Title: Floral visitors differentially respond to local and landscape grassland features

Authors: Elinor M. Lichtenberg[1,2], Jaclyn Heiser[2,3], Kristen A. Baum[4,5], John L. Neff[6],

Shalene Jha[2,7]

1) Department of Biological Sciences and Advanced Environmental Research Institute,

University of North Texas, Denton, Texas, 76203-5017, USA

2) Department of Integrative Biology, The University of Texas at Austin, Austin, TX 78712

USA

3) AMERMIN, Austin, TX 78759, USA

4) Department of Integrative Biology, Oklahoma State University, Stillwater, OK 74078,

USA

5) Kansas Biological Survey & Center for Ecological Research, and Department of Ecology

and Evolutionary Biology, University of Kansas, Lawrence, KS 66047, USA

6) Central Texas Melittological Institute, Austin, TX 78731, USA

7) Lady Bird Johnson Wildflower Center, Austin TX 78739, USA

Corresponding author: Elinor M. Lichtenberg; 1155 Union Circle #310559, Denton, TX

76203-5017, USA; ph +1 (940) 891-6956; elichten@unt.edu

E-mail addresses: Jaclyn Heiser, jaclynnicole97@gmail.com; Kristen A. Baum,

kbaum@ku.edu; John L. Neff, jlnatctmi@yahoo.com; Shalene Jha, sjha@austin.utexas.edu

Abstract

1. Predicting how habitat composition alters communities of mobile ecosystem service providers remains a major challenge in community ecology. This is partially because separate taxonomic groups that provide the same service may respond uniquely to changes in habitat and associated resource availability. Further, the spatial scale at which habitat features impact each group can vary. Failure to account for these differences significantly limits the ability to quantify shared versus contrasting responses to habitat for important ecosystem service-providing groups.

2. We investigated the impacts of local and landscape habitat features on groups of pollinating insects with different basic biologies: Coleoptera, Diptera, Hymenoptera, and Lepidoptera. Habitat features included local flower and shelter resources as well as landscape-scale semi-natural habitat. Relative to floral resource impacts on pollinators, the role of nesting or sheltering habitat has not been well studied.

3. We found that bare ground supported more Hymenoptera and Lepidoptera but fewer Diptera, while more diverse flower communities supported more Hymenoptera but fewer Coleoptera. Interestingly, given that this study occurred in a grassland system, forest cover in the surrounding landscape more strongly affected flower visitor diversity than grassland cover did.

4. Our results highlight that pollinator conservation and sustainable land management depend on understanding the habitat needs of diverse pollinators. Because taxa can have opposite responses to specific habitat features or scales, providing a range of grassland management practices (e.g., variety in the timing and type of biomass removal) may be the most effective approach to support the broader pollinator community.

45   **Keywords:** biodiversity; habitat; community composition; pollinators; Coleoptera; Diptera;

46   Hymenoptera; Lepidoptera; Great Plains

Introduction

Habitat change is causing dramatic alterations to animal biodiversity and its

associated ecosystem functions (Raven & Wagner 2021). Despite the importance of mobile

ecosystem service-providing animals, such as pollinators and pest-predators (Kremen et al.

2007), past research on these organisms has often grouped taxa that provide similar functions

as a monolith. This contrasts with the increasing recognition that ecosystem service providers

with different basic biologies (e.g., different nesting and foraging behaviors), are expected to

show differential responses to habitat change (Lichtenberg et al. 2017; Rader et al. 2020),

even when those taxa serve similar ecological functions. Indeed, research pioneered with

plants highlights that species' traits can be critical determinants of both responses to habitat

alteration and contributions to ecosystem functions (McGill et al. 2006). Despite this

recognition, frameworks for determining community-level response to habitat change often

focus on a single taxonomic group or do not account for key ecological differences across the

taxa that represent a functional group. This significantly limits the ability to quantify the

shared versus contrasting responses to habitat change for important ecosystem service-

providing groups. Differential impacts of habitat may be partly driven by species' differences

in food and shelter resources (Ollerton 2021) or movement restrictions due to body size, wing

morphology, or a need to return to a central nest (Verboven et al. 2014; Lichtenberg et al.

2023), among other factors. Advancing this knowledge requires multi-taxa investigation of

how distinct groups respond to specific resources, and how habitat composition alters the

availability of those resources.

Flower-visiting insects are an ideal system for developing a mechanistic

understanding of the differential impacts of habitat composition on multi-taxa ecosystem-

service-providing groups. Insect pollinators are taxonomically diverse, including organisms

in the orders Hymenoptera (bees, wasps), Coleoptera (beetles), Diptera (flies), Lepidoptera

(butterflies) and others (Rader et al. 2020; Ollerton 2021). Despite this taxonomic breadth,

our understanding of how shelter and food resource availabilities affect the abundance and

diversity of insect pollinators is often limited to our knowledge of how one taxonomic group

(bees, Hymenoptera: Apoidea) responds to floral availability and diversity. Even though

butterflies and flies (Lepidoptera & Diptera) can make up a large fraction of the floral visitor

community (e.g., Cusser et al. 2016), only a small fraction of studies have simultaneously

quantified the impacts of habitat variation on these additional orders. Even less research has

focused on beetles (Coleoptera), despite their potentially similar role in pollination (Rader et

al. 2020; Cusser et al. 2021). These taxonomically diverse flower-visiting insects also provide

ecologically and economically valuable pollination services to natural and human-modified

habitats (Potts et al. 2016), and many of these insects are in decline (Ollerton 2021).

Past work generally finds that pollinator communities respond positively to higher

flower availability or diversity (Kral-O'Brien et al. 2021). What is not known is if this pattern

persists across distinct taxa, especially across environmental conditions. For example, while

flower availability or diversity promotes bees in prairies and urban gardens (Tonietto et al.

2017; Ballare et al. 2019), butterflies in California mountain meadows (Marschalek et al.

2017), and syrphid flies in German agricultural landscapes (Haenke et al. 2009), the

underlying habitat conditions are dramatically different across these systems. The handful of

studies to date that investigated drivers of multiple flower visiting taxa within a single system

found positive impacts of flowers on bees, but a mix of positive and no impacts on

butterflies, beetles, and flies (e.g., Ekroos et al. 2008, 2013; Fründ et al. 2010; Horak 2014;

Gómez-Martínez et al. 2022; He et al. 2022). Indeed, the availability of larval host plants or

other feeding substrates can strongly influence these non-bee flower visitors (Rader et al.

2020). Given the taxonomic diversity of insect pollinators, and the ecological breadth this

diversity encompasses, it is critical to investigate how multiple pollinator groups respond to

the same floral resources.

