Trichoderma afroharzianum T22 induces rhizobia and flavonoid-driven symbiosis to promote iron deficiency tolerance in garden pea

Asha Thapa, Md Rokibul Hasan, Ahmad H. Kabir*
School of Sciences, University of Louisiana at Monroe, Monroe, LA 71209, USA
*e-mail: kabir@ulm.edu

ABSTRACT

Iron (Fe) deficiency is a limiting factor for legumes, yet the role of beneficial fungi in mitigating Fe deficiency in garden pea is understudied. In this study, *Trichoderma afroharzianum* T22 colonized the roots of garden pea cultivars exposed to alkali-mediated Fe-deficiency in a host-specific manner. In Fe-deficient Sugar Snap, T22 improved growth parameters consistent with increased tissue Fe and rhizosphere siderophore. The split-root assay demonstrated that the beneficial effects of T22 on Fe deficiency mitigation are the result of a whole-plant association rather than being confined to the root system alone. RNA-seq analysis showed 575 and 818 differentially expressed genes upregulated and downregulated in the roots inoculated with T22 under Fe deficiency. The upregulated genes were mostly involved in the flavonoid biosynthetic pathway (monooxygenase activity, ammonia-lyase activity, 4-coumarate-CoA ligase), along with genes related to mineral transport and redox homeostasis. Further, a flavonoid precursor restored plant health even in the absence of T22, confirming the role of microbial symbiosis in mitigating Fe deficiency. Interestingly, T22 restored the abundance of rhizobia, particularly *Rhizobium leguminosarum* and *Rhizobium indicum*, along with the induction of *NifA*, *NifD*, and *NifH* in nodules, suggesting a connection between T22 and rhizobia under Fe-starvation. Further, the elevated rhizosphere siderophore, root flavonoid, expression of *PsCoA* (4-coumarate-CoA ligase) as well as the relative abundance of *TaAOX1* and *R. leguminosarum* diminished when T22 was substituted with exogenous Fe. This suggests that exogenous Fe eliminates the need for microbiome-driven Fe mobilization, while T22-mediated Fe mitigation depends on flavonoid-driven symbiosis and *R. leguminosarum* abundance. It was further supported by the positive interaction of T22 on *R. leguminosarum* growth in Fe-deficient media. Thus, the beneficial effect of T22 on rhizobia likely stems from their interactions, not just improved Fe status in plants. This study provides the first mechanistic insights into T22 interactions with host and rhizobia, advancing microbiome strategies to alleviate Fe deficiency in peas and other legumes.

Keywords: Abiotic stress, chlorosis, legume crop, biostimulant, siderophore, symbiosis.

Running title: Mitigating iron deficiency in pea plants using T22

1. INTRODUCTION

Fe (Fe) is an abundant element on earth and is a biologically essential component of every living organism, including plants (Connorton et al. 2017). However, several soil factors can affect the availability of Fe to plants. These include, but are not limited to, high soil pH due to elevated bicarbonate and salt, soil with a high cation exchange capacity, high levels of oxides, and low organic matter in the soil (Zhao et al. 2020; Alcántara et al. 2000; Kroh and Pilon 2019). Fe deficiency in soil may lead to reduced Fe accumulation in crops resulting in leaf chlorosis, photosynthetic disturbance, stunted growth, reduced root development, redox imbalance, and poor yield in crops (Kabir et al. 2022; Sun et al. 2016). Specifically, this deficiency in legume crops can be severe, primarily because of the high levels of Fe required for nodule formation (Brear et al. 2013). Fe deficiency can influence both the legume host and the rhizobia individually, or it can directly affect their interactions (Brear et al. 2013; Dixon and Kahn 2004). It has been observed that *Rhizobium leguminosarum* is a prevalent type of rhizobia capable of forming nitrogen-fixing nodules on pea roots (Boeglin et al. 2022).

Different plants employ various strategies for Fe uptake. Dicot plants utilize what is known as Strategy I for Fe acquisition (Waters et al. 2018; Curie and Briat 2003). Plants following Strategy I typically possess ferric chelate reductase activity, which converts Fe(III) to the more readily available Fe(II) (Kabir et al. 2012). Additionally, these plants acidify the rhizosphere through H^+ secretion, which helps solubilize soil Fe (Molnár et al. 2023; Kabir et al. 2012). Phenolic compounds released through root exudates also play a role by chelating and reducing Fe in response to Fe deficiency (Curie and Mari 2017; Rodríguez-Celma and Schmidt 2013). Once Fe is solubilized, its uptake into the root system is facilitated by metal transporters such as *IRT1* (Fe-regulated transporter), which are commonly found in many dicot species (Kabir et al. 2012; Vert et al. 2001). However, the effectiveness of Strategy I responses largely depends on genetic factors, which can render many high-yielding and popular crop varieties sensitive to Fe deficiency.

Plants can accommodate a vast array of microorganisms in and around their organs which may offer a sustainable solution for fortifying crops with the supply of Fe (Kabir and Bennetzen 2024; Jin et al. 2014). Among the beneficial fungi, arbuscular mycorrhizal fungi (AMF) appear to be almost exclusively studied for their helpful role in plants. However, *Trichoderma* spp. is one of the main groups of rhizosphere mycorrhizal fungi, other than AMF, that can promote plant growth and development through a variety of means (Harman et al. 2004; Qi and Zhao 2013). Of them, *T. afroharzianum* T22 has already demonstrated shreds of evidence for plant growth promotion and abiotic stress tolerance (Bjørk et al. 2023; Kabir et al. 2022). However, there are undoubtedly still a lot of unanswered questions regarding the mechanisms of connection of this fungus with host plants in response to abiotic stresses. In a previous study, we reported that T22 triggered Strategy-I responses in Fe-deficient soybean, leading to the upregulation of *GmFRO2* and *GmIRT1* (Kabir et al. 2022). Recent research has shown that T22 significantly upregulated multiple genes related to Fe uptake, auxin synthesis, nicotianamine synthase 3, and phytosiderophore transporter 1, aiding sorghum plants in coping with Fe deficiency (Kabir and Bennetzen 2024). Studies suggest that fungi-plant symbiosis is involved in local and/or systemic signaling, predominantly depending on host genetic factors (Giovannetti et al. 2024; MacLean et

al. 2017). Symbiosis requires continual signaling between the symbionts and the activation of new transcriptional programs in the plant host, especially in the roots (Bucher et al. 2014). This process triggers changes essential for the plant cell to host the fungal endosymbiont and maintain symbiosis. Of many molecules involved in symbiosis, phytohormones play a key role in the symbiosis between most land plants and fungi (Liao et al. 2018; Lopez-Obando et al. 2015). Other than plant hormones, flavonoids released by plants, have been found to promote microbial colonization and serve as pre-symbiotic signaling molecules in microbial-plant symbiosis (Lidoy et al. 2023; Schliemann et al. 2008). Coumarins, secondary metabolites synthesized via the phenylpropanoid pathway in plants, act as signaling molecules regulating interactions between symbiotic microbes and plants (Harbort et al. 2020; Stringlis et al. 2009). However, this information is still limited to very few crops.

Garden pea (*Pisum sativum* ssp. *hortense*) is an important pulse crop with significant nutritional value, genetic diversity, and available genetic tools, making it a valuable research species (Pandey et al. 2022; Hacisalihoglu et al. 2021; Chaudhari et al. 2020). Addressing Fe deficiency in crops often involves the use of costly and environmentally unsafe soil amendments and fertilizers. Evidence suggests that microbiomes can positively influence Fe status in plants (Kabir and Bennetzen 2024; Kabir et al. 2022; Bhardwaj et al. 2014). However, the specific role and mechanistic basis of T22 in pea plants, particularly its impact on nodule formation under Fe deficiency, remains unclear. Therefore, we investigated whether T22 colonizes garden pea in a host-specific manner and how it induces transcriptional changes that promote symbiotic signaling and Fe deficiency tolerance. Additionally, we sought to determine if the restoration of rhizobia abundance in root nodules is due to the direct effect of T22 on rhizobia or its indirect effect through improving Fe availability in Fe-deficient peas.

2. MATERIALS AND METHODS

2.1 Plant cultivation and growth conditions

The study utilized seeds from six garden pea cultivars: Lillian, Little Marvel, Green Arrow, Sugar Sprint, Sugar Bon, and Sugar Snap. Following initial screening, Sugar Snap was selected for the remaining experiments. The seeds were surface sterilized by soaking in a 4% sodium hypochlorite solution for 5 min, then washed three times with sterile water. Finally, the seeds were placed in a tray and incubated at 25°C for 2 d. After germination, uniform and healthy seedlings were transplanted into pots (containing 500 g of a soil mix) comprising soil collected from a pea field and commercial potting mix in a 1:2 ratio in four different treatments of lime (7.5 g of $NaHCO_3$ and 4.5 g of $CaCO_3$) and *Trichoderma afroharzianum* T22 (1 ×10□ cfu/gram) as follows: (i) control (soil without lime pH ~6.5), (ii) alkali-mediated Fe- (soil mixed with lime pH ~7.8), (iii) alkali-mediated Fe- T22+ (soil mixed with lime pH ~7.8 and T22 inoculum and (iv) T22+ (soil without lime pH ~6.5 and T22 inoculum. The T22 inoculum was cultured on potato dextrose agar (PDA) medium to verify the viability of the fungal spores before inoculation (Supplementary Fig. S1). During the cultivation period, 15 mM $NaHCO_3$ was added weekly to maintain the pH of the Fe-deficient soil ~7.8, inducing alkali-mediated Fe deficiency in the plants. This cultivation process continued for 4 weeks prior to data measurement.

In a targeted experiment, cinnamic acid, a precursor in the flavonoid biosynthetic pathway (Li et al. 2020), was supplemented into the soil (0.5 g/500 g soil) to investigate its potential to induce Fe deficiency tolerance in Sugar Snap in the absence of exogenous T22. Additionally, two new

treatment groups were included: Fe-FeEDDHA+ (soil mixed with lime, pH ~7.8, and FeEDDHA at 0.5 g/500 g soil) and control (soil without lime, pH ~6.5, and Fe-EDDTA at 0.5 g/500 g soil). These targeted studies were conducted for 5 weeks before data analysis. All experiments were performed in a randomized complete block design with 9 biological replications per treatment in a growth chamber with a light regimen of 10 h of light and 14 h of darkness (250 μmol m^2 s^{-1}) at approximately 25°C. The physicochemical characteristics of the soil are detailed in Supplementary Table S1.

2.2 Measurement of morphological features and chlorophyll fluorescence kinetics

The shoot height and root length of each plant were measured using a measuring tape. The harvested shoots and roots were then dried for 3 d at 70°C in an electric oven before recording their dry weights. For nodule assessment, the roots were gently washed to remove soil particles, and nodules were carefully harvested, counted with a magnifying lens, and stored at -80°C for further analysis. Additionally, chlorophyll content and fluorescence kinetics (OJIP), including Fv/Fm (maximal photochemical efficiency of PSII) and Pi_ABS (photosynthetic performance index), were measured on the uppermost fully developed leaves at three different spots. Chlorophyll content was assessed using a handheld SPAD meter (AMTAST, United States). The chlorophyll score refers to the relative measure of chlorophyll content in leaf tissue, obtained using a chlorophyll meter to assess the photosynthetic capacity and health in plants (Uddling et al. 2007. In addition, fluorescence kinetics were measured with a FluorPen FP 110 (Photon Systems Instruments, Czech Republic). For OJIP analysis, leaves were dark-adapted for 1 h before data collection.

2.3 Determination of H_2O_2 in roots

For H_2O_2 determination, root samples were mixed in a solution of 0.1% trichloroacetic acid, as previously described (Alexieva et al. 2001). Briefly, the samples were finely ground into a powder using a mortar and pestle. The extracted fluid underwent centrifugation for 15 min at 10,000 rpm to separate cellular residues. The resulting upper aqueous phase was supplemented with potassium iodide (1 M) and phosphate buffer (10 mM, pH adjusted to 7.0), and then left in darkness for 60 min. Finally, the optical density of the solution was measured at 390 nm using a spectrophotometer (ThermoFisher, Waltham, USA), and the concentration of H_2O_2 was calculated based on the standard curve.

2.4 Molecular detection of T22 colonization in roots

To account for potential variation in T22 colonization across different root sections of garden pea, we performed root partitioning to identify the segment with the highest colonization. The roots were divided into four segments: the mature zone (1–3 cm), the elongation zone (3–6 cm), the meristematic zone (6 cm and beyond), and the root tips, as depicted in Supplementary Fig. S4. The establishment and colonization of T22 in the roots of Sugar Snap cultivated under Fe deficiency were evaluated by measuring the relative abundance of a T22 marker gene (*TaAOX1*: L-amino acid oxidase gene) using DNA-based qPCR (Kabir and Bennetzen 2024). Briefly, root samples were washed twice in fresh sterile phosphate-buffered saline, followed by a brief vortex and two rinses with sterile water. The DNA was extracted on approximately 0.2 g of root samples using CTAB (cetyl trimethylammonium bromide) method (Clarke 2009). Subsequently, DNA was quantified using a NanoDrop ND-1000 Spectrophotometer (Wilmington, USA) and diluted to equal quantities for each DNA sample. The qPCR analysis was conducted using a CFX96

Touch Real-Time PCR Detection System (Bio-Rad, USA), with specific primers (forward: 5′-GTC GGT AGC TGA AAG GGG AT-3′; reverse: 5′-AT TAG AGG CCG GAA ACA CC-3′) designed to amplify a fragment of *TaAOX1* of T22 (Kabir and Bennetzen 2024). Additionally, the relative abundance of *Rhizobium leguminosarum* (RHIZ) was detected in the nodules using specific primer pairs (Boonen et al. 2010). The iTaq™ Universal SYBR® Green Supermix (Bio-Rad, USA) was used in all reactions along with gene-specific primers (Supplementary Table S2) for *PsGAPDH* that served as the internal control for relative quantification using $2^{-\Delta\Delta}$CT method (Livak and Schmittgen 2001).

2.5 Determinations of siderophore levels in the rhizosphere

Siderophore content in the rhizosphere soil was determined using a chrome azurol S (CAS) assay (Himpsl and Mobley, 2019). Briefly, rhizosphere soil samples were homogenized with 80% methanol and centrifuged for 15 min at 10,000 rpm. The supernatant (500 µl) was mixed with 500 µl of CAS solution and incubated at room temperature for 5 min. Absorbance was measured at 630 nm, using 1 ml of CAS reagent as a reference. Siderophore levels per gram of soil sample were calculated with the formula: % siderophore unit = [(Ar - As) / Ar] x 100, where Ar is the absorbance of the reference (CAS reagent only) at 630 nm and As is the absorbance of the sample (CAS reagent + soil sample) at 630 nm.

2.6 Nutrient analysis in root and shoot

Nutrient analysis was performed on both roots and young leaves. The roots were cut, washed with running tap water, and then immersed in 0.1 mM $CaSO_4$ for 10 min, followed by a rinse with deionized water. Young leaves were separately washed with deionized water. All samples were placed in individual envelopes and dried in an electric oven at 75°C for 3d. Elemental concentrations were measured using inductively coupled plasma mass spectrometry (ICP-MS) at the Agricultural and Environment Services Laboratories at the University of Georgia.

2.7 Split-root assay

The plants were cultivated for 2 weeks in sterile vermiculite before conducting the split root assay in soil pots. This was done using a single pot with a partitioned split-root assembly (compartment) with some modifications as previously described (Thilakarathna and Cope 2021). The taproot of each seedling was cut in the middle, and the lateral root system was evenly divided between two compartments within a single pot, filled with natural field soil and commercial potting mix (1:2). The compartments were separated by a solid plastic barrier. Each split-root system was subjected to the same growth conditions, including the presence or absence of Fe deficiency. In one compartment of each split-root system, T22 was inoculated, while the other compartment remained uninoculated, based on the treatment. The split-root plants were cultivated for an additional four weeks before data collection. The split-root assay was performed under three conditions: control/control (-/-), T22/T22 (+/+), and control/T22 (-/+).

2.8 RNA-sequencing and bioinformatics analysis

RNA-seq analysis was conducted on roots collected from the meristematic zone. Before RNA extraction, roots were thoroughly cleaned by rinsing twice with sterile water and vortexing for 10 sec in sterile phosphate-buffered saline to remove surface contaminants. The cleaned roots were then ground into a powder using liquid nitrogen and a pre-cooled pestle and mortar. RNA extraction was performed using the SV Total RNA Isolation System (Promega Corporation,

USA). One microgram of RNA per sample, with a RIN value higher than 8, was used for RNA-seq library preparation. Libraries were prepared with the KAPA RNA HyperPrep Kit with Poly-A Selection (Kapa Biosystems, USA), followed by amplification using the KAPA HiFi HotStart Library Amplification Kit (Kapa Biosystems, USA). Sequencing was carried out on an Illumina NovaSeq 6000 instrument with 0.2 Shared S4 150 bp paired-end sequencing at the RTSF Genomics Core at Michigan State University. A total of 92.5% of reads passed filters with a quality score of ≥ Q30, generating 917.23 Gbp of data.

