Single neuron contributions to the auditory brainstem EEG

Paula T. Kuokkanen[1], Ira Kraemer[2], Christine Koeppl[3],
Catherine E. Carr[2], Richard Kempter[1,4,5]

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Affiliations:

[1] Institute for Theoretical Biology, Humboldt-Universität zu Berlin, 10115 Berlin, Germany

[2] Department of Biology, University of Maryland College Park, College Park, MD 20742

[3] Department of Neuroscience, School of Medicine and Health Sciences, Research Center for Neurosensory Sciences and Cluster of Excellence "Hearing4all" Carl von Ossietzky University, 26129 Oldenburg, Germany

[4] Bernstein Center for Computational Neuroscience Berlin, 10115 Berlin, Germany

[5] Einstein Center for Neurosciences Berlin, 10117 Berlin, Germany

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Abstract

The auditory brainstem response (ABR) is an acoustically evoked EEG potential that is an important diagnostic tool for hearing loss, especially in newborns. The ABR originates from the response sequence of auditory brainstem nuclei, and a click-evoked ABR typically shows three positive peaks ('waves') within the first six milliseconds. However, an assignment of the waves of the ABR to specific sources is difficult, and a quantification of contributions to the ABR waves is not available. Here, we exploit the large size and physical separation of the barn owl first-order cochlear nucleus magnocellularis (NM) to estimate single-cell contributions to the ABR. We simultaneously recorded NM neurons' spikes and the EEG, and found that $\gtrsim 5,000$ spontaneous single-cell spikes are necessary to isolate a significant spike-triggered average response at the EEG electrode. An average single-neuron contribution to the ABR was predicted by convolving the spike-triggered average with the cell's peri-stimulus time histogram. Amplitudes of predicted contributions of single NM cells typically reached 32.9 ± 1.1 nV (mean $\pm$ SE, range: $2.5 - 162.7$ nV), or $0.07 \pm 0.02\%$ (median $\pm$ SE range: $0.01 - 4.0\%$) of the ABR amplitude. The time of the predicted peak coincided best with the peak of the ABR wave II, and this coincidence was independent of the click sound level. Our results suggest that wave II of the ABR is shaped by a small fraction of NM units.

Significance statement

The auditory brainstem response (ABR) is a scalp potential used for the diagnosis of hearing loss, both clinically and in research. We investigated the contribution of single action potentials from auditory brainstem neurons to the ABR and provide direct evidence that action potentials recorded in a first order auditory nucleus, and their EEG contribution, coincide with wave II of the ABR. The study also shows that the contribution of single cells varies strongly across the population.

Introduction

ABRs typically exhibit 3 early peaks, generated in the brainstem by local current sources arising from the auditory nerve as well as first- and second-order auditory nuclei in succession. These local current sources give rise to extracellular field potentials (EFPs) whose origins are not well understood, despite their clinical relevance. Studies of cortical pyramidal cells have led to the widespread assumption that EFPs have their origins mainly in synaptic dipoles (Eccles, 1951; Klee et al., 1965; Creutzfeldt et al., 1966a,b; Nunez and Srinivasan, 2006; da Silva, 2013; Ilmoniemi and Sarvas, 2019). However, other neuronal sources can also contribute, because the source of EFPs depends on the morphology of potential neuronal sources and synchrony of their activity (Gold et al., 2006; Kuokkanen et al., 2010; Lindén et al., 2011; McColgan et al., 2017; Rimehaug et al., 2023). Identifying the sources of brainstem EFPs, and their contributions to the ABR, should both inform models of the ABR and provide further insights into different types of hearing loss. We show here the contributions of single neurons to the ABR.

ABRs were detected first in the 1950s (Dawson, 1954; Geisler et al., 1958), and have been widely used in the clinic for decades (Geisler, 1960; Clark et al., 1961). Furthermore, ABRs are used in common basic hearing tests in animal research (e.g., Zheng et al., 1999; Akil et al., 2016; Kim et al., 2022). Models of the ABR (e.g. Melcher and Kiang, 1996; Ungan et al., 1997; Goksoy et al., 2005; Riedel and Kollmeier, 2006; Colburn et al., 2008; Verhulst et al., 2015, 2018) have helped to clarify ideas about its sources and its binaural components, but have remained difficult to validate experimentally. Most ABR models incorporate the *unitary response* (UR) (Melcher and Kiang, 1996; Dau, 2003; Schaette and McAlpine, 2011; Rønne et al., 2012; Verhulst et al., 2015, 2018), which is the expected average spike-triggered response related to the activation of a single neuronal source at the EEG electrode. The UR typically also includes the full structurally correlated cascade of activations in other brainstem nuclei. When convolved with the peri-stimulus time histogram of that (initial) source, the UR predicts the contribution of that source (and related later sources) to the ABR response. There are, however, many possible UR-solutions to a given ABR waveform, where each solution imposes a set of boundary conditions related to the source of the UR in the cell morphology. Furthermore, URs have been difficult to measure, leading to methods to estimate them indirectly for the whole brainstem by deconvolution from

the ABR and models of firing rates (e.g. Elberling, 1978; Dau, 2003; Rønne et al., 2012). The deconvolution method is adequate for modeling expected ABR responses from various stimuli, but lacks precision about the sources whose activity might be correlated with changes in this UR.

Here, we take a direct approach to measuring single-cell URs from the barn owl's first-order auditory brainstem nucleus magnocellularis (NM). We took advantage of the large size and physical separation of the first-order auditory nuclei in birds (Kubke et al., 1999, 2004). Furthermore, NM units have high spontaneous firing rate (Köppl, 1997a), enabling averaging over tens of thousands of spikes to overcome the noise at the EEG electrode. Measuring the UR directly for NM reveals that URs are highly variable, with amplitudes that can reach several percent of the ABR amplitude, and peaks that coincide with wave II.

Materials and Methods

All the data analysis was done with Matlab 9.0 (version 2016a, MathWorks, Natick, MA). All the data was re-sampled to 50 000 Hz before analysis.

Experimental paradigm

The experiments were conducted in the Department of Biology of the University Maryland. Thirteen barn owls (Tyto furcata) of both sexes were used to collect the data at 27 EEG recording locations and for 151 intracranial recording locations. Many animals were studied in two or three separate physiology experiments, spaced approximately a week apart. Procedures conformed to NIH Guidelines for Animal Research and were approved by the Animal Care and Use Committee of the University of Maryland. Anaesthesia was induced by intramuscular injections of 16 mg/kg ketamine hydrochloride and 3 mg/kg xylazine. Similar supplementary doses were administered to maintain a suitable plane of anaesthesia. Body temperature was maintained at 39°C by a feedback-controlled heating blanket. More details can be found in Carr et al. (2015).

Acoustic stimuli. Recordings were made in a sound-attenuating chamber (IAC, New York). Acoustic stimuli were digitally generated by custom-made software ("Xdphys" written in Dr. M. Konishi's lab at Caltech) driving a signal-processing board (DSP2 (Tucker-Davis

Technologies (TDT), Gainesville, FL). Acoustic signals were calibrated individually at the start of each experiment, using built-in miniature microphones (EM3068; Knowles, Itasca, IL) inserted into the owl's left and right ear canals, respectively. **Tone-pip stimuli** had a duration of 100 ms, including 5 ms ramps. The stimulus level was $40 - 50$ dB SPL. The range of stimulus frequencies was $1 - 9$ kHz, with a typical step size $200 - 500$ Hz, and $3 - 20$ repetitions for each stimulus used. **Clicks** were presented at attenuation levels $55 - 0$ dB, calibrated to correspond to stimulus levels $10 - 65$ dB SPL, respectively ($128 - 3300$ repetitions at each single-unit recording location). Condensation clicks had a rectangular form and a duration of two samples (equivalent to 41.6 μs). **Spontaneous activity** was recorded for about $15 - 60$ minutes for each unit.

Intracranial methods and recording protocol. Tungsten electrodes with impedances $2 - 20$ MΩ were used (F.C. Haer, Bowdoin, ME). A grounded silver chloride pellet, placed under the animal's skin around the incision, served as the reference electrode (WPI, Sarasota, FL). Electrode signals were amplified and band-pass filtered ($10 - 10,000$ Hz) by a custom-built headstage and amplifier. Amplified electrode signals were passed to a threshold discriminator (SD1, TDT) and an analogue-to-digital converter (DD1, TDT) connected to a workstation via an optical interface (OI, TDT). In all experiments, voltage responses were recorded with a sampling frequency of $48,077$ Hz, and saved for off-line analysis.

For an intracranial recording, an electrode was advanced into the brainstem guided by stereotaxic coordinates, and units were characterized based on recorded extracellular spikes. Units were recorded on both sides of the brain. At each recording site, frequency responses were measured for tonal stimuli to each ear, and ITD tuning was measured with binaural tonal stimuli. Recordings confirmed that responses within nucleus magnocellularis (NM) were monaural, as expected. Single unit frequency response curves were recorded for the ipsilateral stimulus: for each recording location, an appropriate range of stimulus frequencies (within $1 - 9$ kHz) was selected to record iso-level frequency response curves. Between single-unit recordings, the electrode was moved typically in steps of 100 μm while searching for the next unit. For some units there were additional control recordings in which the recording from the same unit was continued while moving the intracranial electrode with steps of size $10 - 20$ μm.

