High-throughput screening of more than 30,000 compounds for anthelmintics against gastrointestinal nematode parasites

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Abstract:

Gastrointestinal nematodes (GINs) are amongst the most common parasites of humans,

livestock, and companion animals. GIN parasites infect 1-2 billion people worldwide,

significantly impacting the growth, health, and vitality of hundreds of millions of children,

pregnant women, and adult workers in developing countries, thereby perpetuating poverty. The

cost and losses associated with GIN infections in human and veterinary health are estimated to

be hundreds of billions of dollars worldwide. Two benzimidazoles with suboptimal efficacy are

currently used to treat GINs in humans as part of mass drug administrations, with many

additional instances of lower-than-normal efficacy and possible resistance. Many livestock and

companion animal GINs are resistant to multiple classes of anthelmintic drugs used today. Thus,

new anthelmintics are urgently needed. However, screening methods for new anthelmintics using

human GINs typically have a low throughput. Here, using our novel screening pipeline that starts

with human hookworm parasites, we screened 38,293 conditions in duplicate representing

30,238 unique small molecules from a wide range of compound libraries, including generic

diversity, repurposing, natural derivatives, mechanism of action, and multiple target-focused

libraries (*e.g.*, targeting kinases, GPCRs, and neuronal proteins). We identified 55 compounds

with broad-spectrum activity against adult stages of two evolutionary divergent GINs,

hookworms (*Ancylostoma ceylanicum*) and whipworms (*Trichuris muris*). Based on known

databases, the targets of these 55 compounds were predicted in nematode parasites. One novel

scaffold from the diversity set library, F0317-0202, showed good activity (≥~11% Relative

motility) against both GINs. To better understand this novel scaffold's structure-activity

relationships (SAR), we screened 28 analogs and created SAR models highlighting chemical and

functional groups required for broad-spectrum activity. These studies validate our new and

efficient screening pipeline at the level of tens of thousands of compounds and provide an

important set of new GIN-active compounds for developing novel and broadly-active

anthelmintics.

Keywords:

Introduction:

Human gastrointestinal nematodes (GINs, also known as soil-transmitted helminths) include the roundworm *Ascaris lumbricoides*, the whipworm *Trichuris trichiura*, and the hookworms *Necator americanus*, *Ancylostoma duodenale*, and *Ancylostoma ceylanicum*; they are amongst the most prevalent infections of the so-called Neglected Tropical Diseases (NTDs)[1], affecting people in the world's poorest communities[2-4]. GINs are successful parasites of vertebrates, adapted to parasitize humans, livestock, and companion animals[5]. Morbidities associated with GIN infections in humans include anemia, malnutrition, growth stunting, cognitive impairment, pregnancy complications, and lowered immunity against other infectious diseases[2, 4, 6]. The negative impacts of GINs are hard to measure, but they are minimally responsible for > 2-3 million disability-adjusted life years (DALY)[3, 6, 7], with hookworms accounting for almost one-half of those DALYs[3, 6]. Children who received hookworm treatment had increased school attendance and adult income[4, 8]. GINs also have enormous impacts on the health and productivity of farm animals[9].

The World Health Organization (WHO) has recommended annual or semi-annual Mass Drug Administration (MDA) targeting preschool and school-age children to reduce worm burden and associated morbidity[10]. Four anthelmintics (levamisole, pyrantel pamoate, albendazole, and mebendazole), initially developed for livestock, are currently on the WHO list to control human GINs, with the two benzimidazoles, albendazole and mebendazole, most widely used because of their better efficacies and ease of dosing[11, 12]. In companion and livestock animals, GINs are controlled via frequent and routine treatment using a repertoire of anthelmintics, which has led to widespread and sometimes multi-drug anthelmintic resistance[13-16]. Benzimidazole resistance is

well-established in veterinary parasites[17-20]. Single nucleotide polymorphisms (SNPs) in the beta-

tubulin gene linked to the benzimidazole resistance in veterinary parasites have been found in

human GINs[21-24], indicating that with high drug pressure, there is likely to be an emergence for

benzimidazole resistance in human GINs. In Haiti and Kenya, the frequency of benzimidazole-

resistance codon 200 in *T. trichiura* was significantly increased following local MDA

programs[22]. There are also multiple reports of lowered efficacy of benzimidazoles against human

GINs and warnings against drug resistance[25-27]. Notwithstanding these challenges, the WHO has

set an ambitious goal to eliminate human GINs by 2030[10]. Therefore, potent and broadly active

anthelmintics with new modes of action are urgently needed for long-term control and

elimination. One promising new drug is emodepside, which was discovered >30 years ago [28].

However, it has not yet been approved for human use, and, as with all single anthelmintics,

resistance is a concern[29, 30]. Given the lead time for new drugs to enter the clinic, it is urgent that

new lead anthelmintics be put into the pipeline.

Because understanding of these parasites' biology is still limited, whole organism phenotypic

screening remains a preferred approach for anthelmintic drug discovery[31, 32]. Phenotypic

screening using GIN parasites for anthelmintic has included screening against *ex vivo* adult

parasites, egg hatch assay, larval stages, and infective third-stage larvae[32]. However, regarding

GINs that infect humans, the throughput of these screens has been limited[32-35]. The free-living

nematode *Caenorhabditis elegans* has been used as a surrogate model for GINs that infect

humans[32, 36-39]. However, we found limitations to using *C. elegans* as a model for anthelmintic

screening targeting human GINs, most notably a significant false negative hit rate[31]. Thus, *C.*

elegans is likely to miss some of the most promising compounds. We devised an alternative

screening model based on the larval stages of human hookworm parasites, which we found

predicted better anthelmintic activity against GIN adult parasites than *C. elegans* [31]. In our

pipeline, this is followed by screening against adult stages of evolutionarily divergent hookworm

and whipworm parasites[31]. In a pilot screen, 1,280 compounds were screened as proof of

concept[31].