Bioinformatics analysis of raw FastQ files was carried out using Partek Flow genomic analysis software (Partek, St. Louis, MO, USA). High-quality clean reads were obtained by removing adaptor sequences and low-quality reads using the Trimmomatic tool (Bolger et al. 2014). The clean reads were aligned to the pea reference genome using HISAT2 (Kim Et al. 2015). Read counts mapped to each gene were quantified with HTSeq (Anders et al. 2015). Differentially expressed genes (DEGs) were identified based on an adjusted P-value of <0.05 and a fold change of 2 or greater, using the false discovery rate (FDR) method implemented in the DESeq2 R package (v1.16.1). This analysis was based on FPKM (fragments per kilobase of exon per million mapped fragments). Venn diagrams were created using VennDetail (Hinder Et al. 2017), and heatmaps were generated using pheatmap (Hu et al. 2021) in the R programming environment. Gene enrichment analyses of garden pea DEGs were performed using ShinyGO 0.76.3 (Ge et al. 2020), with gene ontology data sourced from EnsemblPlants resources (Yates et al. 2022).

2.9 Nanopore-based 16S sequencing in root nodules

The nodules were first washed with running tap water, then surface-sterilized in a 3% sodium hypochlorite solution for 3 min and rinsed three times with sterile distilled water. Total DNA was extracted using the CTAB method as described by Clarke (2009). PCR reactions were conducted with 5 ng of DNA from each sample and 16S primers 27F (AGAGTTTGATCMTGGCTCAG) and 1492R (CGGTTACCTTGTTACGACTT) to amplify the near full-length bacterial 16S rRNA gene. Thermal cycling conditions included an initial denaturation at 95°C for 2 min, followed by 30 cycles of denaturation at 95°C for 30 sec, annealing at 60°C for 30 sec, and extension at 72°C for 2 min. A final extension at 72°C for 10 min and a hold at 4°C were performed. Amplicons from each sample were ligated to pooled barcoded reads using the 16S Barcoding Kit 24 V14 (Oxford Nanopore Technologies, Oxford, UK) for library construction. Sequencing was conducted on a MinION portable sequencer (Oxford Nanopore Technologies) using a Flongle flow cell (R10.4.1). MinION™ sequencing data (FASTQ files) were processed using EPI2ME software from Oxford Nanopore Technologies to produce pass reads with an average quality score > 9. Adapter and barcode sequences were trimmed using the EPI2ME Fastq Barcoding workflow (Oxford Nanopore Technologies). Principal coordinate analysis, alpha diversity, and relative abundance of the bacterial community were calculated using the R packages phyloseq and vegan. Non-parametric Kruskal–Wallis tests were used to identify differences in taxa abundance at a 5% significance level (McMurdie and Holmes, 2013).

2.10 Real-time qPCR analysis of genes

Before RNA extraction from pea nodules, surface contaminants were thoroughly removed by washing with PBS buffer. Briefly, RNA extraction and cDNA conversion were performed using the SV Total RNA Isolation System (Promega Corporation, USA) and GoScript™ Reverse

Transcriptase (Promega Corporation, USA) kits, respectively. Subsequently, the expressions of *NifA* (nitrogenase Fe protein regulatory protein), *NifD* (nitrogenase FeMo protein alpha subunit), and *NifH* (nitrogenase FeMo protein beta subunit) associated with the nitrogen fixation process in nodules as well as the expression of *PsCoA* (*Psat2g188920*) gene in roots were studied using gene-specific primers (Supplementary Table S2) in CFX96 Real-Time system (Bio-Rad, USA). The qPCR reaction was conducted with an initial denaturation at 95°C for 3 min, followed by 40 cycles of denaturation at 95°C for 5 sec, annealing at 55°C for 30 sec, and extension at 60°C for 5 min. Expression levels of target genes were analyzed using the $dd^{-\Delta Ct}$ method (Livak and Schmittgen 2001), with normalization to *PsGAPDH*.

2.11 Determination of total flavonoid in roots

The total flavonoid content was measured in pea roots as previously described (Piyanete et al. 2009). Briefly, 0.5 mL of a 70% ethanol extract from the roots was mixed with 2 mL of distilled water and 0.15 mL of a 5% $NaNO_2$ solution. After incubating for 6 min, 0.15 mL of a 10% $AlCl_3$ solution was added, and the mixture was left to stand for another 6 min. Then, 2 mL of a 4% NaOH solution was added. The mixture was diluted to a final volume of 5 mL with methanol and thoroughly mixed. After a 15-min incubation period, the absorbance was measured at 510 nm using a spectrophotometer against a blank. The total flavonoid concentration was expressed in milligrams per gram of dry weight, calculated using a quercetin standard curve.

2.12 Microbial co-culture method

We studied the interactions between T22 and *R. leguminosarum* on nutrient agar under Fe-sufficient and Fe-deficient conditions, supplemented with 15 mM $NaHCO$ to raise the pH to 7.8. The microbial species (5 µl inoculum) were cultured in three different combinations: T22 alone, *R. leguminosarum* alone, and T22 + *R. leguminosarum*. The growth of the microbial species was measured after 7d of culture using ImageJ software (National Institutes of Health, USA). The radius of microbial colonies was measured at three points and then averaged.

2.13 Data analysis

The statistical significance of each variable was assessed using analysis of variance (ANOVA), with the treatment considered as the independent variable. Subsequently, Duncan's Multiple Range Test (DMRT) was performed using SPSS Statistics 20.0 (IBM). Graphical presentations were created with GraphPad Prism 7 and the R package ggplot2.

3. RESULTS

3.1 Effect of T22 on root colonization and morphological features

The colonization efficiency of T22 in roots and its effects on the growth parameters of different garden pea cultivars exposed to Fe deficiency were evaluated. The relative abundance of *TaAOX1* (a marker of T22 colonization) did not differ significantly in the roots of Lillian, Little Marvel, Green Arrow, and Sugar Sprint, whether inoculated with T22 or not, in the presence or absence of alkali-mediated Fe deficiency (Supplementary Fig. S1A). In Sugar Bon and Sugar Snap, Fe deficiency caused no significant changes in *TaAOX1* abundance compared to controls. However, T22 addition under Fe deficiency resulted in a significant increase in *TaAOX1* abundance in these two cultivars. Furthermore, T22 inoculation in Fe-sufficient plants significantly increased the relative abundance of *TaAOX1* in Sugar Bon compared to plants inoculated with T22 under Fe deficiency. In Fe-sufficient conditions, Sugar Snap inoculated with

T22 showed a significant decrease in *TaAOX1* compared to the plants supplemented with T22 under Fe deficiency (Supplementary Fig. S1A). We found that Sugar Snap showed a significant reduction in SPAD scores in leaves under Fe-deficient conditions. However, SPAD scores significantly increased in the leaves of Fe-deficient Sugar Snap inoculated with T22 relative to Fe-deficient plants. Fe-sufficient plants inoculated with T22 had a similar SPAD score to Fe-sufficient controls in Sugar Snap (Supplementary Fig. S1B). Sugar Snap showed a significant decline in root length and root dry weight under Fe-deficient conditions compared to controls. However, T22 supplementation in both Fe-deficient and Fe-sufficient plants led to a significant improvement in root length and root dry weight compared to Fe-deficient plants without T22 (Supplementary Fig. S1C- S1D). Lastly, Sugar Snap exhibited significant declines in both shoot height and shoot dry weight under Fe deficiency compared to controls. Fe-deficient Sugar Snap inoculated with T22 showed a significant increase in these parameters to those of plants solely cultivated under Fe shortage. T22 inoculated in Sugar Snap under Fe sufficiency showed similar shoot height and shoot dry weight to those of controls (Supplementary Fig. S1E-S1F).

In this study, Fe-deficient Sugar Bon inoculated with T22 showed no significant increase in leaf SPAD score, root length, or shoot height compared to plants grown solely under Fe deprivation (Supplementary Fig. S1B, S1C, and S1E). However, the root and shoot dry weights of Fe-deficient Sugar Bon inoculated with T22 significantly increased compared to plants under Fe-deficient conditions alone (Supplementary Fig. S1D-S1F). Among the other genotypes of garden peas, T22 inoculation showed no significant improvement in leaf SPAD score, root length, root dry weight, shoot height, and shoot dry weight in Lillian, Little Marvel, Green Arrow, and Sugar Sprint exposed to Fe deficiency compared to plants grown solely under Fe shortage (Supplementary Fig. S1B- S1F).

3.2 Effect of T22 on photosynthetic parameters and rhizosphere siderophore

In this study, Fe deficiency caused substantial retardation in both aboveground shoot height and belowground root length growth in Sugar Snap (Fig. 1A-1D). However, plants inoculated with T22, regardless of Fe availability, exhibited significant increases in both aboveground and belowground growth parameters compared to plants cultivated solely under Fe deficiency (Fig. 1A-1D). In this study, leaf chlorophyll fluorescence indices showed significant variations in response to the absence or presence of Fe deficiency and T22 (Fig. 1E-1F). The Fv/Fm significantly declined in the leaves due to Fe deficiency compared to controls. The addition of T22 to Fe-deficient soil led to a significant increase in Fv/Fm relative to Fe-starved plants. Plants inoculated with T22 under Fe-sufficient conditions showed similar Fv/Fm values compared to Fe-sufficient controls (Fig. 1E). Additionally, Pi_ABS significantly decreased in response to Fe deficiency compared to Fe-sufficient plants. However, the addition of T22, regardless of Fe availability, resulted in a significant increase in Pi_ABS in the leaves compared to Fe-deficient plants (Fig. 1F).

We also assessed the colonization efficiency of T22 in different root partitions of Sugar Snap plants cultivated under Fe deficiency. The meristematic zone (type C) showed a significant increase in the relative abundance of *TaAOX1* compared to other root partitions (mature zone, elongation zone, root tips) of Sugar Snap (Fig. 1G). Further, Fe deficiency led to a significant increase in siderophore levels in the rhizosphere soil compared to Fe-sufficient controls (Fig. 1H). Interestingly, Fe-deficient plants inoculated with T22 exhibited a significant increase in

siderophore levels in the rhizosphere soil compared to plants solely grown in Fe-deficient soil. The addition of T22 to Fe-sufficient soil resulted in siderophore levels similar to controls, but significantly lower than those under Fe-deficient conditions, with or without T22 inoculation (Fig. 1H).

3.3 Changes in elemental content and hydrogen peroxide

During alkali-mediated Fe deficiency, Fe concentrations significantly decreased in both the roots and leaves of plants compared to Fe-sufficient controls (Table 1). Application of T22 to Fe-deficient plants significantly increased Fe levels in both roots and leaves compared to the plants grown solely in Fe-deficient conditions. Additionally, Zn and B levels showed a significant decrease in the roots in response to Fe deficiency compared to Fe-sufficient controls. Supplementation with T22 did not improve Zn and B levels in the roots of either Fe-deficient or Fe-sufficient plants. Further, Zn and B concentrations in the leaves remained unchanged across all treatment groups (Table 1). Mn levels significantly declined in both the roots and leaves of Fe-deficient plants compared to controls. However, Fe-deficient plants inoculated with T22 exhibited a significant increase in Mn levels compared to Fe-starved plants. Further, Fe-sufficient plants inoculated with T22 showed equivalent Mn levels to those of controls (Table 1). Also, S content significantly decreased in both the roots and leaves of Fe-deficient plants compared to controls. However, no significant changes in S levels were observed when Fe-deficient plants were supplemented with T22. T22 inoculated in Fe-sufficient conditions showed similar S levels in roots, but the leaves showed a significant decrease in S levels compared to control plants (Table 1). Furthermore, no significant differences were found in Mg and P concentrations in both roots and leaves regardless of Fe availability and T22 supplementation. Ca levels in the roots showed no significant changes across different treatment groups. However, Ca levels were significantly reduced in the leaves under Fe-deficient conditions relative to controls. Fe-deficient plants supplemented with T22 showed a significant increase in Ca levels compared to plants solely cultivated under Fe deficiency. Fe-sufficient plants inoculated with T22 had similar Ca levels in the leaves relative to controls (Table 1). Furthermore, hydrogen peroxide was significantly increased in the roots of plants exposed to Fe deficiency compared to nontreated controls (Supplementary Fig. S3). However, Fe-deficient plants inoculated with T22 showed a significant decline in hydrogen peroxide compared to the plants solely cultivated under Fe deficiency. Fe-sufficient plants inoculated with T22 showed similar hydrogen peroxide levels to those of controls (Supplementary Fig. S3).

3.4 Determination of signaling cascade facilitating symbiotic association

We established a split-root system to investigate local versus systemic signaling in the symbiosis between T22 and garden pea (Sugar Snap) under different Fe availabilities (Fig. 2A-2B). Plants cultivated without T22 in both compartments (-/-) showed significant retardation in shoot growth under alkali-mediated Fe deficiency compared to Fe-sufficient controls (Fig. 2C). However, plants inoculated with T22 in both compartments (+/+) or in one compartment (-/+) exhibited significant improvements in shoot growth compared to Fe-deficient plants without T22 in any compartment (-/-). Interestingly, no phenotypic difference in shoot growth was observed between plants inoculated with T22 in both compartments (+/+) and those inoculated in just one compartment (-/+) (Fig. 2C). Similarly, plants cultivated without Fe deficiency, whether inoculated with T22 in both compartments or just one, showed no significant differences in shoot phenotype compared to plants grown under control conditions (Fig. 2C). Fe-deficient plants

grown without T22 in both compartments (-/-) had markedly lower SPAD scores compared to those grown under Fe-sufficient conditions (Fig. 2D). Plants inoculated with T22 in both compartments (+/+) had significantly higher SPAD scores in the leaves compared to Fe-deficient plants that were not inoculated with T22 in any compartment (-/-). However, plants inoculated with T22 in only one compartment (-/+) had relatively lower SPAD scores ($P<0.05$) compared to those inoculated with T22 in both compartments (+/+) (Fig. 2D). Lastly, plants cultivated under Fe sufficiency, whether inoculated with T22 in both (+/+) or just one (-/+) compartment, showed similar SPAD scores to plants grown under control conditions (Fig. 2D).

Plants grown under Fe deficiency without T22 in both compartments (-/-) had significantly lower Fv/Fm levels in their leaves compared to the plants that were cultivated in control conditions (Fig. 2E). Fe-deficient plants inoculated with T22 in both (+/+) or any (-/+) compartment showed a significant increase in SPAD scores relative to Fe-deficient plants not inoculated with T22 in any compartment (Fig. 2E). In contrast, plants cultivated under Fe-sufficient conditions and inoculated with T22 T22 in either both (+/+) or one (-/+) compartment exhibited similar Fv/Fm to those of plants not inoculated with T22 in control conditions (Fig. 2E). Further, Fe concentration in the leaves showed a significant decrease in plants grown under Fe deficiency relative to plants cultivated with sufficient Fe (Fig. 2F). Interestingly, T22 inoculated in in both (+/+) or any (-/+) of the compartment regardless of the Fe availability showed a significant increase in leaf Fe levels compared to the Fe-deficient plants not inoculated with T22 (Fig. 2F). Additionally, plants grown under Fe deficiency without T22 in either compartment (-/-) showed significant decreases in shoot dry weight and shoot height compared to Fe-sufficient plants (Fig. 2G-2H). However, the addition of T22 to either both (+/+) or a single (-/+) compartment under Fe deficiency significantly improved shoot dry weight and shoot height compared to Fe-deficient plants not inoculated with T22. Similarly, T22 inoculated in either both (+/+) or one (-/+) compartment with sufficient Fe in the soil showed similar shoot dry weight and shoot height to those of control plants (Fig. 2G-2H). Lastly, root length did not show significant changes regardless of the exogenous supply of T22 in different compartments in the presence or absence of Fe deficiency (Fig. 2I).

3.5 Effect of T22 on transcriptome and gene enrichment under Fe deficiency

We performed RNA-seq analysis using the meristematic segments of roots to explore the biological mechanisms by which T22 alleviates alkali-mediated Fe deficiency in Sugar Snap. Principal component analysis (PCA) revealed that the first principal component (PC1) accounted for 43.2% of the variance, while the second principal component (PC2) explained 15.9% of the variance (Fig. 3A), indicating substantial variability among the sample groups. The RNA-seq analysis demonstrated significant changes in the root transcriptome (fold-change threshold ≥ 2, $p \leq 0.05$) in response to Fe deficiency, with 364 genes upregulated and 478 genes downregulated (Fig. 3B and 3C). Compared to Fe-starved plants, Fe-deficient plants inoculated with T22 showed significant upregulation of 575 genes and downregulation of 818 genes. Between Fe-T22+ and Fe-deficient conditions, only 3 overlapping genes were differentially upregulated, while 60 genes were differentially downregulated in the roots. In plants with sufficient Fe levels inoculated with T22, 417 genes were significantly upregulated, and 379 genes were downregulated. Of these, 54 genes were shared in the upregulated group and 165 genes in the downregulated group. Furthermore, 442 genes were significantly upregulated, and 367 genes

were significantly downregulated in plants inoculated with T22 under sufficient Fe compared to those under Fe deficiency (Fig. 3B-3C).

