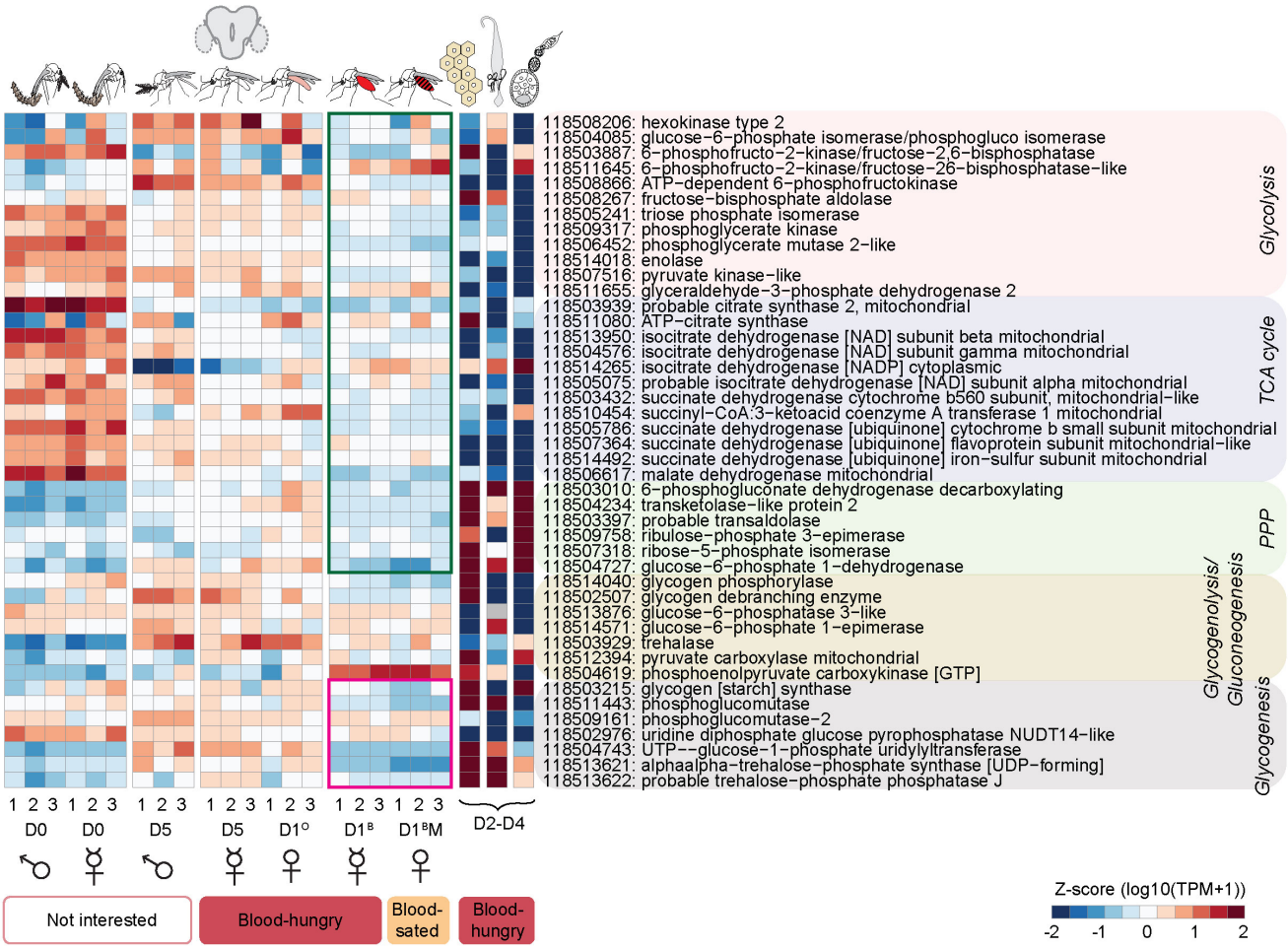
Supplementary Figure 1. Standardization of various behavioural assays to study feeding behaviours in *An. stephensi* A. Detailed schematic of the assay designed to assess the blood-feeding behaviour of virgins and co-housed females across the reproductive cycle. 0-7h old mosquitoes were collected and about 20 females (with or without 20 males) were separated into cups. Each cup was aged appropriately and tested on the relevant day (see methods for more details). Number of blood-fed mosquitoes were visually scored, irrespective of the quantity fed. Representative images of differently fed females are shown in A'. Females were tested for first blood-meal from the day of emergence (D0) to five days of age, every 24h. Spermathecae of co-housed females were dissected post-hoc to determine the mating status. To assay for the second blood-meal, fully fed females were collected and aged appropriately in groups of 20. To assess feeding 1day post-first blood-meal ($D1^B$), Rhodamine B was spiked in the second blood-meal offered to the females. The subsequent days ($D2^B$-$D4^B$) were tested without any such spiking (grey box; see methods for more details). To assay for blood-feeding behaviour in females after oviposition ($D1^O$-$D2^O$), freshly oviposited females were collected and aged in groups of 20 for the appropriate time in cups (blue box; see methods for more details). In all cases, females were presented with host-cues prior to the assay and testing was performed as shown in figure 1B (see methods for more details). B. Standardisation of spiking Rhodamine B dye in blood to assess feeding 24h post-first blood-meal ($D1^B$). To assay for the second blood-meal, blood-fed females were collected, aged for 24h and offered Rhodamine B-spiked blood in a Hemotek. All other assay parameters remained the same as for the first blood-meal (see methods for more details). Widefield fluorescent images of *Ae. aegypti* and *An. stephensi* fed on second blood-meal spiked with Rhodamine B are shown. Images on the left (black box) show mosquitoes not fed and fed on un-spiked blood (without Rhodamine B), as controls for autofluorescence from body tissues and blood itself, respectively. Fluorescence from Rhodamine B added to the second blood-meal enabled identification of fresh blood intake in previously fed mosquitoes reliably. C,D. Comparison of feeding behaviours of *Ae. aegypti* females (C) and *An. stephensi* females (D) fed on blood with and without Rhodamine B at the indicated age. Females fed similarly on both indicating that Rhodamine B itself does not affect feeding behaviour. Unpaired t-test; n.s.: not significant, $p>0.05$. 16-20 females/trial. n= 5 trials/group. D5M: 5days post-emergence, mated; $D2^BM$: 2days post-blood-meal, mated; D8M: 8days post-emergence, mated). E. Determining optimal oviposition timing in *An. stephensi*. Females were tested in groups of 10 and average number of eggs laid per group are shown at the indicated day after the first blood-meal. Dashed lines connect the eggs laid by each group on the indicated day. Majority of the eggs were laid 2days after blood-meal. 10 females/group, n=10 replicates/group. $D0^B$-$D5^BM$: 0-5days post-blood-meal, mated. F. Standardisation of the dual choice assay, to determine the choice between blood and sugar in sugar-sated and sugar-starved *An. stephensi* females, co-housed with males. Widefield fluorescent images of females fed on either blood, sugar or both are shown. White arrowhead marks the fluorescence from sugar (spiked with Rhodamine B) in the abdomens of females engorged on blood, indicating that the female has fed on both. This is not observed in the females fed on blood alone. Females fed on sugar alone can be visualised by the fluorescent lean bellies. Unfed females exhibit no fluorescence or engorgement. G. Widefield fluorescent images of female *An. stephensi* fed on sugar spiked with Rhodamine B. This setup was used to study the sugar-feeding behaviour of virgins and co-housed females across the reproductive cycle. Addition of Rhodamine B to standard sucrose solution enabled visualisation of even the smallest amounts ingested, as indicated by the white arrowheads, under the fluorescent microscope. H. Mating status of sugar-fed and sugar-unfed co-housed females tested in figure 1E, 24-120h post-emergence (D0-D5). n=175-179 females per time-point. I. Percent participation of virgin and co-housed females tested for host-seeking behaviour in the Y-maze olfactometer (figure 2B), at different stages of the reproductive cycle. All females tested participated well. 17-21 females/trial, n=10 trials/group. Kruskal-Wallis, with Dunn's multiple correction test, $p<0.05$. Data labelled with different letters are significantly different.D0: day of emergence ; D5M: 5days post-emergence, mated; D5: 5days post-emergence, virgin; $D3^BM$: 3days post-blood-meal, mated; $D3^B$: 3days post-blood-meal, virgin; $D2^O$: 2days post-oviposition.