EEG methods. An EEG signal was recorded simultaneously with all the intracranial recordings. Recordings were made using two platinum subdermal needle electrodes (Grass F-E2; West Warwick, RI) on the scalp. EEG signals were amplified using a WPI DAM-50 extracellular preamplifier, $0.1 - 10,000$ Hz (World Precision Instruments, Sarasota, FL). The EEG signal was further amplified (100x) using a custom built amplifier, and digitized (DDI, TDT). The voltage responses were recorded with a sampling frequency of $48,077$ Hz and saved for off-line analysis.

The active EEG electrode was always positioned in the dorsal midline, adjacent to the craniotomy, and the EEG reference electrode was positioned behind the ipsilateral ear. EEG electrodes could be slightly repositioned during the recording session to improve the signal.

Intracranial recordings: Data analysis

In addition to custom Matlab scripts, we used the XdPhys script from M. Konishi's lab and the supramagnetic wavelet-based 'Wave-clus' method for spike detection and clustering (Quian Quiroga et al., 2004), as provided as a Matlab script at https://github.com/csn-le/wave_clus.

Spike detection and clustering

We recorded from 151 intracranial locations within the NM cell body region (Fig. 1A) on which the spike detection and clustering was performed. Spikes were detected off-line, and all the data from a single intracranial recording location were combined. After spike detection and clustering, the spikes were ordered by their respective stimulus conditions (tone, click, spontaneous).

Spike detection. For spike detection, the default parameters of the *Wave-clus* method (Quian Quiroga et al., 2004) were modified as follows: The minimum threshold of spike detection (parameter 'std_min') was set manually for each unit depending on its spike size and noise level, and varied between 3.0 and 8.0 standard deviations (SD). Also the polarity of the spikes ('detection') was set manually for each unit upon visual inspection, because our set-up allowed spikes having either polarity. For the spike detection, the band-pass filter setting was $900-6,000$ Hz ('detect_fmin' and 'detect_fmax', respectively). The window length for spike shape was 1 ms before the spike peak and 1.5 ms thereafter, corresponding to 'w_pre' = 50 and 'w_pre' = 75

samples. The refractory time for the detection was set to 0 ms ('ref_ms'), firstly because instantaneous firing rates in NM can be as high as $1,500$ spikes/s (Carr and Boudreau, 1993), and secondly because then we could detect units with spike-doublets. The ISI distribution of each unit was later scrutinized to exclude multi-units and doublet-units (see section 'Prepotentials and doublets').

Spike clustering. The spikes were clustered with the wavelet decomposition method within *Wave-clus* with 5 'scales' in the wavelet decomposition and minimum of 10 inputs per cluster ('min_input'). The radius of the clustering ('template_sdnum') was set to 4.5 SD, and the number of nearest neighbors ('template_k') was set to 10. Otherwise, both for detection and for clustering, the default parameters were used. After visual inspection of the resulting spike shape clusters, the clusters were merged if necessary (typically $2-3$ clusters with an identical spike shape but variability during the onset or offset within the spike-window). Recording sites containing several units (with variable spike waveforms) were discarded from further analysis. In some recordings there was a small number of outliers (detected peaks not fitting any spike cluster) with always $N_{out} < 0.75\%$ of number of spikes in the main cluster(s); typically $N_{out} = 0-50$. These outliers were excluded from the analysis.

Spike separation to stimulus conditions. **Tone-driven spikes**, obtained in response to 100 ms tones and with ≥ 15 dB SPL stimulus level, were included in the analysis when they occurred within $15-95$ ms of the stimulus onset, thus excluding possible onset and offset effects. The **click responses** of the single-unit activity (*peri-stimulus time histogram, PSTH*) were calculated within $0-10$ ms of the click stimulus onset. We considered **spontaneous spikes** to be any activity in trials in which there was no stimulus presented. Additionally, to collect as many spontaneous spikes as possible, we considered spikes to be spontaneous in two scenarios: Spikes occurring in stimulated trials (1) but later than 50 ms after the end of tonal or click stimuli, and (2) during stimuli that did not evoke an elevated sustained response, i.e. low-amplitude tones < 15 dB SPL at frequencies far off from the best frequency, excluding the first 20 ms after the stimulus onset.

Exclusion of recordings. We excluded units using three criteria: (1) Units with too few spontaneous spikes recorded (< 5000) because in this case we could not derive a significant STA EEG. (2) Units for which the single-unit isolation was poor, i.e., the spike waveform SNR was < 8.6 dB. The SNR of the spike waveform was defined by the squared ratio of the spike peak amplitude and the standard deviation of the baseline. (3) Units for which the single-unit isolation broke down at the onset of the click-stimulus as confirmed by a visual inspection (see also 'Click-evoked magnocellular activity' below). After applying these exclusion criteria on the 151 units recorded within NM, 53 single units remained and were further analyzed.

Classification of magnocellular and auditory nerve units

Single units recorded within NM were classified to be either 'AN fibers' or 'NM cell bodies /axons'; classification was based on best frequency (BF) and spontaneous firing rate, which were defined as follows:

BF: Iso-level response curves of the numbers of spikes defined the BF at a recording site as follows (Kuokkanen et al., 2010): a line at half height of a tuning curve was derived from its peak rate and the spontaneous rate. The midpoint of the line at half height yielded the BF. The best frequencies ranged from 1.25 to 7.75 kHz with mean $\pm$ SD: 5.60 ± 1.60 kHz. The tuning was calculated for the sustained activity in the window of $15 - 95$ ms after tone onset, across all repetitions of the stimulus.

Spontaneous rate: Spontaneous rate was defined as the reciprocal of the mean spontaneous inter-spike-interval.

Auditory nerve and NM categories were based on the spontaneous firing rates and the characteristic frequencies (CFs) reported in Köppl (1997a), which provides the fits of CF vs spontaneous rate, S, for AN (S_{AN} [spikes/s] $= 123.8 \cdot \exp(-0.129 \cdot \mathrm{CF}$ [kHz]$)$) and NM units (S_{NM} [spikes/s] $= 255.1 \cdot \exp(-0.0634 \cdot \mathrm{CF}$ [kHz]$)$). We used the separating line of $f \cdot S_{\mathrm{NM}} + (1 - f) \cdot S_{\mathrm{AN}}$ with $f = 0.3$, as this was the best line of separation between the AN / NM classes in Köppl (1997a).

Prepotentials and doublets

Recordings of magnocellular units can exhibit both prepotentials (Zhang and Trussell, 1994) and spike-doublets (Carr and Boudreau, 1993; Kuokkanen et al., 2018). For our analysis, a recording with a prepotential was interpreted as the intracranial electrode being located in the vicinity of an NM cell body and at least one large synapse from AN to this NM cell. In recordings from NM units, also spike-doublets can occur with very short inter-spike-intervals (ISIs $0.22 - 0.5$ ms, Carr and Boudreau (1993), their Fig. 2). However, units with doublets pose a challenge for the estimation of the STA EEG.

Upon visual inspection, 8 NM units were determined to include a large proportion of doublets and were excluded from further analysis.

Click-evoked magnocellular activity

The click-evoked responses of the single units (*peri-stimulus time histogram, PSTH*) were calculated within $0 - 10$ ms after click onset.

The onset delay (or 'click-response latency') of the PSTH characterized the click-evoked responses in NM. We calculated the click-response latency using the same criterion as Köppl (1997a) — the first PSTH bin (with a 50 μs bin size) after the stimulus presentation exceeding the largest spontaneous PSTH bin and being followed by a bin also fulfilling this criterion was defined as the click-response latency.

EEG electrode recordings: Data analysis

Auditory brainstem response (ABR) recordings

We recorded click-elicited responses at the EEG electrode, i.e. the ABR, within either $0 - 10$ ms or $0 - 15$ ms after click onset. ABR waveforms were averaged across stimulus repetitions ($127 - 500$ trials) resulting in a 'trial-averaged ABR' for unchanged recording and stimulus conditions.

ABRs were quantified by the SNR, which was defined as the squared ratio of the peak amplitude of the trial-averaged ABR and the mean RMS of the baseline across ABR trials (5 ms window prior the click onset). The SNRs across the ABR waveforms ranged from -50 dB to $+18$ dB, with

median of +5 dB. After visual inspection, we excluded ABR waveforms with the SNR < -13 dB ($< 7\%$ of ABR waveforms) as well as ABR waveforms not showing three peaks in the waveform ($< 2\%$ of the waveforms). The excluded responses were typically, but not always, recorded with a low stimulus level.

