Novel sex-biased outcomes in neuroblastoma are associated with distinct gene expression and chromosomal loss patterns.

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Abstract

The worst patient outcomes in neuroblastoma are driven by high-risk disease[1,2], which is divided into similarly sized *MYCN* amplified and *MYCN* non-amplified patient subgroups[3]. Male patients have been reported to have slightly worse outcomes than females in all-patient analyses of large patient studies[3,4]. However, we show here that in *MYCN* non-amplified high-risk and stage 4s low-risk disease, female patients have significantly worse overall survival than males. Female *MYCN* non-amplified high-risk patients highly express *H19* and *DLK1*, both of which drive cell growth *in vitro* and are associated with worse outcomes in females but not males. Further, chromosome-specific expression analysis of these patients reveals broad sex disparities in chromosomal patterning, including female-specific retention of chromosome 11q, a pattern typically reserved for *MYCN*-amplified disease[5,6]. Finally, we show that *H19*, a known *let-7* microRNA target[7], sequesters *let-7* in females, providing a rationale for worse female survival and reconciling retention of chromosome 11q. We propose that this novel sex-based outcome disparity is driven by *let-7* inhibition, supporting a model of neuroblastoma development where *let-7* mitigation is central to disease pathology[8].

Introduction

Neuroblastoma (NB) is the most common extracranial solid tumor in children that arises from the neural crest lineage[9,10]. Survival rates in NB are highly dependent upon tumor risk group classification. According to the Children's Oncology Group risk system, NB tumors can be categorized as low-risk (LR), intermediate-risk (IR), or high-risk (HR) groups based on stage, patient age, favorability of histology, *MYCN* status, and presence of other genetic aberrations such as ploidy or chromosomal loss[11,12]. LR and IR NB have favorable overall survival (90-95%), as do stage 4s tumors, which often regress without intervention[10,13,14]. Unfortunately, HR disease is notoriously difficult to treat and has an OS rate below 50%[15,16]. Consequently, HR NB patients endure one of the most intensive genotoxic therapeutic regimens in all of oncology[17], and those that do survive are often left with life-long complications and increased risk for secondary cancers. An increased understanding of the genetic drivers of HR disease will contribute to better classification and stratification of NB patients and to the development of more effective, less genotoxic treatments that improve patient survival and quality of life.

While sex and racial disparities have been previously described [3,4,18–21], the broad-stroke level of interrogation employed is insufficient for understanding cancers with complex genetic subgroupings, such as NB[3,4,18–21]. Large-scale studies of patient outcomes often report that males with NB fair slightly worse than females. While the overarching statistics associated with these conclusions are correct, it is important to consider that the incidence of both NB itself

and HR disease is higher in males. On a per-patient basis, male and female outcomes have not been directly investigated, in particular within the HR NB patient population. This concern spans all cancer types as analysis of patient populations lacking available annotations for categories even as simple as race or sex masks both the source of identified differences in overall outcome, as well as those limited to a distinct subgroup. Comprehensive annotation of cancer patients will facilitate a deeper understanding of the genetic distinctions between subpopulations, enable the identification of more refined subcategories of NB, and lead to improved discovery of novel therapeutic targets.

Patient stratification also occurs based on genetic patterning. For example, HR disease is subdivided by the presence or absence of *MYCN* amplification, referred to as *MYCN*-amplified(*M-Amp*) and *MYCN* non-amplified (*M-Non) disease. MYCN* status is associated with other common chromosomal aberrations, such as deletions of 1p, 11q, 14q, and 17q gain. *MYCN* mRNA is targeted by the *let-7* tumor suppressor microRNA family, which consists of nine *let-7* family members spread across multiple genetic loci[22]. We previously showed that high levels of *MYCN* mRNA in *M-Amp* disease turn the tables on *let-7*, where the 3' UTR of *MYCN* mRNA efficiently sequesters *let-7* and inhibits its function[8,23]. This discovery also explained the longstanding observation that chromosome (Chr) 11q, known to harbor the most highly expressed *let-7* locus (*let-7a-2*), is frequently lost in *M-Non* but not in *M-Amp* disease. This work established that increased levels of Chr 11q-derived *let-7a* are functionally impaired by *MYCN* mRNA. This work was the first to demonstrate that noncoding elements within a protein-coding mRNA can independently contribute to both oncogenicity and selective genetic patterning in cancer through microRNA sequestration.

Here, we assessed potential sex and race biases in NB. We discovered a striking sex bias in *M-Non*/HR NB patient outcomes that associates with deregulation of two known oncogenes, *H19* and *DLK1* [24–27]. Biased outcomes were also identified in *M-Non* stage 4s disease, as well as an unexpected sex bias negatively affecting male African American patients. We identified significant differences in chromosomal loss patterns between female and male *M-Non*/HR NB patients using a novel analytic approach to differential gene expression. Finally, we connected newly identified sex-biased outcomes with chromosomal loss patterns and *let-7* sequestration by *H19*, expanding on our previous report on the intersection between *MYCN* expression, *let-7* inhibition, and chromosomal patterning in *M-Amp* disease[8,23].

Results

Female neuroblastoma patients have worse overall survival in High-Risk MYCN Non-Amplified disease.

Overall, NB incidence and the occurrence of HR disease are slightly higher in males than in females[3,4,28]. Despite this disparity in frequency, an in-depth analysis of female *vs.* male outcomes in HR disease has yet to be reported. We, therefore, first assessed potential sex bias in multiple NB clinical subsets through analysis of a 498-patient cohort (SEQC) with sufficient annotation to simultaneously interrogate risk stratification, *MYCN* status, and patient sex[29]. No difference in overall survival (OS) between the sexes was observed in this and two additional datasets (TARGET, Kocak) when risk status was not considered (***Extended Data Table 1, Extended Data Fig. 1a, 1b, 1c***)[30,31]. SEQC patients with LR disease also exhibited no significant difference in survival between males and females (***Figure 1a***, top right panel). However, among HR patients, females displayed a striking pattern of significantly worse OS (*p = 0.0075*, ***Figure 1a***, top left). Analysis of the *M-Amp* and *M-Non* subgroups within HR disease revealed significantly worse OS in female *M-Non*/HR patients (*p = 0.021*), as opposed to no difference in outcome for females and males in the *M-Amp*/HR subgroup (***Figure 1a***, lower right and left panels, respectively).

