Variation in albumin glycation rates in birds suggests resistance to relative hyperglycaemia rather than conformity to the pace of life syndrome hypothesis

Adrián Moreno-Borrallo[1], Sarahi Jaramillo-Ortiz[1,2], Christine Schaeffer-Reiss[1,2], Benoît

Quintard[3], Benjamin Rey[4], Pierre Bize[5], Vincent A. Viblanc[1], Thierry Boulinier[6], Olivier Chastel[7],

Jorge S. Gutiérrez[8], Jose A. Masero[8], Fabrice Bertile[1,2] *, François Criscuolo[1] *

[1] University of Strasbourg, CNRS, Institut Pluridisciplinaire Hubert Curien, UMR 7178, 67000
Strasbourg, France

[2] National Proteomics Infrastructure, ProFi, FR2048 Strasbourg, France

[3] Parc zoologique et botanique de Mulhouse, 68100 Mulhouse, France

[4] Lyon University 1, UMR CNRS 5558, Laboratoire de Biométrie et Biologie Evolutive, 69622
Villeurbanne, France.

[5] Swiss Ornithological Institute, CH-6207 Sempach, Switzerland

[6] CEFE, Montpellier University, CNRS, EPHE, IRD, Montpellier

[7] Center of Biological Studies of Chizé (CEBC), UMR 7372 CNRS - La Rochelle University, 79360
Villiers-en-Bois, France

[8] Ecology in the Anthropocene, Associated Unit CSIC-UEX, Faculty of Sciences,

University of Extremadura, Badajoz, Spain.

*These authors share senior-authorship

Key words: birds, comparative method, glucose, glycation, life history, longevity

Corresponding author: A. Moreno-Borrallo, adrian.moreno-borrallo@iphc.cnrs.fr

ABSTRACT

The pace of life syndrome hypothesis (POLS) suggests that organisms' life history, physiological and behavioural traits should co-evolve. In this framework, how glycaemia (i.e., blood glucose levels) and its reaction with proteins and other compounds (i.e. glycation) covary with life history traits remain relatively under-investigated, despite the well documented consequences of glucose and glycation on ageing, and therefore potentially on life history evolution. Birds are particularly relevant in this context given that they have the highest blood glucose levels within vertebrates and still higher mass-adjusted longevity when compared to organisms with similar physiology as mammals. We thus performed a comparative analysis on glucose and albumin glycation rates of 88 bird species from 22 orders, in relation to life history traits (body mass, clutch mass, maximum lifespan and developmental time) and diet. Glucose levels correlated positively with albumin glycation rates in a non-linear fashion, suggesting resistance to glycation in species with higher glucose levels. Plasma glucose levels decreased with increasing body mass but, contrary to what is predicted to the POLS hypothesis, glucose levels increased with maximum lifespan before reaching a plateau. Finally, terrestrial carnivores showed higher albumin glycation compared to omnivores despite not showing higher glucose, which we discuss may be related to additional factors as differential antioxidant levels or dietary composition in terms of fibres or polyunsaturated fatty acids. These results increase our knowledge about the diversity of glycaemia and glycation patterns across birds, pointing towards the existence of glycation resistance mechanisms within comparatively high glycaemic birds.

INTRODUCTION

The pace of life (POL) hypothesis [1-3] postulates that organisms behavioural/physiological characteristics and life histories have co-evolved in answer to specific ecological conditions forming pace of life syndromes (POLS) along a fast-slow continuum (but see [4] for the more recent consideration of other axes). According to the classical approach from life history theory, slower organisms would have, e.g. large body mass, late maturation, slow growth rate, small number of large offspring and high longevity, whit faster ones representing the opposite trait values (see [5-8]) within this continuum. Pace of life hypothesis added physiology to this continuum, with studies testing its predictions focusing often on metabolic rate (i.e. higher or lower metabolic rate corresponding to faster or slower pace of life, respectively; see e.g. [9-12]). However, key energy substrates related to metabolic performance, such as glucose

concentrations in tissues, have been largely overlooked (but see [13-16]). Notably, glucose, a central energy source for many organisms, plays a pivotal role in metabolism and there are some indications that its circulating blood or plasma levels correlate positively with metabolic rate per gram of mass at the interspecific level [13,17], and with whole body metabolism at the intraspecific level ([18] for the case of non-diabetic humans).

Research focusing on plasma glucose becomes highly relevant as high glycaemia can entail costs that accelerate ageing. Along with other reducing sugars, glucose can react non-enzymatically with free amino groups of proteins, lipids or nucleic acids, a reaction known as glycation [19][20]. This reaction, after several molecular reorganizations, can lead to the formation of a plethora of molecules called Advanced Glycation End-products (AGEs) [21]. AGEs can form aggregates in tissues that are well known to contribute to several age-related diseases, such as diabetes (e.g., reviews by [22-25]). AGEs are also known to promote a proinflammatory state through their action on specific receptors called RAGEs [26-27].

Remarkably, birds show circulating glucose levels much higher than other vertebrate groups, on average almost twice as high as mammals [28]. These relatively high glucose levels might support the higher metabolic rates [29, 11] and body temperatures observed in birds compared to mammals [30-31]. In addition, elevated glycaemia is thought to be an adaptation to flight, providing birds with rapid access to an easily oxidizable fuel during intense bursts of aerobic exercise (see [32-33]), such as during take-off and short-term flapping flights [34-35]). However, the presence of high glycaemia in birds is also paradoxical, given their remarkable longevity compared to their mammalian counterparts — living up to three times longer than mammals of equivalent body mass [36, 11].

Several non-excluding hypotheses have been proposed to explain how birds apparently resist the pernicious effects of a high glycaemic state. One possibility is that birds might resist protein glycation through a lower exposure of lysine residues at the surface of their proteins [37]. Another hypothesis suggests that increased protein turnover may play a role [38-39]. Additionally, birds might benefit from more effective antioxidant defences [40-42] (although within Passeriformes, [16] shows no coevolution between glycaemia and antioxidant defences) or even the presence of "glycation scavengers" (i. e. molecules that bind glucose avoiding it to react with proteins) such as polyamines which circulate at high concentration [35]. Moreover, it is generally considered that RAGEs are not present in avian tissues, suggesting possible adaptive mechanisms that would allow birds to avoid the inflammatory consequences associated with RAGE activation [43-44]. However, a putative candidate for RAGE and a case of AGE-induced

inflammation in birds were identified by Wein et al. [45], warranting further investigation on the

occurrence of RAGEs in birds.

To date, although protein glycation levels have been measured in birds in several species, these

results were mostly descriptive and concerned a small number of species [46-54]. Additionally,

many of these studies employ commercial kits that were not specifically designed for avian

species, making it challenging to interpret the results [53]. These limitations restrict our ability

to draw general conclusions from these studies. Here, we performed a comparative analysis on

primary data (for glycation and most of glucose values, see methods and ESM6) from 88 bird

species belonging to 22 orders (see ESM6) and assessed whether and how bird glycaemia and

glycation rates are linked to ecological and life-history traits (including body mass). While doing

this, we also checked whether glycation levels are influenced by circulating plasma glucose

concentrations. Finally, as diet can influence glycaemia (see e.g. [15, 55-56] but [14] found no

significant effects) and glycation, we included it in our analyses. We hypothesized that

carnivorous birds would exhibit higher glycaemia and glycation levels than omnivorous or

herbivorous species, as diets with low-carbohydrates, high-proteins and high-fat are associated

with comparatively high glycaemia and reduced insulin sensitivity in some vertebrates (e.g.; [57-

59]). Accordingly, a recent comparative study in birds showed higher blood glucose levels in

carnivorous species [56]. This may be attributed to high levels of constitutive gluconeogenesis,

which has been confirmed in certain raptors [60-61]. Conversely, bird species with high sugar

intake, such as frugivores or nectarivores, are expected to exhibit high glycaemia, as observed

in hummingbirds [49] and Passeriformes with such diet [15] (although the opposite was found

in [56]). In line with POLS hypothesis, we also hypothesized that species with a slower POL should

exhibit a lower glycaemia and higher glycation resistance (glycation levels for a given glycaemia)

compared to species with a faster POL. Hence, we included the following four parameters in our

analyses of glycaemia and glycation levels: body mass, maximum lifespan, clutch mass and

developmental time. Body mass is one of the main factors underlying life history, with higher

body mass associated with "slower" strategies and vice versa [62]. Previous studies have

reported a negative relationship between body mass and glycaemia [63, 34, 13-14, 56](but see

[49, 64] with non-significant trends in birds, and [15] only for temperate species within

Passeriformes). Therefore, we predicted that bird species that live longer, develop more slowly

and invest less per reproductive event (see ESM6 for further justification of the chosen variables)

should show lower plasma glucose levels and albumin glycation rates (after controlling for body

mass, phylogeny, diet and glucose in the case of glycation; see methods).

MATERIALS AND METHODS

<u>Species and sample collection</u>

A total of 484 individuals from 88 species measured were included for this study (see **Table ESM6. 1**). A detailed list with the provenance of the samples, including zoos, a laboratory population, and both designated captures and samples provided by collaborators from wild populations, is provided in ESM3, with a textual description in ESM6.