ABR wave quantification. We quantified the timing and amplitude of 3 positive waves in each waveform objectively as follows: We band-pass filtered ($550 - 4,000$ Hz, Chebyshev type II filter of the order 8) the trial-averaged ABR response, and zero-mean-centered the waveform, to remove the low- and high-frequency noise present in some ABRs. We then used the Matlab algorithm FINDPEAKS.M to find all peaks in this filtered ABR within $0 - 10$ ms after stimulus onset. The algorithm returns the locations and heights of the peaks, and also the Matlab variables 'width' and 'prominence' (width at the half-maximum with respect to the baseline of the individual peak, and the height of the peak with respect to the same baseline). To identify the possible ABR peaks, we included all the maxima exceeding the threshold of 0.4 SD of the trial-averaged preamplifier-filtered ABR response ($0 - 10$ ms after stimulus onset). The threshold was chosen such that at least 2 ABR peaks were detected for all the waveforms. Of all the peaks crossing the threshold, we excluded the peaks with a 'width' narrower than 0.1 ms because typical ABR waves are much wider. If more than three peaks crossed the threshold, we used the three peaks with the highest 'prominence'. If only two peaks were initially detected, we assumed that these would correspond to the peaks of the waves I and II because they typically were the largest peaks of the ABR, whereas the peak of the wave III was often small or even negative with respect to the baseline; thus, we included the largest peak within the period of 0.4 ms after the second found peak's timing (starting point) to 3.0 ms after the first peak's timing (end point). The starting point was selected to ensure that occasional small, local maxima within the wave II were not included, and the end point was selected because 3 ms was the typical duration of the ABR waveform from the wave I peak to the large negativity after wave III. Finally to ensure not to introduce jitter to the peak times because of the filtering, we applied these peak locations to the original preamplifier-filtered, trial-averaged ABR by finding the related maxima, allowing for a change of peak time of at most ± 3 data points. In the end, this algorithm allowed us to quantify three peaks for all the selected ABR recordings.

The peak amplitudes of waves I to III were calculated from the preamplifier-filtered average traces, in comparison to the trial- and time-averaged baseline in the time window from the beginning of the recording ($5 - 10$ ms prior to click onset) to the time point 1 ms prior to wave I peak.

ABR averaging. The trial-averaged ABRs, as just defined, were obtained for different EEG electrode positions, intracranial recording sites, and click levels. After the ABR wave quantification, we averaged the detected peak amplitudes and their timing across trial-averaged ABRs for constant click levels as follows:

1) For the ABR waveform analysis tied to NM single units, all the trial-averaged ABRs recorded simultaneously with the respective single unit responses were used ($1 - 11$ trial-averaged ABRs with median of 1, in total $128 - 3300$ trials, median: 300). For each NM unit the EEG electrode position was kept unchanged.

2) For the ABR waveform analysis unrelated to NM single units, we averaged peak amplitudes and their timings also across intracranial recording sites ($1 - 14$ trial-averaged ABRs with median of 1, in total $128 - 4200$ trials, median 999). In some days the EEG electrode was re-positioned during the experiment. Here, we restricted the ABR waveform analysis to the EEG electrode position with the highest signal-to-noise ratio (SNR), resulting in $N = 24$ EEG electrode positions.

ABR inter-peak-intervals. The inter-peak-intervals of peaks $1 - 2$, $1 - 3$, and $2 - 3$ were calculated based on the delays of peak timings in trial-averaged ABR waveforms and thereafter averaged as described above.

Spike-triggered average EEG

EEG traces recorded in the absence of acoustic stimuli were band-pass filtered ($800 - 3000$ Hz, Chebyshev type II filter of the order 6). Compared to the ABR recordings, a narrower filter was chosen to further reduce noise. The spike-triggered average EEG (STA EEG) was calculated for each NM single unit separately. The STA EEG was derived from 8-ms time windows ($N_t = 402$ data points) of the EEG recording centered at spike times of single units. We used only spontaneous spikes for the STA EEG.

We define the STA EEG mathematically as the average signal at the EEG-electrode, $C(\tau)$, around isolated spikes of a neuron j at times t_i^j where $i = 1, 2, ..., n$ is the spike number and $r(t)$ is the simultaneously recorded EEG:

$$C_j(\tau) = \frac{1}{n} \sum_{i=1}^{n} r(\tau - t_i^j). \tag{1}$$

We excluded from further analysis units by two criteria as follows: 1) To ensure that the EEG signal was high enough for the calculation of the STA EEG and for the calculation of the NM single-cell contribution to the ABR, the SNR of the ABR waveform at the highest click levels was required to be ≥ 1 dB, leading to exclusion of two NM single units. 2) To ensure that there was no cross-talk between the intracranial and EEG electrodes, we excluded the 7 units (out of 31) with an SNR > -18 dB of the STA EEG (range: -79 to $+6$ dB). In two experiments, we accidentally induced electrical cross-talk. This led to an SNR of the STA EEG of > -15 dB. In these units, the average spike waveform of the intracranial electrode and the waveform of the STA EEG were practically identical. After exclusion of 7 NM single units with putative crosstalk in the STA EEG, there were 24 NM single units included in further analysis.

STA EEG waveform significance. The significance of the STA EEG waveform was judged by using two bootstrapping methods. Firstly, the significance of the waveform was estimated with the SNR-based bootstrapping method by Parks et al. (2016). The number of samples was the number of spontaneous spikes, and the SNR distribution was based on 9999 bootstrap samples. The post-window width, for which the *signal* is calculated, was ± 0.25 ms around the spike time, corresponding to a post-window width of 0.5 ms. The pre-window width, from which the respective *noise* level is calculated, was set to 1.75 ms, (from 4 ms to 2.25 ms before the spike time). The 10-percentile lower bound threshold was set to 0 dB based on our SNR distributions. We chose a rather short post-window width to avoid being overly selective about the units left for the prediction of the ABR contributions (see below).

After establishing which of the STA-waveforms as such were significant, the time points (from -1.4 to 1.0 ms wrt. the spike time) at which each was significant were identified as by Teleńczuk

et al. (2015), with the 2-sample bootstrapping method with the confidence interval of 99% of the SE. There was no correction for multiple comparisons.

Control experiment

We conducted control experiments to confirm that electrical cross-talk between the EEG and intracranial electrodes in general did not affect our results. The idea behind these control experiments is as follows: when the intracranial electrode is moved the intracranial spike waveform changes. If there is cross-talk between the intracranial and the EEG electrodes, the STA EEG waveform should change as well. In contrast, if there is no cross-talk, the STA EEG should be independent of the intracranial spike waveform.

We thus moved in an exemplary control experiment the intracranial electrode in ten steps of $10 - 20$ μm over a total distance of 120 μm in the vicinity of an NM cell body. At the initial recording depth, the spike amplitude was 24.21 ± 0.02 μV (mean $\pm$ SE; the spike waveform and the related STA EEG from the initial recording depth is shown in the later Figure 4A). Moving the intracranial electrode deeper into the tissue changed the peak amplitude of the spike. After the first 10 μm-step, the spike amplitude peaked at 26.19 ± 0.04 μV and then decreased monotonically to 14.74 ± 0.03 μV (120 μm away from the first recording position). The amplitude of the prepotential behaved similarly, starting at 4.96 ± 0.02 μV, peaking after 10 μm at 5.39 ± 0.04 μV, and then decreasing monotonically to 2.77 ± 0.03 μV. The relative delay between the prepotential and the spike peak monotonically increased from 460 μs to 660 μs with depth. The spike amplitude and the prepotential amplitude were significantly correlated with the recording depth and with each other: the Pearson correlation coefficient between the spike amplitude and prepotential amplitude was 0.98 ($p = 4.0 \cdot 10^{-7}$), the correlation between spike amplitude and depth was -0.72 ($p = 0.019$), and the correlation between prepotential amplitude and depth was -0.77 ($p = 0.0097$).

By contrast, the STA EEG waveform did not change significantly when the intracranial electrode was moved. There was always a significant positive peak at -190 μs and always a significant negative peak at 130 μs delay ($p < 0.05$ for each intracranial depth, SD bootstrapping method). Interestingly, the peak amplitudes were independent of the intracranial depths: the Pearson

correlation coefficient between the STA EEG amplitude and recording depth was 0.49 ($p = 0.15$) for the positive peak at -190 μs delay and -0.09 ($p = 0.80$) for the negative peak at 130 μs delay.

In summary, the control experiment provides evidence against cross-talk between the intracranial and the EEG electrodes in general, and thus suggests the absence of contamination between the intracranial electrode and the EEG electrode.

Prediction of the single-unit contribution to the ABR

To predict the single-unit contribution to the ABR, we used the recordings from 24 NM units (in 11 owls). From the single-unit recordings obtained for click stimuli, we obtained the trial-averaged peri-stimulus time histograms (PSTHs), which we mathematically describe by the function $\mathrm{PSTH}_j(s)$ for neuron $j = 1, ..., 24$ for $0 \leq s \leq T_j$ with click at time $s = 0$ and recording duration $T_j \in \{10, 15\}$ ms after the click onset). From the EEG recordings, we had obtained the ABR waveforms. And from the combined intracranial and EEG recordings during spontaneous activity, we had derived the STA EEG, i.e. $C_j(\tau)$ for neuron j and $\tau \in [-4, 4]$ ms, in Equation (1).