We subsequently sought to validate this finding within additional datasets. The Kocak NB patient study has annotations for *MYCN* status, patient age, stage, and sex but is not risk status (***Extended Data y Table 1***). As a result, we considered stage 3 or stage 4 patients older than 18 months (approximation of HR disease) [4,4]. compared and again revealed that higher-risk, late-stage female NB patients had significantly worse OS (*p = 0.0037,* **Extended *Data Fig. 2a***). We then assessed female and male OS in the TARGET NB study, which is exceptionally well-annotated (***Extended Data Table 1***). Although an initial comparison of HR patients yielded no difference in outcomes between female vs. male patients or when risk was considered, we reasoned that this may be a result of inherent differences in the types of patients recruited for the study (***Extended Data Fig. 2b***). Whereas the Germany-based Kocak study has a strong Caucasian patient bias (*personal communication*), African American (AA) patients represent the second largest group in the TARGET study (over 12% of subjects). Thus, we reasoned that race may have contributed to our failed validation of female sex bias in the TARGET study. Upon exclusion of AA patients from our TARGET study analysis, we once

again observed the same pattern of significantly worse OS in female *M-Non*/HR patients as the SEQC study (***Extended Data Fig. 2c***). However, when analyzed in isolation, the AA patient group (n = 31) exhibited an inverse pattern, where male HR patients experienced dramatically worse OS (93% DFD) than females (***Figure 1b***).

This striking yet opposite-sex bias in OS led us to evaluate other subgroups within NB. In particular, analysis of *M-Non* stage 4s patients, whose prognosis is overall favorable, also revealed a significant difference in OS between females and males. When analyzed individually, females in the SEQC and Kocak cohorts had significantly worse OS than male *M-Non* stage 4s patients (*p = 0.019, 0.021*, respectively, ***Extended Data Fig. 3a, 3b***). A third study revealed a similar trend but did not achieve significance (*p = 0.17*, **Extended *Data Fig. 3c***). When patient data from these three studies was combined into a single Kaplan-Meier analysis, a strong sex bias emerged where females performed much worse than male 4s patients (*p = 0.00076*, **Figure *1c***). In fact, the only patients who died within this group were female (8/49 patients), while there were no male deaths (0/85 patients) (***Figure 1c, Supporting Data file 1***). This is surprising given that stage 4s disease has an excellent prognosis, with over 90% survival and close to 50% spontaneous regression[32]. Our results suggest a near 100% survival rate in *M-Non* stage 4s male patients, compared to 16.4% death in females (***Figure 1c***). These findings led us to combine HR patients from the SEQC, Kocak, and TARGET (AA patients excluded) studies to increase sample size and power for the previously identified sex bias in *M-Non*/HR disease. From this analysis, we observed an increased statistical significance associated with adverse female-biased OS in HR disease (n = 461 patients, $p = 1.3 \times 10^{-5}$, ***Figure 1d, Supporting Data file 1***) and a stronger representation of sex-biased OS in female *M-Non*/HR disease (n = 271, $p = 4.3 \times 10^{-4}$, ***Figure 1e, Supporting Data file 1***).

H19 and DLK1 are associated with poor survival in female, but not male, neuroblastoma patients.

Differential gene expression analysis of female (n = 14) vs. male (n = 26) *M-Non*/HR/DFD patients (SEQC) indicated that the top five hundred genes among female patients from this group are strongly shifted towards female enriched expression (***Figure 2a***, right panel). In contrast, this set of genes did not exhibit enrichment among female patients in *M-Amp*/HR and *M-Non*/LR living clinical subgroups, indicating female *M-Non*/HR/DFD patients may have a distinct gene expression signature (***Figure 2a***, middle and left panels, respectively). Among the top twenty genes upregulated in female *M-Non*/HR/DFD patients, the long noncoding RNA *H19* and delta-like non-canonical Notch ligand 1 (*DLK1*) emerged as the most upregulated non-Chr X genes (***Figure 2a***, right panel, ***Extended Data Fig. 4a*** left panels, ***Supporting Data file 2***). Further, *H19* and *DLK1* expression is significantly correlated in females (r = 0.52, p = 0.019) but not male *M-Non*/HR patients, suggesting a degree of co-expression in female patients (***Figure 2c, Supporting Data file 2***). A similar analysis of HR-approximated patients in the Kocak study again showed that *H19* and *DLK1* are significantly upregulated in female patients (***Figure 2b, Extended Data Fig. 4a*** right panels, ***Supporting Data file 2***). Further, despite very small patient numbers (n = 5 female, 6 male), a comparison of *H19* and *DLK1* expression in *M-Non* stage 4s patients with progressive disease shows higher, although not significantly so, expression in female patients (***Extended Data Fig. 4b***). Of note, expression of *H19* and *DLK1* in the adrenal medulla is among the highest for all tissues (***Extended Data Fig. 5***), introducing the possibility that high *H19* and *DLK1* expression in female *M-Non*/HR patients may represent sex-specific, retained expression from a common NB tissue of origin. Cox-regression analysis of OS Hazard Ratios of the top twenty female-enriched non-X Chr genes reveals that *DLK1* and *H19* (and its embedded microRNA, *miR-675*) are significantly associated with worse outcomes in only female patients (***Figure 2d, Supporting Data file 2***). This association was validated through Kaplan-Meier analysis of *M-Non* patients in the SEQC and Kocak studies (***Extended Data Fig. 6a, 6b***). No association with female or male outcomes was noted in *M-Amp* disease patients from either study (***Extended Data Fig. 7***).

