Discrete Correlate Summation Clustering of a Dopamine-Angiotensin Network

Brian M. Westwood[1], Liliya M. Yamaleyeva[1] and Rong Chen[2]

[1]Department of Surgery/Hypertension and Vascular Research and [2]Department of Translational Neuroscience

Wake Forest University School of Medicine, Winston-Salem, North Carolina, United States

KEYWORDS - discrete correlate summation clustering, angiotensin, dopamine, framework, contingency test

ABSTRACT: We present a model of directed investigation of KEGG pathway analysis, beginning with discrete correlate summation (DCS) clustering of Swiss-Prot sets. Dopamine and angiotensin related proteins define the boundary for pathway analyses upon a network of proteins also associated with a compositely defined matrix of Swiss-Prot keywords to provide a framework for their biological indices (Figure 1).

Figure 1. Graphical abstract.

INTRODUCTION

This study is a framework. It is designed to visualize a system between two disparate topics, connected by common relationships.

Generally, we categorize genes with differential expression between treatments, to investigate enrichment of an effected gene set, and decide if there is evidence to indicate a concerted alteration of a pathway and not just stochastic changes.

This framework is constructed using the classification structure of the Kyoto Encyclopedia of Genes and Genomes (KEGG) (Kanehisa et al., 2000) molecular network database, evaluated with the ConsensusPathDB-human (CPDB) (Kamburov et al., 2011) over-representation analysis, using very specific starting sets (Swiss-Prot keyword searches) (UniProt Consortium, 2021).

METHODS

This is an investigation of 3309 proteins from the 20 Swiss-Prot keyword searches [oxidative&stress, glutamate, inflammation, memory, glutamatergic, blood&pressure, dopamine, hypertension, blood&brain&barrier, gaba, angiotensin, prefrontal&cortex, dopaminergic, gabaergic, cognition, midbrain, cerebral&vascular, striatum, renin, ageing].

Iterative 'original matrix total array average was calculated right to left' (RL-OriMaTArA) DCS clustering (Bronson et al., 2022) yields 3 clear count domains and 2 singleton searches for the 3309 proteins (Figure 2a, 2b). Briefly, upper cluster of a matrix was determined by sorting the totals of each variable column, then repeating on remaining extra-cluster submatrix.

RL-OriMaTArA produced 3 primary clusters [C, M, Y] and 4 combination sets [CM-blue, MY-red, CY-green, CMY-gray] to search in CPDB for pathway enrichment analysis (Figure 2c, 2d) of KEGG pathways.

The 7 CPDB searches of 3309 proteins yielded a primary output of 137 KEGG pathways with 1991 genes. Area under the curve (AUC) of receiver operating characteristic (ROC) curves and precision-recall F-measures (Fawcett, 2006) were calculated for neighborhoods 1-5 for 51 KEGG pathways and 1045 genes with 4 or more angiotensin or renin Swiss-Prot search proteins/genes and with 4 or more dopamine or dopaminergic Swiss-Prot search proteins/genes in each pathway (Table 1).

For each neighborhood, one through five, surrounding angiotensin/renin (yellow, RAS) nodes, pathways through edges were examined for dopamine/dopaminergic (magenta, DA) node proximity. If the path contained only DA associated (cyan, DA_{assoc}) and/or RAS associated (gray, RAS_{assoc}) nodes, the yellow node was marked with a blue cross (true positive, TP) for 'in' or a cyan cross (false negative, FN) for 'out' of the neighborhood being examined. If the contiguous path contained nodes other than RAS_{assoc} or DA_{assoc} (pale green), the yellow node was marked with a black cross (false positive, FP) for 'in' or a gray cross (true negative, TN) for 'out' of the neighborhood being examined (Figure 3).

All calculations were performed in Excel (Microsoft Corp., WA), ImageJ (Rasband, 1997-2018) was used for tabulation of supplemental gif images and Cytoscape (Shannon et al., 2003) was used for plotting the network in figure 4.

Figure 2. Binary count clustering for Swiss-Prot keyword searches (a); keyword group membership columns by cluster row groups [(b) rows are individual proteins, black cell = membership]; ternary diagram of cluster layout [(c) with underlay of node coloring for associated groups]; groups of clusters organized by index color for KEGG pathway map contingency tests [(d) RAS associated genes (gray) RAS genes (yellow) DA genes (magenta) DA associated genes (cyan)].