Supplementary Figure S2



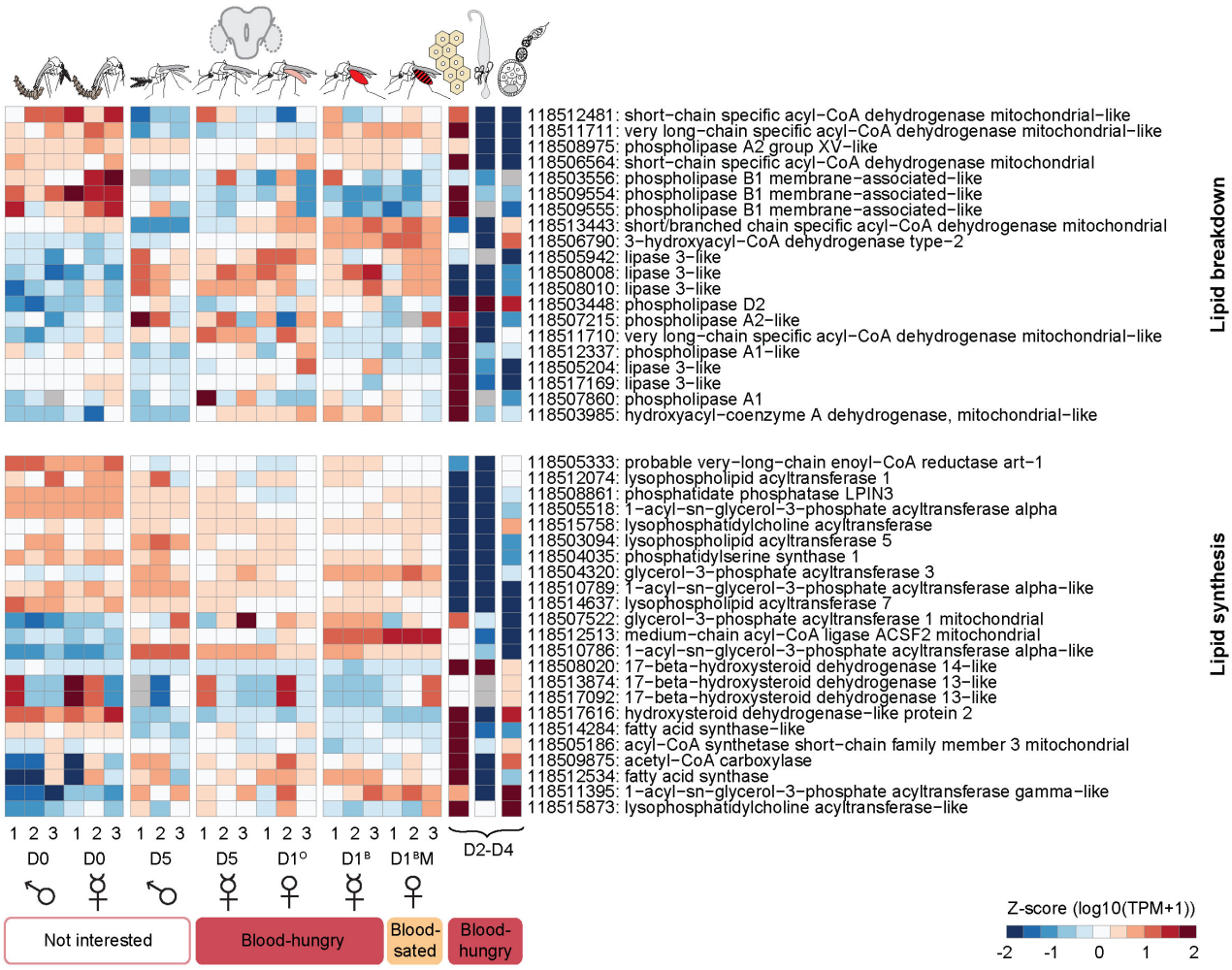
Supplementary Figure 2. Differential expression of genes across various states of blood-hunger and -satiety.. A. Number of genes identified to be differentially up- and down-regulated in the central brain of females across the reproductive cycle (indicated at the bottom), compared to either newly emerged males or females (D0), or blood-fed sated females ($D2^{B}M$). Note that no genes were found to be differentially expressed between mated and virgin blood-fed females, despite their contrasting blood-appetites. B. Top 10 Gene Ontology terms for Biological Processes (GOBP), overrepresented in the samples indicated at the bottom, compared to newly emerged males or females (D0). Size of the dot indicates the p-value significance. Processes involved in protein metabolism (highlighted in red) were found to be highly enriched in blood-fed females. (D0: day of emergence; D5: 5days post-emergence, virgin; $D1^{B}$: 1day post-blood-meal, virgin; $D1^{B}M$: 1day post-blood-meal, mated; $D1^{O}$: 1day post-oviposition)

