A robust relationship between robustness and evolvability

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Data accessibility

Supplementary Information files are available via a GitHub repository (https://github.com/n8-rd/qkModels.git) and will also be deposited in Zenodo.

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Abstract

Here, I consolidate and extend simple models of how genetic robustness affects the evolvability of phenotypes with discrete states. I share three main insights. (1) What we already know about the effects of mutational robustness on evolvability can be readily transferred to environmental robustness. (2) Even without a hard genotype-level trade-off between robustness and evolvability, the optimal level of phenotypic robustness depends on the rate of environmental change. But counter-intuitively, when adaptive landscapes are complex, an increase in environmental stability can increase the frequency of environmentally-robust but mutationally-sensitive genotypes. (3) Even with a fixed probability that a mutation is neutral, populations can evolve along the spectrum of robustness and evolvability by evolving the genotype-determined neighborhood of mutationally-accessible phenotypes. Indeed, because it allows for the evolution of increased evolvability without a concomitant increase in genetic load, selection should favor changes in the phenotypic neighborhood over changes in mutational sensitivity.

Teaser Text

Models of the evolution of discrete phenotypes show that their evolvability is highest when environments fluctuate at intermediate rates, and with intermediate levels of mutational robustness. By merging previous models and relaxing previous assumptions, I show that previous insights are robust. I also show that the same models tell us very different things about the evolvability of discrete traits, namely, that what applies to robustness against mutation is readily transferable to robustness against environmental perturbation, and that selection for evolvability will preferentially work on the neighborhood of mutationally-accessible phenotypes rather than the probability of non-neutral mutation.

39 **1. Introduction**

Phenotypes tend to be robust against much genetic and environmental variation (1,2). If the

targets of selection are stable, such robustness can be adaptive, as it promotes homeostasis and

decreases genetic load (3–6). On the other hand, robustness reduces phenotypic variation (7) so,

if the adaptive landscape varies, selection may promote genotypes that are more sensitive to

mutational or environmental variation, and are thus more evolvable (8–10).

Genetic and environment variation map to phenotypic variation via developmental systems that

can be complex, and fittingly, many studies of the relationship between robustness and

evolvability have used complex models of such mappings (11,12). Examples include

empirically-based models of RNA or protein folding (7,10,13), or more theoretical models of

self-assembling systems (14,15) or gene regulatory interactions (16–19). Although apropos, the

interpretation of such models is fraught due to the fact that robustness is not the only thing

affected by structural variation in a genotype-phenotype map. In particular, such variation can

also affect how a developmental system (1) accumulates cryptic genetic diversity (20), (2)

phenotypically responds to non-neutral mutation (21,22), (3) transitions between ordered and

chaotic dynamics (19), and (4) "learns" from past selective environments (22–24). So, with

complex genotype-phenotype models, when a change in evolvability is observed, it can be

difficult to say that a change in robustness *per se* is the cause.

An alternative approach is to abstract away the details of a developmental system, and model

robustness simply as the probability that a change in the genetic or external environment will

affect a change in phenotype (25,26). Such models have shown, somewhat counter-intuitively,

that evolvability can be facilitated by intermediate levels of robustness (25), and promoted by

intermediate rates of environmental change (26). But the extent to which these insights depend

on specific model assumptions is unclear and therefore so are the boundaries of the theory's

domain of applicability. Our goal here is to relax those assumptions, expose heretofore

unrecognized connections between models, and get a better sense for when phenotypic

robustness is good for the evolvability of discrete traits. Let us start with a quick look at the work

that provides the foundation for what follows.

2. Simple models of the robustness of discrete phenotypes

2.a. Genotype robustness

At the genotype level, robustness and evolvability trade off; robustness decreases phenotypic

variance (10). Meyers et al. (26) elegantly demonstrate that given genotype-level trade-offs

between mutational robustness, environmental robustness, and evolvability (10), the optimal

level of genetic robustness is a function of the rate of environmental change. They ask us to

imagine a pentagonal, bidirectional, mutational network, where each vertex is a genotype, and

each edge is a mutation (Fig. 1a.). A population of genotypes that can mutate along the edges of

this network is exposed to fluctuating selection. Each genotype maps to one of three possible

phenotypes: (1) phenotype A is optimal when the environment is in state a, (2) phenotype B is

optimal when the environment is in state b, and (3) phenotype V has intermediate fitness in both

environmental states (Fig. 1a). To clarify how this model relates to the next we will consider, let

us define a couple new terms. First, let q_i denote the probability that a mutation of genoptype g_i

has no effect on the phenotype. In Fig. 1a, q_i varies from one for genotype g_1, to zero for g_3 and

g_4. Second, let r_i stand for the environmental robustness of genotype g_i, that is, the probability

that a change in the environment has no effect on fitness. In Fig. 1a, it is one for g_3 and less than

one for the rest. Mutational and environmental robustness are both genotype-specific.

Meyers et al. (26) show that given this mutational network, the expected frequency of each

genotype depends on a parameter λ, the number of generations between a change in the selective

environment (Fig. S1): (i) When the environment changes infrequently ($\lambda > 1000$) the most

mutationally robust genotype, g_1, occurs at the highest frequency. (ii) When the environment

changes rapidly ($\lambda < 10$), the most important thing is the environmental robustness of a

genotype, r_i; the generalist genotype g_3 occurs at the highest frequency (8). (iii) When the

environment changes neither too fast nor too slow ($10 < \lambda < 1000$), the population evolves to

maximize evolvability; the highest frequency genotype is g_0, which is the genotype that maps to

phenotype *A* that has the lowest combination of mutational and environmental robustness. In

sum, selection works not just on the observable phenotype {*A, B, V*} but also the robustness and

evolvability of those phenotypes.