Here, we scale up this anthelmintic discovery pipeline and screen 38,293 conditions **(**76,586 in

duplicate) representing 30,238 unique compounds. We identified 55 small molecules with *in*

vitro efficacy against adult hookworms and whipworms. We focused on one novel scaffold and

performed Structure-Activity Relationship (SAR) studies. Based on analogies from other

systems, we propose essential GIN targets for further studies.

Figure 1: Novel anthelmintic discovery pipeline for human GINs. Our pipeline is centered around the zoonotic hookworm, *A. ceylanicum,* as a primary (larval) and secondary (adult) screening model, followed by tertiary screening against the murine whipworm *Trichuris muris* (adult) to identify broadly active compounds. Hits are subjected to deep data mining, and analogs of potential candidates are tested to generate the Structure-Activity Relationships. This report covers the results from the first five stages of the screening pipeline (in blue).

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Results:

We previously described and validated a pilot scale pipeline to discover anthelmintics that

broadly target human GINs (Figure 1)[31]. We scaled up our primary screen with modifications to

include 30,000+ compounds in 13 chemical libraries (Figure 2). Starting with the first larval (L1)

staged *A. ceylanicum* hookworms, we screened these libraries, encompassing many structural

features, including diversity scaffolds, natural product derivatives, repurposing drugs, known

modes of action, and target-based libraries (Table 1). In total, we screened 30,238 unique

151 compounds in duplicate. Some compounds were screened at multiple doses due to plate format

152 availability, making the total number of individual conditions screened 38,293 (76,586 when

153 considered in duplicate). All libraries were initially screened in duplicate at 10 µM. As per our

154 pipeline, primary screen hits (in both duplicate wells) were screened at 30 µM against adult *A.*

155 *ceylanicum* hookworms. Adult hookworm hits were then screened at 30 µM against adult *T.*

156 *muris* whipworms. A breakdown of hits "library by library" is given below and summarized in

157 Table 1.

Figure 2: Primary screening using larval development of *A. ceylanicum* hookworms. To work around variability in egg hatch rates, we found that starting with the first larval stage led to more consistent and robust results (drugs that exclusively targeted egg hatch were very uncommon)[31]. Isolated and purified eggs were allowed to hatch in S-media into synchronized L1. Each 96-well assay plate includes four wells of ivermectin (10µM) as positive control and four wells of 0.25% DMSO as negative controls. After seven days of incubation at 28°C, assay plates were scored under a light microscope. The test compounds were considered active (hit) if at least 90% of all larvae in the well were killed as L1 or failed to develop. Scale bar 1 mm.

Library	Unique Compounds	Screened doses μM	test wells screened in duplicate	*A. ceylanicum* L1 at 10μM (% hits)	*A. ceylanicum* adults 30μM (% hits)	*T. muris* Adults 30μM (% hits)
Life Chemical Diversity Set	15360	10	15360	491 (3.2)	33 (0.21)	7 (0.05)
Broad Institute REPO 1, 2 and 3	6743	10	6743	230 (3.4)	96 (1.42)	36 (0.53)
ICCB Longwood MOA	1245	10, 2, 0.4, 0.08	4980	65 (5.3)	17 (1.36)	9 (0.72)
ICCB Selleck Neuronal signaling	1031	10	1031	29 (2.8)	12 (1.16)	2 (0.19)
ICCB SYNthesis Kinase Inhibitor 2	96	10, 2, 0.4	288	8 (8.3)	1 (1.04)	1 (1.04)
ICCB EMD Kinase Inhibitor 1	244	10	244	10 (4.1)	2 (0.81)	2 (0.81)
ICCB ChemBridge Focused Kinase	250	10	250	2 (0.8)	0	0
ICCB ChemBridge Focused GPCR	250	10	250	2 (0.8)	0	0
ICCB LINCS 1 Kinase inhibitors	88	10, 2, 0.4, 0.08	352	4 (4.5)	2 (2.27)	1 (1.13)
ICCB LINCS 2 Kinase inhibitors	88	10, 2, 0.4, 0.08	352	2 (2.25)	0	0
ICCB LINCS 4 Kinase inhibitors	169	10, 2, 0.4, 0.08	676	8 (4.7)	3 (1.77)	3 (1.7)
SMSF Kinase Inhibitors	492	10	492	13 (3)	5 (1.01)	3 (0.69)
SMSF TEMTEC Natural Compounds	4240	10	4240	14 (0.33)	0	0
Total	**30238**		**38293**	**878 (2.9)**	**171 (0.56)**	**64 (0.21)**

Table 1: Screening summary of 30,238 compounds from 13 different libraries with information on screening doses, the number of hits per screening model, and the hit rates.

Hits from the Generic Diversity Set Library:

Of the 15,360 compounds in the generic diversity set tested against *A. ceylanicum* hookworm

larvae, 491 were active, with a hit rate of 3.2%. These 491 larval hits were tested against the

early adult stages of *A. ceylanicum* hookworm, yielding 33 hookworm adult-active compounds.

An example of screening against the early adult stages of *A. ceylanicum* hookworms is shown in

Figure 3. These 33 *A. ceylanicum* hookworm adult-active compounds were screened against *T.*

muris adult whipworms, yielding seven hits active against the two evolutionarily divergent

parasites. An example of screening against adult *T. muris* whipworms is shown in Figure 4.

Figure 3. Young adult *A. ceylanicum* hookworm screens. The top panel shows healthy and motile worms from the DMSO-treated or nonactive test compounds. The lower panel shows dead or intoxicated worms from ivermectin or active compounds. Because young adult hookworms are much smaller than whipworms, we manually scored these versus using the Worminator. Scale bar 1 mm.

Figure 4: Adult parasite screening of *T. muris* whipworm**.** (A), Adult whipworms, male and female treated with 30µM Levamisole or 1% DMSO. Scale bar 1 mm. (B): Motility readout screenshot from Wormassay-6 software using the Womrinator imaging setup. (C). Example of calculated motility

readout, showing adult-parasite actives with Relative Motility (R.M.) less than 50% compared to the DMSO control.