Supplementary Figure S3



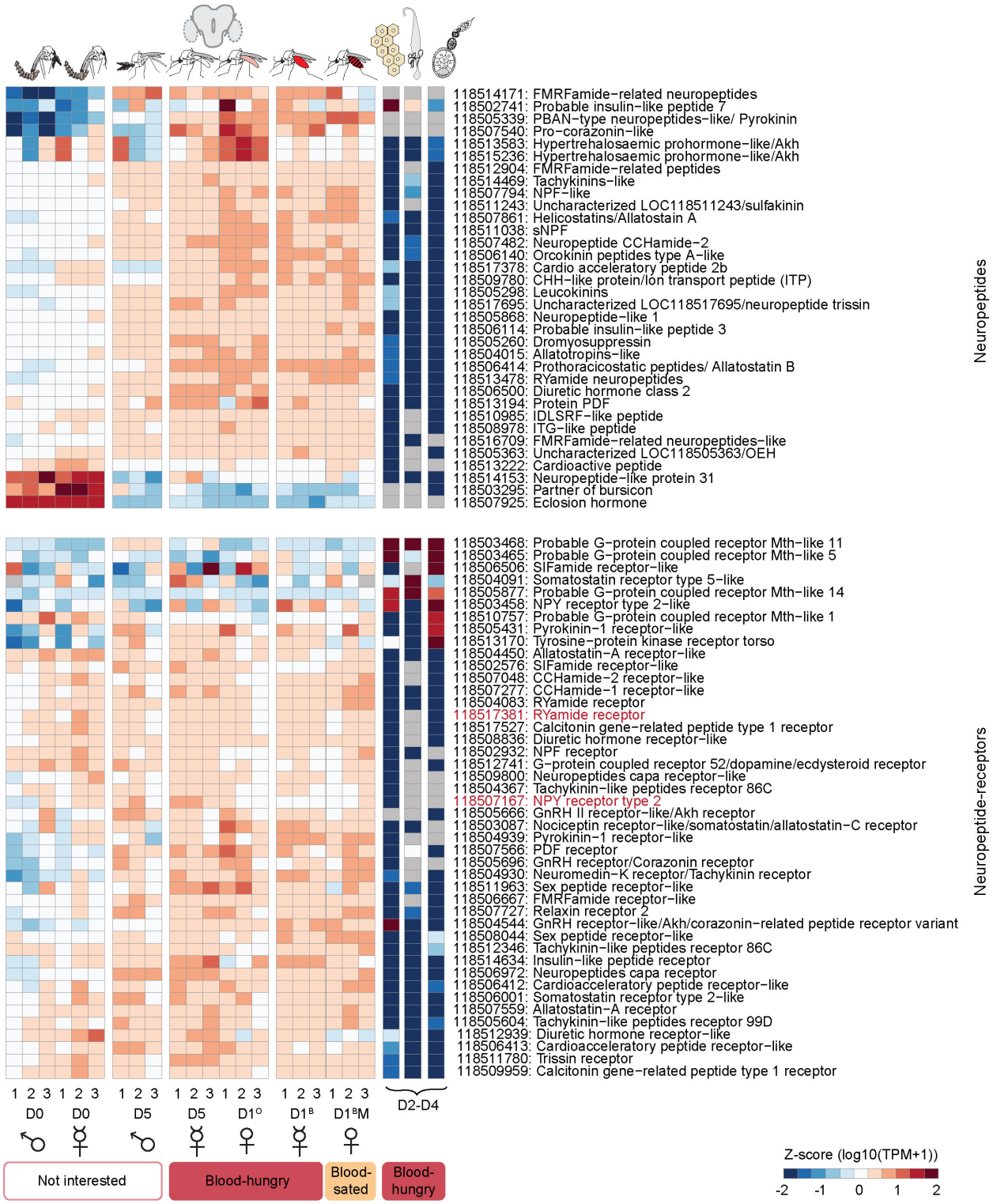
Supplementary Figure 3. Differential expression of genes involved in carbohydrate metabolism across different tissues and states of blood-hunger and -satiety in *An. stephensi* Expression patterns of genes (listed on the right) involved in sugar metabolism via different pathways, across the reproductive cycle in the central brain and in the fat body, midgut and ovary of blood-hungry females, are represented as heatmaps. Green box shows downregulation of genes involved in glucose utilisation and the magenta box represents genes involved in glucose storage. Note that both are low in the brains of blood-fed females indicating that the brain goes into a state of 'sugar rest' after a blood-meal. Analysing publicly available data from the midgut, ovary, and fat body of sugar-fed females suggest that energy expenditure via the non-oxidative branch of the pentose phosphate pathway of glucose metabolism is high in these tissues, likely to prime them for the blood-meal. RNA-seq data from all central brain samples in triplicates and single sample data for fat body, midgut and ovary (taken from Prasad et al., 2017) are represented as z-score normalised log10 (TPM+1) values. TPM values of zero were excluded from z-score transformation and are displayed as grey cells. (D0: day of emergence; D2-D4: 2-4days post emergence; D5: 5days post-emergence, virgin; $D1^{B}$: 1day post-blood-meal, virgin; $D1^{B}M$: 1day post-blood-meal, mated; $D1^{O}$: 1day post-oviposition)

Supplementary Figure S4



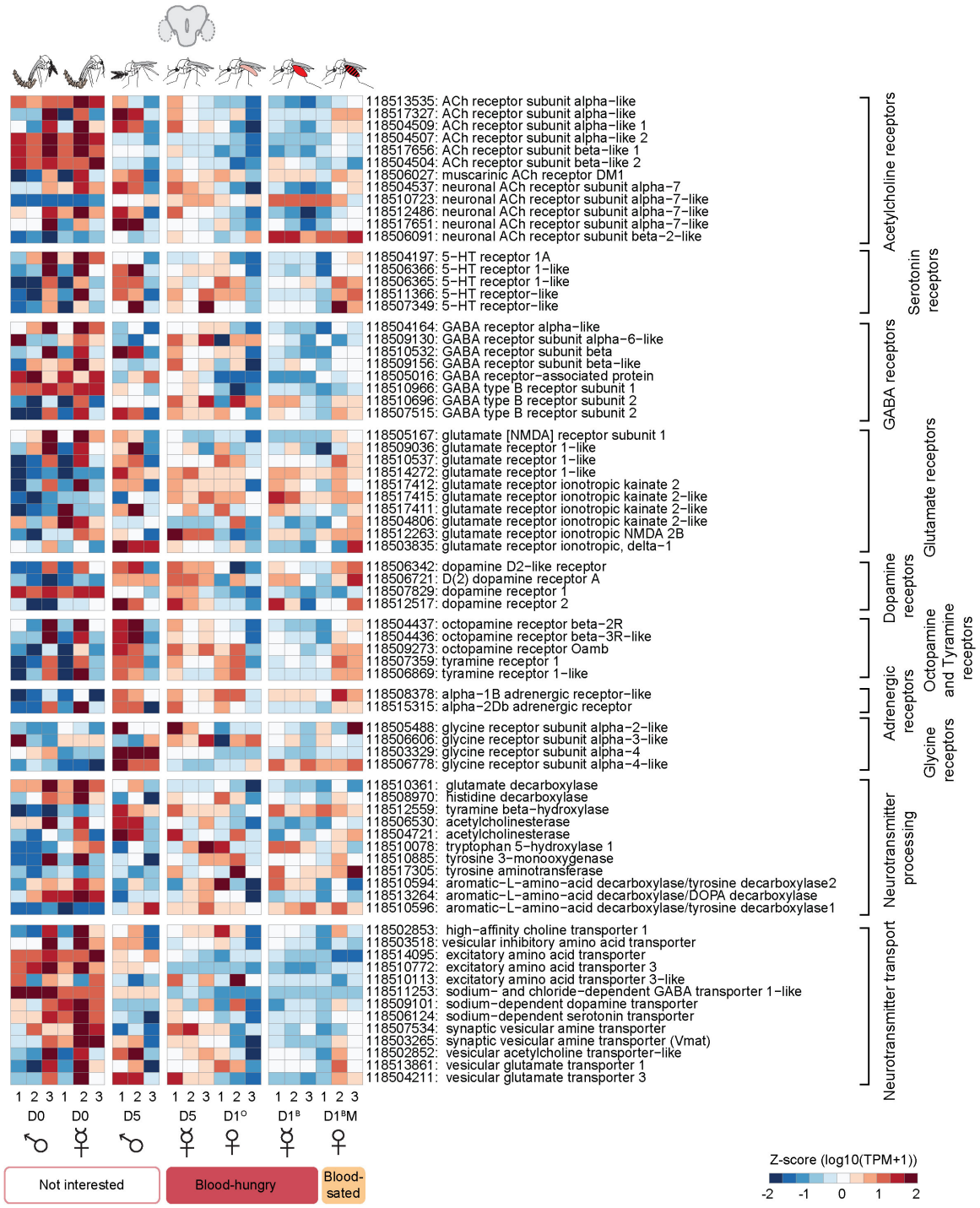
Supplementary Figure 4. Differential expression of genes involved in lipid metabolism across different tissues and states of blood-hunger and -satiety in *An. stephensi*. Expression patterns of genes (listed on the right) involved in lipid metabolism: lipid breakdown (top) and lipid synthesis (bottom), across the reproductive cycle in the central brain and in the fat body, midgut and ovary of blood-hungry females, are represented as heatmaps. No specific patterns were observed in the central brain (pre- and post-blood-meal), midgut or ovaries. As expected, fat body showed high expression of genes involved in both the catabolic and anabolic branches of fatty-acid metabolism. RNA-seq data from all central brain samples in triplicates and single sample data for fat body, midgut and ovary (taken from Prasad et al., 2017) are represented as z-score normalised log10 (TPM+1) values. TPM values of zero were excluded from z-score transformation and are displayed as grey cells. (D0: day of emergence; D2-D4: 2-4days post emergence; D5: 5days post-emergence, virgin; $D1^{B}$: 1day post-blood-meal, virgin; $D1^{B}M$: 1day post-blood-meal, mated; $D1^{O}$: 1day post-oviposition)