H19 and *DLK1* have been implicated in multiple cancers[24–27]. To assess their ability to promote NB tumor growth, we over-expressed both in murine 3T3 and human SH-SY5Y NB cells. Expressing *H19* as either an intact, mature lncRNA sequence (*H19*) or as a full-length genetic locus version containing endogenous introns (*H19-GL*) in both cell types resulted in higher growth rates compared to GFP control cells (***Figure 2e***). Similar results were observed with *DLK1* overexpression in 3T3 and SHSY-5Y (***Figure 2e***). Opposingly, we used *H19 and DLK1* small interfering RNAs (siRNAs) individually or in tandem in *M-Non*, SH-SY5Y cells and noted significant growth inhibition with knockdown (***Extended Data Fig. 8, Supporting Data file 3***). We next used a CRISPR/Cas9-mediated gene disruption growth assay modeled on classical large-scale dropout screens to disrupt *H19* and *DLK1* genetically. Both SHSY-5Y or SK-N-F1 cells displayed a

profound reduction in fluorescence driven by positive control *RPL3* gRNAs (***Figure 2g, Supporting Data file 2***). Similarly, *DLK1* and *H19* targeting gRNAs resulted in significant fluorescence reduction, validating our siRNA results. These data suggest that *H19* and *DLK1* drive more aggressive tumor cell growth, thereby contributing to the poor outcomes we report in *M-Non*/HR female patients.

Differential gene expression analysis reveals striking sex-biased chromosomal loss patterns.

When we noted that the genomic location of the top 500 genes significantly increased in female *M-Non*/HR patients, we identified a strong enrichment for genes residing in the q arm of Chr 11 (15.8%). We then assessed enrichment by chromosomal location for Chr that represented more than 10% of the top 500 genes. Female-biased gene expression was also noted for Chr 4 (10.8%) and Chr 12q (10.2%) (***Figure 3a,*** *bottom panel,* ***Supporting Data file 4***). This prompted us to examine the chromosomal distribution of male-enriched genes, where we observed an overrepresentation of Chr 1p (10.6%) and Chr 19 (11.4%). (***Figure 3a,*** *top panel,* ***Supporting Data file 4***). Thus, we reasoned that these enrichment patterns may suggest actual chromosomal patterning differences between female and male *M-Non*/HR/DFD patients. To examine this directly, we first assessed differential expression patterns in *M-Amp* vs. *M-Non* NB patients, where chromosomal patterns are well established. First, looking at Chr 13, not known to have a differential Chr loss signature in *M-Amp* vs. *M-Non* NB, we observed an expected lack of significant difference in gene expression by volcano plot and paired t-test (***Figure 3b***). Chr 1p, however, known to be lost in almost all *M-Amp* disease [6,33], displayed a significant shift of gene expression toward the *M-Non* group, which is consistent with the known Chr 1p loss pattern ($p = 8.4 \times 10^{-100}$, ***Figure 3c, Supporting Data file 4***). Similarly, Chr 11q, known to be frequently lost in *M-Non* disease[5,6,8,34], exhibited an opposite significant shift in gene expression towards *M-Amp* disease, again consistent with known chromosomal patterning ($p = 0.0028$, ***Figure 3d, Supporting Data file 4***). This analysis shows that we can recapitulate known chromosomal patterning between patient groups, allowing us to use this method to investigate novel chromosomal abnormalities.

Chr 13 genes were not enriched in either female or male patients (1% and 0.6%, respectively (***Figure 3a, Supporting Data file 4 3),*** and we observed no significant sex bias in gene expression in *M-Non*/HR patients from the SEQC study or in stage 4 older than 18-month patients from the Kocak study (***Figure 3e***). Next, we examined Chr 1p and observed a significant shift toward male patients in the SEQC and Kocak study patient groups ($p = 1.8 \times 10^{-34}$, 2.3×10^{-5}, *respectively,* ***Figure 3f, Supporting Data file 4***). Further, analysis of Chr 11q genes showed a significant shift in gene expression towards female patients in both studies ($p = 2.9 \times 10^{-33}$, 1.1×10^{-17}, *respectively,* ***Figure 3g, Supporting Data file 4).*** Other chromosomal enrichment patterns identified across Chr 4, Chr 12q, and Chr 19 (***Figure 3a***) also showed significant sex-biased shifts (***Extended Data Figure 9, Supporting Data file 5***). These results demonstrate for the first time that sex-biased patterns of chromosomal loss exist in NB. While this analysis does not allow for de novo determination of directionality for a given chromosomal imbalance, Chr 1p and 11q aberrations are well-known in NB. Thus, the patterns we observed are likely due to chromosomal arm loss within the deficient group[5,6,34–37]. In addition, Chr 4[35,36,38] and Chr 12q[39–41] loss have been reported to be associated with poor outcomes in NB, which is consistent with the female-biased poor OS we observed in M-Non/HR disease. Whole Chr 19 gain[35] has been reported to be associated with better outcomes, suggesting that we may have identified a pattern of Chr 19 gain in males as opposed to Chr 19 loss in females. These observations show that we have identified significant and novel sex biases within known chromosomal imbalances in NB.

Finally, we examined the local genomic regions near both *DLK1* and *H19*. *We observed* that the Chr 14q region near the *DLK1* locus is associated with a female-enriched shift, suggesting a genetic dose difference favoring females for that region (***Extended Data Figure 10a, Supporting Data file 6***). *H19*, however, showed no significant shift in either direction (***Extended Data Figure 10b Supporting Data file 6***). This discrepancy suggests that *DLK1* elevation in females may be due, at least in part, to differences in gene copy number. In contrast, elevated *H19* expression may be through a more direct mechanism in females.

H19 inhibits let-7 in female M-Non/HR patients in a manner similar to MYCN mRNA in M-Amp disease.

Despite their differences in *MYCN* status, female *M-Non*/HR/DFD patients appear to have a similar Chr 1p/Chr 11q chromosomal pattern to *M-Amp* patients. To explore this further, we repeated our differential chromosome-specific gene

expression analysis of Chr 13, Chr 1p, and Chr 11q in male and female NB patients. *M-Amp* male patients displayed the expected pattern, revealing loss of Chr 1p and retention of Chr 11q compared to the *M-Non* group (*Figure 4a*, upper panels, *Supporting Data file 7*). Female patients showed similar patterns for Chr 13 and Chr 1p but diverged for Chr 11q, showing a mildly significant shift towards *M-Non* patients. This suggests that female *M-Non* patients may indeed retain Chr 11q more than male patients, as we demonstrated in *Figures 3f* and *3g*, and further supports the novel idea that *M-Non*/HR females may display a classically *M-Amp* loss pattern for Chr 1p and Chr 11q.