Hits from the Repurposing and Mechanism of Action (MoA) Libraries

Of the 6,743 compounds tested from the repurposing library, we identified 230 *A. ceylanicum*

hookworm larval-active compounds (hit rate of 3.4%), from which 96 showed activity against

adult *A. ceylanicum* hookworms (hit rate 1.42%). Of these 96 compounds, 36 were active

against adult *T. muris* whipworms with a final hit rate of 0.53%. Similarly, The ICCB-Longwood

MoA library contained 1,245 high-quality compounds with known mechanisms of action. We

identified 65 *A. ceylanicum* hookworm larval-active compounds with a 5.2% hit rate, the highest

among all tested libraries. Seventeen of these compounds carried adulticidal activity against *A.*

ceylanicum hookworms, from which nine were active against *T. muris* whipworm adults, with a

final hit rate of 0.72%.

Hits from the Neuronal and the GPCR libraries:

Of the 1031 compounds with known modulation of neuronal signaling, we identified 29 *A.*

ceylanicum hookworm larval-active compounds (hit rate of 2.8%). Of these 29 compounds, 12

showed activity against adult *A. ceylanicum* hookworms (1.1% of the total), from which two

were active against *T. muris* whipworm adults with a final hit rate of 0.19%. We also screened

250 known GPCR inhibitors and identified two compounds with larval activity against *A.*

ceylanicum; however, these compounds did not show activity against the adult stages of

parasites. One explanation for the lack of success with this library is that these inhibitors were

tailored initially for selective binding to human GPCRs.

Hits from the Kinase Inhibitor Libraries:

Of seven kinase inhibitor libraries with 1427 compounds (Table 1), we identified 47 with *A.*

ceylanicum larval activity, giving a hit rate of 3.2%. Of those 47 compounds, 13 showed

activities against the adult stages of *A. ceylanicum* hookworms (0.9%), of which 10 compounds

were also active against *T. muris* whipworm adults with a hit rate of 0.7%.

Hits from the Natural Products Library:

210   Of 4,240 purified compounds from natural products or derived from natural products, we

211   identified 14 compounds with *A. ceylanicum* larval activity, giving the lowest primary screen hit

212   rate of 0.33%. None of these 14 larvicidal compounds showed activity against the adult stages of

213   *A. ceylanicum* hookworms.

Summary of 30,238 compound screen

Collectively, we identified 878 compounds with *A. ceylanicum* larval activity, giving an overall

primary screen hit rate of 2.9% (Table 1). In our secondary screen, we tested all larval-active

compounds at 30 µM against the adult stages of *A. ceylanicum* hookworm parasites. Of those,

171 (19.5% of the larval hits) had anthelmintic activity toward adult hookworms, representing a

hit rate of 0.5% based on the total number of compounds in the initial libraries. These 171

hookworm-adult actives were tested against adult *T. muris* whipworms at 30 µM. Sixty-four

compounds (37% of the adult hookworm hits and 0.21% based on the total number of

compounds in all the libraries) showed activity against whipworm adults (Table 1). Of the 64

compounds identified from all libraries, nine were redundant (included independently in multiple

libraries), yielding 55 unique active compounds, listed in Table 2.

Screened libraries contained known anthelmintic compounds, and we successfully identified

albendazole, oxibendazole, flubendazole, methiazole, oxfendazole, thiabendazole, ivermectin,

abamectin, dormamectin, emamectine, eprinomectin, selamectin, emodepside, closantel,

monepantel, and moxidectin as part of the primary and secondary *A. ceylanicum* hookworm

screens. However, many of these anthelmintics didn't show activity in our tertiary screening

against the *T. muris* whipworms at 30μM, highlighting the whipworms' resilience to

anthelmintics compared to hookworms. For example, the published 50% inhibitory

concentrations (IC_{50}) of albendazole and ivermectin against *T. muris* whipworms are both >

200μg/ml[40]. Only anthelmintic compounds that showed activity at 30μM against *A. ceylanicum*

and *T. muris* are shown in Table 2.

Data Mining

The 55 broadly active compounds were subjected to a literature search to identify, if known,

their possible mode of action and the eukaryotic drug targets in other organisms and the

orthologous targets in phylum Nematoda, focusing on *C. elegans* in clade V and *T. muris* in

clade I (Table 2). The compounds encompass a wide range of putative target classes including

potassium channel, glutamate-gated chloride channel, acetylcholinesterase, acetylcholine

receptor, tyrosine kinases, aurora kinase, phosphatidylinositol 3-kinase (PI3K), tyrosine

phosphatase, insulin growth factor receptor (IGF1-R), nitric oxide synthase (NOS), Exchange

Protein Activated by Cyclic AMP (EPAC), pro-survival protein (BCL2), ATP synthase,

translation elongation factor, Enoyl-acyl carrier protein reductase (ENR), p38 mitogen-activated

protein Kinase (p38 MAPK), sphingosine 1-phosphate receptor (S1PR), Heat Shock Protein-90

(HSP-90), Dopamine D2 receptor, liver X receptor (LXR), serotonin (5-HT)3 receptor,

chemokine CXCR3 receptor, and ubiquitin specific peptidase 7 (USP-7) and Phosphatase

localized to Mitochondrion 1(PTPMT1). Compounds identified here with well-known targets

and mechanisms of action in other organisms may be used as probes for druggable and essential

ortholog targets in nematode parasites (Table 2). For example, alexidine was shown to induce

selective responses similar to the knockdown of its target PTPMT1[41]. Similarly, T0901317 is a

well-known selective agonist of LXR used in many studies to investigate the physiological

effects of LXR on cholesterol and lipid homeostasis; notably, T0901317 induction of LXR was

found to reduce cholesterol biosynthesis and to induce cholesterol breakdown into bile acids [42-44].

As examples, we discuss putative nematode targets of four of these-- BCL2, NOS, PTPMT1, and

LXRs, selected because these targets are known to be essential (See Supplementary Discussion

Part A).

