Leaf spectroscopy reveals drought response variation in *Fagus sylvatica* saplings from across the species' range

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Abstract

The common European beech (*F. sylvatica*) is sensitive to prolonged droughts, and its natural distribution is expected to shift with climate change. To persist in novel environments, young trees rely on the capacity to express diverse response phenotypes. Several methods exist to study drought effects on trees and their diverse adaptive mechanisms, but these are usually destructive, and challenging to scale to the large sample numbers needed to investigate biological variation.

We conducted a common garden experiment outdoors, but under controlled watering conditions, with 180 potted two-year-old saplings from 16 beech provenances across the species' range, representing three distinct genetic clusters. Drought stress was simulated by interrupting irrigation. We measured leaf reflectance of visible to short-wave infrared electromagnetic radiation. We determined drought-induced changes in biochemical and structural traits derived from spectral indices and a model of leaf optical properties and assessed intraspecific response diversity.

We quantified changes in pigmentation, water balance, nitrogen, lignin, epicuticular wax, and leaf mass per area in drought-treated saplings from different genetic clusters, revealing differences in likely adaptive responses to drought. *F. sylvatica* saplings from the Iberian Peninsula showed signatures of greater drought resistance, i.e., the least trait change in response to the drought treatment. Stomatal conductance and soil moisture were used to assess drought severity.

We demonstrate that high-resolution, broad-range leaf spectroscopy is an effective and non-destructive tool to assess individual drought responses that can characterize functional intraspecific variation among young beech trees.

Keywords: leaf optical properties; common beech; European beech; intraspecific variation; common garden experiment; genetic clusters; pigment composition; drought resistance

1. INTRODUCTION

As a result of climate change, Europe faces prolonged drought periods that will threaten forest ecosystems (Cook et al., 2016; IPCC, 2023). The extended duration of local precipitation deficits will increase the risk of water stress and mortality for trees (Allen et al., 2010), and the series of recent record-breaking summers in Northern Europe in 2003, 2018, and 2022 indicate what Europe's forest ecosystems may experience in the near future (Buras et al., 2020; Hermann et al., 2023; Millán, 2014; Sturm et al., 2022). The common European beech (*Fagus sylvatica*) is a dominant and widespread tree species in Europe (Durrant et al., 2016), but its dominance comes at the cost of considerable drought sensitivity compared to co-occurring broadleaf tree species (Leuschner, 2020; Packham et al., 2012). Consequently, beech faces a disproportionate risk of habitat loss and growth decline, and may ultimately change its natural distribution due to reduced competitiveness (Aranda et al., 2015; Dittmar et al., 2003; Geßler et al., 2006; Kreyling et al., 2014; Leuschner, 2020; Martinez del Castillo et al., 2022; Rigling et al., 2019; Rose et al., 2009; Schmied et al., 2023).

To establish and persist, young trees rely on phenotypic variation, i.e., the potential to express diverse phenotypes given their genetic background and environmental conditions (Arnold et al., 2019; Benito Garzón et al., 2019; Whitman & Ananthakrishnan, 2009). Intra- and interspecific diversity are expected to increase the probability of survival for young trees in communities experiencing novel environmental conditions, such as more intense or prolonged drought (Blondeel et al., 2024; Gazol et al., 2023). Intraspecific variation encompasses a range of physiological and morphological traits providing options for individuals to acclimate, and on which selection can operate, so species can adapt. Different individuals within a species may vary in their drought response, making some more likely to survive (Nguyen et al., 2017). Because drought responses are complex, involving orchestrated changes in physiology, differences among trees in individual traits may not predict differences in drought responses, and multiple strategies are possible (Martínez-Vilalta et al., 2023). Thus, greater within-species variation can serve as a buffer against environmental stress, ensuring that at least some members of a species or population can withstand prolonged periods of drought. The spatial distribution of this variation within versus between populations may determine which, and how many, populations can acclimate and adapt: those differences that are inherited across generations provide material for adaptation.

Acclimation of trees to drought is apparent through several physiological changes in leaf traits (Leuzinger et al., 2005; Pflug et al., 2018; Rukh et al., 2023; Tognetti et al., 1995). As an immedi- ate response, for example, plants can close their stomata to prevent hydraulic failure (McDowell et al., 2008). An anisohydric species (Leuschner, 2020), beech shows high stomatal sensitivity to water deficit and adapts stomatal conductance to optimize photosynthesis in water-scarce situations (Peuke et al., 2002). Photosynthetic pigments such as chlorophyll *a* and b (C_{ab}) are essential for oxygenic photosynthesis in plants, and water deficit and accompanying oxidative stress can reduce chlorophyll content, corresponding to decreased photosynthetic capacity (Gai et al., 2023; Hu et al., 2023; Mafakheri et al., 2010; Wu et al., 2016). Carotenoids (C_{car}) – xanthophyll cycle pigments and carotenes – serve as important photoprotective pigments by absorbing excess solar energy in the blue to green wavelengths, and protect cells from oxidative stress by reacting with singlet oxygen and free radicals, although regeneration e.g. via ascorbic acid is necessary to prevent damage due to oxidized carotenoid products (Baccari et al., 2020; Bacelar et al., 2007; Ben Abdallah et al., 2017; Edge & Truscott, 2018; Havaux, 2014; Sun et al., 2022). Thus, variation in the chlorophyll to

carotenoid ratio C_{ab}/C_{car} may be a good indicator for drought stress in plants (Green & Durnford, 1996). Anthocyanins (C_{ant}), which are generally responsible for the red coloration of senescent leaves, also act as antioxidants and absorb light in the UV, blue and green ranges (Gould, 2004; Saha et al., 2020). Drought stress increases oxidative stress, which can lead to anthocyanin accumulation (Li & Ahammed, 2023). Long-term responses include photoprotective mechanisms, such as an increased ratio of xanthophyll cycle pigments (VAZ) to chlorophylls. Drought-stressed plants also show reversible downregulation of photosystem II (PSII), which results in a decrease of quantum yield and photosynthetic efficiency, but also reduces the generation of free radicals from photosynthesis (Galleé & Feller, 2007; García-Plazaola & Becerril, 2000). Equivalent water thickness (EWT), which is a direct physical representation of water within the leaves, is associated with leaf-level drought tolerance (Féret et al., 2019; Junttila et al., 2022). Leaf water potential (Ψ) includes all energy gradients that move water such as turgor, osmotic and matrix potential and indicates increased stress levels and downregulation of physiological processes with progressing drought (Walthert et al., 2021). The biosynthesis of lignin, which is found in the cell walls of plants and is responsible for the overall stability of plant cells, is upregulated under water deficit (Brinkmann et al., 2003; Hu et al., 2009; Liu et al., 2018). Leaf epicuticular waxes restrict diffusion across the cuticle and reduce oxidative stress by absorbing UV light, and their composition may change in response to environmental factors (Jenks & Ashworth, 1998; Speckert et al., 2023). Constitutive differences in leaf traits can also be related to drought responses. Leaf mass per area (LMA) is usually calculated with leaf dry mass and is determined by many bulk constituents of leaves including starch, sugars, and proteins. LMA is often related to abiotic stress responses such as drought (De La Riva et al., 2016; Quero et al., 2006; Wellstein et al., 2017; Wright et al., 2004), but a sudden withholding of water does not cause LMA variation (Poorter et al., 2009).

Structural and biochemical profiling commonly require destructive sampling and laboratory analysis (Jacquemoud et al., 1996; Pflug et al., 2018). These are usually precise, but difficult to scale due to time and costs, and the requirement to work with harvested leaf tissue can introduce artefacts (Burnett et al., 2021b). High-resolution leaf spectroscopy, a contact measurement closely related to field and imaging spectroscopy, offers a relatively cost-effective, non-destructive, and scalable solution for retrieving leaf traits directly from living leaves on plants (Féret et al., 2019; Jacquemoud & Baret, 1990). Spectroscopy of individual leaves is commonly conducted using a contact probe with a standardized light source and backgrounds. It measures aspects of leaf optical properties (LOP), which describe how leaves physically interact with electromagnetic radiation. Since the biochemical and structural composition of leaves determine how they reflect, transmit, absorb, or fluoresce light, variation in LOP thus carries information about leaf traits (Jacquemoud & Ustin, 2019). Recent studies highlighted that spectral indices do may not entirely reflect leaf structural and biochemical properties (Petibon et al., 2023), however, a growing body of literature links spectra to plant trait variation, e.g. Asner and Martin (2011), D'Odorico et al. (2023), Meireles et al. (2020), and Wang et al. (2022).

Leaf traits are estimated from spectra based on empirical or physical approaches (Féret et al., 2017). Traits are derived empirically using machine learning methods such as partial least squares regression (PLSR), which require training data and yield somewhat, although not exclusively, dataset-specific models (Burnett et al., 2021a; Féret et al., 2019; Ji et al., 2024). A simple physical-based method, spectral indices are validated using laboratory measurements but do not require training data; these comprise reflectance values at different bands and are developed from two or more wavelengths in which the target of interest causes

a relative absorption difference (Jacquemoud & Ustin, 2019; Verrelst et al., 2015). Indices have been found useful to estimate the relative values of traits such as photosynthetic activity (Gamon et al., 2016), and are computationally very efficient (Jacquemoud & Ustin, 2019). Another common physical approach, radiative transfer models (RTMs) such as the extensively used open-source model propriétés spectrales (PROSPECT) (Jacquemoud & Baret, 1990), rely on the physical properties of electromagnetic radiation and its interaction with matter such as absorption and scattering coefficients, and are thus theoretically robust (Féret et al., 2008). In PROSPECT, broadleaves are represented as a generalized plate model with uniform layers (specifically air – cell wall interfaces) quantified as a structure parameter *N*, characterizing the leaf mesophyll structure (Allen et al., 1969; Jacquemoud & Baret, 1990; Spafford et al., 2021; Stokes, 1860), with which directional-hemispherical reflectance and transmittance between 400 nm and 2500 nm are simulated (Schaepman-Strub et al., 2006). One of the most recent PROSPECT developments is called PROSPECT-D(-ynamic). It allows C_{ab}, C_{car}, C_{ant}, N, EWT, and LMA as leaf trait parameter inputs to simulate leaf spectral profiles, and outperforms previous versions (Féret et al., 2008; Féret et al., 2017). The inversion of PROSPECT-D enables the retrieval of leaf traits based on provided reflectance, or transmittance data. The inversion makes use of optimization algorithms with a corresponding merit function (Jacquemoud et al., 1996; Tarantola, 2005). PROSPECT RTMs have been successfully applied across remote sensing and plant ecology research (Asner et al., 2014; Kothari & Schweiger, 2022; Serbin et al., 2014; Sun et al., 2018; Zhao et al., 2014).

To study intraspecific variation in the drought responses of young *F. sylvatica* trees, we conducted a common garden experiment on 180 two-year-old saplings originating from 16 different provenances across the species' range. We determined their population genetic structure, identifying three main genetic clusters. We simulated a drought scenario by restricting water for 14 days in early summer. We measured leaf reflectance with a field spectroradiometer at the end of the drought period to quantify spectral variation between the control (continued watering) and treatment groups, and to derive leaf traits using both spectral indices and an RTM approach (PROSPECT-D inversion). We also measured soil moisture, leaf transpiration, and leaf water content as indicators of drought effects, as well as leaf mass per area and pigmentation, for comparison to spectroscopy measurements. We hypothesized that (i) drought-treated saplings show different responses in spectral indices and simulated leaf traits compared to the control group that can be interpreted in physiological terms, and (ii) there is intraspecific variation in drought responses dependent on the sapling's genetic background.

2. MATERIALS AND METHODS

2.1 Seed collection, germination, and growth conditions

Seeds were collected in 2020 between August and October from well-protected sites across Europe, thus having a relatively high probability of harbouring populations representing natural migration more than introductions for forestry (see supplementary methods). European beech was the dominant tree species at all sites. The majority of seed collections were sampled directly from branches of trees located in an area of 10 x 200 meters. However, we also conducted bulk collections from the ground not more than 1 km from a core area of trees used for leaf spectroscopy measurements in Czyż et al. (2023). Seeds were placed in labelled containers per collection, and upon arrival, dried in mesh envelopes (1/collection) in closed plastic boxes under a gentle nitrogen flow. Once dried, the seeds were transferred to paper

envelopes and preserved in sealed plastic bags at 5° C until February 2021 (for more details, see supplementary methods).

Seeds of 94 collections total from 24 sites (see Table S1) were brought to a nursery on February 25nd, 2020 (47°35'48.9"N, 9°11'40.5"E, Kressibucher AG, Berg Thurgau, Switzerland), where they were imbibed in water for 24 hours, and then sown in 5 x 5 x 20 cm pots in flats (max 4 seeds/pot) with mixed humus and wood chips and no additional fertilizers. A wire grid was installed over the pots and the flats were put outside and exposed to the environment. Damage to the dicotyledons was mitigated by bringing the flats inside a closed shelter when temperatures fell below freezing. Seedlings germinated by July 2021, with seedlings emerging for 43 of the 94 collections, from 16 of the 24 sites, and an overall seedling emergence rate of 3.5% (7.9% for the 43 collections in which seedlings emerged). A summary of the resulting provenances is provided in Table 1.

Table 1. Summary of *F. sylvatica* provenances including sample sizes n, elevation, coordinates, date of collection, mean temperature (T_{mean}) and mean annual precipitation (MAP). Both MAP and T_{mean} were downloaded from WorldClim (Hijmans et al., 2005). Seeds started to germinate in the last week of February 2021.

Label	Country	Locality	*n*	Lat./Lon.	Elevation (m.a.s.l.)	T_{mean} (°C)	MAP (mm)	Collection date
BES	Belgium	Sonian Forest	11	50.751/4.423	168	10.16	805	02.09.2020
CHL	Switzerland	Laegern	8	47.479/8.364	733	9.21	1137	07.08.2020
ESM	Spain	Moncayo Natural Park	11	41.791/-1.810	1479	9.88	556	23.09.2020
ESP	Spain	San Juan de la Pena	31	42.508/-0.676	1200	10.88	720	24.09.2020
FRB	France	Sainte Baume	24	43.328/5.769	950	13.56	715	19.09.2020
FRM	France	Massane	6	42.491/3.032	722	14.20	604	25.08.2020
HBV	Bosnia	Vlasenica	9	44.168/18.919	1050	10.60	1216	2020
HRP	Croatia	Paklenica National Park	18	44.360/15.467	924	7.82	1267	16.09.2020
ITB	Italy	Bracciano Nature Reserve	2	42.174/12.156	584	13.70	234	21.08.2020
ITE	Italy	Etna	2	37.709/15.045	1721	11.41	699	17.08.2020
ITM	Italy	Madonie Natural Park	2	14.042/37.858	1711	13.46	567	16.08.2020
PLB	Poland	Bieszczadski Natural Park	2	49.232/22.472	893	6.01	933	11.09.2020
PLW	Poland	Bukowa Mountain	19	49.669/18.860	662	7.19	925	09.09.2020
ROS	Romania	Şinca	8	45.654/25.164	865	9.80	1182	13.09.2020
RSF	Serbia	Fruška Gora National Park	9	45.138/19.636	474	11.09	645	17.09.2020
SLK	Slovenia	Krokar forest	18	45.543/14.763	1178	7.71	1464	17.09.2020
Total			180					

In November 2021, saplings were transported to the Hauenstein garden center in Rafz, Zurich, Switzerland (47°36'36"N, 8°32'35"E). The plants were transplanted into tall 4 L pots filled with an equally proportioned mixture of peat-free potting soil and low-organic matter sandy soil. The potting soil was enriched with Osmocote Exact Standard NPK (Mg) 12-14M (ICL Group, Tel Aviv-Jaffa, Israel) with micro-nutrients as fertilizer, containing 15% total nitrogen (N), 8% phosphorus pentoxide (P_2O_5), 11% potassium oxide (K_2O) and 2% total magnesium oxide (MgO). During the winter, the saplings were in a winter tunnel, where temperatures were maintained at circa 7° C. The saplings were watered by an overhead irrigation system as needed. Furthermore, Multikraft BB Start (Multikraft, Pichl/Wels, Austria) was used to treat the soil with ectomycorrhizal fungi. Once it was determined that the saplings were no longer at risk of frost damage, the pots were moved outside the winter tunnel (12 May 2022). We controlled aphid infestation by repeatedly spraying NeemAzal-T/S (3ml/L in water, Andermatt Biogarten, Grossdietwil, Switzerland) directly onto the leaves.