We previously established that in *M-Amp* NB, *MYCN* mRNA, which is a *let-7* microRNA target (*Figure 4b*), sequesters *let-7* in *M-Amp* disease. Thereby mitigating its function and relieving the selective pressure to lose Chr 11q, which houses one of the highest expressed *let-7* loci, *let-7a-2*[8] (*Figure 4b, Extended Data Figure 11a, 11b*). This work provided a mechanism by which Chr 11q is retained in *M-Amp* NB but preferentially lost in *M-Non* disease, establishing that noncoding elements within a protein-coding mRNA can have autonomous oncogenic function with genetic consequences. Our current observation, where female M-Non/HR patients have a Chr 11q pattern that matches classical M-Amp patterning, may have consistencies with this original discovery. *H19*, which is highly expressed in these female patients (*Figure 2a, Extended Data Figure 4*), is known to contain four *let-7* sites (*Figure 4d*)[7] and has been reported to sequester *let-7*[42–45]. *H19* shares several characteristics in NB with *MYCN*, a known *let-7* sponge. As expected, *MYCN* correlates with *let-7* levels across *M-Amp* and *M-Non* NB cell lines (*Figure 4c, left panel, Supporting Data file 7*)[8]. Similarly, *H19* correlates with *let-7* levels in *M-Non* NB cell lines (*Figure 4c, right panel*). This relationship holds concerning the *let-7a-2* host gene, *MIR100HG*, in human NB patients, where *MYCN* levels correlate with *MIR100HG* in HR NB patients (*Figure 4b* and *4d, left panel*). *H19* levels also positively correlate with *MIR100HG* expression in *M-Non* HR disease, but only in females (*Figure 4d, right panel*). This suggests that *H19* may play a role in *let-7* inhibition in only female *M-Non*/HR patients, which unites the sex-based disparity in OS with the Chr 11q (*let-7a-2*) patterning we observed.

To test this more directly, we examined the expression levels of a representative set of *let-7* targets in *H19* and *H19-GL* expressing SHSY-5Y cells. Compared to GFP, *H19*-expressing cells all showed significantly higher expression of every *let-7* target tested (*Figure 4e, Supporting Data file 7*). *H19-GL* expressing cells displayed a similar pattern, with all but three primer sets (*LIN28B*, *ZBTB10*, and *SKIL*) registering significantly higher *let-7* target expression (*Figure 4e, Supporting Data file 7*). In general, *H19-GL* resulted in lower levels of *let-7* target enrichment, consistent with lower levels of mature *H19* in *H19-GL* vs. *H19* expressing SHSY-5Y cells (*Extended Data Figure 11c, Supporting Data file 8*). Consistent with these in vitro results, female *M-Non*/HR/DFD NB patients have significantly enriched expression of *let-7* target mRNAs containing two or more *let-7* sites (*n = 133 let-7 targets*, *Extended Data Figure 11d, Supporting Data file 8*). Next, we examined the effect of H19 depletion on *let-7* target mRNA. Using two different siRNAs, we knocked down *H19* in CHLA-42 *M-Non* NB cells (*Extended Data Figure 12a*); we observed significantly decreased expression of all nine *let-7* target mRNAs (*Extended Data Figure 12b, Supporting Data File 9*). These results suggest that *H19* serves to sequester and inhibit *let-7* in female *M-Non*/HR/DFD patients, similar to how *MYCN* sequesters relatively high *let-7* levels in *M-Amp* disease[8]. They similarly provide a plausible rationale for worse female outcomes, mirroring the impact of *M-Amp*-specific *MYCN* mRNA-mediated *let-7* inhibition. This novel role for *H19* in female NB patients provides a selective mechanism by which these female patients tend to retain Chr 11q, a pattern previously only associated with *M-Amp* patients.

Discussion

Neuroblastoma (NB) is the most common pediatric, extracranial solid tumor [9,10]. Although some subgroups, such as LR, IR, and 4S disease, exhibit a positive outlook, HR disease remains difficult to treat and has an OS rate below 50%[15,16]. A genetically heterogeneous malignancy, NB severity highly depends on multiple factors that affect risk and prognosis. Prior to this work, male and female HR NB outcomes had not been directly investigated. Understanding the origins and driving mechanisms behind the sex-based disparity in OS described here may lead to an improved understanding of female-specific cancer genetics and sex-specific approaches to therapy, a first for NB. While primarily male-biased, worse outcomes have been reported in over twenty cancer types[46,47], the majority of these disparities were identified by looking at all patients within a given cancer[47]. Given the results of our own compartmentalized analysis, we predict that sex biases are more widespread in human cancers, that these biases will be more divergent in some patient subgroups, and that the directionality may change depending on the subgroup in question. The results of our analysis provide a

rationale for widespread sub-group analysis across most if not all, tumor types. Important differences within patient subsets can be hidden within all-patient OS analyses. Therefore, it is imperative to consider tractable patient subgroups when analyzing patient outcomes. In addition, to facilitate such subgroup-focused analysis, we must strive to maximize annotation when performing large-scale patient studies. Of eight patient datasets available for possible outcomes analysis on the r2 database, only four listed patient sex within their available annotations[48]. In contrast, the TARGET study with only 249 patients was annotated to the degree that it included patient race, allowing us to discover an additional and opposite-sex bias in African American patients (*Figure 4*).

Both *H19* and *DLK1* have been implicated as oncogenes in multiple cancers[24–27,49–53]. *H19* is a long non-coding RNA with multiple functions, including regulation of gene expression and microRNA sequestration[49,54]. *DLK1*, a transmembrane protein involved in the highly conserved NOTCH signaling pathway, can function as a non-canonical NOTCH ligand to exert control over cell-to-cell communication, proliferation, and differentiation[24,25]. We found similar and robust pro-growth functions for both *H19* and *DLK1* in murine 3T3 embryonic fibroblasts and in human *M-Non* SHSY-5Y NB cells (*Figure 2e, 2f, 2g*, and *Extended Data Figure 8*). The local organization of the *H19* (*h*) and *DLK1* (*d*) loci are similar. They both reside in imprinted regions, and both loci are comprised of a protein-coding gene (*DLK1*(*d*) or *IGF2*(*h*), both imprinted), followed by long noncoding RNA (*MEG3*(*d*) or *H19*(*h*), both imprinted), followed by another protein-coding gene (*RTL1*(*d*) or *MRPL23*(*h*))[55,56]. These similarities suggest the possibility of shared regulation.