We further annotated the functions of DEGs using the Ensembl Genomes 58 database for the pea genome and summarized the top candidate genes due to the presence or absence of Fe deficiency and T22 inoculum (Fig. 3D, Supplementary Table S3). Among the genes associated with symbiosis and phytohormone signaling, the expression of *Psat5g201640* (monooxygenase activity), *Psat3g072160* (ammonia-lyase activity), *Psat3g201240* (UDP-glycosyltransferase activity), *Psat2g001960* (ammonia-lyase activity), *Psat1g068400* (UDP-glycosyltransferase activity), *Psat7g063080* (UDP-glycosyltransferase activity), *Psat1g006320* (auxin-responsive protein), *Psat2g188920* (4-coumarate A ligase), *Psat7g116000* (monooxygenase activity), and *Psat5g190680* (ABC-type transporter activity) showed significant upregulation in the roots of Fe-deficient plants inoculated with T22 compared to Fe-deficient conditions. However, some of these genes either remained unchanged or were significantly downregulated (*Psat5g201640*, *Psat3g072160*, *Psat3g201240*, *Psat1g068400*, *Psat7g063080*) due to Fe deficiency compared to Fe-sufficient controls (Fig. 3D, Supplementary Table S3).

In this study, several genes related to mineral acquisition and transport were significantly upregulated in Fe-deficient plants inoculated with T22 relative to Fe-deficient plants which include: *Psat6g141240* (manganese ion binding), *Psat3g159880* (Fe ion binding), *Psat4g113000* (nitrate transporter activity), *Psat5g286200* (Fe ion binding), *Psat4g000120* (inorganic phosphate transporter), *Psat2g189680* (Zn ion binding), *Psat1g123040* (magnesium ion binding), *Psat1g086520* (oxidoreductase activity), *Psat1g011640* (metal ion transporter activity), *Psat1g171760* (metal ion binding). Most of these transcriptomes showed significant downregulation in the roots of plants exposed to Fe deficiency relative to Fe-sufficient controls (Fig. 3D, Supplementary Table S3). We also found that the expression of *Psat1g168800* (peroxidase activity), *Psat3g121200* (peroxidase activity), *Psat2g065320* (glutamyl−tRNA reductase activity), *Psat3g020440* (glutathione peroxidase activity), *Psat5g186720* (meristem development), *Psat5g198360* (glutathione S-transferase), *Psat7g008120* (calcium ion binding), *Psat2g172320* (growth factor activity), *Psat7g008720* (pyrophosphatase activity), *Psat2g173800* (peroxidase activity) which are related to growth and redox regulation significantly induced in roots of Fe-deficient plants inoculated with T22 in contrast to Fe-deprived plants (Fig. 3D). While *Psat2g065320* (glutamyl-tRNA reductase activity), *Psat3g020440* (glutathione S-transferase), *Psat5g186720* (meristem development), *Psat1g002320* (growth factor activity), *Psat7g008120* (calcium ion binding), *Psat7g008720* (pyrophosphatase activity) and *Psat2g173800* (peroxidase activity) showed significant downregulation, the rest of the genes showed no significant changes in response to Fe deficiency related to controls (Fig. 3D). Furthermore, there were no significant changes in the expression levels of any of top genes discussed under three categories in roots of plants inoculated with T22 under sufficient Fe conditions relative to Fe deficient plants inoculated with T22 (Fig. 3D; Supplementary Table S3).

We also analyzed the biological significance of differentially expressed genes (DEGs) in the roots of Fe-deficient Sugar Snap inoculated with T22 using the ShinyGO (v. 0.80) bioinformatics tool. The pathways significantly enriched in the upregulated DEGs were associated with various biological processes (response to stimulus, organic acid catabolic process, L-phenylalanine catabolic process), molecular functions (ammonia-lyase activity, heme binding), and cellular

components (intracellular organelles, organelles) in plants (Fig. 4A-4C). Additionally, several KEGG pathways were enriched among the significantly upregulated DEGs (flavonoid biosynthesis, biosynthesis of secondary metabolites, phenylalanine metabolism, phenylpropanoid biosynthesis, isoflavonoid biosynthesis) and downregulated DEGs (plant-pathogen interaction, MAPK signaling pathway-plant, plant hormone signal transduction, ribosome) in Fe-deficient plants inoculated with T22 (Fig. 4D). The gene enrichment analysis of the co-expressed network of upregulated DEGs also identified several key pathways with varying degrees of enrichment and statistical significance. The most significantly enriched pathways were photosynthesis-antenna proteins, followed by phenylpropanoid biosynthesis and biosynthesis of secondary metabolites with moderate enrichment, while isoflavonoid biosynthesis and homologous recombination showed the lowest significance (Supplementary Fig. S5).

3.6 Effect of cinnamic acid on the response of Fe-deficient pea plants

Given the RNA-seq analysis indicating the involvement of genes linked to the flavonoid biosynthetic pathway in T22-mediated mitigation of alkali-mediated Fe deficiency, we conducted a targeted study to investigate whether cinnamic acid (CA) could elevate flavonoid levels and induce Fe deficiency tolerance in pea plants without exogenous T22. We observed that Fe-deficient plants exhibited significantly lower SPAD scores and Fv/Fm ratios compared to controls. However, plants treated with either cinnamic acid or T22, regardless of Fe deficiency, showed SPAD scores and Fv/Fm ratios statistically similar to those of control plants (Fig. 5A-5B). Plant height and dry weight (both root and shoot) also declined under Fe deficiency. Nevertheless, Fe-deficient plants supplemented with either T22 or cinnamic acid displayed a significant increase in plant height and dry weight compared to Fe-deficient plants without any treatment. Plants treated with T22 or cinnamic acid under Fe-sufficient conditions showed similar plant height and dry weight to controls (Fig. 5C-5D). Overall, the findings of this study align with the observed upregulation of genes within flavonoid biosynthetic pathway in the RNA-Seq results.

Siderophore levels in the rhizosphere soil showed no significant changes under Fe deficiency compared to Fe-sufficient controls (Fig. 5E). Fe-deficient plants supplemented with T22 or cinnamic acid showed a significant increase in siderophore levels compared to Fe-deficient plants. Siderophore levels in plants with sufficient Fe, treated with either T22 or cinnamic acid, were similar to the control plants (Fig. 5E). Furthermore, total flavonoid content in roots showed no significant changes under Fe deficiency relative to controls. However, Fe-deficient plants supplemented with T22 or cinnamic acid exhibited a significant increase in flavonoid content compared to Fe-deficient plants (Fig. 5F). Interestingly, Fe-sufficient plants inoculated with T22 had significantly lower flavonoid levels compared to all other treatment groups. In contrast, Fe-sufficient plants treated with cinnamic acid showed significantly higher flavonoid levels than those of controls (Fig. 5F).

3.7 Changes in nodulation and rhizobia dynamics in roots nodules

The number of root nodules significantly declined under alkali-mediated Fe-deficient conditions compared to the control. However, the addition of T22 under both Fe-deficient and Fe-sufficient conditions led to a significant increase in the number of root nodules, with levels returning to those similar to the controls (Fig. 6A). Principal component analysis (PCA) revealed substantial variation (88.4%), resulting in distinct separation of sample groups (Fig. 6B). We also analyzed

species richness, diversity, and relative abundance of 16S bacterial communities in root nodules. Species richness (Chao1) and species diversity (Shannon index) showed significant variations in bacterial communities in the root nodules of Sugar Snap, depending on the presence or absence of Fe deficiency and T22 inoculation (Fig. 6C-6D).

Nanopore sequencing revealed the differential relative abundance of bacterial genera, with a notable presence of the *Rhizobium* genus in root nodules influenced by Fe deficiency and T22 inoculation (Fig. 6E). In the 16S analysis, the relative abundance of *Rhizobium* significantly decreased in nodules under Fe deficiency but increased significantly when T22 was supplemented in Fe-deficient soil. Plants inoculated with T22 under Fe-sufficient conditions showed a relative abundance of *Rhizobium* similar to that of the controls (Fig. 6E). Additionally, *Sinocapsa* and *Crinalium* did not show substantial changes in root nodules between the control and Fe-deficient soil, regardless of T22 inoculation. However, plants inoculated with T22 under Fe-sufficient conditions exhibited a slight increase in the relative abundance of *Sinocapsa* and *Crinalium* compared to other treatment groups (Fig. 6E). We observed no changes in the abundance of *Gloeobacter*, *Microcoleus*, *Nodosilinea*, *Oculatella*, *Oscillatoria*, and *Pseudanabaena* in nodules, regardless of Fe availability or T22 inoculation (Fig. 6E). Given that *Rhizobium* species are crucial for legume nodulation, we further analyzed at the species level. The relative abundance of *R. indicum*, *R. leguminosarum*, and *R. redzepovicii* significantly decreased in root nodules under Fe deficiency compared to Fe-sufficient controls (Fig. 6F). However, Fe-deficient plants inoculated with T22 showed a significant increase in the relative abundance of *R. leguminosarum* and *R. redzepovicii* compared to Fe-deficient conditions. Plants grown under Fe-sufficient conditions exhibited a similar relative abundance of *R. indicum*, *R. leguminosarum*, and *R. redzepovicii* in root nodules to Fe-sufficient controls (Fig. 6F).

In this study, alkali-mediated Fe deficiency caused a significant decrease in the expression of *NifA* and *NifD* in nodules compared to controls (Fig. 6G-6H). However, Fe-deficient plants inoculated with T22 showed a significant increase in *NifA* and *NifD* expression compared to Fe-deficient plants without T22. Plants with sufficient Fe supplemented with T22 exhibited similar levels of *NifA* and *NifD* expression in nodules to those of controls (Fig. 6G-6H). Furthermore, the expression of *NifH* significantly increased under Fe deficiency compared to controls (Fig. 6I). The addition of T22 to Fe-deficient plants resulted in a significant increase in *NifH* expression in nodules relative to Fe-deficient plants without T22. Fe-sufficient plants inoculated with T22 showed a similar *NifH* expression pattern to that of controls (Fig. 6I).

3.8 Changes in plant responses supplemented with organic Fe

We cultivated Sugar Snap for 5 weeks to compare the effects of alkali-mediated Fe deficiency treatment with T22 and FeEDDHA supplementation. We observed that leaf SPAD scores significantly declined under Fe deficiency compared to controls. However, the addition of either FeEDDHA or T22 inoculum to the soil resulted in a significant increase in SPAD scores compared to Fe-deficient plants (Fig. 7A). T22 inoculated in Fe-sufficient soil exhibited a similar SPAD score to that of controls and Fe-deficient plants supplemented with FeEDDHA. Notably, Fe-sufficient plants supplemented with FeEDDHA showed a significantly higher SPAD score than all other treatment groups (Fig. 7A). In terms of plant height and biomass, Fe deficiency caused a significant reduction compared to controls. The addition of T22 or FeEDDHA significantly improved these morphological traits relative to plants grown solely under Fe

deficiency (Fig. 7B-7C). Plants with sufficient Fe supplemented with either T22 or FeEDDHA exhibited similar plant height and biomass to controls and Fe-deficient plants supplemented with FeEDDHA (Fig. 7B-7C).

Analysis of rhizosphere siderophore levels revealed a significant decrease under alkali-mediated Fe deficiency compared to controls (Fig. 7D). However, Fe-deficient plants inoculated with T22 showed a significant increase in siderophore levels compared to those grown solely under Fe deficiency or Fe sufficiency. In contrast, Fe-deficient plants supplemented with FeEDDHA exhibited a marked decrease in siderophore levels compared to controls, Fe-deficient plants with T22, and Fe-sufficient plants. Fe-sufficient plants inoculated with T22 showed siderophore levels similar to those observed in Fe-sufficient controls. Notably, FeEDDHA supplementation in Fe-sufficient soil resulted in significantly lower siderophore levels compared to all other treatment groups (Fig. 7D).

In this study, the total flavonoid content in the roots did not show significant changes due to Fe deficiency compared to controls. However, Fe-deficient plants inoculated with T22 exhibited a significant increase in flavonoid content relative to those grown solely in Fe-sufficient or Fe-deficient soil (Fig. 7E). Interestingly, Fe-deficient plants treated with FeEDDHA and Fe-sufficient plants supplemented with either T22 or FeEDDHA showed a significant decrease in flavonoid content compared to the other three treatment groups (control, Fe-, and Fe-T22+) (Fig. 7E). Furthermore, leaf nitrogen content significantly decreased under Fe deficiency compared to controls (Fig. 7F). Fe-deficient plants treated with either T22 or FeEDDHA showed a significant increase in nitrogen content compared to Fe-deficient plants without these treatments. Finally, Fe-sufficient plants inoculated with T22 or supplemented with FeEDDHA exhibited similar leaf nitrogen content to controls. Interestingly, Fe-deficient plants supplemented with Fe-EDDHA showed similar flavonoid levels in the roots to those of Fe-sufficient conditions (Fig. 7F).

We further investigated the expression pattern of *PsCoA*, which showed a significant increase in the roots of Fe-deficient plants inoculated with T22 compared to controls and other treatment groups (Fe-, Fe-FeEDDHA, T22+, and FeEDDHA+) (Fig. 7G). The relative abundance of T22 (*TaAOX1*) in the roots significantly increased when T22 was introduced into the soil, both under Fe-deficient and Fe-sufficient conditions, relative to controls and FeEDDHA-treated soil. Notably, the relative abundance of T22 was higher under Fe-deficient conditions compared to Fe-sufficient soil (Fig. 7H). Furthermore, Fe deficiency led to a significant reduction in the relative abundance of *R. leguminosarum* in nodules, with no significant changes observed with FeEDDHA supplementation (Fig. 7I). However, the addition of T22, either under Fe deficiency or Fe sufficiency, and FeEDDHA with Fe sufficiency, resulted in similar levels of *R. leguminosarum* in the nodules as observed in the controls (Fig. 7I).

We also investigated the interaction between T22 and *R. leguminosarum* under varying Fe availability in nutrient media. In Fe-sufficient media, the growth of both T22 and *R. leguminosarum* was significantly reduced when co-cultured compared to their growth when cultured alone (Supplementary Fig. S6). Under Fe deficiency, T22 growth remained unchanged whether cultured alone or co-cultured with *R. leguminosarum*. In contrast, the growth of *R. leguminosarum* significantly increased when co-cultured with T22 in Fe-deficient media compared to when it was cultured alone (Supplementary Fig. S6).

4. DISCUSSION

Soil mineral deficiency, particularly Fe, presents a significant challenge in agriculture, often resulting in stunted plant growth and reduced yields (Bhat et al. 2024). Fe is essential for legume crops as it is a key component of chlorophyll, crucial for nitrogen fixation, and overall plant health (Brear et al. 2013). Traditional methods to enhance plant stress tolerance, such as genetic modification and chemical fertilizers, may lack sustainability and long-term effectiveness (Yadav et al. 2023; Guzmán et al. 2022). A microbiome-based approach, specifically utilizing beneficial microorganisms like T22, offers a sustainable alternative to enhance crop resilience to Fe deficiency (Kabir et al. 2022; Zhao et al. 2020). This study presents the first mechanistic evidence of how T22 interacts with the host to establish a symbiotic relationship, modulates the host transcriptome, and influences nodule microbial dynamics, thereby helping garden peas cope with alkali-mediated Fe deficiency.

4.1 The colonization efficiency of T22 in mitigating Fe deficiency is host-specific:

Fungal colonization in plant roots is highly host-specific, influenced by factors such as host genetics, immune responses, and the compatibility of fungal species with the plant's physiology (Morán-Diez et al. 2015; Liu et al. 2019). In this study, the abundance analysis of the *TaAOX1* gene indicated that only the Sugar Bon and Sugar Snap cultivars exhibited significant fungal colonization. This suggests a genetic basis for host-specific interactions, which could be utilized to enhance fungal symbiosis and stress tolerance in cultivars that are more efficient with microbiomes. Specifically, Sugar Snap, when inoculated with T22, showed the highest fungal colonization and improved photosynthetic attributes under Fe deficiency, with a notable decrease in colonization when Fe was sufficiently available. This highlights the dependency of T22's beneficial effects on the plant's Fe status and underscores the complex interplay between the fungus, the plant, and soil nutrient availability.

Morán-Diez et al. (2015) analyzed the transcriptomic responses of *Trichoderma virens* interacting with maize and tomato roots, identifying 35 genes with differential expression between the two hosts, including 10 genes with higher expression in tomato roots. In our study, partitioning the Sugar Snap root system revealed the highest T22 colonization in the meristematic zone, as indicated by the abundance of *TaAOX1*. The meristematic zone offers rich nutrients, exudates, and a favorable microenvironment, enhancing microbial colonization compared to other root segments (Tsai et al. 2023; Canarini et al. 2019). The role of the meristematic zone of roots in water and mineral absorption and its variation in fungal colonization across root segments have been reported in other studies (Kabir and Bennetzen 2024; López-Bucio et al. 2003). The high T22 abundance in the meristematic zone may be linked to active cell division and growth, providing optimal conditions for this fungal species (Russo et al. 2019). Root partition studies are essential for understanding microbial colonization and interactions, offering insights into how different root sections influence and are influenced by microbial communities (Bart et al. 2022; Habiyaremye et al. 2020). These findings underscore the importance of screening cultivars for favorable T22 colonization to optimize plant-microbe interactions and enhance stress resilience in crops.