Drug Names	Structure	Source Library	HW M.I. 30µM	WW % R.M. 30µM	Known MoA /Target	Target orthologue in Nematodes
Emodepside		REPO	0	8.6	Anthelminthic, bind to nematode Latrophilin-like receptor (LAT-1), Voltage-gated activated potassium channels (SLO-1) [45, 46]	Lat-1 and slo-1in *C. elegans*, TMP05851 and TMP00134 in *T. muris*
Metrifonate; TRICHLORFON		REPO	0.75	1.3	Acetylcholinesterase (ACHE) inhibitor [47]	ace-1 in *C. elegans*, TMP09383 in *T. muris*
Tyrphostin-A9; Malonoben		REPO, SMSF kinase	0	9.1	Protein tyrosine kinase receptor inhibitor/ Epithelial Growth Factor Receptor (EGFR), Platelet-Derived Growth Factor Receptor (PDGFR) [48]	Let-23, kin-30, in *C. elegans*, TMP05029 and TMP12675 in *T. muris*
Brilliant-green		REPO	0	5.1	--	--
Alexidine		REPO	1.75	1.6	Protein Tyrosine Phosphatase localized to the Mitochondrion 1(PTPMT1) [49]	F28C6.8 in *C. elegans*, TM12441 in *T. muris*
Octenidine		REPO	0.75	7.3	--	--
Walrycin-B		REPO	0	8.6	--	--
EVP4593; QNZ		REPO, ICCB MOA, EMD kinase, LINCS Kinase	0	35.8	TNF-α and NF$_k\beta$ pathway inhibitor [50]	--
AG-1024		REPO	2	7.5	Protein tyrosine kinase receptor inhibitor/ Insulin growth factor receptor inhibitor/ (IGF1R) [51]	Daf-2 in *C. elegans*, TMP10632 in *T. muris*

Tyrphostin AG879		REPO, SMSF kinase	0	20	Protein tyrosine kinase receptor inhibitor/Nerve Growth Factor Receptor (NGFR) [48]	--
Diphenyleneiodonium		REPO	0	15.2	Nitric oxide synthase inhibitor/ NOS2, NOS3 [52]	Lacked in *C. elegans* NOS-Like in *A. suum*
ESI-09		REPO	0.5	16.2	Exchange protein activated by cAMP isoform 2 (EPAC2) inhibitor [53, 54]	epac-1 in *C. elegans*, TMP12712 in *T. muris*
Obatoclax		REPO, ICCB MOA	0.5	10.7	Induce apoptosis via the inhibition of the pro-survival protein/ BCL2[55]	ced-9 in *C. elegans*, TMP02363 in *T. muris*
Oligomycin-A		REPO	1.25	15.9	Mitochondrial ATP synthesis inhibitor/ ATP5A[56]	atp-1 in *C. elegans*, TMP09383 in *T. muris*
Pyrithione-zinc		REPO	0	3.6	KCNQ Kv7 potassium channels agonist [57]	kqt-1 in *C. elegans*, *TMP05091 in T. muris*
NH125		REPO, LINCS 1 kinase	1.25	8	Eukaryotic translation elongation factor 2 kinase (EEF2K) inhibitor [58]	efk-1 in *C. elegans* and TMP13626 in *T. muris*
Tetramisole		REPO	2.5	9.4	Anthelminthic, acetylcholine receptor agonist [59]	acetylcholine receptor agonist
Levamisole		REPO	1	10.1	Anthelminthic, acetylcholine receptor agonist [59]	acetylcholine receptor agonist
Triclocarban		REPO	0	12.7	--	--
Semapimod		REPO	1.75	39.7	Inhibit Toll Like Receptors (TLR) signaling by targeting TLRs chaperone gp96 [60]	*enpl-1* in *C. elegans* and TMP11927 in *T. muris*
Amiselimod		REPO	1	3.3	Sphingosine 1-phosphate receptor (S1PR) modulator [61]	Spin-2 in *C. elegans* and

Compound	Structure				Mechanism	Target
						TMP13635 in *T. muris*
Cetylpyridinium		REPO	0.25	15.1	HSP-90 inhibitor [62]	Hsp-90 in *C. elegans*, TMP11927 in *T. muris*
Domiphen		REPO	0	11.5	Blockade of human ether-a-go-go related gene (HERG) potassium channels [63]	unc-103 in *C. elegans*, TMP12566 in *T. muris*
Sulconazole		REPO	1	3.1	Sterol 14 alpha Demethylase cytochrome (CYP-51) inhibitor [64]	--
MLN8054		REPO	1.5	18	Aurora Kinase inhibitor/ AURKA [65]	Air-2 in *C. elegans*, TMP01390 in *T. muris*
Cetrimonium		REPO	0.25	2.7	--	--
Econazole		REPO	1.2	7.6	Sterol 14 alpha Demethylase cytochrome (CYP-51) inhibitor [64]	--
Crystal-violet		REPO	0.25	5.3	HSP-90 inhibitor [62]	Hsp-90 in C. elegans TMP11927 in *T. muris*
Chlorpromazine		REPO	0.2	2.7	Dopamine D2 receptor antagonist [66]	dop-2 and dop-3 in C. elegans and TMP13188 in *T. muris*
Dioscin		REPO	1.5	15.3	Antagonize chemokine receptor CXCR3 block interaction to IP-10 cytokine [67]	Npr-20 in *C. elegans*, TMP12387 in *T. muris*