NB is a disease of genetic compartmentalization, both in terms of expression patterns of key genes and repeating patterns of chromosomal gains and losses. We identified a surprising sex disparity in chromosomal imbalance across multiple chromosomes, including Chr 1p and Chr 11q (*Figure 3a, 3f,* and *3g*). This observation revealed an unexpected alignment between female *M-Non*/HR and classical *M-Amp* patterning, as both groups appear to lose Chr 1p and retain Chr 11q. This pattern was validated upon sex-directed analysis of *M-Amp* vs. *M-Non* NB patients (*Figure 4a*). Our previous work established that in *M-Amp* NB, *MYCN* mRNA, which contains two *let-7* sites, efficiently sequesters *let-7* and inhibits its function[8,23]. This work provided the first plausible explanation that Chr 11q, which harbors the most highly expressed *let-7* locus (*let-7a-2*), is frequently lost in *M-Non*, but not in *M-Amp* disease. We previously demonstrated that noncoding elements with an oncogenic mRNA can independently contribute to both tumor pathology and selective genetic patterning in cancer[8]. The alignment between female *M-Non*/HR and *M-Amp* patterns led us to discover that *H19*, known to have four *let-7* sites[7], sequesters *let-7* in female *M-Non*/HR patients in a manner mirroring that of *MYCN* in *M-Amp* disease (*Figure 4, Extended Data Figure 11d, 12*). This observation provides a plausible rationale for the female retention of Chr 11q in *M-Non*/HR disease. The mechanistic similarity of noncoding *H19* and *MYCN* mRNA function validates our earlier work. It suggests that sequestration of *let-7* by either 3'UTRs of protein-coding mRNAs or *let-7* sites within long noncoding RNAs may be a fundamental mechanism of *let-7* mitigation in NB and other cancers.

We show here significant sex- and race-based disparities in OS relative to *MYCN* status in NB through analysis of multiple publicly available patient datasets with good to excellent patient annotations. This discovery provides a novel way to consider how incidence and genetic patterning intersect with disease pathology and patient outcomes in a genetically and clinically heterogeneous disease like NB. Our findings demonstrate the value of highly compartmentalized cohort analysis of large, well-annotated patient datasets. Further, given the frequency and consistency of multiple genetic events in NB, these findings establish a novel paradigm of sex-biased outcomes and chromosomal patterning in NB and solidify the centrality of *let-7* mitigation, which has broad implications for human malignancy.

Methods

R2 database: Human NB patient RNA-seq, microarray, and overall survival data sets were obtained from the r2: microarray analysis and visualization platform (http:// r2.amc.nl)[48]. The following data sets were used for analysis: SEQC498 (GEO accession number GSE62564)[29], Kocak649 (GEO accession number GSE45547)[30], OBERTHER251 (E-TAM-38) **OBERTHUER 251**, TARGET249 (r2 internal identifier: ps_avgpres_targetnrbl249_huex10t); results from analysis of the TARGET249 dataset are based upon data generated by the Therapeutically Applicable Research to Generate Effective Treatments (https://www.cancer.gov/ccg/research/genome-sequencing/target) initiative (phs000467)[31].

Secondary analysis: Gene expression graphs, volcano plots, Cox regression analysis, and Kaplan–Meier curves analysis of human NB patients were generated using the data analysis tool suite on the r2: Genomics Analysis and Visualization Platform. Data sets used were listed in the R2 database section of this document. HR NB patients from the Kocak 649 (GSE45547) study were identified as patients with stage 4 disease, >18 months at diagnosis, and patients of any age and stage with *M-Non* tumors. Shifts in differentially expressed chromosome-specific genes were represented as volcano plots for each chromosome. Volcano plots of chromosomal gene expression changes were generated using gene lists from the UC Santa Cruz genome browser. Expression patterns of these gene lists were then obtained from both SEQC-498 (GSE62564) and Kocak 649 (GSE45547) studies on the r2 platform for NB subgroup comparisons.

Cell Culture: The neuroblastoma cell lines, including CHLA-15 (RRID CVCL_6594), CHLA-20 (RRID CVCL_6602), KAN (SMS-KAN, RRID CVCL_7131), KANR (SMS-KANR, RRID CVCL_7132), KCN (SMS-KCN, RRID CVCL_7133), KCNR (SMS-KCNR, RRID CVCL_7134), and SKNF1 (SK-NF-1, RRID CVCL-1702), were obtained from the Childhood Cancer Repository at the Texas Tech University Health Science Center, a partner of the Children's Oncology Group. Additionally, BJ fibroblasts (NHF, ATCC CRL-2522), SH-SY5Y (ATCC CRL-2266), and BALB/3T3 (ATCC CCL-163.2) were acquired from ATCC. All cell lines were cultured in RPMI-1640 media supplemented with 10% heat-inactivated fetal bovine serum (HI-FBS). Cells were incubated at 37°C under 5% CO_2 and routinely tested negative for mycoplasma contamination. All cell lines utilized are not considered commonly misidentified.

RNA-Seq: Total RNA was submitted to LC Sciences (Houston, TX, USA) for library prep and sequencing. Sequencing (without spike-ins) was performed at a depth of 40 million 150 bp paired-end reads. Reads were normalized as Fragments Per Kilobase of transcript per Million mapped reads (FPKM). RNA-Seq data sets are (will be) available under the GEO reference series GSE). CHLA-15, CHLA-20, KAN, KANR, KCN, KCNR, CHLA-42, and SKNF1 were submitted for sequencing in biological duplicate.

Plasmids and Stable Transfection Protocol: Control *GFP* (pPB[Exp]-Puro-EF1A>EGFP); *H19* pPB [ncRNA]-EGFP-EF1A>hH19[NR_002196.2]; *H19-GL*: pBP[ncRNA]-EGFP-EF1a>{h19 full length NR_002196.2}; and *DLK1* (pPB[Exp]-mCherry-SFFV>hDLK1[NM_003836.7]) vectors were designed and purchased from vectorbuilder.com. BALA/3T3 and SH-SY5Y cells were co-transfected with both 1 µg of desired plasmid and 0.5 µg of hyPBase transposase-expressing vector (pRP[Exp]-mCherry-CAG>hyPBase), following the Lipofectamine™ 3000 (Invitrogen™) manufacturer protocol for a 6-well plate. Similarly, SH-SY5Y and SKNF1 cells were stably transfected with a Cas9-expressing Piggybac plasmid (pPB[Exp]-TagBFP2/Neo-hPGK>hCas9) and a transposase plasmid (pRP[Exp]-mCherry-CAG>hyPBase) as described above. Stably transfected cells were then isolated by fluorescence-activated cell sorting (FACS).