Blood samples were collected from the brachial or tarsal vein, or from the feet in the case of swifts, using heparin-lithium capillaries or Microvette® (Sarstedt). As samples were collected mostly by different collaborators, handling times were not always recorded and could not be adjusted for, potentially rendering the results more conservative (for a more detailed discussion on potential stress effects on glucose, see ESM6), Samples were centrifuged at 4°C, 3500 g for 10 min and plasma was aliquoted when a large volume was available. Subsequently, they were transported on dry ice to the IPHC (Institut Pluridisciplinaire Hubert Curien) in Strasbourg and stored at -80º C until analysis. Glycaemia was measured in the laboratory on the remaining plasma after taking an aliquot for glycation assessment, using a Contour Plus® glucometer (Ascensia diabetes solutions, Basel, Switzerland) and expressed in mg/dL. These of point-of-care devices have previously been assessed for its usage in birds [65-66], with several examples of its usage in recent literature related to the topic presented here (e.g. [14, 67-69]. We also performed an assay with this particular brand on a subset of 46 samples from this study, coming from nine species distributed across the whole range of glucose values (three species with "low" values, three with "medium" and three with "high", from both captive and wild populations, with five individuals per species, except one including six), confirming a positive linear correlation ($R^2_{marginal}$=0.66; $R^2_{conditional}$=0.84, for a model including the species as random factor) of this device with Randox GLUC-PAP colourimetry kits (P-value<0.001, unpublished data). Due to technical issues, related to the incapability of the device to determine certain glucose values (not because of the glucose concentration, but perhaps the particular composition of the plasma samples from certain species), we could not determine glycaemia values of 95 individuals of those that we sampled and in which glycation levels were assessed (belonging to 40 species coming from different sources). For these species, if not a single individual had a glucose measurement (which was the case in 13 species), we obtained mean plasma glucose values reported for the species from the ZIMS database from Species360 (Species360 Zoological

153   Information Management System (ZIMS) (2023), zims.Species360.org). This database provides

154   plasma glucose data measured by colorimetric glucose kits on zoo specimens not neccessary

155   corresponding to those in which we measured glycation values. The sample sizes for both

156   glucose and glycation measurements, either from our measured individuals or from the ones

157   from ZIMs, are reported on the ESM5.

Glycation levels

Glycation levels for each individual were determined using liquid chromatography coupled to

mass spectrometry (LC-MS), which is considered the gold standard for the assessment of protein

glycation levels (see e.g. [70]) and has previously been used for birds [52-53, 71]. Given the

relatively high time intrinsically taken by the employed methodology for analysing all the

samples, linked to constraints in the access to the mass spectrometry devices, the whole set of

samples of this study were analysed across several instances within a total timespan of less than

2 years (2021-2023). Only one sample from each individual was measured, given logistic

limitations on the total number of samples that could be processed. Briefly, 3 µl of plasma were

diluted with 22 µl of distilled water containing 0.1% of formic acid, followed by injection of 5 µl

into the system. The glycation values used in the analyses represent the total percentage of

glycated albumin, obtained by adding the percentages of singly and doubly glycated albumin.

These percentages were calculated by dividing the areas of the peaks corresponding to each

glycated molecule form (albumin plus one or two glucose) by the total albumin peak area (sum

of the areas of glycated plus non-glycated molecules) observed in the spectrograms obtained

from the mass spectrometry outcomes. These spectrograms represent the different intensities

of signal for the components differentiated in the sample by their TOF (Time Of Flight), which

depends on the mass to charge ratio (m/z) of the ionized molecules. More detailed information

regarding functioning of the method and data processing can be found in [53]. In cases where

albumin glycation values dropped below the limit of detection, resulting in missing data, these

individuals (10 individuals from 4 species) were excluded from the statistical analyses, as

outlined below.

Data on ecology and life-history

Diets of individual species were extracted from the AVONET dataset ([72], coming from [73], and

those adapted from [74]) and life history traits from the Amniote Database [75]. After

determining that AVONET diet classifications did not align with our research needs, minor

changes were made after consulting the original Wilman et al. [74] database (see ESM6).

For missing data on life history traits after this stage, we extracted values, in order of preference,

from the AnAge database [76] or the Birds of the World encyclopaedia [77], by calculating mean

values if an interval was given, and then averaging if male and female intervals were provided

separately. For European species, maximum lifespan records have always been checked against

the most recent Euring database [78], and from *Chionis minor* and *Eudyptes chrysolophus* (not

available from the previous sources) they were extracted from the Centre d'Etudes Biologiques

de Chizé, Centre National de la Recherche Scientifique (CNRS-CEBC) database. Several species

from the zoo had maximum lifespan values available on the ZIMs from Species 360 (Species360

Zoological Information Management System (ZIMS) (2023), zims.Species360.org), so these were

also compared with the data we had from other sources.

In the case of variables (other than maximum lifespan) for which two different sources provided

different records, an average value was calculated. For the maximum lifespan value, available

sources were cross-checked and the highest value was always used. In the absence of

satisfactory support from another source, values for maximum lifespan indicated as being

anecdotal and of poor quality have been excluded from the analyses. A table with the adequate

citations for each value is provided as part of the online available data (see Data accessibility

section). When the source is AVONET [72], this database is cited, and when it is Amniote

Database [75], we cite the sources provided by them, so the references can be checked in [75].

A list with all the additional references not coming from any of these databases or not provided

by their authors (and that are not already in the main text) is given as ESM4.

Statistical analyses

All analyses were performed in R v.4.3.2 [79]. Alpha (α) ≤0.05 was reported as significant, and

0.05<α≤0.1 as trends. General Linear Mixed Models with a Bayesian approach (*MCMCglmm*

function in R; [80]) were performed. All models were run with 6×10^6 iterations, with a thinning

interval of 100 and a burn-in of 1000. The models were simplified by eliminating quadratic terms

where they were not significant and selecting models with lower AIC (Akaike Information

Criterion) and BIC (Bayesian Information Criterion). Gaussian distribution of the response

variables was assumed (after $\log_{10}$ conversion for glucose; see ESM6 for further discussions). The

priors were established assuming variance proportions of 0.2 for the G matrix and 0.8 for the R

matrix, with v=2. Less informative priors with equal variance proportions for each partition

(residual and random) gave similar results. Lower v values (0.1, 0.2, 0.002) were also tested,

without success (i.e. simulations aborted before the established number of iterations). For the

phylogenetically controlled analyses, consensus trees (each with the species included in the

model) were obtained by using the *consensus.edges* function (from the *phytools* package from

R [81] from a total of 10,000 trees downloaded from Birdtree.org [82] (option "Hackett All

species" [83]). For such purpose, a list of species with the names adapted to the synonyms

available in such website was used (see ESM5), including the change of *Leucocarbo verrucosus*

for *Phalacrocorax campbelli*, as the former was not available and it was the only species from

the order Suliformes in our dataset, so that neither the position in the tree nor the branch length

would be affected by this change. The consensus trees were included as *pedigree* in the models.

We performed models with either glucose or glycation as dependent variables and with the diet,

body mass and life history traits as predicting variables (and glucose in the models with glycation

as a response to assess glycation resistance; see introduction). Generalized Variance Inflation

Factors (GVIFs) were calculated for all the models with more than one predictor variable to

assess the collinearity of them, as it may be expected for life history traits (see results on ESM1),

considering values above 1.6 as slightly concerning, above 2.2 as moderately concerning and

above 3.2 as severely concerning (i.e. indicating high collinearity). Models testing the effects of

age and sex on glucose and glycation levels and the number of exposed lysine residues on

glycation were also carried out. Finally, we performed models on glucose and glycation values

controlling for the taxonomic orders included in the dataset (see ESM5). The effects of

phylogeny on all models was determined by calculating the ratio of the variance estimated for

the "animal" variable, representing the *pedigree* (see ESM6) by the total variance (all random

factors plus *units*). A thorough description of all of the models, including transformations of the

variables and other details is given on the ESM6.

RESULTS

Plasma glucose and albumin glycation variability across the birds' tree

Plasma glucose and albumin glycation values varied considerably across species (**Figure 1**).

Considerable within species repeatability (see ESM6) was observed (Glucose: R = 0.716, SE =

0.042, CI_{95} = [0.619, 0.785], P-value <0.0001; Glycation: R = 0.703, SE = 0.042, CI_{95} = [0.603,

0.768], P-value <0.0001).

We found significant differences between some orders for both glucose and albumin glycation.

ESM1 shows the raw outcomes of the models, showing such significant differences to the

intercept and the Credible Intervals to perform the pairwise comparisons across all groups. To

explore some of the details, we can see for example how Apodiformes showed the highest

average glucose values (species average model: Estimated mean=351.8 mg/dL, CI_{95}[238.9,

514.2]; model with individuals: Estimated mean=349.8 mg/dL, CI_{95}[251.8, 476.7], n=39

individuals, 1 species), followed by Passeriformes (species average model: Estimated

mean=349.2 mg/dL, CI_{95}[279.6, 438.6]; model with individuals: Estimated mean=337.6 mg/dL,

CI_{95}[274.5, 414.5], n=84 individuals, 8 species), while the Suliformes (species average model:

Estimated mean=172 mg/dL, CI_{95}[117.6, 252.9]; model with individuals: Estimated mean=169.5

mg/dL, CI_{95}[120.9, 239.7], n=5 individuals, 1 species), Rheiformes (species average model:

Estimated mean=173.7 mg/dL, CI_{95}[118, 254.7]; model with individuals: Estimated mean=172

mg/dL, CI_{95}[123.7, 239.8], n=9 individuals, 1 species) and Phoenicopteriformes (species average

model: Estimated mean=176 mg/dL, CI_{95}[130, 241.1]; model with individuals: Estimated

mean=141.2 mg/dL, CI_{95}[100.4, 198.2], n=5 individuals, 1 species) show the lowest average

values (for the species average models, the pattern is

Suliformes<Rheiformes<Phoenicopteriformes, while for the individual models is

Phoenicopteriformes<Suliformes<Rheiformes).

For glycation, Strigiformes (species average model: Estimated mean=25.6 %, CI_{95}[20, 31]; model

with individuals: Estimated mean=25.3 %, CI_{95}[19.1, 31.5], n=7 individuals, 2 species),

Apodiformes (species average model: Estimated mean=25.5 %, CI_{95}[18.6, 32.4]; model with

individuals: Estimated mean=24.8 %, CI_{95}[17.9, 32], n=40 individuals, 1 species) and

Coraciiformes (species average model: Estimated mean=24.5 %, CI_{95}[17.9, 31.6]; model with

individuals: Estimated mean=23.9 %, CI_{95}[15.8, 32], n=2 individuals, 1 species), all terrestrial

carnivores as by our sampled species (see discussion below), had the highest average.