Supplementary Figure S5



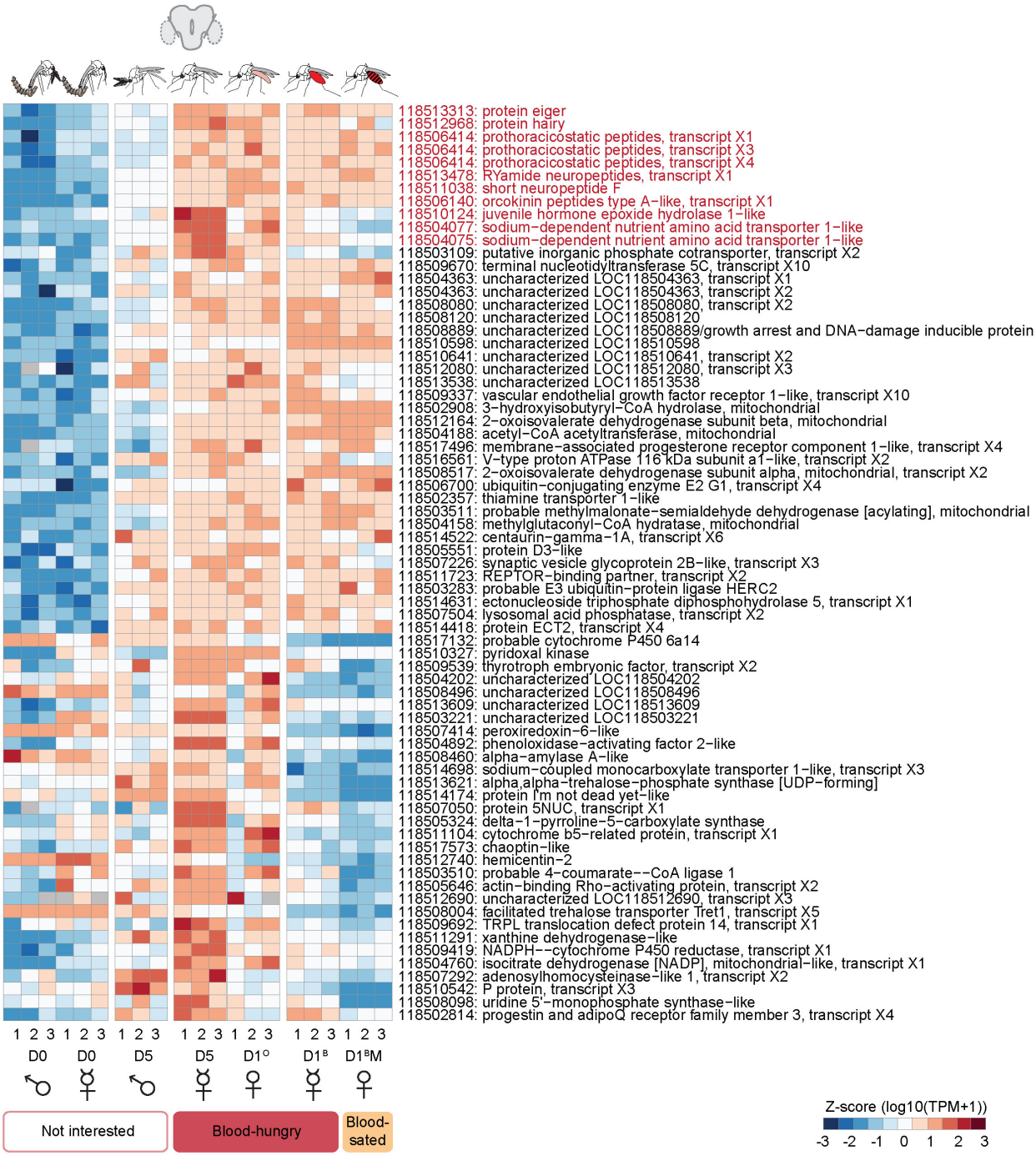
Supplementary Figure 5. Expression patterns of neuropeptides and neuropeptide-receptor across different tissues and states of blood-hunger and -satiety in *An. stephensi*. Expression patterns of neuropeptides (top) and neuropeptide-receptors (bottom), across the reproductive cycle in the central brain and in the fat body, midgut and ovary of blood-hungry females, are represented as heatmaps. While neuropeptides were observed to be downregulated in the central brain of newly emerged males and females (D0), neuropeptide-receptors showed no such patterns. Additionally, both neuropeptides and their receptors were much more abundant in the brain, as compared to fat body, midgut or ovaries. Transcripts for the receptors highlighted in red were visualised *in situ* both in the midgut and brain samples via HCR (figure S10, S11). RNA-seq data from all central brain samples in triplicates and single sample data for fat body, midgut and ovary (taken from Prasad et al., 2017) are represented as z-score normalised log10 (TPM+1) values. TPM values of zero were excluded from z-score transformation and are displayed as grey cells. (D0: day of emergence; D2-D4: 2-4days post emergence; D5: 5days post-emergence, virgin; $D1^B$: 1day post-blood-meal, virgin; $D1^B$M: 1day post-blood-meal, mated; $D1^O$: 1day post-oviposition)