Relative to floral resources, the role of nesting or sheltering habitat in mediating

pollinator community composition has not been well studied. Shelter resources such as bare

ground, leaf litter, or pithy plant stems are critical for insects' survival and reproduction

(Kremen et al. 2007), and these shelter needs often vary dramatically among taxa. For

example, ground-nesting bees often benefit from bare ground availability (Harmon-Threatt

2020), while beetle and fly species that use leaf litter to hunt for food as larvae or overwinter

may benefit from more vegetation cover and less bare ground (e.g., hover flies, Sjödin et al.

2008; pollen beetles, Rusch et al. 2012). In another example, sandy soil often promotes bees

but can reduce beetle abundance, while tall vegetation can promote hover flies and beetles

but not bees (Sjödin et al. 2008). Determining the responses of different taxa to critical

shelter resources is a promising avenue for understanding how entire pollinator communities,

not just single taxonomic groups, are expected to respond to habitat change.

Finally, beyond these key local resources for floral visitors – including flowers, nest

materials, and shelter sites – the larger surrounding landscape composition can also impact

biodiversity patterns and ecological processes within a site (Tscharntke et al. 2012). These

broader landscapes often contain habitat within hundreds of meters of a site. They can

provide resources that may not be locally available or a greater amount of locally available

resources, or impact visitation from the regional species pool (Tscharntke et al. 2012). They

can also connect small habitat patches to each other. Several syntheses find that semi-natural

habitat in the surrounding landscape can have a variety of effects including positive, negative,

or none on pollinator abundance and diversity (Kennedy et al. 2013; Scheper et al. 2013;

Lichtenberg et al. 2017). Responses are hypothesized to be driven partially by mobility

differences across taxa due to variation in body size, dispersal behavior, and whether or not a

species provisions a fixed-location nest (central place foraging) (Ballare et al. 2019; Wong et al. 2019; Lichtenberg et al. 2023). Interestingly, landscapes may be less influential when local habitat patches are large or heterogeneous and thereby provide sufficient resources (Tscharntke et al. 2012). Understanding the interactive effects of local- and landscape-scale habitat availability and quality is particularly important when studying an array of taxa, such as multiple orders of flower-visiting insects.

To understand how distinct taxa within a functional group respond to habitat availability, we investigated the impacts of local and landscape habitat features on flower-visiting insects that vary in their basic biologies, specifically the orders of bees and wasps (Hymenoptera), butterflies (Lepidoptera), beetles (Coleoptera), and flies (Diptera). These habitat features included multiple local flower and shelter resources and the availability of landscape-level semi-natural habitat. We focused on grasslands in the Southern Great Plains, as this is a highly threatened and understudied habitat (Scholtz & Twidwell 2022) where landscapes are heterogeneous at both small and large scales. We addressed two questions. (1) How do flower-visiting insects in distinct orders respond to specific local and landscape habitat features? (2) Beyond insect orders, how do local and landscape habitat features structure the larger pollinator community and individual species' responses? We expected to find differential responses to habitat based primarily on utilized shelter material. For example, we predicted that stem- and cavity-nesting taxa would respond more positively to the amount of woodland in a landscape than ground-nesting taxa. We also expected that central place foragers (here, Hymenoptera) would be more constrained by local floral resources while other taxa would respond to both local resource availability and landscape-scale habitat heterogeneity.

Materials and Methods

Study sites

We conducted our research in the summer of 2017 at 42 sites located in 11 land units

(3-4 sites per land unit) spread across almost 400km of the Cross Timbers ecoregion from

central Texas to south central Oklahoma (Fig. 1). Prior to European settlement, this ecoregion

historically consisted of grasslands interspersed with dense tree belts, but has experienced

increasing levels of woody encroachment and urbanization (Smith et al. 2019). Further,

grasslands in this region are often invaded by two non-native perennial grasses: King Ranch

bluestem (*Bothriochloa ischaemum* (Linnaeus) Keng) and Johnsongrass (*Sorghum halepense*

(L.) Persoon). These grasses were introduced as cattle forage, but can become very dense and

often dominate plant communities after invasion (Fulbright et al. 2013). This may reduce the

abundance of flowering forbs and availability of bare ground that many pollinators require

for food and nesting (Rader et al. 2020; Kral-O'Brien et al. 2021).

Figure 1: (a) Locations of land units (blue dots) in the Cross Timbers ecoregion (gray

outline), and (b) example of a single land unit with four sites (white squares), where circles

indicate the landscape within a 1 km radius around each site and squares approximate the

plots where we sampled.

Because federally supported studies are intended to support a diverse group of land

managers, our sites were a combination of privately owned (4 land units), US Army Corps-

(4 sites), US Fish and Wildlife Service- (1 site), and Oklahoma Department of Wildlife

Conservation- (1 site) managed sites. All sites were characterized by grassland plant

communities and none had been treated with prescribed fire or seeding in the prior 10 years.

Land units were separated by a minimum of 5km (mean: 40.6km, range: 5.1km-69.8km) to

the closest unit and sites were separated by a minimum of 0.7km (mean: 1.9km, range:

0.7km-8.3km) within a land unit, which provided spatial independence of the insect

communities.

Insect sampling

We collected field data within a 10,000m^2 plot (with dimensions 100x100m or

50x200m, depending on land configuration) at each site. In each plot, we sampled flower-

visiting insects through a combination of passive trapping and active netting. We set up 30

pan traps (10 each of bright colors that attract flying insects: white, fluorescent blue,

fluorescent yellow; LeBuhn et al. 2003) in a "+" formation at the center of the plot, and

separated by 5m for maximum effectiveness (Droege et al. 2010). Each trap was a 0.096L

soufflé cup (Solo Brand P325) filled with soapy water. We pushed aside vegetation above

each trap to ensure they were visible to flying insects. Traps were deployed before 0830

hours Central Daylight Savings Time, and left out for 7 hours (as in Tonietto et al. 2017). On

the same day, we collected insects that were visiting flowers in the plot. Four people netted

insects observed in the middle of a flower (and thus likely feeding or serving as a pollinator)

during morning (between 0900 and 1200 hours) and afternoon (between 1300 and 1600

hours) 30-min sessions (four person hours total per site). This region has an extremely short

summer flowering period, limiting repeated visitation, but we ensured each site had a full

four hours of netted monitoring. This duration has been sufficient for characterizing the

community in past pollinator studies (e.g., Winfree et al. 2014). Both the trapping and netting

methods have also previously been successfully implemented in our study region (Ritchie et

al. 2016; Ballare et al. 2019).

We stored all trapped specimen in 95% ethanol and netted specimens in the freezer. In

the lab, we transferred specimens <2mm long to individual vials of ethanol, while we washed

and pinned all other specimens. We then identified each specimen to as fine a taxonomic

resolution as feasible, based on morphological characteristics and available keys (e.g.,

McAlpine 1981; Johnson & Triplehorn 2004; Michener 2007; Schuh et al. 2010; Gibbs et al.

2013; Williams et al. 2014; complete list in Table S1). Specifically, we identified 67% of

individuals to species or species group (closely related species that are hard to differentiate),

12% to genus, 4% to family, and 17% to morphospecies. This is similar resolution to past

studies covering a large breadth of insect taxa (e.g., Samways et al. 2010; Foster et al. 2019).