4.2 Host flavonoid-mediated modulation of factors promoting symbiosis

Microbial networks enable the exchange of signals and cues among plants, inducing plant responses to cope with environmental changes (Gorzelak et al. 2015). Symbiotic signaling

between microbes and plants exposed to abiotic stress is essential for mediating beneficial interactions that improve plant fitness and resilience. In this study, a split-root assay revealed that the presence of T22 in the rhizosphere, even when split into two distinct compartments, is not necessary to induce Fe deficiency tolerance in garden pea. The study found that tolerance to Fe deficiency in Sugar Snap remained unaffected by the absence of T22 inoculation in one of the compartments. This suggests that the mechanism through which T22 induces Fe deficiency tolerance in garden pea relies on the whole plant interactions triggered by the host plant itself, rather than the root-borne associations within the rhizosphere. However, the determinants that aid colonization are influenced by host genetics, which are typically linked to the secretion of metabolites and responses to environmental cues (Wiesmann et al. 2023; Bapaume and Reinhardt 2012).

Our RNA-seq analysis of the meristematic segment of roots revealed the upregulation of several key genes associated with symbiosis in alkali-mediated Fe-deficient pea plants inoculated with T22. Notably, we observed the induction of genes responsible for monooxygenase activity, ammonia-lyase activity, auxin-responsive proteins, and 4-coumarate A ligase in Fe-deficient Sugar Snap plants inoculated with T22. Monooxygenase enzymes are crucial for maintaining symbiotic relationships between plants and microorganisms, playing roles in signaling, secondary metabolite production, and hormone regulation that enhance plant health and resilience in symbiotic systems (Wu et al. 2020; Mnguni et al. 2020; Dasgupta et al. 2011). Similarly, ammonia-lyase enzymes, particularly phenylalanine ammonia-lyase (PAL), are significant in plant-microbe symbioses. PAL is integral to the phenylpropanoid pathway, converting phenylalanine to cinnamic acid, which is then transformed into 4-coumarate - a key component in the biosynthesis of coumarins and flavonoids (Li et al. 2020). Coumarins, synthesized through the phenylpropanoid pathway, act as signaling molecules that regulate symbiotic interactions between microbes and plants (Harbort et al. 2020; Stringlis et al. 2009). In Fe-deficient plants, 4-coumarate is crucial for enhancing Fe uptake, as demonstrated in Arabidopsis, which coordinates coumarin biosynthesis to optimize Fe acquisition (Paffrath et al. 2024). Coumarins can also modify the root microbiota, addressing Fe deficiency in *Arabidopsis* (Harbort et al. 2020). Additionally, flavonoids released by plants promote microbial colonization and act as pre-symbiotic signaling molecules (Lidoy et al. 2023; Schliemann et al. 2008). Consistent with these findings, KEGG pathway analysis of T22-inoculated roots showed enrichment in flavonoid, isoflavonoid, and phenylpropanoid biosynthetic pathways. Flavonoids are crucial for rhizobia nodulation in legumes, acting as signaling molecules to attract rhizobia to the root, with NodD proteins recognizing specific flavonoids secreted by legumes (Wang et al. 2018; Peck et al. 2006). Additionally, flavonoids serve as antioxidants, protecting plants against environmental stresses (Grace 2000). Thus, our findings suggest that T22-mediated induction of ammonia-lyase and 4-coumarate A ligase in pea roots may be crucial for the initial steps of flavonoid biosynthesis. This could lead to the formation of various flavonoid compounds that promote symbiotic associations with T22 and/or rhizobia, helping pea plants cope with Fe deficiency.

Flavonoid accumulation in plants is regulated by genes involved in the flavonoid biosynthetic pathway (Borevitz et al. 2000). To further validate the role of flavonoids in potential symbiotic relationships with soil microbes and their contribution to Fe deficiency tolerance in pea plants, we conducted a targeted study on the effect of cinnamic acid, a precursor in the flavonoid

biosynthesis pathway. Our study showed that pea plants either inoculated with T22 or supplemented with cinnamic acid exhibited significant improvements in photosynthetic parameters and plant biomass compared to Fe-deficient controls. This enhancement in plant health was associated with increased synthesis of rhizosphere siderophores and flavonoids in the roots. These findings suggest that elevated flavonoid levels may stimulate the release of siderophores by rhizosphere microbes, thereby promoting the solubilization of insoluble Fe and aiding pea plants in coping with Fe stress. Zamboni et al. (2012) found that tomato plants respond to Fe deficiency by upregulating the flavonoid-3-hydroxylase gene, indicating a role for flavonoids in influencing Fe bioavailability in soil. Building on these insights, future research could focus on developing biotechnological approaches, such as creating transgenic plants or optimizing microbial inoculants, to enhance the expression of key flavonoid biosynthesis genes or improve T22 colonization. Such advancements could lead to improved Fe acquisition in garden pea and other legume crops.

4.3 Reversal of Fe-deficiency-induced retardation in mineral and redox status

Once a symbiotic relationship between a plant and a microbe is established, host plants often reprogram their responses to better cope with abiotic stress (Kabir and Bennetzen, 2024; Meena et al. 2017). In this study, garden pea plants cultivated under alkali-mediated Fe deficiency exhibited reduced SPAD scores and decreased photosynthetic efficiency, aligning with previous findings on legume crops under Fe stress (Haque et al. 2022; Kabir et al. 2012). In contrast, inoculation with T22 significantly improved chlorophyll synthesis and photosynthetic performance in these Fe-deficient plants. This improvement is consistent with earlier studies reporting enhanced SPAD scores, photosystem II activity, and photosynthesis performance index following T22 application to mineral-deficient plants (Kabir and Bennetzen 2024; Kabir et al. 2022; Singh et al. 2019). Our findings indicate that T22 inoculation mitigated both morphological and physiological damage in Fe-deficient pea plants, corresponding with increased Fe concentrations in roots and shoots. Additionally, T22 supplementation restored Mn levels in plants under Fe-deficient conditions. Previous research showed that *Trichoderma* strains can elevate Fe levels but may reduce concentrations of Cu, Mn, and Zn in some plants grown in calcareous soils (Kabir and Bennetzen 2024; Santiago et al. 2011). These differential mineral levels may result from variations in mineral forms and changes in metal transporter activity in host plants (Ye et al. 2017). The observed enhancement of Fe and Mn status in Fe-deficient plants due to T22 underscores its potential role in inducing key transporters for nutrient acquisition, as evidenced by the upregulation of specific genes in roots. Consistently, RNA-seq analysis identified genes related to manganese ion binding, Fe ion binding, and metal ion binding that were either downregulated or showed no change but were substantially induced by T22 under Fe-deficient conditions. The variability in resistance to Fe deficiency among different plant cultivars is influenced by distinct mechanisms of homeostasis and gene expression patterns (Atencio et al. 2021; Kabir et al. 2012). In our study, Fe-deficient pea plants inoculated with T22 exhibited upregulation of Fe-related genes (*Psat3g159880*, *Psat5g286200*) compared to non-inoculated plants, suggesting a symbiotic advantage in enhancing Fe uptake. This indicates that Fe-deficient plants may either sense a need for increased Fe uptake or are driven to induce Fe uptake when insoluble Fe(III) is solubilized through microbial interactions in the rhizosphere. Elevated siderophore levels in the rhizosphere under Fe deficiency, especially when T22 was supplemented, support this hypothesis. Siderophores are typically released by bacterial and fungal communities to chelate Fe in Fe-deficient conditions (Jin et al. 2010; Crowley et al.

2006). Harzianic acid, a secondary metabolite produced by T22, binds with Fe(III) and enhances Fe solubilization in the soil, benefiting both other microorganisms and the host plant (Vinale et al. 2013). The variability in siderophore levels released by *Trichoderma*, influenced by host cultivar, as observed in our study (Fig. 1A), implies that elevated siderophore levels in the Fe-deficient rhizosphere are closely associated with exogenous T22, particularly in the roots of Sugar Snap.

The accumulation of reactive oxygen species (ROS) due to Fe deficiency leads to oxidative damage in cellular components, including lipids, proteins, and DNA (Tewari et al. 2013). Since Fe is crucial for primary antioxidant enzymes, oxidative stress is exacerbated in plants under Fe-deficient conditions (Halliwell, 2006). In our study, Fe-deficient pea plants inoculated with T22 not only exhibited upregulation of mineral transporters but also showed increased expression of peroxidase and glutathione-related genes. Consistently, the biochemical analysis of H_2O_2 also showed a significant decline in the roots of Fe-deficient plants inoculated with T22. This suggests that the symbiotic relationship with T22 might enhance ROS scavenging in Fe-starved plants. Although the precise mechanisms by which ROS balance is regulated in plants with mycorrhizal associations are still not fully understood, several studies have highlighted the role of arbuscular mycorrhizal fungi (AMF) and *Trichoderma* in inducing antioxidants to protect plants from abiotic stressors, including Fe deficiency (García-Sanchez et al. 2014; Kabir and Bennetzen, 2024). Flavonoids, known for their antioxidant properties, play a crucial role in detoxifying and scavenging ROS produced as by-products of oxidative metabolism (Jan et al. 2021; Nakabayashi et al. 2014). For instance, Mastouri et al. (2012) demonstrated that T22 colonization in drought-stressed tomato plants enhanced ROS scavenging capabilities and the recycling of oxidized ascorbate and glutathione, contributing to improved ROS-scavenging pathways. Additionally, T22 has been shown to boost antioxidant defense in Fe-deficient soybeans by increasing glutathione reductase activity and levels of glutathione and methionine, which stabilize the redox state (Kabir et al. 2022). Although the specific mechanisms of T22 playing a role in ROS scavenging require further investigation, our findings suggest that T22-mediated increases in flavonoid levels may positively influence ROS homeostasis in Fe-deficient pea plants. Furthermore, maintaining ROS balance can improve photosystem II efficiency, ensuring better light capture and energy conversion in plants exposed to abiotic stress (Nadarajah 2020), and support meristem development by minimizing cellular damage (Huang et al. 2019). Excessive buildup of ROS, especially in photosystems I and II, can lead to damage of Fe-S proteins, essential cofactors in plant photosynthesis (Xiong et al. 2021; Murchie and Niyogi 2011). Our findings indicate that Fe deficiency resulted in reduced chlorophyll synthesis and decreased Fv/Fm in the leaves of garden pea. Since Fe-deficient pea inoculated with T22 substantially improved Fe and ROS status, genes related to photosystem II (*Psat1g002320*) and meristem development (*Psat5g186720*) were consistently upregulated under these conditions. This molecular evidence was supported by improved leaf Fv/Fm ratios and root systems in plants supplemented with T22 under Fe deficiency. Research has shown that mycorrhizal symbiosis mediated by AMF in *Robinia pseudoacacia* enhances PSII activity in chloroplasts, leading to improved water use efficiency (Yang et al. 2015). Additionally, studies have shown that T22 can restore chlorophyll levels and photosystem II activity in other mineral-deficient plants (Kabir et al. 2022; Singh et al. 2019). Our study suggests that T22 may play a crucial role in enhancing photosynthetic efficiency, potentially by indirectly modulating ROS homeostasis.

4.4 T22 restores nodule bacterial dynamics in pea exposed to Fe deficiency

Rhizobia is essential for the growth and stress resilience of legume crops, as they form symbiotic relationships that enhance nitrogen fixation and improve overall plant health under various stress conditions (Shumilina et al. 2023). Given the observed changes in nodulation and expression of nitrogen-fixation genes, we further investigated whether T22 impacts the bacterial diversity and dynamics in root nodules of pea plants exposed to alkali-mediated Fe deficiency. Our results demonstrate that Fe deficiency leads to a marked reduction in the number of root nodules. However, the addition of T22 not only mitigated the negative effects of Fe deficiency but also enhanced nodule formation under both Fe-deficient and Fe-sufficient conditions. This suggests a potential biostimulant effect of T22 on nodule development. The synergistic effects of *Rhizobium* and mycorrhizal interactions are still not fully understood. However, rhizobia and AMF can form symbioses with legumes, significantly enhancing plant mineral nutrition, particularly nitrogen and phosphorus uptake (Ossler et al. 2015; Xie et al. 2018). In this study, microbiome analysis in pea nodules showed substantial changes in species richness and diversity in response to Fe availability and T22 inoculation. These findings highlight the profound impact of both Fe deficiency and T22 inoculation on the overall bacterial community structure within root nodules in pea plants. Fe is essential for the functioning of nitrogenase, the enzyme complex responsible for nitrogen fixation in legume plant nodules (Schwember et al. 2019; Brear et al. 2013). Nanopore sequencing further elucidated the changes in the relative abundance of bacterial genera, particularly focusing on *Rhizobium.* The inoculation of pea plants with T22 significantly increased the relative abundance of *Rhizobium* in Fe-deficient soils. These findings suggest the positive impact of T22 on restoring beneficial *Rhizobium* populations under Fe stress, thereby promoting nodulation and overall plant health of peas exposed to Fe deficiency. However, it is unknown at this point whether the improved nodule formation is a direct consequence of T22 on Rhizobia or the effect of the improved nutrient status of Fe-deficient pea plants indirectly mediated by T22. Although Rhizobia-fungi symbiotic associations share common signals and processes, mycorrhizal symbiosis has been reported to increase phosphorus content. This can boost nitrogenase enzyme activity, leading to higher nitrogen fixation by rhizobial symbionts, and, in return, better mycorrhizal development (Ren et al. 2016; Hao et al. 2019). Additionally, AMF can reduce oxidative stress in the nodules (Porcel et al. 2003) and enhance the carbon metabolism within them (Goicoechea et al. 2004). Furthermore, fungi can produce and modulate plant hormones, which promote root and nodule development (Chanclud and Morel 2016).

We further conducted a targeted study to elucidate whether T22-mediated mitigation of alkali-mediated Fe deficiency in pea plants is directly related to associations with rhizosphere microbes, including rhizobia, or indirectly linked to the improved Fe status of the plants. Fe fertilizers are commonly used to prevent the precipitation of Fe in the soil, ensuring its availability even in alkaline or calcareous soils, leading to healthier, more vigorous plant growth and increased yields (Schenkeveld et al. 2010). In this study, we found that both T22 and FeEDDHA were equally effective in mitigating the retardation of chlorophyll synthesis and plant biomass caused by Fe deficiency in pea plants. However, we observed differences in rhizosphere siderophore levels, root flavonoid content, the expression of *PsCoA*, and the relative abundance of *TaAOX1* and *R. leguminosarum* - all of which showed a simultaneous decrease when Fe-deficient pea plants were supplemented with FeEDDHA compared to plants inoculated with T22. This suggests that the elevation of flavonoids and the restoration of *R. leguminosarum* largely depend on the biological interactions with T22 during the symbiotic process, rather than on the

Fe status of the plants supplied through an exogenous source. Specifically, it implies that T22 was involved in siderophore production and host flavonoid content, potentially enhancing the plant's own Fe acquisition processes, while FeEDDHA primarily provides a direct source of available Fe without stimulating these secondary metabolic pathways. Microbial siderophore production is closely linked to Fe availability in the soil (Timofeeva et al. 2022). When soil Fe levels are low, microorganisms increase siderophore production to scavenge Fe more effectively and make it available to both microbes and plants (Ahmed and Holmström 2014). Kabir and Bennetzen (2024) showed that Fe deficiency significantly increased the siderophore-related (*SIT1*) gene in T22, resulting in the solubilization of Fe, which improved Fe status in sorghum. They found the highest siderophore accumulation in sorghum inoculated with T22 under Fe-deficient conditions compared to plants grown with sufficient Fe. Consistently, the elevated siderophore released by T22 in the rhizosphere of Fe-deficient pea plants may be no longer needed when organic Fe fertilizer is available. However, the elevation of siderophores in the rhizosphere of Fe-deficient plants suggests that not only T22 but also other microbes may be involved. Several studies support the idea that bacterial communities, including rhizobia, release siderophores in response to Fe deficiency, thereby improving Fe uptake by plants alongside other beneficial interactions such as nitrogen fixation (Lurthy et al. 2020; Wang et al. 2024; Brear et al. 2013).