To predict a single NM cell's average contribution to the ABR, which we denote as $\mathrm{ABR}_j(t)$ for neuron j, we convolved the PSTH of that neuron with its STA EEG:

$$\mathrm{ABR}_j(t) = \int \mathrm{d}\tau \, C_j(\tau) \, \mathrm{PSTH}_j(t - \tau) \quad .$$

Statistical analysis

All analysis was performed with a custom-written MATLAB code. To estimate the statistical significance of the data, we used the Pearson correlation coefficient and its p-value, N-way analysis of variance (ANOVA), generalized linear models with respective F-statistics, Student's 2-population t-test, and custom bootstrapping methods as explained across Methods. When correction of multiple testing was done, we used the Šidák correction.

Data availability

All the data and codes used to produce the figures in this study are available from the corresponding author upon request.

Results

The aim of this study was to quantify the contribution of the auditory brainstem nucleus magnocellularis (NM) to the auditory brainstem response (ABR). To this end, we determined the contribution of single neurons to the ABR by recording action potentials in NM units simultaneously with the EEG from the scalp. These simultaneous recordings allowed us to estimate the spike-triggered averages (STAs) of NM neurons at the EEG electrode (i.e., the unitary responses). Having measured the click-evoked spike times of the same NM neurons, we could then estimate the neurons' contribution to the click-elicited EEG response, i.e., the ABR.

Classification of single units

To link single cell activity to their contributions to the EEG signal, we analyzed extracellular recordings from 53 single units in 12 owls, obtained within the NM cell body region (Fig. 1A,B). This region also contains auditory nerve (AN) fibers that descend into NM, and NM efferent axons. Thus, AN fibers, NM cell bodies, and NM axons could, in principle, have been recorded at any of the depths used. We classified these units based on their best frequency (BF) and spontaneous firing rate (Fig. 1C,D), since AN units typically have lower spontaneous rates (for each BF) than NM units (Köppl, 1997a). Based on these earlier results, 13 units were putatively classified as AN and 40 units were classified as NM.

We also used the presence and absence of prepotentials (example in Fig. 1B, left) to differentiate between NM cell bodies and AN fibers (see Materials and Methods). Prepotentials have been observed in avian endbulb synapses between AN and NM (Zhang and Trussell, 1994). In the mammalian auditory system, prepotentials originate from the large endbulb of Held synapse between the AN and the anterior ventral cochlear nucleus (AVCN) and from the calyx synapse in the medial nucleus of the trapezoid body (MNTB) (see Discussion, e.g. Pfeiffer, 1966; Kopp-

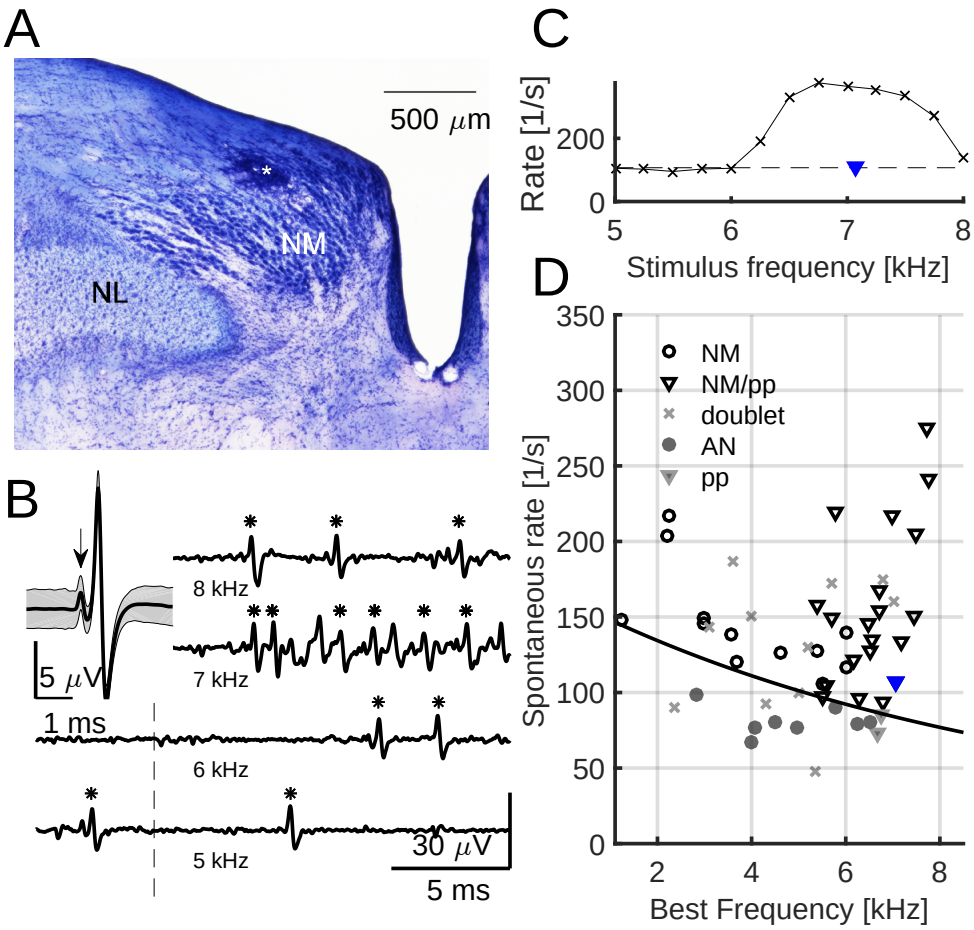

Figure 1: Recordings from NM cell body region. A: Exemplar recording location (lesion, *) in a Nissl-stained coronal slice through the auditory brainstem. The nucleus laminaris (NL) is both ventral and lateral to NM. **B**, Right: Extracellular recordings from an NM neuron in response to tones at different frequencies (tone onsets indicated by vertical dashed line, spikes marked with *). Left: Average waveform of 22 641 spontaneous spikes (black line) ± SD (gray backgound); prepotential indicated by arrow. **C**: Frequency-response tuning curve to pure tones at 50 dB SPL, with a maximum driven spike count rate of 376 spikes/s at 6750 Hz stimulus frequency. The best frequency (BF, marked with a blue triangle) of this unit was 7065 Hz. The dashed line indicates the spontaneous spike count rate 107 spikes/s. **D**: Spontaneous firing rates and BFs of all 53 units. Legend: *NM*: nucleus magnocellularis unit without a prepotential. *MN/pp*: nucleus magnocellularis unit with a prepotential. *AN*: auditory nerve fiber unit. *pp*: a low-spontaneous rate unit with a prepotential. *doublet*: any unit with doublet-spiking. The NM/pp-unit shown in B and C is marked additionally with a blue triangle. Solid line: the decision boundary between NM and AN units (see Methods).

Variable	**mean ± SE**	**range**	N
Number of spontaneous spikes	$43\,700 \pm 1\,100$	[10 558, 140 141]	32
Spontaneous rate (1/mean ISI)	151 ± 2 spikes/s	[94, 275] spikes/s	32
Amplitude of spontaneous spikes	13.4 ± 0.3 μV	[0.7, 28.4] μV	32
SNR of spontaneous spikes	13.86 ± 0.07 dB	[8.91, 18.24] dB	32
Best frequency (BF)	5580 ± 60 Hz	[1250, 7750] Hz	32
Prepotential amplitude	2.20 ± 0.07 μV	[0.11, 4.96] μV	19
Prepotential amplitude, % of spike	13.1 ± 0.3 %	[5.4, 21.2] %	19
Prepotential SD % of spike SD	107.6 ± 1.2 %	[82.5, 174.9] %	19
Prepotential delay wrt. spike	509 ± 6 μs	[340, 820] μs	19
Mode of ISI distribution	900 ± 30 μs	[400, 2100] μs	19

Table 1: Descriptive statistics of the NM population. Abbreviations: SNR: signal-to-noise ratio. ISI: inter-spike interval.

Scheinpflug et al., 2003; Englitz et al., 2009). We concluded that single-units with a prepotential originated, with a high probability, from the vicinity of NM cell bodies (see Table 1 for their properties). Most units with a prepotential (19 out of 21, black downward open triangles in Fig. 1D) aligned with our classification as NM that was based on BF and spontaneous rate. The two units with a low spontaneous rate but showing a prepotential (gray filled downward triangles) were classified as ambiguous (see Materials and Methods).

The stringent classification criteria used so far resulted in the identification of 40 units as originating from NM neurons. Among them, eight units were excluded because of a high proportion of spike doublets (gray crosses; see also Materials and Methods) because it is challenging to determine STAs for such units. Thus, 32 NM units from 12 owls were used in later analyses (black circles and black downward triangles in Fig. 1D; see also Table 1 for properties of these units).

Click-evoked activity in NM

To evaluate the contribution of single units to the ABR, typically evoked by a click stimulus, we recorded peri-stimulus time histograms (PSTHs) of NM units in response to clicks. We recorded responses to a range of click levels for each unit ($10 - 65$ dB SPL, examples in Fig. 2A). To characterize the click-elicited single-unit responses from NM units, we described their single-unit PSTHs by click-response latency (arrowheads in Fig. 2A). This click onset timing could only be identified for clicks at ≥ 30 dB SPL.