Supplementary Figure S6



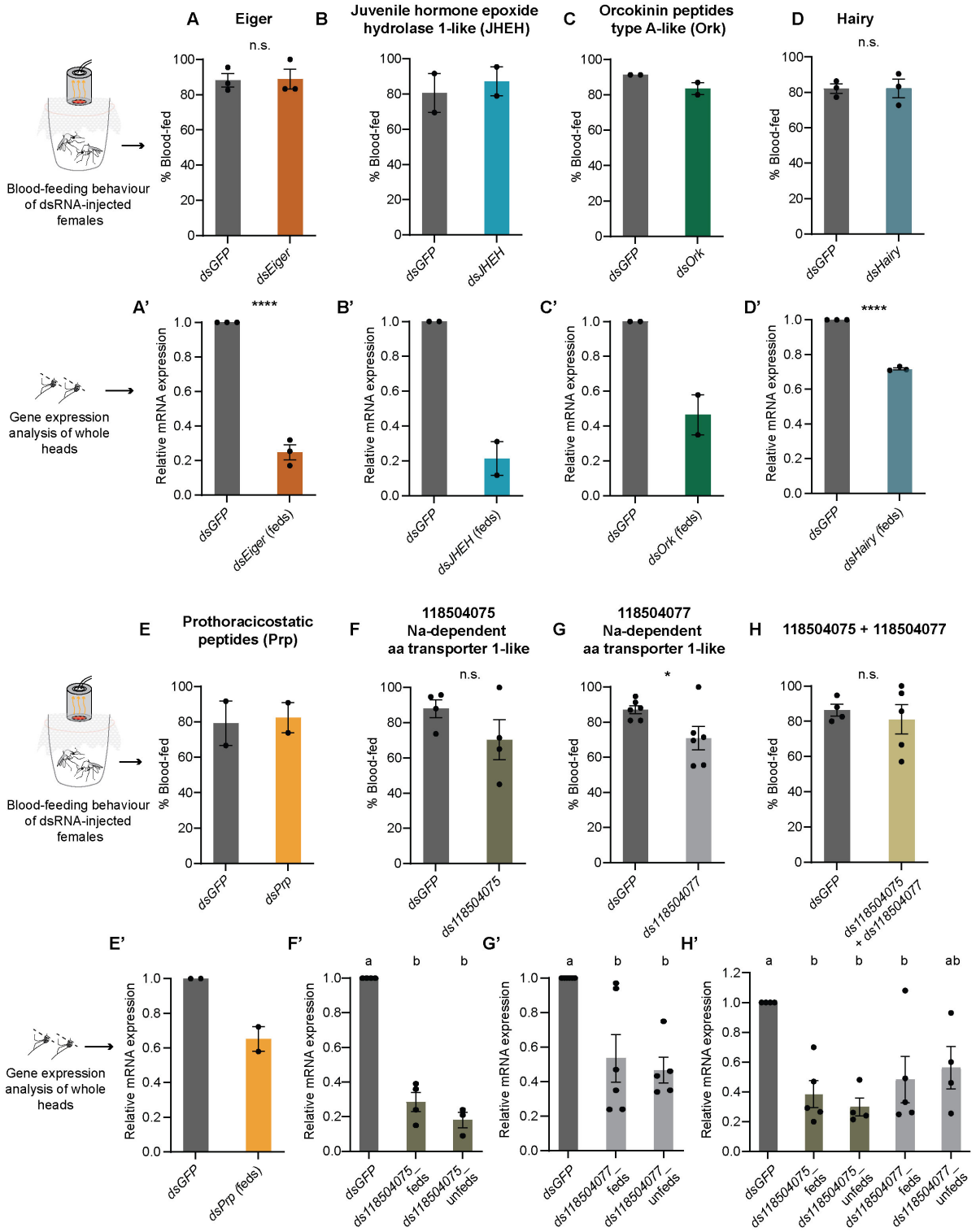
Supplementary Figure 6. Expression patterns of neurotransmitter-related genes in the central brain of *An. stephensi* across different states of blood-hunger and -satiety. Expression patterns of genes (listed on the right) involved in neurotransmitter expression, processing and transport across the reproductive cycle in the central brain, are represented as heatmaps. Interestingly, many of these genes were observed to be up-regulated in the newly emerged (D0) mosquitoes as compared to the older and blood-fed mosquitoes. RNA-seq data from all central brain samples in triplicates are represented as z-score normalised log10 (TPM+1) values. (D0: day of emergence; D5: 5days post-emergence, virgin; $D1^{B}$: 1day post-blood-meal, virgin; $D1^{B}M$: 1day post-blood-meal, mated; $D1^{O}$: 1day post-oviposition)