Figure 1. *Mutational networks with redundant mappings of genotypes to phenotypes. (**a**) The*

network of Meyers et al. (26). Five genotypes are possible, each of which maps to one of three

phenotypes: A (light blue) is optimal in state a environments; B (gold) is optimal in state b

environments; V (gray) has intermediate fitness across environmental states a and b. Back-

mutation is possible and the graph forms a cycle. Robustness varies across genotypes, but the

*robustness and phenotypic neighborhood of each genotype does not change over time. (**b, c**)*

Example of the mutational network of Draghi et al. (25) at two time points. Different vertex

colors denote different phenotypes, with the phenotype p_0 (light blue) being currently optimal.

Potential phenotype-changing mutations are shows with dashed gray edges; the set of vertices

connected by such edges to a genotype constitute that genotype's phenotypic neighborhood, k_i.

Potential neutral mutations are shown as dotted black edges, and realized neutral mutations are

shows as solid black edges. Back-mutation is forbidden, thus, the graph is divergent.

2.b. Phenotype robustness

At the phenotype level, robustness and evolvability need not trade off, since robustness can

increase cryptic genetic diversity (29,30,31,32), an elegant demonstration of which is given by

Draghi et al. (25). Like Meyers et al. (26), they consider an evolving population of genotypes the

relations of which can be visualized as a mutational network (Fig 1b,c). But in this case, instead

of specifying a fixed, cyclical network, they specify a mutational process, and allow a divergent

network of mutations to emerge organically. They let each genotype g_i map to a phenotype p_i,

and have a K-dimension set $\mathbf{k_i}$ of alternative phenotypes that can be reached by a single mutation.

Thus, $\mathbf{k_i}$ is what has been referred to as the phenotypic 1-neighborhood of a genotype (10). (Note

that in the Meyers et al. (26) model, each genotype also has a phenotypic neighborhood, but

there is no requirement that adjacent genotypes have alternative phenotypes, and thus K varies

across genotypes.) Phenotypes for p_i and $\mathbf{k_i}$ are chosen randomly from a set of possible

phenotypes $\mathbf{P}$ of size P. Before time t_x, a population of genotypes is exposed to strong stabilizing

selection about an initial phenotype p_0; all alternative phenotypes confer a fitness of zero.

Mutations at rate μ are neutral with probability q; thus, as in the model of Meyers et al. (26), q

affects mutational robustness, but in this case q does not vary across genotypes. At rate $\mu(1-q)$,

mutation causes a change in the phenotype, with the new phenotype, $p_i{'}$, selected from the

phenotypic neighborhood $\mathbf{k_i}$. Whenever a mutation occurs, regardless of whether or not it is

neutral, a new phenotypic neighborhood, $\mathbf{k_i}$, is randomly sampled from the phenotypic space $\mathbf{P}$.

So, during this initial phase of evolution, while strong stabilizing selection prevents a population

from diversifying phenotypically, the population can grow a network of phenotypically

redundant genotypes, each of which has its own phenotypic neighborhood (Fig. 1b,c).

Figure 2. *The effect of*

mutational robustness, q, on

evolvability, t_a^{-1}, depends on

the difference between the

number of possible

phenotypes, K, and the size

of the phenotypic

neighborhood P. This

reproduces the results of

Draghi et al. (25) but with an individual-based simulation model (Supplementary File S2), with

50 replications for each combination of parameter values, P = 100, K = {5,30,100}, and q =

(0.1, 0.2, …, 1.0). The y-axis is on a log scale. Curves are loess regressions, with a span of 1.0;

shaded areas give 95% confidence intervals. Interpretation*: Unless all phenotypes are*

accessible by one mutation, evolvability is maximized at intermediate levels of mutational

robustness.

At time t_x, a new optimal phenotype, p_x, is chosen from **P**. Draghi et al. (25) examine how, t_a, the

time it takes for the population to evolve p_x depends on q, K, and P. They show that if $K = P$ –

that is, every genotype can with one mutation reach every possible phenotype – increasing

mutational robustness q monotonically increases the expected time for the population to evolve

p_x (Fig. 2). As in Meyers et al. (26), robustness and evolvability are antithetical. But if $K < P$,

mutational robustness can increase the diversity of phenotypic neighborhoods, and the

relationship between q and the expected time for the evolution of p_x is non-monotonic;

evolvability is highest at intermediate values for q (Fig. 2).

3. The evolution of environmental robustness

Although Draghi et al. (25) and Meyers et al. (26) focus on mutational robustness, their models

are readily transferrable to questions about environmental robustness.

Imagine that, as in the Draghi et al. (25) model, there is a world with P possible phenotypes. But

instead of a population of genotypes that explores this world via mutation, suppose there is a

meta-population of genetically-invariant demes that explores it via migration. Phenotypes

emerge from genotype-by-environment interactions, and environments vary such that each deme

has a K-dimensional phenotypic neighborhood, **k_i**, accessible via migration. For a depiction of

this model, Figure 1.b. should suffice; one need only reinterpret the nodes as demes, and the

edges as migration events. Let q_m represent the probability that the environment of a new deme

induces no change of phenotype; in other words, q_m determines the migrational robustness of the

genotype to environmental variation. Since we assume the population is genetically

monomorphic and have replaced mutation with migration, we can not assess the relationship

between robustness and evolvability *per se,* at least not if we follow convention and think about

evolution only in terms of changes in genotype frequencies. But we can assess the relationship

between robustness and plasticity, which is analogous. In the beginning, only phenotype p_0 is

adaptive and only demes that express p_0 survive selection. But migration forms a neutral network

of demes, and the diversity of migrationally-accessible phenotypic neighborhoods grows. If at

some point a new phenotype, p_x, becomes optimal, as long as $K < P$, we know that the

relationship between q_m and rate at which the population evolves via migration to express p_x will

be non-monotonic, with the fastest rates occurring at intermediate values of q_m. This is because

the logic of Draghi et al. (25) model of mutational robustness has not changed, only our

interpretation of some of its variables and parameters. Neutral mutation can boost evolvability by

changing the structure of a mutation network. Neutral migration can boost plasticity by changing

the structure of a population in a heterogeneous environment.