Cell Growth: SH-SY5Y and BALB/3T3 cells expressing *GFP*, *H19*, *H19-GL*, or *DLK1* were seeded in 96-well plates with serum depleted medium (2.5% HI-FBS). The WST-8 cell proliferation assay kit (Cayman Chemical) was used to determine cell growth at 24-, 48-, and 72-hour time points. Growth rates were calculated relative to the 24-hour time point. For assays evaluating loss of *H19* or *DLK1*, SH-SY5Y cells transfected with siRNAs (as described below) were assayed for relative growth at 48 hours post-transfection. Cas9-expressing SH-SY5Y or SK-N-F1 were transfected with relevant gRNA constructs (as described below), and at 24 and 72 hours post-transfection, the level of fluorescence was measured. For all assays in the 96-well format, a seeding density of 5000 cells/well was used.

Let-7 Target enrichment: SH-SY5Y cells expressing *GFP*, H19, or *H19-GL* were plated in 6-well plates. After 48 hours, cells were harvested, and their derived cDNA was used in RT-qPCR (SYBR green system). Relative gene expression of *let-7* targets was determined using the $2^{-\Delta\Delta Ct}$ method. Primer sequences were available in Supplement Data File 10.

Short interfering RNA (siRNA) Transfection: SH-SY5Y cells were seeded in 96-well plates and transfected with 1pmol siRNA according to Lipofectamine™ RNAiMAX transfection reagent (Invitrogen) manufacturer protocol at 48 hours post-transfection relative growth rate was measured. Also, and CHLA-42 cells were seeded in 6 well plates and transfected with 25pmol H19 siRNAs according to Lipofectamine ™ RNAiMAX transfection reagent (Invitrogen) manufacturer protocol. Forty-eight hours post-transfection, cells were harvested, and RNA was isolated using the TRIzol™ reagent. Subsequently, cDNA was synthesized using the Verso™ cDNA kit. For all assays in 6 well format, a seeding density of 5×10^5 cells/well. All of the siRNAs were acquired from IDT™, Control siRNA sense: CGUUAAUCGCGUAUAAUACGCGUAT, antisense: AUACGCGUAUUAUACGCGAUUAACGAC; DLK1 siRNA 13.1 sense: CUCUUAAUGCAUGAUACAGAAUAAT, antisense AUUAUUCUGUAUCAUGCAUUAAGAGAG; DLK1

siRNA 13.2 sense: AAAAUGGAUUCUGCGAGGAUGACAA, antisense UUGUCAUCCUCGCAGAAUCCAUUUUGG; H19 siRNA 13.2 sense: AAAAUGGAUUCUGCGAGGAUGACAA, antisense UCCAGAGCCGAUUCCUGAGUCAGGUAG; H19 siRNA 13.3 sense : CAAGCAUUCCAUUACGCCCCAUCTC, antisense GAGAUGGGGCGUAAUGGAAUGCUUGAA.

CRISPR/Cas-9 gene knockout: Cas9-expressing SH-SY5Y and SK-N-F1 *M-Non* NB cells were transfected with a single guide RNA (gRNA) constructs co-expressing either GFP and a single non-targeting gRNA or mCherry and single gRNAs targeting *RPL3*, *DLK1*, or *H19*. A third *H19*-targeting gRNA directly targets *miR-675* located within the first exon. Specifically, Cas9-expressing SH-SY5Y and SKNF1cells (100,000 cells/well) were transfected with 1 μg of appropriate gRNA vectors in 6-well plates on day 0. Cells were analyzed for percent fluorescence on days 1 and 3, and relative fluorescence on day 3 vs. day 1 was used to determine growth differences. Guide RNAs were designed as previously described[8]; sequences are available in Supplement Data File 10.

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Novel sex-biased outcomes in neuroblastoma are associated with distinct gene expression and chromosomal loss patterns.

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FIGURES

Figure 1

a.

Figure 1. Identification of broad sex-biased outcomes in neuroblastoma. a) Kaplan-Meier (KM) analysis of overall survival (OS) in male vs. female neuroblastoma (NB) patients relative to *MYCN* and risk status. Center bar represents NB patients from the SEQC 498 dataset; HR (n=176), LR (n=322), *M-Amp*/HR (n=92), and *M-Non*/HR (n=83). Data source: SEQC 498 (GSE62564). **b)** KM analysis of OS in female and. male African American high-risk//unfavorable histology NB patients. Data source: TARGET 249 (R2, Asgharzadeh). **c)** KM analysis of OS in male vs. female *M-Non*/stage 4s NB patients. Combined data source: SEQC 498 (GSE62564), Kocak 649 (GSE45547), Oberthuer 251 (E-TABM-38). **d)** KM analysis of OS of male vs. female HR NB patients. Combined data source: SEQC 498 (GSE62564), Kocak 649 (GSE45547; approximation of high-risk constituting stage 3 and 4 patients >18 months old), TARGET 249 (R2, Asgharzadeh). **e)** KM analysis of OS of male vs. female *M-Non*/HR NB patients. Combined data source: SEQC 498 (GSE62564), TARGET 249 (R2, Asgharzadeh), Kocak 649 (GSE45547). All *p* values were determined by log-rank test.