2.2 Common garden experiment

We conducted our experiment with 180 2-year-old saplings from 16 provenances across the natural distribution range in June 2023. The saplings were transported from the Hauenstein nursery to the experiment site at the University of Zurich, Zurich, Switzerland (47°23'44"N, 8°33'05"E; elevation 509 m.a.s.l.). We implemented a 2 x 5 randomized block arrangement, in which each block formed a 3 x 6 grid (Fig. S1). Each block contained 18 saplings, which were placed in the slots of a steel grid, which were spaced approximately 1 m apart to minimize interactions between saplings of different blocks. Pots were placed on black polypropylene fabric for weed control, and secured to the steel grid with hemp cord (Fig. 1b). We installed an all-in-one weather station (ATMOS 41, METER Group, Pullman, WA United States) to measure local weather conditions such as precipitation, humidity, and air temperature.

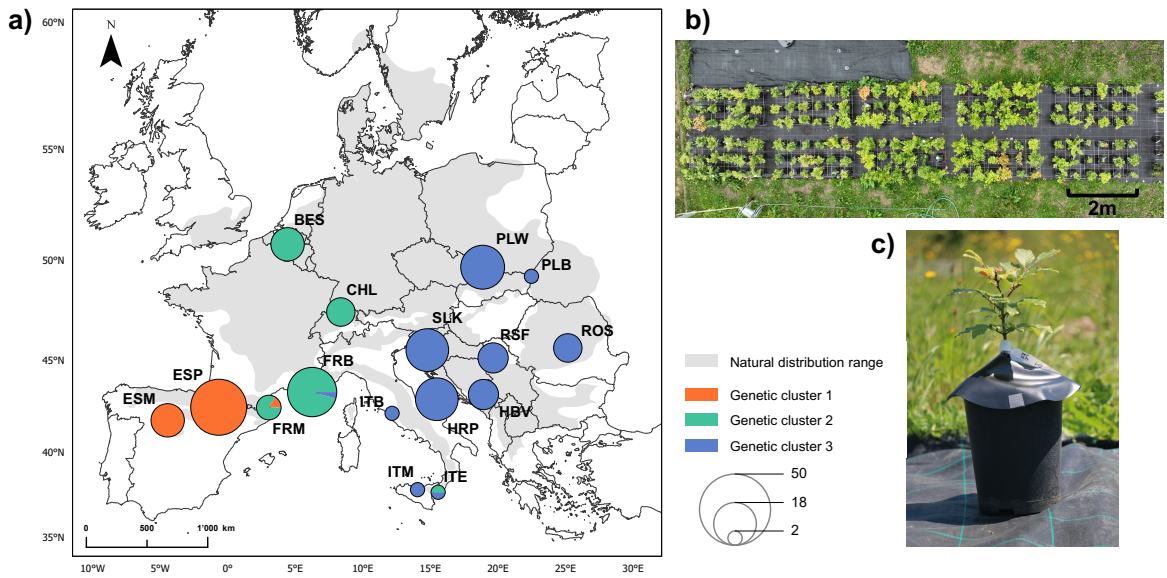

Figure 1. a) European map depicting sampling locations and natural distribution range of *F. sylvatica* (EUFORGEN) (Caudullo et al., 2017). The first two letters in the abbreviations are country identifiers, and the third letter is the site identifier. The provenances are shown as pie charts with the proportions of the samples belonging to the three genetic clusters. The map was created in ArcGIS Pro (v3.1) (ESRI Inc., 2023). b) UAV footage of the experiment site in June 2023. c) Rain cover over the pots used to simulate the drought scenario.

We maximized the diversity of provenances within the blocks, so each provenance was represented in every block whenever possible (Fig. S1a). For each provenance, half of the saplings were assigned to the drought treatment and half to the control (for details, see Fig. S1a). Seed collection was not considered, due to both the limited and unequal number of seed collections per provenance and the limited and unequal number of individuals per collection (see Table S2). The placement of the saplings within the blocks was randomized. In April 2023, we measured the length of the primary shoot using a flexible tape measure from the root collar along the shoot to the highest point in its natural position. We then used these plant height measurements to select individuals to have a similar distribution of heights among the blocks (Fig. S1b).

2.3 Drought treatment

We selected a duration of 14 days as a target to induce realistic drought stress while avoiding tree mortality in the experiment: Switzerland encounters an average of 11 consecutive rain-free days each year, and it is anticipated that these dry periods may extend to as many as 19 days in the midterm future (NCCS, 2018).

During the simulated drought period, both groups were subjected to a controlled exclusion from rain using cone-shaped rain covers which enclosed the entire pot (Fig. 1c), but the control group was watered regularly according to demand and at least every other day. Covers were made of waterproof and UV-resistant PVC black pond foil (Heissner GmbH, Lauterbach, Hessen, Germany) and fixed on the sapling stem with Parafilm (Pechiney Plastic Packaging Inc., Chicago, IL, United States) beneath the first stem node. Three thin wooden sticks were used to elevate the cover to leave a gap between the foil and the pot to enable air circulation. In addition, we inverted the pot trays in the treatment group to avoid the retention of water in the soil.

We prioritized sapling survival by ad hoc irrigation of the drought-treated saplings: when severe wilting was observed, we watered wilting saplings with 600 mL water (see Table S3 for irrigation timeline and details). Wilting was monitored daily and quantified by a wilting index. The wilting index, ranging from 0 to 4, was categorized into quartiles. A score of 0 denoted the absence of wilting, while scores 1-4 indicated wilting levels corresponding to ≤ 25%, > 25% to ≤ 50%, > 50% to ≤ 75%, and > 75% of leaves per saplings, respectively. Our aim was to avoid score 4, as such severe wilting would likely lead to mortality.

To monitor soil moisture and air temperature, we installed 20 TMS-4 probes (TOMST, Prag, Czech Republic) (Wild et al., 2019) in pots from block A to F. The probes covered as many provenances as possible while capturing the full range of plant heights within a single block. Soil moisture and soil temperature were sampled in an interval of 15 minutes during the entire experiment (Fig. S2).

2.4 Leaf spectroscopy

Leaf spectroscopy measurements were taken during the second half of the drought (days 9 to 14, corresponding to June 19th to 24th, 2023). We constrained the measurement time to ±3 hours from solar noon. Therefore, within one day, we measured two blocks (one control, one treatment block) totalling 36 individuals and 72 leaves (2/individual), except for one day when 72 individuals corresponding to 144 leaves were measured. We used the ASD FieldSpec 4 Standard-Res spectrometer (ASD Inc., Boulder, USA), which measures electromagnetic

radiation across a total of 2151 bands covering the spectral range from 350 nm to 2500 nm. We used a plant probe with a leaf clip (model A122317, serial No 455, ASD Inc., Boulder, USA) with a calibrated low-intensity halogen light source. The leaf clip allows the non-destructive handling of the leaves and the isolation of external illumination. We used the spectral acquisition software RS3 (ASD Inc., Boulder, CO, USA) with respective instrument configurations (Table S4). Before taking the measurements, the spectrometer was warmed up for at least 30 minutes to decrease measurement errors due to variation in ambient and internal temperatures (Hueni & Bialek, 2017). For each sapling, we measured two random sun-exposed top-of-canopy leaves. We placed the leaf clip on the leaf adaxial surface, filling the measurement window of the plant probe and avoiding the midrib. Spectral measurements consisted of four successive measurements for each leaf: the white reference (R_w), the white reference with target leaf (T_w), the black reference (R_b), and the black background reference with target leaf (T_b). We gathered 5 readings per measurement, totalling 20 measurements per leaf. We recalibrated (optimized) the sensor gain with the RS3 software against the white reference background of the leaf clip after measuring 10 saplings to correct for differences in temperature sensitivity of the three detectors resulting in deviations of the white reference reflectance value from one (Hueni & Bialek, 2017). The leaf clip and plant probe were regularly checked for dirt or other contamination and cleaned, or white and black reference stickers were replaced as needed.

2.5 Stomatal conductance

During the drought, we measured stomatal conductance (g_s) of a subset of the saplings (n=141) with an SC-1 leaf porometer (METER Group, Pullman, WA United States) according to the manufacturer's protocol. Along with the spectral measurements, we constrained the measurement time to ±3 hours from solar noon due to high variation during the day (Leuschner, 2020). Given this constraint and the significant time required for each measurement, it was not feasible to measure all the beech saplings in a single day. Therefore, we measured two to three blocks each day. To standardize the strong and immediate effects of changing conditions on stomatal conductance during the 6-hour measurement window, we always measured a sapling from the control directly after measuring one from the treatment. With this "paired" approach, the solar radiation and air temperature were on average similar for treatment and control, which allows comparison between them. We calibrated the device according to the manufacturer's manual on each measurement day. When measuring, we took the values as soon as they stabilized. Per sapling, we measured two representative sun-exposed leaves on the adaxial side, where the stomata of beech are found (Peat & Fitter, 1994).

2.6 Validation of modelled pigments and structural traits

To establish a validation dataset for the modelled pigments and structural and biochemical traits (C_{ab}, C_{car}, EWT and LMA), we collected leaves from across the control and treatment group. For pigment extraction, immediately after their spectral measurement, the leaf samples (n=56) were stored in black plastic bags on dry ice to prevent degradation of the pigments and within an hour moved to – 80° C. To measure leaf area and fresh weight, we used a second leaf from the same sapling. To get the fresh weight, the leaf samples (n=56) were weighed immediately after their spectral measurement using a portable field scale. Leaf area was calculated using the image analysis program ImageJ (v1.38) (Schneider et al., 2012) based on a photo taken above parallel to the leaf with a size standard (see Fig. S3 for an example). Dry weight was assessed after 72 h at 70° C using a semi-micro balance (MSU125P, Sartorius, Göttingen, Germany, 0.015 mg repeatability). We then calculated EWT and LMA based on the

equations by Féret et al. (2019). (See also supplementary methods for more details).

For a subset of the collected samples (n=20), the pigment extraction and subsequent HPLC analysis was done according to the protocol proposed by Petibon and Wiesenberg (2022) and conducted under subdued light to prevent pigment degradation. We modified the gradient program and the eluent composition of the HPLC method to optimize peak separation (see supplementary methods). We identified and quantified seven major pigments (chlorophyll a and b, α− and β-carotene, lutein, neoxanthin, and zeaxanthin) based on the measurement of standard solutions containing the respective analyte (see supplementary methods for the standards' sources). From the results we calculated the parameters C_{ab} (sum of chlorophyll a and b) and C_{car} (sum of a/b-carotene, lutein, neoxanthin, zeaxanthin) in $mg.cm^2$ as measure for the chlorophyll and carotenoid concentration. We observed co-elution of α- and β-carotene, for which we used the α-carotene standard calibration. Additionally, we assigned all remaining peaks to one of the following compound classes: pheophytin, chlorophyll-derivatives, carotene-derivatives, or unknown compound, based on their absorption spectra and the decision tree proposed by Petibon and Wiesenberg (2022). For these peaks, no quantification was carried out due to the lack of standard material. Since the method was optimized to retrieve chlorophylls and carotenoids, we were unable to assess anthocyanin concentrations (Fig. S4a, b); however, measurements included one purchased copper beech (a variety with purple leaves due to a mutation that inhibits anthocyanin breakdown (Packham et al., 2012)) as an extreme phenotype for comparison, and anthocyanins were detected in this sample (Fig. S4c), but not in any other samples.

2.7 DNA extraction and whole-genome sequencing

We sampled leaf tissue (one to two mature leaves, i.e. roughly 0.1 g per plant) for genetic analysis by wrapping the leaves in aluminum foil and freezing them on dry ice in June 2022. The samples were stored at -80° C until further processing. Prior to grinding, samples were flash-frozen in liquid nitrogen. Grinding was performed using 3 mm glass beads at 30 Hz for three min in a TissueLyser II (Qiagen). Plant DNA was then extracted using the Norgen Plant and Fungi DNA extraction kit (Norgen Biotek, Thorold, Ontario, Canada), and following the manufacturer's protocol with some optimization steps added to minimize RNA contamination and increase lysis, yield, and purity (described in detail in (Czyż et al., 2023)). The quality and quantity of the purified DNA was assessed with Qubit dsDNA HS Assay Kits (Thermo Fisher Scientific), a DS-11+ microvolume spectrophotometer (DeNovix, supplied by Labgene Scientific SA, Châtel-Saint-Denis, Switzerland), and agarose gel electrophoresis. PCR-based library preparation and Illumina paired-end whole-genome sequencing (n=180 samples) were conducted by Novogene (UK). We obtained 11 GB of raw data per sample, resulting in a mean 20x coverage of the 541 Mbp nuclear reference genome (Mishra et al., 2022).

2.8 Data analyses

2.8.1 Spectral data

To process the spectral data, we used R (v4.3.1) (R Core Team, 2023) with the spectrolab package (v0.0.18) (Meireles & Schweiger, 2020). We spliced the bands at sensor transitions, and interpolated them with the match sensor() function. Due to the low signal:noise ratio and higher absolute uncertainty in the 350-400 nm region, we excluded that range (Petibon et al., 2021).

The measurement per sample consisted of five replicates each of a reference measurement

with both black (R_b) and white backgrounds (R_w) and target leaf with both black (T_b) and white backgrounds (T_w) (see section 2.4). To calculate the total reflectance of each leaf, we took the mean of the five scans for each measurement. The mean reflectance values were then used to calculate the leaf sample reflectance R_l based on the equation by Miller et al. (1987):

$$R_l = \frac{T_b \times R_w - T_w \times R_b}{R_w - R_b}$$

The value is the scaled percentage of reflected radiation, where 1 indicates 100% reflection and 0 means 0% reflection. We derived information about leaf traits by calculating indices with R_l. Fig. 2a shows the mean spectral reflectance across control and treatment group. Fig. 1b-d show individual reflectances depending on their genetic clusters. We calculated previously published spectral indices (Table 2) that are indicative for leaf biochemical traits and structural traits.

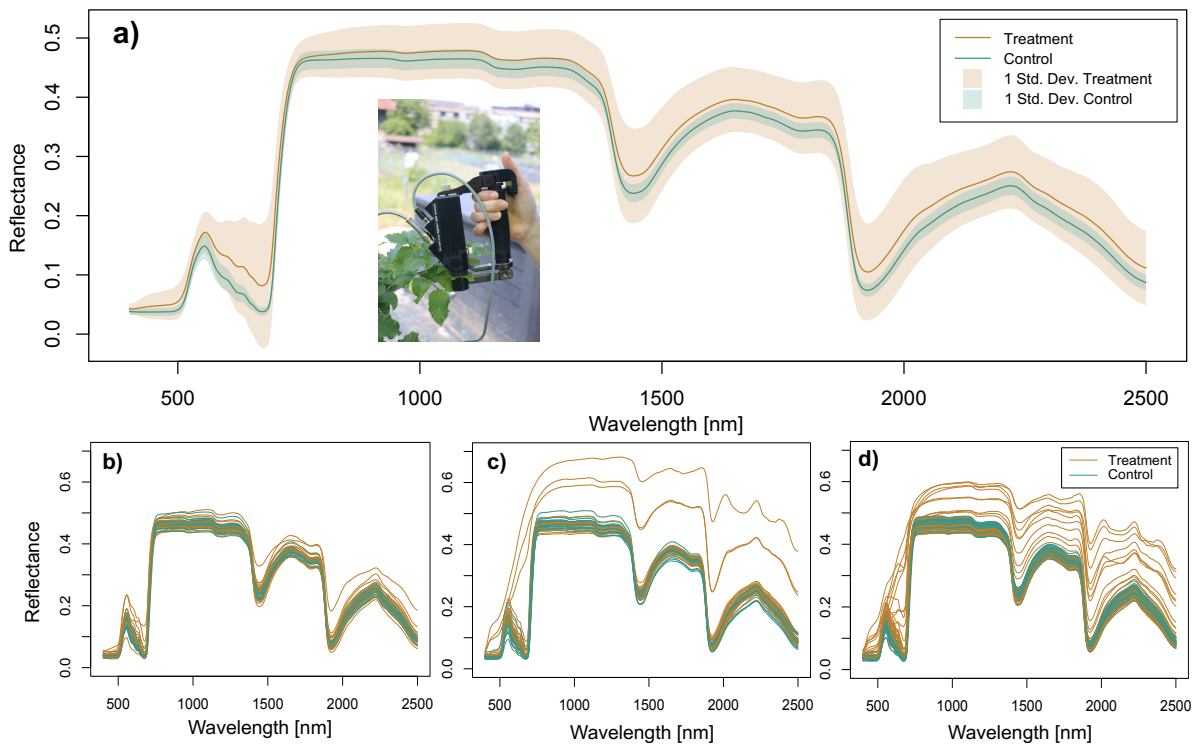

Figure 2. a) Mean reflectance in the spectral range from 400-2500 nm of the leaves from the treatment (orange lines, n=90) and control (green lines, n=90). Shown are calculated reflectance spectra following splice correction (see section 2.8.1). The inset shows the leaf clip used for the spectral measurements. b) Sample reflectances for saplings belonging to genetic cluster 1 (n=43). c) Sample reflectances for saplings from genetic cluster 2 (n=48). d) Sample reflectances for saplings belonging to genetic cluster 3 (n=89).