All specimens will be stored in the Jha Lab until they are deposited in the University of Texas

Entomology Collection.

Floral sampling

We measured flower abundance and richness in 51 1x1m quadrats located along three

transects within the plot (following Ritchie et al. 2016; Ballare et al. 2019). These transects

ran perpendicular to the 25, 50 and 75m marks along a haphazardly-selected side of the plot

for 100x100m plots, and the 50, 100, and 150m marks for the 50x200m plots. We placed the

quadrat every 3m along these transects, where we identified all plants in bloom and counted

the number of flowering inflorescences (e.g., individual flowers, heads, or racemes,

depending on species) of each species (as in Ritchie et al. 2016). We then used these data to

determine the total inflorescence abundance and richness for a site across all 51 quadrats.

Ground cover and grass sampling

In the same 51 quadrats, we measured percent cover of the following variables by

quantifying the portion of 16 squares (each 25x25cm) in each quadrat, to the nearest half

square: bare ground, living grassy or herbaceous vegetation, dead grassy or herbaceous

vegetation, rocks, and wood (e.g., tree trunks, large branches on the ground). We used these

data to calculate the mean proportion of a 1x1m quadrat that contained each habitat feature

(as in Ballare et al. 2019). In two diagonal corners of each quadrat, we also recorded the

presence or absence of the invasive perennial grasses, King Ranch bluestem and

Johnsongrass. We used these data to calculate the proportion of all observed corners that

contained these grasses (invasive grass cover) at a site.

Quantifying landscape context

We used the 2016 National Land Cover Database to determine the amount of

grassland and woodland within a 1km radius of each site. First, we quantified in Fragstats

(McGarigal et al. 2012) the proportion of this area that consisted of each land cover type. We

then summed categories within each habitat to determine the total proportion of area that

consisted of grassland (land cover types: Grassland/Herbaceous and Pasture/Hay) and

woodland (land cover types: Deciduous Forest, Evergreen Forest, Mixed Forest,

Shrub/Scrub, and Woody Wetlands). Grassland and woodland were the primary land uses

around each study site, making up an average 53% and 31% of the overall cover,

respectively. These were followed by agriculture (11%), water (8%), developed land (6%),

and wetland (2%).

Data analysis

We investigated how local and landscape habitat availability impacted abundance,

diversity, and community composition of flower-visiting insects in separate orders,

conducting all analyses in R (R Core Team 2018). First, we calculated abundance, richness,

and evenness (E_{var}, Smith & Wilson 1996) of each order at each site. Because there were

relatively few Lepidoptera, we analyzed presence rather than abundance, richness, or

evenness. We then built negative binomial (abundance data, which were

overdispersed)(MASS package, Venables & Ripley 2002), logistic (Lepidoptera presence),

Poisson (Diptera richness due to heteroscedasticity), or linear (all other richness and evenness

data)(lme4 package, Bates et al. 2015) mixed models with abundance, richness, evenness, or

presence of a given order as the response and land unit as a random effect. The random effect

controlled for sites within a land unit being geographically close to each other and under

similar recent and historic management. To account for heteroscedasticity and an influential

outlier, we log-transformed Diptera evenness data and excluded a site with almost 200 flies

from the Diptera abundance regressions, respectively.

For all models, we selected five fixed-effect predictors that captured a range of

pollinator habitat needs (e.g., Kremen et al. 2007; Harmon-Threatt 2020) within our sites:

floral richness, landscape context, the floral richness-landscape context interaction, bare

ground cover, and invasive grass cover. Initial analyses confirmed that these variables were

not collinear (VIF < 1.25 in all regressions). We did not include floral abundance because it

correlated with flower richness (S=5677.97, r=-0.57, p<0.0001; Table S2) and preliminary

analyses confirmed it was less informative. We scaled floral richness for analyses because all

other predictors were proportions. We measured landscape context as the proportion of the

surrounding landscape that was grassland or woodland. Because these proportions are highly

correlated (S=23186.00, r=-0.75, p<0.0001; Table S2), we only included one per model, and

ran separate models with each landscape type. We then used information-theoretic model

selection to determine the best fit model(s) for each metric and insect order (MuMIn, Bartón

2018), selecting the models with low AICc values (within 2 of the lowest value; Burnham &

Anderson 1998) and focusing our results on these models. We also calculated rarefied

richness and evenness, and estimated richness (Chao1, Gotelli & Colwell 2011). These

estimates were highly correlated with measured richness and evenness (Table S3) and thus

we focus on measured richness and evenness (as in Fortel et al. 2014).

We then investigated impacts of the same set of predictors on community

composition of each order using PERMANOVA (vegan package, Oksanen et al. 2019). To

do so, we categorized each predictor variable into "high" (above the median) and "low"

(below the median) values. Analyses included site as a grouping variable, removed singleton

species to reduce effects of rare taxa, and excluded six sites with fewer than two Coleoptera.

We assessed significance of each term, including the interaction between flower richness and

landscape context, using marginal effects. As with regressions, we ran separate analyses with

landscape context as grassland and woodland. For predictors that significantly affected

community composition, we determined species that are strongly associated with a given type

of habitat using indicator species analysis (indicspecies package, De Cáceres & Legendre

2009) with phi coefficient correction to account for unequal numbers of sites across groups

(Tichý & Chytrý 2006). This provides insight into species' habitat preferences (De Cáceres &

Legendre 2009). To enable indicator species analysis of the interactive effect between flower

richness and landscape context, we created an additional variable that had one level for each

combination of high and low flower richness and landscape context. Prior to performing

indicator species analysis, we ensured homogeneity of multivariate dispersions (vegan

package, Oksanen et al. 2019).

Results

Overall, our sites varied considerably in the local and landscape habitat variables as

well as abundance and diversity of insect orders recorded (Table 1). We collected 4499

individuals in 172 unique insect taxa (species, genus, family, or morphospecies) across all

sites (Table S4, including taxonomic affiliations and authorities). This included 728

Coleoptera in 25 taxa, 1799 Diptera in 47 taxa, 1891 Hymenoptera in 87 taxa, and 81

Lepidoptera in 13 taxa. Ninety percent of Hymenoptera (1700 individuals) were bees, and the

rest were wasps. The most common species were *Ravinia* spp. flies (690 individuals), honey

bees (401 individuals), *Epicauta sericans* beetles (342 individuals), *Asyndetus* spp. flies (288

individuals), *Condylostylus* spp. flies (233 individuals), *Lasioglossum coactus* bees (203

individuals), a *Lasioglossum* morphospecies (139 individuals), and *Bombus pensylvanicus*

bumble bees (112 individuals).