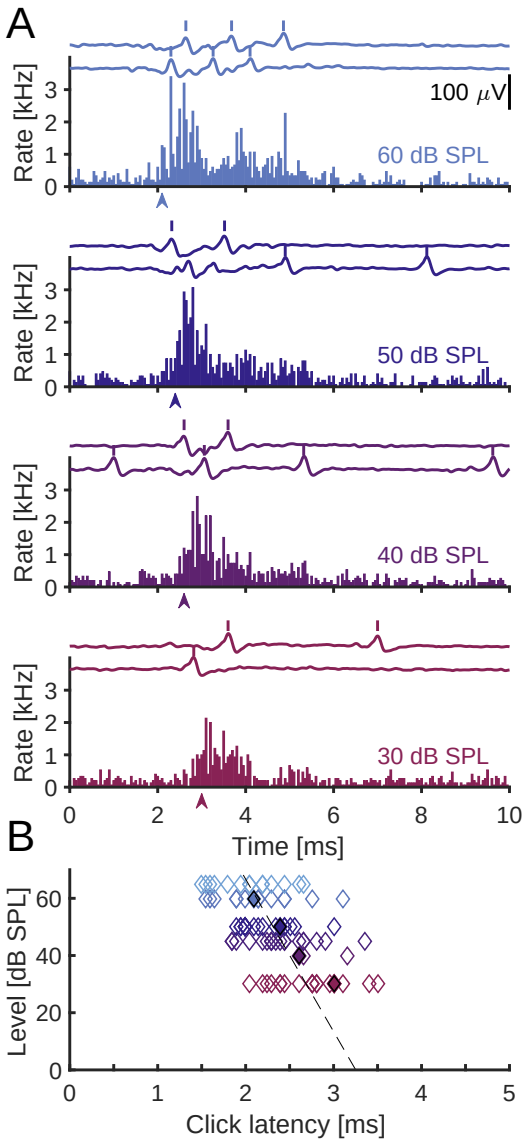

Figure 2: Click-response latency in NM is level- and BF-dependent. A: Examples of click-elicited responses (top) and their peri-stimulus time histograms (PSTHs, bottom) from a single NM unit in response to four different stimulus levels. Bin width: 50 μs. The arrow-heads mark the click-response latency at each level. **B:** Click-response latency decreased with increasing stimulus level and with increasing BF. The examples in A are marked with circles. Dashed line: -19 ± 3 μs/dB$\cdot$level$+3.3\pm0.2$ ms (the GLM for the mean BF = 5.58 kHz). 32 NM units, with 1–4 stimulus levels each, resulting in $N = 91$ click-response latencies.

At the population level, the NM units' click-response latency decreased with increasing level (Fig. 2B) and with increasing BF. A generalized linear model (GLM) showed a significant dependence of click-response latency on both level and BF: -19 ± 3 μs/dB $\cdot$ level $- 90 \pm 20$ μs/kHz $\cdot$ BF $+3.8 \pm 0.2$ ms, with $p(\text{level}) = 1 \cdot 10^{-8}$ and $p(\text{BF}) = 2 \cdot 10^{-5}$ (F-statistics: vs. constant model: $F_{3,88} = 34.4$, $p = 9 \cdot 10^{-12}$; normally distributed residuals, no interaction term between BF and level: $p = 0.61$). If we neglect the dependence on BF, the level dependence of click-response latency had a slope of -19 ± 3 μs/dB (Fig. 2B, dashed line). Köppl (1997b) reported similar values, showing delay-to-level slopes for the tone-elicited delays in 3 NM cells, with slopes ranging from -24 μs/dB to -16 μs/dB (fitted from their Fig. 9, no center frequencies given).

ABR timing: Delays originate in the inner ear

In order to relate the activity of the single units to the EEG, we first measured and quantified the properties of the EEG on its own. We recorded ABRs in response to click stimuli whose sound levels varied from 10 to 65 dB SPL.

ABRs typically contained three positive-going waves within the first 8 ms following the click presentation (Palanca-Castán et al., 2016), and the latencies of the peaks of the three waves increased with decreasing stimulus level (examples in Fig. 3A). To quantify the dependence of the latencies of the peaks on the stimulus level, we analyzed the shift of the three waves as well as their inter-peak-intervals in 27 ABR recordings in 13 owls. The latency of the peak of each wave was indeed level-dependent (all Pearson correlation coefficients < -0.83 with p-values $< 10^{-20}$) across the recordings, and their slopes (Fig. 3B) were not significantly different (GLM with mean-shifted intercepts, all $p = 1$, GLM: $F_{4,225} = 231$, $p = 7 \cdot 10^{-68}$). The level-dependent slope across all peaks was -23.1 ± 0.9 μs/dB, with intercept 2.82 ± 0.05 ms for the first peak, 3.54 ± 0.04 ms for the second peak, and 4.49 ± 0.04 ms for the third peak (GLM: $F_{4,221} = 1230$, $p = 2 \cdot 10^{-137}$, mean $\pm$ SE).

The level-dependent fits for the click-response latency in the NM population (dashed line in Fig. 2B) and the ABR wave II peak delay (solid line in Fig. 3B) were equal within their error margins. We performed an N-way analysis of variance (ANOVA) based on the hypothesis that both groups (ABR wave II peak delay: $N = 75$ and click-response latency: $N = 91$) originated

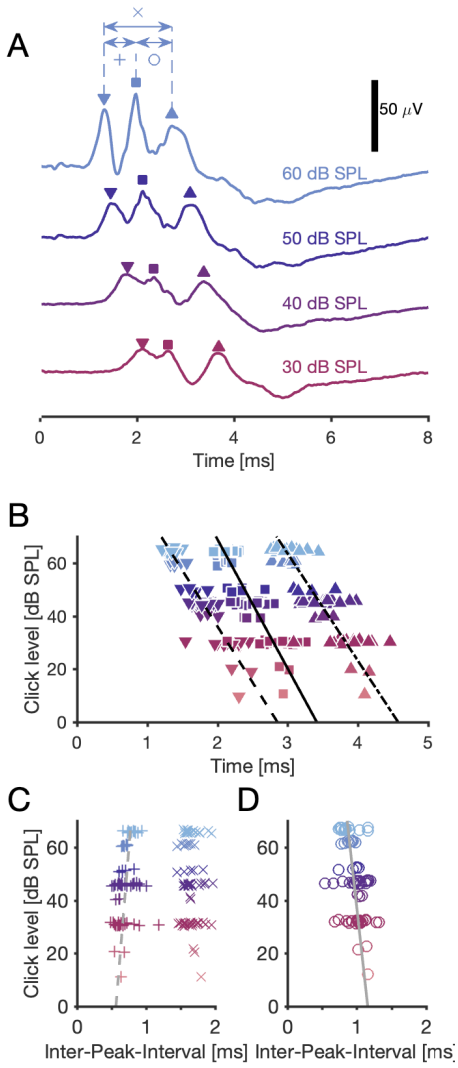

Figure 3: Inner ear dominates the dependence of delay of ABR waves on sound level. A: Examples of an ABR, recorded in response to four different levels of a click with onset at 0 ms. Each curve shows three main peaks (marked with symbols '∇' for wave I, '$\square$' for wave II, and '$\triangle$' for wave III). The inter-peak-intervals are marked with symbols 'x', '+', and 'o'. **B**: ABR waves' peak timing depended significantly on the stimulus level. Linear least-square fits (lines): Wave I peak: -24 μs/dB·level+2.853 ms. Wave II peak: -21 μs/dB $\cdot$ level $+$ 3.414 ms. Wave III peak: -25 μs/dB $\cdot$ level $+$ 4.573 ms. All groups: Pearson correlation coefficients < -0.84 with p-values $< 10^{-20}$, $N = 75$ for each wave. The markers are jittered within 1 dB to reduce overlap. **C**: The inter-peak-interval between peaks 1 and 2 depended on the stimulus level as 3.1 μs/dB $\cdot$ level $+$ 0.561 ms (linear least-square fit), with Pearson correlation coefficient of 0.35 ($p = 0.0022$). The average inter-peak-interval ($\pm$ SE) between peaks 1 and 3 it was 1.67 ± 0.02 ms with no significant correlation with level (Pearson CC: -0.11; $p = 0.34$). **D**: The inter-peak-interval between peaks 2 and 3 depended on the stimulus level as -4 μs/dB $\cdot$ level $+$ 1.159 ms (linear least-square fit), with Pearson correlation coefficient of -0.41 ($p = 0.00034$, $N = 75$). B–D: 24 ABR recordings, with 1–4 stimulus levels each, resulting in $N = 75$ delays and inter-peak-intervals per group.

from the same level-dependent regression model. The group identity had no significant effect on the fit ($F_{1,156} = 1.9$, $p = 0.18$), whereas the level did ($F_{7,156} = 19.2$, $p = 3 \cdot 10^{-18}$), indicating that there was no significant difference between the delays of the ABR wave II peak and the NM cells' click-response latency.