To go beyond plasticity, and connect environmental robustness to evolvability, we need to allow

for mutation as well as migration (Fig. S3). Above, we let phenotypes vary depending on the

interaction between one genotype and many local environments. Suppose instead that each

individual can have one of two genotypes, either g_0 or g_1. Initially, the population is

monomorphic for g_0 but mutation between genotypes occurs at rate μ per individual per

generation. In e_0 environments, genetic variation is cryptic; both genotypes express the optimal

phenotype p_0. Initially, migration to any non-e_0 environment induces strongly deleterious changes

in phenotype. Then at time t_x, a new phenotype p_x – which is expressed only by g_1 genotypes in e_x

environments – becomes adaptive in e_x environments. Now we can ask the question that ties

environmental robustness to evolvability: How does the time it takes a population to evolve

phenotype p_x depend on environmental robustness, q_m? Analysis of this model (with codes given

in Supplementary Document S5) shows that evolvability is highest at intermediate values for q_m

(Figure S4). Robustness is robustness; mutational and migrational robustness have similar effects

on evolvability.

In a similar fashion, the model of Meyers et al. (26) can be transferred to the evolution of

environmental robustness and evolvability; we need only change our interpretation of the edges

and vertices in Fig. 1a. Instead of five genotypes, we have one phenotypically-plastic genotype,

five spatial locations, and three different environment types, such that the genotype can expresses

three different phenotypes depending on its environment. Instead of genotypes connected by

mutation, we have demes connected by migration. If we apply the same kind of fluctuating

selection – with the phenotype expressed in demes g_0, g_1, and g_3 being optimal in temporal state

a, that expressed by deme g_4 being optimal in state b, and that in g_3 with intermediate fitness in

both states – then the proportion of a meta-population in each deme will be a function of the rate

of environmental change. When environmental change is slow, individuals will occur at the

highest frequency in deme g_1, since that minimizes migration load (29). When the world turns

over rapidly, deme g_3 offers a safe harbor of stability. And at intermediate rates of change, the

highest fitness can be realized by hopping back and forth between demes g_0 and g_4.

204 4. Consolidating models

205 In Sections 2 and 3, I pointed out fundamental similarities between the models of Meyers et al.

206 (2005) and Draghi et al. (2010). In this section, I merge those models and relax some their main

207 assumptions. Doing so shows that their main inferences are robust. It also reveals some more

208 nuanced dynamics. I take an individual-based approach, with models developed using the SLiM

209 framework (30)

210 4.a Divergent mutational networks of genotypes varying in mutational
211 and environmental robustness

212 4.a.i. Model description

Imagine a population of haploid individuals in an environment with a carrying capacity of

$N=500$, and for which there is a P-dimensional set **P** of possible phenotypes. Each individual has

three genetically-determined traits: (1) a phenotype p_i, (2) a phenotypic neighborhood $\mathbf{k_i}$ of size

K, and (3) a mutational robustness q_i. The life cycle follows the Wright-Fisher model, with

selection on reproductive success, clonal reproduction with mutation, and non-overlapping

generations. At the start of each simulation, the population is monomorphic, with q_i = 0.1, and an

initial phenotype p_0 and phenotypic neighborhood $\mathbf{k_0}$ selected randomly from $\mathbf{P}$. Mutation

happens at rate μ=2e-3 per individual per generation. Each mutation is neutral with probability q_i.

Whenever a neutral mutation occurs, a new phenotypic neighborhood $\mathbf{k_i}$ is chosen from $\mathbf{P}$. When

a non-neutral mutation occurs, there is an equal chance that it affects either mutational robustness

q_i, or the phenotype p_i. If the former, a new value for q_i is chosen from a random uniform

distribution. If the latter, a new phenotype p_i and phenotypic neighborhood $\mathbf{k_i}$ are randomly

sampled from $\mathbf{P}$.

This population is subjected to fluctuating selection at rate λ^{-1}. When the environment is in state

a, phenotypes in L-dimensional set $\mathbf{O_a}$ are optimal, and when the environment is in state b, a

non-overlapping L-dimensional set $\mathbf{O_b}$ of phenotypes are optimal. Phenotypes in L-dimensional

set $\mathbf{V}$ have intermediate fitness in both environmental states. So,

$w_a(p_i) = \beta_0+s$ for $p_i \in \mathbf{O_a}$; β_0 for $p_i \in \mathbf{O_b}$; β_0+sh for $p_i \in \mathbf{V}$, and

$w_b(p_i) = \beta_0$ for $p_i \in \mathbf{O_a}$; β_0+s for $p_i \in \mathbf{O_b}$; β_0+sh for $p_i \in \mathbf{V}$

where, similar to Meyers et al. (26), w_j gives the fitness of phenotype p_i in environmental state j,

β_0 = 0.6 is the base reproductive fitness, s = 0.4 is the fitness advantage of expressing an

appropriate specialist phenotype, and h = 0.5 determines the intermediacy of the generalist

phenotype.