RESULTS

The 3309 proteins from the 20 Swiss-Prot keyword searches are in Supplementary Table 1. For each entry, membership in a keyword search is tabulated as a '1'. This data was used to calculate the binary count clustering for Swiss-Prot keyword searches and visualize the membership maps in Figure 2.

The contingency testing shown in the 51 supplemental gif images as referenced in Table 1 [1st, 2nd, 3rd, 4th, 5th neighborhoods, unmasked KEGG output] is a graphical representation of the exact F-AUC ROC analysis procedures.

The 51 KEGG pathways in Table 1 are color coded for all F-AUC-3 [specifically (F-AUC)$_{0.7}$-3, maximum AUC or F-measure at least 0.7 and 3 TP nodes (Westwood et al., 2022)] blue for F-AUC of 1, orange for F-AUC at least 0.85 and gray for F-AUC at least 0.7. Of the 25 (F-AUC)$_{0.85}$-5 pathways (bold), there were 160 proteins in at least 4 KEGG pathways (KP). There were 23 RAS (7.7±0.72 KP, mean+sem), 51 RAS$_{assoc}$ (7.5±0.59 KP), 22 DA (7.6±0.98 KP), 64 DA$_{assoc}$ (7.3±0.44 KP) proteins (Chi-square, p=0.44, ns; Figure 4).

Table 1. KEGG pathway contingency test output, sorted by TP then maximum for AUC of ROC curve and F-measure.

KEGG	Pathway	AUC	F	TP
hsa04270	Vascular smooth muscle contraction		0.96	12
hsa04925	Aldosterone synthesis and secretion		1.00	10
hsa04022	cGMP-PKG signaling pathway	0.80	0.90	9
hsa04071	Sphingolipid signaling pathway		1.00	8
hsa04810	Regulation of actin cytoskeleton		1.00	8
hsa04927	Cortisol synthesis and secretion		1.00	8
hsa04750	Inflammatory mediator regulation of TRP channels		0.94	8
hsa04611	Platelet activation		1.00	7
hsa04024	cAMP signaling pathway	0.96	0.93	7
hsa04510	Focal adhesion		0.93	7
hsa04921	Oxytocin signaling pathway	0.89	0.88	7
hsa04062	Chemokine signaling pathway	0.75	0.88	7
hsa04020	Calcium signaling pathway	1.00	1.00	6
hsa04066	HIF-1 signaling pathway		1.00	6
hsa04151	PI3K-Akt signaling pathway		1.00	6
hsa04360	Axon guidance	0.99	0.92	6
hsa04072	Phospholipase D signaling pathway	0.97	0.92	6
hsa04015	Rap1 signaling pathway	0.75	0.92	6
hsa04924	Renin secretion		0.92	6
hsa04926	Relaxin signaling pathway	0.86	0.92	6
hsa04261	Adrenergic signaling in cardiomyocytes		1.00	5
hsa04728	Dopaminergic synapse		1.00	5
hsa04911	Insulin secretion		1.00	5
hsa04913	Ovarian steroidogenesis		1.00	5
hsa04010	MAPK signaling pathway	0.87	0.83	5
hsa04670	Leukocyte transendothelial migration	0.40	0.83	5
hsa04720	Long-term potentiation		1.00	4
hsa04725	Cholinergic synapse		1.00	4
hsa04310	Wnt signaling pathway	0.81	0.89	4
hsa04726	Serotonergic synapse	0.75	0.89	4
hsa04928	Parathyroid hormone synthesis_secretion and action	0.81	0.89	4
hsa04912	GnRH signaling pathway	0.00	0.80	4
hsa04722	Neurotrophin signaling pathway	0.50	0.73	4
hsa04144	Endocytosis		1.00	3
hsa04371	Apelin signaling pathway	1.00	1.00	3
hsa04723	Retrograde endocannabinoid signaling		1.00	3
hsa04727	GABAergic synapse		1.00	3
hsa04014	Ras signaling pathway	0.83	0.86	3
hsa04919	Thyroid hormone signaling pathway	0.75	0.86	3
hsa04613	Neutrophil extracellular trap formation	0.44	0.75	3
hsa04935	Growth hormone synthesis_secretion and action		0.75	3
hsa04724	Glutamatergic synapse		1.00	2
hsa04350	TGF-beta signaling pathway	0.88	0.80	2
hsa04150	mTOR signaling pathway		0.80	2
hsa04370	VEGF signaling pathway	0.75	0.80	2
hsa04657	IL-17 signaling pathway	0.80	0.80	2
hsa04916	Melanogenesis		0.80	2
hsa04929	GnRH secretion	0.55	0.57	2
hsa04080	Neuroactive ligand-receptor interaction		1.00	1
hsa04668	TNF signaling pathway	1.00	1.00	1
hsa04918	Thyroid hormone synthesis		1.00	1