Variable	Min value	Median value	Max value	Regression details table
Flower richness	0	6	12	
Landscape-scale grassland cover	0.17	0.51	0.92	
Landscape-scale woodland cover	0.04	0.25	0.83	
Bare ground cover	0.00	0.003	0.13	
Invasive perennial grass cover	0.00	0.23	1.00	
Coleoptera abundance	0	10	76	S5
Coleoptera richness	0	4	12	S6
Coleoptera evenness	0.20	0.72	1.00	S7
Diptera abundance	4	33	192	S8

Diptera richness	2	8	16	S9
Diptera evenness	0.24	0.50	0.94	S10
Hymenoptera abundance	4	31	178	S11
Hymenoptera richness	3	12	22	S12
Hymenoptera evenness	0.32	0.59	1.00	S13
Lepidoptera presence	NA	NA	NA	S14

Table 1: Habitat characteristics and insect biodiversity at those sites. Lepidoptera were present at 32 (out of 42) sites.

Abundance and diversity

Regressions indicated that local habitat, particularly bare ground cover and floral richness, strongly impacted pollinator communities. Higher bare ground cover led to significantly higher Hymenoptera abundance (Fig. 2a; Table S11) and Lepidoptera presence (Fig. 2c; Table S14), but lower Diptera abundance (Fig. 2d; Table S8) and Hymenoptera evenness (Fig. 2b; Table S13). Increased floral richness led to greater Hymenoptera abundance and richness (Fig. 3a,b; Tables S11, S12), but lower Coleoptera abundance (Fig. 3c; Table S5). Coleoptera evenness also increased with flower richness (Fig. 3d). This increase was greater in landscapes with less grassland in one of three best models (Table S7).

Figure 2: Relationships between bare ground cover at a site and (a) Hymenoptera abundance,

(b) Hymenoptera evenness, (c) Lepidoptera presence/absence, and (d) Diptera abundance.

Fitted lines are from negative binomial (a,d) or general linear (b) regressions.

Figure 3: Relationships between flower richness at a site and (a) Hymenoptera abundance,

(b) Hymenoptera richness, (c) Coleoptera abundance, and (d) Coleoptera evenness. Fitted

lines are from negative binomial (a,c) or general linear (b,d) regressions.

At the landscape scale, woodland cover impacted flower visitors more than grassland

cover did. Sites in landscapes with more woodland had higher Coleoptera richness (Fig. 4a;

Table S6), and lower Coleoptera evenness (Fig. 4c; Table S7) and Diptera richness (Fig. 4b;

Table S9). Sites with greater grassland cover also had greater Coleoptera evenness in one of

three best models (Table S7).

Figure 4: Relationships between the proportion of woodland in the landscape around a site

and (a) Coleoptera richness, (b) Coleoptera evenness, and (c) Diptera richness. Fitted lines

are from negative binomial (a,c) or Poisson (b) regressions.

Cover of the two invasive grass species, King Ranch bluestem and Johnsongrass, did

not correlate with floral richness (Spearman's test: r= 0.18, p=0.25) and did not impact most

pollinator response variables in our regressions. Coleoptera evenness was higher at sites with

more invasive grasses in one of six best models (Table S7).

Community composition

Like our abundance and diversity regressions, local-level flower richness and bare

ground, and landscape-level proportion of woodland, were important predictors of flower-

visiting insect community composition. Diptera community composition differed at sites with

less versus more bare ground, and in landscapes with less versus more woodland (Table S15).

Diptera and Hymenoptera (Table S16) community compositions varied in response to an

interaction between floral richness and the amount of woodland in the landscape (Fig. S1).

Coleoptera community composition was unaffected by any of our local or landscape habitat

variables (Table S17).

Indicator species analyses identified several Diptera and Hymenoptera species that

were associated with specific habitat types (Tables S18-S20). *Condylostylus* long-legged

flies, *Helicobia rapax* flesh flies, and Sciaridae dark-winged fungus gnats were more likely to

be found at sites with little bare ground, while *Geron* bee flies were more likely to occur at

sites with more bare ground and, separately, landscapes with more woodland. *Atherigona*

orientalis tomato flies were more likely to be found in landscapes with less woodland.

Several Diptera and Hymenoptera species, such as *Anoplius nigritus* spider wasps and

Asyndetus flies, were more likely to be found at specific combinations of local floral richness

and landscape woodland availability (Table 2).

	Low woodland	High woodland
Low flower richness	*Augochlorella aurata* (H: G[5], 1.72 ± 0.20 mm) *Chrysotus* spp. (D; 0.57 ± 0.03 mm) *Lasioglossum coreopsis* (H: G[7], 1.04 ± 0.11 mm) *Melissodes communis* (H: G[4], 3.41 ± 0.11 mm)	*Chrysotus* spp. (D; 0.57 ± 0.03 mm) *Geron* spp. (D; 1.44 ± 0.03 mm) *Melissodes communis* (H: G[4], 3.41 ± 0.11 mm) *Epimelissodes petulca* (H: G[6], 3.41 ± 0.13 mm) *Xylocopa virginica* (H: W[2], 6.34 ± 0.20 mm)
High flower richness	*Anoplius nigritus* (H; 1.33 ± 0.03 mm) *Asyndetus* spp. (D; 0.45 ± 0.03 mm)	*Bombus fraternus* (H: G[3], 5.22 ± 0.41 mm) *Chrysotus* spp. (D; 0.57 ± 0.03 mm)

Geron spp. (D; 1.44 ± 0.03 mm)	*Geron* spp. (D; 1.44 ± 0.03 mm)
Melissodes communis (H: G[4], 3.41 ± 0.11 mm)	*Xylocopa virginica* (H: W[2], 6.34 ± 0.20 mm)
M. tepaneca (H: G[4], 3.23 ± 0.10 mm)	
Osmia subfasciata (H: W[1], 2.25 ± 0.10 mm)	
Tachydromiinae spp. (D; 0.59 ± 0.08 mm)	
Tomosvaryella spp. (D; 0.50 ± 0.04 mm)	
Xylocopa virginica (H: W[2], 6.34 ± 0.20 mm)	

Table 2: Habitat associations of indicator taxa. (D) indicates that the taxon is a Diptera, and (H) that it is a Hymenoptera. We indicate nesting location for bees, since these data are available in the literature: (G) indicates ground-nesting and (W) indicates wood-nesting. Body sizes are mean ± standard deviation inter-tegular distances from our collections. (Nesting references are: (1) Cane et al. 2007; (2) Laport & Minckley 2012; (3) Williams et al. 2014; (4) Danforth et al. 2019; (5) Grab et al. 2019; (6) Kilpatrick 2020; (7) Carril & Wilson 2021)

Discussion

We used a multi-taxa approach to examine the impacts of local and landscape habitat features on flower-visiting insects and show that a small number of key habitat features – bare ground cover, floral richness, and landscape-scale forest cover – differentially affected

flower visitor abundance, richness, evenness, and community composition. Specifically, bare

ground promoted Hymenoptera and Lepidoptera, but inhibited Diptera. Further, having a

more diverse flower community promoted Hymenoptera but disfavored Coleoptera.