We simulate evolution by repeating this life cycle for 20K generations, keeping track of the

following model variables: (1) w, the mean population fitness, (2) q_i, the mean population

mutational robustness, (3) D, the mean Simpson's Diversity of the population's phenotypic

neighborhoods, and (4) f_V, the mean frequency of individuals expressing a generalist phenotype,

that is, $p_i \in \mathbf{V}$. Fifty replicated model simulations were run for every combination of model

parameters (Table 1). Model codes are provided as Supplementary Document S3.

Table 1. Divergent network model parameters and variables.

Parameter	Description	Values
μ	Mutation rate	2e-3
N	Population size	500
P	Number of possible phenotypes, **P**	{6, 100}
K	Size of phenotypic neighborhood, $\mathbf{k_i}$	{1, 3, 6} | P=6
		{6, 30, 100} | P=100
L	Number of phenotypes in each set $\mathbf{O_a}$, $\mathbf{O_b}$, and $\mathbf{V}$	1 | P=6
		$K/3$ | P=100
λ	Number of generations between change in environment	{1e+1… 1e+4}
β_0	Viability of specialist genotype in wrong environment	0.6
s	Viability advantage of specialist in right environment	0.4
h	Viability intermediacy of generalist phenotype	0.5
Variable	**Description**	**Range**
w	Population mean fitness	$0 < w < 1$
q_i	Population mean mutational robustness	$0 < q_i < 1$
f_V	Frequency of generalist phenotypes ($p_i \in \mathbf{V}$)	$0 < f_V < 1$
D	Simpson's Diversity of phenotypic neighborhoods, $\mathbf{k_i}$	$0 < D < 1$

243 4.a.ii Model Dynamics

First consider what happens when the set of possible phenotypes is small (P=6) and only one

phenotype is optimal in each environmental state (L=1) (Fig. 3). Regardless of phenotypic

neighborhood size, K, when selection fluctuates rapidly (λ = 10) mean population fitness is ~ 0.8

(Fig. 3a), which is the viability of the generalist phenotype, and indeed, the generalist phenotype

tends to occur at frequencies near one (Fig. 3c). As the rate of fluctuating selection decreases,

mean population fitness rises, and the frequency of the generalist phenotype falls, so that when λ

=10,000, w is near one, and f_V is near zero. (But note that there is a kink in the mapping of λ to f_V,

at ~ λ=100; we will return to this below.) In contrast, the relationships between λ and the other

model statistics, q_i and D, are non-monotonic. Mutational robustness, q_i, is lowest at intermediate

values for λ, with the bend of the curve being more pronounced with larger phenotypic

neighborhoods (Fig. 3b). We see a similar U-shaped mapping of λ to the mean diversity of

phenotypic neighborhoods, D, although when $K=P$, D equals one by definition, and the extent to

which D drops at intermediate values of λ increases with decreasing K. If populations are more

mutationally sensitive and hence more evolvable, repeated bouts of positive selection will tend to

sweep away much of the standing diversity in phenotypic neighborhoods (31). This effect is

especially strong when $K=1$, in which case it takes longer for D to rebound from a sweep. So,

when $K \in \{3, 6\}$ and $P=6$, increased mutational sensitivity $(1-q_i)$ clearly brings about increased

evolvability. On the other hand, when $K=1$, this relationship weakens, as a relatively small dip in

mutational robustness, q_i, comes with a relatively large dip in the diversity of phenotypic

neighborhoods, D; though more mutations are non-neutral, they access a smaller part of the

phenotypic space, P.

Figure 3. *Summary of models where K is fixed and q_i is freely evolvable. Panels show how the*

*number of generations between environmental fluctuations, λ, affects (**a**) population mean*

*fitness, w-bar, (**b**) mean mutational robustness, q_i-bar; (**c**) mean frequency of generalist*

*genotypes f_V-bar, (**d**) mean population diversity of phenotypic neighborhoods, D-bar. Each point*

shows the result for one simulation. Lines show loess regressions. Point and line colors denote

different values for phenotypic neighborhood size, K. Interpretation*: As environments become*

increasingly stable, there is a succession of mutationally robust generalists, evolvable specialists,

and mutationally robust specialists.

When the set of possible phenotypes is comparatively large (*P*=100) and more than one

phenotype is optimal in each environmental state (*L*=*K*/3), what we find is similar to when *P*=6

and *L*=1 (Fig. 4), but when *K*=6, the predominance of specialist genotypes in more stable

environments is much less pronounced, and the kink in the mapping of λ to f_V is yet more

striking. (Fig. 4a). In fact, polynomial regression analysis recovers support for a cubic functional

relationship between f_V and λ, and there is a region in which increasing environmental stability

(λ) increases the frequency of generalist genotypes, f_V. How do we explain this?

Figure 4. *How, when the phenotype space is large, P=100, the number of generations between*

*environmental fluctuations, λ, affects (**a**) the mean frequency of generalist genotypes f_V, (**b**) the*

*mean mutational robustness of genotypes expressing the generalist phenotype V_q and (**c**) the*

mean mutational robustness of specialists, AB_q. Each point shows the result for one simulation.

Lines show loess regressions. Point and line colors denote different values for phenotypic

neighborhood size, K. Interpretation: *When P is large and K is small, it is unlikely that*

populations will evolve well-adapted specialists unless the environment is relatively stable.

Counterintuitively, increasing environment stability can increase the frequency of mutationally

sensitive generalists.

It seems there is an interaction between environmental and mutational robustness. Mean

population mutational robustness, q_i, changes little with λ. But this masks an underlying

divergence in q_i between specialist and generalist genotypes (Fig 4b,c). Specifically, as λ

increases from 10 to 10,000, in generalists q_i drops from 0.45 to 0.13, while in specialists it

increases from 0.34 to 0.60. So, when populations with small phenotypic neighborhoods are

challenged to adapt to a world in which there are many phenotypic possibilities, environmental

stability selects for evolvability in generalists and mutational robustness in specialists. At rates of

environmental change that most favor evolvability ($100 < \lambda < 1000$), generalists tends to be more

evolvable. And so inside this range of values for λ, we see an increase in the mean frequency of

generalist phenotypes.