To run PROSPECT-D, we employed the prospect package (v1.3.0) by Féret et al. (2017) in the R statistical software (v4.3.1) (R Core Team, 2023). To use the inversion and thus to estimate the output parameters, we called the function Invert PROSPECT(), which applies the iterative optimization algorithm in the fmincon() function included in the pracma package (v2.4.2). We used the root-mean-squared error (RMSE) merit function, which minimizes the RMSE between the simulated and measured properties. We employed PROSPECT version D

over previous versions because of its improved trait parametrization over a larger database and the inclusion of relevant traits such as anthocyanin content (Jacquemoud & Ustin, 2019). We followed the recommendations of Spafford et al. (2021) to obtain prior information on the leaf structure parameter N when only R is measured by using the integrated function Get Nprior() because a priori N estimation is expected to improve estimations of leaf traits. We estimated the following leaf traits using PROSPECT-D: C_{ab}, C_{car}, C_{ant}, EWT, and LMA.

2.8.2 Statistical analyses

Since we measured two leaves per sapling, we took the arithmetic mean of the indices and the model outputs. The statistical analysis was conducted using the statistics software R (v4.3.1) (R Core Team, 2023). To ensure we met the model assumptions, we conducted a visual inspection of the histogram using hist(), residuals versus fits and QQ-plots using qqPlot() to identify outliers. We discovered that by excluding all saplings which exhibited strong wilting (wilting index ≥ 2) and outliers in the normalized difference water index (NDWI < 0), we could meet model assumptions, and these were therefore excluded from statistical analyses (n=10). As these were the most drought-affected saplings, this may have led to an underestimation of the magnitude of the drought effects. However, as the excluded individuals belonged to multiple genetic clusters ($n_{cluster2}$ = 3, $n_{cluster3}$ = 7) and sites (1 x BES, 1 x ROS, 3 x RSF, 1 x FRM, 2 x HBV, 1 x PLW, 1 x CHL), their exclusion did not alter the general findings. We used the multiple regression function lm() of the stats package (v3.6.2) to fit the model and subsequently conducted Analysis of Variance (ANOVA) with the function anova() on the model. The response variables were regressed over the predictor variables height, treatment, genetic cluster, and the interaction treatment x cluster. For the ANOVA, we assume randomness in the observations, homogeneous variances in each group and normality in the response variable. We considered $p \leq 0.05$ as our significance threshold.

Next, we calculated effect sizes for each spectral index and modelled trait. Log response ratios (LRR) quantify the log-proportional change of the mean of the treatment and the control group (Lajeunesse, 2016). A strength of this method is that it linearizes its metric, treating the deviations in the numerator and denominator in the same way. To calculate the LRR and to subsequently compute the variations of a leaf trait between the clusters, we adapted the equation from Hedges et al. (1999) and Bakbergenuly et al. (2020):

$$\widehat{\lambda_\iota} = \ln\left(\frac{\bar{X}_T}{\bar{X}_C}\right)$$

where $\widehat{\lambda_\iota}$ indicates the LRR and $\bar{X}$ denotes the observed means of each treatment (T) and control (C) group. Since the logarithm is only defined for positive values and some indices show a series of negative and positive values, we apply an offset transformation by adding a constant c, where c equals to 1 except for ARI, where c equals to 3; we inverted MSI for interpretability (see index definitions in Table 2).

The standard error of the LRR SE was computed as:

$$S_{E_{\widehat{\lambda_\iota}}} = \sqrt{\frac{s_T^2}{n_T X_T^2} + \frac{s_C^2}{n_C X_C^2}}$$

where s denotes standard deviations and n the sample sizes of the groups. We incorporated the outliers in the computation and visualization of the LRR.

Table 2. Summary of spectral indices including equation, author, and year of publication. R_λ indicates reflectance at wavelength λ.

Trait	Index	Short	Formula	Reference
Chlorophyll	Normalized Difference Simple Ratio	**NDSR**	$(R_{800} - R_{680}) / (R_{800} + R_{680})$	Sims et al., 2002
Chlorophyll	Red-Edge Chlorophyll Index	**CIre**	$(R_{783} / R_{704}) - 1$	Gitelson et al., 2005
Chl/Car	Chlorophyll/Carotenoid Index	**CCI**	$(R_{528} - R_{665}) / (R_{528} + R_{665})$	Gamon et al., 2016
Chl/Car	Photochemical Reflectance Index	**PRI**	$(R_{531} - R_{570}) / (R_{531} + R_{570})$	Sims et al., 2002
Anthocyanin	Red:Green Ratio	**RGR**	$\frac{Red}{Green} = \frac{\sum_{i=600}^{699} R_i}{\sum_{i=500}^{599} R_i}$	Gamon et al., 1999
Anthocyanin	Anthocyanin Reflectance Index	**ARI**	$1/R_{550} - 1/R_{700}$	Gitelson et al., 2001
Ψ	Water Balance Index	**WABI**	$(R_{1490} - R_{531}) / (R_{1490} + R_{531})$	Rapaport et al., 2015
Water	Moisture Stress Index	**MSI**	R_{1600} / R_{820}	Hunt Jr., 1989
Water	Normalized Difference Water Index	**NDWI**	$(R_{860} - R_{1240}) / (R_{860} + R_{1240})$	Gao, 1996
Nitrogen	Normalized Difference Nitrogen Index	**NDNI**	$\log(1/R_{1510}) - \log(1/R_{1680}) / \log(1/R_{1510}) - \log(1/R_{1680})$	Serrano et al., 2002
Lignin	Normalized Difference Lignin Index	**NDLI**	$\log(1/R_{1754}) - \log(1/R_{1680}) / \log(1/R_{1754}) - \log(1/R_{1680})$	Serrano et al., 2002
EW	Epicuticular Wax Index	**EWI**	$R_{625} * (1/R_{739} - 1/R_{832})$	Camarillo-Castillo et al., 2021

2.9 Sequencing data and kinship matrix

Reads containing adapter sequences and those with low-quality scores were removed from the raw data using Trimmomatic (v0.39) (Bolger et al., 2014). To avoid bias introduced by using a reference genome, we used a reference-free method. This allows capturing all genetic variation among individuals without losing variation that is not present in the reference genome. Trimmed sequencing reads were analysed using the kmerGWAS pipeline (v0.2) (Voichek & Weigel, 2020) according to the manual found on github (https://github.com/voichek/kmersGWAS). The k-mer database was built using KMC (v3) (Kokot et al., 2017) with the default k-mer size and settings (-k 31 –mac 5 -p 0.2), equivalent to the approach in (Schmid et al., 2024). The k-mer table containing the presence/absence pattern of each 31 bp k-mer in the population was then used to derive the kinship matrix. The kinship matrix (Fig. S5) was calculated using efficient mixed-model association (EMMA) (Kang et al., 2008) with a minor allele frequency (MAF) of 0.05. The resulting matrix showing genetic relatedness was visualized with a heatmap and dendrogram with the ASRgenomics (v1.1.3) package in R using hierarchical clustering and Euclidean distances (Gezan et al., 2022). We then used PLINK (v1.9) (Chang et al., 2015; Purcell et al., 2007) to convert the k-mer table to a bed format and used PLINK to generate a PCA using the .bed file with the following options in effect: double-id –allow-extra-chr –pca –allow-no-sex –out. Visual inspection of the matrix and the PCA indicated the presence of three separate genetic clusters. These corresponded well with a study investigating genetic relatedness based on SNPs among beech trees from many of the same locations as those in this study Czyż et al. (2023), although there is not a perfect overlap between the set of locations used by Czyż et al. (2023) and those of our study (see Table 1). However, the difference in findings is largely due to the used settings (–mac 5) and the MAF being set to 0.05 in this study. This resulted in any k-mers that are private to

populations consisting of fewer than five individuals (ITB, ITM, ITE and PLB) being excluded; however, reducing these threshold values also introduces noise and leads to more complex clustering that is difficult to interpret. Based on the kinship matrix and PCA (Fig. S5, Fig. S6), we assigned every sample (i.e., sapling) a genetic cluster and used these three clusters (annotated as cluster 1, cluster 2 and cluster 3) as predictor variables for further analyses. We note that two samples could not be properly processed using the k-mer pipeline, likely due to insufficient quality of their sequences. Consequently, we assigned them to their genetic clusters based on where their half-siblings were positioned in the kinship matrix. PLW 1 4 was assigned to cluster 3 because all half-sibs clustered together, and ESP 8 6 was assigned to cluster 1 because all its half-sibs were in that cluster. The genetic clusters are visualised geographically in Fig. 1a.

3. RESULTS

3.1 Soil moisture and stomatal conductance

The soil moisture sensors showed significantly lower moisture for the drought treatment (mean TDT value: 1562) compared to the control group (mean TDT value: 1906), indicating that rain exclusion had resulted in 18.0 (± 10.25) percent drier soils in the drought treatment group (Fig. S2a). Furthermore, we found an inverse relationship between soil moisture and tree height for both control and treatment groups, suggesting that taller saplings tended to have less soil moisture in their pots ($R^2 = 0.42$, $p = 0.059$ for the treatment group and $R^2 = 0.52$, $p < 0.05$ for the control group, Fig. S7a).

The drought treatment decreased stomatal conductance (g_s) (($F_{(1,134)} = 43.0879$, $p < 0.001$). As with soil moisture, stomatal conductance was also generally lower for taller saplings (Fig. S7b, ($F_{(1,134)} = 68.6692$, $p < 0.001$). Stomatal conductance did not differ between the genetic clusters (Table 3, $F_{(2,134)} = 1.5842$, $p = 0.21$) and the interaction between genetic cluster and drought treatment was also not significant (Table 3, $F_{(2,134)} = 0.2493$, $p = 0.78$).

3.2 Leaf spectral trait variation in response to drought

The leaf spectra showed on average increased reflectance for the drought-exposed saplings (Fig. 2a). This was more pronounced in genetic clusters 2 and 3 (Fig. 2c, d), and less so in genetic cluster 1 (Fig. 2b). The variation among the individuals also increased in the drought treatment, again, especially for genetic clusters 2 and 3.

We then used the measured leaf reflectance spectra to derive 12 spectral indices and 5 PROSPECT-D modelled leaf traits related to leaf pigments, leaf water content and other structural traits for which we tested the effect of the drought treatment and genetic cluster as well as their interaction. In addition, as we found significant relationships between sapling height and stomatal conductance as well as soil moisture, we included it as a covariate (Table 3). However, sapling height did not differ significantly between the clusters ($F_{(2,177)} = 0.223$, $p = 0.8$).

3.2.1 Leaf pigments

Two out of the three main pigment classes as assessed using spectral indices (Table 2) were significantly reduced in the drought treatment (Table 3, Fig. 2). We observed a negative direction for chlorophyll-related (Fig. 3a) and carotenoid-related indices (Fig. 3c), but the

opposite in anthocyanins derived from indices, which exhibited either a neutral or a positive effect size (ARI and RGR, Fig. 3e). In addition, we found a significant reduction in the PROSPECT-D modelled leaf pigments C_{ab} and C_{car} but, in line with the results from the spectral indices, a non-significant increase in drought-treated saplings for C_{ant}. The pigment ratio between C_{ab} and C_{car} was significantly reduced between the control and the treatment group ($F_{(1,163)} = 8.4248$, $p < 0.01$). CIRE and PRI showed a significant negative association with increasing sapling height, while ARI exhibited a significant positive association with increasing sapling height, indicating more anthocyanin in taller trees.

The genetic cluster did not have a significant effect on any of the leaf pigments, nor did the treatment x genetic cluster interaction (Table 3). There was thus significant variation between the control and treatment groups, but not between genetic clusters and their response to drought, as assessed by leaf spectroscopy. Despite the absence of significant effects in the ANOVA, genetic cluster 1, which is mostly composed of Spanish provenances, showed a lower magnitude of the response to drought in several traits compared to the other genetic clusters (Fig. 3).

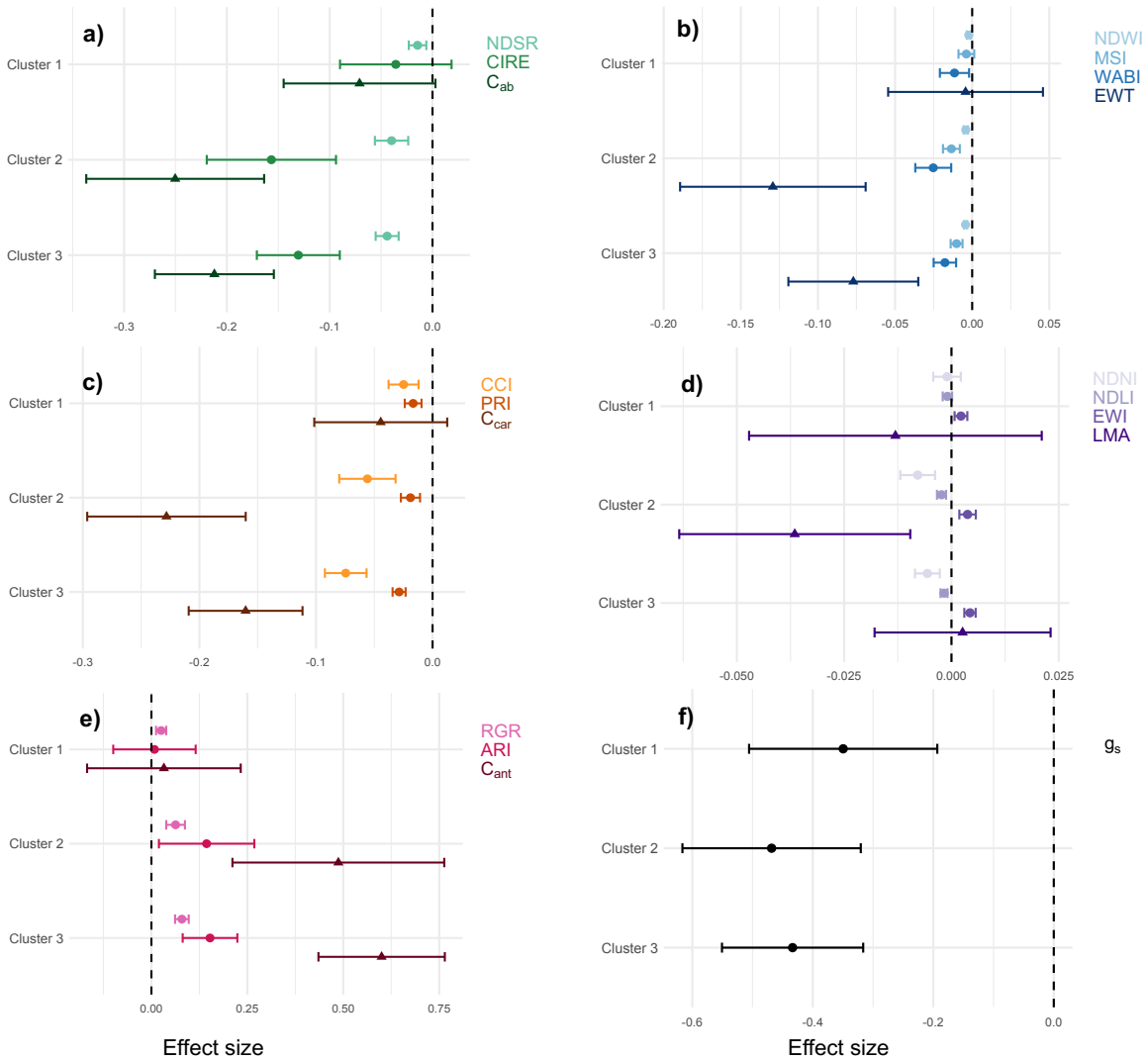

Figure 3. Log response ratios of spectral indices, PROSPECT-D simulated leaf traits and stomatal conductance (mean ± standard error; Table 2 for trait definitions). A negative effect size indicates a negative response to the experimental drought and vice versa. a) Indices related to chlorophylls. NDSR: Normalized Difference Chlorophyll Index, CIRE: Red-edge clorophyll index. C_{ab}: simulated chlorophyll a+b content. b) Traits related to water content and stress. NDWI: Normalized Difference Water Index. MSI: moisture stress. Note that a more negative response reflects greater moisture stress. WABI: Water Balance Index (water potential). EWT: Equivalent Water Thickness.

c) Indices related to carotenoids. CCI: Chlorophyll-Carotenoid Index. PRI: Photochemical Reflectance Index. C_{car}: simulated carotenoid content. d) Leaf structural traits. NDNI: Normalized Difference Nitrogen Index. NDLI: Normalized Difference Lignin Index. EWI: Epicuticular Wax Index. LMA: Leaf Mass per Area. e) Traits related to anthocyanins. RGR: Red:Green Ratio. ARI: Anthocyanin Reflectance Index. C_{ant}: simulated anthocyanin content. f) Stomatal conductance (g_s).