Supplementary Figure S7



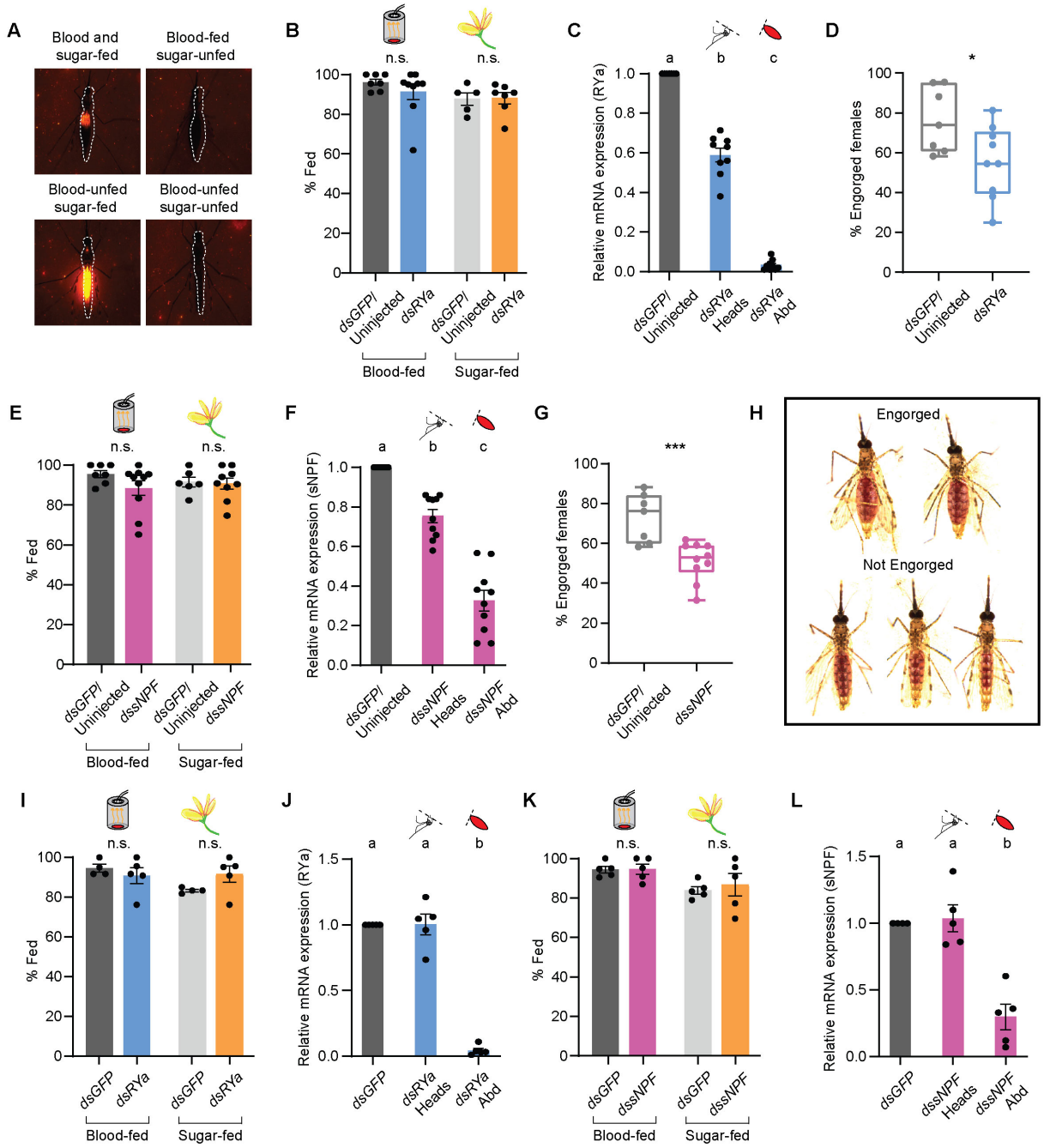
Supplementary Figure 7. Candidates potentially involved in promoting blood-feeding behaviour in *An. stephensi*. 71 candidate genes identified as potential promoters of blood-feeding behaviour are listed. These were identified to be commonly up-regulated in different conditions of blood-hunger (as compared to conditions of satiety or no-interest), and non-overlapping in males. Nine genes shortlisted for further validation are highlighted in red. RNA-seq data from all central brain samples in triplicates are represented as z-score normalised log10 (TPM+1) values. TPM values of zero were excluded from z-score transformation and are displayed as grey cells. (D0: day of emergence; D5: 5days post-emergence, virgin; $D1^{B}$: 1day post-blood-meal, virgin; $D1^{B}M$: 1day post-blood-meal, mated; $D1^{O}$: 1day post-oviposition)

Supplementary Figure S8



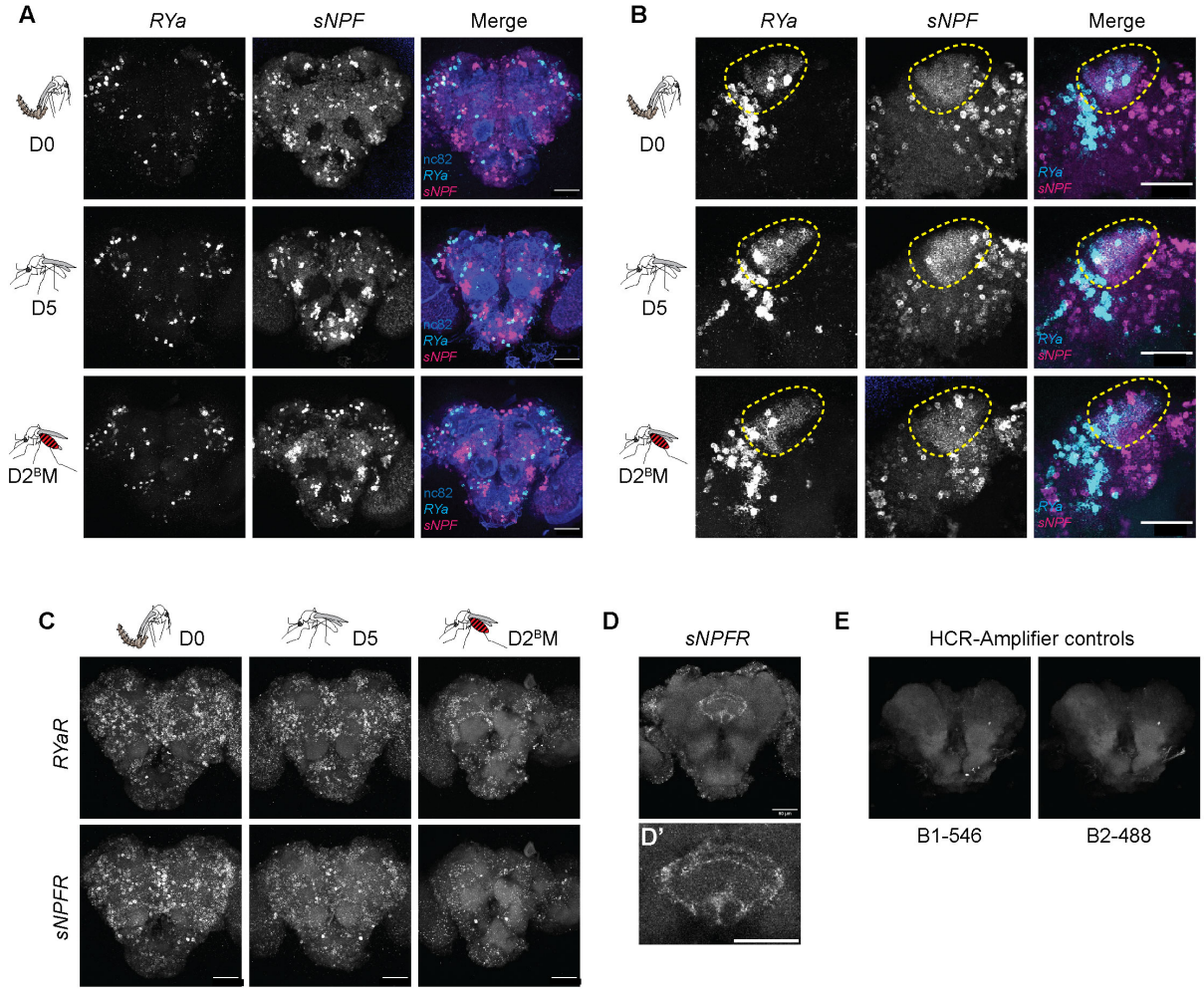
Supplementary Figure 8. Screening of the shortlisted candidates, potentially involved in promoting blood-feeding behaviour in *An. stephensi* via dsRNA-mediated gene knockdown. A-H. Blood-feeding behaviour of virgin females injected with dsRNA against *Eiger* (A), *Juvenile hormone epoxide hydrolase-1 like (JHEH)* (B), *Orcokinin peptides type-A like (Ork)* (C), *Hairy* (D), *Prothoracicostatic peptides (Prp)* (E), *Na-dependent amino acid transporter-1 like 118504075* (F), *Na-dependent amino acid transporter-1 like 118504077* (G), *118504075+118504077* (H). *dsGFP* was injected as a control in each case. No change in the behaviour was observed for any of the candidates tested. 18-25 females/replicate, n=2-6 replicates/group. Unpaired t-test; n.s.: not significant, p>0.05; *p<0.05 (A, D, F-H). A'-H'. Relative mRNA expression in the heads of virgin *An. stephensi* females injected with the indicated dsRNA, as compared to that in *dsGFP* injected females, analysed via qPCR. Efficient knockdown could not be achieved for *Hairy* and *Prp*. Unpaired t-test; ****p<0.0001 (A', D'). One-way ANOVA with Holm-Šídák multiple comparisons test, p<0.05 (F'-H'). For candidates *JHEH* (B), *Ork* (C) and *Prp* (E), significance could not assessed, as statistical testing requires a minimum of three replicates, which were not available for these experiments.