Name	Structure	Source	Value 1	Value 2	Mechanism	Organism
Siramesine; Lu28-179		REPO	2	2.1	σ_2R receptor agonist [68]	--
Selamectin		REPO	1	17.1	Anthelminthic, Neuromuscular paralysis via activation of Glutamate-gated chloride channels [69-71]	avr-14 in *C. elegans*, TMP04454 in *T. muris*
Isoconazole		REPO	0.4	3.1	Sterol 14 alpha Demethylase cytochrome (CYP-51) inhibitor [64]	--
ML281		REPO	1.5	16.7	Serine/threonine kinase inhibitor/ STK33 [72]	--
Abamectin		REPO	0	13.1	Anthelminthic, Neuromuscular paralysis via activation of Glutamate-gated chloride channels [69-71]	avr-14 in *C. elegans*, TMP04454 in *T. muris*
Squalamine		REPO	1	3.1	protein tyrosine phosphatase (ptpn1b) inhibitor [73]	Sdf-1in *C. elegans*, TMP05082 in *T. muris*
Trodusquemine		REPO	0.25	2.1	protein tyrosine phosphatase (ptpn1b) inhibitor[73]	Sdf-1in C. elegans, TMP05082 in *T. muris*
Bafilomycin A1		ICCB MOA	0.5	52.7	Disrupt Autophagic flux by inhibiting V-ATPase and Ca-P60A/SERCA[74]	Vha-19 and sca-1 in *C. elegans*, TMP10618 and TMP09741 in *T. muris*
NMS-873		ICCB MOA	0	34.5	Allosteric inhibitor of the ATPase	cdc-48 in *C. elegans*,

Name	Structure	Library			Target	Organism
					valosin-containing protein (VCP/p97) [75]	TMP02658 in *T. muris*
1-Phenyl-3-[4-(trifluoromethox)phenyl]urea		ICCB MOA	0	50.2	--	--
2,3-Diphenyl-5-[(2-methoxyphenyl)amino]-1,2,4-thiadiazole-2-ium		ICCB MOA	0	41.7	--	--
Furan-2-ylmethyl-(5-phenyl-thieno[2,3-d]pyrimidin-4-yl)-amine		ICCB MOA	0	41.9	--	--
4-Bromo-N-[2,2,2-trichloro-1-(4-chloroanilino)ethyl]benzamide; AMG7160		ICCB MOA	2	27.6	Ca^{2+} Cation channel, transient receptor potential ankyrin 1 (TRPA1) antagonist [76]	Trpa-1 in *C. elegans*, TMP11515 in *T. muris*
T0901317		Selleck Neuronal	2	37.6	Liver X receptor LXR (Nr1H2, Nr1H3) agonist [42-44, 77]	nhr-247 and nhr-8 in *C. elegans*, TMP09010 and TMP06463 in *T. muris*
Vortioxetine; Lu AA21004		Selleck Neuronal	2	30.9	5-HT3-R antagonist [78, 79] and serotonin transporter (SERT) inhibitor [80]	Lgc-7, lgc-8 *and mod-5* in *C. elegans,* TMP06220, TMP00636 and TMP10599 in *T. muris*
Sorafenib		SMSF, SYNthesis2, LINCS 1 kinase	0	20.6	Inhibit Raf-1, BRAF (serine/threonine Kinase), VEGFR, PDGFR, KIT, FLT3, FGFR1(tyrosine kinase) [81]	Lin-45, ver-1, ver-4, kin-30, old-2 and igcm-4 in *C. elegans* TMP03531, TMP12675, TMP13825, TMP00128, TMP02580, TMP02073 in *T. muris*

	Structure	Source	Col1	Col2	Mechanism	Ortholog
Staurosporine		EMD Kinase	1.25	27.7	Promiscuous Protein Kinase inhibitor [82]	
NVP-BGT226; HMSL10349-106		LINCS 1 Kinase	1.5	0	phosphatidylinositol 3-kinase (PI3K) inhibitor and MTOR [83]	age-1 in *C. elegans*, TMP06498 in *T. muris*
F0317-0202		Diversity set	0	10.7	--	--
F1059-0041		Diversity set	1.75	19.5	--	--
F1620-0033		Diversity set	1	13	--	--
F9995-0076		Diversity set	0	3.7	--	--
F9995-0233; dithiazanine iodide		Diversity set	0	1.75	Anthelminthic activity via inhibition of oxygen consumption and interreference with glucose uptake [84-88].	--
F0900-4647		Diversity set	0	3	--	--
F1345-0623; HBX		Diversity set	0.5	9	USP7 inhibitor [89]	math-33 in *C. elegans*, TMP04955 in *T. muris*

Table 2: List of 55 broadly active compounds including chemical structure, source library, *in vitro* activity against *A. ceylanicum* hookworms (HW motility index (M.I.) where 3 is healthy and 0 is completely immotile), *in vitro* activity against *T. muris* whipworms (WW percent Relative Motility (% R.M.),100% is healthy; 0% is non-motile) to the DMSO control, known mode of action in eukaryotic organisms, and the orthologous targets in nematodes if any. (--) no known target.

Novel anthelmintic scaffold:

We identified seven compounds with broad anthelmintic activity from the Life Chemical Diversity Set, which is compiled of *de novo* and newly synthesized molecules developed for initial phenotypic screening. Among these seven actives from this library of novel scaffolds, we identified F0317-0202 as a potential lead for further studies, given its better drug-like structure over the other six. To further investigate, 28 analogs of F0317-0202 were identified using structural similarity online searching tools and purchased from a commercial vendor (Life Chemical), which could serve as training sets for ligand-based optimization and generate the SAR model. We screened this set of analogs against adult *T. muris* whipworms, followed by older and larger *A. ceylanicum* hookworm adults, which could read more easily in the Worminator.

Of the 28 analogs, we identified 12 compounds (41%) that inhibit *T. muris* whipworms motility by $\geq$ 50% relative to the DMSO control (Figure 5, Supplementary Figure 1). All 12 were tested and found to be active against *A. ceylanicum* hookworm adults (Figure 5 and Supplementary Figure 1). Of these 12 broadly active compounds, four (F0317-0013, F0317-0161, F0328-0210, and F0317-0009), along with the parent, showed near-complete motility inhibition at 30µM against the two parasites. Conversely, of the 28 analogs, 16 showed significantly reduced/no activity against whipworm adults.

Based on these data, an F0317-0202 SAR model for potent and broad anthelmintic activity was

built and found to be quite strict— slight modifications could abolish the anthelmintic activity

entirely. The functional core identified in this potent scaffold was the 1,3-benzothiazol-2-

ylidene-3-oxopentanenitrile (Figure 6A) linked to an aromatic ring (phenol or thiophene) with

one of the following electrons withdrawing group replacements: chlorine, bromine, nitrile or 3,5

dimethylpiperidine-1-sulfonyl as indicated in Figure 6B.