Figure 3.   Contingency test and fourth neighborhood example.

Figure 4.   Naive unstructured network of 25 (F-AUC)$_{0.85}$-5 pathways and 160 four KP proteins [RAS|RAS$_{assoc}$|DA|DA$_{assoc}$].

DISCUSSION

This framework ties the RAS and DA systems using targeted Swiss-Prot curated database search output membership in KEGG networks by implementing CPDB, resulting in an enriched dataset with a balanced representation from RAS, RAS$_{assoc}$, DA and DA$_{assoc}$ proteins for further analysis. The utility if this method is in the simplicity of the contingency testing on display in the supplemental gif images. The ontological microenvironment defined by this framework array is broadly applicable to describing biological systems. This Swiss-Prot/KEGG/CPDB conduit can provide foundational information in the investigation of novel networks.

ACKNOWLEDGMENT

Supported by WFSM CVSC Pilot Award, CTSI NCATS UL1TR001420 and Odell Farley Foundation.

ABBREVIATIONS

DCS, discrete correlate summation; KEGG, Kyoto Encyclopedia of Genes and Genomes; CPDB, ConsensusPathDB-human; RL-OriMaTArA, original matrix total array average was calculated right to left; AUC, Area under the curve; ROC, receiver operating characteristic; RAS, angiotensin/renin; DA, dopamine/dopaminergic; DA$_{assoc}$, DA associated; RAS$_{assoc}$, RAS associated; TP, true positive; FN, false negative; FP, false positive; TN, true negative.

REFERENCES

Kanehisa M, Goto S. KEGG: kyoto encyclopedia of genomes. Nucleic Acids Res. 2000 Jan 1;28(1):27-30. doi: 10.1093/nar/28.1.27. PMID: 10592173; PMCID: PMC102409.

Kamburov A, Pentchev K, Galicka H, Wierling C, Lehrach H, Herwig R. ConsensusPathDB: toward a more complete picture of cell biology. Nucleic Acids Res. 2011 Jan;39(Database issue):D712-7. doi: 10.1093/nar/gkq1156. Epub 2010 Nov 11. PMID: 21071422; PMCID: PMC3013724.

The UniProt Consortium , UniProt: the universal protein knowledgebase in 2021, Nucleic Acids Research, Volume 49, Issue D1, 8 January 2021, Pages D480–D489, https://doi.org/10.1093/nar/gkaa1100

Bronson S, Westwood B, Cook K, Emenaker N, Chappell M, Roberts D, Soto-Pantoja D. Discrete Correlation Summation Clustering Reveals Differential Regulation of Liver Metabolism by Thrombospondin-1 in Low-Fat and High-Fat Diet-Fed Mice. Metabolites 2022, 12, 1036. https://doi.org/10.3390/metabo12111036

Fawcett T. An introduction to ROC analysis. Pattern Recogn Lett. 2006;27(8):861-74.

Rasband, W.S., ImageJ, U. S. National Institutes of Health, Bethesda, Maryland, USA, https://imagej.net/ij/, 1997-2018.

Shannon P, Markiel A, Ozier O, Baliga NS, Wang JT, Ramage D, Amin N, Schwikowski B, Ideker T. Cytoscape: a software environment for integrated models of biomolecular interaction networks. Genome Research 2003 Nov; 13(11):2498-504

Westwood B, Patil P, Tallant A, Gallagher P. Microbiome Metabolic Network Expansion Following Muscadine Grape Extract Intervention of Hypertensive Rats SB3C2022-266, 2022. https://archive.sb3c.org/sb3c2022/