Figure 2

Figure 2. *H19* and *DLK1* expression differences in male vs. female neuroblastoma patients. **a)** X-Y plot of differential expression analysis of male vs. female NB patients. Red dots = top 500 genes most significantly upregulated in LR/*M-Non*/alive patients (left); HR/*M-Amp*/DFD patients (center); HR/*M-Non*/DFD patients (right). Data source: SEQC 498 (GSE62564). **b)** X-Y plot of differential expression analysis of male vs. female st4/*M-Non* NB patients. Red dots = top 500 significantly upregulated genes in (2a). Data source: Kocak 649 (GSE45547). **c)** X-Y plot of gene expression correlation between *H19* and *DLK1* expression levels in male and female HR/*M-Non*/DFD patients. Green line, f = females, red line, m = males. p-values determined by ANOVA. Data source: SEQC 498 (GSE62564). **d)** Cox proportional hazard regression analyses for female vs. male *M-Non* NB patients (as labeled). Selected genes are most significantly upregulated non-X Chr-associated genes in *M-Non* NB patients. Extended lines from central dots indicate 95% CI. Blue line = significant correlation > 1.0, red line = significant correlation < 1.0. Data source: SEQC 498 (GSE62564). **(e)** and **(f)** Colorimetric growth assays on 3T3 and SHSY-5Y cells relative to GFP construct-based expression of *H19, H19-GL* and *DLK1*. Significance determined by two-way ANOVA, followed by Dunnett's multiple comparison test; bars represent average +/- SEM; *p<0.05, **p<0.01, <0.001, <0.0001. **g)** CRISPR/Cas9 mediated gene knockout of *H19* and *DLK1* in SHSY-5Y and SK-N-F1 cells. Cells were transfected on d0 with control, *DLK1* and *H19* gRNAs. Loss of *H19* and *DLK1* targeted with gRNAs against negative control gRNA were measured at d1 and d3 via flow cytometry. Significance determined by two-way ANOVA, followed by Sidak's multiple comparison tests; bars represent mean+/-SEM; *p<0.05, **p<0.01, <0.001, <0.0001.

Figure 3

Figure 3. Chromosome-associated differences in gene expression in male vs. female neuroblastoma patients.
a) Chromosomal location of the 500 most highly expressed genes in female vs. male HR/DFD NB patients. Red bars: >20 genes present on respective chromosomes. Data source: SEQC 498 (GSE62564, n=40, composed of 14 females and 26 males). **b), c), d)** Volcano plot of Chr 13, Chr 1p, and Chr 11q genes expression in *M-Amp*/HR and *M-Non*/HR NB patients (n=91). Data source: SEQC 498 (GSE62564) **e), f),** and **g)** Volcano plot of Chr 13, Chr 1p, and Chr 11q genes expression in female and male NB patients. Data source: SEQC 498 (top plots; GSE62564; *M-Non*/HR/DFD subset; n=40) and Kocak 649 (bottom plots; GSE45547; st4 patients >18 months old subset; n=94).

Figure 4

Figure 4

Figure 4. *H19* is strongly associated with let-7 expression in neuroblastoma. **a)** Volcano plots of chromosomal-linked gene expression HR dead from disease NB patients stratified by *MYCN* status and sex. Top row: plots of male *M-Non* (n=26) and *M-Amp* patients (n=26) for stated chromosomal regions; Bottom row: plots for female, *M-Non* (n=14) vs. *M-Amp* (n=25). Yellow dots: genes stemming from specified chromosomes in male or female; red box over chromosome = identified relative chromosome deficiency. Data source: Data source: SEQC 498 (GSE62564). **b)** Schematic of *MYCN* mRNA and *H19* lncRNA *let-7* sites. Green triangles = *let-7* miRNA target sites. Bottom: Location of let-7a-2 host gene (MIR100HG) on chromosome 11. **c)** Correlation plots of *MYCN* vs. *let-7* expression in human *M-Non* and *M-Amp* NB cell lines (n=16; left) and *H19* vs. *let-7* expression in human *M-Non* NB cell lines (n=8; right) were derived from NGS. Data sets will be available under the GEO reference series GSE (http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE (TBD1, TBD2)). p-values were derived by Pearson Product Moment Correlation (PPMC) from correlation coefficient (*r*). **d)** X-Y plots of gene expression correlation between *MYCN* and *MIR100HG* (left) in HR NB patients (left), *H19* and *MIR100HG* in *M-Non*/HR NB patients (right). Green line, f =females; red line, m =males. Data source: SEQC 498 (GSE62564; n=176); *p<0.05, **p<0.01; p-values determined by ANOVA. **e)** Expression levels of *let-7* target genes in *M-Non* SHSY-5Y cells expressing *H19* mature or full-length genetic locus (*H19-GL*). Significance determined by two-way ANOVA, followed by Sidak's multiple comparison tests; bars represent average +/- SEM; *p<0.05, **p<0.01, ***<0.001, ****<0.0001.

Extended Data Table 1

Subject	Annotation	SEQC 498	KOCAK 649	TARGET 249	OBERTHUER 251
	Total Patients	498	649	249	251
Gender	Male	287	344	143	146
	Female	211	268	106	105
	Unknown	-	37	-	-
Age	< 18 months	299	414	31	163
	> 18 months	199	235	218	85
	Unknown	-	-	-	3
MYCN status	Amplified	92	93	68	31
	Non-Amplified	401	550	175	220
	Unknown	5	6	6	-
Risk	Low-risk	322	-	30	-
	High-risk	176	-	217	-
	Unknown	-	649	2	251
Stage	1	121	153	30	68
	2	78	113	-	46
	3	63	91	1	39
	4	183	214	216	67
	4s	53	78	-	31
	Unknown			2	-
Histology	Favorable	181	-	37	-
	Unfavorable	91	-	183	-
	Unknown	226	649	29	251
Death	Yes	105	91	140	37
	No	393	385	107	214

Neuroblastoma patient demographics and available annotations from the studies used in this manuscript.

Extended Data Figure 1

Extended Data Figure 1. Overall survival probability of male and female neuroblastoma patients across datasets. OS in male and female NB from **a)** SEQC 498 (GSE62564) cohort, **b)** TARGET 249 (R2, Asgharzadeh) cohort, and **c)** Kocak 649 (GSE45547) cohort. P-values determined by log-rank test.