Specifically, for genetic cluster 1, the standard errors of the LRRs for all sets of related indices and modelled traits overlapped and LRRs were all close to zero. In contrast, spectral indices responded differently from each other and from the modelled trait they were related to for genetic clusters 2 and especially 3, which had LRRs deviating further from zero. For these clusters, the LRR of CIRE had values closer to the LRR for simulated C_{ab}, with overlapping standard error, whereas the LRR for NDSR was consistently less negative and less variable. In contrast, the LRRs for the modelled C_{car} were more negative and more variable than the LRRs for either carotenoid index (CCI or PRI), i.e., the carotenoid indices did not follow the drought response of modelled carotenoids. The carotenoid indices also behaved differently from each other, with CCI tending to have a more negative LRR with a larger standard error than PRI. Similarly, LRRs for modelled C_{ant} were consistently more positive and more variable than the LRRs for either anthocyanin index (RGR or ARI), although the standard error of the ARI LRR overlapped with the standard error of C_{ant} for cluster 2. The LRRs of RGR and ARI were similar for both clusters, with ARI being more variable.

3.2.2 Leaf water potential and water content

Drought-stressed saplings had lower NDWI and water potential index (WABI), but the moisture stress index (MSI) and modelled EWT were not affected (Table 3, and see Table 2 for indices). Again, we did not find a statistically significant difference between the genetic clusters and their interaction with the drought treatment. We found a negative drought response in these metrics of water content and water potential for all three genetic clusters (Fig. 3b). In line with the results of the leaf pigments, we observed only a slightly negative response in EWT in genetic cluster 1, revealing no water content changes between the control and the treatment saplings, and the MSI was lower in genetic cluster 1 as well (Fig. 3b). As for the pigments, for genetic cluster 1, the standard errors of the LRRs for modelled water-related traits and related indices overlapped and LRRs were all close to zero. In contrast, for clusters 2 and 3, LRRs for modeled EWT were consistently more negative and more variable than for any of the water-related indices, although modelled EWT was not significantly affected by drought due to its variability. NDWI, while significantly affected by drought, had a small magnitude of LRR, leaving WABI, with larger LRRs and a significant change with drought, the clearest water-related indicator of sapling drought response in this dataset.

3.2.3 Other structural and biochemical leaf traits

The drought treatment had a significant effect on lignin content, as assessed with the spectral index NDLI. In contrast, nitrogen content, epicuticular wax (EWI), and LMA were not significantly affected by the drought treatment as assessed by spectral indices and modeling (Fig. 3a, Table 3, and see Table 2 for indices). Again, there was no significant interaction between treatment and genetic cluster. EWI increased with drought in all three genetic clusters, suggesting an increase in epicuticular wax on the leaves, while modelled nitrogen and lignin decreased. For modelled LMA, we found a negative response to drought, although it was not significant. Sapling height was correlated with EWI, but not with any of the other structural leaf spectral traits (Table 3).

Table 3. ANOVA table for the linear models with spectral leaf traits as dependent observations and sapling height, treatment, genetic cluster and treatment x genetic cluster interaction as explanatory factors.

Variable	Factor	DF	F value	P value
NDSR	Height	1	0.0677	0.7950
	Treatment	1	20.2845	1.267e-05 ***
	Cluster	2	0.1387	0.8706
	Treatment x Cluster	2	0.6807	0.5077
	Residuals	163		
CIRE	Height	1	8.7537	0.003550 **
	Treatment	1	15.4489	0.000125 ***
	Cluster	2	0.2193	0.803280
	Treatment x Cluster	2	0.6183	0.540137
	Residuals	163		
CCI	Height	1	1.5056	0.2215783
	Treatment	1	15.6351	0.0001143 ***
	Cluster	2	0.2534	0.7764515
	Treatment x Cluster	2	0.3494	0.7056659
	Residuals	163		
PRI	Height	1	6.5809	0.01121 *
	Treatment	1	33.8565	3.059e-08 ***
	Cluster	2	0.2516	0.77782
	Treatment x Cluster	2	0.2650	0.76757
	Residuals	163		
RGR	Height	1	0.0162	0.8988
	Treatment	1	27.7693	4.278e-07 ***
	Cluster	2	0.1018	0.9033
	Treatment x Cluster	2	0.6076	0.5459
	Residuals	163		
ARI	Height	1	12.4887	0.0007106 ***
	Treatment	1	4.8022	0.0298421 *
	Cluster	2	0.8665	0.4223320
	Treatment x Cluster	2	0.1251	0.8825305
	Residuals	163		
MSI	Height	1	3.7758	0.05372
	Treatment	1	3.2350	0.07393
	Cluster	2	0.2720	0.76219
	Treatment x Cluster	2	0.6506	0.52310
	Residuals	163		
NDWI	Height	1	2.6809	0.10349
	Treatment	1	6.0518	0.01494 *
	Cluster	2	0.0714	0.93116
	Treatment x Cluster	2	0.0756	0.92721
	Residuals	163		
WABI	Height	1	3.9269	0.0492 *
	Treatment	1	20.7369	1.027e-05 ***
	Cluster	2	0.2105	0.8104
	Treatment x Cluster	2	1.0836	0.3408
	Residuals	163		
NDNI	Height	1	1.6635	0.1990
	Treatment	1	0.6213	0.4317
	Cluster	2	1.0973	0.3362

Variable	Factor	DF	F value	P value
	Treatment x Cluster	2	0.5186	0.5963
	Residuals	163		
NDLI	Height	1	0.1488	0.70014
	Treatment	1	4.8308	0.02936 *
	Cluster	2	1.3676	0.25763
	Treatment x Cluster	2	0.5083	0.60245
	Residuals	159		
EWI	Height	1	21.7964	6.299e-06 ***
	Treatment	1	1.5499	0.2149
	Cluster	2	0.1373	0.8718
	Treatment x Cluster	2	0.0009	0.9991
	Residuals	163		
C_{ab}	Height	1	6.9006	0.009439 **
	Treatment	1	21.8055	6.272e-06 ***
	Cluster	2	0.0749	0.927911
	Treatment x Cluster	2	0.8407	0.433285
	Residuals	163		
C_{car}	Height	1	1.4615	0.2284
	Treatment	1	16.2562	8.478e-05 ***
	Cluster	2	0.1950	0.8230
	Treatment x Cluster	2	1.1605	0.3159
	Residuals	163		
C_{ant}	Height	1	3.6239	0.05872
	Treatment	1	0.7938	0.37426
	Cluster	2	0.6886	0.50374
	Treatment x Cluster	2	0.0761	0.92673
	Residuals	163		
EWT	Height	1	0.8154	0.3679
	Treatment	1	1.7213	0.1914
	Cluster	2	0.4006	0.6706
	Treatment x Cluster	2	0.7447	0.4765
	Residuals	163		
LMA	Height	1	0.3761	0.5406
	Treatment	1	2.5040	0.1155
	Cluster	2	0.9626	0.3840
	Treatment x Cluster	2	1.0906	0.3385
	Residuals	163		
g_s	Height	1	68.6692	1.066e-13 ***
	Treatment	1	43.0879	1.053e-9 ***
	Cluster	2	1.5842	0.2089
	Treatment x Cluster	2	0.2493	0.7797
	Residuals	134		

3.3 Validation for PROSPECT-D inversion output

We conducted chromatographic separation for 20 leaf samples, and identified main compounds based on retention times and standard spectra. We regressed all the measured traits to the simulated traits from the PROSPECT-D inversion (Fig. 4). We found a strong and significant relationship in both chlorophylls and carotenoids (chlorophyll (C_{ab}) ($R^2 = 0.73$, $p < 0.001$), carotenoids (C_{car}) ($R^2 = 0.74$, $p < 0.001$), Fig. 4a, b) and a less strong but still significant

positive correlation for EWT and LMA (EWT (R^2 = 0.69, $p < 0.001$), LMA (R^2 = 0.51, $p < 0.001$), Fig. 4c, d). For the pigments, we also found a clear separation of leaf samples from the treatment and control pool, with the drought-treated group exhibiting a smaller pigment concentration per area (Fig. 4). However, there was systemic overestimation in both simulated C_{ab} and C_{car} ($\beta_1 > 1$).

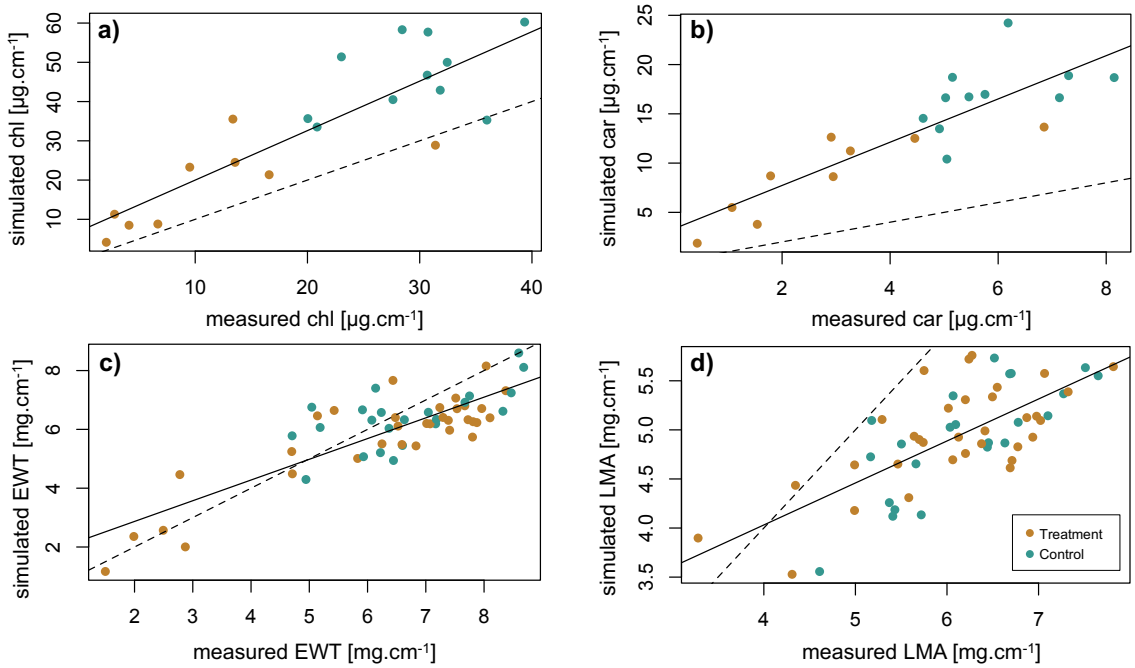

Figure 4. Measured leaf pigments (n=20) and leaf functional traits (n=56) regressed against PROSPECT-D inverse modelled traits. Dashed lines indicate x = y. a) Chlorophyll (C_{ab}) (R^2 = 0.73). b) Carotenoids (C_{car}) (R^2 = 0.74). c) EWT (R^2 = 0.69). d) LMA (R^2 = 0.51).

4. DISCUSSION

4.1 Influence of drought treatment on spectrally derived traits related to pigments

The drought treatment altered the relative abundance of photosynthetically-active pigments estimated from spectral data. Leaf chlorophyll content is well known to be positively correlated with photosynthetic rate (Emerson, 1929; Fleischer, 1935). We observed a significant decrease in chlorophyll-related indices such as NDSR and CIRE, corresponding to a decrease in PROSPECT-D simulated C_{ab} in drought-treated saplings. Previous studies found a similar decrease in chlorophyll content per area in drought-stressed *F. sylvatica* with liquid chromatography or chlorophyll measurements (Arend et al., 2016; Gallé & Feller, 2007; Tognetti et al., 1995). Gallé and Feller (2007), for instance, found that the activity of PSII during drought stress is downregulated, which is consistent with the reduction of chlorophyll in severely drought stressed leaves.

Carotenoid content was also significantly lower in drought-treated saplings compared to the control group. This is in line with previous findings (Gallé & Feller, 2007; Junaid et al., 2023). However, to compensate for water stress, the photosynthetic apparatus is known to change carotenoid composition rather than total content. One may observe an increased ratio of xantophyll cycle pigments (VAZ), which are a division of the carotenoids, to chlorophyll content

during water limitation (Biswal, 1995; Demmig-Adams & Adams, 1992; Merzlyak et al., 1999). VAZ contribute to dissipate excess energy in the apparatus during drought situations and are thus regarded as key pigments for photo-protection in *F. sylvatica* (Munné- Bosch & Alegre, 2000). This is consistent with our findings where the PROSPECT-D modelled ratio C_{chl} /C_{car} declines in drought-treated saplings. Based on these observations, we infer that when plants were water stressed, the content of both chlorophylls and carotenoids declined correspondingly. However, the proportion between them changes differently. In agreement, we found a decline in PRI and CCI, which are spectral indices related to the chlorophyll and carotenoid ratio and linked to photosynthetic capacity (Filella et al., 2009). PRI and CCI both use the 531 nm band, which is indicative of the xanthophyll cycle epoxidation state transition (Gamon et al., 1992). Both were significantly decreased in drought-treated saplings.

Anthocyanins are thought to be involved in many plant protective functions (Gould, 2004; Landi et al., 2015), as they mediate stressors (i.e., ROS) by antioxidant activity and light attenuation reducing excess energy, similar to carotenoids (Feild et al., 2001; Landi et al., 2015; Steyn et al., 2002). However, whether anthocyanins play a key role for plants, in particular beech, under water stress is still unknown, and there is little literature on anthocyanin accumulation in beech except for the copper varieties, which bear a mutated gene preventing them from degrading anthocyanins and are bred for ornamental purposes. We observed a significant increase of the anthocyanin-related indices RGR and ARI in drought-treated saplings, and also an increase of PROSPECT-D modelled C_{ant}. This suggests a potential accumulation of anthocyanins under drought stress. However, the modelled C_{ant} indicated a very low amount of C_{ant} compared to the possible range of variation in PROSPECT-D (Jacquemoud & Ustin, 2019). Consistently, we only detected anthocyanins in a copper beech sample and not in the saplings used for the drought experiment, although our laboratory analysis was not optimized to be sensitive for anthocyanins, and we do not know the limit of detection. It is likely that anthocyanin accumulation in the leaves of our experimental saplings in response to drought was low in comparison to anthocyanin concentrations that have been measured in other plants and other conditions. Gitelson et al. (2009) successfully used ARI in maple (*Acer platanoides*) and dogwood (*Cornus alba*), and the index was also able to predict anthocyanin content in lettuce (*Lactuca sativa*) (Kim & Van Iersel, 2023) and grapevines (Steele et al., 2009). However, the applicability of ARI in beech has not yet been verified, and we therefore do not draw strong conclusions based on this index. Similarly, the RGR has so far, to our knowledge, not yet been applied or validated in *F. sylvatica* exposed to drought or other environmental stress. We thus presume that anthocyanin content was relatively low in our samples during the first drought period, and that we were unable to adequately quantify its change.