By contrast to the latencies of the peaks, the inter-peak-intervals (Fig. 3C,D) showed a much weaker level dependence. The inter-peak interval between peaks 1 and 3 (IPI_{13}) showed no significant level dependence (Pearson correlation coefficient for IPI_{13}: -0.0011 with $p_{1,3} = 0.36$, $N = 75$ in each IPI group), with mean ($\pm$SE) $\mathrm{IPI}_{13} = 1.67 \pm 0.02$ ms. The level dependency of $\mathrm{IPI}_{12} = 3.1\ \mu\mathrm{s/dB} \cdot \mathrm{level} + 0.561$ ms and of $\mathrm{IPI}_{23} = -4\ \mu\mathrm{s/dB}\ +1.116$ ms (linear least-square fits) were nevertheless significant (Pearson correlation coefficients of IPI_{12}: 0.35, $p = 0.0022$ and of IPI_{23}: -0.31, $p = 0.0071$).

Our results so far have implications for the origin(s) of the level dependence of delays in the auditory pathway. ABR wave I is assumed to reflect auditory nerve activity (Melcher and Kiang, 1996). Consistent with this hypothesis, the strong overall level dependence of ABR latency in our data set was mainly defined by the response of the cochlea, which is level dependent. Furthermore, the much weaker dependence of inter-peak-intervals suggests that delays between brainstem nuclei are mainly caused by fixed structural delays, such as synaptic delays and axonal conduction delays, which are basically level independent.

Finally, we also quantified how the peak amplitude of the ABR wave II was modulated by stimulus level. The ABR wave II peak amplitude correlated in the population strongly with the level (Pearson correlation coefficient: 0.65, $p = 4 \cdot 10^{-10}$, $N = 75$) with the slope of 0.47 μV/dB and intercept of -9.3 dB (linear least square fit).

Spontaneous spikes of individual NM neurons were detectable in the EEG signal

To connect the action potentials of single NM cells to the macroscopic EEG, we analyzed the average EEG around the times of spikes. The average contribution of a spike from a single unit is referred to as spike-triggered average (STA) EEG. For this analysis we only used spontaneous

STA EEG Variable	**mean ± SE**	**range**	N	p
Number of spont. spikes (significant)	$58\ 000 \pm 3\ 000$	[12 121, 110 827]	16	0.13
Number of spont. spikes (all)	$50\ 600 \pm 1\ 500$	[12 121, 140 141]	24	
Amplitude of STA EEG peak (significant)	76 ± 4 nV	[25, 267] nV	16	0.31
Amplitude of STA EEG peak (all)	68 ± 3 nV	[25, 267] nV	24	
SD of STA EEG waveform (significant)	5.10 ± 0.09 μV	[3.17, 7.52] μV	16	0.08
SD of STA EEG waveform (all)	5.54 ± 0.08 μV	[3.17, 11.17] μV	24	
SNR of STA EEG ($\mathrm{SNR_{STA}}$) (significant)	-42 ± 1 dB	$[-79, -22]$ dB	16	0.31
SNR of STA EEG ($\mathrm{SNR_{STA}}$) (all)	-39.4 ± 0.7 dB	$[-78.9, -18.3]$ dB	24	
Delay wrt. STA EEG peak (significant)	-95 ± 12 μs	[−690, 110] μs	16	0.77
Delay wrt. STA EEG peak (all)	-105 ± 10 μs	[−830, 270] μs	24	

Table 2: Spike triggered average EEG amplitudes and delays of NM units. The p-values refer to Student's 2-population t-test between the STA EEG populations of significant ($N = 16$) and non-significant ($N = 8$) waveforms.

spikes in order to avoid stimulus-induced correlations among neurons, which would distort the computed STA EEG. Eight NM units (of $N = 32$, Figs. 1 and 2) were excluded from this analysis because their respective EEG recordings failed the stringent inclusion criteria for the EEG; these criteria included both suspected crosstalk between the electrodes and weak ABR responses (see Methods).

Figure 4A,B shows two examples of NM units and their corresponding STA EEG. Two thirds of the analyzed NM neurons (16 out of 24) contributed a statistically significant STA EEG waveform (Fig. 4C) according to the SNR-method by Parks et al. (2016) with an SNR lower bound of 0 dB (see Methods). Yet, averaging over typically 50 000 spontaneous spike times per unit revealed significant waveforms (see text and asterisks next to the waveforms in Fig 4C). Across the population of 16 significant units, there was a large spread both in the amplitudes of the STA EEG peaks and their timing (Fig 4C, D). The peak amplitude of the STA EEG ranged from 25 to 267 nV (mean ± SE: 76 ± 4 nV, see Table 2). The noise in the EEG signal was typically about 3 orders of magnitudes larger than the peak amplitudes of the EEG STA, corresponding to a very low SNR of -42 ± 1 dB (mean ± SE, see Table 2).

Most of the STA EEG maxima occurred slightly prior to the maximum of the extracellular spike waveform, with a mean (± SE) delay of -95 ± 12 μs ($N = 16$; see Table 2 and Fig. 4C, D). The STA EEG peak being close to the spike maximum is consistent with the assumption that we

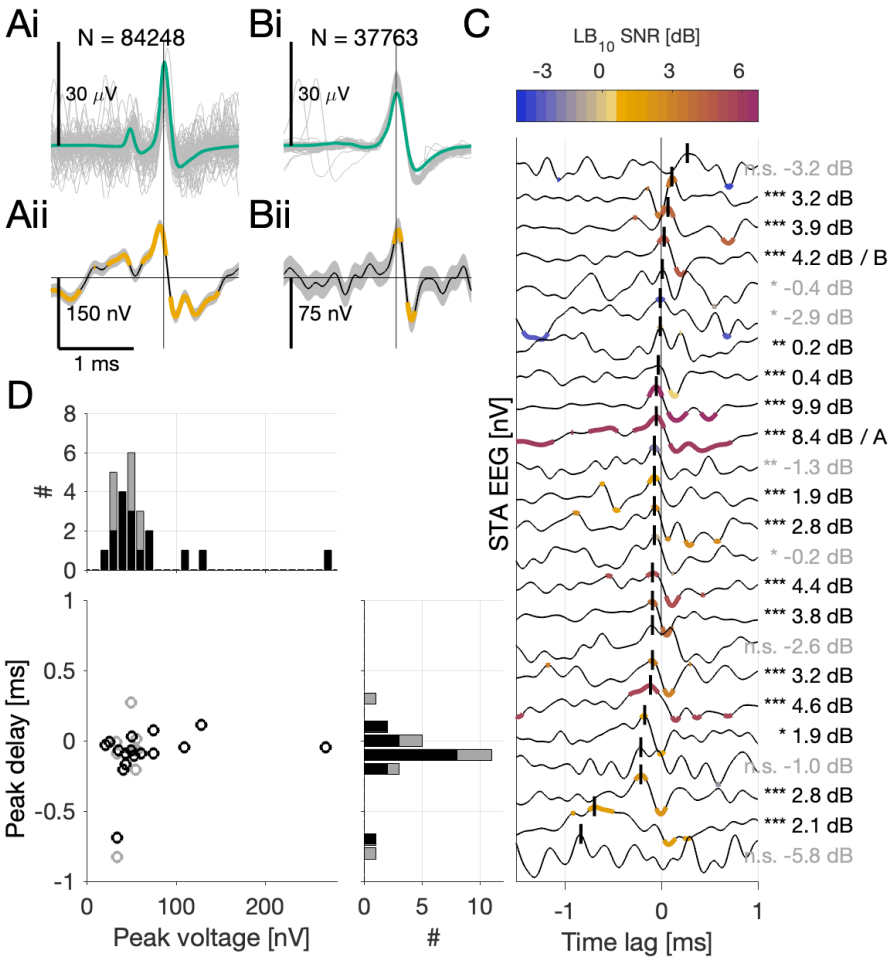

Figure 4: Magnocellular single cell spikes make a detectable contribution at the scalp electrode. **Ai:** Average spike waveform of 84 248 spontaneous spikes of an NM cell (green), recorded extracellularly, and a random selection of 100 spike waveforms thereof (gray). **Aii:** Average waveform at the EEG electrode (STA EEG, black) and SE (shaded), with EEG waveforms aligned to the peaks of the spikes of the NM cell in Ai (thin vertical black line). The parts of the STA EEG marked in orange have a significance level $p < 0.01$, and black portions are non-significant. **B:** Average spike waveform and STA EEG from a different NM unit. **C:** 24 STA EEGs, sorted by the timing of their peaks (vertical black bars) within ± 1.0 ms with respect to the spikes of the respective NM units. Significant curves ($\mathrm{SNR_{LB}} \geq 0$ dB) are highlighted by black numbers of the corresponding values of the $\mathrm{SNR_{LB}}$ ($N = 16$); non-significant curves with gray numbers ($N = 8$). Asterisks indicate the maximum bootstrapped significance of the SDs of curves (*: $p < 0.05$, **: $p < 0.01$, ***: $p < 0.001$, see Methods), and significant parts of the waveforms are colored according to the colorbar at the top. Not significant parts in black. **D:** Peak delays (STA EEG wrt. the spike waveform) and maximum STA EEG amplitudes (peak voltages) were not correlated (Pearson CC: 0.20, $p = 0.35$, $N = 24$). Significant data points ($\mathrm{SNR_{LB}} \geq 0$ dB) are black ($N = 16$), and the non-significant ones are gray ($N = 8$). There was no difference between the two groups neither in the number of spikes, in the peak voltages, in the peak delays nor in the SNR of the STA EEG (see Table 2). Histogram on the top: distribution of the STA EEG peak voltages. Histogram on the right-hand side: distribution of the STA EEG peak delays. Population statistics: see Table 2.