Interestingly, given that this study occurred in a grassland system, grassland cover in the

surrounding landscape minimally affected flower-visiting insect communities. Instead, sites

with greater forest cover had greater Coleoptera richness, but lower Diptera richness and

Coleoptera evenness. Community composition analyses were qualitatively similar to

regressions, although landscape composition more strongly impacted community

composition than abundance or diversity.

Our results highlight a key synergy in management of the two most recognized

pollinator orders: Hymenoptera and Lepidoptera. Both benefited from having more bare

ground, which was typically distributed in small patches (range: 0 to 13 percent cover of

quadrats) and not large swathes. Benefits of bare ground for Hymenoptera likely reflect the

shelter needs of ground-nesting bees. Ninety percent of our Hymenoptera were bees, and the

majority of bees in the study region nest in the ground (Ballare et al. 2019). Studies from

other regions have also shown that sites with more bare ground often have more ground-

nesting bees (reviewed in Harmon-Threatt 2020). Patches of bare ground can also promote

wasps and butterflies (Eilers et al. 2013; Szczepko et al. 2020). In many cases bare patches

have less thatch and thus provide easier nest access for ground-nesting bees and wasps

(Brokaw et al. 2023) and perhaps easier access to soil nutrients needed by butterflies that

exhibit puddling behavior (Ankola et al. 2021). A second hypothesized explanation for

positive response to bare ground is that less vegetative cover creates more favorable

microclimates through increased sun exposure. Warmer soil (Anderson & Harmon-Threatt

2016) and vegetation (via heat reflected off the ground; e.g., Eilers et al. 2013; Dickins et al.

2013) can enhance larval development of bees and butterflies, respectively. Third, the visual

contrast of plants against bare ground may provide easier visual landmarks for bee nest

locations (reviewed in Harmon-Threatt 2020) or facilitate prey detection by predators, which

are often more abundant in sites with less vegetation diversity (Lucatero et al. 2024). Finally,

in grasslands, bare ground may indicate lower grass dominance and thus greater availability

of forb species and the flowers that provide nectar to bees and butterflies (Eilers et al. 2013;

Marschalek et al. 2017). While bare ground cover did not correlate with flower abundance or

richness in our study (Table S2), reduced competition with grasses may support greater forb

access to sunlight and soil nutrients (McCain et al. 2010).

For this same bare ground habitat feature, our results highlight the potential for

tradeoffs with dipterans. Specifically we found fewer Diptera at sites with more bare ground.

This pattern was not driven by syrphids, the most commonly-studied Diptera pollinators,

which comprised only 1% (20 individuals) of our dipterans. Diptera life histories suggest that

adults are largely free living while larvae are often aquatic or parasitic (McAlpine 1981).

Dolichopodid (34% of our specimens) larvae may also live in other humid environments such

as thick mulch (Gill et al. 2011; Cicero et al. 2017) or under bark (Kautz & Gardiner 2019).

This suggests that sites with more bare ground, and thus sparser vegetation or leaf litter, may

not provide sufficient moist microhabitats for fly larvae, thereby limiting populations of flies

that visit flowers as adults. In support of this hypothesis, our community composition

analyses found that *Condylostylus* species (Dolichopodidae) that require humid environments

were found mainly at sites with less bare ground, while *Geron* species that are parasitoids

were found mainly at sites with more bare ground. However, natural histories and local or

landscape drivers of fly communities remain understudied (Cicero et al. 2017; Kautz &

Gardiner 2019), especially compared to bees and butterflies. Our documentation of this key

trade-off between the habitat drivers of Diptera relative to Hymenoptera and Lepidoptera

highlights the urgent need to better study flies to advance more comprehensive insect

conservation strategies.

Similar to the divergent order-level responses we found to ground cover, we found

contrasting responses to local-scale floral diversity. Hymenoptera exhibited a positive

response to floral richness while Coleoptera exhibited a negative response, though they did

exhibit higher evenness in the higher floral richness sites. Bees depend heavily on flowers for

both larval and adult food; past studies have similarly found that bee abundance and richness

increase with floral richness (Venturini et al. 2017; Kral-O'Brien et al. 2021). A diverse pool

of available flowers facilitates diet balancing by floral generalists (Leach & Drummond

2018) and also provides the necessary host plants for floral specialists that collect pollen from

only a small number of plant species (Isbell et al. 2017). Floral-rich sites also tend to provide

floral availability across the entire growing season (Kremen et al. 2007; Kral-O'Brien et al.

2021). Floral richness may be particularly important in grasslands such as our study sites that

have not been under intensive management, where there were fewer flowers than in managed

prairies (Griffin et al. 2021). Beetle responses to floral diversity are less understood. The

several studies that investigate this relationship find positive (Gómez-Martínez et al. 2022) or

no (Sjödin et al. 2008; Toivonen et al. 2022) responses. This lack of consensus may reflect

the small number of studies focusing on flower-visiting beetles or beetles' broad range of

larval food resources (Johnson & Triplehorn 2004).

At the landscape scale, we found two notable patterns. First, although we sampled in

open grasslands, our insect communities were more strongly affected by the amount of

woodland in the landscape than the amount of grassland. Trees provide resources such as

cavities for larval development or overwintering, flowers, and larval food (Rusch et al. 2012;

Söderman et al. 2016; Mola et al. 2021). Consistent with this, our community composition

analyses found *X. virginica* carpenter bees, which make holes in trees in trees to house their

nests, were largely in sites surrounded by more woodland. These bees exhibit signatures of

high natal site fidelity within a few hundred meters, despite being capable of long-distance

flight (Ballare & Jha 2021). Further, 33% of our beetle specimens, representing 72% of the

taxa we found, were in families known to be associated with trees: Anthribidae, Buprestidae,

Cerambicidae, Curculionidae, and Mordellidae (Irmler et al. 2010). On the other hand, the

observed decrease in fly richness in landscapes with more woodland suggests that many fly

species rely largely on open rather than forested habitat for meeting their resource and habitat

needs. Understanding the mechanisms behind this pattern, which differs from syrphids'

reliance on forested landscapes (e.g., Sjödin et al. 2008; Söderman et al. 2016), is hindered by

a lack of research on the non-syrphid fly species that formed the large majority of our

individuals, and on landscape impacts on flies in general (Senapathi et al. 2017; Rader et al.

2020), and requires further research.