On the other hand, Casuariiformes (species average model: Estimated mean=10.8 %, CI_{95}[3.7,

18.1]; model with individuals: Estimated mean=11.4 %, CI_{95}[2.7, 20.6], n=1 individuals, 1

species), Phoenicopteriformes (species average model: Estimated mean=11.2 %, CI_{95}[5.6, 16.7];

model with individuals: Estimated mean=10.5 %, CI_{95}[4.6, 16.3], n=22 individuals, 2 species) and

Suliformes (species average model: Estimated mean=13.8 %, CI_{95}[7, 20.7]; model with

individuals: Estimated mean=13.2 %, CI_{95}[5.6, 20.5], n=5 individuals, 1 species) had the lowest

average levels for the species average models, the pattern is

Casuariiformes<Phoenicopteriformes<Suliformes, while for the individual models is Phoenicopteriformes<Casuariiformes<Suliformes). Graphs with raw data on species average and individual glucose and glycation values by order are shown on ESM2 (**Figure ESM2.3**). Estimates of phylogeny effects on the residuals of the models on glucose and glycation values differ if we consider the models with intraspecific variability or without it, but not so much between the models with and without life history traits (within the previous categories). In the case of species averages, the effect of the tree on residual glucose variation is lower and the estimation is less precise than for residual glycation variation, while for the models considering individual values, it is glycation residuals what shows lower levels of tree-related variance than glucose residuals (see ESM1).

Figure 1. Average plasma glucose values in mg/dL (in grey) and average albumin glycation rate as a percentage of total albumin (in blue) from all the species used in this study (some of them with glucose values coming from ZIMs database; see methods) with the orders they belong to. Glucose and glycation

291 values are standardized in order to be compared, with the dotted lines representing half the maximum

292 and maximum values for each variable (as indicated by the axes in their corresponding colours), from

293 inside out. Tree from a consensus on 10,000 trees obtained from "Hackett All species" on Birdtree.org,

294 including 88 species from 22 orders (see methods).

Bird glycemia appears related to body mass and maximum lifespan

After controlling for intraspecific variation in our analyses, we found that variation in glucose

levels were significantly explained by variations in body mass and both the linear and quadratic

component of residual maximum lifespan (i.e. mass and phylogeny-adjusted maximum lifespan),

but not by variations in diet (**Figure 4.A** with predictions from the model without life history

traits and **Figure ESM2.2.A** with raw individual data from the same dataset), clutch mass and

developmental time (see **Table 1** for the model including life-history trait variables). Heavier

species had lower glucose levels (see **Figure 2.A**, drawn with the estimates from the model

without life history traits, which includes more species). Glucose levels increase with increasing

mass-adjusted lifespan until reaching a plateau (**Figure 2.B**). Models that did not consider

intraspecific variability show no significant effect of any of the aforementioned variables on

glucose levels (see ESM1). The provenance of the samples (wild versus captive) only showed a

trend to a higher glucose levels in the samples from captive individuals in the model without life

history traits (Estimate = 0.058; CI_{95}[-0.008, 0.125]; P-value=0.083).

Table 1. Final glucose model with diet, body mass, life history traits and sample provenance (wild versus captive; see ESM6) as explanatory variables, including the significant quadratic effect of maximum lifespan. Posterior means, CI_{95} and pMCMC from a phylogenetic MCMC GLMM model including n=326 individuals of 58 species. Both glucose and body mass are log_{10} transformed and life history traits are residuals from a pGLS model of log_{10} body mass and log_{10} of the trait in question (see ESM6). Body mass was also centred to better explain the intercept, as 0 body mass would make no biological sense. The intercept corresponds to the Omnivore diet, being used as the reference as it is considered the most diverse and "neutral" group for this purpose. Significant predictors are indicated in bold.

	Estimates	Lower 95% CI	Upper 95% CI	Sampling effort	pMCMC
Intercept (Omnivore)	**2.387**	**2.268**	**2.5**	**59900**	**<0.001**
Diet: Carnivore terrestrial	0.02	-0.056	0.099	59900	0.592
Diet: Aquatic predator	0.034	-0.047	0.112	59900	0.393
Diet: Herbivore	-0.053	-0.177	0.07	59900	0.393
Diet: Frugivore/granivore	-0.055	-0.239	0.135	59900	0.558
Centred Log_{10}Body mass	**-0.061**	**-0.106**	**-0.015**	**59900**	**0.009**
Maximum lifespan	0.107	-0.035	0.253	59900	0.142
Maximum lifespan^2	**-0.616**	**-1.166**	**-0.095**	**59900**	**0.026**
Clutch mass	-0.095	-0.265	0.069	59900	0.258
Developmental time	0.011	-0.185	0.212	59900	0.916
Provenance: Captive	0.034	-0.039	0.106	59900	0.346

Figure 2. Plasma glucose levels (in mg/dL) variation as a function of **A** species mean centred body mass and **B** residual maximum lifespan. Both glucose and body mass are log transformed. Maximum lifespan (in years) is given as the residues of a pGLS model with body mass (in grams), both $\log_{10}$ transformed, so the effects of body mass on longevity are factored out (see ESM6). Different bird orders, are indicated by symbols, as specified on the legends at the right side of the graphs. **Figure 2.A** uses the values and estimates from the glucose model without life history traits (n=389 individuals from 75 species), while **Figure 2.B** uses only the data points employed on the complete model (n=326 individuals of 58 species).

Bird albumin glycation is related to glycemia and diet

After controlling for intraspecific variability, we found that diet was affecting variation in albumin glycation rates, with terrestrial carnivorous species having higher glycation levels than omnivorous species (**Table 2** for complete model; see **Figure 4.B** with predictions of the model without life history traits and **Figure ESM2.2.B** with raw individual data from the same dataset). However, for the models for species averages, there was only a trend on this pattern in the one including life history traits, and no effect in the other (see ESM1). The relation between glycation and glucose levels was positive and significant in all but the model that included life history traits but not intraspecific variation (see **Table 2** for the outcome of the model with individual values and life history traits and ESM1 for the rest; see **Figure 3** with estimates from the model including individual variation but no life history traits, as it contains more species and the estimates are similar). Given the logarithmic relationship between glycation and glucose (see ESM6), the slope lower than one (see **Table 2**) implies that birds with higher glucose levels have relatively lower albumin glycation rates for their glucose, fact that we would be referring to as higher glycation resistance.

Additional analyses looking at the number of exposed lysines in the albumin sequence of a species show no effect of this variable on albumin glycation rates ($\alpha=10.39$: CI_{95} [-3.713, 24.993], $\beta=0.246$: CI_{95}[-0.144, 0.629], $P_{MCMC}=0.196$; ESM1).

Table 2. Final glycation model with diet, body mass, glucose and life history traits as explanatory variables.
Posterior means, CI_{95} and pMCMC from a phylogenetic MCMC GLMM model including n=316 individuals
of 58 species. Glycation, glucose and body mass are $\log_{10}$ transformed and life history traits are residuals
from a linear model of $\log_{10}$ body mass and $\log_{10}$ of the trait in question (see ESM6). Body mass and glucose
were also centred to better explain the intercept. The intercept corresponds to the Omnivore diet, being
used as the reference as it is considered the most diverse and "neutral" group for this purpose. Significant
predictors are indicated in bold and the credible intervals are considered for making pairwise comparisons
between the groups.

	Estimates	Lower 95% CI	Upper 95% CI	Sampling effort	pMCMC
Intercept	**1.232**	**1.112**	**1.351**	**59900**	**<0.001**
Diet: Carnivore terrestrial	**0.101**	**0.017**	**0.187**	**59900**	**0.021**
Diet: Aquatic predator	0.027	-0.062	0.118	59900	0.549
Diet: Herbivore	-0.019	-0.161	0.119	59936	0.781
Diet: Frugivore/granivore	0.095	-0.098	0.292	59900	0.329
Centred $\log_{10}$Body mass	0.004	-0.043	0.05	58765	0.876
$\log_{10}$Glucose	**0.137**	**0.012**	**0.255**	**59900**	**0.027**
Maximum lifespan	0.037	-0.122	0.195	59043	0.648
Clutch mass	0.151	-0.03	0.346	59900	0.114
Developmental time	0.04	-0.188	0.266	59303	0.725

Figure 3. Albumin glycation rates (as a percentage of total albumin, $\log_{10}$ transformed) variation as a
function of individual plasma glucose values (mg/dL, $\log_{10}$ transformed). Different bird orders are
indicated by symbols, as specified on the legends at the right side of the graphs. The values and estimates
used are from the glycation model without life history traits (n=379 individuals from 75 species).

Figure 4. Outcomes of the models on individual data on effects of diet on (**A**) plasma glucose levels and (**B**) albumin glycation in birds. Glucose levels are given in mg/dL, while glycation levels are a percentage of total plasma albumin which is found to be glycated. Terrestrial carnivores showed significantly higher glycation levels than omnivores (Estimate=21.62 %, CI_{95}[18, 25.95], p_{MCMC}=0.049). Models without life history traits, including more individuals, are represented, but the models with life history traits do not show differences in their qualitative predictions (i.e. higher albumin glycation in terrestrial carnivores than in omnivores; see ESM1).

We observed no significant effect of age relative to maximum lifespan nor sex on either glycaemia or glycation (see ESM1). In some models, GVIF for body mass and/or clutch mass were higher than 1.6, and in one case body mass is slightly above 2.2 (2.26, see ESM1). This indicates that there may be moderate collinearity for this variables, but the fact that we used life-history

401 trait covariates that are excluding body mass associated variation, this small effects remain
402 mysterious, but nevertheless probably not worrying.