Why are generalists more evolvable in such environments? One hypothesis would be that

generalists do not persist long enough for selection for mutational robustness to be effective. But

closer analysis reveals evidence to the contrary. In fact, over the λ values at which f_V increases,

generalists tend to persist for several thousand generations (Fig. S2.c). Moreover, they tend to

evolve lower mutational robustness than their specialist ancestors, and this dynamic becomes

more pronounced as λ increases. If we divide the evolutionary history of a population into phases

of generalism ($f_V > 0.9$) and specialism ($f_V <= 0.9$), and then calculate the mean change in q_i over

the course of a generalist phase, Δ_{qV}, the mean of this quantity decreases from -0.04 when $\lambda = 10$,

to -0.18 when $\lambda = 10,000$ (Fig. S2.a). So, the high evolvability of generalists is not because

generalists are ephemeral.

A alternative hypothesis is that selection against mutational sensitivity is less efficient in

generalists, since the expected decrease in fitness from non-neutral mutation is less severe.

Generalists tend to evolve evolvability via genetic drift. Sure enough, whereas increasing

environmental stability, λ, causes the mean of Δ_{qV} to become more negative, the standard

deviation of Δ_{qV} remains consistently positive, regardless of environmental stability (Fig. S2.b).

And this helps us better understand the tendency for Δ_{qV} to be negative: Being a generalist does

not increase evolvability. The evolution of increased evolvability presages the end of generalism.

So in broad strokes, the inferences of Meyers et al. (26) are robust to relaxations of the their

assumptions about genetic architecture. Even if we let mutational sensitivity and phenotypic

neighborhood be freely-evolving genotype properties, over much of the explored parameter

space, as environments become increasingly stable, we find the same succession of mutationally

robust generalists, evolvable specialists, and mutationally robust specialists. On the other hand,

we find some new patterns when phenotypic neighborhood sizes are a small fraction of the total

number of possible phenotypes. In particular, there can be a negative feedback between

mutational sensitivity ($1-q_i$) and phenotypic neighborhood diversity (D) such that more

mutationally sensitive genotypes might not be more evolvable. And counter-intuitively, certain

increases in environmental stability can cause an increase in the frequency of generalist

phenotypes, provided that they are more mutationally sensitive than specialists.

327 4.b. Evolvable cyclical networks of genotypes varying in robustness

328 4.b.i. Model description

Next, let us extend the Meyers et al. (26) model so that the mutational network is a freely

evolvable individual-level property. We mostly follow the original and use the same set of

possible phenotypes, $\mathbf{P} = \{A, B, V\}$, the same fitness functions, and the same regimes of

fluctuating selection ($1 < \lambda < 10{,}000$). But we change the mutational process to allow for two

kinds of effects. With probability $\rho=0.5$, a mutation transforms the genotype of an individual to

one of the adjacent genotypes in their mutational network. With probability $1- \rho$, a mutation

randomly re-assigns the phenotypic mapping of a randomly selected vertex in the individual's

mutational network. To clarify, in the divergent mutational network model of Draghi et al. (25),

the network is a population-level property; an individual's position in that network is determined

by their ancestry and the elements of their phenotypic 1-neighborhood. By contrast, with this

extension of the Meyers et al. (26) cyclical network model, the network is essentially a higher-

order, phenotypic 4-neighborhood, governing what phenotypes are accessible to a genotype via

any number of mutations.

We analyze the dynamics of this model with individual-based simulations, with 20K generations

for each replicate, and 50 replicates per value of λ. Model codes are provided in Supplementary

Document S4. In each generation, we record the following model statistics: (1) w, the mean

population fitness, (2) GP_{max}, the most common genotype-phenotype map, (3) S^{-1}, the population

mean of the inverse richness of mutationally-adjacent phenotypes, which is a measure of

mutational robustness, (4) f_V, the mean frequency of individuals expressing a generalist

phenotype, and (5) f_{Vk}, the mean frequency of phenotypic 1-neighborhoods containing the

generalist phenotype p_V.

4.b.ii. Model dynamics

In broad strokes, with this model, we find the same λ-dependent succession of environmentally-

robust generalists, evolvable specialists, and mutationally robust specialists (Fig. 5). But we also

get a couple of new insights, and a surprise. The first new insight is that when environments turn

over rapidly (λ=10), not only does the generalist phenotype p_V prevail, but mutational networks

are enriched for p_V (Fig6a.). In fact, the most common network, occurring at an average

frequency of ~0.43, is composed exclusively of p_V (Fig 6b.). Populations evolve to be both

environmentally and mutationally robust. And the mutational robustness observed in this case is

more pronounced than anything we see in mutationally-robust specialists in stable environments.

The second new insight is that in stable environments (λ=10,000), mutational robustness is

asymmetrical across specialist phenotypes, with higher average robustness for the phenotype that

was optimal in the first environmental epoch, p_A. Historical contingency can have long-lasting

effects on the evolution of the genotype-phenotype map.

The surprise is that – similar to what we saw for f_V with our extensions of Draghi et al. (25)

model – there is a kink in the functional mapping of variation in λ to f_{Vk}', that is, the mean

population frequency of genotypes containing the generalist phenotype p_V in their phenotypic 1-

neighborhood. A dip in f_{Vk}' happens at $\lambda \sim 100$, where the balance between selection for

evolvability and robustness tips most in favor on evolvability; at $\lambda = 100$, in evolved mutational

networks, a mapping to p_V is rare, and there is relatively even mix of mappings to p_A and p_B (Fig

7a,c.). Beyond $\lambda = 100$, selection against p_V mappings weakens, as mutational robustness again

becomes more valuable. It seems that selection for evolvable specialists more efficiently reduces

the likelihood of generalism-causing mutations than does selection for robust specialists.