459   Additionally, while some studies find that landscape context modulates local habitat

impacts on insect communities (e.g., Batáry et al. 2011; Lichtenberg et al. 2017), in our

system we found that the two scales acted relatively independently. Interactive impacts of

landscape heterogeneity and local management or habitat availability on insect communities

have largely been investigated in cropping systems (Senapathi et al. 2017; but see, e.g.,

Griffin et al. 2021; Lane et al. 2022). Landscape interactions with local conditions may be

less important in areas dominated by semi-natural habitat where resources are overall more

abundant and movement among habitat patches is less restricted. Theoretically, landscape

moderation of field-scale management in agricultural systems is expected to occur mainly in

landscapes that consist of less than 20% non-crop habitat (Tscharntke et al. 2012). In

contrast, the landscapes around our sites were very homogeneous, with 50-100% semi-natural

habitat. Thus, movement patterns resulting from central place foraging may not be mediated

by landscape context in this system. Consistent with movement being constrained at a small

spatial scale due to animals' biologies, we found that Hymenoptera, which are constrained by

the need to provision a fixed-location nest, responded minimally to landscape composition.

Finally, while we found two invasive grass species – King Ranch bluestem and

Johnsongrass – across a number of our sites and often at high densities, their proportion cover

did not impact pollinator abundance or richness. These grasses often dominate plant

communities after invasion, reducing availability of the forbs that pollinators depend on

(Fulbright et al. 2013). While little research has explicitly investigated impacts of invasive

grasses on pollinators (Stout & Morales 2009), there is some evidence that these grasses can

reduce invertebrate (Fulbright et al. 2013) or bee (Pei et al. 2023) abundance and diversity,

and that invasive grass removal can promote butterflies (Gowdy et al. 2022). The present

study did not find evidence that invasive grasses were depressing forbs, as invasive grass

cover did not correlate with flower abundance or richness at our study sites. However, our

study focused on floral metrics and did not examine the effect of invasive grasses on the

entire (flowering and non-flowering) plant community, which may be negatively impacted by

invasive grasses (e.g., Perkins & Hatfield 2014). Additional research is required to evaluate

reproductive versus vegetative impacts of invasive grasses on native plants, especially those

native species which are critical for provisioning floral resources for pollinator communities.

Overall, our study highlights the need to understand how multiple habitat features, at

the local and landscape scales, impact flower-visiting insects across distinct taxonomic

groups. We showed that responses to these habitat features vary systematically both among

insect orders and across individual species. Further, our results indicate that the composition

of semi-natural habitat, at both local and landscape scales, has differential impacts on

members of the pollinator community. Managing systems for diverse pollinator communities

necessitates meeting the resource needs of insects that have sometimes opposite responses to

habitat features such as bare ground or floral richness. Thus, regimes that provide a range of

grassland management practices (e.g., variety in the timing and type of biomass removal)

may be a more effective approach to support the broader pollinator community than

implementing a single management practice at a single point in time. We also highlight that

data-driven pollinator conservation requires more research to understand the responses of a

broader array of flower-visiting taxa and determine which habitat features are most critical to

manage.

Acknowledgements

We thank the US Army Corps of Engineers, Hagerman National Wildlife Refuge, the

Oklahoma Department of Wildlife Conservation, and private landowners for providing access

to field sites. We thank Llewyn Blossfeld, Shannon Dang, Bridget Harter, Katie Strain, and

Sam Wilhelm for field and lab assistance; Llewyn Blossfeld, Parrish Brady, Eliana

Buenaventura, Val Bugh, Shannon Dang, Nicholas Fensler, Jason Hansen, Elizabeth Lopez,

Julia Mlynarek, James Pitts, Mike Quinn, Brian Rabar, Ed Riley, Tín Rodriguez, Anna

Solecki, and Alex Wild for insect identification; Dan Levine for GIS assistance; and Michael

Caballero for fly measurements. This project was funded by US Fish and Wildlife Service

Competitive State Wildlife Grant #492118/494242.

References

Anderson N, Harmon-Threatt A. 2016. The effects of seed mix diversity on soil conditions and nesting of bees in prairie restorations. North American Prairie Conference Proceedings **17**:104–112.

Ankola K, Aishwarya D, Anusha R, Vaishnavi N, Supriya VK, Puttaraju HP. 2021. Ecological significance of puddling as a behavioural phenomenon in the life history of butterfly *Papilio polytes* Linn. (Lepidoptera: Papilionidae). Journal of Asia-Pacific Entomology **24**:1–6.

Ballare KM, Jha S. 2021. Genetic structure across urban and agricultural landscapes reveals evidence of resource specialization and philopatry in the Eastern carpenter bee, *Xylocopa virginica* L. Evolutionary Applications **14**:136–149.

Ballare KM, Neff JL, Ruppel R, Jha S. 2019. Multi-scalar drivers of biodiversity: local management mediates wild bee community response to regional urbanization. Ecological Applications **29**:e01869.

Bartón K. 2018. MuMIn: Multi-model inference.

Batáry P, Báldi A, Kleijn D, Tscharntke T. 2011. Landscape-moderated biodiversity effects of agri-environmental management: a meta-analysis. Proceedings of the Royal Society B: Biological Sciences **278**:1894–1902.

Bates D, Mächler M, Bolker B, Walker S. 2015. Fitting linear mixed-effects models using lme4. Journal of Statistical Software **67**:1–48.

Brokaw J, Portman ZM, Bruninga-Socolar B, Cariveau DP. 2023. Prescribed fire increases the number of ground-nesting bee nests in tallgrass prairie remnants. Insect Conservation and Diversity **16**:355–367.

Burnham KP, Anderson DR. 1998. Model Selection and Multimodel Inference: A Practical Information-Theoretic Approach, 2nd edition. Springer-Verlag, New York.

Cane JH, Griswold T, Parker FD. 2007. Substrates and materials used for nesting by North American *Osmia* bees (Hymenoptera: Apiformes: Megachilidae). Annals of the Entomological Society of America **100**:350–358.

Carril OM, Wilson JS. 2021. Common Bees of Eastern North America | Princeton University Press. Princeton University Press, Princeton, NJ.

Cicero JM, Adair MM, Adair RC, Hunter WB, Avery PB, Mizell RF. 2017. Predatory behavior of long-legged flies (Diptera: Dolichopodidae) and their potential negative effects on the parasitoid biological control agent of the Asian citrus psyllid (Hemiptera: Liviidae). Florida Entomologist **100**:485–487.

Cusser S, Haddad NM, Jha S. 2021. Unexpected functional complementarity from non-bee pollinators enhances cotton yield. Agriculture, Ecosystems & Environment **314**:107415.

Cusser S, Neff JL, Jha S. 2016. Natural land cover drives pollinator abundance and richness, leading to reductions in pollen limitation in cotton agroecosystems. Agriculture, Ecosystems & Environment **226**:33–42.

Danforth BN, Minckley RL, Neff JL. 2019. The Solitary Bees: Biology, Evolution, Conservation. Princeton University Press, Princeton, NJ.

De Cáceres M, Legendre P. 2009. Associations between species and groups of sites: indices and statistical inference. Ecology **90**:3566–3574.

Dickins EL, Yallop AR, Perotto-Baldivieso HL. 2013. A multiple-scale analysis of host plant selection in Lepidoptera. Journal of Insect Conservation **17**:933–939.