4.2 Influence of drought treatment on water content and leaf structural and biochemical traits

Moisture stress and water content were captured with MSI, which was higher in drought-treated saplings, and NDWI, which was lower in drought-treated saplings, as expected. Many studies concerning hydraulic function in leaves observed a similar increase in MSI, suggesting MSI to be a good indicator of altered plant water status (Kim et al., 2015; Zhang & Zhou, 2019). One of the most crucial counteracting mechanisms of plants to water deficiency is the rapid regulation of stomatal conductance to prevent plant dehydration. The closure of stomatal pores allows a decrease of gas exchange through reduced water vapor transpiration, and thus preserving water resources (Aranda et al., 2015; Pflug et al., 2018; Wankmüller & Carminati, 2022).

As a moderately anisohydric species, *F. sylvatica* optimizes stomatal conductance under drought conditions (Peuke et al., 2002; Tardieu & Simonneau, 1998). As EWT was found to be similar in control and drought-treated saplings, this suggests that water content was preserved in the measured leaves. Another measure for hydraulic functioning and water dynamics throughout a drought is the leaf water potential (Ψ), the energy state of water in a system reflecting the water's ability to move. Ψ varies in response to climate and soil conditions, and is strongly influenced by seasonal and diurnal changes (Bartlett et al., 2012; Czajkowski & Bolte, 2006; Dietrich et al., 2019), which resembles the anisohydric behaviour of *F. sylvatica* (Roman et al., 2015). WABI, which was significantly decreased in the drought-treated saplings, relates independent changes in photoprotective pigments involved in non-photochemical quenching (NPQ) (531 nm) and water absorption bands (around 1500 nm) which indicate stress-induced changes by Ψ (Rapaport et al., 2015). Thus, we infer that Ψ was indeed decreased in drought-treated saplings. The discrepancy between changes in Ψ and no changes in water content suggests that the saplings may be able to maintain relatively stable water content through rapid adjustments such as stomatal regulations while simultaneously varying their Ψ.

LMA can be understood as the leaf-level cost of light interception (Grime, 2001). An alternative way to define LMA is the accumulation per area of different classes of compounds, such as lignin or proteins (Poorter & Villar, 1997). We derived LMA through PROSPECT-D inversion and found no effect of drought treatment, implying that leaf structure hardly had time to acclimatize to our induced 14-day drought (Poorter et al., 2009). We also did not find any drought-induced differences in modelled nitrogen content.

At the interface of the leaf towards the atmosphere, epicuticular waxes serve as a protective barrier against ultraviolet light exposure and uncontrolled water diffusion (Speckert et al., 2023). Consistent with our expectations, we observed a slight increase of EWI in drought-treated saplings, although this was not statistically significant. Epicuticular waxes are directly exposed to the environment and thus expected to adjust to abiotic stressors such as drought with the aim to conserve water and resist cellular dehydration in leaves (Shepherd & Wynne Griffiths, 2006). Studies using black pine (Pinus nigra) (Kreyling et al., 2014), grassland plant communities (Srivastava & Wiesenberg, 2018), and several shrubs and coniferous trees (Ofiti et al., 2023) increase renewal rates of waxes, which lead to a reduction of drought stress. However, the biosynthesis of waxes in *F. sylvatica* depends strongly on exposure to sun (Speckert et al., 2023). Deposition of wax as a response to water stress can occur swiftly within a few days (Bengtson et al., 1978). Our findings can thus be potentially explained by alternative water conservation mechanisms such as the aforementioned stomatal regulation to cope with the imposed water stress in *F. sylvatica*, or by changing the composition of the waxes rather than varying the concentration (Sachse et al., 2009).

As a major effect of our experimental conditions, we found that taller saplings tended to have lower soil moisture in both control and treatment groups, potentially due to larger belowground biomass and increased water needs. We thus cannot exclude the possibility that taller saplings on average experienced a more severe drought due to their higher demand for water. We addressed this limitation by controlling for sapling height within our block design and by adding sapling height as a factor in the statistical analyses. We did observe a few significant (but not very strong) associations between sapling height and leaf traits, which would indicate that the responses to drought depended at least in part on sapling height. Along with the experimental conditions, foliar water uptake through permeable cuticles in *F. sylvatica* may have influenced the actual drought conditions of the saplings (Schreel et al., 2020).

4.3 Intraspecific variation in drought stress responses across genetic clusters

There are many studies on the role of intraspecific variation, which captures the diversity of phenotypes, in tree responses to drought (e.g. *F. sylvatica*; Baudis et al., 2014; Carsjens et al., 2014; Cocozza et al., 2016; Dounavi et al., 2016; González De Andrés et al., 2021; Leuschner, 2020; Thom et al., 2023; Wang et al., 2021), with the aim to improve decision-making in forest management and enhance forest resilience to climate change. Our experiment demonstrated a variety of physiological responses to decreased water availability. We hypothesized that there is intraspecific variation in such drought responses depending on the sapling's genetic background. We tested the interaction genetic cluster x treatment for each spectral index and modelled leaf trait. Against expectations, our results reveal that drought-treated saplings did not exhibit statistically different responses based on genetic cluster. However, there were indications of variation depending on the genetic cluster, particularly in genetic cluster 1, which showed a more resistant response in multiple traits. Thus, provenances from the Iberian Peninsula seemed to have increased resistance to the imposed drought. In this context, some studies on intraspecific variation found small performance differences in varying populations of *F. sylvatica* in ecophysiological stress relevant to water stress such as photosynthetic rate and stomatal conductance (Cocozza et al., 2016; González De Andrés et al., 2021), but the variation in traits across species do not reflect a pattern at the intraspecific level (Sánchez-Gómez et al., 2013). Schmeddes et al. (2023) demonstrated the greatest phenotypic variation among *F. sylvatica* seedlings at the most basic intraspecific level, specifically within the progeny of the mother tree. This indicates a significant potential for individual acclimation to climatic variability, and population-level adaptation to drought.

4.4 Leaf spectroscopy can capture leaf phenotypic variation in drought response

Leaf spectroscopy offers a non-destructive, repeatable, and rapid measurement method for leaf traits when compared to traditional methods and is thus a promising method in ecological research (Helsen et al., 2021; Messier et al., 2010; Siefert et al., 2015), with low measurement uncertainties at the leaf level (Petibon et al., 2021). We want to highlight that despite cross-species assessment of leaf traits (i.e., EWT or LMA) (Asner et al., 2014), it remains unclear whether spectrally-derived traits models at the leaf level are accurate enough to reliably capture trait differences within species (Feilhauer et al., 2018; Girard et al., 2020). Yet, despite such uncertainties, we demonstrated that applying spectral indices to high resolution spectral data is a simple method for deriving leaf trait variation in response to an experimental drought, making it a valuable tool in ecological experiments, especially with high replication or experimental units. However, it is crucial to note that spectral indices do not necessarily provide mechanistic explanations: Spectral indices hardly disclose the leaf physiological processes and metabolic systems underpinning the drought responses (Curran, 1989; Rapaport et al., 2015). A different approach is the inclusion of relevant physical processes. The inversion of the PROSPECT RTM simulates leaf traits based on the physical processes of the interaction of electromagnetic radiation with leaves (Jacquemoud & Ustin, 2019). We performed a subsequent extraction and HPLC analysis proposed by Petibon and Wiesenberg (2022) to establish a chromatographic profile of complex photosynthetic pigments of drought-treated and control saplings to estimate chlorophyll and carotenoid concentrations, and also measured leaf mass per area and water content directly for a set of validation samples. Our validation data show a strong relationship between the simulated and extracted traits for chlorophyll and carotenoid content, and a moderate relationship for EWT and LMA. This finding aligns with Spafford et al. (2021), who found similar relationships in their dataset when N is

estimated and only reflectance is used. The robustness of the retrieval of leaf traits may also depend on the availability of transmittance T data, which can be retrieved with an integrating sphere measurement device, for instance (Petibon et al., 2021). Both R and T combined carry the most information for PROSPECT inversion (Spafford et al., 2021); however, the combined measurement of reflectance and transmittance takes at least twice as long per leaf and is a destructive measurement, requiring leaves to be removed for measurement against a fixed integrating sphere (Laliberté & Soffer, 2018). We note that for the assessment of absolute pigment concentration, other techniques are required, such as the extraction methods that we performed on a subset of leaves. However, in an experiment with dozens, hundreds, or even thousands of individuals, as can be the case in ecological studies of species-level, intraspecific and genetic variation, and multiple measurement time points, leaf traits estimated from rapidly acquired spectral data may not only be more practical but also sufficient – or even advantageous, being better able to capture change and support sufficient replication. While the challenge is to meaningfully interpret spectral data, an increasing number of studies, including ours, support this (Cavender-Bares et al., 2016; Czyż et al., 2023; Li et al., 2023; Meireles et al., 2020).

5. CONCLUSIONS

Due to increasing duration and frequency of droughts, there is growing concern that *F. sylvatica* may no longer be adapted to these novel environmental conditions, which challenges the persistence of European beech forests. Thus, understanding the ecophysiological processes that happen in response to increased drought stress is key. We detected signatures of changes in leaf traits such as photosynthetically active pigments and water potential in drought-treated saplings. However, we did not detect significant changes in signatures of leaf structural components such as nitrogen and epicuticular wax. Leaf spectroscopy offered a rapid and non-destructive way to simultaneously assess a suite of relevant traits from individual leaves, and we found the inversion of PROSPECT-D to be a robust approach to quantify changes in leaf traits in European beech saplings exposed to experimental drought. Intraspecific variation in drought responses will likely be an important determinant of beech forest persistence and evolution. With saplings representing three broad genetic clusters from different geographic regions across the species range, we did not find significant variation in the directionality of the drought responses based on the genetic cluster. However, we detected significant variation of the three genetic clusters in the magnitude of their drought response. Together, this is consistent with a developing consensus that beech populations harbour both local plasticity and geographic differentiation in their response to drought, and that the species has the potential to persist in future drier environmental conditions (Cocozza et al., 2016; Wang et al., 2021).

Author contributions

Dave Kurath: Conceptualization, Investigation, Methodology, Formal analysis, Data curation, Visualization, Writing - original draft. **Sofia J. van Moorsel**: Conceptualization, Project administration, Supervision, Investigation, Writing - original draft. **Jolanda Klaver**: Conceptualization, Investigation, Methodology, Data curation. **Tis Voortman**: Investigation, Data curation, Writing - review & editing. **Barbara Siegfried**: Investigation, Methodology, Writing-review&editing. **Yves-Alain Brügger**: Investigation, Methodology. **Abobakr Moradi**: Investigation, Methodology. **Ewa A. Czyż**: Investigation, Writing - review & editing. **Marylaure de La Harpe**: Investigation. **Guido L.B. Wiesenberg**: Methodology, Writing - review & editing. **Michael E. Schaepman**: Funding acquisition. **Meredith C. Schuman**: Conceptualization, Project administration, Supervision, Resources, Writing - review & editing.

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Conflict of interest statement

The authors declare no conflict of interest.

Data availability

Data will be publicly accessible once the manuscript is accepted (forthcoming on Dryad. https://doi.org/10.5061/dryad.cc2fqz6fm). Short reads will be made publicly accessible on the European Nucleotide Archive (project PRJEB77559).

REFERENCES

Allen, C. D., Macalady, A. K., Chenchouni, H., Bachelet, D., McDowell, N., Vennetier, M., Kitzberger, T., Rigling, A., Breshears, D. D., Hogg, E., Gonzalez, P., Fensham, R., Zhang, Z., Castro, J., Demidova, N., Lim, J.-H., Allard, G., Running, S. W., Semerci, A., & Cobb, N. (2010). A global overview of drought and heat-induced tree mortality reveals emerging climate change risks for forests. Forest Ecology and Management, 259(4), 660–684. https://doi.org/10.1016/j.foreco.2009. 09.001

Allen, W. A., Gausman, H. W., Richardson, A. J., & Thomas, J. R. (1969). Interaction of Isotropic Light with a Compact Plant Leaf [Publisher: Optica Publishing Group]. J. Opt. Soc. Am., 59(10), 1376– 1379. https://doi.org/10.1364/JOSA.59.001376

Aranda, I., Cano, F. J., Gasco, A., Cochard, H., Nardini, A., Mancha, J. A., Lopez, R., & Sanchez-Gomez, D. (2015). Variation in photosynthetic performance and hydraulic architecture across European beech (Fagus sylvatica L.) populations supports the case for local adaptation to water stress. Tree Physiology, 35(1), 34–46. https://doi.org/10.1093/treephys/tpu101

Arend, M., Sever, K., Pflug, E., Gessler, A., & Schaub, M. (2016). Seasonal photosynthetic response of European beech to severe summer drought: Limitation, recovery and post-drought stimulation. Agricultural and Forest Meteorology, 220, 83–89. https://doi.org/10.1016/j.agrformet.2016.01.011

Arnold, P. A., Kruuk, L. E. B., & Nicotra, A. B. (2019). How to analyse plant phenotypic plasticity in response to a changing climate. New Phytologist, 222(3), 1235–1241. https://doi.org/10.1111/nph.15656

Asner, G. P., & Martin, R. E. (2011). Canopy phylogenetic, chemical and spectral assembly in a lowland Amazonian forest. New Phytologist, 189(4), 999–1012. https://doi.org/10.1111/j1469-8137.2010.03549.x

Asner, G. P., Martin, R. E., Carranza-Jiménez, L., Sinca, F., Tupayachi, R., Anderson, C. B., & Martinez, P. (2014). Functional and biological diversity of foliar spectra in tree canopies throughout the Andes to Amazon region. New Phytologist, 204(1), 127–139. https://doi.org/10.1111/nph.12895

Baccari, S., Elloumi, O., Chaari-Rkhis, A., Fenollosa, E., Morales, M., Drira, N., Ben Abdallah, F., Fki, L., & Munné-Bosch, S. (2020). Linking Leaf Water Potential, Photosynthesis and Chlorophyll Loss With Mechanisms of Photo- and Antioxidant Protection in Juvenile Olive Trees Subjected to Severe Drought. Frontiers in Plant Science, 11, 614144. https://doi.org/10.3389/fpls.2020.614144

Bacelar, E. A., Santos, D. L., Moutinho-Pereira, J. M., Lopes, J. I., Gonçalves, B. C., Ferreira, T. C., & Correia, C. M. (2007). Physiological behaviour, oxidative damage and antioxidative protection of olive trees grown under different irrigation regimes. Plant and Soil, 292(1-2), 1–12. https://doi.org/10.1007/s11104-006-9088-1

Bakbergenuly, I., Hoaglin, D. C., & Kulinskaya, E. (2020). Estimation in meta-analyses of response ratios. BMC Medical Research Methodology, 20(1), 263. https://doi.org/10.1186/s12874-020-01137-1 Bartlett, M. K., Scoffoni, C., & Sack, L. (2012). The determinants of leaf turgor loss point and prediction of drought tolerance of species and

biomes: A global meta-analysis. Ecology Letters, 15(5), 393–405. https://doi.org/10.1111/j.1461-0248.2012.01751.x

Baudis, M., Ellerbrock, R. H., Felsmann, K., Gessler, A., Gimbel, K., Kayler, Z., Puhlmann, H., Ulrich, A., Weiler, M., Welk, E., & Bruelheide, H. (2014). Intraspecific differences in responses to rainshelter- induced drought and competition of Fagus sylvatica L. across Germany. Forest Ecology and Management, 330, 283–293. https://doi.org/10.1016/j.foreco.2014.07.012