Our follow-up study further demonstrated that Fe-deficient pea plants were able to combat deficiency symptoms with Fe supplementation, without increasing rhizosphere siderophore and root flavonoid content, which is a requirement for T22-mediated improvement of Fe status. The initiation of complex legume-*Rhizobium* development begins with the release of flavonoids (Dong and Song 2020). Since siderophores and flavonoids are involved in symbiotic associations, Fe-deficient plants bypassed this strategy by overcoming Fe deficiency when organic Fe was supplemented in the soil. When Fe is sufficient, bacteria may prioritize other metabolic processes over nodulation to conserve energy, which helps them efficiently allocate resources based on environmental cues, enhancing their overall fitness (Frawley and Fang 2014). Therefore, our findings suggest that the presence of T22, rather than Fe status alone, may be involved in restoring rhizobia dynamics under Fe-deficient conditions, contributing to the mechanistic pathways that mitigate Fe deficiency in pea plants. Thus, T22 may enhance nodulation and support Fe uptake through specific mechanisms that go beyond simply altering the Fe status of the plants. This result is preliminary given the complexity of the soil system, which shapes multifactorial effects on Fe-deficient pea plants. However, our *in vitro* study demonstrated the positive effect of T22 on the growth of *R. leguminosarum* in Fe-deficient nutrient media. These results are consistent with our overall study on the role of T22 in restoring rhizobia dynamics in nodules of Fe-deficient plants. So far, there is a lack of well-documented interaction between T22 and rhizobia in scientific literature. However, it is known that *Trichoderma* can enhance the activities of beneficial bacteria such as Pseudomonas or Bacillus, which contribute to improved plant health and nutrient uptake (Poveda et al. 2022; da Costa et al. 2022). *Trichoderma* species are known to produce a variety of metabolites that can play a role in shaping the composition and dynamics of soil or rhizosphere bacterial communities, influencing overall ecosystem health and plant-microbe interactions (Gao et al. 2023). This correlates with the levels of flavonoids in roots, as rhizobia are also attracted to flavonoid-mediated symbiosis (Shumilina et al. 2023; Li et al. 2019). Furthermore, *Trichoderma* application has been found to shift other fungal communities in the rhizosphere (Asghar and Kataoka 2021; Umadevi et al.

2018). Future research could explore how T22 interacts with rhizobia and identify the specific metabolites responsible for this association in legume plants under varying Fe availability conditions.

CONCLUSION

This study unveils the intricate mechanisms by which T22 interacts with both garden pea hosts and the rhizobia community to mitigate alkali-mediated Fe deficiency. We demonstrated that T22 enhances plant health parameters under Fe deficiency by improving tissue Fe levels and elevating rhizosphere siderophores, which are critical for Fe uptake. Furthermore, T22 fosters the 16S microbial community, particularly enhancing rhizobia abundance and upregulating essential nitrogen fixation genes in nodules (Fig. 8). Our findings also suggest that the beneficial effect of T22 in mitigating Fe deficiency involves whole-plant interactions, not just the root system. RNA-seq analysis revealed significant gene expression changes linked to flavonoid biosynthesis, mineral transport, and redox homeostasis in T22-inoculated pea roots under Fe deficiency. Moreover, the restoration of plant health with cinnamic acid in the absence of T22 reinforces the role of flavonoids in promoting microbial symbiosis. However, the substitution of T22 with Fe supplementation reduced rhizosphere siderophore and flavonoid levels, as well as the abundance of *TaAOX1* and *R. leguminosarum*, nullifying the associations of microbiome-dominated Fe improvement. Overall, our mechanistic insights highlight the potential of T22 as a key player in microbiome-based strategies to alleviate alkali-mediated Fe deficiency in legume crops, paving the way for sustainable agricultural practices. These insights provide a valuable foundation for developing strategies to improve legume growth and stress resilience through targeted microbial interventions.

ACKNOWLEDGMENTS

We express our gratitude to the Genomics Core of Michigan State University. This research was supported by a startup grant (5SFAES-293007) and Elizabeth and Hayden Cutler Endowed Professorship in Biotechnology from the University of Louisiana at Monroe.

AUTHOR CONTRIBUTION

AT was involved in experimental design, plant cultivation, RNA extraction, RNA-seq data analysis, data collection, interpretation, 16S data analysis and prepared the manuscript draft. Md Rokibul Hasan assisted with DNA extraction and microbial co-culture experiments. AHK provided overall supervision, guidance in experimental design, data interpretation, and manuscript revision.

CONFLICT OF INTEREST

We have no conflict of interest.

DATA AVAILABILITY

The data supporting the findings of this study are available in the article and its supplementary information files. Also, the sequencing data used and described in this study has been uploaded to NCBI under the Bioproject accession numbers: PRJNA1133893 (RNA-seq) and PRJNA1134200 (16S).

REFERENCES

Ahmed, E. & Holmström, S. J. (2014) Siderophores in environmental research: roles and applications. *Microbial Biotechnology*, 7(3), 196–208.

Alcántara, E., Romera, F. J., Canete, M. & De la Guardia, M. D. (2000) Effects of bicarbonate and Fe supply on Fe (III) reducing capacity of roots and leaf chlorosis of the susceptible peach rootstock "Nemaguard". *Journal of Plant Nutrition*, 23(11–12), 1607–1617.

Alexieva V, Sergiev, Mapelli S, Karanov E. (2001) The effect of drought and ultraviolet radiation on growth and stress markers in pea and wheat. Plant Cell Environment 24, 1337–1344.

Anders, S., Pyl, P.T. & Huber, W. (2015) HTSeq–a Python framework to work with high-throughput sequencing data. *Bioinformatics*, 31 (2), 166–169.

Asghar, W. & Kataoka, R. (2021) Effect of co-application of *Trichoderma* spp. with organic composts on plant growth enhancement, soil enzymes and fungal community in soil. *Archives of Microbiology*, 203(7), 4281–4291.

Atencio, L.A., Salazar, J.S., Lauter, A.N.M., Gonzales, M.D., O'Rourke, J.A. & Graham, M.A. (2021) Characterizing short- and long-term Fe stress responses in Fe deficiency tolerant and susceptible soybean (*Glycine max* L. Merr.). *Plant Stress*, 2, 100012.

Bapaume, L. & Reinhardt, D. (2012) How membranes shape plant symbioses: signaling and transport in nodulation and arbuscular mycorrhiza. *Frontiers in Plant Science*, 3, 223.

Birt, H. W. G., Tharp, C. L., Custer, G. F. & Dini-Andreote, F. (2022) Root phenotypes as modulators of microbial microhabitats. *Frontiers in Plant Science*, 13, 1003868.

Bhardwaj, D., Ansari, M.W., Sahoo, R.K. & Tuteja, N. (2014) Biofertilizers function as key players in sustainable agriculture by improving soil fertility, plant tolerance, and crop productivity. *Microbial Cell Factories*, 13, 66.

Bhat, M. A., Mishra, A. K., Shah, S. N., Bhat, M. A., Jan, S., Rahman, S., Baek, K. H. & Jan, A. T. (2024) Soil and Mineral Nutrients in Plant Health: A Prospective Study of Iron and Phosphorus in the Growth and Development of Plants. *Current Issues in Molecular Biology*, 46(6), 5194–5222.

Bjørk, P. K., Johansen, N. T., Havshøi, N. W., Rasmussen, S. A., Ipsen, J. Ø., Isbrandt, T., Larsen, T. O. & Fuglsang, A. T. (2023) *Trichoderma harzianum* peptaibols stimulate plant plasma membrane H^+-ATPase activity. *ACS Omega*, 8(38), 34928–34937.

Boeglin, L., Morère Le-Paven, M. C., Clochard, T., Fustec, J. & Limami, A. M. (2022) *Pisum sativum* response to nitrate as affected by *Rhizobium leguminosarum*-derived signals. *Plants*, 11(15), 1966.

Bolger, A.M., Lohse, M. & Usadel, B. (2014) Trimmomatic: a flexible trimmer for Illumina Sequence Data. Bioinforma. btu170.

Boonen, M., Mertens, J., Michiels, J. & Smolders, E. (2010) Quantitative PCR assays to enumerate *Rhizobium leguminosarum* strains in soil also target non-viable cells and overestimate those detected by the plant infection method. *Soil Biology and Biochemistry*, 42(12), 2342-2344

Borevitz, J.O.; Xia, Y.J.; Blount, J.; Dixon, R.A. & (2000) Lamb, C. Activation tagging identifies a conserved MYB regulator of phenylpropanoid biosynthesis. *Plant Cell*, 12, 2383–2393.

Brear, E. M., Day, D. A. & Smith, P. M. (2013) Fe: an essential micronutrient for the legume-rhizobium symbiosis. *Frontiers in Plant Science*, 4, 359.

Bucher M., Hause B., Krajinski F. & Küster H. (2014). Through the doors of perception to function in arbuscular mycorrhizal symbioses. *The New Phytologist*, 204, 833–840.

Callahan, B. J., McMurdie, P. J., Rosen, M. J., Han, A. W., Johnson, A. J. & Holmes, S. P. (2016) DADA2: High-resolution sample inference from Illumina amplicon data. *Nature Methods*, 13(7), 581–583.

Canarini, A., Kaiser, C., Merchant, A., Richter, A. & Wanek, W. (2019) Root exudation of primary metabolites: mechanisms and their roles in plant responses to environment stimuli. *Frontiers in Plant Science*, 10, 157.

Chanclud, E. & Morel, J. B. (2016) Plant hormones: a fungal point of view. *Molecular Plant Pathology*, 17(8), 1289–1297.

Chaudhari, D., Rangappa, K., Das, A., Layek, J., Basavaraj, S., Kandpal, B.K., Shouche, Y., & Rahi, P. (2020) Pea (*Pisum sativum* L.) plant shapes its rhizosphere microbiome for nutrient uptake and stress amelioration in acidic soils of the North-East Region of India. *Frontiers in Microbiology*, 3, 968.

Clarke, J.D. (2009) Cetyltrimethyl ammonium bromide (CTAB) DNA miniprep for plant DNA isolation. Cold Spring Harb. Protoc., pdb.prot5177

Connorton, J. M., Balk, J. & Rodríguez-Celma, J. (2017) Fe homeostasis in plants - a brief overview. *Metallomics: Integrated Biometal Science*, 9(7), 813–823.

Crowley, D.E. (2006) Microbial siderophores in the plant rhizosphere. In: Barton L.L., Abadia J., editors. Fe Nutrition in Plants and Rhizospheric Microorganisms. Springer; Dordrecht, The Netherlands: pp. 169–198.

Curie, C. & Mari, S. (2017) New routes for plant Fe mining. *The New Phytologist*, 214(2), 521–525.

da Costa, S.D.A., Cardoso, A.F., de Castro, G.L.S., Junior, D.D.S., da Silva, T.C. & da Silva, G.B. (2022) Co-inoculation of *Trichoderma asperellum* with *Bacillus subtilis* to promote growth and nutrient absorption in Marandu Grass. *Applied and Environmental Soil Science*, https://doi.org/10.1155/2022/3228594

Dasgupta, K., Ganesan, S., Manivasagam, S. & Ayre, B. G. (2011) A cytochrome P450 monooxygenase commonly used for negative selection in transgenic plants causes growth anomalies by disrupting brassinosteroid signaling. *BMC Plant Biology*, 11, 67.

Dixon, R. & Kahn, D. (2004) Genetic regulation of biological nitrogen fixation. *Nature Reviews Microbiology*, 2, 621–631

Dong, W. & Song, Y. (2020) The Significance of flavonoids in the process of biological nitrogen fixation. *International Journal of Molecular Sciences*, 21(16), 5926.

Frawley, E. R. & Fang, F. C. (2014). The ins and outs of bacterial iron metabolism. *Molecular Microbiology*, 93(4), 609–616.

Gao, P., Qi, K., Han, Y., Ma, L., Zhang, B., Zhang, Y., Guan, X. & Qi, J. (2023) Effect of *Trichoderma viride* on rhizosphere microbial communities and biocontrol of soybean root rot. *Frontiers in Microbiology*, 14, 1204688.

García-S´anchez, M., Palma, J.M., Ocampo, J.A., García-Romera, I. & Aranda, E. (2014) Arbuscular mycorrhizal fungi alleviate oxidative stress induced by ADOR and enhance antioxidant responses of tomato plants. *Journal of Plant Physiology*, 171 (6), 421–428.

Ge, S.X., Jung, D. & Yao, R. (2020) ShinyGO: a graphical gene-set enrichment tool for animals and plants. *Bioinformatics*, 36 (8), 2628–2629.

Giovannetti, M., Binci, F., Navazio, L. & Genre, A. (2024) Nonbinary fungal signals and calcium-mediated transduction in plant immunity and symbiosis. *The New Phytologist*, 241(4), 1393–1400.

Goicoechea, N., Merino, S., & Sanchez-Diaz, M. (2004) Contribution of arbuscular mycorrhizal fungi (AMF) to the adaptations exhibited by the deciduous shrub *Anthyllis cytisoides* L. under water deficit. *Plant Physiology*, 122, 453–464.

Gorzelak, M.A., Asay, A.K., Pickles, B.J. & Simard, S.W. (2015) Inter-plant communication through mycorrhizal networks mediates complex adaptive behaviour in plant communities. *AoB Plants*, 7, plv050.

Grace, S.C. & Logan, B.A. (2000) Energy dissipation and radical scavenging by the plant phenylpropanoid pathway. *Philosophical Transactions of the Royal Society B*, 355, 1499–1510.

Guzmán, M.G, Cellini, F., Fotopoulos, V., Balestrini, R. & Arbona, V. (2022) New approaches to improve crop tolerance to biotic and abiotic stresses. *Physiologia Plantarum*, 174(1), e13547.

Habiyaremye, J. D., Goldmann, K., Reitz, T., Herrmann, S. & Buscot, F. (2020) Tree root zone microbiome: exploring the magnitude of environmental conditions and host tree impact. *Frontiers in Microbiology*, 11, 749.

Hacisalihoglu, G., Beisel, N. S. & Settles, A. M. (2021) Characterization of pea seed nutritional value within a diverse population of *Pisum sativum*. *PloS one*, 16(11), e0259565.

Halliwell, B. (2006) Reactive species and antioxidants. Redox biology is a fundamental theme of aerobic life. *Plant Physiology*, 141 (2), 312–322.

Hao, Z., Xie, W., Jiang, X., Wu, Z., Zhang, X. & Chen, B. (2019) Arbuscular mycorrhizal fungus improves *Rhizobium*–Glycyrrhiza seedling symbiosis under drought stress. *Agronomy*, 9(10), 572.

Haque, A. F. M. M., Rahman, M. A., Das, U., Rahman, M. M., Elseehy, M. M., El-Shehawi, A. M., Parvez, M. S. & Kabir, A. H. (2022) Changes in physiological responses and MTP (metal tolerance protein) transcripts in soybean (*Glycine max*) exposed to differential Fe availability. *Plant Physiology and Biochemistry*, 179, 1–9.

Harbort, C. J., Hashimoto, M., Inoue, H., Niu, Y., Guan, R., Rombolà, A. D., Kopriva, S., Voges, M. J. E. E. E., Sattely, E. S., Garrido-Oter, R. & Schulze-Lefert, P. (2020) Root-secreted coumarins and the microbiota interact to improve Fe nutrition in Arabidopsis. *Cell Host & Microbe*, 28(6), 825–837.e6.

Harman, G. E., Howell, C. R., Viterbo, A., Chet, I. & Lorito, M. (2004) *Trichoderma* species-opportunistic, avirulent plant symbionts. *Nature Reviews Microbiology*, 2(1), 43–56.

Hider, R.C. & Kong, X. (2010) Chemistry and biology of siderophores. *Natural Product Reports*, 27 (5), 637–657.

Himpsl, S.D. & Mobley, H.L.T. (2019) Siderophore detection using chrome azurol S and cross-feeding assays. *Methods in Molecular Biology*, 2021, 97–108.

Hu, K. (2021) Become competent in generating RNA-seq heat maps in one day for novices without prior R experience. *Methods in Molecular Biology*, 2239, 269–303.

Huang, H., Ullah, F., Zhou, D. X., Yi, M. & Zhao, Y. (2019) Mechanisms of ROS Regulation of Plant Development and Stress Responses. *Frontiers in Plant Science*, 10, 800.

Jan, R., Kim, N., Lee, S. H., Khan, M. A., Asaf, S., Lubna, Park, J. R., Asif, S., Lee, I. J. & Kim, K. M. (2021) Enhanced favonoid accumulation reduces combined salt and heat stress through regulation of transcriptional and hormonal mechanisms. *Frontiers in Plant Science*, 12, 796956.

Jin, C.W., Ye, Y.Q. & Zheng, S.J. (2014) An underground tale: contribution of microbial activity to plant iron acquisition via ecological processes. *Annals of Botany*, 113(1), 7-18.

Jin, C. W., Li, G. X., Yu, X. H., and Zheng, S. J. (2010) Plant Fe status affects the composition of siderophore-secreting microbes in the rhizosphere. *Annals of Botany*, 105(5), 835–841.

Kabir, A. H. & Bennetzen, J. L. (2024) Molecular insights into the mutualism that induces iron deficiency tolerance in sorghum inoculated with *Trichoderma harzianum*. *Microbiological Research*, 281, 127630.