403 DISCUSSION

404 Plasma glucose level and albumin glycation rate (co)variation patterns suggests
405 resistance mechanisms

Our findings show that glucose levels vary widely among different bird groups. Interspecific
differences are partly explained by the allometric relationship of glycaemia with body mass
(**Figure 2.A**), which has already been reported in previous studies [56, 63, 34, 13, 14]. Indeed,
Passeriformes, Apodiformes and some Columbiformes (e.g. *Nesoenas mayeri* holds the highest
value in our dataset) are found at the higher end of the glucose level continuum, in accordance
with their relatively small body mass and powered flight, while groups of larger birds such as
Phoenicopteriformes, Anseriformes, Rheiformes and Suliformes tend to show low glycaemia
levels. This pattern is similar for glycation, with some groups of large birds (such as
Phoenicopteriformes, Anseriformes and Suliformes) showing the lowest levels of glycated
albumin, while small birds (such as Apodiformes) had the highest values. Nevertheless, glycation
remains high in some birds in relation to their glucose levels, as in Rheiformes, or low as in
Psittaciformes or Passeriformes. The case of Procellariiformes, which are typically long-lived
birds, is particularly striking, with some species exhibiting some of the highest glycation levels
(*Calonectris diomedea* and *Macronectes giganteus*) and others some of the lowest (*Procellaria*
aequinoctialis). This suggests that, if birds are protected against the deleterious effects of high
glycaemia, certain species may have evolved mechanisms to efficiently prevent proteins to be
glycated at high rates while others may have evolved mechanisms to resist the consequences of
protein glycation. We should also bear in mind that some taxonomic groups may be
underrepresented in our study, or biases in species selection due to availability contingencies
(e.g. species common in zoos or present in European countries) may exist, so further studies
should target this underrepresented groups in order to confirm our predictions.
As expected for a non-enzymatic reaction, just by the law of mass action, bird albumin glycation
rates increase with plasma glucose concentration. However, the logarithmic nature of the
relationship, and the fact that the slope is lower than one, suggest that species with higher
plasma glucose levels exhibit relatively greater resistance to glycation. This finding aligns with
previous research indicating that *in vitro* glycation levels of chicken albumin increase at a slower

rate than those of human albumin when glucose concentration is elevated [83]. Moreover, these

levels are consistently lower than those of bovine serum albumin regardless of glucose

concentration and exposition time [37]. As discussed in previous studies comparing chicken to

bovine serum albumin [37], or zebra finch to human haemoglobin [53], the lower glycation rates

observed in bird proteins may result from a lower number of glycatable amino acids (e.g. lysines)

in their sequence and/or their lesser exposure at the protein surface.

Our analyses do not succeed in indicating a significant positive relationship between average

glycation levels and the number of glycatable lysines in the albumin sequence. This may be

attributed to the limited number of species employed or the weak variation in the number of

glycatable lysine residues, which are mostly ranging from 33 to 39. An interesting exception are

the 18 glycatable lysine residues of flamingos (*Phoenicopterus ruber*), which also shows very low

glycation levels (mean=10.1%). However, the exceptions of 44 in zebra finches and 20 in godwits

(*Limosa lapponica*, used in place of *Limosa limosa*) are nevertheless associated with very similar

average glycation levels of 20.2% and 20.7% respectively.

Plasma glucose relates with longevity and may be influenced by reproductive

strategies

Our results were only in minor agreement with predictions from POLS theory: it holds for body

mass, but not for the other life history variables tested. The only previous comparative study of

glycaemia and life history traits to our knowledge [14] shows no relationship with mass-adjusted

maximum lifespan in passerines. However, our study over 88 bird species on 22 orders revealed

an increase in glucose with mass-adjusted longevity up to a plateau (see **Figure 2.B**). Thus, the

relationship between glucose and maximum lifespan may depend on differences between bird

orders or be tied to specific species particularities not explored in our study. Such species

particularities might involve additional undetermined ecological factors that modify the

relationship of glycaemia with longevity. Further exploration of glucose metabolism in relation

with lifestyle will bring further light on species-specific life-history adaptations concerning

glucose. For example, the species with the lower mass-adjusted maximum lifespan here was a

cormorant (*Leucocarbo verrucosus*), which have quite low glucose values for birds.

Regarding reproductive investment (i.e. clutch mass), our results show no relationship with

glycation (see **Table 2**), while previous studies reported positive relationships with glycaemia in

passerines [14, 83]. Interestingly, most of the species with high clutch mass included in our study

belong to the Anseriformes (**Figure ESM2.1**). While these species exhibit very low glycaemia and

albumin glycation rates, they are also characterized by a particular reproductive strategy

compared to passerines, for whom clutch mass does not imply the same in terms of parental

investment. For instance, Anseriformes, unlike Passeriformes, are precocial and their offspring

are not very dependent on parental care. Furthermore, they are capital breeders, accumulating

the energetic resources required for reproduction in advance rather than collecting them during

reproduction. These species typically have large clutch sizes and incubation is usually carried out

by females, who store internal fat resources to support this main part of the parental

investment. In addition, ducklings are born highly developed, reducing the amount of care

required post-hatching (see e.g. [84-85]). Consequently, their dependence on glucose as a rapid

energy source for reproduction may be lower, with investment in this activity likely more closely

linked to lipid accumulation. This could explain why we did not detect previously reported

effects of clutch mass on glucose levels.

Terrestrial carnivores show a paradoxically increased albumin glycation rate without

increased plasma glucose levels

Contrary to our expectation of finding differences across dietary groups, plasma glucose did not

significantly vary with species diet. This aligns with results previously reported by [14] for

Passeriformes or by [55] for 160 species of vertebrates among which 48 were birds, but not with

[15], that showed glycaemia to be increased with the proportion of fruits/seeds in the diet, or

with [56], that showed higher (mass-adjusted) glucose levels for terrestrial carnivores and

insectivores (those included in our terrestrial carnivores' category) and lower for

frugivorous/nectarivorous. On the other hand, intraspecific data indicates that changes in diet

composition hardly affects glycaemia in birds [86]. These studies suggest that glycaemia is tightly

regulated independently of dietary composition within species, while it probably varies across

species depending on the diet they are adapted to, although depending on the bird groups

included in the analyses and the way of assessing it.

Although terrestrial carnivore species did not have significantly higher glycaemia levels in our

study, they nonetheless demonstrated significantly higher albumin glycation rates, suggesting a

potential susceptibility to protein glycation. Besides unique structural features, this

phenomenon could be due to lower albumin turnover rates in terrestrial carnivores. Studies in

humans have linked higher oxidative damage to albumin with lower turnover rates (reviewed in

[87]), which may extend to other species and post-translational modifications of albumin, such

as bird's albumin glycation. However, the contrary outcome would be anticipated if protein intake were high as seen in carnivorous species [87-88].

Interestingly, the contrast between terrestrial and aquatic predators, which did not show such high glycation rates, suggests the beneficial influence of a substantial difference in food composition or the pre-eminence of another ecological factor, yet to be determined, which may explain lower glycation levels in aquatic predators. In terms of food composition, the proportion of essential n-3 PUFA (polyunsaturated fatty acids), particularly long-chain ones such as DHA and EPA compared with n-6 PUFA and short-chain n-3 PUFA, is different in aquatic and terrestrial environments [89] and therefore in the diet of predators in each environment (e.g. [90]). For instance, low n-6 PUFA/n3- PUFA ratio have been shown to decrease insulin levels and improve insulin resistance in humans [91]. On the other hand, while some studies report that an increase in dietary n3-PUFA reduces the levels of glycated haemoglobin (HbA1c) in humans, this effect was not detected in many other studies and therefore remains inconclusive (reviewed in [92]). The detailed study of the fat composition of the diets of terrestrial and aquatic predators, in particular the ratio between different types of PUFA, merits more attention. In addition, micronutrient content, such as circulating antioxidant defences, should also be taken into consideration. Indeed, a positive relationship between oxidative stress and glycated levels of bovine serum albumin has been reported [93]. One hypothesis is that terrestrial predators have higher systemic oxidative stress levels compared to other species, which may be explained by defects in their antioxidant defences. Uric acid is one of the main non-enzymatic antioxidants in birds [94-96], and uric acid levels are especially high in carnivore species that have a rich protein diet [95, 97-100]. We should therefore take into account other important antioxidants such as vitamin E, vitamin A and carotenoids, which may be less abundant in the diets of terrestrial carnivores (e.g. [101], but see [52]). The question of whether the diet of aquatic carnivores provides a better intake of antioxidants would therefore requires a more detailed description of dietary habits. The herbivorous diet, meanwhile, despite the expected possibility contributing to lower glycaemia and glycation levels due to higher levels of PUFA compared to SFA (saturated fatty acids) (effects reviewed in [92] for humans) and higher fibre content (see [102]), did not lead to significantly lower levels of either of these two parameters. However, the relatively low sample size in that group or the presence of outliers as *Nesoenas mayeri*, with the highest glucose levels of this dataset, makes the interpretation of the obtained results somehow limited. Therefore, further research should be carried out on these species to determine if the expected pattern would emerge with a better sampling.

Finally, differences between captive and wild populations could be considered as a source of

variation in glucose or glycation levels, due to a more sedentary lifestyle in captivity, with lower

activity levels and a higher food intake likely to lead to increased circulating glucose levels in

captive individuals. Additionally, differences in nutrient intake, such as antioxidants, specific

fatty acids or amino acids, between captive and wild populations could contribute to variations

in glycation levels, as we saw above. Nevertheless, we think that this factor is unlikely to

significantly affect our findings regarding diet categories, because our study encompasses

species from both captive and wild populations across various diet groups, particularly those

exhibiting significant differences (e.g., omnivores versus terrestrial carnivores).

CONCLUSION AND PERSPECTIVES

In conclusion, the avian plasma glucose levels measured here are generally higher or in the high

range of values recorded for mammalian species [28]. Our study also concludes that there is

considerable variation in plasma glucose levels and albumin glycation rates between bird

species, with those with the highest glucose levels showing greater resistance to glycation. The

correlation between plasma glucose and life history traits are primarily influenced by its inverse

association with body mass, along with non-linear, mass-independent effects of longevity.