Figure 5. *Dynamics of evolving cyclical mutation networks. How the number of generations*

*between environmental fluctuations, λ, affects (**a**) the population mean fitness, w-bar, (**b**) the*

*mean frequency of a generalist genotypes f_V-bar, (**c**) the mean frequency of mutational networks*

*containing the generalist phenotype f_{Vk}-bar, and (**d**) the mean phenotypic robustness against*

mutation, S^{-1}. Each point shows the result for one simulation. Lines show loess regressions. The

mutation rate μ = 1e-2, and there is an equal probability that a mutation causes an individual to

change to an adjacent genotype, or causes a change in the phenotype-assignment of one of the

genotypes in the network.

Fig 6. *Evolved mutational networks. (a) How the number of generations between environmental fluctuations, λ, affects the average frequency of each phenotype in the population of mutational networks, with the frequency of p_A shown in purple, p_B in green, and p_V in blue. Below, (b-e), the most common mutational network is shown for a subset of λ values. Here genotypes mapping to phenotype p_A are in light blue, p_B in gold, and p_V in gray.* Interpretation: *When we let cyclical mutational networks evolve in volatile environments, populations evolve to be both mutationally and environmentally robust. Increases in evolvability are realized by evolving networks that are an even mix of specialist phenotypes. And in stable environments, historical contingency can cause long-lasting biases in phenotypic robustness.*

5. Robustness via phenotypic neighborhood evolution

In the previous sections, we saw that the evolution of mutational robustness, q_i, and how it affects evolvability depends on $\mathbf{k_i}$, the phenotypic neighborhood. In fact, selection on evolvability could affect q_i or $\mathbf{k_i}$. We show this with another model variant (Supplementary

Document S6). The model set-up is as described in Section 4.a, for $P=6$, except for three

changes: (1) We fix the value of $q \in \{0.0, 0.1, 0.5\}$. (2) We let K evolve freely; hence, each

genotype determines a phenotypic neighborhood size K_i. (3) We change the way that $\mathbf{k_i}$ evolves.

Specifically, at the start of each simulation, the population is monomorphic for a three-

dimensional $\mathbf{k_i}$, with elements randomly sampled from $\mathbf{P}$. Then, as the life cycle turns over,

mutation can with equal probability add or subtract one element from $\mathbf{k_i}$, with the restriction that

$0 < K_i \leq P$. We do not prevent redundancy of the elements of $\mathbf{k_i}$. This is to maximize flexibility

for the design by selection of an optimal phenotypic neighborhood. To further resolve the model

dynamics we also keep track of two additional model statistics, to wit, f_B is the mean frequency

of phenotypic neighborhoods that contain the optimal phenotype for the out-of-phase

environmental state, and f_{Vk} is the mean frequency of phenotypic 1-neighborhoods with the

generalist phenotype.

With simulation analysis, we find that just as when we fix K and let q_i vary across genotypes,

populations appear to evolve maximum evolvability at intermediate values for λ (Fig 7). At

intermediate values for λ, not only is the mean of K_i highest, but so is f_B, the probability that $\mathbf{k_i}$

contains the optimal phenotype of the out-of-phase environmental state, and in fact, the latter

effect is more pronounced. Averaging across values of q, as λ increases from 10 to 100, there is a

13% increase in K_i, and a concomitant 35% increase in f_B; evolvable populations stack their

phenotypic neighborhoods with good options. With increasing λ we also see a decline in the

frequency of the generalist genotype along with f_{Vk}, that is, the frequency of genotypes that have

the generalist phenotype in their neighborhood. These relations are robust to variation in

mutational robustness q, although when q is closer to zero, we see more pronounced reductions

of K_i and f_B at extreme values of λ.

Figure 7. *Summary of models where q is fixed and K_i is freely evolvable. Panels show how changing the number of generations between environmental fluctuations, λ, affects (**a**) mean phenotypic neighborhood size, K_i-bar; (**b**) mean frequency of phenotypic neighborhoods containing the optimal phenotype for the out-of-phase environmental state, f_B-bar; and (**c**) mean frequency of phenotypic neighborhoods containing the generalist phenotype, f_{Vk}-bar. Each point shows the result for one simulation. Lines show loess regressions. Point and line colors denote different values for q.* Interpretation: *At intermediate rates of environmental change, populations evolve to maximize evolvability by enriching their phenotypic neighborhoods with phenotypes that are optimal in the out-of-phase environmental state.*

So, there is more than one way to evolve robustness and evolvability. Selection can shape the probability that a mutation will be neutral. Or it can shape the probabilities of the specific phenotypic effects of non-neutral mutation, as has been demonstrated previously with analyses of more complex genotype-phenotype maps (21–23). In fact, the latter would appear to be favored by selection, since tailoring the phenotypic neighborhood can increase the odds of beneficial mutations without also increasing the odds of deleterious ones. Sure enough, if we simulate the evolution of population in which mutational robustness q_i, phenotypic neighborhood size K_i, and phenotypic neighborhood components $\mathbf{k_i}$ are all free to vary, then environmental stability, λ, has little effect on the means of q_i (which tend to be ~ 0.4) and large effects on means of K_i and f_B , that is, the frequency of phenotypic neighborhood containing the phenotype that is optimal in the out-of-phase environment (Fig. S5). Something to stress here is that for selection on phenotypic neighborhood components to be effective, a genotype-phenotype map needs to have a memory of past selective environments. We made that possible, in an elementary fashion, by implementing a

438 piece-wise mutational routine for adding and removing components of the phenotypic

439 neighborhood, $\mathbf{k_i}$. But more complex genotype-phenotype maps can have better memories

440 (22,24).