Kabir, A. H., Paltridge, N. G., Able, A. J., Paull, J. G. & Stangoulis, J. C. (2012) Natural variation for Fe-efficiency is associated with upregulation of Strategy I mechanisms and enhanced citrate and ethylene synthesis in *Pisum sativum* L. *Planta*, 235(6), 1409–1419.

Kabir, A. H., Rahman, M. A., Rahman, M. M., Brailey-Jones, P., Lee, K. W. & Bennetzen, J. L. (2022) Mechanistic assessment of tolerance to Fe deficiency mediated by *Trichoderma harzianum* in soybean roots. *Journal of Applied Microbiology*, 133(5), 2760–2778.

Kabir, A.H., Rahman, M.M., Haider, S.A. & Paul, N.K. (2015) Mechanisms associated with differential tolerance to Fe deficiency in okra (*Abelmoschus escu*lentus Moench). *Environmental and Experimental Botany*, 112, 16–26.

Kim, D., Langmead, B. & Salzberg, S.L. (2015) HISAT: a fast spliced aligner with low memory requirements. *Nature Methods*, 12 (4), 357–360.

Kroh, G. E. & Pilon, M. (2019) Connecting the negatives and positives of plant Fe homeostasis. *The New Phytologist*, 223(3), 1052–1055.

Li, N., Islam, M. T. & Kang, S. (2019) Secreted metabolite-mediated interactions between rhizosphere bacteria and *Trichoderma* biocontrol agents. *PloS one*, 14(12), e0227228.

Li, S. S., Chang, Y., Li, B., Shao, S. L. & Zhen-Zhu-, Z. (2020) Functional analysis of 4-coumarate: CoA ligase from Dryopteris fragrans in transgenic tobacco enhances lignin and flavonoids. *Genetics and Molecular Biology*, 43(2), e20180355.

Liao, D., Wang, S., Cui, M., Liu, J., Chen, A. & Xu, G. (2018) Phytohormones regulate the development of arbuscular mycorrhizal symbiosis. *International Journal of Molecular Sciences*, 19(10), 3146.

Lidoy, J., Berrio, E., García, M., España-Luque, L., Pozo, M.J., & López-Ráez, J.A. (2023) Flavonoids promote *Rhizophagus irregularis* spore germination and tomato root colonization: A target for sustainable agriculture. *Frontiers in Plant Science*, 5, 1094194.

Liu, A., Zhu, Y., Wang, Y., Wang, T., Zhao, S., Feng, K., Li, L. & Wu, P. (2023) Molecular identification of phenylalanine ammonia lyase-encoding genes *EfPALs* and *EfPAL2*-interacting transcription factors in Euryale ferox. *Frontiers in Plant Science*, 14, 1114345.

Liu, H., Senthilkumar, R., Ma, G., Zou, Q., Zhu, K., Shen, X., Tian, D., Hua, M. S., Oelmüller, R. & Yeh, K. W. (2019) *Piriformospora indica*-induced phytohormone changes and root colonization strategies are highly host-specific. *Plant Signaling and Behavior*, 14(9), 1632688.

Livak, K.J. & Schmittgen, T.D. (2001) Methods. Vol. 25. San Diego, CA: Analysis of relative gene expression data using real-time quantitative PCR and the 2(-Delta Delta C(T)) Method; pp. 402–408.

López-Bucio, J., Cruz-Ramírez, A. & Herrera-Estrella L. (2003) The role of nutrient availability in regulating root architecture. *Current Opinion in Plant Biology*, 6(3), 280–287.

Lopez-Obando, M., Ligerot, Y., Bonhomme, S., Boyer F.D. & Rameau, C. (2015) Strigolactone biosynthesis and signaling in plant development. *Development*, 142, 3615–3619.

Lurthy, T., Cantat, C., Jeudy, C., Declerck, P., Gallardo, K., Barraud, C., Leroy, F., Ourry, A., Lemanceau, P., Salon, C. & Mazurier, S. (2020) Impact of bacterial siderophores on Fe status and ionome in Pea. *Frontiers in Plant Science*, 11, 730.

MacLean, A. M., Bravo, A. & Harrison, M. J. (2017) Plant signaling and metabolic pathways enabling arbuscular mycorrhizal symbiosis. *The Plant Cell*, 29(10), 2319–2335.

Mastouri, F., Bjorkman, T. & Harman, G.E. (2012) *Trichoderma harzianum* enhances antioxidant defense of tomato seedlings and resistance to water deficit. *Molecular Plant-Microbe Interactions*, 25 (9), 1264–1271.

McMurdie, P. J. & Holmes, S. (2013) phyloseq: an R package for reproducible interactive analysis and graphics of microbiome census data. *PloS One*, *8*(4), e61217.

Meena, K. K., Sorty, A. M., Bitla, U. M., Choudhary, K., Gupta, P., Pareek, A., Singh, D. P., Prabha, R., Sahu, P. K., Gupta, V. K., Singh, H. B., Krishanani, K. K. & Minhas, P. S. (2017) Abiotic Stress Responses and Microbe-Mediated Mitigation in Plants: The Omics Strategies. *Frontiers in Plant Science*, 8, 172.

Mnguni, F. C., Padayachee, T., Chen, W., Gront, D., Yu, J. H., Nelson, D. R. & Syed, K. (2020) More P450s are involved in secondary metabolite biosynthesis in *Streptomyces* compared to *Bacillus*, Cyanobacteria, and Mycobacterium. *International Journal of Molecular Sciences*, 21(13), 4814.

Molnár, Z., Solomon, W., Mutum, L. & Janda, T. (2023) Understanding the mechanisms of Fe deficiency in the rhizosphere to promote plant resilience. *Plants*, 12(10), 1945.

Morán-Diez, M. E., Trushina, N., Lamdan, N. L., Rosenfelder, L., Mukherjee, P. K., Kenerley, C. M. & Horwitz, B. A. (2015) Host-specific transcriptomic pattern of *Trichoderma virens* during interaction with maize or tomato roots. *BMC Genomics*, 16(1), 8.

Murchie, E.H. & Niyogi, K.K. (2011) Manipulation of photoprotection to improve plant photosynthesis. *Plant Physiology*, 155, 86–92.

Nadarajah, K. K. (2020) ROS homeostasis in abiotic stress tolerance in plants. *International Journal of Molecular Sciences*, 21(15), 5208.

Nakabayashi, R., Yonekura-Sakakibara, K., Urano, K., Suzuki, M., Yamada, Y., Nishizawa, T., Matsuda, F., Kojima, M., Sakakibara, H., Shinozaki, K., Michael, A. J., Tohge, T., Yamazaki, M. & Saito, K. (2014) Enhancement of oxidative and drought tolerance in Arabidopsis by overaccumulation of antioxidant flavonoids. *The Plant Journal*, 77(3), 367–379.

Ossler, J. N., Zielinski, C. A. & Heath, K. D. (2015) Tripartite mutualism: facilitation or trade-offs between rhizobial and mycorrhizal symbionts of legume hosts. *American Journal of Botany*, 102(8), 1332–1341.

Paffrath, V., Tandron Moya, Y. A., Weber, G., von Wirén, N. & Giehl, R. F. H. (2024) A major role of coumarin-dependent ferric Fe reduction in strategy I-type Fe acquisition in Arabidopsis. *The Plant Cell*, 36(3), 642–664.

Pandey, P.K., Bhowmik, P. & Kagale, S. (2022) Optimized methods for random and targeted mutagenesis in field pea (*Pisum sativum* L.). *Frontiers in Plant Science*, 8, 995542.

Peck, M. C., Fisher, R. F. & Long, S. R. (2006) Diverse flavonoids stimulate NodD1 binding to nod gene promoters in *Sinorhizobium meliloti*. *Journal of Bacteriology*, 188, 5417–5427.

Piyanete, C., Meechai, P. & Nakbanpotecc, W. (2009) Antioxidant activities and phenolic contents of extracts from *Salvinia olesta* and *Eichorniacrassipes*. *Research Journal of Biological Sciences*, 4, 1113–1117.

Porcel, R., Barea, J.M. & Ruiz-Lozano, J.M. (2003) Antioxidant activities in mycorrhizal soybean plants under drought stress and their possible relationship to the process of nodule senescence. *The New Phytologist*, 157, 135–143.

Poveda, J. & Eugui, D. (2022) Combined use of *Trichoderma* and beneficial bacteria (mainly *Bacillus* and *Pseudomonas*): Development of microbial synergistic bio-inoculants in sustainable agriculture. *Biological Control*, 176, 105100.

Qi, W. & Zhao, L. (2013) Study of the siderophore producing *Trichoderma asperellum* Q1 on cucumber growth promotion under salt stress. *Journal of Basic Microbiology*, 53(4), 355–364.

Ren, C.G., Bai, Y.J., Kong, C.C., Bian, B. & Xie, Z.H. (2016) Synergistic interactions between salt-tolerant rhizobia and arbuscular mycorrhizal fungi on salinity tolerance of *Sesbania cannabina* plants. *Journal of Plant Growth Regulation*, 35, 1098–1107.

Rodríguez-Celma, J. and Schmidt, W. (2013) Reduction-based Fe uptake revisited: on the role of secreted Fe-binding compounds. Plant Signal Behav, 8, e26116.

Russo, G., Carotenuto, G., Fiorilli, V., Volpe, V., Chiapello, M., Van Damme, D. & Genre, A. (2019) Ectopic activation of cortical cell division during the accommodation of arbuscular mycorrhizal fungi. *The New Phytologist*, 221(2), 1036-1048.

Santiago, A., Quintero, J.M., Aviles, M. & Delgado, A. (2011) Effect of *Trichoderma asperellum* strain T34 on Fe, copper, manganese, and zinc uptake by wheat grown on a calcareous medium. *Plant and Soil*, 342, 97–104.

Schenkeveld, W. D., Reichwein, A. M., Bugter, M. H., Temminghoff, E. J., & van Riemsdijk, W. H. (2010) Performance of soil-applied FeEDDHA isomers in delivering Fe to soybean plants in relation to the moment of application. *Journal of Agricultural and Food Chemistry*, 58(24), 12833–12839.

Schliemann, W., Ammer, C. & Strack, D. (2008) Metabolite profiling of mycorrhizal roots of *Medicago truncatula*. *Phytochemistry*, 69(1), 112–146.

Shumilina, J., Soboleva, A., Abakumov, E., Shtark, O. Y., Zhukov, V. A. & Frolov, A. (2023) Signaling in Legume-Rhizobia Symbiosis. *International Journal of Molecular Sciences*, 24(24), 17397.

Singh, S.P., Pandey, S., Mishra, N., Giri, V.P., Mahfooz, S., Bhattacharya, A., Kumari, M., Chauhan, P., Verma, P., Nautiyal, C.S. & Mishra, A. (2019) Supplementation of *Trichoderma* improves the alteration of nutrient allocation and transporter genes expression in rice under nutrient deficiencies. *Plant Physiology and Biochemistry*, 143, 351–363.

Stringlis, I. A., de Jonge, R. & Pieterse, C. M. J. (2019) The age of coumarins in plant-microbe Interactions. *Plant and Cell Physiology*, 60(7), 1405–1419.

Sun, C., Wu, T., Zhai, L., Li, D., Zhang, X., Xu, X. & Han, Z. (2016) Reactive oxygen species function to mediate the Fe deficiency response in an Fe-efficient apple Cultivar: An early response mechanism for enhancing reactive oxygen production. *Frontiers in Plant Science*, 7, 1726.

Tewari, R.K., Hadacek, F., Sassmann, S. & Lang, I. (2013) Fe deprivation-induced reactive oxygen species generation leads to non-autolytic PCD in *Brassica napus* leaves. *Environmental and Experimental Botany*, 91, 74–83

Thilakarathna, M. S. & Cope, K. R. (2021) Split-root assays for studying legume-rhizobia symbioses, rhizodeposition, and belowground nitrogen transfer in legumes. *Journal of Experimental Botany*, 72(15), 5285–5299.

Timofeeva, A. M., Galyamova, M. R. & Sedykh, S. E. (2022) Bacterial siderophores: classification, biosynthesis, perspectives of use in agriculture. *Plants*, 11(22), 3065.

Tsai, H. H., Wang, J., Geldner, N. & Zhou, F. (2023) Spatiotemporal control of root immune responses during microbial colonization. *Current Opinion in Plant Biology*, 74, 102369.

Uddling, J., Gelang-Alfredsson, J., Piikki, K. & Pleijel, H. (2007) Evaluating the relationship between leaf chlorophyll concentration and SPAD-502 chlorophyll meter readings. *Photosynthesis Research*, 91(1), 37–46.

Umadevi, P., Anandaraj, M., Srivastav, V. & Benjamin, S. (2018) *Trichoderma harzianum* MTCC 5179 impacts the population and functional dynamics of microbial community in the rhizosphere of black pepper (*Piper nigrum* L.). *Brazilian Journal of Microbiology*, 49(3), 463–470.

Vert, G., Briat, J.F. & Curie, C. (2001) Arabidopsis *IRT2* gene encodes a root-periphery Fe transporter. *The Plant Journal*, 26, 181–189.

Vinale, F., Nigro, M., Sivasithamparam, K., Flematti, G., Ghisalberti, E.L., Ruocco, M. & Lorito, M. (2013) Harzianic acid: a novel siderophore from *Trichoderma harzianum*. *FEMS Microbiology Letters*, 347 (2), 123–129.

Wang, N., Wang, T., Chen, Y., Wang, M., Lu, Q., Wang, K., Dou, Z., Chi, Z., Qiu, W., Dai, J., Niu, L., Cui, J., Wei, Z., Zhang, F., Kümmerli, R. & Zuo, Y. (2024) Microbiome convergence enables siderophore-secreting-rhizobacteria to improve Fe nutrition and yield of peanut intercropped with maize. *Nature Communications*, 15(1), 839.

Wang, Q., Liu, J. & Zhu, H. (2018) Genetic and molecular mechanisms underlying symbiotic specificity in legume-Rhizobium interactions. *Frontiers in Plant Science*, 9, 313.

Wiesmann, C.L., Wang, N.R., Zhang, Y., Liu, Z. & Haney, C.H. (2023) Origins of symbiosis: shared mechanisms underlying microbial pathogenesis, commensalism and mutualism of plants and animals. *FEMS Microbiology Reviews*, 47, fuac048.

Wu, H., Zou, Q., Luo, S., He, D., Li, X., Yan, C. & Cheng, G. (2020) Lack of the slavin-containing monooxygenase FmoA partially impairs the symbiotic interaction of *Mesorhizobium Huakuii* with *Astragalus Sinicus*. *BMC Microbiology*, https://doi.org/10.21203/rs.3.rs-27635/v1

Xie, W., Hao, Z.P., Zhou, X.F., Jiang, X.L., Xu, L.J., Wu, S.L., Zhao, A.H., Zhang, X. & Chen, B.D. (2018) Arbuscular mycorrhiza facilitates the accumulation of glycyrrhizin and liquiritin in *Glycyrrhiza uralensis* under drought stress. *Mycorrhiza*, 28, 285–300.

Xiong, H., Hua, L., Reyna-Llorens, I., Shi, Y., Chen, K.M., Smirnoff, N. & Hibberd, J.M. (2021) Photosynthesis-independent production of reactive oxygen species in the rice bundle sheath during high light is mediated by NADPH oxidase. *Proceedings of the National Academy of Sciences of the United States of America*, 118, e2022702118

Yadav, R. K., Tripathi, M. K., Tiwari, S., Tripathi, N., Asati, R., Chauhan, S., Tiwari, P. N. & Payasi, D. K. (2023) Genome editing and improvement of abiotic stress tolerance in crop plants. *Life*, 13(7), 1456.

Yates, A. D., Allen, J., Amode, R. M., Azov, A. G., Barba, M., Becerra, A., Bhai, J., Campbell, L. I., Carbajo Martinez, M., Chakiachvili, M., Chougule, K., Christensen, M., Contreras-Moreira, B., Cuzick, A., Da Rin Fioretto, L., Davis, P., De Silva, N. H., Diamantakis, S., Dyer, S., Elser, J. & Flicek, P. (2022) Ensembl Genomes 2022: an expanding genome resource for non-vertebrates. *Nucleic Acids Research*, 50(D1), D996–D1003.

Ye, Q., Park, J.E., Gugnani, K., Betharia, S., Pino-Figueroa, A. & Kim, J. (2017) Influence of Fe metabolism on manganese transport and toxicity. *Metallomics*, 16, 1028–1046.

Zamboni, A., Zanin, L., Tomasi, N., Pezzotti, M., Pinton, R., Varanini, Z. & Cesco, S. (2012) Genome-wide microarray analysis of tomato roots showed defined responses to iron deficiency. *BMC Genomics*, 13, 101.

Zhao, L., Wang, Y. & Kong, S. (2020) Effects of *Trichoderma asperellum* and its siderophores on endogenous auxin in *Arabidopsis thaliana* under Fe-deficiency stress. *International Microbiology*, 23(4), 501–509.