Finally, although diet does not explain plasma glucose levels in our study, terrestrial carnivores

have higher albumin glycation rates than omnivores. Whether these intriguing results are

explained by specific effects of certain dietary components, such as PUFAs and/or antioxidants,

remains to be determined. Differences in plasma glucose levels and albumin glycation rates

across bird species do not seem consistent with predictions according to the POL hypothesis,

except for body mass. Further investigation is needed to elucidate the correlation between

these traits and specific life conditions, such as reproductive strategy, migration patterns, flight

mode or more detailed diet composition. In addition, more in-depth exploration of glycation

levels within high glycaemic groups such as the Passeriformes and other small birds, which make

up a significant proportion of avian species, could provide valuable new insights. Similarly,

investigating groups such as the Phoenicopteriformes or Anseriformes, which are at the other

end of the glycaemic-glycation spectrum, could shed light on the origin of differences between

avian orders. Furthermore, notable variations were observed between species within orders, as

in the Procellariiformes, which with their high mass-adjusted longevity, and particularly given

that some of the age known individuals in our dataset (indeed, two specimens of snow petrels,

Pagodroma nivea, showed ages higher than the maximum lifespan reported in the sources we

explored, determining a new record) have still relatively low levels of glycation, may suggest that
some species (the ones that do show higher glycation levels) are not intrinsically resistant to
glycation, but rather to its adverse consequences for health. As the main limitations from this
study, the usage of individuals from both wild and captive populations, sampled at different
periods of the year, and the low sample size for some species, due to logistic constraints, may
have introduced noise in the values reported that should be addressed in future studies by
implementing a stricter sampling protocol. Also, a more thorough and accurate report and
compilation of life history traits from multiple species would allow to increase the number of
species included in this kind of analyses. Future research should also focus on specific species to
unravel the physiological mechanisms mediating the effects of blood glucose and protein
glycation on life-history trade-offs, in particular mechanisms that may vary between taxa and
contribute to characteristic adaptations in birds to mitigate the potentially negative effects of
comparatively high glycaemia.

CONFLICTS OF INTEREST

575 None declared.

DATA ACCESSIBILITY

577 All data used, including references for the values taken from bibliography or databases, will be
578 available as Electronic Supplementary Material (ESM5).

ETHICS STATEMENT

This study followed all the legal considerations, with the ethic authorisations from the French
Ministry of Secondary Education and Research, n°32475 for the zebra finches sampling, the
Swiss Veterinary Office (FSVO) nº 34497 for the Alpine swifts, the Ethics Committee of the
University of Extremadura (licenses112//2020 and 202//2020) and the Government of
Extremadura (licenses CN0012/22/ACA and CN0063/21/ACA) for the godwits and terns from
Spain, and Sampling in Terres Australes et Antarctic Françaises was approved by a Regional
Animal Experimentation Ethical Committee (French Ministry of Secondary Education and
Research permit APAFIS #31773-2019112519421390 v4 and APAFIS#16465–
2018080111195526 v4) and by the Comité de l'Environnement Polaire and CNPN (A-2021-55).

The samples from the Mulhouse zoo were taken by its licensed veterinary with capacity number:

2020-247-SPAE-162.

ACKNOWLEDGMENTS

This research was funded by an ANR (AGEs – ANR21-CE02-0009). We thank Charles-André Bost

(CEBC CNRS) for providing data on some species. We would like to thank the SYLATR Association

for collecting samples of wild Passeriformes as part of the MIGROUILLE program, and Manuela

Forero and Frederic Angelier (CEBC CNRS) for providing some samples of certain

Procellariiformes. We thank the French Parc des Oiseaux (Villars les Dombes), the bird keepers

and veterinarians at Mulhouse zoo for their contribution to the collection of blood samples and

access to individual data from their captive birds. Data on seabirds from the French Southern

Territories was collected within the framework of the ECONERGY (119), ECOPATH (1151) and

ORNITHOECO (109) programs of the French Polar Institute (IPEV). These studies are part of the

long-term Studies in Ecology and Evolution (SEE-Life) program of the CNRS. We are grateful to

Mathilde Lejeune, Natacha Garcin and Camille Lemonnier for their help in collecting those

samples. We are thankful to Orsolya Vincze for producing Figure 1, Adrien Brown for

determining the number of lysines exposed in albumin sequences, Claire Saraux for her help

with the statistics and F. Stephen Dobson for reading and commenting on the manuscript.

Finally, we are thankful to Ascensia Diabetes care® for their generous donation of glucometers

and strips for glucose measurement.

AUTHORS' CONTRIBUTIONS

F. Criscuolo and F. Bertile conceived the idea, directed most of the sample collection and logistics

and contributed significantly to the writing. A. Moreno-Borrallo contributed to the development

of the questions, gathered the diet and life-history data, performed the statistical analyses, and

led the writing. S. Jaramillo Ortiz and C. Schaeffer performed the mass spectrometry analyses

for protein glycation measurements, and S. Jaramillo Ortiz contributed to the glucose measures

and commented on the manuscript. T. Boulinier. and V. A. Viblanc coordinated the ECONERGY

and ECOPATH polar programs, organised the collection of samples on subantarctic seabirds, and

commented on the manuscript. Olivier Chastel collaborated collecting marine bird samples and

commented on the manuscript. B. Rey contributed with the collection of samples from Parc des

Oiseaux (Villars-les-Dombes, France) and commented on the manuscript. P. Bize leads the

monitoring of Alpine swifts' populations from which the samples were obtained, he helped collecting them and commented on the manuscript. J. S Gutiérrez and J. A. Masero contributed with samples from Spain and part of the statistic scripts and commented on the manuscript. B. Quintard leads the health monitoring of Mulhouse zoo bird collection, organized and realized most of Mulhouse zoo samplings and commented on the manuscript. All authors gave their approval for publication.

ORCID

AMB: 0000-0002-2924-1153

SAJ: 0000-0002-9153-4205

CS: 0000-0003-0672-1979

BQ: 0000-0001-7905-886X

BR: 0000-0002-0464-5573

PB: 0000-0002-6759-4371

VV: 0000-0002-4953-659X

TB: 0000-0002-5898-7667

OC: 0000-0002-9435-1041

JSG: 0000-0001-8459-3162

JAM: 0000-0001-5318-4833

FB: 0000-0001-5510-4868

FC: 0000-0001-8997-8184

REFERENCES

[1]. Wikelski, Martin and Robert E. Ricklefs. 2001. The physiology of life histories. *Trends in Ecology & Evolution*, 16 (9): 479–81. DOI:https://doi.org/10.1016/S0169-5347(01)02279-0

[2]. Ricklefs, Robert E. and Martin Wikelski. 2002. The physiology/life-history nexus. Trends in Ecology and Evolution, 17 (10): 462–68. https://doi.org/10.1016/S0169-5347(02)02578-8

[3]. Wikelski, Martin, Laura Spinney, Wendy Schelsky, Alexander Scheuerlein and Eberhard Gwinner. 2003. Slow pace of life in tropical sedentary birds: a common-garden experiment on four stonechat populations from different latitudes. *Proceedings of the Royal Society London B: Biological Sciences*, 270: 2383–88. https://doi.org/10.1098/rspb.2003.2500.

[4]. Stott, Iain, Roberto Salguero-Gómez, Owen R. Jones, Thomas H.G. Ezard, Marlène Gamelon, Shelly Lachish, Jean Dominique Lebreton, Emily G. Simmonds, Jean Michel Gaillard and Dave J. Hodgson. 2024. Life histories are not just fast or slow. *Trends in Ecology and Evolution*, 3319: 1–11. https://doi.org/10.1016/j.tree.2024.06.001.

[5]. Stearns, Stephen C. 1989. Trade-offs in life-history evolution. *Functional Ecology*, 3, (3): 259-268. https://doi.org/10.2307/2389364

[6]. Gaillard, Jean-Michel and Dominique Pontier. 1989. An analysis of demographic tactics in birds and mammals. *Oikos*, 56: 59-76. https://doi.org/10.2307/3566088.

[7]. Stearns, Stephen C. 1992. The evolution of life histories. Oxford university press.

[8]. Vasilieva, N. A. 2022. Pace-of-Life Syndrome (POLS): evolution of the concept. *Biology bulletin*, 49 (7): 750–62, translated by M. Batrukova. https://doi.org/10.1134/S1062359022070238.

[9]. Trevelyan, R., P. H. Harvey and M. D. Pagel. 1990. Metabolic rates and life histories in birds. *Functional Ecology*, 4, (2), 135-141. https://doi.org/10.2307/2389332

[10]. Austad, Steven N. and Kathleen E. Fischer. 1991. Mammalian aging, metabolism, and ecology: evidence from the bats and marsupials. *Journal of Gerontology: Biological sciences*, 46 (2): 47–53. https://doi.org/10.1093/geronj/46.2.B47

[11]. Speakman, John R. 2005. Body size, energy metabolism and lifespan. *Journal of Experimental Biology*, 208 (9): 1717–30. https://doi.org/10.1242/jeb.01556.

[12]. Magalhães, João Pedro de, Joana Costa and George M. Church. 2007. An analysis of
the relationship between metabolism, developmental schedules, and longevity using
Phylogenetic Independent Contrasts. *The Journals of Gerontology: Series A*, 62 (2): 149–
60. https://doi.org/10.1093/gerona/62.2.149

[13]. Kjeld, M and Ö. Ólafsson. 2007. Allometry (scaling) of blood components in mammals:
connection with economy of energy? *Canadian Journal of Zoology*, 899: 890–99.
https://doi.org/10.1139/Z08-061.

[14]. Tomášek, Oldřich, Lukáš Bobek, Tereza Kauzálová, Marie Adamkova and Tomáš
Albrecht. 2019. Fuel for the pace of life: baseline blood glucose concentration co-evolves
with life-history traits in songbirds. *Functional Ecology*, 33 (2): 239–49.
https://doi.org/10.1111/1365-2435.13238

[15]. Tomášek, Oldřich, Lukáš Bobek, Tereza Kauzálová, Ondřej Kauzál, Marie Adámková,
Kryštof Horák, Sampath Anandan Kumar, Judith Pouadjeu Manialeu, Pavel Munclinger,
Eric Djomo Nana, Télesphore Benoît Nguelefack, Ondřej Sedláček, Tomáš Albrecht.
2022. Latitudinal but not elevational variation in blood glucose level is linked to life
history across passerine birds. *Ecology Letters*, 25 (10): 2203-2216.
https://doi.org/10.1111/ele.14097

[16]. Vágási, Csongor I., Orsolya Vincze, Marie Adámkova, Tereza Kauzálova, Ádam Z.
Lendvai, Laura I. Pătras, Janka Pénzes, Péter L. Pap, Tomáš Albrecht and Oldřich
Tomášek. 2024. Songbirds avoid the oxidative stress costs of high blood glucose levels:
a comparative study. *Journal of Experimental Biology*, 227: 1-8.
https://doi.org/10.1242/jeb.246848.