6. Discussion

Here we addressed this question: How does the robustness of a discrete-state phenotype relate to

evolvability when it can vary across genotypes and phenotypes, against variation in both the

genetic and external environment? We improved our intuition by considering some simple

models in which robustness is expressed as q, the probability that some perturbation does not

affect the phenotype, and if a perturbation does affect the phenotype, such effects are mediated

by $\mathbf{k}$, the neighborhood of accessible phenotypes. For ease of discussion, let us call these qk

(pronounced "cuke") models. By merging and extending previously-developed qk models

(25,26), we saw that their inferences are robust and their scope of applicability is broader than

has been appreciated; both tell us as much about the evolution of environmental robustness as

mutational robustness. We also gained two additional key insights. First, counter-intuitively, in

certain genetic and environmental contexts, reducing the rate of environmental change can lead

to selection for more environmentally robust genotypes, that is, generalists. This is because

generalists tend to evolve lower levels of mutational robustness. Second, although the analysis of

qk models has focused on q, the probability of non-neutral mutation, selection for increased

evolvability works preferentially on k, the neighborhood of mutationally-accessible phenotypes.

This is because evolability via phenotypic neighborhood evolution comes without an increase in

mutational load. That selection can affect the distribution of mutational effects has been shown

before with analysis of more complex genotype-phenotype models (19,24,33,34). But to my

knowledge, this is the clearest demonstration that increasing evolvability by evolving the

phenotypic neighborhood evolution is a better option than evolving increased mutational

sensitivity.

Another new insight is that the interaction between different kinds of robustness can cause

counter-intuitive evolutionary dynamics. Specifically, in complex environments, increasing

environmental stability can cause an increase in the frequency of environmentally-robust

genotypes, that is, generalists. We saw that this corresponds to a divergence in the mutational
sensitivities of generalists and specialists, and we traced this divergence to a neutral evolutionary
process; the evolution of evolvability in generalists is accidental, but predictably so. This is yet
another example of how robustness can increase evolvability by facilitating the accumulation of
cryptic genetic diversity (32,35).

There are many things that qk models of the evolution of robustness and evolvability can not do.
In particular, robustness is just one of several properties of a genotype-phenotype map that can
affect evolvability, and even the effects of robustness can depend on other facets of genetic
architecture. Therefore, a richer understanding of the evolution of evolvability requires more
complex models genotype-phenotype maps. Other germane properties include a developmental
system's criticality, that is, the robustness of its robustness (19,36), as well as its capacity for
plasticity and cryptically-neutral genetic diversity (18,20,22), and its capacity to adaptively
remember and generalize from past selective environments (23,24,37). More complex genotype-
phenotype map models can also yield insights into the specific design features that determine
map properties such as robustness, evolvability and memory. A burgeoning body of research has
already revealed several such network features (38), such as modularity (33,39), excitation (17),
and scale-free degree distribution (40,41). Researchers have also begun to detect important
network motifs affecting robustness and evolvability, such as freed-back and feed-forward loops
(42). So, the analysis of more complex genotype-phenotype map models promises to yield a
more multi-faceted and mechanistic understanding of the evolution of evolvability. Nevertheless,
by ignoring the genetic architectural details that might make one genotype-phenotype map more
robust than another, qk-models throw into sharp relief the effects of robustness *per se* on
evolvability.

References

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1. de Visser JAGM, Hermisson J, Wagner GP, Meyers LA, Bagheri-Chaichian H, Blanchard JL, et al. Perspective: Evolution and Detection of Genetic Robustness. Evolution. 2003;57(9):1959–72.

2. Waddington CH. Genetic Assimilation of an Acquired Character. Evolution. 1953;7(2):118–26.

3. Siegal ML, Bergman A. Waddington's canalization revisited: Developmental stability and evolution. Proceedings of the National Academy of Sciences. 2002 Aug 6;99(16):10528–32.

4. Wagner GP, Booth G, Bagheri-Chaichian H. A population genetic theory or canalization. Evolution. 1997 Apr 1;51(2):329–47.

5. Wilke CO, Wang JL, Ofria C, Lenski RE, Adami C. Evolution of digital organisms at high mutation rates leads to survival of the flattest. Nature. 2001 Jul;412(6844):331–3.

6. Wright S. Fisher's Theory of Dominance. The American Naturalist. 1929 May;63(686):274–9.

7. Ancel LW, Fontana W. Plasticity, evolvability, and modularity in RNA. Journal of Experimental Zoology. 2000;288(3):242–83.

8. Kawecki TJ. The Evolution of Genetic Canalization Under Fluctuating Selection. Evolution. 2000;54(1):1–12.

9. Smith JM, Burian R, Kauffman S, Alberch P, Campbell J, Goodwin B, et al. Developmental Constraints and Evolution: A Perspective from the Mountain Lake Conference on Development and Evolution. The Quarterly Review of Biology. 1985 Sep;60(3):265–87.

10. Wagner A. Robustness and evolvability: a paradox resolved. Proceedings of the Royal Society B: Biological Sciences. 2007 Oct 31;275(1630):91–100.

11. Ahnert SE. Structural properties of genotype–phenotype maps. Journal of The Royal Society Interface. 2017 Jul 5;14(132):20170275.

12. Fierst JL, Phillips PC. Modeling the evolution of complex genetic systems: The gene network family tree. Journal of Experimental Zoology Part B: Molecular and Developmental Evolution. 2015;324(1):1–12.