Ben Abdallah, M., Methenni, K., Nouairi, I., Zarrouk, M., & Youssef, N. B. (2017). Drought priming improves subsequent more severe drought in a drought-sensitive cultivar of olive cv. Chétoui. Scientia Horticulturae, 221, 43–52. https://doi.org/10.1016/j.scienta.2017.04.021

Bengtson, C., Larsson, S., & Liljenberg, C. (1978). Effects of Water Stress on Cuticular Transpiration Rate and Amount and Composition of Epicuticular Wax in Seedlings of Six Oat Varieties. Physiologia Plantarum, 44(4), 319–324. https://doi.org/10.1111/j.1399-3054.1978.tb01630.x

Benito Garzón, M., Robson, T. M., & Hampe, A. (2019). Trait SDMs: Species distribution models that account for local adaptation and phenotypic plasticity. New Phytologist, 222(4), 1757–1765. https://doi.org/10.1111/nph.15716

Biswal, B. (1995). Carotenoid catabolism during leaf senescence and its control by light. Journal of Photochemistry and Photobiology B: Biology, 30(1), 3–13. https://doi.org/10.1016/1011-1344(95)07197-A

Blondeel, H., Guillemot, J., Martin-StPaul, N., Druel, A., Bilodeau-Gauthier, S., Bauhus, J., Grossiord, C., Hector, A., Jactel, H., Jensen, J., Messier, C., Muys, B., Serrano-León, H., Auge, H., Barsoum, N., Birhane, E., Bruelheide, H., Cavender-Bares, J., Chu, C., ... Baeten, L. (2024). Tree diversity reduces variability in sapling survival under drought. Journal of Ecology, 112(5), 1164–1180. https://doi.org/10.1111/1365-2745.14294

Bolger, A. M., Lohse, M., & Usadel, B. (2014). Trimmomatic: A flexible trimmer for Illumina sequence data. Bioinformatics, 30(15), 2114–2120. https://doi.org/10.1093/bioinformatics/btu170

Brinkmann, K., Blaschke, L., & Polle, A. (2003). Comparison of Different Methods for Lignin Determi- nation as a Basis for Calibration of Near-Infrared Reflectance Spectroscopy and Implications of Lignoproteins. Journal of chemical ecology, 28, 2483–501. https://doi.org/10.1023/A:1021484002582

Buras, A., Rammig, A., & Zang, C. S. (2020). Quantifying impacts of the 2018 drought on European ecosystems in comparison to 2003. Biogeosciences, 17(6), 1655–1672. https://doi.org/10.5194/bg-17-1655-2020

Burnett, A. C., Anderson, J., Davidson, K. J., Ely, K. S., Lamour, J., Li, Q., Morrison, B. D., Yang, D., Rogers, A., & Serbin, S. P. (2021a). A best-practice guide to predicting plant traits from leaf-level hyperspectral data using partial least squares regression (T. Lawson, Ed.). Journal of Experimental Botany, 72(18), 6175–6189. https://doi.org/10.1093/jxb/erab295

Burnett, A. C., Serbin, S. P., Davidson, K. J., Ely, K. S., & Rogers, A. (2021b). Detection of the metabolic response to drought stress using hyperspectral reflectance (R. Gleadow, Ed.). Journal of Experimental Botany, 72(18), 6474–6489. https://doi.org/10.1093/jxb/erab255

Camarillo-Castillo, F., Huggins, T. D., Mondal, S., Reynolds, M. P., Tilley, M., & Hays, D. B. (2021). High-resolution spectral information enables phenotyping of leaf epicuticular wax in wheat. Plant Methods, 17(1), 58. https://doi.org/10.1186/s13007-021-00759-w

Carsjens, C., Nguyen Ngoc, Q., Guzy, J., Knutzen, F., Meier, I. C., Muller, M., Finkeldey, R., Leuschner, C., & Polle, A. (2014). Intra-specific variations in expression of stress-related genes in beech progenies are stronger than drought-induced responses. Tree Physiology, 34(12), 1348–1361. https://doi.org/10.1093/treephys/tpu093

Caudullo, G., Welk, E., & San-Miguel-Ayanz, J. (2017). Chorological maps for the main European woody species. Data in Brief, 12, 662–666. https://doi.org/10.1016/j.dib.2017.05.007

Cavender-Bares, J., Meireles, J., Couture, J., Kaproth, M., Kingdon, C., Singh, A., Serbin, S., Center, A., Zuniga, E., Pilz, G., & Townsend, P. (2016). Associations of Leaf Spectra with Genetic and Phylogenetic Variation in Oaks: Prospects for Remote Detection of Biodiversity. Remote Sensing, 8(3), 221. https://doi.org/10.3390/rs8030221

Chang, C. C., Chow, C. C., Tellier, L. C., Vattikuti, S., Purcell, S. M., & Lee, J. J. (2015). Second- generation PLINK: Rising to the challenge of larger and richer datasets. GigaScience, 4(1), 7. https://doi.org/10.1186/s13742-015-0047-8

Cocozza, C., De Miguel, M., Pšidová, E., Ditmarová, L., Marino, S., Maiuro, L., Alvino, A., Czajkowski, T., Bolte, A., & Tognetti, R. (2016). Variation in Ecophysiological Traits and Drought Tolerance of Beech (Fagus sylvatica L.) Seedlings from Different Populations. Frontiers in Plant Science, 7. https://doi.org/10.3389/fpls.2016.00886

Cook, J., Oreskes, N., Doran, P. T., Anderegg, W. R. L., Verheggen, B., Maibach, E. W., Carlton, J. S., Lewandowsky, S., Skuce, A. G., Green, S. A., Nuccitelli, D., Jacobs, P., Richardson, M., Winkler, B., Painting, R., & Rice, K. (2016). Consensus on consensus: A synthesis of consensus estimates on human-caused global warming. Environmental Research Letters, 11(4), 048002. https://doi.org/10.1088/1748-9326/11/4/048002

Curran, P. J. (1989). Remote sensing of foliar chemistry. Remote Sensing of Environment, 30(3), 271–278. https://doi.org/10.1016/0034-4257(89)90069-2

Czajkowski, T., & Bolte, A. (2006). Unterschiedliche Reaktion deutscher und polnischer Herkünfte der Buche (Fagus sylvatica L.) auf Trockenheit.

Czyż E. A., Schmid, B., Eppinga, M. B., De La Harpe, M., Moradi, A., Li, C., Schaepman, M. E., & Schuman, M. C. (2023, August). Inferring genetic structure of European beech from observations of spectral phenotypes. https://doi.org/10.1101/2023.08.16.553487

De La Riva, E. G., Olmo, M., Poorter, H., Ubera, J. L., & Villar, R. (2016). Leaf Mass per Area (LMA) and Its Relationship with Leaf Structure and Anatomy in 34 Mediterranean Woody Species along a Water Availability Gradient (C. Armas, Ed.). PLOS ONE, 11(2), e0148788. https://doi.org/10. 1371/journal.pone.0148788

Demmig-Adams, B., & Adams, W. W. (1992). Photoprotection and Other Responses of Plants to High Light Stress. Annual Review of Plant Physiology and Plant Molecular Biology, 43(1), 599–626. https://doi.org/10.1146/annurev.pp.43.060192.003123

Dietrich, L., Delzon, S., Hoch, G., & Kahmen, A. (2019). No role for xylem embolism or

carbohydrate shortage in temperate trees during the severe 2015 drought (G. Battipaglia, Ed.). Journal of Ecology, 107(1), 334–349. https://doi.org/10.1111/1365-2745.13051

Dittmar, C., Zech, W., & Elling, W. (2003). Growth variations of Common beech (Fagus sylvatica L.) under different climatic and environmental conditions in Europe—a dendroecological study. Forest Ecology and Management, 173(1-3), 63–78. https://doi.org/10.1016/S0378-1127(01)00816-7

D'Odorico, P., Schuman, M. C., Kurz, M., & Csilléry, K. (2023). Discerning Oriental from European beech by leaf spectroscopy: Operational and physiological implications. Forest Ecology and Management, 541, 121056. https://doi.org/10.1016/j.foreco.2023.121056

Dounavi, A., Netzer, F., Celepirovic, N., Ivanković, M., Burger, J., Figueroa, A., Schön, S., Simon, J., Cremer, E., Fussi, B., Konnert, M., & Rennenberg, H. (2016). Genetic and physiological differences of European beech provenances (F. sylvatica L.) exposed to drought stress. Forest Ecology and Management, 361, 226–236. https://doi.org/10.1016/j.foreco.2015.11.014

Durrant, T., de Rigo, D., & Caudullo, G. (2016, March). Fagus sylvatica in Europe: Distribution, habitat, usage and threats.

Edge, R., & Truscott, T. (2018). Singlet Oxygen and Free Radical Reactions of Retinoids and Carotenoids—A Review. Antioxidants, 7(1), 5. https://doi.org/10.3390/antiox7010005

Emerson, R. (1929). Chlorophyll content and rate of photosynthesis. Proceedings of the National Academy of Sciences, 15(3), 281–284. https://doi.org/10.1073/pnas.15.3.281

Feild, T. S., Lee, D. W., & Holbrook, N. M. (2001). Why Leaves Turn Red in Autumn. The Role of Anthocyanins in Senescing Leaves of Red-Osier Dogwood. Plant Physiology, 127(2), 566–574. https://doi.org/10.1104/pp.010063

Feilhauer, H., Schmid, T., Faude, U., Sánchez-Carrillo, S., & Cirujano, S. (2018). Are remotely sensed traits suitable for ecological analysis? A case study of long-term drought effects on leaf mass per area of wetland vegetation. Ecological Indicators, 88, 232–240. https://doi.org/10.1016/j.ecolind. 2018.01.012

Féret, J.-B., François, C., Asner, G. P., Gitelson, A. A., Martin, R. E., Bidel, L. P., Ustin, S. L., Le Maire, G., & Jacquemoud, S. (2008). PROSPECT-4 and 5: Advances in the leaf optical properties model separating photosynthetic pigments. Remote Sensing of Environment, 112(6), 3030–3043. https://doi.org/10.1016/j.rse.2008.02.012

Féret, J.-B., Gitelson, A., Noble, S., & Jacquemoud, S. (2017). PROSPECT-D: Towards modeling leaf optical properties through a complete lifecycle. Remote Sensing of Environment, 193, 204–215. https://doi.org/10.1016/j.rse.2017.03.004

Féret, J.-B., Le Maire, G., Jay, S., Berveiller, D., Bendoula, R., Hmimina, G., Cheraiet, A., Oliveira, J., Ponzoni, F., Solanki, T., De Boissieu, F., Chave, J., Nouvellon, Y., Porcar-Castell, A., Proisy, C., Soudani, K., Gastellu-Etchegorry, J.-P., & Lefèvre-Fonollosa, M.-J. (2019). Estimating leaf mass per area and equivalent water thickness based on leaf optical properties: Potential and limitations of physical modeling and machine learning. Remote Sensing of Environment, 231, 110959. https://doi.org/10.1016/j.rse.2018.11.002

Filella, I., Porcar-Castell, A., Munné-Bosch, S., Bäck, J., Garbulsky, M. F., & Peñuelas, J.

(2009). PRI assessment of long-term changes in carotenoids/chlorophyll ratio and short-term changes in de-epoxidation state of the xanthophyll cycle. International Journal of Remote Sensing, 30(17), 4443–4455. https://doi.org/10.1080/01431160802575661

Fleischer, W. E. (1935). The relation between chlorophyll content and rate of photosynthesis. Journal of General Physiology, 18(4), 573–597. https://doi.org/10.1085/jgp.18.4.573

Gai, J., Wang, J., Xie, S., Xiang, L., & Wang, Z. (2023). Spectroscopic determination of chlorophyll content in sugarcane leaves for drought stress detection. Precision Agriculture. https://doi.org/10.1007/ s11119-023-10082-0

Gallé, A., & Feller, U. (2007). Changes of photosynthetic traits in beech saplings (Fagus sylvatica) under severe drought stress and during recovery. Physiologia Plantarum, 131(3), 412–421. https://doi.org/10.1111/j.1399-3054.2007.00972.x

Gamon, J. A., & Surfus, J. S. (1999). Assessing leaf pigment content and activity with a reflectometer. New Phytologist, 143(1), 105–117. https://doi.org/10.1046/j.1469-8137.1999.00424.x

Gamon, J., Peñuelas, J., & Field, C. (1992). A narrow-waveband spectral index that tracks diurnal changes in photosynthetic efficiency. Remote Sensing of Environment, 41(1), 35–44. https://doi.org/10. 1016/0034-4257(92)90059-S

Gamon, J. A., Huemmrich, K. F., Wong, C. Y. S., Ensminger, I., Garrity, S., Hollinger, D. Y., Noormets, A., & Peñuelas, J. (2016). A remotely sensed pigment index reveals photosynthetic phenology in evergreen conifers. Proceedings of the National Academy of Sciences, 113(46), 13087–13092. https://doi.org/10.1073/pnas.1606162113

Gao, B.-c. (1996). NDWI—A normalized difference water index for remote sensing of vegetation liquid water from space. Remote Sensing of Environment, 58(3), 257–266. https://doi.org/10.1016/S0034- 4257(96)00067-3

García-Plazaola, J., & Becerril, J. (2000). Effects of drought on photoprotective mechanisms in European beech (Fagus sylvatica L.) seedlings from different provenances. Trees, 14(8), 485–490. https://doi.org/10.1007/s004680000068

Gazol, A., Fajardo, A., & Camarero, J. J. (2023). Contributions of Intraspecific Variation to Drought Tolerance in Trees. Current Forestry Reports, 9(6), 461–472. https://doi.org/10.1007/s40725-023- 00199-w

Geßler, A., Keitel, C., Kreuzwieser, J., Matyssek, R., Seiler, W., & Rennenberg, H. (2006). Potential risks for European beech (Fagus sylvatica L.) in a changing climate. Trees, 21(1), 1–11. https://doi.org/10.1007/s00468-006-0107-x

Gezan, S., Oliveira, A. A. d., Galli, G., & Murray, D. (2022). ASRgenomics: An R package with comple- mentary genomic functions. VSN International.