[17]. Umminger, Bruce L. 1977. Relation of whole blood sugar concentrations in vertebrates
to standard metabolic rate. *Comparative Biochemistry and Physiology A*, 56, 457-460.

[18]. Guo, Hai, Dilihumaier Duolikun and Qiaoling Yao. 2023. Associations between basal
metabolic rate and insulin resistance in non-diabetic obese adults: evidence from
NHANES 2011–2018. International Journal of Diabetes in Developing Countries.
https://doi.org/10.1007/s13410-023-01190-8.

[19]. Maillard, L. C. 1912. Action des acides amines sur les sucres; formation des melanoidies
par voie methodique. *Comptes rendus de l'Académie des sciences*, 154, 66-68.

[20]. Suji, George and S. Sivakami. 2004. Glucose, glycation and aging. *Biogerontology*, 5:
365–373. https://doi.org/10.1007/s10522-004-3189-0

[21]. Cerami, Anthony, Helen Vlassara and Michael Brownlee. 1986. Role of nonenzymatic
glycosylation in atherogenesis. *Journal of Cellular Biochemistry*, 30:111-120.
https://doi.org/10.1002/jcb.240300203

[22]. Poulsen, Malene W., Rikke V. Hedegaard, Jeanette M. Andersen, Barbora de Courten,
Susanne Bügel, John Nielsen, Leif H. Skibsted and Lars O. Dragsted. 2013. Advanced
Glycation End-products in food and their effects on health. *Food and Chemical*
Toxicology, 60: 10–37. https://doi.org/10.1016/j.fct.2013.06.052.

[23]. Chaudhuri, Jyotiska, Yasmin Bains, Sanjib Guha, Arnold Kahn, David Hall, Neelanjan
Bose, Alejandro Gugliucci and Pankaj Kapahi. 2018. The role of advanced glycation end
products in aging and metabolic diseases: bridging association and causality. *Cell*
Metabolism, 28 (3): 337–52. https://doi.org/10.1016/j.cmet.2018.08.014.

[24]. Khalid, Mariyam, Georg Petroianu and Abdu Adem. 2022. Advanced Glycation End
products and diabetes mellitus: mechanisms and perspectives. *Biomolecules*, 12 (4),
542. https://doi.org/10.3390/biom12040542

[25]. Twarda-Clapa, Aleksandra, Aleksandra Olczak, Aneta M. Białkowska and Maria
Koziołkiewicz. 2022. Advanced Glycation End-products (AGEs): formation, chemistry,
classification, receptors, and diseases related to AGEs. *Cells*, 11(8): 1312.
https://doi.org/10.3390/cells11081312

[26]. Hofmann, Marion A., Steven Drury, Caifeng Fu, Wu Qu, Akihiko Taguchi, Yan Lu, Cecilia
Avila, Neeraja Kambham, Angelika Bierhaus, Peter Nawroth, Markus F. Neurath,
Timothy Slattery, Dale Beach, John McClary, Mariko Nagashima, John Morser, David
Stern and Ann Marie Schmidt. 1999. RAGE mediates a novel proinflammatory axis: a
central cell surface receptor for S100/Calgranulin polypeptides. *Cell*, 97 (7): 889–901.
DOI:https://doi.org/10.1016/S0092-8674(00)80801-6

[27]. Schmidt, Ann Marie, Shi Du Yan, Shi Fang Yan and David M. Stern. 2001. The multiligand
receptor RAGE as a progression factor amplifying immune and inflammatory responses.
Journal of Clinical Investigation, 108 (7): 949–55.
https://doi.org/10.1172/JCI200114002.

[28]. Polakof, Sergio, Thomas P. Mommsen and José L. Soengas. 2011. Glucosensing and
glucose homeostasis: from fish to mammals. *Comparative Biochemistry and Physiology,*
Part B 160 (4): 123–49. https://doi.org/10.1016/j.cbpb.2011.07.006.

[29]. Lasiewski, Robert C. and William R. Dawson. 1967. A re-examination of the relation
between standard metabolic rate and body weight in birds. *The Condor*, 69 (1): 13–23.
https://doi.org/10.2307/1366368

[30]. King, J. R. and Farner, D. S. (1961). Energy metabolism, thermoregulation and body
temperature. In Biology and Comparative Physiology of Birds, vol. II (ed. A. J. Marshall),
pp. 215–288. New York: Academic Press.

[31]. Dawson, Terence J. and A. J. Hulbert. 1970. Standard metabolism, body temperature,
surface areas of australian marsupials. *American Journal of Physiology*, 218 (4).
https://doi.org/10.1152/ajplegacy.1970.218.4.1233

[32]. Marsh, Richard L., David J. Ellerby, Jennifer A. Carr, Havalee T. Henry and Cindy I.
Buchanan. 2004. Partitioning the energetics of walking and running: swinging the limbs
is expensive. *Science*, 303 (5654): 80-83 https://doi.org/10.1126/science.1090704.

[33]. Weber, Jean-Michel. 2011. Metabolic fuels: regulating fluxes to select mix. *Journal*
Experimental Biology, 214 (2): 286–94. https://doi.org/10.1242/jeb.047050.

[34]. Braun, Eldon J. and Karen L. Sweazea. 2008. Glucose regulation in birds. *Comparative*
Biochemistry and Physiology - B Biochemistry and Molecular Biology, 151 (1): 1–9.
https://doi.org/10.1016/j.cbpb.2008.05.007.

[35]. Szwergold, Benjamin S. and Craig B. Miller. 2014b. Potential of birds to serve as
pathology-free models of type 2 diabetes, Part 2: Do high levels of carbonyl-scavenging
amino acids (e.g., taurine) and low concentrations of methylglyoxal limit the production
of Advanced Glycation End-products? *Rejuvenation Research*, 17 (4): 347–58.
https://doi.org/10.1089/rej.2014.1561.

[36]. Lindstedt, Stan L. and William A. Calder. 1976. Body size and longevity in birds. *The*
Condor, 78 (1): 91-94. https://doi.org/10.2307/1366920

[37]. Anthony-Regnitz, Claire M., Amanda E. Wilson, Karen L. Sweazea and Eldon J. Braun.
2020. Fewer exposed lysine residues may explain relative resistance of chicken serum
albumin to in vitro protein glycation in comparison to bovine serum albumin. *Journal of*
Molecular Evolution, 88 (8–9): 653–61. https://doi.org/10.1007/s00239-020-09964-y.

[38]. Muramatsu, Tatsuo. 1990. Nutrition and whole-body protein turnover in the chicken
in relation to mammalian species. *Nutrition Research reviews*, 3: 211-228

[39]. Makino, Ryosuke and Kazumi Kita. 2018. Half-Life of Glycated Tryptophan in the Plasma
of Chickens. *Journal of Poultry Science*, 55 (2): 117–19.
https://doi.org/10.2141/jpsa.0170158.

[40]. Schweigert, F. J., Stephanie Uehlein-Harrell, Giesela v. Hegel and H. Wiesner. 1991. Vitamin A (retinol and retinyl esters), α-tocopherol and lipid levels in plasma of captive wild mammals and birds. *Journal of Veterinary Medicine Series A*, 42: 35–42. https://doi.org/10.1111/j.1439-0442.1991.tb00981.x

[41]. Ku, Hung-Hai and R.S. Sohal. 1993. Comparison of mitochondrial prooxidant generation and antioxidant defenses between rat and pigeon possible basis of variation in longevity and metabolic potential. *Mechanisms of Ageing and Development*, 72: 67–76. https://doi.org/10.1016/0047-6374(93)90132-B

[42]. Hickey, Anthony J.R., Mia Jüllig, Jacqueline Aitken, Kerry Loomes, Mark E. Hauber and Anthony R.J. Phillips. 2012. Birds and longevity: does flight driven aerobicity provide an oxidative sink? *Ageing Research Reviews*, 11 (2): 242–53. https://doi.org/10.1016/j.arr.2011.12.002.

[43]. Eythrib, Jalil. 2013. The search for the receptor for Advanced Glycation End-products in avian vasculature. Bachelor thesis, The University of Arizona.

[44]. Szwergold, Benjamin S. and Craig B. Miller. 2014a. Potential of birds to serve as a pathology-free model of type 2 diabetes, Part 1: Is the apparent absence of the rage gene a factor in the resistance of avian organisms to chronic hyperglycemia? *Rejuvenation Research*, 17 (1): 54–61. https://doi.org/10.1089/rej.2013.1498.

[45]. Wein, Yossi, Enav Bar Shira and Aharon Friedman. 2020. Increased serum levels of advanced glycation end products due to induced molting in hen layers trigger a proinflammatory response by peripheral blood leukocytes. *Poultry Science*, 99 (7): 3452–62. https://doi.org/10.1016/j.psj.2020.04.009.

[46]. Rendell, Marc, P. M. Stephent, Robert Paulsent, J. L. Valentine, Kathy Rasbold, T. I. M. Hestorfe, Susan Eastberg and Daniel C Shintt. 1985. An interspecies comparison of normal levels of glycosylated hemoglobin and glycosylated albumin. *Comparative Biochemistry and physiology. B, Comparative Biochemistry*, 81 (4): 819–22. https://doi.org/10.1016/0305-0491(85)90072-0

[47]. Miksik, I. and Z. Hodny. 1992. Glycated hemoglobin in mute swan (*Cygnus olor*) and rook (*Corvus frugilegus*). *Comparative Biochemistry and Physiology Part B: Comparative Biochemistry*, 103 (3): 553–55. https://doi.org/10.1016/0305-0491(92)90369-3

[48]. Andersson, M. S. and L. Gustafsson. 1995. Glycosylated haemoglobin: a new measure of condition in birds. *Proceedings of the Royal Society B: Biological Sciences*, 260 (1359): 299–303. https://doi.org/10.1098/rspb.1995.0095.