Table 1. ICP-MS analysis of nutrients in the root and leaf of pea plants inoculated with or without *T. afroharzianum* (T22) in the absence or presence of Fe deficiency in the soil. Data represents ± SD of three independent biological samples. Statistically significant differences in the ANOVA test among different treatments ($p < 0.05$) are indicated by different letters at the $P < 0.05$ level.

Elements	Root				Leaf			
	Control	Fe-	Fe-T22+	T22+	Control	Fe-	Fe-T22+	T22+
Fe (ppm)	401.0±14.8a	292.9±53.3b	361.1±53.3a	425.7±27.2a	89.5±14.8a	60.5±53.3b	75.3±53.3c	90.1±27.2a
Zn (ppm)	129.5±0.9a	92.5±35.5b	92.0±35.5b	100.3±24.4b	50.1±0.9a	50.6±35.5a	49.4±35.5a	51.0±24.4a
Mn (ppm)	35.9±8.1a	22.7±18.8b	27.8±18.8c	35.9±7.6a	33.8±8.1a	18.6±18.8b	25.3±18.8c	34.6±7.6a
B (ppm)	17.6±0.4a	9.3±6.8b	11.004±6.8b	11.5±4.4b	11.6±0.4a	10.5±6.8a	10.9±6.8a	10.0±4.4a
S (%)	1.06±0.1a	0.80±0.2b	0.87±0.2b	1.15±0.12a	0.55±0.1a	0.28±0.2b	0.38±0.2b	0.37±0.12b
Mg (%)	0.14±0.0a	0.16±0.1a	0.19±0.1a	0.20±0.07a	0.21±0.1a	0.19±0.1a	0.20±0.1a	0.18±0.07a
Ca (%)	0.69±0.3a	0.66±0.5a	0.66±0.5a	0.64±0.11a	1.02±0.3a	0.39±0.5b	0.58±0.5c	1.05±0.11a
P (%)	0.39±0.0a	0.34±0.5a	0.35±0.5a	0.37±0.30a	0.63±0.01a	0.68±0.5a	0.64±0.5a	0.65±0.30a

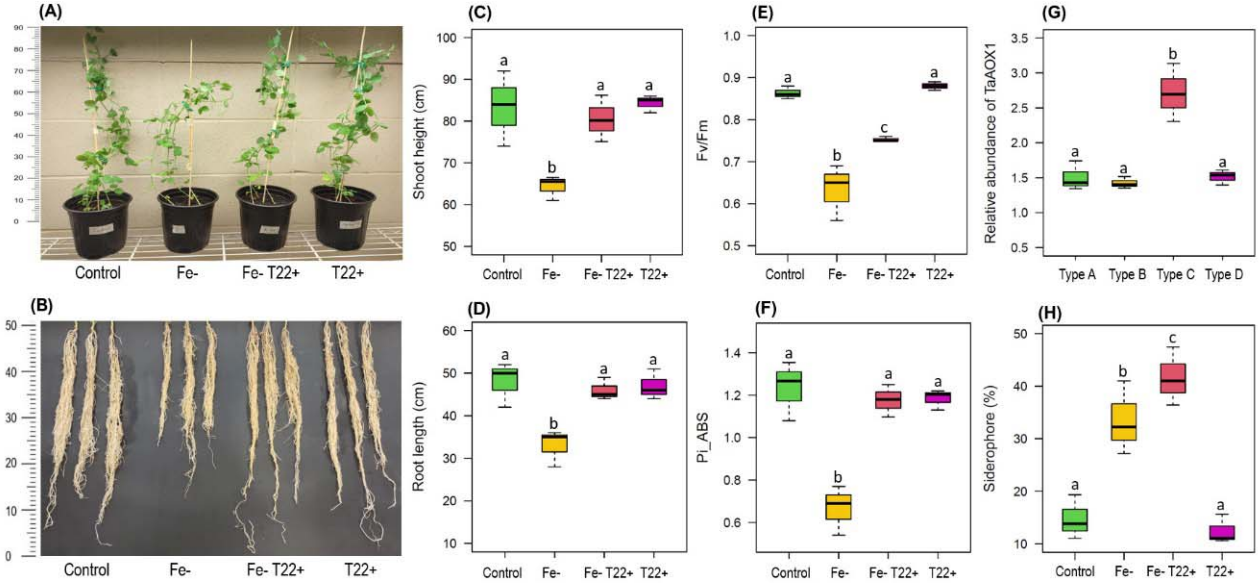

Fig. 1. Changes in aboveground shoot phenotype (A), belowground root phenotype (B), shoot height (C), root length (D), leaf chlorophyll fluorescence indices: *Fv*/*Fm* - maximal quantum yield of PS II (E), Pi_ABS - photosynthetic performance index (F), the relative abundance of *TaAOX1* in different root partitions of Sugar Snap inoculated with T22 under Fe deficiency (type A: mature zone, type B: elongation zone, type C: meristematic zone, type D: root tips) of Sugar Snap (G), and siderophore (%) in rhizosphere soil (H) in Sugar Snap cultivated in the absence or presence of Fe deficiency inoculated with (T22+) or without T22. The data presented are means with standard deviations (n = 3 individual replicates). Statistically significant differences in the ANOVA test among different treatments ($p < 0.05$) are indicated by different letters above the bars, where applicable.

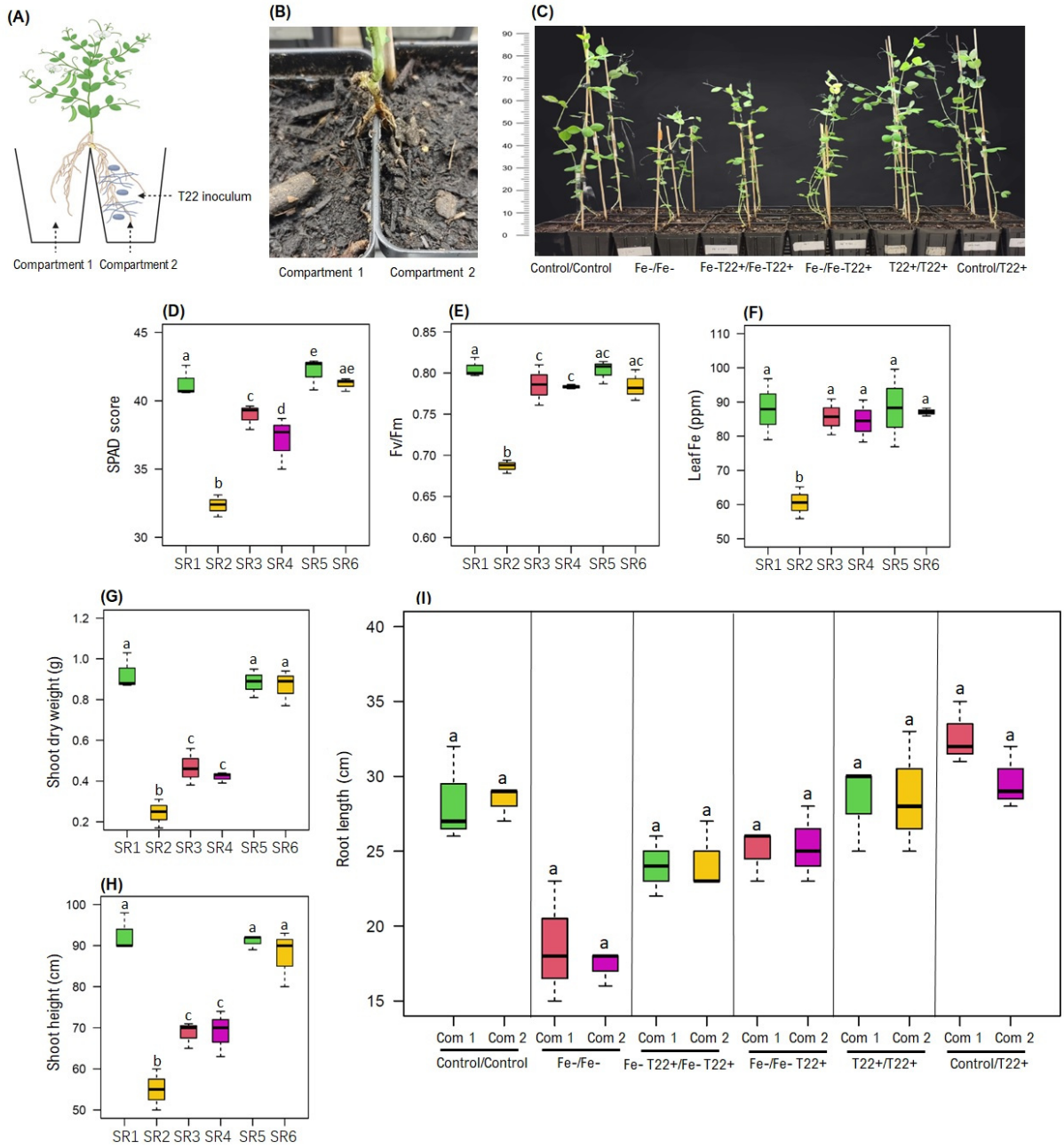

Fig. 2. Schematic diagram of split-root (SR) system (A), establishment of split-root (B), plant phenotype (C), leaf SPAD score (D), leaf *Fv*/*Fm* - maximal quantum yield of PS II (E), leaf Fe (F), shoot dry weight (G), shoot height (H) and root length of plants grown in each compartment (I) in the absence (control) or presence (+) of Fe deficiency and T22 inoculum. The data presented are means with standard deviations (n = 3 individual replicates). Statistically significant differences in the ANOVA test among different treatments ($p < 0.05$) are indicated by different letters above the bars, where applicable. SR1: Control/Control, SR2: Fe-/Fe-, SR3: Fe-T22+/Fe-T22+, SR4: Fe-/Fe-T22+, SR5: T22+/T22+, SR6: Control/T22+.

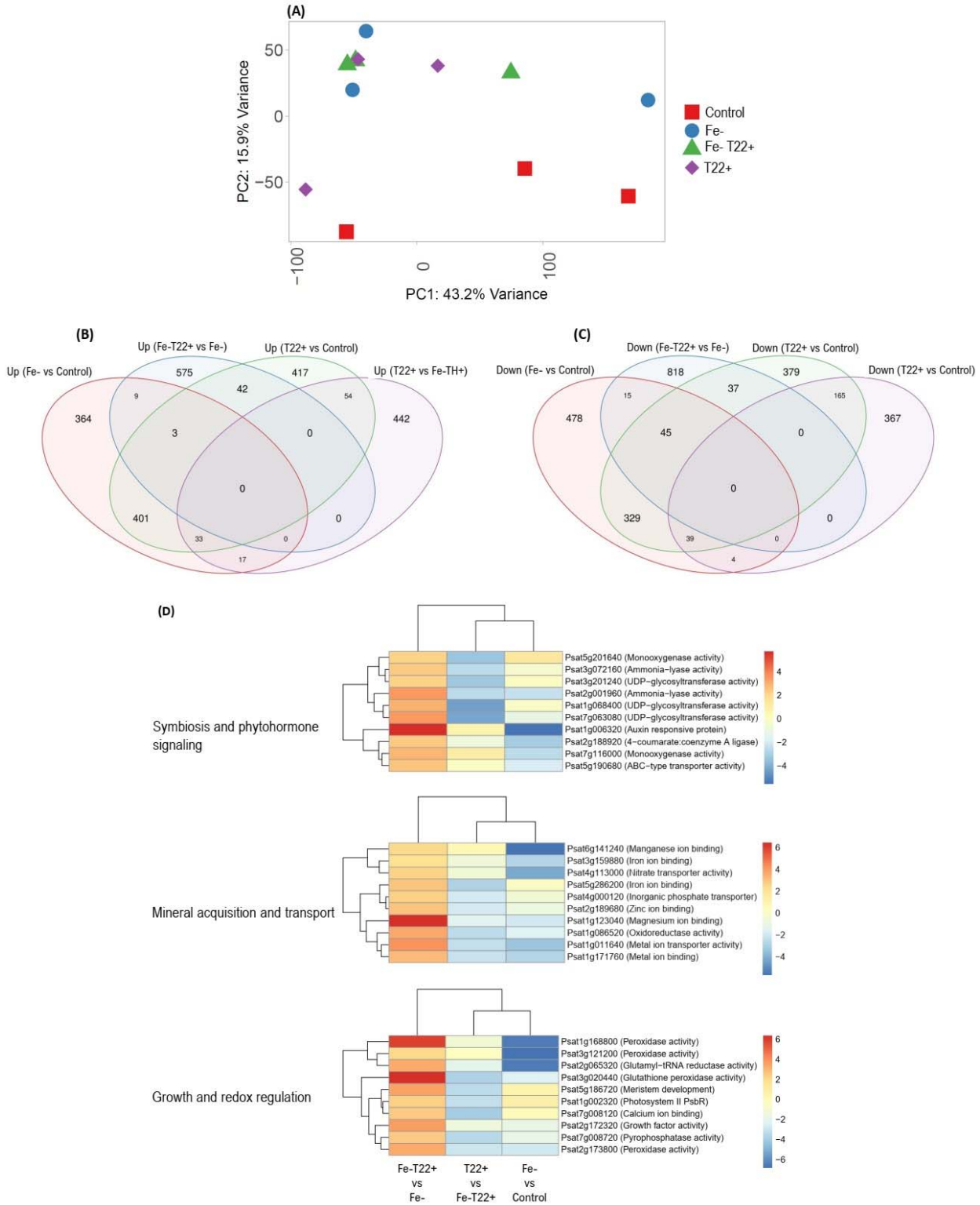

Fig. 3. RNA-seq analysis: principal component analysis of gene variation (A), Venn diagram of differentially expressed (> Log2 fold, < P value 0.05) upregulated (B) and downregulated (C) genes and heatmap of top 10 upregulated genes (D) associated with different functions

(symbiosis and phytohormone signaling, mineral acquisition and transport and growth and redox regulations) in pea plants inoculated with T22.

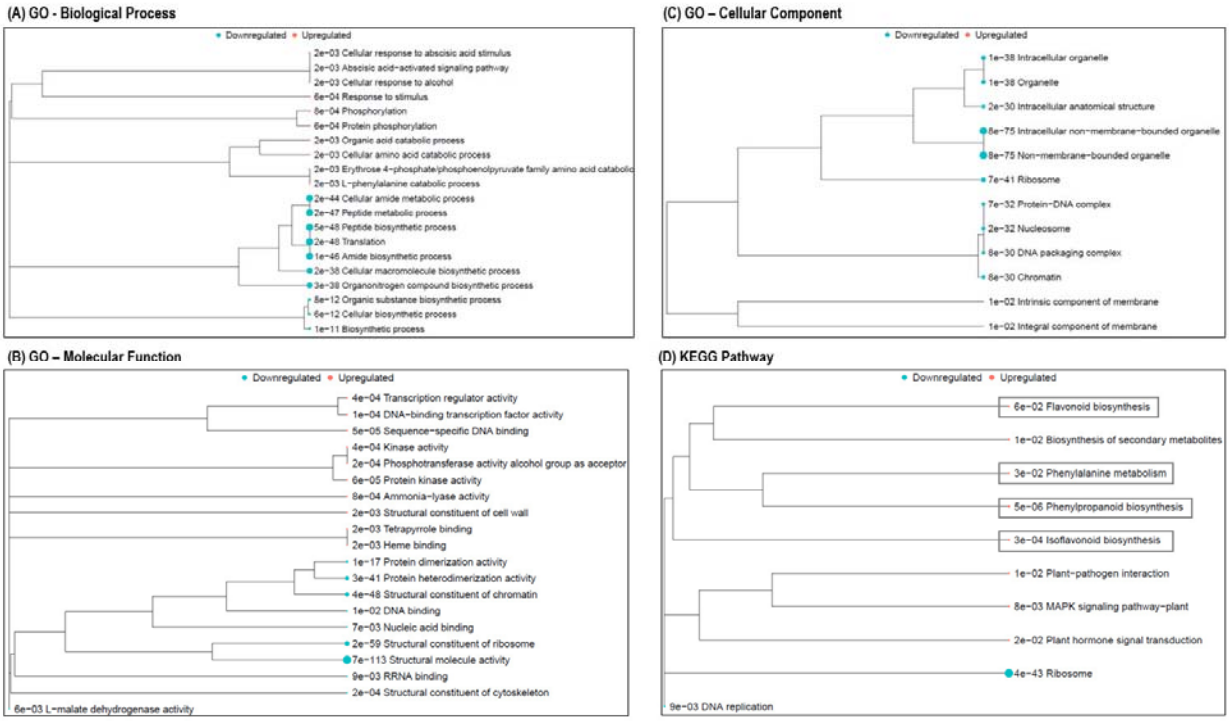

Fig. 4. Gene enrichment and functional analysis of differentially expressed genes (DEGs) in pea roots cultivated under Fe deficiency inoculated with T22: (A) biological process, (B) molecular function, (c) cellular component (d) KEGG pathway analysis. The analysis was performed using ShinyGO 0.76.3, with separate categories for upregulated and downregulated genes indicated by blue and red dots, respectively.