Extended Data Figure 2

Extended Data Figure 2. Overall survival of male and female neuroblastoma patients in additional patient studies. a) KM analysis of OS in male vs. female st 3 and 4 NB patients > 18 months old (approximating HR). Data source: Kocak 649 (GSE45547). **b)** KM analysis of OS in male and female *M-Amp*/HR, *M-Non*/HR neuroblastoma patients with unfavorable histology (UH). Subgroup determinants are detailed on y-axis for each graph; data source: TARGET 249 (R2, Asgharzadeh). **c)** KM analysis of OS in non-African American male and female HR, *M-Amp* and HR, *M-Non* neuroblastoma patients with unfavorable histology (UH). Subgroup determinants are detailed on y-axis for each graph; data source: TARGET 249 (R2, Asgharzadeh). All p-values were determined by the log-rank test.

b.

All Patients

High-Risk (HR) Unf. Hist. (UH)
male (100)
female (83)
p = 0.41

HR, UH MYCN-Amplified
male (28)
female (33)
p = 0.68

HR, UH Non-Amplified
male (72)
female (50)
p = 0.25

c.

Excluding African American Patients

High-Risk (HR) Unf. Hist. (UH)
male (74)
female (64)
p = 0.015

HR, UH MYCN-Amplified
male (22)
female (26)
p = 0.57

HR, UH Non-Amplified
male (52)
female (38)
p = 0.0098

Extended Data Figure 3

Extended Data Figure 3. Sex-biased outcomes in M-Non, stage 4s neuroblastoma. OS in male and female NB patients from **a)** SEQC 498 (GSE62564) cohort, **b)** Kocak 649 (GSE45547) cohort, and **c)**. Oberthuer 251 (E-TABM-38) cohort. p values determined by log-rank test.

Extended Data Figure 4

Extended Data Figure 4. *H19* and *DLK1* expression in female and male M-Non/HR patients. a) Boxplots of *H19* (top) and *DLK1* (bottom) expression in male (m; red dots) vs. female (f; green dots) patients from subsets from two datasets: SEQC 498 (left; HR/*M-Non*/DFD subset; GSE62564) and Kocak 649 (right; *M-Non*/st4 and >18 months old subset; GSE45547). *p<0.05, ***p<0.001; determined by ANOVA. **b)** Boxplots of *H19* and *DLK1* expression in male and female M-Non stage4s progressed disease patients: Data source SEQC 498. p values determined by ANOVA

Extended Data Figure 5

Extended Data Figure 5. ***H19*** **and** ***DLK1*** **expression levels in normal human tissues.** Expression Values are shown in Transcripts per Million (TPM). Box plots are shown as median and 25th and 75th percentiles; outliers ± 1.5x the interquartile range. Data Source: GTEx Analysis Release V8 (dbGaP Accession phs000424.v8.p2)

Extended Data Figure 6

Extended Data Figure 6. Overall survival in male and female *MYCN* non-amplified neuroblastoma patients relative to *H19* or *DLK1* expression. KM plots for males (top) and females (bottom) relative to expression (high or low) levels of **a)** *H19* or **b)** *DLK1* in *M-Non* NB. Data source: SEQC 498 (GSE62564) and Kocak 649 (GSE45547); source of the graph as listed on the respective figure. Red lines = low expression; blue lines = high expression; cutoff for high/low is average expression. p-values calculated by log-rank method.

Extended Data Figure 7

Extended Data Figure 7. _H19_ and _DLK1_ are not associated with overall survival in _MYCN_-amplified disease. KM analaysis of OS in two cohorts (as labeled) of male and female _M-Amp_ NB patients relative to **a,b)** _H19_ or **c,d)** _DLK1_ expression. In each panel, plots contain males + females (left), males only (center) and females only (right). Red line = low expression; blue line = high expression; high/low cutoff is average expression. Data source: SEQC 498 (GSE62564), Kocak 649 (GSE45547), R2; p-values calculated by log-rank method.

Extended Data Figure 8

Extended Data Figure 8. Growth effects of *H19* and *DLK1* knockdown in SHSY-5Y cells. Stated siRNAs were transfected on day 0 and assayed for growth (WST-8) 48 hours later.

Extended Data Figure 9

Extended Data Figure 9. Additional chromosomal-linked gene expression data from male and female high-risk NB patients dead from disease. Volcano gene expression plots of male vs. female HR, NB patients from **a)** Chr 19, bp 1 – q/ter, and **b)** Chr 4, bp 1 – q/ter and **c)** Chr 12q, bp 67e6 – q/ter. In all panels, green dots = genes stemming from specified chromosome; red box over chromosome = relative chromosome deficiency in neuroblastoma. Data source: SEQC 498 (GSE62564, n=40) and Kocak 649 (GSE45547, n=94), as labeled.

Extended Data Figure 10

Extended Data Figure 10: Volcano plots of genes located along **a)** Chr 14 50e6 -q/ter, and **b)** chr 11q 1 – 12e6 in male vs. female HR NB patients. In all panels, green dots = genes stemming from specified chromosome; red box over chromosome = relative chromosome deficiency in neuroblastoma. Data source as labeled on the graph: SEQC 498 (top; GSE62564, *M-Non*/HR/DFD subset; n=40) and Kocak 649 (Bottom; GSE45547, st4 patients >18 months old subset; n=94).

Extended Data Figure 11

Extended Data Figure 11. a) *Let-7* family member expression in the adrenal medulla, the tissue of origin for NB. Bars represent mean+/-standard deviation. Data source is GSE29742 including the following samples (n=6): GSM737418, GSM737419, GSM737420, GSM737421, GSM737422, GSM737423. p-values determined via Dunnett's T3 Multiple Comparisons Test with *let-7a* as control; *p<0.05, **p<0.01; **b)** Mean expression of *let-7* family members in NB. Data source: Bell (GSE155945, n=97); error bars represent SEM. **c)** *H19* expression in SHSY-5Y cells transfected with *H19 and H19-GL* plasmid constructs (see *Methods*). **p<0.01 **d)** Volcano plot of differentially expressed *let-7* target mRNAs (containing >2 *let-7* sites) between female and male NB patients Data source: SEQC 498 (GSE62564; *M-Non*/HR/DFD subset; *n=40*).

Extended Data Figure 12

Extended Data Figure 12. a) H19 expression in *M-Non* CHLA-42 cells in response to control siRNA or *H19* targeting siRNA transfection. **b)** Expression levels of *let-7* target genes in CHLA-42 cells, respectively, treated as in (a)Significance determined by two-way ANOVA, followed by Tukey's multiple comparison tests; bars represent average +/- SEM; **p<0.01