[49]. Beuchat, Carol A. and Curtis R. Chong. 1998. Hyperglycemia in hummingbirds and its
consequences for hemoglobin glycation. *Comparative Biochemistry and Physiology - A*
Molecular and Integrative Physiology, 120 (3): 409–16. https://doi.org/10.1016/S1095-
6433(98)10039-9.

[50]. Ardia, Daniel R. 2006. Glycated hemoglobin and albumin reflect nestling growth and
condition in american kestrels. *Comparative Biochemistry and Physiology - A Molecular*
and Integrative Physiology, 143 (1): 62–66. https://doi.org/10.1016/j.cbpa.2005.10.024.

[51]. Récapet, Charlotte, Adélaïde Sibeaux, Laure Cauchard, Blandine Doligez and Pierre
Bize. 2016. Selective disappearance of individuals with high levels of glycated
haemoglobin in a free-living bird. *Biology Letters*, 12: 20160243.
https://doi.org/10.1098/rsbl.2016.0243.

[52]. Ingram, Tana, Jessica Zuck, Chad R. Borges, Patrick Redig and Karen L. Sweazea. 2017.
Variations in native protein glycation and plasma antioxidants in several birds of prey.
Comparative Biochemistry and Physiology Part - B: Biochemistry and Molecular Biology,
210 (January): 18–28. https://doi.org/10.1016/j.cbpb.2017.05.004.

[53]. Brun, Charlotte, Oscar Hernandez-Alba, Agnès Hovasse, François Criscuolo, Christine
Schaeffer-Reiss and Fabrice Bertile. 2022. Resistance to glycation in the zebra finch:
mass spectrometry-based analysis and its perspectives for evolutionary studies of aging.
Experimental Gerontology, 164 111811. https://doi.org/10.1016/j.exger.2022.111811.

[54]. Borger, M. J. (2024). How to prime your offspring: putting behavioural ecology to the
test. PhD thesis. University of Groningen. https://doi.org/10.33612/diss.849300689

[55]. Kapsetaki, Stefania E., Anthony J. Basile, Zachary T. Compton, Shawn M. Rupp, G. Duke,
Amy M. Boddy, Tara M. Harrison, Karen L. Sweazea and Carlo C. Maley. 2023. The
relationship between diet, plasma glucose, and cancer prevalence across vertebrates.
Version 1. bioRxiv. Preprint. 2023 Aug 2. doi: 10.1101/2023.07.31.551378

[56]. Szarka E.Z. and Lendvai Á.Z. Trophic guilds differ in blood glucose concentrations: a
phylogenetic comparative analysis in birds. 2024. *Proceedings of the Royal Society B:*
Biological Sciences, 291: 20232655

[57]. Coulson, Roland A. and Thomas Hernandez. 1983. Alligator Metabolism studies on
chemical reactions in vivo. *Comparative Biochemistry and Physiology Part B:*
Comparative Biochemistry, 74(1), 1–175. https://doi.org/10.1016/0305-
0491(83)90418-2

[58]. Dobbs, J. C. (1985). Avian carnivore: food recognition, nutrient intakes, and utilization
of nitrogen. PhD thesis, University of California-Davis, Davis, CA.

[59]. Verbrugghe, Adronie and Myriam Hesta. 2017. Cats and carbohydrates: the carnivore
fantasy? *Veterinary Sciences*, 4 (55): 1–22. https://doi.org/10.3390/vetsci4040055.

[60]. Migliorini, R. H., J. C. Linder, L. Moura and J. A. S. Veiga. 1973. Gluconeogenesis in a
carnivorous bird (black vulture). *American journal of physiology*, 225 (6): 1389–1392.
https://doi.org/10.1152/ajplegacy.1973.225.6.1389

[61]. Myers, Merrick R. and Kirk C. Klasing. 1999. Low glucokinase activity and high rates of
gluconeogenesis contribute to hyperglycemia in barn owls (*Tyto alba*) after a glucose
challenge. *The Journal of Nutrition*, 129 (10): 1896–1904.
https://doi.org/10.1093/jn/129.10.1896

[62]. Stearns, Stephen C. 1983. The Influence of size and phylogeny on patterns of
covariation among life-history traits in the mammals. *Oikos*, 41 (2): 173-187.

[63]. Umminger, Bruce L. 1975. Body size and whole blood sugar concentrations in
mammals. *Comparative Biochemistry and Physiology A*, 52, 455-458.

[64]. Scanes, Colin G. 2016. A re-evaluation of allometric relationships for circulating
concentrations of glucose in mammals. *Food and Nutrition Sciences*, 7 (4): 240–51. DOI:
10.4236/fns.2016.74026

[65]. Mohsenzadeh, Mahdieh Sadat, Mahdieh Zaeemi, Jamshid Razmyar and Mohammad
Azizzadeh. 2015. Comparison of a point-of-care glucometer and a laboratory
autoanalyzer for measurement of blood glucose concentrations in domestic pigeons
(*Columba livia domestica*). *Journal of Avian Medicine and Surgery*, 29, 3: 181–86.
https://doi.org/10.1647/2014-020.

[66]. Morales, Ana, Barbara Frei, Casey Leung, Rodger Titman, Shannon Whelan, Z. Morgan
Benowitz-Fredericks and Kyle H. Elliott. 2020. Point-of-care blood analyzers measure
the nutritional state of eighteen free-living bird species. *Comparative Biochemistry and*
Physiology -Part A: Molecular and Integrative Physiology, 240, 110594: 1095–6433.
https://doi.org/10.1016/j.cbpa.2019.110594.

[67]. Downs, Colleen T., Andrea E. Wellmann and Mark Brown. 2010. Diel variations in
plasma glucose concentrations of malachite sunbirds *Nectarinia famosa*. *Journal of*
Ornithology, 151: 235–39. https://doi.org/10.1007/s10336-009-0439-6.

[68]. Breuner, Creagh W., Brendan Delehanty and Rudy Boonstra. 2013. Evaluating stress
in natural populations of vertebrates: total CORT is not good enough. *Functional*

868   *Ecology*, 27: 24–36. https://doi.org/10.1111/1365-2435.12016.

869 [69]. McGraw, Kevin J., Katherine Chou, Annika Bridge, Hannah C. McGraw, Peyton R.
870   McGraw and Richard K. Simpson. 2020. Body condition and poxvirus infection predict
871   circulating glucose levels in a colorful songbird that inhabits urban and rural
872   environments. *Journal of Experimental Zoology*, 333: 561–68.
873   https://doi.org/10.1002/jez.2391.

874 [70]. Priego-Capote, F., Ramirez-Boo, M., Finamore, F., Gluck, F. and Sanchez, J.C. 2014.
875   Quantitative analysis of glycated proteins. *Journal of Proteome Research*, 13, 2: 336–
876   347.

877 [71]. Zuck, Jessica, Chad R. Borges, Eldon J. Braun and Karen L. Sweazea. 2017. Chicken
878   albumin exhibits natural resistance to glycation. *Comparative Biochemistry and*
879   *Physiology Part - B: Biochemistry and Molecular Biology*, 203: 108–114.
880   https://doi.org/10.1016/j.cbpb.2016.10.003.

881 [72]. Tobias, Joseph A., Catherine Sheard, Alex L. Pigot, Adam J. M. Devenish, Jingyi Yang,
882   Ferran Sayol, Montague H. C. Neate-Clegg, Nico Alioravainen, Thomas L. Weeks, Robert
883   A. Barber, Patrick A. Walkden, Hannah E. A. MacGregor, Samuel E. I. Jones, Claire
884   Vincent, Anna G. Phillips, Nicola M. Marples, Flavia A. Montaño-Centellas, Victor
885   Leandro-Silva, Santiago Claramunt, Bianca Darski, Benjamin G. Freeman, Tom P.
886   Bregman, Christopher R. Cooney, Emma C. Hughes, Elliot J. R. Capp, Zoë K. Varley,
887   Nicholas R. Friedman, Heiko Korntheuer, Andrea Corrales-Vargas, Christopher H. Trisos,
888   Brian C. Weeks, Dagmar M. Hanz, Till Töpfer, Gustavo A. Bravo, Vladimír Remeš, Larissa
889   Nowak, Lincoln S. Carneiro, Amilkar J. Moncada R., Beata Matysioková, Daniel T.
890   Baldassarre, Alejandra Martínez-Salinas, Jared D. Wolfe, Philip M. Chapman, Benjamin
891   G. Daly, Marjorie C. Sorensen, Alexander Neu, Michael A. Ford, Rebekah J. Mayhew, Luis
892   Fabio Silveira, David J. Kelly, Nathaniel N. D. Annorbah, Henry S. Pollock, Ada M.
893   Grabowska-Zhang, Jay P. McEntee, Juan Carlos T. Gonzalez, Camila G. Meneses, Marcia
894   C. Muñoz, Luke L. Powell, Gabriel A. Jamie, Thomas J. Matthews, Oscar Johnson,
895   Guilherme R. R. Brito, Kristof Zyskowski, Ross Crates, Michael G. Harvey, Maura Jurado
896   Zevallos, Peter A. Hosner, Tom Bradfer-Lawrence, James M. Maley, F. Gary Stiles,
897   Hevana S. Lima, Kaiya L. Provost, Moses Chibesa, Mmatjie Mashao, Jeffrey T. Howard,
898   Edson Mlamba, Marcus A. H. Chua, Bicheng Li, M. Isabel Gómez, Natalia C. García,
899   Martin Päckert, Jérôme Fuchs, Jarome R. Ali, Elizabeth P. Derryberry, Monica L. Carlson,
900   Rolly C. Urriza, Kristin E. Brzeski, Dewi M. Prawiradilaga, Matt J. Rayner, Eliot T. Miller,

Rauri C. K. Bowie, René-Marie Lafontaine, R. Paul Scofield, Yingqiang Lou, Lankani Somarathna, Denis Lepage, Marshall Illif, Eike Lena Neuschulz, Mathias Templin, D. Matthias Dehling, Jacob C. Cooper, Olivier S. G. Pauwels, Kangkuso Analuddin, Jon Fjeldså, Nathalie Seddon, Paul R. Sweet, Fabrice A. J. DeClerck, Luciano N. Naka, Jeffrey D. Brawn, Alexandre Aleixo, Katrin Böhning-Gaese, Carsten Rahbek, Susanne A. Fritz, Gavin H. Thomas and Matthias Schleuning. 2022. AVONET: Morphological, ecological and geographical data for all birds. *Ecology Letters*, 25: 581–97. https://doi.org/10.1111/ele.13898

[73]. Pigot, Alex L., Catherine Sheard, Eliot T. Miller, Tom P. Bregman, Benjamin G. Freeman, Uri Roll, Nathalie Seddon, Christopher H. Trisos, Brian C. Weeks and Joseph A. Tobias. 2020. Macroevolutionary convergence connects morphological form to ecological function in birds. *Nature Ecology & Evolution*, 4: 230–239. https://doi.org/10.1038/s41559-019-1070-4.