Figure 5: F0317-0202 broadly active analogs: Chemical structure of the top tested analogs and the % of relative motility to the DMSO control (R.M.) of 30μM relative to DMSO the *in vitro* activity, against *A. ceylanicum* hookworms (HW) and *T. muris* whipworms (WW).

Figure 6: F0317-0202 Structure-activity relationship (SAR) model. (A) The bioactive group in F0317-0202 is 1,3-benzothiazol-2-ylidene-3-oxopentanenitrile (blue). (B) Structure-activity relationship (SAR) model generated for F0317-0202.

Discussion

Human GINs are among the most prevalent infections of the NTDs. Because understanding of

these parasites' basic biology is still limited, whole organism screening (*i.e.*, phenotypic

screening vs. target-based screening) remains the most effective approach in finding effective

anthelmintics. We previously validated a novel screening pipeline using a set of 1280 FDA-

approved compounds from a single library [31]. This screening pipeline utilizes two

phylogenetically diverse nematode parasites, *A. ceylanicum* hookworms and *T. muris*

whipworms, located at opposite ends of the phylum Nematoda phylogenetic tree and that are

either a human parasite (*A. ceylanicum*) or closely related to a human parasite (*T. muris* vs *T.*

trichiura). By selecting these diverse nematodes, we aim to identify compounds with broad

anthelmintic activity. Here, using much larger and more diverse sets of compound libraries, we

used our pipeline to screen more than 30,000 compounds, which is unprecedented for targeting

the GINs of humans.

The life chemical diversity set library comprises 15,360 compounds representing 1,717 chemical

scaffolds from novel and newly synthesized small molecules. Using our screening pipeline, we

uncovered seven molecules with broad activity against parasitic nematodes, giving a hit rate of

0.045%. Among these seven, four are polymethine cyanine dyes: F1059-0041, F9995-0076,

F0900-4647, and dithiazanine iodide (F9995-0233). Dithiazanine iodide is known to be active

against hookworms, threadworms, roundworms, pinworms, and whipworms [84-88], providing

additional validation for our pipeline. Of the other three non-polymethine cyanine dyes, F1345-

0623 has been identified as a potential inhibitor (IC_{50} in the sub-micromolar range) of the

deubiquitinase USP7 [89]; homologous proteins have been found to play essential roles in *C.*

elegans (math-33)[90] and *Schistosoma mansoni* (SmUSP-7) [91].

Another compound, F0317-0202, has an excellent drug-like structure, and was selected for

further studies. F0317-0202 includes two aromatic ring systems (thiophene and benzothiazole),

listed among the most frequently used ring systems from small molecules in the FDA Orange

book [92]. The compound also has two functional groups (OH and CN), listed among the most

frequent functional groups occurring in bioactive molecules [93]. The compound's

physicochemical property agrees with Lipinski's rule of five for drug-likeness, with a molecular

weight of 318.8g/mol (less than 500), one hydrogen bond donor (less than five), five hydrogen

bond acceptor (less than 10), and lipophilicity score (Log P) of 4.3 (less than five)[94]. Out of 28

tested analogs of F0317-0202, 12 compounds (42%) showed potent activity against whipworm

and hookworm parasites, with four compounds showing near-complete inhibition in motility in

the two parasites. We consider these five novel compounds (F0317-0202 and four of its analogs,

F0317-0013, F0317-0161, F0328-0210, and F0317-0009) as well-positioned for further *in vitro*

characterization, lead optimization, and *in vivo* validation.

In contrast to screening diversity sets or random compound collections, we also screened

repurposing libraries, which contain compounds already approved or associated with clinical

trials and which can serve as a source for novel therapeutics for new indications [95]. The Broad

Institute repurposing library, which we used in our screen, contains 6,743 compounds from

molecules with high structural diversity and with 2006 known protein targets, including but not

limited to the G protein-coupled receptors (GPCR), acetylcholine receptors, protein kinases,

oxygenases, voltage-gated channels, and nuclear hormone receptors[96]. In addition, screening

compounds with known targets can be highly beneficial, especially against NTDs where a deep

understanding of their biology is still lacking. Compounds with known modes of action offer a

significant advantage in drug development because they can act as probes for essential targets,

assuming the targets are known. For this reason, we also screened the ICCB-Longwood

mechanism of action library containing 1,245 high-quality compounds with known targets.

The total number of compounds tested from both libraries was 7,988, uncovering 45 compounds

with putative broad activity against parasitic nematodes, giving a hit rate of 0.56%, 12.4 times

that obtained from the generic diversity set library. These results are the first to thoroughly

compare a generic dissimilarity-based library type with repurposing/known mode of action

compounds against parasitic nematodes using the same screening conditions and pipeline. Our

results indicate that repurposing/known mode of action libraries are significantly more (>10X;

P< 0.00001 Fisher's exact test) likely to uncover active compounds.

The plethora of the chemical universe is expected to contain 10^{12} to 10^{180} drug-like compounds[97],

which are impossible to screen against any single organism, not to mention GINs. Therefore,

techniques and strategies to design chemical libraries covering essential pharmacological targets

are highly recommended. Among these approaches is the compilation of a target-focused library

of compounds, which can be fitted by molecular docking to bind and inhibit the function of a

specific target or protein family[98, 99]. This strategy has been previously validated using a kinase-

targeted library of 1440 compounds tailored to fit in a total of 41 kinases from five different

families, showing a 6.7-fold increase in the overall hits compared to the generic collection of

molecules[100]. Target-focused screening improves efficiency by excluding compounds unlikely to

modulate known target proteins of interest and is associated with increased potency and

specificity[101].