Supplementary Figure S9



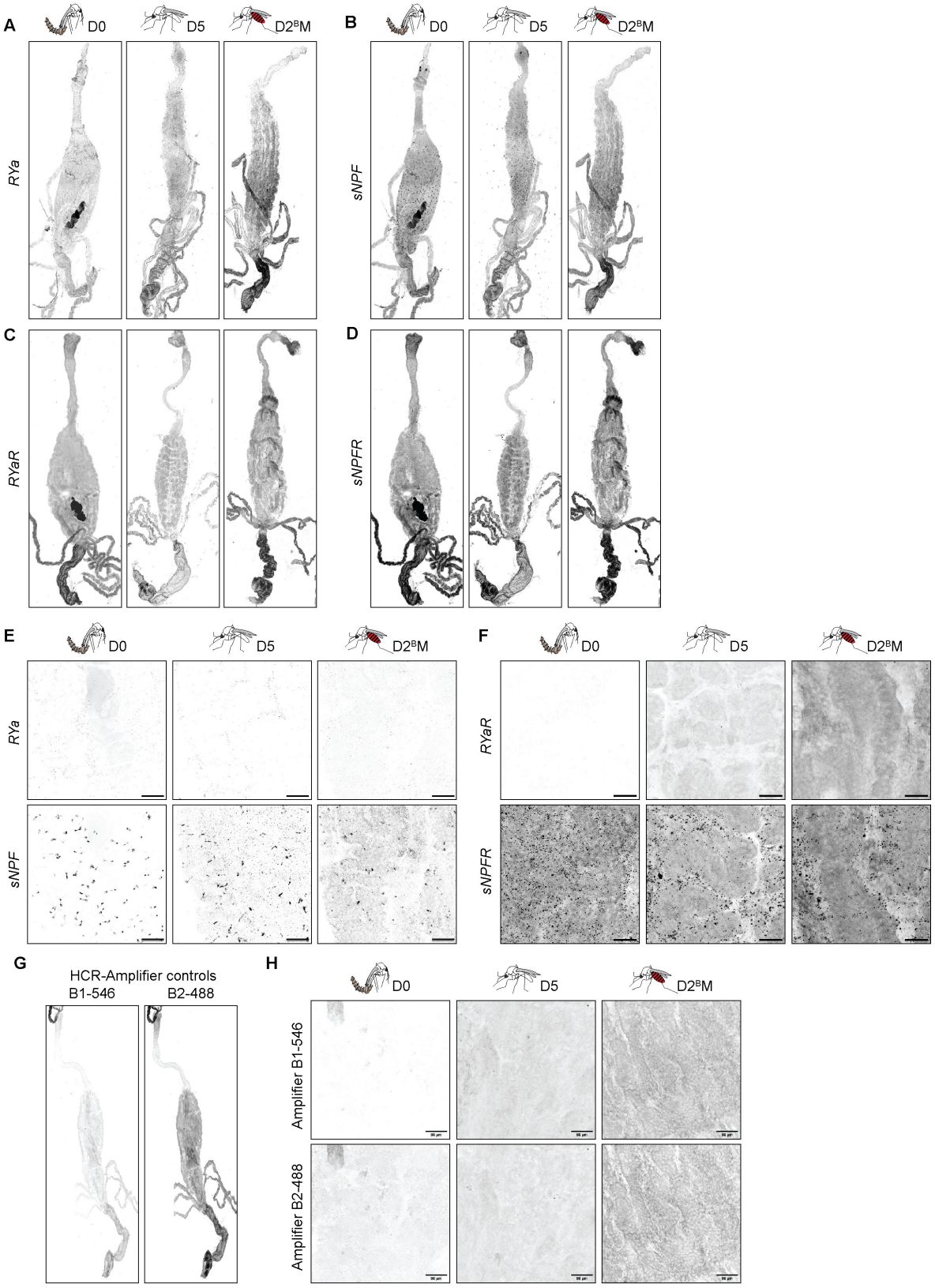
Supplementary Figure 9. Abdominal knockdown of *RYa* or *sNPF* alone does not contribute to the blood-feeding behaviour in *An. stephensi*. A. Widefield fluorescent images of females assayed for both sugar and blood appetite, post-dsRNA injections. Females were injected as described earlier (see methods for details) and offered Rhodamine B-spiked sugar 24h prior to the assay. Blood-feeding behaviour was tested as described earlier. Representative images of females fed on both blood and sugar, blood only, sugar only or none, are shown. B-H. Blood-feeding and sugar-feeding behaviour of virgin females injected with *dsRYa* (B) and *dssNPF* (E). *dsGFP* injected or uninjected females were used as controls. Relative mRNA expression in the heads and abdomens of the injected females were analysed via qPCR (C, F) to assay for the knockdown efficiency in both tissues. Percent females engorged on blood upon injection with *dsRYa* and *dssNPF*, as compared to control females are shown in D and G respectively. Representative widefield images of females considered to be engorged or not, are shown in H. While the mRNA levels in the abdomens were significantly downregulated, efficient knockdown could not be achieved in the heads. Under these conditions, no change in the blood- or sugar-feeding behaviour was observed for either of the targets. However, fewer females were able to fully engorge on blood in both cases and instead took smaller meals. 16-26 females/replicate, n=5-10 replicates/group. Unpaired t-test; n.s.: not significant, p>0.05; *p<0.05, ***p<0.001 (B,D,E,G). One-way ANOVA with Holm-Šídák multiple comparisons test, p<0.05 (C,F). I-L. Blood-feeding and sugar-feeding behaviour of females when *RYa* and *sNPF* are knocked down only in the abdomen. Virgin females were injected with dsRNA against *RYa* (H) and *sNPF* (J) and assayed for both sugar- and blood-appetite, as described before. *dsGFP* injected females were used as controls. Relative mRNA expression in the heads and abdomens of *RYa* (I) and *sNPF* (K) injected females were analysed via qPCR to assay for the knockdown efficiency in both tissues. No change in the blood- or sugar-feeding behaviour was observed when *RYa* or *sNPF* levels were significantly downregulated in the abdomens without altering the levels in the heads, suggesting that neuronal expression is necessary to promote the blood-feeding behaviour. 19-24 females/replicate, n=4-5 replicates/group. Unpaired t-test; n.s.: not significant, p>0.05 (I, K). One-way ANOVA with Holm-Šídák multiple comparisons test, p<0.05 (J,L).