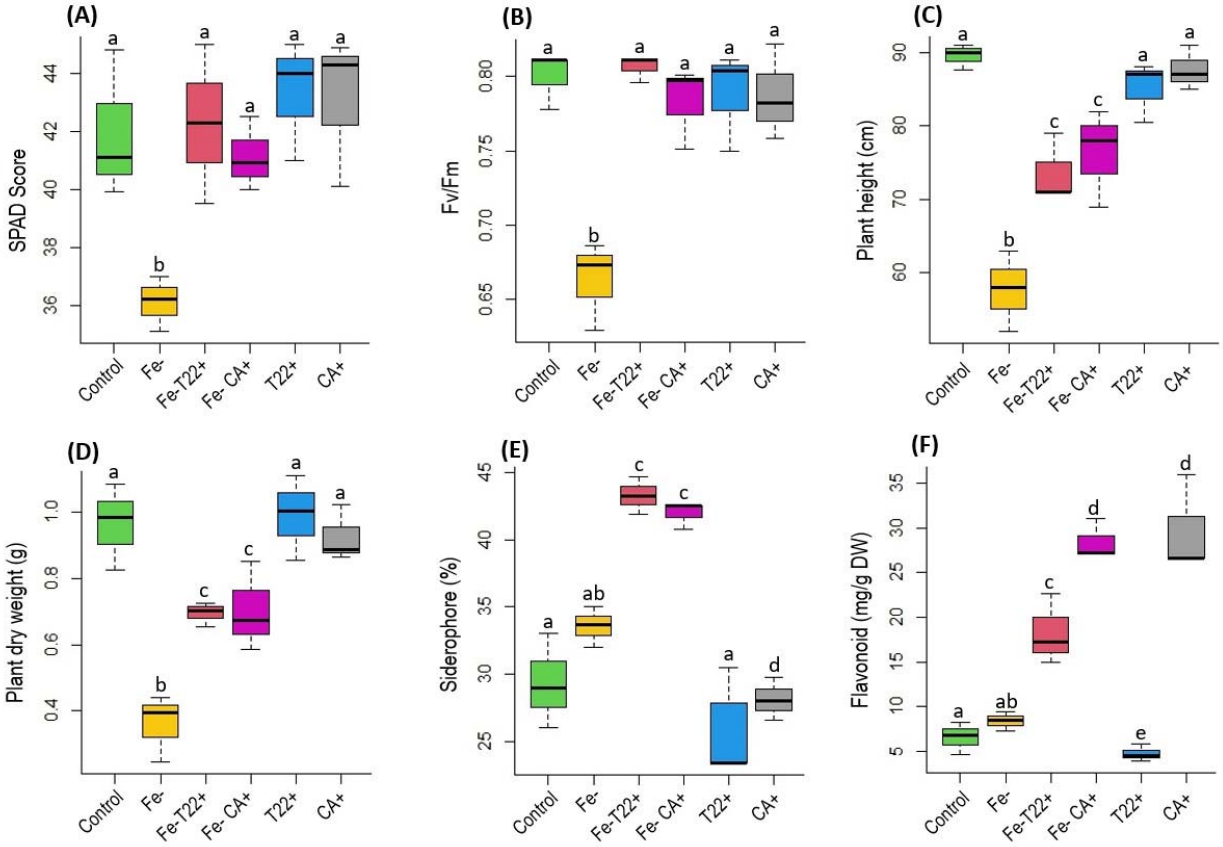

Fig. 5. Effect of cinnamic acid (CA) on leaf SPAD score (A), Fv/Fm: maximal quantum yield of PS II (B), plant height (C), plant dry weight (D), rhizosphere siderophore (E) and root flavonoid content (F) in pea cultivated with or without the absence (control) or presence of Fe deficiency (Fe-) and T22. The data presented are means with standard deviations (n = 3 individual replicates). Statistically significant differences in the ANOVA test among different treatments ($p < 0.05$) are indicated by different letters above the bars.

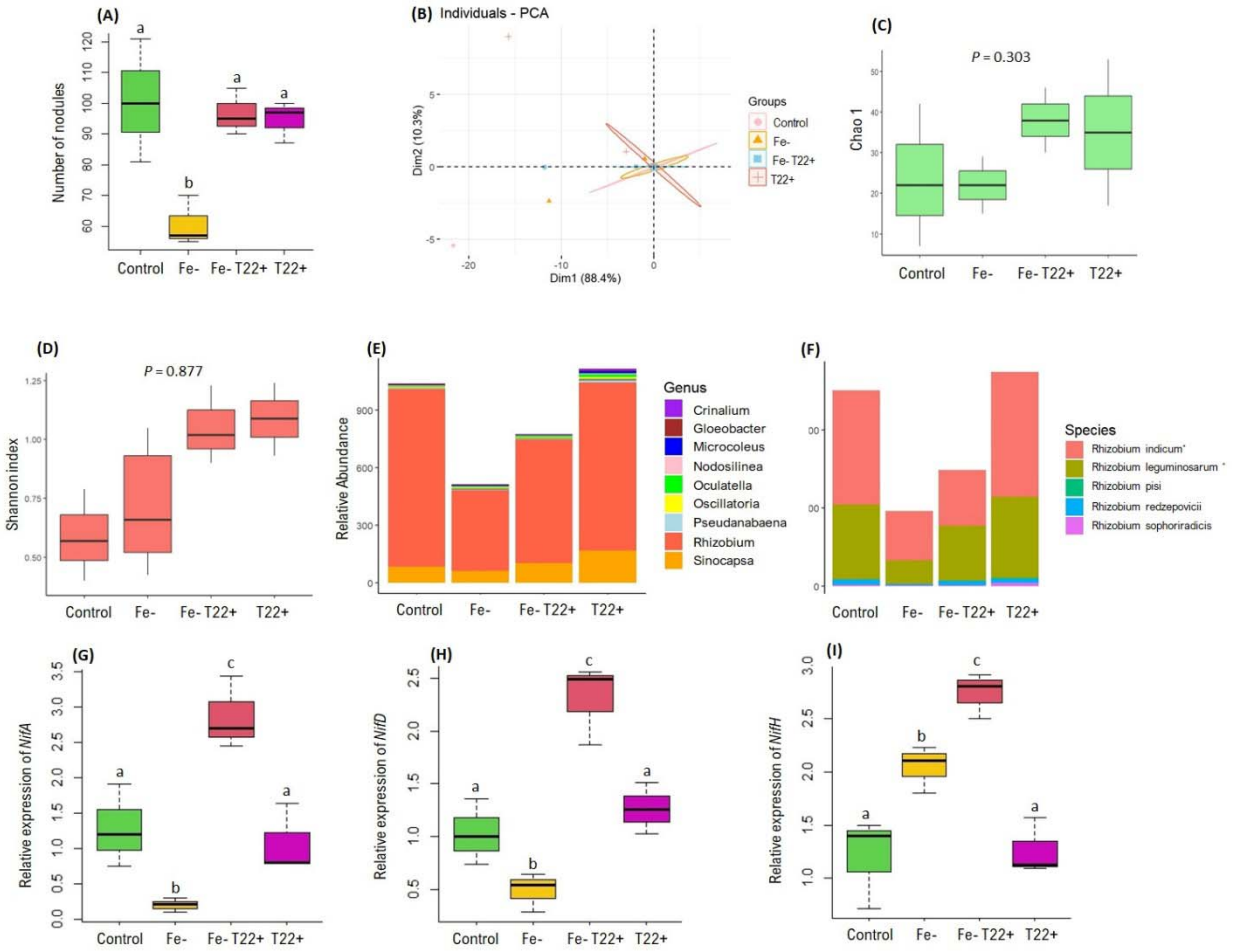

Fig. 6. Changes in nodule number (A), principal component analysis (B), Chao 1 (C), Shannon index (D), relative abundance of bacterial genera (E), relative abundance of *Rhizobium* species (F) as well as the expression of *NifA* (G), *NifD* (H) and *NifH* (I) in nodules of pea plants inoculated with or without T22 cultivated in the absence or presence of Fe deficiency. The data presented are means with standard deviations (n = 3 individual replicates). Statistically significant differences in the ANOVA test among different treatments ($p < 0.05$) are indicated by different letters above the bars, where applicable.

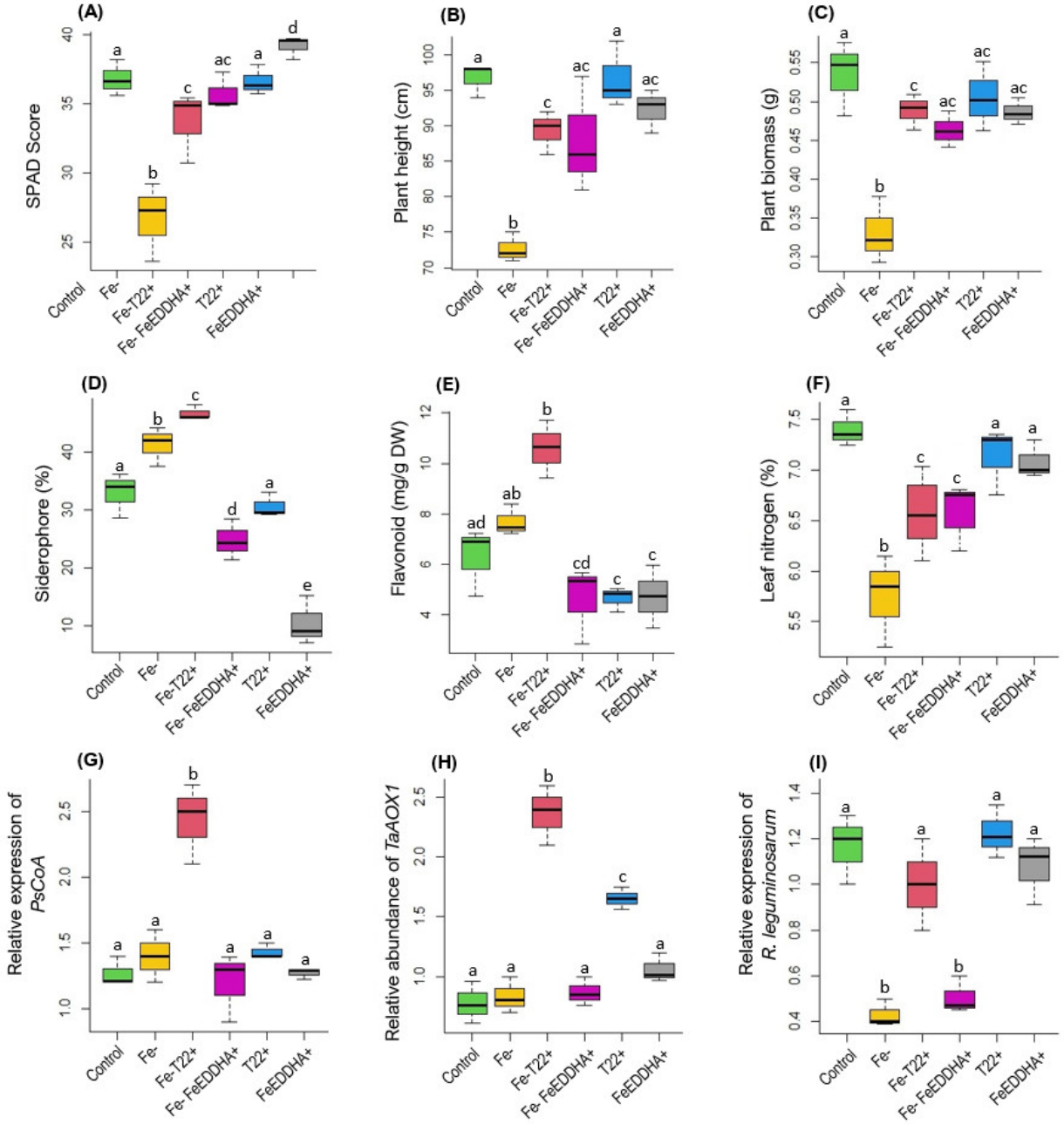

Fig. 7. Effect of FeEDDHA on leaf SPAD score (A), plant height (B), plant biomass (C), rhizosphere siderophore (D), root flavonoid content (E) and leaf nitrogen (F), relative expression of *PsCoA* in roots (G), relative abundance of *TaAOX1* (H) and *R. leguminosarum* (I) in nodules of pea plants cultivated with or without the absence (control) or presence of Fe deficiency (Fe-) and T22. The data presented are means with standard deviations (n = 3 individual replicates). Statistically significant differences in the ANOVA test among different treatments ($p < 0.05$) are indicated by different letters above the bars.

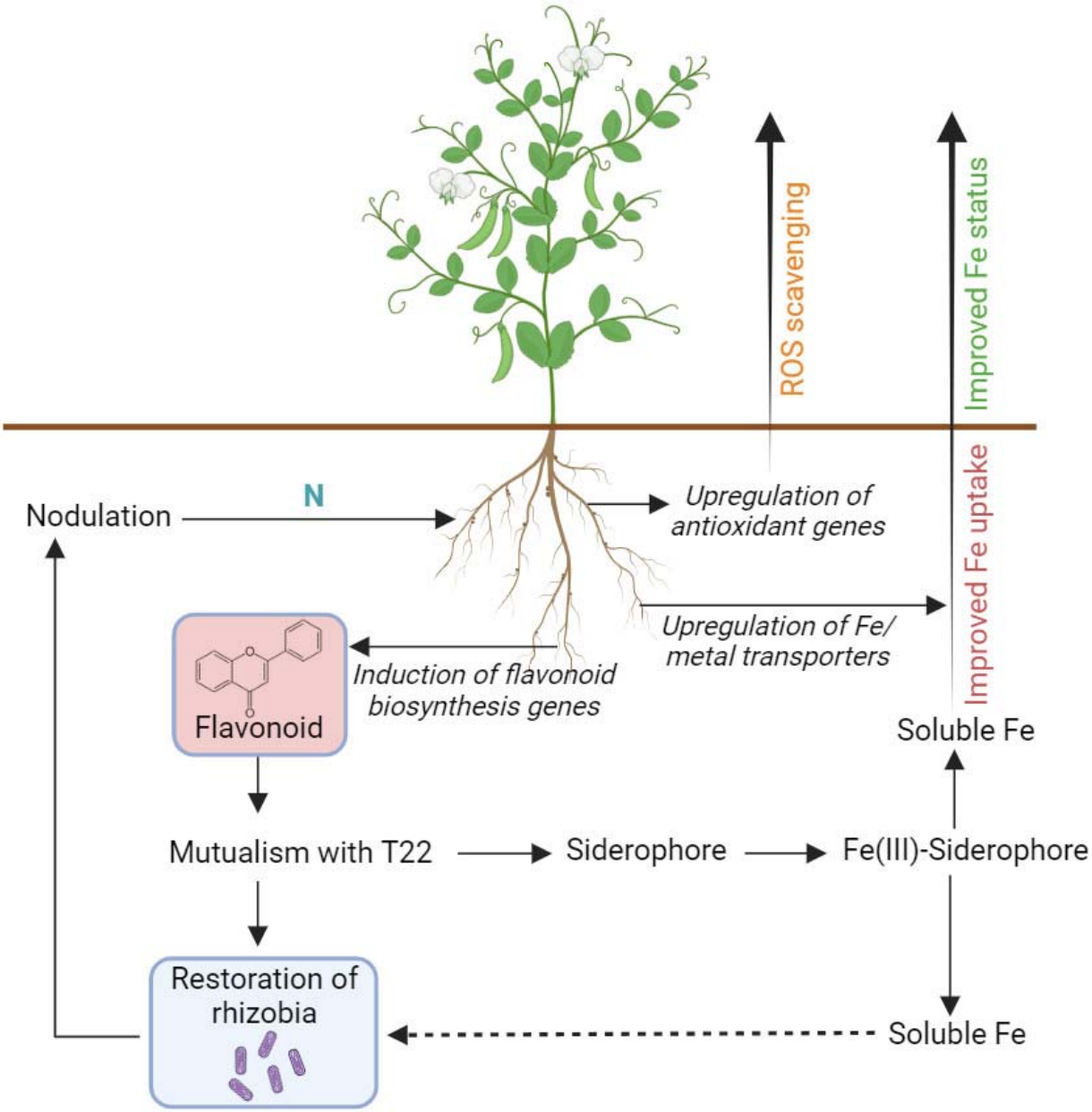

Fig. 8. Model of interactions by which T22 influences the symbiotic processes to help pea plants cope with Fe deficiency. Flavonoids, induced by specific biosynthesis genes in host plants, play a crucial role in mutualistic interactions with T22. This mutualism promotes the production of siderophores, which chelate Fe(III) and convert it into a soluble form that can be taken up by the induction of plant Fe/metal transporters. Additionally, the presence of T22 helps restore rhizobia populations through direct co-interaction or a Fe-mediated indirect pathway, facilitating N assimilation. T22 induces antioxidant genes in plants to regulate ROS status. Improved Fe uptake., combined with mutualistic interactions, helps the pea plant cope with Fe deficiency.