Prediction variable	**median ± SE**	**range**
Prediction amplitude, *mean	$32.9^* \pm 1.1$ nV	[2.5, 162.7] nV
Prediction amp./ spike amp.	0.07 ± 0.02 %	[0.009, 3.993] %
P1-P_{pred}	-950 ± 30 μs	$[-3260, -400]$ μs
P2-P_{pred}	-300 ± 20 μs	$[-2520, 200]$ μs
P3-P_{pred}	710 ± 20 μs	$[-1360, 1140]$ μs
P_c-P_{pred}	-160 ± 11 μs	$[-1360, 380]$ μs
P1-P_c	-780 ± 13 μs	$[-1900, -480]$ μs
P2-P_c	0 ± 13 μs	$[-1160, 0]$ μs
P3-P_c	870 ± 13 μs	[0, 1280] μs

Table 3: Prediction amplitudes and relative prediction delays of NM units. $N = 38$ predictions.

typically recorded intracranially close to the cell bodies and that the (far-field) dipoles originating from these neurons would have a similar but not necessarily equal peak time at the scalp.

Predicted NM contribution matches the peak latency of the ABR wave II

In order to establish a direct connection between click-elicited NM single cell activity (Fig. 2) and the ABR (i.e., click-elicited EEG response, Fig. 3), we recorded them simultaneously and used the STA EEG (Fig. 4) to predict the single-cell contribution to the ABR (Fig. 5).

For each unit, we convolved its peri-stimulus time histogram (PSTH) with its spontaneous STA EEG (Fig. 5A). This procedure results in an average (across trials) contribution of this individual cell to the EEG because in every single trial an NM unit typically produces several spikes, and the STA-EEG contributions add up. Averaging such EEG contributions across many single trials is equivalent to averaging the spiking responses of an NM unit, resulting in the PSTH, and then convolving the PSTH with the STA EEG.

The predicted contribution of the NM exemplary unit (Fig. 5A, green) had a 162 nV peak amplitude. The prediction peak was aligned in time with the peak of wave II (P_2) of the click-driven ABR response with a difference of 240 μs. The click-driven ABR response had an amplitude of 47 μV (Fig. 5A, yellow), and thus this NM unit contributed about $0.28 \pm 0.02\%$ to the ABR wave II amplitude.

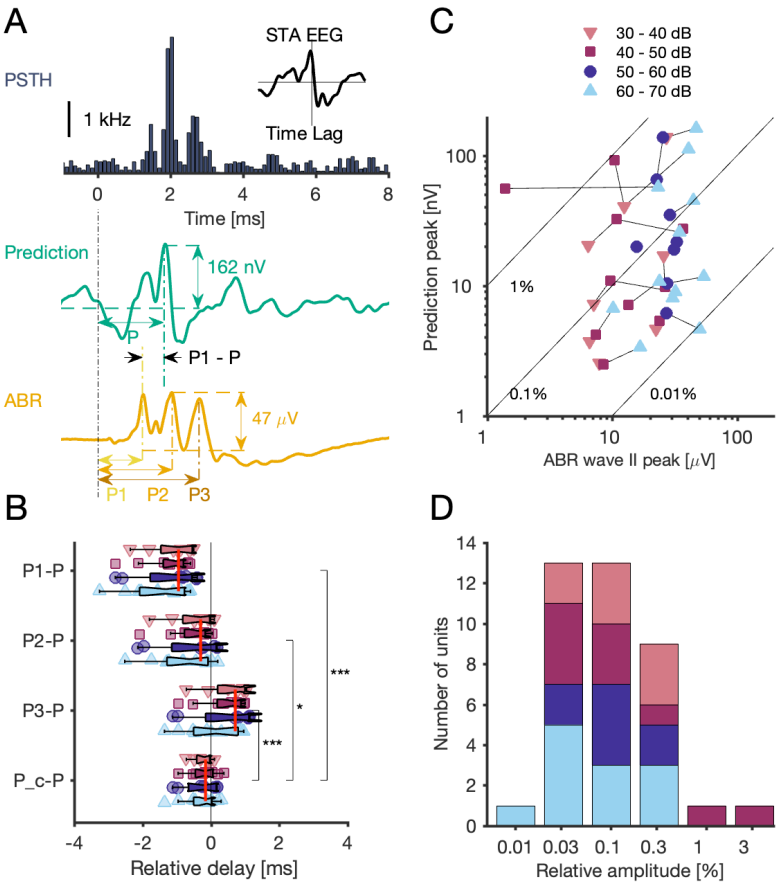

Figure 5: Predicted NM single-cell contribution aligns best with peak of ABR wave II. A, Top: PSTH (gray bars) in response to a click stimulus at 0 ms. Inset: STA EEG of the spontaneous spikes ($N = 84\,248$; see Fig. 4Aii). Middle: Prediction (green) of the single-unit contribution to the ABR, calculated as the convolution of the STA EEG with the PSTH (both shown above); peak amplitude of prediction: 162 nV (wrt. average level at click onset ± 1 ms). Delay of peak indicated by 'P'. Bottom: ABR (yellow) in response to the click stimulus; peak-to-peak amplitude of ABR wave II: 47 μV (wrt. lowest neighboring minimum). Delay of peak indicated by 'P2'. All parts of this panel share the same time scale, and the click onset is marked with a vertical line at 0 ms. **B–D**: Population data from 38 EEG recordings (at variable click levels) and from 16 NM cells. Plots share the same color schema with respect to stimulus levels (see legend in C). **B:** Boxplots and data points of the relative delays wrt. each ABR peak and for each level group. The relative delay is the difference between the delay P of the predicted single-cell ABR contribution peak and one of the delays (P1, P2, or P3) of a peak of ABR waves I through III; we also show the relative delay of the predicted peak and the closest ABR wave's peak ($P_c - P$; *: $p = 0.011$, ***: $p < 0.0001$, 2-population t-tests). The vertical red lines mark the medians of each relative delay across levels. **C:** Amplitude of predicted peak vs. amplitude of ABR wave II. Short lines connect data points obtained from the same NM cell but at different click levels. Long diagonal lines indicate fixed relative amplitude, i.e. ratio of predicted and observed amplitudes of peaks. **D:** Histogram of relative amplitudes.

Also for the population, the peak of ABR wave II (P2) was closest to the predicted peak (Fig. 5B), with a median ($\pm$ SE) relative delay of -300 ± 20 μs ($N = 38$ considering all stimulus levels, see also Table 3). The distribution of these relative P2-delays was nevertheless significantly different from the distribution of closest possible relative delays (-250 ± 11 μs, 2-population t-test, Šidák-corrected for multiple comparisons: $p = 0.011$). However, most of the closest delays stemmed from the wave II peak (23 out of 38), and a minority from the wave III peak (15 out of 38), with no significant difference in the stimulus levels between these groups (2-population t-test, $p = 0.12$). By contrast, the distributions of the relative delays for P1 and P3 (-1290 ± 20 μs and 340 ± 20 μs, respectively) were both highly significantly different from the distribution of closest possible relative delays ($p < 0.0001$, 2-population t-tests).

We previously showed that the level dependence was strong both for the ABR peak delays (Fig. 3B) and for the click-response delays (Fig. 2B), and that at the population level the slopes were indistinguishable. However, these slopes are insufficient to establish that at the single-cell level the relative timing between the prediction and the ABR peak(s) is level-independent. For example, the peak (but not the onset latency) of the PSTH will dominate the timing of the prediction peak. We therefore performed an N-way ANOVA based on the hypothesis that the delay of the ABR wave II peak and the delay of the prediction peak (both: $N = 38$) originated from the same level-dependent regression model. The group identity had no significant effect on the fit ($F_{1,69} = 1.14$, $p = 0.35$), whereas the click level did ($F_{5,69} = 6.92$, $p = 0.011$), indicating that there was no significant difference between the delay of the ABR wave II peak and the delay of the prediction peak. Furthermore there was no significant correlation between the level and the relative prediction delay with respect to the wave II peak delay ($p = 0.35$, $N = 38$; see Fig. 5B group P2-P). Such a level independence, additionally to the large spread of the relative delays in population, thus means that the ABR wave II peak delay can not be predicted reliably by a single NM unit, but that the wave II is expected to arise only when averaging over a large population of such predictions.

The predicted amplitudes were broadly distributed ('prediction peaks' in Fig. 5C), and the relative amplitudes of predictions ranged from 0.01% to 1% of the ABR wave II peak amplitudes with a median ($\pm$ SE) of 0.07 ± 0.02 % (Fig. 5C, D). One outlier (about 4%) was attributed to an

unusually small ABR wave II peak amplitude. Neither the absolute amplitudes of the predictions (in nV), nor their relative amplitudes were significantly dependent on the stimulus level as a population (Pearson correlation coefficients: $CC = 0.14$, $p = 0.40$ and $CC = -0.07$, $p = 0.64$, respectively). Predicted amplitudes were, however, significantly dependent on stimulus level for 9 out of 16 NM units, when considering the logarithms of both 'prediction peaks' and the 'ABR wave II peaks', and using NM units' identity as a random effect (GLM: $F_{17,21} = 43$, $p = 2 \cdot 10^{-12}$). All in all, the stimulus level was not a good predictor of a unit's relative contribution to the ABR wave II amplitude nor in population (Pearson correlation coefficient: 0.25, $p = 0.13$), neither when using individual owls as a random effect (GLM: $F_{17,21} = 1.69$, $p = 0.13$). This means that the relative peak amplitude of a given single unit to the ABR did not change reliably with stimulus level.