Supplementary Figure S10



Supplementary Figure 10. Transcripts of neuropeptides *sNPF*, *RYa* and their receptors are expressed in *An. stephensi* central brain. A, B. *RYa* and *sNPF* mRNA expression in the central brain of females uninterested in blood (D0), blood-hungry females (D5) and blood-sated females ($D2^BM$), analysed via HCR. Transcripts of both neuropeptides are expressed in several clusters across the central brain (A). While the clusters are largely non-overlapping, co-expression was observed in the Kenyon cells of the mushroom body, indicated by the marked area (B). Maximum intensity projections of confocal stacks are shown. Scale bar, 50µm. C. *RYa receptor (RYaR)* and *sNPF receptor (sNPFR)* mRNA expression in the central brain of females uninterested in blood (D0), blood-hungry females (D5) and blood-sated ($D2^BM$), analysed via HCR. Transcripts of both receptors were found to be expressed in multiple cells. Maximum intensity projections of confocal stacks are shown. Scale bar, 50µm. D. *sNPFR* mRNA expression in the central complex. A magnified image is shown in D'. Maximum intensity projection of selected confocal stacks is shown. Scale bar, 50µm. E. central brain samples incubated with HCR-Amplifiers only, to serve as a control for background signal in the absence of HCR probes. No signal was observed for either of the amplifiers used (B1 labelled with 546 fluorophore (B1-546) and B2 labelled with 488 fluorophore (B2-488)). (D0: day of emergence; D5: 5days post-emergence, virgin; $D2^BM$: 2days post-blood-meal, mated)

Supplementary Figure S11



Supplementary Figure 11. Transcripts of *sNPF* and *sNPFR* are expressed in *An. stephensi* posterior midgut. A-F. *RYa* (A), *sNPF* (B), *RYaR* (C) and *sNPFR* (D) mRNA expression in the guts of females uninterested in blood (D0), blood-hungry females (D5) and blood-sated females ($D2^{B}M$), analysed via HCR. Transcripts of both *sNPF* and its receptor *sNPFR* are expressed in the posterior midgut, at all stages analysed. No expression of *RYa* or *RYaR* was observed. Magnified images of transcript expression of neuropeptides and their receptors are shown in E and F, respectively. Scale bar, 50µm. G,H. Gut samples incubated with HCR-Amplifiers only, to serve as a control for background signal in the absence of HCR probes. No signal was observed for either of the amplifiers used (B1 labelled with 546 fluorophore (B1-546) and B2 labelled with 488 fluorophore (B2-488)). Scale bar, 50µm. (D0: day of emergence; D5: 5days post-emergence, virgin; $D2^{B}M$: 2days post-blood-meal, mated)

Table 1. List of primers used in the study for *dsRNA*-mediated gene knockdown and quantitative PCR (qPCR)

S.No.	Primer name	Primer Sequence
1.	T7-GFP_F_RNAi	TAATACGACTCACTATAGGGAGACTGGTCGAGCTGGACGGCGA
2.	T7-GFP_R_RNAi	TAATACGACTCACTATAGGGAGACTTCTCGTTGGGGTCTTTGCTCAGG
3.	T7-RYamide_F_RNAi	TAATACGACTCACTATAGGGATGACCTGGCGAACGATGAAAC
4.	T7-RYamide_R_RNAi	TAATACGACTCACTATAGGGGGTCGCCATTCTCGTTGAACTGG
5.	RYamide_qPCR_F	ACCGATGCTACAGTCGTGATTCG
6.	RYamide_qPCR_R	TAGGCTAGTTGCATGGATCGTACC
7.	T7-sNPF_F_RNAi	TAATACGACTCACTATAGGGTTTGATGTCGGAGTCACTGCATCC
8.	T7-sNPF_R_RNAi	TAATACGACTCACTATAGGGTCACTCGCATCGTTAACCAACTGC
9.	sNPF_qPCR_F	CAATGAGCATCAGCTAGCACCG
10.	sNPF_qPCR_R	TCTAGCGCTTTATCCTCGGAGG
11.	T7-Hairy_RNAi_F	TAATACGACTCACTATAGGGCTCGTATCAACAACTGTCTGAACG
12.	T7-Hairy_RNAi_R	TAATACGACTCACTATAGGGGTACTTGGGTGGTGAGCGTTTGG
13.	Hairy_qPCR_F	AATCGTCGGAGCAACAAACCGATC
14.	Hairy_qPCR_R	GATGCTTCACCGTCATCTCCAGG
15.	T7-Eiger_RNAi_F	TAATACGACTCACTATAGGGCTGGACGATAGCGAGGAAAAGG
16.	T7-Eiger_RNAi_R	TAATACGACTCACTATAGGGGCTCATTCGGTTCAGTATTTGTGC
17.	Eiger_qPCR_F	GGGCGTAAGACATCACTCACGC
18.	Eiger_qPCR_R	TACGATTTCCTTCTGCGTTCCAGG
19.	T7-JHEH_RNAi_F	TAATACGACTCACTATAGGGGATTATTGGGGACCCGGAAATGG
20.	T7-JHEH_RNAi_R	TAATACGACTCACTATAGGGCGTTTGTGACCGATGCGTTCC
21.	JHEH_qPCR_F	ATCGCCAGTGCCAGTTGAGC
22.	JHEH_qPCR_R	TTGCTCCATTTCCGGGTCCC
23.	T7-Prp_RNAi_F	TAATACGACTCACTATAGGGAATGTATCGCGGTCTGCTTTGC
24.	T7-Prp_RNAi_R	TAATACGACTCACTATAGGGCGACTAATTTCCTGCCGTTTGCC
25.	Prp_qPCR_F	AAGTTCGGTGCGGCCTGG
26.	Prp_qPCR_R	TCTGCTGCTGCTGCTGCTG
27.	T7-4075_RNAi_F	TAATACGACTCACTATAGGGGATACGGTCACTTCGCTGGTGG
28.	T7-4075_RNAi_R	TAATACGACTCACTATAGGGATGAACAGTACGGTGATGGGCG
29.	4075_qPCR_F	CCTTCGCCTGGATCTATGGCG
30.	4075_qPCR_R	CGAGCGCATTGTAACCGAGTGG
31.	T7-4077_RNAi_F	TAATACGACTCACTATAGGGCTCGACACATTCACCTCCATCG
32.	T7-4077_RNAi_R	TAATACGACTCACTATAGGGATAAGAATGAGCAGTGTGGCGG
33.	4077_qPCR_F	ATCTACGGCGTGGATCGTATCTGC
34.	4077_qPCR_R	CCTAAAACGTTGTAGCCGATCGGG
35.	T7-Orcokinin_F_RNAi	TAATACGACTCACTATAGGGGTCTGTGCCGTACTGCTGTTCG
36.	T7-Orcokinin_R_RNAi	TAATACGACTCACTATAGGGGATCGTGCTTCAGGTTGTTACCGG
37.	Orcokinin_qPCR_F	GCTGAACGATGGCCAACTAAAGC
38.	Orcokinin_qPCR_R	CTTGTCGTACATCGGTGCCAATCC
39.	RpS7_qPCR_F	AGGTTGTCGGCAAGCGTATCC
40.	RpS7_qPCR_R	AGCTTCTTGTACACCGACGCG