Droege S, Tepedino VJ, Lebuhn G, Link W, Minckley RL, Chen Q, Conrad C. 2010. Spatial patterns of bee captures in North American bowl trapping surveys. Insect Conservation and Diversity **3**:15–23.

Eilers S, Pettersson LB, Öckinger E. 2013. Micro-climate determines oviposition site selection and abundance in the butterfly *Pyrgus armoricanus* at its northern range margin. Ecological Entomology **38**:183–192.

Ekroos J, Piha M, Tiainen J. 2008. Role of organic and conventional field boundaries on boreal bumblebees and butterflies. Agriculture, Ecosystems & Environment **124**:155–159.

Ekroos J, Rundlöf M, Smith HG. 2013. Trait-dependent responses of flower-visiting insects to distance to semi-natural grasslands and landscape heterogeneity. Landscape Ecology **28**:1283–1292.

Fortel L, Henry M, Guilbaud L, Guirao AL, Kuhlmann M, Mouret H, Rollin O, Vaissière BE. 2014. Decreasing abundance, increasing diversity and changing structure of the wild bee community (Hymenoptera: Anthophila) along an urbanization gradient. PLoS ONE **9**:e104679.

Foster CW, Neumann JL, Holloway GJ. 2019. Linking mesoscale landscape heterogeneity and biodiversity: gardens and tree cover significantly modify flower-visiting beetle communities. Landscape Ecology **34**:1081–1095.

Fründ J, Linsenmair KE, Blüthgen N. 2010. Pollinator diversity and specialization in relation to flower diversity. Oikos **119**:1581–1590.

Fulbright TE, Hickman KR, Hewitt DG. 2013. Exotic grass invasion and wildlife abundance and diversity, south-central United States. Wildlife Society Bulletin **37**:503–509.

Gibbs J, Packer L, Dumesh S, Danforth BN. 2013. Revision and reclassification of *Lasioglossum* (*Evylaeus*), *L.* (*Hemihalictus*) and *L.* (*Sphecodogastra*) in eastern North America (Hymenoptera: Apoidea: Halictidae). Zootaxa **3672**:1–117.

Gill HK, McSorley R, Branham M. 2011. Effect of organic mulches on soil surface insects and other arthropods. Florida Entomologist **94**:226–232.

Gómez-Martínez C, González-Estévez MA, Cursach J, Lázaro A. 2022. Pollinator richness, pollination networks, and diet adjustment along local and landscape gradients of resource diversity. Ecological Applications **32**:e2634.

Gotelli NJ, Colwell RK. 2011. Estimating species richness. Pages 39–54 in Magurran AE, McGill BJ, editors. Biological Diversity. Oxford University Press, New York.

Gowdy G, Hernández F, Fulbright T, Grahmann E, Wester D, Vreugdenhil E, Henehan A, Smith F, Hehman M. 2022. Plant, avian, and butterfly response to a native-grassland restoration in southern Texas. Ecological Restoration **40**:44–52.

Grab H, Brokaw J, Anderson E, Gedlinske L, Gibbs J, Wilson J, Loeb G, Isaacs R, Poveda K. 2019. Habitat enhancements rescue bee body size from the negative effects of landscape simplification. Journal of Applied Ecology **56**:2144–2154.

Griffin SR, Bruninga-Socolar B, Gibbs J. 2021. Bee communities in restored prairies are structured by landscape and management, not local floral resources. Basic and Applied Ecology **50**:144–154.

Haenke S, Scheid B, Schaefer M, Tscharntke T, Thies C. 2009. Increasing syrphid fly diversity and density in sown flower strips within simple vs. complex landscapes. Journal of Applied Ecology **46**:1106–1114.

Harmon-Threatt A. 2020. Influence of nesting characteristics on health of wild bee communities. Annual Review of Entomology **65**:39–56.

He M, Ran N, Jiang H, Han Z, Dian Y, Li X, Xie D, Bowler PA, Wang H. 2022. Effects of landscape and local factors on the diversity of flower-visitor groups under an urbanization gradient, a case study in Wuhan, China. Diversity **14**:208.

Horak J. 2014. Insect taxa with similar habitat requirements may differ in response to the environment in heterogeneous patches of traditional fruit orchards. Journal of Insect Conservation **18**:637–642.

Irmler U, Arp H, Nötzold R. 2010. Species richness of saproxylic beetles in woodlands is
affected by dispersion ability of species, age and stand size. Journal of Insect
Conservation **14**:227–235.
Isbell F et al. 2017. Benefits of increasing plant diversity in sustainable agroecosystems.
Journal of Ecology **105**:871–879.
Johnson NF, Triplehorn CA. 2004. Borror and DeLong's Introduction to the Study of Insects.
7th edition. Cengage Learning, Australia.
Kautz AR, Gardiner MM. 2019. Agricultural intensification may create an attractive sink for
Dolichopodidae, a ubiquitous but understudied predatory fly family. Journal of Insect
Conservation **23**:453–465.
Kennedy CM et al. 2013. A global quantitative synthesis of local and landscape effects on
wild bee pollinators in agroecosystems. Ecology Letters **16**:584–599.
Kilpatrick LD. 2020. Establishing Naturalized Areas in Managed Turfgrass Systems in North
Carolina to Evaluate Their Potential to Offset the Effects of Urbanization and Support
Biodiverse Pollinator Communities. M.Sc. North Carolina State University, United
States -- North Carolina.
Kral-O'Brien KC, O'Brien PL, Hovick TJ, Harmon JP. 2021. Meta-analysis: higher plant
richness supports higher pollinator richness across many land use types. Annals of the
Entomological Society of America **114**:267–275.
Kremen C et al. 2007. Pollination and other ecosystem services produced by mobile
organisms: a conceptual framework for the effects of land-use change. Ecology
Letters **10**:299–314.
Lane IG, Portman ZM, Herron-Sweet CH, Pardee GL, Cariveau DP. 2022. Differences in bee
community composition between restored and remnant prairies are more strongly
linked to forb community differences than landscape differences. Journal of Applied
Ecology **59**:129–140.
Laport RG, Minckley RL. 2012. Occupation of active *Xylocopa virginica* nests by the
recently invasive *Megachile sculpturalis* in upstate New York. Journal of the Kansas
Entomological Society **85**:384–386.
Leach ME, Drummond F. 2018. A review of native wild bee nutritional health. International
Journal of Ecology **2018**:e9607246.
LeBuhn G et al. 2003. A standardized method for monitoring Bee Populations – The Bee
Inventory (BI) Plot.
Lichtenberg EM et al. 2017. A global synthesis of the effects of diversified farming systems
on arthropod diversity within fields and across agricultural landscapes. Global Change
Biology **23**:4946–4957.
Lichtenberg EM, Milosavljević I, Campbell AJ, Crowder DW. 2023. Differential effects of
soil conservation practices on arthropods and crop yields. Journal of Applied
Entomology **147**:931–940.
Marschalek DA, Faulkner DK, Deutschman DH. 2017. Livestock grazing shapes the
vegetation structure and subsequent habitat use by the endangered skipper *Pyrgus*
ruralis lagunae (Lepidoptera: Hesperiidae). Environmental Entomology **46**:445–453.
McAlpine JF. 1981. Manual of Nearctic Diptera. Agriculture Canada, Ottawa, ON.
McCain KNS, Baer SG, Blair JM, Wilson GWT. 2010. Dominant grasses suppress local
diversity in restored tallgrass prairie. Restoration Ecology **18**:40–49.
McGarigal K, Cushman S, Ene E. 2012. FRAGSTATS. University of Massachusetts,
Amherst, Amherst, MA.
McGill BJ, Enquist BJ, Weiher E, Westoby M. 2006. Rebuilding community ecology from
functional traits. Trends in Ecology & Evolution **21**:178–185.