We found that target-based libraries could serve as good sources for anthelmintic discovery. We

screened nine target-focused libraries, including one for neuronal targets, one for GPCRs, and

seven libraries targeting protein kinases. Neuronal signaling and the neuromuscular system are

essential for nematodes' survival, including feeding, mating, egg laying, and locomotion; thus,

many anthelmintics drugs target the nematode nervous system[102]. Neuropeptide GPCRs have

been identified as potential and druggable targets and were found to play critical roles in the

helminth neuromuscular function[103]. Kinases are the second most targeted protein class after

GPCRs, with over 200 kinase inhibitors having reached the clinical phases and 52 FDA-

approved drugs targeting protein kinases[36, 104].

Together, these nine libraries contain 2,708 compounds, and within these, we identified 12

compounds with putative broad activity, giving a hit rate of 0.44%, statistically the same as that

obtained from the combined repurposing and mode of action libraries (P=0.54, Fisher's exact

test). It is worth noting that three of the nine target-based libraries resulted in no hits with activity

against hookworms and whipworms: the ChemBridge Focused kinase, GPCR, and the LINCS 2

kinase inhibitors, which could be explained by the lack of specificity to the parasitic kinases and

GPCRs, as these libraries were fitted to selectively binding specific human targets.

Historically, natural products have been the primary source of remedies for human ailments, and

purified natural molecules or direct derivatives have contributed to at least 34% of all drugs

approved by the FDA between 1981 and 2010[105]. Using our novel pipeline, we screened the

TimTec library of 4,240 purified compounds from natural products or direct derivatives and

identified 14 compounds with *A. ceylanicum* larval activity. However, none of these larvicidal

compounds carried activity against adult parasites. This result reflects may reflect biases of this

particular library.

In summary, we have developed a novel high throughput screening pipeline to identify

anthelmintic compounds for GIN parasites and screened 13 different libraries with a total of

30,238 compounds that led to the discovery of 55 compounds with potent activity against two

species of evolutionary distant nematode parasites. Repurposing, mode of action, and target-

based libraries were more profitable than the diversity set for anthelmintic screening. For these

55 compounds, we reported the known modes of action and related targets in other organisms,

and potential counterparts in the nematode, if available. As an example, we highlighted four

nematode targets as potential candidates for target-based drug screening. We also identified

thiazolyl oxopropanenitrile as a novel chemical scaffold with broader anthelmintic activity

identified from the diversity set library and established the SAR models for potent activity

against hookworms and whipworms. Our findings open the doors for multiple follow-up

approaches, including target identification, target-based screening, and ligand or structure-based

pharmacophore optimization for developing safe and parasite-selective molecules.

Materials and Methods:

Ethics statement:

All experiments involving animals were conducted per the recommendation of the National

Institute of Health (USA) Guide for Care and Use of Laboratory Animals and the Animal

Welfare Act. The animal protocols used in this study (PROTO202000044 and

PROTO202000071) were approved by the Institutional Animal Care and Use Committee

(IACUC) of UMASS Chan Medical School.

Parasite maintenance in laboratory animals: Following standard protocols, *A. ceylanicum*

hookworm parasites were maintained in golden Syrian hamsters[106], and *T. muris* whipworm

parasites were maintained in STAT6-/- mice [107].

Primary screening using Larval Developmental Assay:

Feces from *A. ceylanicum*-infected hamsters were collected ~18 days post-infection and about 20

grams of feces were processed for egg isolation and purification following salt/sugar flotation

protocol[108, 109]. Purified and surface sterilized eggs were incubated in S-medium[110] at 28 °C,

without any food source for about 20 hours, allowing eggs to hatch into synchronized first-stage

larvae [109]. Newly hatched L1 were washed three times in a S-medium. About 30-35 L1 larvae

were dispensed into each well of 96-well assay plates containing 90µL of S-media supplemented

with 100U/ml of penicillin, 100µg/ml streptomycin, and 2.5µg/ml amphotericin, *Escherichia coli*

OP50 with final OD600 of 0.3 and 0.15µl Dimethylsulfoxide (DMSO). Library plates were

diluted in S-media to 100µM and 1%DMSO. Using a multichannel pipette, 10µL of the library

plates were transferred into each well of the assay plates, giving a final concentration of 10µM of

test drug and 0.25% DMSO. Plates were prepared in duplicates. Test plates were incubated at 28

°C for seven days and then scored for development under a dissecting microscope. Test drugs

were considered hit if > 90% of L1 larvae failed to develop in duplicate wells. Each assay plate

includes four wells of 10µM ivermectin as positive control and four wells of 0.25% DMSO as

negative control.

Secondary screening using young adults of hookworm parasites:

We next screened hits from the primary screening (larval development) against the adult stages

of hookworm parasites. *A. ceylanicum* hookworm young adults were harvested 10-11 days post-

inoculation from the small intestines of infected hamsters. Harvested worms were washed and

incubated in prewarmed hookworm culture medium (RPMI1640 of pH 7.2 supplemented with

10% fetal bovine serum, 100U penicillin, 100µg/ml streptomycin, 2.5µg/ml amphotericin). Two

adults of *A. ceylanicum* hookworm were manually picked into each well of the 96-well assay

plates, containing 70µL of hookworm medium supplemented with 0.7µl DMSO. Using a

multichannel pipette, 30µL from the library plates diluted in hookworm medium to a final

concentration of 100µM and 1% DMSO were dispensed into screening wells, giving a final

concentration in assay plates of 30µM of test compounds and 1% DMSO with duplicate

wells/drug. Assay plates were incubated for 48 hours at 37 °C and 5% CO_2. Drug activity was

determined using the standard motility index [31, 107]. A motility index of 3 was given to vigorous

healthy adults, 2 for motile but slow adults, 1 for immotile adults that moved after stimulation by

touching, and 0 for non-motile adults even after touching. The drug was considered a hit if the

motility index of at least three of the four *A. ceylanicum* hookworm adults were scored ≤1 and/or

if all the parasites in the well had significant morphological defects (*e.g.*, shrunken cuticles,

darkened intestines (Figure 3). Each assay plate included four positive control wells (30μM

ivermectin) and four negative control wells (1% DMSO).