Girard, A., Schweiger, A. K., Carteron, A., Kalacska, M., & Laliberté, E. (2020). Foliar Spectra and Traits of Bog Plants across Nitrogen Deposition Gradients. Remote Sensing, 12(15), 2448. https://doi.org/10.3390/rs12152448

Gitelson, A. A. (2005). Remote estimation of canopy chlorophyll content in crops. Geophysical Research Letters, 32(8), L08403. https://doi.org/10.1029/2005GL022688

Gitelson, A. A., Chivkunova, O. B., & Merzlyak, M. N. (2009). Nondestructive estimation of anthocyanins and chlorophylls in anthocyanic leaves. American Journal of Botany, 96(10), 1861–1868. https://doi.org/10.3732/ajb.0800395

Gitelson, A. A., Merzlyak, M. N., & Chivkunova, O. B. (2001). Optical Properties and Nondestructive Estimation of Anthocyanin Content in Plant Leaves. Photochemistry and Photobiology, 74(1), 38. https://doi.org/10.1562/0031-8655(2001)074⟨0038:OPANEO⟩2.0.CO;2

González De Andrés, E., Rosas, T., Camarero, J. J., & Martínez-Vilalta, J. (2021). The intraspecific variation of functional traits modulates drought resilience of European beech and pubescent oak. Journal of Ecology, 109(10), 3652–3669. https://doi.org/10.1111/1365-2745.13743

Gould, K. S. (2004). Nature's Swiss Army Knife: The Diverse Protective Roles of Anthocyanins in Leaves. Journal of Biomedicine and Biotechnology, 2004(5), 314–320. https://doi.org/10.1155/ S1110724304406147

Green, B. R., & Durnford, D. G. (1996). The chlorophyll-carotenoid proteins of oxygenic photosynthesis. Annual Review of Plant Physiology and Plant Molecular Biology, 47(1), 685–714. https://doi.org/ 10.1146/annurev.arplant.47.1.685

Grime, J. (2001, January). Plant Strategies, Vegetation Processes, and Ecosystem Properties [Journal Abbreviation: Biological Conservation - BIOL CONSERV]. In Biological Conservation - BIOL CONSERV (Vol. 107). https://doi.org/10.1016/S0006-3207(02)00055-1

Havaux, M. (2014). Carotenoid oxidation products as stress signals in plants. The Plant Journal, 79(4), 597–606. https://doi.org/10.1111/tpj.12386

Hedges, L. V., Gurevitch, J., & Curtis, P. S. (1999). The meta-analysis of response ration in experimental ecology. Ecology, 80(4), 1150–1156. https://doi.org/10.1890/0012-9658(1999)080[1150 : TMAORR]2.0.CO;2

Helsen, K., Bassi, L., Feilhauer, H., Kattenborn, T., Matsushima, H., Van Cleemput, E., Somers, B., & Honnay, O. (2021). Evaluating different methods for retrieving intraspecific leaf trait variation from hyperspectral leaf reflectance. Ecological Indicators, 130, 108111. https://doi.org/10.1016/j. ecolind.2021.108111

Hermann, M., Röthlisberger, M., Gessler, A., Rigling, A., Senf, C., Wohlgemuth, T., & Wernli, H. (2023). Meteorological history of low-forest-greenness events in Europe in 2002–2022. Biogeosciences, 20(6), 1155–1180. https://doi.org/10.5194/bg-20-1155-2023

Hijmans, R. J., Cameron, S. E., Parra, J. L., Jones, P. G., & Jarvis, A. (2005). Very high resolution interpolated climate surfaces for global land areas. International Journal of Climatology, 25(15), 1965–1978. https://doi.org/10.1002/joc.1276

Hu, F., Zhang, Y., & Guo, J. (2023). Effects of drought stress on photosynthetic physiological characteristics, leaf microstructure, and related gene expression of yellow horn. Plant Signaling & Behavior, 18(1), 2215025. https://doi.org/10.1080/15592324.2023.2215025

Hu, Y., Li, W. C., Xu, Y. Q., Li, G. J., Liao, Y., & Fu, F. (2009). Differential expression of candidate genes for lignin biosynthesis under drought stress in maize leaves. Journal of Applied Genetics, 50(3), 213–223. https://doi.org/10.1007/BF03195675

Hueni, A., & Bialek, A. (2017). Cause, Effect, and Correction of Field Spectroradiometer Interchannel Radiometric Steps. IEEE Journal of Selected Topics in Applied Earth Observations and Remote Sensing, 10(4), 1542–1551. https://doi.org/10.1109/JSTARS.2016.2625043

Hunt Jr., E., & Rock, B. (1989). Detection of changes in leaf water content using Near- and Middle-Infrared reflectances. Remote Sensing of Environment, 30(1), 43–54. https://doi.org/10.1016/0034-4257(89)90046-1

IPCC. (2023, June). Climate Change 2022 – Impacts, Adaptation and Vulnerability: Working Group II Contribution to the Sixth Assessment Report of the Intergovernmental Panel on Climate Change (1st ed.). Cambridge University Press. https://doi.org/10.1017/9781009325844

Jacquemoud, S., & Baret, F. (1990). PROSPECT: A model of leaf optical properties spectra. Remote.Sensing of Environment, 34(2), 75–91. https://doi.org/10.1016/0034-4257(90)90100-Z

Jacquemoud, S., Ustin, S., Verdebout, J., Schmuck, G., Andreoli, G., & Hosgood, B. (1996). Estimating leaf biochemistry using the PROSPECT leaf optical properties model. Remote Sensing of Environment, 56(3), 194–202. https://doi.org/10.1016/0034-4257(95)00238-3

Jacquemoud, S., & Ustin, S. (2019, September). Leaf Optical Properties (1st ed.). Cambridge University Press. https://doi.org/10.1017/9781108686457

Jenks, M. A., & Ashworth, E. N. (1998, December). Plant Epicuticular Waxes: Function, Production, and Genetics. In J. Janick (Ed.), Horticultural Reviews (1st ed., pp. 1–68). Wiley. https://doi.org/10. 1002/9780470650752.ch1

Ji, F., Li, F., Hao, D., Shiklomanov, A. N., Yang, X., Townsend, P. A., Dashti, H., Nakaji, T., Kovach, K. R., Liu, H., Luo, M., & Chen, M. (2024). Unveiling the transferability of PLSR models for leaf trait estimation: Lessons from a comprehensive analysis with a novel global dataset. New Phytologist, 243(1), 111–131. https://doi.org/10.1111/ nph.19807

Junaid, M. D., Öztürk, Z. N., & Gökçe, A. F. (2023). Drought stress effects on morphophysiological and quality characteristics of commercial carrot cultivars. Turkish Journal of Botany, 47(2), 111–126. https://doi.org/10.55730/1300-008X.2750

Junttila, S., Hölttä, T., Saarinen, N., Kankare, V., Yrttimaa, T., Hyyppä, J., & Vastaranta, M. (2022). Close-range hyperspectral spectroscopy reveals leaf water content dynamics. Remote Sensing of Environment, 277, 113071. https://doi.org/10.1016/j.rse.2022.113071

Kang, H. M., Zaitlen, N. A., Wade, C. M., Kirby, A., Heckerman, D., Daly, M. J., & Eskin, E. (2008). Efficient Control of Population Structure in Model Organism Association Mapping. Genetics, 178(3), 1709–1723. https://doi.org/10.1534/genetics.107.080101

Kim, C., & Van Iersel, M. W. (2023). Image-based phenotyping to estimate anthocyanin concentrations in lettuce. Frontiers in Plant Science, 14, 1155722. https://doi.org/10.3389/fpls.2023.1155722

Kim, D. M., Zhang, H., Zhou, H., Du, T., Wu, Q., Mockler, T. C., & Berezin, M. Y. (2015). Highly sensitive image-derived indices of water-stressed plants using hyperspectral imaging in SWIR and histogram analysis. Scientific Reports, 5, 15919. https://doi.org/10.1038/srep15919

Kokot, M., Długosz, M., & Deorowicz, S. (2017). KMC 3: Counting and manipulating k -mer statistics (B. Berger, Ed.). Bioinformatics, 33(17), 2759–2761. https://doi.org/10.1093/bioinformatics/btx304

Kothari, S., & Schweiger, A. K. (2022). Plant spectra as integrative measures of plant phenotypes. Journal of Ecology, 110(11), 2536–2554. https://doi.org/10.1111/1365-2745.13972

Kreyling, J., Buhk, C., Backhaus, S., Hallinger, M., Huber, G., Huber, L., Jentsch, A., Konnert, M., Thiel, D., Wilmking, M., & Beierkuhnlein, C. (2014). Local adaptations to frost in marginal and central populations of the dominant forest tree Fagus sylvatica L. as affected by temperature and extreme drought in common garden experiments. Ecology and Evolution, 4(5), 594–605. https://doi.org/10.1002/ece3.971

Lajeunesse, M. J. (2016). Bias and correction for the log response ratio in ecological meta-analysis [Publisher: Figshare]. https://doi.org/10.6084/M9.FIGSHARE.C.3307854.V1

Laliberté, E., & Soffer, R. (2018, June). Measuring spectral reflectance and transmittance (350-2500 nm) of small and/or narrow leaves using the Spectra Vista Corporation (SVC) DC-R/T Integrating Sphere v3. https://doi.org/10.17504/protocols.io.q56dy9e

Landi, M., Tattini, M., & Gould, K. S. (2015). Multiple functional roles of anthocyanins in plant-environment interactions. Environmental and Experimental Botany, 119, 4–17. https://doi.org/10.1016/j.envexpbot.2015.05.012

León-Lobos, P., & Ellis, R. H. (2002). Seed storage behaviour of Fagus sylvatica and Fagus crenata. Seed Science Research, 12(1), 31–37. https://doi.org/10.1079/SSR200195

Leuschner, C. (2020). Drought response of European beech (Fagus sylvatica L.)—A review. Perspectives in Plant Ecology, Evolution and Systematics, 47, 125576. https://doi.org/10.1016/j.ppees.2020.125576

Leuzinger, S., Zotz, G., Asshoff, R., & Korner, C. (2005). Responses of deciduous forest trees to severe drought in Central Europe. Tree Physiology, 25(6), 641–650. https://doi.org/10.1093/treephys/25.6.641

Li, C., Czyż, E. A., Halitschke, R., Baldwin, I. T., Schaepman, M. E., & Schuman, M. C. (2023). Evaluating potential of leaf reflectance spectra to monitor plant genetic variation. Plant Methods, 19(1), 108. https://doi.org/10.1186/s13007-023-01089-9

Li, Z., & Ahammed, G. J. (2023). Plant stress response and adaptation via anthocyanins: A review. Plant Stress, 10, 100230. https://doi.org/10.1016/j.stress.2023.100230

Liu, Q., Luo, L., & Zheng, L. (2018). Lignins: Biosynthesis and Biological Functions in Plants. International Journal of Molecular Sciences, 19(2), 335. https://doi.org/10.3390/ijms19020335

Mafakheri, A., Siosemardeh, A., Bahramnejad, B., Struik, P., & Sohrabi, Y. (2010). Effect of Drought Stress on Yield, Proline and Chlorophyll Contents in Three Chickpea Cultivars [Place: Lismore, N.S.W. Publisher: Southern Cross Publishers]. Australian Journal of Crop Science, 4(8), 580–585. Retrieved January 4, 2024, from https://search.informit.org/doi/10.3316/informit.857341254680658

Martinez del Castillo, E., Zang, C. S., Buras, A., Hacket-Pain, A., Esper, J., Serrano-Notivoli, R., Hartl, C., Weigel, R., Klesse, S., Resco de Dios, V., Scharnweber, T., Dorado-Liñán, I., van

der Maaten- Theunissen, M., van der Maaten, E., Jump, A., Mikac, S., Banzragch, B.-E., Beck, W., Cavin, L., ... de Luis, M. (2022). Climate-change-driven growth decline of European beech forests. Communications Biology, 5(1), 163. https://doi.org/10.1038/s42003-022-03107-3

Martínez-Vilalta, J., García-Valdés, R., Jump, A., Vilà-Cabrera, A., & Mencuccini, M. (2023). Accounting for trait variability and coordination in predictions of drought-induced range shifts in woody plants. New Phytologist, 240(1), 23–40. https://doi.org/10.1111/nph.19138

McDowell, N., Pockman, W. T., Allen, C. D., Breshears, D. D., Cobb, N., Kolb, T., Plaut, J., Sperry, J., West, A., Williams, D. G., & Yepez, E. A. (2008). Mechanisms of plant survival and mortality during drought: Why do some plants survive while others succumb to drought? New Phytologist, 178(4), 719–739. https://doi.org/10.1111/j.1469-8137.2008.02436.x

Meireles, J. E., & Schweiger, A. (2020, July). Meireles/spectrolab: CRAN-v0.0.9. https://doi.org/10.5281/ZENODO.3934575

Meireles, J. E., Cavender-Bares, J., Townsend, P. A., Ustin, S., Gamon, J. A., Schweiger, A. K., Schaepman, M. E., Asner, G. P., Martin, R. E., Singh, A., Schrodt, F., Chlus, A., & O'Meara, B. C. (2020). Leaf reflectance spectra capture the evolutionary history of seed plants. New Phytologist, 228(2), 485–493. https://doi.org/10.1111/nph.16771

Merzlyak, M. N., Gitelson, A. A., Chivkunova, O. B., & Rakitin, V. Y. (1999). Non-destructive optical detection of pigment changes during leaf senescence and fruit ripening. Physiologia Plantarum, 106(1), 135–141. https://doi.org/10.1034/j.1399-3054.1999.106119.x

Messier, J., McGill, B. J., & Lechowicz, M. J. (2010). How do traits vary across ecological scales? A case for trait-based ecology. Ecology Letters, 13(7), 838–848. https://doi.org/10.1111/j.1461- 0248.2010.01476.x

Millán, M. M. (2014). Extreme hydrometeorological events and climate change predictions in Europe. Journal of Hydrology, 518, 206–224. https://doi.org/10.1016/j.jhydrol.2013.12.041

Miller, J. R., Hare, E. W., Hollinger, A. B., & Sturgeon, D. R. (1987, January). Imaging Spectrometry As A Tool For Botanical Mapping. In G. Vane (Ed.). https://doi.org/10.1117/12.942289

Mishra, B., Ulaszewski, B., Meger, J., Aury, J.-M., Bodénès, C., Lesur-Kupin, I., Pfenninger, M., Da Silva, C., Gupta, D. K., Guichoux, E., Heer, K., Lalanne, C., Labadie, K., Opgenoorth, L., Ploch, S., Le Provost, G., Salse, J., Scotti, I., Wötzel, S., ... Thines, M. (2022). A Chromosome-Level Genome Assembly of the European Beech (Fagus sylvatica) Reveals Anomalies for Organelle DNA Integration, Repeat Content and Distribution of SNPs. Frontiers in Genetics, 12, 691058. https://doi.org/10.3389/fgene.2021.691058

Munné-Bosch, S., & Alegre, L. (2000). Changes in carotenoids, tocopherols and diterpenes during drought and recovery, and the biological significance of chlorophyll loss in Rosmarinus officinalis plants. Planta, 210(6), 925–931. https://doi.org/10.1007/s004250050699

NCCS. (2018). CH2018 - Klimaszenarien für die Schweiz.

Nguyen, Q. N., Polle, A., & Pena, R. (2017). Intraspecific variations in drought response and fitness traits of beech (Fagus sylvatica L.) seedlings from three provenances differing in annual precipitation. Trees, 31(4), 1215–1225. https://doi.org/10.1007/s00468-017-1539-1

Ofiti, N. O., Altermatt, M., Petibon, F., Warren, J. M., Malhotra, A., Hanson, P. J., & Wiesenberg, G. L. (2023). Warming and elevated CO2 induced shifts in carbon partitioning and lipid composition within an ombrotrophic bog plant community. Environmental and Experimental Botany, 206, 105182. https://doi.org/10.1016/j.envexpbot.2022.105182

Packham, J. R., Thomas, P. A., Atkinson, M. D., & Degen, T. (2012). Biological Flora of the British Isles: Fagus sylvatica. Journal of Ecology, 100(6), 1557–1608. https://doi.org/10.1111/j.1365-2745.2012.02017.x

Peat, H. J., & Fitter, A. H. (1994). A comparative study of the distribution and density of stomata in the British flora. Biological Journal of the Linnean Society, 52(4), 377–393. https://doi.org/10.1111/j.1095-8312.1994.tb00999.x

Pedregosa, F., Varoquaux, G., Gramfort, A., Michel, V., Thirion, B., Grisel, O., Blondel, M., Prettenhofer, P., Weiss, R., Dubourg, V., Vanderplas, J., Passos, A., Cournapeau, D., Brucher, M., Perrot, M., Duchesnay, E., & Louppe, G. (2012). Scikit-learn: Machine Learning in Python. Journal of Machine Learning Research, 12.

Petibon, F., Schuman, M., Wiesenberg, G., Schaepman, M., & Schmidt, M. (2023). Seeing the Forest for the Trees: European Beech Through the Lenses of Spectroscopy and Metabolomics. Universität Zürich.

Petibon, F., Czyż, E. A., Ghielmetti, G., Hueni, A., Kneubühler, M., Schaepman, M. E., & Schuman, M. C. (2021). Uncertainties in measurements of leaf optical properties are small compared to the biological variation within and between individuals of European beech. Remote Sensing of Environment, 264, 112601. https://doi.org/10.1016/j.rse.2021.112601

Petibon, F., & Wiesenberg, G. L. B. (2022). Characterization of complex photosynthetic pigment profiles in European deciduous tree leaves by sequential extraction and reversed-phase high-performance liquid chromatography. Frontiers in Plant Science, 13, 957606. https://doi.org/10.3389/fpls.2022.957606

Petrovic, S., Zvezdanović, J., Andelković, T., & Marković, D. (2012). The identification of chlorophyll and its derivatives in the pigment mixtures: HPLC-chromatography, visible and mass spectroscopy studies. Savremene tehnologije, 1, 16–24.