13. Bloom JD, Labthavikul ST, Otey CR, Arnold FH. Protein stability promotes evolvability. Proceedings of the National Academy of Sciences. 2006 Apr 11;103(15):5869–74.

14. Ahnert SE, Johnston IG, Fink TMA, Doye JPK, Louis AA. Self-assembly, modularity, and physical complexity. Phys Rev E. 2010 Aug 27;82(2):026117.

15. Greenbury SF, Johnston IG, Louis AA, Ahnert SE. A tractable genotype–phenotype map modelling the self-assembly of protein quaternary structure. Journal of The Royal Society Interface. 2014 Jun 6;11(95):20140249.

16. Ciliberti S, Martin OC, Wagner A. Robustness Can Evolve Gradually in Complex Regulatory Gene Networks with Varying Topology. PLOS Computational Biology. 2007 Feb 2;3(2):e15.

17. Draghi JA, Wagner GP. The evolutionary dynamics of evolvability in a gene network model. Journal of Evolutionary Biology. 2009 Mar 1;22(3):599–611.

18. Fierst JL. A history of phenotypic plasticity accelerates adaptation to a new environment. Journal of Evolutionary Biology. 2011 Sep 1;24(9):1992–2001.

19. Torres-Sosa C, Huang S, Aldana M. Criticality Is an Emergent Property of Genetic Networks that Exhibit Evolvability. PLOS Computational Biology. 2012 Sep 6;8(9):e1002669.

20. Rutherford SL. From genotype to phenotype: buffering mechanisms and the storage of genetic information. BioEssays. 2000;22(12):1095–105.

21. Crombach A, Hogeweg P. Evolution of Evolvability in Gene Regulatory Networks. PLOS Computational Biology. 2008 Jul 11;4(7):e1000112.

22. Draghi JA, Whitlock MC. Phenotypic plasticity facilitqates mutational variance, genetic variance, and evolvability along the major axis of environemental variation. Evolution. 2012 Sep 1;66(9):2891–902.

23. Espinosa-Soto C, Wagner A. Specialization Can Drive the Evolution of Modularity. PLOS Computational Biology. 2010 Mar 26;6(3):e1000719.

24. Watson RA, Wagner GP, Pavlicev M, Weinreich DM, Mills R. The evolution of phenotypic correlations and "developmental memory." Evolution. 2014 Apr 1;68(4):1124–38.

25. Draghi JA, Parsons TL, Wagner GP, Plotkin JB. Mutational robustness can facilitate adaptation. Nature. 2010 Jan;463(7279):353–5.

26. Meyers LA, Ancel FD, Lachmann M. Evolution of Genetic Potential. PLOS Computational Biology. 2005 Aug 26;1(3):e32.

27. Fontana W, Schuster P. Continuity in Evolution: On the Nature of Transitions. Science. 1998 May 29;280(5368):1451–5.

28. Schuster P, Fontana W, Stadler PF, Hofacker IL. From sequences to shapes and back: a case study in RNA secondary structures. Proceedings of the Royal Society of London Series B: Biological Sciences. 1997 Jan;255(1344):279–84.

29. Bolnick DI, Nosil P. Natural selection in populations subject to migration load. Evolution. 2007 Sep 1;61(9):2229–43.

30. Haller BC, Messer PW. SLiM 4: Multispecies Eco-Evolutionary Modeling. The American Naturalist. 2023 May;201(5):E127–39.

31. Smith JM, Haigh J. The hitch-hiking effect of a favourable gene. Genetics Research. 1974 Feb;23(1):23–35.

32. Masel J. Cryptic Genetic Variation Is Enriched for Potential Adaptations. Genetics. 2006 Mar 1;172(3):1985–91.

33. Hernández U, Posadas-Vidales L, Espinosa-Soto C. On the effects of the modularity of gene regulatory networks on phenotypic variability and its association with robustness. Biosystems. 2022 Feb 1;212:104586.

34. Oikonomou P, Cluzel P. Effects of topology on network evolution. Nature Phys. 2006 Aug;2(8):532–6.

35. Paaby AB, Rockman MV. Cryptic genetic variation: evolution's hidden substrate. Nat Rev Genet. 2014 Apr;15(4):247–58.

36. Manicka S, Marques-Pita M, Rocha LM. Effective connectivity determines the critical dynamics of biochemical networks. Journal of The Royal Society Interface. 2022 Jan 19;19(186):20210659.

37. Aldana M, Balleza E, Kauffman S, Resendiz O. Robustness and evolvability in genetic regulatory networks. Journal of Theoretical Biology. 2007 Apr 7;245(3):433–48.

38. Kitano H. Biological robustness. Nat Rev Genet. 2004 Nov;5(11):826–37.

39. Verd B, Monk NA, Jaeger J. Modularity, criticality, and evolvability of a developmental gene regulatory network. Altenberg L, Wittkopp PJ, editors. eLife. 2019 Jun 6;8:e42832.

40. Greenbury SF, Johnston IG, Smith MA, Doye JPK, Louis AA. The effect of scale-free topology on the robustness and evolvability of genetic regulatory networks. Journal of Theoretical Biology. 2010 Nov 7;267(1):48–61.

41. Seo CH, Kim JR, Kim MS, Cho KH. Hub genes with positive feedbacks function as master switches in developmental gene regulatory networks. Bioinformatics. 2009 Aug 1;25(15):1898–904.

42. Kadelka C, Butrie TM, Hilton E, Kinseth J, Schmidt A, Serdarevic H. A meta-analysis of Boolean network models reveals design principles of gene regulatory networks. Science Advances. 2024 Jan 12;10(2):eadj0822.