Tertiary screening using adult whipworm parasites:

We next screened *A. ceylanicum* hookworm adult hits against *T. muris* whipworm parasites. *T.*

muris adult whipworms were harvested from the cecum and large intestines of infected STAT6 -

/- mice. Single worms were manually picked into each well of a 48-well plate, six wells/ test

drug, and a final volume of 291μL of whipworm culture medium with 2.1μl DMSO (like

hookworm culture medium but supplemented with 5% fetal bovine serum). 9μL from library

plates diluted in whipworm medium to a final concentration of 1mM and 10% DMSO were

dispensed into each well of the screening plates, giving a final concentration of 30μM test drug

and 1% DMSO. Assay plates were incubated at 37 °C, and 5% CO_2. *T. muris* whipworm

motility was measured using an in-house assembled worminator and worm assay software[111] for

3 minutes after 24 and 48 hours. Drug activity was determined using the percent of relative

motility compared to DMSO control. Drugs were considered active if the 24- and/or 48-hour

motility was reduced by at least 50% compared to negative control (Figure 4).

Chemical Libraries Screened

Life Chemical 15K diversity set:

Life Chemical pre-plated diversity set library was obtained from the Small Molecules Screening

Facility (SMSF), UMASS Chan Medical School. The library contains 15,360 compounds

representing 1,717 chemical scaffolds with an average Tanimoto similarity index of 0.409.

Library compounds were plated in 96-well plates with 1ul of 10mM stock concentration. Hits

from the primary and secondary screening were cherrypicked into 96-well plates for *A.*

ceylanicum hookworms or *T. muris* whipworms screening.

Broad Institute Repurposing Hub:

The drug repurposing hub includes 6,743 compounds which were from a curated and annotated

collection of preclinical drugs in phase 1 (REPO 1) or phases 2-3 (REPO 2) and FDA-approved

and launched drugs (REPO 3). REPO drugs cover a wide range of disease areas, including but

not limited to infectious diseases, allergies, cardiology, dermatology, endocrinology,

metabolism, neurology, and gastroenterology. The library covers 1,133 different modes of action

and 2,183 different drug targets[96].

ICCB Screening libraries:

The Institute of Chemistry and Cell Biology-Longwood (ICCB-L) Screening Program at Harvard

Medical School offers a wide range of well-annotated drug screening libraries. We have screened

multiple ICCB-L libraries, including mechanism of action, neuronal targets, GPCR-based

inhibitors, and six libraries of kinase inhibitors.

ICCB-L MOA: the mechanism of the action library is a collection of high-quality annotated

compounds with biological activity against known targets with target-compound pairs based on

high target specificity and potency. The library contains 1,245 unique compounds, each plated in

4 doses. *ICCB-L Selleck Neuronal Targets:* the neuronal targets library was initially composed of

Selleck chemicals from FDA-approved compound sets that were determined to have biological

activity in neurologic research and neurological assays. *ICCB-L ChemBridge focused GPCR:*

This focused library of 250 GPCR inhibitors was selected from 13,000 GPCR inhibitors

assembled by ChemBridge. The library has 11 different scaffolds, each containing 19-24

compounds.

Kinase inhibitor libraries: in total, we screened focused sets of 1428 kinase inhibitors from 7

different kinase libraries, some of which were screened at multiple doses.

ICCB-L SYNthesis kinase inhibitor: this library was assembled by SYNthesis Med Chem of 96

compounds known to inhibit different kinases. *ICCB-L EMD Kinase inhibitors:* this library was

built by EMD of 244 protein kinase inhibitors. *ICCB-L ChemBridge Focused Kinase inhibitors:*

This library of 250 compounds was created and selected from the ChemBridge kinase-biased

library of 6000 compounds. These compounds were selected based on the pharmacophore

properties of known kinase inhibitors or from compounds predicted to interact with the ATP

binding site of kinases. In total, these compounds represent 16 different scaffolds that cover 34

kinase targets. *ICCB-L LINCS 1,2 and 4 Kinase inhibitors:* these three libraries were assembled

by LINCS together, including 345 compounds with known or predicted kinase inhibitors.

SMSF kinase inhibitor library: This library includes 429 kinase inhibitors with activity and

safety confirmed by preclinical research and clinical trials.

Natural Products: we screened the TimTec library of 4240 purified compounds from natural

products or derivatives. The TimTec library was obtained from the SMSF and includes two types

of compounds: *Natural Products Library (NPL):* The NPL library includes 1200 purified natural

compounds from plant, bacteria, fungi, and animal sources. *Natural Derivatives Library (NDL):*

The NDL library includes 3040 semi-natural compounds derived from natural molecules or

synthetic compounds with natural-like structures.

Structure-Activity Relationship (SAR) of anthelmintic hits: using vendors' databases and

online tools, analogs of F0317-0202 were identified and purchased from Life Chemicals. Each

analog has slight structural modification compared to the parent molecule. These analogs were

tested against the adult stages of *T. muris* whipworm and *A. ceylanicum* hookworm parasites at

30µM and scored using the Worminator at 24 and 48 hours. Hence, *A. ceylanicum* hookworm

parasites used in testing F0317-0202 analogs for SAR model were mature adults harvested 18

days post-infection.

Authors contributions:

M.A.E and R.V.A., conceived, designed, supervised, and interpreted the experiments. M.A.E and

R.V.A. wrote the manuscript. M.A.E, Y.M.K., and E.G. performed the *in vitro* screening. M.A.E

and S.S. performed data mining and analyzed SAR results. P.R.T. and L.B. contributed libraries,

technical advice, and inputs on the manuscript. M.A.E. designed the graphic abstract. R.V.A.

secured funding.

Supporting information:

A Additional details for the potential nematode targets (PDF)

B Additional experimental details for F0317-0202 tested analogs, chemical structures, and

motility inhibition (PDF)

Acknowledgments:

This work was financially supported by the National Institute of Health (USA) – National

Institute of Allergy and Infectious Diseases grants R01 AI150866 and R01 AI056189 both to

R.V.A.

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