Michener CD. 2007. The Bees of the World, 2nd edition. The Johns Hopkins University
Press, Baltimore.
Mola JM, Hemberger J, Kochanski J, Richardson LL, Pearse IS. 2021. The importance of
forests in bumble bee biology and conservation. BioScience **71**:1234–1248.
Oksanen J et al. 2019. vegan: Community Ecology Package.
Ollerton J. 2021. Pollinators and Pollination: Nature and Society. Pelagic Publishing, Exeter.
Available from https://pelagicpublishing.com/products/pollinators-and-pollination-
ollerton-9781784272289.
Pei CK, Hovick TJ, Limb RF, Harmon JP, Geaumont BA. 2023. Invasive grass and litter
accumulation constrain bee and plant diversity in altered grasslands. Global Ecology
and Conservation **41**:e02352.
Perkins LB, Hatfield G. 2014. Competition, legacy, and priority and the success of three
invasive species. Biological Invasions **16**:2543–2550.
Potts SG, Imperatriz-Fonseca VL, Ngo HT, editors. 2016. The Assessment Report of the
Intergovernmental Science-Policy Platform on Biodiversity and Ecosystem Services
on Pollinators, Pollination, and Food Production. Secretariat of the Intergovernmental
Science-Policy Platform on Biodiversity and Ecosystem Services, Bonn, Germany.
R Core Team. 2018. R: A language and environment for statistical computing. R Foundation
for Statistical Computing, Vienna.
Rader R, Cunningham SA, Howlett BG, Inouye DW. 2020. Non-bee insects as visitors and
pollinators of crops: biology, ecology, and management. Annual Review of
Entomology **65**:391–407.
Raven PH, Wagner DL. 2021. Agricultural intensification and climate change are rapidly
decreasing insect biodiversity. Proceedings of the National Academy of Sciences
118:e2002548117.
Ritchie AD, Ruppel R, Jha S. 2016. Generalist behavior describes pollen foraging for
perceived oligolectic and polylectic bees. Environmental Entomology **45**:909–919.
Rusch A, Valantin-Morison M, Roger-Estrade J, Sarthou J-P. 2012. Local and landscape
determinants of pollen beetle abundance in overwintering habitats. Agricultural and
Forest Entomology **14**:37–47.
Samways MJ, McGeoch MA, New TR. 2010. Insect Conservation: A Handbook of
Approaches and Methods. Oxford University Press, Oxford.
Scheper J, Holzschuh A, Kuussaari M, Potts SG, Rundlöf M, Smith HG, Kleijn D. 2013.
Environmental factors driving the effectiveness of European agri-environmental
measures in mitigating pollinator loss – a meta-analysis. Ecology Letters **16**:912–920.
Scholtz R, Twidwell D. 2022. The last continuous grasslands on Earth: Identification and
conservation importance. Conservation Science and Practice **4**:e626.
Schuh RT, Hewson-Smith S, Ascher JS. 2010. Specimen databases: a case study in
entomology using web-based software. American Entomologist **56**:206–216.
Senapathi D, Goddard MA, Kunin WE, Baldock KCR. 2017. Landscape impacts on
pollinator communities in temperate systems: evidence and knowledge gaps.
Functional Ecology **31**:26–37.
Sjödin NE, Bengtsson J, Ekbom B. 2008. The influence of grazing intensity and landscape
composition on the diversity and abundance of flower-visiting insects. Journal of
Applied Ecology **45**:763–772.
Smith B, Wilson JB. 1996. A consumer's guide to evenness indices. Oikos **76**:70–82.
Smith LA, Lopez RR, Lund AA, Wegner BN, Cathey JC, Lopez A, Anderson RE, Powers
GW, Skow KL, Crawford MA. 2019. Status Update and Trends of Texas Working
Lands: 1997-2017. Page 20. Texas A&M Natural Resources Institute (NRI), College
Station, TX.

Söderman AME, Ekroos J, Hedlund K, Olsson O, Smith HG. 2016. Contrasting effects of
field boundary management on three pollinator groups. Insect Conservation and
Diversity **9**:427–437.
Stout JC, Morales CL. 2009. Ecological impacts of invasive alien species on bees.
Apidologie **40**:388–409.
Szczepko K, Kruk A, Wiśniowski B. 2020. Local habitat conditions shaping the assemblages
of vespid wasps (Hymenoptera: Vespidae) in a post-agricultural landscape of the
Kampinos National Park in Poland. Scientific Reports **10**:1424.
Tichý L, Chytrý M. 2006. Statistical determination of diagnostic species for site groups of
unequal size. Journal of Vegetation Science **17**:809–818.
Toivonen M, Karimaa A-E, Herzon I, Kuussaari M. 2022. Flies are important pollinators of
mass-flowering caraway and respond to landscape and floral factors differently from
honeybees. Agriculture, Ecosystems & Environment **323**:107698.
Tonietto RK, Ascher JS, Larkin DJ. 2017. Bee communities along a prairie restoration
chronosequence: similar abundance and diversity, distinct composition. Ecological
Applications **27**:705–717.
Tscharntke T et al. 2012. Landscape moderation of biodiversity patterns and processes - eight
hypotheses. Biological Reviews **87**:661–685.
Venables WN, Ripley BD. 2002. Modern Applied Statistics with S, 4th edition. Springer,
NY.
Venturini EM, Drummond FA, Hoshide AK, Dibble AC, Stack LB. 2017. Pollination
reservoirs for wild bee habitat enhancement in cropping systems: a review.
Agroecology and Sustainable Food Systems **41**:101–142.
Verboven HAF, Uyttenbroeck R, Brys R, Hermy M. 2014. Different responses of bees and
hoverflies to land use in an urban–rural gradient show the importance of the nature of
the rural land use. Landscape and Urban Planning **126**:31–41.
Williams PH, Thorp RW, Richardson LL, Colla SR. 2014. Bumble Bees of North America:
An Identification Guide. Princeton University Press, Princeton.
Wong MKL, Guénard B, Lewis OT. 2019. Trait-based ecology of terrestrial arthropods.
Biological Reviews **94**:999–1022.
744