[74]. Wilman, Hamish, Marcelo M. Rivadeneira Jonathan Belmaker, Jennifer Simpson, Carolina de la Rosa and Walter Jetz. 2014. EltonTraits 1.0: species-level foraging attributes of the world's birds and mammals. *Ecology*, 95(7), 2027. https://doi.org/10.1890/13-1917.1

[75]. Myhrvold NP, Baldridge E, Chan B, Sivam D, Freeman DL, Ernest SKM. An amniote life-history database to perform comparative analyses with birds, mammals, and reptiles. *Ecology*. 2015. 96:3109.

[76]. Tacutu, Robi, Daniel Thornton, Emily Johnson, Arie Budovsky, Diogo Barardo, Thomas Craig, Eugene Diana, Gilad Lehmann, Dmitri Toren, Jingwei Wang, Vadim E. Fraifeld and João P. de Magalhães. 2018. Human Ageing Genomic Resources: new and updated databases. *Nucleic Acids Research*, 46 (D1): D1083–D1090. https://doi.org/10.1093/nar/gkx1042

[77]. Billerman, S. M., B. K. Keeney, P. G. Rodewald, and T. S. Schulenberg (Editors) (2022). Birds of the World. Cornell Laboratory of Ornithology, Ithaca, NY, USA. https://birdsoftheworld.org/bow/home

[78]. Fransson, T., Kolehmainen, T., Moss, D. & Robinson, R. 2023. EURING list of longevity records for European birds.

[79]. R: A language and environment for statistical computing. 2023. R Foundation for Statistical Computing, Vienna, Austria, https://www.R-project.org/

[80]. Hadfield, Jarrod D. 2010. MCMC Methods for multi-response generalized linear mixed

models: the MCMCglmm R package. Journal of Statistical Software, 33, 2.

[81]. Revell, L. J. 2024. phytools 2.0: an updated R ecosystem for phylogenetic comparative

methods (and other things). PeerJ, 12:e16505. https://doi.org/10.7717/peerj.16505

[82]. Jetz, W., G. H. Thomas, J. B. Joy, K. Hartmann and A. O. Mooers. 2012. The global dive
rsity of birds in space and time. *Nature*, 491, 444-448.

[83]. Hackett, Shannon J., Rebecca T. Kimball, Sushma Reddy, Rauri C. K. Bowie, Edward L.

Braun, Michael J. Braun, Jena L. Chojnowski, W. Andrew Cox, Kin-lan Han, John

Harshman, Christopher J. Huddleston, Ben D. Marks, Kathleen J. Miglia, William S.

Moore, Frederick H. Sheldon, David W. Steadman, Christopher C. Witt and Tamaki Yuri.

2008. A phylogenomic study of birds reveals their evolutionary history. *Science*, 320,

5884: 1763–68. DOI: 10.1126/science.1157704

[84]. Moreno, J. 1989. Strategies of mass change in breeding birds. *Biological Journal of the*

Linnean Society, 37 (4): 297–310. https://doi.org/10.1111/j.1095-8312.1989.tb01907.x

[85]. Winkler, D. W. 2016. Breeding biology of birds. In: *Handbook of bird biology.* Edited by

Irby J. Lovette, John W. Fitzpatrick. Cornell laboratory of ornithology, 407-450.

[86]. Basile, Anthony J., Kavita C. Singh, Deborah F. Watson and Karen L. Sweazea. 2022.

Effect of macronutrient and micronutrient manipulation on avian blood glucose

concentration: a systematic review. *Comparative Biochemistry and Physiology, Part A*,

272 (July). https://doi.org/10.1016/j.cbpa.2022.111279

[87]. Wada, Yasuaki, Yasuhiro Takeda and Masashi Kuwahata. 2018. Potential role of amino

acid/protein nutrition and exercise in serum albumin redox state. *Nutrients,* 10 (1), 17,

1–11. https://doi.org/10.3390/nu10010017.

[88]. Honma, Ayaka, Chiaki Ogawa, Misaki Sugahara, Shinobu Fujimura and Kazumi Kita.

2017. Influence of varying dietary protein levels on glycation of albumin, tryptophan and

valine in the plasma of chickens. *Journal of Poultry Science*, 54 (3): 242–46.

https://doi.org/10.2141/jpsa.0160146.

[89]. Colombo, Stefanie M., Alexander Wacker, Christopher C. Parrish, J. Martin, Michael T.

Arts, Stefanie M. Colombo, Alexander Wacker, Christopher C. Parrish, Martin J. Kainz

and Michael T. Arts. 2017. A fundamental dichotomy in long-chain polyunsaturated fatty

acid abundance between and within marine and terrestrial ecosystems. *Environmental*

Reviews, 25 (2): 163–74. https://doi.org/10.1139/er-2016-0062

[90]. Charles, Apostolos-Manuel Koussoroplis, Lemarchand Alexandre, Philippe Berny and Gilles Bourdier. 2008. From aquatic to terrestrial food webs: decrease of the docosahexaenoic acid/linoleic acid ratio. *Lipids*, 43:461–66. https://doi.org/10.1007/s11745-008-3166-5.

[91]. Li, Na, Hao Yue, Min Jia, Wei Liu, Bin Qiu, Hanxue Hou, Fenghong Huang and Tongcheng Xu. 2019. Effect of low-ratio n-6/n-3 pufa on blood glucose: a meta-analysis. *Food and Function*, 10 (8): 4557–65. https://doi.org/10.1039/c9fo00323a.

[92]. Vibeke H. Telle-Hansen, Line Gaundal and Mari C.W. Myhrstad. 2019. Polyunsaturated fatty acids and glycemic control in type 2 diabetes. *Nutrients*, 11 (1067): 12–15. doi:10.3390/nu11051067

[93]. Bavkar, Laxman N., Rahul S. Patil, Sheetalnath B. Rooge, Megha L. Nalawade and Akalpita U. Arvindekar. 2019. Acceleration of protein glycation by oxidative stress and comparative role of antioxidant and protein glycation inhibitor. *Molecular and Cellular Biochemistry*. https://doi.org/10.1007/s11010-019-03550-7.

[94]. Klandorf, H., I. L. Probert and M. Iqbal. 1999. In the defence against hyperglycamiea: an avian strategy. *World's Poultry Science Journal*, 55 (3): 251–68. https://doi.org/10.1079/WPS19990019.

[95]. Machín, M., M. F. Simoyi, K. P. Blemings and H. Klandorf. 2004. Increased dietary protein elevates plasma uric acid and is associated with decreased oxidative stress in rapidly-growing broilers. *Comparative Biochemistry and Physiology Part B*, 137: 383–90. https://doi.org/10.1016/j.cbpc.2004.01.002.

[96]. Stinefelt, Beth, Stephen S. Leonard, Kenneth P. Blemings, Xianglin Shi and Hillar Klandorf. 2005. Free radical scavenging, dna protection, and inhibition of lipid peroxidation mediated by uric acid. *Anals of clinical and laboratory science*, 35 (1): 37–45.

[97]. Harr, Kendal E. 2002. Clinical chemistry of companion avian species: a review. *Veterinary Clinical Pathology* 31 (3). DOI:10.18637/jss.v033.i02

[98]. Smith, Susan B., Scott R. Mcwilliams and Christopher G. Guglielmo. 2007. Effect of diet composition on plasma metabolite profiles in a migratory songbird. *The Condor*, 109 (1): 48–58, https://doi.org/10.1093/condor/109.1.48

[99]. Cohen, Alan A., Kevin J. McGraw and W. Douglas Robinson. 2009. Serum antioxidant levels in wild birds vary in relation to diet, season, life history strategy, and species. *Oecologia*, 161: 673–83. https://doi.org/10.1007/s00442-009-1423-9.

[100]. Alan, Rebecca R. and Scott R. Mcwilliams. 2013. Oxidative stress, circulating
antioxidants, and dietary preferences in songbirds. *Comparative Biochemistry and*
Physiology, Part B, 164 (3): 185–93. https://doi.org/10.1016/j.cbpb.2012.12.005.

[101]. Schneeberger, Karin, Gábor Á. Czirják and Christian C. Voigt. 2014. Frugivory is
associated with low measures of plasma oxidative stress and high antioxidant
concentration in free-ranging bats. *Naturwissenschaften*, 101: 285–90.
https://doi.org/10.1007/s00114-014-1155-5.

[102]. Goff, H. Douglas, Nikolay Repin, Hrvoje Fabek, Dalia El Khoury and Michael J. Gidley.
2018. Dietary fibre for glycaemia control: towards a mechanistic understanding.
Bioactive Carbohydrates and Dietary Fibre, 14, 39-53.
http://dx.doi.org/10.1016/j.bcdf.2017.07.005