Peuke, A. D., Schraml, C., Hartung, W., & Rennenberg, H. (2002). Identification of drought-sensitive beech ecotypes by physiological parameters. New Phytologist, 154(2), 373–387. https://doi.org/ 10.1046/j.1469-8137.2002.00400.x

Pflug, E. E., Buchmann, N., Siegwolf, R. T. W., Schaub, M., Rigling, A., & Arend, M. (2018). Resilient Leaf Physiological Response of European Beech (Fagus sylvatica L.) to Summer Drought and Drought Release. Frontiers in Plant Science, 9, 187. https://doi.org/10.3389/fpls.2018.00187

Poorter, H., Niinemets, Ü., Poorter, L., Wright, I. J., & Villar, R. (2009). Causes and consequences of variation in leaf mass per area (LMA): A meta-analysis. New Phytologist, 182(3), 565–588. https://doi.org/10.1111/j.1469-8137.2009.02830.x

Poorter, H., & Villar, R. (1997). The fate of acquired carbon in plants: Chemical composition and construction costs. https://api.semanticscholar.org/CorpusID:194329

Purcell, S., Neale, B., Todd-Brown, K., Thomas, L., Ferreira, M. A., Bender, D., Maller, J.,

Sklar, P., De Bakker, P. I., Daly, M. J., & Sham, P. C. (2007). PLINK: A Tool Set for Whole-Genome Association and Population-Based Linkage Analyses. The American Journal of Human Genetics, 81(3), 559–575. https://doi.org/10.1086/519795

Quero, J. L., Villar, R., Marañón, T., & Zamora, R. (2006). Interactions of drought and shade effects on seedlings of four Quercus species: Physiological and structural leaf responses. New Phytologist, 170(4), 819–834. https://doi.org/10.1111/j.1469-8137.2006.01713.x

R Core Team. (2023). R: A Language and Environment for Statistical Computing. https : / / www. R- project.org/

Rapaport, T., Hochberg, U., Shoshany, M., Karnieli, A., & Rachmilevitch, S. (2015). Combining leaf physiology, hyperspectral imaging and partial least squares-regression (PLS-R) for grapevine water status assessment. ISPRS Journal of Photogrammetry and Remote Sensing, 109, 88–97. https://doi.org/10.1016/j.isprsjprs.2015.09.003

Rigling, A., Etzold, S., Bebi, P., Brang, P., Ferretti, M., Forrester, D., Gärtner, H., Gessler, A., Ginzler, C., Moser, B., Schaub, M., Stroheker, S., Trotsiuk, V., Walthert, L., Zweifel, R., & Wohlgemuth, T. (2019). Wie viel Trockenheit ertragen unsere Wälder? Lehren aus extremen Trockenjahren. Lernen aus Extremereignissen, 39–51. https://doi.org/10.55419/wsl:19983

Roman, D. T., Novick, K. A., Brzostek, E. R., Dragoni, D., Rahman, F., & Phillips, R. P. (2015). The role of isohydric and anisohydric species in determining ecosystem-scale response to severe drought. Oecologia, 179(3), 641–654. https://doi.org/10.1007/s00442-015-3380-9

Rose, L., Leuschner, C., Köckemann, B., & Buschmann, H. (2009). Are marginal beech (Fagus sylvatica L.) provenances a source for drought tolerant ecotypes? European Journal of Forest Research, 128(4), 335–343. https://doi.org/10.1007/s10342-009-0268-4

Rukh, S., Sanders, T. G. M., Krüger, I., Schad, T., & Bolte, A. (2023). Distinct Responses of European Beech (Fagus sylvatica L.) to Drought Intensity and Length—A Review of the Impacts of the 2003 and 2018–2019 Drought Events in Central Europe. Forests, 14(2), 248. https://doi.org/10. 3390/f14020248

Sachse, D., Kahmen, A., & Gleixner, G. (2009). Significant seasonal variation in the hydrogen isotopic com- position of leaf-wax lipids for two deciduous tree ecosystems (Fagus sylvativa and Acerpseudoplatanus). Organic Geochemistry, 40(6), 732–742. https://doi.org/10.1016/j.orggeochem.2009.02.008

Saha, S., Singh, J., Paul, A., Sarkar, R., Khan, Z., & Banerjee, K. (2020). Anthocyanin Profiling Using UV-Vis Spectroscopy and Liquid Chromatography Mass Spectrometry. Journal of AOAC INTERNATIONAL, 103(1), 23–39. https://doi.org/10.5740/jaoacint.19-0201

Sánchez-Gómez, D., Robson, T. M., Gascó, A., Gil-Pelegrín, E., & Aranda, I. (2013). Differences in the leaf functional traits of six beech (Fagus sylvatica L.) populations are reflected in their response to water limitation. Environmental and Experimental Botany, 87, 110–119. https://doi.org/10.1016/j.envexpbot.2012.09.011

Schaepman-Strub, G., Schaepman, M., Painter, T., Dangel, S., & Martonchik, J. (2006). Reflectance quantities in optical remote sensing—definitions and case studies. Remote Sensing of Environment, 103(1), 27–42. https://doi.org/10.1016/j.rse.2006.03.002

Schmeddes, J., Muffler, L., Barbeta, A., Beil, I., Bolte, A., Holm, S., Karitter, P., Klisz, M., Löf,

M., Nicolas, M., Peñuelas, J., Vitasse, Y., Weigel, R., & Kreyling, J. (2023). High phenotypic variation found within the offspring of each mother tree in Fagus sylvatica regardless of the environment or source population. Global Ecology and Biogeography, geb.13794. https://doi.org/10.1111/geb.13794

Schmid, M. W., Moradi, A., Leigh, D. M., Schuman, M. C., & Van Moorsel, S. J. (2024, February). Covering the bases: Population genomic structure of Lemna minor and the cryptic species L. japonica in Switzerland. https://doi.org/10.1101/2024.02.14.580260

Schmied, G., Pretzsch, H., Ambs, D., Uhl, E., Schmucker, J., Fäth, J., Biber, P., Hoffmann, Y.-D., Sěho, M., Mellert, K. H., & Hilmers, T. (2023). Rapid beech decline under recurrent drought stress: Individual neighborhood structure and soil properties matter. Forest Ecology and Management, 545, 121305. https://doi.org/10.1016/j.foreco.2023.121305

Schneider, C. A., Rasband, W. S., & Eliceiri, K. W. (2012). NIH Image to ImageJ: 25 years of image analysis. Nature Methods, 9(7), 671–675. https://doi.org/10.1038/nmeth.2089

Schreel, J. D. M., Leroux, O., Goossens, W., Brodersen, C., Rubinstein, A., & Steppe, K. (2020). Identifying the pathways for foliar water uptake in beech (Fagus sylvatica L.): A major role for trichomes. The Plant Journal, 103(2), 769–780. https://doi.org/10.1111/tpj.14770

Serbin, S. P., Singh, A., McNeil, B. E., Kingdon, C. C., & Townsend, P. A. (2014). Spectroscopic determination of leaf morphological and biochemical traits for northern temperate and boreal tree species. Ecological Applications, 24(7), 1651–1669. https://doi.org/10.1890/13-2110.1

Serrano, L., Peñuelas, J., & Ustin, S. L. (2002). Remote sensing of nitrogen and lignin in Mediterranean vegetation from AVIRIS data. Remote Sensing of Environment, 81(2-3), 355–364. https://doi.org/ 10.1016/S0034-4257(02)00011-1

Shepherd, T., & Wynne Griffiths, D. (2006). The effects of stress on plant cuticular waxes. New Phytologist, 171(3), 469–499. https://doi.org/10.1111/j.1469-8137.2006.01826.x

Siefert, A., Violle, C., Chalmandrier, L., Albert, C. H., Taudiere, A., Fajardo, A., Aarssen, L. W., Baraloto, C., Carlucci, M. B., Cianciaruso, M. V., De L. Dantas, V., De Bello, F., Duarte, L. D. S., Fonseca, C. R., Freschet, G. T., Gaucherand, S., Gross, N., Hikosaka, K., Jackson, B., ... Wardle, D. A. (2015). A global meta-analysis of the relative extent of intraspecific trait variation in plant communities (J. Chase, Ed.). Ecology Letters, 18(12), 1406–1419. https://doi.org/10.1111/ele. 12508

Sims, D. A., & Gamon, J. A. (2002). Relationships between leaf pigment content and spectral reflectance across a wide range of species, leaf structures and developmental stages. Remote Sensing of Environment, 81(2-3), 337–354. https://doi.org/10.1016/S0034-4257(02)00010-X

Spafford, L., Le Maire, G., MacDougall, A., De Boissieu, F., & Féret, J.-B. (2021). Spectral subdomains and prior estimation of leaf structure improves PROSPECT inversion on reflectance or transmittance alone. Remote Sensing of Environment, 252, 112176. https://doi.org/10.1016/j.rse.2020.112176

Speckert, T. C., Petibon, F., & Wiesenberg, G. L. B. (2023). Late-season biosynthesis of leaf fatty acids and n-alkanes of a mature beech (Fagus sylvatica) tree traced via13CO2 pulse-chase labelling and compound-specific isotope analysis. Frontiers in Plant Science, 13, 1029026. https://doi.org/10.3389/fpls.2022.1029026

Srivastava, K., & Wiesenberg, G. L. (2018). Severe drought-influenced composition and 13C of plant and soil n-alkanes in model temperate grassland and heathland ecosystems. Organic Geochemistry, 116, 77–89. https://doi.org/10.1016/j.orggeochem.2017.11.002

Steele, M. R., Gitelson, A. A., Rundquist, D. C., & Merzlyak, M. N. (2009). Nondestructive Estimation of Anthocyanin Content in Grapevine Leaves. American Journal of Enology and Viticulture, 60(1), 87–92. https://doi.org/10.5344/ajev.2009.60.1.87

Steyn, W. J., Wand, S. J. E., Holcroft, D. M., & Jacobs, G. (2002). Anthocyanins in vegetative tissues: A proposed unified function in photoprotection. New Phytologist, 155(3), 349–361. https://doi.org/ 10.1046/j.1469-8137.2002.00482.x

Stokes, G. G. (1860). On the Intensity of the Light Reflected from or Transmitted through a Pile of Plates [Publisher: The Royal Society]. Proceedings of the Royal Society of London, 11, 545–556. Retrieved January 6, 2024, from http://www.jstor.org/stable/111970

Sturm, J., Santos, M. J., Schmid, B., & Damm, A. (2022). Satellite data reveal differential responses of Swiss forests to unprecedented 2018 drought. Global Change Biology, 28(9), 2956–2978. https://doi.org/10.1111/gcb.16136

Sun, J., Shi, S., Yang, J., Du, L., Gong, W., Chen, B., & Song, S. (2018). Analyzing the performance of PROSPECT model inversion based on different spectral information for leaf biochemical properties retrieval. ISPRS Journal of Photogrammetry and Remote Sensing, 135, 74–83. https://doi.org/10.1016/j.isprsjprs.2017.11.010

Sun, T., Rao, S., Zhou, X., & Li, L. (2022). Plant carotenoids: Recent advances and future perspectives. Molecular Horticulture, 2(1), 3. https://doi.org/10.1186/s43897-022-00023-2

Tarantola, A. (2005). Inverse problem theory and methods for model parameter estimation [OCLC: ocm56672375]. Society for Industrial; Applied Mathematics.

Tardieu, F., & Simonneau, T. (1998). Variability among species of stomatal control under fluctuating soil water status and evaporative demand: Modelling isohydric and anisohydric behaviours. Journal of Experimental Botany, 49(Special), 419–432. https://doi.org/10.1093/jxb/49.Special Issue.419

Thom, D., Buras, A., Heym, M., Klemmt, H.-J., & Wauer, A. (2023). Varying growth response of Central European tree species to the extraordinary drought period of 2018 – 2020. Agricultural and Forest Meteorology, 338, 109506. https://doi.org/10.1016/j.agrformet.2023.109506

Tognetti, R., Johnson, J., & Michelozzi, M. (1995). The response of European beech (Fagus sylvatica L.) seedlings from two Italian populations to drought and recovery. Trees, 9(6). https://doi.org/10.1007/BF00202499

Verrelst, J., Camps-Valls, G., Muñoz-Marí, J., Rivera, J. P., Veroustraete, F., Clevers, J. G., & Moreno, J. (2015). Optical remote sensing and the retrieval of terrestrial vegetation bio-geophysical properties – A review. ISPRS Journal of Photogrammetry and Remote Sensing, 108, 273–290. https://doi.org/10.1016/j.isprsjprs.2015.05.005

Voichek, Y., & Weigel, D. (2020). Identifying genetic variants underlying phenotypic variation in plants without complete genomes. Nature Genetics, 52(5), 534–540. https://doi.org/10.1038/s41588- 020-0612-7

Walthert, L., Ganthaler, A., Mayr, S., Saurer, M., Waldner, P., Walser, M., Zweifel, R., & Von Arx, G. (2021). From the comfort zone to crown dieback: Sequence of physiological stress thresholds in mature European beech trees across progressive drought. Science of The Total Environment, 753, 141792. https://doi.org/10.1016/j.scitotenv.2020.141792

Wang, F., Israel, D., Ramírez-Valiente, J.-A., Sánchez-Gómez, D., Aranda, I., Aphalo, P. J., & Robson, T. M. (2021). Seedlings from marginal and core populations of European beech (Fagus sylvatica L.) respond differently to imposed drought and shade. Trees, 35(1), 53–67. https://doi.org/10.1007/s00468-020-02011-9

Wang, Z., Townsend, P. A., & Kruger, E. L. (2022). Leaf spectroscopy reveals divergent inter- and intra- species foliar trait covariation and trait–environment relationships across NEON domains. New Phytologist, 235(3), 923–938. https://doi.org/10.1111/nph.18204

Wankmüller, F. J. P., & Carminati, A. (2022). Stomatal regulation prevents plants from critical water potentials during drought: Result of a model linking soil–plant hydraulics to abscisic acid dynamics. Ecohydrology, 15(5), e2386. https://doi.org/10.1002/eco.2386

Wellstein, C., Poschlod, P., Gohlke, A., Chelli, S., Campetella, G., Rosbakh, S., Canullo, R., Kreyling, J., Jentsch, A., & Beierkuhnlein, C. (2017). Effects of extreme drought on specific leaf area of grassland species: A meta-analysis of experimental studies in temperate and sub-Mediterranean systems. Global Change Biology, 23(6), 2473–2481. https://doi.org/10.1111/gcb.13662

Whitman, D., & Ananthakrishnan, T. (Eds.). (2009, January). Phenotypic Plasticity of Insects: Mechanisms and Consequences. Science Publishers. https://doi.org/10.1201/b10201

Wild, J., Kopecký, M., Macek, M., Šanda, M., Jankovec, J., & Haase, T. (2019). Climate at ecologically relevant scales: A new temperature and soil moisture logger for long-term microclimate measurement. Agricultural and Forest Meteorology, 268, 40–47. https://doi.org/10.1016/j.agrformet.2018.12.018

Wright, I. J., Reich, P. B., Westoby, M., Ackerly, D. D., Baruch, Z., Bongers, F., Cavender-Bares, J., Chapin, T., Cornelissen, J. H. C., Diemer, M., Flexas, J., Garnier, E., Groom, P. K., Gulias, J., Hikosaka, K., Lamont, B. B., Lee, T., Lee, W., Lusk, C., ... Villar, R. (2004). The worldwide leaf economics spectrum. Nature, 428(6985), 821–827. https://doi.org/10.1038/nature02403

Wu, X., Yuan, J., Luo, A., Chen, Y., & Fan, Y. (2016). Drought stress and re-watering increase secondary metabolites and enzyme activity in dendrobium moniliforme. Industrial Crops and Products, 94, 385–393. https://doi.org/10.1016/j.indcrop.2016.08.041

Zhang, F., & Zhou, G. (2019). Estimation of vegetation water content using hyperspectral vegetation indices: A comparison of crop water indicators in response to water stress treatments for summer maize. BMC Ecology, 19(1), 18. https://doi.org/10.1186/s12898-019-0233-0

Zhao, F., Guo, Y., Huang, Y., Reddy, K. N., Lee, M. A., Fletcher, R. S., & Thomson, S. J. (2014). Early detection of crop injury from herbicide glyphosate by leaf biochemical parameter inversion. International Journal of Applied Earth Observation and Geoinformation, 31, 78–85. https://doi.org/10.1016/j.jag.2014.03.010