Discussion

Simultaneous recordings of ABRs and single units in barn owl NM demonstrated that individual spikes can make detectable contributions to the EEG with amplitude 76 ± 4 nV (range $25 - 267$ nV). The median single-unit contribution to the click-driven ABR was $\approx 0.1\%$ of the elicited ABR wave II peak.

The time lag of the peak of the single-cell spike-triggered average (STA) EEG typically coincided with the rising phase of the extracellular NM spike waveform ($-95 \pm 12\,\mu$s). However, the range of time lags was large (from -300 to $+110$ μs excluding one outlier, Fig. 4D). This could be due to the variable position of the intracranial electrode: the peak of the STA EEG is locked to the spike generation at the soma, but the propagation of the spike from the soma along the axon to the location at which the intracranial recording electrode is closest adds a variable delay. The longer this delay the more negative the 'time lag'. Furthermore, NM neurons have a variable spatial orientation, and this variable dipole axis can add variability to the time lag of the peak of the STA EEG. In contrast to the often negative time lag and the large variability we found, Teleńczuk et al. (2010) reported cortical STA EEGs for which the peak either coincides with the spike peak time, or for which the STA EEG has a rising phase at the spike time; and there was only a $100\,\mu$s-range delay between the peaks. The grand average peak had some $50 - 100\,\mu$s

positive delay wrt the spike peak. This may be explained by intracranial electrodes always being close to the soma and a preferred orientation of the dipole of pyramidal cells.

Let us compare the magnitude of obtained STA EEGs with those in other systems. We estimated the dipole moment Q of a spike generated by an NM cell based on the STA EEG peak potential V_{STA} with the dipole approximation (Malmivuo and Plonsey, 1995)

$$Q = \frac{4\pi}{\eta} \frac{V_{\mathrm{STA}}(r,\theta) \cdot r^2}{\cos(\theta)}$$

with constant tissue resistivity $\eta = 2.47\ \Omega\mathrm{m}$ (Logothetis et al., 2007), angle θ with respect to the dipole axis, and distance r of the EEG electrode from the source. The average intracranial recording depth below the dura was 10.2 ± 0.7 mm (mean $\pm$ SD). The active EEG electrode was positioned in the bone at $\approx 1-2$ mm above the dura and ≈ 5 mm away from the intracranial electrode, which leads to $r \approx 12$ mm. Furthermore, we assumed $\theta = 0$ for the active EEG electrode. Thus, for the range of our STA EEG peak amplitudes ($25 - 267\,\mathrm{nV}$), the dipole moments range $\approx 20 - 200$ nA mm. These dipole moments are larger than the dipole moments reported for cortical pyramidal neurons: Murakami and Okada (2006) found $Q = 0.78 - 2.97$ nA mm, which matches to data from pyramidal neurons of macaque monkeys (Teleńczuk et al., 2010) as well as to modeling results for rat and human cortical neurons (Næss et al., 2021).

The estimated dipole moment for spikes of NM cells highly depends on the (unknown) spatial orientation of the dipole. Furthermore, the dipole moment depends on cell morphology (e.g. Næss et al., 2021), including the turns of the axon (Stegeman et al., 1987; Jewett et al., 1990), distribution of synaptic inputs (Gold et al., 2006; Lindén et al., 2010), spike generation site (Telenczuk et al., 2017), possible after-hyperpolarizing currents (Storm, 1987), and possible back-propagation of the spike (Gold et al., 2006; Telenczuk et al., 2017). Better understanding the differences of cortical and brainstem single-cell contributions to EEG calls for further modeling studies.

We predicted the average contribution of a single NM cell to the ABR by convolving the STA EEG with the cell's click-elicited PSTH. This led to the amplitude $32.9 \pm 1.1\,\mathrm{nV}$ (range $2.5 - 162.7\,\mathrm{nV}$) or about half the average amplitude of STA EEGs. Although each NM neuron fires several spikes

in response to a click, which could in principle increase the summed amplitude, the temporal dispersion of the spikes, as visible in PSTHs, leads to a reduction in amplitude. The largest predicted single-cell contributions were $\approx 1\%$ of the min-max amplitude of ABR wave II, and the median was $\approx 0.1\%$ (Fig. 5C). Such large contributions were unexpected because NM has around $26,000$ neurons (Han et al., Submitted). Therefore, an NM neuron should contribute only $\approx 1/26,000^{th}$ of the total ABR wave II response. There are several potential causes for this discrepancy: even though the peaks of the predicted contributions of individual NM neurons aligned best with wave II of the ABR, the peaks showed temporal jitter (from -2.5 to $+0.2$ ms, Table 3), which reduces the amplitude of the peak of the summed (across many NM neurons) ABR. Some units even made a negative contribution to the peak II. Furthermore, we selected statistically significant STA EEGs, which could have biased the amplitudes to large values.

The compound effect of a neuronal population to the ABR depends on the synchronization of the cells within the population (Kuokkanen et al., 2010; Ahlfors et al., 2010a,b; Lindén et al., 2011). Temporal synchrony is famously precise in the auditory brainstem (Kuokkanen et al., 2010; McColgan et al., 2017) leading to macroscopic signals that can be recorded at the scalp more than a centimeter from their source. Note that the ABR, exhibiting several waves, is a sum of several subsequently activated neural populations. Thus, assumptions of the populations' spatial alignment and temporal synchronization underlie, at least implicitly, all ABR models (Melcher and Kiang, 1996; Ungan et al., 1997; Dau, 2003; Goksoy et al., 2005; Riedel and Kollmeier, 2006; Colburn et al., 2008; Schaette and McAlpine, 2011; Rønne et al., 2012; Verhulst et al., 2015, 2018). NM responses alone are sufficient to produce wave II, but a thorough quantification would require additional modeling to consider the variable geometry of NM cells. Furthermore, other sources, such as nucleus angularis (Takahashi and Konishi, 1988; Köppl and Carr, 2003) are likely to contribute to wave II. Nucleus angularis, like NM, is a first-order auditory nucleus with similar average onset latencies as NM (Köppl and Carr, 2003), and its contributions are thus expected to be temporally aligned with the ABR wave II. However, the observed variation in onset latencies ($\approx 1.5 - 4.5$ ms for $20 - 35$ dB tones, Köppl and Carr, 2003) and in response types in nucleus angularis raises questions about their coherence in generating a collective ABR peak (Sachs and Sinnott, 1978; Soares et al., 2002; Köppl and Carr, 2003).

Other brainstem structures, such as the nucleus laminaris and the superior olivary nucleus can be excluded as wave II sources because they have longer response latencies than NM (Lachica et al., 1994; Yang et al., 1999; Monsivais et al., 2000; Burger et al., 2005). McColgan et al. (2017) estimated that the branching patterns of the NM axons in NL could collectively contribute microvolt excursions in the scalp EEG recordings. This contribution is expected to be more aligned with ABR peak III than peak II, considering a conduction delay of about 1.2 to 1.5 ms between the NM cell body response and the responses from their axonal arbors in the NL (Carr and Konishi, 1990; Köppl, 1997b).

There are clear differences between the *unitary response* (UR), as used in the ABR modelling, and the STA EEG (and its convolution with the PSTH) that we have measured, despite the fact that the UR is defined as the expected average spike-triggered response of a single neuronal source at the EEG electrode. For one, the UR, as often used in ABR models, is typically derived from the driven responses (ABRs) by deconvolution, and thus includes the structurally correlated cascade of activation of any neuronal sources associated with the spike in a single auditory nerve fiber (Dau, 2003; Rønne et al., 2012; Verhulst et al., 2015, 2018). By contrast, we tried to minimize such correlations in our STA EEG by using spontaneous spikes, and show only the scalp contribution of single NM cells. Secondly, the UR has the same average waveform for all sources, disregarding any variation in the neuron population or even between neuron types. By contrast, our STA EEGs included the large variability present in the NM cell population. Thus, defining the STA EEG for group of single neurons in a single nucleus will help limit the number of possible realistic unitary responses. Given the wide range of the STA EEG responses, our data suggest that it is unlikely that a single NM spike-triggered average EEG waveform represents the UR. Nevertheless, an NM UR can be derived from the sum of the STA EEG responses.

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Disclosures

No conflicts of interest, financial or otherwise, are declared by the authors.

Author Contributions

Author contributions: P.T.K., C.E.C., and R.K. conception and design of research; C.E.C. and I.K. performed experiments; P.T.K. analyzed the data; P.T.K., C.E.C., C.K., I.K., and R.K. interpreted results of experiments; P.T.K. prepared figures; P.T.K. drafted manuscript; all authors edited and revised manuscript; all authors approved final version of manuscript.

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